British Pharmacopoeia 2020

Volume I

The British Pharmacopoeia Commission has caused this British Pharmacopoeia 2020 to be prepared under regulation 317(1) of the Human Medicines Regulations 2012 and, in accordance with regulation 317(4), the Ministers have arranged for it to be published. It has been notified in draft to the European Commission in accordance with Directive 98/34/EEC.

The monographs of the Ninth Edition of the European Pharmacopoeia (2016), as amended by Supplements 9.1 to 9.8, published by the Council of Europe are reproduced either in this edition of the British Pharmacopoeia or in the associated edition of the British Pharmacopoeia (Veterinary).

See General Notices

Effective date: 1 January 2020

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Foreword

Patients rightly expect that their medicines work and are acceptably safe and this is underpinned by the assurance of medicines quality. The British Pharmacopoeia (BP), as part of the Medicines and Healthcare Products Regulatory Agency (MHRA), plays an important role in the assurance of medicines quality and supporting innovation through the development of its quality standards. The BP shares the MHRA's commitment to working with our stakeholders, from our partners in industry to international regulatory and pharmacopoeial peers.

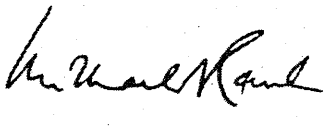
The BP excels at collaboration and this can be seen across its work with international partners, support for European standards via its role as a National Pharmacopoeial Authority to the European Pharmacopoeia Commission and its constructive approach to consultation, with recent examples being the BP's consultation on Dissolution testing and its input into the MHRA's Strategy for Pharmacopoeial Public Quality Standards for Biological Medicines.

Just as the BP's partners and stakeholders are diverse and varied, so too is the breadth of expertise needed to produce standards that ensure patients' expectations of quality medicines are met.

Some of our most important partners are the independent experts that form the BP Commission and its working groups, panels and parties. These experts volunteer from a wide range of backgrounds such as Industry, Academia and the UK's National Health Service (NHS) and bring a diverse array of skills, including pharmaceutics, synthetic chemistry, biology and statistics, to our work.

It is these experts who work to help us overcome challenges and develop the standards and policies that continue to ensure that our work meets the needs of our users, and ultimately, patients. They all share our commitment to the assurance of quality and the role of standards in supporting this.

It is only by working together collectively with our experts, peers and stakeholders that we can realise our shared mission to protect public health.



Professor Sir Michael Rawlins

Chairman
Medicines and Healthcare products Regulatory Agency

Contents

Notices

Monographs of the European Pharmacopoeia are distinguished by a chaplet of stars against the title. The term European Pharmacopoeia, used without qualification, means the ninth edition of the European Pharmacopoeia comprising, unless otherwise stated, the main volume, published in 2016, as amended by any subsequent supplements and revisions.

Patents

In this Pharmacopoeia certain drugs and preparations have been included notwithstanding the existence of actual or potential patent rights. In so far as such substances are protected by Letters Patent their inclusion in this Pharmacopoeia neither conveys, nor implies, licence to manufacture.

Effective dates

New and revised monographs of national origin enter into force on 1 January 2020. The monographs are brought into effect under regulation 320(2) of the Human Medicines Regulations 2012.

Monographs of the European Pharmacopoeia have previously been published by the European Directorate for the Quality of Medicines & HealthCare, in accordance with the Convention on the Elaboration of a European Pharmacopoeia, and have been brought into effect under European Directives 2001/82/EC, 2001/83/EC and 2003/63/EC, as amended, on medicines for human and veterinary use.

Preface

The British Pharmacopoeia Commission has caused this British Pharmacopoeia 2020 to be prepared under regulation 317(1) of the Human Medicines Regulations 2012 and, in accordance with regulation 317(4), the Ministers have arranged for it to be published.

The British Pharmacopoeia 2020 contributes significantly to the quality control of medicinal products for human use. It contains publicly available, legally enforceable standards that provide an authoritative statement of the quality that a product, material or article is expected to meet at any time during its period of use. The Pharmacopoeial standards are designed to complement and assist the licensing and inspection processes and are part of the overall system for safeguarding the health of purchasers and users of medicinal products in the UK.

The British Pharmacopoeia Commission wishes to record its appreciation of the services of all those who have contributed to this important work.

British Pharmacopoeia Commission

The British Pharmacopoeia Commission is appointed, on behalf of the Secretary of State for Health and Social Care, by the Department of Health and Social Care's Appointments Team who are responsible for appointments to all of the Advisory Bodies appointed under the Human Medicines Regulations 2012.

Under the terms of the Human Medicines Regulations 2012, the duties of the British Pharmacopoeia Commission are as follows:

(a) the preparation and publication of any new edition of the British Pharmacopoeia [regulations 317(1) and 317(4)];

(b) the preparation and publication of any compendium containing information relating to substances and articles which are or may be used in the practice of veterinary medicine or veterinary surgery [regulations 317(3)(b) and 317(4)];

(c) the preparation and publication of a list of names to be used as the headings to monographs in the British Pharmacopoeia [regulations 318 (1) and 318(2)];

(d) the preparation of any amendments to the above publications [regulation 317(5)(a)].

Members of the British Pharmacopoeia Commission are appointed for a renewable term of 4 years and, under the requirements laid down by the Office of the Commissioner for Public Appointments, can serve for a maximum of 10 years.

In order to ensure that the British Pharmacopoeia Commission fulfils its duties under the Human Medicines Regulations 2012, the members also have the following duties:

(1) to frame clear and unequivocal technical advice in order to discharge the Commission's responsibilities both for the British Pharmacopoeia, the British Pharmacopoeia (Veterinary) and British Approved Names and as the national pharmacopoeial authority with respect to the European Pharmacopoeia;

(2) to develop clear policies for the preparation and publication of the British Pharmacopoeia and its related publications;

(3) to serve on one or more Expert Advisory Groups or Panels of Experts of the BP Commission, usually in the position of Chair or Vice-Chair;

(4) to approve new and revised text for inclusion in new editions of the British Pharmacopoeia and British Pharmacopoeia (Veterinary);

(5) to approve new and revised names for inclusion in new editions of British Approved Names and its annual supplements.

In addition to the duties listed above, the Chair of the British Pharmacopoeia Commission has the following additional duties:

(1) To chair all scheduled and unscheduled meetings;

(2) To carry out members appraisals in accordance with policies and timelines laid down by the Department of Health and Social Care;

(3) To participate in the process to appoint/re-appoint members of the British Pharmacopoeia Commission.

Expert Advisory Groups, Panels of Experts and Working Parties

Members of Expert Advisory Groups, Panels of Experts and Working Parties are appointed by the British Pharmacopoeia Commission.

The duties of the members are as follows:

(a) to collaborate in the preparation and revision of Monographs, Appendices and Supplementary Chapters for inclusion in the British Pharmacopoeia and British Pharmacopoeia (Veterinary);

(b) to collaborate in the preparation and revision of Monographs, Methods and General Chapters of the European Pharmacopoeia;

(c) to review reports from the British Pharmacopoeia Laboratory in terms of technical content and, where possible, provide independent experimental data to assist in decision making;

(d) to collaborate in the preparation and revision of the list of names to be used as titles for monographs of the British Pharmacopoeia and British Pharmacopoeia (Veterinary).

Members of Expert Advisory Groups, Panels of Experts and Working Parties are usually appointed for a renewable term of 4 years.

Code of Practice

Members of the British Pharmacopoeia Commission and its supporting Expert Advisory Groups, Panels of Experts and Working Parties are required to comply with a Code of Practice on Declaration of Interests in the Pharmaceutical Industry.

British Pharmacopoeia Commission

The Chair and members of the British Pharmacopoeia Commission are required to make a full declaration of interests on appointment and annually thereafter. They must also inform the BP Secretariat promptly of any changes to these interests during the year. These interests are published in the Medicines Advisory Bodies Annual Reports.

Relevant interests must be declared at meetings and are recorded in the Minutes.

Expert Advisory Groups, Panels of Experts and Working Parties

Chairs and members are required to make a full declaration of interests on appointment and to update the Secretariat if these interests change during their term of office. A record is kept of those experts who have declared specific interests, but these are not published.

Relevant interests must be declared at meetings and are recorded in the Minutes.

Membership of the British Pharmacopoeia Commission

The list below includes those members who served during the period 2018 to 2019.

Chair Professor Kevin M G Taylor BPharm PhD FRPharmS
Professor of Clinical Pharmaceutics, UCL School of Pharmacy

Vice-Chair Professor Alastair G Davidson BSc PhD FRPharmS
Visiting Professor of Pharmaceutical Sciences, University of Strathclyde

Professor Matthew Almond BSc DPhil DSc CChem FRSC PFHEA NTF
Professor of Chemistry Education, University of Reading

Dr Jon Beaman BSc PhD MBA CChem MRSC
Head of Development Analytical Group, Pfizer UK

Dr Anna-Maria Brady BSc PhD
Former Head of Biologicals and Administration, Veterinary Medicines Directorate

Dr Graham D Cook BPharm PhD MRPharmS
Senior Director, Process Knowledge/Quality by Design, Pfizer

Dr Andrew Coulson[1] BVetMed MSc MRCVS MA
Member of the Royal College of Veterinary Surgeons; Non-Executive Director, Veterinary Medicines Directorate; former Superintending Inspector, Science & Research Group, The Home Office

Dr Alison Gleadle BSc PhD (*Lay representative*)
Former Group Product Risk Director, Tesco Stores Ltd.

Dr Gerard Lee BPharm PhD FRPharmS MRSC CChem
Former Group Manager, British Pharmacopoeia and Laboratory Services, MHRA; former Secretary & Scientific Director of the British Pharmacopoeia Commission

Mr Robert Lowe BPharm FRPharmS
Director of Pharmacy Quality Assurance Specialist Services, NHS East of England & Northamptonshire

Dr Brian R Matthews[2] BPharm PhD FRPharmS FTOPRA
Consultant on pharmaceutical and medical device regulatory affairs; former Senior Director, EC Registration, Alcon Laboratories

Professor John Miller MSc PhD MRSC CChem
Visiting Professor, Strathclyde Institute of Pharmacy and Biomedical Sciences; former Head of the EDQM Laboratory

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[1] *Deceased.*
[2] *Retired, 31 December 2018.*

Ms Sharon Palser MSc (*Lay Representative*)
Former Director of Development, NHS Plymouth

Professor Monique Simmonds OBE JP BSc PhD FLS FBS FRES FWIF
Deputy Director of Science, Royal Botanic Gardens, Kew

Dr Ronald Torano BSc PhD MRSC CChem
Pharmacopoeial Intelligence and Advisory Specialist; GlaxoSmithKline

Dr Paul Varley BSc PhD
Vice President of Biopharmaceutical Development, Medimmune Limited

Secretary and Scientific Director

Mr James Pound BSc
Group Manager, British Pharmacopoeia and Laboratory Services, MHRA

Membership of Expert Advisory Groups, Panels of Experts and Working Parties

The Commission appointed the following Expert Advisory Groups, Panels of Experts and Working Parties to advise it in carrying out its duties. Membership has changed from time to time; the lists below include all who have served during the period 2018 to 2019.

EXPERT ADVISORY GROUPS

ABS: Antibiotics

R L Horder (*Chair*), G Cook (*Vice-Chair*), G Blake, E Flahive, V Jaitely, W Mann, J Miller, M Pires, J Sumal, I R Williams

BIO: Biological and Biotechnological Products

P Varley (*Chair*), A-M Brady (*Vice-Chair*), L Bisset[1], C. Braxton, C Burns, K Chidwick[1], A Cook[1], J Cook[1], B Cowper, S Gill, E Griffiths, C Jones[1], V Loh, A Kippen, K Nordgren, B Patel, A M Pickett[1], T Pronce, L Randon, I Rees[1], S Schepelmann[1], P Sheppard, P Stickings[1], A H Thomas[2], R Thorpe, L Tsang, M Wadhwa[1], W Zunic

HCM: Herbal and Complementary Medicines

M Simmonds (*Chair*), R Middleton (*Vice-Chair*), L A Anderson, P Anderson, A Booker, C Etheridge, C Leon, B Moore, M Pires, E Reich, M Rowan, A Slater, K Strohfeldt-Venables, J Sumal[1], C Welham, E Williamson, K Zhao
(*Corresponding members* SS Handa, A Krauss, Z-T Wang)

MC1: Medicinal Chemicals

A G Davidson (*Chair*), D Cairns (*Vice-Chair*), S Bale, H Batchelor, J C Berridge, E Bush, A J Caws, D Deutsch, P Fleming, E Gray, W J Lough, D Malpas, S Nolan

MC2: Medicinal Chemicals

G Cook (*Chair*), C T Goddard (*Vice-Chair*), J Birchall, K Bracht, J Cowie, D Edwards, K Foster, E Hook, J Lim, J Miller, P Murray[2], A Ruggiero, M Turgoose, N Wynne
(*Corresponding members* M Brits, W Sherwin)

MC3: Medicinal Chemicals

M Almond (*Chair*), J Beach (*Vice-Chair, from 1 January 2019*), J Beaman, K Foster, C T Goddard, P Hampshire, W K L Pugh, B Rackstraw, R Torano, M Tubby, I R Williams

NOM: Nomenclature

J K Aronson (*Chair*), L Tsang (*Vice-Chair, until 31 December 2018*), M Ahmed, A McFarlane, D Mehta, G P Moss, R Thorpe
(*Corresponding members* R G Balocco Mattavelli, J S Robertson)

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[1] *Specialist member.*
[2] *Deceased.*

PCY: Pharmacy

R L Horder (*Chair*), B R Matthews (*Vice-Chair, until 31 December 2018*), R Lowe (*Vice-Chair, from 1 January 2019*), M Ahmed[1], E Baker, J Beach, D Elder, J Lim[1], J MacDonald, A McFarlane, J F McGuire, T Purewal, K M G Taylor, S Wicks
(*Corresponding member* J Churchill)

ULM: Unlicensed Medicines

M G Lee (*Chair*), V Fenton-May (*Vice-Chair*), A Bosley, S Branch, D Caulfield, M Godber, W Goddard, S Hartley, S Ho, J Rickard, D Kirby, M Santillo, J Ramada-Magalhaes, J Smith, A Sully, P Weir, M Westwood

PANELS OF EXPERTS

BLP: Blood Products

K Chidwick, A R Hubbard, J More, P Varley

CX: Excipients

B R Matthews (*Chair, until 31 December 2018*), C Mroz (*Vice-Chair*), H Batchelor, R Cawthorne, D Deutsch

DNA: Identification Techniques

A Slater (*Chair*), M Carine, I Feavers, J Hawkins, E Mee, E Williamson

(Disbanded, 30 June 2019)

IGC: Inorganic and General Chemicals

C T Goddard (*Chair*), M Almond, S Atherton, S Boland, D Caulfield, P Henrys, G Lay

MIC: Microbiology

V Fenton-May (*Chair*), B Alexander, S Denyer, P Hargreaves, C Iverson, V Jaitely, B R Matthews, J Silva

RAD: Radioactive Materials

I Boros, J Brain, D Graham, G Inwards, R D Pickett, R Smith

VET: Veterinary Medicines

E Williamson (*Chair*), A Coulson[2] (*Vice-Chair*), A Cairns, S Cockbill, D Evans, E Flahive, B Ward

VIP: Veterinary Immunological Products

A M Brady (*Chair*), R Banks, R Cooney, M Johnson, K Redhead, J Salt, C Stirling, R Woodland

WORKING PARTIES

AQbD: Analytical Quality by Design

G Cook (*Chair*), S Brown, M Chatfield, S Ellison, C Gray, M Hanna-Brown, S Jones, P Nethercote, E Razzano
(*Corresponding members* K Barnett, B Harrington, W Sherwin)

BIO-DPS: Documentary and Physical Standards*

P Varley (*Chair*), A-M Brady (*Vice-Chair*), B Cowper, C Burns, N Czeloth, L Duhau, V Ganeva, C E Giartosio, A Ramzan, B Rellahan, M Wild

* *BIO-DPS: Alternative Approaches for Documentary and Physical Standards for Biotechnological Products*

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[1] *Specialist member.*
[2] *Deceased.*

MCS: Microscopy E Williamson (*Chair*), R Arroo, R Fleck, K Helliwell, K MacLellan Gibson
(Disbanded, 31 December 2018)

AD-HOC GROUP

New Analytical Technologies M Almond, J Beaman, G Cook, J Miller, R Torano, M Simmonds

Current British Pharmacopoeia Staff

Secretariat J Pound (*Secretary and Scientific Director*)

A Gibb (*Editor-in-Chief*)

S Young (*Head of Analytical Science*)

H Ashraf, H Corns, P Crowley, L Elanganathan, A Evans, A Gardiner, S Gomersal, G Kemp, C Lenihan, G Li-Ship, S Maddocks, H Makwana, F J Swanson, M-L Wall, M Whaley

NIBSC-based Staff L Gibson, C Gkouva, C Howard, C Lockie-Williams

Administrative F Chughtai, B F Delahunty, J Paine, U Rothna, N Siddika

ISO 9001
FS 27268

Current British Pharmacopoeia Laboratory Staff

R Adams (*Operations Manager*)

D Ballottin, C Balsa, O Bernabe, A Biesenbruch, M Boardman, H Bowden, K Busuttil, A Cepeda, S Choudhury, A Ciesluk, J Couzins, Y El Dabh, S Doyle, S Ganguli, M Goode, R Griffiths, D Holcombe, N Ionescu, P Makhomu, K Meyer de Figueiredo, W Mohammed, G Naar, M Nanasi, A Paul, M Petrova, L Piare, S Planou, R Ravishankar, S Reeves, I Reydellet, D Rutty, M Sciberras, G Searle, C Smart, B Smith, C Thompson, M Threadgold, V Vekereya

ISO 9001
FS 27613

Current Staff of the Publisher of the British Pharmacopoeia

A Prince (*Business Director*)

P Allard (*Service Delivery Manager*)

A Allen (*Director, Parliament and Publishing*)

N Billington, C Cole, A Dampier, C Gaines, N Griffiths, A Hughes, I Ichongiri, N Joisa, J Kharuna, S Page, M Parka, N Pope, P Relfe, J Stoker, V Verma, T Wheeler

ISO 9001
FS 22428

Introduction

British Pharmacopoeia 2020

The British Pharmacopoeia 2020 supersedes the British Pharmacopoeia 2019. It has been prepared by the British Pharmacopoeia Commission, with the collaboration and support of its Expert Advisory Groups, Panels of Experts and Working Parties and contains approximately 4000 monographs for substances, preparations and articles used in the practice of medicine. Some of these monographs are of national origin and have been elaborated or revised under the auspices of the British Pharmacopoeia Commission whilst others (indicated to users by a chaplet of stars) have been elaborated, or revised, under the auspices of the European Pharmacopoeia Commission, supported by its Groups of Experts and Working Parties, and are reproduced from the European Pharmacopoeia. This edition, together with its companion volume, the British Pharmacopoeia (Veterinary) 2020, incorporates all the monographs of the 9th Edition of the European Pharmacopoeia, as amended by Supplements 9.1 to 9.8. Users of the British Pharmacopoeia thereby benefit by finding within this comprehensively indexed compendium all current United Kingdom pharmacopoeial standards for medicines for human use.

The BP 2020 comprises six volumes as follows.

Volumes I and II	Medicinal Substances
Volume III	Formulated Preparations: General Monographs Formulated Preparations: Specific Monographs
Volume IV	Herbal Drugs, Herbal Drug Preparations and Herbal Medicinal Products Materials for use in the Manufacture of Homoeopathic Preparations Blood-related Products Immunological Products Radiopharmaceutical Preparations Surgical Materials
Volume V	Infrared Reference Spectra Appendices Supplementary Chapters Index
Volume VI	British Pharmacopoeia (Veterinary) 2020

Effective Date The effective date for British Pharmacopoeia monographs in this edition is 1 January 2020.

National monographs omitted from this or earlier editions of the British Pharmacopoeia remain effective in accordance with Regulation 252(2)(c) of the Human Medicines Regulations 2012.

Implementation dates regarding European Pharmacopoeia publications are provided in Supplementary Chapter IV B: Dates of Implementation. European Pharmacopoeia monographs are identified by a chaplet of stars alongside the title.

Additions

A list of monographs included for the first time in the British Pharmacopoeia 2020 is given at the end of this introduction. It includes 35 new monographs of national origin and 40 new monographs reproduced from the 9th Edition of the European Pharmacopoeia, as amended by Supplements 9.1 to 9.8.

Pharmacopoeial Public Quality Standards for Biological Medicines

The Medicines and Healthcare products Regulatory Agency (MHRA) has continued to implement its strategy for pharmacopoeial public quality standards for biological medicines as published in 2017[1]. The strategy acknowledges the importance of biological medicines, the value of pharmacopoeial public quality standards and the unique position of the MHRA to lead in this field through its alignment of regulatory, documentary (BP) and physical (NIBSC) standard setting functions.

Part of the published strategy was to investigate alternative approaches to standards for biological medicines. This has led to the establishment of the Alternative Approaches for Documentary and Physical Standards for Biotechnological Products Working Party (WP BIO-DPS) in 2018. WP BIO-DPS aims to better understand the challenges of implementing pharmacopoeial standards and how alternative approaches can respond to these issues whilst continuing to support innovation throughout the product lifecycle. The working party are exploring ideas for new standards concepts, including performance and class based standards, and how to assess the value and potential implementation of such concepts.

The strategy also highlighted the need to investigate and take forward documentary and physical standard setting opportunities for Advanced Therapy Medicinal Products (ATMPs). The MHRA has engaged with groups across the ATMP community to improve its understanding of the challenges and needs faced by the ATMP community and the role of standards in supporting innovation and the assurance of product quality.

Dissolution Consultation

The dissolution consultation response was published in January 2019 and incorporates the feedback from stakeholders and proposals put forward by the Pharmacy Expert Advisory Group and British Pharmacopoeia Commission. The response document outlines a revised policy on dissolution testing. The BP has addressed each of the points raised with a clear action plan that defines how this policy will be implemented.

https://www.pharmacopoeia.com/content/file/Dissolution-Testing-Consultation-Response-Document.pdf

The BP 2020 includes an initial revision to Supplementary Chapter I E (Dissolution Testing of Solid Oral Dosage Forms) and a change to the order of the appendices. These initial changes to the supporting information for dissolution testing in the pharmacopoeia are the first stage of the implementation of this revised policy.

[1] *The strategy and work programme can be found on the following webpage: https://www.gov.uk/government/consultations/strategy-for-pharmacopoeial-public-quality-standards-for-biological-medicines.*

Traditional Herbal Medicines; Homoeopathic Preparations

Two new BP monographs for herbal medicines are included in this edition. They reflect a continued commitment to providing standards for herbal drugs commonly used in the UK and for those known to be used for the preparation of traditional medicines.

Appendix XI V: Deoxyribonucleic Acid (DNA) Based Identification Techniques for Herbal Drugs has been updated to include a worked example of the DNA-based identification of *Tribulus terrestris* Fruit. Supplementary Chapter VII D: DNA Barcoding as a tool for Botanical Identification of Herbal Drugs has also been updated to include the reference sequences for *Tribulus terrestris* Fruit and *Galium aparine* L. (Clivers). Where individual monographs have a reference sequence, a non-mandatory annex has been included in the monograph to inform users of the published reference sequence in Supplementary Chapter VII D.

Unlicensed Medicines

With this new edition, a further 5 monographs for unlicensed formulations have been added. All monographs for such formulations are characterised by a statement that the monograph has been prepared to cover unlicensed formulations. The general and individual monographs are intended to apply to all types of Unlicensed Medicines, that is, those formulations prepared under a Manufacturer's 'Specials' Licence and those prepared extemporaneously under the supervision of a pharmacist.

The General Monograph on Unlicensed Medicines has been amended to reflect that alternative approaches to the BP test for Sterility might be required for sterile preparations that are prepared extemporaneously or in small batches.

Analytical Quality by Design (AQbD)

The British Pharmacopoeia, working with the MHRA and stakeholders, is investigating the application of the Quality by Design concept to analytical methods and the pharmacopoeia. Several AQbD concepts have been practically assessed in conjunction with the British Pharmacopoeia Laboratory, including: risk analysis, Design of Experiments (DoE) and Analytical Target Profiles (ATPs). The Australian Therapeutic Goods Administration have also been a key collaborator in the project.

The MHRA recognises the importance of alignment between regulators, pharmacopoeias and stakeholders in the development of new and innovative policy, such as the application of AQbD principles. Therefore, the outcomes of this work are to be published ahead of the publication of the BP 2020. The MHRA will also publish an accompanying consultation to seek stakeholders' views on how AQbD concepts could be applied in a pharmacopoeial context.

Revisions

A significant number (331, comprising 103 technical revisions and 228 editorial revisions) of national monographs have been amended by means of this edition. Of these monographs, those with major technical revisions are listed at the end of this Introduction. For the benefit of the reader this list indicates the section, or sections, of each monograph which has/have been revised.

The list of revisions appended to this Introduction is as comprehensive as practicable. However, to ensure that the reader uses the current standard, it is essential to refer to the full text of each individual monograph.

For those texts reproduced from the European Pharmacopoeia, the European Directorate for the Quality of Medicines & HealthCare (EDQM) database (see below, under Websites) provides information on revisions of the monographs or other texts on a historical basis, beginning from the 5th Edition of the European Pharmacopoeia.

British Pharmacopoeia Chemical Reference Substances (BPCRS)

The British Pharmacopoeia continues to expand the catalogue of BPCRS which are essential parts of the published monographs. The catalogue currently contains well over 800 items. The British Pharmacopoeia Commission Laboratory continuously strives to improve the percentage of BPCRS in stock and is working towards making the BPCRS to support new monographs for the BP 2020 and future editions available for our users at the same time as the publication becomes available and ahead of the implementation date.

Title Changes

108 monograph titles have been amended in this edition. In accordance with the British Pharmacopoeia Commission decision to amend monograph titles that include a split in the standard term, changes have been made to affected monograph titles in this edition and are included within the appended list.

Omissions

73 monographs have been omitted from the British Pharmacopoeia 2020. The list of omissions is appended at the end of this Introduction.

Infrared Reference Spectra

As with the previous edition, the reference spectra are placed in alphabetical order within this edition. Six new spectra have been added to the collection.

Appendices

Three new Appendices to harmonise with the European Pharmacopoeia were first published in the British Pharmacopoeia 2019 electronic updates. These have been consolidated in the new edition as follows:
Appendix II M. Direct Amperometric and Pulsed Electrochemical Detection (Ph. Eur. Method 2.2.63);
Appendix XI O. Foam Index (Ph. Eur. Method 2.8.24);
Appendix XVI H. Microbiological Examination of Live Biotherapeutic Products (Ph. Eur. Methods 2.6.36 and 2.6.38).

European Pharmacopoeia

Co-operation Agreement

As a consequence of the Co-operation Agreement with the EDQM of the Council of Europe, the British Pharmacopoeia Commission is pleased to note the integration of European Pharmacopoeia texts for the British Pharmacopoeia 2019 in-year online updates and for this edition of the British Pharmacopoeia.

In accordance with previous practice, all monographs and requirements of the European Pharmacopoeia are reproduced in this edition of the British Pharmacopoeia or, where appropriate, within its companion edition, the British Pharmacopoeia (Veterinary) 2020.

Where a monograph has been reproduced from the European Pharmacopoeia, this is signified by the presence of a chaplet of stars alongside its title. Additionally, reference to the European Pharmacopoeia monograph number is included immediately below the title in italics in the form '*Ph. Eur. monograph xxxx*'. Where the title in the British

Pharmacopoeia is different from that in the European Pharmacopoeia, an approved synonym has been created (see Appendix XXI B) and the European Pharmacopoeia title is included before the monograph number. The entire European Pharmacopoeia text is delineated by two horizontal lines bearing the symbol '*Ph. Eur.*'.

The European Pharmacopoeia texts have been reproduced in their entirety but, where deemed appropriate, additional statements of relevance to UK usage have been added (e.g. action and use statement, a list of British Pharmacopoeia preparations). It should be noted, however, that in the event of doubt of interpretation in any text of the European Pharmacopoeia, the text published in English under the direction of the Council of Europe should be consulted.

Correspondence between the general methods of the European Pharmacopoeia and the appendices of the British Pharmacopoeia is indicated in each appendix and by inclusion of a list at the beginning of the appendices section.

Pharmacopoeial Requirements

It should be noted that any article intended for medicinal use which is described by a name at the head of a monograph in the current edition of the Pharmacopoeia must comply with that monograph '*whether or not it is referred to as BP*'.

It is also important to note that no requirement of the Pharmacopoeia can be taken in isolation. A valid interpretation of any particular requirement depends upon it being read in the context of (i) the monograph as a whole, (ii) the specified method of analysis, (iii) the relevant General Notices and, where appropriate, (iv) the relevant General Monograph(s). Familiarity with the General Notices of the Pharmacopoeia will facilitate the correct application of the requirements. Additional guidance and information on the basis of pharmacopoeial requirements is provided in Supplementary Chapter I. This non-mandatory text describes the general underlying philosophy and current approaches to particular aspects of pharmacopoeial control.

Code of Practice

Members of the British Pharmacopoeia Commission and its supporting Expert Advisory Groups, Panels of Experts and Working Parties are required to comply with a Code of Practice on Declaration of Interests in the pharmaceutical industry. Details of the Code are published on the website (pharmacopoeia.com).

Websites

British Pharmacopoeia Website

The British Pharmacopoeia website, pharmacopoeia.com, contains information relating to the British Pharmacopoeia. It allows subscribers to access the British Pharmacopoeia 2020 and British Pharmacopoeia (Veterinary) 2020 online and British Approved Names publications. All users are also able to view and purchase BPCRS products through the website.

Chromatograms for information to support new monographs published in the British Pharmacopoeia 2020 have been added to the example test results gallery to aid users of British Pharmacopoeia monographs. This service will increase year-on-year to allow users to examine chromatograms

obtained during the practical evaluation of new monographs by the British Pharmacopoeia Commission Laboratory.

A regular review schedule for draft texts is included on the website, with draft new and revised monographs being posted at the start of each quarter and available for comment for a period of three months thereafter. This free service allows greater visibility of the BP's work programme and enables stakeholder contributions to monograph development.

Subscribers to the BP online will find that draft texts and example test results are also linked with relevant texts and directly accessible from the BP online content. Additionally, BPCRS products are also linked with relevant BP monographs and subscribers to the BP online will be able to purchase these directly from the BP online. BPCRS customers are able to make purchases through invoice or credit card orders.

An email subscription feature allows users to keep abreast with BP news. Additionally, users can subscribe to receive BPCRS updates, which are now posted monthly.

Access to previous editions of the BP is available as a BP archive product for purchase by new and existing BP online subscribers. The content of the archive starts from the BP 2014 onwards and grows year-on-year as superseded editions are added to the archive.

The British Pharmacopoeia is committed to continuously improving the user experience of our products and services. Since 2019 the publication process has been upgraded. User benefits include searchable tables and equations, better linking functionality and higher formatting consistency. Additionally, the reagent requirements have been harmonised with the European Pharmacopoeia where possible, making it easier for users to ensure compliance.

A policy of continuous improvement allows the BP website to keep up to date and respond to users. Customers are therefore invited to provide the Secretariat with feedback on their experience. Independent user researchers have developed a programme of research to further understand our users needs in order to enhance BP products. This has informed the development of a short guide on how to use the BP with further developments to be released over the year.

European Pharmacopoeia Websites

For those texts reproduced from the European Pharmacopoeia, the EDQM website provides access to a database (the Knowledge database: https://extranet.edqm.eu/publications/recherches_sw.shtml) containing information of various sorts related to monographs and intended to facilitate their proper use. Information is provided on chromatographic columns used in monograph development, suppliers of reagents and equipment that may be difficult to find for some users, the status of monographs (in development, adopted, published, under revision), revisions of the monographs on a historical basis, beginning from the 5th Edition of the European Pharmacopoeia as well as other useful information.

The European Pharmacopoeia Forum, Pharmeuropa, is published quarterly as an aid for the elaboration of monographs and as a vehicle for information

on pharmacopoeial and related matters. Pharmeuropa is available as a free online publication: https://pharmeuropa.edqm.eu/home

International Collaboration

Therapeutic Goods Administration, Australia The British Pharmacopoeia Commission is pleased to continue its long-standing co-operation with the Australian Department of Health Therapeutic Goods Administration (TGA). The TGA continues to provide advice to British Pharmacopoeia Commission Expert Advisory Groups, to participate in inter-laboratory evaluation of British Pharmacopoeia monographs and to review data jointly. This collaboration has enabled the production of robust, high quality monographs for users.

Chinese Pharmacopoeia The British Pharmacopoeia Commission is pleased to continue its collaboration with the Chinese Pharmacopoeia on the development of monographs and staff exchanges to support mutually agreed projects.

The Croatian Agency for Medicinal Products and Medical Devices ("HALMED") The Cooperation Agreement between the Medicines and Healthcare products Regulatory Agency and HALMED provides a licence for the use of information in the British Pharmacopoeia on unlicensed medicines.

The Japanese Pharmacopoeia The British Pharmacopoeia has collaborated with the Japanese Pharmacopeia for the development of informally harmonised standards and knowledge sharing in a number of areas of mutual interest.

State Pharmacopoeia of the Republic of Kazakhstan Following the signing of a Collaboration Agreement in April 2016, the Medicines and Healthcare products Regulatory Agency has granted the Committee on Surveillance of Medical and Pharmaceutical Activities of the Ministry of Health of the Republic of Kazakhstan a licence to continue to use relevant contents of the British Pharmacopoeia in the State Pharmacopoeia of the Republic of Kazakhstan.

State Pharmacopoeia of Ukraine Following the signing of a Collaboration Agreement in 2016, the Medicines and Healthcare products Regulatory Agency has continued to grant the Ukrainian Scientific Pharmacopoeial Center for Quality of Medicines a licence to use relevant contents of the British Pharmacopoeia in the State Pharmacopoeia of Ukraine.

United States Pharmacopeia Close collaboration with the United States Pharmacopeia continues, building on the success of the programme of work to jointly develop and revise drug product monographs, with joint participation in conferences and symposia and knowledge sharing in areas of mutual interest.

World Health Organization The collaboration agreement between the British Pharmacopoeia and the International Pharmacopoeia continues to support the work of the WHO, including collaboration and information exchange, contribution to the International Meeting of World Pharmacopoeias, and the international non-proprietary names programme.

Forward Look

In-year Updates The British Pharmacopoeia 2020 online updates will be published on the website, pharmacopoeia.com, to enable users to keep up to date with monographs published in the European Pharmacopoeia. These updates will be integrated annually with the publication of the main edition of the British Pharmacopoeia.

Acknowledgements

The British Pharmacopoeia Commission is greatly indebted to the members of its Expert Advisory Groups, Panels of Experts and Working Parties for their dedicated enthusiasm and assistance in the preparation of this edition. In particular Dr Brian Matthews who, after 9 years of providing support and advice, has ended his term on the British Pharmacopoeia Commission. We would also like to acknowledge the contributions of Dr Andrew Coulson, Mr Peter Murray and Dr Adrian Thomas who sadly passed away during the last year.

Close co-operation has continued with many organisations at home and overseas. These include the Medicines and Healthcare products Regulatory Agency, the Veterinary Medicines Directorate, the Royal Pharmaceutical Society, the Association of the British Pharmaceutical Industry, the British Association of Homoeopathic Manufacturers, the United Kingdom Herbal Forum, The China Food and Drug Administration, the Chinese Pharmacopoeia Commission, the European Pharmacopoeia Commission and the European Directorate for the Quality of Medicines & HealthCare, the Therapeutic Goods Administration (Australia), the Health Products and Food Branch of Health Canada, the United States Pharmacopeia, the Quality Assurance and Safety: Medicines Department of the World Health Organization (WHO), the Health Sciences Authority of Singapore and the Royal Botanic Gardens, Kew.

The British Pharmacopoeia Commission wishes to thank the European Directorate for the Quality of Medicines & HealthCare for their support and assistance in the reproduction of the European Pharmacopoeia texts and monographs. The British Pharmacopoeia Commission acknowledges the importance of the work of the European Pharmacopoeia Commission and its Groups of Experts and Working Parties. The British Pharmacopoeia Commission is also grateful for the generous contribution by the UK experts to the work of the Groups of Experts and Working Parties of the European Pharmacopoeia Commission.

The British Pharmacopoeia Commission acknowledges the contribution of Professor Frederick A Senese, Department of Chemistry, Frostburg State University, USA, for his kind permission to reproduce the indicator colour chart.

The British Pharmacopoeia Commission also acknowledges and appreciates the advice of the publishing team at The Stationery Office, in particular, Mr Paul Allard, Mr Andrew Allen, Ms Nichola Billington, Mr Chris Cole, Mr Nagaraja Joisa, Mr Steve Page, Mr Paul Relfe and Mr Ian Webb, in the production of this edition.

The British Pharmacopoeia Commission acknowledges the contribution of two members of the Civil Service Fast Stream programme, Mr Toby Gladwin and Ms Naomi Clothier who spent six and twelve months, respectively, working with the British Pharmacopoeia Secretariat.

Additions The following monographs of the British Pharmacopoeia 2020 were not included in the British Pharmacopoeia 2019.

Medicinal and Pharmaceutical Substances
Atazanavir Sulfate[1]
Boldine[1]
Dexamfetamine Sulfate[1]
Everolimus[1]
Fingolimod Hydrochloride[1]
Concentrated Solutions for Haemofiltration and Haemodiafiltration[1]
Levofloxacin Hemihydrate[1]
Infliximab Concentrated Solution[1]
Magnesium Aluminometasilicate[1]
Mebeverine Hydrochloride[1]
Nilotinib Hydrochloride Monohydrate[1]
Phenoxymethylpenicillin (Benzathine) Tetrahydrate[1]
Phytomenadione[1]
Podophyllotoxin[1]
Regorafenib Monohydrate[1]
Rotigotine[1]
Sulfobutylbetadex Sodium[1]
Terpin Monohydrate[1]
Zoledronic Acid Monohydrate[1]

Formulated Preparations: General Monographs
Live Biotherapeutic Products for Human Use[1]

Formulated Preparations: Specific Monographs
Amitriptyline Oral Solution
Azithromycin Eye Drops
Cabergoline Tablets
Calcium Carbonate Oral Suspension
Capecitabine Tablets
Celecoxib Capsules
Cilastatin and Imipenem for Infusion
Colistimethate Inhalation Powder, Hard Capsule
Deferiprone Oral Solution[1]
Deferiprone Tablets[1]
Diltiazem Oral Suspension
Ferric Chloride Injection
Filgrastim Injection[1]
Galantamine Prolonged-release Capsules
Galantamine Oral Solution
Galantamine Tablets
Ibuprofen Orodispersible Tablets
Lacosamide Infusion[1]
Lacosamide Oral Solution[1]
Lacosamide Tablets[1]
Leflunomide Tablets
Letrozole Tablets
Metoprolol Oral Suspension
Minocycline Capsules

[1] *denotes a monograph of the European Pharmacopoeia*

Moxonidine Tablets
Risedronate Sodium Tablets
Ritonavir Oral Solution
Ritonavir Tablets
Rotigotine Transdermal Patches
Salmeterol Inhalation Powder, pre-metered
Salmeterol Pressurised Inhalation, Suspension
Temozolomide Capsules
Temozolomide for Injection
Tobramycin Eye Drops
Tobramycin and Dexamethasone Eye Drops
Tobramycin Inhalation Powder, Hard Capsule

Herbal Drugs, Herbal Drug Preparations and Herbal Medicinal Products
Achyranthes Bidendata Root[1]
Clivers
Corydalis Rhizome[1]
Gastrodia Elata Rhizome[1]
Ligusticum Root and Rhizome[1]
Dwarf Lilyturf Tuber[1]
Indian Sandalwood Oil
Sophora Flavescens Root[1]
Typhae Pollen[1]

Materials for use in the Manufacture of Homoeopathic Preparations
Digitalis for Homoeopathic Preparations[1]

Immunological Products
Meningococcal Group A, C, W135 and Y Conjugate Vaccine[1]

Radiopharmaceutical Preparations
Flurodopa (^{18}F) (Prepared by Nucleophilic Substitution) Injection[1]
Yttrium (^{90}Y) Chloride Solution for Radiolabelling[1]

Omissions

The following monographs of the British Pharmacopoeia 2019 are not included in the British Pharmacopoeia 2020.

Medicinal and Pharmaceutical Substances
Aloxiprin
Azapropazone
Benorilate
Carbaryl
Chlormethine Hydrochloride
Chlorpropamide[2]
Debrisoquine Sulfate
Desoxycortone Acetate[3]
Dextropropoxyphene Napsilate
Dihydroergotamine Tartrate[4]
Diloxanide Furoate
Dipipanone Hydrochloride

[1] *denotes a monograph of the European Pharmacopoeia*
[2] *Monograph suppressed by the European Pharmacopoeia Commission on 1st April 2019*
[3] *Monograph suppressed by the European Pharmacopoeia Commission on 1st January 2019.*
[4] *Monograph suppressed by the European Pharmacopoeia Commission on 1st July 2019.*

Emetine Hydrochloride Pentahydrate[1]
Gliquidone
Isometheptene Mucate
Methoxamine Hydrochloride
Mexenone
Nicotinyl Alcohol Tartrate
Oxprenolol Hydrochloride[2]
Pentagastrin
Phenindamine Tartrate
Phytomenadione[1,3]
Poldine Metilsulfate
Polythiazide
Quinidine Bisulfate
Ritodrine Hydrochloride
Tolazamide
Highly Purified Water[2]

Formulated Preparations: Specific Monographs
Aloxiprin Tablets
Azapropazone Capsules
Benorilate Oral Suspension
Brompheniramine Tablets
Carbaryl Lotion
Chlormethine Injection
Chloroquine Sulfate Injection
Chlorpropamide Tablets
Cyclopenthiazide Tablets
Dextromoramide Tablets
Dextropropoxyphene Capsules
Digitoxin Tablets
Diloxanide Tablets
Dipipanone and Cyclizine Tablets
Estradiol Injection
Estropipate Tablets
Etodolac Tablets
Fenbufen Capsules
Flucytosine Tablets
Gliquidone Tablets
Guanethidine Tablets
Hydroflumethiazide Tablets
Idoxuridine Eye Drops
Interferon Alfa-2b Injection
Levodopa Capsules
Levodopa Tablets
Mepyramine Tablets
Methyldopate Injection
Nandrolone Decanoate Injection
Nicotinyl Alcohol Tablets

[1] *Monograph suppressed by the European Pharmacopoeia Commission on 1st January 2019.*
[2] *Monograph suppressed by the European Pharmacopoeia Commission on 1st April 2019*
[3] *The monograph for Phytomenadione was replaced by the monograph for Racemic Phytomenadione in Supplement 9.6 of the 9th Edition of the European Pharmacopoeia, but the title "Phytomenadione" has been retained in the British Pharmacopoeia.*

Emetine Hydrochloride Pentahydrate[1]
Gliquidone
Isometheptene Mucate
Methoxamine Hydrochloride
Mexenone
Nicotinyl Alcohol Tartrate
Oxprenolol Hydrochloride[2]
Pentagastrin
Phenindamine Tartrate
Phytomenadione[1,3]
Poldine Metilsulfate
Polythiazide
Quinidine Bisulfate
Ritodrine Hydrochloride
Tolazamide
Highly Purified Water[2]

Formulated Preparations: Specific Monographs
Aloxiprin Tablets
Azapropazone Capsules
Benorilate Oral Suspension
Brompheniramine Tablets
Carbaryl Lotion
Chlormethine Injection
Chloroquine Sulfate Injection
Chlorpropamide Tablets
Cyclopenthiazide Tablets
Dextromoramide Tablets
Dextropropoxyphene Capsules
Digitoxin Tablets
Diloxanide Tablets
Dipipanone and Cyclizine Tablets
Estradiol Injection
Estropipate Tablets
Etodolac Tablets
Fenbufen Capsules
Flucytosine Tablets
Gliquidone Tablets
Guanethidine Tablets
Hydroflumethiazide Tablets
Idoxuridine Eye Drops
Interferon Alfa-2b Injection
Levodopa Capsules
Levodopa Tablets
Mepyramine Tablets
Methyldopate Injection
Nandrolone Decanoate Injection
Nicotinyl Alcohol Tablets

[1] *Monograph suppressed by the European Pharmacopoeia Commission on 1st January 2019.*
[2] *Monograph suppressed by the European Pharmacopoeia Commission on 1st April 2019*
[3] *The monograph for Phytomenadione was replaced by the monograph for Racemic Phytomenadione in Supplement 9.6 of the 9th Edition of the European Pharmacopoeia, but the title "Phytomenadione" has been retained in the British Pharmacopoeia.*

Amitriptyline Tablets	*Identification; Dissolution; Related substances;Production*
Benzatropine Injection	*Production*
Benzatropine Tablets	*Production*
Benzylpenicillin Injection	*Requirements for ready-to-use solution – deleted; Benzylpenicillin for Injection – Identification; Related substances; Assay*
Bisacodyl Gastro-resistant Tablets	*Title change; Dissolution*
Bisoprolol Tablets	*Content of bisoprolol fumarate; Identification test A; Dissolution; Related substances; Assay*
Bromocriptine Capsules	*Production*
Bromocriptine Tablets	*Production*
Caffeine Citrate Injection	*Related substances; Assay*
Caffeine Citrate Oral Solution	*Related substances; Assay*
Calcium Chloride Injection	*Assay*
Candesartan Tablets	*Related substances*
Carbimazole Tablets	*Related substances*
Ceftriaxone Injection	*Assay*
Clindamycin Capsules	*Identification test A; Dissolution*
Clobazam Oral Suspension	*Dissolution; Related substances*
Co-amoxiclav Injection	*Related substances*
Co-beneldopa Capsules	*Related substances test A*
Co-beneldopa Dispersible Tablets	*Title change; Related substances test A*
Colchicine Tablets	*Related substances*
Dalteparin Sodium Injection	*Identification test A; Related substances*
Desferrioxamine Injection	*Production*
Dobutamine Infusion	*Related substances*
Enoxaparin Sodium Injection	*Identification test A; Related substances*
Ferrous Fumarate Capsules	*Dissolution*
Fluvoxamine Tablets	*Related substances*
Griseofulvin Tablets	*Identification; Related substances; Assay*
Heparin Injection	*Related substances*
Hydrochlorothiazide Tablets	*Related substances*
Hydroxycarbamide Capsules	*Hydroxylamine*
Hyoscine Butylbromide Tablets	*Dissolution; Related substances*
Ibuprofen Prolonged-release Capsules	*Title change; Related substances*
Ibuprofen Gel	*Identification; Acidity or alkalinity; Related substances; Assay*
Ibuprofen Tablets	*Dissolution; Related substances*
Ibuprofen Prolonged-release Tablets	*Title change; Related substances*
Interferon Beta-1a Injection	*Content of interferon beta-1a; Identification test C; Oxidised forms; Dimers and related substances of higher molecular weight (Method A)*
Ipratropium Pressurised Inhalation, Solution	*Title change; Content of ipratropium bromide; Identification tests A and B; Uniformity of delivered dose; Impurity A; Related substances; Assay*
Ketamine Nasal Spray	*Identification test C; Related substances; Assay*
Ketoprofen Capsules	*Related substances*

Ketoprofen Gel	*Related substances; Impurities*
Loprazolam Tablets	*Production*
Lorazepam Injection	*Related substances*
Lorazepam Tablets	*Related substances*
Minocycline Prolonged-release Capsules	*Title change; Content of minocycline; Identification; Related substances; Assay*
Minocycline Tablets	*Content of minocycline; Identification; Dissolution; Related substances; Assay*
Olmesartan Tablets	*Assay*
Pantoprazole Gastro-resistant Tablets	*Dissolution; Assay*
Phenobarbital Injection	*Related substances*
Phenoxymethylpenicillin Oral Solution	*Related substances; Assay*
Phenoxymethylpenicillin Tablets	*Related substances; Assay*
Phentolamine Injection	*Production*
Phenytoin Capsules	*Production; Content of phenytoin sodium; Identification; Related substances; Assay; Impurities*
Phenytoin Injection	*Identification; Benzil and benzophenone (deleted); Ethanol (deleted); Propylene Glycol (deleted); Ethanol and propylene glycol (added); Related substances; Assay; Impurities*
Phenytoin Oral Suspension	*Content of phenytoin; Identification; Benzil and benzophenone (deleted); Related substances; Assay; Impurities*
Phenytoin Tablets	*Production; Identification; Related substances; Assay; Impurities*
Prochlorperazine Injection	*Production*
Prochlorperazine Oral Solution	*Production*
Progesterone Injection	*Related substances; Assay*
Pyrimethamine Tablets	*Content of pyrimethamine; Dissolution; Related substances; Impurities*
Ranitidine Oral Solution	*Related substances*
Salbutamol Pressurised Inhalation, Suspension	*Title change; Identification; Uniformity of delivered dose; Related substances; Assay; Impurities*
Sildenafil Orodispersible Films	*Related substances*
Sildenafil Injection	*Related substances*
Sildenafil Powder for Oral Suspension	*Related substances*
Sildenafil Tablets	*Related substances; Assay*
Sildenafil Chewable Tablets	*Related substances*
Sildenafil Orodispersible Tablets	*Related substances*
Silver Nitrate Sterile Solution	*Title change; Definition; Italic opening statement; Sterility; Labelling (deleted)*
Simvastatin Oral Suspension	*Related substances*
Simvastatin Tablets	*Related substances*
Compound Sodium Lactate Infusion	*Identification test C*
Sotalol Injection	*Identification test B*
Sumatriptan Injection	*Related substances*
Sumatriptan Nasal Spray	*Related substances*
Tamoxifen Tablets	*Identification; Related substances; Assay*
Tenoxicam Injection	*Impurities*

Tenoxicam Tablets	*Impurities*
Tinzaparin Sodium Injection	*Identification test A; Related substances*
Triamcinolone Acetonide Injection	*Identification test B; Related substances; Assay*
Triamcinolone Oromucosal Paste	*Related substances; Assay*
Zuclopenthixol Tablets	*Related substances; Assay*

Herbal Drugs, Herbal Drug Preparations and Herbal Medicinal Products

Anethum Graveolens Sowa Fruit	*Apiole; DNA Reference sequence*
Holy Basil Leaf	*DNA Reference sequence*
Glehnia Littoralis Root	*DNA Reference sequence*
Nutmeg	*Foreign matter; DNA Reference sequence*
Phellodendron Amurense Bark	*DNA Reference sequence*
Phellodendron Chinense Bark	*DNA Reference sequence*
Tribulus Terrestris Fruit	*DNA Reference sequence*

In addition to the changes listed above, the following changes have also been made to BP monographs in this edition.

The opening statements in monographs for unlicensed medicines have been amended to reflect that such monographs have been developed to cover unlicensed formulations. This will avoid any confusion should such formulations become licensed in the future.

The titles of a large number of monographs have been updated to reflect the decision of the British Pharmacopoeia Commission to avoid including split standard terms in the title.

Changes in Title

The following list gives the alterations in the titles of monographs of the British Pharmacopoeia 2019 that have been retained in the British Pharmacopoeia 2020.

BRITISH PHARMACOPOEIA 2019	BRITISH PHARMACOPOEIA 2020
Medicinal and Pharmaceutical Substances	
Spray-dried Acacia	Acacia, Dried Dispersion
Alfentanil Hydrochloride	Alfentanil Hydrochloride Hydrate
Benzathine Benzylpenicillin	Benzathine Benzylpenicillin Tetrahydrate
Calcifediol	Calcifediol Monohydrate
Calcium Folinate	Calcium Folinate Hydrate
Calcium Levofolinate Pentahydrate	Calcium Levofolinate Hydrate
Dipotassium Clorazepate	Dipotassium Clorazepate Monohydrate
Lidocaine Hydrochloride	Lidocaine Hydrochloride Monohydrate
Methylthioninium Chloride	Methylthioninium Chloride Hydrate
Procaine Benzylpenicillin	Benzylpenicillin (Procaine) Monohydrate
Hydrated Valaciclovir Hydrochloride	Valaciclovir Hydrochloride Hydrate

Formulated Preparations: Specific Monographs

Gastro-resistant Acamprosate Tablets	Acamprosate Gastro-resistant Tablets
Dispersible Aciclovir Tablets	Aciclovir Dispersible Tablets
Prolonged-release Alfuzosin Tablets	Alfuzosin Prolonged-release Tablets
Chewable Aluminium Hydroxide Tablets	Aluminium Hydroxide Chewable Tablets
Prolonged-release Aminophylline Tablets	Aminophylline Prolonged-release Tablets
Sterile Arginine Hydrochloride Concentrate	Arginine Hydrochoride Sterile Concentrate
Chewable Ascorbic Acid Tablets	Ascorbic Acid Chewable Tablets
Dispersible Aspirin Tablets	Aspirin Dispersible Tablets
Effervescent Soluble Aspirin Tablets	Aspirin Effervescent Soluble Tablets
Prolonged-release Bezafibrate Tablets	Bezafibrate Prolonged-release Tablets
Gastro-resistant Bisacodyl Tablets	Bisacodyl Gastro-resistant Tablets
Bumetanide and Prolonged-release Potassium Tablets	Bumetanide and Potassium Prolonged-release Tablets
Chewable Calcium and Colecalciferol Tablets	Calcium and Colecalciferol Chewable Tablets
Chewable Calcium and Ergocalciferol Tablets	Calcium and Ergocalciferol Chewable Tablets
Chewable Calcium Carbonate Tablets	Calcium Carbonate Chewable Tablets
Chewable Calcium Carbonate and Heavy Magnesium Carbonate Tablets	Calcium Carbonate and Heavy Magnesium Carbonate Chewable Tablets
Chewable Calcium Gluconate Tablets	Calcium Gluconate Chewable Tablets
Effervescent Calcium Gluconate Tablets	Calcium Gluconate Effervescent Tablets
Chewable Carbamazepine Tablets	Carbamazepine Chewable Tablets
Prolonged-release Carbamazepine Tablets	Carbamazepine Prolonged-release Tablets
Prolonged-release Cefaclor Tablets	Cefaclor Prolonged-release Tablets
Prolonged-release Clarithromycin Tablets	Clarithromycin Prolonged-release Tablets
Prolonged-release Clomipramine Tablets	Clomipramine Prolonged-release Tablets
Dispersible Co-amoxiclav Tablets	Co-amoxiclav Dispersible Tablets
Prolonged-release Co-beneldopa Capsules	Co-beneldopa Prolonged-release Capsules
Dispersible Co-beneldopa Tablets	Co-beneldopa Dispersible Tablets
Effervescent Co-codamol Tablets	Co-codamol Effervescent Tablets
Dispersible Co-codaprin Tablets	Co-codaprin Dispersible Tablets
Dispersible Co-trimoxazole Tablets	Co-trimoxazole Dispersible Tablets
Prolonged-release Diclofenac Capsules	Diclofenac Prolonged-release Capsules
Gastro-resistant Diclofenac Tablets	Diclofenac Gastro-resistant Tablets
Prolonged-release Diclofenac Tablets	Diclofenac Prolonged-release Tablets
Prolonged-release Diltiazem Tablets	Diltiazem Prolonged-release Tablets
Prolonged-release Dipyridamole Capsules	Dipyridamole Prolonged-release Capsules

Gastro-resistant Erythromycin Capsules	Erythromycin Gastro-resistant Capsules
Gastro-resistant Erythromycin Tablets	Erythromycin Gastro-resistant Tablets
Prolonged-release Felodipine Tablets	Felodipine Prolonged-release Tablets
Prolonged-release Ferrous Sulfate Tablets	Ferrous Sulfate Prolonged-release Tablets
Prolonged-release Fluvastatin Tablets	Fluvastatin Prolonged-release Tablets
Prolonged-release Ibuprofen Capsules	Ibuprofen Prolonged-release Capsules
Prolonged-release Ibuprofen Tablets	Ibuprofen Prolonged-release Tablets
Prolonged-release Indapamide Tablets	Indapamide Prolonged-release Tablets
Ipratropium Pressurised Inhalation	Ipratropium Pressurised Inhalation, Solution
Prolonged-release Isosorbide Mononitrate Capsules	Isosorbide Mononitrate Prolonged-release Capsules
Prolonged-release Isosorbide Mononitrate Tablets	Isosorbide Mononitrate Prolonged-release Tablets
Dispersible Lamotrigine Tablets	Lamotrigine Dispersible Tablets
Gastro-resistant Lansoprazole Capsules	Lansoprazole Gastro-resistant Capsules
Gastro-resistant Lansoprazole Tablets	Lansoprazole Gastro-resistant Tablets
Prolonged-release Lithium Carbonate Tablets	Lithium Carbonate Prolonged-release Tablets
Orodispersible Loperamide Tablets	Loperamide Orodispersible Tablets
Chewable Magnesium Glycerophosphate Tablets	Magnesium Glycerophosphate Chewable Tablets
Chewable Compound Magnesium Trisilicate Tablets	Compound Magnesium Trisilicate Chewable Tablets
Prolonged-release Metformin Tablets	Metformin Prolonged-release Tablets
Prolonged-release Metoprolol Tartrate Tablets	Metoprolol Tartrate Prolonged-release Tablets
Prolonged-release Minocycline Capsules	Minocycline Prolonged-release Capsules
Orodispersible Mirtazapine Tablets	Mirtazapine Orodispersible Tablets
Chewable Montelukast Tablets	Montelukast Chewable Tablets
Prolonged-release Morphine Capsules	Morphine Prolonged-release Capsules
Prolonged-release Morphine Tablets	Morphine Prolonged-release Tablets
Gastro-resistant Naproxen Tablets	Naproxen Gastro-resistant Tablets
Prolonged-release Nifedipine Capsules	Nifedipine Prolonged-release Capsules
Prolonged-release Nifedipine Tablets	Nifedipine Prolonged-release Tablets
Orodispersible Olanzapine Tablets	Olanzapine Orodispersible Tablets
Prolonged-release Oxybutynin Tablets	Oxybutynin Prolonged-release Tablets
Gastro-resistant Pancreatin Tablets	Pancreatin Gastro-resistant Tablets
Soluble Paracetamol and Caffeine Tablets	Paracetamol and Caffeine Soluble Tablets
Dispersible Paracetamol Tablets	Paracetamol Dispersible Tablets
Effervescent Paracetamol Tablets	Paracetamol Effervescent Tablets
Soluble Paracetamol Tablets	Paracetamol Soluble Tablets
Chewable Piperazine Phosphate Tablets	Piperazine Phosphate Chewable Tablets
Effervescent Potassium Chloride Tablets	Potassium Chloride Effervescent Tablets

Prolonged-release Potassium Chloride Tablets	Potassium Chloride Prolonged-release Tablets
Prolonged-release Pramipexole Tablets	Pramipexole Prolonged-release Tablets
Prolonged-release Propranolol Capsules	Propranolol Prolonged-release Capsules
Effervescent Ranitidine Tablets	Ranitidine Effervescent Tablets
Prolonged-release Salbutamol Capsules	Salbutamol Prolonged-release Capsules
Salbutamol Pressurised Inhalation	Salbutamol Pressurised Inhalation, Suspension
Sterile Sodium Acetate Concentrate	Sodium Acetate Sterile Concentrate
Sterile Sodium Benzoate Concentrate	Sodium Benzoate Sterile Concentrate
Soluble Sodium Chloride Tablets	Sodium Chloride Soluble Tablets
Prolonged-release Sodium Valproate Capsules	Sodium Valproate Prolonged-release Capsules
Prolonged-release Sodium Valproate Tablets	Sodium Valproate Prolonged-release Tablets
Sterile Silver Nitrate Solution	Silver Nitrate Sterile Solution
Gastro-resistant Sulfasalazine Tablets	Sulfasalazine Gastro-resistant Tablets
Prolonged-release Tamsulosin Capsules	Tamsulosin Prolonged-release Capsules
Prolonged-release Tamsulosin Tablets	Tamsulosin Prolonged-release Tablets
Prolonged-release Theophylline Tablets	Theophylline Prolonged-release Tablets
Prolonged-release Tramadol Capsules	Tramadol Prolonged-release Capsules
Prolonged-release Tramadol Tablets	Tramadol Prolonged-release Tablets
Prolonged-release Trospium Chloride Capsules	Trospium Chloride Prolonged-release Capsules
Prolonged-released Venlafaxine Capsules	Venlafaxine Prolonged-released Capsules
Prolonged-released Venlafaxine Tablets	Venlafaxine Prolonged-released Tablets
Prolonged-released Verapamil Capsules	Verapamil Prolonged-released Capsules
Prolonged-released Verapamil Tablets	Verapamil Prolonged-released Tablets

Herbal Drugs, Herbal Drug Preparations and Herbal Medicinal Products

Gastro-resistant Peppermint Oil Capsules	Peppermint Oil Gastro-resistant Capsules

Radiopharmaceutical Preparations

Fluorodopa (^{18}F) Injection	Fluorodopa (^{18}F) (Prepared by Electrophilic Substitution) Injection

General Notices

CONTENTS OF THE GENERAL NOTICES

Part I

The British Pharmacopoeia comprises the entire text within this publication. The word 'official' is used in the Pharmacopoeia to signify 'of the Pharmacopoeia'. It applies to any title, substance, preparation, method or statement included in the general notices, monographs and appendices of the Pharmacopoeia. The abbreviation for British Pharmacopoeia is BP.

European Pharmacopoeia

Monographs of the European Pharmacopoeia are reproduced in this edition of the British Pharmacopoeia by incorporation of the text published under the direction of the Council of Europe (Partial Agreement) in accordance with the Convention on the Elaboration of a European Pharmacopoeia (Treaty Series No. 32 (1974) CMND 5763) as amended by the Protocol to the Convention (Treaty Series No. MISC16 (1990) CMND 1133). They are included for the convenience of users of the British Pharmacopoeia. In cases of doubt or dispute reference should be made to the Council of Europe text.

Monographs of the European Pharmacopoeia are distinguished by a chaplet of stars against the title and by reference to the European Pharmacopoeia monograph number included immediately below the title in italics. The beginning and end of text from the European Pharmacopoeia are denoted by means of horizontal lines with the symbol '*Ph Eur*' ranged left and right, respectively.

The general provisions of the European Pharmacopoeia relating to different types of dosage form are included in the appropriate general monograph in that section of the British Pharmacopoeia entitled Monographs: Formulated Preparations. These general provisions apply to all dosage forms of the type defined, whether or not an individual monograph is included in the British Pharmacopoeia. In addition, the provisions of the European Pharmacopoeia General Monograph for Pharmaceutical Preparations apply to all dosage forms, whether or not an individual monograph is included in the British Pharmacopoeia.

Texts of the European Pharmacopoeia are governed by the General Notices of the European Pharmacopoeia. These are reproduced as Part III of these notices.

Part II

The following general notices apply to the statements made in the monographs of the British Pharmacopoeia other than those reproduced from the European Pharmacopoeia and to the statements made in the Appendices of the British Pharmacopoeia other than when a method, test or other matter described in an appendix is invoked in a monograph reproduced from the European Pharmacopoeia.

Official Standards The requirements stated in the monographs of the Pharmacopoeia apply to articles that are intended for medicinal use but not necessarily to articles that may be sold under the same name for other purposes. An article intended for medicinal use that is described by means of an official title must comply with the requirements of the relevant monograph. A formulated preparation must comply throughout its assigned shelf-life (period of validity). The subject of any other monograph must comply throughout its period of use.

A monograph is to be construed in accordance with any general monograph or notice or any appendix, note or other explanatory material that is contained in this edition and that is applicable to that monograph. All statements contained in the monographs, except where a specific general notice indicates otherwise and with the exceptions given below, constitute standards for the official articles. An article is not of pharmacopoeial quality unless it complies with all of the requirements stated. This does not imply that a manufacturer is obliged to perform all the tests in a monograph in order to assess compliance with the Pharmacopoeia before release of a product. The manufacturer may assure himself that a product is of pharmacopoeial quality by other means, for example, from data derived from validation studies of the manufacturing process, from in-process controls or from a combination of the two. Parametric release in appropriate circumstances is thus not precluded by the need to comply with the Pharmacopoeia. The general notice on Assays and Tests indicates that analytical methods other than those described in the Pharmacopoeia may be employed for routine purposes.

Requirements in monographs have been framed to provide appropriate limitation of potential impurities rather than to provide against all possible impurities. Material found to contain an impurity not detectable by means of the prescribed tests is not of pharmacopoeial quality if the nature or amount of the impurity found is incompatible with good pharmaceutical practice.

The status of any statement given under the headings Definition, Production, Characteristics, Storage, Labelling or Action and use is defined within the general notice relating to the relevant heading. In addition to any exceptions indicated by one of the general notices referred to above, the following parts of a monograph do not constitute standards: (a) a graphic or molecular formula given at the beginning of a monograph; (b) a molecular weight; (c) a Chemical Abstracts Service Registry Number; (d) any information given at the end of a monograph concerning impurities known to be limited by that monograph; (e) information in any annex to a

monograph. Any statement containing the word 'should' constitutes non-mandatory advice or recommendation.

The expression 'unless otherwise justified and authorised' means that the requirement in question has to be met, unless a competent authority authorises a modification or exemption where justified in a particular case. The term 'competent authority' means the national, supranational or international body or organisation vested with the authority for making decisions concerning the issue in question. It may, for example, be a licensing authority or an official control laboratory. For a formulated preparation that is the subject of monograph in the British Pharmacopoeia any justified and authorised modification to, or exemption from, the requirements of the relevant general monograph of the European Pharmacopoeia is stated in the individual monograph. For example, the general monograph for Tablets requires that Uncoated Tablets, except for chewable tablets, disintegrate within 15 minutes; for Calcium Lactate Tablets a time of 30 minutes is permitted.

Many of the general monographs for formulated preparations include statements and requirements additional to those of the European Pharmacopoeia that are applicable to the individual monographs of the British Pharmacopoeia. Such statements and requirements apply to all monographs for that dosage form included in the Pharmacopoeia unless otherwise indicated in the individual monograph.

Where a monograph on a biological substance or preparation refers to a strain, a test, a method, a substance, etc., using the qualifications 'suitable' or 'appropriate' without further definition in the text, the choice of such strain, test, method, substance, etc., is made in accordance with any international agreements or national regulations affecting the subject concerned.

Definition of Terms

Where the term 'about' is included in a monograph or test it should be taken to mean approximately (fairly correct or accurate; near to the actual value).

Where the term 'corresponds' is included in a monograph or test it should be taken to mean similar or equivalent in character or quantity.

Where the term 'similar' is included in a monograph or test it should be taken to mean alike though not necessarily identical.

Further qualifiers (such as numerical acceptance criteria) for the above terms are not included in the BP. The acceptance criteria for any individual case is set based on the range of results obtained from known reference samples, the level of precision of the equipment or apparatus used and the level of accuracy required for the particular application. The user should determine the variability seen in his/her own laboratory and set in-house acceptance criteria that he/she judges to be appropriate based on the local operating conditions.

Expression of Standards

Where the standard for the content of a substance described in a monograph is expressed in terms of the chemical formula for that substance an upper limit exceeding 100% may be stated. Such an upper limit applies to the result of the assay calculated in terms of the equivalent content of the specified chemical formula. For example, the statement 'contains not less than 99.0% and not more than 101.0% of $C_{20}H_{24}N_2O_2,HCl$' implies that the result of the assay is not less than 99.0% and not more than 101.0%, calculated in terms of the equivalent content of $C_{20}H_{24}N_2O_2,HCl$.

Where the result of an assay or test is required to be calculated with reference to the dried, anhydrous or ignited substance, the substance free from a specified solvent or to the peptide content, the determination of loss on drying, water content, loss on ignition, content of the specified solvent or peptide content is carried out by the method prescribed in the relevant test in the monograph.

Temperature The Celsius thermometric scale is used in expressing temperatures.

Weights and Measures The metric system of weights and measures is employed; SI Units have generally been adopted. Metric measures are required to have been graduated at 20° and all measurements involved in the analytical operations of the Pharmacopoeia are intended, unless otherwise stated, to be made at that temperature. Graduated glass apparatus used in analytical operations should comply with Class A requirements of the appropriate International Standard issued by the International Organization for Standardization. The abbreviation for litre is 'L' throughout the Pharmacopoeia. In line with European Directive 80/181/EEC, the abbreviation 'l' is also permitted for use.

Atomic Weights The atomic weights adopted are the values given in the Table of Relative Atomic Weights 2001 published by the International Union of Pure and Applied Chemistry (Appendix XXV).

Constant Weight The term 'constant weight', used in relation to the process of drying or the process of ignition, means that two consecutive weighings do not differ by more than 0.5 mg, the second weighing being made after an additional period of drying or ignition under the specified conditions appropriate to the nature and quantity of the residue (1 hour is usually suitable).

Expression of Concentrations The term 'per cent' or more usually the symbol '%' is used with one of four different meanings in the expression of concentrations according to circumstances. In order that the meaning to be attached to the expression in each instance is clear, the following notation is used:

Per cent w/w (% w/w) (percentage weight in weight) expresses the number of grams of solute in 100 g of product.

Per cent w/v (% w/v) (percentage weight in volume) expresses the number of grams of solute in 100 mL of product.

Per cent v/v (% v/v) (percentage volume in volume) expresses the number of millilitres of solute in 100 mL of product.

Per cent v/w (% v/w) (percentage volume in weight) expresses the number of millilitres of solute in 100 g of product.

Usually the strength of solutions of solids in liquids is expressed as percentage weight in volume, of liquids in liquids as percentage volume in volume and of gases in liquids as percentage weight in weight.

When the concentration of a solution is expressed as parts per million (ppm), it means weight in weight, unless otherwise specified.

When the concentration of a solution is expressed as parts of dissolved substance in parts of the solution, it means parts by weight (g) of a solid in parts by volume (mL) of the final solution; or parts by volume (mL) of a liquid in parts by volume (mL) of the final solution; or parts by weight (g) of a gas in parts by weight (g) of the final solution.

When the concentration of a solution is expressed in molarity designated by the symbol M preceded by a number, it denotes the number of moles of the stated solute contained in sufficient Purified Water (unless otherwise stated) to produce 1 litre of solution.

Water Bath The term 'water bath' means a bath of boiling water, unless water at some other temperature is indicated in the text. An alternative form of heating may be employed providing that the required temperature is approximately maintained but not exceeded.

Reagents The reagents required for the assays and tests of the Pharmacopoeia are defined in appendices. The descriptions set out in the appendices do not imply that the materials are suitable for use in medicine.

Indicators Indicators, the colours of which change over approximately the same range of pH, may be substituted for one another but in the event of doubt or dispute as to the equivalence of indicators for a particular purpose, the indicator specified in the text is alone authoritative.

The quantity of an indicator solution appropriate for use in acid-base titrations described in assays or tests is 0.1 mL unless otherwise stated in the text.

Any solvent required in an assay or test in which an indicator is specified is previously neutralised to the indicator, unless a blank test is prescribed.

Caution Statements A number of materials described in the monographs and some of the reagents specified for use in the assays and tests of the Pharmacopoeia may be injurious to health unless adequate precautions are taken. The principles of good laboratory practice and the provisions of any appropriate regulations such as those issued in the United Kingdom in accordance with the Health and Safety at Work *etc.* Act 1974 should be observed at all times in carrying out the assays and tests of the Pharmacopoeia.

Attention is drawn to particular hazards in certain monographs by means of an italicised statement; the absence of such a statement should not however be taken to mean that no hazard exists.

Titles Subsidiary titles, where included, have the same significance as the main titles. An abbreviated title constructed in accordance with the directions given in Appendix XXI A has the same significance as the main title.

Titles that are derived by the suitable inversion of words of a main or subsidiary title, with the addition of a preposition if appropriate, are also official titles. Thus, the following are all official titles: Aspirin Tablets, Tablets of Aspirin; Atropine Injection, Injection of Atropine.

A title of a formulated preparation that includes the full nonproprietary name of the active ingredient or ingredients, where this is not included in the title of the monograph, is also an official title. For example, the title Promethazine Hydrochloride Oral Solution has the same significance as Promethazine Oral Solution and the title Brompheniramine Maleate Tablets has the same significance as Brompheniramine Tablets.

Where the English title at the head of a monograph in the European Pharmacopoeia is different from that at the head of the text incorporated into the British Pharmacopoeia, an Approved Synonym has been created on the recommendation of the British Pharmacopoeia Commission. Approved Synonyms have the same significance as the main title and are thus official

titles. A cumulative list of such Approved Synonyms is provided in Appendix XXI B.

Where the names of pharmacopoeial substances, preparations and other materials occur in the text they are printed with capital initial letters and this indicates that materials of Pharmacopoeial quality must be used. Words in the text that name a reagent or other material, a physical characteristic or a process that is described or defined in an appendix are printed in italic type, for example, *methanol*, *absorbance*, *gas chromatography*, and these imply compliance with the requirements specified in the appropriate appendix.

Chemical Formulae When the chemical composition of an official substance is known or generally accepted, the graphic and molecular formulae, the molecular weight and the Chemical Abstracts Service Registry Number are normally given at the beginning of the monograph for information. This information refers to the chemically pure substance and is not to be regarded as an indication of the purity of the official material. Elsewhere, in statements of standards of purity and strength and in descriptions of processes of assay, it is evident from the context that the formulae denote the chemically pure substances.

Where the absolute stereochemical configuration is specified, the International Union of Pure and Applied Chemistry (IUPAC) *R*/*S* and *E*/*Z* systems of designation have been used. If the substance is an enantiomer of unknown absolute stereochemistry the sign of the optical rotation, as determined in the solvent and under the conditions specified in the monograph, has been attached to the systematic name. An indication of sign of rotation has also been given where this is incorporated in a trivial name that appears on an IUPAC preferred list.

All amino acids, except glycine, have the L-configuration unless otherwise indicated. The three-letter and one-letter symbols used for amino acids in peptide and protein sequences are those recommended by the Joint Commission on Biochemical Nomenclature of the International Union of Pure and Applied Chemistry and the International Union of Biochemistry and Molecular Biology.

In the graphic formulae the following abbreviations are used:

Me	$-CH_3$	Bu^s	$-CH(CH_3)CH_2CH_3$
Et	$-CH_2CH_3$	Bu^n	$-CH_2CH_2CH_2CH_3$
Pr^i	$-CH(CH_3)_2$	Bu^t	$-C(CH_3)_3$
Pr^n	$-CH_2CH_2CH_3$	Ph	$-C_6H_5$
Bu^i	$-CH_2CH(CH_3)_2$	Ac	$-COCH_3$

Definition Statements given under the heading Definition constitute an official definition of the substance, preparation or other article that is the subject of the monograph. They constitute instructions or requirements and are mandatory in nature.

Certain medicinal or pharmaceutical substances and other articles are defined by reference to a particular method of manufacture. A statement that a substance or article *is* prepared or obtained by a certain method constitutes part of the official definition and implies that other methods are not permitted. A statement that a substance *may be* prepared or obtained by a certain method, however, indicates that this is one possible method and does not imply that other methods are proscribed.

Additional statements concerning the definition of formulated preparations are given in the general notice on Manufacture of Formulated Preparations.

Production Statements given under the heading Production draw attention to particular aspects of the manufacturing process but are not necessarily comprehensive. They constitute mandatory instructions to manufacturers. They may relate, for example, to source materials, to the manufacturing process itself and its validation and control, to in-process testing or to testing that is to be carried out by the manufacturer on the final product (bulk material or dosage form) either on selected batches or on each batch prior to release. These statements cannot necessarily be verified on a sample of the final product by an independent analyst. The competent authority may establish that the instructions have been followed, for example, by examination of data received from the manufacturer, by inspection or by testing appropriate samples.

The absence of a section on Production does not imply that attention to features such as those referred to above is not required. A substance, preparation or article described in a monograph of the Pharmacopoeia is to be manufactured in accordance with the principles of good manufacturing practice and in accordance with relevant international agreements and supranational and national regulations governing medicinal products.

Where in the section under the heading Production a monograph on a vaccine defines the characteristics of the vaccine strain to be used, any test methods given for confirming these characteristics are provided as examples of suitable methods. The use of these methods is not mandatory.

Additional statements concerning the production of formulated preparations are given in the general notice on Manufacture of Formulated Preparations.

Manufacture of Formulated Preparations Attention is drawn to the need to observe adequate hygienic precautions in the preparation and dispensing of pharmaceutical formulations. The principles of good pharmaceutical manufacturing practice should be observed.

The Definition in certain monographs for pharmaceutical preparations is given in terms of the principal ingredients only. Any ingredient, other than those included in the Definition, must comply with the general notice on Excipients and the product must conform with the Pharmacopoeial requirements.

The Definition in other monographs for pharmaceutical preparations is presented as a full formula. No deviation from the stated formula is permitted except those allowed by the general notices on Colouring Agents and Antimicrobial Preservatives. Where additionally directions are given under the heading Extemporaneous Preparation these are intended for the extemporaneous preparation of relatively small quantities for short-term supply and use. When so prepared, no deviation from the stated directions is permitted. If, however, such a pharmaceutical preparation is manufactured on a larger scale with the intention that it may be stored, deviations from the stated directions are permitted provided that the final product meets the following criteria:

(1) compliance with all of the requirements stated in the monograph;
(2) retention of the essential characteristics of the preparation made strictly in accordance with the directions of the Pharmacopoeia.

Monographs for yet other pharmaceutical preparations include both a Definition in terms of the principal ingredients and, under the side-heading Extemporaneous Preparation, a full formula together with, in some cases, directions for their preparation. Such full formulae and directions are intended for the extemporaneous preparation of relatively small quantities for short-term supply and use. When so prepared, no deviation from the stated formula and directions is permitted. If, however, such a pharmaceutical preparation is manufactured on a larger scale with the intention that it may be stored, deviations from the formula and directions stated under the heading Extemporaneous Preparation are permitted provided that any ingredient, other than those included in the Definition, complies with the general notice on Excipients and that the final product meets the following criteria:

(1) accordance with the Definition stated in the monograph;
(2) compliance with all of the requirements stated in the monograph;
(3) retention of the essential characteristics of the preparation made strictly in accordance with the formula and directions of the Pharmacopoeia.

In the manufacture of any official preparation on a large scale with the intention that it should be stored, in addition to following any instruction under the heading Production, it is necessary to ascertain that the product is satisfactory with respect to its physical and chemical stability and its state of preservation over the claimed shelf-life. This applies irrespective of whether the formula of the Pharmacopoeia and any instructions given under the heading Extemporaneous Preparation are followed precisely or modified. Provided that the preparation has been shown to be stable in other respects, deterioration due to microbial contamination may be inhibited by the incorporation of a suitable antimicrobial preservative. In such circumstances the label states appropriate storage conditions, the date after which the product should not be used and the identity and concentration of the antimicrobial preservative.

Freshly and Recently Prepared The direction, given under the heading Extemporaneous Preparation, that a preparation must be freshly prepared indicates that it must be made not more than 24 hours before it is issued for use. The direction that a preparation should be recently prepared indicates that deterioration is likely if the preparation is stored for longer than about 4 weeks at 15° to 25°.

Methods of Sterilisation The methods of sterilisation used in preparing the sterile materials described in the Pharmacopoeia are given in Appendix XVIII. For aqueous preparations, steam sterilisation (heating in an autoclave) is the method of choice wherever it is known to be suitable. Any method of sterilisation must be validated with respect to both the assurance of sterility and the integrity of the product and to ensure that the final product complies with the requirements of the monograph.

Water The term water used without qualification in formulae for formulated preparations means either potable water freshly drawn direct from the public supply and suitable for drinking or freshly boiled and cooled Purified

Water. The latter should be used if the public supply is from a local storage tank or if the potable water is unsuitable for a particular preparation.

Excipients Where an excipient for which there is a pharmacopoeial monograph is used in preparing an official preparation it shall comply with that monograph. Any substance added in preparing an official preparation shall be innocuous, shall have no adverse influence on the therapeutic efficacy of the active ingredients and shall not interfere with the assays and tests of the Pharmacopoeia. Particular care should be taken to ensure that such substances are free from harmful organisms.

Colouring Agents If in a monograph for a formulated preparation defined by means of a full formula a specific colouring agent or agents is prescribed, suitable alternatives approved in the country concerned may be substituted.

Antimicrobial Preservatives When the term 'suitable antimicrobial preservative' is used it is implied that the preparation concerned will be effectively preserved according to the appropriate criteria applied and interpreted as described in the test for *efficacy of antimicrobial preservation* (Appendix XVI C). In certain monographs for formulated preparations defined by means of a full formula, a specific antimicrobial agent or agents may be prescribed; suitable alternatives may be substituted provided that their identity and concentration are stated on the label.

Characteristics Statements given under the heading Characteristics are not to be interpreted in a strict sense and are not to be regarded as official requirements. Statements on taste are provided only in cases where this property is a guide to the acceptability of the material (for example, a material used primarily for flavouring). The status of statements on solubility is given in the general notice on Solubility.

Solubility Statements on solubility given under the heading Characteristics are intended as information on the approximate solubility at a temperature between 15° and 25°, unless otherwise stated, and are not to be considered as official requirements.

Statements given under headings such as Solubility in ethanol express exact requirements and constitute part of the standards for the substances under which they occur.

The following table indicates the meanings of the terms used in statements of approximate solubilities.

Descriptive term	Approximate volume of solvent in millilitres per gram of solute
very soluble	less than 1
freely soluble	from 1 to 10
soluble	from 10 to 30
sparingly soluble	from 30 to 100
slightly soluble	from 100 to 1000
very slightly soluble	from 1000 to 10 000
practically insoluble	more than 10 000

The term 'partly soluble' is used to describe a mixture of which only some of the components dissolve.

Identification The tests described or referred to under the heading Identification are not necessarily sufficient to establish absolute proof of identity. They provide a means of verifying that the identity of the material being examined is in accordance with the label on the container.

Unless otherwise prescribed, identification tests are carried out at a temperature between 15° and 25°.

Reference spectra Where a monograph refers to an infrared reference spectrum, this spectrum is provided in a separate section of the Pharmacopoeia. A sample spectrum is considered to be concordant with a reference spectrum if the transmission minima (absorption maxima) of the principal bands in the sample correspond in position, relative intensities and shape to those of the reference. Instrumentation software may be used to calculate concordance with a previously recorded reference spectrum.

When tests for infrared absorption are applied to material extracted from formulated preparations, strict concordance with the specified reference spectrum may not always be possible, but nevertheless a close resemblance between the spectrum of the extracted material and the specified reference spectrum should be achieved.

Assays and Tests The assays and tests described are the official methods upon which the standards of the Pharmacopoeia depend. The analyst is not precluded from employing alternative methods, including methods of micro-analysis, in any assay or test if it is known that the method used will give a result of equivalent accuracy. Local reference materials may be used for routine analysis, provided that these are calibrated against the official reference materials. In the event of doubt or dispute, the methods of analysis, the reference materials and the reference spectra of the Pharmacopoeia are alone authoritative.

Where the solvent used for a solution is not named, the solvent is Purified Water.

Unless otherwise prescribed, the assays and tests are carried out at a temperature between 15° and 25°.

A temperature in a test for Loss on drying, where no temperature range is given, implies a range of ±2° about the stated value.

Visual comparative tests, unless otherwise prescribed, are carried out using identical tubes of colourless, transparent, neutral glass with a flat base. The volumes of liquid prescribed are for use with tubes 16 mm in internal diameter; tubes with a larger internal diameter may be used but the volume of liquid examined must be increased so that the depth of liquid in the tubes is not less than that obtained when the prescribed volume of liquid and tubes 16 mm in internal diameter are used. Equal volumes of the liquids to be compared are examined down the vertical axis of the tubes against a white background or, if necessary, against a black background. The examination is carried out in diffuse light.

Where a direction is given that an analytical operation is to be carried out 'in subdued light', precautions should be taken to avoid exposure to direct sunlight or other strong light. Where a direction is given that an analytical operation is to be carried out 'protected from light', precautions should be taken to exclude actinic light by the use of low-actinic glassware, working in a dark room or similar procedures.

For preparations other than those of fixed strength, the quantity to be taken for an assay or test is usually expressed in terms of the active ingredient. This means that the quantity of the active ingredient expected to be present and the quantity of the preparation to be taken are calculated from the strength stated on the label.

In assays the approximate quantity to be taken for examination is indicated but the quantity actually used must not deviate by more than 10% from that stated. The quantity taken is accurately weighed or measured and the result of the assay is calculated from this exact quantity. Reagents are measured and the procedures are carried out with an accuracy commensurate with the degree of precision implied by the standard stated for the assay.

In tests the stated quantity to be taken for examination must be used unless any divergence can be taken into account in conducting the test and calculating the result. The quantity taken is accurately weighed or measured with the degree of precision implied by the standard or, where the standard is not stated numerically (for example, in tests for Clarity and colour of solution), with the degree of precision implied by the number of significant figures stated. Reagents are measured and the procedures are carried out with an accuracy commensurate with this degree of precision.

The limits stated in monographs are based on data obtained in normal analytical practice; they take account of normal analytical errors, of acceptable variations in manufacture and of deterioration to an extent considered acceptable. No further tolerances are to be applied to the limits prescribed to determine whether the article being examined complies with the requirements of the monograph.

In determining compliance with a numerical limit, the calculated result of a test or assay is first rounded to the number of significant figures stated, unless otherwise prescribed. The last figure is increased by 1 when the part rejected is equal to or exceeds one half-unit, whereas it is not modified when the part rejected is less than a half-unit.

In certain tests, the concentration of impurity is given in parentheses either as a percentage or in parts per million by weight (ppm). In chromatographic tests such concentrations are stated as a percentage irrespective of the limit. In other tests they are usually stated in ppm unless the limit exceeds 500 ppm. In those chromatographic tests in which a secondary spot or peak in a chromatogram obtained with a solution of the substance being examined is described as corresponding to a named impurity and is compared with a spot or peak in a chromatogram obtained with a reference solution of the same impurity, the percentage given in parentheses indicates the limit for that impurity. In those chromatographic tests in which a spot or peak in a chromatogram obtained with a solution of the substance being examined is described in terms other than as corresponding to a named impurity (commonly, for example, as any (other) *secondary spot* or *peak*) but is compared with a spot or peak in a chromatogram obtained with a reference solution of a named impurity, the percentage given in parentheses indicates an impurity limit expressed in terms of a nominal concentration of the named impurity. In chromatographic tests in which a comparison is made between spots or peaks in chromatograms obtained with solutions of different concentrations of the substance being examined, the percentage given in parentheses indicates an impurity limit expressed in terms of a nominal concentration of the medicinal substance itself. In some monographs, in particular those for

certain formulated preparations, the impurity limit is expressed in terms of a nominal concentration of the active moiety rather than of the medicinal substance itself. Where necessary for clarification the terms in which the limit is expressed are stated within the monograph.

In all cases where an impurity limit is given in parentheses, the figures given are approximations for information only; conformity with the requirements is determined on the basis of compliance or otherwise with the stated test.

The use of a proprietary designation to identify a material used in an assay or test does not imply that another equally suitable material may not be used.

Biological Assays and Tests

Methods of assay described as Suggested methods are not obligatory, but when another method is used its precision must be not less than that required for the Suggested method.

For those antibiotics for which the monograph specifies a microbiological assay the potency requirement is expressed in the monograph in International Units (IU) per milligram. The material is not of pharmacopoeial quality if the upper fiducial limit of error is less than the stated potency. For such antibiotics the required precision of the assay is stated in the monograph in terms of the fiducial limits of error about the estimated potency.

For other substances and preparations for which the monograph specifies a biological assay, unless otherwise stated, the precision of the assay is such that the fiducial limits of error, expressed as a percentage of the estimated potency, are within a range not wider than that obtained by multiplying by a factor of 10 the square roots of the limits given in the monograph for the fiducial limits of error about the stated potency.

In all cases fiducial limits of error are based on a probability of 95% ($P = 0.95$).

Where the biological assay is being used to ascertain the purity of the material, the stated potency means the potency stated on the label in terms of International Units (IU) or other Units per gram, per milligram or per millilitre. When no such statement appears on the label, the stated potency means the fixed or minimum potency required in the monograph. This interpretation of stated potency applies in all cases except where the monograph specifically directs otherwise.

Where the biological assay is being used to determine the total activity in the container, the stated potency means the total number of International Units (IU) or other Units stated on the label or, if no such statement appears, the total activity calculated in accordance with the instructions in the monograph.

Wherever possible the primary standard used in an assay or test is the respective International Standard or Reference Preparation established by the World Health Organization for international use and the biological activity is expressed in International Units (IU).

In other cases, where Units are referred to in an assay or test, the Unit for a particular substance or preparation is, for the United Kingdom, the specific biological activity contained in such an amount of the respective primary standard as the appropriate international or national organisation indicates. The necessary information is provided with the primary standard.

Unless otherwise directed, animals used in an assay or a test are healthy animals, drawn from a uniform stock, that have not previously been treated

with any material that will interfere with the assay or test. Unless otherwise stated, guinea-pigs weigh not less than 250 g or, when used in systemic toxicity tests, not less than 350 g. When used in skin tests they are white or light coloured. Unless otherwise stated, mice weigh not less than 17 g and not more than 22 g.

Certain of the biological assays and tests of the Pharmacopoeia are such that in the United Kingdom they may be carried out only in accordance with the Animals (Scientific Procedures) Act 1986. Instructions included in such assays and tests in the Pharmacopoeia, with respect to the handling of animals, are therefore confined to those concerned with the accuracy and reproducibility of the assay or test.

Reference Substances and Reference Preparations

Certain monographs require the use of a reference substance, a reference preparation or a reference spectrum. These are chosen with regard to their intended use as prescribed in the monographs of the Pharmacopoeia and are not necessarily suitable in other circumstances.

Any information necessary for proper use of the reference substance or reference preparation is given on the label or in the accompanying leaflet or brochure. Where no drying conditions are stated in the leaflet or on the label, the substance is to be used as received. No certificate of analysis or other data not relevant to the prescribed use of the product are provided. The products are guaranteed to be suitable for use for a period of three months from dispatch when stored under the appropriate conditions. The stability of the contents of opened containers cannot be guaranteed. The current lot is listed in the BP Laboratory website catalogue. Additional information is provided in Supplementary Chapter III E.

Chemical Reference Substances The abbreviation BPCRS indicates a Chemical Reference Substance established by the British Pharmacopoeia Commission. The abbreviation CRS or EPCRS indicates a Chemical Reference Substance established by the European Pharmacopoeia Commission. Some Chemical Reference Substances are used for the microbiological assay of antibiotics and their activity is stated, in International Units, on the label or on the accompanying leaflet and defined in the same manner as for Biological Reference Preparations.

Biological Reference Preparations The majority of the primary biological reference preparations referred to are the appropriate International Standards and Reference Preparations established by the World Health Organisation. Because these reference materials are usually available only in limited quantities, the European Pharmacopoeia has established Biological Reference Preparations (indicated by the abbreviation BRP or EPBRP) where appropriate. Where applicable, the potency of the Biological Reference Preparations is expressed in International Units. For some Biological Reference Preparations, where an international standard or reference preparation does not exist, the potency is expressed in European Pharmacopoeia Units.

Storage

Statements under the side-heading Storage constitute non-mandatory advice. The substances and preparations described in the Pharmacopoeia are to be stored under conditions that prevent contamination and, as far as possible, deterioration. Unless otherwise stated in the monograph, the substances and preparations described in the Pharmacopoeia are kept in well-closed containers and stored at a temperature not exceeding 25°. Precautions that should be taken in relation to the effects of the

atmosphere, moisture, heat and light are indicated, where appropriate, in the monographs. Further precautions may be necessary when some materials are stored in tropical climates or under other severe conditions.

The expression 'protected from moisture' means that the product is to be stored in an airtight container. Care is to be taken when the container is opened in a damp atmosphere. A low moisture content may be maintained, if necessary, by the use of a desiccant in the container provided that direct contact with the product is avoided.

The expression 'protected from light' means that the product is to be stored either in a container made of a material that absorbs actinic light sufficiently to protect the contents from change induced by such light or in a container enclosed in an outer cover that provides such protection or stored in a place from which all such light is excluded.

The expression 'tamper-evident container' means a closed container fitted with a device that reveals irreversibly whether the container has been opened, whereas, the expression 'tamper-proof container' means a closed container in which access to the contents is prevented under normal conditions of use. The two terms are considered to be synonymous by the European Pharmacopoeia Commission.

Labelling The labelling requirements of the Pharmacopoeia are not comprehensive, and the provisions of regulations issued in accordance with the requirements of the territory in which the medicinal product is to be used should be met.

Licensed medicines intended for use within the United Kingdom must comply with the requirements of The Human Medicines Regulations 2012 and European Directive 2001/83/EC, Title V (as amended) in respect of their labelling and package leaflets, together with those regulations for the labelling of hazardous materials.

Best practice guidance on the labelling and packaging of medicines for use in the United Kingdom advises that certain items of information are deemed critical for the safe use of the medicine (see "Best Practice Guidance on the Labelling and Packaging of Medicines" issued by the MHRA, 2012). Further information and guidance on the labelling of medicinal products can be found in Supplementary Chapter I G.

Such matters as the exact form of wording to be used and whether a particular item of information should appear on the primary label and additionally, or alternatively, on the package or exceptionally in a leaflet are, in general, outside the scope of the Pharmacopoeia. When the term 'label' is used in Labelling statements of the Pharmacopoeia, decisions as to where the particular statement should appear should therefore be made in accordance with relevant legislation.

The label of every official formulated preparation other than those of fixed strength also states the content of the active ingredient or ingredients expressed in the terms required by the monograph. Where the content of active ingredient is required to be expressed in terms other than the weight of the official medicinal substance used in making the formulation, this is specifically stated under the heading Labelling. Unless otherwise stated in the monograph, the content of the active ingredient is expressed in terms of the official medicinal substance used in making the formulation.

These requirements do not necessarily apply to unlicensed preparations supplied in accordance with a prescription. For requirements for unlicensed medicines see the general monograph on Unlicensed Medicines.

Action and Use The statements given under this heading in monographs are intended only as information on the principal pharmacological actions or the uses of the materials in medicine or pharmacy. It should not be assumed that the substance has no other action or use. The statements are not intended to be binding on prescribers or to limit their discretion.

Crude Drugs; Traditional Herbal and Complementary Medicines *Herbal and complementary medicines are classed as medicines under European Directive 2001/83/EC as amended. It is emphasised that, although requirements for the quality of the material are provided in the monograph to assist the registration scheme by the UK Licensing Authority, the British Pharmacopoeia Commission has not assessed the safety or efficacy of the material in traditional use.*

Monograph Title For traditional herbal medicines, the monograph title is a combination of the binomial name together with a description of use. Monographs for the material that has not been processed (the herbal drug) and the processed material (the herbal drug preparation) are published where possible. To distinguish between the two, the word 'Processed' is included in the relevant monograph title.

Definition Under the heading Definition, the botanical name together with any synonym is given. Where appropriate, for material that has not been processed, information on the collection/harvesting and/or treatment/drying of the whole herbal drug may be given. For processed materials, the method of processing, where appropriate, will normally be given in a separate section.

Characteristics References to odour are included only where this is highly characteristic. References to taste are not included.

Control methods Where applicable, the control methods to be used in monographs are:

(a) macroscopical and microscopical descriptions and chemical/chromatographic tests for identification

(b) tests for absence of any related species

(c) microbial test to assure microbial quality

(d) tests for inorganic impurities and non-specific purity tests, including extractive tests, Sulfated ash and Heavy metals, where appropriate

(e) test for Loss on drying or Water

(f) wherever possible, a method for assaying the active constituent(s) or suitable marker constituent(s).

The macroscopical characteristics include those features that can be seen by the unaided eye or by the use of a hand lens. When two species/subspecies of the same plant are included in the Definition, individual differences between the two are indicated where possible.

The description of the microscopical characteristics of the powdered drug includes information on the dominant or the most specific characters. Where it is considered to be an aid to identification, illustrations of the powdered drug may be provided.

The following aspects are controlled by the general monograph for Herbal Drugs: they are required to be free from moulds, insects, decay, animal matter and animal excreta. Unless otherwise prescribed the amount of foreign matter is not more than 2% w/w. Microbial contamination should be minimal.

In determining the content of the active constituents or the suitable marker substances measurements are made with reference to the dried or

anhydrous herbal drug. In the tests for Acid-insoluble ash, Ash, Extractive soluble in ethanol, Loss on drying, Sulfated ash, Water, Water-soluble ash and Water-soluble extractive of herbal drugs, the calculations are made with reference to the herbal drug that has not been specifically dried unless otherwise prescribed in the monograph.

Homoeopathic Medicines

Homoeopathic medicines are classed as medicines under European Directive 2001/83/EC as amended. It is emphasised that, although requirements for the quality of the material are provided in the relevant monograph in order to assist the simplified registration scheme by the UK Licensing Authority, the British Pharmacopoeia Commission has not assessed the safety or efficacy of the material in use.

All materials used for the production of homoeopathic medicines, including excipients, must comply with European Pharmacopoeia or British Pharmacopoeia monographs for those materials. Where such European Pharmacopoeia or British Pharmacopoeia monographs do not exist, each material used for the production of homoeopathic medicines must comply with an official national pharmacopoeia of a Member State.

British Pharmacopoeia monographs for homoeopathic medicines apply to homoeopathic stocks and mother tinctures only, but may be prefaced by a section which details the quality requirements applicable to the principle component where there is no European Pharmacopoeia or British Pharmacopoeia monograph for the material. These monographs also include either general statements on the methods of preparation or refer to specific methods of preparation given in the European Pharmacopoeia. Homoeopathic stocks and mother tinctures undergo the further process referred to as potentisation. Potentisation is a term specific to homoeopathic medicine and is a process of dilution of stocks and mother tinctures to produce the final product.

Identification tests are established for the components in homoeopathic stocks and usually relate to those applied to the materials used in the production of the homoeopathic stocks. An assay is included for the principal component(s) where possible. For mother tinctures, an identification test, usually chromatographic, is established and, where applicable, an assay for the principle component(s); where appropriate, other tests, related to the solvent, dry matter or known adulterants, are included.

Specifications have not been set for final homoeopathic products due to the high dilution used in their preparation and the subsequent difficulty in applying analytical methodology.

Statements under Crude Drugs; Traditional Herbal and Complementary Medicines also apply to homoeopathic stocks and mother tinctures, when appropriate.

Unlicensed Medicines

The General Monograph for Unlicensed Medicines applies to those formulations used in human medicine that are prepared under a Manufacturer's 'Specials' Licence or prepared extemporaneously under the supervision of a pharmacist, whether or not there is a published monograph for the specific dosage form.

An article intended for medicinal use that is described by means of an official title must comply with the requirements of the relevant monograph. A formulated preparation must comply throughout its assigned shelf-life

(period of validity). The subject of any other monograph must comply throughout its period of use.

Unlicensed medicines that are prepared under a Manufacturer's 'Specials' Licence comply with the requirements of the General Monograph for Pharmaceutical Preparations, the requirements of the General Monograph for Unlicensed Medicines and, where applicable, the requirements of the individual monograph for the specific dosage form.

Unlicensed medicines prepared extemporaneously under the supervision of a pharmacist comply with the requirements of the General Monograph for Pharmaceutical Preparations, the requirements of the General Monograph for Unlicensed Medicines and, where applicable, the requirements of the individual monograph for the specific dosage form. While it is expected that extemporaneous preparations will demonstrate pharmacopoeial compliance when tested, it is recognised that it might not be practicable to carry out the pharmacopoeial tests routinely on such formulations. In the event of doubt or dispute, the methods of analysis, the reference materials and the reference spectra of the Pharmacopoeia are alone authoritative.

Part III

Monographs and other texts of the European Pharmacopoeia that are incorporated in this edition of the British Pharmacopoeia are governed by the general notices of the European Pharmacopoeia; these are reproduced below.

GENERAL NOTICES OF THE EUROPEAN PHARMACOPOEIA

1.1. GENERAL STATEMENTS

The General Notices apply to all monographs and other texts of the European Pharmacopoeia.

The official texts of the European Pharmacopoeia are published in English and French. Translations in other languages may be prepared by the signatory States of the European Pharmacopoeia Convention. In case of doubt or dispute, the English and French versions are alone authoritative.

In the texts of the European Pharmacopoeia, the word 'Pharmacopoeia' without qualification means the European Pharmacopoeia. The official abbreviation Ph. Eur. may be used to indicate the European Pharmacopoeia.

The use of the title or the subtitle of a monograph implies that the article complies with the requirements of the relevant monograph. Such references to monographs in the texts of the Pharmacopoeia are shown using the monograph title and reference number in *italics*.

A preparation must comply throughout its period of validity; a distinct period of validity and/or specifications for opened or broached containers may be decided by the competent authority. The subject of any other monograph must comply throughout its period of use. The period of validity that is assigned to any given article and the time from which that period is to be calculated are decided by the competent authority in light of experimental results of stability studies.

Unless otherwise indicated in the General Notices or in the monographs, statements in monographs constitute mandatory requirements. General chapters become mandatory when referred to in a monograph, unless such reference is made in a way that indicates that it is not the intention to make the text referred to mandatory but rather to cite it for information.

The active substances, excipients, pharmaceutical preparations and other articles described in the monographs are intended for human and veterinary use (unless explicitly restricted to one of these uses).

Quality systems The quality standards represented by monographs are valid only where the articles in question are produced within the framework of a suitable quality system. The quality system must assure that the articles consistently meet the requirements of the Pharmacopoeia.

Alternative methods The tests and assays described are the official methods upon which the standards of the Pharmacopoeia are based. With the agreement of the competent authority, alternative methods of analysis may be used for control purposes, provided that the methods used enable an unequivocal decision to be made as to whether compliance with the standards of the

monographs would be achieved if the official methods were used. In the event of doubt or dispute, the methods of analysis of the Pharmacopoeia are alone authoritative.

Demonstration of compliance with the Pharmacopoeia

(1) An article is not of Pharmacopoeia quality unless it complies with all the requirements stated in the monograph. This does not imply that performance of all the tests in a monograph is necessarily a prerequisite for a manufacturer in assessing compliance with the Pharmacopoeia before release of a product. The manufacturer may obtain assurance that a product is of Pharmacopoeia quality on the basis of its design, together with its control strategy and data derived, for example, from validation studies of the manufacturing process.

(2) An enhanced approach to quality control could utilise process analytical technology (PAT) and/or real-time release testing (including parametric release) strategies as alternatives to end-product testing alone. Real-time release testing in circumstances deemed appropriate by the competent authority is thus not precluded by the need to comply with the Pharmacopoeia.

(3) Reduction of animal testing: the European Pharmacopoeia is dedicated to phasing out the use of animals for test purposes, in accordance with the 3Rs (Replacement, Reduction, Refinement) set out in the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes. In demonstrating compliance with the Pharmacopoeia as indicated above (1), manufacturers may consider establishing additional systems to monitor consistency of production. With the agreement of the competent authority, the choice of tests performed to assess compliance with the Pharmacopoeia when animal tests are prescribed is established in such a way that animal usage is minimised as much as possible.

Grade of materials

Certain materials that are the subject of a pharmacopoeial monograph may exist in different grades suitable for different purposes. Unless otherwise indicated in the monograph, the requirements apply to all grades of the material. In some monographs, particularly those on excipients, a list of functionality-related characteristics that are relevant to the use of the substance may be appended to the monograph for information. Test methods for determination of one or more of these characteristics may be given, also for information.

General monographs

Substances and preparations that are the subject of an individual monograph are also required to comply with relevant, applicable general monographs. Cross-references to applicable general monographs are not normally given in individual monographs.

General monographs apply to all substances and preparations within the scope of the Definition section of the general monograph, except where a preamble limits the application, for example to substances and preparations that are the subject of a monograph of the Pharmacopoeia.

General monographs on dosage forms apply to all preparations of the type defined. The requirements are not necessarily comprehensive for a given specific preparation and requirements additional to those prescribed in the general monograph may be imposed by the competent authority.

General monographs and individual monographs are complementary. If the provisions of a general monograph do not apply to a particular product, this is expressly stated in the individual monograph.

Validation of pharmacopoeial methods The test methods given in monographs and general chapters have been validated in accordance with accepted scientific practice and current recommendations on analytical validation. Unless otherwise stated in the monograph or general chapter, validation of the test methods by the analyst is not required.

Implementation of pharmacopoeial methods When implementing a pharmacopoeial method, the user must assess whether and to what extent the suitability of the method under the actual conditions of use needs to be demonstrated according to relevant monographs, general chapters and quality systems.

Conventional terms The term 'competent authority' means the national, supranational or international body or organisation vested with the authority for making decisions concerning the issue in question. It may, for example, be a national pharmacopoeia authority, a licensing authority or an official control laboratory.

The expression 'unless otherwise justified and authorised' means that the requirements have to be met, unless the competent authority authorises a modification or an exemption where justified in a particular case.

Statements containing the word 'should' are informative or advisory.

In certain monographs or other texts, the terms 'suitable' and 'appropriate' are used to describe a reagent, micro-organism, test method etc.; if criteria for suitability are not described in the monograph, suitability is demonstrated to the satisfaction of the competent authority.

Medicinal product (a) Any substance or combination of substances presented as having properties for treating or preventing disease in human beings and/or animals; or (b) any substance or combination of substances that may be used in or administered to human beings and/or animals with a view either to restoring, correcting or modifying physiological functions by exerting a pharmacological, immunological or metabolic action, or to making a medical diagnosis.

Herbal medicinal product Any medicinal product, exclusively containing as active ingredients one or more herbal drugs or one or more herbal drug preparations, or one or more such herbal drugs in combination with one or more such herbal drug preparations.

Active substance Any substance intended to be used in the manufacture of a medicinal product and that, when so used, becomes an active ingredient of the medicinal product. Such substances are intended to furnish a pharmacological activity or other direct effect in the diagnosis, cure, mitigation, treatment or prevention of disease, or to affect the structure and function of the body.

Excipient (auxiliary substance). Any constituent of a medicinal product that is not an active substance. Adjuvants, stabilisers, antimicrobial preservatives, diluents, antioxidants, for example, are excipients.

Interchangeable methods Certain general chapters contain a statement that the text in question is harmonised with the corresponding text of the Japanese Pharmacopoeia and/or the United States Pharmacopeia and that these texts are interchangeable. This implies that if a substance or preparation is found to

comply with a requirement using an interchangeable method from one of these pharmacopoeias it complies with the requirements of the European Pharmacopoeia. In the event of doubt or dispute, the text of the European Pharmacopoeia is alone authoritative.

References to regulatory documents

Monographs and general chapters may contain references to documents issued by regulatory authorities for medicines, for example directives and notes for guidance of the European Union. These references are provided for information for users for the Pharmacopoeia. Inclusion of such a reference does not modify the status of the documents referred to, which may be mandatory or for guidance.

1.2. OTHER PROVISIONS APPLYING TO GENERAL CHAPTERS AND MONOGRAPHS

Quantities

In tests with numerical limits and assays, the quantity stated to be taken for examination is approximate. The amount actually used, which may deviate by not more than 10 per cent from that stated, is accurately weighed or measured and the result is calculated from this exact quantity. In tests where the limit is not numerical, but usually depends upon comparison with the behaviour of a reference substance in the same conditions, the stated quantity is taken for examination. Reagents are used in the prescribed amounts.

Quantities are weighed or measured with an accuracy commensurate with the indicated degree of precision. For weighings, the precision corresponds to plus or minus 5 units after the last figure stated (for example, 0.25 g is to be interpreted as 0.245 g to 0.255 g). For the measurement of volumes, if the figure after the decimal point is a zero or ends in a zero (for example, 10.0 mL or 0.50 mL), the volume is measured using a pipette, a volumetric flask or a burette, as appropriate; otherwise, a graduated measuring cylinder or a graduated pipette may be used. Volumes stated in microlitres are measured using a micropipette or microsyringe.

It is recognised, however, that in certain cases the precision with which quantities are stated does not correspond to the number of significant figures stated in a specified numerical limit. The weighings and measurements are then carried out with a sufficiently improved accuracy.

Apparatus and procedures

Volumetric glassware complies with Class A requirements of the appropriate International Standard issued by the International Organisation for Standardisation.

Unless otherwise prescribed, analytical procedures are carried out at a temperature between 15 °C and 25 °C.

Unless otherwise prescribed, comparative tests are carried out using identical tubes of colourless, transparent, neutral glass with a flat base; the volumes of liquid prescribed are for use with tubes having an internal diameter of 16 mm, but tubes with a larger internal diameter may be used provided the volume of liquid used is adjusted (*2.1.5*). Equal volumes of the liquids to be compared are examined down the vertical axis of the tubes against a white background, or if necessary against a black background. The examination is carried out in diffuse light.

Any solvent required in a test or assay in which an indicator is to be used is previously neutralised to the indicator, unless a blank test is prescribed.

Water-bath The term 'water-bath' means a bath of boiling water unless water at another temperature is indicated. Other methods of heating may be substituted provided the temperature is near to but not higher than 100 °C or the indicated temperature.

Drying and ignition to constant mass The terms 'dried to constant mass' and 'ignited to constant mass' mean that 2 consecutive weighings do not differ by more than 0.5 mg, the 2nd weighing following an additional period of drying or of ignition respectively appropriate to the nature and quantity of the residue.

Where drying is prescribed using one of the expressions 'in a desiccator' or '*in vacuo*', it is carried out using the conditions described in chapter *2.2.32. Loss on drying*.

Reagents The proper conduct of the analytical procedures described in the Pharmacopoeia and the reliability of the results depend, in part, upon the quality of the reagents used. The reagents are described in general chapter 4. It is assumed that reagents of analytical grade are used; for some reagents, tests to determine suitability are included in the specifications.

Solvents Where the name of the solvent is not stated, the term 'solution' implies a solution in water.

Where the use of water is specified or implied in the analytical procedures described in the Pharmacopoeia or for the preparation of reagents, water complying with the requirements of the monograph *Purified water (0008)* is used, except that for many purposes the requirements for bacterial endotoxins (*Purified water in bulk*) and microbial contamination (*Purified water in containers*) are not relevant. The term 'distilled water' indicates purified water prepared by distillation.

The term 'ethanol' without qualification means anhydrous ethanol. The term 'alcohol' without qualification means ethanol (96 per cent). Other dilutions of ethanol are indicated by the term 'ethanol' or 'alcohol' followed by a statement of the percentage by volume of ethanol (C_2H_6O) required.

Expression of content In defining content, the expression 'per cent' is used according to circumstances with one of 2 meanings:

— per cent *m/m* (percentage, mass in mass) expresses the number of grams of substance in 100 g of final product;

— per cent *V/V* (percentage, volume in volume) expresses the number of millilitres of substance in 100 mL of final product.

The expression 'parts per million' (or ppm) refers to mass in mass, unless otherwise specified.

Temperature Where an analytical procedure describes temperature without a figure, the general terms used have the following meaning:

— in a deep-freeze: below −15 °C;

— in a refrigerator: 2 °C to 8 °C;

— cold or cool: 8 °C to 15 °C;

— room temperature: 15 °C to 25 °C.

1.3. GENERAL CHAPTERS

Containers Materials used for containers are described in general chapter *3.1*. General names used for materials, particularly plastic materials, each cover a range

of products varying not only in the properties of the principal constituent but also in the additives used. The test methods and limits for materials depend on the formulation and are therefore applicable only for materials whose formulation is covered by the preamble to the specification. The use of materials with different formulations, and the test methods and limits applied to them, are subject to agreement by the competent authority.

The specifications for containers in general chapter *3.2* have been developed for general application to containers of the stated category, but in view of the wide variety of containers available and possible new developments, the publication of a specification does not exclude the use, in justified circumstances, of containers that comply with other specifications, subject to agreement by the competent authority.

Reference may be made within the monographs of the Pharmacopoeia to the definitions and specifications for containers provided in chapter *3.2. Containers*. The general monographs for pharmaceutical dosage forms may, under the heading Definition/Production, require the use of certain types of container; certain other monographs may, under the heading Storage, indicate the type of container that is recommended for use.

1.4. MONOGRAPHS

Titles Monograph titles are in English and French in the respective versions and there is a Latin subtitle.

Relative Atomic And Molecular Masses The relative atomic mass (A_r) or the relative molecular mass (M_r) is shown, as and where appropriate, at the beginning of each monograph. The relative atomic and molecular masses and the molecular and graphic formulae do not constitute analytical standards for the substances described.

Chemical Abstracts Service (CAS) Registry Number CAS registry numbers are included for information in monographs, where applicable, to provide convenient access to useful information for users. CAS Registry Number® is a registered trademark of the American Chemical Society.

Definition Statements under the heading Definition constitute an official definition of the substance, preparation or other article that is the subject of the monograph.

Limits of content Where limits of content are prescribed, they are those determined by the method described under Assay.

Herbal drugs In monographs on herbal drugs, the definition indicates whether the subject of the monograph is, for example, the whole drug or the drug in powdered form. Where a monograph applies to the drug in several states, for example both to the whole drug and the drug in powdered form, the definition states this.

Production Statements under the heading Production draw attention to particular aspects of the manufacturing process but are not necessarily comprehensive. They constitute mandatory requirements for manufacturers, unless otherwise stated. They may relate, for example, to source materials; to the manufacturing process itself and its validation and control; to in-process testing; or to testing that is to be carried out by the manufacturer on the final article, either on selected batches or on each batch prior to release. These statements cannot necessarily be verified on a sample of the final article by an independent analyst. The competent authority may establish

that the instructions have been followed, for example, by examination of data received from the manufacturer, by inspection of manufacture or by testing appropriate samples.

The absence of a Production section does not imply that attention to features such as those referred to above is not required.

Choice of vaccine strain, Choice of vaccine composition The Production section of a monograph may define the characteristics of a vaccine strain or vaccine composition. Unless otherwise stated, test methods given for verification of these characteristics are provided for information as examples of suitable methods. Subject to approval by the competent authority, other test methods may be used without validation against the method shown in the monograph.

Potential Adulteration Due to the increasing number of fraudulent activities and cases of adulteration, information may be made available to Ph. Eur. users to help detect adulterated materials (i.e. active substances, excipients, intermediate products, bulk products and finished products).

To this purpose, a method for the detection of potential adulterants and relevant limits, together with a reminder that all stages of production and sourcing are subjected to a suitable quality system, may be included in this section of monographs on substances for which an incident has occurred or that present a risk of deliberate contamination. The frequency of testing by manufacturers or by users (e.g. manufacturers of intermediate products, bulk products and finished products, where relevant) depends on a risk assessment, taking into account the level of knowledge of the whole supply chain and national requirements.

This section constitutes requirements for the whole supply chain, from manufacturers to users (e.g. manufacturers of intermediate products, bulk products and finished products, where relevant). The absence of this section does not imply that attention to features such as those referred to above is not required.

Characters The statements under the heading Characters are not to be interpreted in a strict sense and are not requirements.

Solubility In statements of solubility in the Characters section, the terms used have the following significance, referred to a temperature between 15 °C and 25 °C.

Descriptive term	Approximate volume of solvent in millilitres per gram of solute			
Very soluble	less than	1		
Freely soluble	from	1	to	10
Soluble	from	10	to	30
Sparingly soluble	from	30	to	100
Slightly soluble	from	100	to	1000
Very slightly soluble	from	1000	to	10 000
Practically insoluble	more than			10 000

The term 'partly soluble' is used to describe a mixture where only some of the components dissolve. The term 'miscible' is used to describe a liquid that is miscible in all proportions with the stated solvent.

Identification *Scope* The tests given in the Identification section are not designed to give a full confirmation of the chemical structure or composition of the product; they are intended to give confirmation, with an acceptable degree of assurance, that the article conforms to the description on the label.

First and second identifications Certain monographs have subdivisions entitled 'First identification' and 'Second identification'. The test or tests that constitute the 'First identification' may be used in all circumstances. The test or tests that constitute the 'Second identification' may be used in pharmacies provided it can be demonstrated that the substance or preparation is fully traceable to a batch certified to comply with all the other requirements of the monograph.

Certain monographs give two or more sets of tests for the purpose of the first identification, which are equivalent and may be used independently. One or more of these sets usually contain a cross-reference to a test prescribed in the Tests section of the monograph. It may be used to simplify the work of the analyst carrying out the identification and the prescribed tests. For example, one identification set cross-refers to a test for enantiomeric purity while the other set gives a test for specific optical rotation: the intended purpose of the two is the same, that is, verification that the correct enantiomer is present.

Powdered herbal drugs Monographs on herbal drugs may contain schematic drawings of the powdered drug. These drawings complement the description given in the relevant identification test.

Tests And Assays *Scope* The requirements are not framed to take account of all possible impurities. It is not to be presumed, for example, that an impurity that is not detectable by means of the prescribed tests is tolerated if common sense and good pharmaceutical practice require that it be absent. See also below under Impurities.

Calculation Where the result of a test or assay is required to be calculated with reference to the dried or anhydrous substance or on some other specified basis, the determination of loss on drying, water content or other property is carried out by the method prescribed in the relevant test in the monograph. The words 'dried substance' or 'anhydrous substance' etc. appear in parentheses after the result.Where a quantitative determination of a residual solvent is carried out and a test for loss on drying is not carried out, the content of residual solvent is taken into account for the calculation of the assay content of the substance, the specific optical rotation and the specific absorbance. No further indication is given in the specific monograph.

Limits The limits prescribed are based on data obtained in normal analytical practice; they take account of normal analytical errors, of acceptable variations in manufacture and compounding and of deterioration to an extent considered acceptable. No further tolerances are to be applied to the limits prescribed to determine whether the article being examined complies with the requirements of the monograph.

In determining compliance with a numerical limit, the calculated result of a test or assay is first rounded to the number of significant figures stated, unless otherwise prescribed. The limits, regardless of whether the values are expressed as percentages or as absolute values, are considered significant to the last digit shown (for example 140 indicates 3 significant figures). The last figure of the result is increased by one when the part rejected is equal to

or exceeds one half-unit, whereas it is not modified when the part rejected is less than a half-unit.

Indication of permitted limit of impurities The acceptance criteria for related substances are expressed in monographs either in terms of comparison of peak areas (comparative tests) or as numerical values. For comparative tests, the approximate content of impurity tolerated, or the sum of impurities, may be indicated in brackets for information only. Acceptance or rejection is determined on the basis of compliance or non-compliance with the stated test. If the use of a reference substance for the named impurity is not prescribed, this content may be expressed as a nominal concentration of the substance used to prepare the reference solution specified in the monograph, unless otherwise described.

Herbal Drugs For herbal drugs, the sulfated ash, total ash, water-soluble matter, alcohol-soluble matter, water content, content of essential oil and content of active principle are calculated with reference to the drug that has not been specially dried, unless otherwise prescribed in the monograph.

Equivalents Where an equivalent is given, for the purposes of the Pharmacopoeia only the figures shown are to be used in applying the requirements of the monograph.

Culture media The culture media described in monographs and general chapters have been found to be satisfactory for the intended purpose. However, the components of media, particularly those of biological origin, are of variable quality, and it may be necessary for optimal performance to modulate the concentration of some ingredients, notably:

— peptones and meat or yeast extracts, with respect to their nutritive properties;
— buffering substances;
— bile salts, bile extract, deoxycholate, and colouring matter, depending on their selective properties;
— antibiotics, with respect to their activity.

Storage The information and recommendations given under the heading Storage do not constitute a pharmacopoeial requirement but the competent authority may specify particular storage conditions that must be met.

The articles described in the Pharmacopoeia are stored in such a way as to prevent contamination and, as far as possible, deterioration. Where special conditions of storage are recommended, including the type of container (see section 1.3. General chapters) and limits of temperature, they are stated in the monograph.

The following expressions are used in monographs under Storage with the meaning shown.

In an airtight container Means that the product is stored in an airtight container (3.2). Care is to be taken when the container is opened in a damp atmosphere. A low moisture content may be maintained, if necessary, by the use of a desiccant in the container provided that direct contact with the product is avoided.

Protected from light Means that the product is stored either in a container made of a material that absorbs actinic light sufficiently to protect the contents from change induced by such light, or in a container enclosed in an outer cover that provides such protection, or is stored in a place from which all such light is excluded.

Labelling In general, labelling of medicines is subject to supranational and national regulation and to international agreements. The statements under the heading Labelling are not therefore comprehensive and, moreover, for the purposes of the Pharmacopoeia only those statements that are necessary to demonstrate compliance or non-compliance with the monograph are mandatory. Any other labelling statements are included as recommendations. When the term 'label' is used in the Pharmacopoeia, the labelling statements may appear on the container, the package, a leaflet accompanying the package, or a certificate of analysis accompanying the article, as decided by the competent authority.

Warnings Materials described in monographs and reagents specified for use in the Pharmacopoeia may be injurious to health unless adequate precautions are taken. The principles of good quality control laboratory practice and the provisions of any appropriate regulations are to be observed at all times. Attention is drawn to particular hazards in certain monographs by means of a warning statement; absence of such a statement is not to be taken to mean that no hazard exists.

Impurities A list of all known and potential impurities that have been shown to be detected by the tests in a monograph may be given. See also chapter *5.10. Control of impurities in substances for pharmaceutical use*. The impurities are designated by a letter or letters of the alphabet. Where a letter appears to be missing, the impurity designated by this letter has been deleted from the list during monograph development prior to publication or during monograph revision.

Functionality-related Characteristics of Excipients Monographs on excipients may have a section on functionality-related characteristics. The characteristics, any test methods for determination and any tolerances are not mandatory requirements; they may nevertheless be relevant for use of the excipient and are given for information (see also section 1.1. General statements).

Reference Standards Certain monographs require the use of reference standards (chemical reference substances, herbal reference standards, biological reference preparations, reference spectra). See also chapter *5.12. Reference standards*. The European Pharmacopoeia Commission establishes the official reference standards, which are alone authoritative in case of arbitration. These reference standards are available from the European Directorate for the Quality of Medicines & HealthCare (EDQM). Information on the available reference standards and a batch validity statement can be obtained via the EDQM website.

1.5. ABBREVIATIONS AND SYMBOLS

A	Absorbance	mp	Melting point
$A_{1\,cm}^{1\ per\ cent}$	Specific absorbance	n_D^{20}	Refractive index
A_r	Relative atomic mass	Ph. Eur. U.	European Pharmacopoeia Unit
$[\alpha]_D^{20}$	Specific optical rotation	ppb	Parts per billion (micrograms per kilogram)
bp	Boiling point	ppm	Parts per million (milligrams per kilogram)
BRP	Biological reference preparation	R	Substance or solution defined under 4. *Reagents*
CRS	Chemical reference substance	R_F	Retardation factor (see chapter *2.2.46*)
d_{20}^{20}	Relative density	R_x	Used in chromatography to indicate the ratio of the distance travelled by a substance to the distance travelled by a reference substance
λ	Wavelength		
HRS	Herbal reference standard	RV	Substance used as a primary standard in volumetric analysis (chapter *4.2.1*)
IU	International Unit		
M	Molarity		
M_r	Relative molecular mass		

Abbreviations used in the monographs on immunoglobulins, immunosera and vaccines

CFU	Colony-forming units	Lo/10 dose	The largest quantity of a toxin that, in the conditions of the test, when mixed with 0.1 IU of antitoxin and administered by the specified route, does not cause symptoms of toxicity in the test animals within a given period
LD_{50}	The statistically determined quantity of a substance that, when administered by the specified route, may be expected to cause the death of 50 per cent of the test animals within a given period	Lf dose	The quantity of toxin or toxoid that flocculates in the shortest time with 1 IU of antitoxin
MLD	Minimum lethal dose	$CCID_{50}$	The statistically determined quantity of virus that may be expected to infect 50 per cent of the cell cultures to which it is added
L+/10 dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 0.1 IU of antitoxin and administered by the specified route, causes the death of the test animals within a given period	EID_{50}	The statistically determined quantity of virus that may be expected to infect 50 per cent of the fertilised eggs into which it is inoculated
L+ dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 1 IU of antitoxin and administered by the specified route, causes the death of the test animals within a given period	ID_{50}	The statistically determined quantity of a virus that may be expected to infect 50 per cent of the animals into which it is inoculated
lr/100 dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 0.01 IU of antitoxin and injected intracutaneously causes a characteristic reaction at the site of injection within a given period	PD_{50}	The statistically determined dose of a vaccine that, in the conditions of the test, may be expected to protect 50 per cent of the animals against a challenge dose of the micro-organisms or toxins against which it is active
Lp/10 dose	The smallest quantity of toxin that, in the conditions of the test, when mixed with 0.1 IU of antitoxin and administered by the specified route, causes paralysis in the test animals within a given period	ED_{50}	The statistically determined dose of a vaccine that, in the conditions of the test, may be expected to induce specific antibodies in 50 per cent of the animals for the relevant vaccine antigens
		PFU	Pock-forming units or plaque-forming units
		SPF	Specified-pathogen-free

Collections of micro-organisms

ATCC — American Type Culture Collection
10801 University Boulevard
Manassas, Virginia 20110-2209, USA

C.I.P. — Collection de Bactéries de l'Institut Pasteur
B.P. 52, 25 rue du Docteur Roux
75724 Paris Cedex 15, France

IMI — International Mycological Institute
Bakeham Lane
Surrey TW20 9TY, Great Britain

I.P. — Collection Nationale de Culture de Microorganismes (C.N.C.M.)
Institut Pasteur
25, rue du Docteur Roux
75724 Paris Cedex 15, France

NCIMB — National Collection of Industrial and Marine Bacteria Ltd
23 St Machar Drive
Aberdeen AB2 1RY, Great Britain

NCPF — National Collection of Pathogenic Fungi
London School of Hygiene and Tropical Medicine
Keppel Street
London WC1E 7HT, Great Britain

NCTC — National Collection of Type Cultures
Central Public Health Laboratory
Colindale Avenue
London NW9 5HT, Great Britain

NCYC — National Collection of Yeast Cultures
AFRC Food Research Institute
Colney Lane
Norwich NR4 7UA, Great Britain

NITE — Biological Resource Center
Department of Biotechnology
National Institute of Technology and Evaluation
2-5-8 Kazusakamatari, Kisarazu-shi, Chiba, 292-0818
Japan

S.S.I. — Statens Serum Institut
80 Amager Boulevard, Copenhagen, Denmark

1.6. UNITS OF THE INTERNATIONAL SYSTEM (SI) USED IN THE PHARMACOPOEIA AND EQUIVALENCE WITH OTHER UNITS

International System Of Units (SI)

The International System of Units comprises 3 classes of units, namely base units, derived units and supplementary units[1]. The base units and their definitions are set out in Table 1.6-1.

The derived units may be formed by combining the base units according to the algebraic relationships linking the corresponding quantities. Some of these derived units have special names and symbols. The SI units used in the Pharmacopoeia are shown in Table 1.6-2.

Some important and widely used units outside the International System are shown in Table 1.6-3.

The prefixes shown in Table 1.6-4 are used to form the names and symbols of the decimal multiples and submultiples of SI units.

Table 1.6.-1. – *SI base units*

Quantity		Unit		Definition
Name	Symbol	Name	Symbol	
Length	*l*	metre	m	The metre is the length of the path travelled by light in a vacuum during a time interval of 1/299 792 458 of a second.
Mass	*m*	kilogram	kg	The kilogram is equal to the mass of the international prototype of the kilogram.
Time	*t*	second	s	The second is the duration of 9 192 631 770 periods of the radiation corresponding to the transition between the two hyperfine levels of the ground state of the caesium-133 atom.
Electric current	*I*	ampere	A	The ampere is that constant current which, maintained in two straight parallel conductors of infinite length, of negligible circular cross-section and placed 1 metre apart in vacuum would produce between these conductors a force equal to 2×10^{-7} newton per metre of length.
Thermodynamic temperature	*T*	kelvin	K	The kelvin is the fraction 1/273.16 of the thermodynamic temperature of the triple point of water.
Amount of substance	*n*	mole	mol	The mole is the amount of substance of a system containing as many elementary entities as there are atoms in 0.012 kilogram of carbon-12*.
Luminous intensity	I_v	candela	cd	The candela is the luminous intensity in a given direction of a source emitting monochromatic radiation with a frequency of 540×10^{12} hertz and whose energy intensity in that direction is 1/683 watt per steradian.

* When the mole is used, the elementary entities must be specified and may be atoms, molecules, ions, electrons, other particles or specified groups of such particles.

[1] *The definitions of the units used in the International System are given in the booklet 'Le Systeme International d'Unites (SI)', published by the Bureau International des Poids et Mesures, Pavillon de Breteuil, F-92310 Sevres.*

Table 1.6.-2. *SI units used in the European Pharmacopoeia and equivalence with other units*

Quantity		Unit				
Name	**Symbol**	**Name**	**Symbol**	**Expression in SI base units**	**Expression in other SI units**	**Conversion of other units into SI units**
Wave number	ν	one per metre	l/m	m^{-1}		
Wavelength	λ	micrometre	µm	10^{-6} m		
		nanometre	nm	10^{-9} m		
Area	A, S	square metre	m^2	m^2		
Volume	V	cubic metre	m^3	m^3		1 mL = 1 cm^3 = 10^{-6} m^3
Frequency	ν	hertz	Hz	s^{-1}		
Density	ρ	kilogram per cubic metre	kg/m^3	$kg \cdot m^{-3}$		1 g/mL = 1 g/cm^3 = 10^3 $kg \cdot m^{-3}$
Velocity	v	metre per second	m/s	$m \cdot s^{-1}$		
Force	F	newton	N	$m \cdot kg \cdot s^{-2}$		1 dyne = 1 $g \cdot cm \cdot s^{-2}$ = 10^{-5} N 1 kp = 9.806 65 N
Pressure	p	pascal	Pa	$m^{-1} \cdot kg \cdot s^{-2}$	$N \cdot m^{-2}$	1 dyne/cm^2 = 10^{-1} Pa = 10^{-1} $N \cdot m^{-2}$ 1 atm = 101 325 Pa = 101.325 kPa 1 bar = 10^5 Pa = 0.1 MPa 1 mm Hg = 133.322 387 Pa 1 Torr = 133.322 368 Pa 1 psi = 6.894 757 kPa
Dynamic viscosity	η	pascal second	Pa·s	$m^{-1} \cdot kg \cdot s^{-1}$	$N \cdot s \cdot m^{-2}$	1 P = 10^{-1} Pa·s = 10^{-1} $N \cdot s \cdot m^{-2}$ 1 cP = 1 mPa·s
Kinematic viscosity	ν	square metre per second	m^2/s	$m^2 \cdot s^{-1}$	$Pa \cdot s \cdot m^3 \cdot kg^{-1}$ $N \cdot m \cdot s \cdot kg^{-1}$	1 St = 1 $cm^2 \cdot s^{-1}$ = 10^{-4} $m^2 \cdot s^{-1}$
Energy	W	joule	J	$m^2 \cdot kg \cdot s^{-2}$	N·m	1 erg = 1 $cm^2 \cdot g \cdot s^{-2}$ = 1 dyne·cm = 10^{-7} J 1 cal = 4.1868 J
Power Radiant flux	P	watt	W	$m^2 \cdot kg \cdot s^{-3}$	$N \cdot m \cdot s^{-1}$ $J \cdot s^{-1}$	1 erg/s = 1 dyne·cm·s^{-1} = 10^{-7} W = 10^{-7} $N \cdot m \cdot s^{-1}$ = 10^{-7} $J \cdot s^{-1}$
Absorbed dose (of radiant energy)	D	gray	Gy	$m^2 \cdot s^{-2}$	$J \cdot kg^{-1}$	1 rad = 10^{-2} Gy
Electric potential, electromotive force	U	volt	V	$m^2 \cdot kg \cdot s^{-3} \cdot A^{-1}$	$W \cdot A^{-1}$	
Electric resistance	R	ohm	Ω	$m^2 \cdot kg \cdot s^{-3} \cdot A^{-2}$	$V \cdot A^{-1}$	
Amount of electricity	Q	coulomb	C	A·s		
Activity of a radionuclide	A	becquerel	Bq	s^{-1}		1 Ci = $37 \cdot 10^9$ Bq = $37 \cdot 10^9$ s^{-1}

Quantity		Unit				
Name	Symbol	Name	Symbol	Expression in SI base units	Expression in other SI units	Conversion of other units into SI units
Concentration (of amount of substance), molar concentration	c	mole per cubic metre	mol/m^3	$mol \cdot m^{-3}$		$1\ mol/L = 1\ M = 1\ mol/dm^3 = 10^3\ mol \cdot m^{-3}$
Mass concentration	ρ	kilogram per cubic metre	kg/m^3	$kg \cdot m^{-3}$		$1\ g/L = 1\ g/dm^3 = 1\ kg \cdot m^{-3}$

Table 1.6.-3. – *Units used with the International System*

Quantity	Unit		Value in SI units
	Name	Symbol	
Time	minute	min	1 min = 60 s
	hour	h	1 h = 60 min = 3600 s
	day	d	1 d = 24 h = 86 400 s
Plane angle	degree	°	$1° = (\pi/180)$ rad
Volume	litre	L	$1\ L = 1\ dm^3 = 10^{-3}\ m^3$
Mass	tonne	t	$1\ t = 10^3\ kg$
Rotational frequency	revolution per minute	r/min	$1\ r/min = (1/60)\ s^{-1}$

Table 1.6.-4. – *Decimal multiples and sub-multiples of units*

Factor	Prefix	Symbol	Factor	Prefix	Symbol
10^{18}	exa	E	10^{-1}	deci	d
10^{15}	peta	P	10^{-2}	centi	c
10^{12}	tera	T	10^{-3}	milli	m
10^{9}	giga	G	10^{-6}	micro	μ
10^{6}	mega	M	10^{-9}	nano	n
10^{3}	kilo	k	10^{-12}	pico	p
10^{2}	hecto	h	10^{-15}	femto	f
10^{1}	deca	da	10^{-18}	atto	a

Notes

1. In the Pharmacopoeia, the Celsius temperature is used (symbol t). This is defined by the following equation:

$$t = T - T_0$$

where T_0 = 273.15 K by definition. The Celsius or centigrade temperature is expressed in degrees Celsius (symbol °C). The unit 'degree Celsius' is equal to the unit 'kelvin'.

2. The practical expressions of concentrations used in the Pharmacopoeia are defined in the General Notices.

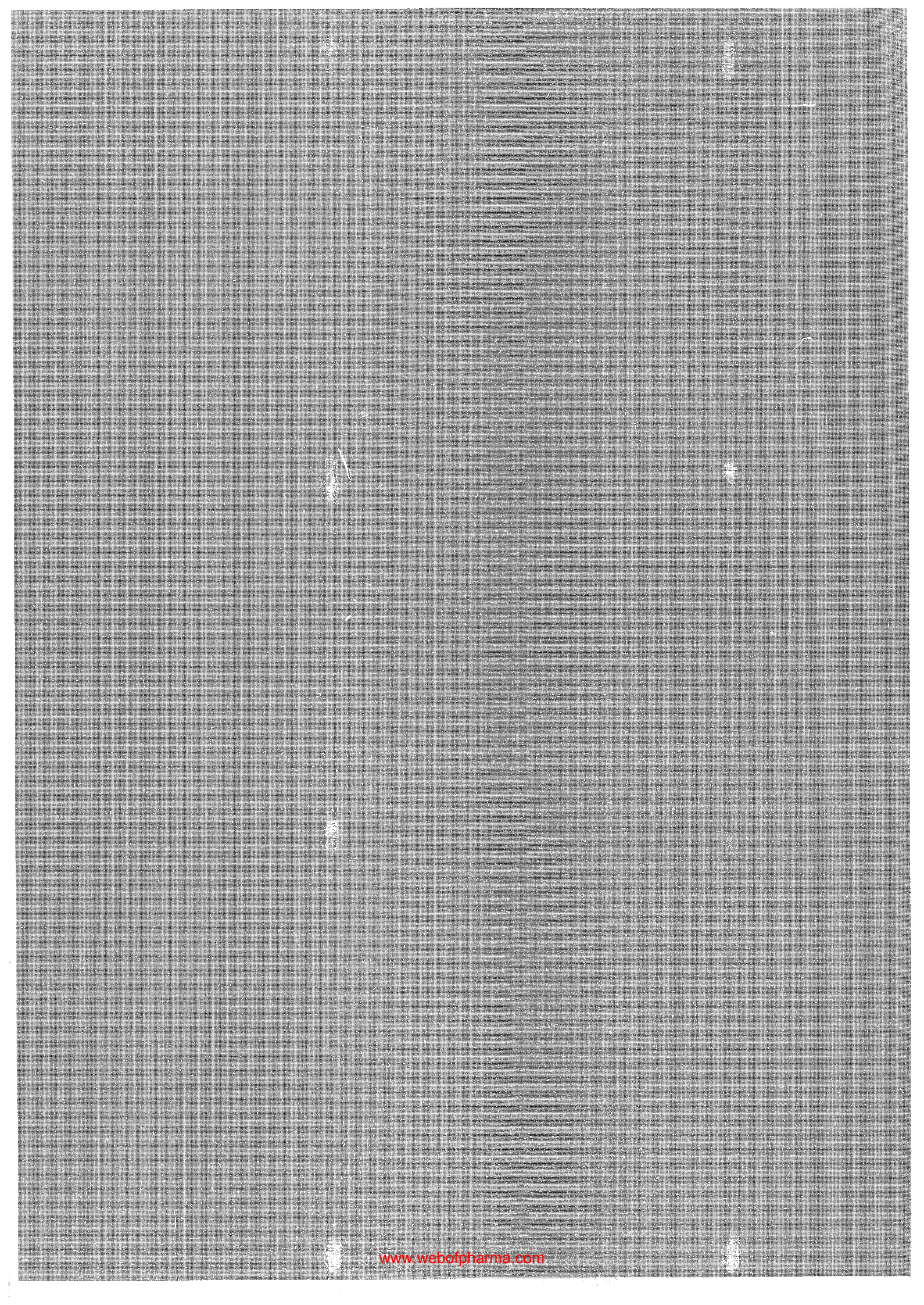
3. The radian is the plane angle between two radii of a circle that cut off on the circumference an arc equal in length to the radius.

4. In the Pharmacopoeia, conditions of centrifugation are defined by reference to the acceleration due to gravity (g):

$$g = 9.806\ 65\ m{\cdot}s^{-2}$$

5. Certain quantities without dimensions are used in the Pharmacopoeia: relative density (*2.2.5*), absorbance (*2.2.25*), specific absorbance (*2.2.25*) and refractive index (*2.2.6*).

6. The microkatal is defined as the enzymic activity that, under defined conditions, produces the transformation (e.g. hydrolysis) of 1 micromole of the substrate per second.



Monographs

Medicinal and Pharmaceutical Substances (A to I)

MEDICINAL AND PHARMACEUTICAL SUBSTANCES

Substances for Pharmaceutical Use

(Ph. Eur. monograph 2034)

Ph Eur

DEFINITION

Substances for pharmaceutical use are any organic or inorganic substances that are used as active substances or excipients for the production of medicinal products for human or veterinary use. They may be obtained from natural sources or produced by extraction from raw materials, fermentation or synthesis.

This general monograph does not apply to herbal drugs, herbal drugs for homoeopathic preparations, herbal drug preparations, herbal drug extracts, or mother tinctures for homoeopathic preparations, which are the subject of separate general monographs (*Herbal drugs (1433)*, *Herbal drugs for homoeopathic preparations (2045)*, *Herbal drug preparations (1434)*, *Herbal drug extracts (0765)*, *Mother tinctures for homoeopathic preparations (2029)*). It does not apply to raw materials for homoeopathic preparations, except where there is an individual monograph for the substance in the non-homoeopathic part of the Pharmacopoeia.

This monograph does not apply to chemical precursors for radiopharmaceutical preparations which are the subject of a separate monograph (*Chemical precursors for radiopharmaceutical preparations (2902)*).

Where a substance for pharmaceutical use not described in an individual monograph of the Pharmacopoeia is used in a medicinal product prepared for the special needs of individual patients, the need for compliance with the present general monograph is decided in the light of a risk assessment that takes account of the available quality of the substance and its intended use.

Where medicinal products are manufactured using substances for pharmaceutical use of human or animal origin, the requirements of chapter *5.1.7. Viral safety* apply.

Substances for pharmaceutical use may be used as such or as starting materials for subsequent formulation to prepare medicinal products. Depending on the formulation, certain substances may be used either as active substances or as excipients. Solid substances may be compacted, coated, granulated, powdered to a certain fineness, or processed in other ways. A monograph is applicable to a substance processed with an excipient only where such processing is mentioned in the definition section of the monograph.

Substance for pharmaceutical use of special grade Unless otherwise indicated or restricted in the individual monographs, a substance for pharmaceutical use is intended for human and veterinary use, and is of appropriate quality for the manufacture of all dosage forms in which it can be used.

Polymorphism Individual monographs do not usually specify crystalline or amorphous forms, unless bioavailability is affected. All forms of a substance for pharmaceutical use comply with the requirements of the monograph, unless otherwise indicated.

PRODUCTION

Substances for pharmaceutical use are manufactured by procedures that are designed to ensure a consistent quality and comply with the requirements of the individual monograph or approved specification.

The manufacture of active substances must take place under conditions of good manufacturing practice.

The provisions of general chapter *5.10* apply to the control of impurities in substances for pharmaceutical use.

Whether or not it is specifically stated in the individual monograph that the substance for pharmaceutical use:

— is a recombinant protein or another substance obtained as a direct gene product based on genetic modification, where applicable, the substance also complies with the requirements of the general monograph *Products of recombinant DNA technology (0784)*;

— is obtained from animals susceptible to transmissible spongiform encephalopathies other than by experimental challenge, where applicable, the substance also complies with the requirements of the general monograph *Products with risk of transmitting agents of animal spongiform encephalopathies (1483)*;

— is a substance derived from a fermentation process, whether or not the micro-organisms involved are modified by traditional procedures or recombinant DNA (rDNA) technology, where applicable, the substance also complies with the requirements of the general monograph *Products of fermentation (1468)*.

If solvents are used during production, they are of suitable quality. In addition, their toxicity and their residual level are taken into consideration (*5.4*). If water is used during production, it is of suitable quality.

The identity of elemental impurities derived from intentionally added catalysts and reagents is known, and strategies for controlling them should be established using the principles of risk management.

If substances are produced or processed to yield a certain form or grade, that specific form or grade of the substance complies with the requirements of the monograph. Certain functionality-related tests may be described to control properties that may influence the suitability of the substance and subsequently the properties of dosage forms prepared from it.

Powdered substances May be processed to obtain a certain degree of fineness (*2.9.35*).

Compacted substances Are processed to increase the particle size or to obtain particles of a specific form and/or to obtain a substance with a higher bulk density.

Coated active substances Consist of particles of the active substance coated with one or more suitable excipients.

Granulated active substances Are particles of a specified size and/or form produced from the active substance by granulation directly or with one or more suitable excipients.

If substances are processed with excipients, these excipients comply with the requirements of the relevant monograph or, where no such monograph exists, the approved specification.

Where active substances have been processed with excipients to produce, for example, coated or granulated substances, the processing is carried out under conditions of good manufacturing practice and the processed substances are regarded as intermediates in the manufacture of a medicinal product.

accordance with the recommendations of general chapter *5.1.10. Guidelines for using the test for bacterial endotoxins.*

Pyrogens (*2.6.8*)
If the test for pyrogens is justified rather than the test for bacterial endotoxins and if a pyrogen-free grade is offered, the substance for pharmaceutical use complies with the test for pyrogens. The limit and test method are stated in the individual monograph or approved by the competent authority. Based on appropriate test validation for bacterial endotoxins and pyrogens, the test for bacterial endotoxins may replace the test for pyrogens.

Additional properties
Control of additional properties (e.g. physical characteristics, functionality-related characteristics) may be necessary for individual manufacturing processes or formulations. Grades (such as sterile, endotoxin-free, pyrogen-free) may be produced with a view to manufacture of preparations for parenteral administration or other dosage forms and appropriate requirements may be specified in an individual monograph.

ASSAY
Unless justified and authorised, contents of substances for pharmaceutical use are determined. Suitable methods are used.

LABELLING
In general, labelling is subject to supranational and national regulation and to international agreements. The statements under the heading Labelling therefore are not comprehensive and, moreover, for the purposes of the Pharmacopoeia only those statements that are necessary to demonstrate compliance or non-compliance with the monograph are mandatory. Any other labelling statements are included as recommendations. When the term 'label' is used in the Pharmacopoeia, the labelling statements may appear on the container, the package, a leaflet accompanying the package or a certificate of analysis accompanying the article, as decided by the competent authority.

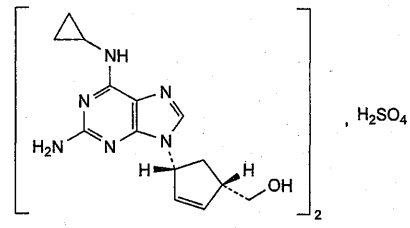
Where appropriate, the label states that the substance is:
— intended for a specific use;
— of a distinct crystalline form;
— of a specific degree of fineness;
— compacted;
— coated;
— granulated;
— sterile;
— free from bacterial endotoxins;
— free from pyrogens;
— containing gliding agents.

Where applicable, the label states:
— the degree of hydration;
— the name and concentration of any excipient.

Ph Eur

Abacavir Sulfate

(Ph. Eur. monograph 2589)



$C_{28}H_{38}N_{12}O_6S$ 671 *188062-50-2*

Action and use
Nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Preparations
Abacavir Oral Solution
Abacavir Tablets
Abacavir, Zidovudine and Lamivudine Tablets
Abacavir and Lamivudine Tablets

Ph Eur

DEFINITION
Bis[[(1*S*,4*R*)-4-[2-amino-6-(cyclopropylamino)-9*H*-purin-9-yl]cyclopent-2-enyl]methanol] sulfate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

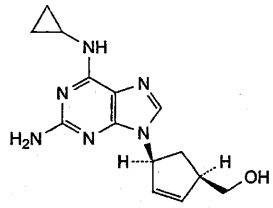
CHARACTERS
Appearance
White or almost white powder.

Solubility
Soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

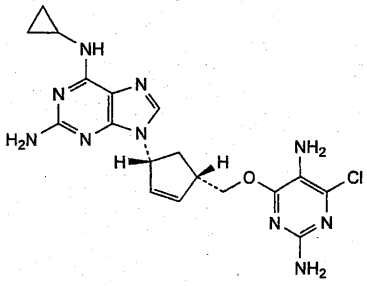
IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *abacavir sulfate CRS.*
B. Enantiomeric purity (see Tests).
C. Solution S (see Tests) gives reaction (a) of sulfates (*2.3.1*).

TESTS
Solution S
Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

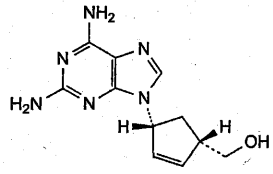
Enantiomeric purity
Liquid chromatography (*2.2.29*).
Solution A Mix 0.5 mL of *trifluoroacetic acid R* and 100 mL of *methanol R.*
Solution B Mix 30 volumes of *methanol R*, 30 volumes of *2-propanol R* and 40 volumes of *heptane R.*
Test solution Dissolve 40 mg of the substance to be examined in 30 mL of solution A. Sonicate until dissolution is complete. Add 30 mL of *2-propanol R* and dilute to 100.0 mL with *heptane R.*
Reference solution (a) Dissolve 2 mg of *abacavir for system suitability CRS* (containing impurities A and D) in 1.5 mL of solution A. Sonicate until dissolution is complete. Add 1.5 mL of *2-propanol R* and dilute to 5.0 mL with *heptane R.*



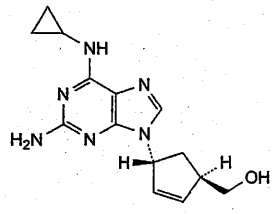
A. [(1*R*,4*S*)-4-[2-amino-6-(cyclopropylamino)-9*H*-purin-9-yl] cyclopent-2-enyl]methanol,



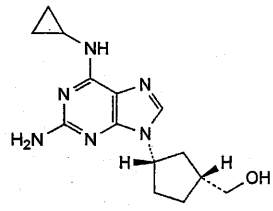
B. 6-(cyclopropylamino)-9-[(1*R*,4*S*)-4-[[(2,5-diamino-6-chloropyrimidin-4-yl)oxy]methyl]cyclopent-2-enyl]-9*H*-purine-2-amine,



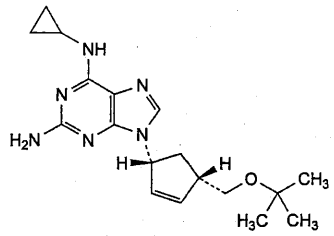
C. [(1*S*,4*R*)-4-(2,6-diamino-9*H*-purin-9-yl)cyclopent-2-enyl] methanol,



D. [(1*R*,4*R*)-4-[2-amino-6-(cyclopropylamino)-9*H*-purin-9-yl] cyclopent-2-enyl]methanol,



E. [(1*R*,3*S*)-3-[2-amino-6-(cyclopropylamino)-9*H*-purin-9-yl] cyclopentyl]methanol,



F. 6-(cyclopropylamino)-9-[(1*R*,4*S*)-4-[[(1,1-dimethylethyl) oxy]methyl]cyclopent-2-enyl]-9*H*-purine-2-amine.

Ph Eur

Acacia

(Ph. Eur. monograph 0307)

Action and use

Bulk-forming laxative; excipient.

When Powdered Acacia is prescribed or demanded, material complying with the requirements below with the exception of Identification test A shall be dispensed or supplied.

Ph Eur

DEFINITION

Air-hardened, gummy exudate flowing naturally from or obtained by incision of the trunk and branches of *Acacia senegal* L. Willd. (syn. *Senegalia senegal* (L.) Britton), other species of *Acacia* of African origin and *Acacia seyal* Delile.

CHARACTERS

It is almost completely but very slowly soluble, after about 2 h, in twice its mass of water leaving only a very small residue of vegetable particles; the liquid obtained is colourless or yellowish, dense, viscous, adhesive, translucent and weakly acid to blue litmus paper. It is practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. It occurs as yellowish-white, yellow or pale amber, sometimes with a pinkish tint, friable, opaque, spheroidal, oval or reniform pieces (tears) of a diameter from about 1-3 cm, frequently with a cracked surface, easily broken into irregular, whitish or slightly yellowish angular fragments with a conchoidal fracture and a glassy and transparent appearance. In the centre of an unbroken tear there is sometimes a small cavity.

B. Microscopic examination (*2.8.23*). The powder is white or yellowish-white. Examine under a microscope using *ethanol (96 per cent) R*. The powder shows the following diagnostic characters: angular, irregular, colourless, transparent fragments. Only traces of starch or plant tissues are visible. No stratified membrane is apparent.

C. Examine the chromatograms obtained in the test for glucose and fructose.

Results See below the sequence of zones present in the chromatograms obtained with reference solution (a) and the test solution.

Top of the plate	
	3 blue zones, very faint
Rhamnose: a greenish-brown zone	A greenish-brown zone, very faint to equivalent (rhamnose)
Xylose: a brownish-grey zone	
Arabinose: a brownish-grey zone	A brownish-grey zone, intense (arabinose)
Glucose: a greyish-blue zone	
Galactose: a greyish-blue zone	A greyish-blue zone, intense (galactose)
	1 or 2 brownish-grey zones, very faint to equivalent
	1 or 2 blue zones, faint to equivalent
Reference solution (a)	**Test solution**

D. Dissolve 1 g of the powdered herbal drug (355) (*2.9.12*) in 2 mL of *water R* by stirring frequently for 2 h. Add 2 mL of *ethanol (96 per cent) R*. After shaking, a white gelatinous mucilage is formed that becomes fluid upon addition of 10 mL of *water R*.

TESTS

Solution S

Dissolve 3.0 g of the powdered herbal drug (355) (*2.9.12*) in 25 mL of *water R* by stirring for 30 min. Allow to stand for 30 min and dilute to 30 mL with *water R*.

Insoluble matter

Maximum 0.5 per cent.

To 5.0 g of the powdered herbal drug (355) (*2.9.12*) add 100 mL of *water R* and 14 mL of *dilute hydrochloric acid R*, boil gently for 15 min, shaking frequently and filter while hot through a tared sintered-glass filter (*2.1.2*). Wash with hot *water R* and dry at 100-105 °C. The residue weighs a maximum of 25 mg.

Glucose and fructose

High-performance thin-layer chromatography (*2.8.25*).

Test solution To 0.1 g of the powdered herbal drug (355) (*2.9.12*) in a thick-walled centrifuge tube, add 2 mL of a 100 g/L solution of *trifluoroacetic acid R* and shake vigorously. Stopper the tube and heat the mixture at 120 °C for 1 h. Centrifuge, transfer 1 mL of the clear supernatant into a 10 mL flask and add 5 mL of *methanol R*.

Reference solution (a) Dissolve 5 mg of *arabinose R*, 5 mg of *galactose R*, 5 mg of *glucose R*, 5 mg of *rhamnose R* and 5 mg of *xylose R* in 1 mL of *water R* and dilute to 10.0 mL with *methanol R*.

Reference solution (b) Dilute 2.5 mL of reference solution (a) to 10.0 mL with *methanol R*.

Reference solution (c) Dissolve 5 mg of *galactose R* and 5 mg of *glucose R* in 1 mL of *water R* and dilute to 10 mL with *methanol R*.

Intensity marker Galactose.

Plate *TLC silica gel F_{254} plate R* (2-10 µm).

Mobile phase *water R*, *acetonitrile R* (15:85 *V/V*).

Application 4 µL of the test solution and reference solutions (a) and (b), and 2 µL of reference solution (c), as bands of 8 mm.

Development A 70 mm from the lower edge of the plate, in an unsaturated tank.

Drying A In air.

Development B 70 mm from the lower edge of the plate, in an unsaturated tank, using freshly prepared mobile phase.

Drying B In air.

Detection Treat with a solution prepared as follows: dissolve 4 g of *diphenylamine R* and 4 mL of *aniline R* in 160 mL of *acetone R* and add *phosphoric acid R* until the precipitate formed dissolves again (about 30 mL). Heat at 120 °C for 5-10 min and examine in daylight.

System suitability Reference solution (c):
— the chromatogram shows in the middle third 2 distinct zones, which may be touching; the lower zone (galactose) and the upper zone (glucose) are greyish-blue.

Results The chromatogram obtained with the test solution shows no greyish-blue zone and no reddish zone between the zones due to galactose and arabinose in the chromatogram obtained with reference solution (a).

Starch, dextrin and agar

To 10 mL of solution S, previously boiled and cooled, add 0.1 mL of *0.05 M iodine*. No blue or reddish-brown colour develops.

Sterculia gum

A. Place 0.2 g of the powdered herbal drug (355) (*2.9.12*) in a 10 mL ground-glass-stoppered cylinder graduated in 0.1 mL. Add 10 mL of *ethanol (60 per cent V/V) R* and shake. Any gel formed occupies a maximum of 1.5 mL.

B. To 1.0 g of the powdered herbal drug (355) (*2.9.12*) add 100 mL of *water R* and shake. Add 0.1 mL of *methyl red solution R*. Not more than 5.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Tannins

To 10 mL of solution S add 0.1 mL of *ferric chloride solution R1*. A gelatinous precipitate is formed, but neither the precipitate nor the liquid is dark blue.

Tragacanth

Examine the chromatograms obtained in the test for glucose and fructose.

Results The chromatogram obtained with the test solution shows no faint to intense brownish-grey zone corresponding to the zone due to xylose in the chromatogram obtained with reference solution (a).

Loss on drying (*2.2.32*)

Maximum 15.0 per cent, determined on 1.000 g of the powdered herbal drug (355) (*2.9.12*) by drying in an oven at 105 °C.

Total ash (*2.4.16*)
Maximum 4.0 per cent.

Microbial contamination
TAMC: acceptance criterion 10^4 CFU/g (*2.6.12*).
TYMC: acceptance criterion 10^4 CFU/g (*2.6.12*).
Absence of *Escherichia coli* (*2.6.13*).
Absence of *Salmonella* (*2.6.13*).

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for acacia used as a viscosity-increasing agent and/or suspending agent in aqueous preparations.

Apparent viscosity
Determine the dynamic viscosity using a capillary viscometer (*2.2.9*) or a rotating viscometer (*2.2.10*) on a 100 g/L solution of acacia (dried substance).

Ph Eur

Acacia, Dried Dispersion

Spray-dried Acacia
(Ph. Eur. monograph 0308)

Ph Eur

DEFINITION
Powder obtained from a dispersion of *Acacia (0307)* after a drying process.

CHARACTERS
It dissolves completely, after about 20 min, in twice its mass of water. The liquid obtained is colourless or yellowish, dense, viscous, adhesive, translucent and weakly acid to blue litmus paper. It is practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Examine under a microscope using *ethanol (96 per cent) R* as the mounting medium. The preparation to be examined consists of predominantly spheroidal or irregular and angular particles varying in size (4-500 µm), with 1 or more rounded cavities containing 1 or several air bubbles; a few flat fragments are also present. Only traces of starch granules are visible and no plant tissue is observed.

B. Examine the chromatograms obtained in the test for glucose and fructose.

Results See below the sequence of zones present in the chromatograms obtained with reference solution (a) and the test solution.

Top of the plate	
	3 blue zones, very faint
Rhamnose: a greenish-brown zone	A greenish-brown zone, very faint to equivalent (rhamnose)
Xylose: a brownish-grey zone	
Arabinose: a brownish-grey zone	A brownish-grey zone, intense (arabinose)
Glucose: a greyish-blue zone	
Galactose: a greyish-blue zone	A greyish-blue zone, intense (galactose)
	1 or 2 brownish-grey zones, very faint to equivalent
	1 or 2 blue zones, faint to equivalent
Reference solution (a)	**Test solution**

C. Dissolve 1 g of the preparation to be examined in 2 mL of *water R* by stirring frequently for 20 min. Add 2 mL of *ethanol (96 per cent) R*. After shaking, a white gelatinous mucilage is formed that becomes fluid upon addition of 10 mL of *water R*.

TESTS
Solution S
Dissolve 3.0 g of the preparation to be examined in 25 mL of *water R* by stirring for 10 min. Allow to stand for 20 min and dilute to 30 mL with *water R*.

Glucose and fructose
High-performance thin-layer chromatography (*2.8.25*)

Test solution To 0.1 g in a thick-walled centrifuge tube add 2 mL of a 100 g/L solution of *trifluoroacetic acid R* and shake vigorously. Stopper the tube and heat the mixture at 120 °C for 1 h. Centrifuge, transfer 1 mL of the clear supernatant into a 10 mL flask and add 5 mL of *methanol R*.

Reference solution (a) Dissolve 5 mg of *arabinose R*, 5 mg of *galactose R*, 5 mg of *glucose R*, 5 mg of *rhamnose R* and 5 mg of *xylose R* in 1 mL of *water R* and dilute to 10.0 mL with *methanol R*.

Reference solution (b) Dilute 2.5 mL of reference solution (a) to 10.0 mL with *methanol R*.

Reference solution (c) Dissolve 5 mg of *galactose R* and 5 mg of *glucose R* in 1 mL of *water R* and dilute to 10 mL with *methanol R*.

Intensity marker Galactose.

Plate *TLC silica gel F_{254} plate R* (2-10 µm).

Mobile phase *water R, acetonitrile R* (15:85 *V/V*).

Application 4 µL of the test solution and reference solutions (a) and (b), and 2 µL of reference solution (c), as bands of 8 mm.

Development A 70 mm from the lower edge of the plate, in an unsaturated tank.

Drying A In air.

Development B 70 mm from the lower edge of the plate, in an unsaturated tank, using freshly prepared mobile phase.

Drying B In air.

Detection Treat with a solution prepared as follows: dissolve 4 g of *diphenylamine R* and 4 mL of *aniline R* in 160 mL of *acetone R* and add *phosphoric acid R* until the precipitate formed dissolves again (about 30 mL). Heat at 120 °C for 5-10 min and examine in daylight.

System suitability Reference solution (c):

— the chromatogram shows in the middle third 2 distinct zones, which may be touching; the lower zone (galactose) and the upper zone (glucose) are greyish-blue.

Results The chromatogram obtained with the test solution shows no greyish-blue zone and no reddish zone between the zones due to galactose and arabinose in the chromatogram obtained with reference solution (a).

Starch, dextrin and agar

To 10 mL of solution S, previously boiled and cooled, add 0.1 mL of *0.05 M iodine*. No blue or reddish-brown colour develops.

Sterculia gum

A. Place 0.2 g in a 10 mL ground-glass-stoppered cylinder graduated in 0.1 mL. Add 10 mL of *ethanol (60 per cent V/V) R* and shake. Any gel formed occupies not more than 1.5 mL.

B. To 1.0 g add 100 mL of *water R* and shake. Add 0.1 mL of *methyl red solution R*. Not more than 5.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Tannins

To 10 mL of solution S add 0.1 mL of *ferric chloride solution R1*. A gelatinous precipitate is formed, but neither the precipitate nor the liquid is dark blue.

Tragacanth

Examine the chromatograms obtained in the test for glucose and fructose.

Results The chromatogram obtained with the test solution shows no faint to intense brownish-grey zone corresponding to the zone due to xylose in the chromatogram obtained with reference solution (a).

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Total ash (*2.4.16*)

Maximum 4.0 per cent.

Microbial contamination

TAMC: acceptance criterion 10^4 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for acacia dried dispersion used as a viscosity-increasing agent and/or suspending agent in aqueous preparations.

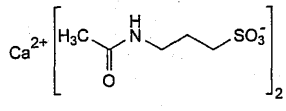
Apparent viscosity

Determine the dynamic viscosity using a capillary viscometer (*2.2.9*) or a rotating viscometer (*2.2.10*) on a 100 g/L solution of acacia, dried dispersion (dried substance).

Ph Eur

Acamprosate Calcium

(Ph. Eur. monograph 1585)



$C_{10}H_{20}CaN_2O_8S_2$ 400.5 *77337-73-6*

Action and use

Treatment of alcoholism.

Preparation

Acamprosate Gastro-resistant Tablets

Ph Eur

DEFINITION

Calcium bis[3-(acetylamino)propane-1-sulfonate].

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of acamprosate calcium.*

B. It gives reaction (a) of calcium (*2.3.1*).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
5.5 to 7.0 for solution S.

Impurity A
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.40 g of the substance to be examined in *distilled water R* and dilute to 20.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with *borate buffer solution pH 10.4 R*. Place 3.0 mL of the solution obtained in a 25 mL ground-glass-stoppered tube. Add 0.15 mL of a freshly prepared 5 g/L solution of *fluorescamine R* in *acetonitrile R*. Shake immediately and vigorously for 30 s. Place in a water-bath at 50 °C for 30 min. Cool under a stream of cold water. Centrifuge and filter the supernatant through a suitable membrane filter (nominal pore size 0.45 µm), 25 mm in diameter.

Reference solution Dissolve 50 mg of *acamprosate impurity A CRS* in *distilled water R* and dilute to 200.0 mL with the same solvent. Dilute 0.4 mL of the solution to 100.0 mL with *borate buffer solution pH 10.4 R*. Treat 3.0 mL of this solution in the same way as the test solution

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm) with a specific surface area of 170 m^2/g and a pore size of 12 nm.

Mobile phase *acetonitrile R, methanol R*, 0.1 M phosphate buffer solution pH 6.5 R (10:10:80 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 261 nm.

Injection 20 µL.

Run time 6 times the retention time of impurity A

Retention times Fluorescamine = about 4 min; impurity A = about 8 min; acamprosate is not detected by this system.

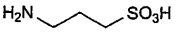
Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.4 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
To 4 g of *cation-exchange resin R* (75-150 µm) add 20 mL of *distilled water R* and stir magnetically for 10 min. Introduce this suspension into a glass column, 45 cm long and 2.2 cm in internal diameter, equipped with a polytetrafluoroethylene flow cap covered by a glass-wool plug. Allow a few millilitres of this solution to flow, then place a plug of glass wool over the resin. Pass 50 mL of *1 M hydrochloric acid* through the column. The pH of the eluate is close to 1. Wash with 3 quantities, each of 200 mL, of *distilled water R* to obtain an eluate at pH 6. Dissolve 0.100 g of the substance to be examined in 15 mL of *distilled water R*. Pass through the column and wash with 3 quantities, each of 25 mL, of *distilled water R*, collecting the eluate. Allow to elute until an eluate at pH 6 is obtained. Titrate the solution obtained with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* corresponds to 20.02 mg of $C_{10}H_{20}CaN_2O_8S_2$.

IMPURITIES

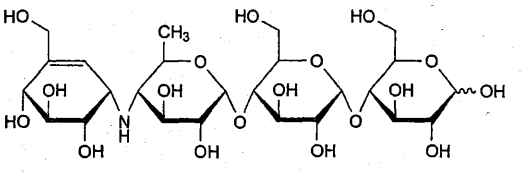


A. 3-aminopropane-1-sulfonic acid (homotaurine).

Ph Eur

Acarbose

(Ph. Eur. monograph 2089)



$C_{25}H_{43}NO_{18}$ 646 *56180-94-0*

Action and use
Alpha-glucosidase inhibitor; treatment of diabetes mellitus.

Ph Eur

DEFINITION
O-4,6-Dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-D-glucopyranose, which is produced by certain strains of *Actinoplanes utahensis*.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or yellowish, hygroscopic, amorphous powder.

Solubility
Very soluble in water, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acarbose for identification CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS
Solution S
Dissolve 1.00 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

pH (*2.2.3*)
5.5 to 7.5 for solution S.

Specific optical rotation (*2.2.7*)
+ 168 to + 183 (anhydrous substance).

Dilute 2.0 mL of solution S to 10.0 mL with *water R*.

Absorbance (*2.2.25*)
Maximum 0.15 at 425 nm for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.200 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of *acarbose CRS* in 5.0 mL of *water R*.

Reference solution (b) Dissolve the contents of a vial of *acarbose for peak identification CRS* (acarbose containing impurities A, B, C, D, E, F and G) in 1 mL of *water R*.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *aminopropylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase Mix 750 volumes of *acetonitrile R1* and 250 volumes of a solution containing 0.60 g/L of *potassium dihydrogen phosphate R* and 0.35 g/L of *disodium hydrogen phosphate dihydrate R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of acarbose.

Identification of impurities Use the chromatogram supplied with *acarbose for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F and G.

Relative retention With reference to acarbose (retention time = about 16 min): impurity D = about 0.5; impurity B = about 0.8; impurity A = about 0.9; impurity C = about 1.2; impurity E = about 1.7; impurity F = about 1.9; impurity G = about 2.2.

System suitability Reference solution (b):
— the chromatogram obtained is similar to the chromatogram supplied with *acarbose for peak identification CRS*;
— *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to acarbose.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.63; impurity D = 0.75; impurity E = 1.25; impurity F = 1.25; impurity G = 1.25;
— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.5 per cent);
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *impurity A*: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.6 per cent);
— *impurity B*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurities F, G*: for each impurity, not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurity E*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{25}H_{43}NO_{18}$ taking into account the assigned content of *acarbose CRS*.

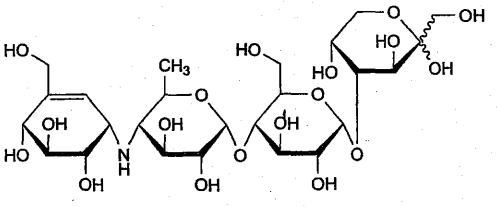
STORAGE

In an airtight container.

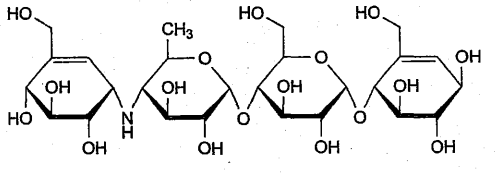
IMPURITIES

Specified impurities *A, B, C, D, E, F, G.*

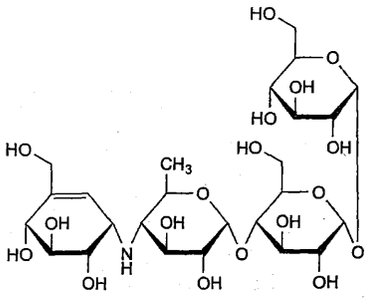
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H.*



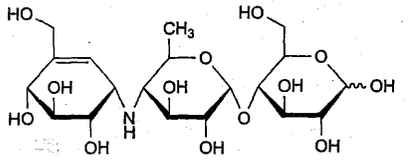
A. *O*-4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-D-*arabino*-hex-2-ulopyranose,



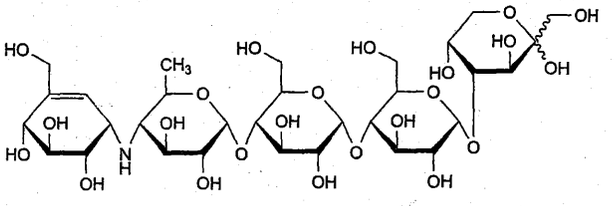
B. (1*R*,4*R*,5*S*,6*R*)-4,5,6-trihydroxy-2-(hydroxymethyl) cyclohex-2-enyl 4-*O*-[4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl] amino]-α-D-glucopyranosyl]-α-D-glucopyranoside,



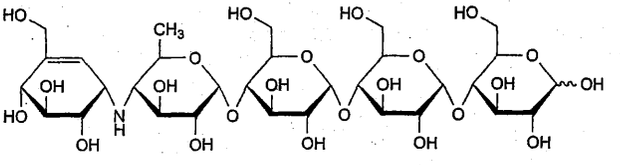
C. α-D-glucopyranosyl 4-*O*-[4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl]-α-D-glucopyranoside,



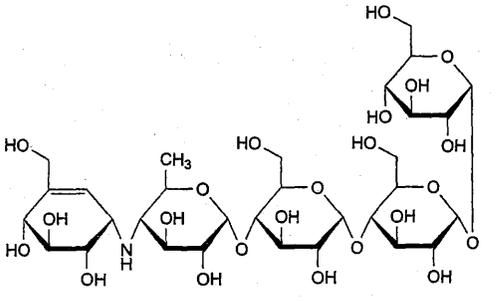
D. 4-*O*-[4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl]-D-glucopyranose,



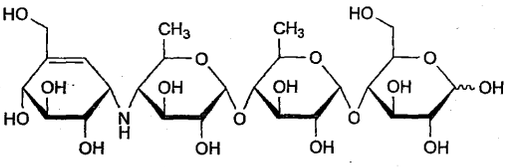
E. *O*-4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-D-*arabino*-hex-2-ulopyranose (4-*O*-α-acarbosyl-D-fructopyranose),



F. *O*-4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-D-glucopyranose (4-*O*-α-acarbosyl-D-glucopyranose),



G. α-D-glucopyranosyl *O*-4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranosyl-(1→4)-*O*-α-D-glucopyranoside (α-D-glucopyranosyl α-acarboside),

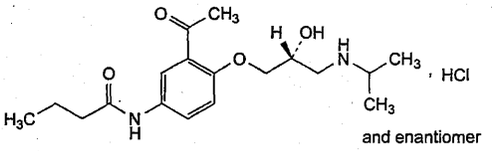


H. *O*-4,6-dideoxy-4-[[(1*S*,4*R*,5*S*,6*S*)-4,5,6-trihydroxy-3-(hydroxymethyl)cyclohex-2-enyl]amino]-α-D-glucopyranosyl-(1→4)-*O*-6-deoxy-α-D-glucopyranosyl-(1→4)-D-glucopyranose.

Ph Eur

Acebutolol Hydrochloride

(Ph. Eur. monograph 0871)



and enantiomer

$C_{18}H_{29}ClN_2O_4$ 372.9 *34381-68-5*

Action and use
Beta-adrenoceptor antagonist.

Preparations
Acebutolol Capsules
Acebutolol Tablets

Ph Eur

DEFINITION

N-[3-Acetyl-4-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino]propoxy]phenyl]butanamide hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in ethanol (96 per cent), very slightly soluble in acetone and in methylene chloride.

mp
About 143 °C.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 20.0 mg in a 0.1 per cent *V/V* solution of *hydrochloric acid R* and dilute to 100.0 mL with the same acid solution. Dilute 5.0 mL of this solution to 100.0 mL with a 0.1 per cent *V/V* solution of *hydrochloric acid R*.

Spectral range 220-350 nm.

Absorption maxima At 233 nm and 322 nm.

Specific absorbance at the absorption maximum 555 to 605 at 233 nm.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *acebutolol hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 20 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *acebutolol hydrochloride CRS* in *methanol R* and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *pindolol CRS* in *methanol R* and dilute to 20 mL with the same solvent. To 1 mL of this solution add 1 mL of reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *perchloric acid R, methanol R, water R* (5:395:600 *V/V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 0.5 g in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)

5.0 to 7.0.

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 20.0 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A. Dilute 0.5 mL of this solution to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *acebutolol impurity I CRS* in 1.0 mL of mobile phase A.

Reference solution (c) Mix 2.0 mL of reference solution (a) and 1.0 mL of reference solution (b) and dilute to 10.0 mL with mobile phase A.

Reference solution (d) Dissolve 5.0 mg of *acebutolol impurity C CRS* in 10 mL of *acetonitrile R* and dilute to 25.0 mL with mobile phase A. Dilute 0.5 mL of this solution to 50.0 mL with mobile phase A.

Reference solution (e) Dissolve 5.0 mg of *acebutolol impurity B CRS* in 10.0 mL of *acetonitrile R* and dilute to 25.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 50.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.125 m, Ø = 4 mm,
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm),
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: mix 2.0 mL of *phosphoric acid R*, and 3.0 mL of *triethylamine R* and dilute to 1000 mL with *water R*;
- *mobile phase B*: mix equal volumes of *acetonitrile R* and mobile phase A;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	98	2
2 - 30.5	98 → 10	2 → 90
30.5 - 41	10	90

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 25 µL.

System suitability Reference solution (c):
- *resolution*: minimum 7.0 between the peaks due to impurity I and acebutolol.

Limits:
- *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.2 per cent);
- *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.1 per cent);
- *impurity I*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of *ethanol (96 per cent) R* and add 1 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

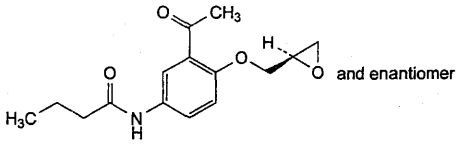
1 mL of *0.1 M sodium hydroxide* is equivalent to 37.29 mg of $C_{18}H_{29}ClN_2O_4$.

STORAGE

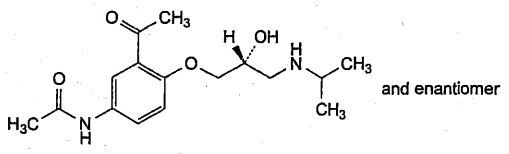
Protected from light.

IMPURITIES

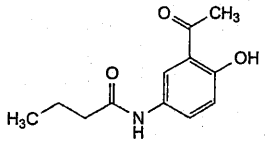
Specified impurities *A, B, C, D, E, F, G, H, I, J, K.*



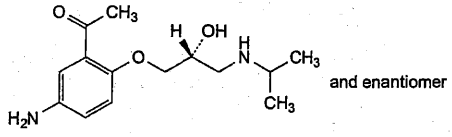
A. *N*-[3-acetyl-4-[(2*RS*)-oxiran-2-ylmethoxy] phenyl]butanamide,



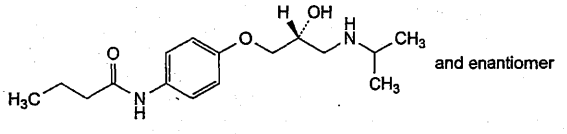
B. *N*-[3-acetyl-4-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino] propoxy]phenyl]acetamide (diacetolol),



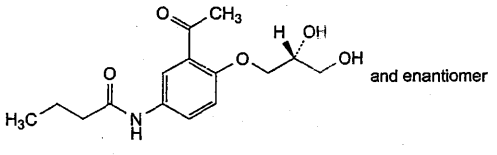
C. *N*-(3-acetyl-4-hydroxyphenyl)butanamide,



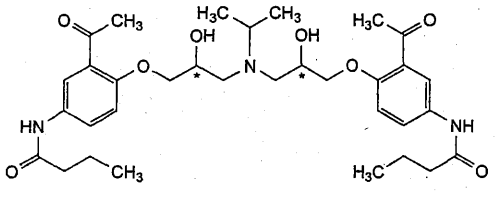
D. 1-[5-amino-2-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino] propoxy]phenyl]ethanone,



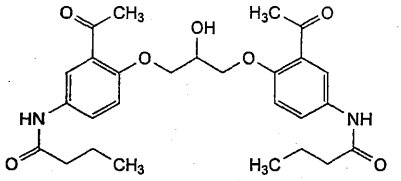
E. *N*-[4-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino]propoxy] phenyl]butanamide,



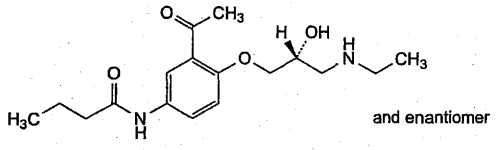
F. *N*-[3-acetyl-4-[(2*RS*)-2,3-dihydroxypropoxy] phenyl]butanamide,



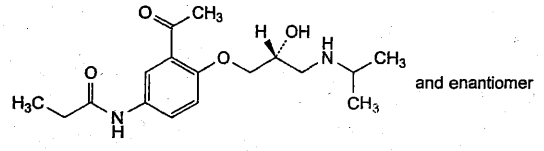
G. *N*,*N*′-[[(1-methylethyl)imino]bis[(2-hydroxypropane-1,3-diyl)oxy(3-acetyl-1,4-phenylene)]]dibutanamide (biamine),



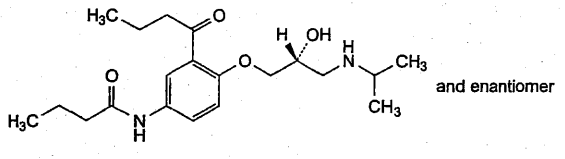
H. *N*,*N*′-[(2-hydroxypropane-1,3-diyl)bis[oxy(3-acetyl-1,4-phenylene)]]dibutanamide,



I. *N*-[3-acetyl-4-[(2*RS*)-3-(ethylamino)-2-hydroxypropoxy] phenyl]butanamide,



J. *N*-[3-acetyl-4-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino] propoxy]phenyl]propanamide,

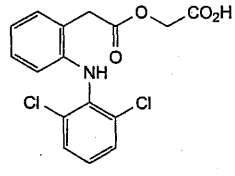


K. *N*-[3-butanoyl-4-[(2*RS*)-2-hydroxy-3-[(1-methylethyl)amino]propoxy]phenyl]butanamide.

Ph Eur

Aceclofenac

(Ph. Eur. monograph 1281)



$C_{16}H_{13}Cl_2NO_4$ 354.2 *89796-99-6*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
[[[2-[(2,6-Dichlorophenyl)amino]phenyl]acetyl]oxy]acetic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 50.0 mL with *methanol R*.

Spectral range 220-370 nm.

Absorption maximum 275 nm.

Specific absorbance at the absorption maximum 320 to 350.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison: Ph. Eur. reference spectrum of aceclofenac.

C. Dissolve about 10 mg in 10 mL of *ethanol (96 per cent) R*. To 1 mL of the solution, add 0.2 mL of a mixture, prepared immediately before use, of equal volumes of a 6 g/L solution of *potassium ferricyanide R* and a 9 g/L solution of *ferric chloride R*. Allow to stand protected from light for 5 min. Add 3 mL of a 10.0 g/L solution of *hydrochloric acid R*. Allow to stand protected from light for 15 min. A blue colour develops and a precipitate is formed.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mobile phase A, mobile phase B (30:70 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 21.6 mg of *diclofenac sodium CRS* (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 2.0 mL of the test solution to 10.0 mL with the solvent mixture.

Reference solution (c) Mix 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b) and dilute to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 4.0 mg of *aceclofenac impurity F CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (e) Dissolve 2.0 mg of *aceclofenac impurity H CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (f) Mix 1.0 mL of reference solution (b), 1.0 mL of reference solution (d) and 1.0 mL of reference solution (e) and dilute to 100.0 mL with the solvent mixture.

Reference solution (g) Dissolve 5.0 mg of *aceclofenac impurity I CRS* in the solvent mixture and dilute to 50.0 mL with solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (h) Dissolve 4 mg of *aceclofenac for peak identification CRS* (containing impurities B, C, D, E and G) in 2 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 10 nm and a carbon loading of 19 per cent;
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: 1.12 g/L solution of *phosphoric acid R* adjusted to pH 7.0 with a 42 g/L solution of *sodium hydroxide R*;
— *mobile phase B*: *water R, acetonitrile R* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	70 → 50	30 → 50
25 - 30	50 → 20	50 → 80
30 - 50	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL of the test solution and reference solutions (c), (d), (e), (f), (g) and (h).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A; use the chromatogram supplied with *aceclofenac for peak identification CRS* and the chromatogram obtained with reference solution (h) to identify the peaks due to impurities B, C, D, E and G; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity F; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity H; use the chromatogram obtained with reference solution (g) to identify the peak due to impurity I.

Relative retention With reference to aceclofenac (retention time = about 11 min): impurity A = about 0.8; impurity G = about 1.3; impurity H = about 1.5; impurity I = about 2.3; impurity D = about 3.1; impurity B = about 3.2; impurity E = about 3.3; impurity C = about 3.5; impurity F = about 3.7.

System suitability Reference solution (c):
— *resolution*: minimum 5.0 between the peaks due to impurity A and aceclofenac.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *impurities B, C, D, E, G*: for each impurity, not more than the area of the peak due to aceclofenac in the chromatogram obtained with reference solution (f) (0.2 per cent);
— *impurity F*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.2 per cent);
— *impurity H*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.15 per cent);
— *impurity I*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (g) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the peak due to aceclofenac in the chromatogram obtained with reference solution (f) (0.10 per cent);
— *total*: maximum 0.7 per cent;
— *disregard limit*: 0.25 times the area of the peak due to aceclofenac in the chromatogram obtained with reference solution (f) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

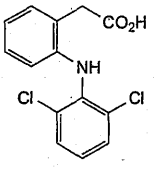
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 40 mL of *methanol R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

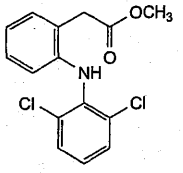
1 mL of *0.1 M sodium hydroxide* is equivalent to 35.42 mg of $C_{16}H_{13}Cl_2NO_4$.

STORAGE
Protected from light.

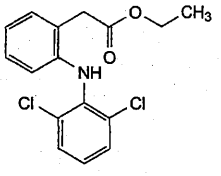
IMPURITIES
Specified impurities *A, B, C, D, E, F, G, H, I.*



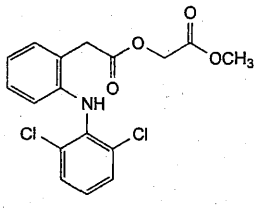
A. [2-[(2,6-dichlorophenyl)amino]phenyl]acetic acid (diclofenac),



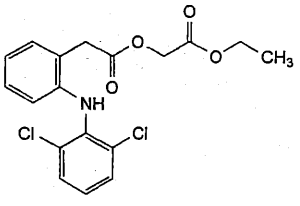
B. methyl [2-[(2,6-dichlorophenyl)amino]phenyl]acetate (methyl ester of diclofenac),



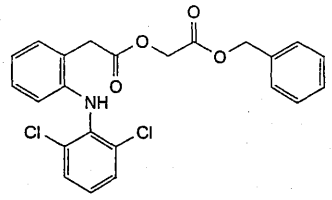
C. ethyl [2-[(2,6-dichlorophenyl)amino]phenyl]acetate (ethyl ester of diclofenac),



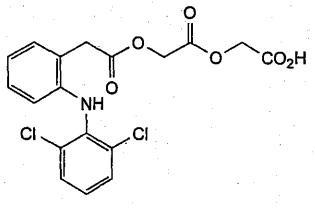
D. methyl [[[2-[(2,6-dichlorophenyl)amino]phenyl]acetyl]oxy] acetate (methyl ester of aceclofenac),



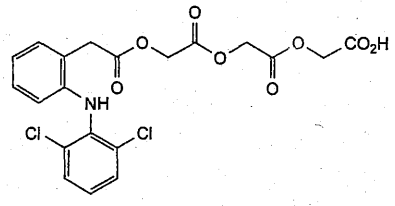
E. ethyl [[[2-[(2,6-dichlorophenyl)amino]phenyl]acetyl]oxy] acetate (ethyl ester of aceclofenac),



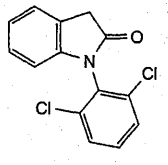
F. benzyl [[[2-[(2,6-dichlorophenyl)amino]phenyl]acetyl]oxy] acetate (benzyl ester of aceclofenac),



G. [[[[[2-[(2,6-dichlorophenyl)amino]phenyl]acetyl]oxy] acetyl]oxy]acetic acid (acetic aceclofenac),



H. [[[[[[[2-[(2,6-dichlorophenyl)amino]phenyl]acetyl]oxy] acetyl]oxy]acetyl]oxy]acetic acid (diacetic aceclofenac),

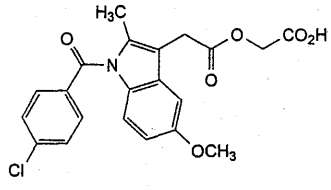


I. 1-(2,6-dichlorophenyl)-1,3-dihydro-2*H*-indol-2-one.

Ph Eur

Acemetacin

(Ph. Eur. monograph 1686)



$C_{21}H_{18}ClNO_6$ 415.8 *53164-05-9*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
[[[1-(4-Chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl] acetyl]oxy]acetic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow or greenish-yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone, slightly soluble in anhydrous ethanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acemetacin CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *acetonitrile for chromatography R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 50.0 mL with *acetonitrile for chromatography R*. Dilute 1.0 mL of this solution to 100.0 mL with *acetonitrile for chromatography R*.

Reference solution (b) Dissolve 5.0 mg of *acemetacin impurity A CRS* and 10.0 mg of *indometacin CRS* (impurity B) in *acetonitrile for chromatography R*, and dilute to 50.0 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 20.0 mL with *acetonitrile for chromatography R*.

Reference solution (d) To 1 mL of reference solution (b), add 10 mL of the test solution and dilute to 20 mL with *acetonitrile for chromatography R*.

Reference solution (e) Dissolve the contents of a vial of *acemetacin impurity mixture CRS* (containing impurities C, D, E and F) in 1.0 mL of the test solution.

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: dissolve 1.0 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 6.5 with *1 M sodium hydroxide* and dilute to 1000 mL with *water R*;
- *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	95	5
5 - 9	95 → 65	5 → 35
9 - 16	65	35
16 - 28	65 → 20	35 → 80
28 - 34	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 µL.

Identification of impurities:
- use the chromatogram supplied with *acemetacin impurity mixture CRS* and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities C, D, E and F;
- use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to acemetacin (retention time = about 15 min): impurity A = about 0.7; impurity B = about 0.9; impurity F = about 1.2; impurity C = about 1.3; impurity D = about 1.5; impurity E = about 2.2.

System suitability Reference solution (d):
- *peak-to-valley ratio*: minimum 15, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to acemetacin.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.3; impurity D = 1.4; impurity F = 1.3;
- *impurity E*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
- *impurities C, D, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 20 mL of *acetone R* and add 10 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

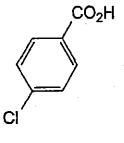
1 mL of *0.1 M sodium hydroxide* is equivalent to 41.58 mg of $C_{21}H_{18}ClNO_6$.

STORAGE

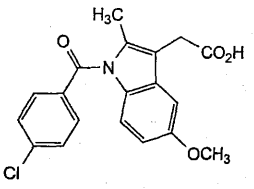
Protected from light.

IMPURITIES

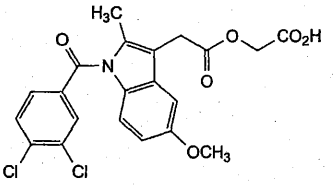
Specified impurities *A, B, C, D, E, F.*



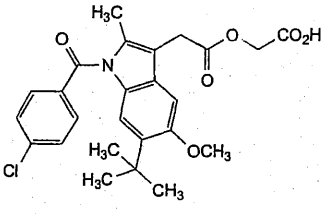
A. 4-chlorobenzoic acid,



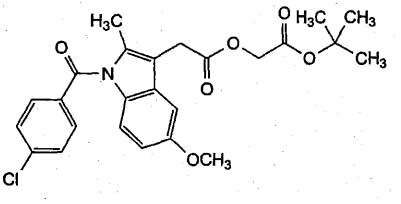
B. [1-(4-chlorobenzoyl)-5-methoxy-2-methylindol-3-yl]acetic acid (indometacin),



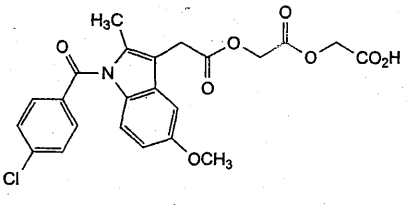
C. [[[1-(3,4-dichlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetyl]oxy]acetic acid,



D. [[[1-(4-chlorobenzoyl)-6-(1,1-dimethylethyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetyl]oxy]acetic acid,



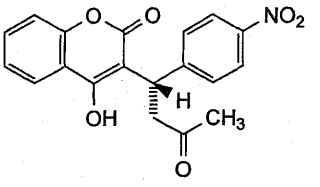
E. 1,1-dimethylethyl [[[1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetyl]oxy]acetate,



F. [[[[[1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetyl]oxy]acetyl]oxy]acetic acid.

Ph Eur

Acenocoumarol



and enantiomer

$C_{19}H_{15}NO_6$ 353.3 *152-72-7*

Action and use
Vitamin K epoxide reductase inhibitor; oral anticoagulant.

Preparation
Acenocoumarol Tablets

DEFINITION
Acenocoumarol is (*RS*)-4-hydroxy-3-(1-*p*-nitrophenyl-3-oxobutyl)coumarin. It contains not less than 98.5% and not more than 100.5% of $C_{19}H_{15}NO_6$, calculated with reference to the dried substance.

CHARACTERISTICS
An almost white to buff powder.

Practically insoluble in *water* and in *ether*; slightly soluble in *ethanol (96%)*. It dissolves in aqueous solutions of the alkali hydroxides. It exhibits polymorphism.

IDENTIFICATION
The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of acenocoumarol *(RS 001)*. If the spectra are not concordant, dissolve 0.1 g of the substance being examined in 10 mL of *acetone* and add *water* drop wise until the solution becomes turbid. Heat on a water bath until the solution is clear and allow to stand. Filter, wash the crystals with a mixture of equal volumes of *acetone* and *water* and dry at 100° at a pressure of 2 kPa for 30 minutes. Prepare a new spectrum of the residue.

TESTS
Clarity and colour of solution
A. A 2.0% w/v solution in *acetone* is *clear*, Appendix IV A.

B. The *absorbance* of a 4-cm layer of a 2.0% w/v solution in *acetone* at 460 nm is not more than 0.12, Appendix II B.

C. A 2.0% w/v solution in 0.1M *sodium hydroxide* is *clear*, Appendix IV A, and yellow.

Light absorption
Absorbance of a 0.001% w/v solution in a mixture of 1 volume of *1M hydrochloric acid* and 9 volumes of *methanol* at the maximum at 306 nm, 0.50 to 0.54, calculated with reference to the dried substance, Appendix II B.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in *acetone*.

(1) 2.0% w/v of the substance being examined.

(2) 0.0020% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating *silica gel* GF_{254}.

(b) Use the mobile phase as described below.

(c) Apply 20 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, allow it to dry in air and immediately examine under *ultraviolet light (254 nm)*.

MOBILE PHASE

20 volumes of *glacial acetic acid*, 50 volumes of *cyclohexane* and 50 volumes of *dichloromethane*.

LIMITS

Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.1%).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

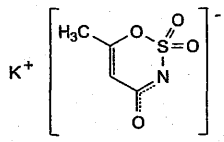
Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.6 g in 50 mL of *acetone* and titrate with *0.1M sodium hydroxide VS* using *bromothymol blue solution R3* as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of sodium hydroxide required. Each mL of *0.1M sodium hydroxide VS* is equivalent to 35.33 mg of $C_{19}H_{15}NO_6$.

Acesulfame Potassium

(Ph. Eur. monograph 1282)



$C_4H_4KNO_4S$ 201.2 *55589-62-3*

Action and use

Sweetening agent.

Ph Eur

DEFINITION

Potassium 6-methyl-1,2,3-oxathiazin-4-olate 2,2-dioxide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Soluble in water, very slightly soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acesulfame potassium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *acesulfame potassium CRS* in *water R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *acesulfame potassium CRS* and 5 mg of *saccharin sodium R* in *water R* and dilute to 5 mL with the same solvent.

Plate *cellulose for chromatography R* as the coating substance.

Mobile phase *concentrated ammonia R, acetone R, ethyl acetate R* (10:60:60 *V/V/V*).

Application 5 µL as bands.

Development Twice over 2/3 of the plate.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated zones.

Results The principal zone in the chromatogram obtained with the test solution is similar in position and size to the principal zone in the chromatogram obtained with reference solution (a).

C. 0.5 mL of solution S (see Tests) gives reaction (b) of potassium (*2.3.1*).

TESTS

Solution S

Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 20 mL of solution S add 0.1 mL of *bromothymol blue solution R1*. Not more than 0.2 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.80 g of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *acetylacetamide R* (impurity A) in *water R* and dilute to 25 mL with the same solvent. To 5 mL of the solution add 45 mL of *water R* and dilute to 100 mL with *methanol R*.

Reference solution (b) To 10 mL of reference solution (a) add 1 mL of the test solution and dilute to 20 mL with *methanol R*.

Plate *TLC silica gel plate R.*

Mobile phase *water R, ethanol (96 per cent) R, ethyl acetate R* (2:15:74 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air until the solvents are completely removed.

Detection Spray with *phosphoric vanillin solution R* and heat at 120 °C for about 10 min; examine in daylight.

System suitability The chromatogram obtained with reference solution (a) shows a clearly visible spot and the chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Limit:

— *impurity A*: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.125 per cent).

Impurity B

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 4.0 mg of *acesulfame potassium impurity B CRS* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 200.0 mL with *water R*.

Reference solution (b) Dissolve 0.100 g of the substance to be examined in reference solution (a) and dilute to 10.0 mL with the same solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Mix 40 volumes of *acetonitrile R* and 60 volumes of a 3.3 g/L solution of *tetrabutylammonium hydrogen sulfate R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 234 nm.

Injection 20 µL.

Run time Twice the retention time of acesulfame.

Relative retention With reference to acesulfame (retention time = about 5.3 min): impurity B = about 1.6.

System suitability:
— *signal-to-noise ratio*: minimum 10 for the peak due to impurity B in the chromatogram obtained with reference solution (a);
— *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to acesulfame, in the chromatogram obtained with reference solution (b).

Limit:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (20 ppm).

Fluorides
Maximum 3 ppm.

Potentiometry (*2.2.36, Method I*).

Test solution Dissolve 3.000 g of the substance to be examined in *distilled water R*, add 15.0 mL of *total-ionic-strength-adjustment buffer R1* and dilute to 50.0 mL with *distilled water R*.

Reference solutions To 0.5 mL, 1.0 mL, 1.5 mL and 3.0 mL of *fluoride standard solution (10 ppm F) R* add 15.0 mL of *total-ionic-strength-adjustment buffer R1* and dilute to 50.0 mL with *distilled water R*.

Indicator electrode Fluoride-selective.

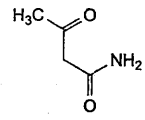
Reference electrode Silver-silver chloride.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

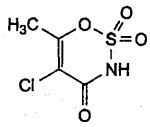
ASSAY
Dissolve 0.150 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 20.12 mg of $C_4H_4KNO_4S$.

IMPURITIES
Specified impurities *A, B.*



A. 3-oxobutanamide (acetylacetamide),

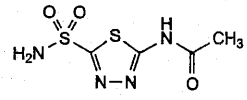


B. 5-chloro-6-methyl-1,2,3-oxathiazin-4(3*H*)-one 2,2-dioxide.

Ph Eur

Acetazolamide

(Ph. Eur. monograph 0454)



$C_4H_6N_4O_3S_2$ 222.2 *59-66-5*

Action and use
Carbonic anhydrase inhibitor; diuretic; treatment of glaucoma and ocular hypertension; treatment of mountain sickness.

Preparation
Acetazolamide Tablets

Ph Eur

DEFINITION
N-(5-Sulfamoyl-1,3,4-thiadiazol-2-yl)acetamide.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Solution A Dissolve 30.0 mg in *0.01 M sodium hydroxide* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *0.01 M sodium hydroxide*.

Solution B Dilute 25.0 mL of solution A to 100.0 mL with *0.01 M sodium hydroxide*.

Spectral range 230-260 nm for solution A; 260-350 nm for solution B.

Absorption maximum At 240 nm for solution A; at 292 nm for solution B.

Specific absorbance at the absorption maximum 162 to 176 for solution A; 570 to 620 for solution B.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acetazolamide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *ethanol (96 per cent) R*, evaporate to dryness and record new spectra using the residues.

C. Introduce about 20 mg into a test-tube and add 4 mL of *dilute hydrochloric acid R* and 0.2 g of *zinc powder R*. Immediately place a piece of *lead acetate paper R* over the mouth of the tube. The paper shows a brownish-black colour.

D. Dissolve about 25 mg in a mixture of 0.1 mL of *dilute sodium hydroxide solution R* and 5 mL of *water R*. Add 0.1 mL of *copper sulfate solution R*. A greenish-blue precipitate is formed.

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution Y_5 or BY_5 (*2.2.2, Method II*).

Dissolve 1.0 g in 10 mL of *1 M sodium hydroxide*.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 40 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *acetazolamide for system suitability CRS* (containing impurities A, B, C, D, E and F) in 1.0 mL of the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped propoxybenzene silica gel for chromatography R* (4 µm).

Mobile phase *acetonitrile for chromatography R*, 6.8 g/L solution of *potassium dihydrogen phosphate R* (10:90 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 25 µl.

Run time 3.5 times the retention time of acetazolamide.

Identification of impurities Use the chromatogram supplied with *acetazolamide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E and F.

Relative retention With reference to acetazolamide (retention time = about 8 min): impurity E = about 0.3; impurity D = about 0.4; impurity B = about 0.6; impurity C = about 1.4; impurity A = about 2.1; impurity F = about 2.6.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to impurities E and D.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 2.3; impurity C = 2.6; impurity D = 1.6;
- *impurities A, B, C, D, E, F*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfates (*2.4.13*)

Maximum 500 ppm.

To 0.4 g add 20 mL of *distilled water R* and dissolve by heating to boiling. Allow to cool with frequent shaking and filter.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

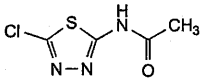
Dissolve 0.200 g in 25 mL of *dimethylformamide R*. Titrate with *0.1 M ethanolic sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 22.22 mg of $C_4H_6N_4O_3S_2$.

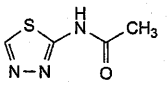
IMPURITIES

Specified impurities *A, B, C, D, E, F.*

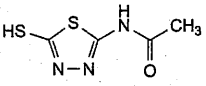
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G.*



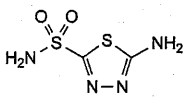
A. *N*-(5-chloro-1,3,4-thiadiazol-2-yl)acetamide,



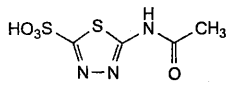
B. *N*-(1,3,4-thiadiazol-2-yl)acetamide,



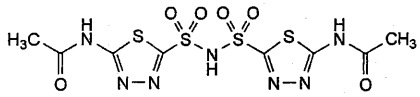
C. *N*-(5-sulfanyl-1,3,4-thiadiazol-2-yl)acetamide,



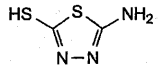
D. 5-amino-1,3,4-thiadiazole-2-sulfonamide,



E. 5-acetamido-1,3,4-thiadiazole-2-sulfonic acid,



F. *N*-[5-[(5-acetamido-1,3,4-thiadiazol-2-yl) sulfonyl]sulfamoyl-1,3,4-thiadiazol-2-yl]acetamide,

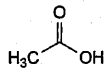


G. 5-amino-1,3,4-thiadiazole-2-thiol.

Ph Eur

Glacial Acetic Acid

(Ph. Eur. monograph 0590)



$C_2H_4O_2$	60.1	*64-19-7*

Ph Eur

DEFINITION

Content

99.0 per cent *m/m* to 100.5 per cent *m/m*.

CHARACTERS

Appearance

Crystalline mass or clear, colourless, volatile liquid.

Solubility

Miscible with water, with ethanol (96 per cent) and with methylene chloride.

IDENTIFICATION

A. A 100 g/L solution is strongly acid (*2.2.4*).

B. To 0.03 mL add 3 mL of *water R* and neutralise with *dilute sodium hydroxide solution R*. The solution gives reaction (b) of acetates (*2.3.1*).

TESTS

Solution S

Dilute 20 mL to 100 mL with *distilled water R*.

Appearance

The substance to be examined is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Freezing point (*2.2.18*)

Minimum 14.8 °C.

Reducing substances

Dilute 2.0 mL to 10.0 mL with *water R*. Add 0.1 mL of *0.02 M potassium permanganate*. Heat on a water-bath for 1 min, the colour remains pink.

Chlorides (*2.4.4*)

Maximum 25 mg/L.

Dilute 10 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 50 mg/L, determined on solution S.

Iron (*2.4.9*)

Maximum 5 ppm.

Dissolve the residue obtained in the test for residue on evaporation by heating with 2 quantities, each of 15 mL, of *water R* and dilute to 50.0 mL with *water R*. Dilute 5.0 mL of the solution to 10.0 mL with *water R*.

Residue on evaporation

Maximum 0.01 per cent.

Evaporate 20 g to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 2.0 mg.

ASSAY

Weigh accurately a conical flask with a ground-glass stopper containing 25 mL of *water R*. Add 1.0 mL of the substance to be examined and weigh again accurately. Add 0.5 mL of *phenolphthalein solution R* and titrate with *1 M sodium hydroxide*.

1 mL of *1 M sodium hydroxide* is equivalent to 60.1 mg of $C_2H_4O_2$.

STORAGE

In an airtight container.

Ph Eur

Acetic Acid (6 per cent)

Dilute Acetic Acid

DEFINITION

Acetic Acid (6 per cent) contains not less than 5.7% and not more than 6.3% w/w of acetic acid, $C_2H_4O_2$. It may be prepared by mixing 182 g of Acetic Acid (33 per cent) with 818 g of Purified Water.

IDENTIFICATION

A. Strongly acidic.

B. When neutralised, yields the reactions characteristic of *acetates*, Appendix VI.

TESTS

Weight per mL

About 1.005 g, Appendix V G.

Chloride

Dilute 5.0 mL with sufficient *water* to produce 100 mL. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (70 ppm).

Sulfate

12.5 mL of the solution used in the test for Chloride, diluted to 15 mL with *water*, complies with the *limit test for sulfates*, Appendix VII (240 ppm).

Aldehydes

Distil 75 mL. To the first 5 mL of the distillate add 10 mL of a 5% w/v solution of *mercury(II) chloride*, make alkaline with 5M *sodium hydroxide*, allow to stand for 5 minutes and acidify with 1M *sulfuric acid*. The solution shows not more than a faint turbidity.

Formic acid and oxidisable impurities

Mix 5 mL with 6 mL of *sulfuric acid* and cool to 20°. Add 0.4 mL of 0.0167M *potassium dichromate VS*, allow to stand for 1 minute, add 25 mL of *water* and 1 mL of freshly prepared *dilute potassium iodide solution* and titrate the liberated iodine with 0.1M *sodium thiosulfate VS* using *starch mucilage* as indicator. Not less than 0.2 mL of 0.1M *sodium thiosulfate VS* is required.

Readily oxidisable impurities
To 25 mL add 0.2 mL of 0.02M *potassium permanganate VS* and allow to stand for 1 minute. The pink colour is not entirely discharged.

Non-volatile matter
When evaporated to dryness and dried at 105°, leaves not more than 0.01% w/w of residue.

ASSAY
Add 30 mL of *water* to 20 g in a stopper flask and titrate with *1M sodium hydroxide VS* using *phenolphthalein solution R1* as indicator. Each mL of *1M sodium hydroxide VS* is equivalent to 60.05 mg of $C_2H_4O_2$.

Acetic Acid (33 per cent)

Acetic Acid

Preparation
Acetic Acid (6 per cent)

DEFINITION
Acetic Acid (33 per cent) contains not less than 32.5% and not more than 33.5% w/w of acetic acid, $C_2H_4O_2$.

CHARACTERISTICS
A clear, colourless liquid.

Miscible with *water*, with *ethanol (96%)* and with *glycerol*.

IDENTIFICATION
A. Strongly acidic, even when diluted freely.

B. When neutralised, yields the reactions characteristic of *acetates*, Appendix VI.

TESTS
Weight per mL
1.040 to 1.042 g, Appendix V G.

Chloride
Dilute 5.0 mL with sufficient *water* to produce 100 mL. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (70 ppm).

Sulfate
12.5 mL of the solution used in the test for Chloride, diluted to 15 mL with *water*, complies with the *limit test for sulfates*, Appendix VII (240 ppm).

Aldehydes
Distil 15 mL. To the first 5 mL of the distillate add 10 mL of a 5% w/v solution of *mercury(II) chloride*, make alkaline with 5M *sodium hydroxide*, allow to stand for 5 minutes and make acidic with 1M *sulfuric acid*. The solution shows not more than a faint turbidity.

Formic acid and oxidisable impurities
Mix 5 mL with 6 mL of *sulfuric acid* and cool to 20°. Add 2 mL of 0.0167M *potassium dichromate VS*, allow to stand for 1 minute, add 25 mL of *water* and 1 mL of freshly prepared *dilute potassium iodide solution* and titrate the liberated iodine with 0.1M *sodium thiosulfate VS* using *starch mucilage* as indicator. Not less than 1.0 mL of 0.1M *sodium thiosulfate VS* is required.

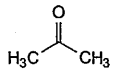
Readily oxidisable impurities
To 5.0 mL add 20 mL of *water* and 0.2 mL of 0.02M *potassium permanganate VS* and allow to stand for 1 minute. The pink colour is not entirely discharged.

Non-volatile matter
When evaporated to dryness and dried at 105°, leaves not more than 0.01% w/w of residue.

ASSAY
Weigh 5 g into a stopper flask containing 50 mL of *water* and titrate with *1M sodium hydroxide VS* using *phenolphthalein solution R1* as indicator. Each mL of *1M sodium hydroxide VS* is equivalent to 60.05 mg of $C_2H_4O_2$.

Acetone

(Ph. Eur. monograph 0872)



C_3H_6O 58.08 *67-64-1*

Ph Eur

DEFINITION
Propanone.

CHARACTERS
Appearance
Volatile, clear, colourless liquid.

Solubility
Miscible with water and with ethanol (96 per cent).

The vapour is flammable.

IDENTIFICATION
A. Relative density (see Tests).

B. To 1 mL, add 3 mL of *dilute sodium hydroxide solution R* and 0.3 mL of a 25 g/L solution of *sodium nitroprusside R*. An intense red colour is produced which becomes violet with the addition of 3.5 mL of *acetic acid R*.

C. To 10 mL of a 0.1 per cent *V/V* solution of the substance to be examined in *ethanol (50 per cent V/V) R*, add 1 mL of a 10 g/L solution of *nitrobenzaldehyde R* in *ethanol (50 per cent V/V) R* and 0.5 mL of *strong sodium hydroxide solution R*. Allow to stand for about 2 min and acidify with *acetic acid R*. A greenish-blue colour is produced.

TESTS
Appearance of solution
To 10 mL add 10 mL of *water R*. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 5 mL add 5 mL of *carbon dioxide-free water R*, 0.15 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M sodium hydroxide*. The solution is pink. Add 0.7 mL of *0.01 M hydrochloric acid* and 0.05 mL of *methyl red solution R*. The solution is red or orange.

Relative density (*2.2.5*)
0.790 to 0.793.

Reducing substances
To 30 mL add 0.1 mL of *0.02 M potassium permanganate* and allow to stand in the dark for 2 h. The mixture is not completely decolourised.

Related substances
Gas chromatography (*2.2.28*).

Test solution The substance to be examined.

Reference solution (a) To 0.5 mL of *methanol R* add 0.5 mL of *2-propanol R* and dilute to 100.0 mL with the test solution. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Reference solution (b) Dilute 100 µL of *benzene R* to 100.0 mL with the test solution. Dilute 0.20 mL of this solution to 100.0 mL with the test solution.

Column:
— *material*: fused silica,
— *size*: *l* = 50 m, Ø = 0.3 mm,
— *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Linear velocity 21 cm/s.

Split ratio 1:50.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 11	45 → 100
	11 - 20	100
Injection port		150
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Retention time Impurity C = about 7.5 min.

System suitability:
— *resolution*: minimum 5.0 between the peak due to impurity A (2nd peak) and the peak due to impurity B (3rd peak) in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 5 for the peak due to impurity C in the chromatogram obtained with reference solution (b).

Limits:
— *impurities A, B*: for each impurity, not more than the difference between the areas of the corresponding peaks in the chromatogram obtained with reference solution (a) and the areas of the corresponding peaks in the chromatogram obtained with the test solution (0.05 per cent *V/V*),
— *impurity C*: not more than the difference between the area of the peak due to impurity C in the chromatogram obtained with reference solution (b) and the area of the corresponding peak in the chromatogram obtained with the test solution (2 ppm *V/V*),
— *any other impurity*: for each impurity, not more than the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution (0.05 per cent *V/V*).

Matter insoluble in water
Dilute 1.0 mL to 20 mL with *water R*. The solution is clear (*2.2.1*).

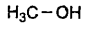
Residue on evaporation
Maximum 50 ppm.

Evaporate 20.0 g to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 1 mg.

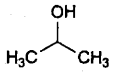
Water (*2.5.12*)
Maximum 3 g/L, determined on 10.0 mL.

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C.*



A. methanol,



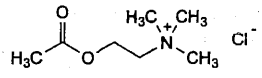
B. propan-2-ol (isopropanol),

C. benzene.

Ph Eur

Acetylcholine Chloride

(Ph. Eur. monograph 1485)



$C_7H_{16}ClNO_2$ 181.7 *60-31-1*

Action and use
Cholinoceptor agonist.

Ph Eur

DEFINITION
2-(Acetyloxy)-*N,N,N*-trimethylethanaminium chloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white crystalline powder or colourless crystals, very hygroscopic.

Solubility
Very soluble in water, freely soluble in alcohol, slightly soluble in methylene chloride.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Melting point (*2.2.14*): 149 °C to 152 °C.

Introduce the substance to be examined into a capillary tube. Dry in an oven at 100-105 °C for 3 h. Seal the tube and determine the melting point.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acetylcholine chloride CRS.*

C. Examine the chromatograms obtained in the test for related substances.

Results The principal zone in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal zone in the chromatogram obtained with reference solution (b).

D. To 15 mg add 10 mL of *dilute sodium hydroxide solution R*, 2 mL of *0.02 M potassium permanganate* and heat. The vapours formed change the colour of *red litmus paper R* to blue.

E. 0.5 mL of solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 or BY_6 (*2.2.2, Method II*).

Acidity

Dilute 1 mL of solution S to 10 mL with *carbon dioxide-free water R*. Add 0.05 mL of *phenolphthalein solution R*. Not more than 0.4 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances

Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 0.30 g of the substance to be examined in *methanol R* and dilute to 3.0 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dilute 1 mL of test solution (a) to 100 mL with *methanol R*.

Reference solution (b) Dissolve 20.0 mg of *acetylcholine chloride CRS* in *methanol R* and dilute to 2.0 mL with the same solvent.

Reference solution (c) Dissolve 20 mg of *choline chloride R* in *methanol R*, add 0.4 mL of test solution (a) and dilute to 2.0 mL with *methanol R*.

Plate *TLC silica gel plate R.*

Mobile phase Mix 20 volumes of a 40 g/L solution of *ammonium nitrate R*, 20 volumes of *methanol R* and 60 volumes of *acetonitrile R*.

Application 5 µL as bands of 10 mm by 2 mm.

Development Over 2/3 of the plate.

Detection Spray with *potassium iodobismuthate solution R3*.

System suitability The chromatogram obtained with reference solution (c) shows 2 clearly separated zones.

Limits:

— *any impurity*: any zones in the chromatogram obtained with test solution (a), apart from the principal zone, are not more intense than the principal zone in the chromatogram obtained with reference solution (a) (1 per cent).

Trimethylamine

Dissolve 0.1 g in 10 mL of *sodium carbonate solution R* and heat to boiling. No vapours appear which turn *red litmus paper R* blue.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

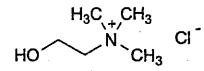
Dissolve 0.200 g in 20 mL of *carbon dioxide-free water R*. Neutralise with *0.01 M sodium hydroxide* using 0.15 mL of *phenolphthalein solution R* as indicator. Add 20.0 mL of *0.1 M sodium hydroxide* and allow to stand for 30 min. Titrate with *0.1 M hydrochloric acid*.

1 mL of *0.1 M sodium hydroxide* is equivalent to 18.17 mg of $C_7H_{16}ClNO_2$.

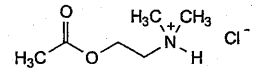
STORAGE

In ampoules, protected from light.

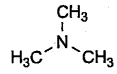
IMPURITIES



A. 2-hydroxy-*N*,*N*,*N*-trimethylethanaminium chloride (choline chloride),



B. 2-(acetyloxy)-*N*,*N*-dimethylethanaminium chloride,

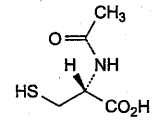


C. *N*,*N*-dimethylmethanamine.

Ph Eur

Acetylcysteine

(Ph. Eur. monograph 0967)



$C_5H_9NO_3S$ 163.2 *616-91-1*

Action and use

Sulfydryl donor; antidote to paracetamol poisoning; mucolytic.

Preparations

Acetylcysteine eye drops

Acetylcysteine Injection

Ph Eur

DEFINITION

(2*R*)-2-(Acetylamino)-3-sulfanylpropanoic acid.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water and in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, C.

Second identification: A, B, D, E.

A. Specific optical rotation (see Tests).

B. Melting point (*2.2.14*): 104 °C to 110 °C.

C. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium bromide R*.

Comparison *acetylcysteine CRS.*

D. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (b).

E. To 0.5 mL of solution S (see Tests) add 0.05 mL of a 50 g/L solution of *sodium nitroprusside R* and 0.05 mL of *concentrated ammonia R*. A dark violet colour develops.

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

2.0 to 2.8.

To 2 mL of solution S add 8 mL of *carbon dioxide-free water R* and mix.

Specific optical rotation (*2.2.7*)

+ 21.0 to + 27.0 (dried substance).

In a 25 mL volumetric flask, mix 1.25 g with 1 mL of a 10 g/L solution of *sodium edetate R*. Add 7.5 mL of a 40 g/L solution of *sodium hydroxide R*, mix and dissolve. Dilute to 25.0 mL with *phosphate buffer solution pH 7.0 R2*.

Related substances

Liquid chromatography (*2.2.29*). *Except where otherwise prescribed, prepare the solutions immediately before use.*

Test solution (a) Suspend 0.80 g of the substance to be examined in 1 mL of *1 M hydrochloric acid* and dilute to 100.0 mL with *water R*.

Test solution (b) Dilute 5.0 mL of test solution (a) to 100.0 mL with *water R*. Dilute 5.0 mL of this solution to 50.0 mL with *water R*.

Test solution (c) Use test solution (a) after storage for at least 1 h.

Reference solution (a) Suspend 4.0 mg of *acetylcysteine CRS*, 4.0 mg of *L-cystine R* (impurity A), 4.0 mg of *L-cysteine R* (impurity B), 4.0 mg of *acetylcysteine impurity C CRS* and 4.0 mg of *acetylcysteine impurity D CRS* in 1 mL of *1 M hydrochloric acid* and dilute to 100.0 mL with *water R*.

Reference solution (b) Suspend 4.0 mg of *acetylcysteine CRS* in 1 mL of *1 M hydrochloric acid* and dilute to 100.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Stir 3 volumes of *acetonitrile R* and 97 volumes of *water R* in a beaker; adjust to pH 3.0 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL, 3 times; inject *0.01 M hydrochloric acid* as a blank.

Run time 5 times the retention time of acetylcysteine (about 30 min).

Retention time Impurity A = about 2.2 min; impurity B = about 2.4 min; 2-methyl-2-thiazoline-4-carboxylic acid, originating in test solution (c) = about 3.3 min; acetylcysteine = about 6.4 min; impurity C = about 12 min; impurity D = about 14 min.

System suitability Reference solution (a):
- *resolution*: minimum 1.5 between the peaks due to impurities A and B and minimum 2.0 between the peaks due to impurities C and D.

From the chromatogram obtained with test solution (a), calculate the percentage content of the known impurities (T_1) and the unknown impurities (T_2) using the following equations:

$$T_1 = \frac{A_1 \times m_2 \times 100}{A_2 \times m_1}$$

$$T_2 = \frac{A_3 \times m_3 \times 100}{A_4 \times m_1}$$

A_1 = peak area of individual impurity (impurity A, impurity B, impurity C and impurity D) in the chromatogram obtained with test solution (a);
A_2 = peak area of the corresponding individual impurity (impurity A, impurity B, impurity C and impurity D) in the chromatogram obtained with reference solution (a);
A_3 = peak area of unknown impurity in the chromatogram obtained with test solution (a);
A_4 = peak area of acetylcysteine in the chromatogram obtained with reference solution (b);
m_1 = mass of the substance to be examined in test solution (a);
m_2 = mass of the individual impurity in reference solution (a);
m_3 = mass of acetylcysteine in reference solution (b).

Limits:
- *impurities A, B, C, D*: for each impurity, maximum 0.5 per cent;
- *any other impurity*: for each impurity, maximum 0.5 per cent;
- *total*: maximum 0.5 per cent;
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak with a retention time of about 3.3 min due to 2-methyl-2-thiazoline-4-carboxylic acid.

Zinc

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Dissolve 1.00 g in *0.001 M hydrochloric acid* and dilute to 50.0 mL with the same acid.

Reference solutions Prepare the reference solutions using *zinc standard solution (5 mg/mL Zn) R*, diluting with *0.001 M hydrochloric acid*.

Source Zinc hollow-cathode lamp.

Wavelength 213.8 nm.

Atomisation device Air-acetylene flame.

Use a correction procedure for non-specific absorption.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven *in vacuo* at 70 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.140 g in 60 mL of *water R* and add 10 mL of *dilute hydrochloric acid R*. After cooling in iced water, add 10 mL of *potassium iodide solution R* and titrate with *0.05 M iodine*, using 1 mL of *starch solution R* as indicator.

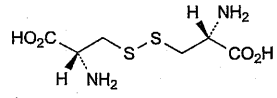
1 mL of *0.05 M iodine* is equivalent to 16.32 mg of $C_5H_9NO_3S$.

STORAGE

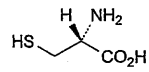
Protected from light.

IMPURITIES

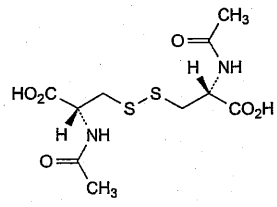
Specified impurities *A, B, C, D.*



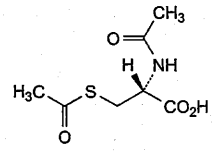
A. 3,3′-disulfanediylbis[(2*R*)-2-aminopropanoic acid] (L-cystine),



B. (2*R*)-2-amino-3-sulfanylpropanoic acid (L-cysteine),



C. (2*R*,2′*R*)-3,3′-disulfanediylbis[2-(acetylamino)propanoic acid] (*N*,*N*′-diacetyl-L-cystine),

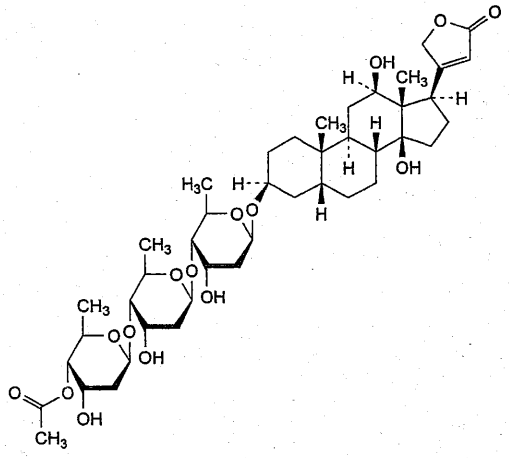


D. (2*R*)-2-(acetylamino)-3-(acetylsulfanyl)propanoic acid (*N*,*S*-diacetyl-L-cysteine).

Ph Eur

Acetyldigoxin

(β-Acetyldigoxin, Ph. Eur. monograph 2168)



$C_{43}H_{66}O_{15}$ 823 *5355-48-6*

Action and use

Cardiac Glycoside.

Ph Eur

DEFINITION

3β-[(4-*O*-Acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *β-acetyldigoxin CRS.*

TESTS

Specific optical rotation (*2.2.7*)

+ 26.2 to + 28.2 (dried substance).

Dissolve 0.50 g in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 25.0 mL with the same mixture of solvents.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mix equal volumes of *methanol R2* and *acetonitrile for chromatography R.*

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 10.0 mg of *β-acetyldigoxin CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *gitoxin CRS* (impurity D) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. To 5.0 mL of this solution, add 0.5 mL of reference solution (a) and dilute to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5.0 mg of *β-acetyldigoxin for peak identification CRS* (containing impurities A and B) in 10.0 mL of the solvent mixture.

Column:

— *size: l* = 0.125 m, Ø = 4.0 mm;

— *stationary phase: octadecylsilyl silica gel for chromatography R* (4 µm).

Mobile phase:

— *mobile phase A: water for chromatography R;*

— *mobile phase B: acetonitrile for chromatography R;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	70	30
10 - 20	70 → 35	30 → 65
20 - 20.1	35 → 70	65 → 30
20.1 - 25	70	30

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatograms obtained with reference solutions (c) and (d) to identify the peaks due to impurities A, B and D.

Relative retention With reference to β-acetyldigoxin (retention time = about 9 min): impurity B = about 0.3; impurity A = about 0.7; impurity D = about 1.2.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to β-acetyldigoxin and impurity D;
— *symmetry factor*: maximum 2.5 for the peak due to β-acetyldigoxin.

Limits:
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurity D*: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *any other impurity*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *sum of impurities other than A, B and D*: not more than 1.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.6 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Loss on drying (*2.2.32*)
Maximum 1.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{43}H_{66}O_{15}$ from the declared content of *β-acetyldigoxin CRS*.

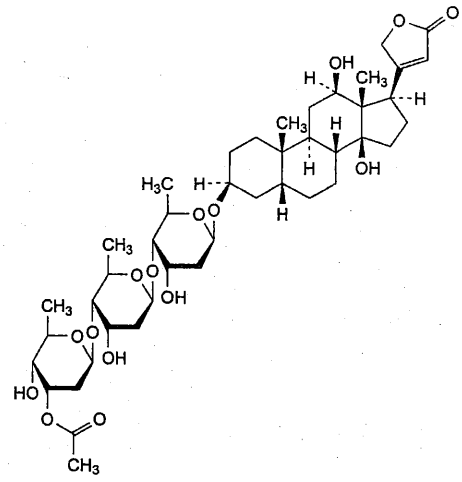
STORAGE

Protected from light.

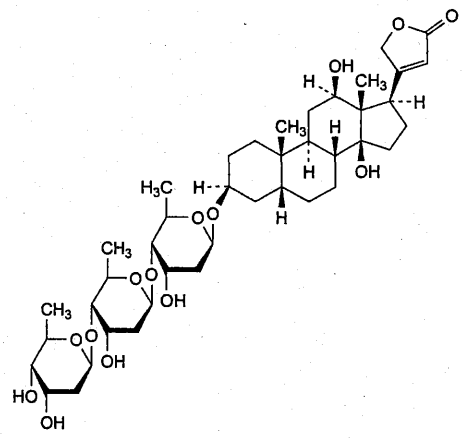
IMPURITIES

Specified impurities *A, B, D.*

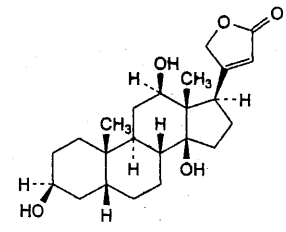
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, F, G, H.*



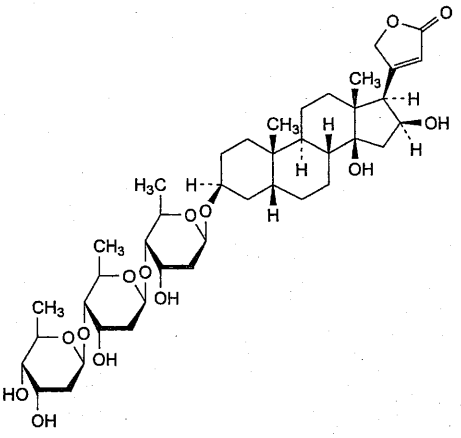
A. 3β-[(3-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (α-acetyldigoxin),



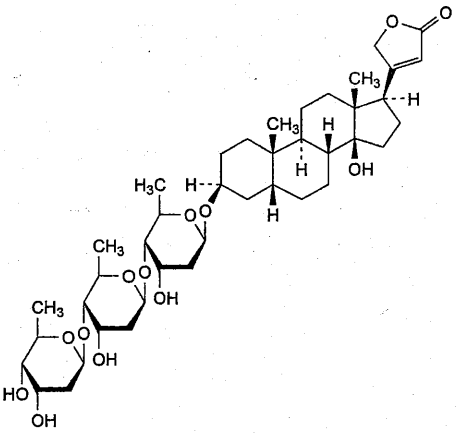
B. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (digoxin),



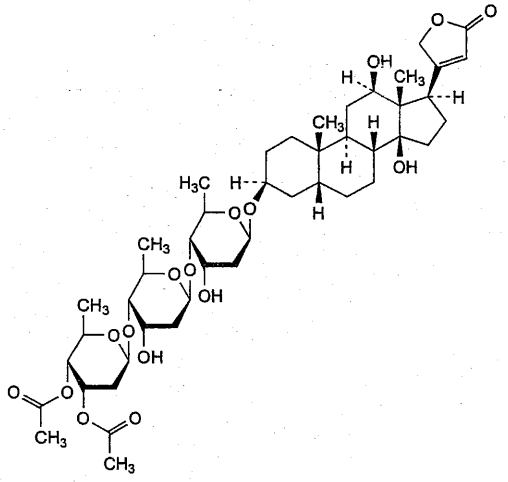
C. 3β,12β,14-trihydroxy-5β-card-20(22)-enolide (digoxigenin),



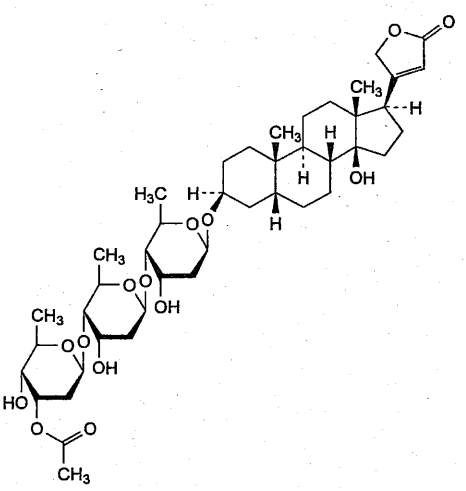
D. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14,16β-dihydroxy-5β-card-20(22)-enolide (gitoxin),



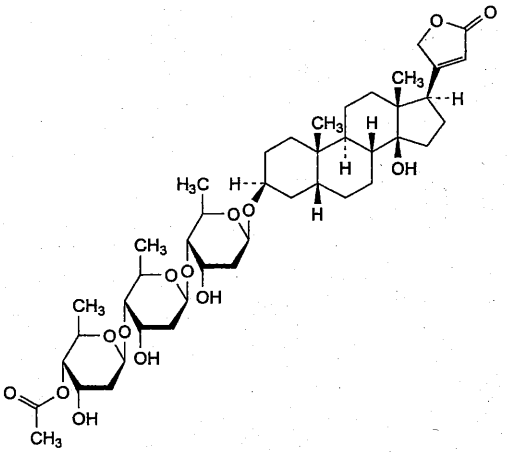
E. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14-hydroxy-5β-card-20(22)-enolide (digitoxin),



F. 3β-[(3,4-*O*-diacetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (diacetyldigoxin),



G. 3β-[(3-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14-hydroxy-5β-card-20(22)-enolide (α-acetyldigitoxin),



H. 3β-[(4-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14-hydroxy-5β-card-20(22)-enolide (β-acetyldigitoxin).

Ph Eur

Acetylene Intermix (1 per cent) in Nitrogen

(Ph. Eur. monograph 2903)

Ph Eur

DEFINITION

A mixture containing 1 per cent *V/V* of acetylene in *Low-oxygen nitrogen (1685)*.

Content

0.95 per cent *V/V* to 1.05 per cent *V/V* of acetylene (C_2H_2) in nitrogen (N_2).

This monograph applies to acetylene intermix (1 per cent) in nitrogen used in the preparation of lung function test gas mixtures for medicinal use.

PRODUCTION

The acetylene used in the manufacturing process is produced by hydrolysis of calcium carbide.

Prior to using the gas in the manufacturing process, the acetylene may be passed through an activated charcoal filter.

The acetylene is stored in cylinders, which may be filled with a porous mass with acetone the only solvent permitted.

CHARACTERS

Appearance

Colourless gas.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The peak due to acetylene in the chromatogram obtained with the gas to be examined is similar in retention time and size to the peak due to acetylene in the chromatogram obtained with the reference gas.

B. Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas *Nitrogen R1.*

Column:
- *material*: stainless steel;
- *size*: *l* = 2 m, Ø = 2 mm;
- *stationary phase*: *molecular sieve for chromatography R* (0.5 nm).

Carrier gas *helium for chromatography R.*

Flow rate 20 mL/min.

Temperature:
- *column*: 80 °C;
- *detector*: 130 °C.

Detection Thermal conductivity.

Injection 10 µL.

Retention time Nitrogen = about 2 min.

Results The principal peak in the chromatogram obtained with the gas to be examined is similar in retention time to the principal peak in the chromatogram obtained with the reference gas.

TESTS

Acetone

Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas Mixture containing 250 ppm *V/V* of *acetone R* in *nitrogen R.*

Column:
- *material*: fused silica;
- *size*: *l* = 10 m, Ø = 0.53 mm;
- *stationary phase*: *polyorganosiloxane for oxygen-containing compounds R* (film thickness 10 µm).

Carrier gas *helium for chromatography R.*

Flow rate 50 mL/min.

Temperature:
- *column*: 200 °C;
- *injection port*: 240 °C;
- *detector*: 250 °C.

Detection Flame ionisation.

Injection 25 µL.

Retention time Acetone = about 1 min.

Calculation of percentage content:
- use the concentration of acetone in the reference gas.

Limit:
- *acetone*: maximum 250 ppm *V/V.*

Arsine

Maximum 0.25 ppm *V/V*, determined using an arsine detector tube (*2.1.6*).

Phosphine

Maximum 0.2 ppm *V/V*, determined using a phosphine detector tube (*2.1.6*).

Hydrogen sulfide

Maximum 0.2 ppm *V/V*, determined using a hydrogen sulfide detector tube (*2.1.6*).

Water (*2.5.28*)

Maximum 10 ppm *V/V.*

ASSAY

Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas Mixture containing 1.0 per cent *V/V* of *acetylene R* in *nitrogen R1.*

Column:
- *material*: stainless steel;
- *size*: *l* = 2 m, Ø = 2 mm;
- *stationary phase*: 3 per cent *squalane R* on alumina.

Carrier gas *helium for chromatography R.*

Flow rate 20 mL/min.

Temperature:
- *column*: 100 °C;
- *detector*: 250 °C.

Detection Flame ionisation.

Injection 100 µL.

Retention time Acetylene = about 6 min.

Calculate the percentage content of C_2H_2.

STORAGE

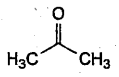
As a compressed gas, in appropriate high-pressure cylinders complying with the legal regulations.

LABELLING

The label states the nominal content, in per cent *V/V*, of acetylene in nitrogen.

IMPURITIES

Specified impurities *A, B, C, D, E.*



A. propan-2-one (acetone),

B. arsane (arsine),

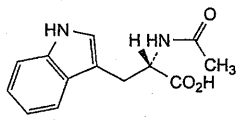
C. phosphane (phosphine),

D. hydrogen sulfide,

E. water.

Acetyltryptophan

(N-Acetyltryptophan, Ph. Eur. monograph 1383)



and enantiomer

$C_{13}H_{14}N_2O_3$ 246.3 *87-32-1*

Ph Eur

DEFINITION

(*RS*)-2-Acetylamino-3-(1*H*-indol-3-yl)propanoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

Tryptophan used for the production of *N*-acetyltryptophan complies with the test for impurity A and other related substances in the monograph on *Tryptophan (1272)*.

CHARACTERS

Appearance

White or almost white, crystalline powder, or colourless crystals.

Solubility

Slightly soluble in water, very soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

mp

About 205 °C.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D, E.

A. Optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *N-acetyltryptophan CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in 0.2 mL of *concentrated ammonia R* and dilute to 10 mL with *water R.*

Reference solution (a) Dissolve 50 mg of *N-acetyltryptophan CRS* in 0.2 mL of *concentrated ammonia R* and dilute to 10 mL with *water R.*

Reference solution (b) Dissolve 10 mg of *tryptophan R* in the test solution and dilute to 2 mL with the test solution.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (25:25:40 *V/V/V*).

Application 2 µL.

Development Over a path of 10 cm.

Drying In an oven at 100-105 °C for 15 min.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 2 mg in 2 mL of *water R.* Add 2 mL of *dimethylaminobenzaldehyde solution R6.* Heat on a water-bath. A blue or greenish-blue colour develops.

E. It gives the reaction of acetyl (*2.3.1*). Proceed as described for substances hydrolysable only with difficulty.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 or GY_7 (*2.2.2, Method II*).

Dissolve 1.0 g in a 40 g/L solution of *sodium hydroxide R* and dilute to 100 mL with the same alkaline solution.

Optical rotation (*2.2.7*)

−0.1° to + 0.1°.

Dissolve 2.50 g in a 40 g/L solution of *sodium hydroxide R* and dilute to 25.0 mL with the same alkaline solution.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the test and reference solutions immediately before use.*

Buffer solution pH 2.3 Dissolve 3.90 g of *sodium dihydrogen phosphate R* in 1000 mL of *water R.* Add about 700 mL of a 2.9 g/L solution of *phosphoric acid R* and adjust to pH 2.3 with the same acid solution.

Solvent mixture *acetonitrile R, water R* (10:90 *V/V*).

Test solution Dissolve 0.10 g of the substance to be examined in a mixture of 50 volumes of *acetonitrile R* and 50 volumes of *water R* and dilute to 20.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 4.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *1,1′-ethylidenebistryptophan CRS* in 1 mL of reference solution (b).

Column:

— *size: l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature:* 40 °C.

Mobile phase:

— *mobile phase A: acetonitrile R,* buffer solution pH 2.3 (115:885 *V/V*);

— *mobile phase B: acetonitrile R,* buffer solution pH 2.3 (350:650 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100	0
10 - 45	100 → 0	0 → 100
45 - 65	0	100

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (a) and (c).

Retention time *N*-acetyltryptophan = about 29 min; 1,1′-ethylidenebis(tryptophan) = about 34 min.

System suitability Reference solution (c):

— *resolution:* minimum 8.0 between the peaks due to *N*-acetyltryptophan and 1,1′-ethylidenebis(tryptophan); if necessary, adjust the time programme for the elution gradient (an increase in the duration of elution with mobile phase A produces longer retention times and a better resolution);

— *symmetry factor*: maximum 3.5 for the peak due to 1,1′-ethylidenebistryptophan in the chromatogram obtained with reference solution (c).

Limits:

— *impurities A, B, C, D, E, F, G, H, I, J, K, L*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);

— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.01 times the area of the principal peak the chromatogram obtained with reference solution (a) (0.01 per cent).

Ammonium (*2.4.1, Method B*)
Maximum 200 ppm, determined on 0.10 g.

Prepare the standard using 0.2 mL of *ammonium standard solution (100 ppm NH_4) R*.

Iron (*2.4.9*)
Maximum 10 ppm.

Dissolve 1.0 g in 50 mL of *hydrochloric acid R1*, with heating at 50 °C. Allow to cool. In a separating funnel, shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Examine the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 5 mL of *methanol R*. Add 50 mL of *anhydrous ethanol R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

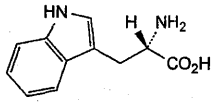
1 mL of *0.1 M sodium hydroxide* is equivalent to 24.63 mg of $C_{13}H_{14}N_2O_3$.

STORAGE

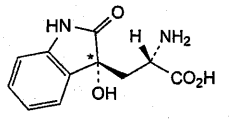
Protected from light.

IMPURITIES

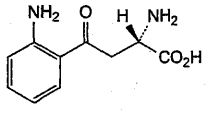
Specified impurities *A, B, C, D, E, F, G, H, I, J, K, L.*



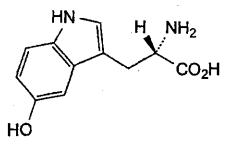
A. (*S*)-2-amino-3-(1*H*-indol-3-yl)propanoic acid (tryptophan),



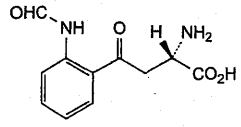
B. (*S*)-2-amino-3-[(3*RS*)-3-hydroxy-2-oxo-2,3-dihydro-1*H*-indol-3-yl]propanoic acid (dioxyindolylalanine),



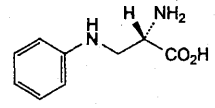
C. (*S*)-2-amino-4-(2-aminophenyl)-4-oxobutanoic acid (kynurenine),



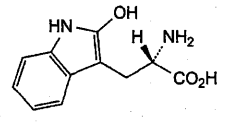
D. (*S*)-2-amino-3-(5-hydroxy-1*H*-indol-3-yl)propanoic acid (5-hydroxytryptophan),



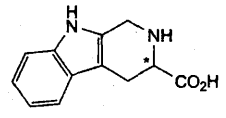
E. (*S*)-2-amino-4-[2-(formylamino)phenyl]-4-oxobutanoic acid (*N*-formylkynurenine),



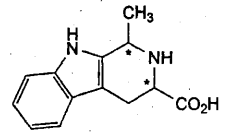
F. (*S*)-2-amino-3-(phenylamino)propanoic acid (3-phenylaminoalanine),



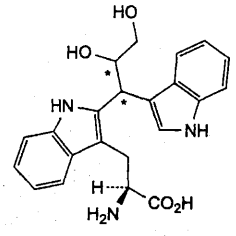
G. (*S*)-2-amino-3-(2-hydroxy-1*H*-indol-3-yl)propanoic acid (2-hydroxytryptophan),



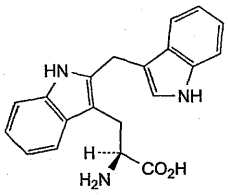
H. (3*RS*)-1,2,3,4-tetrahydro-9*H*-β-carboline-3-carboxylic acid,



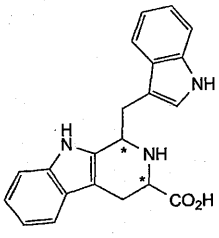
I. 1-methyl-1,2,3,4-tetrahydro-9*H*-β-carboline-3-carboxylic acid,



J. (*S*)-2-amino-3-[2-[2,3-dihydroxy-1-(1*H*-indol-3-yl) propyl]-1*H*-indol-3-yl]propanoic acid,



K. (*S*)-2-amino-3-[2-(1*H*-indol-3-ylmethyl)-1*H*-indol-3-yl] propanoic acid,

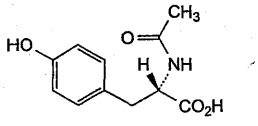


L. 1-(1*H*-indol-3-ylmethyl)-1,2,3,4-tetrahydro-9*H*-β-carboline-3-carboxylic acid.

Ph Eur

Acetyltyrosine

(N-Acetyltyrosine, Ph. Eur. monograph 1384)



$C_{11}H_{13}NO_4$ 223.2 *537-55-3*

Ph Eur

DEFINITION

(2*S*)-2-(Acetylamino)-3-(4-hydroxyphenyl)propanoic acid.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, practically insoluble in cyclohexane.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *N-acetyltyrosine CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 80 mg of the substance to be examined in a mixture of 3 volumes of *glacial acetic acid R*, 3 volumes of *water R* and 94 volumes of *anhydrous ethanol R*, and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 80 mg of *N-acetyltyrosine CRS* in a mixture of 3 volumes of *glacial acetic acid R*, 3 volumes of *water R* and 94 volumes of *anhydrous ethanol R*, and dilute to 10 mL with the same mixture of solvents.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *water R, glacial acetic acid R, ethyl acetate R* (10:15:75 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Solution S (see Tests) is strongly acid (*2.2.4*).

TESTS

Solution S

Dissolve 2.50 g in *water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 46 to + 49 (dried substance).

Dilute 10.0 mL of solution S to 25.0 mL with *water R*.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 20.0 mg of *tyrosine CRS* (impurity A) in 2 mL of a 40 g/L solution of *sodium hydroxide R* and dilute to 20.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with mobile phase A.

Reference solution (d) Dilute 1.0 mL of reference solution (b) to 20.0 mL with the test solution.

Column:

— *size*: *l* = 0.15 m, Ø = 3 mm;
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: mix 1.0 mL of *phosphoric acid R* and 1000 mL of *water for chromatography R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	97	3
2 - 15	97 → 62	3 → 38

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 219 nm.

Injection 2 µL of the test solution and reference solutions (a), (c) and (d).

Relative retention With reference to *N*-acetyltyrosine (retention time = about 6 min): impurity A = about 0.5.

System suitability Reference solution (d):

— *resolution*: minimum 5.0 between the principal peak and the peak due to impurity A.

Limits:
- — *impurity A*: not more than 0.8 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.8 per cent);
- — *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- — *total*: maximum 1.0 per cent;
- — *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 10 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 200 ppm.

Dissolve 1.0 g in *distilled water R* and dilute to 20 mL with the same solvent.

Ammonium (*2.4.1, Method B*)
Maximum 200 ppm, determined on 0.100 g.

Prepare the standard using 0.2 mL of *ammonium standard solution (100 ppm NH_4) R*.

Iron (*2.4.9*)
Maximum 20 ppm.

In a separating funnel, dissolve 0.5 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. The aqueous layer complies with the test.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 25 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

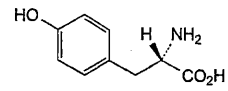
ASSAY
Dissolve 0.180 g in 50 mL of *carbon dioxide-free water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 22.32 mg of $C_{11}H_{13}NO_4$.

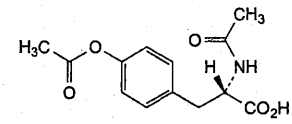
STORAGE
Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES
Specified impurities *A*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B*.



A. (2*S*)-2-amino-3-(4-hydroxyphenyl)propanoic acid (tyrosine),

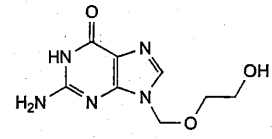


B. (2*S*)-2-(acetylamino)-3-[4-(acetoxy)phenyl]propanoic acid (diacetyltyrosine).

Ph Eur

Aciclovir

(Ph. Eur. monograph 0968)



$C_8H_{11}N_5O_3$ 225.2 *59277-89-3*

Action and use
Purine nucleoside analogue; antiviral (herpesviruses).

Preparations
Aciclovir Cream
Aciclovir Eye Ointment
Aciclovir Infusion
Aciclovir Oral Suspension
Aciclovir Tablets
Aciclovir Dispersible Tablets

Ph Eur

DEFINITION
2-Amino-9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6*H*-purin-6-one.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in heptane. It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *aciclovir CRS*.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 0.25 g in a 4 g/L solution of *sodium hydroxide R* and dilute to 25 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *dimethyl sulfoxide R, water R* (20:80 *V/V*).

Phosphate buffer solution pH 2.5 Dissolve 3.48 g of *dipotassium hydrogen phosphate R* in 1000 mL of *water R* and adjust to pH 2.5 with *phosphoric acid R*.

Phosphate buffer solution pH 3.1 Dissolve 3.48 g of *dipotassium hydrogen phosphate R* in 1000 mL of *water R* and adjust to pH 3.1 with *phosphoric acid R*.

Test solution Dissolve 25 mg of the substance to be examined in 5.0 mL of *dimethyl sulfoxide R* and dilute to 25.0 mL with *water R*.

Reference solution (a) Dissolve 5 mg of *aciclovir for system suitability CRS* (containing impurities A, B, J, K, N, O and P) in 1 mL of *dimethyl sulfoxide R* and dilute to 5.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *aciclovir for peak identification 1 CRS* (containing impurities C and I) in 200 µL of *dimethyl sulfoxide R* and dilute to 1.0 mL with *water R*.

Reference solution (d) Dissolve the contents of a vial of *aciclovir for peak identification 2 CRS* (containing impurities F and G) in 1.0 mL of reference solution (a).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: *acetonitrile R*, phosphate buffer solution pH 3.1 (1:99 *V/V*);
- *mobile phase B*: *acetonitrile R*, phosphate buffer solution pH 2.5 (50:50 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 27	100 → 80	0 → 20
27 - 40	80	20

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *aciclovir for peak identification 1 CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and I; use the chromatogram supplied with *aciclovir for peak identification 2 CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, F, G, J, K, N, O and P.

Relative retention With reference to aciclovir (retention time = about 13 min): impurity B = about 0.4; impurity P = about 0.7; impurity C = about 0.9; impurity N = about 1.37; impurities O and Q = about 1.42; impurity I = about 1.57; impurity J = about 1.62; impurity F = about 1.7; impurity A = about 1.8; impurities K and R = about 2.5; impurity G = about 2.6.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to impurity C and aciclovir in the chromatogram obtained with reference solution (c); minimum 1.5 between the peaks due to impurities F and A and minimum 1.5 between the peaks due to impurities K and G in the chromatogram obtained with reference solution (d).

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity I by 1.5;
- *impurity B*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
- *sum of impurities O and Q*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *sum of impurities K and R*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *impurities A, G, J, N, P*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *impurities C, F, I*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
- *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
- *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

Water (*2.5.12*)

Maximum 6.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14, Method D*)

Less than 0.50 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

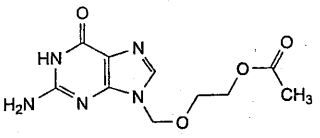
Dissolve 0.150 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of *0.1 M perchloric acid* is equivalent to 22.52 mg of $C_8H_{11}N_5O_3$.

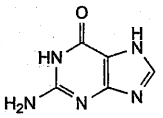
IMPURITIES

Specified impurities *A, B, C, F, G, I, J, K, N, O, P, Q, R.*

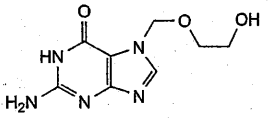
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *L, M.*



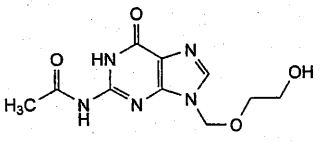
A. 2-[(2-amino-6-oxo-1,6-dihydro-9*H*-purin-9-yl)methoxy] ethyl acetate,



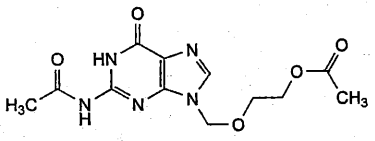
B. 2-amino-1,7-dihydro-6*H*-purin-6-one (guanine),



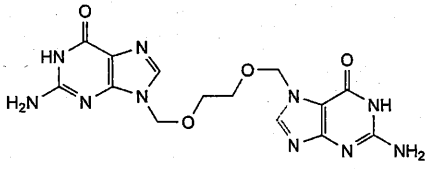
C. 2-amino-7-[(2-hydroxyethoxy)methyl]-1,7-dihydro-6*H*-purin-6-one,



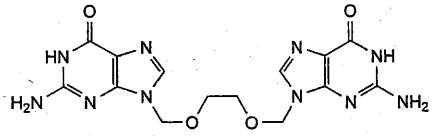
F. *N*-[9-[(2-hydroxyethoxy)methyl]-6-oxo-6,9-dihydro-1*H*-purin-2-yl]acetamide,



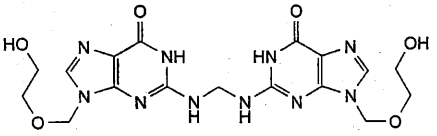
G. 2-[[2-(acetylamino)-6-oxo-1,6-dihydro-9*H*-purin-9-yl] methoxy]ethyl acetate,



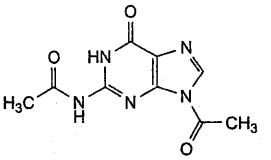
I. 2-amino-7-[[2-[(2-amino-6-oxo-1,6-dihydro-9*H*-purin-9-yl)methoxy]ethoxy]methyl]-1,7-dihydro-6*H*-purin-6-one,



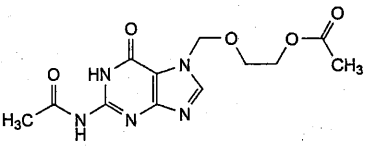
J. 9,9′-[ethylenebis(oxymethylene)]bis(2-amino-1,9-dihydro-6*H*-purin-6-one),



K. 2,2′-(methylenediimino)bis[9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6*H*-purin-6-one],



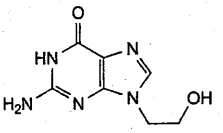
L. *N*-(9-acetyl-6-oxo-6,9-dihydro-1*H*-purin-2-yl)acetamide (N^2,9-diacetylguanine),



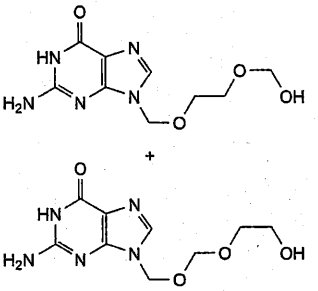
M. 2-[[2-(acetylamino)-6-oxo-1,6-dihydro-7*H*-purin-7-yl] methoxy]ethyl acetate,

N. unknown structure,

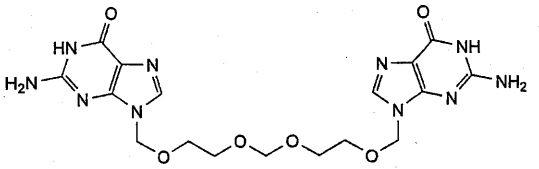
O. unknown structure,



P. 2-amino-9-(2-hydroxyethyl)-1,9-dihydro-6*H*-purin-6-one,



Q. mixture of 2-amino-9-[[2-(hydroxymethoxy) ethoxy]methyl]-1,9-dihydro-6*H*-purin-6-one and 2-amino-9-[[2-(hydroxyethoxy)methoxy]methyl]-1,9-dihydro-6*H*-purin-6-one,

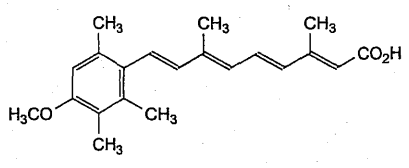


R. 9,9′-[methylenebis(oxyethyleneoxymethylene)]bis(2-amino-1,9-dihydro-6*H*-purin-6-one).

Ph Eur

Acitretin

(Ph. Eur. monograph 1385)



$C_{21}H_{26}O_3$ 326.4 *55079-83-9*

Action and use
Vitamin A analogue (retinoid); treatment of psoriasis; ichthyosis; Darier's disease.

Preparation
Acitretin Capsules

Ph Eur

DEFINITION

(2*E*,4*E*,6*E*,8*E*)-9-(4-Methoxy-2,3,6-trimethylphenyl)-3,7-dimethylnona-2,4,6,8-tetraenoic acid

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
Yellow or greenish-yellow, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in tetrahydrofuran, slightly soluble in acetone and in ethanol (96 per cent), very slightly soluble in cyclohexane.

It is sensitive to air, heat and light, especially in solution.

It shows polymorphism (*5.9*).

Carry out all operations as rapidly as possible and avoid exposure to actinic light; use freshly prepared solutions.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acitretin CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *2-propanol R* by heating under reflux; filter, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Test solution (a) Dissolve 25.0 mg of the substance to be examined in 5 mL of *tetrahydrofuran R* and dilute to 100.0 mL with *anhydrous ethanol R.*

Test solution (b) Dilute 10.0 mL of test solution (a) to 25.0 mL with *anhydrous ethanol R.*

Reference solution (a) Dissolve 25.0 mg of *acitretin CRS* in 5 mL of *tetrahydrofuran R* and dilute to 100.0 mL with *anhydrous ethanol R.* Dilute 10.0 mL of the solution to 25.0 mL with *anhydrous ethanol R.*

Reference solution (b) Dissolve 1 mg of *tretinoin CRS* in *anhydrous ethanol R* and dilute to 20.0 mL with the same solvent. Mix 5.0 mL of the solution with 2.5 mL of reference solution (a) and dilute to 100.0 mL with *anhydrous ethanol R.*

Reference solution (c) Dilute 1.0 mL of the test solution (a) to 100.0 mL with *anhydrous ethanol R.* Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R.*

Reference solution (d) Dissolve 2.5 mg of *acitretin for impurity A identification CRS* in 0.5 mL of *tetrahydrofuran R* and dilute to 10.0 mL with *anhydrous ethanol R.*

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: octadecylsilyl silica gel for chromatography for separation of polycyclic aromatic hydrocarbons R (5 µm);
— *temperature*: 25 °C.

Mobile phase 0.3 per cent *V/V* solution of *glacial acetic acid R* in a mixture of 8 volumes of *water for chromatography R* and 92 volumes of *anhydrous ethanol R.*

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 360 nm.

Autosampler Set at 4 °C.

Injection 10 µL of test solution (a) and reference solutions (b), (c) and (d).

Run time 2.5 times the retention time of acitretin.

Identification of impurities Use the chromatogram supplied with *acitretin for impurity A identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity A.

Relative retention With reference to acitretin (retention time = about 6 min): impurity A = about 0.8; tretinoin = about 0.85.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to tretinoin and acitretin.

Calculation of percentage contents:
— for each impurity, use the concentration of acitretin in reference solution (c).

Limits:
— *impurity A*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 100 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{21}H_{26}O_3$ taking into account the assigned content of *acitretin CRS.*

STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

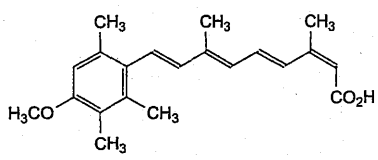
It is recommended that the contents of an opened container be used as soon as possible and any unused part be protected by an atmosphere of inert gas.

IMPURITIES

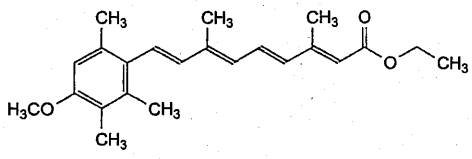
Specified impurities *A.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests

in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) B.



A. (2*Z*,4*E*,6*E*,8*E*)-9-(4-methoxy-2,3,6-trimethylphenyl)-3,7-dimethylnona-2,4,6,8-tetraenoic acid,

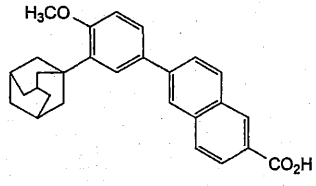


B. ethyl (2*E*,4*E*,6*E*,8*E*)-9-(4-methoxy-2,3,6-trimethylphenyl)-3,7-dimethylnona-2,4,6,8-tetraenoate.

Ph Eur

Adapalene

(Ph. Eur. monograph 2445)



$C_{28}H_{28}O_3$ 412.5 *106685-40-9*

Action and use
Vitamin A analogue (retinoid); treatment of acne.

Preparations
Adapalene Cream
Adapalene Gel

Ph Eur

DEFINITION
6-(4-Methoxy-3-tricyclo[3.3.1.1^{3,7}]dec-1-ylphenyl) naphthalene-2-carboxylic acid.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, sparingly soluble in tetrahydrofuran, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *adapalene CRS*.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).
Dissolve 0.2 g in *tetrahydrofuran R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *tetrahydrofuran R, acetonitrile R, water R* (20:37:43 *V/V/V*).
Test solution (a) Dissolve 40.0 mg of the substance to be examined in 10 mL of *tetrahydrofuran R*, add 7 mL of the solvent mixture and dilute to 20.0 mL with *tetrahydrofuran R*.
Test solution (b) Dissolve 20.0 mg of the substance to be examined in 50 mL of *tetrahydrofuran R*, add 35 mL of the solvent mixture and dilute to 100.0 mL with *tetrahydrofuran R*. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.
Reference solution (a) Dilute 1.0 mL of test solution (a) to 10.0 mL with *tetrahydrofuran R*. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.
Reference solution (b) Dissolve 2.4 mg of *adapalene impurity C CRS* in 2 mL of *tetrahydrofuran R* and dilute to 20.0 mL with the same solvent. Dilute 2.0 mL of the solution to 20.0 mL with the solvent mixture. To 2.0 mL of this solution add 2.0 mL of reference solution (a) and dilute to 20.0 mL with the solvent mixture.
Reference solution (c) Dissolve the contents of a vial of *adapalene for peak identification CRS* (containing impurities A, C and D) in 0.5 mL of *tetrahydrofuran R* and dilute to 1.0 mL with the solvent mixture.
Reference solution (d) Dissolve 20.0 mg of *adapalene CRS* in 50 mL of *tetrahydrofuran R*, add 35 mL of the solvent mixture and dilute to 100.0 mL with *tetrahydrofuran R*. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.
Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped phenylsilyl silica gel for chromatography R* (5 µm) with a carbon loading of 7.5 per cent;
— *temperature*: 30 °C.
Mobile phase:
— *mobile phase A*: *glacial acetic acid R, water R* (0.1:100 *V/V*);
— *mobile phase B*: *tetrahydrofuran R, acetonitrile R* (35:65 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2.5	50	50
2.5 - 40	50 → 28	50 → 72
40 - 42	28	72

Flow rate 1.2 mL/min.
Detection Spectrophotometer at 270 nm.
Injection 25 µL of test solution (a) and reference solutions (a), (b) and (c).
Identification of impurities Use the chromatogram supplied with *adapalene for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, C and D.

Relative retention With reference to adapalene (retention time = about 20 min): impurity A = about 0.3; impurity C = about 0.9; impurity D = about 1.9.

System suitability Reference solution (b):
- *resolution*: minimum 4.5 between the peaks due to impurity C and adapalene;
- *signal-to-noise ratio*: minimum 10 for the peak due to impurity C.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7; impurity C = 7; impurity D = 1.4;
- *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

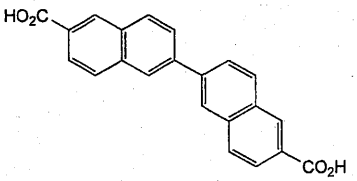
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

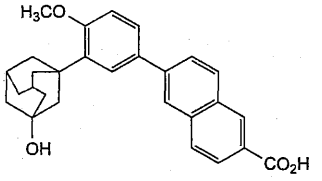
Calculate the percentage content of adapalene from the declared content of *adapalene CRS*.

IMPURITIES
Specified impurities *A, C, D.*

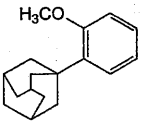
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) B.



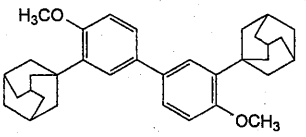
A. 2,2′-binaphthalene-6,6′-dicarboxylic acid,



B. 6-[3-(3-hydroxytricyclo[3.3.1.1^{3,7}]dec-1-yl)-4-methoxyphenyl]naphthalene-2-carboxylic acid,



C. 1-(2-methoxyphenyl)tricyclo[3.3.1.1^{3,7}]decane,

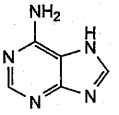


D. 1,1′-[4,4′-bis(methoxy)biphenyl-3,3′-diyl]bis(tricyclo[3.3.1.1^{3,7}]decane).

Ph Eur

Adenine

(Ph. Eur. monograph 0800)



$C_5H_5N_5$ 135.1 *73-24-5*

Action and use
Constituent of anticoagulant and preservative solutions for blood.

Ph Eur

DEFINITION
Adenine contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of 7*H*-purin-6-amine, calculated with reference to the dried substance.

CHARACTERS
A white or almost white powder, very slightly soluble in water and in alcohol. It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *adenine CRS*. Examine the substances prepared as discs.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 1 g add 3.5 mL of *propionic anhydride R* and boil for 15 min with stirring. Cool. To the resulting crystalline mass

add 15 mL of *light petroleum R* and heat to boiling with vigorous stirring. Cool and filter. Wash the precipitate with two quantities, each of 5 mL, of *light petroleum R*. Dissolve the precipitate in 10 mL of *water R* and boil for 1 min. Filter the mixture at 30 °C to 40 °C. Allow to cool. Filter, and dry the precipitate at 100 °C to 105 °C for 1 h. The melting point (*2.2.14*) of the precipitate is 237 °C to 241 °C.

TESTS

Solution S

Suspend 2.5 g in 50 mL of *distilled water R* and boil for 3 min. Cool and dilute to 50 mL with *distilled water R*. Filter. Use the filtrate as solution S.

Appearance of solution

Dissolve 0.5 g in *dilute hydrochloric acid R* and dilute to 50 mL with the same acid. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *bromothymol blue solution R1* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is blue. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is yellow.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *dilute acetic acid R*, with heating if necessary, and dilute to 10 mL with the same acid.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *dilute acetic acid R*.

Reference solution (a) Dissolve 10 mg of *adenine CRS* in *dilute acetic acid R*, with heating if necessary, and dilute to 10 mL with the same acid.

Reference solution (b) Dilute 1 mL of test solution (b) to 20 mL with *dilute acetic acid R*.

Reference solution (c) Dissolve 10 mg of *adenine CRS* and 10 mg of *adenosine R* in *dilute acetic acid R*, with heating if necessary, and dilute to 10 mL with the same acid.

Apply to the plate 5 µL of each solution. Develop over a path of 12 cm using a mixture of 20 volumes of *concentrated ammonia R*, 40 volumes of *ethyl acetate R* and 40 volumes of *propanol R*. Dry the plate in a current of warm air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated spots.

Chlorides (*2.4.4*)

To 10 mL of solution S add 1 mL of *concentrated ammonia R* and 3 mL of *silver nitrate solution R2*. Filter. Wash the precipitate with a little *water R* and dilute the filtrate to 15 mL with *water R*. The solution complies with the limit test for chlorides (100 ppm). When carrying out the test, add 2 mL of *dilute nitric acid R* instead of 1 mL of *dilute nitric acid R*.

Sulfates (*2.4.13*)

Dilute 10 mL of solution S to 15 mL with *distilled water R*. The solution complies with the limit test for sulfates (300 ppm).

Ammonium

Prepare a cell consisting of two watch-glasses 60 mm in diameter placed edge to edge. To the inner wall of the upper watch-glass stick a piece of *red litmus paper R* 5 mm square and wetted with a few drops of *water R*. Finely powder the substance to be examined, place 0.5 g in the lower watch-glass and suspend in 0.5 mL of *water R*. To the suspension add 0.30 g of *heavy magnesium oxide R*. Briefly triturate with a glass rod. Immediately close the cell by putting the two watch-glasses together. Heat at 40 °C for 15 min. The litmus paper is not more intensely blue coloured than a standard prepared at the same time and in the same manner using 0.05 mL of *ammonium standard solution (100 ppm* NH_4*) R*, 0.5 mL of *water R* and 0.30 g of *heavy magnesium oxide R* (10 ppm).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

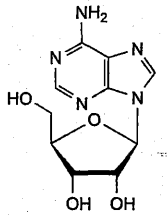
Dissolve 0.100 g in a mixture of 20 mL of *acetic anhydride R* and 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 13.51 mg of $C_5H_5N_5$.

Ph Eur

Adenosine

(Ph. Eur. monograph 1486)



$C_{10}H_{13}N_5O_4$ 267.2 *58-61-7*

Action and use

Antiarrhythmic.

Ph Eur

DEFINITION

9-β-D-Ribofuranosyl-9*H*-purin-6-amine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, soluble in hot water, practically insoluble in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute mineral acids.

mp

About 234 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *adenosine CRS*.

TESTS

Solution S

Suspend 5.0 g in 100 mL of *distilled water R* and heat to boiling. Allow to cool, filter with the aid of vacuum and dilute to 100 mL with *distilled water R*.

Appearance of solution

Solution S is colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 10 mL of solution S, add 0.1 mL of *bromocresol purple solution R* and 0.1 mL of *0.01 M hydrochloric acid*. The solution is yellow. Add 0.4 mL of *0.01 M sodium hydroxide*. The solution is violet-blue.

Specific optical rotation (*2.2.7*)

−45 to −49 (dried substance).

Dissolve 1.25 g in *1 M hydrochloric acid* and dilute to 50.0 mL with the same acid. Examine within 10 min of preparing the solution.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Dissolve 6.8 g of *potassium hydrogen sulfate R* and 3.4 g of *tetrabutylammonium hydrogen sulfate R* in *water R*, adjust to pH 6.5 with a 60 g/L solution of *potassium hydroxide R* and dilute to 1000 mL with the same solvent. Use a freshly prepared solvent mixture.

Test solution Dissolve 20 mg of the substance to be examined in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *adenine R* (impurity A) and 5 mg of *inosine R* (impurity G) in the mobile phase and dilute to 50 mL with the mobile phase. Dilute 4 mL of this solution to 100 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water R*, solvent mixture (40:60 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 1.5 times the retention time of adenosine.

Relative retention With reference to adenosine (retention time = about 13 min): impurity A = about 0.3; impurity G = about 0.4.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the peaks due to impurities A and G.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity G = 1.4;

— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurity G*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)

Maximum 100 ppm.

Dilute 10 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 200 ppm, determined on solution S.

Ammonium (*2.4.1, Method B*)

Maximum 10 ppm, determined on 0.5 g.

Prepare the standard using 5 mL of *ammonium standard solution (1 ppm NH_4) R*.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

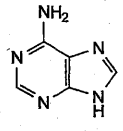
Dissolve 0.200 g, warming slightly if necessary, in a mixture of 20 mL of *acetic anhydride R* and 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 26.72 mg of $C_{10}H_{13}N_5O_4$.

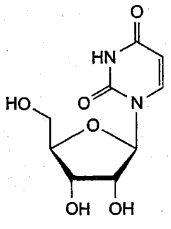
IMPURITIES

Specified impurities *A, G.*

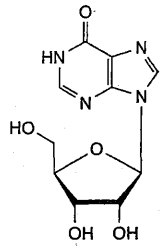
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, H.*



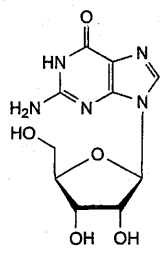
A. 7*H*-purin-6-amine (adenine),



F. 1-β-D-ribofuranosylpyrimidine-2,4(1*H*,3*H*)-dione (uridine),



G. 9-β-D-ribofuranosyl-1,9-dihydro-6*H*-purin-6-one (inosine),

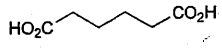


H. 2-amino-9-β-D-ribofuranosyl-1,9-dihydro-6*H*-purin-6-one (guanosine).

Ph Eur

Adipic Acid

(Ph. Eur. monograph 1586)



$C_6H_{10}O_4$ 146.1 *124-04-9*

Action and use
Excipient.

Ph Eur

DEFINITION
Hexanedioic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, soluble in boiling water, freely soluble in ethanol (96 per cent) and in methanol, soluble in acetone.

IDENTIFICATION
A. Melting point (*2.2.14*): 151 °C to 154 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *adipic acid CRS.*

TESTS

Solution S
Dissolve 5.0 g with heating in *distilled water R* and dilute to 50 mL with the same solvent. Allow to cool and to crystallise. Filter through a sintered-glass filter (40) (*2.1.2*). Wash the filter with *distilled water R*. Collect the filtrate and the washings until a volume of 50 mL is obtained.

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.20 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 20 mg of *glutaric acid R* in 1.0 mL of the test solution and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase, dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm,
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm) with a specific surface area of 350 m^2/g and a pore size of 10 nm,
— *temperature*: 30 °C.

Mobile phase Mix 3 volumes of *acetonitrile R* and 97 volumes of a 24.5 g/L solution of *dilute phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 209 nm.

Injection 20 µL.

Run time 3 times the retention time of adipic acid.

System suitability Reference solution (a):
— *resolution*: minimum 9.0 between the peaks due to glutaric acid and adipic acid.

Limits:
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with *water R*.

Nitrates
Maximum 30 ppm.

To 1 mL of solution S add 2 mL of *concentrated ammonia R*, 0.5 mL of a 10 g/L solution of manganese sulfate R, 1 mL of a 10 g/L solution of *sulfanilamide R* and dilute to 20 mL with *water R*. Add 0.10 g of *zinc powder R* and cool in iced water for 30 min; shake from time to time. Filter and cool 10 mL of the filtrate in iced water. Add 2.5 mL of *hydrochloric acid R1* and 1 mL of a 10 g/L solution of *naphthylethylenediamine dihydrochloride R*. Allow to stand at room temperature. After 15 min the mixture is not more intensely coloured than a standard prepared at the same time and in the same manner, using 1.5 mL of *nitrate standard solution (2 ppm NO_3) R* instead of 1 mL of solution S. The test is invalid if a blank solution prepared at the same time and in the same manner, using 1 mL of *water R* instead of 1 mL of solution S, is more intensely coloured than a 2 mg/L solution of *potassium permanganate R*.

Sulfates (*2.4.13*)
Maximum 500 ppm.

Dilute 3 mL of solution S to 15 mL with *distilled water R*.

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S.

Loss on drying (*2.2.32*)
Maximum 0.2 per cent, determined on 1.000 g by drying in an oven at 105 °C.

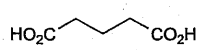
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent.

Melt 1.0 g completely over a gas burner, then ignite the melted substance with the burner. After ignition, lower or remove the flame in order to prevent the substance from boiling and keep it burning until completely carbonised. Carry out the test for sulfated ash using the residue.

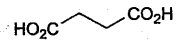
ASSAY
Dissolve 60.0 mg in 50 mL of *water R*. Add 0.2 mL of *phenolphthalein solution R* and titrate with *0.1 M sodium hydroxide*.

1 mL of *0.1 M sodium hydroxide* is equivalent to 7.31 mg of $C_6H_{10}O_4$.

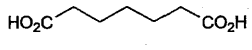
IMPURITIES



A. pentanedioic acid (glutaric acid),



B. butanedioic acid (succinic acid),

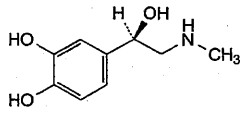


C. heptanedioic acid (pimelic acid).

Ph Eur

Adrenaline / Epinephrine

(Ph. Eur. monograph 2303)



$C_9H_{13}NO_3$ 183.2 *51-43-4*

Action and use
Adrenoceptor agonist.

Preparations
Adrenaline Eye Drops/Epinephrine Eye Drops

Dilute Adrenaline Injection (1 in 10,000)/Dilute Epinephrine Injection (1 in 10,000)

Ph Eur

DEFINITION
4-[(1*R*)-1-Hydroxy-2-(methylamino)ethyl]benzene-1,2-diol.

Synthetic product.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white crystalline powder, becoming coloured on exposure to air and light.

Solubility
Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride. It dissolves in hydrochloric acid.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *adrenaline CRS*.

B. Specific optical rotation (see Tests).

TESTS
Solution S
Dissolve 1.000 g in a 25.75 g/L solution of *hydrochloric acid R* and dilute to 50.0 mL with the same solvent. Examine the solution immediately.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
−50.0 to −54.0 (dried substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions protected from light.*

Solvent mixture A Dissolve 5.0 g of *potassium dihydrogen phosphate R* and 2.6 g of *sodium octanesulfonate R* in *water for chromatography R* and dilute to 1000 mL with the same solvent (it is usually necessary to stir for at least 30 min to achieve complete dissolution). Adjust to pH 2.8 with *phosphoric acid R*.

Solvent mixture B *acetonitrile R1*, solvent mixture A (13:87 *V/V*).

Test solution Dissolve 40 mg of the substance to be examined in 5 mL of *0.1 M hydrochloric acid* and dilute to 50.0 mL with solvent mixture B.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solvent mixture B. Dilute 1.0 mL of this solution to 10.0 mL with solvent mixture B.

Reference solution (b) Dissolve 1.5 mg of *noradrenaline tartrate CRS* (impurity B) and 1.5 mg of *adrenalone hydrochloride R* (impurity C) in solvent mixture B, add 1.0 mL of the test solution and dilute to 100 mL with solvent mixture B.

Reference solution (c) Dissolve the contents of a vial of *adrenaline impurity mixture CRS* (containing impurities D and E) in 1.0 mL of the blank solution.

Reference solution (d) Dissolve 4 mg of *adrenaline with impurity F CRS* in 0.5 mL of *0.1 M hydrochloric acid* and dilute to 5 mL with solvent mixture B.

Blank solution *0.1 M hydrochloric acid*, solvent mixture B (1:9 *V/V*).

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, solvent mixture A (5:95 *V/V*);
— *mobile phase B*: *acetonitrile R1*, solvent mixture A (45:55 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	92 → 50	8 → 50
15 - 20	50 → 92	50 → 8
20 - 25	92	8

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *adrenaline impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D and E; use the chromatogram supplied with *adrenaline with impurity F CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity F.

Relative retention With reference to adrenaline (retention time = about 4 min): impurity F = about 0.2; impurity B = about 0.8; impurity C = about 1.3; impurity D = about 3.3; impurity E = about 3.7.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to impurity B and adrenaline.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.7; impurity E = 0.6;
— *impurities B, C, F*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurities D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 18 h.

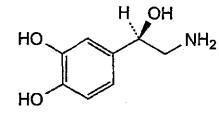
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

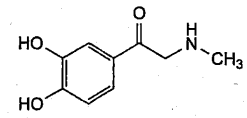
1 mL of *0.1 M perchloric acid* is equivalent to 18.32 mg of $C_9H_{13}NO_3$.

STORAGE
Under nitrogen, protected from light.

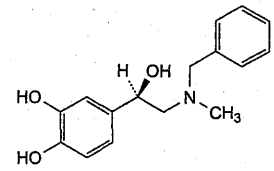
IMPURITIES
Specified impurities *B, C, D, E, F.*



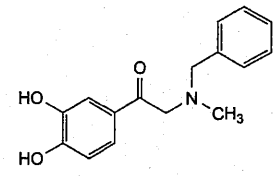
B. (1*R*)-2-amino-1-(3,4-dihydroxyphenyl)ethanol (noradrenaline),



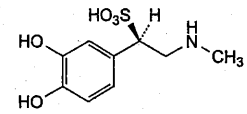
C. 1-(3,4-dihydroxyphenyl)-2-(methylamino)ethanone (adrenalone),



D. 4-[(1*R*)-2-(benzylmethylamino)-1-hydroxyethyl]benzene-1,2-diol,



E. 2-(benzylmethylamino)-1-(3,4-dihydroxyphenyl)ethanone,

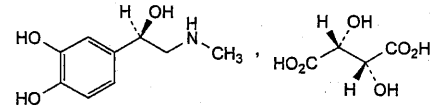


F. (1*R*)-1-(3,4-dihydroxyphenyl)-2-(methylamino) ethanesulfonic acid.

Ph Eur

Adrenaline Acid Tartrate / Epinephrine Acid Tartrate

(Adrenaline Tartrate, Ph. Eur. monograph 0254)



$C_{13}H_{19}NO_9$ 333.3 *51-42-3*

Action and use
Adrenoceptor agonist.

Preparations
Adrenaline Injection/Epinephrine Injection

Dilute Adrenaline Injection (1 in 10,000)/Dilute Epinephrine Injection (1 in 10,000)

Adrenaline Solution/Epinephrine Solution

Adrenaline and Cocaine Intranasal Solution

Bupivacaine and Adrenaline Injection/Bupivacaine and Epinephrine Injection

Lidocaine and Adrenaline Injection/Lidocaine and Epinephrine Injection

Ph Eur

DEFINITION

(1*R*)-1-(3,4-Dihydroxyphenyl)-2-(methylamino)ethanol hydrogen (2*R*,3*R*)-2,3-dihydroxybutanedioate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or greyish-white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 5 g in 50 mL of a 5 g/L solution of *sodium metabisulfite R* and make alkaline by addition of *ammonia R*. Keep the mixture at room temperature for at least 15 min and filter. Reserve the filtrate for identification test C. Wash the precipitate with 3 quantities, each of 10 mL, of *methanol R*. Dry at 80 °C. The specific optical rotation (*2.2.7*) of the residue (adrenaline base) is −53.5 to −50, determined using a 20.0 g/L solution in *0.5 M hydrochloric acid*.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of adrenaline base prepared as described under identification test A.

Comparison Use adrenaline base prepared as described under identification test A from 50 mg of *adrenaline tartrate CRS* dissolved in 5 mL of a 5 g/L solution of *sodium metabisulfite R*. Keep the mixture at room temperature for at least 30 min. Filter through a sintered-glass filter (*2.1.2*).

C. 0.2 mL of the filtrate obtained in identification test A gives reaction (b) of tartrates (*2.3.1*).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 0.5 g in *water R* and dilute to 10 mL with the same solvent. Examine the solution immediately.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions protected from light.*

Solvent mixture A Dissolve 5.0 g of *potassium dihydrogen phosphate R* and then 2.6 g of *sodium octanesulfonate R* in *water for chromatography R*, and dilute to 1000 mL with the same solvent (it is usually necessary to stir for at least 30 min to achieve complete dissolution). Adjust to pH 2.8 with *phosphoric acid R*.

Solvent mixture B *acetonitrile R1*, solvent mixture A (130:870 *V/V*).

Test solution Dissolve 75 mg of the substance to be examined in 5 mL of *0.1 M hydrochloric acid* and dilute to 50 mL with solvent mixture B.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solvent mixture B. Dilute 1.0 mL of this solution to 10.0 mL with solvent mixture B.

Reference solution (b) Dissolve 1.5 mg of *noradrenaline tartrate CRS* (impurity B) and 1.5 mg of *adrenalone hydrochloride R* (impurity C) in solvent mixture B, add 1.0 mL of the test solution and dilute to 100.0 mL with solvent mixture B.

Reference solution (c) Dissolve the contents of a vial of *adrenaline impurity mixture CRS* (impurities D and E) in 0.1 mL of *0.1 M hydrochloric acid* and 0.9 mL of solvent mixture B.

Reference solution (d) Dissolve 7.5 mg of *adrenaline tartrate with impurity A CRS* in 0.5 mL of *0.1 M hydrochloric acid* and dilute to 5.0 mL with solvent mixture B.

Blank solution *0.1 M hydrochloric acid*, solvent mixture B (1:9 *V/V*).

Column:

- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 50 °C.

Mobile phase:

- *mobile phase A*: *acetonitrile R1*, solvent mixture A (5:95 *V/V*);
- *mobile phase B*: *acetonitrile R1*, solvent mixture A (45:55 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	92 → 50	8 → 50
15 - 20	50 → 92	50 → 8
20 - 25	92	8

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *adrenaline impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D and E; use the chromatogram supplied with *adrenaline tartrate with impurity A CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity A.

Relative retention With reference to adrenaline (retention time = about 4 min): impurity B = about 0.8; impurity C = about 1.3; impurity A = about 3.2; impurity D = about 3.3; impurity E = about 3.7.

System suitability Reference solution (b):

- *resolution*: minimum 3.0 between the peaks due to impurity B and adrenaline.

Limits:

- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.7; impurity E = 0.6;
- *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurities B, C*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurities D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* for 18 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of *anhydrous acetic acid R*, heating gently if necessary. Titrate with *0.1 M perchloric acid* until a bluish-green colour is obtained, using 0.1 mL of *crystal violet solution R* as indicator.

1 mL of *0.1 M perchloric acid* is equivalent to 33.33 mg of $C_{13}H_{19}NO_9$.

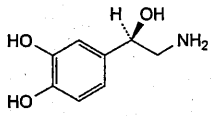
STORAGE

In an airtight container, or preferably in a sealed tube under vacuum or under an inert gas, protected from light.

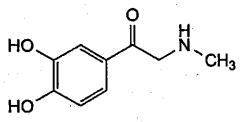
IMPURITIES

Specified impurities *A, B, C, D, E.*

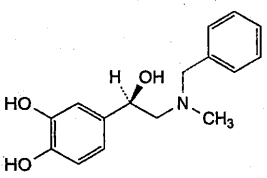
A. unknown structure,



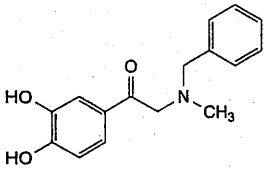
B. (1*R*)-2-amino-1-(3,4-dihydroxyphenyl)ethanol (noradrenaline),



C. 1-(3,4-dihydroxyphenyl)-2-(methylamino)ethanone (adrenalone),



D. 4-[(1*R*)-2-(benzylmethylamino)-1-hydroxyethyl]benzene-1,2-diol,



E. 2-(benzylmethylamino)-1-(3,4-dihydroxyphenyl)ethanone.

Ph Eur

Agar

(Ph. Eur. monograph 0310)

Action and use
Excipient.

Ph Eur

DEFINITION

Polysaccharides from various species of Rhodophyceae mainly belonging to the genus *Gelidium*. It is prepared by treating the algae with boiling water; the extract is filtered whilst hot, concentrated and dried.

CHARACTERS

Appearance
Powder or crumpled strips 2-5 mm wide or sometimes flakes, colourless or pale yellow, translucent, somewhat tough and difficult to break, becoming more brittle on drying.

Mucilaginous taste.

IDENTIFICATION

A. Examine under a microscope. When mounted in 0.005 M iodine, the strips or flakes are partly stained brownish-violet. Magnified 100 times, they show the following diagnostic characters: numerous minute, colourless, ovoid or rounded grains on an amorphous background; occasional brown, round or ovoid spores with a reticulated surface, measuring up to 60 µm, may be present. Reduce to a powder, if necessary. The powder is yellowish-white. Examine under a microscope using 0.005 M iodine. The powder presents angular fragments with numerous grains similar to those seen in the strips and flakes; some of the fragments are stained brownish-violet.

B. Dissolve 0.1 g with heating in 50 mL of *water R*. Cool. To 1 mL of the mucilage carefully add 3 mL of *water R* so as to form 2 separate layers. Add 0.1 mL of *0.05 M iodine*. A dark brownish-violet colour appears at the interface. Mix. The liquid becomes pale yellow.

C. Heat 5 mL of the mucilage prepared for identification test B on a water-bath with 0.5 mL of *hydrochloric acid R* for 30 min. Add 1 mL of *barium chloride solution R1*. A white turbidity develops within 30 min.

D. Heat 0.5 g with 50 mL of *water R* on a water-bath until dissolved. Only a few fragments remain insoluble. During cooling, the solution gels between 35 °C and 30 °C. Heat the gel thus obtained on a water-bath; it does not liquefy below 80 °C.

TESTS

Swelling index (*2.8.4*)
Minimum 10 and within 10 per cent of the value stated on the label, determined on the powdered herbal drug (355) (*2.9.12*).

Insoluble matter
Maximum 1.0 per cent.

To 5.00 g of the powdered herbal drug (355) (*2.9.12*) add 100 mL of *water R* and 14 mL of *dilute hydrochloric acid R*. Boil gently for 15 min with frequent stirring. Filter the hot liquid through a tared, sintered-glass filter (160) (*2.1.2*), rinse the filter with hot *water R* and dry at 100-105 °C.
The residue weighs a maximum of 50 mg.

Gelatin
To 1.00 g add 100 mL of *water R* and heat on a water-bath until dissolved. Allow to cool to 50 °C. To 5 mL of this

solution add 5 mL of *picric acid solution R*. No turbidity appears within 10 min.

Loss on drying (*2.2.32*)
Maximum 20.0 per cent, determined on 1.000 g of the powdered herbal drug (355) (*2.9.12*) by drying in an oven at 105 °C.

Total ash (*2.4.16*)
Maximum 5.0 per cent.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

LABELLING
The label states the swelling index.

Ph Eur

Medical Air

(Medicinal Air, Ph. Eur. monograph 1238)

When Medical Air is intended for use in a room in which magnetic resonance imaging (MRI) is being performed, the cylinder and fittings should be made from suitable non-ferromagnetic materials and labelled accordingly.

Ph Eur

DEFINITION
Compressed ambient air.

Content
20.4 per cent *V/V* to 21.4 per cent *V/V* of oxygen (O_2).

CHARACTERS

Appearance
Colourless gas.

Solubility
At 20 °C at a pressure of 101 kPa, 1 volume dissolves in about 50 volumes of water.

PRODUCTION

Carbon dioxide
Maximum 500 ppm *V/V*, determined using an infrared analyser (*2.5.24*).

Gas to be examined Filter the substance to be examined to avoid stray light phenomena.

Reference gas (a) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing less than 1 ppm *V/V* of *carbon dioxide R1*.

Reference gas (b) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing 500 ppm *V/V* of *carbon dioxide R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon dioxide in the gas to be examined.

Carbon monoxide
Maximum 5 ppm *V/V*, determined using an infrared analyser (*2.5.25*).

Gas to be examined Filter the substance to be examined to avoid stray light phenomena.

Reference gas (a) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing less than 1 ppm *V/V* of *carbon monoxide R*.

Reference gas (b) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing 5 ppm *V/V* of *carbon monoxide R*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon monoxide in the gas to be examined.

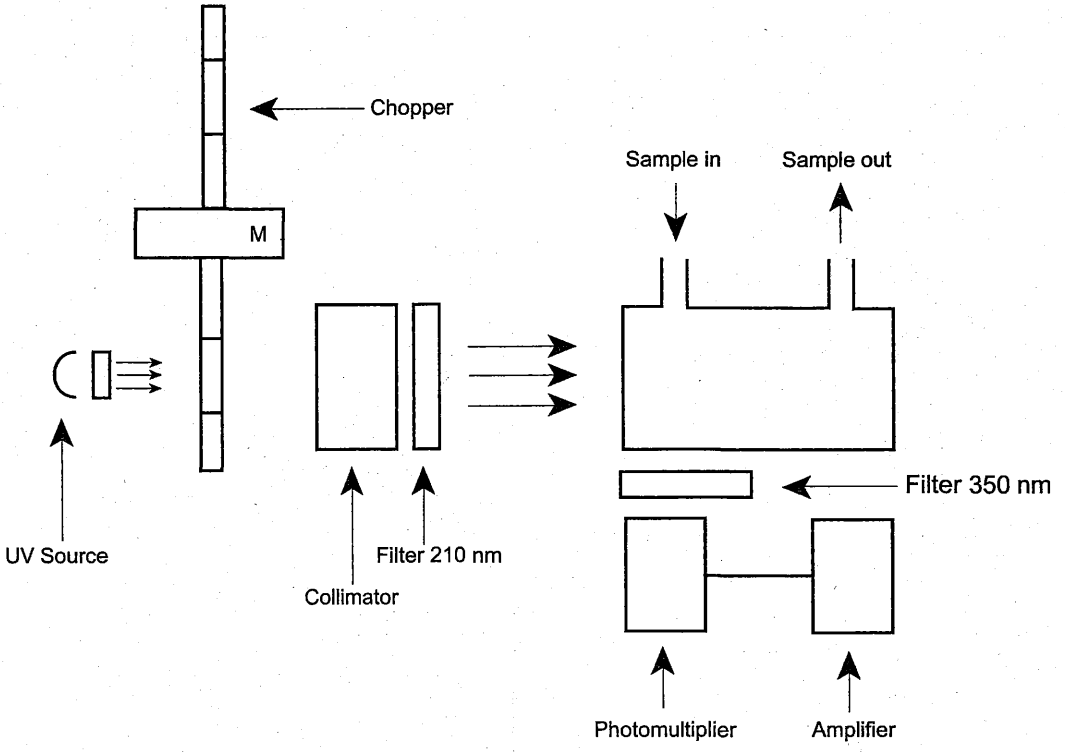


Figure 1238.-1. – *UV fluorescence analyser*

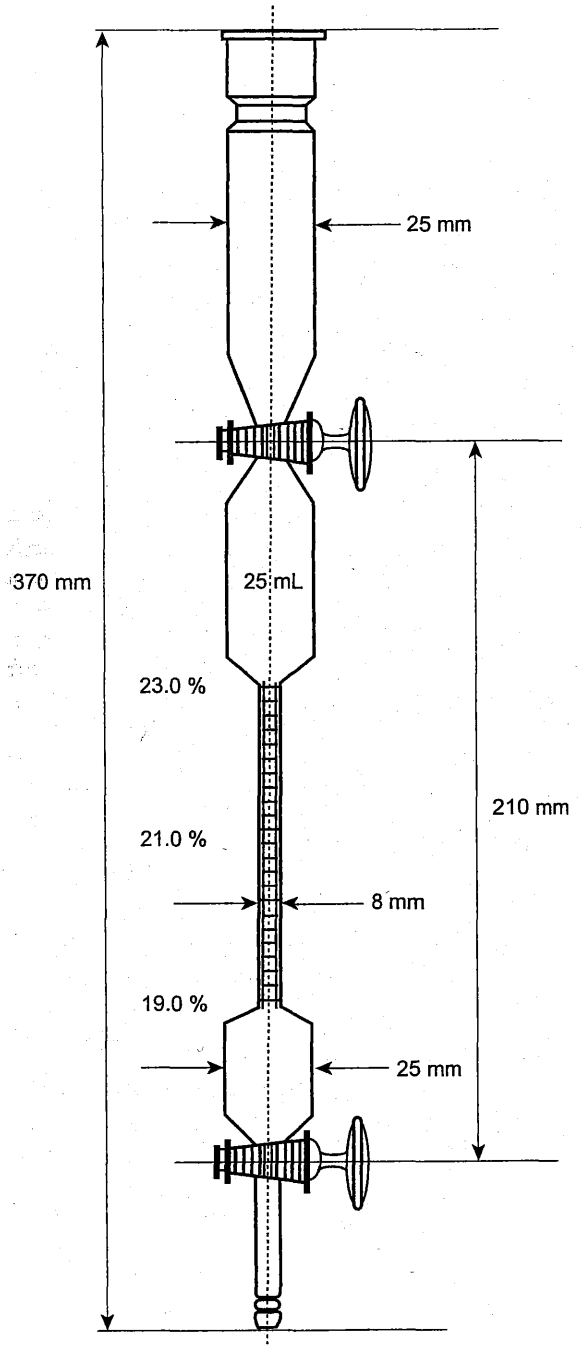


Figure 1238.-2. – *Gas burette*

Sulfur dioxide

Maximum 1 ppm *V/V*, determined using an ultraviolet fluorescence analyser (Figure 1238.-1).

The apparatus consists of the following:

— a system generating ultraviolet radiation with a wavelength of 210 nm, made up of an ultraviolet lamp, a collimator, and a selective filter; the beam is blocked periodically by a chopper rotating at high speeds;

— a reaction chamber, through which flows the gas to be examined;

— a system that detects radiation emitted at a wavelength of 350 nm, made up of a selective filter, a photomultiplier tube and an amplifier.

Gas to be examined Filter the substance to be examined.

Reference gas (a) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*.

Reference gas (b) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing 0.5 ppm *V/V* to 2 ppm *V/V* of *sulfur dioxide R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of sulfur dioxide in the gas to be examined.

Oil

Maximum 0.1 mg/m^3, determined using an oil detector tube (*2.1.6*), when an oil-lubricated compressor is used for the production.

Nitrogen monoxide and nitrogen dioxide

Maximum 2 ppm *V/V* in total, determined using a chemiluminescence analyser (*2.5.26*).

Gas to be examined The substance to be examined.

Reference gas (a) Use a mixture of 21 per cent *V/V* of *oxygen R* and 79 per cent *V/V* of *nitrogen R1*, containing less than 0.05 ppm *V/V* of nitrogen monoxide and nitrogen dioxide.

Reference gas (b) Use a mixture of 2 ppm *V/V* of *nitrogen monoxide R* in *nitrogen R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of nitrogen monoxide and nitrogen dioxide in the gas to be examined.

Water

Maximum 67 ppm *V/V*, determined using an electrolytic hygrometer (*2.5.28*), except where the competent authority decides that the following limit applies to medicinal air generated on-site and distributed in pipe-line systems operating at a pressure not greater than 10 bar and a temperature not less than 5 °C: maximum 870 ppm *V/V*, determined using an electrolytic hygrometer (*2.5.28*).

Assay

Determine the concentration of oxygen in air using a paramagnetic analyser (*2.5.27*).

IDENTIFICATION

First identification: C.

Second identification: A, B.

A. In a conical flask containing the substance to be examined, place a glowing wood splinter. The splinter remains glowing.

B. Use a gas burette (Figure 1238.-2) of 25 mL capacity in the form of a chamber in the middle of which is a tube graduated in 0.2 per cent between 19.0 per cent and 23.0 per cent, and isolated at each end by a tap with a conical barrel. The lower tap is joined to a tube with an olive-shaped nozzle and is used to introduce the gas into the apparatus. A cylindrical funnel above the upper tap is used to introduce the absorbent solution. Wash the burette with *water R* and dry. Open the 2 taps. Connect the nozzle to the source of the gas to be examined and set the flow rate to 1 L/min. Flush the burette by passing the gas to be examined through it for 1 min. Close the lower tap of the burette and immediately afterwards the upper tap. Rapidly disconnect the burette from the source of the gas to be examined. Rapidly give a half turn to the upper tap to eliminate any excess pressure in the burette. Keeping the burette vertical, fill the funnel with a freshly prepared mixture of 21 mL of a 560 g/L solution of *potassium hydroxide R* and 130 mL of a 200 g/L solution of *sodium dithionite R*. Open the upper tap slowly. The solution absorbs the oxygen and enters the burette. Allow to stand for 10 min without shaking. Read the level of the liquid meniscus on the graduated part of the burette.

This figure represents the percentage *V/V* of oxygen.
The value read is 20.4 to 21.4.

C. It complies with the limits of the assay.

TESTS

Carbon dioxide
Maximum 500 ppm *V/V*, determined using a carbon dioxide detector tube (*2.1.6*).

Sulfur dioxide
Maximum 1 ppm *V/V*, determined using a sulfur dioxide detector tube (*2.1.6*).

Oil
Maximum 0.1 mg/m^3, determined using an oil detector tube (*2.1.6*), when an oil-lubricated compressor is used for the production.

Nitrogen monoxide and nitrogen dioxide
Maximum 2 ppm *V/V*, determined using a nitrogen monoxide and nitrogen dioxide detector tube (*2.1.6*).

Carbon monoxide
Maximum 5 ppm *V/V*, determined using a carbon monoxide detector tube (*2.1.6*).

Water vapour
Maximum 67 ppm *V/V*, determined using a water vapour detector tube (*2.1.6*), except where the competent authority decides that the following limit applies to medicinal air generated on-site and distributed in pipe-line systems operating at a pressure not greater than 10 bar and a temperature not less than 5 °C: maximum 870 ppm *V/V*, determined using a water vapour detector tube (*2.1.6*).

STORAGE
As a gas, in suitable containers complying with the legal regulations or as a gas supplied by a pipe network.

LABELLING
Where applicable, the label states the production method, as regards to the use of an oil - lubricated compression.

IMPURITIES
A. CO_2: carbon dioxide,
B. SO_2: sulfur dioxide,
C. NO: nitrogen monoxide,
D. NO_2: nitrogen dioxide,
E. oil,
F. CO: carbon monoxide,
G. H_2O: water.

Ph Eur

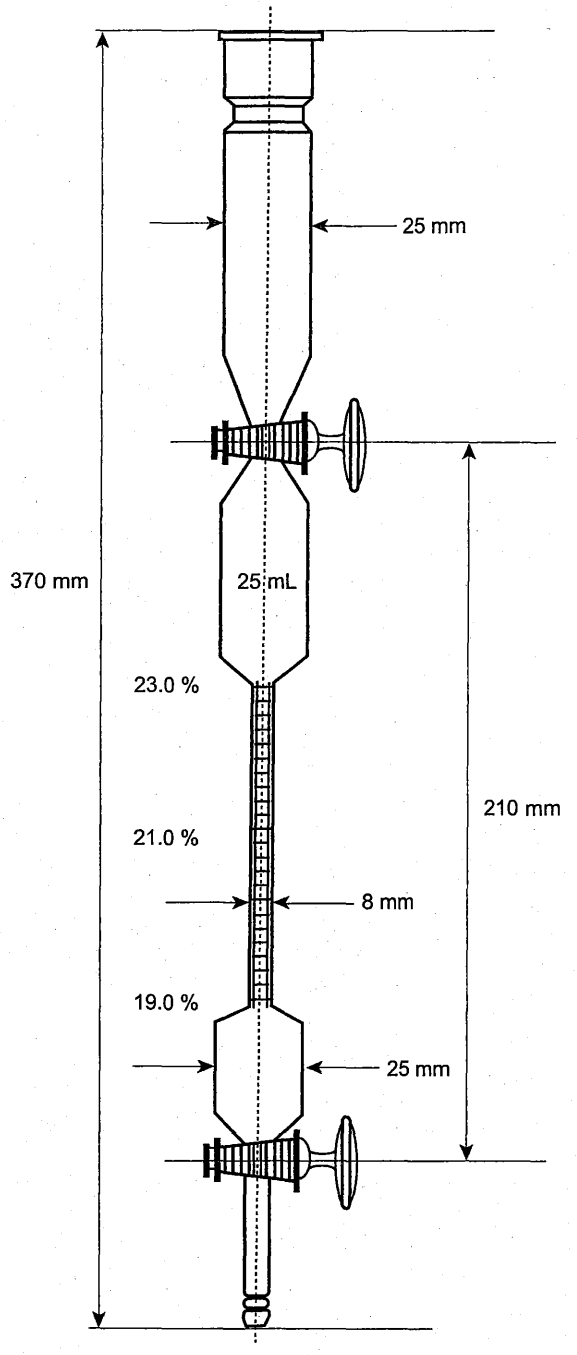


Figure 1684.-1.– *Gas burette*

Synthetic Air

(Synthetic Medicinal Air, Ph. Eur. monograph 1684)

When Synthetic Air is intended for use in a room in which magnetic resonance imaging (MRI) is being performed, the cylinder and fittings should be made from suitable non-ferromagnetic materials and labelled accordingly.

Ph Eur

DEFINITION
Mixture of *Nitrogen (1247)* and *Oxygen (0417)*.

Content
95.0 per cent to 105.0 per cent of the nominal value which is between 21.0 per cent *V/V* to 22.5 per cent *V/V* of oxygen (O_2).

CHARACTERS
Colourless and odourless gas.

Solubility
At a temperature of 20 °C and a pressure of 101 kPa, 1 volume dissolves in about 50 volumes of water.

PRODUCTION

Water (*2.5.28*)
Maximum 67 ppm *V/V*.

Assay (*2.5.27*)
Carry out the determination of oxygen in gases.

IDENTIFICATION
First identification: C.
Second identification: A, B.

A. In a conical flask containing the substance to be examined, place a glowing splinter of wood. The splinter remains glowing.

B. Use a gas burette (Figure 1684.-1) of 25 mL capacity in the form of a chamber, in the middle of which is a tube graduated in 0.2 per cent between 19.0 per cent and 23.0 per cent, and isolated at each end by a tap with a conical barrel. The lower tap is joined to a tube with an olive-shaped nozzle and is used to introduce the gas into the apparatus. A cylindrical funnel above the upper tap is used to introduce the absorbent solution. Wash the burette with *water R* and dry. Open both taps. Connect the nozzle to the source of the substance to be examined and set the flow rate to 1 L/min. Flush the burette by passing the substance to be examined through it for 1 min. Close the lower tap of the burette and immediately afterwards the upper tap. Rapidly disconnect the burette from the source of the substance to be examined. Rapidly give a half turn of the upper tap to eliminate any excess pressure in the burette. Keeping the burette vertical, fill the funnel with a freshly prepared mixture of 21 mL of a 560 g/L solution of *potassium hydroxide R* and 130 mL of a 200 g/L solution of *sodium dithionite R*. Open the upper tap slowly. The solution absorbs the oxygen and enters the burette. Allow to stand for 10 min without shaking. Read the level of the liquid meniscus on the graduated part of the burette. This figure represents the percentage *V/V* of oxygen. The value read is 95.0 per cent to 105.0 per cent of the nominal value.

C. It complies with the limits of the assay.

TESTS

Water vapour

Maximum 67 ppm *V/V*, determined using a water vapour detector tube (*2.1.6*).

STORAGE

As a compressed gas in suitable containers complying with the legal regulations or as a compressed gas supplied by a pipe network, after mixing of the components.

LABELLING

The label states the nominal content of O_2 in per cent *V/V*.

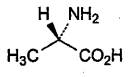
IMPURITIES

A. H_2O: water.

Ph Eur

Alanine

(Ph. Eur. monograph 0752)



$C_3H_7NO_2$ 89.1 *56-41-7*

Action and use

Amino acid.

Ph Eur

DEFINITION

(2*S*)-2-Aminopropanoic acid.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alanine CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of *alanine CRS* in *water R* and dilute to 50 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 0.5 g in a mixture of 0.25 mL of *hydrochloric acid R1*, 0.5 mL of a 100 g/L solution of *sodium nitrite R* and 1 mL of *water R*. Shake; gas is given off. Add 2 mL of *dilute sodium hydroxide solution R*, followed by 0.25 mL of *iodinated potassium iodide solution R*. After about 30 min, a yellow precipitate is formed.

TESTS

Solution S

Dissolve 2.5 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dilute 10 mL of solution S to 20 mL with *water R*.

Specific optical rotation (*2.2.7*)

+ 13.5 to + 15.5 (dried substance).

Dissolve 2.50 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances

Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *dilute hydrochloric acid R1* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of alanine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *any ninhydrin-positive substance*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

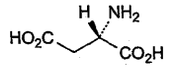
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 80.0 mg in 3 mL of *anhydrous formic acid R*. Add 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

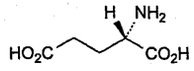
1 mL of *0.1 M perchloric acid* is equivalent to 8.91 mg of $C_3H_7NO_2$.

STORAGE
Protected from light.

IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*



A. (2*S*)-2-aminobutanedioic acid (aspartic acid),

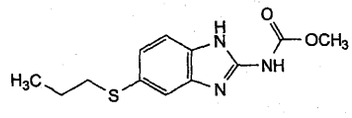


B. (2*S*)-2-aminopentanedioic acid (glutamic acid).

Ph Eur

Albendazole

(Ph. Eur. monograph 1386)



$C_{12}H_{15}N_3O_2S$ 265.3 *54965-21-8*

Action and use
Benzimidazole antihelminthic.

Preparations
Albendazole Oral Suspension
Albendazole Oral Suspension with Minerals

Ph Eur

DEFINITION
Methyl *N*-[5-(propylsulfanyl)-1*H*-benzimidazol-2-yl] carbamate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellowish powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous formic acid, very slightly soluble in methylene chloride, practically insoluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *albendazole CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.10 g in a mixture of 10 volumes of *anhydrous formic acid R* and 90 volumes of *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *sulfuric acid R, methanol R* (1:99 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in 5 mL of the solvent mixture and immediately dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *albendazole for system suitability CRS* (containing impurities B, C, E, F and H) in 1 mL of the solvent mixture and dilute to 10 mL with the mobile phase.

Reference solution (c) Dilute 1 mL of the solvent mixture to 10 mL with the mobile phase. Use 1 mL of this solution to dissolve the contents of a vial of *albendazole impurity mixture CRS* (containing impurities A and D).

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 1.67 g/L solution of *ammonium dihydrogen phosphate R, methanol R* (30:70 *V/V*).

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time Twice the retention time of albendazole.

Identification of impurities Use the chromatogram supplied with *albendazole impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and D; use the chromatogram supplied with *albendazole for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B + C, E, F and H.

Relative retention With reference to albendazole (retention time = about 11 min): impurity D = about 0.35; impurities B and C = about 0.40; impurity E = about 0.45; impurity A = about 0.48; impurity F = about 0.57; impurity H = about 0.66.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurities B + C and E.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.7; impurities B and C = 1.4; impurity D = 1.9; impurity E = 1.4;
— for each impurity, use the concentration of albendazole in reference solution (a).

Limits:
— *impurity H*: maximum 0.6 per cent;
— *impurity F*: maximum 0.5 per cent;
— *impurity A*: maximum 0.4 per cent;
— *sum of impurities B and C*: maximum 0.4 per cent;
— *impurity E*: maximum 0.3 per cent;
— *impurity D*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.3 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.250 g in 3 mL of *anhydrous formic acid R* and add 40 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 26.53 mg of $C_{12}H_{15}N_3O_2S$.

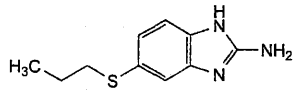
STORAGE

Protected from light.

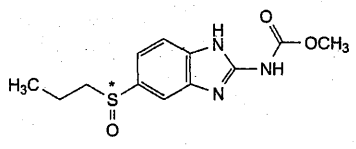
IMPURITIES

Specified impurities *A, B, C, D, E, F, H.*

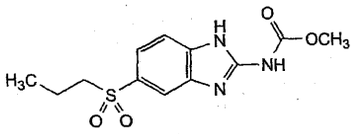
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, I, J, K, L.*



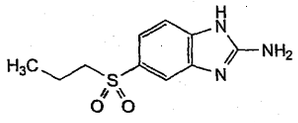
A. 5-(propylsulfanyl)-1*H*-benzimidazol-2-amine,



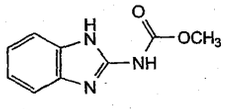
B. methyl *N*-[5-(propylsulfinyl)-1*H*-benzimidazol-2-yl]carbamate,



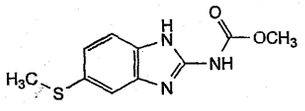
C. methyl *N*-[5-(propylsulfonyl)-1*H*-benzimidazol-2-yl]carbamate,



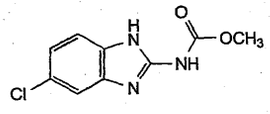
D. (2-amino-1*H*-benzimidazol-5-yl)propyl-λ^6-sulfanedione,



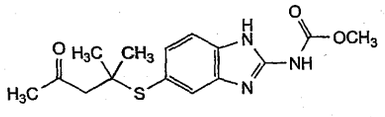
E. methyl *N*-(1*H*-benzimidazol-2-yl)carbamate,



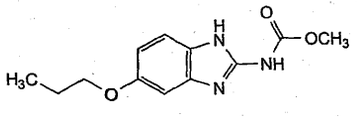
F. methyl *N*-[5-(methylsulfanyl)-1*H*-benzimidazol-2-yl]carbamate,



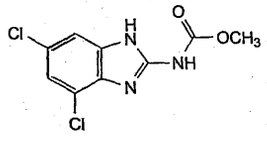
G. methyl *N*-(5-chloro-1*H*-benzimidazol-2-yl)carbamate,



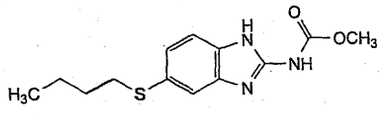
H. methyl *N*-[5-[(2-methyl-4-oxopentan-2-yl)sulfanyl]-1*H*-benzimidazol-2-yl]carbamate,



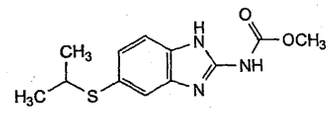
I. methyl *N*-(5-propoxy-1*H*-benzimidazol-2-yl)carbamate,



J. methyl *N*-(4,6-dichloro-1*H*-benzimidazol-2-yl)carbamate,



K. methyl *N*-[5-(butylsulfanyl)-1*H*-benzimidazol-2-yl]carbamate,

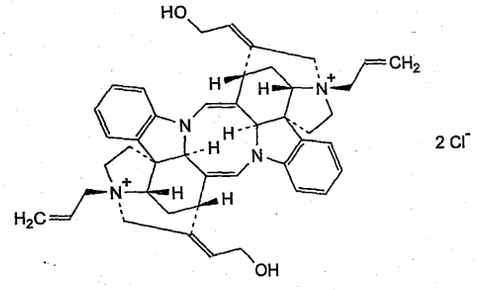


L. methyl *N*-[5-[(propan-2-yl)sulfanyl]-1*H*-benzimidazol-2-yl]carbamate.

Ph Eur

Alcuronium Chloride

(Ph. Eur. monograph 1285)



$C_{44}H_{50}Cl_2N_4O_2$ 738 *15180-03-7*

Action and use
Non-depolarizing neuromuscular blocker.

Ph Eur

DEFINITION

(1*R*,3a*S*,10*S*,11a*S*,12*R*,14a*S*,19a*S*,20b*S*,21*S*,22a*S*,23*E*,26*E*)-23,26-bis(2-Hydroxyethylidene)-1,12-bis(prop-2-enyl)-2,3,11,11a,13,14,22,22a-octahydro-10*H*,21*H*-1,21:10,12-diethano-19a*H*,20b*H*-[1,5]diazocino[1,2,3-*lm*:5,6,7-*l'm'*]dipyrrolo[2',3'-*d*:2'',3''-*d'*]dicarbazolediium dichloride (4,4'-didesmethyl-4,4'-bis(prop-2-enyl)toxiferin I dichloride).

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly greyish-white, crystalline powder.

Solubility
Freely soluble in water and in methanol, soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

Carry out the identification, tests and assay as rapidly as possible avoiding exposure to actinic light.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alcuronium chloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *alcuronium chloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase Mix 15 volumes of a 58.4 g/L solution of *sodium chloride R*, 35 volumes of *dilute ammonia R2* and 50 volumes of *methanol R.*

Application 10 µL.

Development Over a path of 15 cm.

Drying In air for 10 min.

Detection Spray with *0.1 M ammonium and cerium nitrate.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 0.250 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6, BY_6 or B_6 (*2.2.2, Method I*).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M hydrochloric acid.* The solution is red. Add 0.4 mL of *0.01 M sodium hydroxide.* The solution is yellow.

Specific optical rotation (*2.2.7*)

−430 to −451 (anhydrous substance), determined on solution S.

Propan-2-ol (*2.4.24, System A*)

Maximum 1.0 per cent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mix 100 mL of *methanol R*, 200 mL of *acetonitrile R* and 200 mL of a 6.82 g/L solution of *potassium dihydrogen phosphate R.* Dissolve 1.09 g of *sodium laurylsulfonate for chromatography R* in the mixture and adjust the apparent pH to 8.0 with a 100 g/L solution of *sodium hydroxide R.*

Test solution Dissolve 0.20 g of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 4.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (d) To 5.0 mL of the test solution add 5.0 mg of *allylstrychnine bromide CRS,* dissolve in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Column:

— *size: l* = 0.25 m, Ø = 4 mm;

— *stationary phase: octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 200 mL of *methanol R*, 400 mL of *acetonitrile R* and 400 mL of a 6.82 g/L solution of *potassium dihydrogen phosphate R.* Dissolve 2.18 g of *sodium laurylsulfonate for chromatography R* in the mixture and adjust the apparent pH to 5.4 with a 100 g/L solution of *phosphoric acid R.*

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time Twice the retention time of alcuronium.

System suitability Reference solution (d):

— *resolution*: minimum 4.0 between the peaks due to *N*-allylstrychnine and alcuronium.

Limits:

— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent) and not more than one of the peaks has an area greater than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);

— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g by stirring in 70 mL of *acetic anhydride R* for 1 min. Titrate with *0.1 M perchloric acid* until the colour changes from violet-blue to greenish-blue, using 0.1 mL of *crystal violet solution R* as indicator.

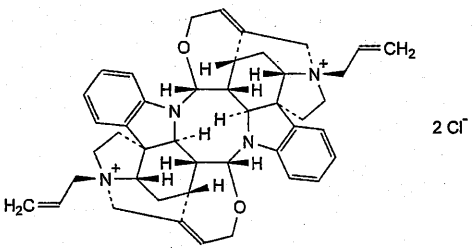
1 mL of *0.1 M perchloric acid* is equivalent to 36.9 mg of $C_{44}H_{50}Cl_2N_4O_2$.

STORAGE

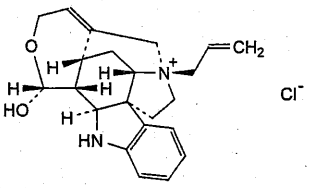
In an airtight container under nitrogen, protected from light, at a temperature of 2 °C to 8 °C.

IMPURITIES

Specified impurities *A, B.*



A. (1*R*,3a*S*,9*R*,9a*R*,10*R*,11a*S*,12*R*,14a*S*,19a*S*,20*R*,20a*R*,20b*S*,21*R*,22a*S*)-1,12-bis(prop-2-enyl)-2,3,9a,11,11a,13,14,19a,20a,21,22,22a-dodecahydro-10*H*,20b*H*-1,23:12,27-dimethano-9,10:20,21-bis(epoxyprop[2]eno)-9*H*,20*H*-[1,5]diazocino[1,2,3-*lm*:5,6,7-*l'm'*]dipyrrolo[2',3'-*d*:2'',3'':*d'*]dicarbazolediium dichloride (4,4'-diallylcaracurin V dichloride),

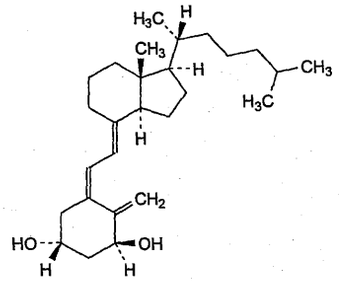


B. (4b*S*,7*R*,7a*S*,8a*R*,13*R*,13a*R*,13b*S*)-13-hydroxy-7-(prop-2-enyl)-5,6,7a,8,8a,11,13,13a,13b,14-decahydro-7,9-methano-7*H*-oxepino[3,4-*a*]pyrrolo[2,3-*d*]carbazolium chloride ((4*R*,17*R*)-4-allyl-17,18-epoxy-17-hydroxy-19,20-didehydrocuranium chloride).

Ph Eur

Alfacalcidol

(Ph. Eur. monograph 1286)



$C_{27}H_{44}O_2$ 400.6 *41294-56-8*

Action and use
Vitamin D analogue.

Ph Eur

DEFINITION

(5*Z*,7*E*)-9,10-Secocholesta-5,7,10(19)-triene-1α,3β-diol.

Content
97.0 per cent to 102.0 per cent.

A reversible isomerisation to pre-alfacalcidol takes place in solution, depending on temperature and time. The activity is due to both compounds (see Assay).

CHARACTERS

Appearance
White or almost white crystals.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in fatty oils.

It is sensitive to air, heat and light.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of alfacalcidol.*

B. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Carry out the test as rapidly as possible, avoiding exposure to light and air.*

Test solution Dissolve 1.0 mg of the substance to be examined without heating in 10.0 mL of the mobile phase.

Reference solution (a) Dissolve 1.0 mg of *alfacalcidol CRS* without heating in 10.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) In order to prepare pre-alfacalcidol *in situ*, dissolve the contents of a vial of *alfacalcidol for system suitability CRS* (containing impurities A and B) in 25 mL of the mobile phase, heat in a water-bath at 80 °C under a reflux condenser for 2 h and cool.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *ammonia R, water R, acetonitrile R* (1:200:800 *V/V/V*).

Flow rate 2.6 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 100 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of alfacalcidol.

Identification of impurities Use the chromatogram supplied with *alfacalcidol for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to alfacalcidol (retention time = about 21 min): pre-alfacalcidol = about 0.88; impurity A = about 0.93; impurity B = about 1.1.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to pre-alfacalcidol and impurity A and minimum 1.5 between the peaks due to impurity A and alfacalcidol.

Limits:
— *impurities A, B*: for each impurity, maximum 0.5 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to pre-alfacalcidol.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solutions (a) and (c).

System suitability Reference solution (c):
— *repeatability*: maximum relative standard deviation of 1 per cent for the peak due to alfacalcidol after 6 injections.

Calculate the percentage content of $C_{27}H_{44}O_2$ taking into account the assigned content of *alfacalcidol CRS* and, if necessary, the peak due to pre-alfacalcidol.

STORAGE

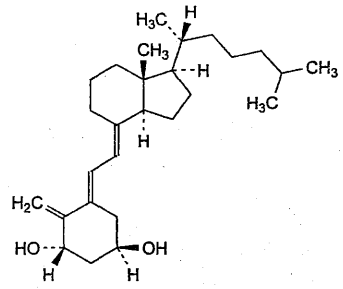
Under nitrogen, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

The contents of an opened container are to be used immediately.

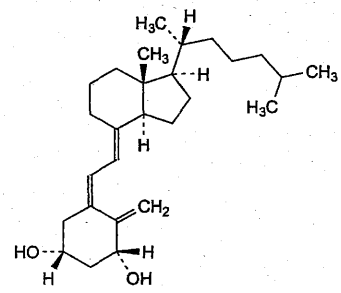
IMPURITIES

Specified impurities *A, B.*

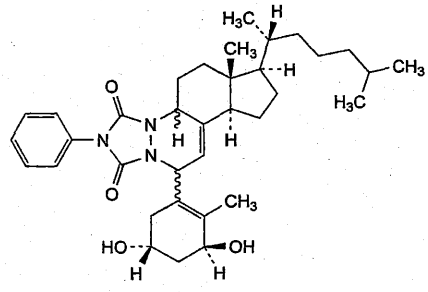
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



A. (5*E*,7*E*)-9,10-secocholesta-5,7,10(19)-triene-1α,3β-diol (*trans*-alfacalcidol),



B. (5*Z*,7*E*)-9,10-secocholesta-5,7,10(19)-triene-1β,3β-diol (1β-calcidol),



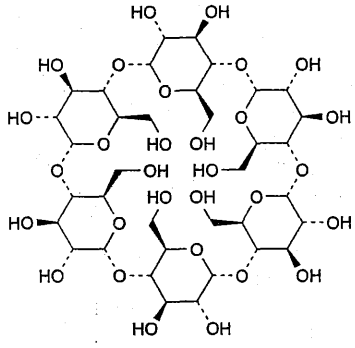
C. 6ξ-[(3*S*,5*R*)-3,5-dihydroxy-2-methylcyclohex-1-en-1-yl]-2-phenyl-2,5,10-triaza-4,19-dinor-9ξ-cholest-7-ene-1,3-dione.

Ph Eur

Alfadex

Alphacyclodextrin

(Ph. Eur. monograph 1487)



$[C_6H_{10}O_5]_6$ 973 *10016-20-3*

Action and use
Cyclodextran; carrier molecule for drug delivery systems.

Ph Eur

DEFINITION
Cyclohexakis-(1→4)-(α-D-glucopyranosyl) (cyclomaltohexaose or α-cyclodextrin).

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, amorphous or crystalline powder.

Solubility
Freely soluble in water, slightly soluble in propylene glycol, practically insoluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

C. Dissolve 0.2 g in 2 mL of *iodine solution R4* by warming on a water-bath, and allow to stand at room temperature; a yellowish-brown precipitate is formed.

TESTS

Solution S
Dissolve 1.000 g in *carbon dioxide-free water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

pH (*2.2.3*)
5.0 to 8.0.

Mix 1 mL of a 223.6 g/L solution of *potassium chloride R* and 30 mL of solution S.

Specific optical rotation (*2.2.7*)
+ 147 to + 152 (dried substance), determined on solution S.

Reducing sugars
Maximum 0.2 per cent.

Test solution To 1 mL of solution S add 1 mL of *cupri-tartaric solution R4*. Heat on a water-bath for 10 min, cool to room temperature. Add 10 mL of *ammonium molybdate reagent R1* and allow to stand for 15 min.

Reference solution Prepare a reference solution at the same time and in the same manner as the test solution, using 1 mL of a 0.02 g/L solution of *glucose R*.

Measure the absorbance (*2.2.25*) of the test solution and the reference solution at the absorption maximum at 740 nm using *water R* as the compensation liquid. The absorbance of the test solution is not greater than that of the reference solution.

Light-absorbing impurities
Examine solution S between 230 nm and 750 nm. Between 230 nm and 350 nm, the absorbance (*2.2.25*) is not greater than 0.10. Between 350 nm and 750 nm, the absorbance (*2.2.25*) is not greater than 0.05.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.25 g of the substance to be examined in *water R* with heating, cool and dilute to 25.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with *water R*.

Reference solution (a) Dissolve 25.0 mg of *betadex CRS* (impurity A), 25.0 mg of *gammacyclodextrin CRS* (impurity B) and 50.0 mg of *alfadex CRS* in *water R*, then dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with *water R*.

Reference solution (c) Dissolve 25.0 mg of *alfadex CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase *methanol R, water R* (10:90 *V/V*).

Flow rate 1.5 mL/min.

Detection Differential refractometer.

Equilibration With the mobile phase for about 3 h.

Injection 50 µL of test solution (a) and reference solutions (a) and (b).

Run time 3.5 times the retention time of alfadex.

Relative retention With reference to alfadex (retention time = about 10 min): impurity B = about 0.7; impurity A = about 2.2.

System suitability Reference solution (a):
- *resolution*: minimum 1.5 between the peaks due to impurity B and alfadex; if necessary, adjust the concentration of methanol in the mobile phase.

Limits:
- *impurities A, B*: for each impurity, not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
- *sum of impurities other than A and B*: not more than 0.5 times the area of the peak due to alfadex in the chromatogram obtained with reference solution (b) (0.5 per cent).

Loss on drying (*2.2.32*)
Maximum 11 per cent, determined on 1.000 g by drying in an oven at 120 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

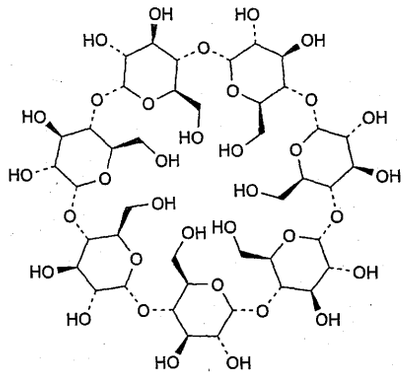
Injection Test solution (b) and reference solutions (a) and (c).

System suitability:
- *repeatability*: maximum relative standard deviation of 2.0 per cent for the peak due to alfadex after 5 injections of reference solution (a).

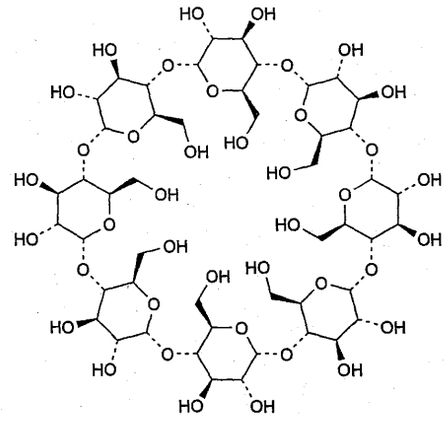
Calculate the percentage content of $[C_6H_{10}O_5]_6$ from the assigned content of *alfadex CRS*.

STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B.*



A. cycloheptakis-(1→4)-(α-D-glucopyranosyl) (betadex or cyclomaltoheptaose or β-cyclodextrin),

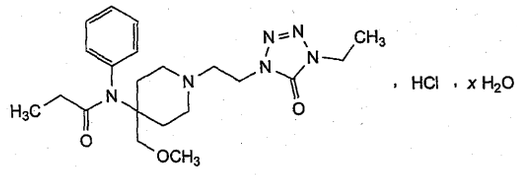


B. cyclooctakis-(1→4)-(α-D-glucopyranosyl) (cyclomaltooctaose or γ-cyclodextrin).

Ph Eur

Alfentanil Hydrochloride Hydrate

Alfentanil Hydrochloride
(Ph. Eur. monograph 1062)



$C_{21}H_{33}ClN_6O_3, xH_2O$ 453.0 (anhydrous substance)

Anhydrous alfentanil hydrochloride *69049-06-5*

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION
N-[1-[2-(4-Ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl)ethyl]-4-(methoxymethyl)piperidin-4-yl]-*N*-phenylpropanamide hydrochloride hydrate.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).
It contains a variable quantity of water.

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, in ethanol (96 per cent) and in methanol.

mp
About 140 °C, with decomposition.
It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *alfentanil hydrochloride hydrate CRS.*
If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Dissolve 50 mg in a mixture of 0.4 mL of *ammonia R* and 2 mL of *water R*. Mix, allow to stand for 5 min and filter. Acidify the filtrate with *dilute nitric acid R*. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).
Dissolve 0.2 g in *water R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*
Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.
Reference solution (a) In order to prepare impurity E *in situ*, dissolve 10 mg of the substance to be examined in 10.0 mL of *dilute hydrochloric acid R*. Heat on a water-bath under a reflux condenser for 4 h. Neutralise with 10.0 mL of *dilute sodium hydroxide solution R* and evaporate to dryness on a water-bath. Cool and take up the residue in 10 mL of *methanol R*. Filter.
Reference solution (b) Dissolve the contents of a vial of *alfentanil impurity D CRS* in 1 mL of *methanol R*.
Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.
Blank solution *methanol R.*
Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm).
Mobile phase:
— *mobile phase A*: 5 g/L solution of *ammonium carbonate R* in a mixture of 10 volumes of *tetrahydrofuran R* and 90 volumes of *water for chromatography R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	90 → 40	10 → 60
15 - 20	40	60
20 - 25	40 → 90	60 → 10

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 220 nm.
Injection 10 µL.
Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.
Relative retention With reference to alfentanil (retention time = about 8 min): impurity D = about 0.8; impurity E = about 0.9.
System suitability Reference solution (a):
— *resolution*: minimum 4.0 between the peaks due to impurity E and alfentanil.
Calculation of percentage contents:
— for each impurity, use the concentration of alfentanil hydrochloride hydrate in reference solution (c).
Limits:
— *impurity D*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
3.0 per cent to 4.0 per cent, determined on 0.500 g.

ASSAY

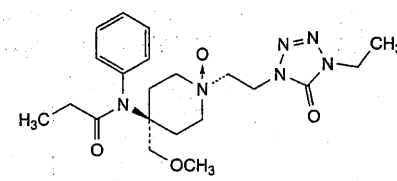
Dissolve 0.350 g in 50 mL of a mixture of 1 volume of *ethanol (96 per cent) R* and 4 volumes of *water R* and add 5.0 mL of *0.01 M hydrochloric acid*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.
1 mL of *0.1 M sodium hydroxide* is equivalent to 45.30 mg of $C_{21}H_{33}ClN_6O_3$.

STORAGE

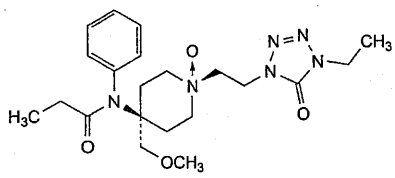
Protected from light.

IMPURITIES

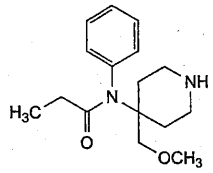
Specified impurities *D.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E, F, G, H.*



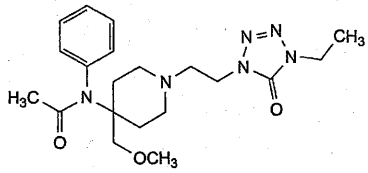
A. (1*s*,4*s*)-1-[2-(4-ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl) ethyl]-4-(methoxymethyl)-4-(*N*-phenylpropanamido) piperidine 1-oxide,



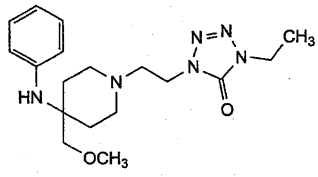
B. (1*r*,4*r*)-1-[2-(4-ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl) ethyl]-4-(methoxymethyl)-4-(*N*-phenylpropanamido) piperidine 1-oxide,



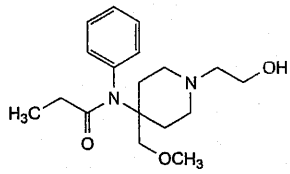
C. *N*-[4-(methoxymethyl)piperidin-4-yl]-*N*-phenylpropanamide,



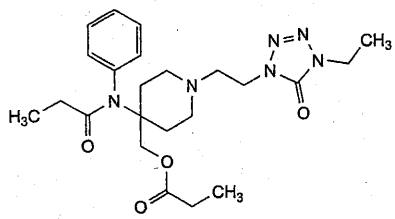
D. *N*-[1-[2-(4-ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl) ethyl]-4-(methoxymethyl)piperidin-4-yl]-*N*-phenylacetamide,



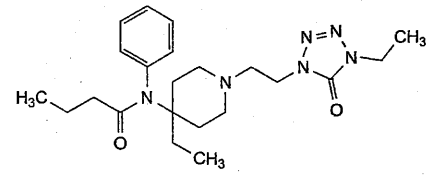
E. 1-ethyl-4-[2-[4-(methoxymethyl)-4-(phenylamino) piperidin-1-yl]ethyl]-1,4-dihydro-5*H*-tetrazol-5-one,



F. *N*-[1-(2-hydroxyethyl)-4-(methoxymethyl)piperidin-4-yl]-*N*-phenylpropanamide,



G. *N*-[1-[2-(4-ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl) ethyl]-4-[(propanoyloxy)methyl]piperidin-4-yl]-*N*-phenylpropanamide,

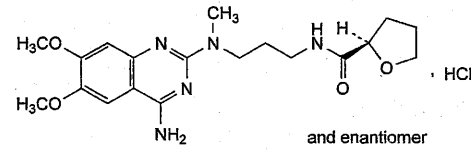


H. *N*-[1-[2-(4-ethyl-5-oxo-4,5-dihydro-1*H*-tetrazol-1-yl) ethyl]-4-(methoxymethyl)piperidin-4-yl]-*N*-phenylbutanamide.

Ph Eur

Alfuzosin Hydrochloride

(Ph. Eur. monograph 1287)



$C_{19}H_{28}ClN_5O_4$ 425.9 *81403-68-1*

Action and use
Alpha$_1$-adrenoceptor antagonist.

Preparations
Alfuzosin Tablets
Alfuzosin Prolonged-release Tablets

Ph Eur

DEFINITION
(2*RS*)-*N*-[3-[(4-Amino-6,7-dimethoxyquinazolin-2-yl) methylamino]propyl]oxolan-2-carboxamide hydrochloride.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder, slightly hygroscopic.

Solubility
Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *alfuzosin hydrochloride CRS.*
B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
pH (*2.2.3*)
4.0 to 5.5.
Dissolve 0.500 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent. Use a freshly prepared solution.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 40 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 4 mg of *alfuzosin for system suitability A CRS* (containing impurities B, F and G) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 4 mg of *alfuzosin for peak identification CRS* (containing impurity D) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 25 °C; if necessary, increase the temperature slightly to achieve the required resolution between the peaks due to impurity G and alfuzosin.

Mobile phase Mix 1 volume of *tetrahydrofuran R*, 20 volumes of *acetonitrile R* and 80 volumes of a solution prepared as follows: dilute 5.0 mL of *perchloric acid R* in 900 mL of *water for chromatography R*, adjust to pH 3.5 with *dilute sodium hydroxide solution R* and dilute to 1000 mL with *water for chromatography R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time Twice the retention time of alfuzosin.

Identification of impurities Use the chromatogram supplied with *alfuzosin for system suitability A CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, F and G; use the chromatogram supplied with *alfuzosin for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to alfuzosin (retention time = about 9 min): impurity D = about 0.4; impurity B = about 0.57; impurity F = about 0.63; impurity G = about 0.9.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurities B and F; minimum 1.5 between the peaks due to impurity G and alfuzosin.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity F by 0.6;
- *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurity F*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

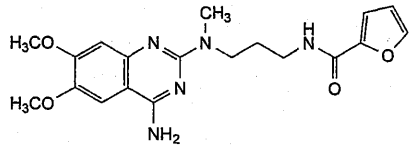
ASSAY
Dissolve 0.300 g in a mixture of 40 mL of *anhydrous acetic acid R* and 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 42.59 mg of $C_{19}H_{28}ClN_5O_4$.

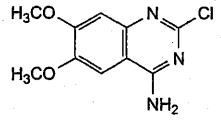
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *D, F.*

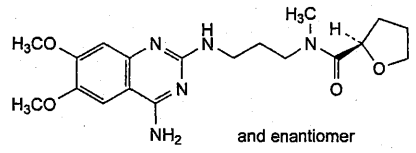
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E, G.*



A. *N*-[3-[(4-amino-6,7-dimethoxyquinazolin-2-yl) methylamino]propyl]furan-2-carboxamide,

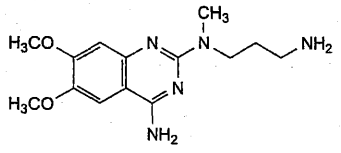


B. 2-chloro-6,7-dimethoxyquinazolin-4-amine,

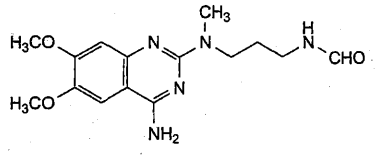


and enantiomer

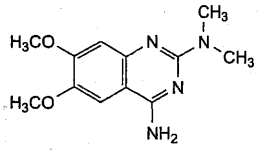
C. (2*RS*)-*N*-[3-[(4-amino-6,7-dimethoxyquinazolin-2-yl) amino]propyl]-*N*-methyloxolan-2-carboxamide,



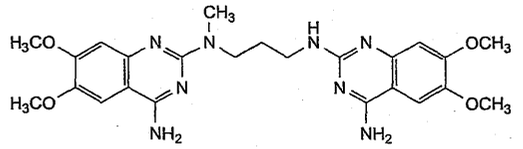
D. N^2-(3-aminopropyl)-6,7-dimethoxy-N^2-methylquinazolin-2,4-diamine,



E. *N*-[3-[(4-amino-6,7-dimethoxyquinazolin-2-yl) methylamino]propyl]formamide,



F. 6,7-dimethoxy-N^2,N^2-dimethylquinazoline-2,4-diamine,



G. N^2-[3-[(4-amino-6,7-dimethoxyquinazolin-2-yl)amino] propyl]-6,7-dimethoxy-N^2-methylquinazoline-2,4-diamine.

Ph Eur

Alginic Acid

(Ph. Eur. monograph 0591)

Action and use
Treatment of gastro-oesophageal reflux disease; excipient; thickening agent.

Ph Eur

DEFINITION
Mixture of polyuronic acids $[(C_6H_8O_6)_n]$ composed of residues of D-mannuronic and L-guluronic acids, obtained mainly from algae belonging to the Phaeophyceae. A small proportion of the carboxyl groups may be neutralised.

Content
19.0 per cent to 25.0 per cent of carboxyl groups ($-CO_2H$) (dried substance).

CHARACTERS
Appearance
White or pale yellowish-brown, crystalline or amorphous powder.

Solubility
Very slightly soluble or practically insoluble in ethanol (96 per cent), practically insoluble in organic solvents. It swells in water but does not dissolve; it dissolves in solutions of alkali hydroxides.

IDENTIFICATION
A. To 0.2 g add 20 mL of *water R* and 0.5 mL of *sodium carbonate solution R*. Shake and filter. To 5 mL of the filtrate add 1 mL of *calcium chloride solution R*. A voluminous gelatinous mass is formed.

B. To 5 mL of the filtrate obtained in identification test A add 0.5 mL of a 123 g/L solution of *magnesium sulfate R*. No voluminous gelatinous mass is formed.

C. To 5 mg add 5 mL of *water R*, 1 mL of a freshly prepared 10 g/L solution of *1,3-dihydroxynaphthalene R* in *ethanol (96 per cent) R* and 5 mL of *hydrochloric acid R*. Boil gently for 3 min, cool, add 5 mL of *water R*, and shake with 15 mL of *di-isopropyl ether R*. Carry out a blank test. The upper layer obtained with the substance to be examined exhibits a deeper bluish-red colour than that obtained with the blank.

TESTS
Chlorides
Maximum 1.0 per cent.

To 2.50 g add 50 mL of *dilute nitric acid R*, shake for 1 h and dilute to 100.0 mL with *dilute nitric acid R*. Filter. To 50.0 mL of the filtrate add 10.0 mL of *0.1 M silver nitrate* and 5 mL of *toluene R*. Titrate with *0.1 M ammonium thiocyanate*, using 2 mL of *ferric ammonium sulfate solution R2* as indicator and shaking vigorously towards the end-point.

1 mL of *0.1 M silver nitrate* is equivalent to 3.545 mg of Cl.

Loss on drying *(2.2.32)*
Maximum 15.0 per cent, determined on 0.1000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash *(2.4.14)*
Maximum 8.0 per cent (dried substance), determined on 0.100 g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g *(2.6.12)*.

Absence of *Escherichia coli* *(2.6.13)*.

Absence of *Salmonella* *(2.6.13)*.

ASSAY
To 0.2500 g add 25 mL of *water R*, 25.0 mL of *0.1 M sodium hydroxide* and 0.2 mL of *phenolphthalein solution R*. Titrate with *0.1 M hydrochloric acid*.

1 mL of *0.1 M sodium hydroxide* is equivalent to 4.502 mg of carboxyl groups ($-CO_2H$).

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for alginic acid used as disintegrant and/or binder.

Particle-size distribution *(2.9.31* or *2.9.38)*

Settling volume
Place 75 mL of *water R* in a 100 mL graduated cylinder and add 1.5 g of the substance to be examined in 0.5 g portions, shaking vigorously after each addition. Dilute to 100.0 mL with *water R* and shake again until the substance is homogeneously distributed. Allow to stand for 4 h and determine the volume of the settled mass.

The following characteristic may be relevant for alginic acid used as gelling agent or viscosity-increasing agent.

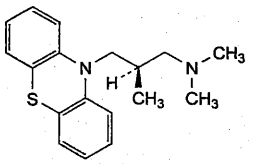
Apparent viscosity
Determine the dynamic viscosity using a rotating viscometer *(2.2.10)*.

Prepare a 20 g/L suspension of alginic acid (dried substance) and add *0.1 M sodium hydroxide* until a solution is obtained.

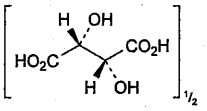
Ph Eur

Alimemazine Tartrate

(Alimemazine Hemitartrate Ph. Eur. monograph 2650)



and enantiomer ,



$C_{20}H_{25}N_2O_3S$ 373.5 *4330-99-8*

Action and use
Histamine H_1, receptor antagonist; sedative.

Preparations
Paediatric Alimemazine Oral Solution
Strong Paediatric Alimemazine Oral Solution
Alimemazine Tablets

Ph Eur

DEFINITION

(*2RS*)-*N*,*N*,2-Trimethyl-3-(10*H*-phenothiazin-10-yl)propan-1-amine hemi[(2*R*,3*R*)-2,3-dihydroxybutanedioate].

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or very slightly yellowish powder.

Solubility
Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in toluene.

It deteriorates when exposed to air and light.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alimemazine hemitartrate CRS.*

TESTS

Appearance of solution
The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 1.0 g in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)
5.0 to 6.5. *Carry out the test protected from light and use a freshly prepared solution.*

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and use freshly prepared solutions.*

Solvent mixture *acetonitrile R, water R* (20:80 *V/V*).

Test solution Dissolve 35 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 3.5 mg of *alimemazine for system suitability CRS* (containing impurities A, B and C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 40 °C.

Mobile phase *acetonitrile R, methanol R*, 3.854 g/L solution of *ammonium acetate R* (10:40:50 *V/V/V*).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 253 nm.

Injection 20 µL.

Run time Twice the retention time of alimemazine.

Identification of impurities Use the chromatogram supplied with *alimemazine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to alimemazine (retention time = about 27 min): impurity A = about 0.1; impurity B = about 0.5; impurity C = about 1.4.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to alimemazine and impurity C.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 4.4; impurity C = 0.4;
— for each impurity, use the concentration of alimemazine in reference solution (a).

Limits:
— *impurity B*: maximum 0.3 per cent;
— *impurities A, C*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

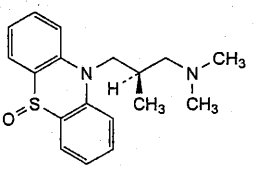
1 mL of *0.1 M perchloric acid* is equivalent to 37.35 mg of $C_{20}H_{25}N_2O_3S$.

STORAGE

In an airtight container, protected from light.

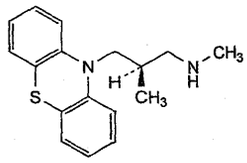
IMPURITIES

Specified impurities *A, B, C.*



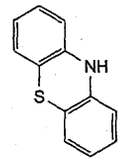
and enantiomer

A. (2*RS*)-*N*,*N*,2-trimethyl-3-(5-oxido-10*H*-phenothiazin-10-yl)propan-1-amine,



and enantiomer

B. (2*RS*)-*N*,2-dimethyl-3-(10*H*-phenothiazin-10-yl)propan-1-amine,

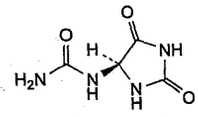


C. 10*H*-phenothiazine.

Ph Eur

Allantoin

(Ph. Eur. monograph 1288)



and enantiomer

$C_4H_6N_4O_3$ 158.1 *97-59-6*

Action and use
Astringent; keratolytic.

Ph Eur

DEFINITION
Allantoin contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of (*RS*)-(2,5-dioxoimidazolidin-4-yl)urea.

CHARACTERS
A white or almost white, crystalline powder, slightly soluble in water, very slightly soluble in alcohol.

It melts at about 225 °C, with decomposition.

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *allantoin CRS*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Boil 20 mg with a mixture of 1 mL of *dilute sodium hydroxide solution R* and 1 mL of *water R*. Allow to cool. Add 1 mL of *dilute hydrochloric acid R*. To 0.1 mL of the solution add 0.1 mL of a 100 g/L solution of *potassium bromide R*, 0.1 mL of a 20 g/L solution of *resorcinol R* and 3 mL of *sulfuric acid R*. Heat for 5 min to 10 min on a water-bath. A dark blue colour develops, which becomes red after cooling and pouring into about 10 mL of *water R*.

D. Heat about 0.5 g. Ammonia vapour is evolved, which turns *red litmus paper R* blue.

TESTS
Solution S
Dissolve 0.5 g in *carbon dioxide-free water R*, with heating if necessary, and dilute to 100 mL with the same solvent.

Acidity or alkalinity
To 5 mL of solution S add 5 mL of *carbon dioxide-free water R*, 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is red.

Optical rotation (*2.2.7*)
The angle of optical rotation, determined on solution S, is −0.10° to + 0.10°.

Reducing substances
Shake 1.0 g with 10 mL of *water R* for 2 min. Filter. Add 1.5 mL of *0.02 M potassium permanganate*. The solution must remain violet for at least 10 min.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using a suitable *cellulose for chromatography R* as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in 5.0 mL of *water R* with heating. Allow to cool. Dilute to 10 mL with *methanol R*. *Use the solution immediately after preparation.*

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of *methanol R* and 1 volume of *water R*.

Reference solution (a) Dissolve 10 mg of *allantoin CRS* in a mixture of 1 volume of *methanol R* and 1 volume of *water R* and dilute to 10 mL with the same mixture of solvents.

Reference solution (b) Dissolve 10 mg of *urea R* in 10 mL of *water R*. Dilute 1 mL of this solution to 10 mL with *methanol R*.

Reference solution (c) Mix 1 mL of reference solution (a) and 1 mL of reference solution (b).

Apply to the plate 10 µL of test solution (a) and 5 µL each of test solution (b), reference solution (a), reference solution (b) and reference solution (c). Develop over a path of 10 cm using a mixture of 15 volumes of *glacial acetic acid R*, 25 volumes of *water R* and 60 volumes of *butanol R*. Allow the plate to dry in air. Spray the plate with a 5 g/L solution of *dimethylaminobenzaldehyde R* in a mixture of 1 volume of *hydrochloric acid R* and 3 volumes of *methanol R*. Dry the plate in a current of hot air. Examine in daylight after 30 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated principal spots.

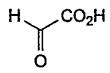
Loss on drying (*2.2.32*)
Not more than 0.1 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

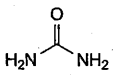
ASSAY
Dissolve 120.0 mg in 40 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 15.81 mg of $C_4H_6N_4O_3$.

IMPURITIES



A. glyoxylic acid,



B. carbamide (urea).

Ph Eur

Allergen Products

(Ph. Eur. monograph 1063)

Ph Eur

This monograph does not apply to: chemicals that are used solely for diagnosis of contact dermatitis; chemically synthesised products; allergens derived by recombinant DNA technology. It does not necessarily apply to allergen products for veterinary use.

DEFINITION

Allergen products are pharmaceutical preparations derived from extracts of naturally occurring source materials containing allergens, which are substances that lead to and/or provoke allergic reactions. The allergenic components are most often of a proteinaceous nature. Allergen products are intended for *in vivo* diagnosis or treatment of allergic diseases attributed to these allergens.

Allergen products are available as finished products, and as finished products used on a named-patient basis. Allergen products are generally presented as parenteral preparations, eye preparations, preparations for inhalation, preparations for oral use, sublingual preparations or preparations for skin tests.

For *in vivo* diagnostic use, allergen products are usually prepared as unmodified extracts in a 50 per cent *V/V* solution of glycerol for skin testing. For intradermal diagnosis or for provocation tests by nasal, ocular or bronchial administration, suitable dilutions of allergen products may be prepared by dilution of aqueous or glycerinated extracts, or by reconstitution of unmodified freeze-dried extracts.

For *specific immunotherapy*, allergen products may be either unmodified extracts or extracts modified chemically and/or by adsorption onto different carriers (for example, aluminium hydroxide, calcium phosphate or tyrosine).

PRODUCTION

Where allergen products or source materials are manufactured using materials of human or animal origin, the requirements of general chapter *5.1.7. Viral safety* apply.

SOURCE MATERIALS

Source materials are obtained from qualified suppliers. The source materials comply with the requirements of the appropriate individual monographs (where a relevant monograph exists) and the statements in this section are intended to be read in conjunction with the individual monographs.

Source materials for the preparation of allergen products are products of animal or vegetable origin, mostly pollens, moulds, mites, animal epithelia and outgrowths, and Hymenoptera venoms. Other source materials include certain insects and foods.

The source materials are defined, where possible, by their origin, nature, method of collection or production and pre-treatment. Control methods and acceptance criteria relating to identity and purity are established. The acceptance criteria must ensure the consistency of the allergenic source material from a qualitative and quantitative point of view. The source materials are stored under controlled conditions justified by stability data.

The collection or production, as well as the handling of the source materials, are such that consistent composition is ensured from batch to batch.

When applicable, pesticides, heavy metals and residual solvents are limited according to the principles defined in general chapters *2.8.13. Pesticide residues*, *2.4.27. Heavy metals in herbal drugs and herbal drug preparations* and *2.4.24. Identification and control of residual solvents*, respectively.

Microbial contamination of the source material may be unavoidable and should be monitored according to a justified sampling plan; if a determination of microbial contamination is not applicable, this must be justified.

The scientific name (species, variety, strain etc.) of the source material is indicated and the part used is stated, if applicable.

Foods must be of a quality suitable for human consumption. The origin of the food stuff as well as its processing stage is stated.

MANUFACTURING PROCESS

Allergen products are generally obtained by extraction, and may be purified, from the source materials using appropriate methods shown to preserve the allergenic properties of the components. Allergens for which there are not enough patients to determine the total allergenic activity *in vivo* or *in vitro*, the extraction ratio indicating the relative proportions (*m/V*) of allergenic source materials and solvents is a minimum requirement. Allergen products presented as parenteral preparations, eye preparations, preparations for inhalation and preparations for skin testing are manufactured under aseptic conditions.

In the manufacture, packaging, storage and distribution of allergen products intended for administration by other routes, suitable measures are taken to ensure their microbial quality; recommendations on this aspect are provided in general chapter *5.1.4. Microbiological quality of non-sterile pharmaceutical preparations and substances for pharmaceutical use*.

All allergen preparations are manufactured under conditions designed to minimise exogenous and endogenous enzymatic degradation.

Any purification procedure is designed to minimise the content of any potential irritant low-molecular-mass components and non-allergenic components.

Allergen products may contain suitable antimicrobial preservatives. The nature and concentration of the antimicrobial preservatives have to be justified and their efficacy complies with chapter *5.1.3. Efficacy of antimicrobial preservation*.

The manufacturing process comprises various stages:

— source material;
— active substance: it is generally a modified or an unmodified allergen extract; where applicable it is stored under conditions ensuring its stability, for example freeze-dried;
— finished product.

All other stages of the manufacturing process are considered as intermediates.

IN-HOUSE REFERENCE PREPARATION

An appropriate representative preparation is selected as the in-house reference preparation (IHRP), characterised and used to verify batch-to-batch consistency. The IHRP is stored in suitably sized aliquots under conditions ensuring its stability, for example freeze-dried.

Characterisation of the in-house reference preparation

The extent of characterisation of the IHRP depends on the source material, knowledge of the allergenic components and availability of suitable reagents, as well as the intended use. The characterised IHRP is used as the reference in the batch control of active substances and intermediates and, if possible, in the batch control of finished products.

The IHRP is characterised by the protein content determination and a protein profile using appropriate methods (such as isoelectric focusing, sodium dodecyl sulfate polyacrylamide gel electrophoresis, immunoelectrophoresis, capillary electrophoresis, chromatographic techniques and mass spectrometry).

Allergenic components may be detected by appropriate methods (for example, immunoblotting or crossed radio-immunoelectrophoresis). Characterisation of the allergenic components may include identification of relevant allergens based on serological or other techniques using pooled or individual sera from allergic patients, or allergen-specific polyclonal or monoclonal antibodies.

Determination of the content of relevant allergens is performed wherever possible. This determination may be made using individual allergen-specific reference standards, when available. The choice of the relevant allergen components subjected to the determination must be justified. Individual allergens are identified and named according to internationally established nomenclature wherever possible.

The biological potency of the first IHRP is determined in patients by *in vivo* techniques such as skin testing, and expressed in units of biological activity, except when not enough patients are available. In this case, the potency of the first IHRP is determined by an *in vitro* method. Subsequently, the biological activity of future IHRPs is demonstrated by *in vitro* methods by comparison with the results obtained with the first IHRP. The *in vitro* potency may be measured by a suitable immunoassay (for example, an assay based on the inhibition of the binding capacity of specific immunoglobulin E antibodies).

IDENTIFICATION

The tests for identification are performed as late as possible in the manufacturing process. In the case of products used on a named-patient basis, the control is performed on the active substance and/or at the intermediate stage between the active substance and the finished product.

Identity is confirmed by comparison with the IHRP using protein profiling by appropriate methods (for example, isoelectric focusing, sodium dodecyl sulfate polyacrylamide gel electrophoresis, immunoelectrophoresis, immunoblotting, liquid chromatography or mass spectrometry).

In exceptional cases, if no IHRP is available, a representative batch may be used to confirm identity.

Identity may also be confirmed by comparison with individual allergen-specific reference standards, when available.

TESTS

The tests are performed as late as possible in the manufacturing process. In the case of products used on a named-patient basis, the control is performed on the active substance and/or at the intermediate stage between the active substance and the finished product.

Various biochemical and immunological tests have been developed in order to characterise allergens qualitatively and quantitatively. In those cases where such methods cannot be applied, particularly for the determination of allergenic activity and allergen and/or protein profile, justification must be provided.

Water (*2.5.12* or *2.5.32*) or **loss on drying** (*2.2.32*)

Maximum 5 per cent for freeze-dried products. In the case of oral lyophilisates, the water content may be higher than 5 per cent, where justified and authorised.

Sterility (*2.6.1*)

Allergen products presented as parenteral preparations, eye preparations, preparations for inhalation or preparations for skin testing comply with the test for sterility.

Microbial contamination

For non-sterile allergen products, recommendations are provided in general chapter *5.1.4. Microbiological quality of non-sterile pharmaceutical preparations and substances for pharmaceutical use.*

Protein content (*2.5.33*)

80 per cent to 120 per cent of the stated content, unless otherwise justified and authorised. If the biological potency can be determined then the test for protein content is performed as a batch-to-batch consistency test and the protein content is within 50 per cent to 150 per cent of the stated content. When the finished product contains proteinaceous excipients, the test for protein content is performed as late as possible during production before addition of the proteinaceous excipient.

Protein profile

The protein profile determined by suitable methods corresponds to that of the IHRP. The presence of relevant allergen components is verified, where possible. The choice of relevant allergen components to be tested for must be justified.

Various additional tests, some with increasing selectivity, depending on the allergen product concerned can be applied, but in any case for allergen products intended for therapeutic use, a validated test measuring the potency (total allergenic activity, determination of individual allergens or any other justified tests) must be applied.

Aluminium (*2.5.13*)

80 per cent to 120 per cent of the stated amount but in any case not more than 1.25 mg per human dose unless otherwise justified and authorised, when aluminium hydroxide or aluminium phosphate is used as adsorbent.

Calcium (*2.5.14*)

80 per cent to 120 per cent of the stated amount when calcium phosphate is used as adsorbent.

Allergen profile

Relevant allergenic components are identified by means of suitable techniques using allergen-specific human or animal antibodies.

Total allergenic activity

50 per cent to 150 per cent of the stated amount as assayed by inhibition of the binding capacity of specific

immunoglobulin E antibodies or a suitable equivalent *in vitro* method.

Individual allergens
50 per cent to 200 per cent of the stated amount of each relevant allergen component, determined by a suitable method.

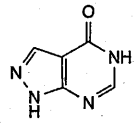
STORAGE
Adsorbed allergen products are not to be frozen, unless otherwise justified and authorised.

LABELLING
The label states:
— the name of the allergen product;
— the biological potency and/or the protein content and/or the extraction concentration;
— the route of administration and the intended use;
— the storage conditions;
— where applicable, the name and amount of added antimicrobial preservative;
— where applicable, for freeze-dried preparations:
  — the name, composition and volume of the reconstituting liquid to be added;
  — the period of time within which the preparation is to be used after reconstitution;
— where applicable, that the preparation is sterile;
— where applicable, the name and amount of adsorbent.

Ph Eur

Allopurinol

(Ph. Eur. monograph 0576)



$C_5H_4N_4O$ 136.1 *315-30-0*

Action and use
Xanthine oxidase inhibitor; treatment of gout and hyperuricaemia.

Preparations
Allopurinol Oral Suspension
Allopurinol Tablets

Ph Eur

DEFINITION
1,5-Dihydro-4*H*-pyrazolo[3,4-*d*]pyrimidin-4-one.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Very slightly soluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: B.
Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 10 mg in 1 mL of a 4 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*. Dilute 10.0 mL of this solution to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*.

Spectral range 220-350 nm.

Absorption maximum At 250 nm.

Absorption minimum At 231 nm.

Absorbance ratio A_{231}/A_{250} = 0.52 to 0.62.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *allopurinol CRS*.

C. Dissolve 0.3 g in 2.5 mL of *dilute sodium hydroxide solution R* and add 50 mL of *water R*. Add slowly and with shaking 5 mL of *silver nitrate solution R1*. A white precipitate is formed which does not dissolve on the addition of 5 mL of *ammonia R*.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *concentrated ammonia R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *allopurinol CRS* in *concentrated ammonia R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous ethanol R, methylene chloride R* (40:60 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Use freshly prepared solutions. Store and inject them at 8 °C, using a cooled autosampler.*

Test solution (a) Dissolve 25.0 mg of the substance to be examined in 2.5 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 50.0 mL with the mobile phase.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in 5.0 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 250.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *allopurinol impurity A CRS*, 5 mg of *allopurinol impurity B CRS* and 5.0 mg of *allopurinol impurity C CRS* in 5.0 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 20.0 mg of *allopurinol CRS* in 5.0 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 250.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 1.25 g/L solution of *potassium dihydrogen phosphate R*.

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Run time Twice the retention time of allopurinol.

Elution order Impurity A, impurity B, impurity C, allopurinol.

Retention time Allopurinol = about 10 min.

System suitability Reference solution (b):
— *resolution*: minimum 1.1 between the peaks due to impurities B and C.

Limits:
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *impurity C*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than A, B and C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurities D and E

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions. Store and inject them at 8 °C, using a cooled autosampler.*

Solution A 1.25 g/L solution of *potassium dihydrogen phosphate R*.

Test solution Dissolve 50.0 mg of the substance to be examined in 5.0 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 100.0 mL with solution A.

Reference solution Dissolve 5.0 mg of *allopurinol impurity D CRS* and 5.0 mg of *allopurinol impurity E CRS* in 5.0 mL of a 4 g/L solution of *sodium hydroxide R* and dilute immediately to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase *methanol R*, 1.25 g/L solution of *potassium dihydrogen phosphate R* (10:90 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 1.5 times the retention time of impurity E.

Retention times Impurity D = about 3.6 min; impurity E = about 4.5 min.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to impurities D and E.

Limits:
— *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent);
— *impurity E*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Impurity F

Liquid chromatography (*2.2.29*).

Under the following conditions, any hydrazine in the sample reacts with benzaldehyde to give benzaldehyde azine.

Solvent mixture Mix equal volumes of *dilute sodium hydroxide solution R* and *methanol R*.

Solution A Dissolve 2.0 g of *benzaldehyde R* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Prepare immediately before use.

Test solution Dissolve 250.0 mg of the substance to be examined in 5 mL of the solvent mixture. Add 4 mL of solution A, mix and allow to stand for 2.5 h at room temperature. Add 5.0 mL of *hexane R* and shake for 1 min. Allow the layers to separate and use the upper layer.

Reference solution Dissolve 10.0 mg of *hydrazine sulfate R* in the solvent mixture by sonicating for about 2 min and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture. To 5.0 mL of the solution obtained, add 4 mL of solution A, mix and allow to stand for 2.5 h at room temperature. Add 5.0 mL of *hexane R* and shake for 1 min. Allow the layers to separate and use the upper layer.

Blank solution To 5 mL of the solvent mixture add 4 mL of solution A, mix and allow to stand for 2.5 h at room temperature. Add 5.0 mL of *hexane R* and shake for 1 min. Allow the layers to separate and use the upper layer.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *cyanosilyl silica gel for chromatography R* (5 µm) with a pore size of 10 nm;
— *temperature*: 30 °C.

Mobile phase *2-propanol R*, *hexane R* (5:95 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 310 nm.

Injection 20 µL.

Relative retention With reference to benzaldehyde (retention time = about 2.8 min): benzaldehyde azine = about 0.8.

System suitability Reference solution:
— *resolution*: minimum 2 between the peaks due to benzaldehyde azine and benzaldehyde;
— *signal-to-noise ratio*: minimum 20 for the peak due to benzaldehyde azine.

Limit:
— *impurity F*: the area of the peak due to benzaldehyde azine in the chromatogram obtained with the test solution is not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (10 ppm of hydrazine sulfate equivalent to 2.5 ppm of hydrazine).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

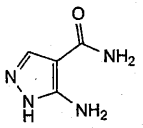
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

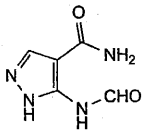
Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_5H_4N_4O$ from the declared content of *allopurinol CRS*.

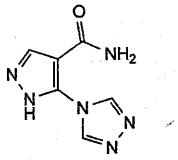
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



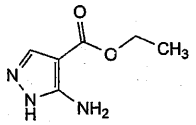
A. 5-amino-1*H*-pyrazole-4-carboxamide,



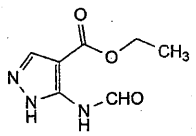
B. 5-(formylamino)-1*H*-pyrazole-4-carboxamide,



C. 5-(4*H*-1,2,4-triazol-4-yl)-1*H*-pyrazole-4-carboxamide,



D. ethyl 5-amino-1*H*-pyrazole-4-carboxylate,



E. ethyl 5-(formylamino)-1*H*-pyrazole-4-carboxylate,

F. diazane (hydrazine).

Ph Eur

Almagate

(Ph. Eur. monograph 2010)

$Al_2Mg_6C_2O_{20}H_{14}4H_2O$ 630 *66827-12-1*

Action and use
Antacid.

Ph Eur

DEFINITION
Hydrated aluminium magnesium hydroxycarbonate.

Content
— aluminium: 15.0 per cent to 17.0 per cent (calculated as Al_2O_3),
— magnesium: 36.0 per cent to 40.0 per cent (calculated as MgO),
— carbonic acid: 12.5 per cent to 14.5 per cent (calculated as CO_2).

CHARACTERS
Appearance
White or almost white, fine, crystalline powder.

Solubility
Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride. It dissolves with effervescence and heating in dilute mineral acids.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of almagate.*

B. Dissolve 0.15 g in *dilute hydrochloric acid R* and dilute to 20 mL with the same acid. 2 mL of the solution gives the reaction of aluminium (*2.3.1*).

C. 2 mL of the solution prepared under identification test B gives the reaction of magnesium (*2.3.1*).

TESTS
pH (*2.2.3*)
9.1 to 9.7.

Disperse 4.0 g in 100 mL of *carbon dioxide-free water R*, stir for 2 min and filter.

Neutralising capacity
Carry out the test at 37 °C Disperse 0.5 g in 100 mL of *water R*, heat, add 100.0 mL of *0.1 M hydrochloric acid*, previously heated and stir continuously; the pH (*2.2.3*) of the solution between 5 min and 20 min is not less than 3.0 and not greater than 4.5. Add 10.0 mL of *0.5 M hydrochloric acid*, previously heated, stir continuously for 1 h and titrate with *0.1 M sodium hydroxide* to pH 3.5; not more than 20.0 mL of *0.1 M sodium hydroxide* is required.

Chlorides (*2.4.4*)
Maximum 0.1 per cent.

Dissolve 0.33 g in 5 mL of *dilute nitric acid R* and dilute to 100 mL with *water R*. Prepare simultaneously the standard by diluting 0.7 mL of *dilute nitric acid R* to 5 mL with *water R* and adding 10 mL of *chloride standard solution (5 ppm Cl) R*.

Sulfates (*2.4.13*)
Maximum 0.4 per cent.

Dissolve 0.25 g in 5 mL of *dilute hydrochloric acid R* and dilute to 100 mL with *distilled water R*. Prepare simultaneously the standard by adding 0.8 mL of *dilute hydrochloric acid R* to 15 mL of *sulfate standard solution (10 ppm SO_4) R*.

Sodium
Maximum 150 ppm.
Atomic absorption spectrometry (*2.2.23, Method I*).
Test solution Dissolve 0.25 g in 50 mL of a 103 g/L solution of *hydrochloric acid R*.
Reference solutions Prepare the reference solutions using *sodium standard solution (200 ppm Na) R*, diluted as necessary with a 103 g/L solution of *hydrochloric acid R*.

Loss on ignition
43.0 per cent to 49.0 per cent, determined on 1.000 g by ignition at 900 ± 50 °C.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).
TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).
Absence of *Escherichia coli* (*2.6.13*).
Absence of *Pseudomonas aeruginosa* (*2.6.13*).

ASSAY
Aluminium
Dissolve 1.000 g in 5 mL of *hydrochloric acid R*, heating if necessary. Allow to cool to room temperature and dilute to 100.0 mL with *water R* (solution A). Introduce 10.0 mL of solution A into a 250 mL conical flask, add 25.0 mL of 0.05 M sodium edetate, 20 mL of *buffer solution pH 3.5 R*, 40 mL of *ethanol R* and 2 mL of a freshly prepared 0.25 g/L solution of *dithizone R* in *ethanol R*. Titrate the excess of sodium edetate with *0.05 M zinc sulfate* until the colour changes from greenish-violet to pink.
1 mL of 0.05 M sodium edetate is equivalent to 2.549 mg of Al_2O_3.

Magnesium
Introduce 10.0 mL of solution A prepared in the assay of aluminium into a 500 mL conical flask, add 200 mL of *water R*, 20 mL of *triethanolamine R* with shaking, 10 mL of *ammonium chloride buffer solution pH 10.0 R* and 50 mg of *mordant black 11 triturate R*. Titrate with 0.05 M sodium edetate until the colour changes from violet to pure blue.
1 mL of 0.05 M sodium edetate is equivalent to 2.015 mg of MgO.

Carbonic acid
12.5 per cent to 14.5 per cent.
Test sample Place 7.00 mg of the substance to be examined in a tin capsule. Seal the capsule.
Reference sample Place 7.00 mg of *almagate CRS* in a tin capsule. Seal the capsule.
Introduce separately the test sample and the reference sample into a combustion chamber of a CHN analyser purged with *helium for chromatography R* and maintained at a temperature of 1020 °C. Simultaneously, introduce *oxygen R* at a pressure of 40 kPa and a flow rate of 20 mL/min and allow complete combustion of the sample. Sweep the combustion gases through a reduction reactor and separate the gases formed by gas chromatography (*2.2.28*).
Column:
— *size*: *l* = 2 m, Ø = 4 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R1*.
Carrier gas *helium for chromatography R*.
Flow rate 100 mL/min.
Temperature:
— *column*: 65 °C;
— *detector*: 190 °C.
Detection Thermal conductivity.
Run time 16 min.
System suitability:
— average percentage of carbon in 5 reference samples must be within ± 0.2 per cent of the value assigned to the CRS; the difference between the upper and the lower values of the percentage of carbon in these samples must be below 0.2 per cent.

Calculate the percentage content of carbonic acid in the test sample according to the following formula:

$$C \times K \times \frac{A}{m}$$

C = percentage content of carbonic acid in the reference sample;
K = mean value for the 5 reference samples of the ratio of the mass in milligrams to the area of the peak due to carbonic acid;
A = area of the peak due to carbonic acid in the chromatogram obtained with the test sample;
m = sample mass, in milligrams.

STORAGE
In an airtight container.

Ph Eur

Virgin Almond Oil

Almond Oil
(Ph. Eur. monograph 0261)

Preparation
Almond Oil Ear Drops

Ph Eur

DEFINITION
Fatty oil obtained by cold expression from the ripe seeds of *Prunus dulcis* (Mill.) D.A.Webb var. *dulcis* or *Prunus dulcis* (Mill.) D.A.Webb var. *amara* (DC.) Buchheim or a mixture of both varieties.

CHARACTERS
Appearance
Yellow, clear liquid.

Solubility
Slightly soluble in ethanol (96 per cent), miscible with light petroleum.

Relative density
About 0.916.
It solidifies at about −18 °C.

IDENTIFICATION
First identification: A, C.
Second identification: A, B.
A. Absorbance (see Tests).
B. Identification of fatty oils by thin-layer chromatography (*2.3.2*).
Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.
C. Composition of fatty acids (see Tests).

TESTS
Specific absorbance (*2.2.25*)
Maximum 0.2, determined at the absorption maximum at 270 nm. The ratio of the absorbance measured at 232 nm to that measured at 270 nm is greater than 7.
To 0.100 g add *cyclohexane R* and dilute to 10.0 mL with the same solvent. Adapt the concentration of the solution so that

the absorbance lies between 0.5 and 1.5, measured in a 1 cm cell.

Acid value (*2.5.1*)
Maximum 2.0, determined on 5.0 g.

Peroxide value (*2.5.5, Method A*)
Maximum 15.0.

Unsaponifiable matter (*2.5.7*)
Maximum 0.9 per cent, determined on 5.0 g.

Composition of fatty acids (*2.4.22, Method A*)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty-acid fraction of the oil:
— *saturated fatty acids of chain length less than C_{16}*: maximum 0.1 per cent,
— *palmitic acid*: 4.0 per cent to 9.0 per cent,
— *palmitoleic acid*: maximum 0.8 per cent,
— *margaric acid*: maximum 0.2 per cent,
— *stearic acid*: maximum 3.0 per cent,
— *oleic acid*: 62.0 per cent to 86.0 per cent,
— *linoleic acid*: 20.0 per cent to 30.0 per cent,
— *linolenic acid*: maximum 0.4 per cent,
— *arachidic acid*: maximum 0.2 per cent,
— *eicosenoic acid*: maximum 0.3 per cent,
— *behenic acid*: maximum 0.2 per cent,
— *erucic acid*: maximum 0.1 per cent.

Sterols (*2.4.23*)
Composition of sterol fraction of the oil:
— *cholesterol*: maximum 0.7 per cent,
— *campesterol*: maximum 4.0 per cent,
— *stigmasterol*: maximum 3.0 per cent,
— *β-sitosterol*: 73.0 per cent to 87.0 per cent,
— *Δ5-avenasterol*: minimum 10.0 per cent,
— *Δ7-stigmastenol*: maximum 3.0 per cent,
— *Δ7-avenasterol*: maximum 3.0 per cent,
— *brassicasterol*: maximum 0.3 per cent.

Water (*2.5.32*)
Maximum 0.1 per cent, determined on 1.00 g.

STORAGE
In a well-filled container, protected from light.

Ph Eur

Refined Almond Oil

(Ph. Eur. monograph 1064)

Ph Eur

DEFINITION
Fatty oil obtained from the ripe seeds of *Prunus dulcis* (Mill.) D.A. Webb var. *dulcis* or *Prunus dulcis* (Mill.) D.A. Webb var. *amara* (DC.) Buchheim or a mixture of both varieties by cold expression. It is then refined. A suitable antioxidant may be added.

CHARACTERS

Appearance
Pale yellow, clear liquid.

Solubility
Slightly soluble in ethanol (96 per cent), miscible with light petroleum.

Relative density
About 0.916.

It solidifies at about −18 °C.

IDENTIFICATION
A. Identification of fatty oils by thin-layer chromatography (*2.3.2*).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

B. Composition of fatty acids (see Tests).

TESTS

Specific absorbance (*2.2.25*)
0.2 to 6.0, determined at the absorption maximum at 270 nm.

To 0.100 g add *cyclohexane R* and dilute to 10.0 mL with the same solvent. Adapt the concentration of the solution so that the absorbance lies between 0.5 and 1.5, measured in a 1 cm cell.

Acid value (*2.5.1*)
Maximum 0.5, determined on 5.0 g.

Peroxide value (*2.5.5, Method A*)
Maximum 5.0.

Unsaponifiable matter (*2.5.7*)
Maximum 0.9 per cent, determined on 5.0 g.

Composition of fatty acids (*2.4.22, Method A*)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty-acid fraction of the oil:
— *saturated fatty acids of chain length less than C_{16}*: maximum 0.1 per cent;
— *palmitic acid*: 4.0 per cent to 9.0 per cent;
— *palmitoleic acid*: maximum 0.8 per cent;
— *margaric acid*: maximum 0.2 per cent;
— *stearic acid*: maximum 3.0 per cent;
— *oleic acid*: 62.0 per cent to 86.0 per cent;
— *linoleic acid*: 20.0 per cent to 30.0 per cent;
— *linolenic acid*: maximum 0.4 per cent;
— *arachidic acid*: maximum 0.2 per cent;
— *eicosenoic acid*: maximum 0.3 per cent;
— *behenic acid*: maximum 0.2 per cent;
— *erucic acid*: maximum 0.1 per cent.

Sterols (*2.4.23*)
Composition of the sterol fraction of the oil:
— *cholesterol*: maximum 0.7 per cent;
— *campesterol*: maximum 5.0 per cent;
— *stigmasterol*: maximum 4.0 per cent;
— *β-sitosterol*: 73.0 per cent to 87.0 per cent;
— *Δ5-avenasterol*: minimum 5.0 per cent;
— *Δ7-stigmastenol*: maximum 3.0 per cent;
— *Δ7-avenasterol*: maximum 3.0 per cent;
— *brassicasterol*: maximum 0.3 per cent.

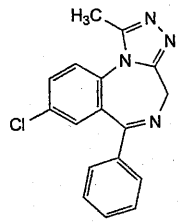
Water (*2.5.32*)
Maximum 0.1 per cent, determined on 1.00 g.

STORAGE
In a well-filled container, protected from light.

Ph Eur

Alprazolam

(Ph. Eur. monograph 1065)



$C_{17}H_{13}ClN_4$ 308.8 *28981-97-7*

Action and use
Benzodiazepine.

Ph Eur

DEFINITION
8-Chloro-1-methyl-6-phenyl-4*H*-[1,2,4]triazolo[4,3-*a*][1,4]benzodiazepine.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in acetone and in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Dissolve the substance to be examined in the smallest necessary quantity of *ethyl acetate R* and evaporate to dryness on a water-bath. Thoroughly mix 5.0 mg of the substance to be examined with 5.0 mg of *alprazolam CRS*. The melting point (*2.2.14*) of the mixture does not differ by more than 2 °C from the melting point of the substance to be examined.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *alprazolam CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethyl acetate R*, evaporate to dryness on a water-bath and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *alprazolam CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *alprazolam CRS* and 10 mg of *midazolam CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, methanol R, ethyl acetate R* (2:15:20:80 *V/V/V/V*).

Application 5 µL.

Development Over a path of 12 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separately spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 7.7 g of *ammonium acetate R* in 1000 mL of *water R* and adjust to pH 4.2 with *glacial acetic acid R.*

Test solution Dissolve 0.100 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of *alprazolam CRS* and 2 mg of *triazolam CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with *dimethylformamide R.* Dilute 0.5 mL of this solution to 10.0 mL with *dimethylformamide R.*

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: phenylsilyl silica gel for chromatography R1* (5 µm).

Mobile phase:
— *mobile phase A*: buffer solution, *methanol R* (44:56 *V/V*);
— *mobile phase B*: buffer solution, *methanol R* (5:95 *V/V*);
— *temperature*: 40 °C;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	98	2
15 - 35	98 → 1	2 → 99
35 - 40	1	99

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL; inject *dimethylformamide R* as a blank.

Retention time Triazolam = about 9 min; alprazolam = about 10 min.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to triazolam and alprazolam.

Limits:
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

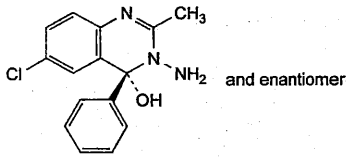
Dissolve 0.140 g in 50 mL of a mixture of 2 volumes of *acetic anhydride R* and 3 volumes of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Titrate to the 2nd point of inflexion.

1 mL of *0.1 M perchloric acid* is equivalent to 15.44 mg of $C_{17}H_{13}ClN_4$.

STORAGE

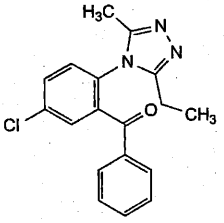
Protected from light.

IMPURITIES

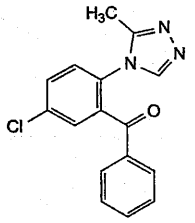


and enantiomer

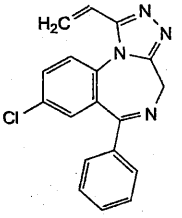
A. (4*RS*)-3-amino-6-chloro-2-methyl-4-phenyl-3,4-dihydroquinazolin-4-ol,



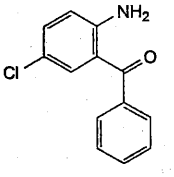
B. [5-chloro-2-[3-(hydroxymethyl)-5-methyl-4*H*-1,2,4-triazol-4-yl]phenyl]phenylmethanone,



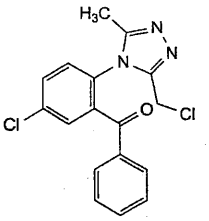
C. [5-chloro-2-[3-methyl-4*H*-1,2,4-triazol-4-yl]phenyl] phenylmethanone,



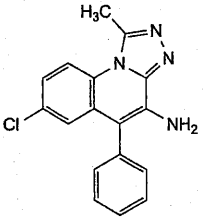
D. 8-chloro-1-ethenyl-6-phenyl-4*H*-[1,2,4]triazolo[4,3-*a*] [1,4]benzodiazepine,



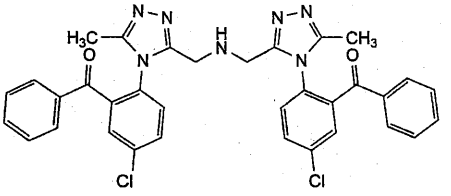
E. (2-amino-5-chlorophenyl)phenylmethanone,



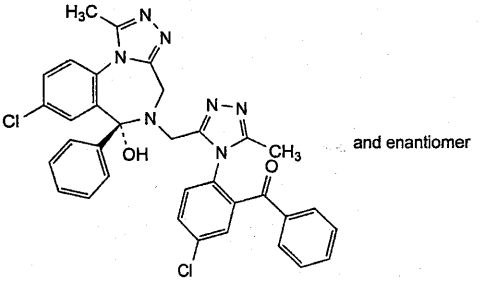
F. [5-chloro-2-[3-(chloromethyl)-5-methyl-4*H*-1,2,4-triazol-4-yl]phenyl]phenylmethanone,



G. 7-chloro-1-methyl-5-phenyl[1,2,4]triazolo[4,3-*a*]quinolin-4-amine,

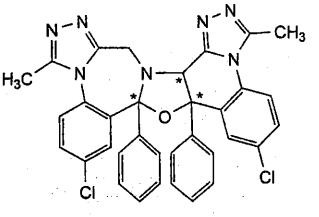


H. bis[[4-(2-benzoyl-4-chlorophenyl)-5-methyl-4*H*-1,2,4-triazol-3-yl]methyl]amine,



and enantiomer

I. [5-chloro-2-[3-[[(6*RS*)-8-chloro-6-hydroxy-1-methyl-6-phenyl-4*H*-[1,2,4]triazolo[4,3-*a*][1,4]benzodiazepin-5 (6*H*)-yl]methyl]-5-methyl-4*H*-1,2,4-triazol-4-yl]phenyl] phenylmethanone,

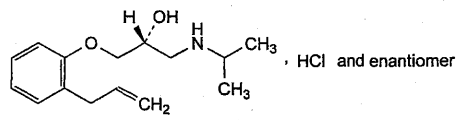


J. 2,17-dichloro-6,13-dimethyl-18b,19a-diphenyl-8b,19adihydro-10*H*,18b*H*-[1,2,4]triazolo [4‴,3‴:1″,2″]quinolo[3″,4″:4′,5′]oxazolo[3′,2′-*d*]-1,2,4-triazolo[4,3-*a*][1,4]benzodiazepine.

Ph Eur

Alprenolol Hydrochloride

(Ph. Eur. monograph 0876)



$C_{15}H_{24}ClNO_2$ 285.8 *13707-88-5*

Action and use
Beta-adrenoceptor antagonist.

Ph Eur

DEFINITION

(2*RS*)-1-[(1-Methylethyl)amino]-3-[2-(prop-2-enyl) phenoxy]propan-2-ol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 108 °C to 112 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alprenolol hydrochloride CRS.*

C. Examine the chromatograms obtained in the test for impurity D.

Detection Examine in daylight, after exposure to iodine vapour for 30 min.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_9 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.2 mL of *methyl red solution R* and 0.2 mL of *0.01 M hydrochloric acid*; the solution is red. Add 0.4 mL of *0.01 M sodium hydroxide*; the solution is yellow.

Impurity C
Maximum 0.1 per cent.

Dissolve 0.25 g in *ethanol (96 per cent) R* and dilute to 25 mL with the same solvent. The absorbance (*2.2.25*) measured at 297 nm is not greater than 0.20.

Impurity D
Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.50 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with *methanol R.*

Reference solution (a) Dissolve 10 mg of *alprenolol hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *alprenolol hydrochloride CRS* and 10 mg of *oxprenolol hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dilute 5 mL of test solution (b) to 50 mL with *methanol R.*

Plate *TLC silica gel G plate R.*

Mobile phase Place 2 beakers each containing 30 mL of *ammonia R* at the bottom of the tank containing a mixture of 5 volumes of *methanol R* and 95 volumes of *ethyl acetate R.*

Application 5 µL.

Development Over a path of 15 cm in a tank saturated for at least 1 h.

Drying At 100 °C for 15 min.

Detection Expose to iodine vapour for up to 6 h.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Limits Test solution (a):
— *impurity D*: any spot with an R_F value greater than that of the principal spot is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 4.0 mg of *alprenolol hydrochloride CRS* and 0.8 mg of *4-isopropylphenol R* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 4.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 0.656 g of *sodium octanesulfonate R* with 150 mL of *acetonitrile R* and dilute to 500 mL with phosphate buffer pH 2.8 prepared as follows: mix 1.78 g of *phosphoric acid R* and 15.6 g of *sodium dihydrogen phosphate R* and dilute to 2000 mL with *water R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Equilibration With the mobile phase for about 1 h.

Injection 20 µL.

Run time Twice the retention time of alprenolol.

Retention time Alprenolol = about 11 min; 4-isopropylphenol = about 18 min.

System suitability Reference solution (a):
— *resolution*: minimum 5 between the peaks due to alprenolol and 4-isopropylphenol; if necessary, adjust the concentration of sodium octanesulfonate and/or acetonitrile in the mobile phase (increase the

concentration of sodium octanesulfonate to increase the retention time of alprenolol and increase the concentration of acetonitrile to decrease the retention times of both compounds).

Limits:

— *unspecified impurities*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.04 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide R* at a pressure not exceeding 2.7 kPa.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 25 mL of a mixture of equal volumes of *anhydrous ethanol R* and *water R*. Add 10 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 28.58 mg of $C_{15}H_{24}ClNO_2$.

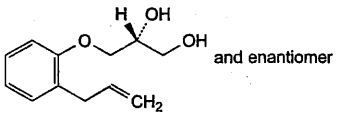
STORAGE

Protected from light.

IMPURITIES

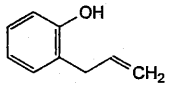
Specified impurities *C, D.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*

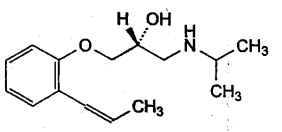


and enantiomer

A. (2*RS*)-3-[2-(prop-2-enyl)phenoxy]propan-1,2-diol,

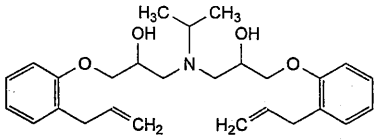


B. 2-(prop-2-enyl)phenol,



and enantiomer

C. (2*RS*)-1-[(1-methylethyl)amino]-3-[2-(prop-1-enyl) phenoxy]propan-2-ol,

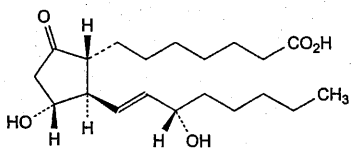


D. 1,1′-[(1-methylethyl)imino]bis[3-[2-(prop-2-enyl)phenoxy] propan-2-ol].

Ph Eur

Alprostadil

(Ph. Eur. monograph 1488)



$C_{20}H_{34}O_5$ 354.5 *745-65-3*

Action and use
Prostaglandin E_1 (PGE_1).

Ph Eur

DEFINITION

7-[(1*R*,2*R*,3*R*)-3-Hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoic acid.

Content
95.0 per cent to 102.5 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellowish, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in acetone, slightly soluble in ethyl acetate.

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): −70 to −60 (anhydrous substance).

Immediately before use, dissolve 50 mg in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alprostadil CRS.*

C. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions protected from light.*

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R1* and *water R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 100 µL of the test solution to 20.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

Reference solution (b) Dissolve 1.0 mg of *dinoprostone impurity C CRS* (alprostadil impurity H) and 1.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R1* and *water R* and dilute to 20.0 mL with the same mixture of solvents.

Reference solution (c) In order to prepare impurities A and B *in situ*, dissolve 1 mg of the substance to be examined in 100 µL of *1 M sodium hydroxide* (the solution becomes brownish-red), wait for 3 min and add 100 µL of a 112 g/L solution of *phosphoric acid R* (yellowish-white opalescent solution); dilute to 5.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

System A

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (4 µm) with a pore size of 6 nm;
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.9 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1.0 L with the same solvent; adjust to pH 2.5 with a 2.9 g/L solution of *phosphoric acid R* (approximately 600 mL is required); to 740 mL of the buffer solution add 260 mL of *acetonitrile R1*;
— *mobile phase B*: dissolve 3.9 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1.0 L with the same solvent; adjust to pH 2.5 with a 2.9 g/L solution of *phosphoric acid R* (approximately 600 mL is required); to 200 mL of the buffer solution add 800 mL of *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 75	100	0
75 - 76	100 → 0	0 → 100
76 - 86	0	100
86 - 87	0 → 100	100 → 0
87 - 102	100	0

Flow rate 1 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

Retention time Alprostadil = about 63 min.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to impurity H and alprostadil in the chromatogram obtained with reference solution (b).

System B

Use the same conditions as for system A with the following mobile phase and elution programme:
— *mobile phase A*: dissolve 3.9 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1.0 L with the same solvent; adjust to pH 2.5 with a 2.9 g/L solution of *phosphoric acid R* (approximately 600 mL is required); to 600 mL of the buffer solution add 400 mL of *acetonitrile R1*;
— *mobile phase B*: use mobile phase B as described under system A;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 50	100	0
50 - 51	100 → 0	0 → 100
51 - 61	0	100
61 - 62	0 → 100	100 → 0
62 - 72	100	0

Relative retention With reference to alprostadil (retention time = about 7 min): impurity A = about 2.4; impurity B = about 2.6.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to impurity A and impurity B in the chromatogram obtained with reference solution (c).

Carry out the test according to system A and B.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the impurities listed in Table 1488.-1 by the corresponding correction factor;

Table 1488.-1.

Impurity	Relative retention (system A)	Relative retention (system B)	Correction factor
impurity G	0.80	-	0.7
impurity F	0.88	-	0.8
impurity D	0.90	-	1.0
impurity H	0.96	-	0.7
impurity E	1.10	-	0.7
impurity C	-	1.36	1.9
impurity K	-	1.85	0.06
impurity A	-	2.32	0.7
impurity B	-	2.45	1.5
impurity I	-	4.00	1.0
impurity J	-	5.89	1.0

— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *any other impurity*: not more than 1.8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.9 per cent), and not more than 1 such peak has an area greater than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent). Evaluate impurities appearing at relative retentions less than 1.2 by system A and impurities appearing at relative retentions greater than 1.2 by system B;
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.32*)

Maximum 0.5 per cent, determined on 50 mg.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances, system A. *Prepare the solutions protected from light.*

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R1* and

water R and dilute to 25.0 mL with the same mixture of solvents. Dilute 3.0 mL of the solution to 20.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

Reference solution Dissolve 5.0 mg of *alprostadil CRS* in a mixture of equal volumes of *acetonitrile R1* and *water R* and dilute to 25.0 mL with the same mixture of solvents. Dilute 6.0 mL of the solution to 20.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

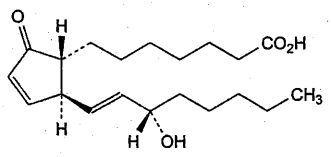
Injection 20 µL.

Calculate the percentage content of $C_{20}H_{34}O_5$ taking into account the assigned content of *alprostadil CRS*.

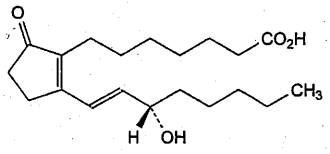
STORAGE

At a temperature of 2 °C to 8 °C.

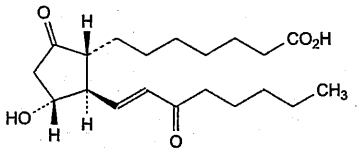
IMPURITIES



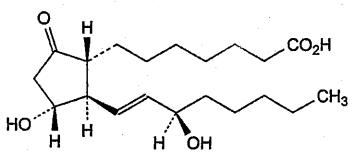
A. 7-[(1*R*,2*S*)-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopent-3-enyl]heptanoic acid (prostaglandin A_1),



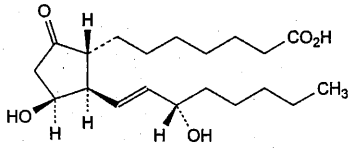
B. 7-[2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopent-1-enyl]heptanoic acid (prostaglandin B_1),



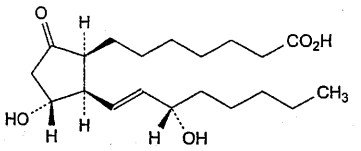
C. 7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*)-3-oxooct-1-enyl]-5-oxocyclopentyl]heptanoic acid (15-oxoprostaglandin E_1),



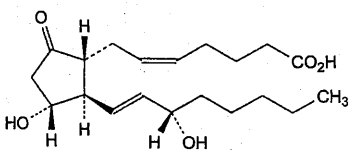
D. 7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*R*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoic acid (15-epiprostaglandin E_1),



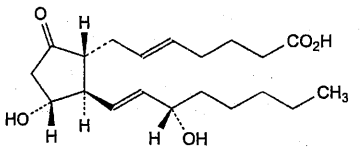
E. 7-[(1*R*,2*R*,3*S*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoic acid (11-epiprostaglandin E_1),



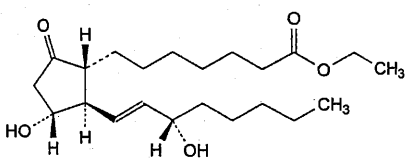
F. 7-[(1*S*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoic acid (8-epiprostaglandin E_1),



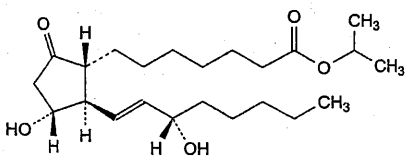
G. (5*Z*)-7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (dinoprostone),



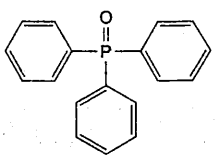
H. (5*E*)-7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid ((5*E*)-prostaglandin E_2),



I. ethyl 7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoate (prostaglandin E_1, ethyl ester),



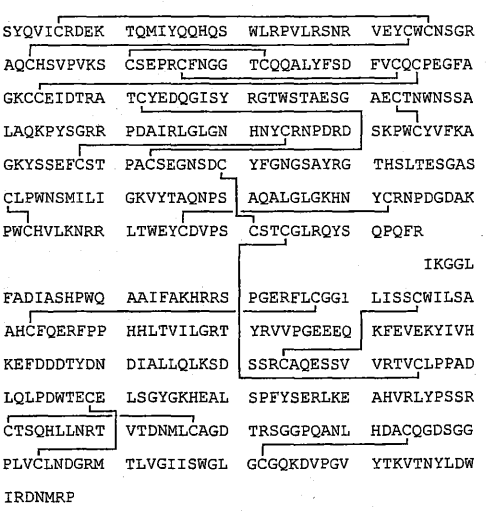
J. 1-methylethyl 7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(1*E*,3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]heptanoate (prostaglandin E_1, isopropyl ester),



K. triphenylphosphine oxide.

Alteplase for Injection

(Ph. Eur. monograph 1170)



Action and use
Tissue-type plasminogen activator; fibrinolytic.

Ph Eur

DEFINITION

Alteplase for injection is a sterile, freeze-dried preparation of alteplase, a tissue plasminogen activator produced by recombinant DNA technology. It has a potency of not less than 500 000 IU per milligram of protein.

Tissue plasminogen activator binds to fibrin clots and activates plasminogen, leading to the generation of plasmin and to the degradation of fibrin clots or blood coagulates.

Alteplase consists of 527 amino acids with a calculated relative molecular mass of 59 050 without consideration of the carbohydrate moieties attached at positions Asn 117, Asn 184 and Asn 448. The total relative molecular mass is approximately 65 000. Alteplase is cleaved by plasmin between amino-acids 275 and 276 into a two-chain form (A chain and B chain) that are connected by a disulfide bridge between Cys 264 and Cys 395. The single-chain form and the two-chain form show comparable fibrinolytic activity *in vitro*.

PRODUCTION

Alteplase is produced by recombinant DNA synthesis in cell culture; the fermentation takes place in serum-free medium.

The purification process is designed to remove efficiently potential impurities, such as antibiotics, DNA and protein contaminants derived both from the host cell and from the production medium, and potential viral contaminants.

If alteplase is stored in bulk form, stability (maintenance of potency) in the intended storage conditions must be demonstrated.

The production, purification and product consistency are checked by a number of analytical methods described below, carried out routinely as in-process controls.

Protein content
The protein concentration of alteplase solutions is determined by measuring the absorbance (*2.2.25*) of the protein solution at 280 nm and at 320 nm, using formulation buffer as the compensation liquid. If dilution of alteplase samples is necessary, the samples are diluted in formulation buffer. For the calculation of the alteplase concentration, the absorbance value ($A_{280} - A_{320}$) is divided by the specific absorption coefficient for alteplase of 1.9.

Potency
The potency of alteplase is determined in an *in vitro* clot-lysis assay as described under Assay. The specific activity of bulk alteplase is approximately 580 000 IU per milligram of alteplase.

***N*-terminal sequence**
N-terminal sequencing is applied to determine the correct *N*-terminal sequence and to determine semiquantitatively additional cleavage sites in the alteplase molecule, for example at position AA 275-276 or at position AA 27-28. The *N*-terminal sequence must conform with the sequence of human tissue plasminogen activator.

Isoelectric focusing
The consistency in the microheterogeneity of glycosylation of the alteplase molecule can be demonstrated by isoelectric focusing (IEF). A complex banding pattern with 10 major and several minor bands in the pH range 6.5-8.5 is observed. Denaturing conditions are applied to achieve a good separation of differently charged variants of alteplase. The broad charge distribution observed is due to a population of molecules, which differ in the fine structure of biantenary and triantenary complex-type carbohydrate residues, with different degrees of substitution with sialic acids. The banding pattern of alteplase test samples must be consistent with the pattern of alteplase reference standard.

Single-chain alteplase content
The alteplase produced by CHO (Chinese hamster ovary) cells in serum-free medium is predominantly single-chain alteplase. The single-chain form can be separated from the two-chain form by gel-permeation liquid chromatography under reducing conditions as described under Single-chain content (see Tests). The single-chain alteplase content in bulk samples must be higher than 60 per cent.

Tryptic-peptide mapping
The primary structure of the alteplase molecule is verified by tryptic-peptide mapping as described under Identification B. The reduced and carboxymethylated molecule is cleaved by trypsin into about 50 peptides, which are separated by reverse-phase liquid chromatography. A characteristic chromatogram (fingerprint) is obtained. The identity of the tryptic-peptide map of a given alteplase sample with the profile of a well-characterised reference standard is an indirect confirmation of the amino-acid sequence, because even single amino-acid exchanges in individual peptides can be detected by this sensitive technique. In addition, complex peaks of the glycopeptides can be isolated from the tryptic-peptide map and separated in a second dimension, either by reverse-phase liquid chromatography under modified conditions or by capillary electrophoresis. By this two-dimensional separation of glycopeptide variants, lot-to-lot consistency of the microheterogeneity of glycosylation can be demonstrated.

The tryptic-peptide map of alteplase samples must be consistent with the tryptic-peptide map of alteplase reference standard.

Monomer content
The monomer content of alteplase is measured by gel-permeation liquid chromatography under non-reduced conditions as described under Monomer content (see Tests). The monomer content of alteplase bulk samples must be higher than 95 per cent.

Type I/Type II alteplase content
CHO cells produce 2 glycosylation variants of alteplase. Type I alteplase contains 1 polymannose-type glycosylation at position Asn 117 and 2 complex-type glycosylation sites at positions Asn 184 and Asn 448. Type II alteplase is only glycosylated at positions Asn 117 and Asn 448.

The ratio of Type I/Type II alteplase is constant in the range of 45 to 65 per cent of Type I and 35 to 55 per cent of Type II. The content of alteplase Type I and Type II can be determined by a densitometric scan of SDS-PAGE (sodium dodecyl sulfate polyacrylamide gel electrophoresis) gel. Plasmin-treated samples of alteplase, which are reduced and carboxymethylated before loading on the gel, are separated into 3 bands: Type I alteplase A-chain (AA 1-275), Type II alteplase A-chain (AA 1-275) and alteplase B-chain (AA 276-527). The ratio of Type I/Type II alteplase is determined from a calibration curve, which is obtained by a densitometric scan of defined mixtures of purified Type I alteplase and Type II alteplase standards.

SDS-PAGE
SDS-PAGE (silver staining) is used to demonstrate purity of the alteplase bulk material and the integrity of the alteplase molecule. For alteplase bulk samples, no additional protein bands compared to reference standard or degradation products must occur in SDS-PAGE gels at a loading amount of 2.5 µg alteplase protein per lane and a limit of detection of 5 ng per protein (BSA) band.

Bacterial endotoxins (*2.6.14*)
Less than 1 IU per milligram of alteplase.

Sialic acids
Proceed using a suitable validated method developed according to general chapter *2.2.59. Glycan analysis of glycoproteins*. The sialic acids content for the test samples must be in the range of 70 to 130 per cent compared to alteplase reference standard, which contains about 3 moles of sialic acids per mole of alteplase.

Neutral sugars
Dilute alteplase samples and the reference standard in the assay buffer, containing 34.8 g/L of *arginine R*, 0.1 g/L of *polysorbate 80 R* and adjusted to pH 7.4 with *phosphoric acid R*, to a protein concentration of 50 µg/mL. Prepare the following concentrations of mannose in the same assay buffer for a calibration curve: 20, 30, 40, 50 and 60 µg/mL. Pipette 2 mL of alteplase samples and reference standard, as well as 2 mL of each mannose concentration in duplicate in reagent tubes. Add 50 µL of *phenol R*, followed by 5 mL of *sulfuric acid R*, in each reagent tube. Incubate the mixture for 30 min at room temperature. Measure the absorbance at 492 nm for each tube. Read the content of neutral sugars from the mannose calibration curve. The neutral sugar content is expressed in moles of neutral sugar per mole of alteplase, taking into account the dilution factor for alteplase samples and reference standard and using a relative molecular mass of 180.2 for mannose and a relative molecular mass of 59 050 for the alteplase protein moiety. The neutral sugar content of the alteplase samples must be in the range of 70 to 130 per cent compared to alteplase reference standard, which contains about 12 moles of neutral sugar per mole of alteplase.

CHARACTERS
White or slightly yellow powder or solid friable mass.

Reconstitute the preparation as stated on the label immediately before carrying out the Identification, Tests (except those for solubility and water) and Assay.

IDENTIFICATION
A. The assay serves also to identify the preparation.

B. Tryptic-peptide mapping. Examine by liquid chromatography (*2.2.29*).

Test solution Dilute the preparation to be examined with *water R* to obtain a solution containing about 1 mg of alteplase per millilitre. Dialyse about 2.5 mL of the solution for at least 12 h into a solution containing 480 g/L of *urea R*, 44 g/L of *tris(hydroxymethyl)aminomethane R* and 1.5 g/L of *sodium edetate R* and adjusted to pH 8.6, using a membrane with a cut-off point corresponding to a relative molecular mass of 10 000 for globular proteins. Measure the volume of the solution, transfer it to a clean test-tube and add per millilitre 10 µL of a 156 g/L solution of *dithiothreitol R*. Allow to stand for 4 h, cool in iced water and add per millilitre of solution 25 µL of a freshly prepared 190 g/L solution of *iodoacetic acid R*. Allow to stand in the dark for 30 min. Add per millilitre 50 µL of dithiothreitol solution to stop the reaction. Dialyse for 24 h against an 8 g/L solution of *ammonium hydrogen carbonate R*. Add 1 part of *trypsin for peptide mapping R* to 100 parts of the protein and allow to stand for 6 h to 8 h. Repeat the addition of trypsin and allow to stand for a total of 24 h.

Reference solution Prepare as for the test solution using a suitable reference standard instead of the preparation to be examined.

The chromatographic procedure may be carried out using:
- a column 0.1 m long and 4.6 mm in internal diameter packed with *octadecylsilyl silica gel for chromatography R* (5 µm to 10 µm);

  Mobile phase A 8 g/L solution of *sodium dihydrogen phosphate R*, adjusted to pH 2.85 with *phosphoric acid R*, filtered and degassed;

  Mobile phase B 75 per cent *V/V* solution of *acetonitrile R* in mobile phase A;
- as detector a spectrophotometer set at 210 nm.

Equilibrate the system with mobile phase A at a flow rate of 1 mL/min. After injection of the solution, increase the proportion of mobile phase B at a rate of 0.44 per cent per minute until the ratio of mobile phase A to mobile phase B is 60:40, then increase the proportion of mobile phase B at a rate of 1.33 per cent per minute until the ratio of mobile phase A to mobile phase B is 20:80 and then continue elution with this mixture for a further 10 min. Record the chromatogram for the reference solution: the test is not valid unless the resolution of peaks 6 (peptides 268-275) and 7 (peptides 1-7) is at least 1.5; w_{h1} and w_{h2} are not more than 0.4 min. Inject about 100 µL of the test solution and record the chromatogram. Verify the identity of the peaks by comparison with the chromatograms of the reference solution. There should not be any additional significant peaks or shoulders, a significant peak or shoulder being defined as one with an area response equal to or greater than 5 per cent of peak 19 (peptides 278-296); no significant peak is missing. A type chromatogram for identification of the peaks cited is shown in Figure 1170.-1.

TESTS
Appearance of solution
The reconstituted preparation is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)
7.1 to 7.5.

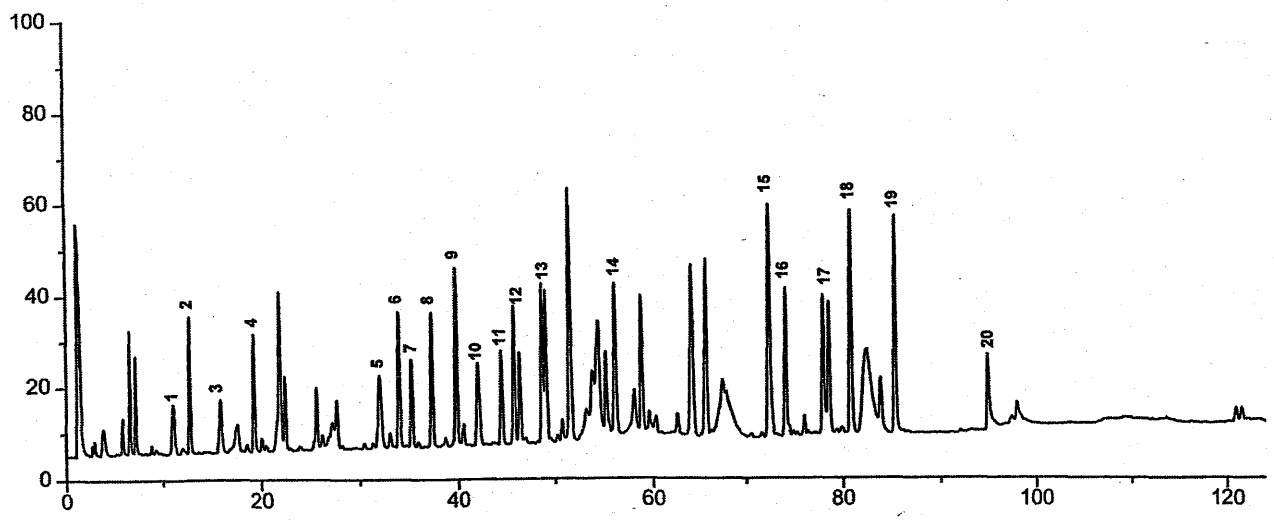


Figure 1170.-1. – *Chromatogram for tryptic-peptide mapping of alteplase*

Solubility
Add the volume of the liquid stated on the label.
The preparation dissolves completely within 2 min at 20 °C to 25 °C.

Protein content
Prepare a solution of the substance to be examined with an accurately known concentration of about 1 g/L. Using a 34.8 g/L solution of *arginine R* adjusted to pH 7.3 with *phosphoric acid R*, dilute an accurately measured volume of the solution of the substance to be examined so that the absorbance measured at the maximum at about 280 nm is 0.5 to 1.0 (*test solution*). Measure the absorbance (*2.2.25*) of the solution at the maximum at about 280 nm and at 320 nm using the arginine solution as the compensation liquid. Calculate the protein content in the portion of alteplase taken from the following expression:

$$\frac{V(A_{280} - A_{320})}{1.9}$$

in which V is the volume of the test solution, A_{280} is the absorbance at the maximum at about 280 nm and A_{320} is the absorbance at 320 nm.

Single-chain content
Examine by liquid chromatography (*2.2.29*).

Test solution Dissolve the preparation to be examined in *water R* to obtain a solution containing about 1 mg of alteplase per millilitre. Place about 1 mL of the solution in a tube, add 3 mL of a 3 g/L solution of *dithiothreitol R* in the mobile phase, place a cap on the tube and heat at about 80 °C for 3 min to 5 min.

The chromatographic procedure may be carried out using:
— a column 0.6 m long and 7.5 mm in internal diameter packed with silica-based, rigid, hydrophilic gel with spherical particles 10 µm to 13 µm in diameter, suitable for size-exclusion chromatography;
— as mobile phase at a flow rate of 0.5 mL/min a solution containing 30 g/L of *sodium dihydrogen phosphate R* and 1 g/L of *sodium dodecyl sulfate R*, adjusted to pH 6.8 with *dilute sodium hydroxide solution R*;
— as detector a spectrophotometer set at 214 nm.

Inject about 50 µL of the test solution and record the chromatogram. The chromatogram shows 2 major peaks corresponding to single-chain and two-chain alteplase.

Calculate the relative amount of single-chain alteplase from the peak area values.

The test is not valid unless: the number of theoretical plates calculated on the basis of the single-chain alteplase peak is at least 1000. The content of single-chain alteplase is not less than 60 per cent of the total amount of alteplase-related substances found.

Monomer content
Examine by liquid chromatography (*2.2.29*).

Test solution Reconstitute the preparation to be examined to obtain a solution containing about 1 mg per millilitre.

The chromatographic procedure may be carried out using:
— a column 0.6 m long and 7.5 mm in internal diameter packed with silica-based rigid, hydrophilic gel with spherical particles 10 µm to 13 µm in diameter, suitable for size-exclusion chromatography;
— as mobile phase at a flow rate of 0.5 mL/min a solution containing 30 g/L of *sodium dihydrogen phosphate R* and 1 g/L of *sodium dodecyl sulfate R*, adjusted to pH 6.8 with *dilute sodium hydroxide solution R*;
— as detector a spectrophotometer set at 214 nm.

Inject the test solution and record the chromatogram.
The test is not valid unless the number of theoretical plates calculated for the alteplase monomer peak is at least 1000. Measure the response for all peaks, i.e. peaks corresponding to alteplase species of different molecular masses. Calculate the relative content of monomer from the area values of these peaks. The monomer content for alteplase must be at least 95 per cent.

Water (*2.5.12*)
Not more than 4.0 per cent, determined by the semi-micro determination of water.

Bacterial endotoxins (*2.6.14*)
Less than 1 IU per milligram of protein.

Sterility (*2.6.1*)
It complies with the test for sterility.

ASSAY
The potency of alteplase is determined by comparing its ability to activate plasminogen to form plasmin with the same capacity of a reference preparation calibrated in International Units. The formation of plasmin is measured by the determination of the lysis time of a fibrin clot in given conditions.

The International Unit is the activity of a stated quantity of the International Standard of alteplase. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Solvent buffer A solution containing 1.38 g/L of *sodium dihydrogen phosphate monohydrate R*, 7.10 g/L of *anhydrous disodium hydrogen phosphate R*, 0.20 g/L of *sodium azide R* and 0.10 g/L of *polysorbate 80 R*.

Human thrombin solution A solution of *human thrombin R* containing 33 IU/mL in solvent buffer.

Human fibrinogen solution A 2 g/L solution of *fibrinogen R* in solvent buffer.

Human plasminogen solution A 1 g/L solution of *human plasminogen R* in solvent buffer.

Test solutions Using a solution of the substance to be examined containing 1 g/L, prepare serial dilutions using solvent buffer, for example 1:5000, 1:10 000, 1:20 000.

Reference solutions Using a solution of a suitable reference standard having an accurately known concentration of about 1 g/L (580 000 IU of alteplase per millilitre), prepare 5 serial dilutions using *water R* to obtain reference solutions having known concentrations in the range 9.0 IU/mL to 145 IU/mL.

To each of a set of labelled glass test-tubes, add 0.5 mL of human thrombin solution. Allocate each test and reference solution to a separate tube and add to each tube 0.5 mL of the solution allocated to it. To each of a second set of labelled glass tubes, add 20 µL of human plasminogen solution, and 1 mL of human fibrinogen solution, mix and store on ice. Beginning with the reference/thrombin mixture containing the lowest number of International Units per millilitre, record the time and separately add 200 µL of each of the thrombin mixtures to the test tubes containing the plasminogen-fibrinogen mixture. Using a vortex mixer, intermittently mix the contents of each tube for a total of 15 s and carefully place in a rack in a circulating water-bath at 37 °C. A visibly turbid clot forms within 30 s and bubbles subsequently form within the clot. Record the clot-lysis time as the time between the first addition of alteplase solution and the moment when the last bubble rises to the surface. Using a least-squares fit, determine the equation of the line using the logarithms of the concentrations of the reference preparation in International Units per millilitre versus the logarithms of the values of their clot-lysis times in seconds, according to the following equation:

$$\log_{10} t = a + b(\log_{10} U_S)$$

in which t is the clot-lysis time, U_S the activity in International Units per millilitre of the reference preparation, b is the slope and a the y-intercept of the line. The test is not valid unless the correlation coefficient is −0.9900 to −1.0000. From the line equation and the clot-lysis time for the test solution, calculate the logarithm of the activity U_A from the following equation:

$$\log_{10} U_A = \frac{[(\log_{10} t) - a]}{b}$$

Calculate the alteplase activity in International Units per millilitre from the following expression:

$$D \times U_A$$

in which D is the dilution factor for the test solution.

Calculate the specific activity in the portion of the substance to be examined from the following expression:

$$\frac{U_A}{P}$$

in which P is the concentration of protein obtained in the test for protein content.

The estimated potency is not less than 90 per cent and not more than 110 per cent of the stated potency.

STORAGE

Store in a colourless, glass container, under vacuum or under an inert gas, protected from light, at a temperature of 2 °C to 30 °C.

LABELLING

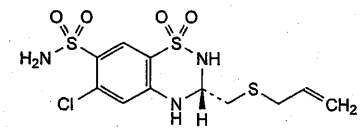
The label states:

— the number of International Units per container;
— the amount of protein per container;
— the name and volume of the liquid to be used for reconstitution.

Ph Eur

Altizide

(Ph. Eur. monograph 2185)



and enantiomer

$C_{11}H_{14}ClN_3O_4S_3$ 383.9 *5588-16-9*

Action and use
Thiazide diuretic.

Ph Eur

DEFINITION

(3*RS*)-6-Chloro-3-[(prop-2-enylsulfanyl)methyl]-3,4-dihydro-2*H*-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, soluble in methanol, practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *altizide CRS*.

If the spectra obtained show differences, dissolve 50 mg of the substance to be examined and 50 mg of the reference substance separately in 2 mL of *acetone R* and evaporate the solvent. Precipitate by adding 1 mL of *methylene chloride R*. Evaporate to dryness and record new spectra using the residues.

TESTS

Impurity B
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.200 g of the substance to be examined in *acetone R* and dilute to 2.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *altizide impurity B CRS* in *acetone R* and dilute to 25.0 mL with the same solvent.

Reference solution (b) To 1.0 mL of reference solution (a) add 1.0 mL of the test solution.

Reference solution (c) Dilute 5.0 mL of reference solution (a) to 10.0 mL with *acetone R*.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *acetone R, methylene chloride R* (25:75 *V/V*).

Application 10 µL of the test solution and reference solutions (b) and (c).

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a mixture of equal volumes of a 10 g/L solution of *potassium permanganate R* and a 50 g/L solution of *sodium carbonate R*, prepared immediately before use. Allow to stand for 30 min and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Limit Any spot due to impurity B is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use, except reference solution (b).*

Test solution Dissolve 50 mg of the substance to be examined in 5 mL of *acetonitrile R* and dilute to 25 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) In order to produce impurity A *in situ*, dissolve 50 mg of the substance to be examined in 5 mL of *acetonitrile R* and dilute to 25 mL with *water R*. Allow to stand for 30 min.

Reference solution (c) Dissolve 4 mg of *furosemide CRS* in 2 mL of *acetonitrile R*, add 2 mL of the test solution and dilute to 100 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *acetonitrile R, water R* previously adjusted to pH 2.0 with *perchloric acid R* (25:75 *V/V*).

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 5 µL.

Run time Twice the retention time of altizide.

Relative retention With reference to altizide (retention time = about 25 min): impurity A = about 0.15; furosemide = about 1.05.

System suitability Reference solution (c):
— *resolution*: minimum 1.0 between the peaks due to altizide and furosemide.

Limits:
— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 50.0 mg.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

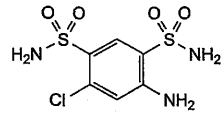
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances, with the following modifications.

Test solution Dissolve 25.0 mg of the substance to be examined in 2 mL of *acetonitrile R* and dilute to 25.0 mL with the mobile phase.

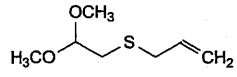
Reference solution Dissolve 25.0 mg of *altizide CRS* in 2 mL of *acetonitrile R* and dilute to 25.0 mL with the mobile phase.

Calculate the percentage content of $C_{11}H_{14}ClN_3O_4S_3$ from the declared content of *altizide CRS*.

IMPURITIES
Specified impurities *A, B.*



A. 4-amino-6-chlorobenzene-1,3-disulfonamide,



B. 3-[(2,2-dimethoxyethyl)sulfanyl]prop-1-ene.

Ph Eur

Alum

Potash Alum
Aluminium Potassium Sulphate
Aluminium Potassium Sulfate
(Ph. Eur. monograph 0006)

$AlK(SO_4)_2,12H_2O$ 474.4 *7784-24-9*

Action and use
Astringent.

Ph Eur

DEFINITION
Content
99.0 per cent to 100.5 per cent of $AlK(SO_4)_2,12H_2O$.

CHARACTERS
Appearance
Granular powder or colourless, transparent, crystalline masses.

Solubility
Freely soluble in water, very soluble in boiling water, soluble in glycerol, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Solution S (see Tests) gives the reactions of sulfates (*2.3.1*).

B. Solution S gives the reaction of aluminium (*2.3.1*).

C. Shake 10 mL of solution S with 0.5 g of *sodium hydrogen carbonate R* and filter. The filtrate gives reaction (a) of potassium (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in *water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
3.0 to 3.5.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Ammonium (*2.4.1*)
Maximum 0.2 per cent.

To 1 mL of solution S add 4 mL of *water R*. Dilute 0.5 mL of this solution to 14 mL with *water R*.

Iron (*2.4.9*)
Maximum 100 ppm.

Dilute 2 mL of solution S to 10 mL with *water R*. Use in this test 0.3 mL of *thioglycollic acid R*.

ASSAY
Dissolve 0.900 g in 20 mL of *water R* and carry out the complexometric titration of aluminium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 47.44 mg of $AlK(SO_4)_2,12H_2O$.

Ph Eur

Aluminium Chloride Hexahydrate

(Ph. Eur. monograph 0971)

$AlCl_3,6H_2O$ 241.4 *7784-13-6*

Action and use
Astringent.

Preparation
Aluminium Chloride Solution

Ph Eur

DEFINITION
Content
95.0 per cent to 101.0 per cent.

CHARACTERS
Appearance
White or slightly yellow, crystalline powder or colourless crystals, deliquescent.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent), soluble in glycerol.

IDENTIFICATION
A. Dilute 0.1 mL of solution S2 (see Tests) to 2 mL with *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

B. Dilute 0.3 mL of solution S2 to 2 mL with *water R*. The solution gives the reaction of aluminium (*2.3.1*).

TESTS
Solution S1
Dissolve 10.0 g in *distilled water R* and dilute to 100 mL with the same solvent.

Solution S2
Dilute 50 mL of solution S1 to 100 mL with *water R*.

Appearance of solution
Solution S2 is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).

Sulfates (*2.4.13*)
Maximum 100 ppm, determined on solution S1.

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S1.

Alkali and alkaline-earth metals
Maximum 0.5 per cent.

To 20 mL of solution S2 add 100 mL of *water R* and heat to boiling. To the hot solution add 0.2 mL of *methyl red solution R*. Add *dilute ammonia R1* until the colour of the indicator changes to yellow and dilute to 150 mL with *water R*. Heat to boiling and filter. Evaporate 75 mL of the filtrate to dryness on a water-bath and ignite to constant mass. The residue weighs a maximum of 2.5 mg.

Water (*2.5.12*)
42.0 per cent to 48.0 per cent, determined on 50.0 mg.

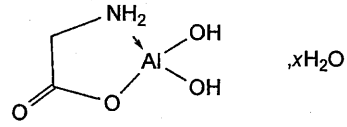
ASSAY
Dissolve 0.500 g in 25.0 mL of *water R*. Carry out the complexometric titration of aluminium (*2.5.11*). Titrate with *0.1 M zinc sulfate* until the colour of the indicator changes from greyish-green to pink. Carry out a blank titration.

1 mL of *0.1 M sodium edetate* is equivalent to 24.14 mg of $AlCl_3,6H_2O$.

STORAGE
In an airtight container.

Ph Eur

Aluminium Glycinate



$,xH_2O$

$C_2H_6AlNO_4,xH_2O$ 135.1 *41354-48-7*

Action and use
Antacid.

DEFINITION
Aluminium Glycinate is a basic aluminium monoglycinate, partly hydrated. It contains not less than 34.5% and not more than 38.5% of Al_2O_3 and not less than 9.9% and not more than 10.8% of N, both calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white powder.

Practically insoluble in *water* and in organic solvents. It dissolves in dilute mineral acids and in aqueous solutions of the alkali hydroxides.

IDENTIFICATION

A. Add 0.1 g to 10 mL of a solution prepared by dissolving 0.84 g of *citric acid* in 8 mL of *1M sodium hydroxide* and diluting to 20 mL with *water*. Add 0.5 mL of a 0.1% w/v solution of *ninhydrin* in *methanol* and warm. A purple colour is produced.

B. Suspend 1 g in 25 mL of *0.5M hydrochloric acid* and heat gently until a clear solution is produced. Reserve half of the solution. To 2 mL of the solution add 0.15 mL of *liquefied phenol*, shake and add carefully without shaking 5 mL of *dilute sodium hypochlorite solution*. A blue colour is produced.

C. The solution reserved in test B yields the reaction characteristic of *aluminium salts*, Appendix VI.

TESTS

Acidity or alkalinity

pH of a suspension of 1 g in 25 mL of *carbon dioxide-free water*, 6.5 to 7.5, Appendix V L.

Neutralising capacity

Shake 0.2 g vigorously with 25 mL of *0.1M hydrochloric acid* for 5 minutes and allow to stand for 5 minutes. The pH of the mixture is greater than 3.0, Appendix V L.

Arsenic

Dissolve 2.0 g in 18 mL of *brominated hydrochloric acid* and 32 mL of *water*. 25 mL of the resulting solution complies with the *limit test for arsenic*, Appendix VII (1 ppm).

Mercuric salts

Dissolve 2.0 g in 10 mL of *1M sulfuric acid*, transfer to a separating funnel with the aid of *water*, dilute to about 50 mL with *water* and add 50 mL of *0.5M sulfuric acid*. Add 100 mL of *water*, 2 g of *hydroxylamine hydrochloride*, 1 mL of *0.05M disodium edetate* and 1 mL of *glacial acetic acid*. Add 5 mL of *chloroform*, shake, allow to separate and discard the chloroform layer. Titrate the aqueous layer with a solution of *dithizone* in *chloroform* containing 8 µg per mL until the chloroform layer remains green. After each addition, shake vigorously, allow the layers to separate and discard the chloroform layer. Repeat the operation using a solution prepared by diluting 1 mL of *mercury standard solution (5 ppm Hg)* to 100 mL with *0.5M sulfuric acid* and beginning at the words 'Add 100 mL of *water*...'. The volume of the dithizone solution required by the substance being examined does not exceed that required by the mercury standard solution.

Chloride

Dissolve 1.0 g in 10 mL of *2M nitric acid* and dilute to 100 mL with *water*. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (330 ppm).

Loss on drying

When dried to constant weight at 130°, loses not more than 12.0% of its weight. Use 1 g.

ASSAY

For Al_2O_3

Dissolve 0.25 g in a mixture of 3 mL of *1M hydrochloric acid* and 50 mL of *water*, add 50 mL of *0.05M disodium edetate VS* and neutralise with *1M sodium hydroxide* using *methyl red solution* as indicator. Heat the solution to boiling, allow to stand for 10 minutes on a water bath, cool rapidly, add about 50 mg of *xylenol orange triturate* and 5 g of *hexamine* and titrate the excess of disodium edetate with *0.05M lead nitrate VS* until the solution becomes red. Each mL of *0.05M disodium edetate VS* is equivalent to 2.549 mg of Al_2O_3.

Hydrated Aluminium Hydroxide for Adsorption

(Ph. Eur. monograph 1664)

$[AlO(OH)]_x,xH_2O$

Ph Eur

DEFINITION

Content

90.0 per cent to 110.0 per cent of the content of aluminium stated on the label.

NOTE: shake the gel vigorously for at least 30 s immediately before examining.

CHARACTERS

Appearance

White or almost white, translucent, viscous, colloidal gel. A supernatant may be formed upon standing.

Solubility

A clear or almost clear solution is obtained with alkali hydroxide solutions and mineral acids.

IDENTIFICATION

Solution S (see Tests) gives the reaction of aluminium.

To 10 mL of solution S add about 0.5 mL of *dilute hydrochloric acid R* and about 0.5 mL of *thioacetamide reagent R*. No precipitate is formed. Add dropwise 5 mL of *dilute sodium hydroxide solution R*. Allow to stand for 1 h. A gelatinous white precipitate is formed which dissolves upon addition of 5 mL of *dilute sodium hydroxide solution R*. Gradually add 5 mL of *ammonium chloride solution R* and allow to stand for 30 min. The gelatinous white precipitate is re-formed.

TESTS

Solution S

Add 1 g to 4 mL of *hydrochloric acid R*. Heat at 60 °C for 1 h, cool, dilute to 50 mL with *distilled water R* and filter if necessary.

pH *(2.2.3)*

5.5 to 8.5.

Adsorption power

Dilute the substance to be examined with *distilled water R* to obtain an aluminium concentration of 5 mg/mL. Prepare *bovine albumin R* solutions with the following concentrations of bovine albumin: 0.5 mg/mL, 1 mg/mL, 2 mg/mL, 3 mg/mL, 5 mg/mL and 10 mg/mL. If necessary, adjust the gel and the *bovine albumin R* solutions to pH 6.0 with *dilute hydrochloric acid R* or *dilute sodium hydroxide solution R*.

For adsorption, mix 1 part of the diluted gel with 4 parts of each of the solutions of *bovine albumin R* and allow to stand at room temperature for 1 h. During this time shake the mixture vigorously at least 5 times. Centrifuge or filter through a non-protein-retaining filter. Immediately determine the protein content *(2.5.33, Method 2)* of either the supernatant or the filtrate.

It complies with the test if no bovine albumin is detectable in the supernatant or filtrate of the 2 mg/mL *bovine albumin R* solution (maximum level of adsorption) and in the supernatant or filtrate of *bovine albumin R* solutions of lower concentrations. Solutions containing 3 mg/mL, 5 mg/mL and 10 mg/mL *bovine albumin R* may show bovine albumin in the supernatant or filtrate, proportional to the amount of bovine albumin in the solutions.

Sedimentation
If necessary, adjust the substance to be examined to pH 6.0 using *dilute hydrochloric acid R* or *dilute sodium hydroxide solution R*. Dilute with *distilled water R* to obtain an aluminium concentration of approximately 5 mg/mL. If the aluminium content of the substance to be examined is lower than 5 mg/mL, adjust to pH 6.0 and dilute with a 9 g/L solution of *sodium chloride R* to obtain an aluminium concentration of about 1 mg/mL. After shaking for at least 30 s, place 25 mL of the preparation in a 25 mL graduated cylinder and allow to stand for 24 h.

It complies with the test if the volume of the clear supernatant is less than 5 mL for the gel with an aluminium content of about 5 mg/mL.

It complies with the test if the volume of the clear supernatant is less than 20 mL for the gel with an aluminium content of about 1 mg/mL.

Chlorides (*2.4.4*)
Maximum 0.33 per cent.

Dissolve 0.5 g in 10 mL of *dilute nitric acid R* and dilute to 500 mL with *water R*.

Nitrates
Maximum 100 ppm.

Place 5 g in a test-tube immersed in ice-water, add 0.4 mL of a 100 g/L solution of *potassium chloride R*, 0.1 mL of *diphenylamine solution R* and, dropwise with shaking, 5 mL of *sulfuric acid R*. Transfer the tube to a water-bath at 50 °C. After 15 min, any blue colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 5 mL of *nitrate standard solution (100 ppm NO_3) R*.

Sulfates (*2.4.13*)
Maximum 0.5 per cent.

Dilute 2 mL of solution S to 20 mL with *water R*.

Ammonium (*2.4.1, Method B*)
Maximum 50 ppm, determined on 1.0 g.

Prepare the standard using 0.5 mL of *ammonium standard solution (100 ppm NH_4) R*.

Arsenic (*2.4.2, Method A*)
Maximum 1 ppm, determined on 1 g.

Iron (*2.4.9*)
Maximum 15 ppm, determined on 0.67 g.

Bacterial endotoxins (*2.6.14*)
Less than 5 IU of endotoxin per milligram of aluminium, if intended for use in the manufacture of an adsorbed product without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Dissolve 2.50 g in 10 mL of *hydrochloric acid R*, heating for 30 min at 100 °C on a water-bath. Cool and dilute to 20 mL with *water R*. To 10 mL of the solution, add *concentrated ammonia R* until a precipitate is obtained. Add the smallest quantity of *hydrochloric acid R* needed to dissolve the precipitate and dilute to 20 mL with *water R*. Carry out the complexometric titration of aluminium (*2.5.11*). Carry out a blank titration.

STORAGE
At a temperature not exceeding 30 °C. Do not allow to freeze. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING
The label states the declared content of aluminium.

Ph Eur

Dried Aluminium Hydroxide

(Hydrated Aluminium Oxide, Ph. Eur. monograph 0311)

Action and use
Antacid.

Preparations
Aluminium Hydroxide Chewable Tablets
Aluminium Hydroxide Oral Suspension
Compound Magnesium Trisilicate Chewable Tablets
Co-magaldrox Oral Suspension
Co-magaldrox Tablets

Ph Eur

DEFINITION
Content
47.0 per cent to 60.0 per cent of Al_2O_3 (M_r 102.0).

CHARACTERS
Appearance
White or almost white, amorphous powder.

Solubility
Practically insoluble in water. It dissolves in dilute mineral acids and in solutions of alkali hydroxides.

IDENTIFICATION
Solution S (see Tests) gives the reaction of aluminium (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in 15 mL of *hydrochloric acid R*, heating on a water-bath. Dilute to 100 mL with *distilled water R*.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).

Alkaline impurities
Shake 1.0 g with 20 mL of *carbon dioxide-free water R* for 1 min and filter. To 10 mL of the filtrate add 0.1 mL of *phenolphthalein solution R*. Any pink colour disappears on the addition of 0.3 mL of *0.1 M hydrochloric acid*.

Neutralising capacity
Carry out the test at 37 °C Disperse 0.5 g in 100 mL of *water R*, heat, add 100.0 mL of *0.1 M hydrochloric acid*, previously heated, and stir continuously; the pH (*2.2.3*) of the solution after 10 min, 15 min and 20 min is not less than 1.8, 2.3 and 3.0 respectively and is at no time greater than 4.5. Add 10.0 mL of *0.5 M hydrochloric acid*, previously heated, stir continuously for 1 h and titrate with *0.1 M sodium hydroxide* to pH 3.5; not more than 35.0 mL of *0.1 M sodium hydroxide* is required.

Chlorides (*2.4.4*)
Maximum 1 per cent.

Dissolve 0.1 g with heating in 10 mL of *dilute nitric acid R* and dilute to 100 mL with *water R*. Dilute 5 mL of the solution to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 1 per cent.
Dilute 4 mL of solution S to 100 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 4 ppm, determined on 10 mL of solution S.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).
TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).
Absence of bile-tolerant gram-negative bacteria (*2.6.13*).
Absence of *Escherichia coli* (*2.6.13*).

ASSAY

Dissolve 0.800 g in 10 mL of *hydrochloric acid R1*, heating on a water-bath. Cool and dilute to 50.0 mL with *water R*. To 10.0 mL of the solution add *dilute ammonia R1* until a precipitate begins to appear. Add the smallest quantity of *dilute hydrochloric acid R* needed to dissolve the precipitate and dilute to 20 mL with *water R*. Carry out the complexometric titration of aluminium (*2.5.11*).
1 mL of *0.1 M sodium edetate* is equivalent to 5.098 mg of Al_2O_3.

STORAGE

In an airtight container, at a temperature not exceeding 30 °C.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hydrated aluminium oxide used as adsorbent.

Particle-size distribution (*2.9.31*)

Specific surface area (*2.9.26*)

Ph Eur

Aluminium Magnesium Silicate

(Ph. Eur. monograph 1388)

Action and use
Excipient.

Ph Eur

DEFINITION

Mixture of colloidal-size particles of montmorillonite and saponite, practically free from grit and non-swellable ore.
The requirements for viscosity and ratio of aluminium content to magnesium content differ for the several types of aluminium magnesium silicate, as shown in the table below.

Type	Viscosity (mPa·s)		Al content / Mg content	
	min.	max.	min.	max.
IA	225	600	0.5	1.2
IB	150	450	0.5	1.2
IC	800	2200	0.5	1.2
IIA	100	300	1.4	2.8

CHARACTERS

Appearance
Almost white, coarse powder, granules or plates (types IA, IC and IIA); almost white, fine powder (type IB).

Solubility
Practically insoluble in water and in organic solvents.
It swells in water to produce a colloidal dispersion.

IDENTIFICATION

Carry out either tests A, B, C, F or tests D, E.

A. In a platinum crucible mix 1.0 g with 5.0 g of *anhydrous lithium metaborate R*. Heat slowly at first and ignite at 1000-1200 °C for 15 min. Allow to cool and crush the residue. 0.25 g of the residue gives the reaction of silicates (*2.3.1*).

B. Dissolve 1.0 g of the residue obtained in identification test A in a mixture of 5 mL of *dilute hydrochloric acid R* and 10 mL of *water R*. Filter to obtain a clear solution and add *ammonium chloride buffer solution pH 10.0 R*. A white, gelatinous precipitate is formed. Centrifuge and keep the supernatant for identification test C. Dissolve the precipitate in *dilute hydrochloric acid R*. Add dropwise *dilute sodium hydroxide solution R*. A white gelatinous precipitate is formed. Filter and add a few drops of *phenolphthalein solution R* to the residue. The residue turns pink. Wash the residue with *water R* until the pink colour is completely discharged and the residue remains white upon addition of a drop of *phenolphthalein solution R*. Sprinkle a few crystals of *sodium fluoride R* on the residue. The residue, in contact with the crystals, turns pink again in a short time.

C. To 2 mL of the supernatant obtained after centrifugation in identification test B, add 1 mL of *dilute ammonia R1* and 1 mL of *ammonium chloride solution R*. Upon the addition of *dilute ammonia R1* a white precipitate may form, which dissolves after addition of the *ammonium chloride solution R*. Add 1 mL of *disodium hydrogen phosphate solution R*. A white precipitate is formed.

D. X-ray diffraction (*2.9.33*), oriented sample.

Add 2 g in small portions to 100 mL of *water R*, with vigorous shaking. Allow to stand for at least 12 h to ensure complete hydration. Place 2 mL of the resulting mixture on a suitable glass slide and allow to dry in air at room temperature to produce an oriented film. Place the slide in a vacuum desiccator over *ethylene glycol R*. Evacuate the desiccator and close the stopcock so that the ethylene glycol saturates the chamber. Allow to stand for at least 12 h. Record the X-ray diffraction pattern and calculate the *d* values: the largest peak corresponds to a *d* value between 1.5 nm and 1.72 nm.

E. X-ray diffraction (*2.9.33*), random sample.

Prepare a random powder sample, record the X-ray diffraction pattern and determine the *d* values in the region between 0.148 nm and 0.154 nm. Peaks are found between 0.1492 nm and 0.1504 nm and between 0.1510 nm and 0.1540 nm.

F. It complies with the limits of the assay.

TESTS

pH (*2.2.3*)

9.0 to 10.0.

Disperse 5.0 g in 100 mL of *carbon dioxide-free water R*.

Viscosity (*2.2.10*)

Weigh a quantity of the substance to be examined equivalent to 25.0 g of the dried substance and immediately transfer to a suitable 1 L blender jar containing a quantity of *water R*, at 25 ± 2 °C, that is sufficient to produce a mixture weighing 500 g. Blend for exactly 3 min, at 14 000-15 000 r/min. The heat generated during blending causes the temperature to rise to above 30 °C. Transfer the contents of the blender to a 600 mL beaker and allow to stand for 5 min. The sample temperature should be 33 ± 3 °C.

Using a suitable rotating viscometer equipped with a spindle as specified below, operate the viscometer at 60 r/min for exactly 6 min and record the scale reading.

For type IA, use a spindle with a cylinder 1.87 cm in diameter and 0.69 cm high attached to a shaft 0.32 cm in diameter, the distance from the top of the cylinder to the lower tip of the shaft being 2.54 cm, and an immersion depth of 5.00 cm (No. 2 spindle); if the scale reading is greater than 90 per cent of the full scale, repeat the measurement using a spindle similar to the No. 2 spindle but with a cylinder 1.27 cm in diameter and 0.16 cm high (No. 3 spindle).

For type IC, use a No. 3 spindle; if the scale reading is greater than 90 per cent of the full scale, repeat the measurement using a spindle with a cylindrical shaft 0.32 cm in diameter and an immersion depth of 4.05 cm (No. 4 spindle).

For types IB and IIA, use a No. 2 spindle.

Type	Viscosity (mPa·s)	
	min.	max.
IA	225	600
IB	150	450
IC	800	2200
IIA	100	300

Arsenic (*2.4.2, Method A*)

Maximum 3 ppm.

Transfer 16.6 g to a 250 mL beaker containing 100 mL of *dilute hydrochloric acid R*. Mix, cover with a watch glass and boil gently, with occasional stirring, for 15 min. Allow the insoluble matter to settle and decant the supernatant through a rapid-flow filter paper into a 250 mL volumetric flask, retaining as much sediment as possible in the beaker. To the residue in the beaker add 25 mL of hot *dilute hydrochloric acid R*, stir, heat to boiling, allow the insoluble matter to settle and decant the supernatant through the filter into the volumetric flask. Repeat the extraction with 4 additional quantities, each of 25 mL, of hot *dilute hydrochloric acid R*, decanting each supernatant through the filter into the volumetric flask. At the last extraction, transfer as much of the insoluble matter as possible onto the filter. Allow the combined filtrates to cool to room temperature and dilute to 250.0 mL with *dilute hydrochloric acid R*. Dilute 5.0 mL of this solution to 25.0 mL with *dilute hydrochloric acid R*.

Lead

Maximum 15 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Transfer 10.0 g to a 250 mL beaker containing 100 mL of *dilute hydrochloric acid R*. Mix, cover with a watch glass and boil for 15 min. Allow to cool to room temperature and allow the insoluble matter to settle. Decant the supernatant through a rapid-flow filter paper into a 400 mL beaker. To the insoluble matter in the 250 mL beaker add 25 mL of hot *water R*. Stir, allow the insoluble matter to settle and decant the supernatant through the filter into the 400 mL beaker. Repeat the extraction with 2 additional quantities, each of 25 mL, of *water R*, decanting each time the supernatant through the filter into the 400 mL beaker. Wash the filter with 25 mL of hot *water R*, collecting this filtrate in the 400 mL beaker. Concentrate the combined filtrates to about 20 mL by gently boiling. If a precipitate appears, add about 0.1 mL of *nitric acid R*, heat to boiling and allow to cool to room temperature. Filter the concentrated extracts through a rapid-flow filter paper into a 50 mL volumetric flask. Transfer the remaining contents of the 400 mL beaker through the filter paper and into the flask with *water R*. Dilute this solution to 50.0 mL with *water R*.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R*, diluted as necessary with *water R*.

Source Lead hollow-cathode lamp.

Wavelength 217 nm.

Atomisation device Oxidising air-acetylene flame.

Loss on drying (*2.2.32*)

Maximum 8.0 per cent, determined on 1.000 g by drying in an oven at 110 °C.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

ASSAY

Aluminium

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution In a platinum crucible mix 0.200 g with 1.0 g of *anhydrous lithium metaborate R*. Heat slowly at first and ignite at 1000-1200 °C for 15 min. Allow to cool, place the crucible in a 100 mL beaker containing 25 mL of a 50 g/L solution of *nitric acid R* and add 50 mL of a 50 g/L solution of *nitric acid R*, filling and submerging the crucible. Place a polytetrafluoroethylene-coated magnetic stirring bar in the crucible and stir gently with a magnetic stirrer until dissolution is complete. Transfer the solution to a 200 mL volumetric flask, wash the beaker, crucible and magnetic stirrer bar with *water R*, collecting the washings in the volumetric flask, and dilute to 200.0 mL with *water R* (solution A). To 20.0 mL of solution A add 20 mL of a 10 g/L solution of *sodium chloride R* and dilute to 100.0 mL with *water R*.

Reference solutions Dissolve, with gentle heating, 1.000 g of *aluminium R* in a mixture of 10 mL of *hydrochloric acid R* and 10 mL of *water R*. Allow to cool, then dilute to 1000.0 mL with *water R* (1 mg of aluminium per millilitre). Into 4 identical volumetric flasks, each containing 0.20 g of *sodium chloride R*, introduce 1.0 mL, 2.0 mL, 3.0 mL and 4.0 mL of this solution respectively, and dilute to 100.0 mL with *water R*.

Blank solution Dissolve 0.20 g of *sodium chloride R* in *water R* and dilute to 100.0 mL with the same solvent.

Source Aluminium hollow-cathode lamp.

Wavelength 309 nm.

Atomisation device Acetylene-nitrous oxide flame.

Magnesium

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dilute 25.0 mL of solution A, prepared in the assay for aluminium, to 50.0 mL with *water R*. To 5.0 mL of this solution add 20.0 mL of *lanthanum chloride solution R* and dilute to 100.0 mL with *water R*.

Reference solutions Place 1.000 g of *magnesium R* in a 250 mL beaker containing 20 mL of *water R* and carefully add 20 mL of *hydrochloric acid R*, warming if necessary to dissolve. Transfer the solution to a volumetric flask and dilute to 1000.0 mL with *water R* (1 mg of magnesium per millilitre). Dilute 5.0 mL of this solution to 500.0 mL with *water R*. Into 4 identical volumetric flasks, introduce 5.0 mL, 10.0 mL, 15.0 mL and 20.0 mL of the solution respectively. To each flask add 20.0 mL of *lanthanum chloride solution R* and dilute to 100.0 mL with *water R*.

Blank solution Dilute 20 mL of *lanthanum chloride solution R* to 100.0 mL with *water R*.

Source Magnesium hollow-cathode lamp.

Wavelength 285 nm.

Atomisation device Air-acetylene flame.

LABELLING

The label states the ratio of aluminium content to magnesium content, the viscosity and the corresponding type (see Definition).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for aluminium magnesium silicate used as viscosity-increasing agent and stabiliser.

Acid demand

Weigh a quantity of the substance to be examined equivalent to 5.00 g of the dried substance and disperse in 500 mL of *water R* using a suitable blender fitted with a 1 L jar. With constant mixing, add 3.0 mL portions of *0.1 M hydrochloric acid* at 5 s, 65 s, 125 s, 185 s, 245 s, 305 s, 365 s, 425 s, 485 s, 545 s, 605 s, 665 s and 725 s and add a 1.0 mL portion at 785 s. Determine the pH potentiometrically at 840 s. The pH is not greater than 4.0.

Viscosity

(see Tests).

Ph Eur

Dried Aluminium Phosphate

(Aluminium Phosphate, Hydrated, Ph. Eur. monograph 1598)

$AlPO_4,xH_2O$ 122.0
(anhydrous substance)

Action and use

Antacid.

Ph Eur

DEFINITION

Content

94.0 per cent to 102.0 per cent of $AlPO_4$ (M_r 122.0) (ignited substance).

It contains a variable quantity of water.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION

A. Solution S (see Tests) gives reaction (b) of phosphates (*2.3.1*).

B. Solution S gives the reaction of aluminium (*2.3.1*).

TESTS

Solution S

Dissolve 2.00 g in *dilute hydrochloric acid R* and dilute to 100 mL with the same acid.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

5.5 to 7.2

Shake 4.0 g with *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Chlorides (*2.4.4*)

Maximum 1.3 per cent.

Dissolve 50.0 mg in 10 mL of *dilute nitric acid R* and dilute to 200 mL with *water R*.

Soluble phosphates

Maximum 1.0 per cent, calculated as PO_4^{3-}.

Test solution Stir 5.0 g with 150 mL of *water R* for 2 h. Filter and wash the filter with 50 mL of *water R*. Combine the filtrate and the washings and dilute to 250.0 mL with *water R*. Dilute 10.0 mL of this solution to 100.0 mL with *water R*.

Reference solution (a) Dissolve 2.86 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 100 mL with the same solvent.

Reference solution (b) Dilute 1 mL of reference solution (a) to 5 mL with *water R*.

Reference solution (c) Dilute 3 mL of reference solution (a) to 5 mL with *water R*.

Treat each solution as follows. To 5.0 mL add 4 mL of *dilute sulfuric acid R*, 1 mL of *ammonium molybdate solution R*, 5 mL of *water R* and 2 mL of a solution containing 0.10 g of *4-methylaminophenol sulfate R*, 0.5 g of *anhydrous sodium sulfite R* and 20.0 g of *sodium metabisulfite R* in 100 mL of *water R*. Shake and allow to stand for 15 min. Dilute to

25.0 mL with *water R* and allow to stand for a further 15 min. Measure the absorbance (*2.2.25*) at 730 nm. Calculate the content of soluble phosphates from a calibration curve prepared using reference solutions (a), (b) and (c) after treatment.

Sulfates (*2.4.13*)
Maximum 0.6 per cent.

Dilute 8 mL of solution S to 100 mL with *distilled water R*.

Arsenic (*2.4.2*)
Maximum 1 ppm.

1.0 g complies with limit test A.

Loss on ignition
10.0 per cent to 20.0 per cent, determined on 1.000 g at 800 ± 50 °C.

Neutralising capacity
Add 0.50 g to 30 mL of *0.1 M hydrochloric acid* previously heated to 37 °C and maintain at this temperature for 15 min while stirring. The pH (*2.2.3*) of the mixture after 15 min at 37 °C is 2.0 to 2.5.

ASSAY
Dissolve 0.400 g in 10 mL of *dilute hydrochloric acid R* and dilute to 100.0 mL with *water R*. To 10.0 mL of the solution, add 10.0 mL of *0.1 M sodium edetate* and 30 mL of a mixture of equal volumes of *ammonium acetate solution R* and *dilute acetic acid R*. Boil for 3 min, then cool. Add 25 mL of *ethanol (96 per cent) R* and 1 mL of a freshly prepared 0.25 g/L solution of *dithizone R* in *ethanol (96 per cent) R*. Titrate the excess of sodium edetate with *0.1 M zinc sulfate* until the colour changes to pink.

1 mL of *0.1 M sodium edetate* is equivalent to 12.20 mg of $AlPO_4$.

STORAGE
In an airtight container.

Ph Eur

Aluminium Phosphate Gel

(Ph. Eur. monograph 2166)

Action and use
Antacid; vaccine adjuvant.

Ph Eur

DEFINITION
Hydrated $AlPO_4$ in gel form.

Content
19.0 per cent to 21.0 per cent of $AlPO_4$.

CHARACTERS
Appearance
Gel.

Solubility
Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute solutions of mineral acids.

IDENTIFICATION
A. Solution S (see Tests) gives reaction (b) of phosphates (*2.3.1*).

B. Solution S gives the reaction of aluminium (*2.3.1*).

C. It complies with the assay.

TESTS
Solution S
Dissolve 2.00 g in *dilute hydrochloric acid R* and dilute to 100 mL with the same acid.

pH (*2.2.3*)
6.0 to 8.0.

Peroxides
Maximum 150 ppm, expressed as hydrogen peroxide.

Test solution Dissolve with heating 1.0 g of the substance to be examined in 5 mL of *dilute hydrochloric acid R*, then add 5 mL of *water R* and 2 mL of *divanadium pentoxide solution in sulfuric acid R*.

Reference solution Dilute 1.0 mL of *dilute hydrogen peroxide solution R* to 200.0 mL with *water R*. To 1 mL of this solution add 9 mL of *water R* and 2 mL of *divanadium pentoxide solution in sulfuric acid R*.

The test solution is not more intensely coloured than the reference solution.

Chlorides (*2.4.4*)
Maximum 500 ppm.

Dissolve 1.3 g in 5 mL of *dilute nitric acid R* and dilute to 200 mL with *water R*.

Soluble phosphates
Maximum 0.5 per cent, expressed as PO_4.

Test solution Centrifuge 10.0 g until a clear supernatant is obtained. To 2.00 mL of the supernatant add 20.0 mL of a 10.3 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with *water R*. To 10.0 mL of this solution add 10.0 mL of nitro-molybdovanadic reagent R and dilute to 50.0 mL with *water R*. Allow to stand protected from light for 15 min.

Reference solution Add 10.0 mL of nitro-molybdovanadic reagent R to 10.0 mL of a 143 mg/L solution of *potassium dihydrogen phosphate R* and dilute to 50.0 mL with *water R*. Allow to stand protected from light for 15 min.

Measure the absorbances (*2.2.25*) of the 2 solutions at 400 nm. The absorbance of the test solution is not greater than that of the reference solution.

Sulfates (*2.4.13*)
Maximum 0.2 per cent.

Dilute 25 mL of solution S to 100 mL with *distilled water R*.

Soluble aluminium
Maximum 50 ppm.

To 16.0 g add 50 mL of *water R*. Heat to boiling for 5 min. Cool and centrifuge. Separate the supernatant. Wash the residue with 20 mL of *water R* and centrifuge. Separate the supernatant and add to the first supernatant. To the combined supernatants add 5 mL of *hydrochloric acid R* and 20 mL of *water R*. Introduce all of this solution into a 500 mL conical flask and carry out the complexometric titration of aluminium (*2.5.11*) using *0.01 M sodium edetate*.

Arsenic (*2.4.2, Method A*)
Maximum 1 ppm, determined on 1.0 g.

Acid neutralising capacity
Add 2.0 g to 30 mL of *0.1 M hydrochloric acid* heated to 37 °C and maintain at 37 °C while shaking. Determine the pH after 15 min. The pH (*2.2.3*) of the mixture is 2.0 to 2.5.

Residue on ignition
19.0 per cent to 23.0 per cent.

Heat 0.500 g at 50 °C for 5 hours, then ignite at 500 ± 50 °C until constant mass.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of bile-tolerant gram-negative bacteria (*2.6.13*).

Absence of *Escherichia coli* (*2.6.13*).

ASSAY

Dissolve with heating 0.300 g in 5 mL of *dilute hydrochloric acid R*. Add 45 mL of *water R*, 10.0 mL of *0.1 M sodium edetate* and 30 mL of a mixture of equal volumes of *ammonium acetate solution R* and *dilute acetic acid R*. Heat to boiling and maintain boiling for 3 min. Cool, then add 25 mL of *ethanol (96 per cent) R*. Titrate with *0.1 M zinc sulfate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M zinc sulfate* is equivalent to 12.2 mg of $AlPO_4$.

STORAGE

In an airtight container.

Ph Eur

Aluminium Powder

Al 26.98 *7429-90-5*

Action and use

Topical protective.

Preparation

Compound Aluminium Paste

DEFINITION

Aluminium Powder consists mainly of metallic aluminium in the form of very small flakes, usually with an appreciable proportion of aluminium oxide; it is lubricated with stearic acid to prevent oxidation. It contains not less than 86.0% of Al, calculated with reference to the substance freed from lubricant and volatile matter.

CHARACTERISTICS

A silvery grey powder.

Practically insoluble in *water* and in *ethanol (96%)*.

It dissolves in dilute acids and in aqueous solutions of alkali hydroxides, with the evolution of hydrogen.

IDENTIFICATION

A solution in *2M hydrochloric acid* yields the reaction characteristic of *aluminium salts*, Appendix VI.

TESTS

Surface-covering power

Not less than 4000 cm^2 per g when determined by the following method. Fill with *water* a shallow trough measuring approximately 60 cm × 12 cm × 1.5 cm, fitted with a movable partition so constructed that it is a sliding fit and can be used to divide the trough into two rectangular areas. Place the movable partition near one end and sprinkle 50 mg of the substance being examined on the surface of the liquid confined in the smaller area. Using a glass rod, spread the powder evenly over the liquid surface until an unbroken film covers the entire surface. Move the partition so as to increase the area confined and again spread the powder to cover the increased surface. Continue this process and determine the maximum unbroken surface area obtained. The surface-covering power is the area covered per g of the powder at the breaking point of the film.

Iron

Dissolve 10 mg in 20 mL of *2M hydrochloric acid* and dilute to 100 mL with *water*. 10 mL of the resulting solution complies with the *limit test for iron*, Appendix VII (1.0%).

Lead

Use two solutions prepared in the following manner.

For solution (1) boil 0.40 g with 20 mL of *2M hydrochloric acid* and 10 mL of *water* until effervescence ceases, add 0.5 mL of *nitric acid*, boil for 30 seconds and cool; add 2 g of *ammonium chloride* and 2 g of *ammonium thiocyanate*, extract with three 10 mL quantities of a mixture of equal volumes of *amyl alcohol* and *ether*, discard the extracts and add 2 g of *citric acid*. For solution (2) dissolve 2 g of *citric acid* in 10 mL of *2M hydrochloric acid* and add 4 mL of *lead standard solution (10 ppm Pb)*. Make solutions (1) and (2) alkaline with *5M ammonia* and to each add 1 mL of *potassium cyanide solution PbT*. The solutions should be not more than faintly opalescent. If the colours of the solutions differ, equalise by the addition of about 0.2 mL of a highly diluted solution of burnt sugar or other non-reactive substance. Dilute each solution to 50 mL with *water*, add 0.1 mL of a 10% w/v solution of *sodium sulfide* to each and mix thoroughly.

The colour produced in solution (1) is not more intense than that produced in solution (2), when viewed against a white background (100 ppm).

Other metals

Dissolve 2 g in 40 mL of *2M hydrochloric acid*. Dilute 20 mL of the solution to 100 mL with *water*, make alkaline to *litmus paper* by the addition of *5M ammonia*, boil and filter. Evaporate the filtrate to dryness, add 0.05 mL of *sulfuric acid* and ignite. The residue weighs not more than 2 mg.

Lubricant

To 2 g add 100 mL of hot *water*, cover and add, drop wise, sufficient of a mixture of equal volumes of *hydrochloric acid* and *water* to dissolve the metal almost completely. Heat to complete dissolution, cool, filter through a hardened filter paper and wash the vessel and filter paper thoroughly with *water*; dry both the vessel and paper at room temperature. Extract the paper with three 100-mL quantities of boiling, freshly distilled *acetone*, using the original vessel to contain the solvent and then wash the paper with five 10-mL quantities of freshly distilled *acetone*. Evaporate the combined filtrate and washings to dryness using a rotary evaporator. The residue, after drying at 105° for 30 minutes and allowing to cool, weighs 10 to 60 mg.

When the basin containing the residue is floated in a beaker of water suitably stirred and heated, the residue melts between 40° and 60°. The residue is almost completely soluble, with effervescence, in hot *dilute sodium carbonate solution*.

Volatile matter

When heated to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

ASSAY

Transfer 0.2 g, previously freed from lubricant by successive washing with *acetone* and drying, to a three-necked 500 mL flask fitted with a 150 mL dropping funnel, an inlet tube connected to a cylinder of *carbon dioxide* and an outlet tube dipping into a water trap. Add 60 mL of *water* and disperse the substance being examined; replace the air by *carbon dioxide* and add 100 mL of a solution containing 56 g of *ammonium iron(III) sulfate* and 7.5 mL of *sulfuric acid* in *water*. While maintaining an atmosphere of *carbon dioxide* in the flask, heat to boiling, boil for 5 minutes after the sample has dissolved, cool rapidly to 20° and dilute to 250 mL with *water*. To 50 mL add 15 mL of *orthophosphoric acid* and titrate with *0.02M potassium permanganate VS*. Each mL of *0.02M potassium permanganate VS* is equivalent to 0.8994 mg of Al.

Aluminium Sodium Silicate

(Ph. Eur. monograph 1676)

Ph Eur

DEFINITION

Silicic acid aluminium sodium salt of synthetic origin.

Content

— *aluminium* (Al; M_r 26.98): 2.7 per cent to 7.9 per cent (dried substance);
— *sodium* (Na; M_r 22.99): 3.7 per cent to 6.3 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, fine, light, amorphous powder.

Solubility

Practically insoluble in water and in organic solvents.

IDENTIFICATION

A. Transfer 1.0 g to a 100 mL beaker and add 10 mL of *dilute hydrochloric acid R*. Mix, cover with a watch glass and boil for 15 min. Allow to cool to room temperature, mix and centrifuge the solution. 2 mL of the supernatant gives the reaction of aluminium (*2.3.1*).

B. 2 mL of the supernatant obtained in identification test A gives reaction (a) of sodium (*2.3.1*).

C. 0.2 g gives the reaction of silicates (*2.3.1*).

TESTS

pH (*2.2.3*)

9.5 to 11.5.

Disperse 5.0 g in 100 mL of *carbon dioxide-free water R*.

Arsenic (*2.4.2, Method A*)

Maximum 3 ppm.

Transfer 8.3 g to a 250 mL beaker containing 50 mL of *dilute hydrochloric acid R*. Mix, cover with a watch glass and boil gently, with occasional stirring, for 15 min. Centrifuge, and decant the supernatant through a rapid-flow filter paper into a 250 mL volumetric flask. To the residue in the beaker, add 25 mL of hot *dilute hydrochloric acid R*, stir, centrifuge, and decant the supernatant through the same filter into the volumetric flask. Repeat the extraction with 3 additional quantities, each of 25 mL, of hot *dilute hydrochloric acid R*, filtering each supernatant through this filter into the volumetric flask. Allow the combined filtrates to cool to room temperature and dilute to 250.0 mL with *dilute hydrochloric acid R*. Dilute 10.0 mL of the solution to 25.0 mL with *water R*.

Lead

Maximum 5 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Transfer 5.0 g to a 250 mL beaker containing 50 mL of *dilute hydrochloric acid R*. Mix, cover with a watch glass and boil for 15 min. Allow to cool to room temperature. Centrifuge, and decant the supernatant through a rapid-flow filter paper into a 250 mL beaker. To the insoluble matter add 25 mL of hot *water R*. Stir vigorously, centrifuge, and decant the supernatant through the same filter into the beaker. Repeat the extraction with 2 additional quantities, each of 25 mL, of hot *water R*, decanting each supernatant through the filter into the beaker. Wash the filter with 25 mL of hot *water R*, collecting the filtrate in the beaker. Concentrate the combined filtrates by gently boiling to about 15 mL. Add about 0.05 mL of *heavy metal-free nitric acid R*, heat to boiling and allow to cool to room temperature. Filter the concentrated extracts through a rapid-flow filter paper into a 25 mL volumetric flask. Transfer the remaining contents of the beaker through the filter paper and into the volumetric flask with *water R* and dilute to 25.0 mL with the same solvent.

Reference solutions Into 4 separate 100 mL volumetric flasks, introduce respectively 3.0 mL, 5.0 mL, 10.0 mL and 15.0 mL of *lead standard solution (10 ppm Pb) R*, add 0.20 mL of *heavy metal-free nitric acid R* and dilute to 100.0 mL with *water R*.

Source Lead hollow-cathode lamp.

Wavelength 217.0 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)

Maximum 8.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Loss on ignition

5.0 per cent to 11.0 per cent (dried substance), determined on 1.000 g by ignition in a platinum crucible to constant mass at 1000 ± 25 °C.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

ASSAY

Aluminium

Atomic absorption spectrometry (*2.2.23, Method I*).

Acid mixture Add 50 mL of *nitric acid R* to 500 mL of *water R*. Dissolve in this solution 17 g of *tartaric acid R* and dilute to 1000 mL with *water R*.

Blank solution Dissolve 1.4 g of *anhydrous lithium metaborate R* in 60 mL of the acid mixture and dilute to 200 mL with *water R*.

Test solution In a platinum crucible mix 0.200 g with 1.4 g of *anhydrous lithium metaborate R*. Heat slowly at first and ignite at 1100 ± 25 °C for 15 min. Cool, then place the crucible in a 100 mL beaker containing 60 mL of the acid mixture. Place a polytetrafluoroethylene-coated magnetic stirring bar in the crucible and stir gently with a magnetic stirrer for 16 h. Transfer the contents of the crucible into a 200 mL volumetric flask. Wash the crucible, the magnetic stirring bar and the beaker with *water R* and dilute to 200.0 mL with the same solvent (solution A). To 10.0 mL of this solution, add 1.0 mL of *lanthanum chloride solution R* and dilute to 50.0 mL with *water R*.

Reference solutions Into 5 separate 50 mL volumetric flasks, introduce respectively 1.0 mL, 2.5 mL, 5.0 mL, 7.5 mL and 10.0 mL of *aluminium standard solution (100 ppm Al) R*, add 1 mL of *lanthanum chloride solution R* and 10 mL of the blank solution, and dilute to 50.0 mL with *water R*.

Source Aluminium hollow-cathode lamp.

Wavelength 309.3 nm.

Atomisation device Acetylene-nitrous oxide flame.

Sodium

Atomic emission spectrometry (*2.2.22, Method I*).

Test solution To 2.0 mL of solution A, prepared in the assay of aluminium, add 1 mL of a 12.5 g/L solution of *caesium chloride R* and dilute to 20.0 mL with *water R*.

Reference solutions Into 5 separate 200 mL volumetric flasks, each containing 10 mL of a 12.5 g/L solution of *caesium chloride R*, introduce respectively 1.0 mL, 2.0 mL, 4.0 mL,

6.0 mL and 10.0 mL of *sodium standard solution (200 ppm Na) R* and dilute to 200.0 mL with *water R*.

Wavelength 589.0 nm.

Ph Eur

Aluminium Stearate

(Ph. Eur. monograph 1663)

Ph Eur

DEFINITION

Aluminium salts of a mixture of solid organic acids consisting mainly of variable proportions of aluminium stearate and aluminium palmitate. The organic acids are obtained from sources of vegetable or animal origin.

Content

— *aluminium* (Al; A_r 26.98): 3.0 per cent to 9.0 per cent (dried substance);
— *stearic acid in the fatty acid fraction*: minimum 40.0 per cent;
— *sum of stearic acid and palmitic acid in the fatty acid fraction*: minimum 90.0 per cent.

CHARACTERS

Appearance

White or almost white, very fine, light powder.

Solubility

Practically insoluble in water and in anhydrous ethanol.

IDENTIFICATION

First identification: C, D.

Second identification: A, B, D.

A. Freezing point (*2.2.18*): minimum 53 °C, determined on the residue obtained in the preparation of solution S (see Tests).

B. Acid value (*2.5.1*): 195 to 210.

Dissolve 0.200 g of the residue obtained in the preparation of solution S in 25 mL of the prescribed mixture of solvents.

C. Examine the chromatograms obtained in the assay of stearic acid and palmitic acid.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 2 principal peaks in the chromatogram obtained with the reference solution.

D. 1 mL of solution S gives the reaction of aluminium (*2.3.1*). The addition of 0.5 mL of *dilute hydrochloric acid R* described in the general method is omitted.

TESTS

Solution S

To 5.0 g add 50 mL of *peroxide-free ether R*, 20 mL of *dilute nitric acid R* and 20 mL of *distilled water R* and heat gently under a reflux condenser until dissolution is complete. Allow to cool. In a separating funnel, separate the aqueous layer and shake the ether layer with 2 quantities, each of 4 mL, of *distilled water R*. Combine the aqueous layers, wash with 15 mL of *peroxide-free ether R* and dilute to 50.0 mL with *distilled water R* (solution S). Evaporate the ether layer to dryness and dry the residue at 100-105 °C. Keep the residue for identification tests A and B.

Acidity or alkalinity

To 1.0 g add 20 mL of *carbon dioxide-free water R* and boil for 1 min with continuous shaking. Cool and filter.

To 10 mL of the filtrate add 0.05 mL of *bromothymol blue solution R4*. Not more than 0.05 mL of *0.1 M hydrochloric acid* or *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Chlorides (*2.4.4*)

Maximum 0.1 per cent.

Dilute 0.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 0.5 per cent.

Dilute 0.3 mL of solution S to 15 mL with *distilled water R*.

Cadmium

Maximum 3 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a strong-acid, strong-base, mixed-bed ion-exchange resin before use. Select all reagents to have as low a content of cadmium, lead and nickel as practicable and store all reagent solutions in containers of borosilicate glass. Clean glassware before use by soaking in warm 8 M nitric acid for 30 min and by rinsing with deionised water.

Blank solution Dilute 25 mL of *cadmium- and lead-free nitric acid R* to 100.0 mL with *water R*.

Modifier solution Dissolve 20 g of *ammonium dihydrogen phosphate R* and 1 g of *magnesium nitrate R* in *water R* and dilute to 100 mL with the same solvent. Alternatively, use an appropriate matrix modifier as recommended by the graphite furnace atomic absorption (GFAA) spectrometer manufacturer.

Test solution Place 0.100 g of the substance to be examined in a polytetrafluoroethylene digestion bomb and add 2.5 mL of *cadmium- and lead-free nitric acid R*. Close and seal the bomb according to the manufacturer's operating instructions. *When using a digestion bomb, be thoroughly familiar with the safety and operating instructions. Carefully follow the bomb manufacturer's instructions regarding care and maintenance of these digestion bombs. Do not use metal-jacketed bombs or liners that have been used with hydrochloric acid due to contamination from corrosion of the metal jacket by hydrochloric acid.* Heat the bomb in an oven at 170 °C for 3 h. Cool the bomb slowly in air to room temperature according to the bomb manufacturer's instructions. Place the bomb in a fume cupboard and open carefully as corrosive gases may be expelled. Dissolve the residue in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution Prepare a solution containing 0.00165 µg/mL of *cadmium nitrate tetrahydrate R* in the blank solution (equivalent to 0.006 µg/mL of Cd).

Dilute 1.0 mL of the test solution to 10.0 mL with the blank solution. Prepare mixtures of this solution, the reference solution and the blank solution in the following proportions: (1.0:0:1.0 *V/V/V*), (1.0:0.25:0.75 *V/V/V*), (1.0:0.5:0.5 *V/V/V*), (1.0:0.75:0.25 *V/V/V*). To each mixture add 50 µL of the modifier solution and mix. These solutions contain respectively 0 µg, 0.0015 µg, 0.0030 µg and 0.0045 µg of cadmium per millilitre from the reference solution. Keep the remaining test solution for use in the test for lead and nickel.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Furnace.

Platform Pyrolytically coated with integrated tube.

Operating conditions Use the temperature programme recommended for cadmium by the GFAA manufacturer.

An example of temperature parameters for GFAA analysis of cadmium is shown below.

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	600	10	30
Atomisation	1800	0	5

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a strong-acid, strong-base, mixed-bed ion-exchange resin before use. Select all reagents to have as low a content of cadmium, lead and nickel as practicable and store all reagent solutions in containers of borosilicate glass. Clean glassware before use by soaking in warm 8 M nitric acid for 30 min and by rinsing with deionised water.

Blank solution Use the solution described in the test for cadmium.

Modifier solution Use the solution described in the test for cadmium.

Test solution Use the solution described in the test for cadmium.

Reference solution Prepare a solution of 0.100 µg/mL of Pb by suitable dilutions of *lead standard solution (100 ppm Pb) R* with the blank solution.

Prepare mixtures of the test solution, the reference solution and the blank solution in the following proportions: (1.0:0:1.0 *V/V/V*), (1.0:0.5:0.5 *V/V/V*), (1.0:1.0:0 *V/V/V*). To each mixture add 50 µL of the modifier solution and mix. These solutions contain respectively 0 µg, 0.025 µg and 0.05 µg of lead per millilitre from the reference solution.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm.

Atomisation device Furnace.

Platform Pyrolytically coated with integrated tube.

Operating conditions Use the temperature programme recommended for lead by the GFAA manufacturer.
An example of temperature parameters for GFAA analysis of lead is shown below.

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	450	10	30
Atomisation	2000	0	5

Nickel

Maximum 5 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a strong-acid, strong-base, mixed-bed ion-exchange resin before use. Select all reagents to have as low a content of cadmium, lead and nickel as practicable and store all reagent solutions in containers of borosilicate glass. Clean glassware before use by soaking in warm 8 M nitric acid for 30 min and by rinsing with deionised water.

Blank solution Use the solution described in the test for cadmium.

Modifier solution Dissolve 20 g of *ammonium dihydrogen phosphate R* in *water R* and dilute to 100 mL with the same solvent. Alternatively, use an appropriate matrix modifier as recommended by the GFAA spectrometer manufacturer.

Test solution Use the solution described in the test for cadmium.

Reference solution Prepare a solution of 0.050 µg/mL of Ni by suitable dilutions of a 0.2477 µg/mL solution of *nickel nitrate hexahydrate R* with the blank solution.

Prepare mixtures of the test solution, the reference solution and the blank solution in the following proportions: (1.0:0:1.0 *V/V/V*), (1.0:0.5:0.5 *V/V/V*), (1.0:1.0:0 *V/V/V*). To each mixture add 50 µL of the modifier solution and mix. These solutions contain respectively 0 µg, 0.0125 µg and 0.025 µg of nickel per millilitre from the reference solution.

Source Nickel hollow-cathode lamp.

Wavelength 232.0 nm.

Atomisation device Furnace.

Platform Pyrolytically coated with integrated tube.

Operating conditions Use the temperature programme recommended for nickel by the GFAA manufacturer.
An example of temperature parameters for GFAA analysis of nickel is shown below.

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	1000	20	30
Atomisation	2300	0	5

Loss on drying (*2.2.32*)

Maximum 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

ASSAY

Aluminium

To 0.250 g in a 250 mL conical flask add 20 mL of *methanol R* and, slowly, 2 mL of *sulfuric acid R*. Heat the solution for 30 min under reflux on a water-bath, swirling frequently. Allow to cool. Add 100 mL of *water R* and adjust to about pH 1 by adding approximately 12 mL of *dilute sodium hydroxide solution R*. Add 20.0 mL of *0.1 M sodium edetate* and adjust to between pH 5 and pH 6 by the addition of *sodium acetate R*. Add 70 mg of *xylenol orange triturate R* and titrate immediately and quickly with *0.1 M zinc sulfate* until the colour changes from yellow to pinkish-violet.

1 mL of *0.1 M sodium edetate* is equivalent to 2.698 mg of Al.

Stearic acid and palmitic acid

Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution In a conical flask fitted with a reflux condenser, dissolve 0.100 g of the substance to be examined in 5 mL of *boron trifluoride-methanol solution R*. Boil under a reflux condenser for 10 min. Add 4 mL of *heptane R* through the condenser and boil again under a reflux condenser for 10 min. Allow to cool. Add 20 mL of *saturated sodium chloride solution R*. Shake and allow the layers to separate. Dry the organic layer over 0.1 g of *anhydrous sodium sulfate R*

previously washed with *heptane R*. Dilute 1.0 mL of the solution to 10.0 mL with *heptane R*.

Reference solution Prepare the reference solution in the same manner as the test solution using 50.0 mg of *palmitic acid CRS* and 50.0 mg of *stearic acid CRS* instead of the substance to be examined.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.32 mm;
- *stationary phase*: *macrogol 20 000 R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R*.

Flow rate 2.4 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	70
	2 - 36	70 → 240
	36 - 41	240
Injection port		220
Detector		260

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to methyl stearate: methyl palmitate = about 0.9.

System suitability Reference solution:
- *resolution*: minimum 5.0 between the peaks due to methyl palmitate and methyl stearate;
- *repeatability*: maximum relative standard deviation of 3.0 per cent for the areas of the peaks due to methyl palmitate and methyl stearate after 6 injections; maximum relative standard deviation of 1.0 per cent for the ratio of the areas of the peaks due to methyl palmitate to the areas of the peaks due to methyl stearate after 6 injections.

Ph Eur

Aluminium Sulfate

Aluminium Sulphate

(Ph. Eur. monograph 0165)

$Al_2(SO_4)_3,xH_2O$ 342.1 (anhydrous substance)

Preparation
Aluminium Acetate Ear Drops

Ph Eur

DEFINITION
Content
51.0 per cent to 59.0 per cent of $Al_2(SO_4)_3$.

It contains a variable quantity of water of crystallisation.

CHARACTERS
Appearance
Colourless, lustrous crystals or crystalline masses.

Solubility
Soluble in cold water, freely soluble in hot water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Solution S (see Tests) gives reaction (a) of sulfates (*2.3.1*).

B. Solution S gives the reaction of aluminium (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in *water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension III (*2.2.1*) and is colourless (*2.2.2, Method II*).

pH (*2.2.3*)
2.5 to 4.0.

Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Alkali and alkaline-earth metals
Maximum 0.4 per cent.

To 20 mL of solution S add 100 mL of *water R*, heat and add 0.1 mL of *methyl red solution R*. Add *dilute ammonia R1* until the colour of the indicator changes to yellow. Dilute to 150 mL with *water R*, heat to boiling and filter. Evaporate 75 mL of the filtrate to dryness on a water-bath and ignite. The residue weighs a maximum of 2 mg.

Ammonium (*2.4.1*)
Maximum 500 ppm.

Dilute 0.4 mL of solution S to 14 mL with *water R*.

Iron (*2.4.9*)
Maximum 100 ppm.

Dilute 2 mL of solution S to 10 mL with *water R*.
Use 0.3 mL of *thioglycollic acid R* in this test.

ASSAY
Dissolve 0.500 g in 20 mL of *water R*. Carry out the complexometric titration of aluminium (*2.5.11*).

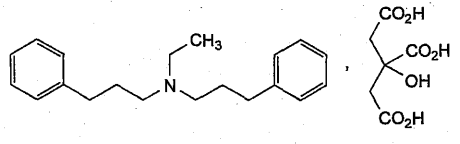
1 mL of *0.1 M sodium edetate* is equivalent to 17.11 mg of $Al_2(SO_4)_3$.

STORAGE
In an airtight container.

Ph Eur

Alverine Citrate

(Ph. Eur. monograph 2156)



$C_{26}H_{35}NO_7$ 473.6 *5560-59-8*

Action and use
Smooth muscle relaxant; antispasmodic.

Preparation
Alverine Capsules

Ph Eur

DEFINITION
N-Ethyl-3-phenyl-*N*-(3-phenylpropyl)propan-1-amine dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water and in methylene chloride, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *alverine citrate CRS.*

TESTS

pH (*2.2.3*)
3.5 to 4.5.

Dissolve 0.250 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Related substances
Gas chromatography (*2.2.28*): use the normalisation procedure. *Use freshly prepared solutions.*

Test solution Dissolve 0.250 g of the substance to be examined in *water R* and dilute to 20 mL with the same solvent. Add 2 mL of *concentrated ammonia R* and shake with 3 quantities, each of 15 mL, of *methylene chloride R.* To the combined lower layers add *anhydrous sodium sulfate R,* shake, filter, and evaporate the filtrate by suitable means at a temperature not exceeding 30 °C. Take up the residue with *methylene chloride R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *alverine impurity D CRS* (impurity D citrate) in 5 mL of *water R,* add 1 mL of *concentrated ammonia R* and shake with 3 quantities, each of 5 mL, of *methylene chloride R.* To the combined lower layers add *anhydrous sodium sulfate R,* shake, filter, and evaporate the filtrate by suitable means at a temperature not exceeding 30 °C. Take up the residue with *methylene chloride R,* add 0.2 mL of the test solution and dilute to 2.0 mL with *methylene chloride R.*

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methylene chloride R.* Dilute 1.0 mL of this solution to 20.0 mL with *methylene chloride R.*

Reference solution (c) Dissolve 20 mg of *alverine impurity C CRS* in *methylene chloride R* and dilute to 20.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *methylene chloride R.*

Column:
— *material:* fused silica;
— *size:* *l* = 25 m, Ø = 0.32 mm;
— *stationary phase:* *poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.45 µm).

Carrier gas *helium for chromatography R.*

Flow rate 2.2 mL/min.

Split ratio 1:11.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 7	120
	7 - 13	120 → 240
	13 - 21	240
	21 - 24	240 → 290
	24 - 39	290
Injection port		290
Detector		290

Detection Flame ionisation.

Injection 1 µL.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to alverine (retention time = about 18 min): impurity C = about 0.5; impurity D = about 0.97.

System suitability Reference solution (a):
— *resolution:* minimum 3.0 between the peaks due to impurity D and alverine.

Limits:
— *impurities C, D:* for each impurity, maximum 0.15 per cent;
— *unspecified impurities:* for each impurity, maximum 0.10 per cent;
— *total:* maximum 0.5 per cent;
— *reporting threshold:* 0.05 per cent (reference solution (b)).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 80 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.375 g in 50 mL of *anhydrous acetic acid R.* Titrate with *0.1 M perchloric acid,* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 47.36 mg of $C_{26}H_{35}NO_7$.

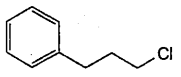
STORAGE

Protected from light.

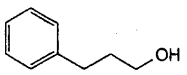
IMPURITIES

Specified impurities *C, D.*

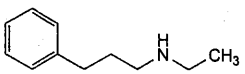
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E.*



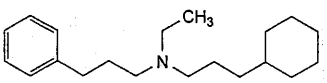
A. 1-chloro-3-phenylpropane,



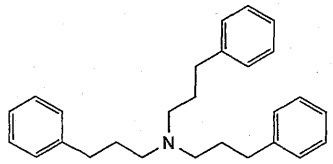
B. 3-phenylpropan-1-ol,



C. *N*-ethyl-3-phenylpropan-1-amine,



D. *N*-(3-cyclohexylpropyl)-*N*-ethyl-3-phenylpropan-1-amine,

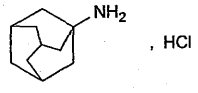


E. 3-phenyl-*N*,*N*-bis(3-phenylpropyl)propan-1-amine.

Ph Eur

Amantadine Hydrochloride

(Ph. Eur. monograph 0463)



$C_{10}H_{18}ClN$ 187.7 *665-66-7*

Action and use
Viral replication inhibitor (influenza A); dopamine receptor agonist; treatment of influenza and Parkinson's disease.

Preparations
Amantadine Capsules
Amantadine Oral Solution

Ph Eur

DEFINITION
Tricyclo[3.3.1.$1^{3,7}$]decan-1-amine hydrochloride.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in ethanol (96 per cent).
It sublimes on heating.

IDENTIFICATION
First identification: A, D.
Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *amantadine hydrochloride CRS.*

B. To 0.1 g add 1 mL of *pyridine R*, mix and add 0.1 mL of *acetic anhydride R*. Heat to boiling for about 10 s. Pour the hot solution into 10 mL of *dilute hydrochloric acid R*, cool to 5 °C and filter. The precipitate, washed with *water R* and dried *in vacuo* at 60 °C for 1 h, melts (*2.2.14*) at 147 °C to 151 °C.

C. Dissolve 0.2 g in 1 mL of *0.1 M hydrochloric acid.* Add 1 mL of a 500 g/L solution of *sodium nitrite R*. A white precipitate is formed.

D. 1 mL of solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Acidity or alkalinity
Dilute 2 mL of solution S to 10 mL with *carbon dioxide-free water R*. Add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid.* The solution is red.

Related substances
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.500 g of *adamantane R* in *methylene chloride R* and dilute to 10.0 mL with the same solvent.

Test solution Weigh 0.5 g of the substance to be examined into a centrifuge tube. Add 9 mL of *methylene chloride R* and 10 mL of a 210 g/L solution of *sodium hydroxide R*. Shake for 10 min. Discard the upper layer. Dry the lower layer over *anhydrous sodium sulfate R*. Filter and collect the filtrate in a volumetric flask. Add 0.1 mL of the internal standard solution and dilute to 10.0 mL with *methylene chloride R*.

Reference solution Weigh 5 mg of *amantadine hydrochloride CRS* into a centrifuge tube. Add 9 mL of *methylene chloride R* and 10 mL of a 210 g/L solution of *sodium hydroxide R*. Shake for 10 min. Discard the upper layer. Dry the lower layer over *anhydrous sodium sulfate R*. Filter and collect the filtrate in a volumetric flask. Add 1.0 mL of the internal standard solution and dilute to 100.0 mL with *methylene chloride R*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *base-deactivated poly(dimethyl) (diphenyl) siloxane R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 4 mL/min.

Split ratio 1:50.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 5	70
	5 - 23	70 → 250
	23 - 40	250
Injection port		220
Detector		300

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to amantadine (retention time = about 14 min): internal standard = about 0.8.

System suitability Reference solution:
— *resolution*: minimum 5.0 between the peaks due to the internal standard and amantadine.

Limits:
— *unspecified impurities*: calculate the ratio (R_1) of the area of the peak due to amantadine to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with the test solution, calculate the ratio of the area of any peak, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than R_1 (0.10 per cent);
— *total*: calculate the ratio (R_2) of 3 times the area of the peak due to amantadine to the area of the peak due to the internal standard from the chromatogram obtained with

the reference solution; from the chromatogram obtained with the test solution, calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than R_2 (0.3 per cent);

— *disregard limit*: calculate the ratio (R_3) of 0.5 times the area of the peak due to amantadine to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with the test solution, calculate the ratio of the area of any peak, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: disregard any peak with a ratio less than R_3 (0.05 per cent).

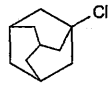
Water (*2.5.12*)
Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

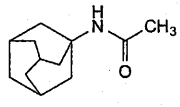
ASSAY
Dissolve 0.150 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 18.77 mg of $C_{10}H_{18}ClN$.

IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*



A. 1-chlorotricyclo[3.3.1.1^{3,7}]decane,

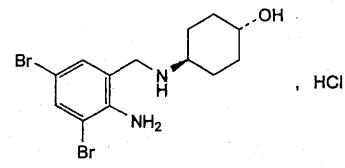


B. *N*-(tricyclo[3.3.1.1^{3,7}]dec-1-yl)acetamide.

Ph Eur

Ambroxol Hydrochloride

(Ph. Eur. monograph 1489)



$C_{13}H_{19}Br_2ClN_2O$ 414.6 *23828-92-4*

Action and use
Mucolytic expectorant.

Ph Eur

DEFINITION
trans-4-[(2-Amino-3,5-dibromobenzyl)amino]cyclohexanol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish, crystalline powder.

Solubility
Sparingly soluble in water, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 20.0 mg in *dilute sulfuric acid R1* and dilute to 100.0 mL with the same acid. Dilute 2.0 mL of the solution to 10.0 mL with *dilute sulfuric acid R1*.

Spectral range 200-350 nm.

Absorption maxima At 245 nm and 310 nm.

Absorbance ratio A_{245}/A_{310} = 3.2 to 3.4.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ambroxol hydrochloride CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 50 mg of *ambroxol hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R, propanol R, ethyl acetate R, hexane R* (1:10:20:70 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

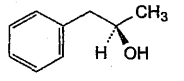
Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 25 mg in 2.5 mL of *water R*, mix with 1.0 mL of *dilute ammonia R1* and allow to stand for 5 min. Filter and

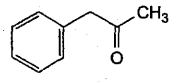
IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, C, D.

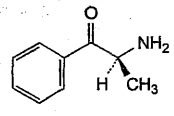


and enantiomer

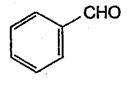
A. (2*RS*)-1-phenylpropan-2-ol,



B. 1-phenylpropan-2-one,



C. (2*S*)-2-amino-1-phenylpropan-1-one (cathinone),

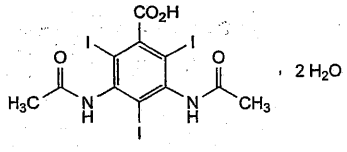


D. benzaldehyde.

Ph Eur

Amidotrizoic Acid Dihydrate

(Ph. Eur. monograph 0873)



$C_{11}H_9I_3N_2O_4,2H_2O$ 650 *50978-11-5*

Action and use

Iodinated contrast medium.

Preparation

Meglumine Amidotrizoate Injection

Ph Eur

DEFINITION

3,5-Bis(acetylamino)-2,4,6-triiodobenzoic acid dihydrate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amidotrizoic acid dihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in a 3 per cent *V/V* solution of *ammonia R* in *methanol R* and dilute to 5 mL with the same solution.

Reference solution Dissolve 25 mg of *amidotrizoic acid dihydrate CRS* in a 3 per cent *V/V* solution of *ammonia R* in *methanol R* and dilute to 5 mL with the same solution.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *anhydrous formic acid R, methyl ethyl ketone R, toluene R* (20:25:60 *V/V/V*).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air until the solvents have evaporated.

Detection In ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Heat 50 mg gently in a small porcelain dish over a naked flame. Violet vapour is evolved.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in *dilute sodium hydroxide solution R* and dilute to 20 mL with the same solution.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Dissolve 0.250 g of *sodium hydroxide R* and 0.860 g of *sodium dihydrogen phosphate R* in 50 mL of *water R* and dilute to 1000 mL with the same solvent.

Test solution Dissolve 40.0 mg of the substance to be examined in 10.0 mL of the solvent mixture with the aid of ultrasound.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *amidotrizoic acid for system suitability CRS* (impurities A, B, C and D) in 1.0 mL of the solvent mixture.

Column:

— *size: l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 3.4 g of *tetrabutylammonium hydrogen sulfate R* in a mixture of 230 mL of *acetonitrile R* and 770 mL of *water R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 236 nm.

Injection 20 µL.

Run time 4 times the retention time of amidotrizoic acid.

Identification of impurities Use the chromatogram supplied with *amidotrizoic acid for system suitability CRS* and the

chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to amidotrizoic acid (retention time = about 5 min): impurity B = about 0.8; impurity C = about 0.9; impurity A = about 1.4; impurity D = about 1.8.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to impurities B and C in the chromatogram obtained with reference solution (c);
- *signal-to-noise ratio*: minimum 25 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
- *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *impurities A, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.01 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
- *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent), except for the peaks due to impurities A and D.

Halides expressed as chlorides (*2.4.4*)
Maximum 150 ppm.

Dissolve 0.55 g in a mixture of 4 mL of *dilute sodium hydroxide solution R* and 15 mL of *water R*. Add 6 mL of *dilute nitric acid R* and filter.

Free aromatic amines
Maintain the solutions and reagents in iced water, protected from bright light To 0.50 g in a 50 mL volumetric flask add 15 mL of *water R*. Shake and add 1 mL of *dilute sodium hydroxide solution R*. Cool in iced water, add 5 mL of a freshly prepared 5 g/L solution of *sodium nitrite R* and 12 mL of *dilute hydrochloric acid R*. Shake gently and allow to stand for exactly 2 min after adding the hydrochloric acid. Add 10 mL of a 20 g/L solution of *ammonium sulfamate R*. Allow to stand for 5 min, shaking frequently, and add 0.15 mL of a 100 g/L solution of *α-naphthol R* in *ethanol (96 per cent) R*. Shake and allow to stand for 5 min. Add 3.5 mL of *buffer solution pH 10.9 R*, mix and dilute to 50.0 mL with *water R*. The absorbance (*2.2.25*), measured within 20 min at 485 nm using as the compensation liquid a solution prepared at the same time and in the same manner but omitting the substance to be examined, is not greater than 0.30.

Loss on drying (*2.2.32*)
4.5 per cent to 7.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

To 0.150 g in a 250 mL round-bottomed flask add 5 mL of *strong sodium hydroxide solution R*, 20 mL of *water R*, 1 g of *zinc powder R* and a few glass beads. Boil under a reflux condenser for 30 min. Allow to cool and rinse the condenser with 20 mL of *water R*, adding the rinsings to the flask. Filter through a sintered-glass filter (*2.1.2*) and wash the filter with several quantities of *water R*. Collect the filtrate and washings. Add 40 mL of *dilute sulfuric acid R* and titrate immediately with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 20.47 mg of $C_{11}H_9I_3N_2O_4$.

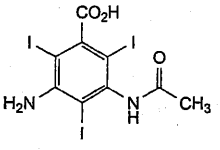
STORAGE

Protected from light.

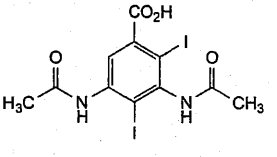
IMPURITIES

Specified impurities *A, B, D.*

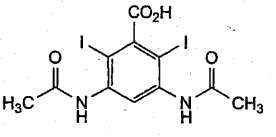
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E.*



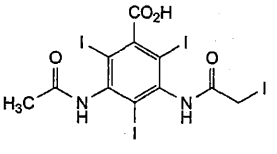
A. 3-(acetylamino)-5-amino-2,4,6-triiodobenzoic acid,



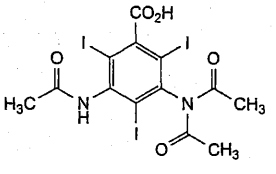
B. 3,5-bis(acetylamino)-2,4-diiodobenzoic acid,



C. 3,5-bis(acetylamino)-2,6-diiodobenzoic acid,



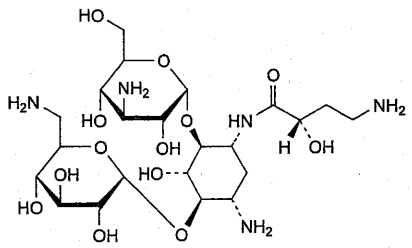
D. 3-(acetylamino)-5-[(iodoacetyl)amino]-2,4,6-triiodobenzoic acid,



E. 3-(acetylamino)-5-(diacetylamino)-2,4,6-triiodobenzoic acid.

Amikacin

(Ph. Eur. monograph 1289)



$C_{22}H_{43}N_5O_{13}$ 585.6 *37517-28-5*

Action and use
Aminoglycoside antibacterial.

Ph Eur

DEFINITION

6-*O*-(3-Amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine.

Antimicrobial substance obtained from kanamycin A.

Semi-synthetic product derived from a fermentation product.

Content
96.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Sparingly soluble in water, slightly soluble in methanol, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amikacin CRS*.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of *amikacin CRS* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *kanamycin monosulfate CRS* in 1 mL of the test solution and dilute to 10 mL with *water R*.

Plate *TLC silica gel plate R*.

Mobile phase *methylene chloride R, ammonia R, methanol R* (25:30:40 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

pH (*2.2.3*)
9.5 to 11.5.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 97 to + 105 (anhydrous substance).

Dissolve 0.50 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *amikacin for system suitability CRS* (containing impurities A, B, F and H) in mobile phase A and dilute to 10 mL with mobile phase A.

Reference solution (d) Dissolve 5.0 mg of *amikacin impurity I CRS* in mobile phase A and dilute to 20.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: a mixture prepared with *carbon dioxide-free water R*, containing 1.8 g/L of *sodium octanesulfonate R*, 20 g/L of *anhydrous sodium sulfate R1*, 1.4 per cent *V/V* of *tetrahydrofuran R*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas;
— *mobile phase B*: a mixture prepared with *carbon dioxide-free water R*, containing 1.8 g/L of *sodium octanesulfonate R*, 28 g/L of *anhydrous sodium sulfate R1*, 1.4 per cent *V/V* of *tetrahydrofuran R*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 38.0	100 → 30	0 → 70
38.0 - 38.1	30 → 0	70 → 100
38.1 - 68	0	100

Flow rate 1.0 mL/min.

Post-column solution Mixture of 1 volume of *carbonate-free sodium hydroxide solution R* and 24 volumes of previously degassed *carbon dioxide-free water R*, which is added in a pulseless manner to the column effluent using a 375 µL polymeric mixing coil.

Flow rate of post-column solution 0.3 mL/min.

Detection Pulsed amperometric detector or equivalent with a gold indicator electrode, a silver-silver chloride reference electrode, and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V

oxidation and − 0.15 V reduction potentials, with pulse durations according to the instrument used.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *amikacin for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, F and H; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity I.

Relative retention With reference to amikacin (retention time = about 28 min): impurity I = about 0.13; impurity F = about 0.92; impurity B = about 0.95; impurity A = about 1.62; impurity H = about 1.95.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to amikacin; if necessary, adjust the volume of tetrahydrofuran in the mobile phase.

Calculation of percentage contents:
— for impurity I, use the concentration of impurity I in reference solution (d);
— for impurities other than I, use the concentration of amikacin in reference solution (a).

Limits:
— *impurities A, B, F, H, I*: for each impurity, maximum 0.5 per cent;
— *any other impurity*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.1 per cent.

Water (*2.5.12*)
Maximum 8.5 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution Dissolve 50.0 mg of *amikacin CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase A mixture prepared with *carbon dioxide-free water R*, containing 1.8 g/L of *sodium octanesulfonate R*, 20 g/L of *anhydrous sodium sulfate R1*, 5.8 per cent *V/V* of *acetonitrile R1*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

Run time 1.3 times the retention time of amikacin.

Retention time Amikacin = about 30 min.

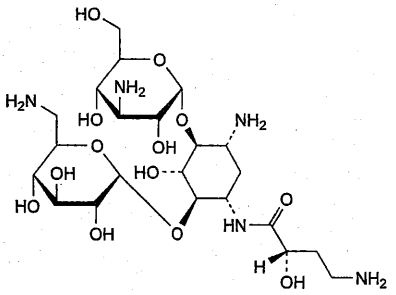
System suitability Reference solution:
— *symmetry factor*: maximum 1.5 for the peak due to amikacin; if necessary, adjust the amount of *acetonitrile R1* in the mobile phase; peak splitting may be observed when the retention time becomes too short;
— *repeatability*: maximum relative standard deviation of 1.5 per cent after 6 injections.

Calculate the percentage content of $C_{22}H_{43}N_5O_{13}$ taking into account the assigned content of *amikacin CRS*.

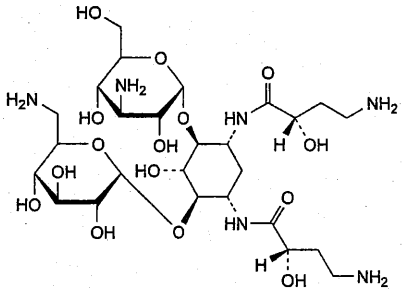
IMPURITIES

Specified impurities *A, B, F, H, I.*

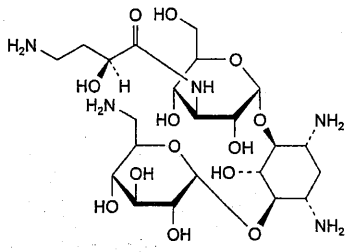
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, G.*



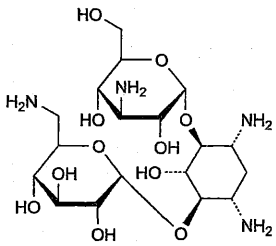
A. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-L-streptamine,



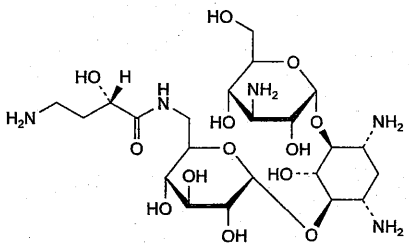
B. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1,3-*N*-bis[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-L-streptamine,



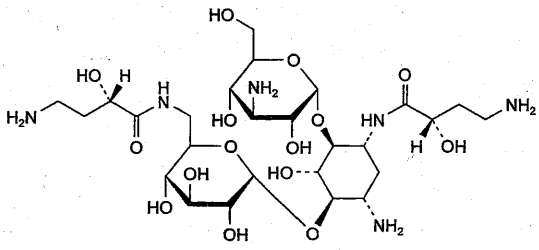
C. 4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-6-*O*-[3-[[(2*S*)-4-amino-2-hydroxybutanoyl]amino]-3-deoxy-α-D-glucopyranosyl]-2-deoxy-D-streptamine,



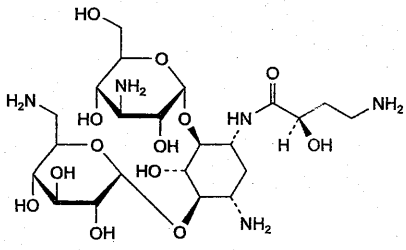
D. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine (kanamycin),



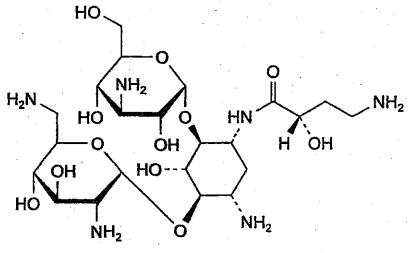
E. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-[6-[[(2*S*)-4-amino-2-hydroxybutanoyl]amino]-6-deoxy-α-D-glucopyranosyl]-2-deoxy-L-streptamine,



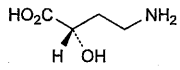
F. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-[6-[(2*S*)-4-amino-2-hydroxybutanoyl]amino-6-deoxy-α-D-glucopyranosyl]-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine,



G. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*R*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine,



H. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine,



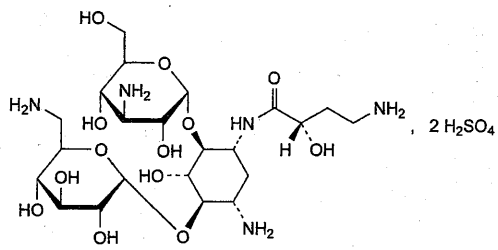
I. (2*S*)-4-amino-2-hydroxybutanoic acid.

Ph Eur

Amikacin Sulfate

Amikacin Sulphate

(Ph. Eur. monograph 1290)



$C_{22}H_{47}N_5O_{21}S_2$ 782 *39831-55-5*

Action and use
Aminoglycoside antibacterial.

Preparation
Amikacin Injection

Ph Eur

DEFINITION

6-*O*-(3-Amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine sulfate.

Antimicrobial substance obtained from kanamycin A.

Semi-synthetic product derived from a fermentation product.

Content
96.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amikacin sulfate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of *amikacin sulfate CRS* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *kanamycin monosulfate CRS* in 1 mL of the test solution and dilute to 10 mL with *water R.*

Plate *TLC silica gel plate R.*

Mobile phase *methylene chloride R, ammonia R, methanol R* (25:30:40 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

pH (*2.2.3*)
2.0 to 4.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 76 to + 84 (dried substance).

Dissolve 0.50 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 33 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *amikacin for system suitability CRS* (containing impurities A, B, F and H) in mobile phase A and dilute to 10 mL with mobile phase A.

Reference solution (d) Dissolve 6.6 mg of *amikacin impurity I CRS* in mobile phase A and dilute to 20.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: a mixture prepared with *carbon dioxide-free water R*, containing 1.8 g/L of *sodium octanesulfonate R*, 20 g/L of *anhydrous sodium sulfate R1*, 1.4 per cent *V/V* of *tetrahydrofuran R*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas;
— *mobile phase B*: a mixture prepared with *carbon dioxide-free water R*, containing 1.8 g/L of *sodium octanesulfonate R*, 28 g/L of *anhydrous sodium sulfate R1*, 1.4 per cent *V/V* of *tetrahydrofuran R*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 38.0	100 → 30	0 → 70
38.0 - 38.1	30 → 0	70 → 100
38.1 - 68	0	100

Flow rate 1.0 mL/min.

Post-column solution Mixture of 1 volume of *carbonate-free sodium hydroxide solution R* and 24 volumes of previously degassed *carbon dioxide-free water R*, which is added in a pulseless manner to the column effluent using a 375 µL polymeric mixing coil.

Flow rate of post-column solution 0.3 mL/min.

Detection Pulsed amperometric detector or equivalent with a gold indicator electrode, a silver-silver chloride reference electrode, and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V oxidation and − 0.15 V reduction potentials, with pulse durations according to the instrument used.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *amikacin for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, F and H; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity I.

Relative retention With reference to amikacin (retention time = about 28 min): impurity I = about 0.13; impurity F = about 0.92; impurity B = about 0.95; impurity A = about 1.62; impurity H = about 1.95.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to amikacin; if necessary, adjust the volume of tetrahydrofuran in the mobile phase.

Calculation of percentage contents:
— for impurity I, use the concentration of impurity I in reference solution (d);
— for impurities other than I, use the concentration of amikacin sulfate in reference solution (a).

Limits:
— *impurities A, B, F, H, I*: for each impurity, maximum 0.5 per cent;
— *any other impurity*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.1 per cent.

Sulfate
23.3 per cent to 25.8 per cent (dried substance).

Dissolve 0.250 g in 100 mL of *water R* and adjust the solution to pH 11 using *concentrated ammonia R*. Add 10.0 mL of *0.1 M barium chloride* and about 0.5 mg of *phthalein purple R*. Titrate with *0.1 M sodium edetate* adding 50 mL of *ethanol (96 per cent) R* when the colour of the solution begins to change and continue the titration until the violet-blue colour disappears.

1 mL of *0.1 M barium chloride* is equivalent to 9.606 mg of sulfate (SO_4).

Loss on drying (*2.2.32*)
Maximum 13.0 per cent, determined on 0.500 g by drying in an oven at 105 °C at a pressure not exceeding 0.7 kPa for 3 h.

Pyrogens (*2.6.8*)
If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of pyrogens, it complies with the test for pyrogens. Inject per kilogram of the rabbit's mass 5 mL of a solution containing 25 mg of the substance to be examined in *water for injections R*.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution Dissolve 37.4 mg of *amikacin CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mixture containing 1.8 g/L of *sodium octanesulfonate R*, 20 g/L of *anhydrous sodium sulfate R1*, 5.8 per cent *V/V* of *acetonitrile R1*, and 5 per cent *V/V* of *0.2 M potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *dilute phosphoric acid R*; degas.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

Run time 1.3 times the retention time of amikacin.

Retention time Amikacin = about 30 min.

System suitability Reference solution:
— *symmetry factor*: maximum 1.5 for the peak due to amikacin; if necessary, adjust the amount of *acetonitrile R1* in the mobile phase; peak splitting may be observed when the retention time becomes too short;
— *repeatability*: maximum relative standard deviation of 1.5 per cent after 6 injections.

Calculate the percentage content of $C_{22}H_{47}N_5O_{21}S_2$ taking into account the assigned content of *amikacin CRS* and a correction factor of 1.335.

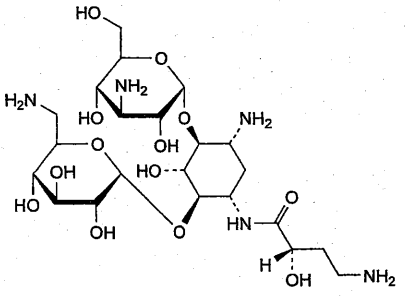
STORAGE

In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

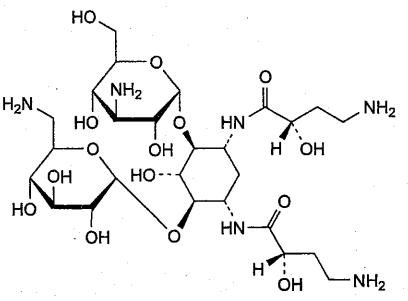
IMPURITIES

Specified impurities *A, B, F, H, I.*

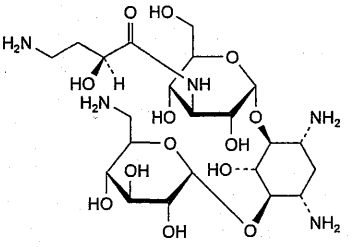
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, G.*



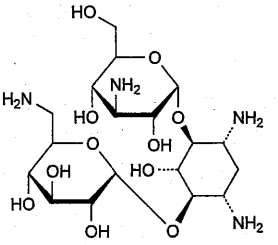
A. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-L-streptamine,



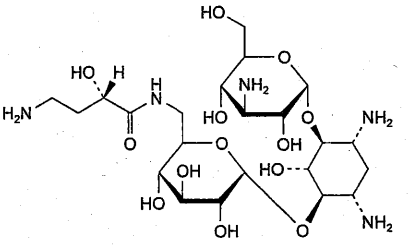
B. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1,3-*N*-bis[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-L-streptamine,



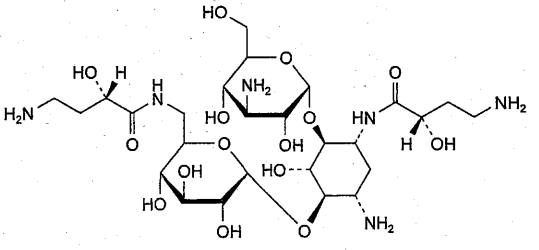
C. 4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-6-*O*-[3-[[(2*S*)-4-amino-2-hydroxybutanoyl]amino]-3-deoxy-α-D-glucopyranosyl]-2-deoxy-D-streptamine,



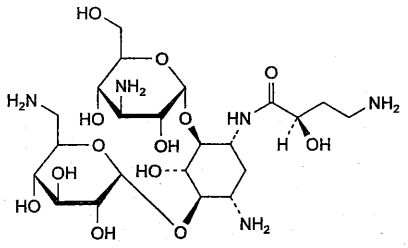
D. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine (kanamycin),



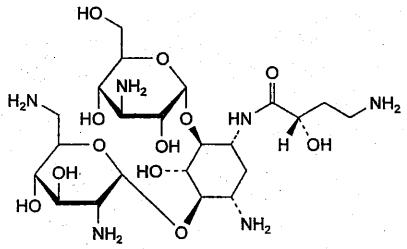
E. 4-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-6-*O*-[6-[[(2*S*)-4-amino-2-hydroxybutanoyl]amino]-6-deoxy-α-D-glucopyranosyl]-2-deoxy-L-streptamine,



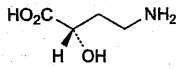
F. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-[6-[(2*S*)-4-amino-2-hydroxybutanoyl]amino-6-deoxy-α-D-glucopyranosyl]-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine,



G. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-4-*O*-(6-amino-6-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*R*)-4-amino-2-hydroxybutanoyl]-2-deoxy-D-streptamine,



H. 6-*O*-(3-amino-3-deoxy-α-D-glucopyranosyl)-1-*N*-[(2*S*)-4-amino-2-hydroxybutanoyl]-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine,

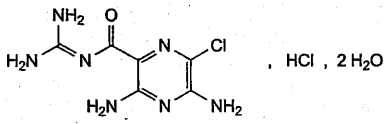


I. (2*S*)-4-amino-2-hydroxybutanoic acid.

Ph Eur

Amiloride Hydrochloride Dihydrate

(Ph. Eur. monograph 0651)



$C_6H_9Cl_2N_7O,2H_2O$ 302.1 *17440-83-4*

Action and use
Sodium channel blocker; potassium-sparing diuretic.

Preparations
Amiloride Tablets
Co-amilofruse Tablets
Co-amilozide Oral Solution
Co-amilozide Tablets

Ph Eur

DEFINITION
3,5-Diamino-6-chloro-*N*-(diaminomethylidene)pyrazine-2-carboxamide hydrochloride dihydrate.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
Pale yellow or greenish-yellow powder.

Solubility
Slightly soluble in water and in anhydrous ethanol, practically insoluble in heptane.

IDENTIFICATION
First identification: A, C, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amiloride hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 5 mg of *amiloride hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R, propanol R* (30:70 *V/V*).

Application 5 µL; the volume may be adapted according to the type of plate used.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 25 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent. 2 mL of the solution gives reaction (a) of chlorides (*2.3.1*); acidify with 5 mL of *dilute acetic acid R*, instead of *dilute nitric acid R*.

D. Water (see Tests).

TESTS
Free acid
Dissolve 1.0 g in a mixture of 50 mL of *methanol R* and 50 mL of *water R* and titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Not more than 0.3 mL of *0.1 M sodium hydroxide* is required to reach the end-point.

Related substances
Liquid chromatography (*2.2.29*).

Solution A Dissolve 2.76 g of *sodium dihydrogen phosphate monohydrate R* in 850 mL of *water for chromatography R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1.0 L with *water for chromatography R*.

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *methanol R1* and dilute to 10.0 mL with solution A.

Reference solution (a) Dissolve 2 mg of *amiloride impurity A CRS* in 0.5 mL of *methanol R1*, add 0.5 mL of the test solution and dilute to 10.0 mL with solution A.

Reference solution (b) Dissolve 4 mg of *amiloride for peak identification CRS* (containing impurity C) in 0.5 mL of *methanol R1* and dilute to 2.0 mL with solution A.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated end-capped octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 0.8 g of *sodium hexanesulfonate monohydrate R* in a mixture of 80 mL of *acetonitrile R1* and 920 mL of solution A.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Run time Twice the retention time of amiloride.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *amiloride for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to amiloride (retention time = about 10 min): impurity C = about 0.5; impurity A = about 0.8.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurity A and amiloride.

Calculation of percentage contents:
— for each impurity, use the concentration of amiloride hydrochloride dihydrate in reference solution (c).

Limits:
— *impurity C*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
11.0 per cent to 13.0 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

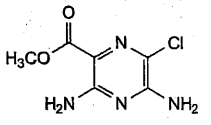
ASSAY
Dissolve 0.200 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 26.61 mg of $C_6H_9Cl_2N_7O$.

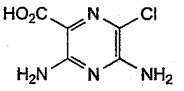
STORAGE
Protected from light.

IMPURITIES
Specified impurities *C*.

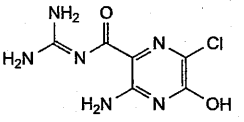
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*



A. methyl 3,5-diamino-6-chloropyrazine-2-carboxylate,



B. 3,5-diamino-6-chloropyrazine-2-carboxylic acid,

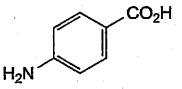


C. 3-amino-6-chloro-*N*-(diaminomethylidene)-5-hydroxypyrazine-2-carboxamide.

Ph Eur

Aminobenzoic Acid

(4-Aminobenzoic Acid, Ph. Eur. monograph 1687)



$C_7H_7NO_2$ 137.1 *150-13-0*

Action and use
Skin protective.

Ph Eur

DEFINITION
4-Aminobenzoic acid.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in alcohol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 186 °C to 189 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *4-aminobenzoic acid CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 20 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *4-aminobenzoic acid CRS* in *methanol R* and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *4-nitrobenzoic acid R* in 10 mL of reference solution (a).

Plate Suitable silica gel with a fluorescent indicator having an optimal intensity at 254 nm as the coating substance.

Mobile phase *glacial acetic acid R, hexane R, methylene chloride R* (5:20:75 *V/V/V*).

Application 1 µL.

Development Over a path of 10 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_5 (*2.2.2, Method II*).

Dissolve 1.0 g in *alcohol R* and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution Dissolve 25.0 mg of *4-nitrobenzoic acid R* and 25.0 mg of *benzocaine R* in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.12 m, Ø = 4.0 mm,
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 20 volumes of a mixture of 70 volumes of *acetonitrile R* and 80 volumes of *methanol R*, and 80 volumes of a solution containing 1.5 g/L of *potassium dihydrogen phosphate R* and 2.5 g/L of *sodium octanesulfonate R* adjusted to pH 2.2 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 µL.

Run time 11 times the retention time of 4-aminobenzoic acid.

Relative retention With reference to 4-aminobenzoic acid (retention time = about 3 min): impurity A = about 4; impurity B = about 9.

Limits:
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.2 per cent),
- *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.2 per cent),
- *any other impurity*: not more than 0.5 times the area of the peak due to impurity A in the chromatogram obtained with the reference solution (0.1 per cent),
- *total*: not more than 2.5 times the area of the peak due to impurity A in the chromatogram obtained with the reference solution (0.5 per cent),
- *disregard limit*: 0.1 times the area of the peak due to impurity A in the chromatogram obtained with the reference solution (0.02 per cent).

Impurity C and impurity D

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 20.0 mg of *lauric acid R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 1.000 g of the substance to be examined in 10.0 mL of an 84 g/L solution of *sodium hydroxide R* and extract with 2 quantities, each of 10 mL, of *methylene chloride R*. Combine and wash with 5 mL of *water R*; filter through *anhydrous sodium sulfate R*. Wash the filter with *methylene chloride R*. Evaporate in a water-bath at 50-60 °C to obtain a volume of about 1-5 mL. Add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *methylene chloride R*.

Reference solution (a) Dissolve 20.0 mg of *aniline R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 20.0 mg of *p-toluidine R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dilute 0.50 mL of reference solution (a), 0.50 mL of reference solution (b) and 10.0 mL of the internal standard solution to 100.0 mL with *methylene chloride R*.

Column:
- *material*: fused silica,
- *size*: *l* = 30 m, Ø = 0.32 mm,
- *stationary phase*: *poly[methyl(95)phenyl(5)]siloxane R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1.0 mL/min.

Split ratio 1:10.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 4	130
	4 - 6.5	130 → 180
	6.5 - 11.5	180
Injection port		280
Detector		300

Detection Flame ionisation.

Injection 2 µL; inject the test solution and reference solution (c).

Retention time Internal standard = about 9.5 min.

Limits:
- *impurity C*: calculate the ratio (*R*) of the area of the peak due to impurity C to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (c); calculate the ratio of the area of the peak due to impurity C to the area of the peak due to the internal standard from the chromatogram obtained with the test solution: this ratio is not greater than *R* (10 ppm),
- *impurity D*: calculate the ratio (*R*) of the area of the peak due to impurity D to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (c); calculate the ratio of the area of the peak due to impurity D to the area of the peak due to the internal standard from the chromatogram obtained with the test solution: this ratio is not greater than *R* (10 ppm).

Iron (*2.4.9*)

Maximum 40 ppm.

Dissolve 0.250 g in 3 mL of *alcohol R* and dilute to 10.0 mL with *water R*.

Water (*2.5.12*)

Maximum 0.2 per cent, determined on 1.00 g.

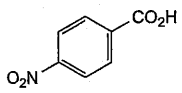
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.100 g with heating in 50 mL of *carbon dioxide-free water R*. Titrate with *0.1 M sodium hydroxide* determining the end-point potentiometrically (*2.2.20*).

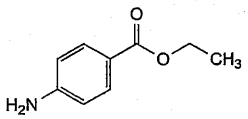
1 mL of *0.1 M sodium hydroxide* is equivalent to 13.71 mg of $C_7H_7NO_2$.

STORAGE
Protected from light.

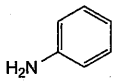
IMPURITIES



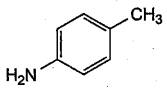
A. 4-nitrobenzoic acid,



B. ethyl 4-aminobenzoate (benzocaine),



C. aniline,

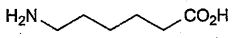


D. 4-methylaniline (*p*-toluidine).

Ph Eur

Aminocaproic Acid

(Ph. Eur. monograph 0874)



$C_6H_{13}NO_2$ 131.2 *60-32-2*

Action and use
Antifibrinolytic.

Ph Eur

DEFINITION
Aminocaproic acid contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of 6-aminohexanoic acid, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder or colourless crystals, freely soluble in water, slightly soluble in alcohol.

It melts at about 205 °C with decomposition.

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *aminocaproic acid CRS*. Examine the substances prepared as discs.

B. Examine the chromatograms obtained in the test for ninhydrin-positive substances. The principal spot in the chromatogram obtained with the test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.5 g in 4 mL of a mixture of equal volumes of *dilute hydrochloric acid R* and *water R*. Evaporate to dryness by heating on a water-bath. Dry the residue in a desiccator. Dissolve the residue in about 2 mL of boiling *ethanol R*. Allow to cool and maintain at 4 °C to 8 °C for 3 h. Filter under reduced pressure. The residue washed with about 10 mL of *acetone R* and dried at 60 °C for 30 min, melts (*2.2.14*) at 131 °C to 133 °C.

D. Dissolve about 5 mg in 0.5 mL of *distilled water R*. Add 3 mL of *dimethylformamide R* and 2 mL of *ascorbic acid solution R*. Heat on a water-bath. An orange colour develops.

TESTS

Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is colourless (*2.2.2, Method II*) and remains clear (*2.2.1*) on standing for 24 h.

pH (*2.2.3*)
The pH of solution S is 7.5 to 8.0.

Absorbance (*2.2.25*)
A. The absorbance of solution S at 287 nm is not more than 0.10 and at 450 nm is not more than 0.03.

B. Place 2.0 g in an even layer in a shallow dish 9 cm in diameter, cover and allow to stand at 98 °C to 102 °C for 72 h. Dissolve in *water R* and dilute to 10.0 mL with the same solvent. The absorbance of the solution at 287 nm is not more than 0.15 and at 450 nm is not more than 0.03.

Ninhydrin-positive substances
Examine by thin-layer chromatography (*2.2.27*), using a suitable silica gel as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with *water R*.

Reference solution (a) Dissolve 10 mg of *aminocaproic acid CRS* in *water R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (b) to 20 mL with *water R*.

Reference solution (c) Dissolve 10 mg of *aminocaproic acid CRS* and 10 mg of *leucine CRS* in *water R* and dilute to 25 mL with the same solvent.

Apply separately to the plate 5 µL of each solution. Allow the plate to dry in air. Develop over a path of 15 cm using a mixture of 20 volumes of *glacial acetic acid R*, 20 volumes of *water R* and 60 volumes of *butanol R*. Dry the plate in a current of warm air. Spray with *ninhydrin solution R* and heat at 100 °C to 105 °C for 15 min. Any spot in the chromatogram obtained with the test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram

obtained with reference solution (c) shows two clearly separated principal spots.

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

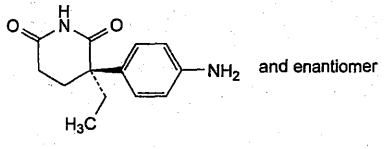
ASSAY
Dissolve 0.100 g in 20 mL of *anhydrous acetic acid R*. Using 0.1 mL of *crystal violet solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from bluish-violet to bluish-green.

1 mL of *0.1 M perchloric acid* is equivalent to 13.12 mg of $C_6H_{13}NO_2$.

Ph Eur

Aminoglutethimide

(Ph. Eur. monograph 1291)



and enantiomer

$C_{13}H_{16}N_2O_2$ 232.3 *125-84-8*

Action and use
Inhibitor of adrenal corticosteroid synthesis; used in chemical adrenalectomy.

Ph Eur

DEFINITION
(3*RS*)-3-(4-Aminophenyl)-3-ethylpiperidine-2,6-dione.

Content
98.0 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in methanol.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 150 °C to 154 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *aminoglutethimide CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *acetone R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of *aminoglutethimide CRS* in *acetone R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of *aminoglutethimide CRS* and 25 mg of *glutethimide CRS* in *acetone R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, methanol R, ethyl acetate R* (0.5:15:85 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separed spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Solution S
Dissolve 1.0 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, acetate buffer solution pH 5.0 R* (50:50 *V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5.0 mg of *aminoglutethimide impurity A CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of the test solution to 10.0 mL with reference solution (a).

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (4 µm);
— *temperature*: 40 °C.

Mobile phase Mix 27 volumes of *methanol R* and 73 volumes of *acetate buffer solution pH 5.0 R*.

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Run time 4 times the retention time of aminoglutethimide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to aminoglutethimide (retention time = about 9 min): impurity A = about 1.3.

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to aminoglutethimide and impurity A.

Limits:
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *sum of impurities other than A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *total*: maximum 2.0 per cent for the sum of the contents of all impurities;
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Impurity D

Liquid chromatography (*2.2.29*). *Carry out the test protected from light. Use shaking, not sonication or heat, to dissolve the reference substance and the substance to be examined.*

Test solution Dissolve 0.100 g of the substance to be examined in *dimethyl sulfoxide R* and dilute to 100.0 mL with the same solvent.

Reference solution Dissolve 3.0 mg of *aminoglutethimide impurity D CRS* in *dimethyl sulfoxide R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *dimethyl sulfoxide R*.

Column:
— *size*: *l* = 0.12 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 0.285 g of *sodium edetate R* in *water R*, add 7.5 mL of *dilute acetic acid R* and 50 mL of *0.1 M potassium hydroxide* and dilute to 1000 mL with *water R*; adjust to pH 5.0 with *glacial acetic acid R*; mix 350 mL of this solution with 650 mL of *methanol R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 328 nm.

Injection 10 µL.

System suitability Test solution:
— *number of theoretical plates*: minimum 3300, calculated for the principal peak;
— *mass distribution ratio*: 2.0 to 5.0 for the principal peak;
— *symmetry factor*: maximum 1.2 for the principal peak.

Limit:
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (300 ppm).

Sulfates (*2.4.13*)

Maximum 500 ppm.

Dilute 6 mL of solution S to 15 mL with *distilled water R*.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

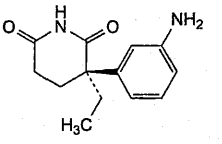
Dissolve 0.180 g in 50 mL of *anhydrous acetic acid R* and titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 23.23 mg of $C_{13}H_{16}N_2O_2$.

IMPURITIES

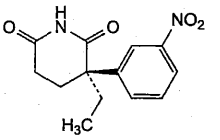
Specified impurities *A, D.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C.*



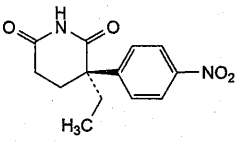
and enantiomer

A. (3*RS*)-3-(3-aminophenyl)-3-ethylpiperidine-2,6-dione (3-aminoglutethimide),



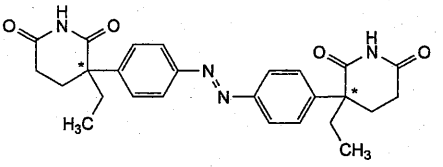
and enantiomer

B. (3*RS*)-3-ethyl-3-(3-nitrophenyl)piperidine-2,6-dione,



and enantiomer

C. (3*RS*)-3-ethyl-3-(4-nitrophenyl)piperidine-2,6-dione,

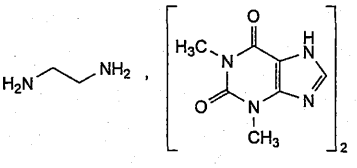


D. 3,3′-[diazenediylbis(4,1-phenylene)]bis(3-ethylpiperidine-2,6-dione) (azoglutethimide).

Ph Eur

Aminophylline

(Theophylline-Ethylenediamine, Ph. Eur. monograph 0300)



$C_{16}H_{24}N_{10}O_4$ 420.4 *317-34-0*

Action and use

Non-selective phosphodiesterase inhibitor; treatment of reversible airways obstruction.

Preparations

Aminophylline Injection

Aminophylline Tablets

Aminophylline Prolonged-release Tablets

Ph Eur

DEFINITION

Content

— *theophylline* ($C_7H_8N_4O_2$; M_r 180.2): 84.0 per cent to 87.4 per cent (anhydrous substance);
— *ethylenediamine* ($C_2H_8N_2$; M_r 60.1): 13.5 per cent to 15.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly yellowish powder, sometimes granular, hygroscopic.

Solubility

Freely soluble in water (the solution becomes cloudy through absorption of carbon dioxide), practically insoluble in anhydrous ethanol.

IDENTIFICATION

First identification: B, C, E.

Second identification: A, C, D, E, F.

Dissolve 1.0 g in 10 mL of *water R* and add 2 mL of *dilute hydrochloric acid R* dropwise with shaking. Filter. Use the precipitate for identification tests A, B, D and F and the filtrate for identification test C.

A. Melting point (*2.2.14*): 270 °C to 274 °C, determined after washing the precipitate with *water R* and drying at 105 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Precipitate, washed with *water R* and dried at 105 °C.

Comparison *theophylline CRS.*

C. To the filtrate add 0.2 mL of *benzoyl chloride R*, make alkaline with *dilute sodium hydroxide solution R* and shake vigorously. Filter the precipitate, wash with 10 mL of *water R*, dissolve in 5 mL of hot *ethanol (96 per cent) R* and add 5 mL of *water R*. A precipitate is formed, which, when washed and dried at 105 °C, melts (*2.2.14*) at 248 °C to 252 °C.

D. Heat about 10 mg of the precipitate with 1.0 mL of a 360 g/L solution of *potassium hydroxide R* in a water-bath at 90 °C for 3 min, then add 1.0 mL of *diazotised sulfanilic acid solution R*. A red colour slowly develops. Carry out a blank test.

E. Water (see Tests).

F. The precipitate gives the reaction of xanthines (*2.3.1*).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).

Dissolve 0.5 g with gentle warming in 10 mL of *carbon dioxide-free water R.*

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 47 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *theobromine R* (impurity G) in the mobile phase, add 5 mL of the test solution and dilute to 100 mL with the mobile phase. Dilute 5 mL of this solution to 50 mL with the mobile phase.

Column:

— *size: l* = 0.25 m, Ø = 4 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase Mix 7 volumes of *acetonitrile for chromatography R* and 93 volumes of a 1.36 g/L solution of *sodium acetate R* containing 0.50 per cent *V/V* of *glacial acetic acid R.*

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 µL.

Run time 3.5 times the retention time of theophylline.

Relative retention With reference to theophylline (retention time = about 6 min): impurity G = about 0.6.

System suitability Reference solution (b):

— *resolution*: minimum 2.0 between the peaks due to impurity G and theophylline.

Limits:

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

Maximum 1.5 per cent, determined on 0.50 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Ethylenediamine

Dissolve 0.250 g in 30 mL of *water R*. Add 0.1 mL of *bromocresol green solution R*. Titrate with *0.1 M hydrochloric acid* until a green colour is obtained.

1 mL of *0.1 M hydrochloric acid* is equivalent to 3.005 mg of $C_2H_8N_2$.

Theophylline

Heat 0.200 g to constant mass in an oven at 135 °C. Dissolve the residue with heating in 100 mL of *water R*, allow to cool, add 20 mL of *0.1 M silver nitrate* and shake. Add 1 mL of *bromothymol blue solution R1*. Titrate with *0.1 M sodium hydroxide.*

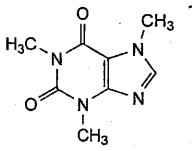
1 mL of *0.1 M sodium hydroxide* is equivalent to 18.02 mg of $C_7H_8N_4O_2$.

STORAGE

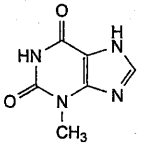
In an airtight container, protected from light.

IMPURITIES

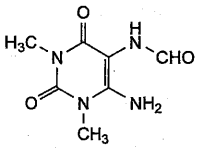
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



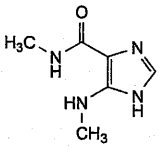
A. 1,3,7-trimethyl-3,7-dihydro-1*H*-purine-2,6-dione (caffeine),



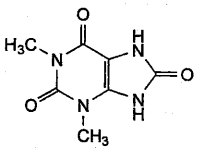
B. 3-methyl-3,7-dihydro-1*H*-purine-2,6-dione,



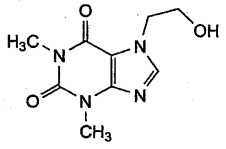
C. *N*-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide,



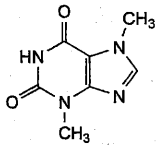
D. *N*-methyl-5-(methylamino)-1*H*-imidazole-4-carboxamide,



E. 1,3-dimethyl-7,9-dihydro-1*H*-purine-2,6,8(3*H*)-trione,



F. 7-(2-hydroxyethyl)-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (etofylline),

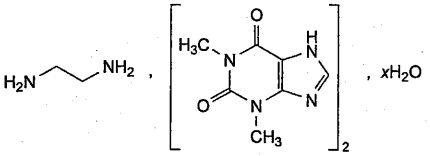


G. 3,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theobromine).

Ph Eur

Aminophylline Hydrate

(Theophylline-Ethylenediamine Hydrate, Ph. Eur. monograph 0301)



$C_{16}H_{24}N_{10}O_4,xH_2O$ 420.4 *72487-55-9*
(anhydrous substance)

Action and use
Non-selective phosphodiesterase inhibitor; treatment of reversible airways obstruction.

Preparations
Aminophylline Injection
Aminophylline Tablets
Aminopylline Prolonged-release Tablets

Ph Eur

DEFINITION

Content
— *theophylline* ($C_7H_8N_4O_2$; M_r 180.2): 84.0 per cent to 87.4 per cent (anhydrous substance);
— *ethylenediamine* ($C_2H_8N_2$; M_r 60.1): 13.5 per cent to 15.0 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS

Appearance
White or slightly yellowish powder, sometimes granular.

Solubility
Freely soluble in water (the solution becomes cloudy through absorption of carbon dioxide), practically insoluble in anhydrous ethanol.

IDENTIFICATION

First identification: B, C, E.

Second identification: A, C, D, E, F.

Dissolve 1.0 g in 10 mL of *water R* and add 2 mL of *dilute hydrochloric acid R* dropwise with shaking. Filter. Use the precipitate for identification tests A, B, D and F and the filtrate for identification test C.

A. Melting point (*2.2.14*): 270 °C to 274 °C, determined after washing the precipitate with *water R* and drying at 105 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Precipitate, washed with *water R* and dried at 105 °C.

Comparison *theophylline CRS*.

C. To the filtrate add 0.2 mL of *benzoyl chloride R*, make alkaline with *dilute sodium hydroxide solution R* and shake vigorously. Filter the precipitate, wash with 10 mL of *water R*, dissolve in 5 mL of hot *ethanol (96 per cent) R* and add 5 mL of *water R*. A precipitate is formed which, when washed and dried at 105 °C, melts (*2.2.14*) at 248 °C to 252 °C.

D. Heat about 10 mg of the precipitate with 1.0 mL of a 360 g/L solution of *potassium hydroxide R* in a water-bath at 90 °C for 3 min, then add 1.0 mL of *diazotised sulfanilic acid*

solution R. A red colour slowly develops. Carry out a blank test.

E. Water (see Tests).

F. The precipitate gives the reaction of xanthines (*2.3.1*).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).

Dissolve 0.5 g with gentle warming in 10 mL of *carbon dioxide-free water R*.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *theobromine R* (impurity G) in the mobile phase, add 5 mL of the test solution and dilute to 100 mL with the mobile phase. Dilute 5 mL of this solution to 50 mL with the mobile phase.

Column:
- *size*: l = 0.25 m, Ø = 4 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase Mix 7 volumes of *acetonitrile for chromatography R* and 93 volumes of a 1.36 g/L solution of *sodium acetate R* containing 0.50 per cent *V/V* of *glacial acetic acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 µL.

Run time 3.5 times the retention time of theophylline.

Relative retention With reference to theophylline (retention time = about 6 min): impurity G = about 0.6.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to impurity G and theophylline.

Limits:
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

3.0 per cent to 8.0 per cent, determined on 0.50 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Ethylenediamine

Dissolve 0.250 g in 30 mL of *water R*. Add 0.1 mL of *bromocresol green solution R*. Titrate with *0.1 M hydrochloric acid* until a green colour is obtained.

1 mL of *0.1 M hydrochloric acid* is equivalent to 3.005 mg of $C_2H_8N_2$.

Theophylline

Heat 0.200 g to constant mass in an oven at 135 °C. Dissolve the residue with heating in 100 mL of *water R*, allow to cool, add 20 mL of *0.1 M silver nitrate* and shake. Add 1 mL of *bromothymol blue solution R1*. Titrate with *0.1 M sodium hydroxide*.

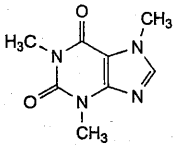
1 mL of *0.1 M sodium hydroxide* is equivalent to 18.02 mg of $C_7H_8N_4O_2$.

STORAGE

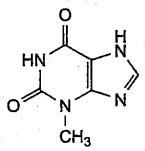
In a well-filled, airtight container, protected from light.

IMPURITIES

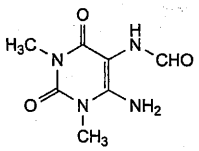
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



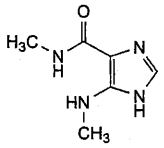
A. 1,3,7-trimethyl-3,7-dihydro-1*H*-purine-2,6-dione (caffeine),



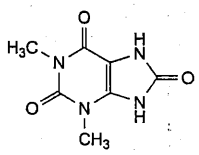
B. 3-methyl-3,7-dihydro-1*H*-purine-2,6-dione,



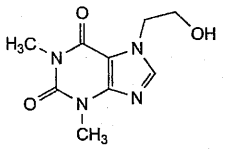
C. *N*-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide,



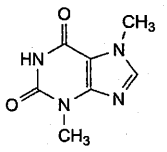
D. *N*-methyl-5-(methylamino)-1*H*-imidazole-4-carboxamide,



E. 1,3-dimethyl-7,9-dihydro-1*H*-purine-2,6,8(3*H*)-trione,



F. 7-(2-hydroxyethyl)-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (etofylline),

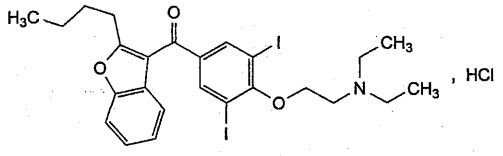


G. 3,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theobromine).

Ph Eur

Amiodarone Hydrochloride

(Ph. Eur. monograph 0803)



$C_{25}H_{30}ClI_2NO_3$ 682 *19774-82-4*

Action and use
Potassium channel blocker; class III antiarrhythmic.

Preparations
Amiodarone Intravenous Infusion
Amiodarone Oral Suspension
Amiodarone Tablets

Ph Eur

DEFINITION
(2-Butylbenzofuran-3-yl)[4-[2-(diethylamino)ethoxy]-3,5-diiodophenyl]methanone hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, fine, crystalline powder.

Solubility
Very slightly soluble in water, freely soluble in methylene chloride, soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *amiodarone hydrochloride CRS*.

B. It gives reaction (b) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 or BY_5 (*2.2.2, Method II*).
Dissolve 1.0 g in *methanol R* and dilute to 20 mL with the same solvent.

pH (*2.2.3*)
3.2 to 3.8.
Dissolve 1.0 g in *carbon dioxide-free water R*, heating at 80 °C, cool and dilute to 20 mL with the same solvent.

Impurity H
Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use and keep protected from bright light.*
Test solution Dissolve 0.500 g of the substance to be examined in *methylene chloride R* and dilute to 5.0 mL with the same solvent.
Reference solution (a) Dissolve 10.0 mg of *(2-chloroethyl) diethylamine hydrochloride R* (impurity H) in *methylene chloride R* and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 20.0 mL with *methylene chloride R*.
Reference solution (b) Mix 2.0 mL of the test solution and 2.0 mL of reference solution (a).
Plate *TLC silica gel F_{254} plate R*.
Mobile phase *anhydrous formic acid R, methanol R, methylene chloride R* (5:10:85 *V/V/V*).
Application 50 µL of the test solution and reference solution (a); 100 µL of reference solution (b).
Development Over 2/3 of the plate.
Drying In a current of cold air.
Detection Spray with *potassium iodobismuthate solution R1* and then with *dilute hydrogen peroxide solution R*; examine immediately in daylight.
System suitability Reference solution (b):
— the spot due to impurity H is clearly visible.
Limit:
— *impurity H*: any spot due to impurity H is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.02 per cent).

Related substances
Liquid chromatography (*2.2.29*).
Buffer solution pH 4.9 To 800 mL of *water R* add 3.0 mL of *glacial acetic acid R*, adjust to pH 4.9 with *dilute ammonia R1* and dilute to 1000 mL with *water R*.
Test solution Dissolve 0.125 g of the substance to be examined in a mixture of equal volumes of *acetonitrile R* and *water R* and dilute to 25.0 mL with the same mixture of solvents.
Reference solution Dissolve 5 mg of *amiodarone impurity D CRS*, 5 mg of *amiodarone impurity E CRS* and 5.0 mg of *amiodarone hydrochloride CRS* in *methanol R* and dilute to 25.0 mL with the same solvent. Dilute 1.0 mL of the solution to 20.0 mL with a mixture of equal volumes of *acetonitrile R* and *water R*.
Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.
Mobile phase Buffer solution pH 4.9, *methanol R*, *acetonitrile R* (30:30:40 *V/V/V*).
Flow rate 1 mL/min.
Detection Spectrophotometer at 240 nm.
Injection 10 µL.
Run time Twice the retention time of amiodarone.
Relative retention With reference to amiodarone (retention time = about 24 min): impurity A = about 0.26;

impurity D = about 0.29; impurity E = about 0.37; impurity B = about 0.49; impurity C = about 0.55; impurity G = about 0.62; impurity F = about 0.69.

System suitability Reference solution:
— *resolution*: minimum 3.5 between the peaks due to impurities D and E.

Limits:
— *impurities A, B, C, D, E, F, G*: for each impurity, not more than the area of the peak due to amiodarone in the chromatogram obtained with the reference solution (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the peak due to amiodarone in the chromatogram obtained with the reference solution (0.10 per cent);
— *total*: not more than 2.5 times the area of the peak due to amiodarone in the chromatogram obtained with the reference solution (0.5 per cent);
— *disregard limit*: 0.25 times the area of the peak due to amiodarone in the chromatogram obtained with the reference solution (0.05 per cent).

Iodides
Maximum 150 ppm.

Prepare the test and reference solutions simultaneously.

Solution A Add 1.50 g of the substance to be examined to 40 mL of *water R* at 80 °C and shake until completely dissolved. Cool and dilute to 50.0 mL with *water R*.

Test solution To 15.0 mL of solution A add 1.0 mL of *0.1 M hydrochloric acid* and 1.0 mL of *0.05 M potassium iodate*. Dilute to 20.0 mL with *water R*. Allow to stand protected from light for 4 h.

Reference solution To 15.0 mL of solution A add 1.0 mL of *0.1 M hydrochloric acid*, 1.0 mL of an 88.2 mg/L solution of *potassium iodide R* and 1.0 mL of *0.05 M potassium iodate*. Dilute to 20.0 mL with *water R*. Allow to stand protected from light for 4 h.

Measure the absorbances (*2.2.25*) of the solutions at 420 nm, using a mixture of 15.0 mL of solution A and 1.0 mL of *0.1 M hydrochloric acid* diluted to 20.0 mL with *water R* as the compensation liquid. The absorbance of the test solution is not greater than half the absorbance of the reference solution.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying at 50 °C at a pressure not exceeding 0.3 kPa for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.600 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 75 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

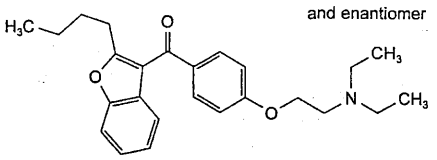
1 mL of *0.1 M sodium hydroxide* is equivalent to 68.18 mg of $C_{25}H_{30}ClI_2NO_3$.

STORAGE

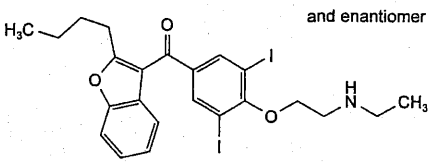
Protected from light, at a temperature not exceeding 30 °C.

IMPURITIES

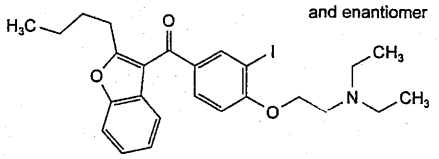
Specified impurities *A, B, C, D, E, F, G, H.*



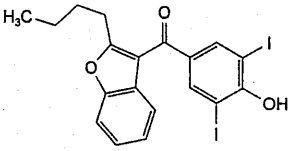
A. (2-butylbenzofuran-3-yl)[4-[2-(diethylamino)ethoxy]phenyl]methanone,



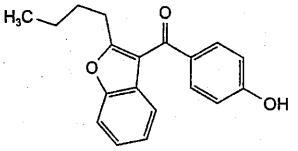
B. (2-butylbenzofuran-3-yl)[4-[2-(ethylamino)ethoxy]-3,5-diiodophenyl]methanone,



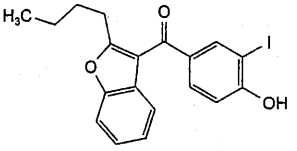
C. (2-butylbenzofuran-3-yl)[4-[2-(diethylamino)ethoxy]-3-iodophenyl]methanone,



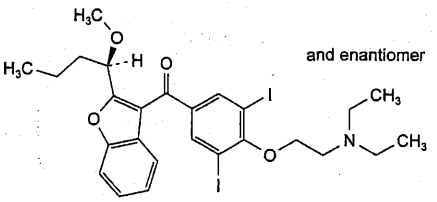
D. (2-butylbenzofuran-3-yl)(4-hydroxy-3,5-diiodophenyl)methanone,



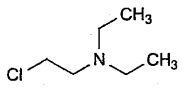
E. (2-butylbenzofuran-3-yl)(4-hydroxyphenyl)methanone,



F. (2-butylbenzofuran-3-yl)(4-hydroxy-3-iodophenyl) methanone,



G. [4-[2-(diethylamino)ethoxy]-3,5-diiodophenyl][2-[(1*RS*)-1-methoxybutyl]benzofuran-3-yl]methanone,

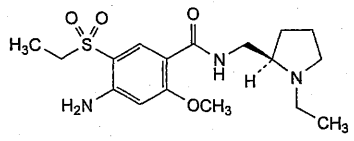


H. 2-chloro-*N*,*N*-diethylethanamine (2-chlorotriethylamine, (2-chloroethyl)diethylamine).

Ph Eur

Amisulpride

(Ph. Eur. monograph 1490)



and enantiomer

$C_{17}H_{27}N_3O_4S$ 369.5 *71675-85-9*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparations
Amisulpride Oral Solution
Amisulpride Tablets

Ph Eur

DEFINITION
4-Amino-*N*-[[(2*RS*)-1-ethylpyrrolidin-2-yl]methyl]-5-(ethylsulfonyl)-2-methoxybenzamide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in anhydrous ethanol.

mp
About 126 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *amisulpride CRS.*

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 1.0 g in 3 mL of a mixture of 1 volume of *acetic acid R* and 4 volumes of *water R*, and dilute to 20 mL with *water R*.

Impurity A
Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 0.20 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.
Reference solution (a) Dissolve 5 mg of *sulpiride impurity A CRS* (amisulpride impurity A) in *methanol R* and dilute to 25 mL with the same solvent. Dilute 2 mL of the solution to 20 mL with *methanol R*.
Reference solution (b) Dilute 1 mL of the test solution to 10 mL with reference solution (a).
Plate *TLC silica gel G plate R.*
Mobile phase 50 per cent *V/V* solution of *concentrated ammonia R, anhydrous ethanol R, di-isopropyl ether R* (10:25:65 *V/V/V*); use the upper layer obtained after shaking the mixture.
Application 10 µL.
Development Over 2/3 of the plate.
Drying In air.
Detection Spray with *ninhydrin solution R* and heat at 100-105 °C for 15 min.
Retardation factors Impurity A = about 0.2; amisulpride = about 0.5.
System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.
Limit:
— *impurity A*: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram obtained with reference solution (a) (0.1 per cent).

Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *acetonitrile R, methanol R,* mobile phase A (12:16:72 *V/V/V*).
Test solution Dissolve 0.100 g of the substance to be examined in 16 mL of *methanol R*, add 12 mL of *acetonitrile R* and dilute to 100.0 mL with mobile phase A.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.
Reference solution (b) Dissolve the contents of a vial of *amisulpride for system suitability CRS* (containing impurity B) in 1 mL of the solvent mixture.
Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.
Mobile phase:
— *mobile phase A*: dissolve 0.7 g of *sodium octanesulfonate R* in 930 mL of *water for chromatography R* and add 45.0 mL of a 5 per cent *V/V* solution of *dilute sulfuric acid R*; adjust to pH 2.3 with *dilute sulfuric acid R* and dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: *methanol R1* ;
— *mobile phase C*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 18	72	16	12
18 - 35	72 → 50	16 → 38	12

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 225 nm.
Injection 10 µL.
Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.
Relative retention With reference to amisulpride (retention time = about 17 min): impurity B = about 1.1.
System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to amisulpride.

Calculation of percentage contents:
— for each impurity, use the concentration of amisulpride in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.3 per cent;
— *reporting threshold*: 0.05 per cent.

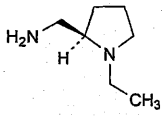
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g with shaking in a mixture of 5 mL of *acetic anhydride R* and 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

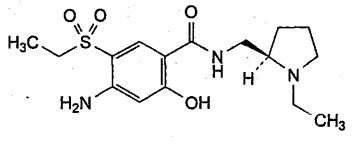
1 mL of *0.1 M perchloric acid* is equivalent to 36.95 mg of $C_{17}H_{27}N_3O_4S$.

IMPURITIES
Specified impurities *A.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H.*



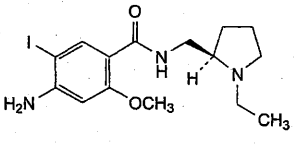
and enantiomer

A. [(2*RS*)-1-ethylpyrrolidin-2-yl]methanamine,



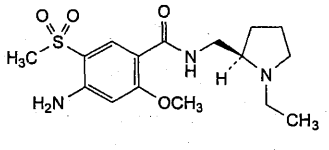
and enantiomer

B. 4-amino-*N*-[[(2*RS*)-1-ethylpyrrolidin-2-yl]methyl]-5-(ethylsulfonyl)-2-hydroxybenzamide,



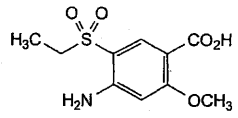
and enantiomer

C. 4-amino-*N*-[[(2*RS*)-1-ethylpyrrolidin-2-yl]methyl]-5-iodo-2-methoxybenzamide,

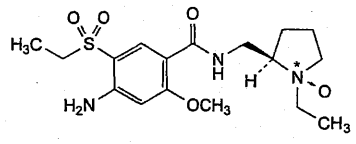


and enantiomer

D. 4-amino-*N*-[[(2*RS*)-1-ethylpyrrolidin-2-yl]methyl]-2-methoxy-5-(methylsulfonyl)benzamide,

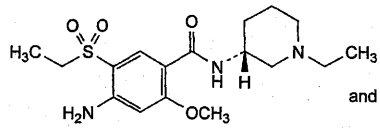


E. 4-amino-5-(ethylsulfonyl)-2-methoxybenzoic acid,



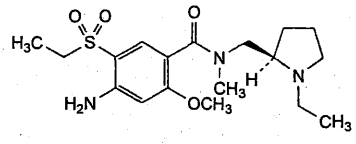
and enantiomer

F. 4-amino-*N*-[[(2*RS*)-1-ethyl-1-oxidopyrrolidin-2-yl]methyl]-5-(ethylsulfonyl)-2-methoxybenzamide,



and enantiomer

G. 4-amino-*N*-[(3*RS*)-1-ethylpiperidin-3-yl]-5-(ethylsulfonyl)-2-methoxybenzamide,



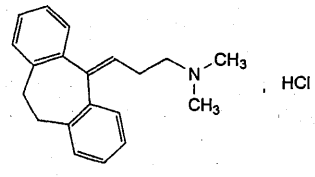
and enantiomer

H. 4-amino-*N*-[[(2*RS*)-1-ethylpyrrolidin-2-yl]methyl]-5-(ethylsulfonyl)-2-methoxy-*N*-methylbenzamide.

Ph Eur

Amitriptyline Hydrochloride

(Ph. Eur. monograph 0464)



$C_{20}H_{24}ClN$ 313.9 *549-18-8*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparations
Amitriptyline Tablets
Amitriptyline Oral Solution

Ph Eur

DEFINITION
3-(10,11-Dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ylidene)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder or colourless crystals.

Solubility
Freely soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amitriptyline hydrochloride CRS.*

B. 20 mg gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).

Dissolve 1.25 g in *water R* and dilute to 25 mL with the same solvent.

Acidity or alkalinity

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide.*
The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid.* The solution is red.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *dibenzosuberone CRS* (impurity A) and 5.0 mg of *cyclobenzaprine hydrochloride CRS* (impurity B) in 5.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 5.23 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 7.0 with *phosphoric acid R.*

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 3 times the retention time of amitriptyline.

Relative retention With reference to amitriptyline (retention time = about 14 min): impurity B = about 0.9; impurity A = about 2.2.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity B and amitriptyline.

Limits:
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *impurity A*: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the peak due to amitriptyline in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the peak due to amitriptyline in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the peak due to amitriptyline in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 30 mL of *ethanol (96 per cent) R.* Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 31.39 mg of $C_{20}H_{24}ClN$.

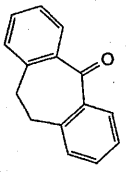
STORAGE

Protected from light.

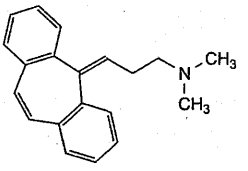
IMPURITIES

Specified impurities *A, B.*

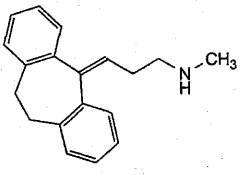
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, F, G.*



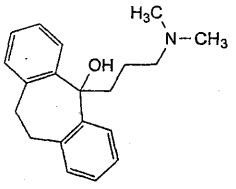
A. 10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-one (dibenzosuberone),



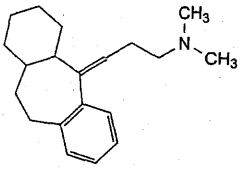
B. 3-(5*H*-dibenzo[*a*,*d*][7]annulen-5-ylidene)-*N*,*N*-dimethylpropan-1-amine (cyclobenzaprine),



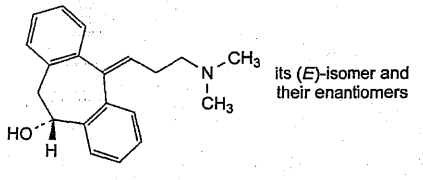
C. 3-(10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ylidene)-*N*-methylpropan-1-amine (nortriptyline),



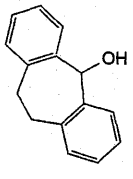
D. 5-[3-(dimethylamino)propyl]-10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ol,



E. *N*,*N*-dimethyl-3-(1,2,3,4,4a,10,11,11a-octahydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ylidene)propan-1-amine,



F. (5*EZ*,10*RS*)-5-[3-(dimethylamino)propylidene]-10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-10-ol,

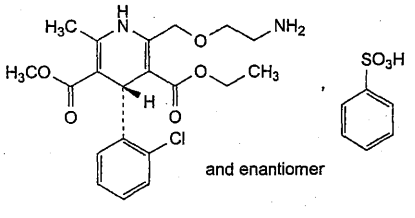


G. 10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ol (dibenzosuberol).

Ph Eur

Amlodipine Besilate

(Ph. Eur. monograph 1491)



$C_{26}H_{31}ClN_2O_8S$ 567.1 *111470-99-6*

Action and use
Calcium channel blocker.

Preparations
Amlodipine Oral Solution
Amlodipine Besilate Tablets

Ph Eur

DEFINITION
3-Ethyl 5-methyl (4*RS*)-2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-6-methyl-1,4-dihydropyridine-3,5-dicarboxylate benzenesulfonate.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION
It is considered that alkyl benzenesulfonate esters are genotoxic and are potential impurities in amlodipine besilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general method *2.5.41. Methyl, ethyl and isopropyl benzenesulfonate in active substances* is available to assist manufacturers.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Slightly soluble in water, freely soluble in methanol, sparingly soluble in anhydrous ethanol, slightly soluble in 2-propanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *amlodipine besilate CRS.*

TESTS
Optical rotation (*2.2.7*)
−0.10° to + 0.10°.
Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*
Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Test solution (b) Dilute 5.0 mL of test solution (a) to 100.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (b) Dissolve 2.5 mg of *amlodipine impurity B CRS* and 2.5 mg of *amlodipine impurity G CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 2.5 mg of *amlodipine for peak identification CRS* (containing impurities D, E and F) in 5 mL of the mobile phase.
Reference solution (d) Dissolve 5.0 mg of *amlodipine impurity A CRS* in *acetonitrile R* and dilute to 5.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (e) Dissolve 50.0 mg of *amlodipine besilate CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 100.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase 2.3 g/L solution of *ammonium acetate R*, *methanol R* (30:70 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 237 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b), (c) and (d).

Run time Twice the retention time of amlodipine.

Identification of impurities Use the chromatogram supplied with *amlodipine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D, E and F; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity A.

Relative retention With reference to amlodipine (retention time = about 20 min): impurity G = about 0.21; impurity B = about 0.25; impurity D = about 0.5; impurity F = about 0.8; impurity E = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities G and B.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 1.7; impurity F = 0.7;
— *impurity D*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity A*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.15 per cent);
— *impurities E, F*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: maximum 0.8 per cent;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to benzene sulfonate (relative retention = about 0.14).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (e).

Calculate the percentage content of $C_{26}H_{31}ClN_2O_8S$ taking into account the assigned content of *amlodipine besilate CRS*.

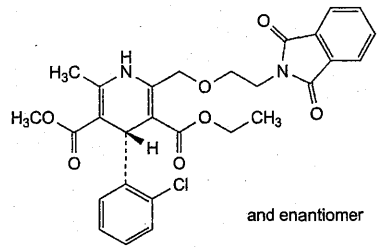
STORAGE

In an airtight container, protected from light.

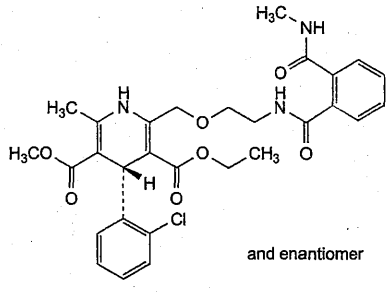
IMPURITIES

Specified impurities *A, D, E, F.*

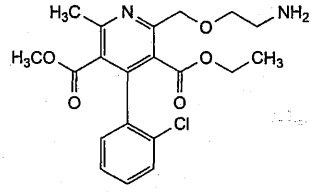
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, G, H.*



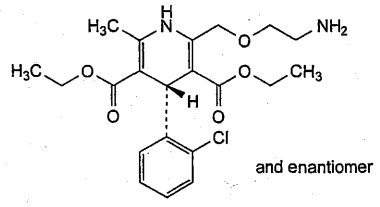
A. 3-ethyl 5-methyl (4*RS*)-4-(2-chlorophenyl)-2-[[2-(1,3-dioxo-1,3-dihydro-2*H*-isoindol-2-yl)ethoxy]methyl]-6-methyl-1,4-dihydropyridine-3,5-dicarboxylate,



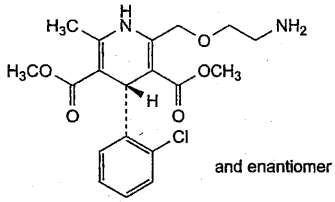
B. 3-ethyl 5-methyl (4*RS*)-4-(2-chlorophenyl)-6-methyl-2-[[2-[[2-(methylcarbamoyl)benzoyl]amino]ethoxy]methyl]-1,4-dihydropyridine-3,5-dicarboxylate,



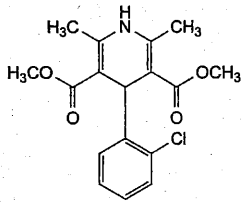
D. 3-ethyl 5-methyl 2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-6-methylpyridine-3,5-dicarboxylate,



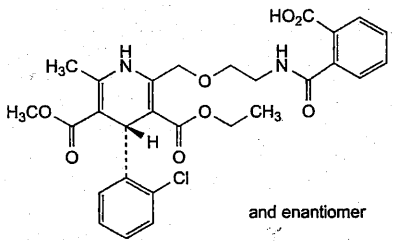
E. diethyl (4*RS*)-2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-6-methyl-1,4-dihydropyridine-3,5-dicarboxylate,



F. dimethyl (4*RS*)-2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-6-methyl-1,4-dihydropyridine-3,5-dicarboxylate,



G. dimethyl 4-(2-chlorophenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate,



H. 2-[[2-[[(4*RS*)-4-(2-chlorophenyl)-3-(ethoxycarbonyl)-5-(methoxycarbonyl)-6-methyl-1,4-dihydropyridin-2-yl]methoxy]ethyl]carbamoyl]benzoic acid.

Ph Eur

Strong Ammonia Solution

(Ammonia Solution, Concentrated, Ph. Eur. monograph 0877)

NH_3 17.03

Preparation
Dilute Ammonia Solution

Ph Eur

DEFINITION

Content
25.0 per cent *m/m* to 30.0 per cent *m/m*.

CHARACTERS

Appearance
Clear, colourless liquid, very caustic.

Solubility
Miscible with water and with ethanol (96 per cent).

IDENTIFICATION

A. Relative density (*2.2.5*): 0.892 to 0.910.

B. It is strongly alkaline (*2.2.4*).

C. To 0.5 mL add 5 mL of *water R*. Bubble air through the solution and lead the gaseous mixture obtained over the surface of a solution containing 1 mL of *0.1 M hydrochloric acid* and 0.05 mL of *methyl red solution R*. The colour changes from red to yellow. Add 1 mL of *sodium cobaltinitrite solution R*. A yellow precipitate is formed.

TESTS

Solution S
Evaporate 220 mL almost to dryness on a water-bath. Cool, add 1 mL of *dilute acetic acid R* and dilute to 20 mL with *distilled water R*.

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

To 2 mL add 8 mL of *water R*.

Oxidisable substances
Cautiously add, whilst cooling, 8.8 mL to 100 mL of *dilute sulfuric acid R*. Add 0.75 mL of *0.002 M potassium permanganate*. Allow to stand for 5 min. The solution remains faintly pink.

Pyridine and related substances
Maximum 2 ppm, calculated as pyridine.

Measure the absorbance (*2.2.25*) at 252 nm using *water R* as the compensation liquid. The absorbance is not greater than 0.06.

Carbonates
Maximum 60 ppm.

To 10 mL in a test-tube with a ground-glass neck add 10 mL of *calcium hydroxide solution R*. Stopper immediately and mix. Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of a 0.1 g/L solution of *anhydrous sodium carbonate R*.

Chlorides (*2.4.4*)
Maximum 1 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 5 ppm.

Dilute 3 mL of solution S to 15 mL with *distilled water R*.

Iron (*2.4.9*)
Maximum 0.25 ppm.

Dilute 4 mL of solution S to 10 mL with *water R*.

Residue on evaporation
Maximum 20 mg/L.

Evaporate 50 mL to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs a maximum of 1 mg.

ASSAY

Weigh accurately a flask with a ground-glass neck containing 50.0 mL of *1 M hydrochloric acid*. Add 2 mL of the substance to be examined and re-weigh. Add 0.1 mL of *methyl red solution R* as indicator. Titrate with *1 M sodium hydroxide* until the colour changes from red to yellow.

1 mL of *1 M hydrochloric acid* is equivalent to 17.03 mg of NH_3.

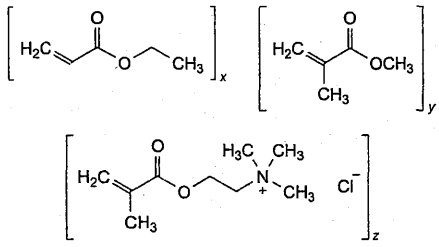
STORAGE

Protected from air, at a temperature not exceeding 20 °C.

Ph Eur

Ammonio Methacrylate Copolymer (Type A)

(Ph. Eur. monograph 2081)



Action and use
Excipient.

Ph Eur

DEFINITION

Poly[ethyl propenoate-*co*-methyl 2-methylprop-2-enoate-*co*-*N*,*N*,*N*-trimethyl-2-[(2-methylprop-2-enoyl)oxy]ethan-1-aminium chloride] having a mean relative molecular mass of about 150 000.

The ratio of ethyl acrylate (ethyl propenoate) groups to methyl methacrylate (methyl 2-methylprop-2-enoate) groups to ammonio methacrylate (*N*,*N*,*N*-trimethyl-2-[(2-methylprop-2-enoyl)oxy]ethan-1-aminium chloride) groups is about 1:2:0.2.

Content of ammonio methacrylate groups 8.9 per cent to 12.3 per cent (dried substance).

CHARACTERS

Appearance
Colourless to white or almost white granules or powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol and in methylene chloride giving clear to cloudy solutions. Due to the polymeric nature of the substance, a stirring time of up to 5 h may be necessary.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *ammonio methacrylate copolymer CRS.*

B. Viscosity (see Tests).

C. It complies with the limits of the assay.

TESTS

Solution S
Dissolve a quantity of the substance to be examined corresponding to 12.5 g of the dried substance in a mixture of 35.0 g of *acetone R* and 52.5 g of *2-propanol R*.

Viscosity (*2.2.10*)
Maximum 15 mPa·s, determined on solution S.

Apparatus Rotating viscometer.

Dimensions:
— *spindle*: diameter = 25.15 mm; height = 90.74 mm; shaft diameter = 4.0 mm;
— *cylinder*: diameter = 27.62 mm; height = 0.135 m.

Stirring speed 30 r/min.

Volume of solution 16 mL of solution S.

Temperature 20 °C.

Appearance of a film
Spread 2 mL of solution S evenly on a glass plate. Upon drying a clear film is formed.

Monomers
Liquid chromatography (*2.2.29*).

Solution A Dissolve 3.5 g of *sodium perchlorate R* in *water R* and dilute to 100 mL with the same solvent.

Test solution Dissolve 5.00 g of the substance to be examined in *methanol R* and dilute to 50.0 mL with the same solvent. To 10.0 mL of this solution add 5.0 mL of solution A, dropwise, while continuously stirring. Remove the precipitated polymer by centrifugation. Use the clear supernatant solution.

Reference solution Dissolve 50.0 mg of *ethyl acrylate R* and 10.0 mg of *methyl methacrylate R* in *methanol R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *methanol R*. Add 10 mL of this solution to 5 mL of solution A.

Column:
— *size*: *l* = 0.12 m, Ø = 4.6 mm;
— *stationary phase*: irregular *end-capped octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase Mix 20 volumes of *methanol R2* and 80 volumes of *water for chromatography R* previously ajusted to pH 2.0 with *phosphoric acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 µL.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to impurity A and impurity B.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (100 ppm);
— *impurity B*: not more than 2.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (50 ppm).

Methanol (*2.4.24, System A*)
Maximum 1.5 per cent.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 5 h.

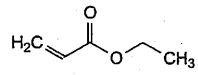
ASSAY

Dissolve 1.000 g in 75 mL of *glacial acetic acid R* at about 50 °C within about 30 min. Allow to cool to room temperature and add 25 mL of a 6 g/L solution of *copper acetate R* in *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

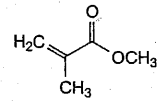
1 mL of *0.1 M perchloric acid* is equivalent to 20.77 mg of $C_9H_{18}O_2NCl$ (ammonio methacrylate groups).

IMPURITIES

Specified impurities *A, B.*



A. ethyl propenoate (ethyl acrylate),

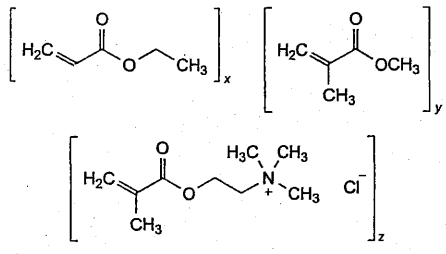


B. methyl 2-methylprop-2-enoate (methyl methacrylate).

Ph Eur

Ammonio Methacrylate Copolymer (Type B)

(Ph. Eur. monograph 2082)



Action and use
Excipient.

Ph Eur

DEFINITION

Poly[ethyl propenoate-*co*-methyl 2-methylprop-2-enoate-*co*-*N*,*N*,*N*-trimethyl-2-[(2-methylprop-2-enoyl)oxy]ethan-1-aminium chloride] having a mean relative molecular mass of about 150 000.

The ratio of ethyl acrylate (ethyl propenoate) groups to methyl methacrylate (methyl 2-methylprop-2-enoate) groups to ammonio methacrylate (*N*,*N*,*N*-trimethyl-2-[(2-methylprop-2-enoyl)oxy]ethan-1-aminium chloride) groups is about 1:2:0.1.

Content of ammonio methacrylate groups 4.5 per cent to 7.0 per cent (dried substance).

CHARACTERS

Appearance
Colourless to white or almost white granules or powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol and in methylene chloride giving clear to cloudy solutions. Due to the polymeric nature of the substance, a stirring time of up to 5 h may be necessary.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ammonio methacrylate copolymer CRS.*

B. Viscosity (see Tests).

C. It complies with the limits of the assay.

TESTS

Solution S
Dissolve a quantity of the substance to be examined corresponding to 12.5 g of the dried substance in a mixture of 35.0 g of *acetone R* and 52.5 g of *2-propanol R*.

Viscosity (*2.2.10*)
Maximum 15 mPa·s, determined on solution S.

Apparatus Rotating viscometer.

Dimensions:
— *spindle*: diameter = 25.15 mm; height = 90.74 mm; shaft diameter = 4.0 mm;
— *cylinder*: diameter = 27.62 mm; height = 0.135 m.

Stirring speed 30 r/min.

Volume of solution 16 mL of solution S.

Temperature 20 °C.

Appearance of a film
Spread 2 mL of solution S evenly on a glass plate. Upon drying a clear film is formed.

Monomers
Liquid chromatography (*2.2.29*).

Solution A Dissolve 3.5 g of *sodium perchlorate R* in *water R* and dilute to 100 mL with the same solvent.

Test solution Dissolve 5.00 g of the substance to be examined in *methanol R* and dilute to 50.0 mL with the same solvent. To 10.0 mL of this solution add 5.0 mL of solution A, dropwise, while continuously stirring. Remove the precipitated polymer by centrifugation. Use the clear supernatant solution.

Reference solution Dissolve 50.0 mg of *ethyl acrylate R* and 10.0 mg of *methyl methacrylate R* in *methanol R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *methanol R*. Add 10 mL of this solution to 5 mL of solution A.

Column:
— *size*: *l* = 0.12 m, Ø = 4.6 mm;
— *stationary phase*: irregular *end-capped octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase Mix 20 volumes of *methanol R2* and 80 volumes of *water for chromatography R* previously ajusted to pH 2.0 with *phosphoric acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 µL.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to impurity A and impurity B.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (100 ppm);
— *impurity B*: not more than 2.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (50 ppm).

Methanol (*2.4.24, System A*)
Maximum 1.5 per cent.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 5 h.

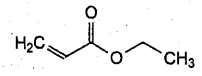
ASSAY

Dissolve 2.000 g in 75 mL of *glacial acetic acid R* at about 50 °C within about 30 min. Allow to cool to room temperature and add 25 mL of a 6 g/L solution of *copper acetate R* in *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

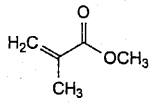
1 mL of *0.1 M perchloric acid* is equivalent to 20.77 mg of $C_9H_{18}O_2NCl$ (ammonio methacrylate groups).

IMPURITIES

Specified impurities *A, B.*



A. ethyl propenoate (ethyl acrylate),



B. methyl 2-methylprop-2-enoate (methyl methacrylate).

Ph Eur

Ammonium Bicarbonate

(Ammonium Hydrogen Carbonate, Ph. Eur. monograph 1390)

NH_4HCO_3 79.1 *1066-33-7*

Action and use
Expectorant.

Preparations
Aromatic Ammonia Solution
Strong Ammonium Acetate Solution
Aromatic Ammonia Spirit

Ph Eur

DEFINITION

Content
98.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
Fine, white or almost white, crystalline powder or white or almost white crystals, slightly hygroscopic.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent).

It volatilises rapidly at 60 °C. The volatilisation takes place slowly at ambient temperatures if the substance is slightly moist. It is in a state of equilibrium with ammonium carbamate.

IDENTIFICATION

A. It gives the reaction of carbonates and bicarbonates (*2.3.1*).

B. Dissolve 50 mg in 2 mL of *water R*. The solution gives the reaction of ammonium salts (*2.3.1*).

TESTS

Solution S
Dissolve 14.0 g in 100 mL of *distilled water R*. Boil to remove the ammonia, allow to cool and dilute to 100.0 mL with *distilled water R*.

Chlorides (*2.4.4*)
Maximum 70 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 70 ppm, determined on solution S.

Iron (*2.4.9*)
Maximum 40 ppm.

Dilute 1.8 mL of solution S to 10 mL with *water R*.

ASSAY

Dissolve cautiously 0.500 g in 50 mL of *carbon dioxide-free water R*. Titrate with *1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*). Read the volume added at the 2nd point of inflection, or at the point of inflection if only 1 point is detected.

1 mL of *1 M hydrochloric acid* is equivalent to 79.1 mg of NH_4HCO_3.

STORAGE

In an airtight container.

Ph Eur

Ammonium Bromide

(Ph. Eur. monograph 1389)

NH_4Br 97.9 *12124-97-9*

Ph Eur

DEFINITION

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, hygroscopic.

Solubility
Freely soluble in water, sparingly soluble in ethanol (96 per cent).

It becomes yellow when exposed to light or air.

IDENTIFICATION

A. It gives reaction (a) of bromides (*2.3.1*).

B. 10 mL of solution S (see Tests) gives the reaction of ammonium salts (*2.3.1*).

TESTS

Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.05 mL of *methyl red solution R*. Not more than 0.5 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Bromates
To 10 mL of solution S add 1 mL of *starch solution R*, 0.1 mL of a 100 g/L solution of *potassium iodide R* and 0.25 mL of *0.5 M sulfuric acid* and allow to stand protected from light for 5 min. No blue or violet colour develops.

Chlorides and sulfates
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.400 g of the substance to be examined in 50 mL of *water for chromatography R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 25.0 mL of test solution (a) to 50.0 mL with *water for chromatography R*.

Reference solution (a) To 25.0 mL of test solution (a) add 1.0 mL of *sulfate standard solution (10 ppm SO_4) R* and 12.0 mL of *chloride standard solution (50 ppm Cl) R* and dilute to 50.0 mL with *water for chromatography R*.

Reference solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with *water for chromatography R*. To 2.0 mL of this solution add 8.0 mL of *chloride standard solution (50 ppm Cl) R* and dilute to 20.0 mL with *water for chromatography R*.

Blank solution *water for chromatography R*.

Column:
— *size*: *l* = 0.25 m, Ø = 2 mm;
— *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (13 µm).

Mobile phase Dissolve 0.600 g of *potassium hydroxide R* in *water for chromatography R* and dilute to 1000.0 mL with the same solvent.

Flow rate 0.4 mL/min.

Detection Conductivity detector equipped with a suitable ion suppressor.

Injection 50 µL of test solution (b), reference solutions (a) and (b) and the blank solution.

Run time 2.5 times the retention time of bromide.

Retention time Chloride = about 5 min; bromide = about 8 min; sulfate = about 16 min.

System suitability Reference solution (b):
— *resolution*: minimum 8.0 between the peaks due to chloride and bromide.

Calculation of percentage contents:
— for chlorides, use the concentration of chloride in reference solution (a); correct the area of the peak due to chloride in the chromatogram obtained with reference solution (a) by subtracting the area of the peak due to chloride in the chromatogram obtained with test solution (b);
— for sulfates, use the concentration of sulfate in reference solution (a); correct the area of the peak due to sulfate in the chromatogram obtained with reference solution (a) by subtracting the area of the peak due to sulfate in the chromatogram obtained with test solution (b).

Limits:
— *chlorides*: maximum 0.6 per cent;
— *sulfates*: maximum 0.01 per cent.

Iodides
To 5 mL of solution S add 0.15 mL of *ferric chloride solution R1* and 2 mL of *methylene chloride R*. Shake and allow to separate. The lower layer is colourless (*2.2.2, Method I*).

Iron (*2.4.9*)
Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with *water R*.

Magnesium and alkaline-earth metals (*2.4.7*)
Maximum 200 ppm, calculated as Ca.

10.0 g complies with the test for magnesium and alkaline-earth metals. The volume of *0.01 M sodium edetate* used does not exceed 5.0 mL.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 80.0 mg in *water R*, add 5 mL of *dilute nitric acid R* and dilute to 50 mL with *water R*. Titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 9.794 mg of NH_4Br.

Calculate the percentage content of NH_4Br using the following expression:

$$a - 2.763\ b$$

a = percentage content of NH_4Br and NH_4Cl obtained in the assay and calculated as NH_4Br;
b = percentage content of Cl obtained in the test for chlorides.

STORAGE
In an airtight container, protected from light.

Ph Eur

Ammonium Chloride

(Ph. Eur. monograph 0007)

NH_4Cl 53.49 *12125-02-9*

Action and use
Used for the acidification of urine and to correct metabolic alkalosis.

Preparation
Ammonium Chloride Mixture

Ph Eur

DEFINITION
Content
99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water.

IDENTIFICATION
A. It gives the reactions of chlorides (*2.3.1*).

B. 10 mL of solution S (see Tests) gives the reaction of ammonium salts (*2.3.1*).

TESTS
Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.05 mL of *methyl red solution R*. Not more than 0.5 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Bromides and iodides
To 10 mL of solution S add 0.1 mL of *dilute hydrochloric acid R* and 0.05 mL of *chloramine solution R*. After 1 min, add 2 mL of *chloroform R* and shake vigorously. The chloroform layer remains colourless (*2.2.2, Method I*).

Sulfates (*2.4.13*)
Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

Calcium (*2.4.3*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *distilled water R*.

Iron (*2.4.9*)
Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.00 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 2.0 g.

ASSAY
Dissolve 1.000 g in 20 mL of *water R* and add a mixture of 5 mL of *formaldehyde solution R*, previously neutralised to *phenolphthalein solution R*, and 20 mL of *water R*. After

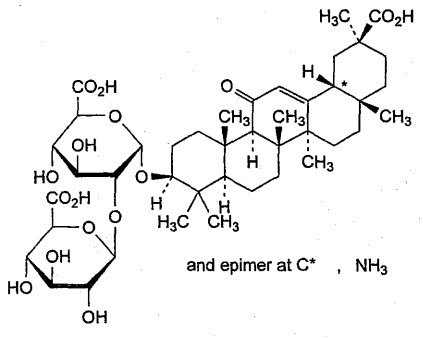
1-2 min, titrate slowly with *1 M sodium hydroxide*, using a further 0.2 mL of the same indicator.

1 mL of *1 M sodium hydroxide* is equivalent to 53.49 mg of NH_4Cl.

Ph Eur

Ammonium Glycyrrhizinate

(Ammonium Glycyrrhizate, Ph. Eur. monograph 1772)



$C_{42}H_{65}NO_{16}$ 840 *53956-04-0*

Ph Eur

DEFINITION

Mixture of ammonium 18α- and 18β-glycyrrhizate (ammonium salt of (20β)-3β-[[2-*O*-(β-D-glucopyranosyluronic acid)-α-D-glucopyranosyluronic acid] oxy]-11-oxoolean-12-en-29-oic acid), the 18β-isomer being the main component.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish-white, hygroscopic powder.

Solubility

Slightly soluble in water, very slightly soluble in anhydrous ethanol, practically insoluble in acetone. It dissolves in dilute solutions of acids and of alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ammonium glycyrrhizate CRS*.

B. Dissolve 0.1 g in 20 mL of *water R*, add 2 mL of *dilute sodium hydroxide solution R* and heat cautiously. On heating, the solution gives off vapours that may be identified by the alkaline reaction of wet litmus paper (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method I*).

Dissolve 1.0 g in *ethanol (20 per cent V/V) R* and dilute to 100.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)

+ 49.0 to + 54.0 (anhydrous substance).

Dissolve 0.5 g in *ethanol (50 per cent V/V) R* and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 50 mg of *ammonium glycyrrhizate CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 20.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase *glacial acetic acid R, acetonitrile R, water R* (6:380:614 *V/V/V*).

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 3 times the retention time of 18β-glycyrrhizic acid.

Relative retention With reference to 18β-glycyrrhizic acid (retention time = about 8 min): impurity A = about 0.8; 18α-glycyrrhizic acid = about 1.2.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to 18β-glycyrrhizic acid and 18α-glycyrrhizic acid.

Limits:
- *18α-glycyrrhizic acid*: not more than twice the sum of the areas of the peaks in the chromatogram obtained with reference solution (a) (10.0 per cent),
- *impurity A*: not more than the sum of the areas of the peaks in the chromatogram obtained with reference solution (a) (5.0 per cent),
- *any other impurity*: for each impurity, not more than 0.4 times the sum of the areas of the peaks in the chromatogram obtained with reference solution (a) (2.0 per cent),
- *sum of other impurities*: not more than 1.4 times the sum of the areas of the peaks in the chromatogram obtained with reference solution (a) (7.0 per cent),
- *disregard limit*: 0.04 times the sum of the areas of the peaks in the chromatogram obtained with reference solution (a) (0.2 per cent).

Water (*2.5.12*)

Maximum 6.0 per cent, determined on 0.250 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

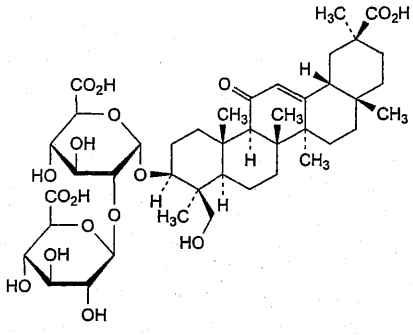
Dissolve 0.600 g in 60 mL of *anhydrous acetic acid R* heating at 80 °C if necessary. Cool. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 84.0 mg of $C_{42}H_{65}NO_{16}$.

STORAGE

In an airtight container.

IMPURITIES

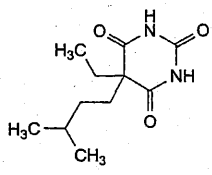


A. (4β,20β)-3β-[[2-*O*-(β-D-glucopyranosyluronic acid)-α-D-glucopyranosyluronic acid]oxy]-23-hydroxy-11-oxoolean-12-en-29-oic acid (24-hydroxyglycyrrhizinic acid).

Ph Eur

Amobarbital

(Ph. Eur. monograph 0594)



$C_{11}H_{18}N_2O_3$ 226.3 *57-43-2*

Action and use
Barbiturate.

Ph Eur

DEFINITION
Amobarbital contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 5-ethyl-5-(3-methylbutyl)pyrimidin-2,4,6(1*H*,3*H*,5*H*)-trione, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, very slightly soluble in water, freely soluble in alcohol, soluble in methylene chloride. It forms water-soluble compounds with alkali hydroxides and carbonates and with ammonia.

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Determine the melting point (*2.2.14*) of the substance to be examined. Mix equal parts of the substance to be examined and *amobarbital CRS* and determine the melting point of the mixture. The difference between the melting points (which are about 157 °C) is not greater than 2 °C.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *amobarbital CRS*.

C. Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 0.1 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dissolve 0.1 g of *amobarbital CRS* in *alcohol R* and dilute to 100 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 18 cm using the lower layer from a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of non-nitrogen substituted barbiturates (*2.3.1*).

TESTS

Appearance of solution
Dissolve 1.0 g in a mixture of 4 mL of *dilute sodium hydroxide solution R* and 6 mL of *water R*. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 1.0 g add 50 mL of *water R* and boil for 2 min. Allow to cool and filter. To 10 mL of the filtrate add 0.15 mL of *methyl red solution R* and 0.1 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.2 mL of *0.01 M hydrochloric acid*. The solution is red.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 1.0 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dilute 0.5 mL of the test solution to 100 mL with *alcohol R*.

Apply separately to the plate 20 µL of each solution. Develop over a path of 15 cm using the lower layer from a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine the plate immediately in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution. Spray with *diphenylcarbazone mercuric reagent R*. Allow the plate to dry in air and spray with freshly prepared *alcoholic potassium hydroxide solution R* diluted 1 in 5 with *aldehyde-free alcohol R*. Heat at 100 °C to 105 °C for 5 min and examine immediately. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

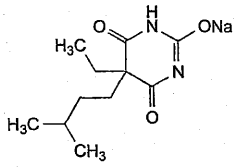
ASSAY
Dissolve 0.100 g in 5 mL of *pyridine R*. Add 0.5 mL of *thymolphthalein solution R* and 10 mL of *silver nitrate solution in pyridine R*. Titrate with *0.1 M ethanolic sodium hydroxide* until a pure blue colour is obtained. Carry out a blank titration.

1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 11.31 mg of $C_{11}H_{18}N_2O_3$.

Ph Eur

Amobarbital Sodium

(Ph. Eur. monograph 0166)



$C_{11}H_{17}N_2NaO_3$ 248.3 *64-43-7*

Action and use
Barbiturate.

Ph Eur

DEFINITION
Amobarbital sodium contains not less than 98.5 per cent and not more than the equivalent of 102.0 per cent of sodium derivative of 5-ethyl-5-(3-methylbutyl)pyrimidin-2,4,6 (1*H*,3*H*,5*H*)-trione, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, granular powder, hygroscopic, very soluble in carbon dioxide-free water (a small fraction may be insoluble), freely soluble in alcohol.

IDENTIFICATION
First identification: A, B, E.

Second identification: A, C, D, E.

A. Acidify 10 mL of solution S (see Tests) with *dilute hydrochloric acid R* and shake with 20 mL of *ether R*. Separate the ether layer, wash with 10 mL of *water R*, dry over *anhydrous sodium sulfate R* and filter. Evaporate the filtrate to dryness and dry the residue at 100 °C to 105 °C (test residue). Repeat the operations using 0.1 g of *amobarbital sodium CRS* (reference residue). Determine the melting point (*2.2.14*) of the test residue. Mix equal parts of the test residue and the reference residue and determine the melting point of the mixture. The difference between the melting points (which are about 157 °C) is not greater than 2 °C.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing the spectrum obtained with the reference residue prepared from *amobarbital sodium CRS* with that obtained with the test residue (see identification test A).

C. Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 0.1 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dissolve 0.1 g of *amobarbital sodium CRS* in *alcohol R* and dilute to 100 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 18 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of non-nitrogen substituted barbiturates (*2.3.1*).

E. It gives reaction (a) of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g in *alcohol (50 per cent V/V) R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent. Disregard any slight residue. The pH of the solution is not more than 11.0.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 1.0 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dilute 0.5 mL of the test solution to 100 mL with *alcohol R*.

Apply separately to the plate 20 µL of each solution. Develop over a path of 15 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine the plate immediately in ultraviolet light at 254 nm. Spray with *diphenylcarbazone mercuric reagent R*. Allow the plate to dry in air and spray with freshly prepared *alcoholic potassium hydroxide solution R* diluted 1 in 5 with *aldehyde-free alcohol R*. Heat at 100 °C to 105 °C for 5 min and examine immediately. When examined in ultraviolet light and after spraying, any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent). Disregard any spot at the point of application.

Loss on drying (*2.2.32*)
Not more than 3.0 per cent, determined on 0.50 g by drying in an oven at 130 °C.

ASSAY
Dissolve 0.200 g in 5 mL of *ethanol R*. Add 0.5 mL of *thymolphthalein solution R* and 10 mL of *silver nitrate solution in pyridine R*. Titrate with *0.1 M ethanolic sodium hydroxide* until a pure blue colour is obtained. Carry out a blank titration.

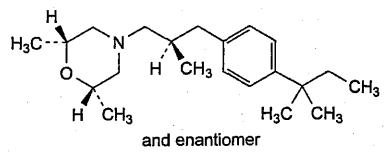
1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 24.83 mg of $C_{11}H_{17}N_2NaO_3$.

STORAGE
Store in an airtight container.

Ph Eur

Amorolfine Hydrochloride

(Ph. Eur. monograph 2756)



and enantiomer

$C_{21}H_{36}ClNO$ 354.0 *78613-38-4*

Action and use
Antifungal.

Ph Eur

DEFINITION

(2*RS*,6*SR*)-2,6-Dimethyl-4-[(2*RS*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]morpholine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, soluble in methanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amorolfine hydrochloride CRS.*

B. To 20 mg add 4.0 mL of *water R*, and acidify with *dilute nitric acid R*. A precipitate is formed. Centrifuge, and to 2 mL of the supernatant add 0.4 mL of *silver nitrate solution R1*. A curdled, white precipitate is formed. Centrifuge and wash the precipitate with 3 quantities, each of 1 mL, of *water R*. Suspend the precipitate in 2 mL of *water R* and add 1.5 mL of *ammonia R*. The precipitate dissolves easily with the possible exception of a few large particles which dissolve slowly.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 3.5 g of *dipotassium hydrogen phosphate R* in 1000 mL of *water for chromatography R* and adjust to pH 7.0 with *phosphoric acid R*.

Test solution Dissolve 20 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dissolve 4 mg of *amorolfine for system suitability CRS* (containing impurities D, E, I and J) in mobile phase A and dilute to 5 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 4 mg of *amorolfine for peak identification CRS* (containing impurity M) in mobile phase A and dilute to 5 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped amidohexadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, buffer solution, *methanol R2* (5:35:60 *V/V/V*);
— *mobile phase B*: buffer solution, *acetonitrile R1*, *methanol R2* (10:30:60 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	90	10
2 - 25	90 → 0	10 → 100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *amorolfine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities D, E, I and J; use the chromatogram supplied with *amorolfine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurity M (peaks 1 and 2).

Relative retention With reference to amorolfine (retention time = about 15 min): impurity M (peak 1) = about 0.56; impurity M (peak 2) = about 0.60; impurity D = about 0.85; impurity J = about 0.97; impurity I = about 1.05; impurity E (peak 1) = about 1.14; impurity E (peak 2) = about 1.17.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to impurity J and amorolfine in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 20 for the principal peak in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
— for each impurity, use the concentration of amorolfine hydrochloride in reference solution (b).

Limits:
— *impurity D*: maximum 0.2 per cent;
— *impurity E*: maximum 0.2 per cent for the sum of the areas of the 2 peaks;
— *impurity I*: maximum 0.15 per cent;
— *impurity M* : maximum 0.15 per cent for each peak;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 10.0 mL of *0.01 M hydrochloric acid* and 40 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

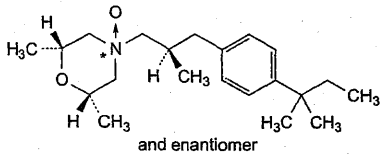
1 mL of *0.1 M sodium hydroxide* is equivalent to 35.40 mg of $C_{21}H_{36}ClNO$.

IMPURITIES

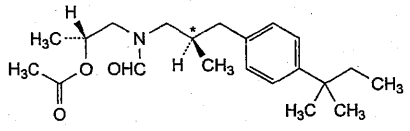
Specified impurities *D, E, I, M.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests

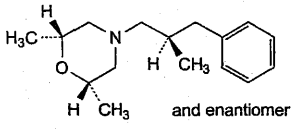
in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, C, F, G, H, J, K, L, O.



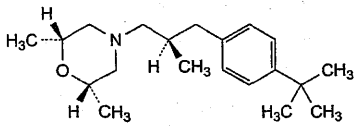
A. (2*RS*,4Ξ,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]morpholine 4-oxide,



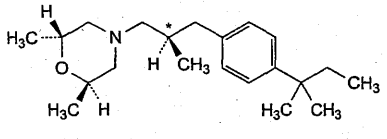
B. mixture of (2*RS*)-1-[*N*-[(2*R*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]formamido]propan-2-yl acetate and (2*RS*)-1-[*N*-[(2*S*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]formamido]propan-2-yl acetate,



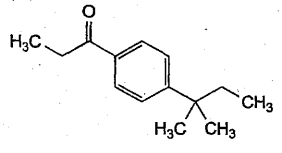
C. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-phenylpropyl]morpholine,



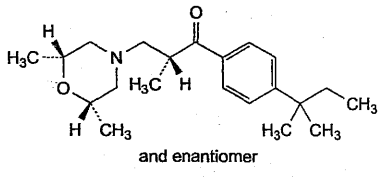
D. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-3-(4-*tert*-butylphenyl)-2-methylpropyl]morpholine,



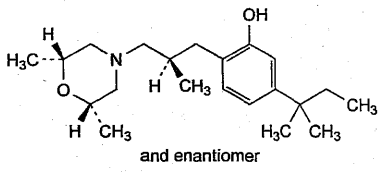
E. mixture of (2*RS*,6*RS*)-2,6-dimethyl-4-[(2*R*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]morpholine and (2*RS*,6*RS*)-2,6-dimethyl-4-[(2*S*)-2-methyl-3-[4-(2-methylbutan-2-yl)phenyl]propyl]morpholine,



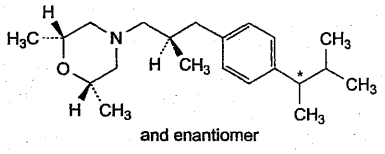
F. 1-[4-(2-methylbutan-2-yl)phenyl]propan-1-one,



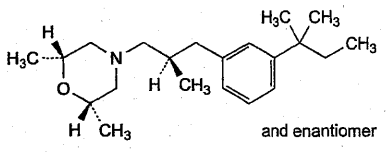
G. (2*RS*)-3-[(2*RS*,6*SR*)-2,6-dimethylmorpholin-4-yl]-1-[4-(2-methylbutan-2-yl)phenyl]-2-methylpropan-1-one,



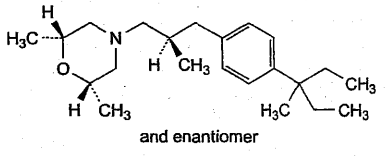
H. 2-[(2*RS*)-3-[(2*RS*,6*SR*)-2,6-dimethylmorpholin-4-yl]-2-methylpropyl]-5-(2-methylbutan-2-yl)phenol,



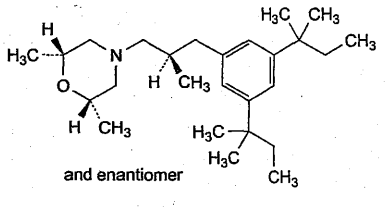
I. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-[4-[(2Ξ)-3-methylbutan-2-yl]phenyl]propyl]morpholine,



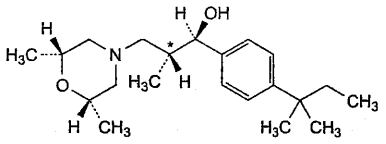
J. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-[3-(2-methylbutan-2-yl)phenyl]propyl]morpholine,



K. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-[4-(3-methylpentan-3-yl)phenyl]propyl]morpholine,

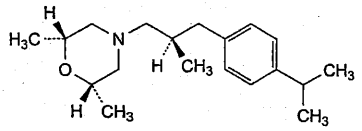


L. (2*RS*,6*SR*)-4-[(2*RS*)-3-[3,5-bis(2-methylbutan-2-yl)phenyl]-2-methylpropyl]-2,6-dimethylmorpholine,



its epimer at C* and their enantiomers

M. mixture of (1*RS*,2*RS*)-3-[(2*RS*,6*SR*)-2,6-dimethylmorpholin-4-yl]-2-methyl-1-[4-(2-methylbutan-2-yl)phenyl]propan-1-ol and (1*RS*,2*SR*)-3-[(2*RS*,6*SR*)-2,6-dimethylmorpholin-4-yl]-2-methyl-1-[4-(2-methylbutan-2-yl)phenyl]propan-1-ol,



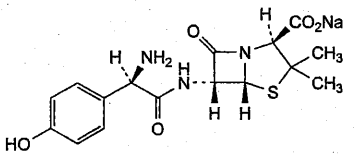
and enantiomer

O. (2*RS*,6*SR*)-2,6-dimethyl-4-[(2*RS*)-2-methyl-3-[4-(propan-2-yl)phenyl]propyl]morpholine.

Ph Eur

Amoxicillin Sodium

(Ph. Eur. monograph 0577)



$C_{16}H_{18}N_3NaO_5S$ 387.4 *34642-77-8*

Action and use
Penicillin antibacterial.

Preparations
Amoxicillin Injection
Co-amoxiclav Injection

Ph Eur

DEFINITION

Sodium (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
89.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, very hygroscopic, powder.

Solubility
Very soluble in water, sparingly soluble in anhydrous ethanol, very slightly soluble in acetone.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 0.250 g in 5 mL of *water R*, add 0.5 mL of *dilute acetic acid R*, swirl and allow to stand for 10 min in iced water. Filter the crystals and wash with 2-3 mL of a mixture of 1 volume of *water R* and 9 volumes of *acetone R*, then dry in an oven at 60 °C for 30 min.

Comparison *amoxicillin trihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (a) Dissolve 25 mg of *amoxicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (b) Dissolve 25 mg of *amoxicillin trihydrate CRS* and 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of *acetone R* and 90 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Appearance of solution
The solution is not more opalescent than reference suspension II (*2.2.1*), it may show an initial, but transient, pink colour, and after 5 min, its absorbance (*2.2.25*) at 430 nm is not greater than 0.20.

Dissolve 1.0 g in *water R* and dilute to 10.0 mL with the same solvent. Examine immediately after dissolution.

pH (*2.2.3*)
8.0 to 10.0.

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 240 to + 290 (anhydrous substance).

Dissolve 62.5 mg in a 4 g/L solution of *potassium hydrogen phthalate R* and dilute to 25.0 mL with the same solution.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A. *Prepare immediately before use.*

Reference solution (a) Dissolve 30.0 mg of *amoxicillin trihydrate CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 4.0 mg of *cefadroxil CRS* in mobile phase A and dilute to 50 mL with mobile phase A. To 5.0 mL of this solution add 5.0 mL of reference solution (a) and dilute to 100 mL with mobile phase A.

Reference solution (c) Dilute 2.0 mL of reference solution (a) to 20.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 20.0 mL with mobile phase A.

Reference solution (d) To 0.20 g of *amoxicillin trihydrate R* add 1.0 mL of *water R*. Shake and add dropwise *dilute sodium hydroxide solution R* to obtain a solution. The pH of the solution is about 8.5. Store the solution at room temperature for 4 h. Dilute 0.5 mL of this solution to 50.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 1 volume of *acetonitrile R* and 99 volumes of a 25 per cent *V/V* solution of *0.2 M potassium dihydrogen phosphate R* adjusted to pH 5.0 with *dilute sodium hydroxide solution R*;
— *mobile phase B*: mix 20 volumes of *acetonitrile R* and 80 volumes of a 25 per cent *V/V* solution of *0.2 M potassium dihydrogen phosphate R* adjusted to pH 5.0 with *dilute sodium hydroxide solution R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	92	8
t_R - (t_R + 25)	92 → 0	8 → 100
(t_R + 25) - (t_R + 40)	0	100
(t_R + 40) - (t_R + 55)	92	8

t_R = retention time of amoxicillin determined with reference solution (c)

If the mobile phase has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 µL of reference solutions (b) and (c) with isocratic elution at the initial mobile phase composition and 50 µL of test solution (b) and reference solution (d) according to the elution gradient described under Mobile phase; inject mobile phase A as a blank according to the elution gradient described under Mobile phase.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the 3 principal peaks eluted after the main peak corresponding to impurity C, amoxicillin dimer (impurity J; *n* = 1) and amoxicillin trimer (impurity J; *n* = 2).

Relative retention With reference to amoxicillin: impurity C = about 3.4; impurity J (*n* = 1) = about 4.1; impurity J (*n* = 2) = about 4.5.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to amoxicillin and cefadroxil; if necessary, adjust the ratio A: B of the mobile phase.

Limits:
— *impurity J* (*n* = 1): not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3 per cent);
— *any other impurity*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (2 per cent);
— *total*: not more than 9 times the area of the principal peak in the chromatogram obtained with reference solution (c) (9 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

N,N-Dimethylaniline (*2.4.26, Method A or B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.8 per cent *m/m*.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 0.400 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

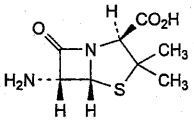
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of amoxicillin sodium by multiplying the percentage content of amoxicillin by 1.060.

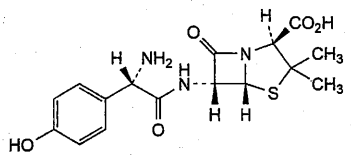
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

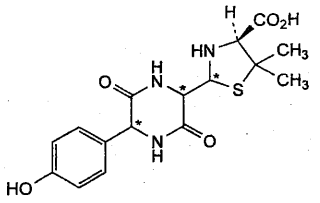
IMPURITIES



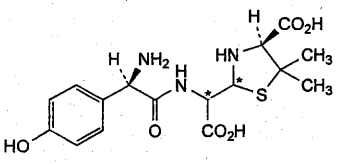
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



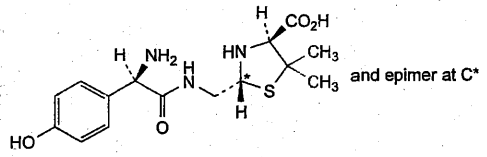
B. (2*S*,5*R*,6*R*)-6-[[(2*S*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-amoxicillin),



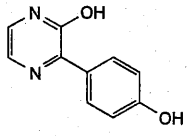
C. (4*S*)-2-[5-(4-hydroxyphenyl)-3,6-dioxopiperazin-2-yl]-5,5-dimethylthiazolidine-4-carboxylic acid (amoxicillin diketopiperazines),



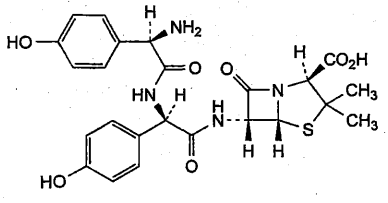
D. (4*S*)-2-[[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]carboxymethyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of amoxicillin),



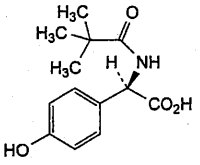
E. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of amoxicillin),



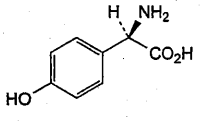
F. 3-(4-hydroxyphenyl)pyrazin-2-ol,



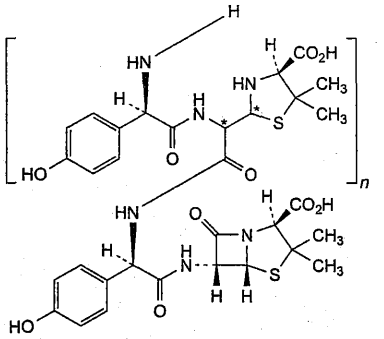
G. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane2-carboxylic acid (D-(4-hydroxyphenyl)glycylamoxicillin),



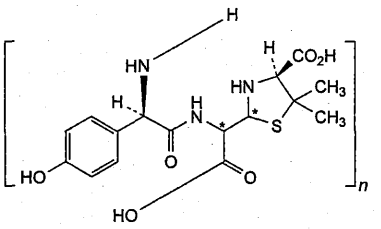
H. (2*R*)-2-[(2,2-dimethylpropanoyl)amino]-2-(4-hydroxyphenyl)acetic acid,



I. (2*R*)-2-amino-2-(4-hydroxyphenyl)acetic acid,



J. co-oligomers of amoxicillin and penicilloic acids of amoxicillin,

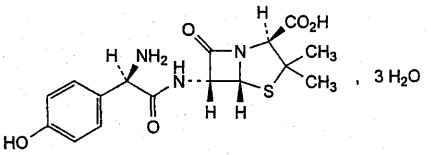


K. oligomers of penicilloic acids of amoxicillin.

Ph Eur

Amoxicillin Trihydrate

(Ph. Eur. monograph 0260)



$C_{16}H_{19}N_3O_5S,3H_2O$ 419.4 *61336-70-7*

Action and use
Penicillin antibacterial.

Preparations
Amoxicillin Capsules
Amoxicillin Oral Suspension
Co-amoxiclav Oral Suspension
Co-amoxiclav Tablets
Co-amoxiclav Dispersible Tablets

Ph Eur

DEFINITION
(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-Amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid trihydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in fatty oils. It dissolves in dilute acids and dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amoxicillin trihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (a) Dissolve 25 mg of *amoxicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (b) Dissolve 25 mg of *amoxicillin trihydrate CRS* and 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of *acetone R* and 90 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

TESTS

Solution S
With the aid of ultrasound or gentle heating, dissolve 0.100 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

pH (*2.2.3*)
3.5 to 5.5 for solution S.

Specific optical rotation (*2.2.7*)
+ 290 to + 315 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Buffer solution pH 5.0 To 250 mL of *0.2 M potassium dihydrogen phosphate R* add *dilute sodium hydroxide solution R* to pH 5.0 and dilute to 1000.0 mL with *water R.*

Test solution (a) Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A. *Prepare immediately before use.*

Reference solution (a) Dissolve 30.0 mg of *amoxicillin trihydrate CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 4.0 mg of *cefadroxil CRS* in mobile phase A and dilute to 50 mL with mobile phase A. To 5.0 mL of this solution add 5.0 mL of reference solution (a) and dilute to 100 mL with mobile phase A.

Reference solution (c) Dilute 2.0 mL of reference solution (a) to 20.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 20.0 mL with mobile phase A.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A: acetonitrile R*, buffer solution pH 5.0 (1:99 *V/V*);
— *mobile phase B: acetonitrile R*, buffer solution pH 5.0 (20:80 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	92	8
t_R - (t_R + 25)	92 → 0	8 → 100
(t_R + 25) - (t_R + 40)	0	100
(t_R + 40) - (t_R + 55)	92	8

t_R = retention time of amoxicillin determined with reference solution (c)

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 µL of reference solutions (b) and (c) with isocratic elution at the initial mobile phase composition and 50 µL of test solution (b) according to the elution gradient described under Mobile phase; inject mobile phase A as a blank according to the elution gradient described under Mobile phase.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to amoxicillin and cefadroxil; if necessary, adjust the ratio A: B of the mobile phase.

Limit:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1 per cent).

***N,N*-Dimethylaniline** (*2.4.26, Method A or B*)
Maximum 20 ppm.

Water (*2.5.12*)
11.5 per cent to 14.5 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

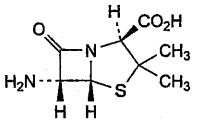
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of $C_{16}H_{19}N_3O_5S$ taking into account the assigned content of *amoxicillin trihydrate CRS*.

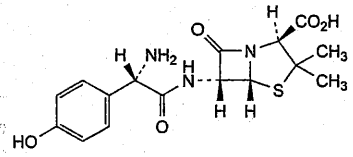
STORAGE

In an airtight container.

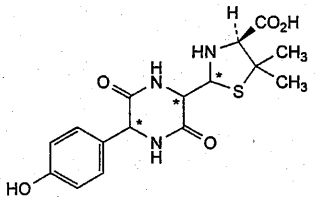
IMPURITIES



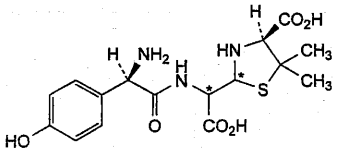
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



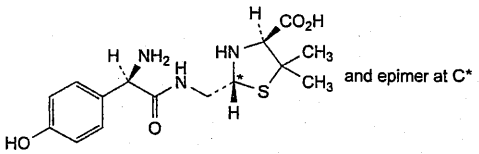
B. (2*S*,5*R*,6*R*)-6-[[(2*S*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-amoxicillin),



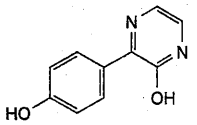
C. (4*S*)-2-[5-(4-hydroxyphenyl)-3,6-dioxopiperazin-2-yl]-5,5-dimethylthiazolidine-4-carboxylic acid (amoxicillin diketopiperazines),



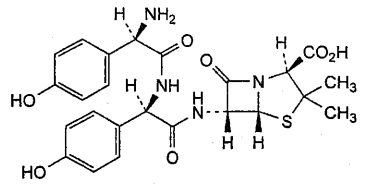
D. (4*S*)-2-[[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]carboxymethyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of amoxicillin),



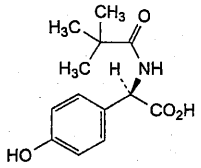
E. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of amoxicillin),



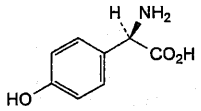
F. 3-(4-hydroxyphenyl)pyrazin-2-ol,



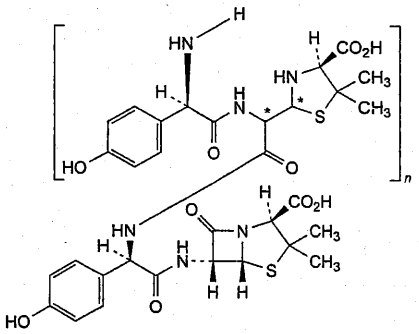
G. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-2-(4-hydroxy-phenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (D-(4-hydroxyphenyl) glycylamoxicillin),



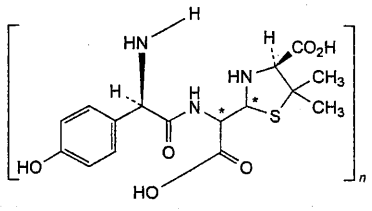
H. (2*R*)-2-[(2,2-dimethylpropanoyl)amino]-2-(4-hydroxyphenyl)acetic acid,



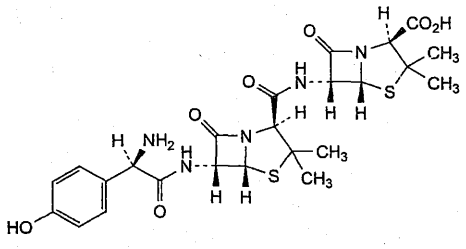
I. (2*R*)-2-amino-2-(4-hydroxyphenyl)acetic acid,



J. co-oligomers of amoxicillin and of penicilloic acids of amoxicillin,



K. oligomers of penicilloic acids of amoxicillin,

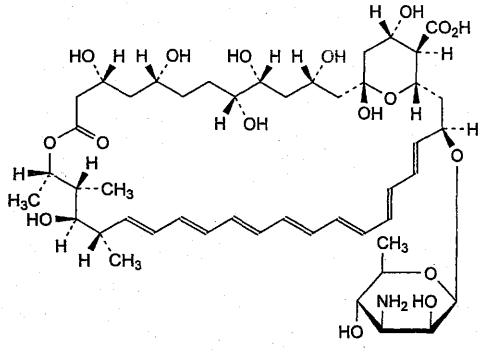


L. (2*S*,5*R*,6*R*)-6-[[(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-APA amoxicillin amide).

Ph Eur

Amphotericin

(Amphotericin B, Ph. Eur. monograph 1292)



$C_{47}H_{73}NO_{17}$ 924 *1397-89-3*

Action and use
Antifungal.

Preparation
Amphotericin for Infusion

Ph Eur

DEFINITION

Mixture of antifungal polyenes produced by the growth of certain strains of *Streptomyces nodosus* or obtained by any other means. It consists mainly of amphotericin B which is (1*R*,3*S*,5*R*,6*R*,9*R*,11*R*,15*S*,16*R*,17*R*,18*S*,19*E*,21*E*,23*E*,25*E*,27*E*,29*E*,31*E*,33*R*,35*S*,36*R*,37*S*)-33-[(3-amino-3,6-dideoxy-β-D-mannopyranosyl)oxy]-1,3,5,6,9,11,17,37-octahydroxy-15,16,18-trimethyl-13-oxo-14,39-dioxa-bicyclo[33.3.1]nonatriaconta-19,21,23,25,27,29,31-heptaene-36-carboxylic acid.

Content
Minimum 750 IU/mg (dried substance).

CHARACTERS

Appearance
Yellow or orange, hygroscopic powder.

Solubility
Practically insoluble in water, soluble in dimethyl sulfoxide and in propylene glycol, slightly soluble in dimethylformamide, very slightly soluble in methanol, practically insoluble in ethanol (96 per cent).

It is sensitive to light in dilute solutions.

IDENTIFICATION

First identification: B, D.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 25 mg in 5 mL of *dimethyl sulfoxide R* and dilute to 50 mL with *methanol R*. Dilute 2 mL of the solution to 200 mL with *methanol R*.

Spectral range 300-450 nm.

Absorption maxima At 362 nm, 381 nm and 405 nm.

Absorbance ratios:
- A_{362}/A_{381} = 0.57 to 0.61;
- A_{381}/A_{405} = 0.87 to 0.93.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *amphotericin B CRS*.

If the spectra obtained show differences, dry the substance to be examined and reference substance at 60 °C at a pressure not exceeding 0.7 kPa for 1 h and record new spectra.

C. To 1 mL of a 0.5 g/L solution in *dimethyl sulfoxide R*, add 5 mL of *phosphoric acid R* to form a lower layer, avoiding mixing the 2 liquids. A blue ring is immediately produced at the junction of the liquids. Mix, an intense blue colour is produced. Add 15 mL of *water R* and mix; the solution becomes pale yellow.

D. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with the test solution at 383 nm is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Protect the solutions from light and use within 24 h of preparation, except for reference solution (c) which should be injected immediately after its preparation.*

Solvent mixture 10 g/L solution of *ammonium acetate R*, *N-methylpyrrolidone R*, *methanol R* (1:1:2 *V/V/V*).

Test solution Dissolve 20.0 mg of the substance to be examined in 15 mL of *N-methylpyrrolidone R* and within 2 h dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of *amphotericin B CRS* in 15 mL of *N-methylpyrrolidone R* and within 2 h dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 25.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 20.0 mg of *nystatin CRS* in 15 mL of *N-methylpyrrolidone R* and within 2 h dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 25.0 mL with reference solution (a). Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (d) In order to prepare impurities B and C, dissolve 10 mg of the substance to be examined in 5 mL of *N-methylpyrrolidone R* and within 2 h add 35 mL of a mixture of 1 volume of *methanol R* and 4 volumes of *anhydrous ethanol R*. Add 0.10 mL of *dilute hydrochloric acid R*, mix and incubate at 25 °C for 2.5 h. Add 10 mL of 10 g/L solution of *ammonium acetate R* and mix.

Reference solution (e) Dissolve 4 mg of *amphotericin B for peak identification CRS* (containing impurities A and B) in 5 mL of *N-methylpyrrolidone R* and within 2 h dilute to 50 mL with the solvent mixture.

Blank solution The solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 20 °C.

Mobile phase:
- *mobile phase A*: mix 1 volume of *methanol R*, 3 volumes of *acetonitrile R* and 6 volumes of a 4.2 g/L solution of *citric acid monohydrate R* previously adjusted to pH 4.7 using *concentrated ammonia R*;
- *mobile phase B*: mix 12 volumes of *methanol R*, 20 volumes of a 4.2 g/L solution of *citric acid monohydrate R* previously adjusted to pH 3.9 using *concentrated ammonia R* and 68 volumes of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 23	100 → 70	0 → 30
23 - 33	70 → 0	30 → 100
33 - 40	0	100

Flow rate 0.8 mL/min.

Detection Spectrophotometer:
- at 303 nm: detection of tetraenes;
- at 383 nm: detection of heptaenes.

Injection 20 µL of the test solution and reference solutions (b), (c), (d) and (e).

Identification of impurities Use the chromatograms supplied with *amphotericin B for peak identification CRS* and the chromatograms obtained with reference solution (e) to identify the peaks due to impurities A and B.

Relative retention With reference to amphotericin B (retention time = about 16 min): impurity B = about 0.75; impurity A = about 0.8; nystatin = about 0.85.

System suitability at 383 nm Reference solution (d):
- *resolution*: minimum 1.5 between the 2 peaks presenting a relative retention of about 0.7.

Limits:
- *impurity A at 303 nm*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (5.0 per cent); if intended for use in the manufacture of parenteral preparations: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent);
- *any other impurity at 303 nm*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
- *impurity B at 383 nm*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (4.0 per cent);
- *any other impurity at 383 nm*: for each impurity, not more than 2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
- *total at 303 and 383 nm*: maximum 15.0 per cent;
- *disregard limit at 303 nm*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
- *disregard limit at 383 nm*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.7 kPa.

Sulfated ash (*2.4.14*)
Maximum 3.0 per cent, determined on 1.0 g; if intended for use in the manufacture of parenteral preparations: maximum 0.5 per cent.

Bacterial endotoxins (*2.6.14*)
Less than 1.0 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Protect all solutions from light throughout the assay Dissolve 25.0 mg in *dimethyl sulfoxide R* and dilute, with shaking, to 25.0 mL with the same solvent. Under constant stirring of this stock solution, dilute with *dimethyl sulfoxide R* to obtain solutions of appropriate concentrations (the following concentrations have been found suitable: 44.4, 66.7 and 100 IU/mL). Prepare final solutions by diluting 1:20 with 0.2 M phosphate buffer solution pH 10.5 so that they all contain 5 per cent *V/V* of dimethyl sulfoxide. Prepare the reference and the test solutions simultaneously. Carry out the microbiological assay of antibiotics (*2.7.2*).

STORAGE

Protected from light, at a temperature of 2 °C to 8 °C in an airtight container. If the substance is sterile, store in a sterile, tamper-proof container.

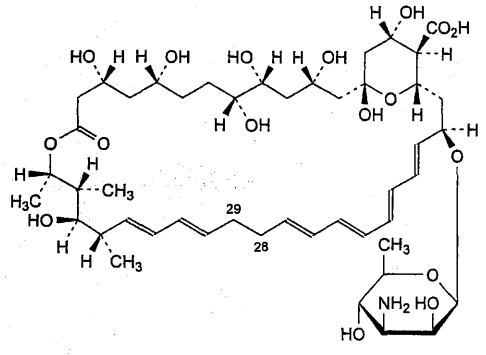
LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

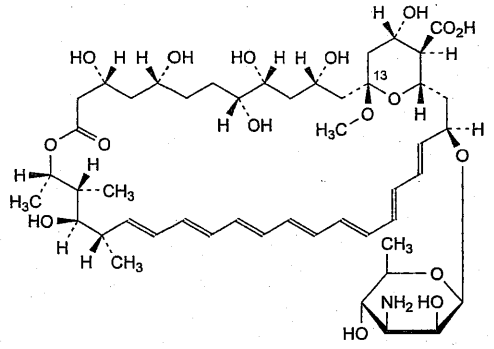
IMPURITIES

Specified impurities *A, B.*

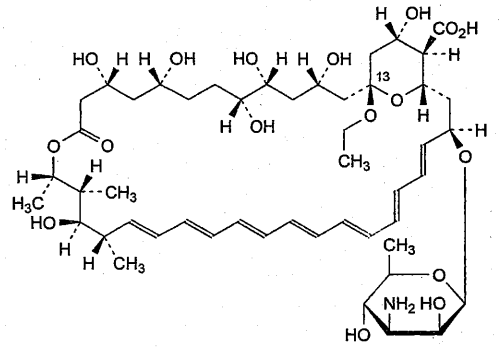
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



A. amphotericin A (28,29-dihydro-amphotericin B),



B. amphotericin X1 (13-*O*-methyl-amphotericin B),

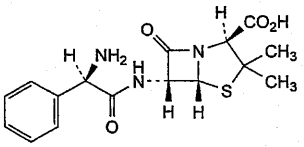


C. amphotericin X2 (13-*O*-ethyl-amphotericin B).

Ph Eur

Ampicillin

(Ph. Eur. monograph 0167)



$C_{16}H_{19}N_3O_4S$ 349.4 *69-53-4*

Action and use
Penicillin antibacterial.

Preparations
Ampicillin Capsules
Ampicillin Oral Suspension

Ph Eur

DEFINITION

(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-Amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, practically insoluble in acetone, in ethanol (96 per cent) and in fatty oils. It dissolves in dilute solutions of acids and of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium bromide R.*

Comparison *anhydrous ampicillin CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (a) Dissolve 25 mg of *anhydrous ampicillin CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (b) Dissolve 25 mg of *amoxicillin trihydrate CRS* and 25 mg of *anhydrous ampicillin CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of *acetone R* and 90 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

D. Water (see Tests).

TESTS

Appearance of solution
The solutions are not more opalescent than reference suspension II (*2.2.1*).

Dissolve 1.0 g in 10 mL of *1 M hydrochloric acid.* Separately dissolve 1.0 g in 10 mL of *dilute ammonia R2.* Examine immediately after dissolution.

pH (*2.2.3*)
3.5 to 5.5.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 40 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 280 to + 305 (anhydrous substance).

Dissolve 62.5 mg in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 27.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) *Prepare immediately before use.* Dissolve 27.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 27.0 mg of *anhydrous ampicillin CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 2.0 mg of *cefradine CRS* in mobile phase A and dilute to 50 mL with mobile phase A. To 5.0 mL of this solution add 5.0 mL of reference solution (a).

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 20.0 mL with mobile phase A.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 0.5 mL of *dilute acetic acid R*, 50 mL of *0.2 M potassium dihydrogen phosphate R* and 50 mL of *acetonitrile R*, then dilute to 1000 mL with *water R*;
— *mobile phase B*: mix 0.5 mL of *dilute acetic acid R*, 50 mL of *0.2 M potassium dihydrogen phosphate R* and 400 mL of *acetonitrile R*, then dilute to 1000 mL with *water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	85	15
t_R - (t_R + 30)	85 → 0	15 → 100
(t_R + 30) - (t_R + 45)	0	100
(t_R + 45) - (t_R + 60)	85	15

t_R = retention time of ampicillin determined with reference solution (c)

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 µL of reference solutions (b) and (c) with isocratic elution at the initial mobile phase composition and 50 µL of test solution (b) according to the elution gradient described under Mobile phase; inject mobile phase A as a blank according to the elution gradient described under Mobile phase.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to ampicillin and cefradin; if necessary, adjust the ratio A:B of the mobile phase.

Limit:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)
Maximum 20 ppm.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

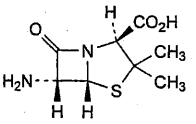
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of $C_{16}H_{19}N_3O_4S$ from the declared content of *anhydrous ampicillin CRS*.

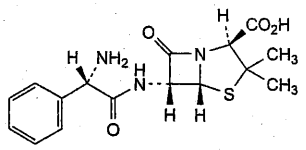
STORAGE

In an airtight container, at a temperature not exceeding 30 °C.

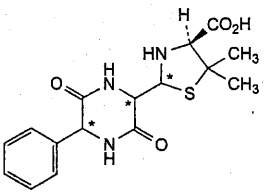
IMPURITIES



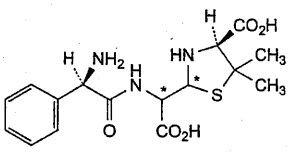
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



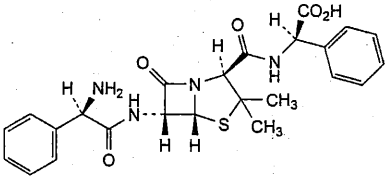
B. (2*S*,5*R*,6*R*)-6-[[(2*S*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-ampicillin),



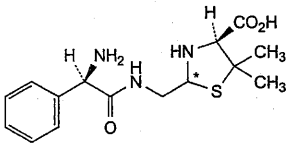
C. (4*S*)-2-(3,6-dioxo-5-phenylpiperazin-2-yl)-5,5-dimethylthiazolidine-4-carboxylic acid (diketopiperazines of ampicillin),



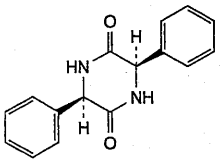
D. (4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]carboxymethyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acids of ampicillin),



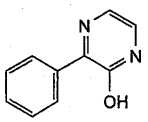
E. (2*R*)-2-[[[(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-2-phenylacetic acid (ampicillinyl-D-phenylglycine),



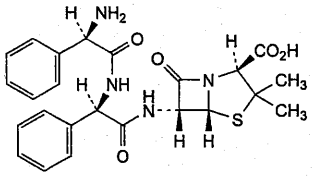
F. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penilloic acids of ampicillin),



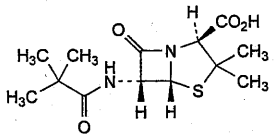
G. (3*R*,6*R*)-3,6-diphenylpiperazine-2,5-dione,



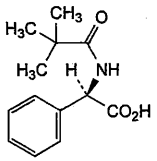
H. 3-phenylpyrazin-2-ol,



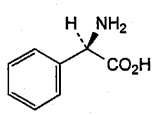
I. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (D-phenylglycylampicillin),



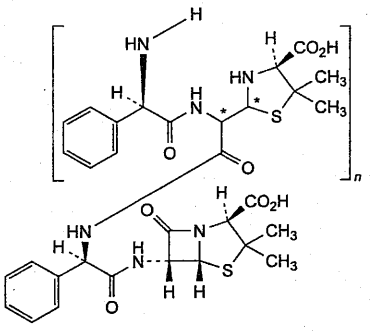
J. (2*S*,5*R*,6*R*)-6-[(2,2-dimethylpropanoyl)amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,



K. (2*R*)-2-[(2,2-dimethylpropanoyl)amino]-2-phenylacetic acid,



L. (2*R*)-2-amino-2-phenylacetic acid (D-phenylglycine),

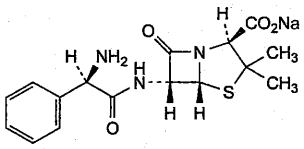


M. co-oligomers of ampicillin and of penicilloic acids of ampicillin.

Ph Eur

Ampicillin Sodium

(Ph. Eur. monograph 0578)



$C_{16}H_{18}N_3NaO_4S$ 371.4 *69-52-3*

Action and use
Penicillin antibacterial.

Preparation
Ampicillin Injection

Ph Eur

DEFINITION
Sodium (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
91.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder, hygroscopic.

Solubility
Freely soluble in water, sparingly soluble in acetone, practically insoluble in fatty oils and in liquid paraffin.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 0.250 g in 5 mL of *water R*, add 0.5 mL of *dilute acetic acid R*, swirl and allow to stand for 10 min in iced water. Filter the crystals through a small sintered-glass filter (40) (*2.1.2*), applying suction, wash with 2-3 mL of a mixture of 1 volume of *water R* and 9 volumes of *acetone R*, then dry in an oven at 60 °C for 30 min.

Comparison *ampicillin trihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (a) Dissolve 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (b) Dissolve 25 mg of *amoxicillin trihydrate CRS* and 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of *acetone R* and 90 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Appearance of solution

Solutions A and B are not more opalescent than reference suspension II (*2.2.1*) and the absorbance (*2.2.25*) of solution B at 430 nm is not greater than 0.15.

Place 1.0 g in a conical flask and add slowly and with continuous swirling 10 mL of *1 M hydrochloric acid* (solution A). Separately dissolve 1.0 g in *water R* and dilute to 10.0 mL with the same solvent (solution B). Examine immediately after dissolution.

pH (*2.2.3*)

8.0 to 10.0.

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent. Measure 10 min after dissolution.

Specific optical rotation (*2.2.7*)

+ 258 to + 287 (anhydrous substance).

Dissolve 62.5 mg in a 4 g/L solution of *potassium hydrogen phthalate R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 31.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dissolve 31.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A. *Prepare immediately before use.*

Reference solution (a) Dissolve 27.0 mg of *anhydrous ampicillin CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 2.0 mg of *cefradine CRS* in mobile phase A and dilute to 50 mL with mobile phase A. To 5.0 mL of this solution add 5.0 mL of reference solution (a).

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 20.0 mL with mobile phase A.

Reference solution (d) To 0.20 g of the substance to be examined add 1.0 mL of *water R.* Heat the solution at 60 °C for 1 h. Dilute 0.5 mL of this solution to 50.0 mL with mobile phase A.

Column:
— *size:* l = 0.25 m, Ø = 4.6 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A:* mix 0.5 mL of *dilute acetic acid R,* 50 mL of *0.2 M potassium dihydrogen phosphate R* and 50 mL of *acetonitrile R,* then dilute to 1000 mL with *water R;*
— *mobile phase B:* mix 0.5 mL of *dilute acetic acid R,* 50 mL of *0.2 M potassium dihydrogen phosphate R* and 400 mL of *acetonitrile R,* then dilute to 1000 mL with *water R;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	85	15
t_R - (t_R + 30)	85 → 0	15 → 100
(t_R + 30) - (t_R + 45)	0	100
(t_R + 45) - (t_R + 60)	85	15

t_R = retention time of ampicillin determined with reference solution (c)

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 µL of reference solutions (b) and (c) with isocratic elution at the initial mobile phase composition and 50 µL of test solution (b) and reference solution (d) according to the elution gradient described under Mobile phase; inject mobile phase A as a blank according to the elution gradient described under Mobile phase.

Identification of peaks Use the chromatogram obtained with reference solution (d) to identify the peaks due to ampicillin and ampicillin dimer.

Relative retention With reference to ampicillin: ampicillin dimer = about 2.8.

System suitability Reference solution (b):
— *resolution:* minimum 3.0 between the peaks due to ampicillin and cefradin; if necessary adjust the ratio A:B of the mobile phase.

Limits:
— *ampicillin dimer:* not more than 4.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (4.5 per cent);
— *any other impurity:* for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (2 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)

Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)

Maximum 0.8 per cent *m/m.*

Methylene chloride

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 1.0 mL of *ethylene chloride R* in *water R* and dilute to 500.0 mL with the same solvent.

Test solution (a) Dissolve 1.0 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Test solution (b) Dissolve 1.0 g of the substance to be examined in *water R*, add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*.

Reference solution Dissolve 1.0 mL of *methylene chloride R* in *water R* and dilute to 500.0 mL with the same solvent. To 1.0 mL of this solution add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*.

Column:
— *material*: glass;
— *size*: *l* = 1.5 m, Ø = 4 mm;
— *stationary phase*: diatomaceous earth for gas chromatography R impregnated with 10 per cent *m/m* of macrogol 1000 R.

Carrier gas *nitrogen for chromatography R*.

Flow rate 40 mL/min.

Temperature:
— *column*: 60 °C;
— *injection port*: 100 °C;
— *detector*: 150 °C.

Detection Flame ionisation.

Calculate the content of methylene chloride taking its density at 20 °C to be 1.325 g/mL.

Limit:
— *methylene chloride*: maximum 0.2 per cent *m/m*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.300 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.15 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

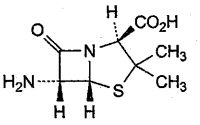
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of ampicillin sodium by multiplying the percentage content of ampicillin by 1.063.

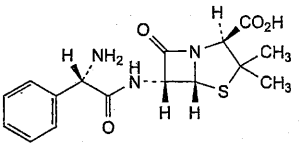
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

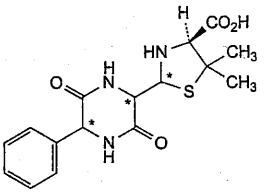
IMPURITIES



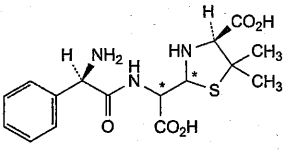
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



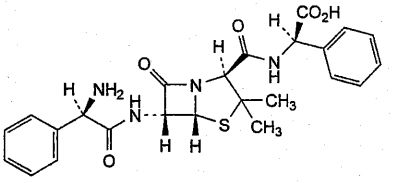
B. (2*S*,5*R*,6*R*)-6-[[(2*S*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-ampicillin),



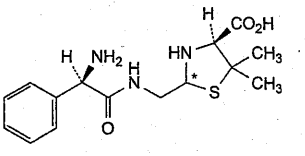
C. (4*S*)-2-(3,6-dioxo-5-phenylpiperazin-2-yl)-5,5-dimethylthiazolidine-4-carboxylic acid (diketopiperazines of ampicillin),



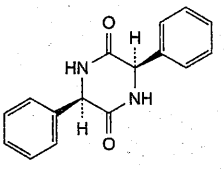
D. (4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]carboxymethyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of ampicillin),



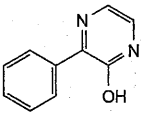
E. (2*R*)-2-[[[(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-2-phenylacetic acid (ampicillinyl-D-phenylglycine),



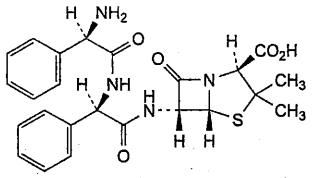
F. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of ampicillin),



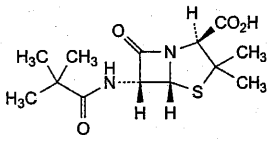
G. (3*R*,6*R*)-3,6-diphenylpiperazine-2,5-dione,



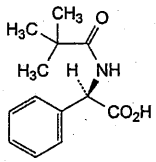
H. 3-phenylpyrazin-2-ol,



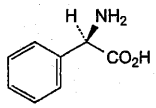
I. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia1-azabicyclo[3.2.0]heptane-2-carboxylic acid (D-phenylglycylampicillin),



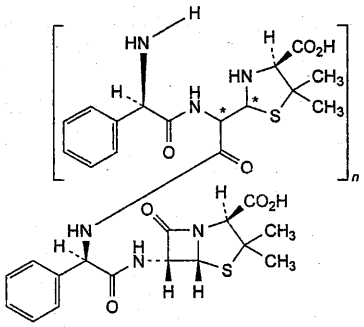
J. (2*S*,5*R*,6*R*)-6-[(2,2-dimethylpropanoyl)amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,



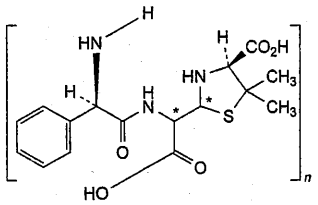
K. (2*R*)-2-[(2,2-dimethylpropanoyl)amino]-2-phenylacetic acid,



L. (2*R*)-2-amino-2-phenylacetic acid (D-phenylglycine),



M. co-oligomers of ampicillin and of penicilloic acids of ampicillin,

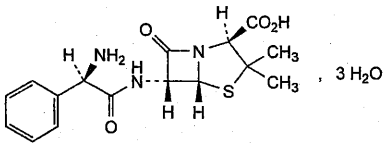


N. oligomers of penicilloic acids of ampicillin.

Ph Eur

Ampicillin Trihydrate

(Ph. Eur. monograph 0168)



$C_{16}H_{19}N_3O_4S,3H_2O$ 403.5 *7177-48-2*

Action and use
Penicillin antibacterial.

Preparations
Ampicillin Capsules
Ampicillin Oral Suspension
Co-fluampicil Capsules
Co-fluampicil Oral Suspension

Ph Eur

DEFINITION

(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-Amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid trihydrate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, practically insoluble in ethanol (96 per cent) and in fatty oils. It dissolves in dilute solutions of acids and of alkali hydroxides.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ampicillin trihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (a) Dissolve 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Reference solution (b) Dissolve 25 mg of *amoxicillin trihydrate CRS* and 25 mg of *ampicillin trihydrate CRS* in 10 mL of *sodium hydrogen carbonate solution R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of *acetone R* and 90 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R*. Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

D. Water (see Tests).

TESTS

Appearance of solution

The solutions are not more opalescent than reference suspension II (*2.2.1*).

Dissolve 1.0 g in 10 mL of *1 M hydrochloric acid*. Separately dissolve 1.0 g in 10 mL of *dilute ammonia R2*. Examine immediately after dissolution.

pH (*2.2.3*)

3.5 to 5.5.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 40 mL with the same solvent.

Specific optical rotation (*2.2.7*)

+ 280 to + 305 (anhydrous substance).

Dissolve 62.5 mg in *water R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 31.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dissolve 31.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A. *Prepare immediately before use.*

Reference solution (a) Dissolve 27.0 mg of *anhydrous ampicillin CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 2 mg of *cefradine CRS* in mobile phase A and dilute to 50 mL with mobile phase A. To 5 mL of this solution, add 5 mL of reference solution (a).

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 20.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: mix 0.5 mL of *dilute acetic acid R*, 50 mL of *0.2 M potassium dihydrogen phosphate R* and 50 mL of *acetonitrile R*, then dilute to 1000 mL with *water R*;
- *mobile phase B*: mix 0.5 mL of *dilute acetic acid R*, 50 mL of *0.2 M potassium dihydrogen phosphate R* and 400 mL of *acetonitrile R*, then dilute to 1000 mL with *water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	85	15
t_R - (t_R + 30)	85 → 0	15 → 100
(t_R + 30) - (t_R + 45)	0	100
(t_R + 45) - (t_R + 60)	85	15

t_R = retention time of ampicillin determined with reference solution (c)

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 µL of reference solutions (b) and (c) with isocratic elution at the initial mobile phase composition and 50 µL of test solution (b) according to the elution gradient described under Mobile phase; inject mobile phase A as a blank according to the elution gradient described under Mobile phase.

System suitability Reference solution (b):
- *resolution*: minimum 3.0 between the peaks due to ampicillin and cefradin; if necessary, adjust the ratio A:B of the mobile phase.

Limit:
- *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)

Maximum 20 ppm.

Water (*2.5.12*)

12.0 per cent to 15.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

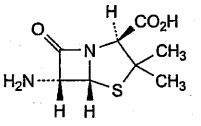
System suitability Reference solution (a):
- *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of ampicillin from the declared content of *anhydrous ampicillin CRS*.

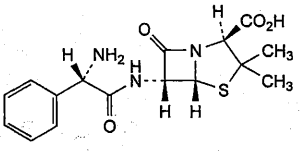
STORAGE

In an airtight container.

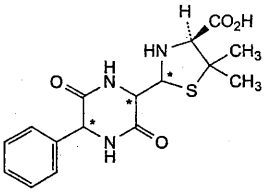
IMPURITIES



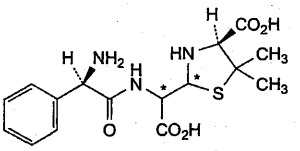
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



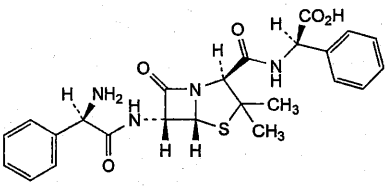
B. (2*S*,5*R*,6*R*)-6-[[(2*S*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-ampicillin),



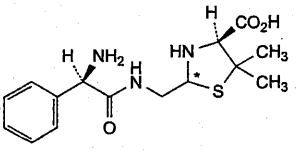
C. (4*S*)-2-(3,6-dioxo-5-phenylpiperazin-2-yl)-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (diketopiperazines of ampicillin),



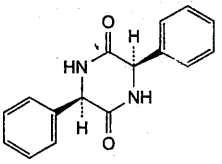
D. (4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino] carboxymethyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acids of ampicillin),



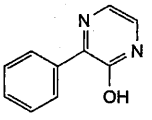
E. (2*R*)-2-[[[(2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-2-phenylacetic acid (ampicillinyl-D-phenylglycine),



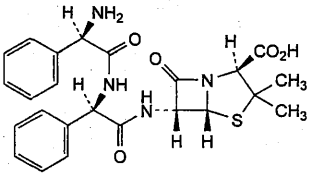
F. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl] amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penilloic acids of ampicillin),



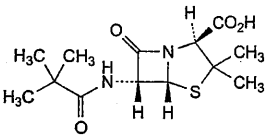
G. (3*R*,6*R*)-3,6-diphenylpiperazine-2,5-dione,



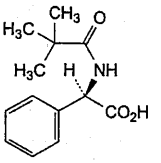
H. 3-phenylpyrazin-2-ol,



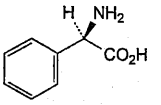
I. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (D-phenylglycylampicillin),



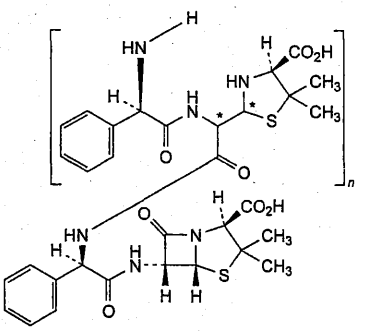
J. (2*S*,5*R*,6*R*)-6-[(2,2-dimethylpropanoyl)amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,



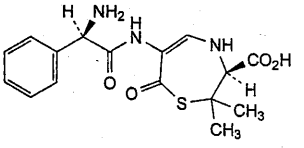
K. (2*R*)-2-[(2,2-dimethylpropanoyl)amino]-2-phenylacetic acid,



L. (2*R*)-2-amino-2-phenylacetic acid (D-phenylglycine),



M. co-oligomers of ampicillin and of penicilloic acids of ampicillin,

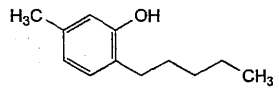


N. (3*S*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2,2-dimethyl-7-oxo-2,3,4,7-tetrahydro-1,4-thiazepine-3-carboxylic acid.

Ph Eur

Amylmetacresol

(Ph. Eur. monograph 2405)



$C_{12}H_{18}O$ 178.3 *1300-94-3*

Action and use
Antiseptic.

Ph Eur

DEFINITION
5-Methyl-2-pentylphenol.

Content
98.0 per cent to 102.0 per cent.

CHARACTERS
Appearance
Clear or almost clear liquid, or solid crystalline mass, colourless or slightly yellow when freshly prepared.
The substance changes colour during storage by darkening and/or discolouration to dark yellow, brownish-yellow or pink.

Solubility
Practically insoluble in water, very soluble in acetone and in ethanol (96 per cent).

It solidifies at about 22 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Preparation Film between 2 plates of *potassium bromide R*.

Comparison *amylmetacresol CRS*.

TESTS
Related substances
Gas chromatography (*2.2.28*): use the normalisation procedure.

Internal standard solution Dissolve 0.100 g of *butylhydroxytoluene R* in *2-propanol R* and dilute to 10.0 mL with the same solvent.

Test solution (a) Dissolve 0.1000 g of the substance to be examined in *2-propanol R* and dilute to 10.0 mL with the same solvent.

Test solution (b) To 2.0 mL of test solution (a) add 2.0 mL of the internal standard solution and dilute to 10.0 mL with *2-propanol R*.

Reference solution (a) Dissolve 10 mg of *m-cresol R* (impurity B) and 10 mg of *p-cresol R* (impurity D) in *2-propanol R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve the contents of a vial of *amylmetacresol for peak identification CRS* (containing impurities A, G and K) in 1.0 mL of *2-propanol R*.

Reference solution (c) Dissolve 0.1000 g of *amylmetacresol CRS* in *2-propanol R* and dilute to 10.0 mL with the same solvent. To 2.0 mL of this solution add 2.0 mL of the internal standard solution and dilute to 10.0 mL with *2-propanol R*.

Reference solution (d) Dilute 1.0 mL of test solution (a) to 100.0 mL with *2-propanol R*. Dilute 1.0 mL of this solution to 20.0 mL with *2-propanol R*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R*.

Linear velocity 33 cm/s.

Split ratio 1:30.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 17.5	100 → 240
	17.5 - 32.5	240
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1.0 µL of test solution (a) and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram supplied with *amylmetacresol for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, G and K.

Relative retention With reference to amylmetacresol (retention time = about 16 min): impurity G (diastereoisomer 1) = about 0.51; impurity G (diastereoisomer 2) = about 0.53; impurity D = about 0.77; impurity B = about 0.78; impurity K = about 0.95; impurity A = about 0.99.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurities D and B.

Limits:
— *impurity A*: maximum 0.6 per cent;
— *impurities G* (sum of the 2 diastereoisomers), *K*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *disregard limit*: the area of the peak due to amylmetacresol in the chromatogram obtained with reference solution (d) (0.05 per cent).

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*) as described in the test for related substances with the following modification.

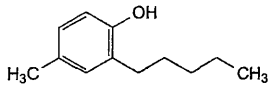
Injection 1.0 µL of test solution (b) and reference solution (c).

Calculate the percentage content of $C_{12}H_{18}O$ from the declared content of *amylmetacresol CRS*.

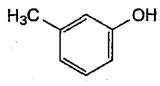
STORAGE
In an airtight, non-metallic container, protected from light.

IMPURITIES
Specified impurities *A, G, K.*

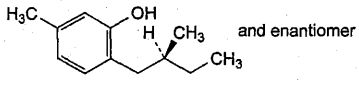
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, H, I, J.*



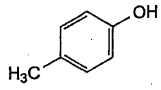
A. 4-methyl-2-pentylphenol,



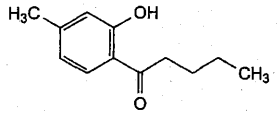
B. 3-methylphenol (*m*-cresol),



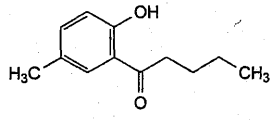
C. 5-methyl-2-[(2*RS*)-2-methylbutyl]phenol,



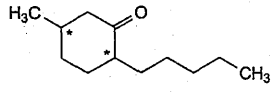
D. 4-methylphenol (*p*-cresol),



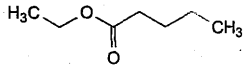
E. 1-(2-hydroxy-4-methylphenyl)pentan-1-one,



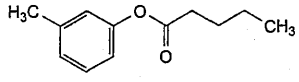
F. 1-(2-hydroxy-5-methylphenyl)pentan-1-one,



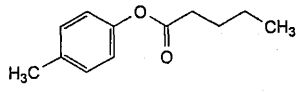
G. 5-methyl-2-pentylcyclohexanone,



H. ethyl pentanoate,



I. 3-methylphenyl pentanoate,



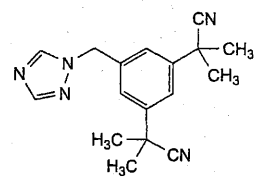
J. 4-methylphenyl pentanoate,

K. unknown structure.

Ph Eur

Anastrozole

(Ph. Eur. monograph 2406)



$C_{17}H_{19}N_5$ 293.4 *120511-73-1*

Action and use
Aromotase inhibitor; treatment of breast carcinoma.

Preparation
Anastrozole Tablets

Ph Eur

DEFINITION

2,2′-[5-(1*H*-1,2,4-Triazol-1-ylmethyl)benzene-1,3-diyl]bis(2-methylpropanenitrile).

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, freely soluble in anhydrous ethanol, practically insoluble in cyclohexane.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *anastrozole CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water for chromatography R* (50:50 *V/V*).

Test solution (a) Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 200.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.5 mg of *anastrozole impurity E CRS* in 20.0 mL of the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with test solution (a).

Reference solution (c) Dissolve 25.0 mg of *anastrozole CRS* in the solvent mixture and dilute to 200.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase: end-capped ethylene-bridged polar-embedded octadecylsilyl silica gel for chromatography (hybrid material) R* (3.5 µm).

Mobile phase:
— *mobile phase A: phosphoric acid R, water for chromatography R* (0.1:100 *V/V*);
— *mobile phase B: phosphoric acid R, acetonitrile R1* (0.1:100 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	95	5
2 - 54	95 → 35	5 → 65

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity E.

Relative retention With reference to anastrozole (retention time = about 29 min): impurity E = about 1.05.

System suitability Reference solution (b):
— *resolution*: minimum 3.5 between the peaks due to anastrozole and impurity E.

Calculation of percentage contents:
— for each impurity, use the concentration of anastrozole in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)
Maximum 0.3 per cent, determined on 50.0 mg.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

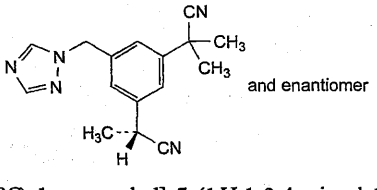
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

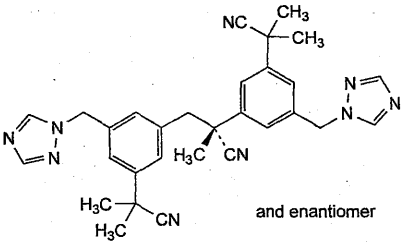
Calculate the percentage content of $C_{17}H_{19}N_5$ taking into account the assigned content of *anastrozole CRS*.

IMPURITIES

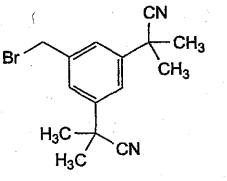
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I.*



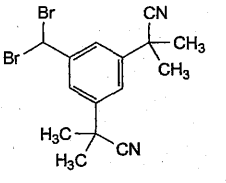
A. 2-[3-[(1*RS*)-1-cyanoethyl]-5-(1*H*-1,2,4-triazol-1-ylmethyl) phenyl]-2-methylpropanenitrile,



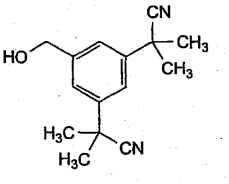
B. (2*RS*)-2,3-bis[3-(1-cyano-1-methylethyl)-5-(1*H*-1,2,4-triazol-1-ylmethyl)phenyl]-2-methylpropanenitrile,



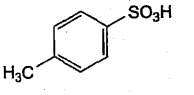
C. 2,2′-[5-(bromomethyl)benzene-1,3-diyl]bis(2-methylpropanenitrile),



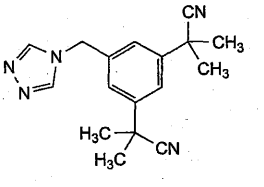
D. 2,2′-[5-(dibromomethyl)benzene-1,3-diyl]bis(2-methylpropanenitrile),



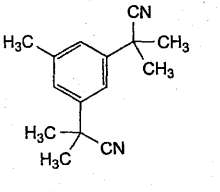
E. 2,2′-[5-(hydroxymethyl)benzene-1,3-diyl]bis(2-methylpropanenitrile),



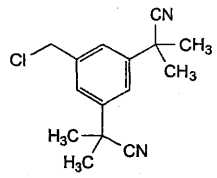
F. 4-methylbenzenesulfonic acid,



G. 2,2′-[5-(4*H*-1,2,4-triazol-4-ylmethyl)benzene-1,3-diyl]bis (2-methylpropanenitrile),



H. 2,2′-(5-methylbenzene-1,3-diyl)bis(2-methylpropanenitrile),



I. 2,2′-[5-(chloromethyl)benzene-1,3-diyl]bis(2-methylpropanenitrile).

Ph Eur

Animal Epithelia and Outgrowths for Allergen Products

(Ph. Eur. monograph 2621)

Ph Eur

DEFINITION

Animal epithelia and outgrowths for allergen products consist of hair, epithelium fragments, dander, feathers and other structures that grow from the epidermis of mammals or birds.

Animal epithelia and outgrowths may contain proteins deposited from the saliva and/or secretions from the sebaceous glands of the animal. They may be further processed (e.g. cut or washed) using qualified methods or are unprocessed.

PRODUCTION

Animal epithelia and outgrowths for allergen products are obtained from healthy animals selected to avoid possible transmissible agents of disease. The exact species and/or variety of animal is stated. Typical production steps, including animal management, source material collection and purification, are specified. The origin, quality, and traceability of the source material must be demonstrated.

It is expected that, where applicable, the animal care and husbandry follows the principles described for the protection of vertebrate animals used for experimental and other scientific purposes. A responsible veterinarian or another competent person confirms the identity of the species and that the animals are healthy. It is verified that the skin is visibly clean and intact before harvest and that the animals have not been recently treated with preparations for cutaneous application, such as antiparasitic drugs.

The collection of animal epithelia and outgrowths must be performed without injuring the skin of the animal. Confirmation that measures are in place to prevent cross-contamination by animal epithelia and outgrowths from other animals is provided, including during animal management, collection and processing. Methods involving the grinding of whole skin and/or pelts must not be used.

Where major changes to the production of the animal epithelia and outgrowths take place (e.g. when a new process or supplier is introduced), such changes are qualified.

Microbial contamination of the animal epithelia and outgrowths may be unavoidable and should be monitored on a representative number of batches of source material according to a justified sampling plan and each time a new supplier and/or a new process for the source material production is introduced; if a determination of microbial contamination is not applicable, this must be justified. Microbial contamination values and potential increases in microbial contamination are monitored during stability studies, in order to assess this aspect along with the source material characteristics upon storage.

Control methods and acceptance criteria relating to identity and purity of the animal epithelia and outgrowths are established. The acceptance criteria must ensure the consistency of the animal epithelia and outgrowths source material from a qualitative and quantitative point of view. The animal epithelia and outgrowths source material is stored under controlled conditions justified by stability data. The collection and production, as well as the handling of the source material, are such that consistent composition is ensured from batch to batch.

ANIMAL EPITHELIA AND OUTGROWTHS FOR ALLERGEN PRODUCTS REFERENCE BATCH

An appropriate reference batch is established for each animal epithelia and outgrowths source material. The nature of the reference batch depends on the testing approach to verify batch-to-batch consistency and to establish acceptable quality. The reference batch may be, for example, an internal reference preparation (if available), a source material extract or a sample of a production batch. Its characterisation must be described. The extent of characterisation of the reference batch depends on the nature of the animal epithelia and outgrowths source material, knowledge of the allergenic components and availability of suitable reagents.
The reference batch is stored under controlled conditions ensuring its stability.

BATCH-TO-BATCH CONSISTENCY

To establish batch-to-batch consistency, one or more of the following tests are performed on each batch. The choice of tests must be justified.

Total protein *(2.5.33)*

Protein profile
Determined by using suitable electrophoresis methods *(2.2.31, 2.2.54)*.

Allergen profile
Relevant allergenic components are identified by means of suitable techniques using allergen-specific antibodies.

Major allergen content
Determined by using suitable immunochemical methods *(2.7.1)* such as enzyme-linked immunosorbent assay (ELISA).

Total allergenic activity
Determined by testing inhibition of the binding capacity of specific immunoglobulin E antibodies or by a suitable equivalent *in vitro* method.

CHARACTERS

Animal epithelia and outgrowths for allergen products are supplied as coloured powders or other materials such as feathers, dander or hairs.

IDENTIFICATION

The identity of animal epithelia and outgrowths is confirmed by their relevant macroscopic and microscopic characteristics in comparison to those of a reference batch or reference documents. Identity may also be confirmed using other methods such as ELISA or by genetic identification, if performed by generally accepted methods.

TESTS

Foreign matter
Foreign matter is defined as vermin (e.g. mites and fleas), dirt, and foreign animal epithelia and outgrowths. Foreign matter is determined by appropriate tests (e.g. microscopic examination, ELISA), visual inspection and/or tactile

inspection. Foreign matter is below a predefined and justified limit.

Water (*2.5.12* or *2.5.32*) or **loss on drying** (*2.2.32*)
The water content of dried material is determined; specification limits must be supported by batch analysis and stability data.

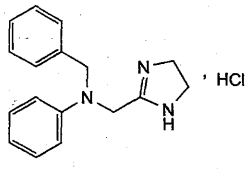
STORAGE
The source materials are stored under controlled conditions justified by stability data.

LABELLING
The label states:
— the species of the source animal;
— the nature of the animal epithelia and outgrowths.

Ph Eur

Antazoline Hydrochloride

(Ph. Eur. monograph 0972)



$C_{17}H_{20}ClN_3$ 301.8 *2508-72-7*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION
Antazoline hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of *N*-benzyl-*N*-[(4,5-dihydro-1*H*-imidazol-2-yl)methyl]aniline hydrochloride, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, sparingly soluble in water, soluble in alcohol, slightly soluble in methylene chloride.

It melts at about 240 °C, with decomposition.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *antazoline hydrochloride CRS*. Examine the substances as discs prepared using *potassium chloride R*.

B. Examine the chromatograms obtained in the test for related substances in daylight after spraying. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (b).

C. To 5 mL of solution S (see Tests) add, drop by drop, *dilute sodium hydroxide solution R* until an alkaline reaction is produced. Filter. The precipitate, washed with two quantities, each of 10 mL, of *water R* and dried in a desiccator under reduced pressure, melts (*2.2.14*) at 119 °C to 123 °C.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 2.0 g in *carbon dioxide-free water R* prepared from *distilled water R*, heating at 60 °C if necessary. Allow to cool and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.2 mL of *methyl red solution R*. Not more than 0.1 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance. Heat the plate at 110 °C for 15 min before using.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 5 mL with *methanol R*.

Reference solution (a) Dilute 0.5 mL of test solution (a) to 100 mL with *methanol R*.

Reference solution (b) Dissolve 20 mg of *antazoline hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (c) Dissolve 20 mg of *xylometazoline hydrochloride CRS* in 1 mL of test solution (a) and dilute to 5 mL with *methanol R*.

Apply to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 5 volumes of *diethylamine R*, 10 volumes of *methanol R* and 85 volumes of *ethyl acetate R*. Dry the plate in a current of warm air for 15 min. Examine in ultraviolet light at 254 nm. The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated principal spots. Spray with a mixture of equal volumes of a 200 g/L solution of *ferric chloride R* and a 5 g/L solution of *potassium ferricyanide R*. Examine immediately in daylight. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent).

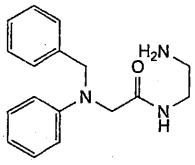
Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY
Dissolve 0.250 g in 100 mL of *alcohol R*. Add 0.1 mL of *phenolphthalein solution R1*. Titrate with *0.1 M alcoholic potassium hydroxide*.

1 mL of *0.1 M alcoholic potassium hydroxide* is equivalent to 30.18 mg of $C_{17}H_{20}ClN_3$.

IMPURITIES

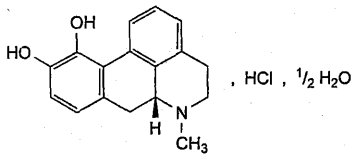


A. *N*-(2-aminoethyl)-2-(benzylphenylamino)acetamide.

Ph Eur

Apomorphine Hydrochloride Hemihydrate

(Ph. Eur. monograph 0136)



$C_{17}H_{18}ClNO_2,½H_2O$ 312.8 *41372-20-7*

Action and use
Dopamine receptor agonist; treatment of Parkinson's disease.

Preparation
Apomorphine Hydrochloride for Homoeopathic Preparations

Ph Eur

DEFINITION
(6a*R*)-6-Methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline-10,11-diol hydrochloride hemihydrate.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or slightly yellowish-brown or green-tinged greyish, crystalline powder or crystals; on exposure to air and light, the green tinge becomes more pronounced.

Solubility
Sparingly soluble in water and in ethanol (96 per cent), practically insoluble in toluene.

IDENTIFICATION
First identification: B, D.
Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 10.0 mg in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with the same acid solution. Dilute 10.0 mL of the solution to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*.

Spectral range 230-350 nm

Absorption maximum At 273 nm.

Shoulder At 300-310 nm.

Specific absorbance at the absorption maximum 530 to 570.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *apomorphine hydrochloride hemihydrate CRS.*

C. To 5 mL of solution S (see Tests) add a few millilitres of *sodium hydrogen carbonate solution R* until a permanent, white precipitate is formed. The precipitate slowly becomes greenish. Add 0.25 mL of *0.05 M iodine* and shake. The precipitate becomes greyish-green. Collect the precipitate. The precipitate dissolves in *methylene chloride R* giving a violet-blue solution and in *ethanol (96 per cent) R* giving a blue solution.

D. To 2 mL of solution S (see Tests) add 0.1 mL of *nitric acid R*. Mix and filter. The filtrate gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.25 g without heating in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 or GY_5(*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)
−52 to −48 (dried substance).

Dissolve 0.25 g in a 2.06 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same acid solution.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in a 1 per cent *V/V* solution of *glacial acetic acid R* and dilute to 20.0 mL with the same solution.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with a 1 per cent *V/V* solution of *glacial acetic acid R*. Dilute 1.0 mL of this solution to 10.0 mL with a 1 per cent *V/V* solution of *glacial acetic acid R*.

Reference solution (b) Dissolve 12.5 mg of *apomorphine impurity B CRS* in a 1 per cent *V/V* solution of *glacial acetic acid R* and dilute to 10.0 mL with the same solution.

Reference solution (c) Dilute 2.0 mL of reference solution (b) to 10.0 mL with a 1 per cent *V/V* solution of *glacial acetic acid R*. Dilute 2.0 mL of this solution to 100.0 mL with a 1 per cent *V/V* solution of *glacial acetic acid R*.

Reference solution (d) Dissolve 25 mg of *boldine R* in a 1 per cent *V/V* solution of *glacial acetic acid R* and dilute to 10.0 mL with the same solution. To 1 mL of this solution add 1 mL of the test solution and dilute to 10.0 mL with a 1 per cent *V/V* solution of *glacial acetic acid R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: 1.1 g/L solution of *sodium octanesulfonate R*, adjusted to pH 2.2 with a 50 per cent *m/m* solution of *phosphoric acid R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	85	15
2 - 32	85 → 68	15 → 32
32 - 37	68	32

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to apomorphine (retention time = about 18 min): impurity B = about 0.4; boldine = about 0.9.

System suitability Reference solution (d):

— *resolution*: minimum 2.5 between the peaks due to boldine and apomorphine.

Limits:

— *impurity B*: not more than 0.75 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: maximum 0.5 per cent;

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
2.5 per cent to 4.2 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

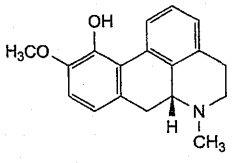
ASSAY
Dissolve 0.250 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the first 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 30.38 mg of $C_{17}H_{18}ClNO_2$.

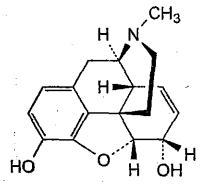
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *B.*

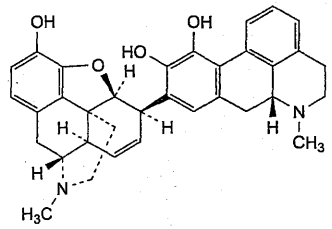
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C.*



A. (6a*R*)-10-methoxy-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinolin-11-ol (apocodeine),



B. 7,8-didehydro-4,5α-epoxy-17-methylmorphinan-3,6α-diol (morphine),

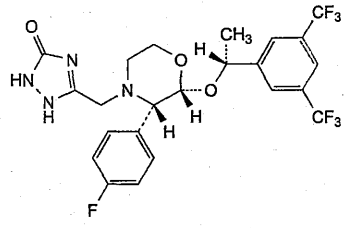


C. (6a*R*)-9-[7,8-didehydro-4,5α-epoxy-3-hydroxy-17-methylmorphinan-6α-yl]-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline-10,11-diol (morphine-apomorphine dimer).

Ph Eur

Aprepitant

(Ph. Eur. monograph 2757)



$C_{23}H_{21}F_7N_4O_3$ 534.4 *170729-80-3*

Action and use
Neurokinin-1 (NK_1) receptor antagonist; prevention of nausea and vomiting associated with emetogenic chemotherapy.

Preparation
Aprepitant Capsules

Ph Eur

DEFINITION
5-[[(2*R*,3*S*)-2-[(1*R*)-1-[3,5-Bis(trifluoromethyl)phenyl]ethoxy]-3-(4-fluorophenyl)morpholin-4-yl]methyl]-1,2-dihydro-3*H*-1,2,4-triazol-3-one.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, sparingly soluble in anhydrous ethanol, practically insoluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *aprepitant CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)

+ 66.0 to + 70.0 (anhydrous substance), measured at 25 °C.

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Solvent mixture *acetonitrile R1, water for chromatography R* (50:50 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 40.0 mg of *aprepitant CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 4 mg of *aprepitant for system suitability CRS* (containing impurity A) in 5.0 mL of the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: 0.1 per cent *V/V* solution of *phosphoric acid R*;
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	65	35
2 - 22	65 → 20	35 → 80
22 - 32	20	80

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *aprepitant for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to aprepitant (retention time = about 15 min): impurity A = about 0.97.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to impurity A and aprepitant.

Limits:
- *impurity A*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.2 per cent;
- *reporting threshold*: 0.05 per cent (reference solution (b)).

Water (*2.5.32*)

Maximum 0.2 per cent, determined on 0.200 g by direct sample introduction.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

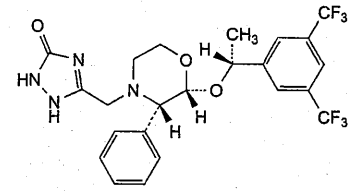
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{23}H_{21}F_7N_4O_3$ taking into account the assigned content of *aprepitant CRS*.

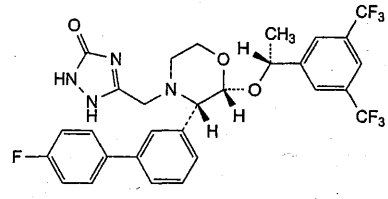
IMPURITIES

Specified impurities *A.*

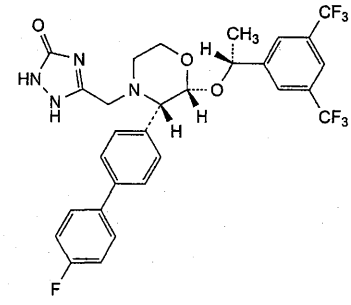
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C.*



A. 5-[[(2*R*,3*S*)-2-[(1*R*)-1-[3,5-bis(trifluoromethyl) phenyl]ethoxy]-3-phenylmorpholin-4-yl]methyl]-1,2-dihydro-3*H*-1,2,4-triazol-3-one,



B. 5-[[(2*R*,3*S*)-2-[(1*R*)-1-[3,5-bis(trifluoromethyl) phenyl]ethoxy]-3-(4′-fluorobiphenyl-3-yl)morpholin-4-yl]methyl]-1,2-dihydro-3*H*-1,2,4-triazol-3-one,

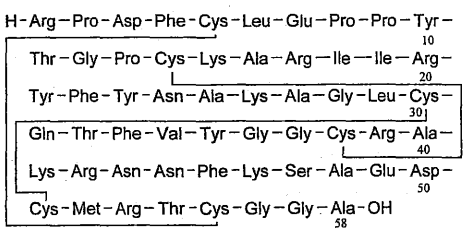


C. 5-[[(2*R*,3*S*)-2-[(1*R*)-1-[3,5-bis(trifluoromethyl) phenyl]ethoxy]-3-(4′-fluorobiphenyl-4-yl)morpholin-4-yl]methyl]-1,2-dihydro-3*H*-1,2,4-triazol-3-one.

Ph Eur

Aprotinin

(Ph. Eur. monograph 0580)



$C_{284}H_{432}N_{84}O_{79}S_7$ 6511 *9087-70-1*

Action and use
Antifibrinolytic.

Ph Eur

DEFINITION

Aprotinin is a polypeptide consisting of a chain of 58 amino acids. It inhibits stoichiometrically the activity of several proteolytic enzymes such as chymotrypsin, kallikrein, plasmin and trypsin. It contains not less than 3.0 Ph. Eur. U. of aprotinin activity per milligram, calculated with reference to the dried substance.

PRODUCTION

The animals from which aprotinin is derived must fulfil the requirements for the health of animals suitable for human consumption.

The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Histamine *(2.6.10)*
Maximum 0.2 µg of histamine base per 3 Ph. Eur. U.

CHARACTERS

Appearance
Almost white hygroscopic powder.

Solubility
Soluble in water and in isotonic solutions, practically insoluble in organic solvents.

IDENTIFICATION

A. Thin-layer chromatography *(2.2.27)*.

Test solution Solution S (see Tests).

Reference solution Dilute *aprotinin solution BRP* in *water R* to obtain a concentration of 15 Ph. Eur. U./mL.

Plate *TLC silica gel G plate R.*

Mobile phase *water R, glacial acetic acid R* (80:100 *V/V*) containing 100 g/L of *sodium acetate R.*

Application 10 µL.

Development Over a path of 12 cm.

Drying In air.

Detection Spray with a solution of 0.1 g of *ninhydrin R* in a mixture of 6 mL of a 10 g/L solution of *cupric chloride R*, 21 mL of *glacial acetic acid R* and 70 mL of *anhydrous ethanol R*. Dry the plate at 60 °C.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Determine the ability of the substance to be examined to inhibit trypsin activity using the method described below.

Test solution Dilute 1 mL of solution S to 50 mL with *buffer solution pH 7.2 R.*

Trypsin solution Dissolve 10 mg of *trypsin BRP* in *0.002 M hydrochloric acid* and dilute to 100 mL with the same acid.

Casein solution Dissolve 0.2 g of *casein R* in *buffer solution pH 7.2 R* and dilute to 100 mL with the same buffer solution.

Precipitating solution *glacial acetic acid R, water R, anhydrous ethanol R* (1:49:50 *V/V/V*).

Mix 1 mL of the test solution with 1 mL of the trypsin solution. Allow to stand for 10 min and add 1 mL of the casein solution. Incubate at 35 °C for 30 min. Cool in iced water and add 0.5 mL of the precipitating solution. Shake and allow to stand at room temperature for 15 min.
The solution is cloudy. Carry out a blank test under the same conditions using *buffer solution pH 7.2 R* instead of the test solution. The solution is not cloudy.

TESTS

Solution S
Prepare a solution of the substance to be examined containing 15 Ph. Eur. U./mL, calculated from the activity stated on the label.

Appearance of solution
Solution S is clear *(2.2.1)*.

Absorbance *(2.2.25)*
Maximum 0.80 by measuring at the absorption maximum at 277 nm.

Prepare a solution of the substance to be examined containing 3.0 Ph. Eur. U./mL.

Des-Ala-aprotinin and des-Ala-des-Gly-aprotinin
Capillary zone electrophoresis *(2.2.47)*: use the normalisation procedure.

Test solution Prepare a solution of the substance to be examined in *water R* containing not less than 1 Ph. Eur. U./mL.

Reference solution Dilute *aprotinin solution BRP* in *water R* to obtain the same concentration as the test solution.

Capillary:
— *material*: uncoated fused silica;
— *size*: effective length = 45-60 cm, Ø = 75 µm.

Temperature 25 °C.

CZE buffer Dissolve 8.21 g of *potassium dihydrogen phosphate R* in 400 mL of *water R*, adjust to pH 3.0 with *phosphoric acid R*, dilute to 500.0 mL with *water R* and filter through a membrane filter (nominal pore size 0.45 µm).

Detection Spectrophotometer at 214 nm.

Between-run rinsing Rinse the capillary for at least 1 min with *0.1 M sodium hydroxide* filtered through a membrane filter (nominal pore size 0.45 µm) and for 2 min with the CZE buffer.

Injection Under pressure or vacuum (for example, 3 s at a differential pressure of 3.5 kPa).

Migration Apply a field strength of 0.2 kV/cm, using the CZE buffer as the electrolyte in both buffer reservoirs.

Run time 30 min.

Identification of impurities Use the electropherogram supplied with *aprotinin solution BRP* and the electropherogram obtained with the reference solution to identify the peaks due to impurities A and B.

Relative migration With reference to aprotinin (migration time = about 22 min): impurity A = about 0.98; impurity B = about 0.99.

System suitability Reference solution after at least 6 injections:
— *migration time*: aprotinin = 19.0 min to 25.0 min;
— *resolution*: minimum 0.8 between the peaks due to impurities A and B; minimum 0.5 between the peaks due to impurity B and aprotinin;
— *peak distribution*: the electropherogram obtained is qualitatively and quantitatively similar to the electropherogram supplied with *aprotinin solution BRP*;
— *height of the principal peak*: at least 1000 times the height of the baseline noise. If necessary, adjust the sample load to give peaks of sufficient height.

Limits:
— *impurity A*: maximum 8.0 per cent;
— *impurity B*: maximum 7.5 per cent.

Pyroglutamyl-aprotinin and related compounds
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Prepare a solution of the substance to be examined in mobile phase A, containing about 5 Ph. Eur. U./mL.

Reference solution Dissolve the contents of a vial of *aprotinin for system suitability CRS* in 2.0 mL of mobile phase A.

Column:
— *size*: *l* = 0.075 m, Ø = 7.5 mm;
— *stationary phase*: *strong cation-exchange silica gel for chromatography R* (10 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.52 g of *potassium dihydrogen phosphate R* and 7.26 g of *disodium hydrogen phosphate dihydrate R* in 1000 mL of *water for chromatography R*; filter and degas;
— *mobile phase B*: dissolve 3.52 g of *potassium dihydrogen phosphate R*, 7.26 g of *disodium hydrogen phosphate dihydrate R* and 66.07 g of *ammonium sulfate R* in 1000 mL of *water for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 21	92 → 64	8 → 36
21 - 30	64 → 0	36 → 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 40 µL.

Relative retention With reference to aprotinin (retention time = 17.0 min to 20.0 min): impurity C = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to impurity C and aprotinin;
— *symmetry factor*: maximum 1.3 for the peak due to aprotinin.

Limits:
— *impurity C*: maximum 1.0 per cent;
— *any other impurity*: maximum 0.5 per cent;
— *sum of impurities other than C*: maximum 1.0 per cent.

Aprotinin oligomers
Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Prepare a solution of the substance to be examined in *water R* containing about 5 Ph. Eur. U./mL.

Reference solution Treat the substance to be examined to obtain about 2 per cent aprotinin oligomers. For example, heat freeze-dried aprotinin at about 110 °C for about 4 h. Then dissolve in *water R* to obtain a concentration of about 5 Ph. Eur. U./mL.

Column 3 columns coupled in series:
— *size*: *l* = 0.30 m, Ø = 7.8 mm;
— *stationary phase*: *hydrophilic silica gel for chromatography R* of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 20 000 to 10 000 000 (8 µm).

Mobile phase *acetonitrile R, glacial acetic acid R, water for chromatography R* (2:2:6 *V/V/V*); filter and degas.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 277 nm.

Injection 100 µL.

Run time 40 min.

Relative retention With reference to aprotinin monomer (retention time = 24.5 min to 25.5 min): aprotinin dimer = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 1.3 between the peaks due to aprotinin dimer and monomer;
— *symmetry factor*: maximum 2.5 for the peak due to aprotinin monomer.

Limit:
— *total*: maximum 1.0 per cent.

Loss on drying (*2.2.32*)
Maximum 6.0 per cent, determined on 0.100 g by drying *in vacuo*.

Bacterial endotoxins (*2.6.14*)
Less than 0.14 IU per European Pharmacopoeia Unit of aprotinin, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

The activity of aprotinin is determined by measuring its inhibitory action on a solution of trypsin of known activity. The inhibiting activity of the aprotinin is calculated from the difference between the initial activity and the residual activity of the trypsin.

The inhibiting activity of aprotinin is expressed in European Pharmacopoeia Units. 1 Ph. Eur. U. inhibits 50 per cent of the enzymatic activity of 2 microkatals of trypsin.

Use a reaction vessel with a capacity of about 30 mL, provided with:
— a device that will maintain a temperature of 25 ± 0.1 °C;
— a stirring device, such as a magnetic stirrer;
— a lid with 5 holes for accommodating the electrodes, the tip of a burette, a tube for the admission of nitrogen and the introduction of the reagents.

An automatic or manual titration apparatus may be used. In the latter case the burette is graduated in 0.05 mL and the pH-meter is provided with a wide reading scale and glass-silver-silver chloride or other suitable electrodes.

Test solution Prepare a solution of the substance to be examined in *0.0015 M borate buffer solution pH 8.0 R* expected to contain 1.67 Ph. Eur. U./mL (about 0.6 mg (*m* mg) per millilitre).

Trypsin solution Prepare a solution of *trypsin BRP* containing about 0.8 microkatals per millilitre (about 1 mg/mL), using *0.001 M hydrochloric acid* as the solvent. Use a freshly prepared solution and keep in iced water.

Trypsin and aprotinin solution To 4.0 mL of the trypsin solution add 1.0 mL of the test solution. Dilute immediately to 40.0 mL with *0.0015 M borate buffer solution pH 8.0 R*. Allow to stand at room temperature for 10 min and then keep in iced water. Use within 6 h of preparation.

Dilute trypsin solution Dilute 0.5 mL of the trypsin solution to 10.0 mL with *0.0015 M borate buffer solution pH 8.0 R*. Allow to stand at room temperature for 10 min and then keep in iced water.

Maintain an atmosphere of nitrogen in the reaction flask and stir continuously; introduce 9.0 mL of *0.0015 M borate buffer solution pH 8.0 R* and 1.0 mL of a freshly prepared 6.9 g/L solution of *benzoylarginine ethyl ester hydrochloride R*. Adjust to pH 8.0 with *0.1 M sodium hydroxide*. When the temperature has reached equilibrium at 25 ± 0.1 °C, add 1.0 mL of the trypsin and aprotinin solution and start a timer. Maintain at pH 8.0 by the addition of *0.1 M sodium hydroxide* and note the volume added every 30 s. Continue the reaction for 6 min. Determine the number of millilitres of *0.1 M sodium hydroxide* used per second (n_1 mL). Carry out, under the same conditions, a titration using 1.0 mL of the dilute trypsin solution. Determine the number of millilitres of *0.1 M sodium hydroxide* used per second (n_2 mL).

Calculate the aprotinin activity in European Pharmacopoeia Units per milligram using the following expression:

$$\frac{4000(2n_2 - n_1)}{m}$$

The estimated activity is not less than 90 per cent and not more than 110 per cent of the activity stated on the label.

STORAGE

In an airtight, tamper-proof container, protected from light.

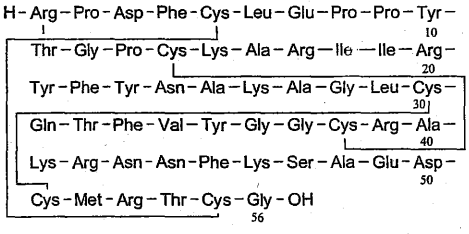
LABELLING

The label states:

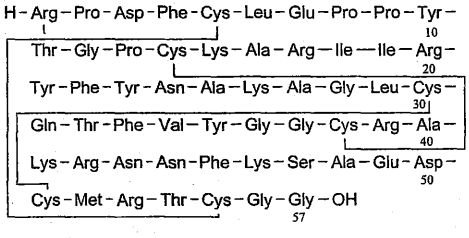
— the number of European Pharmacopoeia Units of aprotinin activity per milligram;

— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

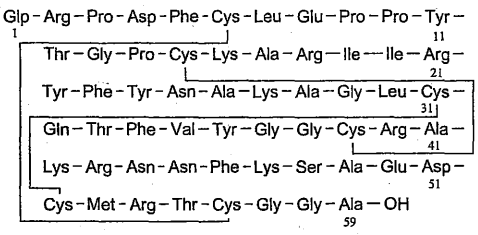
IMPURITIES



A. aprotinin-(1-56)-peptide,



B. aprotinin-(1-57)-peptide,



C. (5-oxoprolyl)aprotinin (pyroglutamylaprotinin).

Ph Eur

Aprotinin Concentrated Solution

(Ph. Eur. monograph 0579)

$C_{284}H_{432}N_{84}O_{79}S_7$ 6511

Action and use
Antifibrinolytic.

Ph Eur

DEFINITION

Aprotinin concentrated solution is a solution of aprotinin, a polypeptide consisting of a chain of 58 amino acids, which inhibits stoichiometrically the activity of several proteolytic enzymes such as chymotrypsin, kallikrein, plasmin and trypsin. It contains not less than 15.0 Ph. Eur. U. of aprotinin activity per millilitre.

PRODUCTION

The animals from which aprotinin is derived must fulfil the requirements for the health of animals suitable for human consumption.

The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Histamine *(2.6.10)*
Maximum 0.2 µg of histamine base per 3 Ph. Eur. U.

CHARACTERS

Appearance
Clear, colourless liquid.

IDENTIFICATION

A. Thin-layer chromatography *(2.2.27)*.

Test solution Solution S (see Tests).

Reference solution Dilute *aprotinin solution BRP* in *water R* to obtain a concentration of 15 Ph. Eur. U./mL.

Plate *TLC silica gel G plate R.*

Mobile phase *water R, glacial acetic acid R* (80:100 *V/V*) containing 100 g/L of *sodium acetate R.*

Application 10 µL.

Development Over a path of 12 cm.

Drying In air.

Detection Spray with a solution of 0.1 g of *ninhydrin R* in a mixture of 6 mL of a 10 g/L solution of *cupric chloride R*, 21 mL of *glacial acetic acid R* and 70 mL of *anhydrous ethanol R*. Dry the plate at 60 °C.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Determine the ability of the preparation to be examined to inhibit trypsin activity using the method described below.

Test solution Dilute 1 mL of solution S to 50 mL with *buffer solution pH 7.2 R*.

Trypsin solution Dissolve 10 mg of *trypsin BRP* in *0.002 M hydrochloric acid* and dilute to 100 mL with the same acid.

Casein solution Dissolve 0.2 g of *casein R* in *buffer solution pH 7.2 R* and dilute to 100 mL with the same buffer solution.

Precipitating solution *glacial acetic acid R, water R, anhydrous ethanol R* (1:49:50 *V/V/V*).

Mix 1 mL of the test solution with 1 mL of the trypsin solution. Allow to stand for 10 min and add 1 mL of the casein solution. Incubate at 35 °C for 30 min. Cool in iced water and add 0.5 mL of the precipitating solution. Shake and allow to stand at room temperature for 15 min.
The solution is cloudy. Carry out a blank test under the same conditions using *buffer solution pH 7.2 R* instead of the test solution. The solution is not cloudy.

TESTS

Solution S

Prepare a solution containing 15 Ph. Eur. U./mL, if necessary by dilution, on the basis of the activity stated on the label.

Appearance of solution

Solution S is clear (*2.2.1*).

Absorbance (*2.2.25*)

Maximum 0.80 by measuring at the absorption maximum at 277 nm.

Prepare a solution containing 3.0 Ph. Eur. U./mL.

Des-Ala-aprotinin and des-Ala-des-Gly-aprotinin

Capillary zone electrophoresis (*2.2.47*): use the normalisation procedure.

Test solution Dilute the preparation to be examined in *water R* to obtain a concentration of not less than 1 Ph Eur. U./mL.

Reference solution Dilute *aprotinin solution BRP* in *water R* to obtain the same concentration as the test solution.

Capillary:
— *material*: uncoated fused silica;
— *size*: effective length = 45-60 cm, Ø = 75 µm.

Temperature 25 °C.

CZE buffer Dissolve 8.21 g of *potassium dihydrogen phosphate R* in 400 mL of *water R*, adjust to pH 3.0 with *phosphoric acid R*, dilute to 500.0 mL with *water R* and filter through a membrane filter (nominal pore size 0.45 µm).

Detection Spectrophotometer at 214 nm.

Between-run rinsing Rinse the capillary for at least 1 min with *0.1 M sodium hydroxide* filtered through a membrane filter (nominal pore size 0.45 µm) and for 2 min with the CZE buffer.

Injection Under pressure or vacuum (for example, 3 s at a differential pressure of 3.5 kPa).

Migration Apply a field strength of 0.2 kV/cm, using the CZE buffer as the electrolyte in both buffer reservoirs.

Run time 30 min.

Identification of impurities Use the electropherogram supplied with *aprotinin solution BRP* and the electropherogram obtained with the reference solution to identify the peaks due to impurities A and B.

Relative migration With reference to aprotinin (migration time = about 22 min): impurity A = about 0.98; impurity B = about 0.99.

System suitability Reference solution after at least 6 injections:
— *migration time*: aprotinin = 19.0 min to 25.0 min;
— *resolution*: minimum 0.8 between the peaks due to impurities A and B; minimum 0.5 between the peaks due to impurity B and aprotinin;
— *peak distribution*: the electropherogram obtained is qualitatively and quantitatively similar to the electropherogram supplied with *aprotinin solution BRP*;
— *height of the principal peak*: at least 1000 times the height of the baseline noise. If necessary, adjust the sample load to give peaks of a sufficient height.

Limits:
— *impurity A*: maximum 8.0 per cent;
— *impurity B*: maximum 7.5 per cent.

Pyroglutamyl-aprotinin and related compounds

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dilute the preparation to be examined in mobile phase A to a concentration of about 5 Ph. Eur. U./mL.

Reference solution Dissolve the contents of a vial of *aprotinin for system suitability CRS* in 2.0 mL of mobile phase A.

Column:
— *size*: *l* = 0.075 m, Ø = 7.5 mm;
— *stationary phase*: *strong cation-exchange silica gel for chromatography R* (10 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.52 g of *potassium dihydrogen phosphate R* and 7.26 g of *disodium hydrogen phosphate dihydrate R* in 1000 mL of *water for chromatography R*; filter and degas;
— *mobile phase B*: dissolve 3.52 g of *potassium dihydrogen phosphate R*, 7.26 g of *disodium hydrogen phosphate dihydrate R* and 66.07 g of *ammonium sulfate R* in 1000 mL of *water for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 21	92 → 64	8 → 36
21 - 30	64 → 0	36 → 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 40 µL.

Relative retention With reference to aprotinin (retention time = 17.0 min to 20.0 min): impurity C = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to impurity C and aprotinin;
— *symmetry factor*: maximum 1.3 for the peak due to aprotinin.

Limits:
— *impurity C*: maximum 1.0 per cent;
— *any other impurity*: maximum 0.5 per cent;
— *sum of impurities other than C*: maximum 1.0 per cent.

Aprotinin oligomers
Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Dilute the preparation to be examined in *water R* to obtain a concentration of about 5 Ph. Eur. U./mL.

Reference solution Treat the substance to be examined to obtain about 2 per cent aprotinin oligomers. For example, heat freeze-dried aprotinin at about 110 °C for about 4 h. Then dissolve in *water R* to obtain a concentration of about 5 Ph. Eur. U./mL.

Column 3 columns coupled in series:
— *size*: l = 0.30 m, Ø = 7.8 mm;
— *stationary phase*: *hydrophilic silica gel for chromatography R* of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 20 000 to 10 000 000 (8 µm).

Mobile phase *acetonitrile R, glacial acetic acid R, water for chromatography R* (2:2:6 *V/V/V*); filter and degas.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 277 nm.

Injection 100 µL.

Run time 40 min.

Relative retention With reference to aprotinin monomer (retention time = 24.5 min to 25.5 min): aprotinin dimer = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 1.3 between the peaks due to aprotinin dimer and monomer;
— *symmetry factor*: maximum 2.5 for the peak due to aprotinin monomer.

Limit:
— *total*: maximum 1.0 per cent.

Specific activity of the dry residue
Minimum 3.0 Ph. Eur. U. of aprotinin activity per milligram of dry residue.

Evaporate 25.0 mL to dryness in a water-bath, dry the residue at 110 °C for 15 h and weigh. From the mass of the residue and the activity determined as described below, calculate the number of European Pharmacopoeia Units per milligram of dry residue.

Bacterial endotoxins (*2.6.14*)
Less than 0.14 IU per European Pharmacopoeia Unit of aprotinin, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
The activity of aprotinin is determined by measuring its inhibitory action on a solution of trypsin of known activity. The inhibiting activity of the aprotinin is calculated from the difference between the initial activity and the residual activity of the trypsin.

The inhibiting activity of aprotinin is expressed in European Pharmacopoeia Units. 1 Ph. Eur. U. inhibits 50 per cent of the enzymatic activity of 2 microkatals of trypsin.

Use a reaction vessel with a capacity of about 30 mL, provided with:
— a device that will maintain a temperature of 25 ± 0.1 °C;
— a stirring device, such as a magnetic stirrer;
— a lid with 5 holes for accommodating the electrodes, the tip of a burette, a tube for the admission of nitrogen and the introduction of the reagents.

An automatic or manual titration apparatus may be used. In the latter case the burette is graduated in 0.05 mL and the pH-meter is provided with a wide reading scale and glass-silver-silver chloride or other suitable electrodes.

Test solution With *0.0015 M borate buffer solution pH 8.0 R* prepare an appropriate dilution (D) of the aprotinin concentrated solution expected, on the basis of the stated potency, to contain 1.67 Ph. Eur. U./mL.

Trypsin solution Prepare a solution of *trypsin BRP* containing about 0.8 microkatals per millilitre (about 1 mg/mL), using *0.001 M hydrochloric acid* as the solvent. Use a freshly prepared solution and keep in iced water.

Trypsin and aprotinin solution To 4.0 mL of the trypsin solution add 1.0 mL of the test solution. Dilute immediately to 40.0 mL with *0.0015 M borate buffer solution pH 8.0 R*. Allow to stand at room temperature for 10 min and then keep in iced water. Use within 6 h of preparation.

Dilute trypsin solution Dilute 0.5 mL of the trypsin solution to 10.0 mL with *0.0015 M borate buffer solution pH 8.0 R*. Allow to stand at room temperature for 10 min and then keep in iced water.

Maintain an atmosphere of nitrogen in the reaction flask and stir continuously; introduce 9.0 mL of *0.0015 M borate buffer solution pH 8.0 R* and 1.0 mL of a freshly prepared 6.9 g/L solution of *benzoylarginine ethyl ester hydrochloride R*. Adjust to pH 8.0 with *0.1 M sodium hydroxide*. When the temperature has reached equilibrium at 25 ± 0.1 °C, add 1.0 mL of the trypsin and aprotinin solution and start a timer. Maintain at pH 8.0 by the addition of *0.1 M sodium hydroxide* and note the volume added every 30 s. Continue the reaction for 6 min. Determine the number of millilitres of *0.1 M sodium hydroxide* used per second (n_1 mL). Carry out, under the same conditions, a titration using 1.0 mL of the dilute trypsin solution. Determine the number of millilitres of *0.1 M sodium hydroxide* used per second (n_2 mL).

Calculate the aprotinin activity in European Pharmacopoeia Units per millilitre using the following expression:

$$4000(2n_2 - n_1) \times D$$

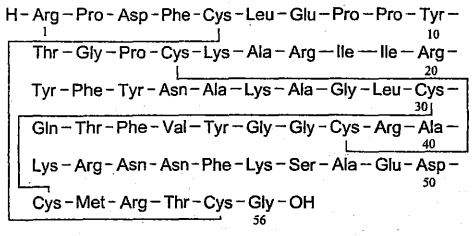
D = dilution factor of the aprotinin concentrated solution to be examined in order to obtain a solution containing 1.67 Ph. Eur. U./mL.

The estimated activity is not less than 90 per cent and not more than 110 per cent of the activity stated on the label.

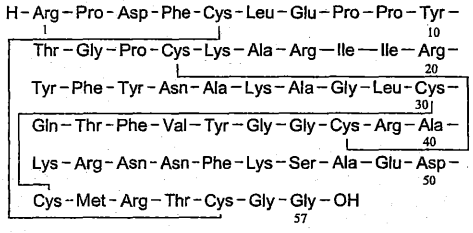
STORAGE
In an airtight, tamper-proof container, protected from light.

LABELLING
The label states:
— the number of European Pharmacopoeia Units of aprotinin activity per millilitre;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

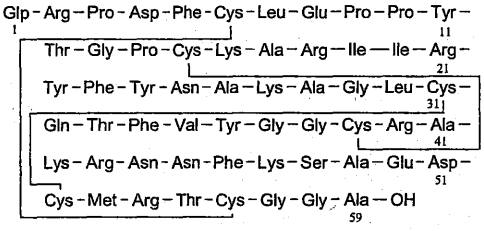
IMPURITIES



A. aprotinin-(1-56)-peptide,



B. aprotinin-(1-57)-peptide,



C. (5-oxoprolyl)aprotinin (pyroglutamylaprotinin).

Ph Eur

Arachis Oil

Peanut Oil

(Refined Arachis Oil, Ph. Eur. monograph 0263)

Preparation

Arachis Oil Enema

Ph Eur

DEFINITION

The refined fatty oil obtained from the shelled seeds of *Arachis hypogaea* L. A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Very slightly soluble in ethanol (96 per cent), miscible with light petroleum.

Relative density

About 0.915.

It solidifies at about 2 °C.

IDENTIFICATION

First identification: B.

Second identification: A.

A. Identification of fatty oils by thin-layer chromatography (*2.3.2*).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

B. Composition of fatty acids (see Tests).

TESTS

Acid value (*2.5.1*)

Maximum 0.5, determined on 10.0 g.

Peroxide value (*2.5.5, Method A*)

Maximum 5.0.

Unsaponifiable matter (*2.5.7*)

Maximum 1.0 per cent, determined on 5.0 g.

Alkaline impurities (*2.4.19*)

It complies with the test.

Composition of fatty acids (*2.4.22, Method A*)

Use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty-acid fraction of the oil:

- *saturated fatty acids of chain length less than C_{16}*: maximum 0.4 per cent;
- *palmitic acid*: 5.0 per cent to 14.0 per cent;
- *stearic acid*: 1.3 per cent to 6.5 per cent;
- *oleic acid*: 35.0 per cent to 76.0 per cent;
- *linoleic acid*: 8.0 per cent to 43.0 per cent;
- *linolenic acid*: maximum 0.6 per cent;
- *arachidic acid*: 0.5 per cent to 3.0 per cent;
- *eicosenoic acid*: 0.5 per cent to 3.0 per cent;
- *behenic acid*: 1.0 per cent to 5.0 per cent;
- *erucic acid*: maximum 0.5 per cent;
- *lignoceric acid*: 0.5 per cent to 3.0 per cent.

Water (*2.5.32*)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

In a well-filled container, protected from light.

Ph Eur

Hydrogenated Arachis Oil

Hydrogenated Peanut Oil

(Ph. Eur. monograph 1171)

Ph Eur

DEFINITION

Oil obtained by refining, bleaching, hydrogenating and deodorising oil obtained from the shelled seeds of *Arachis hypogaea* L. Each type of hydrogenated arachis oil is characterised by its nominal drop point.

CHARACTERS

Appearance

White or faintly yellowish, soft mass which melts to a clear, pale yellow liquid when heated.

Solubility

Practically insoluble in water, freely soluble in methylene chloride and in light petroleum (bp: 65-70 °C), very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, C.

Second identification: A, B.

A. Drop point (see Tests).

B. Identification of fatty oils by thin-layer chromatography (*2.3.2*).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

C. Composition of fatty acids (see Tests).

TESTS

Drop point (*2.2.17*)
32 °C to 43 °C, and within 3 °C of the nominal value.

Acid value (*2.5.1*)
Maximum 0.5.

Dissolve 10.0 g in 50 mL of the prescribed solvent by heating on a water-bath.

Peroxide value (*2.5.5, Method A*)
Maximum 5.0.

Dissolve 5.0 g in 30 mL of the prescribed solvent by heating on a water-bath.

Unsaponifiable matter (*2.5.7*)
Maximum 1.0 per cent.

Alkaline impurities (*2.4.19*)
It complies with the test.

Composition of fatty acids (*2.4.22, Method A*)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.25 mm;
— *stationary phase*: *poly(cyanopropyl)siloxane R* (film thickness 0.2 µm).

Carrier gas *helium for chromatography R.*

Flow rate 0.7 mL/min.

Split ratio 1:100.

Temperature:
— *column*: 180 °C for 20 min;
— *injection port and detector*: 250 °C.

Detection Flame ionisation.

Composition of the fatty-acid fraction of the oil:
— *saturated fatty acids of chain length less than C_{14}*: maximum 0.5 per cent;
— *myristic acid*: maximum 0.5 per cent;
— *palmitic acid*: 7.0 per cent to 16.0 per cent;
— *stearic acid*: 3.0 per cent to 19.0 per cent;
— *oleic acid and isomers*: 54.0 per cent to 78.0 per cent;
— *linoleic acid and isomers*: maximum 10.0 per cent;
— *arachidic acid*: 1.0 per cent to 3.0 per cent;
— *eicosenoic acids*: maximum 2.1 per cent;
— *behenic acid*: 1.0 per cent to 5.0 per cent;
— *erucic acid and isomers*: maximum 0.5 per cent;
— *lignoceric acid*: 0.5 per cent to 3.0 per cent.

Nickel
Maximum 1 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Into a platinum or silica crucible previously tared after ignition introduce 5.0 g. Cautiously heat and introduce into the substance a wick formed from twisted ashless filter paper. Ignite the wick. When the substance has ignited stop heating. After combustion, ignite in a muffle furnace at about 600 ± 50 °C. Continue ignition until white ash is obtained. After cooling, take up the residue with 2 quantities, each of 2 mL, of *dilute hydrochloric acid R* and transfer into a 25 mL graduated flask. Add 0.3 mL of *nitric acid R* and dilute to 25.0 mL with *water R*.

Reference solutions Prepare 3 reference solutions by adding 1.0 mL, 2.0 mL and 4.0 mL of *nickel standard solution (0.2 ppm Ni) R* to 2.0 mL of the test solution and diluting to 10.0 mL with *water R*.

Source Nickel hollow-cathode lamp.

Wavelength 232 nm.

Atomisation device Graphite furnace.

Carrier gas *argon R.*

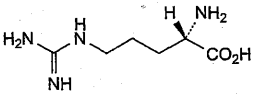
STORAGE
Protected from light.

LABELLING
The label states the nominal drop point.

Ph Eur

Arginine

(Ph. Eur. monograph 0806)



$C_6H_{14}N_4O_2$ 174.2 *74-79-3*

Action and use
Amino acid; nutrient.

Ph Eur

DEFINITION
(2*S*)-2-Amino-5-guanidinopentanoic acid.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, hygroscopic.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, C.

Second identification: A, B, D, E.

A. Specific optical rotation (see Tests).

B. Solution S (see Tests) is strongly alkaline (*2.2.4*).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *arginine CRS.*

If the spectra obtained show differences, dry the substance to be examined and the reference substance in an oven at 105 °C and record new spectra.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 50 mL with the same solution.

Reference solution Dissolve 10 mg of *arginine CRS* in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 50 mL with the same solution.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, 2-propanol R* (30:70 *V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 105 °C until the ammonia disappears completely.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

E. Dissolve about 25 mg in 2 mL of *water R*. Add 1 mL of *α-naphthol solution R* and 2 mL of a mixture of equal volumes of *strong sodium hypochlorite solution R* and *water R*. A red colour develops.

TESTS

Solution S

Dissolve 2.5 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 25.5 to + 28.5 (dried substance).

Dissolve 2.00 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances

Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *water R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
- *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
- for any ninhydrin-positive substance detected at 570 nm, use the concentration of arginine in reference solution (a);
- for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
- *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
- *total*: maximum 0.5 per cent;
- *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides (*2.4.4*)

Maximum 200 ppm.

To 5 mL of solution S add 0.5 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 300 ppm.

To 10 mL of solution S, add 1.7 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Ammonium

Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
- *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 50 mL of *water R*. Titrate with *0.1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M hydrochloric acid* is equivalent to 17.42 mg of $C_6H_{14}N_4O_2$.

STORAGE

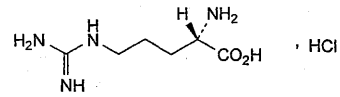
In an airtight container, protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*

Arginine Hydrochloride

(Ph. Eur. monograph 0805)



$C_6H_{15}ClN_4O_2$ 210.7 *1119-34-2*

Action and use
Amino acid; nutrient.

Preparations
Arginine Hydrochloride Infusion
Arginine Hydrochloride Oral Suspension
Arginine Hydrochloride Sterile Concentrate

Ph Eur

DEFINITION

(2*S*)-2-Amino-5-guanidinopentanoic acid hydrochloride.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, E.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *arginine hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of *arginine hydrochloride CRS* in *water R* and dilute to 50 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, 2-propanol R* (30:70 *V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 105 °C until the ammonia disappears completely.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 25 mg in 2 mL of *water R*. Add 1 mL of *α-naphthol solution R* and 2 mL of a mixture of equal volumes of *strong sodium hypochlorite solution R* and *water R*. A red colour develops.

E. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 2.5 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
+ 21.0 to + 23.5 (dried substance).

Dissolve 2.00 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances
Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *water R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of arginine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Flow rate 70 mL/min.

Temperature:
— *column*: 80 °C;
— *detector*: 40 °C.

Detection Discharge ionisation.

Injection 1 mL.

Sample rate 100 mL/min.

Relative retention With reference to impurity C (retention time = about 4.7 min): impurity A = about 0.4; impurity B = about 0.7.

System suitability Reference gas:
— *resolution*: minimum 3.0 between the peaks due to impurities A and B and minimum 2.0 between the peaks due to impurities B and C.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (5.0 ppm *V/V*);
— *total*: maximum 0.0040 per cent of the sum of the areas of all the peaks (40.0 ppm *V/V*).

Water (*2.5.28*)
Maximum 10.0 ppm *V/V*, determined using an electrolytic hygrometer.

STORAGE
In gaseous or liquid state, in suitable containers, complying with the legal regulations.

IMPURITIES
Specified impurities *A, D.*

Other detectable impurities *B, C.*

A. oxygen,

B. nitrogen,

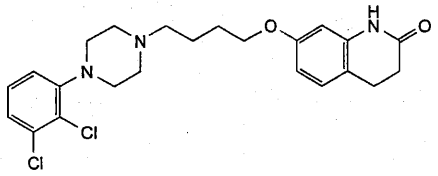
C. methane,

D. water.

Ph Eur

Aripiprazole

(Ph. Eur. monograph 2617)



$C_{23}H_{27}Cl_2N_3O_2$ 448.4 *129722-12-9*

Action and use
Dopamine D_2 receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
7-[4-[4-(2,3-Dichlorophenyl)piperazin-1-yl]butoxy]-3,4-dihydroquinolin-2(1*H*)-one.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white crystals or crystalline powder.

Solubility
Practically insoluble in water, soluble in methylene chloride, very slighty soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *aripiprazole CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

TESTS
Appearance of solution
If intended for use in the manufacture of parenteral preparations, the solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 (*2.2.2, Method II*).

Dissolve 0.5 g in a mixture of 10 volumes of *acetic acid R* and 90 volumes of *anhydrous ethanol R* and dilute to 20 mL with the same mixture of solvents. Sonicate for about 15 min, shaking occasionally, until dissolution is complete.

Related substances
Liquid chromatography (*2.2.29*). *Protect the solutions from light.*

Solvent mixture *acetic acid R, methanol R, acetonitrile R, water R* (1:10:30:60 *V/V/V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *aripiprazole impurity F CRS* in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL of the solution to 50 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *aripiprazole CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: *acetonitrile R*, 0.05 per cent *V/V* solution of *trifluoroacetic acid R* (10:90 *V/V*);
— *mobile phase B*: 0.05 per cent *V/V* solution of *trifluoroacetic acid R, acetonitrile R* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	80	20
2 - 10	80 → 65	20 → 35
10 - 20	65 → 10	35 → 90
20 - 25	10	90

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (a) and (b).

Relative retention With reference to aripiprazole (retention time = about 11 min): impurity F = about 1.1.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to aripiprazole and impurity F.

Calculation of percentage contents:
— for each impurity, use the concentration of aripiprazole in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

Dissolve 1.0 mg of the substance to be examined in 20 mL of a 5.17 g/L solution of *hydrochloric acid R*.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (c).

System suitability Reference solution (c):
— *symmetry factor*: maximum 2.0.

Calculate the percentage content of $C_{23}H_{27}Cl_2N_3O_2$ taking into account the assigned content of *aripiprazole CRS*.

STORAGE

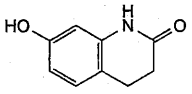
Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

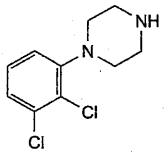
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES

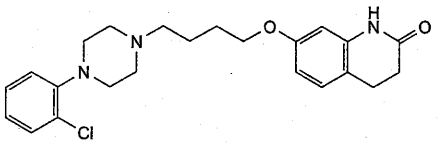
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



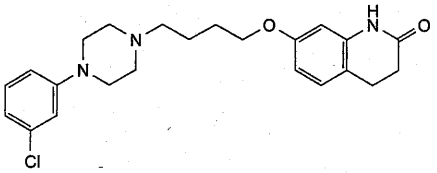
A. 7-hydroxy-3,4-dihydroquinolin-2(1*H*)-one,



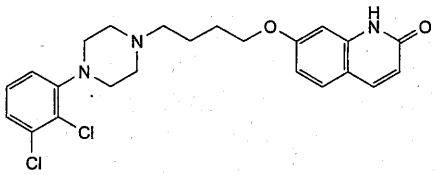
B. 1-(2,3-dichlorophenyl)piperazine,



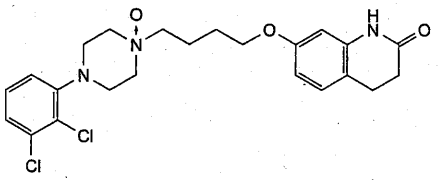
C. 7-[4-[4-(2-chlorophenyl)piperazin-1-yl]butoxy]-3,4-dihydroquinolin-2(1*H*)-one,



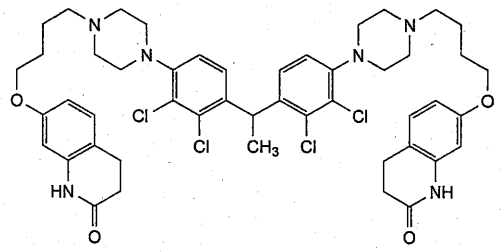
D. 7-[4-[4-(3-chlorophenyl)piperazin-1-yl]butoxy]-3,4-dihydroquinolin-2(1*H*)-one,



E. 7-[4-[4-(2,3-dichlorophenyl)piperazin-1-yl]butoxy]quinolin-2(1*H*)-one,



F. 7-[4-[4-(2,3-dichlorophenyl)-1-oxidopiperazin-1-yl]butoxy]-3,4-dihydroquinolin-2(1*H*)-one,

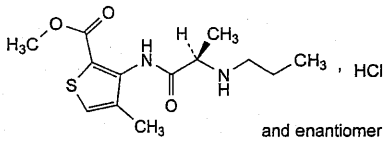


G. 7,7′-[ethane-1,1-diylbis[(2,3-dichlorobenzene-4,1-diyl)piperazine-4,1-diylbutane-4,1-diyloxy]]bis[3,4-dihydroquinolin-2(1*H*)-one].

Ph Eur

Articaine Hydrochloride

(Ph. Eur. monograph 1688)



$C_{13}H_{21}ClN_2O_3S$ 320.8 *23964-57-0*

Action and use
Local anaesthetic.

Ph Eur

DEFINITION

Methyl 4-methyl-3-[[(2*RS*)-2-(propylamino) propanoyl]amino]thiophene-2-carboxylate hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Dissolve 50.0 mg in a 1 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of the solution to 100.0 mL with a 1 g/L solution of *hydrochloric acid R*. Examined between 200 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 272 nm.
The specific absorbance at the maximum is 290 to 320.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Place dropwise 20 µL of the test solution on 300 mg discs.

Test solution Dissolve 0.1 g in 5 mL of *water R*, add 3 mL of a saturated solution of *sodium hydrogen carbonate R* and shake twice with 2 mL of *methylene chloride R*. Combine the methylene chloride layers, dilute to 5.0 mL with *methylene chloride R* and dry over *anhydrous sodium sulfate R*.

Comparison *articaine hydrochloride CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 5 mL of *ethanol (96 per cent) R*.

Reference solution Dissolve 20 mg of *articaine hydrochloride CRS* in 5 mL of *ethanol (96 per cent) R*.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *triethylamine R, ethyl acetate R, heptane R* (10:35:65 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.50 g in *water R* and dilute to 10 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method I*).

pH (*2.2.3*)
4.2 to 5.2.

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *articaine impurity A CRS* and 2.5 mg of *articaine impurity E CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 45 °C.

Mobile phase Mix 25 volumes of *acetonitrile R* and 75 volumes of a solution prepared as follows: dissolve 2.02 g of *sodium heptanesulfonate R* and 4.08 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 1000 mL with the same solvent. Adjust to pH 2.0 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 10 µL.

Run time 5 times the retention time of articaine.

Relative retention With reference to articaine (retention time = about 9 min): impurity A = about 0.8; impurity E = about 0.86.

System suitability Reference solution (b):
— *resolution*: minimum 1.2 between the peaks due to impurities A and E.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurites other than A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 5 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R.* Carry out a potentiometric titration (*2.2.20*) using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 32.08 mg of $C_{13}H_{21}ClN_2O_3S$.

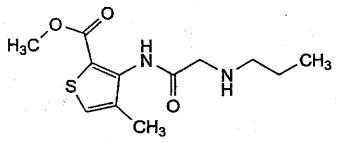
STORAGE

Protected from light.

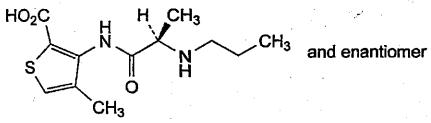
IMPURITIES

Specified impurities *A.*

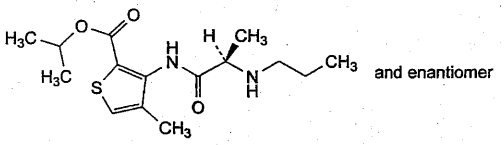
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H, I, J.*



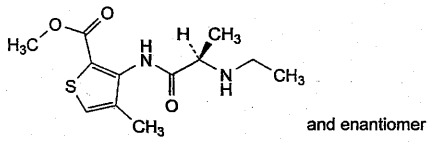
A. methyl 4-methyl-3-[[2-(propylamino)acetyl]amino] thiophene-2-carboxylate (acetamidoarticaine),



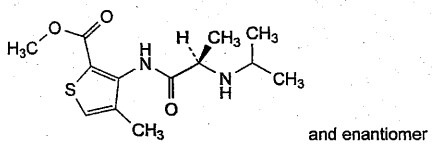
B. 4-methyl-3-[[(2*RS*)-2-(propylamino)propanoyl]amino] thiophene-2-carboxylic acid (articaine acid),



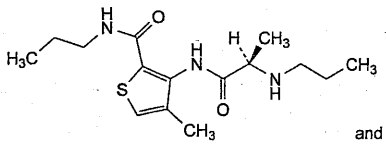
C. 1-methylethyl 4-methyl-3-[[(2*RS*)-2-(propylamino) propanoyl]amino]thiophene-2-carboxylate (articaine isopropyl ester),



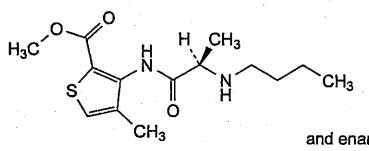
D. methyl 3-[[(2*RS*)-2-(ethylamino)propanoyl]amino]-4-methylthiophene-2-carboxylate (ethylarticaine),



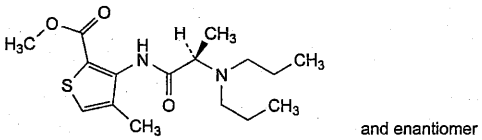
E. methyl 4-methyl-3-[[(2*RS*)-2-[(1-methylethyl)amino] propanoyl]amino]thiophene-2-carboxylate (isopropylarticaine),



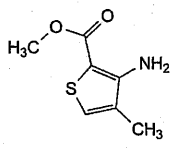
F. 4-methyl-*N*-propyl-3-[[(2*RS*)-2-(propylamino)propanoyl] amino]thiophene-2-carboxamide (articaine acid propionamide),



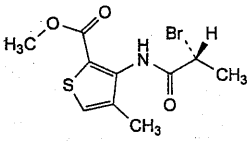
G. methyl 3-[[(2*RS*)-2-(butylamino)propanoyl]amino]-4-methylthiophene-2-carboxylate (butylarticaine),



H. methyl 3-[[(2*RS*)-2-(dipropylamino)propanoyl]amino]-4-methylthiophene-2-carboxylate (dipropylarticaine),



I. methyl 3-amino-4-methylthiophene-2-carboxylate (3-aminoarticaine),

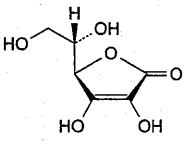


J. methyl 3-[[(2*RS*)-2-bromopropanoyl]amino]-4-methylthiophene-2-carboxylate (bromo compound).

Ph Eur

Ascorbic Acid

(Ph. Eur. monograph 0253)



$C_6H_8O_6$ 176.1 *50-81-7*

Action and use

Vitamin C.

Preparations

Ascorbic Acid Injection

Ascorbic Acid Tablets

Ascorbic Acid Chewable Tablets

Paediatric Vitamins A, C and D Oral Drops

Potassium Ascorbate Eye Drops

Vitamins B and C Injection

When Vitamin C is prescribed or demanded, Ascorbic Acid shall be dispensed or supplied.

Ph Eur

DEFINITION

(5*R*)-5-[(1*S*)-1,2-Dihydroxyethyl]-3,4-dihydroxyfuran-2(5*H*)-one.

Content

99.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals, becoming discoloured on exposure to air and moisture.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent).

mp

About 190 °C, with decomposition.

IDENTIFICATION

First identification: B, C.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.10 g in *water R* and dilute immediately to 100.0 mL with the same solvent. Add 1.0 mL of the solution to 10 mL of a 10.3 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with *water R*.

Absorption maximum At 243 nm, determined immediately after dissolution.

Specific absorbance at the absorption maximum 545 to 585.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ascorbic acid CRS*.

C. pH (*2.2.3*): 2.1 to 2.6 for solution S (see Tests).

D. To 1 mL of solution S add 0.2 mL of *dilute nitric acid R* and 0.2 mL of *silver nitrate solution R2*. A grey precipitate is formed.

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 20.5 to + 21.5.

Dissolve 2.50 g in *water R* and dilute to 25.0 mL with the same solvent.

Impurity E

Maximum 0.2 per cent.

Test solution Dissolve 0.25 g in 5 mL of *water R*. Neutralise using *dilute sodium hydroxide solution R*, then add 1 mL of *dilute acetic acid R* and 0.5 mL of *calcium chloride solution R*.

Reference solution Dissolve 70 mg of *oxalic acid R* (dihydrate of impurity E) in *water R* and dilute to 500 mL with the same solvent; to 5 mL of the solution add 1 mL of *dilute acetic acid R* and 0.5 mL of *calcium chloride solution R*. Allow the solutions to stand for 1 h. Any opalescence in the test solution is not more intense than that in the reference solution.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Phosphate buffer solution Dissolve 6.8 g of *potassium dihydrogen phosphate R* in *water for chromatography R* and dilute to about 175 mL with the same solvent. Filter through a membrane filter (nominal pore size 0.45 µm) and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 0.500 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *ascorbic acid impurity C CRS* in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *ascorbic acid impurity D CRS* and 5.0 mg of *ascorbic acid CRS* in the mobile phase, add 2.5 mL of reference solution (a) and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1 mL of the test solution to 200 mL with the mobile phase. Mix 1 mL of this solution and 1 mL of reference solution (a).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *aminopropylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 45 °C.

Mobile phase Phosphate buffer solution, *acetonitrile R1* (25:75 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of ascorbic acid.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and D.

Relative retention With reference to ascorbic acid (retention time = about 11 min): impurity D = about 0.4; impurity C = about 1.7.

System suitability:
- *resolution*: minimum 3.0 between the peaks due to ascorbic acid and impurity C in the chromatogram obtained with reference solution (c);
- *signal-to-noise ratio*: minimum 20 for the peak due to impurity C in the chromatogram obtained with reference solution (b).

Limits:
- *impurities C, D*: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the peak due to ascorbic acid in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *sum of impurities other than C and D*: not more than twice the area of the peak due to ascorbic acid in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *disregard limit*: 0.5 times the area of the peak due to ascorbic acid in the chromatogram obtained with reference solution (b) (0.05 per cent).

Copper
Maximum 5 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 2.0 g in *0.1 M nitric acid* and dilute to 25.0 mL with the same acid.

Reference solutions Prepare the reference solutions (0.2 ppm, 0.4 ppm and 0.6 ppm) using *copper standard solution (10 ppm Cu) R*, diluting with *0.1 M nitric acid*.

Source Copper hollow-cathode lamp.

Wavelength 324.8 nm.

Atomisation device Air-acetylene flame.

Adjust the zero of the apparatus using *0.1 M nitric acid*.

Iron
Maximum 2 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 5.0 g in *0.1 M nitric acid* and dilute to 25.0 mL with the same acid.

Reference solutions Prepare the reference solutions (0.2 ppm, 0.4 ppm and 0.6 ppm) using *iron standard solution (20 ppm Fe) R*, diluting with *0.1 M nitric acid*.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Adjust the zero of the apparatus using *0.1 M nitric acid*.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

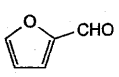
ASSAY
Dissolve 0.150 g in a mixture of 10 mL of *dilute sulfuric acid R* and 80 mL of *carbon dioxide-free water R*. Add 1 mL of *starch solution R*. Titrate with *0.05 M iodine* until a persistent violet-blue colour is obtained.

1 mL of *0.05 M iodine* is equivalent to 8.81 mg of $C_6H_8O_6$.

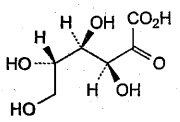
STORAGE
In a non-metallic container, protected from light.

IMPURITIES
Specified impurities *C, D, E.*

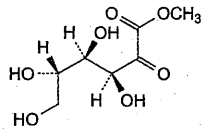
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, F, G, H.*



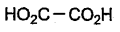
A. furan-2-carbaldehyde,



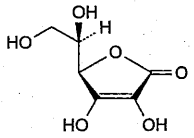
C. L-*xylo*-hex-2-ulosonic acid (L-sorbosonic acid),



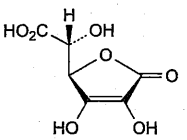
D. methyl L-*xylo*-hex-2-ulosonate (methyl L-sorbosonate),



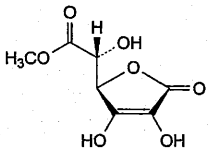
E. oxalic acid,



F. (5*R*)-5-[(1*R*)-1,2-dihydroxyethyl]-3,4-dihydroxyfuran-2(5*H*)-one,



G. (*R*)-[(2*R*)-3,4-dihydroxy-5-oxo-2,5-dihydrofuran-2-yl]hydroxyacetic acid,

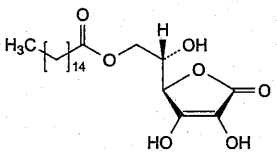


H. methyl (*R*)-[(2*R*)-3,4-dihydroxy-5-oxo-2,5-dihydrofuran-2-yl]hydroxyacetate.

Ph Eur

Ascorbyl Palmitate

(Ph. Eur. monograph 0807)



$C_{22}H_{38}O_7$ 414.5 *137-66-6*

Action and use
Excipient.

Ph Eur

DEFINITION
(2*S*)-2-[(2*R*)-3,4-Dihydroxy-5-oxo-2,5-dihydrofuran-2-yl]-2-hydroxyethyl hexadecanoate.

Content
98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish-white powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent) and in methanol, practically insoluble in methylene chloride and in fatty oils.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ascorbyl palmitate CRS.*

C. Dissolve about 10 mg in 5 mL of *methanol R.*
The solution decolourises *dichlorophenolindophenol standard solution R.*

TESTS
Solution S
Dissolve 2.50 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_4 (*2.2.2, Method I*).

Specific optical rotation (*2.2.7*)
+ 21 to + 24 (dried substance), determined on solution S.

Related substances
The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 5 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 50 mL of *ethanol (96 per cent) R.*
Add 30 mL of *water R* and titrate with *0.05 M iodine* until a yellow colour is obtained.

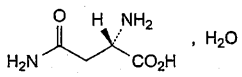
1 mL of *0.05 M iodine* is equivalent to 20.73 mg of $C_{22}H_{38}O_7$.

STORAGE
In an airtight container, protected from light.

Ph Eur

Asparagine Monohydrate

(Ph. Eur. monograph 2086)



$C_4H_8N_2O_3,H_2O$ 150.1 *5794-13-8*

Action and use
Amino acid.

Ph Eur

DEFINITION
(2*S*)-2,4-Diamino-4-oxobutanoic acid monohydrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Slightly soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: A, B, D.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *asparagine monohydrate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *asparagine monohydrate CRS* in *water R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (25:25:50 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 110 °C for 15 min.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Loss on drying (see Tests).

TESTS
Solution S
Dissolve with heating 2.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 6.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 33.7 to + 36.0 (dried substance).
Dissolve 2.50 g in a 309.0 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same acid.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R.*

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *water R.*

Reference solution (c) Dissolve 5.0 mg of *aspartic acid R* (impurity A) in *water R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *water R.*

Reference solution (d) Dissolve 3.0 mg of *asparagine impurity C CRS* in 40 mL of *water R* using sonification and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *water R*.

Reference solution (e) Mix 5.0 mL of reference solution (c) with 2.5 mL of reference solution (a) and dilute to 10.0 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 25 °C.

Mobile phase Dissolve 13.6 g of *potassium dihydrogen phosphate R* and 2.16 g of *sodium octanesulfonate R* in about 900 mL of *water for chromatography R*. Adjust to pH 2.2 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*. Add 5 mL of *acetonitrile R1*.

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Run time Twice the retention time of asparagine.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to asparagine (retention time = about 6.6 min): impurity C = about 0.6; impurity A = about 1.2.

System suitability Reference solution (e):
— *resolution*: minimum 5.0 between the peaks due to asparagine and impurity A.

Calculation of percentage contents:
— for impurity A, use the concentration of impurity A in reference solution (c);
— for impurity C, use the concentration of impurity C in reference solution (d);
— for impurities other than A and C, use the concentration of asparagine monohydrate in reference solution (b).

Limits:
— *impurity A*: maximum 0.5 per cent;
— *impurity C*: maximum 0.1 per cent;
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.8 per cent;
— *reporting threshold*: 0.03 per cent.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 12.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 200 ppm.

To 0.75 g add 2.5 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*. Examine after 30 min.

Ammonium (*2.4.1, Method B*)
Maximum 0.1 per cent, determined on 10 mg.

Prepare the standard using 0.1 mL of *ammonium standard solution (100 ppm NH_4) R*.

Iron (*2.4.9*)
Maximum 10 ppm.

Dissolve 1.0 g in *dilute hydrochloric acid R* and dilute to 10 mL with the same acid. Shake 3 times with 10 mL of *methyl isobutyl ketone R1* for 3 min. Wash the combined organic phases with 10 mL of *water R* for 3 min. The aqueous phase complies with the limit test for iron.

Loss on drying (*2.2.32*)
10.5 per cent to 12.5 per cent, determined on 1.000 g by drying in an oven at 130 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

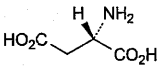
Dissolve 0.110 g in 5 mL of *anhydrous formic acid R*. Add 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 13.21 mg of $C_4H_8N_2O_3$.

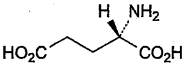
IMPURITIES

Specified impurities *A, C.*

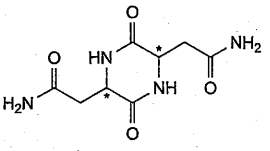
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, E, F, G, H.*



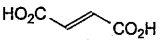
A. (2*S*)-2-aminobutanedioic acid (aspartic acid),



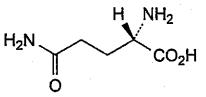
B. (2*S*)-2-aminopentanedioic acid (glutamic acid),



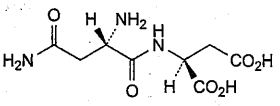
C. 2,2′-[(2Ξ,5Ξ)-3,6-dioxopiperazine-2,5-diyl]diacetamide,



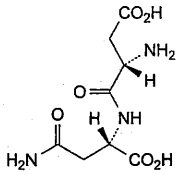
D. (2*E*)-but-2-enedioic acid (fumaric acid),



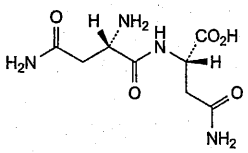
E. (2*S*)-2,5-diamino-5-oxopentanoic acid (glutamine),



F. (2*S*)-2-[[(2*S*)-2,4-diamino-4-oxobutanoyl]amino] butanedioic acid (asparaginylaspartic acid),



G. (2*S*)-4-amino-2-[[(2*S*)-2-amino-3-carboxypropanoyl] amino]-4-oxobutanoic acid (α-aspartylasparagine),

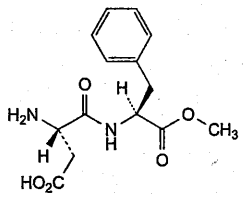


H. (2*S*)-4-amino-2-[[(2*S*)-2,4-diamino-4-oxobutanoyl] amino]-4-oxobutanoic acid (asparaginylasparagine).

Ph Eur

Aspartame

(Ph. Eur. monograph 0973)



$C_{14}H_{18}N_2O_5$ 294.3 *22839-47-0*

Action and use
Sweetening agent.

Ph Eur

DEFINITION

(3*S*)-3-Amino-4-[[(2*S*)-1-methoxy-1-oxo-3-phenylpropan-2-yl]amino]-4-oxobutanoic acid (methyl α-L-aspartyl-L-phenylalaninate).

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, slightly hygroscopic, crystalline powder.

Solubility
Sparingly soluble or slightly soluble in water and in ethanol (96 per cent), practically insoluble in hexane and in methylene chloride.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.1 g in *ethanol (96 per cent) R* and dilute to 100 mL with the same solvent.

Spectral range 230-300 nm.

Absorption maxima At 247 nm, 252 nm, 258 nm and 264 nm.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *aspartame CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 15 mg of the substance to be examined in 2.5 mL of *water R* and dilute to 10 mL with *acetic acid R.*

Reference solution Dissolve 15 mg of *aspartame CRS* in 2.5 mL of *water R* and dilute to 10 mL with *acetic acid R.*

Plate *TLC silica gel G plate R.*

Mobile phase *water R, anhydrous formic acid R, methanol R, methylene chloride R* (2:4:30:64 *V/V/V/V*).

Application 20 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 100-105 °C for 15 min.

Results The spot in the chromatogram obtained with the test solution is similar in position, colour and size to the spot in the chromatogram obtained with the reference solution.

D. Dissolve about 20 mg in 5 mL of *methanol R* and add 1 mL of *alkaline hydroxylamine solution R1.* Heat on a water-bath for 15 min. Allow to cool and adjust to about pH 2 with *dilute hydrochloric acid R.* Add 0.1 mL of *ferric chloride solution R1.* A brownish-red colour is produced.

TESTS

Solution S
Dissolve 0.8 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).

Conductivity (*2.2.38*)
Maximum 30 µS·cm^{-1}.

Dissolve 0.80 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution (C_1) and that of the water used for preparing the solution (C_2). The readings must be stable within 1 per cent over a period of 30 s.

Calculate the conductivity of the solution of the substance to be examined using the following expression:

$$C_1 - 0.992\ C_2$$

Specific optical rotation (*2.2.7*)
+ 14.5 to + 16.5 (dried substance).

Dissolve 2.00 g in a 690 g/L solution of *anhydrous formic acid R* and dilute to 50.0 mL with the same solution. Measure within 30 min of preparation.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.60 g of the substance to be examined in a mixture of 1.5 volumes of *glacial acetic acid R* and 98.5 volumes of *water R* and dilute to 100.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 4.5 mg of *aspartame impurity A CRS* in a mixture of 1.5 volumes of *glacial acetic acid R* and 98.5 volumes of *water R* and dilute to 50.0 mL with the same mixture of solvents.

Reference solution (b) Dissolve 30.0 mg of *phenylalanine R* (impurity C) in a mixture of 15 volumes of *glacial acetic*

acid R and 85 volumes of *water R* and dilute to 100.0 mL with the same mixture of solvents. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (c) Dilute 5.0 mL of the test solution to 10.0 mL with *water R*. Dilute 3.0 mL of this solution to 100.0 mL with *water R*.

Reference solution (d) Dissolve 30.0 mg of *L-aspartyl-L-phenylalanine R* (impurity B) in a mixture of 15 volumes of *glacial acetic acid R* and 85 volumes of *water R* and dilute to 100.0 mL with the same mixture of solvents. Dilute 1.0 mL of the solution to 10.0 mL with *water R*. Mix 1.0 mL of this solution with 1.0 mL of reference solution (b).

Column
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase Mix 10 volumes of *acetonitrile R* and 90 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.7 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time Twice the retention time of aspartame.

System suitability Reference solution (d):
— *resolution*: minimum 3.5 between the peaks due to impurities B and C.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *sum of impurities other than A and C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.5 per cent);
— *disregard limit*: disregard any peak due to the solvent.

Loss on drying (*2.2.32*)
Maximum 4.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 1.5 mL of *anhydrous formic acid R* and 60 mL of *anhydrous acetic acid R*. Titrate immediately with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 29.43 mg of $C_{14}H_{18}N_2O_5$.

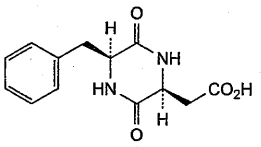
STORAGE

In an airtight container.

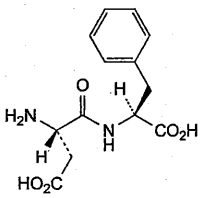
IMPURITIES

Specified impurities *A, C.*

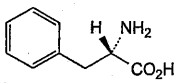
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. 2-[(2*S*,5*S*)-5-benzyl-3,6-dioxopiperazin-2-yl]acetic acid,



B. (3*S*)-3-amino-4-[[(1*S*)-1-carboxy-2-phenylethyl]amino]-4-oxobutanoic acid (α-L-aspartyl-L-phenylalanine),

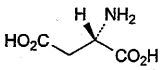


C. (2*S*)-2-amino-3-phenylpropanoic acid (L-phenylalanine).

Ph Eur

Aspartic Acid

(Ph. Eur. monograph 0797)



$C_4H_7NO_4$ 133.1 *56-84-8*

Action and use
Amino acid.

Ph Eur

DEFINITION

(2*S*)-2-Aminobutanedioic acid (L-aspartic acid).

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Slightly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification *[carry out either tests A, C or tests C, D.]*

Second identification: A, B, E.

A. Specific optical rotation (*2.2.7*): + 24.0 to + 26.0 (dried substance).

Dissolve 2.00 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

B. A suspension of 1 g in 10 mL of *water R* is strongly acid (*2.2.4*).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *aspartic acid CRS.*

D. Enantiomeric purity (see Tests).

E. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 2 mL of *dilute ammonia R1* and dilute to 50 mL with *water R*.

Reference solution Dissolve 10 mg of *aspartic acid CRS* in 2 mL of *dilute ammonia R1* and dilute to 50 mL with *water R*.

Plate *TLC silica gel plate R*.

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.5 g in a 103 g/L solution of *hydrochloric acid R* and dilute to 10 mL with the same acid.

Enantiomeric purity

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 0.100 g of *D-aspartic acid R* (impurity I) in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 0.100 g of the substance to be examined in 90 mL of *water R*, add 0.3 mL of reference solution (a) and dilute to 100.0 mL with *water R*.

Reference solution (c) Dilute 0.3 mL of reference solution (a) to 100.0 mL with *water R*.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *L-penicillamine coated silica gel for chiral separations R* (5 µm);
- *temperature*: 30 °C.

Mobile phase *2-propanol R*, 0.5 g/L solution of *copper sulfate pentahydrate R* (5:95 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Relative retention With reference to aspartic acid (retention time = about 12 min): impurity I = about 0.85.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to impurity I and aspartic acid.

Calculation of percentage content:
- for impurity I, use the concentration of impurity I in reference solution (c).

Limit:
- *impurity I*: maximum 0.3 per cent.

Other dicarboxylic acids

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.500 g of the substance to be examined in 2.0 mL of a 618 g/L solution of *hydrochloric acid R* and dilute to 10.0 mL with *water R*.

Reference solution (a) Dissolve 20.0 mg of *malic acid R* (impurity A) in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dissolve 10.0 mg of *maleic acid R* (impurity H) in *water R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *water R*.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with reference solution (a).

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *water R*.

Reference solution (e) Dissolve 10.0 mg of *fumaric acid R* (impurity B) in *water R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *water R*.

Column:
- *size*: *l* = 0.30 m, Ø = 7.8 mm;
- *stationary phase*: *cation-exchange resin R* (9 µm);
- *temperature*: 30 °C.

Mobile phase 0.39 g/L solution of *sulfuric acid R*.

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 10 µL.

Run time 4 times the retention time of impurity H.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and H; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity B.

Relative retention With reference to impurity H (retention time = about 7.5 min): impurity A = about 1.2; impurity B = about 2.0.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to impurities H and A.

Calculation of percentage contents:
- for impurity A, use the concentration of impurity A in reference solution (d);
- for impurities B and H, use the concentration of impurity H in reference solution (b);
- for impurities other than B and H, use the concentration of impurity A in reference solution (d).

Limits:
- *impurity A*: maximum 0.2 per cent;
- *impurity B*: maximum 0.10 per cent;
- *impurity H*: maximum 0.10 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *reporting threshold*: 0.05 per cent.

Ninhydrin-positive substances

Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test and reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *dilute hydrochloric acid R1* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Reference solution (e) Dissolve 30.0 mg of *alanine R* (impurity D), 60.0 mg of *asparagine R* (impurity G) and 30.0 mg of *glutamic acid R* (impurity C) in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test solution, blank solution and reference solutions (a), (b), (d) and (e) into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for impurities C, D and G, use the concentration of each impurity in reference solution (e);
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of aspartic acid in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *impurities C, D, G*: for each impurity, maximum 0.2 per cent;
— *any ninhydrin-positive substance*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold*: 0.05 per cent.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dissolve 0.25 g in 3 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*. Add 1 mL of *water R* instead of 1 mL of *dilute nitric acid R*.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dissolve 0.5 g in 4 mL of *hydrochloric acid R* and dilute to 15 mL with *distilled water R*. Carry out the evaluation of the test after 30 min.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modification.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 50 mL of *carbon dioxide-free water R*, with slight heating if necessary. Cool and titrate with *0.1 M sodium hydroxide* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 13.31 mg of $C_4H_7NO_4$.

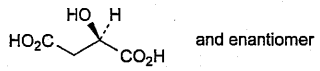
STORAGE

Protected from light.

IMPURITIES

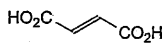
Specified impurities *A, B, C, D, H, G, I.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F.*

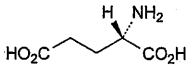


and enantiomer

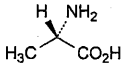
A. (2*RS*)-2-hydroxybutanedioic acid (malic acid),



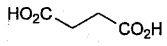
B. (2*E*)-but-2-enedioic acid (fumaric acid),



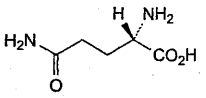
C. (2*S*)-2-aminopentanedioic acid (glutamic acid),



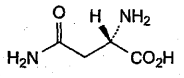
D. (2*S*)-2-aminopropanoic acid (alanine),



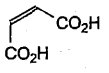
E. butanedioic acid (succinic acid),



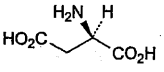
F. (2*S*)-2,5-diamino-5-oxopentanoic acid (L-glutamine),



G. (2*S*)-2,4-diamino-4-oxobutanoic acid (asparagine),



H. (2*Z*)-but-2-enedioic acid (maleic acid),

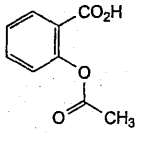


I. (2*R*)-2-aminobutanedioic acid (D-aspartic acid).

Ph Eur

Aspirin

(Acetylsalicylic Acid, Ph. Eur. monograph 0309)



$C_9H_8O_4$ 180.2 *50-78-2*

Action and use
Salicylate; non-selective cyclo-oxygenase inhibitor; antipyretic; analgesic; anti-inflammatory.

Preparations
Aspirin Tablets
Aspirin Dispersible Tablets
Aspirin Effervescent Soluble Tablets
Aspirin Gastro-resistant Tablets
Aspirin and Caffeine Tablets
Co-codaprin Tablets
Co-codaprin Dispersible Tablets

Ph Eur

DEFINITION
2-(Acetyloxy)benzoic acid.

Content
99.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Slightly soluble in water, freely soluble in ethanol (96 per cent).

mp
About 143 °C (instantaneous method).

IDENTIFICATION
First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *acetylsalicylic acid CRS.*

B. To 0.2 g add 4 mL of *dilute sodium hydroxide solution R* and boil for 3 min. Cool and add 5 mL of *dilute sulfuric acid R*. A crystalline precipitate is formed. Filter, wash the precipitate and dry at 100-105 °C. The melting point (*2.2.14*) is 156 °C to 161 °C.

C. In a test tube mix 0.1 g with 0.5 g of *calcium hydroxide R*. Heat the mixture and expose to the fumes produced a piece of filter paper impregnated with 0.05 mL of *nitrobenzaldehyde solution R*. A greenish-blue or greenish-yellow colour develops on the paper. Moisten the paper with *dilute hydrochloric acid R*. The colour becomes blue.

D. Dissolve with heating about 20 mg of the precipitate obtained in identification test B in 10 mL of *water R* and cool. The solution gives reaction (a) of salicylates (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in 9 mL of *ethanol (96 per cent) R*.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in *acetonitrile for chromatography R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *salicylic acid R* (impurity C) in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *salicylic acid R* (impurity C) in the mobile phase and dilute to 10.0 mL with the mobile phase. To 1.0 mL of the solution add 0.2 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve with the aid of ultrasound the contents of a vial of *acetylsalicylic acid for peak identification CRS* (containing impurities A, B, D, E and F) in 1.0 mL of *acetonitrile R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *phosphoric acid R, acetonitrile for chromatography R, water R* (2:400:600 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 237 nm.

Injection 10 µL.

Run time 7 times the retention time of acetylsalicylic acid.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C; use the chromatogram supplied with *acetylsalicylic acid for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, D, E and F.

Relative retention With reference to acetylsalicylic acid (retention time = about 5 min): impurity A = about 0.7; impurity B = about 0.8; impurity C = about 1.3;

impurity D = about 2.3; impurity E = about 3.2; impurity F = about 6.0.

System suitability Reference solution (b):
— *resolution*: minimum 6.0 between the peaks due to acetylsalicylic acid and impurity C.

Limits:
— *impurities A, B, C, D, E, F*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);
— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Loss on drying *(2.2.32)*
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo*.

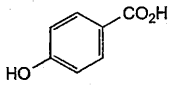
Sulfated ash *(2.4.14)*
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
In a flask with a ground-glass stopper, dissolve 1.000 g in 10 mL of *ethanol (96 per cent) R*. Add 50.0 mL of *0.5 M sodium hydroxide*. Close the flask and allow to stand for 1 h. Using 0.2 mL of *phenolphthalein solution R* as indicator, titrate with *0.5 M hydrochloric acid*. Carry out a blank titration.

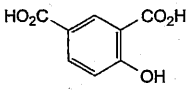
1 mL of *0.5 M sodium hydroxide* is equivalent to 45.04 mg of $C_9H_8O_4$.

STORAGE
In an airtight container.

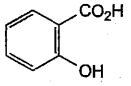
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



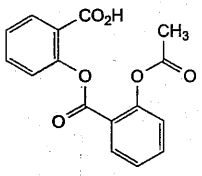
A. 4-hydroxybenzoic acid,



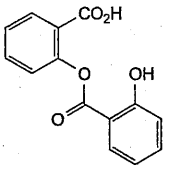
B. 4-hydroxybenzene-1,3-dicarboxylic acid (4-hydroxyisophthalic acid),



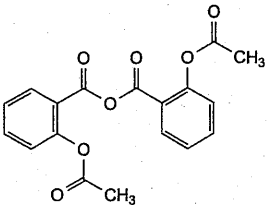
C. 2-hydroxybenzenecarboxylic acid (salicylic acid),



D. 2-[[2-(acetyloxy)benzoyl]oxy]benzoic acid (acetylsalicylsalicylic acid),



E. 2-[(2-hydroxybenzoyl)oxy]benzoic acid (salsalate, salicylsalicylic acid),

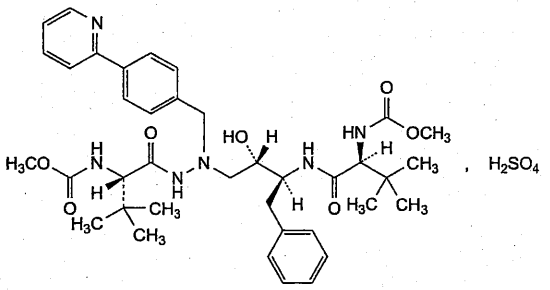


F. 2-(acetyloxy)benzoic anhydride (acetylsalicylic anhydride).

Ph Eur

Atazanavir Sulfate

(Ph. Eur. monograph 2898)



$C_{38}H_{54}N_6O_{11}S$ 803 *229975-97-7*

Action and use
Antiviral (HIV).

Ph Eur

DEFINITION
Methyl [(5*S*,10*S*,11*S*,14*S*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-8-[[4-(pyridin-2-yl)phenyl]methyl]-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate sulfate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or pale yellow, slightly hygroscopic, crystalline powder that may contain agglomerates.

Solubility
Slightly soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in heptane.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry *(2.2.24)*.
Comparison *atazanavir sulfate CRS*.

C. It gives reaction (a) of sulfates *(2.3.1)*.

TESTS

Specific optical rotation (*2.2.7*)

−44 to −40 (anhydrous substance), measured at 25 °C.

Dissolve 0.100 g in 8 mL of *methanol R*, using sonication if necessary, and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mix equal volumes of *acetonitrile R1* and a freshly prepared 2.73 g/L solution of *potassium dihydrogen phosphate R* in *water for chromatography R* previously adjusted to pH 3.5 with *dilute phosphoric acid R*.

Test solution (a) Dissolve 20.0 mg of the substance to be examined in 40 mL of the solvent mixture, sonicate for 3 min and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in 40 mL of the solvent mixture, sonicate for 3 min and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of *atazanavir sulfate CRS* in 40 mL of the solvent mixture, sonicate for 3 min and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 4 mg of *atazanavir for system suitability CRS* (containing impurity F) in 8 mL of the solvent mixture, sonicate for 3 min and dilute to 10 mL with the solvent mixture.

Reference solution (d) Dissolve 2.0 mg of *atazanavir impurity K CRS* in 9 mL of the solvent mixture, sonicate for 3 min and dilute to 10.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 100.0 mL with the solvent mixture. Dilute 3.0 mL of this solution to 20.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.0 µm);
- *temperature*: 25 °C.

Mobile phase:
- *mobile phase A*: mix 25 volumes of *acetonitrile R1* and 75 volumes of a freshly prepared 2.73 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.5 with *phosphoric acid R*;
- *mobile phase B*: mix 25 volumes of a freshly prepared 2.73 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.5 with *phosphoric acid R*, and 75 volumes of *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 45	100 → 0	0 → 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL of test solution (a) and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *atazanavir for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to atazanavir (retention time = about 30 min): impurity F = about 0.99.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to impurity F and atazanavir.

Calculation of percentage contents:
- for each impurity, use the concentration of atazanavir sulfate in reference solution (b).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.5 per cent;
- *reporting threshold*: 0.05 per cent; disregard any peak with a relative retention with reference to atazanavir of less than 0.2.

Impurity K

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	95	5
5 - 8	95 → 0	5 → 100
8 - 14	0	100

Injection 20 µL of test solution (b) and reference solution (d).

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peak due to impurity K.

Relative retention With reference to atazanavir (retention time = about 10 min): impurity K = about 0.4.

Calculation of percentage content:
- for impurity K, use the concentration of impurity K in reference solution (d).

Limit:
- *impurity K*: maximum 0.15 per cent.

Water (*2.5.32*)

Maximum 2.5 per cent, determined on 0.100 g by direct sample introduction.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Solvent mixture.

Injection Test solution (a) and reference solutions (a) and (c).

Run time 1.6 times the retention time of atazanavir.

Relative retention With reference to atazanavir (retention time = about 9.5 min): impurity F = about 0.94.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to impurity F and atazanavir.

Calculate the percentage content of $C_{38}H_{54}N_6O_{11}S$ using the chromatogram obtained with reference solution (a) and taking into account the assigned content of *atazanavir sulfate CRS*.

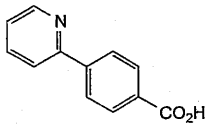
STORAGE

In an airtight container.

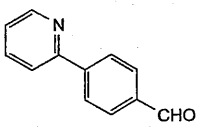
IMPURITIES

Specified impurities *K*.

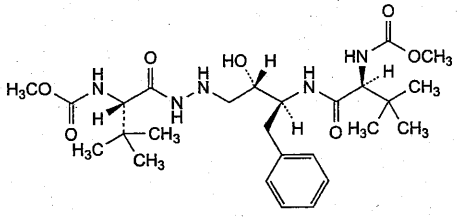
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, C, D, E, F, G, H, I, J.



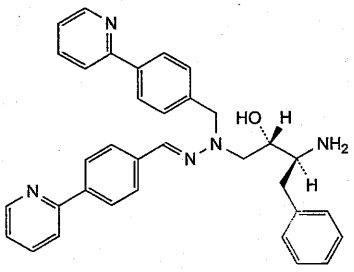
A. 4-(pyridin-2-yl)benzoic acid,



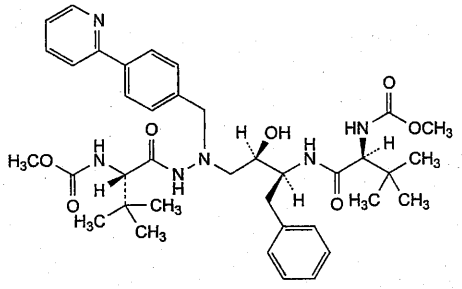
B. 4-(pyridin-2-yl)benzaldehyde,



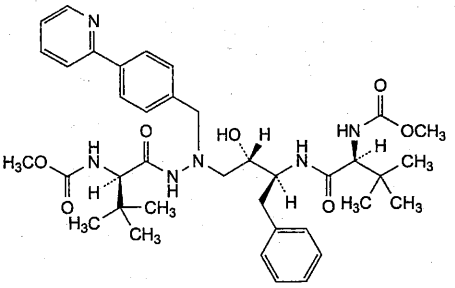
C. methyl [(5*S*,10*S*,11*S*,14*S*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate,



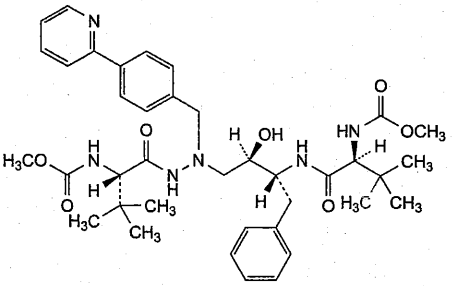
D. (2*S*,3*S*)-3-amino-4-phenyl-1-[(*E*)-1-[[4-(pyridin-2-yl)phenyl]methyl]-2-[[4-(pyridin-2-yl)phenyl]methylidene]hydrazin-1-yl]butan-2-ol,



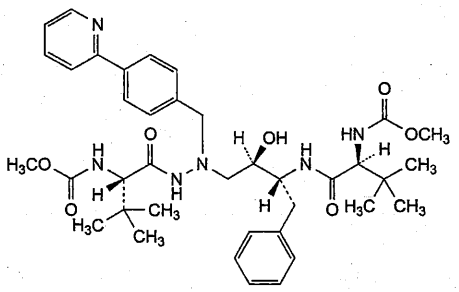
E. methyl [(5*S*,10*R*,11*S*,14*S*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-8-[[4-(pyridin-2-yl)phenyl]methyl]-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate,



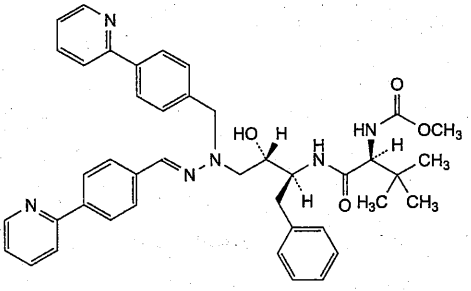
F. methyl [(5*R*,10*S*,11*S*,14*S*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-8-[[4-(pyridin-2-yl)phenyl]methyl]-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate,



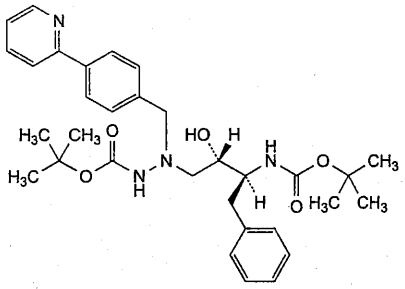
G. methyl [(5*S*,10*S*,11*S*,14*R*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-8-[[4-(pyridin-2-yl)phenyl]methyl]-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate,



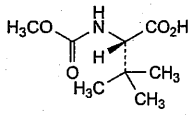
H. methyl [(5*S*,10*R*,11*R*,14*S*)-11-benzyl-5-*tert*-butyl-10-hydroxy-15,15-dimethyl-3,6,13-trioxo-8-[[4-(pyridin-2-yl)phenyl]methyl]-2-oxa-4,7,8,12-tetraazahexadecan-14-yl]carbamate,



I. methyl [(2*S*)-1-[[(2*S*,3*S*)-3-hydroxy-1-phenyl-4-[(*E*)-1-[[4-(pyridin-2-yl)phenyl]methyl]-2-[[4-(pyridin-2-yl)phenyl]methylidene]hydrazin-1-yl]butan-2-yl]amino]-3,3-dimethyl-1-oxobutan-2-yl]carbamate,



J. *tert*-butyl 2-[(2*S*,3*S*)-3-(*tert*-butoxyformamido)-2-hydroxy-4-phenylbutyl]-2-[[4-(pyridin-2-yl)phenyl]methyl] hydrazine-1-carboxylate,

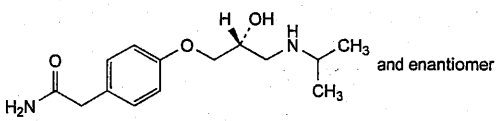


K. (2*S*)-2-(methoxyformamido)-3,3-dimethylbutanoic acid.

Ph Eur

Atenolol

(Ph. Eur. monograph 0703)



$C_{14}H_{22}N_2O_3$ 266.3 *29122-68-7*

Action and use
Beta-adrenoceptor antagonist.

Preparations
Atenolol Injection
Atenolol Oral Solution
Atenolol Tablets
Co-tenidone Tablets

Ph Eur

DEFINITION
2-[4-[(2*RS*)-2-Hydroxy-3-[(propan-2-yl)amino] propoxy]phenyl]acetamide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Sparingly soluble in water, soluble in anhydrous ethanol, slightly soluble in methylene chloride.

IDENTIFICATION
First identification: C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 152 °C to 155 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.100 g in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *methanol R*.

Spectral range 230-350 nm.

Absorption maxima At 275 nm and 282 nm.

Absorbance ratio A_{275}/A_{282} = 1.15 to 1.20.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *atenolol CRS*.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 1.0 mL of *methanol R*.

Reference solution Dissolve 10 mg of *atenolol CRS* in 1.0 mL of *methanol R*.

Plate *TLC silanised silica gel F_{254} plate R*.

Mobile phase *concentrated ammonia R1, methanol R* (1:99 *V/V*).

Application 10 µL.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Solution S
Dissolve 0.10 g in *water R* and dilute to 10 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than degree 6 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
+ 0.10° to −0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in 20 mL of the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *atenolol for system suitability CRS* (containing impurities B, F, G, I and J) in 1.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 1.0 g of *sodium octanesulfonate R* and 0.4 g of *tetrabutylammonium hydrogen sulfate R* in 1 L of a mixture of 20 volumes of *tetrahydrofuran R*, 180 volumes of *methanol R2* and 800 volumes of a 3.4 g/L solution of *potassium dihydrogen phosphate R*; adjust the apparent pH to 3.0 with *phosphoric acid R*.

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 226 nm.

Injection 10 µL.

Run time 5 times the retention time of atenolol.

Identification of impurities Use the chromatogram supplied with *atenolol for system suitability CRS* and the chromatogram

obtained with reference solution (a) to identify the peaks due to impurities B, F, G, I and J.

Relative retention With reference to atenolol (retention time = about 8 min): impurity B = about 0.3; impurity J = about 0.7; impurity I = about 0.8; impurity F = about 2.0 (pair of peaks); impurity G = about 3.5.

System suitability Reference solution (a):
— *resolution*: minimum 1.4 between the peaks due to impurities J and I.

Limits:
— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurities F, G, I, J*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 0.1 per cent.

Dissolve 50 mg in a mixture of 1 mL of *dilute nitric acid R* and 15 mL of *water R*. The solution, without further addition of *dilute nitric acid R*, complies with the test.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

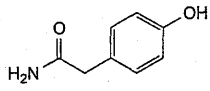
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 80 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

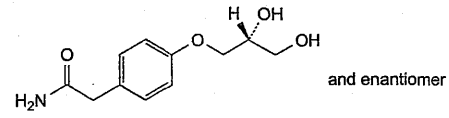
1 mL of *0.1 M perchloric acid* is equivalent to 26.63 mg of $C_{14}H_{22}N_2O_3$.

IMPURITIES
Specified impurities *B, F, G, I, J.*

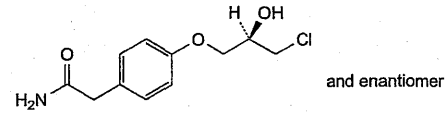
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, H.*



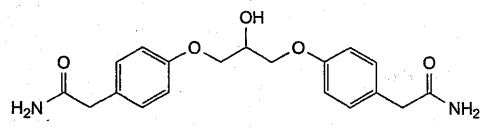
A. 2-(4-hydroxyphenyl)acetamide,



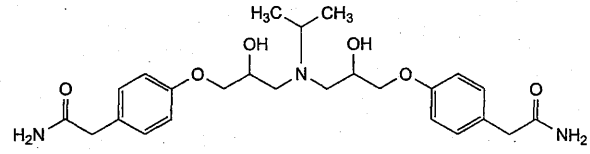
B. 2-[4-[(2*RS*)-2,3-dihydroxypropoxy]phenyl]acetamide,



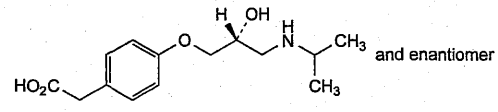
D. 2-[4-[(2*RS*)-3-chloro-2-hydroxypropoxy]phenyl]acetamide,



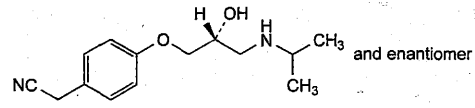
E. 2,2′-[(2-hydroxypropane-1,3-diyl)bis(oxy-4,1-phenylene)] diacetamide,



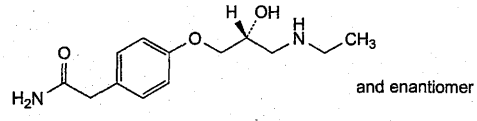
F. 2,2′-[[(propan-2-yl)azanediyl]bis[(2-hydroxypropane-3,1-diyl)oxy-4,1-phenylene]]diacetamide,



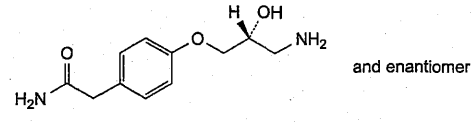
G. [4-[(2*RS*)-2-hydroxy-3-[(propan-2-yl)amino]propoxy] phenyl]acetic acid,



H. [4-[(2*RS*)-2-hydroxy-3-[(propan-2-yl)amino]propoxy] phenyl]acetonitrile,



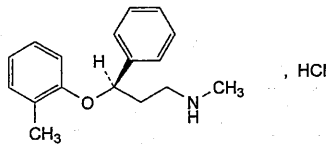
I. 2-[4-[(2*RS*)-3-(ethylamino)-2-hydroxypropoxy]phenyl] acetamide,



J. 2-[4-[(2*RS*)-3-amino-2-hydroxypropoxy]phenyl]acetamide.

Atomoxetine Hydrochloride

(Ph. Eur. monograph 2640)



$C_{17}H_{22}ClNO$ 291.8 *82248-59-7*

Action and use
Noradrenaline reuptake inhibitor; treatment of attention deficit hyperactivity disorder (ADHD).

Ph Eur

DEFINITION

(3*R*)-*N*-Methyl-3-(2-methylphenoxy)-3-phenylpropan-1-amine hydrochloride.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Sparingly soluble in water, soluble in anhydrous ethanol, practically insoluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *atomoxetine hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

B. Isomeric purity (see Tests).

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Isomeric purity
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 35.0 mg of the substance to be examined in 2.5 mL of *anhydrous ethanol R*, sonicate until dissolution is complete and dilute to 10.0 mL with *heptane R*.

Reference solution (a) Dissolve 3.5 mg of *atomoxetine impurity B CRS* and 1 mg of *atomoxetine impurity D CRS* in 5 mL of *anhydrous ethanol R*, sonicate until dissolution is complete and dilute to 20.0 mL with *heptane R*.

Reference solution (b) Dissolve 35.0 mg of the substance to be examined in 2.5 mL of *anhydrous ethanol R*. Add 1.0 mL of reference solution (a) and dilute to 10.0 mL with *heptane R*.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *heptane R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *cellulose derivative of silica gel for chiral separation R* (5 µm).

Mobile phase Mix 1.5 mL of *diethylamine R*, 2.0 mL of *trifluoroacetic acid R* and 150.0 mL of *2-propanol R* and dilute to 1000 mL with *heptane R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 1.3 times the retention time of atomoxetine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and D.

Relative retention With reference to atomoxetine (retention time = about 12 min): impurity B = about 0.5; impurity D = about 0.6.

System suitability Reference solution (b):
— *resolution*: minimum 1.8 between the peaks due to impurities B and D.

Limits:
— *impurity B*: maximum 0.5 per cent;
— *impurity D*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *disregard limit*: the area of the peak due to impurity B in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak with a relative retention with reference to atomoxetine of about 0.7 (impurity A).

Related substances
Liquid chromatography (*2.2.29*).

Solution A Dissolve 5.9 g of *sodium octanesulfonate monohydrate R* in 1000 mL of a 2.9 g/L solution of *phosphoric acid R* previously adjusted to pH 2.5 with a 280 g/L solution of *potassium hydroxide R*.

Test solution (a) Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 7.5 mg of *3-(methylamino)-1-phenylpropan-1-ol R* (impurity H) and 5 mg of *mandelic acid R* (impurity E) in test solution (b) and dilute to 50 mL with test solution (b).

Reference solution (c) Dissolve 5 mg of *atomoxetine for impurity A identification CRS* in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (d) Dissolve 25.0 mg of *atomoxetine hydrochloride CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 40 °C.

Mobile phase *propanol R*, solution A (27:73 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (c).

Run time 2.5 times the retention time of atomoxetine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to

impurities E and H; use the chromatogram supplied with *atomoxetine for impurity A identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to atomoxetine (retention time = about 10 min): impurity E = about 0.2; impurity H = about 0.3; impurity A = about 0.7.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurities E and H.

Calculation of percentage contents:
— for each impurity, use the concentration of atomoxetine hydrochloride in reference solution (a).

Limits:
— *impurity A*: maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

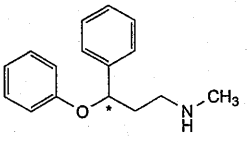
Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{17}H_{22}ClNO$ taking into account the assigned content of *atomoxetine hydrochloride CRS*.

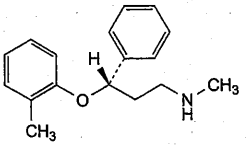
IMPURITIES

Specified impurities *A, B, D.*

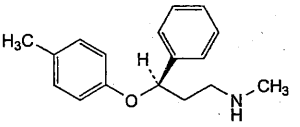
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, F, G, H.*



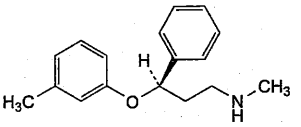
A. *N*-methyl-3-phenoxy-3-phenylpropan-1-amine,



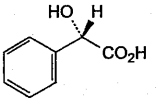
B. (3*S*)-*N*-methyl-3-(2-methylphenoxy)-3-phenylpropan-1-amine,



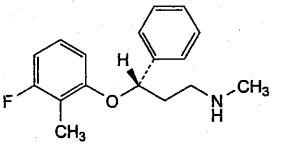
C. (3*R*)-*N*-methyl-3-(4-methylphenoxy)-3-phenylpropan-1-amine,



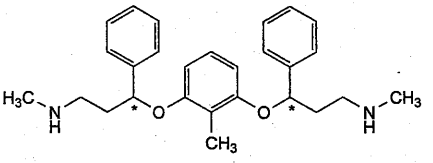
D. (3*R*)-*N*-methyl-3-(3-methylphenoxy)-3-phenylpropan-1-amine,



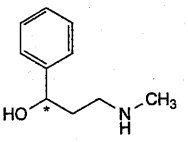
E. (2*S*)-2-hydroxy-2-phenylacetic acid (L-mandelic acid),



F. (3*S*)-3-(3-fluoro-2-methylphenoxy)-*N*-methyl-3-phenylpropan-1-amine,



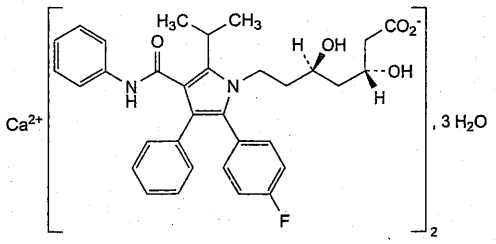
G. 3,3′-[(2-methylbenzene-1,3-diyl)bis(oxy)]bis(*N*-methyl-3-phenylpropan-1-amine),



H. 3-(methylamino)-1-phenylpropan-1-ol.

Atorvastatin Calcium Trihydrate

(Ph. Eur. monograph 2191)



$C_{66}H_{68}CaF_2N_4O_{10},3H_2O$ 1209 *344423-98-9*

Action and use
HMG Co-A reductase inhibitor; lipid-regulating drug.

Ph Eur

DEFINITION

Calcium (3*R*,5*R*)-7-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-3,5-dihydroxyheptanoate trihydrate.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *atorvastatin calcium trihydrate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Enantiomeric purity (see Tests).

C. Water (see Tests).

D. Ignite. The residue gives reaction (b) of calcium (*2.3.1*). Filtration may be necessary in case the residue does not completely dissolve.

TESTS

Enantiomeric purity
Liquid chromatography (*2.2.29*).

Solvent mixture *anhydrous ethanol R, methanol R* (50:50 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in 4 mL of the solvent mixture and dilute to 10.0 mL with *hexane R*.

Reference solution (a) Dissolve 2 mg of *atorvastatin impurity E CRS* in *methanol R* and dilute to 20.0 mL with the same solvent (solution A). Dissolve 10 mg of the substance to be examined in 1.25 mL of *methanol R*, add 0.75 mL of solution A and 2 mL of *anhydrous ethanol R* and dilute to 10.0 mL with *hexane R*.

Reference solution (b) To 2.0 mL of the test solution add 40.0 mL of the solvent mixture and dilute to 100.0 mL with *hexane R*. To 3.0 mL of this solution add 5 mL of the solvent mixture and dilute to 20.0 mL with *hexane R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *amylose derivative of silica gel for chromatography R* (10 µm).

Mobile phase *trifluoroacetic acid R, anhydrous ethanol R, hexane R* (0.1:6:94 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 244 nm.

Injection 20 µL.

Run time 1.2 times the retention time of atorvastatin.

Relative retention With reference to atorvastatin (retention time = about 44 min): impurity E = about 0.8.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity E and atorvastatin.

Limit:
— *impurity E*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 40.0 mg of the substance to be examined in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dissolve 50 mg of the substance to be examined in *dimethylformamide R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 40.0 mg of *atorvastatin calcium trihydrate CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (b) to 100.0 mL with *dimethylformamide R*. Dilute 1.0 mL of this solution to 10.0 mL with *dimethylformamide R*.

Reference solution (c) Dissolve 2.5 mg of *atorvastatin impurity A CRS*, 2.5 mg of *atorvastatin impurity B CRS*, 2.5 mg of *atorvastatin impurity C CRS*, 2.5 mg of *atorvastatin impurity D CRS* and 2.5 mg of the substance to be examined in *dimethylformamide R* and dilute to 50.0 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: *tetrahydrofuran R, acetonitrile R*, 3.9 g/L solution of *ammonium acetate R* adjusted to pH 5.0 with *glacial acetic acid R* (12:21:67 *V/V/V*);
— *mobile phase B*: *tetrahydrofuran R*, 3.9 g/L solution of *ammonium acetate R* adjusted to pH 5.0 with *glacial acetic acid R, acetonitrile R* (12:27:61 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 40	100	0
40 - 70	100 → 20	0 → 80
70 - 85	20 → 0	80 → 100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 244 nm.

Injection 20 µL of test solution (b) and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to atorvastatin (retention time = about 33 min): impurity A = about 0.8; impurity B = about 0.9; impurity C = about 1.2; impurity D = about 2.1.

If necessary, adjust the mobile phase by increasing or decreasing the percentage of acetonitrile or the pH of the ammonium acetate solution to achieve a retention time of about 33 min for atorvastatin. For example, raising the pH would decrease the retention time of atorvastatin.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity B and atorvastatin.

Limits:
— *impurities A, B*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurities C, D*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to dimethylformamide.

Sodium

Maximum 0.4 per cent (anhydrous substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Solvent mixture *hydrochloric acid R, water R, methanol R* (2:25:75 *V/V/V*).

Test solution Dissolve 5.0 mg in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solutions Prepare the reference solutions using *sodium standard solution (50 ppm Na) R*, diluting with the solvent mixture.

Source Sodium hollow-cathode lamp.

Wavelength 589.0 nm.

Atomisation device Air-acetylene flame.

Water (*2.5.12*)

3.5 per cent to 5.5 per cent, determined on 0.130 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

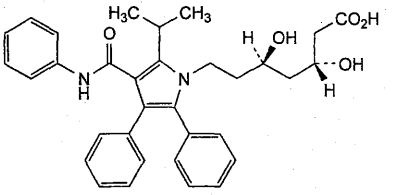
Injection Test solution (a) and reference solution (a).

Calculate the percentage content of $C_{66}H_{68}CaF_2N_4O_{10}$ from the declared content of *atorvastatin calcium trihydrate CRS*.

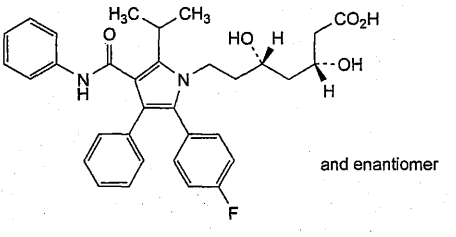
IMPURITIES

Specified impurities *A, B, C, D, E.*

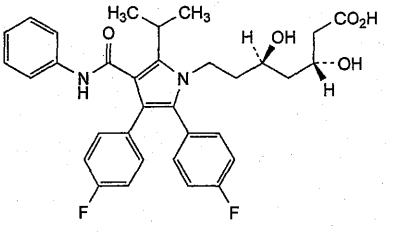
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G, H.*



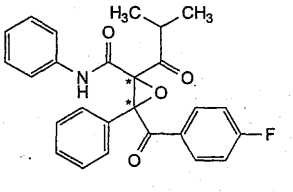
A. (3*R*,5*R*)-3,5-dihydroxy-7-[5-(1-methylethyl)-2,3-diphenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]heptanoic acid (desfluoroatorvastatin),



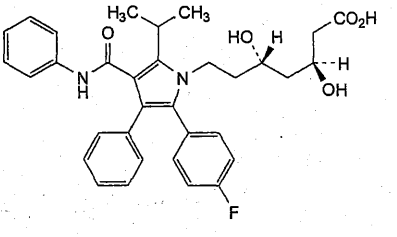
B. (3*RS*,5*SR*)-7-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-3,5-dihydroxyheptanoic acid,



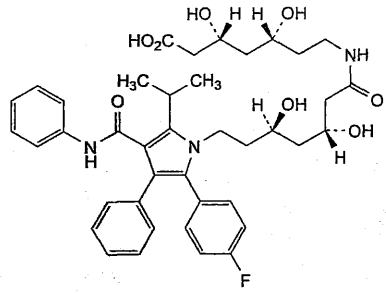
C. (3*R*,5*R*)-7-[2,3-bis(4-fluorophenyl)-5-(1-methylethyl)-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-3,5-dihydroxyheptanoic acid (fluoroatorvastatin),



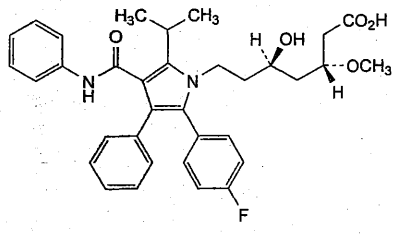
D. 3-[(4-fluorophenyl)carbonyl]-2-(2-methylpropanoyl)-*N*,3-diphenyloxirane-2-carboxamide,



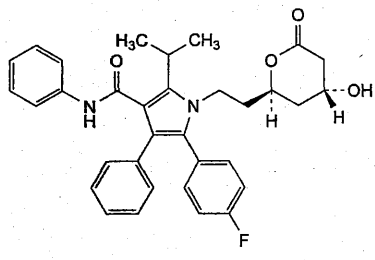
E. (3*S*,5*S*)-7-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-3,5-dihydroxyheptanoic acid (*ent*-atorvastatin),



F. (3*R*,5*R*)-7-[[(3*R*,5*R*)-7-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-3,5-dihydroxyheptanoyl]amino]-3,5-dihydroxyheptanoic acid,



G. (3*R*,5*R*)-7-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]-5-hydroxy-3-methoxyheptanoic acid (3-*O*-methylatorvastatin),

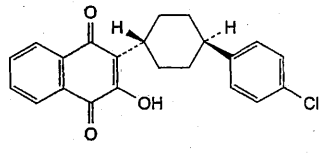


H. (4*R*,6*R*)-6-[2-[2-(4-fluorophenyl)-5-(1-methylethyl)-3-phenyl-4-(phenylcarbamoyl)-1*H*-pyrrol-1-yl]ethyl]-4-hydroxytetrahydro-2*H*-pyran-2-one.

Ph Eur

Atovaquone

(Ph. Eur. monograph 2192)



$C_{22}H_{19}ClO_3$ 366.8 *95233-18-4*

Action and use
Antiprotozoal (malaria).

Ph Eur

DEFINITION
2-[*trans*-4-(4-Chlorophenyl)cyclohexyl]-3-hydroxynaphthalene-1,4-dione.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
Yellow, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, very slightly soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *atovaquone CRS.*

If the spectra obtained show differences, dissolve 0.1 g of the substance to be examined and 0.1 g of the reference substance separately in 2.5 mL of a 50 g/L solution of *potassium hydroxide R* in *methanol R*. Filter the solutions and add each filtrate dropwise to a mixture of 0.8 mL of *acetic acid R* and 1.5 mL of *methanol R*, stirring continuously. Filter, wash the residues with *methanol R* and then with *water R*, and dry under vacuum at 55 °C. Record new spectra using the residues.

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture *water R, acetonitrile R1* (20:80 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of *atovaquone CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.5 mg of *atovaquone for system suitability CRS* (containing impurities B and C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *phosphoric acid R, methanol R2, water for chromatography R, acetonitrile R1* (0.5:17.5:30:52.5 *V/V/V/V*).

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of atovaquone.

Identification of impurities Use the chromatogram supplied with *atovaquone for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

Relative retention With reference to atovaquone (retention time = about 15 min): impurity B = about 0.85; impurity C = about 0.90.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity C and atovaquone;
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to impurity B.

Calculation of percentage contents:
— for each impurity, use the concentration of atovaquone in reference solution (c).

Limits:
— *impurity B*: maximum 0.5 per cent;
— *impurity C*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.6 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)
Maximum 0.3 per cent, determined on 0.100 g using the evaporation technique:
— *temperature*: 160 °C;
— *heating time*: 3 min;
— *flow rate*: 50 mL/min.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

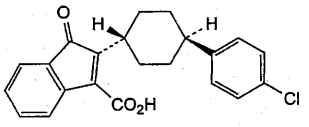
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

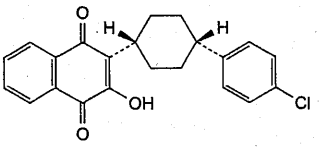
Calculate the percentage content of $C_{22}H_{19}ClO_3$ taking into account the assigned content of *atovaquone CRS*.

IMPURITIES
Specified impurities *B, C.*

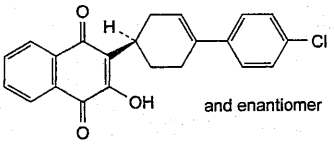
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D.*



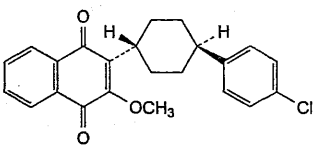
A. 2-[*trans*-4-(4-chlorophenyl)cyclohexyl]-1-oxo-1*H*-indene-3-carboxylic acid,



B. 2-[*cis*-4-(4-chlorophenyl)cyclohexyl]-3-hydroxynaphthalene-1,4-dione,



C. 2-[(1*RS*)-4-(4-chlorophenyl)cyclohex-3-en-1-yl]-3-hydroxynaphthalene-1,4-dione,

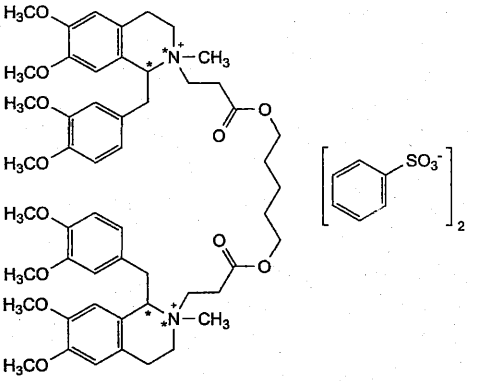


D. 2-[*trans*-4-(4-chlorophenyl)cyclohexyl]-3-methoxynaphthalene-1,4-dione.

Ph Eur

Atracurium Besilate

(Ph. Eur. monograph 1970)



$C_{65}H_{82}N_2O_{18}S_2$ 1243 *64228-81-5*

Action and use
Non-depolarizing neuromuscular blocker.

Ph Eur

DEFINITION
Mixture of the *cis-cis*, *cis-trans* and *trans-trans* isomers of 2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-1,3-diyl)]]bis[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium] dibenzenesulfonate.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION
It is considered that alkyl benzenesulfonate esters are genotoxic and are potential impurities in atracurium besilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general method *2.5.41. Methyl, ethyl and isopropyl benzenesulfonate in active substances* is available to assist manufacturers.

CHARACTERS
Appearance
White or yellowish-white, slightly hygroscopic powder.

Solubility
Soluble in water, very soluble in acetonitrile, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *atracurium besilate CRS.*

B. Examine the chromatograms obtained in the assay.

Results The 3 principal isomeric peaks in the chromatogram obtained with test solution (a) are similar in retention time to

those in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 1.00 g in *water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 50.0 mg of *atracurium besilate CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with mobile phase A.

Reference solution (c) Dissolve 20.0 mg of *methyl benzenesulfonate R* in *acetonitrile R* and dilute to 100.0 mL with the same solvent. Dilute 50 µL of the solution to 100.0 mL with mobile phase A.

Reference solution (d) Dissolve 2.0 mg of *atracurium for peak identification CRS* (containing impurities A1, A2, B, C1, C2, D1, D2, E, G and K) in 2.0 mL of mobile phase A.

Reference solution (e) Dissolve 2.0 mg of *atracurium for impurity F identification CRS* in 2.0 mL of mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: mix 5 volumes of *methanol R*, 20 volumes of *acetonitrile R* and 75 volumes of a 10.2 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.1 with *phosphoric acid R*;
- *mobile phase B*: mix 20 volumes of *acetonitrile R*, 30 volumes of *methanol R* and 50 volumes of a 10.2 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.1 with *phosphoric acid R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	80	20
5 - 15	80 → 40	20 → 60
15 - 25	40	60
25 - 30	40 → 0	60 → 100
30 - 45	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b), (d) and (e).

Identification of impurities Use the chromatogram obtained with reference solution (d) and the chromatogram supplied with *atracurium for peak identification CRS* to identify the peaks due to impurities A1, A2, B, C1, C2, D1, D2, E, G and K; use the chromatogram obtained with reference solution (e) and the chromatogram supplied with *atracurium for impurity F identification CRS* to identify the peak due to impurity F.

Relative retention With reference to the atracurium *cis-cis* isomer (retention time = about 30 min): impurity E = about 0.2; impurity F = about 0.25; impurity G = about 0.3; impurity D1 = about 0.45; impurity D2 = about 0.5; atracurium *trans-trans* isomer = about 0.8; atracurium *cis-trans* isomer = about 0.9; impurity A1 = about 1.04; impurity I1 = about 1.07; impurity H1 = about 1.07 (shoulder on the front of peak A2); impurity A2 (major isomer) = about 1.08; impurity K1 = about 1.09 (shoulder on the tail of peak A2); impurity I2 (major isomer) = about 1.12; impurity H2 (major isomer) = about 1.12; impurity K2 (major isomer) = about 1.12; impurity B = about 1.15; impurity C1 = about 1.2; impurity C2 (major isomer) = about 1.3.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to the atracurium *trans-trans* isomer and the atracurium *cis-trans* isomer, and minimum 1.5 between the peaks due to the atracurium *cis-trans* isomer and the atracurium *cis-cis* isomer in the chromatogram obtained with reference solution (a);
- *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak due to impurity A1 and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the atracurium *cis-cis* isomer in the chromatogram obtained with reference solution (d).

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity G by 0.5;
- *impurity E*: not more than 1.5 times the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (1.5 per cent);
- *impurities A, D*: for each impurity, for the sum of the areas of the 2 isomer peaks, not more than 1.5 times the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (1.5 per cent);
- *impurity C*: for the sum of the areas of the 2 isomer peaks, not more than the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *impurities F, G*: for each impurity, not more than the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *impurities H, I, K*: for the sum of the areas of the isomer peaks of these impurities, not more than the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *unspecified impurities*: for each impurity, not more than 0.1 times the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 3.5 times the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (3.5 per cent);

— *disregard limit*: 0.05 times the sum of the areas of the peaks due to the atracurium *cis-cis*, *trans-trans* and *cis-trans* isomers in the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity J

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	80	20
5 - 15	80 → 75	20 → 25
15 - 25	75	25
25 - 30	75 → 55	25 → 45
30 - 38	55 → 0	45 → 100
38 - 45	0	100

Detection Spectrophotometer at 217 nm.

Injection 100 µL of test solution (b) and reference solution (c).

Retention time Impurity J = about 25 min; atracurium *trans-trans* isomer = about 38 min.

Limit:

— *impurity J*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (10 ppm).

Isomer composition

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification. Use the normalisation procedure.

Injection Test solution (a).

Limits:

— *atracurium cis-cis isomer*: 55.0 per cent to 60.0 per cent,
— *atracurium cis-trans isomer*: 34.5 per cent to 38.5 per cent,
— *atracurium trans-trans isomer*: 5.0 per cent to 6.5 per cent.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (a) and reference solution (a).

Calculate the percentage content of $C_{65}H_{82}N_2O_{18}S_2$ from the sum of the areas of the peaks due to the 3 isomers.

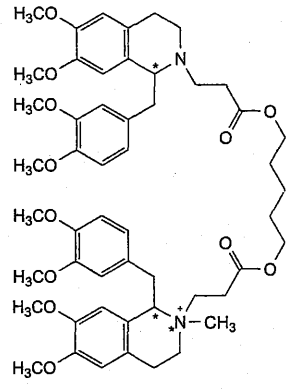
STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

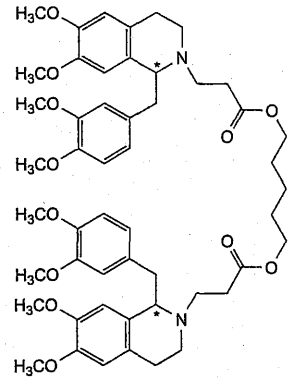
IMPURITIES

Specified impurities *A, C, D, E, F, G, H, I, J, K.*

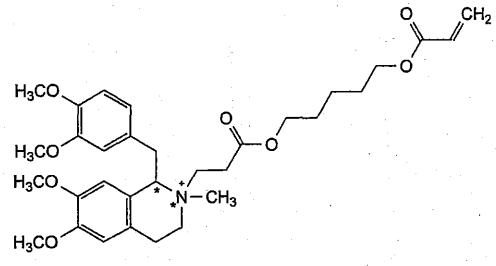
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



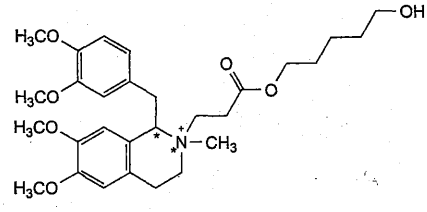
A. 1-(3,4-dimethoxybenzyl)-2-[13-[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl]-3,11-dioxo-4,10-dioxatridecyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium (A1 = *trans* isomer, A2 = *cis* isomer),



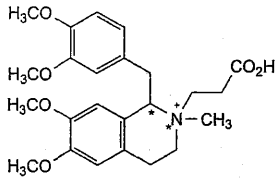
B. pentane-1,5-diyl bis[3-[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl]propanoate],



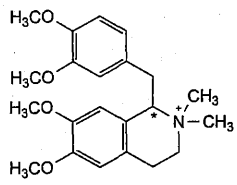
C. 1-(3,4-dimethoxybenzyl)-2-(3,11-dioxo-4,10-dioxatridec-12-enyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium (C1 = *trans* isomer, C2 = *cis* isomer),



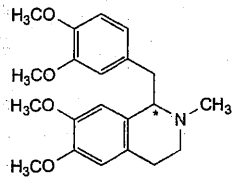
D. 1-(3,4-dimethoxybenzyl)-2-[3-[(5-hydroxypentyl)oxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium (D1 = *trans* isomer, D2 = *cis* isomer),



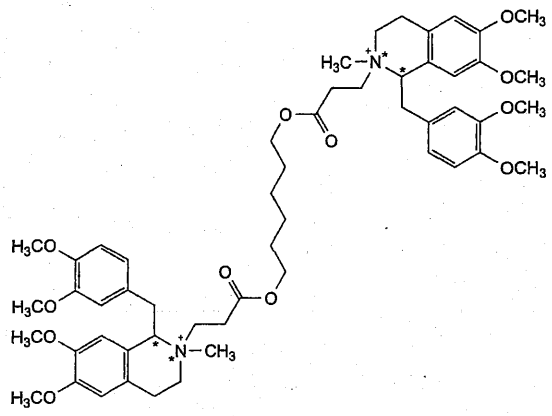
E. 2-(2-carboxyethyl)-1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



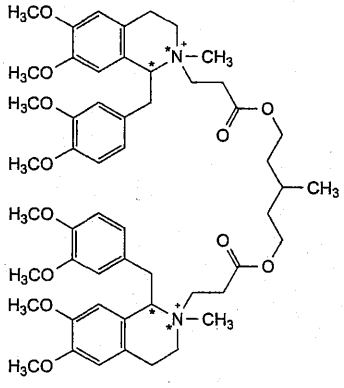
F. 1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2,2-dimethyl-1,2,3,4-tetrahydroisoquinolinium,



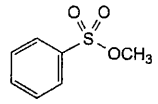
G. 1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinoline,



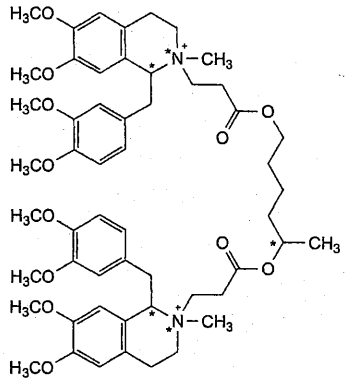
H. 2,2′-[hexane-1,6-diylbis[oxy(3-oxopropane-1,3-diyl)]]bis[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium] (H1 = *cis-trans* isomer, H2 = *cis-cis* isomer),



I. 2,2′-[(3-methylpentane-1,5-diyl)bis[oxy(3-oxopropane-1,3-diyl)]]bis[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium] (I1 = *cis-trans* isomer, I2 = *cis-cis* isomer),



J. methyl benzenesulfonate,

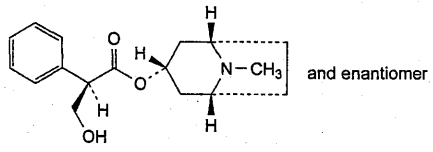


K. 2,2′-[hexane-1,5-diylbis[oxy(3-oxopropane-1,3-diyl)]]bis[1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium].

Ph Eur

Atropine

(Ph. Eur. monograph 2056)



$C_{17}H_{23}NO_3$ 289.4 *51-55-8*

Action and use
Anticholinergic.

Ph Eur

DEFINITION

(1*R*,3*R*,5*S*)-8-Methyl-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: A, B, E.

Second identification: A, C, D, E.

A. Melting point (*2.2.14*): 115 °C to 119 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison atropine CRS.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *atropine CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, water R, acetone R* (3:7:90 *V/V/V*).

Application 10 µL.

Development Over half of the plate.

Drying At 100-105 °C for 15 min.

Detection After cooling, spray with *dilute potassium iodobismuthate solution R.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Place about 3 mg in a porcelain crucible and add 0.2 mL of *fuming nitric acid R.* Evaporate to dryness on a water-bath. Dissolve the residue in 0.5 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*; a violet colour develops.

E. Optical rotation (see Tests).

TESTS

Optical rotation (*2.2.7*)

−0.70° to + 0.05° (measured in a 2 dm tube).

Dissolve 1.25 g in *ethanol (96 per cent) R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 24 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *atropine impurity B CRS* in the test solution and dilute to 20.0 mL with the test solution. Dilute 5.0 mL of this solution to 25.0 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of *atropine for peak identification CRS* (containing impurities A, D, E, F, G and H) in 1.0 mL of mobile phase A.

Reference solution (d) Dissolve 5 mg of *tropic acid R* (impurity C) in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R* (3 µm).

Mobile phase:
- *mobile phase A*: dissolve 3.5 g of *sodium dodecyl sulfate R* in 606 mL of a 7.0 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.3 with a 5.8 g/L solution of *phosphoric acid R*, and mix with 320 mL of *acetonitrile R1*;
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	95	5
2 - 20	95 → 70	5 → 30

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *atropine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, D, E, F, G and H; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to atropine (retention time = about 11 min): impurity C = about 0.2; impurity E = about 0.67; impurity D = about 0.73; impurity F = about 0.8; impurity B = about 0.89; impurity H = about 0.93; impurity G = about 1.1; impurity A = about 1.7.

System suitability Reference solution (b):
- *resolution*: minimum 2.5 between the peaks due to impurity B and atropine.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity C = 0.6;
- *impurities E, H*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurities A, B, C, D, F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.2 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

ASSAY

Dissolve 0.250 g in 40 mL of *anhydrous acetic acid R*, heating if necessary, and allow to cool. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

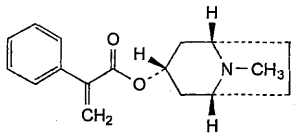
1 mL of *0.1 M perchloric acid* is equivalent to 28.94 mg of $C_{17}H_{23}NO_3$.

STORAGE

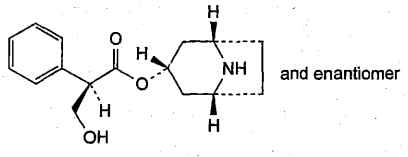
Protected from light.

IMPURITIES

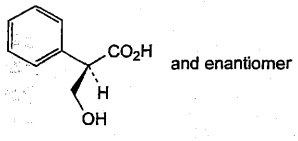
Specified impurities *A, B, C, D, E, F, G, H.*



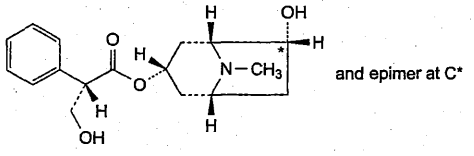
A. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl 2-phenylpropenoate (apoatropine),



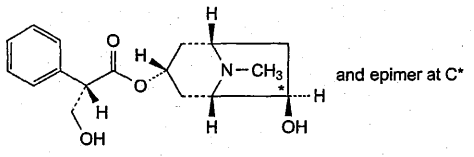
B. (1*R*,3*r*,5*S*)-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate (noratropine),



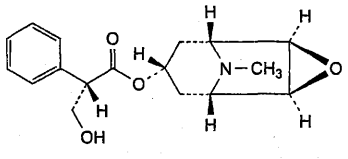
C. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (tropic acid),



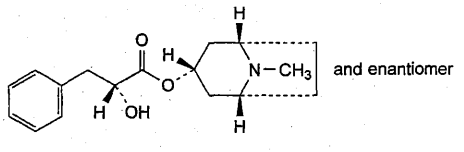
D. (1*R*,3*S*,5*R*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (6-hydroxyhyoscyamine),



E. (1*S*,3*R*,5*S*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (7-hydroxyhyoscyamine),



F. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscine),



G. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-2-hydroxy-3-phenylpropanoate (littorine),

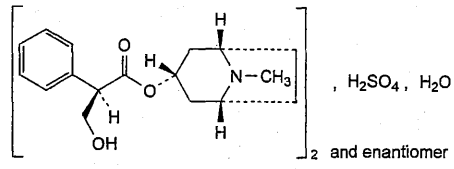
H. unknown structure.

Ph Eur

Atropine Sulfate

Atropine Sulphate

(Ph. Eur. monograph 0068)



$C_{34}H_{48}N_2O_{10}S,H_2O$ 695 *5908-99-6*

Action and use
Anticholinergic.

Preparations
Atropine Eye Drops
Atropine Eye Ointment
Atropine Injection
Atropine Tablets

Ph Eur

DEFINITION

Bis[(1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[*3.2.1*]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate] sulfate monohydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, E.

Second identification: C, D, E, F.

A. Optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison atropine sulfate CRS.

C. Dissolve about 50 mg in 5 mL of *water R* and add 5 mL of *picric acid solution R*. The precipitate, washed with *water R* and dried at 100-105 °C for 2 h, melts (*2.2.14*) at 174 °C to 179 °C.

D. To about 1 mg add 0.2 mL of *fuming nitric acid R* and evaporate to dryness in a water-bath. Dissolve the residue in 2 mL of *acetone R* and add 0.1 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*. A violet colour develops.

E. It gives the reactions of sulfates (*2.3.1*).

F. It gives the reaction of alkaloids (*2.3.1*).

TESTS

pH (*2.2.3*)
4.5 to 6.2.

Dissolve 0.6 g in *carbon dioxide-free water R* and dilute to 30 mL with the same solvent.

Optical rotation (*2.2.7*)
−0.50° to + 0.05° (measured in a 2 dm tube).

Dissolve 2.50 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 24 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *atropine impurity B CRS* in the test solution and dilute to 20 mL with the test solution. Dilute 5 mL of this solution to 25 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of *atropine for peak identification CRS* (containing impurities A, D, E, F, G and H) in 1 mL of mobile phase A.

Reference solution (d) Dissolve 5 mg of *tropic acid R* (impurity C) in mobile phase A and dilute to 10 mL with mobile phase A. Dilute 1 mL of the solution to 100 mL with mobile phase A. Dilute 1 mL of this solution to 10 mL with mobile phase A.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R* (3 µm).

Mobile phase:
— *mobile phase A*: dissolve 3.5 g of *sodium dodecyl sulfate R* in 606 mL of a 7.0 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.3 with a 5.8 g/L solution of *phosphoric acid R*, and mix with 320 mL of *acetonitrile R1*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	95	5
2 - 20	95 → 70	5 → 30

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *atropine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, D, E, F, G and H. Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B, and use the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to atropine (retention time = about 11 min): impurity C = about 0.2; impurity E = about 0.67; impurity D = about 0.73; impurity F = about 0.8; impurity B = about 0.89; impurity H = about 0.93; impurity G = about 1.1; impurity A = about 1.7.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurity B and atropine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity C = 0.6;
— *impurities E, H*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities A, B, C, D, F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
2.0 per cent to 4.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.500 g in 30 mL of *anhydrous acetic acid R*, warming if necessary. Cool the solution. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

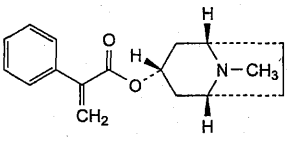
1 mL of *0.1 M perchloric acid* is equivalent to 67.68 mg of $C_{34}H_{48}N_2O_{10}S$.

STORAGE

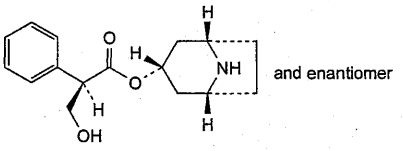
Protected from light.

IMPURITIES

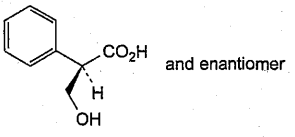
Specified impurities *A, B, C, D, E, F, G, H.*



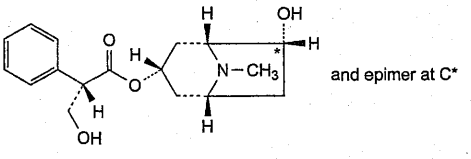
A. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[*3.2.1*]oct-3-yl 2-phenylpropenoate (apoatropine),



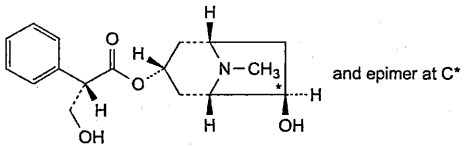
B. (1*R*,3*r*,5*S*)-8-azabicyclo[*3.2.1*]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate (noratropine),



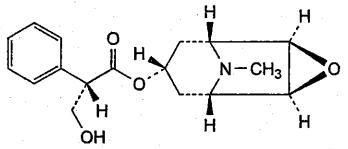
C. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (tropic acid),



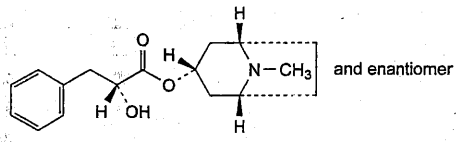
D. (1*R*,3*S*,5*R*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[*3.2.1*]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (6-hydroxyhyoscyamine),



E. (1*S*,3*R*,5*S*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[*3.2.1*]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (7-hydroxyhyoscyamine),



F. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscine),



G. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[*3.2.1*]oct-3-yl (2*RS*)-2-hydroxy-3-phenylpropanoate (littorine).

H. unknown structure.

Ph Eur

Attapulgite

Action and use
Excipient.

DEFINITION
Attapulgite is a purified native hydrated magnesium aluminium silicate essentially consisting of the clay mineral palygorskite.

CHARACTERISTICS
A light, cream or buff, very fine powder, free or almost free from gritty particles.

IDENTIFICATION
A. Ignite 0.5 g with 2 g of *anhydrous sodium carbonate* for 20 minutes, cool and extract with 25 mL of boiling *water*. Cool, filter, wash the residue with *water* and add the washings to the filtrate. Reserve the residue for test B. Cautiously acidify the combined filtrate and washings with *hydrochloric acid*, evaporate to dryness, moisten the residue with 0.2 mL of *hydrochloric acid*, add 10 mL of *water* and stir. A white, gelatinous precipitate is produced.

B. Wash the residue reserved in test A with *water* and dissolve in 10 mL of 2M *hydrochloric acid*. To 2 mL of the solution add a 10% w/v solution of *ammonium thiocyanate*. An intense red colour is produced.

C. To 2 mL of the solution obtained in test B add 1 mL of *strong sodium hydroxide solution* and filter. To the filtrate add 3 mL of *ammonium chloride solution*. A gelatinous white precipitate is produced.

D. To 2 mL of the solution obtained in test B add *ammonium chloride* and an excess of 13.5M *ammonia* and filter. To the filtrate add 0.15 mL of *magneson reagent* and an excess of 5M *sodium hydroxide*. A blue precipitate is produced.

TESTS
Acidity or alkalinity
pH of a 5% w/v suspension in *carbon dioxide-free water*, after shaking for 5 minutes, 7.0 to 9.5, Appendix V L.

Adsorptive capacity
Moisture adsorption, 5 to 14% when determined by the following method. Dry in air and powder a sufficient quantity of the substance being examined and pass through a sieve with a nominal mesh aperture of 150 µm. Spread 0.5 g as a thin layer on a previously weighed piece of aluminium foil (60 mm × 50 mm) of nominal gauge 17.5 µm and transfer to a desiccator containing a dish of sodium chloride crystals partially immersed in saturated brine at 25°. After 4 hours, remove from the desiccator and weigh immediately. Dry in an oven at 110° for 4 hours, allow to cool in a desiccator and weigh. The *moisture adsorption* is the gain in weight of the substance being examined expressed as a percentage of its oven-dried weight.

Arsenic
To 0.13 g add 5 mL of *water*, 2 mL of *sulfuric acid* and 10 mL of *sulfur dioxide solution* and evaporate on a water bath until the sulfur dioxide solution is removed and the volume reduced to about 2 mL. Transfer the solution to the generator flask with the aid of 5 mL of *water*. The resulting solution complies with the *limit test for arsenic*, Appendix VII (8 ppm).

Acid-soluble matter
Boil 2 g with 100 mL of *0.2M hydrochloric acid* under a reflux condenser for 5 minutes, cool and filter. Evaporate 50 mL of the filtrate to dryness. The residue, after ignition at about 600° for 30 minutes, weighs not more than 0.25 g.

Water-soluble matter
Boil 10 g with 100 mL of *water* under a reflux condenser for 5 minutes, cool and filter. Evaporate 50 mL of the filtrate to dryness. The residue, after ignition at 600° for 30 minutes, weighs not more than 50 mg.

Loss on drying
When dried to constant weight at 105°, loses not more than 17.0% of its weight. Use 1 g.

Loss on ignition
When ignited at 600°, loses 15.0 to 27.0% of its weight. Use 1 g.

Activated Attapulgite

Action and use
Antidiarrhoeal.

DEFINITION
Activated Attapulgite is a purified native hydrated magnesium aluminium silicate essentially consisting of the clay mineral palygorskite that has been carefully heated to increase its adsorptive capacity.

CHARACTERISTICS
A light, cream or buff, very fine powder, free or almost free from gritty particles.

IDENTIFICATION
A. Ignite 0.5 g with 2 g of *anhydrous sodium carbonate* for 20 minutes, cool and extract with 25 mL of boiling *water*. Cool, filter, wash the residue with *water* and add the washings to the filtrate. Reserve the residue for test B. Cautiously acidify the combined filtrate and washings with

hydrochloric acid, evaporate to dryness, moisten the residue with 0.2 mL of *hydrochloric acid*, add 10 mL of *water* and stir. A white, gelatinous precipitate is produced.

B. Wash the residue reserved in test A with *water* and dissolve in 10 mL of 2M *hydrochloric acid*. To 2 mL of the solution add a 10% w/v solution of *ammonium thiocyanate*. An intense red colour is produced.

C. To 2 mL of the solution obtained in test B add 1 mL of *strong sodium hydroxide solution* and filter. To the filtrate add 3 mL of *ammonium chloride solution*. A gelatinous white precipitate is produced.

D. To 2 mL of the solution obtained in test B add *ammonium chloride* and an excess of 13.5M *ammonia* and filter. To the filtrate add 0.15 mL of *magneson reagent* and an excess of 5M *sodium hydroxide*. A blue precipitate is produced.

TESTS

Acidity or alkalinity

pH of a 5% w/v suspension in *carbon dioxide-free water*, after shaking for 5 minutes, 7.0 to 9.5, Appendix V L.

Arsenic

To 0.13 g add 5 mL of *water*, 2 mL of *sulfuric acid* and 10 mL of *sulfur dioxide solution* and evaporate on a water bath until the sulfur dioxide solution is removed and the volume reduced to about 2 mL. Transfer the solution to the generator flask with the aid of 5 mL of *water*. The resulting solution complies with the *limit test for arsenic*, Appendix VII (8 ppm).

Acid-soluble matter

Boil 2 g with 100 mL of *0.2M hydrochloric acid* under a reflux condenser for 5 minutes, cool and filter. Evaporate 50 mL of the filtrate to dryness. The residue, after ignition at about 600° for 30 minutes, weighs not more than 0.25 g.

Water-soluble matter

Boil 10 g with 100 mL of *water* under a reflux condenser for 5 minutes, cool and filter. Evaporate 50 mL of the filtrate to dryness. The residue, after ignition at 600° for 30 minutes, weighs not more than 50 mg.

Adsorptive capacity

In a stoppered bottle shake 1.0 g, in *very fine powder*, with 50 mL of a 0.12% w/v solution of *methylene blue* for 5 minutes, allow to settle and centrifuge. The colour of the clear supernatant solution is not more intense than that of a 0.0012% w/v solution of *methylene blue*.

Loss on drying

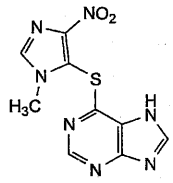
When dried to constant weight at 105°, loses not more than 4.0% of its weight. Use 1 g.

Loss on ignition

When ignited at 600°, loses not more than 9.0% of its weight. Use 1 g.

Azathioprine

(Ph. Eur. monograph 0369)



$C_9H_7N_7O_2S$ 277.3 *446-86-6*

Action and use

Immunosuppressant.

Preparations

Azathioprine Tablets

Azathioprine Oral Suspension

Ph Eur

DEFINITION

6-[(1-Methyl-4-nitro-1*H*-imidazol-5-yl)sulfanyl]-7*H*-purine.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Pale-yellow powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It is soluble in dilute solutions of alkali hydroxides and sparingly soluble in dilute mineral acids.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *azathioprine CRS*.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solution A 2.76 g/L solution of *sodium dihydrogen phosphate monohydrate R* adjusted to pH 2.5 with *phosphoric acid R*.

Test solution Dissolve 10 mg of the substance to be examined in 35 mL of a 0.8 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with solution A.

Reference solution (a) Dissolve 5 mg of *azathioprine impurity A CRS* and 5 mg of *mercaptopurine R* (impurity B) in 8.75 mL of a 0.8 g/L solution of *sodium hydroxide R* and dilute to 25.0 mL with solution A. To 1.0 mL of this solution, add 35 mL of a 0.8 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with solution A.

Reference solution (b) Dissolve 2.5 mg of *azathioprine impurity G CRS* and 2.5 mg of the substance to be examined in 8.8 mL of a 0.8 g/L solution of *sodium hydroxide R* and dilute to 25.0 mL with solution A. To 1.0 mL of this solution, add 17.5 mL of a 0.8 g/L solution of *sodium hydroxide R* and dilute to 50.0 mL with solution A.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, solution A (5:95 *V/V*);
— *mobile phase B*: solution A, *methanol R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 15	100 → 0	0 → 100
15 - 20	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B. Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity G.

Relative retention With reference to azathioprine (retention time = about 15 min): impurity A = about 0.3; impurity B = about 0.4; impurity G = about 0.97.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to impurities A and B in the chromatogram obtained with reference solution (a); minimum 2.0 between the peaks due to impurity G and azathioprine in the chromatogram obtained with reference solution (b).

Limits:
— *impurities A, B*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 25 mL of *dimethylformamide R*. Titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 27.73 mg of $C_9H_7N_7O_2S$.

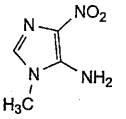
STORAGE

Protected from light.

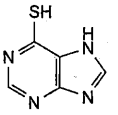
IMPURITIES

Specified impurities *A, B.*

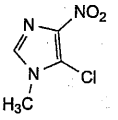
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, F, G.*



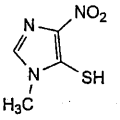
A. 1-methyl-4-nitro-1*H*-imidazol-5-amine,



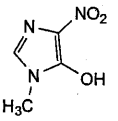
B. 7*H*-purine-6-thiol (mercaptopurine),



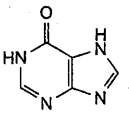
C. 5-chloro-1-methyl-4-nitro-1*H*-imidazole,



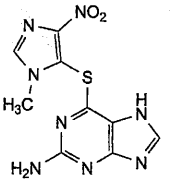
D. 1-methyl-4-nitro-1*H*-imidazole-5-thiol,



E. 1-methyl-4-nitro-1*H*-imidazol-5-ol,



F. 1,7-dihydro-6*H*-purin-6-one (hypoxanthine),

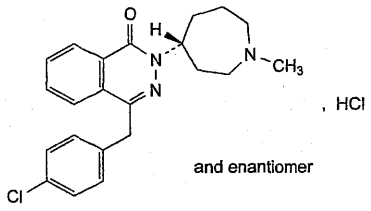


G. 6-[(1-methyl-4-nitro-1*H*-imidazol-5-yl)sulfanyl]-7*H*-purin-2-amine (thiamiprine).

Ph Eur

Azelastine Hydrochloride

(Ph. Eur. monograph 1633)



$C_{22}H_{25}Cl_2N_3O$ 418.4 *79307-93-0*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION

4-[(4-Chlorophenyl)methyl]-2-[(4*RS*)-1-methylhexahydro-1*H*-azepin-4-yl]phthalazin-1(2*H*)-one hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *azelastine hydrochloride CRS.*

B. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Acidity or alkalinity
To 10 mL of solution S add 0.2 mL of *bromothymol blue solution R1*. Not more than 0.1 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water for chromatography R* (45:55 *V/V*).

Test solution Dissolve 0.125 g of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 1 mg of *benzohydrazide R* (impurity A), 1 mg of *azelastine impurity B CRS*, 1 mg of *2-[2-(4-chlorophenyl)acetyl]benzoic acid R* (impurity C), 1 mg of *azelastine impurity D CRS* and 1 mg of *azelastine impurity E CRS* in the test solution and dilute to 20.0 mL with the test solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *cyanosilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 2.16 g of *sodium octanesulfonate R* and 0.68 g of *potassium dihydrogen phosphate R* in 740 mL of *water for chromatography R*; adjust to pH 3.0-3.1 with *dilute phosphoric acid R*, add 260 mL of *acetonitrile R1* and mix.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Run time 1.5 times the retention time of azelastine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and E. The elution order may vary, especially for impurity E, but the peak area of each impurity in reference solution (b) is different, so a clear identification of the impurities is possible.

Relative retention With reference to azelastine (retention time = about 10 min): impurity A = about 0.2; impurity B = about 0.3; impurity C = about 0.4; impurity D = about 0.6; impurity E = about 0.8.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to impurity E and azelastine and minimum 1.5 between the peaks due to impurities C and D in the chromatogram obtained with reference solution (b);
— *signal-to-noise ratio*: minimum 90 for the principal peak in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 2.6; impurity B = 4.5; impurity C = 2.0; impurity E = 2.8;
— for each impurity, use the concentration of azelastine in reference solution (a).

Limits:
— *impurities A, B, C, E*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.300 g in 5 mL of *anhydrous formic acid R*. Add 30 mL of *acetic anhydride R*. Titrate quickly with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

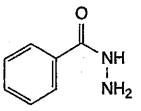
1.0 mL of *0.1 M perchloric acid* is equivalent to 41.84 mg of $C_{22}H_{25}Cl_2N_3O$.

IMPURITIES

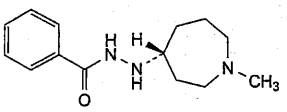
Specified impurities *A, B, C, E.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) D.

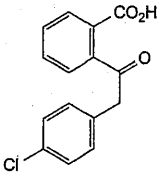


A. benzohydrazide (benzoyldiazane),

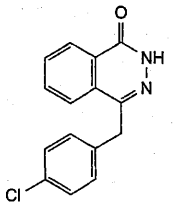


and enantiomer

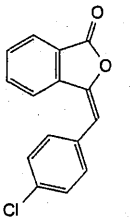
B. *N'*-[(4*RS*)-1-methylhexahydro-1*H*-azepin-4-yl]benzohydrazide,



C. 2-[2-(4-chlorophenyl)acetyl]benzoic acid,



D. 4-[(4-chlorophenyl)methyl]phthalazin-1(2*H*)-one,



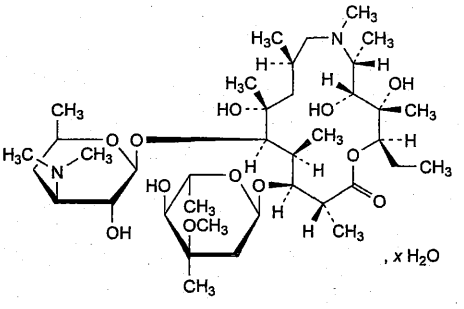
and (*Z*)-isomer

E. (3*E*)-3-[(4-chlorophenyl)methylidene]-2-benzofuran-1(3*H*)-one.

Ph Eur

Azithromycin

(Ph. Eur. monograph 1649)



$C_{38}H_{72}N_2O_{12}xH_2O$ with x = 1 or 2

749 (anhydrous substance)

Azithromycin monohydrate	*121470-24-4*
Azithromycin dihydrate	*117772-70-0*

Action and use
Macrolide antibacterial.

Preparations
Azithromycin Capsules
Azithromycin Eye Drops
Azithromycin for Infusion
Azithromycin Oral Suspension
Azithromycin Tablets

Ph Eur

DEFINITION

(2*R*,3*S*,4*R*,5*R*,8*R*,10*R*,11*R*,12*S*,13*S*,14*R*)-13-[(2,6-Dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-2-ethyl-3,4,10-trihydroxy-3,5,6,8,10,12,14-heptamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-1-oxa-6-azacyclopentadecan-15-one. The degree of hydration is 1 or 2.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison azithromycin CRS.

If the spectra obtained in the solid state show differences, prepare further spectra using 90 g/L solutions in *methylene chloride R.*

TESTS

Solution S
Dissolve 0.500 g in *anhydrous ethanol R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
9.0 to 11.0.

Dissolve 0.100 g in 25.0 mL of *methanol R* and dilute to 50.0 mL with *carbon dioxide-free water R*.

Specific optical rotation (*2.2.7*)
−49 to −45 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Prepare a 1.73 g/L solution of *ammonium dihydrogen phosphate R* adjusted to pH 10.0 with *ammonia R*. Transfer 350 mL of this solution to a suitable container. Add 300 mL of *acetonitrile R* and 350 mL of *methanol R*. Mix well.

Test solution Dissolve 0.200 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *azithromycin for system suitability CRS* (containing impurities F, H and J) in 1.0 mL of the solvent mixture and sonicate for 5 min.

Reference solution (c) Dissolve 8.0 mg of *azithromycin for peak identification CRS* (containing impurities A, B, C, E, F, G, I, J, L, M, N, O and P) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (5 µm);
— *temperature*: 60 °C.

Mobile phase:
— *mobile phase A*: 1.80 g/L solution of *anhydrous disodium hydrogen phosphate R* adjusted to pH 8.9 with *dilute phosphoric acid R* or with *dilute sodium hydroxide solution R*;
— *mobile phase B*: *methanol R2, acetonitrile R1* (25:75 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	50 → 45	50 → 55
25 - 30	45 → 40	55 → 60
30 - 80	40 → 25	60 → 75
80 - 81	25 → 50	75 → 50
81 - 93	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram supplied with *azithromycin for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, E, F, G, I, J, L, M, N, O and P; use the chromatogram supplied with *azithromycin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity H.

Relative retention With reference to azithromycin (retention time = 45-50 min): impurity L = about 0.29; impurity M = about 0.37; impurity E = about 0.43; impurity F = about 0.51; impurity D = about 0.54; impurity J = about 0.54; impurity Q = about 0.54; impurity I = about 0.61; impurity C = about 0.73; impurity N = about 0.76; impurity H = about 0.79; impurity A = about 0.83; impurity P = about 0.92; impurity O = about 1.23; impurity G = about 1.26; impurity B = about 1.31.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 1.4, where H_p = height above the baseline of the peak due to impurity J and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity F.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity F = 0.3; impurity G = 0.2; impurity H = 0.1; impurity L = 2.3; impurity M = 0.6; impurity N = 0.7;
— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent);
— *impurities A, C, E, F, H, I, L, M, N, O, P*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *sum of impurities D, J and Q*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity G*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); disregard the peaks eluting before impurity L and after impurity B.

Water (*2.5.12*)
1.8 per cent to 6.5 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*).

Solution A Mix 60 volumes of *acetonitrile R* and 40 volumes of a 6.7 g/L solution of *dipotassium hydrogen phosphate R* adjusted to pH 8.0 with *phosphoric acid R*.

Test solution Dissolve 53.0 mg of the substance to be examined in 2 mL of *acetonitrile R* and dilute to 100.0 mL with solution A.

Reference solution (a) Dissolve 53.0 mg of *azithromycin CRS* in 2 mL of *acetonitrile R* and dilute to 100.0 mL with solution A.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *azithromycin impurity A CRS* in 0.5 mL of *acetonitrile R* and dilute to 10 mL with solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl vinyl polymer for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 60 volumes of *acetonitrile R1* and 40 volumes of a 6.7 g/L solution of *dipotassium hydrogen*

phosphate R adjusted to pH 11.0 with a 560 g/L solution of *potassium hydroxide R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Run time 1.5 times the retention time of azithromycin.

Retention time Azithromycin = about 10 min.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to impurity A and azithromycin.

Calculate the percentage content of $C_{38}H_{72}N_2O_{12}$ taking into account the assigned content of *azithromycin CRS*.

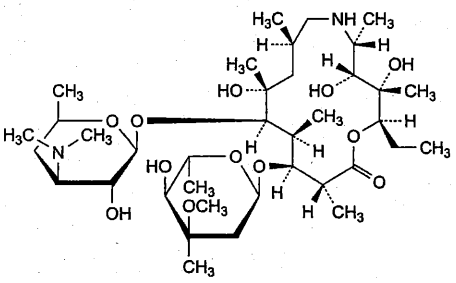
STORAGE

In an airtight container.

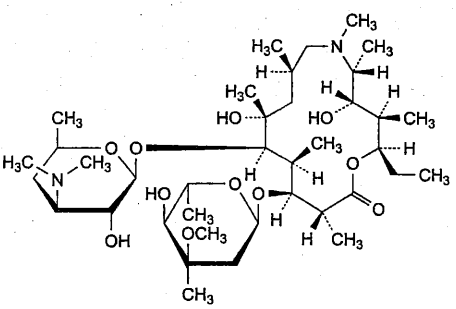
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H, I, J, L, M, N, O, P, Q.*

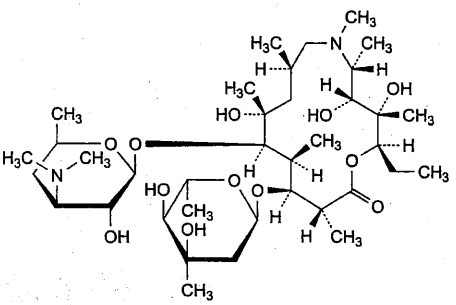
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) K.



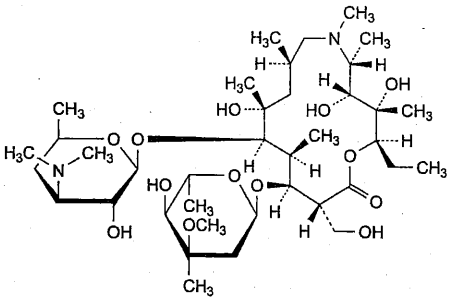
A. 6-demethylazithromycin,



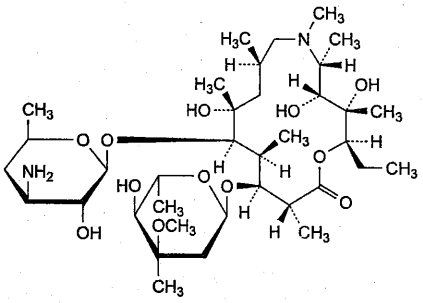
B. 3-deoxyazithromycin (azithromycin B),



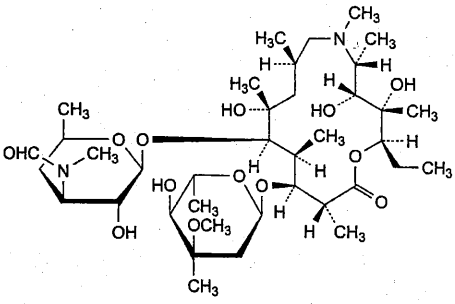
C. 3″-*O*-demethylazithromycin (azithromycin C),



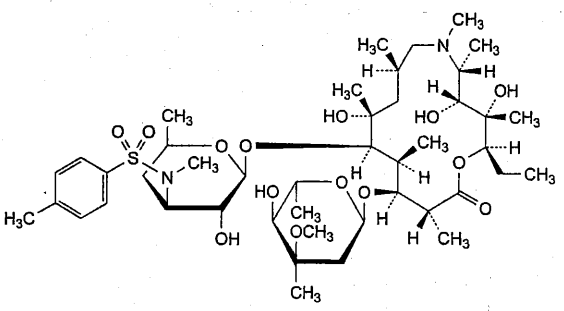
D. 14-demethyl-14-(hydroxymethyl)azithromycin (azithromycin F),



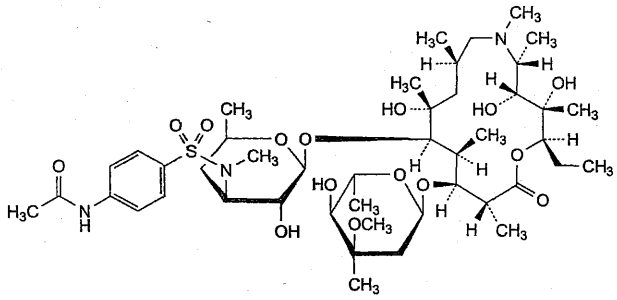
E. 3′-(*N*,*N*-didemethyl)azithromycin (aminoazithromycin),



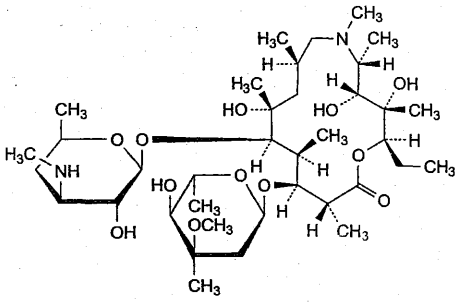
F. 3′-*N*-demethyl-3′-*N*-formylazithromycin,



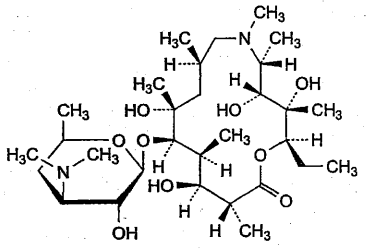
G. 3′-*N*-demethyl-3′-*N*-[(4-methylphenyl) sulfonyl]azithromycin,



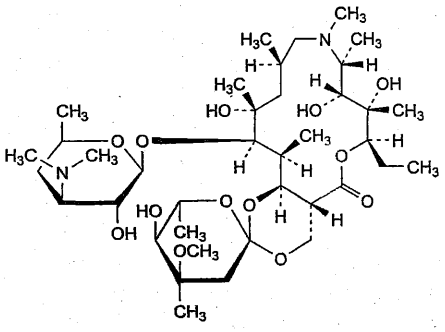
H. 3′-*N*-[[4-(acetylamino)phenyl]sulfonyl]-3′-*N*-demethylazithromycin,



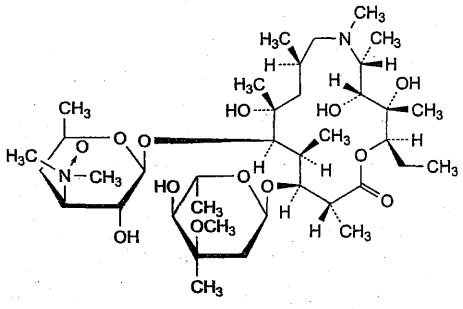
I. 3′-*N*-demethylazithromycin,



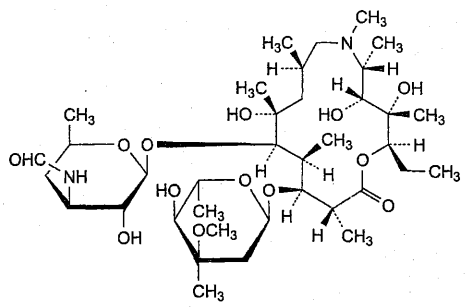
J. 13-*O*-decladinosylazithromycin,



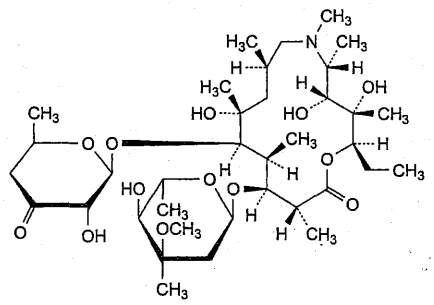
K. C^{14},1″-epoxyazithromycin (azithromycin E),



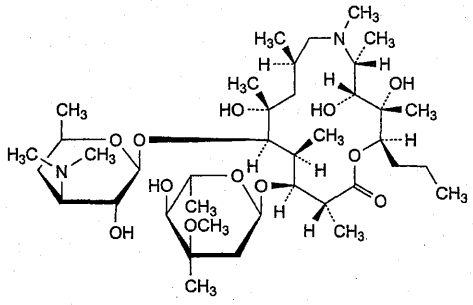
L. azithromycin 3′-*N*-oxide,



M. 3′-(*N*,*N*-didemethyl)-3′-*N*-formylazithromycin,

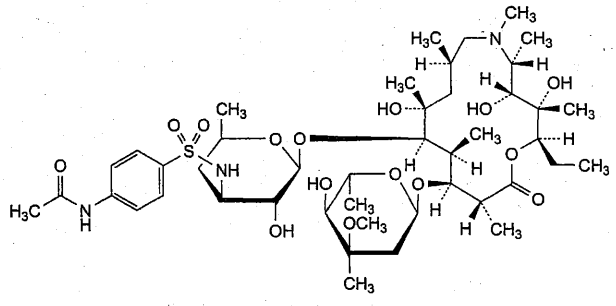


N. 3′-de(dimethylamino)-3′-oxoazithromycin,



O. 2-desethyl-2-propylazithromycin,

P. unknown structure,

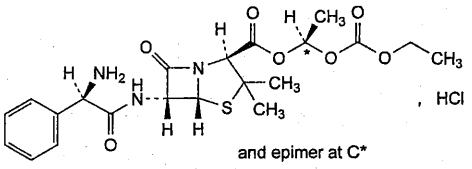


Q. 3′-*N*-[[4-(acetylamino)phenyl]sulfonyl]-3′-(*N*,*N*-didemethyl)azithromycin.

Ph Eur

Bacampicillin Hydrochloride

(Ph. Eur. monograph 0808)



$C_{21}H_{28}ClN_3O_7S$ 502.0 *37661-08-8*

Action and use
Penicillin antibacterial.

Ph Eur

DEFINITION

(1*RS*)-1-[(Ethoxycarbonyl)oxy]ethyl (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate hydrochloride.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder or granules, hygroscopic.

Solubility
Soluble in water, freely soluble in ethanol (96 per cent), soluble in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bacampicillin hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 2 mL of *methanol R.*

Reference solution (a) Dissolve 10 mg of *bacampicillin hydrochloride CRS* in 2 mL of *methanol R.*

Reference solution (b) Dissolve 10 mg of *bacampicillin hydrochloride CRS*, 10 mg of *talampicillin hydrochloride CRS* and 10 mg of *pivampicillin CRS* in 2 mL of *methanol R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 10 volumes of a 272 g/L solution of *sodium acetate R* adjusted to pH 5.0 with *glacial acetic acid R*, 40 volumes of *water R* and 50 volumes of *ethanol (96 per cent) R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In a current of warm air.

Detection Spray with *ninhydrin solution R1* and heat at 60 °C for 10 min.

System suitability Reference solution (b):
— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube on a water-bath for 1 min; a dark yellow colour develops.

D. Dissolve about 25 mg in 2 mL of *water R.* Add 2 mL of *dilute sodium hydroxide solution R* and shake. Wait a few minutes and add 3 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R1.* A white precipitate is formed. Add 0.5 mL of *concentrated ammonia R.* The precipitate dissolves.

TESTS

Appearance of solution
Dissolve 0.200 g in 20 mL of *water R*; the solution is not more opalescent than reference suspension II (*2.2.1*).
Dissolve 0.500 g in 10 mL of *water R*; the absorbance (*2.2.25*) of the solution at 430 nm is not greater than 0.10.

pH (*2.2.3*)
3.0 to 4.5.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 175 to + 195 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the test solution and reference solutions (a), (b) and (d) immediately before use.*

Phosphate buffer A Dissolve 1.4 g of *sodium dihydrogen phosphate monohydrate R* in *water R* and dilute to about 800 mL with the same solvent. Adjust to pH 3.0 with *dilute phosphoric acid R* and dilute to 1000.0 mL with *water R.*

Phosphate buffer B Dissolve 2.75 g of *sodium dihydrogen phosphate monohydrate R* and 2.3 g of *disodium hydrogen phosphate dihydrate R* in *water R* and dilute to about 1800 mL with the same solvent. Adjust to pH 6.8, if necessary, using *dilute phosphoric acid R* or *dilute sodium hydroxide solution R* and dilute to 2000.0 mL with *water R.*

Test solution Dissolve 30.0 mg of the substance to be examined in phosphate buffer A and dilute to 100.0 mL with phosphate buffer A.

Reference solution (a) Dissolve 30.0 mg of *bacampicillin hydrochloride CRS* in phosphate buffer A and dilute to 100.0 mL with phosphate buffer A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with phosphate buffer A.

Reference solution (c) Dissolve 30 mg of the substance to be examined in phosphate buffer B and dilute to 100 mL with phosphate buffer B. Heat at 80 °C for about 30 min.

Reference solution (d) Dissolve 20 mg of *ampicillin trihydrate CRS* (impurity I) in phosphate buffer A and dilute to 250 mL with phosphate buffer A. Dilute 5 mL of this solution to 100 mL with phosphate buffer A.

Column:
— *size: l* = 0.05 m, Ø = 3.9 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 30 volumes of *acetonitrile R1* and 70 volumes of a 0.06 per cent *m/m* solution of *tetrahexylammonium hydrogen sulfate R* in phosphate buffer B.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time 3.5 times the retention time of bacampicillin.

System suitability:
— the peak due to impurity I is separated from the peaks due to the solvent in the chromatogram obtained with reference solution (d);
— *relative retention* with reference to bacampicillin: degradation product eluting just after bacampicillin = 1.12 to 1.38 in the chromatogram obtained with reference solution (c); if necessary, adjust the concentration of tetrahexylammonium hydrogen sulfate in the mobile phase.

Limits:
— *any impurity*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Butyl acetate and ethyl acetate (*2.4.24, System A*)
Maximum 2.0 per cent of butyl acetate, maximum 4.0 per cent of ethyl acetate and maximum 5.0 per cent for the sum of the contents.

Sample solution Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Use the method of standard additions.

Static head-space conditions that may be used:
— *equilibration temperature*: 60 °C;
— *equilibration time*: 20 min.

***N,N*-Dimethylaniline** (*2.4.26, Method A*)
Maximum 20 ppm.

Water (*2.5.12*)
Maximum 0.8 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)
Maximum 1.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

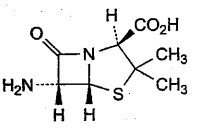
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of $C_{21}H_{28}ClN_3O_7S$ from the declared content of *bacampicillin hydrochloride CRS*.

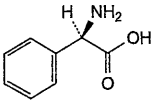
STORAGE

In an airtight container.

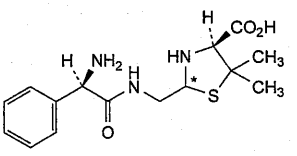
IMPURITIES



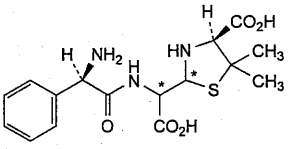
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



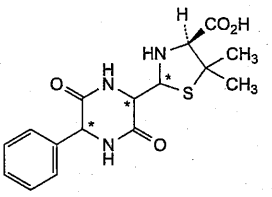
B. (2*R*)-2-amino-2-phenylacetic acid (D-phenylglycine),



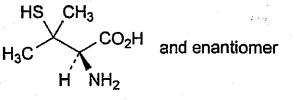
C. (2*RS*,4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of ampicillin),



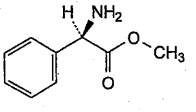
D. (4*S*)-2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]carboxymethyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of ampicillin),



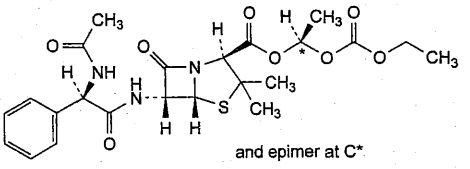
E. (4*S*)-2-(3,6-dioxo-5-phenylpiperazin-2-yl)-5,5-dimethylthiazolidine-4-carboxylic acid (diketopiperazines of ampicillin),



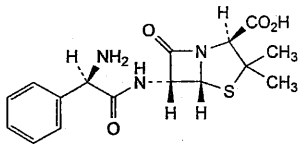
F. (2*RS*)-2-amino-3-methyl-3-sulfanylbutanoic acid (DL-penicillamine),



G. methyl (2*R*)-2-amino-2-phenylacetate (methyl D-phenylglycinate),



H. (1*RS*)-1-[(ethoxycarbonyl)oxy]ethyl (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-(acetylamino)-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (*N*-acetylbacampicillin),

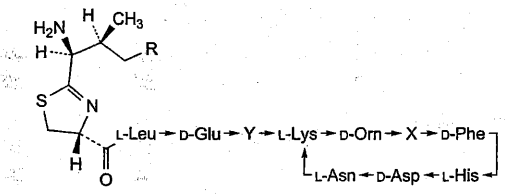


I. (2*S*,5*R*,6*R*)-6-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (ampicillin).

Ph Eur

Bacitracin

(Ph. Eur. monograph 0465)



Name	Mol. Formula	X	Y	R
Bacitracin A	$C_{66}H_{103}N_{17}O_{16}S$	L-Ile	L-Ile	CH_3
Bacitracin B1	$C_{65}H_{101}N_{17}O_{16}S$	L-Ile	L-Ile	H
Bacitracin B2	$C_{65}H_{101}N_{17}O_{16}S$	L-Val	L-Ile	CH_3
Bacitracin B3	$C_{65}H_{101}N_{17}O_{16}S$	L-Ile	L-Val	CH_3

1405-87-4

Action and use
Polypeptide antibacterial.

Ph Eur

DEFINITION
Mixture of antimicrobial polypeptides produced by certain strains of *Bacillus licheniformis* or *Bacillus subtilis*, the main components being:
- 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin A);
- 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B1);
- 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B2);
- 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B3).

Content
Minimum 60 IU/mg (dried substance).

CHARACTERS
Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION
First identification: B, C.

Second identification: A, C.

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a 3.4 g/L solution of *hydrochloric acid R* and dilute to 1.0 mL with the same solution.

Reference solution Dissolve 10 mg of *bacitracin zinc CRS* in a 3.4 g/L solution of *hydrochloric acid R* and dilute to 1.0 mL with the same solution.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (14:29:57 *V/V/V*).

Application 10 µL.

Development Over half of the plate.

Drying At 100-105 °C.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

Results The spots in the chromatogram obtained with the test solution are similar in position, size and colour to the spots in the chromatogram obtained with the reference solution.

B. Composition (see Tests).

C. Ignite 0.2 g. There is no significant yellow-coloured residue at high temperature. Allow to cool. Dissolve the residue in 0.1 mL of *dilute hydrochloric acid R*. Add 5 mL of *water R* and 0.2 mL of *strong sodium hydroxide solution R*. No white precipitate is formed.

TESTS
Solution S
Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

pH (*2.2.3*)
6.0 to 7.0 for solution S.

Composition
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Solution A 40 g/L solution of *sodium edetate R* adjusted to pH 7.0 with *dilute sodium hydroxide solution R*.

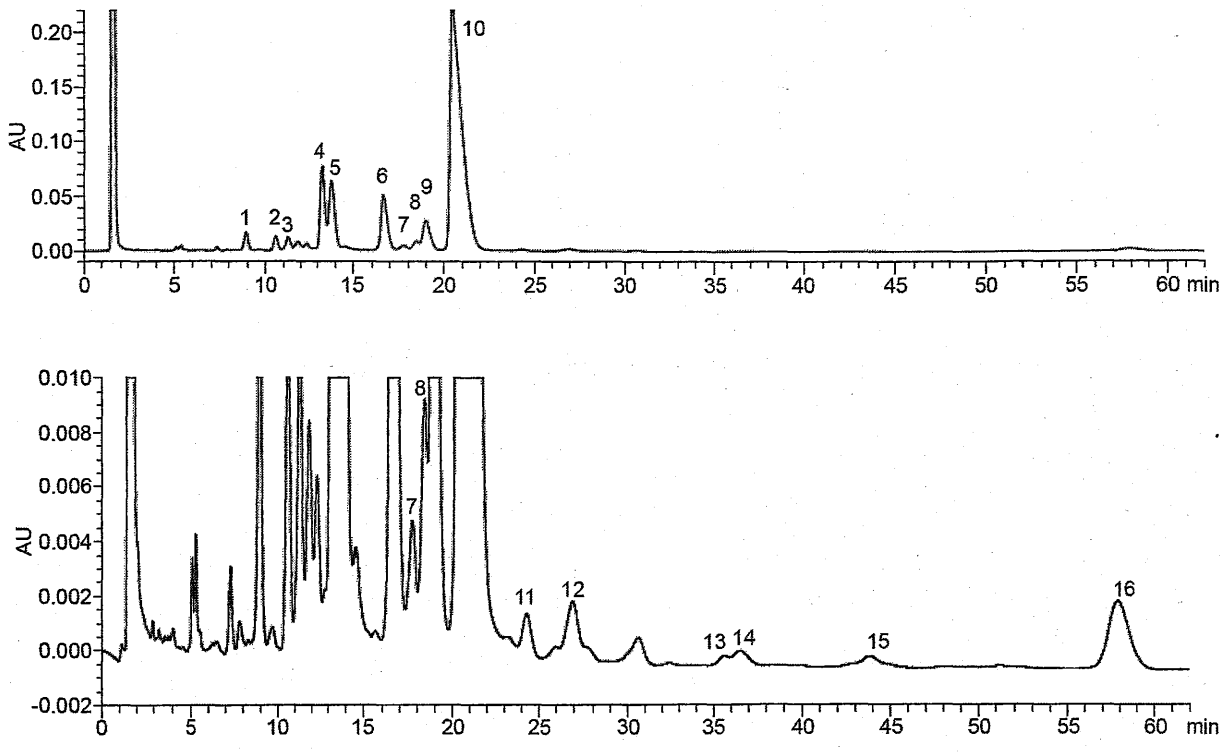
Solution B In a volumetric flask, dissolve 54.4 g of *potassium dihydrogen phosphate R* in *water for chromatography R* and dilute to 2000 mL with the same solvent. Adjust to pH 6.0 with a 34.8 g/L solution of *dipotassium hydrogen phosphate R* and filter through a membrane filter (nominal pore size 0.45 µm).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of *bacitracin for system suitability CRS* in solution A and dilute to 10.0 mL with the same solution.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (c) In order to prepare impurities E, F, G and H *in situ*, heat about 4 mL of reference solution (a) in a water-bath for 30 min. Cool to room temperature.



1. impurity A
2. impurity B
3. impurity C
4. bacitracin B1
5. bacitracin B2
6. bacitracin B3
7. impurity M
8. impurity N
9. impurity L
10. bacitracin A
11. impurity O
12. impurities P and Q
13. impurity F
14. impurity G
15. impurity H
16. impurity E

Figure 0465.-1. – *Chromatogram for the test for composition of bacitracin: test solution*

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped, charged surface, ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R* (3.5 µm);
— *temperature*: 28 ± 2 °C.

Mobile phase *acetonitrile R*, solution B, *water for chromatography R, methanol R1* (43:100:300:557 *V/V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 100 µL of the test solution and reference solutions (a) and (b).

Run time 3 times the retention time of bacitracin A.

Identification of peaks Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurity M and bacitracins A, B1, B2 and B3 (see Figure 0465.-1).

Relative retention With reference to bacitracin A (retention time = about 20 min): impurity A = about 0.44; impurity B = about 0.52; impurity C = about 0.55; bacitracin B1 = about 0.65; bacitracin B2 = about 0.67; bacitracin B3 = about 0.81; impurity M = about 0.87; impurity N = about 0.90; impurity L = about 0.93; impurity O = about 1.2; impurities P and Q = about 1.3; impurity F = about 1.6; impurity G = about 1.8; impurity H = about 2.1; impurity E = about 2.8.

If necessary, adjust the composition of the mobile phase by changing the amount of organic modifier whilst keeping the ratio constant between methanol and acetonitrile.

System suitability:
— *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak due to bacitracin B2 and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bacitracin B1 in the chromatogram obtained with reference solution (a);
— *peak-to-valley ratio*: minimum 1.1, where H_p = height above the baseline of the peak due to impurity M and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bacitracin B3 in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 50 for the peak due to bacitracin A in the chromatogram obtained with reference solution (b).

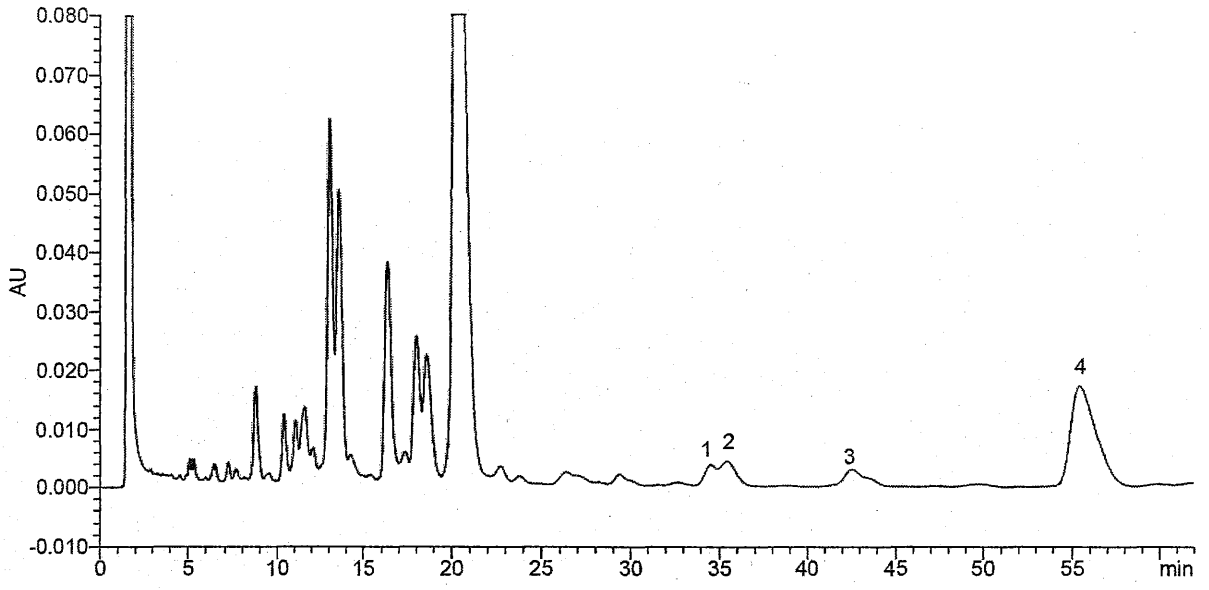
Limits:
— *bacitracin A*: minimum 45.0 per cent;
— *sum of bacitracins A, B1, B2 and B3*: minimum 77.0 per cent.

Related substances

Liquid chromatography (*2.2.29*) as described in the test for composition with the following modifications. Use the normalisation procedure.

Injection Test solution and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, L, M, N, O, P and Q (see Figure 0465.-1); use the chromatogram obtained with



1. impurity F 2. impurity G 3. impurity H 4. impurity E

Figure 0465.-2. – *Chromatogram for the test for related substances of bacitracin: reference solution (c)*

reference solution (c) to identify the peaks due to impurities E, F, G and H (see Figure 0465.-2).

Limits:

— *sum of impurities L and N*: maximum 8.0 per cent;
— *impurity E*: maximum 4.0 per cent;
— *impurity A*: maximum 3.5 per cent;
— *impurities B, M*: for each impurity, maximum 3.0 per cent;
— *impurity C*: maximum 2.5 per cent;
— *sum of impurities O, P and Q*: maximum 2.5 per cent;
— *sum of impurities F and G*: maximum 2.0 per cent;
— *impurity H*: maximum 1.0 per cent;
— *any other impurity*: for each impurity, maximum 2.0 per cent;
— *total*: maximum 23.0 per cent;
— *reporting threshold*: 0.25 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.1 kPa for 3 h.

Sulfated ash (*2.4.14*)

Maximum 1.0 per cent, determined on 1.0 g.

ASSAY

Carry out the microbiological assay of antibiotics (*2.7.2*). Use *bacitracin zinc CRS* as the reference substance.

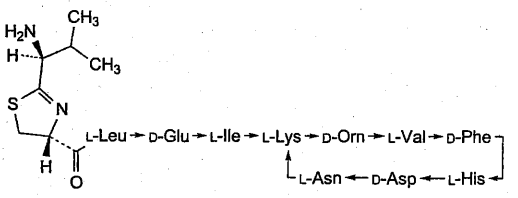
STORAGE

In an airtight container at 2 °C to 8 °C. If the substance is sterile, the container is also sterile and tamper-proof.

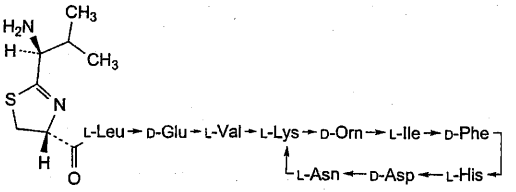
IMPURITIES

Specified impurities A, B, C, E, F, G, H, L, M, N, O, P, Q.

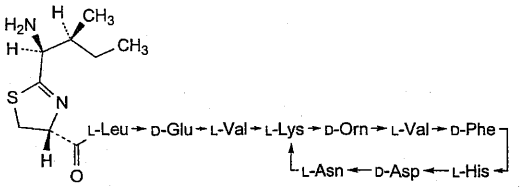
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) D, I, J, K.



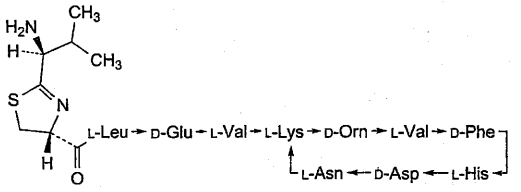
A. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D1, bacitracin C2),



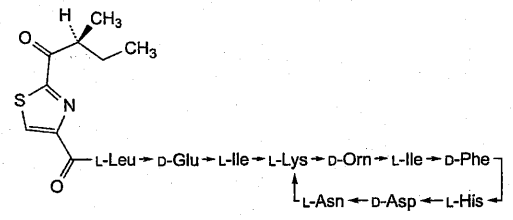
B. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D2, bacitracin C3),



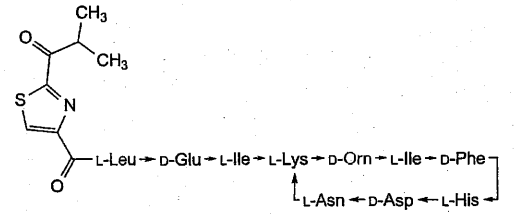
C. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D3, bacitracin C1a),



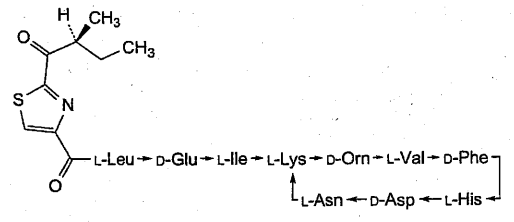
D. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin E),



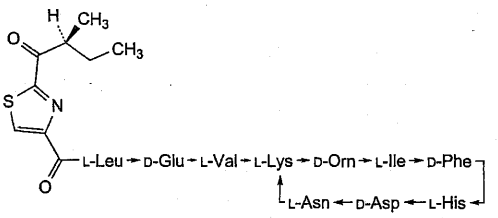
E. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin F),



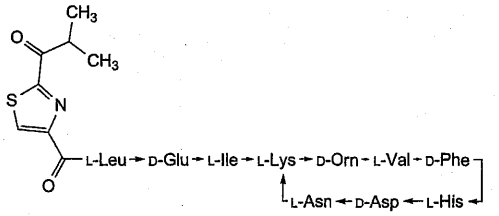
F. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H1),



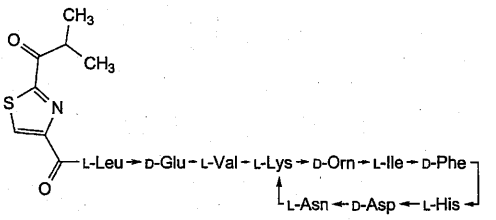
G. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H2),



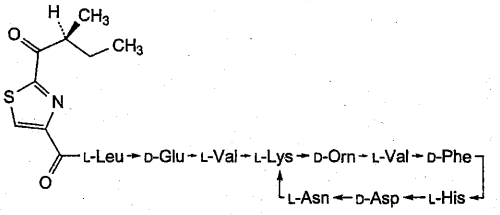
H. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H3),



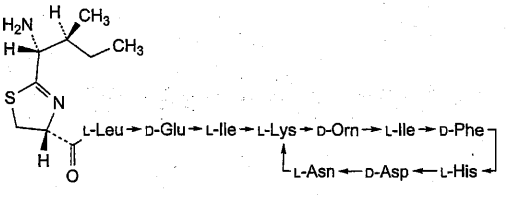
I. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I1),



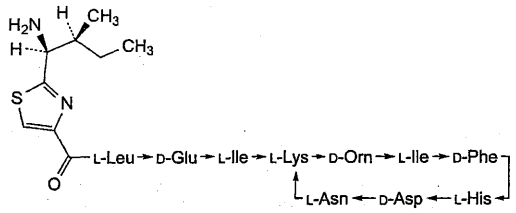
J. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I2),



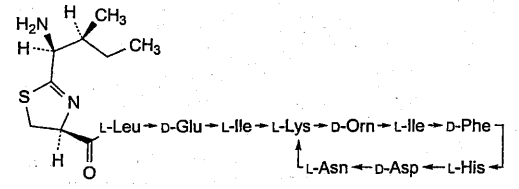
K. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I3),



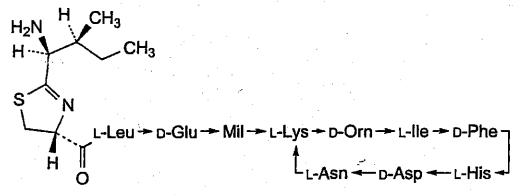
L. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*R*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin X),



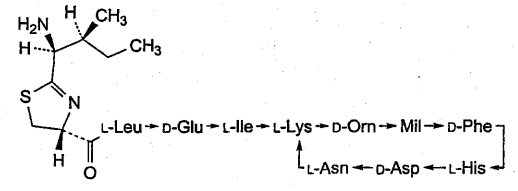
M. 4,10-anhydro[*N*-[[2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin Y),



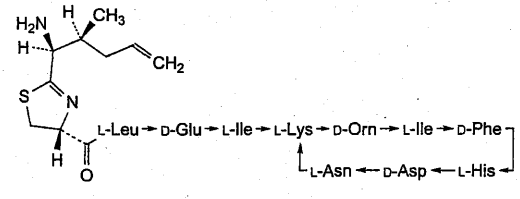
N. 4,10-anhydro[*N*-[[(4*S*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin Z),



O. Mil = 5-methylene-L-isoleucine: 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-5-methylene-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J1),



P. Mil = 5-methylene-L-isoleucine: 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-5-methylene-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J2),

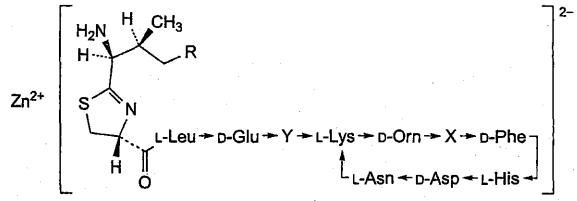


Q. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylpent-4-en-1-yl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J3).

Ph Eur

Bacitracin Zinc

(Ph. Eur. monograph 0466)



Name	Mol. Formula	X	Y	R
Bacitracin A	$C_{66}H_{103}N_{17}O_{16}S$	L-Ile	L-Ile	CH_3
Bacitracin B1	$C_{65}H_{101}N_{17}O_{16}S$	L-Ile	L-Ile	H
Bacitracin B2	$C_{65}H_{101}N_{17}O_{16}S$	L-Val	L-Ile	CH_3
Bacitracin B3	$C_{65}H_{101}N_{17}O_{16}S$	L-Ile	L-Val	CH_3

1405-89-6

Action and use
Polypeptide antibacterial.

Preparation
Polymyxin and Bacitracin Ointment

Ph Eur

DEFINITION

Zinc complex of bacitracin, which consists of a mixture of antimicrobial polypeptides produced by certain strains of *Bacillus licheniformis* or *Bacillus subtilis*, the main components being:

— 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin A);

— 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B1);

— 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B2);

— 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin B3).

Content
Minimum 60 IU/mg (dried substance).

CHARACTERS

Appearance
White or light yellowish-grey, hygroscopic powder.

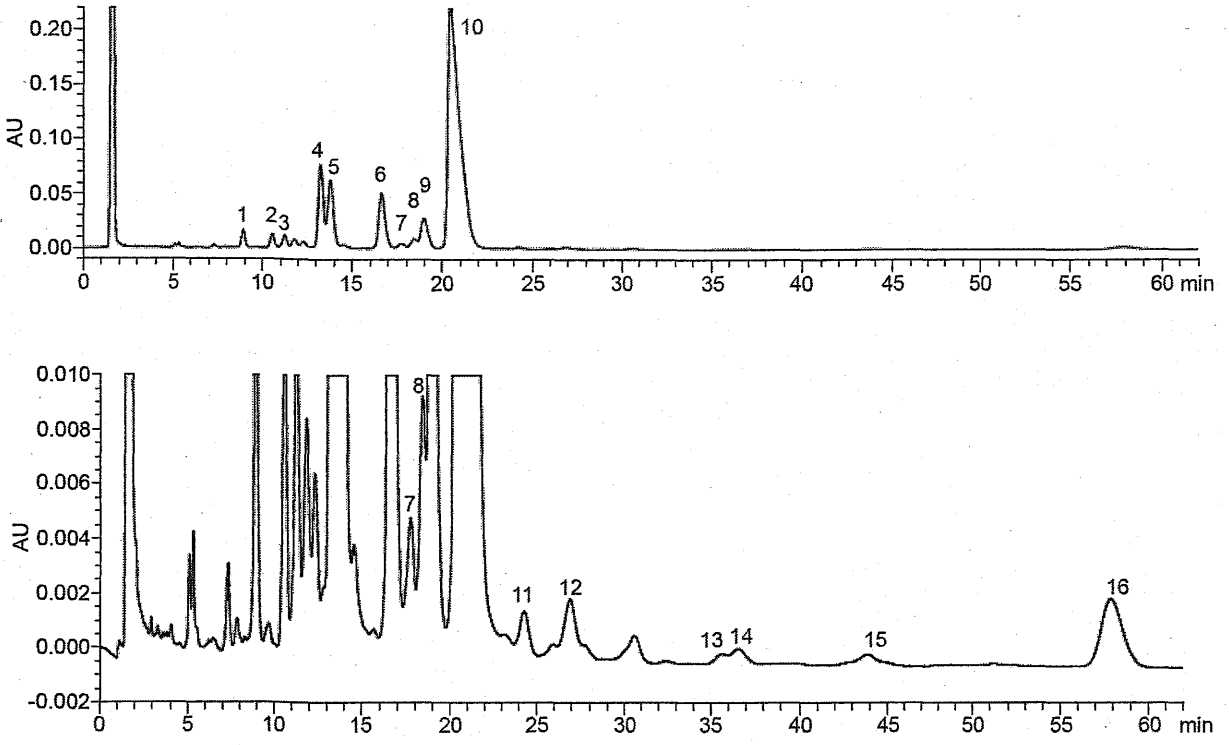
Solubility
Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification: B, C.

Second identification: A, C.

A. Thin-layer chromatography (*2.2.27*).



1. impurity A
2. impurity B
3. impurity C
4. bacitracin B1
5. bacitracin B2
6. bacitracin B3
7. impurity M
8. impurity N
9. impurity L
10. bacitracin A
11. impurity O
12. impurities P and Q
13. impurity F
14. impurity G
15. impurity H
16. impurity E

Figure 0466.-1. – *Chromatogram for the test for composition of bacitracin zinc: test solution*

Test solution Dissolve 10 mg of the substance to be examined in 0.5 mL of *dilute hydrochloric acid R* and dilute to 1.0 mL with *water R*.

Reference solution Dissolve 10 mg of *bacitracin zinc CRS* in 0.5 mL of *dilute hydrochloric acid R* and dilute to 1.0 mL with *water R*.

Plate *TLC silica gel plate R*.

Mobile phase *glacial acetic acid R, water R, butanol R* (14:29:57 *V/V/V*).

Application 10 µL.

Development Over half of the plate.

Drying At 100-105 °C.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

Results The spots in the chromatogram obtained with the test solution are similar in position, size and colour to the spots in the chromatogram obtained with the reference solution.

B. Composition (see Tests).

C. Ignite about 0.15 g, allow to cool and dissolve the residue in 1 mL of *dilute hydrochloric acid R*. Add 4 mL of *water R*. The solution gives the reaction of zinc (*2.3.1*).

TESTS

pH (*2.2.3*)

6.0 to 7.5.

Shake 1.0 g for about 1 min with 10 mL of *carbon dioxide-free water R* and filter.

Composition

Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Solution A 40 g/L solution of *sodium edetate R* adjusted to pH 7.0 with *dilute sodium hydroxide solution R*.

Solution B In a volumetric flask, dissolve 54.4 g of *potassium dihydrogen phosphate R* in *water for chromatography R* and dilute to 2000 mL with the same solvent. Adjust to pH 6.0 with a 34.8 g/L solution of *dipotassium hydrogen phosphate R* and filter through a membrane filter (nominal pore size 0.45 µm).

Test solution Dissolve 0.100 g of the substance to be examined in solution A and dilute to 50.0 mL with the same solution.

Reference solution (a) Dissolve 20.0 mg of *bacitracin for system suitability CRS* in solution A and dilute to 10.0 mL with the same solution.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (c) In order to prepare impurities E, F, G and H *in situ*, heat about 4 mL of reference solution (a) in a water-bath for 30 min. Cool to room temperature.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped, charged surface, ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R* (3.5 µm);

— *temperature*: 28 ± 2 °C.

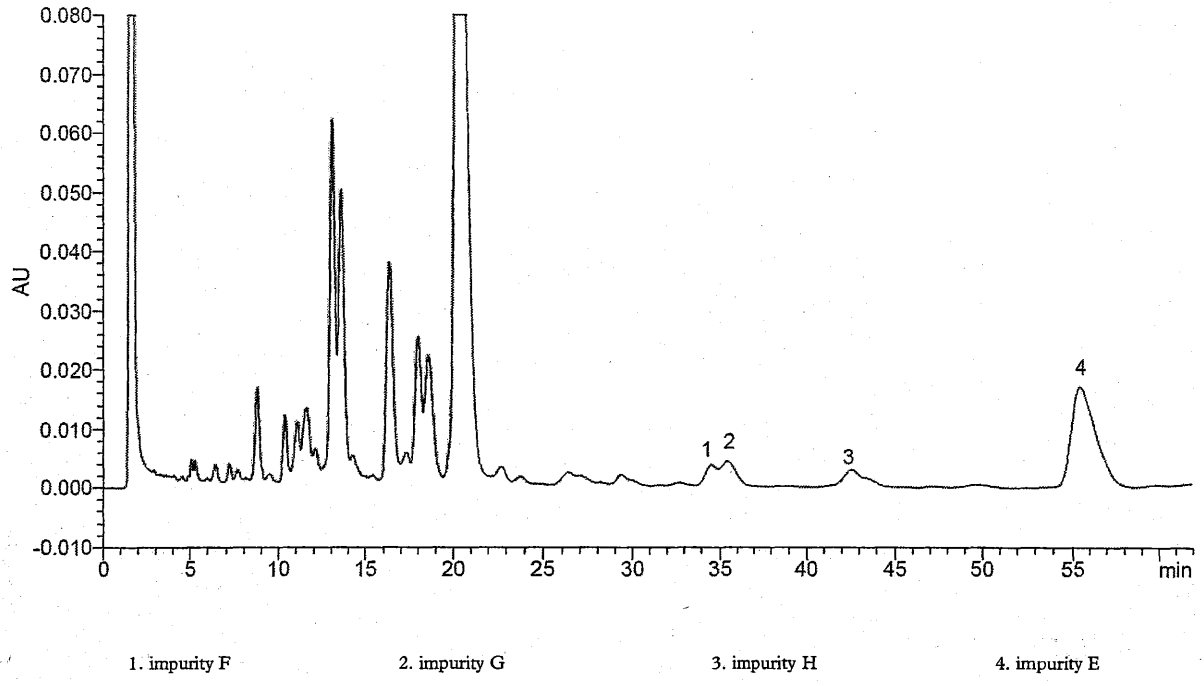


Figure 0466.-2. – *Chromatogram for the test for related substances of bacitracin zinc: reference solution (c)*

Mobile phase *acetonitrile R*, solution B, *water for chromatography R, methanol R1* (43:100:300:557 *V/V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 100 µL of the test solution and reference solutions (a) and (b).

Run time 3 times the retention time of bacitracin A.

Identification of peaks Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurity M and bacitracins A, B1, B2 and B3 (see Figure 0466.-1).

Relative retention With reference to bacitracin A (retention time = about 20 min): impurity A = about 0.44; impurity B = about 0.52; impurity C = about 0.55; bacitracin B1 = about 0.65; bacitracin B2 = about 0.67; bacitracin B3 = about 0.81; impurity M = about 0.87; impurity N = about 0.90; impurity L = about 0.93; impurity O = about 1.2; impurities P and Q = about 1.3; impurity F = about 1.6; impurity G = about 1.8; impurity H = about 2.1; impurity E = about 2.8.

If necessary, adjust the composition of the mobile phase by changing the amount of organic modifier whilst keeping the ratio constant between methanol and acetonitrile.

System suitability:

— *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak due to bacitracin B2 and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bacitracin B1 in the chromatogram obtained with reference solution (a);

— *peak-to-valley ratio*: minimum 1.1, where H_p = height above the baseline of the peak due to impurity M and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bacitracin B3 in the chromatogram obtained with reference solution (a);

— *signal-to-noise ratio*: minimum 50 for the peak due to bacitracin A in the chromatogram obtained with reference solution (b).

Limits:

— *bacitracin A*: minimum 45.0 per cent;

— *sum of bacitracins A, B1, B2 and B3*: minimum 77.0 per cent.

Related substances

Liquid chromatography (*2.2.29*) as described in the test for composition with the following modifications. Use the normalisation procedure.

Injection Test solution and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, L, M, N, O, P and Q (see Figure 0466.-1); use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities E, F, G, and H (see Figure 0466.-2).

Limits:

— *sum of impurities L and N*: maximum 8.0 per cent;

— *impurity E*: maximum 4.0 per cent;

— *impurity A*: maximum 3.5 per cent;

— *impurities B, M*: for each impurity, maximum 3.0 per cent;

— *impurity C*: maximum 2.5 per cent;

— *sum of impurities O, P and Q*: maximum 2.5 per cent;

— *sum of impurities F and G*: maximum 2.0 per cent;

— *impurity H*: maximum 1.0 per cent;

— *any other impurity*: for each impurity, maximum 2.0 per cent;

— *total*: maximum 23.0 per cent;

— *reporting threshold*: 0.25 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Zinc

3.5 per cent to 5.5 per cent (dried substance).

Dissolve 0.200 g in a mixture of 2.5 mL of *dilute acetic acid R* and 2.5 mL of water. Add 50 mL of *water R*, 50 mg of *xylenol orange triturate R* and sufficient *hexamethylenetetramine R* to produce a red colour. Add 2 g of *hexamethylenetetramine R* in excess. Titrate with *0.01 M sodium edetate* until a yellow colour is obtained.

1 mL of *0.01 M sodium edetate* is equivalent to 0.654 mg of Zn.

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.1 kPa for 3 h.

ASSAY

Suspend 50.0 mg in 5 mL of *water R*, add 0.5 mL of *dilute hydrochloric acid R* and dilute to 100.0 mL with *water R*. Allow the solution to stand for 30 min. Carry out the microbiological assay of antibiotics (*2.7.2*).

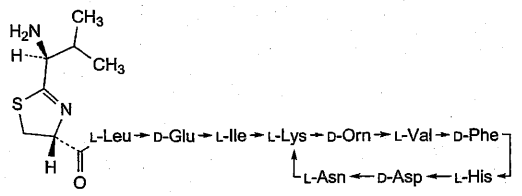
STORAGE

In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

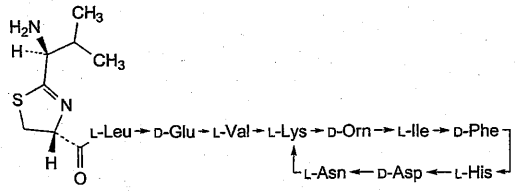
IMPURITIES

Specified impurities *A, B, C, E, F, G, H, L, M, N, O, P, Q.*

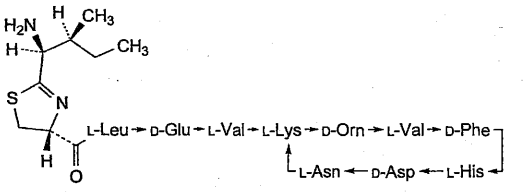
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, I, J, K.*



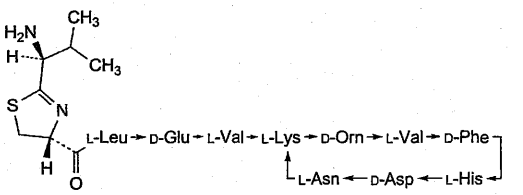
A. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D1, bacitracin C2),



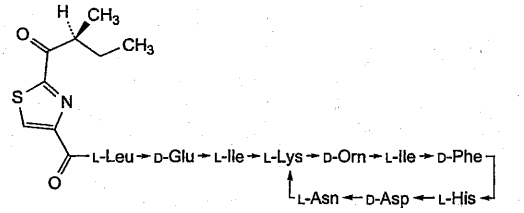
B. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D2, bacitracin C3),



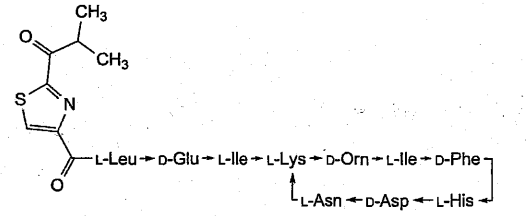
C. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin D3, bacitracin C1a),



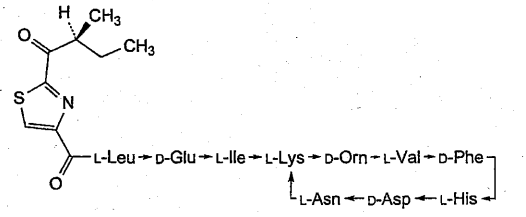
D. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*)-1-amino-2-methylpropyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin E),



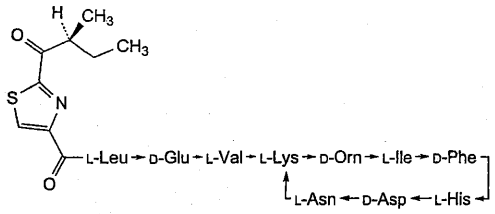
E. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine], (bacitracin F),



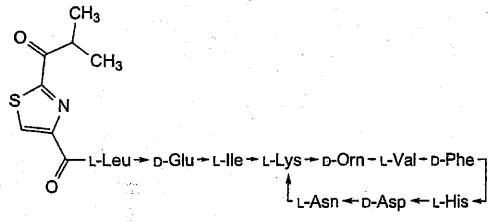
F. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H1),



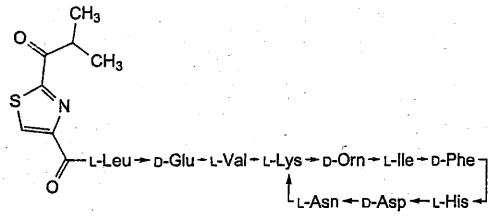
G. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H2),



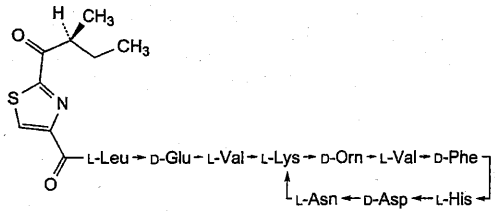
H. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin H3),



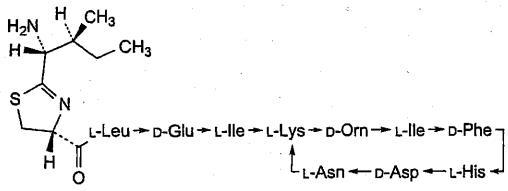
I. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I1),



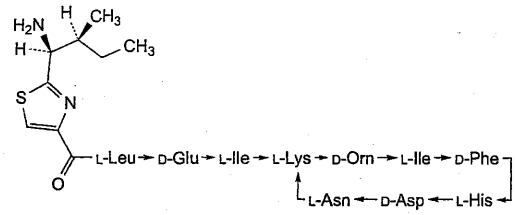
J. 4,10-anhydro[*N*-[[2-(2-methyl-1-oxopropyl)-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I2),



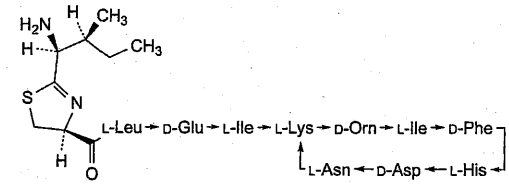
K. 4,10-anhydro[*N*-[[2-[(2*S*)-2-methyl-1-oxobutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-valyl-L-lysyl-D-ornithyl-L-valyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin I3),



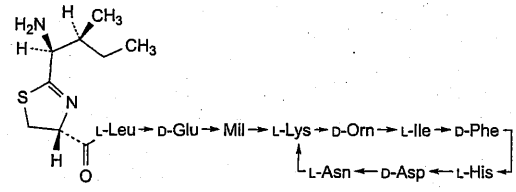
L. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*R*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin X),



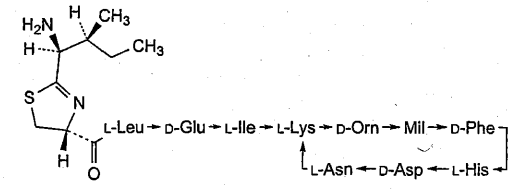
M. 4,10-anhydro[*N*-[[2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin Y),



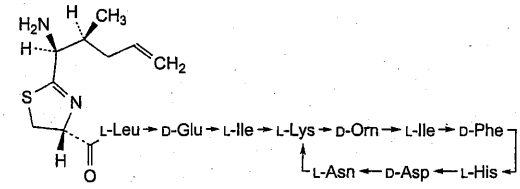
N. 4,10-anhydro[*N*-[[(4*S*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin Z),



O. Mil = 5-methylene-L-isoleucine: 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-5-methylene-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J1),



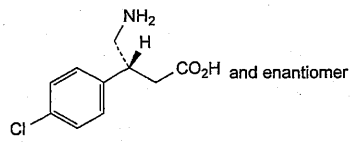
P. Mil = 5-methylene-L-isoleucine: 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylbutyl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-5-methylene-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J2),



Q. 4,10-anhydro[*N*-[[(4*R*)-2-[(1*S*,2*S*)-1-amino-2-methylpent-4-en-1-yl]-4,5-dihydro-1,3-thiazol-4-yl]carbonyl]-L-leucyl-D-α-glutamyl-L-isoleucyl-L-lysyl-D-ornithyl-L-isoleucyl-D-phenylalanyl-L-histidyl-D-α-aspartyl-L-asparagine] (bacitracin J3).

Baclofen

(Ph. Eur. monograph 0653)



$C_{10}H_{12}ClNO_2$ 213.7 *1134-47-0*

Action and use
Skeletal muscle relaxant.

Preparations
Baclofen Oral Solution
Baclofen Tablets

Ph Eur

DEFINITION
(3*RS*)-4-Amino-3-(4-chlorophenyl)butanoic acid.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in acetone. It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 70 mg in *water R* and dilute to 100.0 mL with the same solvent.

Spectral range 220-320 nm.

Absorption maxima At 259 nm, 266 nm and 275 nm.

Resolution (*2.2.25*): minimum 1.5 for the absorbance ratio.

Specific absorbance at the absorption maxima:
— at 259 nm: 9.8 to 10.8;
— at 266 nm: 11.5 to 12.7;
— at 275 nm: 8.4 to 9.3.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs prepared using 3 mg of substance and 300 mg of *potassium bromide R*.

Comparison *baclofen CRS*.

If the spectra obtained in the solid state show differences, dissolve 0.1 g of each of the substances separately in 1 mL of *dilute sodium hydroxide solution R* and add 10 mL of *ethanol (96 per cent) R* and 1 mL of *dilute acetic acid R*. Allow to stand for 1 h. Filter, wash the precipitate with *ethanol (96 per cent) R* and dry *in vacuo*. Prepare new discs and record the spectra.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution Dissolve 10 mg of *baclofen CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Plate *TLC silica gel G plate R.*

Mobile phase *anhydrous formic acid R, water R, methanol R, chloroform R, ethyl acetate R* (5:5:20:30:40 *V/V/V/V/V*).

Application 5 µL.

Development Over a path of 12 cm.

Drying Allow the solvents to evaporate.

Detection Spray with *ninhydrin solution R3* until the plate is slightly wet. Place in an oven maintained at 100 °C for 10 min. Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Appearance of solution
The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 0.50 g in *1 M sodium hydroxide* and dilute to 25 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *baclofen impurity A CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 2.0 mL of the test solution and 2.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase Dissolve 1.822 g of *sodium hexanesulfonate R* in 1 L of a mixture of 560 volumes of *water R*, 440 volumes of *methanol R* and 5 volumes of *glacial acetic acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time 5 times the retention time of baclofen.

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to baclofen and impurity A.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent).

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.000 g.

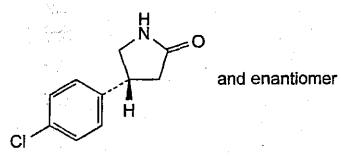
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.1500 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

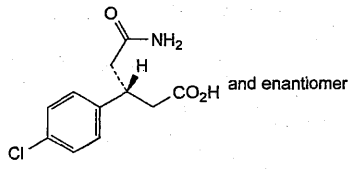
1 mL of *0.1 M perchloric acid* is equivalent to 21.37 mg of $C_{10}H_{12}ClNO_2$.

IMPURITIES
Specified impurities *A.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. (4*RS*)-4-(4-chlorophenyl)pyrrolidin-2-one,

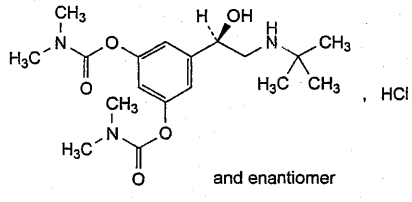


B. (3*RS*)-5-amino-3-(4-chlorophenyl)-5-oxopentanoic acid.

Ph Eur

Bambuterol Hydrochloride

(Ph. Eur. monograph 1293)



$C_{18}H_{30}ClN_3O_5$ 403.9 *81732-46-9*

Action and use
$Beta_2$-adrenoceptor agonist; bronchodilator.

Ph Eur

DEFINITION
5-[(1*RS*)-2-[(1,1-Dimethylethyl)amino]-1-hydroxyethyl]-1,3-phenylene bis(dimethylcarbamate) hydrochloride.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *bambuterol hydrochloride CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in a mixture of 1 volume of *water R* and 6 volumes of *acetone R*, cool in ice to precipitate and dry both precipitates *in vacuo* at 50 °C to constant weight. Record new spectra using the residues.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 4.0 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Acidity or alkalinity
To 10 mL of solution S add 0.2 mL of *methyl red solution R* and 0.2 mL of *0.01 M hydrochloric acid.* The solution is red. Add 0.4 mL of *0.01 M sodium hydroxide.* The solution is yellow.

Optical rotation (*2.2.7*)
−0.10° to + 0.10°.

Dilute 1 mL of solution S to 10 mL with *carbon dioxide-free water R.*

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 5.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 1.0 mg of *formoterol fumarate dihydrate CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Mix 0.8 mL of this solution with 0.4 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 1.3 g of *sodium octanesulfonate R* in 430 mL of a mixture of 25 volumes of *acetonitrile R1* and 75 volumes of *methanol R*; then mix this solution with 570 mL of 0.050 M phosphate buffer pH 3.0 prepared as follows: dissolve 6.90 g of *sodium dihydrogen phosphate monohydrate R* in *water R* and dilute to 1000 mL with *water R*, adjust to pH 3.0 with a 50 g/L solution of *dilute phosphoric acid R.*

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL; inject the mobile phase as a blank.

Run time 1.5 times the retention time of bambuterol.

Retention time Formoterol = about 7 min; bambuterol = about 9 min. If necessary, adjust the composition of the mobile phase; increase the content of phosphate buffer to increase the retention time.

System suitability Reference solution (a):

— *resolution*: minimum 5.0 between the peaks due to bambuterol and formoterol.

Limits:

— *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.6 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to the mobile phase.

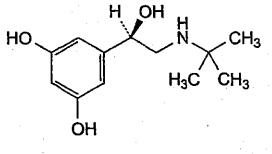
Water (*2.5.12*)
Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

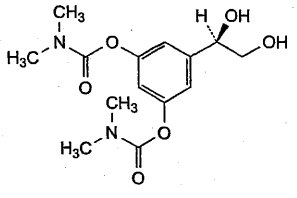
ASSAY
Dissolve 0.320 g in 50 mL of *ethanol (96 per cent) R* and add 5 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 40.39 mg of $C_{18}H_{30}ClN_3O_5$.

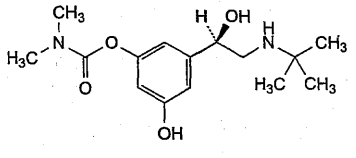
IMPURITIES
Specified impurities *A, B, C, D, E, F.*


and enantiomer

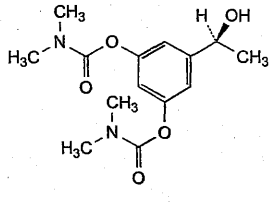
A. (1*RS*)-1-(3,5-dihydroxyphenyl)-2-[(1,1-dimethylethyl)amino]ethanol (terbutaline),


and enantiomer

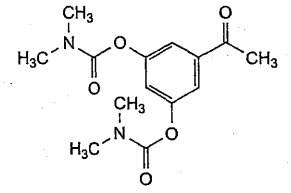
B. 5-[(1*RS*)-1,2-dihydroxyethyl]-1,3-phenylene bis (dimethylcarbamate),


and enantiomer

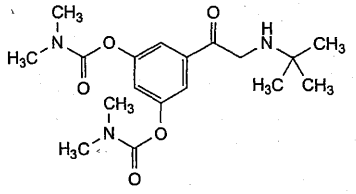
C. 3-[(1*RS*)-2-[(1,1-dimethylethyl)amino]-1-hydroxyethyl]-5-hydroxyphenyl dimethylcarbamate,


and enantiomer

D. 5-[(1*RS*)-1-hydroxyethyl]-1,3-phenylene bis (dimethylcarbamate),



E. 5-acetyl-1,3-phenylene bis(dimethylcarbamate),

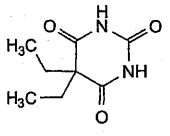


F. 5-[[(1,1-dimethylethyl)amino]acetyl]-1,3-phenylene bis (dimethylcarbamate).

Ph Eur

Barbital

(Ph. Eur. monograph 0170)



$C_8H_{12}N_2O_3$ 184.2 *57-44-3*

Action and use
Barbiturate.

Ph Eur

DEFINITION
Barbital contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 5,5-diethylpyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder or colourless crystals, slightly soluble in water, soluble in boiling water and in alcohol. It forms water-soluble compounds with alkali hydroxides and carbonates and with ammonia.

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Determine the melting point (*2.2.14*) of the substance to be examined. Mix equal parts of the substance to be examined and *barbital CRS* and determine the melting point of the mixture. The difference between the melting points (which are about 190 °C) is not greater than 2 °C.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *barbital CRS*.

C. Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 75 mg of the substance to be examined in *alcohol R* and dilute to 25 mL with the same solvent.

Reference solution Dissolve 75 mg of *barbital CRS* in *alcohol R* and dilute to 25 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 18 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of non-nitrogen substituted barbiturates (*2.3.1*).

TESTS

Appearance of solution

Dissolve 1.0 g in a mixture of 4 mL of *dilute sodium hydroxide solution R* and 6 mL of *water R*. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity

Boil 1.0 g with 50 mL of *water R* for 2 min, allow to cool and filter. To 10 mL of the filtrate add 0.15 mL of *methyl red solution R*. The solution is orange-yellow. Not more than 0.1 mL of *0.1 M sodium hydroxide* is required to produce a pure yellow colour.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 1.0 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dilute 0.5 mL of the test solution to 100 mL with *alcohol R*.

Apply separately to the plate 20 µL of each solution. Develop over a path of 15 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. Spray with *diphenylcarbazone mercuric reagent R*. Allow the plate to dry in air and spray with freshly prepared *alcoholic potassium hydroxide solution R* diluted 1 in 5 with *aldehyde-free alcohol R*. Heat at 100 °C to 105 °C for 5 min and examine immediately. When examined in ultraviolet light and after spraying, any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 1.00 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 85.0 mg in 5 mL of *pyridine R*. Add 0.5 mL of *thymolphthalein solution R* and 10 mL of *silver nitrate solution in pyridine R*. Titrate with *0.1 M ethanolic sodium hydroxide* until a pure blue colour is obtained. Carry out a blank titration.

1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 9.21 mg of $C_8H_{12}N_2O_3$.

Ph Eur

Barium Sulfate

(Ph. Eur. monograph 0010)

$BaSO_4$ 233.4 *7727-43-7*

Action and use

Radio-opaque substance used in the investigation of the gastro-intestinal tract.

Preparation

Barium Sulfate for Suspension

Ph Eur

CHARACTERS

Appearance

Fine, white or almost white powder, free from gritty particles.

Solubility

Practically insoluble in water and in organic solvents. It is very slightly soluble in acids and in solutions of alkali hydroxides.

IDENTIFICATION

A. Boil a suspension of 0.2 g with 5 mL of a 500 g/L solution of *sodium carbonate R* for 5 min, add 10 mL of *water R*, filter and acidify a part of the filtrate with *dilute hydrochloric acid R*. The solution gives the reactions of sulfates (*2.3.1*).

B. Wash the residue collected in the preceding test with 3 successive small quantities of *water R*. To the residue add 5 mL of *dilute hydrochloric acid R*, filter and add to the filtrate 0.3 mL of *dilute sulfuric acid R*. A white precipitate is formed that is insoluble in *dilute sodium hydroxide solution R*.

TESTS

Solution S

To 20.0 g add 40 mL of *distilled water R* and 60 mL of *dilute acetic acid R*. Boil for 5 min, filter and dilute the cooled filtrate to 100 mL with *distilled water R*.

Acidity or alkalinity

Heat 5.0 g with 20 mL of *carbon dioxide-free water R* on a water-bath for 5 min and filter. To 10 mL of the filtrate add 0.05 mL of *bromothymol blue solution R1*. Not more than 0.5 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Acid-soluble substances

Maximum 0.3 per cent.

Evaporate 25 mL of solution S to dryness on a water-bath and dry to constant mass at 100-105 °C. The residue weighs a maximum of 15 mg.

Oxidisable sulfur compounds

Shake 1.0 g with 5 mL of *water R* for 30 s and filter. To the filtrate add 0.1 mL of *starch solution R*, dissolve 0.1 g of *potassium iodide R* in the mixture, add 1.0 mL of a freshly prepared 3.6 mg/L solution of *potassium iodate R* and 1 mL of *1 M hydrochloric acid* and shake well. The colour of the solution is more intense than that of a standard prepared at the same time and in the same manner, but omitting the potassium iodate.

Soluble barium salts

Maximum 10 ppm.

To 2.5 mL of a 0.2 mg/L solution of *barium nitrate R* in a mixture of 30 volumes of *ethanol (96 per cent) R* and 70 volumes of *water R*, add 10 mL of *dilute sulfuric acid R*. Shake and allow to stand for 5 min. To 1 mL of this solution add 10 mL of solution S. Prepare a standard in the same

manner using 10 mL of *barium standard solution (2 ppm Ba) R* instead of solution S.

After 10 min, any opalescence in the test solution is not more intense than that in the standard.

Loss on ignition
Maximum 2.0 per cent, determined on 1.0 g at 600 ± 50 °C.

Ph Eur

Barium Sulfate for Suspension

Barium Sulphate for Suspension

Action and use
Radio-opaque preparation used in the investigation of the gastro-intestinal tract.

Preparation
Barium Sulfate Oral Suspension

DEFINITION
Barium Sulfate for Suspension is a dry mixture of Barium Sulfate with a suitable dispersing agent and may contain suitable flavours and suitable antimicrobial preservatives.

Content of barium sulfate, $BaSO_4$
90.0 to 110.0% of the stated amount.

CHARACTERISTICS
A fine, white or creamy white powder.

IDENTIFICATION
A. Ignite 1 g to constant weight. To 0.2 g of the residue add 5 mL of a 50% w/v solution of *sodium carbonate* and boil for 5 minutes. Add 10 mL of *water* and filter. Reserve the residue for test B. Acidify a portion of the filtrate with 2M *hydrochloric acid*. The solution yields the reactions characteristic of *sulfates*, Appendix VI.

B. Wash the residue reserved in test A with *water*, add 5 mL of *2M hydrochloric acid*, mix well and filter. Add 0.3 mL of *1M sulfuric acid* to the filtrate. A white precipitate is produced which is insoluble in 2M *hydrochloric acid*.

TESTS
Acidity or alkalinity
pH of an aqueous suspension containing the equivalent of 60% w/w of Barium Sulfate or, for lower strengths, the aqueous suspension at the strength of intended use, 3.5 to 8.5, Appendix V L.

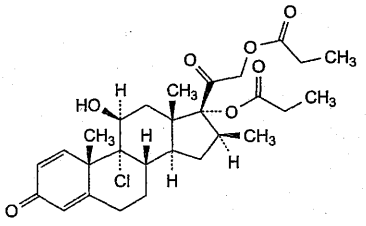
Loss on drying
When dried at 105° for 4 hours, loses not more than 1.0% of its weight. Use 1 g.

ASSAY
To a quantity containing 0.6 g of Barium Sulfate in a platinum dish add 5 g of *sodium carbonate* and 5 g of *potassium carbonate sesquihydrate* and mix. Heat to 1000° and maintain at this temperature for 15 minutes. Allow to cool and suspend the residue in 150 mL of *water*. Wash the dish with 2 mL of *6M acetic acid* and add the washings to the suspension. Cool in ice and decant the supernatant liquid, transferring as little of the solid matter as possible to the filter. Wash the residue with successive quantities of a 2% w/v solution of *sodium carbonate* until the washings are free from sulfate and discard the washings. Add 5 mL of *2M hydrochloric acid* to the filter, wash through into the vessel containing the bulk of the solid matter with *water*, add 5 mL of *hydrochloric acid* and dilute to 100 mL with *water*.

Add 10 mL of a 40% w/v solution of *ammonium acetate*, 25 mL of a 10% w/v solution of *potassium dichromate* and 10 g of *urea*. Cover and digest in a hot-air oven at 80° to 85° for 16 hours. Filter whilst still hot through a sintered-glass filter (ISO 4793, porosity grade 4, is suitable), washing the precipitate initially with a 0.5% w/v solution of *potassium dichromate* and finally with 2 mL of *water*. Dry to constant weight at 105°. Each g of the residue is equivalent to 0.9213 g of barium sulfate, $BaSO_4$.

Beclometasone Dipropionate

Anhydrous Beclometasone Dipropionate
(Ph. Eur. monograph 0654)



$C_{28}H_{37}ClO_7$ 521.0 *5534-09-8*

Action and use
Glucocorticoid.

Preparations
Beclometasone Cream
Beclometasone Aqueous Nasal Spray
Beclometasone Inhalation Powder
Beclometasone Inhalation Powder, pre-metered
Beclometasone Ointment
Beclometasone Pressurised Inhalation

Ph Eur

DEFINITION
9-Chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate.

Content
96.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison anhydrous beclometasone dipropionate CRS.

B. Treat 25 mg by the oxygen-flask method (*2.5.10*). Use a mixture of 1 mL of *1 M sodium hydroxide* and 20 mL of *water R* to absorb the combustion products. The solution gives reaction (a) of chlorides (*2.3.1*).

C. Loss on drying (see Tests).

TESTS
Specific optical rotation (*2.2.7*)
+ 108 to + 115 (dried substance).

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, mobile phase B (45:55 *V/V*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in 28 mL of mobile phase B and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of test solution (b) to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *beclometasone dipropionate for system suitability CRS* (containing impurity D) in 3 mL of mobile phase B and dilute to 5 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *beclometasone dipropionate for peak identification CRS* (containing impurities A, B, C, L and M) in 3 mL of mobile phase B and dilute to 5 mL with mobile phase A. Use 1 mL of this solution to dissolve the contents of a vial of *beclometasone dipropionate impurities F and N CRS*.

Reference solution (d) Dissolve 50.0 mg of *anhydrous beclometasone dipropionate CRS* in 28 mL of mobile phase B and dilute to 50.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: spherical difunctional bonded *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.35 with *phosphoric acid R*;
- *mobile phase B*: *tetrahydrofuran R, acetonitrile R, methanol R* (5:23:25 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	40	60
4 - 12	40 → 45	60 → 55
12 - 59	45	55

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µl of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with *beclometasone dipropionate for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, F, L, M and N; use the chromatogram supplied with *beclometasone dipropionate for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to beclometasone dipropionate (retention time = about 25 min): impurity A = about 0.3; impurity B = about 0.6; impurity D = about 1.1; impurity M = about 1.2; impurity L = about 1.3; impurity C = about 1.8; impurity N = about 2.0; impurity F = about 2.2.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to beclometasone dipropionate.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity F = 1.3; impurity M = 2.0;
- *impurity L*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
- *impurities B, F, M*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurities A, D, N*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

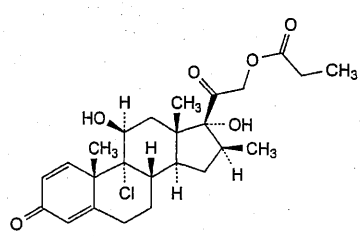
Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{28}H_{37}ClO_7$ from the declared content of *anhydrous beclometasone dipropionate CRS*.

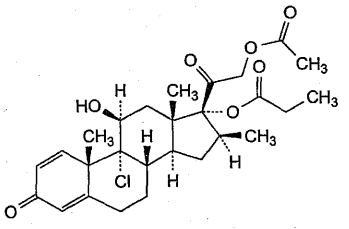
IMPURITIES

Specified impurities *A, B, C, D, F, L, M, N.*

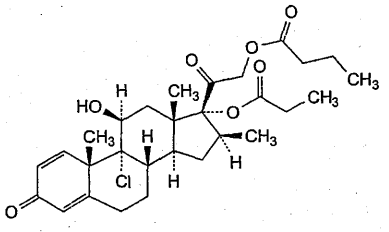
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, H, I, J, O, Q, R, S, U, V.*



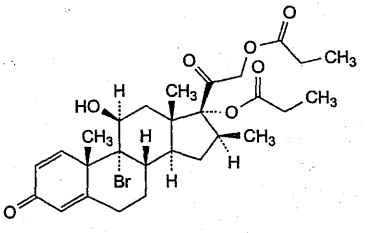
A. 9-chloro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl propanoate (beclometasone 21-propionate),



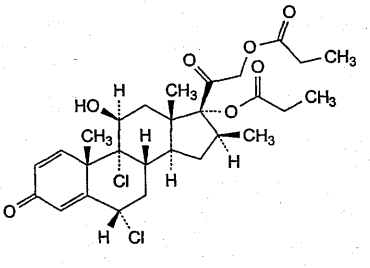
B. 21-(acetyloxy)-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (beclometasone 21-acetate 17-propionate),



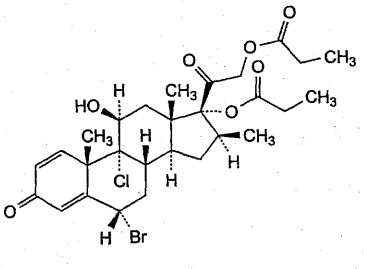
C. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxo-17-(propanoyloxy)-pregna-1,4-dien-21-yl butanoate (beclometasone 21-butyrate 17-propionate),



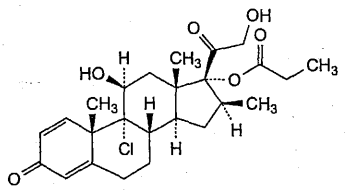
D. 9-bromo-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



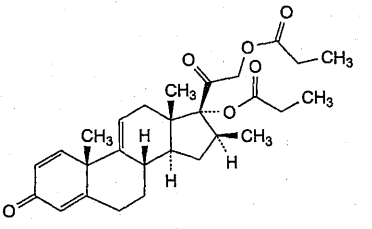
E. 6α,9-dichloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



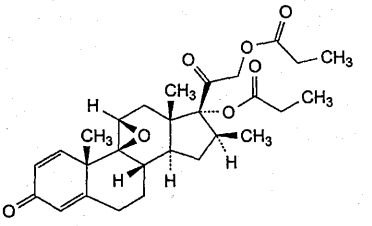
F. 6α-bromo-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



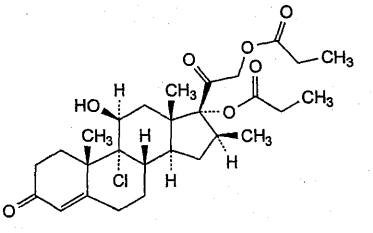
H. 9-chloro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (beclometasone 17-propionate),



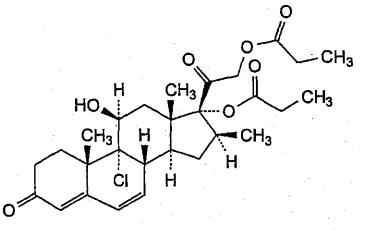
I. 16β-methyl-3,20-dioxopregna-1,4,9(11)-triene-17,21-diyl dipropanoate,



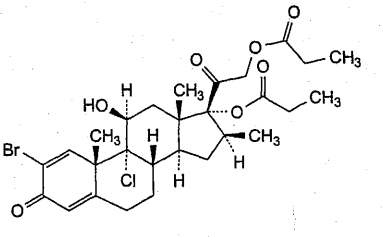
J. 9,11β-epoxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-diene-17,21-diyl dipropanoate,



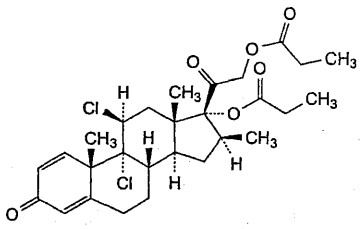
L. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregn-4-ene-17,21-diyl dipropanoate,



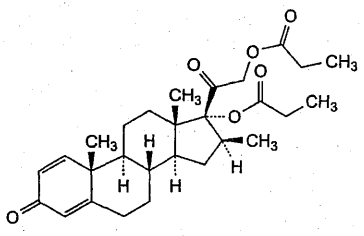
M. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-4,6-diene-17,21-diyl dipropanoate,



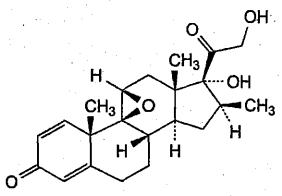
N. 2-bromo-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



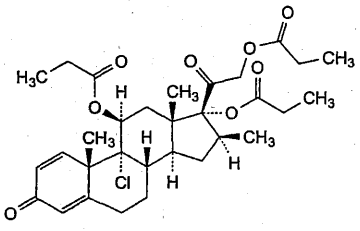
O. 9,11β-dichloro-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



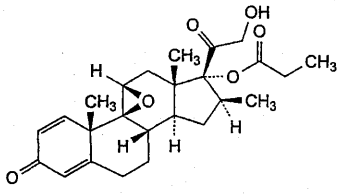
Q. 16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



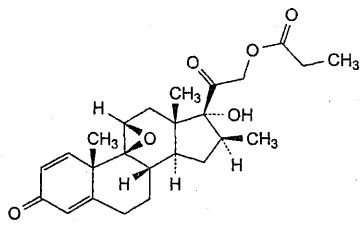
R. 9,11β-epoxy-17,21-dihydroxy-16β-methyl-9β-pregna-1,4-diene-3,20-dione,



S. 9-chloro-16β-methyl-3,20-dioxopregna-1,4-diene-11β,17,21-triyl tripropanoate (beclometasone tripropionate),



U. 9,11β-epoxy-21-hydroxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-dien-17-yl propanoate,

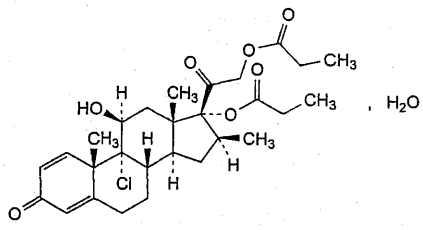


V. 9,11β-epoxy-17-hydroxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-dien-21-yl propanoate.

Ph Eur

Beclometasone Dipropionate Monohydrate

(Ph. Eur. monograph 1709)



$C_{28}H_{37}ClO_7,H_2O$ 539.1

Action and use
Glucocorticoid.

Preparations
Beclometasone Aqueous Nasal Spray
Beclometasone Inhalation Powder
Beclometasone Inhalation Powder, pre-metered

Ph Eur

DEFINITION

9-Chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate monohydrate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in acetone, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison beclometasone dipropionate monohydrate CRS.

B. Treat 25 mg by the oxygen-flask method (*2.5.10*). Use a mixture of 1 mL of *1 M sodium hydroxide* and 20 mL of *water R* to absorb the combustion products. The solution gives reaction (a) of chlorides (*2.3.1*).

C. Loss on drying (see Tests).

TESTS

Specific optical rotation (*2.2.7*)
+ 108 to + 115 (dried substance).
Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, mobile phase B (45:55 *V/V*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in 28 mL of mobile phase B and dilute to 50.0 mL with mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of test solution (b) to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *beclometasone dipropionate for system suitability CRS* (containing impurity D) in 3 mL of mobile phase B and dilute to 5 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *beclometasone dipropionate for peak identification CRS* (containing impurities B, C and L) in 3 mL of mobile phase B and dilute to 5 mL with mobile phase A. Use 1 mL of this solution to dissolve the contents of a vial of *beclometasone dipropionate impurities F and N CRS*.

Reference solution (d) Dissolve 50.0 mg of *anhydrous beclometasone dipropionate CRS* in 28 mL of mobile phase B and dilute to 50.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: spherical difunctional bonded *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.35 with *phosphoric acid R*;
- *mobile phase B*: *tetrahydrofuran R, acetonitrile R, methanol R* (5:23:25 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	40	60
4 - 12	40 → 45	60 → 55
12 - 59	45	55

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µl of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with *beclometasone dipropionate for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C, F and L; use the chromatogram supplied with *beclometasone dipropionate for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to beclometasone dipropionate (retention time = about 25 min): impurity B = about 0.6; impurity D = about 1.1; impurity L = about 1.3; impurity C = about 1.8; impurity F = about 2.2.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to beclometasone dipropionate.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity F by 1.3;
- *impurity B*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurities C, F, L*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

2.8 per cent to 3.8 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

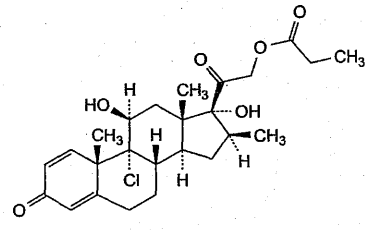
Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{28}H_{37}ClO_7$ from the declared content of *anhydrous beclometasone dipropionate CRS*.

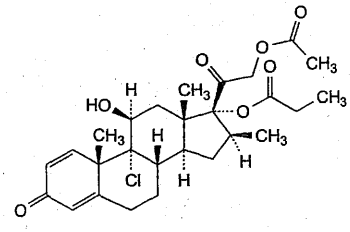
IMPURITIES

Specified impurities *B, C, F, L.*

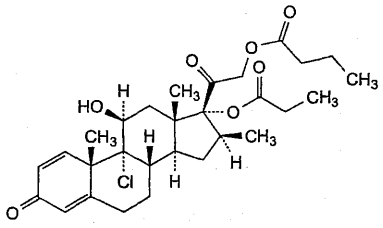
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, H, I, J, M, N, O, Q, R, S, U, V.*



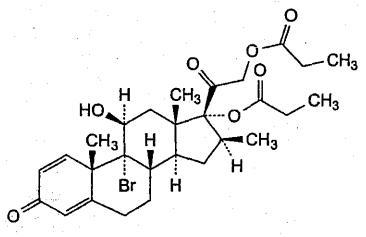
A. 9-chloro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl propanoate (beclometasone 21-propionate),



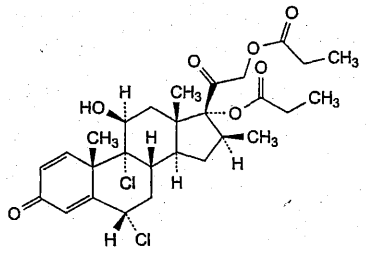
B. 21-(acetyloxy)-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (beclometasone 21-acetate 17-propionate),



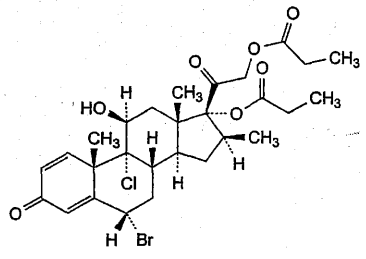
C. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxo-17-(propanoyloxy)-pregna-1,4-dien-21-yl butanoate (beclometasone 21-butyrate 17-propionate),



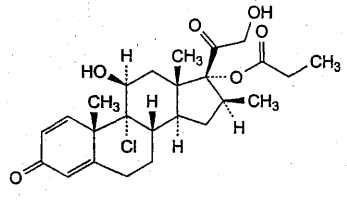
D. 9-bromo-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



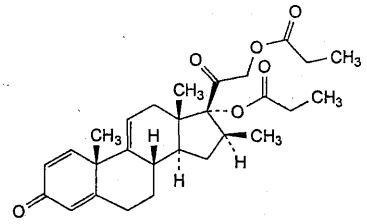
E. 6α,9-dichloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



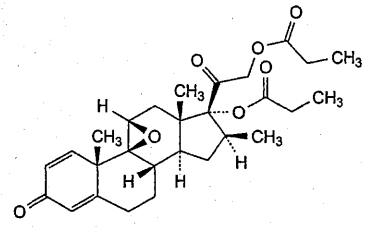
F. 6α-bromo-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



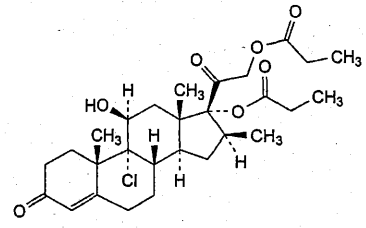
H. 9-chloro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (beclometasone 17-propionate),



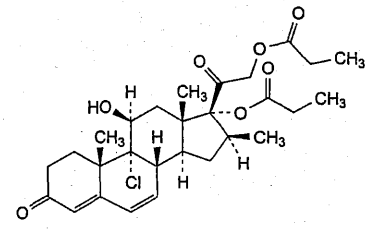
I. 16β-methyl-3,20-dioxopregna-1,4,9(11)-triene-17,21-diyl dipropanoate,



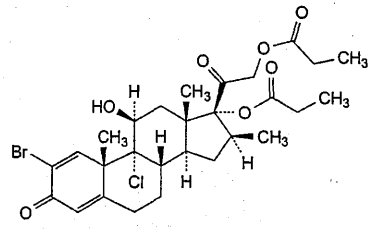
J. 9,11β-epoxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-diene-17,21-diyl dipropanoate,



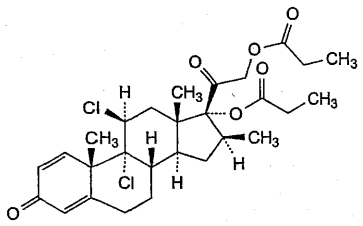
L. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregn-4-ene-17,21-diyl dipropanoate,



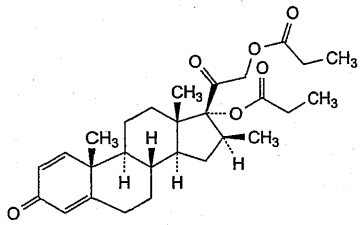
M. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-4,6-diene-17,21-diyl dipropanoate,



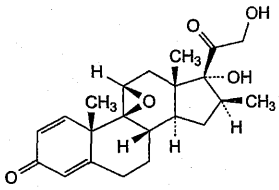
N. 2-bromo-9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



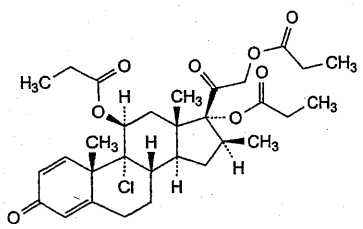
O. 9,11β-dichloro-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



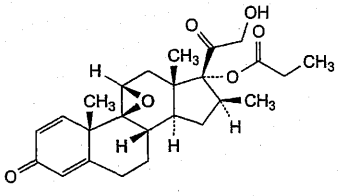
Q. 16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate,



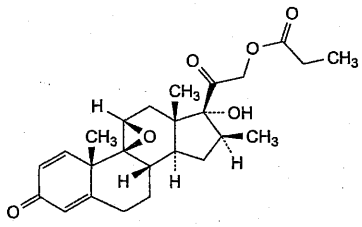
R. 9,11β-epoxy-17,21-dihydroxy-16β-methyl-9β-pregna-1,4-diene-3,20-dione,



S. 9-chloro-16β-methyl-3,20-dioxopregna-1,4-diene-11β,17,21-triyl tripropanoate (beclometasone tripropionate),



U. 9,11β-epoxy-21-hydroxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-dien-17-yl propanoate,



V. 9,11β-epoxy-17-hydroxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-dien-21-yl propanoate.

Ph Eur

White Beeswax

(Ph. Eur. monograph 0069)

Action and use
Excipient.

Ph Eur

DEFINITION
Wax obtained by bleaching yellow beeswax.

CHARACTERS
Appearance
White or yellowish-white pieces or plates, translucent when thin, with a fine-grained, matt and non-crystalline fracture; when warmed in the hand they become soft and malleable.

It has an odour similar to that of yellow beeswax, though fainter and never rancid. It is tasteless and does not stick to the teeth.

Solubility
Practically insoluble in water, partially soluble in hot ethanol (90 per cent *V/V*) and completely soluble in fatty and essential oils.

Relative density
About 0.960.

TESTS
Drop point *(2.2.17)*
61 °C to 66 °C.

Melt the beeswax by heating on a water-bath, pour onto a glass plate and allow to cool to a semi-solid mass. Fill the metal cup by inserting the wider end into the beeswax and repeating the procedure until beeswax extrudes from the narrow opening. Remove the excess with a spatula and insert the thermometer immediately. Remove the beeswax displaced. Allow to stand at room temperature for at least 12 h before determining the drop point.

Acid value
17.0 to 24.0.

To 2.00 g (*m* g), in a 250 mL conical flask fitted with a reflux condenser, add 40 mL of *xylene R* and a few glass beads. Heat until the substance is dissolved. Add 20 mL of *ethanol (96 per cent) R* and 0.5 mL of *phenolphthalein solution R1* and titrate the hot solution with 0.5 M alcoholic potassium hydroxide until a red colour persists for at least 10 s (n_1 mL). Carry out a blank test (n_2 mL).

$$\text{Acid value} = \frac{28.05(n_1 - n_2)}{m}$$

Ester value *(2.5.2)*
70 to 80.

Saponification value
87 to 104.

To 2.00 g (*m* g), in a 250 mL conical flask fitted with a reflux condenser, add 30 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *xylene R* and a few glass beads. Heat until the substance is dissolved. Add 25.0 mL of 0.5 M alcoholic potassium hydroxide and heat under a reflux condenser for 3 h. Titrate the hot solution immediately with *0.5 M hydrochloric acid*, using 1 mL of *phenolphthalein solution R1* as indicator (n_1 mL). Reheat the solution to boiling several times during the course of the titration. Carry out a blank test (n_2 mL).

$$\text{Saponification value} = \frac{28.05(n_2 - n_1)}{m}$$

Ceresin, paraffins and certain other waxes
To 3.0 g, in a 100 mL round-bottomed flask, add 30 mL of a 40 g/L solution of *potassium hydroxide R* in *aldehyde-free alcohol R* and boil gently under a reflux condenser for 2 h. Remove the condenser and immediately insert a thermometer. Place the flask in a water-bath at 80 °C and allow to cool, swirling the solution continuously.
No precipitate is formed until 65 °C, although the solution may be slightly opalescent. Beginning at 65 °C, the solution may become cloudy and precipitates may be formed.
At 59 °C, the solution is cloudy.

Glycerol and other polyols
Maximum 0.5 per cent *m/m*, calculated as glycerol.

To 0.20 g add 10 mL of *alcoholic potassium hydroxide solution R* and heat on a water-bath under a reflux condenser for 30 min. Add 50 mL of *dilute sulfuric acid R*, cool and filter. Rinse the flask and the filter with *dilute sulfuric acid R*. Combine the filtrate and washings and dilute to 100.0 mL with *dilute sulfuric acid R*. Place 1.0 mL of the solution in a test-tube, add 0.5 mL of a 10.7 g/L solution of *sodium periodate R*, mix and allow to stand for 5 min. Add 1.0 mL of *decolorised fuchsin solution R* and mix. Any precipitate disappears. Place the tube in a beaker containing water at 40 °C. During cooling observe for 10-15 min. Any violet-blue colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 1.0 mL of a 10 mg/L solution of *glycerol R* in *dilute sulfuric acid R*.

Ph Eur

Yellow Beeswax

(Ph. Eur. monograph 0070)

Action and use
Excipient.

Ph Eur

DEFINITION
Wax obtained by melting the walls of the honeycomb made by the honey-bee, *Apis mellifera* L., with hot water and removing foreign matter.

CHARACTERS
Appearance
Yellow or light brown pieces or plates with a fine-grained, matt and non-crystalline fracture; when warmed in the hand they become soft and malleable. It has a faint odour, characteristic of honey. It is tasteless and does not stick to the teeth.

Solubility
Practically insoluble in water, partially soluble in hot ethanol (90 per cent *V/V*) and completely soluble in fatty and essential oils.

Relative density
About 0.960.

TESTS
Drop point *(2.2.17)*
61 °C to 66 °C.

Melt the beeswax by heating on a water-bath, pour onto a glass plate and allow to cool to a semi-solid mass. Fill the metal cup by inserting the wider end into the beeswax and repeating the procedure until beeswax extrudes from the narrow opening. Remove the excess with a spatula and insert the thermometer immediately. Remove the beeswax displaced. Allow to stand at room temperature for at least 12 h before determining the drop point.

Acid value
17.0 to 22.0.

To 2.00 g (*m* g), in a 250 mL conical flask fitted with a reflux condenser, add 40 mL of *xylene R* and a few glass beads. Heat until the substance is dissolved. Add 20 mL of *ethanol (96 per cent) R* and 0.5 mL of *phenolphthalein solution R1* and titrate the hot solution with 0.5 M alcoholic potassium hydroxide until a red colour persists for at least 10 s (n_1 mL). Carry out a blank test (n_2 mL).

$$\text{Acid value} = \frac{28.05(n_1 - n_2)}{m}$$

Ester value *(2.5.2)*
70 to 80.

Saponification value
87 to 102.

To 2.00 g (*m* g), in a 250 mL conical flask fitted with a reflux condenser, add 30 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *xylene R* and a few glass beads. Heat until the substance is dissolved. Add 25.0 mL of 0.5 M alcoholic potassium hydroxide and heat under a reflux condenser for 3 h. Titrate the hot solution immediately with *0.5 M hydrochloric acid*, using 1 mL of *phenolphthalein solution R1* as indicator (n_1 mL). Reheat the solution to boiling several times during the course of the titration. Carry out a blank test (n_2 mL).

$$\text{Saponification value} = \frac{28.05(n_2 - n_1)}{m}$$

Ceresin, paraffins and certain other waxes
To 3.0 g, in a 100 mL round-bottomed flask, add 30 mL of a 40 g/L solution of *potassium hydroxide R* in *aldehyde-free alcohol R* and boil gently under a reflux condenser for 2 h. Remove the condenser and immediately insert a thermometer. Place the flask in a water-bath at 80 °C and allow to cool, swirling the solution continuously.
No precipitate is formed until 65 °C, although the solution may be slightly opalescent. Beginning at 65 °C, the solution may become cloudy and precipitates may be formed.
At 59 °C, the solution is cloudy.

Glycerol and other polyols
Maximum 0.5 per cent *m/m*, calculated as glycerol.

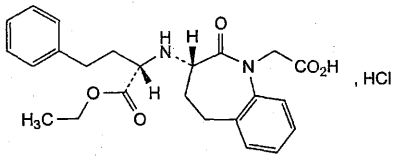
To 0.20 g add 10 mL of *alcoholic potassium hydroxide solution R* and heat on a water-bath under a reflux condenser

for 30 min. Add 50 mL of *dilute sulfuric acid R*, cool and filter. Rinse the flask and the filter with *dilute sulfuric acid R*. Combine the filtrate and washings and dilute to 100.0 mL with *dilute sulfuric acid R*. Place 1.0 mL of the solution in a test-tube, add 0.5 mL of a 10.7 g/L solution of *sodium periodate R*, mix and allow to stand for 5 min. Add 1.0 mL of *decolorised fuchsin solution R* and mix. Any precipitate disappears. Place the tube in a beaker containing water at 40 °C. During cooling observe for 10-15 min. Any violet-blue colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 1.0 mL of a 10 mg/L solution of *glycerol R* in *dilute sulfuric acid R*.

Ph Eur

Benazepril Hydrochloride

(Ph. Eur. monograph 2388)



$C_{24}H_{29}ClN_2O_5$ 461.0 *86541-74-4*

Action and use
Angiotensin converting enzyme inhibitor.

Ph Eur

DEFINITION
[(3*S*)-3-[[(1*S*)-1-(Ethoxycarbonyl)-3-phenylpropyl]amino]-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetic acid hydrochloride.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder, hygroscopic.

Solubility
Slightly soluble in water, freely soluble in anhydrous ethanol, very slightly soluble in ethyl acetate, practically insoluble in cyclohexane.

It shows polymorphism (*5.9*).

IDENTIFICATION
Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (*2.2.7*): −141 to −136 (dried substance).

Dissolve 1.000 g in *anhydrous ethanol R* and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benazepril hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

C. Enantiomeric purity (see Tests).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *benazepril hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *benazepril for system suitability CRS* (containing impurities B, C, D, E, F and G) in 1.0 mL of test solution (a).

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.30 m, Ø = 3.9 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase Add 0.2 mL of *glacial acetic acid R* to 1000 mL of a mixture of 360 volumes of *water R* and 640 volumes of *methanol R2*; add 0.81 g of *tetrabutylammonium bromide R* and stir to dissolve.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 25 µL of test solution (a) and reference solutions (b) and (c).

Run time 3 times the retention time of benazepril.

Relative retention With reference to benazepril (retention time = about 6 min): impurity E = about 0.3; impurity F = about 0.4; impurity C = about 0.5; impurity B = about 1.8; impurity D = about 2.0; impurity G = about 2.5.

Identification of impurities Use the chromatogram supplied with *benazepril for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D, E, F and G.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to benazepril and impurity B and minimum 1.5 between the peaks due to impurities E and F.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 0.5; impurity F = 0.7;
— *impurity B*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurities D, E, F, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Enantiomeric purity

Liquid chromatography (*2.2.29*).

Buffer solution pH 6.0 Dissolve 3.58 g of *disodium hydrogen phosphate dodecahydrate R* and 9.66 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 1000.0 mL with the same solvent.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *benazepril impurity A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Column:
— *size*: *l* = 0.10 m, Ø = 4.0 mm;
— *stationary phase*: spherical *silica gel AGP for chiral chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *methanol R2*, buffer solution pH 6.0 (20:80 *V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 50 µL of the test solution and reference solutions (b) and (c).

Run time 3.5 times the retention time of benazepril.

Relative retention With reference to benazepril (retention time = about 6 min): impurity A = about 1.9.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to benazepril.

Limit:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying (*2.2.32*)

Maximum 1.5 per cent, determined on 1.000 g by drying *in vacuo* at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

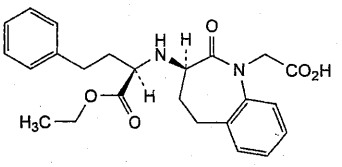
Calculate the percentage content of $C_{24}H_{29}ClN_2O_5$ from the declared content of *benazepril hydrochloride CRS*.

STORAGE

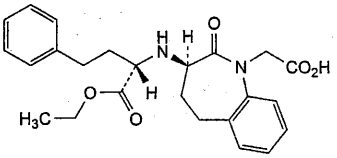
Protected from light, in an airtight container.

IMPURITIES

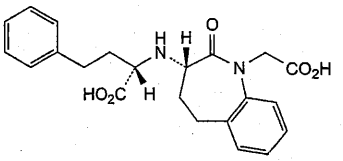
Specified impurities *A, B, C, D, E, F, G.*



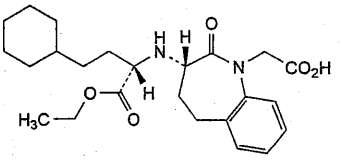
A. [(3*R*)-3-[[(1*R*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetic acid,



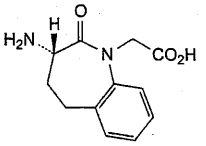
B. [(3*RS*)-3-[[(1*SR*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetic acid,



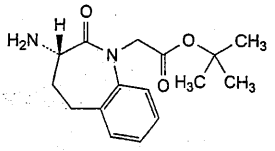
C. (2*S*)-2-[[(3*S*)-1-(carboxymethyl)-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-3-yl]amino]-4-phenylbutanoic acid,



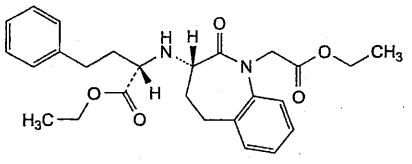
D. [(3*S*)-3-[[(1*S*)-3-cyclohexyl-1-(ethoxycarbonyl) propyl]amino]-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetic acid,



E. [(3*S*)-3-amino-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetic acid,



F. 1,1-dimethylethyl [(3*S*)-3-amino-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-1-yl]acetate,

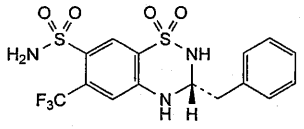


G. ethyl (2*S*)-2-[[(3*S*)-1-(2-ethoxy-2-oxoethyl)-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzazepin-3-yl]amino]-4-phenylbutanoate.

Ph Eur

Bendroflumethiazide

(Ph. Eur. monograph 0370)



and enantiomer

$C_{15}H_{14}F_3N_3O_4S_2$ 421.4 *73-48-3*

Action and use
Thiazide diuretic.

Preparations
Bendroflumethiazide Tablets
Bendroflumethiazide Oral Suspension

Ph Eur

DEFINITION
(3*RS*)-3-Benzyl-6-(trifluoromethyl)-3,4-dihydro-2*H*-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *bendroflumethiazide CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mix 40 volumes of *methanol R* and 60 volumes of a 2.0 g/L solution of *citric acid monohydrate R.*

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of *bendroflumethiazide impurity A CRS* and 2.5 mg of *altizide CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture. Mix 1 mL of this solution with 1 mL of the test solution and dilute to 100 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 15 volumes of *tetrahydrofuran R*, 25 volumes of *methanol R* and 60 volumes of a 2.0 g/L solution of *citric acid monohydrate R.*

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 µL.

Run time Twice the retention time of bendroflumethiazide.

Relative retention With reference to bendroflumethiazide (retention time = about 8 min): impurity A = about 0.2; altizide = about 0.5.

System suitability Reference solution (a):
— *resolution*: minimum 10 between the peaks due to altizide and bendroflumethiazide.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

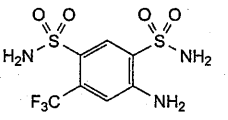
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 50 mL of *dimethyl sulfoxide R.* Titrate to the 2nd point of inflexion with *0.1 M tetrabutylammonium hydroxide in 2-propanol*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of *0.1 M tetrabutylammonium hydroxide in 2-propanol* is equivalent to 21.07 mg of $C_{15}H_{14}F_3N_3O_4S_2$.

IMPURITIES
Specified impurities *A.*

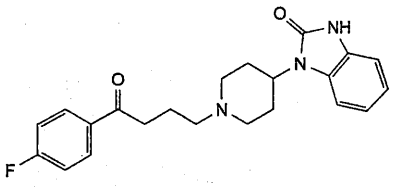


A. 4-amino-6-(trifluoromethyl)benzene-1,3-disulfonamide.

Ph Eur

Benperidol

(Ph. Eur. monograph 1172)



$C_{22}H_{24}FN_3O_2$ 381.4 *2062-84-2*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION

1-[1-[4-(4-Fluorophenyl)-4-oxobutyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in dimethylformamide, soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benperidol CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *methyl isobutyl ketone R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (a) Dissolve 30 mg of *benperidol CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of *benperidol CRS* and 30 mg of *droperidol CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetone R, methanol R* (10:90 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 5 mL of *anhydrous ethanol R*. Add 0.5 mL of *dinitrobenzene solution R* and 0.5 mL of *2 M alcoholic potassium hydroxide R*. A violet colour is produced which becomes brownish-red after 20 min.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.10 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of *benperidol CRS* and 2.5 mg of *droperidol CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethylformamide R*. Dilute 5.0 mL of this solution to 20.0 mL with *dimethylformamide R*.

Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 10 g/L solution of *tetrabutylammonium hydrogen sulfate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100 → 60	0 → 40
15 - 20	60	40
20 - 25	100	0

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL.

Relative retention With reference to benperidol (retention time = about 6.5 min): impurity A = about 0.2; impurity B = about 0.9; droperidol = about 1.1; impurity D = about 1.2; impurity E = about 1.3; impurity C = about 1.5.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to benperidol and droperidol.

Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

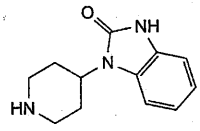
1 mL of *0.1 M perchloric acid* is equivalent to 38.14 mg of $C_{22}H_{24}FN_3O_2$.

STORAGE

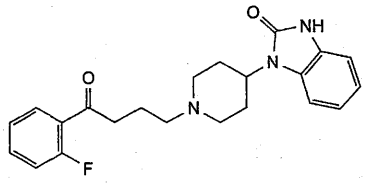
Protected from light.

IMPURITIES

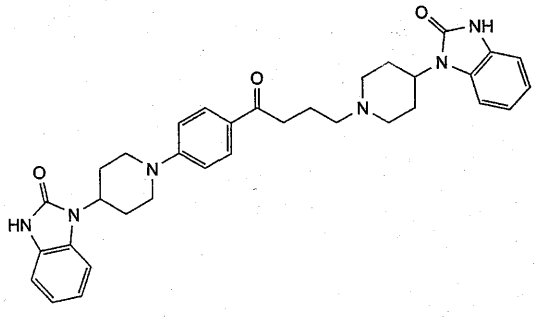
Specified impurities *A, B, C, D, E.*



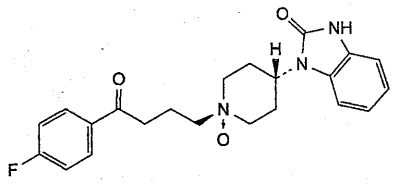
A. 1-(piperidin-4-yl)-1,3-dihydro-2*H*-benzimidazol-2-one,



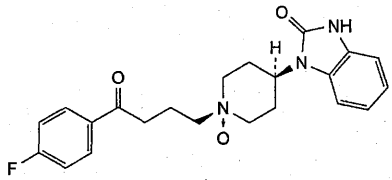
B. 1-[1-[4-(2-fluorophenyl)-4-oxobutyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one,



C. 1-[1-[4-oxo-4-[4-[4-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)piperidin-1-yl]phenyl]butyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one,



D. *cis*-1-[1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl 1-oxide]-1,3-dihydro-2*H*-benzimidazol-2-one,

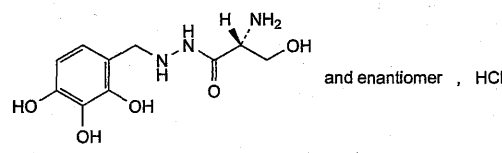


E. *trans*-1-[1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl 1-oxide]-1,3-dihydro-2*H*-benzimidazol-2-one.

Ph Eur

Benserazide Hydrochloride

(Ph. Eur. monograph 1173)



$C_{10}H_{16}ClN_3O_5$ 293.7 *14919-77-8*

Action and use
Dopa decarboxylase inhibitor.

Preparations
Co-beneldopa Capsules
Co-beneldopa Dispersible Tablets
Co-beneldopa Prolonged-release Capsules

Ph Eur

DEFINITION

(2*RS*)-2-Amino-3-hydroxy-2′-(2,3,4-trihydroxybenzyl) propanohydrazide hydrochloride.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or yellowish-white or orange-white, crystalline powder.

Solubility
Freely soluble in water, very slightly soluble in anhydrous ethanol, practically insoluble in acetone.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benserazide hydrochloride CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in hot *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Solution S (see Tests) gives reaction (b) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)

4.0 to 5.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

All solutions must be injected immediately or stored at 4 °C.

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R2* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *benserazide impurity A CRS*, 5.0 mg of *benserazide impurity C CRS* and 5.0 mg of *benserazide hydrochloride CRS* in *methanol R2* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of this solution to 50.0 mL with *methanol R2*.

Reference solution (b) Dilute 2.0 mL of reference solution (a) to 10.0 mL with *methanol R2*.

Reference solution (c) Dissolve 5 mg of *benserazide for peak identification CRS* (containing impurities A, B and C) in *methanol R2* and dilute to 5.0 mL with the same solvent.

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: dissolve 2.2 g of *sodium heptanesulfonate monohydrate R* and 6.8 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, add 50 mL of *methanol R2* and adjust to pH 3.5 with *phosphoric acid R*;
- *mobile phase B*: dissolve 2.2 g of *sodium heptanesulfonate monohydrate R* and 6.8 g of *potassium dihydrogen phosphate R* in 500 mL of *water R*, adjust to pH 3.5 with *phosphoric acid R* and add 500 mL of *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100 → 0	0 → 100
15 - 25	0	100

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *benserazide for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C; doubling of the peak due to impurity C, related to separation of the (*EZ*)-isomers, may be observed.

Relative retention With reference to benserazide (retention time = about 9 min): impurity A = about 0.6; impurity C = about 1.2; impurity B = about 1.5.

System suitability Reference solution (a):
- *resolution*: minimum 5.0 between the peaks due to benserazide and impurity C; use the 1st peak of impurity C if 2 peaks occur.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity B by 0.7;
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurity B*: not more than the area of the peak due to benserazide in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurity C*: not more than the area of the corresponding peak or pair of peaks in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the peak due to benserazide in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *sum of impurities other than A*: not more than twice the area of the peak due to benserazide in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the peak due to benserazide in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.250 g in 5 mL of *anhydrous formic acid R*. Add 70 mL of *anhydrous acetic acid R*. Titrate immediately with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

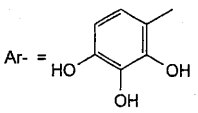
1 mL of *0.1 M perchloric acid* is equivalent to 29.37 mg of $C_{10}H_{16}ClN_3O_5$.

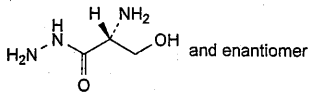
STORAGE

Protected from light.

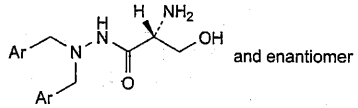
IMPURITIES

Specified impurities *A, B, C.*

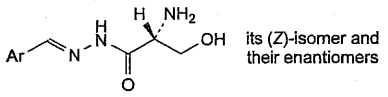




A. (2*RS*)-2-amino-3-hydroxypropanohydrazide,



B. (2*RS*)-2-amino-3-hydroxy-2′,2′-bis(2,3,4-trihydroxybenzyl)propanohydrazide,



its (Z)-isomer and their enantiomers

C. (2*RS*)-2-amino-3-hydroxy-2′-[(1*EZ*)-(2,3,4-trihydroxybenzylidene)]propanohydrazide.

Ph Eur

Bentonite

(Ph. Eur. monograph 0467)

Ph Eur

DEFINITION

Natural clay containing a high proportion of montmorillonite, a native hydrated aluminium silicate in which some aluminium and silicon atoms may be replaced by other atoms such as magnesium and iron.

CHARACTERS

Appearance

Very fine, homogeneous, greyish-white powder with a more or less yellowish or pinkish tint.

Solubility

Practically insoluble in water and in aqueous solutions.

It swells with a little water forming a malleable mass.

IDENTIFICATION

A. To 0.5 g in a metal crucible add 1 g of *potassium nitrate R* and 3 g of *sodium carbonate R* and heat until the mixture melts. Allow to cool. To this residue add 20 mL of boiling *water R*, mix and filter. Wash the insoluble residue with 50 mL of *water R*. To this residue add 1 mL of *hydrochloric acid R* and 5 mL of *water R*. Filter. To the filtrate add 1 mL of *strong sodium hydroxide solution R* and filter. To this filtrate add 3 mL of *ammonium chloride solution R*. A gelatinous white precipitate is formed.

B. Add 2.0 g in 20 portions to 100 mL of a 10 g/L solution of *sodium laurilsulfate R* in a 100 mL graduated cylinder about 30 mm in diameter. Allow 2 min between additions for each portion to settle. Allow to stand for 2 h.
The apparent volume of the sediment is not less than 22 mL.

C. 0.25 g gives the reaction of silicates (*2.3.1*).

TESTS

Alkalinity

To 2 g add 100 mL of *carbon dioxide-free water R* and shake for 5 min. To 5 mL of this suspension add 0.1 mL of *thymolphthalein solution R*. The liquid becomes bluish. Add 0.1 mL of *0.1 M hydrochloric acid*. The liquid is decolourised within 5 min.

Coarse particles

Maximum 0.5 per cent.

To 20 g add 1000 mL of *water R* and mix for 15 min using a high-speed mixer capable of operating at not less than 5000 r/min. Transfer the suspension to a wet sieve (75), tared after drying at 100-105 °C. Wash with 3 quantities, each of 500 mL, of *water R*, ensuring that any agglomerates have been dispersed. Dry the sieve at 100-105 °C and weigh. The particles on the sieve weigh a maximum of 0.1 g.

Loss on drying (*2.2.32*)

Maximum 15 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for bentonite used as viscosity-increasing agent or suspending agent.

Sedimentation volume

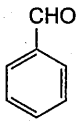
To 6.0 g add 200 mL of *water R* and mix for 20 min using a high-speed mixer capable of operating at 10 000 r/min. Transfer 100 mL of this suspension to a graduated cylinder. Allow to stand for 24 h. The volume of the clear supernatant is not greater than 2 mL.

Swelling power with water

See Identification B.

Ph Eur

Benzaldehyde



C_7H_6O 106.1 *100-52-7*

Action and use

Flavour.

DEFINITION

Benzaldehyde contains not less than 98.0% w/w and not more than 100.5% w/w of C_7H_6O.

CHARACTERISTICS

A clear, colourless liquid.

Slightly soluble in *water*; miscible with *ethanol (96%)* and with *ether*.

TESTS

Refractive index

1.544 to 1.546, Appendix V E.

Weight per mL

1.043 to 1.049 g, Appendix V G.

Free acid

Not more than 1.0% w/v, calculated as benzoic acid, $C_7H_6O_2$, when determined by the following method.
To 10 mL add 20 mL of *ethanol (96%)* previously neutralised to *phenolphthalein solution R1* and titrate with *0.1M sodium hydroxide VS* using *phenolphthalein solution R1* as

indicator. Each mL of *0.1M sodium hydroxide VS* is equivalent to 12.21 mg of $C_7H_6O_2$.

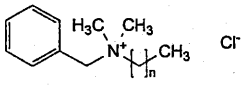
Chlorinated compounds
Not more than 0.05% w/v, calculated as Cl, when determined by the following method. To 5 mL add 50 mL of *isoamyl alcohol* and 3 g of *sodium* and boil under a reflux condenser for 1 hour. Cool, add 50 mL of *water* and 15 mL of *nitric acid*, cool, add 5 mL of *0.1M silver nitrate VS*, shake and titrate the excess silver nitrate with *0.1M ammonium thiocyanate VS* using *ammonium iron(III) sulfate solution R2* as indicator. Repeat the procedure without the substance being examined. The difference between the titrations represents the amount of silver nitrate required. Each mL of *0.1M silver nitrate VS* is equivalent to 3.545 mg of Cl.

ASSAY
Carry out the method for *determination of aldehydes*, Appendix X K, using 0.5 g. Each mL of *0.5M potassium hydroxide in ethanol (60%) VS* is equivalent to 53.06 mg of C_7H_6O.

STORAGE
Benzaldehyde should be kept in a well-filled container, protected from light and stored at a temperature not exceeding 15°.

Benzalkonium Chloride

(Ph. Eur. monograph 0372)



8001-54-5

Action and use
Antiseptic.

Ph Eur

DEFINITION
Mixture of alkylbenzyldimethylammonium chlorides, the alkyl groups mainly having chain lengths of C_{12}, C_{14} and C_{16}.

Content
95.0 per cent to 104.0 per cent of alkylbenzyldimethylammonium chlorides (anhydrous substance) calculated using the average relative molecular mass (see Tests).

CHARACTERS
Appearance
White or yellowish-white powder or gelatinous, yellowish-white fragments, hygroscopic. On heating it forms a clear molten mass.

Solubility
Very soluble in water and in ethanol (96 per cent). An aqueous solution froths copiously when shaken.

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 80 mg in *water R* and dilute to 100.0 mL with the same solvent.

Spectral range 220-350 nm.

Absorption maxima At 257 nm, 263 nm and 269 nm.

Shoulder At about 250 nm.

B. Examine the chromatograms obtained in the test for average relative molecular mass and ratio of alkyl components.

Results The principal peaks in the chromatogram obtained with the test solution are similar in retention time to the principal peaks in the chromatogram obtained with the reference solution.

C. To 2 mL of solution S (see Tests) add 0.1 mL of *glacial acetic acid R* and, dropwise, 1 mL of *sodium tetraphenylborate solution R*. A white precipitate is formed. Filter. Dissolve the precipitate in a mixture of 1 mL of *acetone R* and 5 mL of *ethanol (96 per cent) R*, heating to not more than 70 °C. Add *water R* dropwise to the warm solution until a slight opalescence forms. Heat gently until the solution is clear and allow to cool. White crystals separate. Filter, wash with 3 quantities, each of 10 mL, of *water R* and dry *in vacuo* over *diphosphorus pentoxide R* or *anhydrous silica gel R* at a temperature not exceeding 50 °C. The crystals melt (*2.2.14*) at 127 °C to 133 °C.

D. To 5 mL of *dilute sodium hydroxide solution R* add 0.1 mL of *bromophenol blue solution R1* and 5 mL of *methylene chloride R* and shake. The methylene chloride layer is colourless. Add 0.1 mL of solution S and shake. The methylene chloride layer becomes blue.

E. To 2 mL of solution S add 1 mL of *dilute nitric acid R*. A white precipitate is formed which dissolves on the addition of 5 mL of *ethanol (96 per cent) R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 50 mL of solution S add 0.1 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.1 M hydrochloric acid* or *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Average relative molecular mass and ratio of alkyl components
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.400 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution Dissolve the contents of a vial of *benzalkonium chloride for system suitability CRS* in 5.0 mL of *water R*.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped nitrile silica gel for chromatography R* (5 µm).

Mobile phase Mix 45 volumes of *acetonitrile R* and 55 volumes of a 13.6 g/L solution of *sodium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of homologues Use the chromatogram supplied with *benzalkonium chloride for system suitability CRS* and the chromatogram obtained with the reference solution to identify the peaks due to C_{12}, C_{14} and C_{16}.

Relative retention With reference to C_{12} homologue (retention time = about 6 min): C_{14} homologue = about 1.3; C_{16} homologue = about 1.7.

System suitability Reference solution:

— *resolution*: minimum 1.5 between the peaks due to the C_{12} and C_{14} homologues.

Calculate the average relative molecular mass of the sample by summing the products for each homologue, using the following expression:

$$W\left(\frac{A}{B}\right)$$

A = area of the peak due to the given homologue in the chromatogram obtained with the test solution;

B = sum of the areas of the peaks due to all homologues in the chromatogram obtained with the test solution;

W = relative molecular mass for the given homologue: 340, 368 and 396 for the C_{12}, C_{14} and C_{16} homologues, respectively.

Calculate the percentage of each homologue, using the following expression:

$$100\left(\frac{C}{D}\right)$$

C = product of the relative molecular mass of the given homologue and the area of the corresponding peak in the chromatogram obtained with the test solution;

D = sum of the C values for all homologues quantified.

Limits:

— *C_{12} homologue*: minimum 40 per cent;

— *C_{14} homologue*: minimum 20 per cent;

— *sum of C_{12} and C_{14} homologues*: minimum 70 per cent.

Impurities A, B and C

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.50 g of the substance to be examined in *methanol R1* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 25.0 mg of *benzyl alcohol CRS* (impurity A) in *methanol R1* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 75.0 mg of *benzaldehyde CRS* (impurity B) in *methanol R1* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R1*.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *methanol R1*.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase:

— *mobile phase A*: dissolve 1.09 g of *sodium hexanesulfonate R* and 6.9 g of *sodium dihydrogen phosphate monohydrate R* in *water R*; adjust to pH 3.5 with *phosphoric acid R* and dilute to 1000.0 mL with the same solvent;

— *mobile phase B*: *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	80	20
10 - 14	80 → 50	20 → 50
14 - 35	50	50
35 - 36	50 → 20	50 → 80
36 - 55	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm for impurities A and C, and at 257 nm for impurity B.

Injection 20 µL.

Relative retention With reference to impurity A (retention time = about 10 min): impurity B = about 1.3; impurity C = about 2.4.

System suitability At 210 nm:

— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (c);

— *symmetry factor*: minimum 0.6 for the peak due to impurity A in the chromatogram obtained with reference solution (a).

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.3;

— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *impurity C*: not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Amines and amine salts

Dissolve 5.0 g with heating in 20 mL of a mixture of 3 volumes of *1 M hydrochloric acid* and 97 volumes of *methanol R* and add 100 mL of *2-propanol R*. Pass a stream of *nitrogen R* slowly through the solution. Titrate with up to 12.0 mL of 0.1 M tetrabutylammonium hydroxide and record the potentiometric titration curve (*2.2.20*). If the curve shows 2 points of inflexion, the volume of titrant added between the 2 points is not greater than 5.0 mL. If the curve shows no point of inflexion, the substance to be examined does not comply with the test. If the curve shows 1 point of inflexion, repeat the test but add 3.0 mL of a 25.0 g/L solution of *dimethyldecylamine R* in *2-propanol R* before the titration. If the titration curve after addition of 12.0 mL of the titrant shows only 1 point of inflexion, the substance to be examined does not comply with the test.

Water (*2.5.12*)

Maximum 10 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 2.00 g in *water R* and dilute to 100.0 mL with the same solvent. Transfer 25.0 mL of the solution to a separating funnel, add 25 mL of *methylene chloride R*, 10 mL of *0.1 M sodium hydroxide* and 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R*. Shake well, allow to separate and discard the methylene chloride layer. Shake the aqueous layer with 3 quantities, each of 10 mL, of *methylene chloride R* and discard the methylene chloride layers. To the aqueous layer add 40 mL of *hydrochloric acid R*, allow to cool and titrate with *0.05 M potassium iodate* until the deep-brown

colour is almost discharged. Add 5 mL of *methylene chloride R* and continue the titration, shaking vigorously, until the methylene chloride layer no longer changes colour. Carry out a blank titration on a mixture of 10.0 mL of the freshly prepared 50 g/L solution of *potassium iodide R*, 20 mL of *water R* and 40 mL of *hydrochloric acid R*.

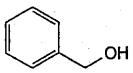
1 mL of *0.05 M potassium iodate* is equivalent to $\frac{x}{10}$ mg of benzalkonium chloride where x is the average relative molecular mass of the sample.

STORAGE

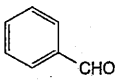
In an airtight container.

IMPURITIES

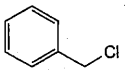
Specified impurities *A, B, C.*



A. benzyl alcohol,



B. benzaldehyde,



C. (chloromethyl)benzene.

Ph Eur

Benzalkonium Chloride Solution

(Ph. Eur. monograph 0371)

Action and use

Antiseptic.

Ph Eur

DEFINITION

Aqueous solution of a mixture of alkylbenzyldimethylammonium chlorides, the alkyl groups mainly having chain lengths of C_{12}, C_{14} and C_{16}.

Content

475 g/L to 525 g/L of alkylbenzyldimethylammonium chlorides, calculated using the average relative molecular mass (see Tests). The solution may contain ethanol (96 per cent).

CHARACTERS

Appearance

Clear, colourless or slightly yellowish liquid.

Solubility

Miscible with water and with ethanol (96 per cent).

It froths copiously when shaken.

IDENTIFICATION

First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dilute 0.3 mL to 100.0 mL with *water R*.

Spectral range 220-350 nm.

Absorption maxima At 257 nm, 263 nm and 269 nm.

Shoulder At about 250 nm.

B. Examine the chromatograms obtained in the test for average relative molecular mass and ratio of alkyl components.

Results The principal peaks in the chromatogram obtained with the test solution are similar in retention time to the principal peaks in the chromatogram obtained with the reference solution.

C. To 0.05 mL add 2 mL of *water R*, 0.1 mL of *glacial acetic acid R* and, dropwise, 1 mL of *sodium tetraphenylborate solution R*. A white precipitate is formed. Filter. Dissolve the precipitate in a mixture of 1 mL of *acetone R* and 5 mL of *ethanol (96 per cent) R*, heating to not more than 70 °C. Add *water R* dropwise to the warm solution until a slight opalescence forms. Heat gently until the solution is clear and allow to cool. White crystals separate. Filter, wash with 3 quantities, each of 10 mL, of *water R* and dry *in vacuo* over *diphosphorus pentoxide R* or *anhydrous silica gel R* at a temperature not exceeding 50 °C. The crystals melt (*2.2.14*) at 127 °C to 133 °C.

D. To 5 mL of *dilute sodium hydroxide solution R* add 0.1 mL of *bromophenol blue solution R1* and 5 mL of *methylene chloride R* and shake. The methylene chloride layer is colourless. Add 0.05 mL of the solution to be examined and shake. The methylene chloride layer becomes blue.

E. To 0.05 mL add 1 mL of *dilute nitric acid R*. A white precipitate is formed which dissolves on the addition of 5 mL of *ethanol (96 per cent) R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dilute 2.0 g to 100 mL with *carbon dioxide-free water R*.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity

To 50 mL of solution S add 0.1 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.1 M hydrochloric acid* or *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Average relative molecular mass and ratio of alkyl components

Liquid chromatography (*2.2.29*).

Test solution Determine the density (*2.2.5*) of the solution to be examined. Dilute a quantity of the solution to be examined equivalent to about 0.400 g of benzalkonium chloride to 100.0 mL with *water R*.

Reference solution Dissolve the contents of a vial of *benzalkonium chloride for system suitability CRS* in 5.0 mL of *water R*.

Column:

— *size: l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase: end-capped nitrile silica gel for chromatography R* (5 µm).

Mobile phase Mix 45 volumes of *acetonitrile R* and 55 volumes of a 13.6 g/L solution of *sodium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of homologues Use the chromatogram supplied with *benzalkonium chloride for system suitability CRS* and the chromatogram obtained with the reference solution to identify the peaks due to homologues C_{12}, C_{14} and C_{16}.

Relative retention With reference to C_{12} homologue (retention time = about 6 min): C_{14} homologue = about 1.3; C_{16} homologue = about 1.7.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to the C_{12} and C_{14} homologues.

Calculate the average relative molecular mass of the sample by summing the products for each homologue, using the following expression:

$$W\left(\frac{A}{B}\right)$$

A = area of the peak due to the given homologue in the chromatogram obtained with the test solution;
B = sum of the areas of the peaks due to all homologues in the chromatogram obtained with the test solution;
W = relative molecular mass for the given homologue: 340, 368 and 396 for the C_{12}, C_{14} and C_{16} homologues, respectively.

Calculate the percentage of each homologue, using the following expression:

$$100\left(\frac{C}{D}\right)$$

C = product of the relative molecular mass of the given homologue and the area of the corresponding peak in the chromatogram obtained with the test solution;
D = sum of the C values for all homologues quantified.

Limits:
— *C_{12} homologue*: minimum 40 per cent;
— *C_{14} homologue*: minimum 20 per cent;
— *sum of C_{12} and C_{14} homologues*: minimum 70 per cent.

Impurities A, B and C

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Determine the density (*2.2.5*) of the solution to be examined. Dilute a quantity of the solution to be examined equivalent to 2.5 g of benzalkonium chloride to 50.0 mL with *methanol R1*.

Reference solution (a) Dissolve 25.0 mg of *benzyl alcohol CRS* (impurity A) in *methanol R1* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 75.0 mg of *benzaldehyde CRS* (impurity B) in *methanol R1* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R1*.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *methanol R1*.

Column:
— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.09 g of *sodium hexanesulfonate R* and 6.9 g of *sodium dihydrogen phosphate monohydrate R* in *water R*; adjust to pH 3.5 with *phosphoric acid R* and dilute to 1000.0 mL with the same solvent;
— *mobile phase B*: *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	80	20
10 - 14	80 → 50	20 → 50
14 - 35	50	50
35 - 36	50 → 20	50 → 80
36 - 55	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm for impurities A and C, and at 257 nm for impurity B.

Injection 20 µL.

Relative retention With reference to impurity A (retention time = about 10 min): impurity B = about 1.3; impurity C = about 2.4.

System suitability At 210 nm:
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (c);
— *symmetry factor*: minimum 0.6 for the peak due to impurity A in the chromatogram obtained with reference solution (a).

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.3;
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *impurity C*: not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Amines and amine salts

Mix 10.0 g, while heating, with 20 mL of a mixture of 3 volumes of *1 M hydrochloric acid* and 97 volumes of *methanol R* and add 100 mL of *2-propanol R*. Pass a stream of *nitrogen R* slowly through the solution. Titrate with up to 12.0 mL of 0.1 M tetrabutylammonium hydroxide and record the potentiometric titration curve (*2.2.20*). If the curve shows 2 points of inflexion, the volume of titrant added between the 2 points is not greater than 5.0 mL. If the curve shows no point of inflexion, the solution to be examined does not comply with the test. If the curve shows 1 point of inflexion, repeat the test but add 3.0 mL of a 25.0 g/L solution of *dimethyldecylamine R* in *2-propanol R* before the titration. If the titration curve after the addition of 12.0 mL of the titrant shows only 1 point of inflexion, the solution to be examined does not comply with the test.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Determine the density (*2.2.5*) of the solution to be examined. Dilute 4.00 g to 100.0 mL with *water R*. Transfer 25.0 mL of the solution to a separating funnel, add 25 mL of *methylene chloride R*, 10 mL of *0.1 M sodium hydroxide* and 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R*. Shake well, allow to separate and discard the methylene chloride layer. Shake the aqueous layer with 3 quantities, each of 10 mL, of *methylene chloride R* and discard the methylene chloride layers. To the aqueous layer add 40 mL of *hydrochloric acid R*, allow to cool and titrate with *0.05 M potassium iodate* until the deep-brown colour is almost discharged. Add 5 mL of *methylene chloride R* and

continue the titration, shaking vigorously, until the methylene chloride layer no longer changes colour. Carry out a blank titration on a mixture of 10.0 mL of the freshly prepared 50 g/L solution of *potassium iodide R*, 20 mL of *water R* and 40 mL of *hydrochloric acid R*.

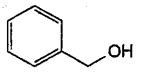
1 mL of *0.05 M potassium iodate* is equivalent to $\frac{x}{10}$ mg of benzalkonium chloride where x is the average relative molecular mass of the sample.

LABELLING

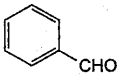
The label states the content of ethanol (96 per cent), if any.

IMPURITIES

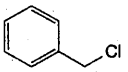
Specified impurities *A, B, C.*



A. benzyl alcohol,



B. benzaldehyde,



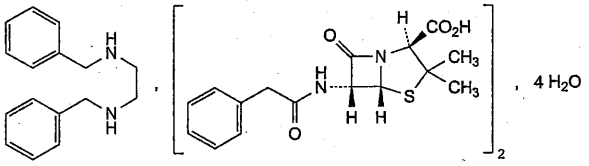
C. (chloromethyl)benzene.

Ph Eur

Benzathine Benzylpenicillin Tetrahydrate

Benzathine Benzylpenicillin

(Benzylpenicillin (Benzathine) Tetrahydrate, Ph. Eur. monograph 0373)



$C_{48}H_{56}N_6O_8S_2,4H_2O$ 981 *41372-02-5*

Action and use

Penicillin antibacterial.

Ph Eur

DEFINITION

N^1,N^2-Dibenzylethane-1,2-diamine bis[(2*S*,5*R*,6*R*)-3,3-dimethyl-7-oxo-6-(2-phenylacetamido)-4-thia-1-azabicyclo [3.2.0]heptane-2-carboxylate] tetrahydrate.

Salt obtained from *Benzylpenicillin sodium (0114)* or *Benzylpenicillin potassium (0113)* produced by the growth of certain strains of *Penicillium notatum* or related micro-organisms.

Content

— *benzathine benzylpenicillin*: 94.5 per cent to 102.0 per cent (anhydrous substance) without correction for dispersing or suspending agents;

— *benzathine*: 24.0 per cent to 27.0 per cent (anhydrous substance).

Dispersing or suspending agents (e.g. lecithin and polysorbate 80) may be added.

CHARACTERS

Appearance

White or almost white, slightly hygroscopic powder.

Solubility

Very slightly soluble in water, freely soluble in dimethylformamide and in formamide, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzathine benzylpenicillin CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *methanol R.*

Reference solution Dissolve 25 mg of *benzathine benzylpenicillin CRS* in 5 mL of *methanol R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 7.0 with *ammonia R.*

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution:

— the chromatogram shows 2 clearly separated spots.

Results The 2 principal spots in the chromatogram obtained with the test solution are similar in position, colour and size to the 2 principal spots in the chromatogram obtained with the reference solution.

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube on a water-bath for 1 min; a reddish-brown colour develops.

D. To 0.1 g add 2 mL of *1 M sodium hydroxide* and shake for 2 min. Shake the mixture with 2 quantities, each of 3 mL, of *ether R.* Evaporate the combined ether layers to dryness and dissolve the residue in 1 mL of *ethanol (50 per cent V/V) R.* Add 5 mL of *picric acid solution R*, heat at 90 °C for 5 min and allow to cool slowly. Separate the crystals and recrystallise from *ethanol (25 per cent V/V) R* containing 10 g/L of *picric acid R.* The crystals melt (*2.2.14*) at about 214 °C.

TESTS

Acidity or alkalinity

To 0.50 g add 100 mL of *carbon dioxide-free water R* and shake for 5 min. Filter through a sintered-glass filter (*2.1.2*). To 20 mL of the filtrate add 0.1 mL of *bromothymol blue solution R1.* The solution is green or yellow. Not more than 0.2 mL of *0.02 M sodium hydroxide* is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and by diluting to volume immediately after dissolution.*

Solution A Prepare a solution containing 1.3 g/L of *disodium hydrogen phosphate dodecahydrate R* and 6.8 g/L of *potassium dihydrogen phosphate R*.

Test solution (a) Dissolve 40.0 mg of the substance to be examined in 50 mL of *methanol R* and dilute to 100.0 mL with solution A.

Test solution (b) Dissolve 70.0 mg of the substance to be examined in 25 mL of *methanol R* and dilute to 50.0 mL with solution A.

Reference solution (a) Dissolve 40.0 mg of *benzathine benzylpenicillin CRS* in 50 mL of *methanol R* and dilute to 100.0 mL with solution A.

Reference solution (b) Dissolve 3 mg of *benzathine benzylpenicillin for peak identification CRS* (containing impurities A, B, C, D, E, F, G, H, I, J and K) in 1 mL of *methanol R* and dilute to 2 mL with solution A.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 20.0 mL with a mixture of equal volumes of *methanol R* and solution A.

Reference solution (d) Dilute 3.0 mL of reference solution (c) to 100.0 mL with a mixture of equal volumes of *methanol R* and solution A.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: mix 10 volumes of a 34 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.3 with *phosphoric acid R*, 30 volumes of *methanol R1* and 60 volumes of *water for chromatography R*;
- *mobile phase B*: mix 5 volumes of a 34 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.3 with *phosphoric acid R*, 25 volumes of *water for chromatography R* and 70 volumes of *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	85	15
2 - 16	85 → 0	15 → 100
16 - 26	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (b) and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *benzathine benzylpenicillin for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G, H, I, J and K.

Relative retention With reference to benzylpenicillin (retention time = about 7 min): impurity A = about 0.18; benzathine = about 0.30; impurity D = about 0.36; impurity G = about 0.38; impurity J = about 0.44; impurity E = about 0.51 and 0.60; impurity B = about 0.69; impurity F = about 0.84 and 0.88; impurity H = about 1.22; impurity I = about 1.42; impurity C = about 1.75; impurity K = about 2.90.

System suitability:
- *resolution*: minimum 1.0 between the peaks due to the epimers of impurity F and minimum 1.5 between the peaks due to impurities D and G in the chromatogram obtained with reference solution (b);
- *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (d).

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 1.9; impurity F = 1.5;
- for each impurity, use the concentration of benzylpenicillin in reference solution (c).

Limits:
- *impurity C*: maximum 2.0 per cent;
- *impurity K*: maximum 1.0 per cent;
- *impurity J*: maximum 0.5 per cent;
- *impurities E (sum of isomers), F (sum of epimers)*: for each impurity, maximum 0.3 per cent;
- *impurities A, B, D, G, H, I*: for each impurity, maximum 0.2 per cent;
- *any other impurity*: for each impurity, maximum 0.2 per cent;
- *total*: maximum 3.5 per cent;
- *reporting threshold*: 0.05 per cent; disregard the peak due to benzathine.

Water (*2.5.12*)

5.0 per cent to 8.0 per cent, determined on 0.200 g.

Bacterial endotoxins (*2.6.14, Method E*)

Less than 0.13 IU/mL, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

Suspend 20 mg in 20 mL of a solution of *0.1 M sodium hydroxide* diluted 1 to 100, shake thoroughly and centrifuge. Examine the supernatant.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase B, mobile phase A (15:85 *V/V*).

Injection Test solution (a) and reference solution (a).

System suitability Reference solution (a):
- *symmetry factor*: maximum 1.8 for the peak due to benzylpenicillin.

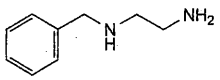
Calculate the percentage contents of benzathine ($C_{16}H_{20}N_2$) and benzathine benzylpenicillin ($C_{48}H_{56}N_6O_8S_2$) taking into account the assigned content of *benzathine benzylpenicillin CRS*.

STORAGE

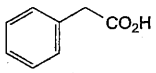
In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

IMPURITIES

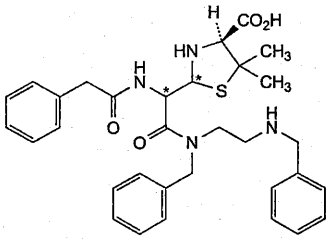
Specified impurities *A, B, C, D, E, F, G, H, I, J, K.*



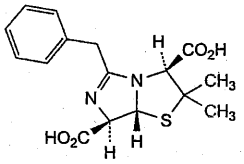
A. N^1-benzylethane-1,2-diamine,



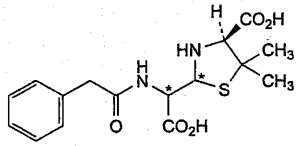
B. phenylacetic acid,



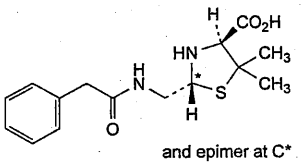
C. (2*E*,4*S*)-2-[(1*E*)-2-[benzyl[2-(benzylamino)ethyl]amino]-2-oxo-1-(2-phenylacetamido)ethyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (benzylpenicilloic acids benzathide),



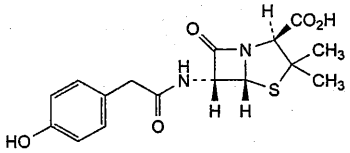
D. (3*S*,7*R*,7a*R*)-5-benzyl-2,2-dimethyl-2,3,7,7a-tetrahydroimidazo[5,1-*b*][1,3]thiazole-3,7-dicarboxylic acid (penillic acid of benzylpenicillin),



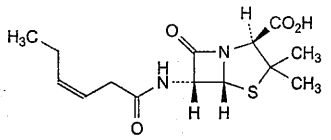
E. (2*E*,4*S*)-2-[(*E*)-carboxy(2-phenylacetamido)methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acids of benzylpenicillin),



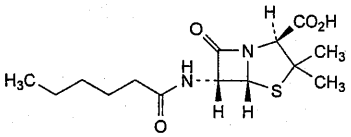
F. (2*RS*,4*S*)-5,5-dimethyl-2-[(2-phenylacetamido)methyl]-1,3-thiazolidine-4-carboxylic acid (penilloic acids of benzylpenicillin),



G. (2*S*,5*R*,6*R*)-6-[2-(4-hydroxyphenyl)acetamido]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,

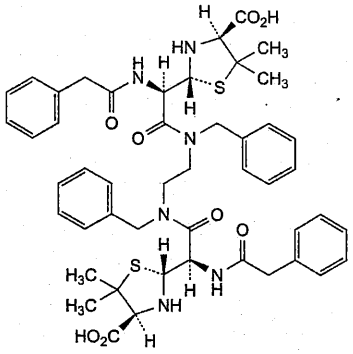


H. (2*S*,5*R*,6*R*)-6-[(3*Z*)-hex-3-enamido]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (isopenicillin F),



I. (2*S*,5*R*,6*R*)-6-hexanamido-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (dihydropenicillin F),

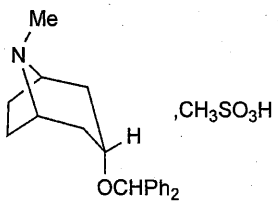
J. unknown structure,



K. (2*R*,2′*R*,4*S*,4′*S*)-2,2′-[(4*R*,11*R*)-6,9-dibenzyl-2,5,10,13-tetraoxo-1,14-diphenyl-3,6,9,12-tetraazatetradecane-4,11-diyl]bis(5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid).

Ph Eur

Benzatropine Mesilate



$C_{21}H_{25}NO,CH_4O_3S$ 403.5 *132-17-2*

Action and use
Anticholinergic.

Preparations
Benzatropine Injection
Benzatropine Tablets

DEFINITION
Benzatropine Mesilate is (1*R*,3*R*,5*S*)-3-benzhydryloxytropane methanesulfonate. It contains not less than 98.0% and not more than 100.5% of $C_{21}H_{25}NO,CH_4O_3S$, calculated with reference to the dried substance.

PRODUCTION

Risk assessment should be used to evaluate the potential for genotoxic methanesulfonate esters to be formed in the presence of low molecular weight alcohols. If a risk of methanesulfonate ester formation is identified through risk assessment, these impurities should not exceed the threshold of toxicological concern.

CHARACTERISTICS

A white, crystalline powder. It melts at about 144°.

Very soluble in *water*; freely soluble in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION

A. Dry the substance at 105° for 3 hours. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of benzatropine mesilate (*RS 026*).

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.1% w/v solution in *2M hydrochloric acid* exhibits two maxima, at 253 and 258 nm. The *absorbance* at 253 nm is about 0.96 and at 258 nm is about 1.1.

C. Dissolve 10 mg in 2 mL of *water*, pour into 5 mL of hot *picric acid solution R1* and allow to cool. The *melting point* of the precipitate, after drying at 105°, is about 185°, Appendix V A.

TESTS

Tropine

Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and a mixture of 75 volumes of *ethanol (96%)* and 15 volumes of *13.5M ammonia* as the mobile phase. Apply separately to the plate 10 µL of each of two solutions in *acetone* containing (1) 4.0% w/v of the substance being examined and (2) 0.020% w/v of *tropine*. After removal of the plate, allow it to dry in air and spray with *sodium iodobismuthate solution* and then with a 0.4% w/v solution of *sulfuric acid*. Any spot corresponding to tropine in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions. For solution (1) mix with the aid of ultrasound 50 mg of the substance being examined with 15 mL of mobile phase A, dilute to 50 mL with the same solvent and filter. For solution (2) dilute 1 volume of solution (1) to 100 volumes with mobile phase A and further dilute 1 volume of the resulting solution to 5 volumes with the same solvent. For solution (3) mix with the aid of ultrasound 50 mg of *desmethyl benzatropine hydrochloride BPCRS* with 15 mL of mobile phase A, dilute to 100 mL and dilute 1 volume of the resulting solution to 100 volumes with the same solvent. Solution (4) contains 0.01% w/v each of *benzatropine mesilate BPCRS* and *desmethyl benzatropine hydrochloride BPCRS* in mobile phase A.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with *phenylsilyl silica gel for chromatography* (5 µm) (Zorbax SB-Phenyl 5µ is suitable). Carry out a linear gradient elution with a flow rate of 1 mL per minute using the following conditions. Use a detection wavelength of 220 nm.

Mobile phase A A mixture of 5 volumes of a 1M potassium phosphate buffer prepared as described for mobile phase B, 20 volumes of *acetonitrile* and 75 volumes of *water*.

Mobile phase B A mixture of 35 volumes of *water*, 60 volumes of *acetonitrile* and 5 volumes of a 1M potassium phosphate buffer prepared in the following manner: dissolve 136.1 g of *potassium dihydrogen orthophosphate* in 900 mL of *water*, add 5 mL of *orthophosphoric acid (85%)* and dilute to 1000 mL.

Time (Minutes)	Mobile phase A (% v/v)	Mobile phase B (% v/v)	Comment
0-20	70→30	30→70	linear gradient
20-30	30→0	70→100	linear gradient
30-55	0	100	isocratic
55-65	70	30	isocratic

Inject 20 µL of solution (4). The test is not valid unless the *resolution factor* between the two principal peaks is at least 1. If necessary adjust the concentration of *acetonitrile* or adjust the time program of the linear gradient elution.

Inject separately 20 µL of mobile phase A as a blank and 20 µL each of solutions (1), (2) and (3). In the chromatogram obtained with solution (1) the area of any peak corresponding to desmethyl benzatropine is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.5%), the area of any other *secondary peak* is not greater that the area of the principal peak in the chromatogram obtained with solution (2) (0.2%) and the sum of the areas of any such peaks is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%). In solution (1) disregard any peaks corresponding to the peaks in the chromatogram obtained with the blank solution.

Loss on drying

When dried to constant weight at 105°, loses not more than 5.0% of its weight. Use 1 g.

Sulfated ash

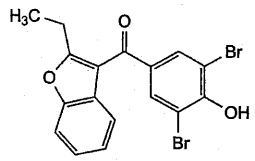
Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.6 g in 25 mL of *water*, add 5 mL of *dilute sodium carbonate solution* and extract with four 10 mL quantities of *chloroform*. Wash the combined extracts with 10 mL of *water*, extract the washings with 5 mL of *chloroform* and add the chloroform to the combined extracts. Filter and wash the filter with 5 mL of *chloroform*. To the combined filtrate and washings add 25 mL of *1,4-dioxan* and titrate with *0.1M perchloric acid VS* using 0.15 mL of a 0.1% w/v solution of *methyl red* in *methanol* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 40.35 mg of $C_{21}H_{25}NO$, CH_4O_3S.

Benzbromarone

(Ph. Eur. monograph 1393)



$C_{17}H_{12}Br_2O_3$ 424.1 *3562-84-3*

Action and use
Uricosuric; treatment of hyperuricaemia.

Ph Eur

DEFINITION
(3,5-Dibromo-4-hydroxyphenyl)(2-ethylbenzofuran-3-yl)methanone.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in methylene chloride, sparingly soluble in ethanol (96 per cent).

mp
About 152 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzbromarone CRS*.

B. By means of a copper wire, previously ignited, introduce a small amount of the substance to be examined into the non-luminous part of a flame. The colour of the flame becomes green.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 1.25 g in *dimethylformamide R* and dilute to 25 mL with the same solvent.

Acidity or alkalinity
Shake 0.5 g with 10 mL of *carbon dioxide-free water R* for 1 min and filter. To 2.0 mL of the filtrate add 0.1 mL of *methyl red solution R* and 0.1 mL of *0.01 M hydrochloric acid*. The solution is red. Add 0.3 mL of *0.01 M sodium hydroxide*. The solution is yellow.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.125 g of the substance to be examined in 30 mL of *methanol R* and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *benzarone CRS* (impurity C) in the mobile phase and dilute to 20 mL with the mobile phase. To 5 mL of this solution add 1 mL of the test solution and dilute to 100 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *glacial acetic acid R, acetonitrile R, water R, methanol R* (5:25:300:990 *V/V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 231 nm.

Injection 20 µL.

Run time 2.5 times the retention time of benzbromarone.

Relative retention With reference to benzbromarone: impurity A = about 0.6; impurity B = about 2.

System suitability Reference solution (b):
— *resolution*: minimum 10.0 between the peaks due to impurity C (1st peak) and benzbromarone (2nd peak).

Limits:
— *impurity A*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *impurity B*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than A and B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Halides expressed as chlorides (*2.4.4*)
Maximum 400 ppm.

Shake 1.25 g with a mixture of 5 mL of *dilute nitric acid R* and 15 mL of *water R*. Filter. Rinse the filter with *water R* and dilute the filtrate to 25 mL with the same solvent. Dilute 2.5 mL of this solution to 15 mL with *water R*.

Iron (*2.4.9*)
Maximum 125 ppm.

Moisten the residue obtained in the test for sulfated ash with 2 mL of *hydrochloric acid R* and evaporate to dryness on a water-bath. Add 0.05 mL of *hydrochloric acid R* and 10 mL of *water R*, heat to boiling and maintain boiling for 1 min. Allow to cool. Rinse the crucible with *water R*, collect the rinsings and dilute to 25 mL with *water R*. Dilute 2 mL of this solution to 10 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 50 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

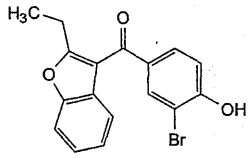
ASSAY
Dissolve 0.300 g in 60 mL of *methanol R*. Stir until completely dissolved and add 10 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 42.41 mg of $C_{17}H_{12}Br_2O_3$.

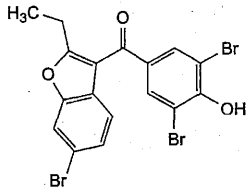
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B.*

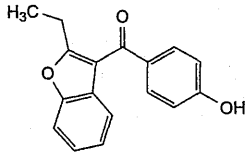
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) C.



A. (3-bromo-4-hydroxyphenyl)(2-ethylbenzofuran-3-yl) methanone,



B. (6-bromo-2-ethylbenzofuran-3-yl)(3,5-dibromo-4-hydroxyphenyl)methanone,

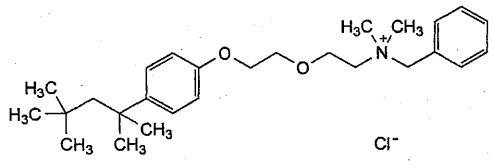


C. (2-ethylbenzofuran-3-yl)(4-hydroxyphenyl)methanone (benzarone).

Ph Eur

Benzethonium Chloride

(Ph. Eur. monograph 0974)



$C_{27}H_{42}ClNO_2$ 448.1 *121-54-0*

Action and use
Antiseptic.

Ph Eur

DEFINITION
N-Benzyl-*N*,*N*-dimethyl-2-[2-[4-(1,1,3,3-tetramethylbutyl)phenoxy]ethoxy]ethanaminium chloride.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish-white powder.

Solubility
Very soluble in water and in ethanol (96 per cent), freely soluble in methylene chloride.

An aqueous solution froths copiously when shaken.

IDENTIFICATION
A. Melting point (*2.2.14*): 158 °C to 164 °C, after drying at 105 °C for 4 h.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 25 mg of *benzethonium chloride CRS* in *water R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, methanol R* (5:5:100 *V/V/V*).

Application 20 µL.

Development Over a path of 12 cm.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 5 mL of *dilute sodium hydroxide solution R* add 0.1 mL of *bromophenol blue solution R1* and 5 mL of *methylene chloride R* and shake. The lower layer is colourless. Add 0.1 mL of solution S (see Tests) and shake. A blue colour develops in the lower layer.

D. To 2 mL of solution S add 1 mL of *dilute nitric acid R*. A white precipitate is formed which dissolves upon addition of 5 mL of *ethanol (96 per cent) R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 25 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution is colourless. Add 0.3 mL of *0.01 M sodium hydroxide*. The solution is pink. Add 0.1 mL of *methyl red solution R* and 0.5 mL of *0.01 M hydrochloric acid*. The solution is orange-red.

Volatile bases and salts of volatile bases (*2.4.1, Method B*)
Maximum 50 ppm, determined on 0.20 g.

Prepare the standard using 0.1 mL of *ammonium standard solution (100 ppm NH_4) R*. Replace heavy magnesium oxide by 2.0 mL of *strong sodium hydroxide solution R*.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 2.000 g in *water R* and dilute to 100.0 mL with the same solvent. Transfer 25.0 mL of the solution to a

separating funnel, add 10 mL of a 4 g/L solution of *sodium hydroxide R*, 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R* and 25 mL of *methylene chloride R*. Shake vigorously, allow to separate and discard the lower layer. Shake the upper layer with 3 quantities, each of 10 mL, of *methylene chloride R* and discard the lower layers. To the upper layer add 40 mL of *hydrochloric acid R*, allow to cool and titrate with *0.05 M potassium iodate* until the deep brown colour is almost discharged. Add 4 mL of *methylene chloride R* and continue the titration, shaking vigorously, until the lower layer is no longer brown. Carry out a blank titration using a mixture of 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R*, 20 mL of *water R* and 40 mL of *hydrochloric acid R*.

1 mL of *0.05 M potassium iodate* is equivalent to 44.81 mg of $C_{27}H_{42}ClNO_2$.

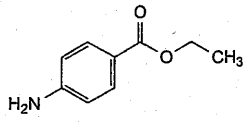
STORAGE

Protected from light.

Ph Eur

Benzocaine

(Ph. Eur. monograph 0011)



$C_9H_{11}NO_2$ 165.2 *94-09-7*

Action and use

Local anaesthetic.

Ph Eur

DEFINITION

Ethyl 4-aminobenzoate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzocaine CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1*, *water for chromatography R* (50:50 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in 5 mL of *acetonitrile R1* and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *4-nitrobenzoic acid R* (impurity E) in 10.0 mL of the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.10 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (3 µm);

— *temperature*: 35 °C.

Mobile phase:

— *mobile phase A*: dilute 1 mL of *perchloric acid R* to 100 mL with *water for chromatography R*; dilute 1 mL of the solution to 100 mL with *water for chromatography R*; mix 9 volumes of this solution and 1 volume of *acetonitrile R1*;

— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 15	100 → 38.5	0 → 61.5

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity E.

Relative retention With reference to benzocaine (retention time = about 10 min): impurity E = about 0.9.

System suitability Reference solution (b):

— *resolution*: minimum 5.0 between the peaks due to impurity E and benzocaine.

Calculation of percentage contents:

— for each impurity, use the concentration of benzocaine in reference solution (a).

Limits:

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 0.2 per cent;

— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo*.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Carry out the determination of primary aromatic amino-nitrogen (*2.5.8*), using 0.400 g dissolved in a mixture of 25 mL of *hydrochloric acid R* and 50 mL of *water R*.

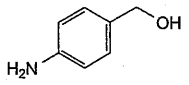
1 mL of *0.1 M sodium nitrite* is equivalent to 16.52 mg of $C_9H_{11}NO_2$.

STORAGE

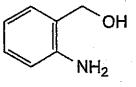
Protected from light.

IMPURITIES

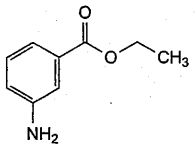
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H.*



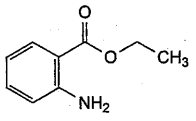
A. (4-aminophenyl)methanol,



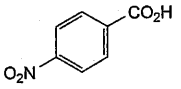
B. (2-aminophenyl)methanol,



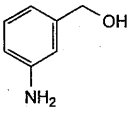
C. ethyl 3-aminobenzoate,



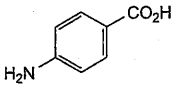
D. ethyl 2-aminobenzoate,



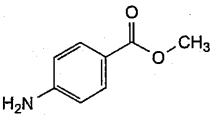
E. 4-nitrobenzoic acid,



F. (3-aminophenyl)methanol,



G. 4-aminobenzoic acid,

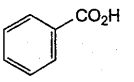


H. methyl 4-aminobenzoate.

Ph Eur

Benzoic Acid

(Ph. Eur. monograph 0066)



$C_7H_6O_2$ 122.1 *65-85-0*

Action and use
Antimicrobial preservative.

Preparations
Compound Benzoic Acid Ointment
Benzoic Acid Solution

Ph Eur

DEFINITION
Benzenecarboxylic acid.

Content
99.0 per cent to 100.5 per cent.

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Slightly soluble in water, soluble in boiling water, freely soluble in ethanol (96 per cent) and in fatty oils.

IDENTIFICATION
A. Melting point (*2.2.14*): 121 °C to 124 °C.
B. Solution S (see Tests) gives reaction (a) of benzoates (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *ethanol (96 per cent) R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Carbonisable substances
Dissolve 0.5 g with shaking in 5 mL of *sulfuric acid R*. After 5 min, the solution is not more intensely coloured than reference solution Y_5 (*2.2.2, Method I*).

Oxidisable substances
Dissolve 0.2 g in 10 mL of boiling *water R*. Cool, shake and filter. To the filtrate add 1 mL of *dilute sulfuric acid R* and 0.2 mL of *0.02 M potassium permanganate*. After 5 min, the solution is still coloured pink.

Halogenated compounds and halides
Maximum 300 ppm.

All glassware used must be chloride-free and may be prepared by soaking overnight in a 500 g/L solution of nitric acid R, rinsed with water R and stored full of water R. It is recommended that glassware be reserved for this test.

Solution (a) Dissolve 6.7 g in a mixture of 40 mL of *1 M sodium hydroxide* and 50 mL of *ethanol (96 per cent) R* and dilute to 100.0 mL with *water R*. To 10.0 mL of this solution add 7.5 mL of *dilute sodium hydroxide solution R* and 0.125 g of *nickel-aluminium alloy R* and heat on a water-bath for 10 min. Allow to cool to room temperature, filter into a 25 mL volumetric flask and wash with 3 quantities, each of 2 mL, of *ethanol (96 per cent) R*. Dilute the filtrate and

washings to 25.0 mL with *water R*. This solution is used to prepare solution A.

Solution (b) In the same manner, prepare a similar solution without the substance to be examined. This solution is used to prepare solution B.

In four 25 mL volumetric flasks, place separately 10 mL of solution (a), 10 mL of solution (b), 10 mL of *chloride standard solution (8 ppm Cl) R* (used to prepare solution C) and 10 mL of *water R*. To each flask add 5 mL of *ferric ammonium sulfate solution R5*, mix and add dropwise and with swirling 2 mL of *nitric acid R* and 5 mL of *mercuric thiocyanate solution R*. Shake. Dilute the contents of each flask to 25.0 mL with *water R* and allow the solutions to stand in a water-bath at 20 °C for 15 min. Measure at 460 nm the absorbance (*2.2.25*) of solution A using solution B as the compensation liquid, and the absorbance of solution C using the solution obtained with 10 mL of *water R* as the compensation liquid. The absorbance of solution A is not greater than that of solution C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

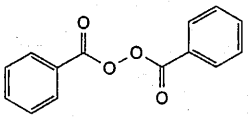
ASSAY
Dissolve 0.200 g in 20 mL of *ethanol (96 per cent) R* and titrate with *0.1 M sodium hydroxide*, using 0.1 mL of *phenol red solution R* as indicator, until the colour changes from yellow to violet-red.

1 mL of *0.1 M sodium hydroxide* is equivalent to 12.21 mg of $C_7H_6O_2$.

Ph Eur

Hydrous Benzoyl Peroxide

(Ph. Eur. monograph 0704)



$C_{14}H_{10}O_4$ 242.2 *94-36-0*
(anhydrous substance)Anhydrous benzoyl peroxide

Action and use
Used topically in the treatment of acne.

Preparations
Benzoyl Peroxide and Clindamycin Gel
Benzoyl Peroxide Cream
Benzoyl Peroxide Gel
Benzoyl Peroxide Lotion
Potassium Hydroxyquinoline Sulfate and Benzoyl Peroxide Cream

Ph Eur

DEFINITION
Content
— *dibenzoyl peroxide*: 70.0 per cent to 77.0 per cent;
— *water*: minimum 20.0 per cent.

CHARACTERS
Appearance
White or almost white, amorphous or granular powder.

Solubility
Practically insoluble in water, soluble in acetone, soluble in methylene chloride with the separation of water, slightly soluble in ethanol (96 per cent).
It loses water rapidly on exposure to air with a risk of explosion.
Mix the entire sample thoroughly before carrying out the following tests.

IDENTIFICATION
First identification: B
Second identification: A, C, D.
A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).
Solution A Dissolve 80.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *ethanol (96 per cent) R*.
Solution B Dilute 10.0 mL of solution A to 100.0 mL with *ethanol (96 per cent) R*.
Spectral ranges 250-300 nm for solution A; 220-250 nm for solution B.
Absorption maxima At 274 nm for solution A; at 235 nm for solution B.
Shoulder At about 282 nm for solution A.
Absorbance ratio A_{235}/A_{274} = 1.17 to 1.21.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of hydrous benzoyl peroxide.*
C. Dissolve about 25 mg in 2 mL of *acetone R*. Add 1 mL of a 10 g/L solution of *diethylphenylenediamine sulfate R* and mix. A red colour develops which quickly darkens and becomes dark violet within 5 min.
D. To 1 g add 5 mL of *ethanol (96 per cent) R*, 5 mL of *dilute sodium hydroxide solution R* and 10 mL of *water R*. Boil the mixture under reflux for 20 min. Cool. The solution gives reaction (c) of benzoates (*2.3.1*).

TESTS
Acidity
Dissolve a quantity of the substance to be examined containing the equivalent of 1.0 g of dibenzoyl peroxide in 25 mL of *acetone R*, add 75 mL of *water R* and filter. Wash the residue with two quantities, each of 10 mL, of *water R*. Combine the filtrate and the washings and add 0.25 mL of *phenolphthalein solution R1*. Not more than 1.25 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator. Carry out a blank test.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*
Test solution Dissolve a quantity of the substance to be examined containing the equivalent of 0.10 g of dibenzoyl peroxide in *acetonitrile R* and dilute to 50 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R*.
Reference solution (b) Dissolve 30.0 mg of *benzoic acid R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 50.0 mg of *ethyl benzoate R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve 50.0 mg of *benzaldehyde R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (e) Dissolve 30.0 mg of *benzoic acid R* and 30.0 mg of *benzaldehyde R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase *glacial acetic acid R, acetonitrile R, water R* (1:500:500 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 µL loop injector.

Run time 2 times the retention time of dibenzoyl peroxide.

Relative retention With reference to dibenzoyl peroxide (retention time = about 28.4 min): impurity B = about 0.15; impurity A = about 0.2; impurity C = about 0.4.

System suitability Reference solution (e):
— *resolution*: minimum 6 between the peaks due to benzoic acid and benzaldehyde.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.25 per cent);
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Chlorides (*2.4.4*)
Maximum 0.4 per cent.

Dissolve a quantity of the substance to be examined containing the equivalent of 0.5 g of dibenzoyl peroxide in 15 mL of *acetone R*. Add, while stirring, 50 mL of 0.05 M nitric acid. Allow to stand for 10 min and filter. Wash the residue with 2 quantities, each of 10 mL, of 0.05 M nitric acid. Combine the filtrate and the washings and dilute to 100 mL with 0.05 M nitric acid. Dilute 2.5 mL of the solution to 15.0 mL with *water R*.

ASSAY

Solution (a) Dissolve 2.500 g immediately before use in 75 mL of *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Dibenzoyl peroxide
To 5.0 mL of solution (a) add 20 mL of *acetone R* and 3 mL of a 500 g/L solution of *potassium iodide R* and mix. Allow to stand for 1 min. Titrate with *0.1 M sodium thiosulfate* using 1 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 12.11 mg of $C_{14}H_{10}O_4$.

Water (*2.5.12*)
Carry out the semi-micro determination of water, using 5.0 mL of solution (a). Use as the solvent a mixture of 20.0 mL of *anhydrous methanol R* and 3.0 mL of a 100 g/L solution of *potassium iodide R* in *dimethylformamide R*. After adding solution (a), stir for 5 min before starting the titration. Carry out a blank determination.

Calculate the percentage content of water using the following expression:

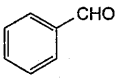
$$\frac{(n_1 - n_2) \times w \times 2}{m} + (p \times 0.0744)$$

n_1 = number of millilitres of *iodosulfurous reagent R* used in the sample determination,
n_2 = number of millilitres of *iodosulfurous reagent R* used in the blank determination,
w = water equivalent of *iodosulfurous reagent R* in milligrams of water per millilitre of reagent,
m = mass of the substance to be examined used for the preparation of solution (a) in grams,
p = percentage content of dibenzoyl peroxide.

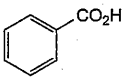
STORAGE

In a container that has been treated to reduce static discharge and that has a device for release of excess pressure, at a temperature of 2 °C to 8 °C, protected from light.

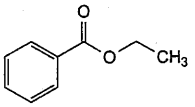
IMPURITIES



A. benzaldehyde,



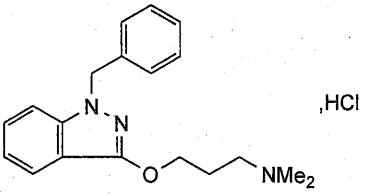
B. benzoic acid,



C. ethyl benzoate.

Ph Eur

Benzydamine Hydrochloride



$C_{19}H_{23}N_3O,HCl$ 345.9 *132-69-4*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Benzydamine Cream
Benzydamine Mouthwash
Benzydamine Oromucosal Spray

DEFINITION

Benzydamine Hydrochloride is 3-(1-benzylindazol-3-yloxy) propyldimethylamine hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{19}H_{23}N_3O$,HCl, calculated with reference to the dried substance.

PRODUCTION

The method of manufacture is such that the level of 3-chloropropyl(dimethyl)amine hydrochloride is not more than 5 ppm when determined by a suitable method.

CHARACTERISTICS

A white crystalline powder.

Very soluble in *water*; freely soluble in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of benzydamine hydrochloride *(RS 027)*.

B. Yields reaction A characteristic of *chlorides*, Appendix VI.

TESTS

Clarity and colour of solution

A 10.0% w/v solution is *clear*, Appendix IV A, and not more intensely coloured than *reference solution* Y_6, Appendix IV B, Method II.

Acidity

pH of a 10% w/v solution, 4.0 to 5.5, Appendix V L.

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol (50%)*. Solution (1) contains 0.25% w/v of the substance being examined. Solution (2) contains 0.0005% w/v of *3-dimethylaminopropyl 2-benzylaminobenzoate hydrochloride BPCRS* (impurity A) and 0.00125% w/v of *3-(1,5-dibenzyl-1H-indazole-3-yl)oxypropyldimethylamine hydrochloride BPCRS* (impurity B). Solution (3) contains 0.00025% w/v of *1-benzyl-1H-indazol-3-ol BPCRS* (impurity C). Solution (4) contains 0.00025% w/v of the substance being examined. Solution (5) contains equal volumes of solutions (1), (2) and (3).

The chromatographic procedure may be carried out using a stainless steel column (25 cm × 4.6 mm) packed with *amido-alkylsilyl silica gel for chromatography* (5 µm) (Suplex pKB-100 is suitable), fitted with a stainless steel guard column (2 cm × 4.6 mm) packed with the same material. Use as the initial mobile phase with a flow rate of 1.5 mL per minute, a mixture of 50 volumes of solution A and 50 volumes of *methanol*. Solution A contains 0.01M *potassium dihydrogen orthophosphate* and 0.005M *sodium octyl sulfate* in *water*, adjusted to pH 3.0 ± 0.1 with *orthophosphoric acid*. Carry out a linear gradient elution increasing the percentage of *methanol* to 70% over 20 minutes from the moment of injection and then decreasing the percentage of *methanol* to 50% over 2 minutes and maintain the final mobile phase until the completion of that run. Use a detection wavelength of 320 nm.

Inject 20 µL of solution (5) and modify the rate of change of the mobile phase, if necessary, to obtain a retention time of about 10 minutes for the substance being examined.

The test is not valid unless, in the chromatogram obtained with solution (5), the *resolution factor* between any two adjacent peaks is at least 2.5.

Inject 20 µL of solution (1) and allow the chromatography to proceed for 30 minutes. In the chromatogram obtained the areas of any peak corresponding to impurity A or impurity B is not greater than the area of the corresponding peak in the chromatogram obtained with solution (2) (0.2% of impurity A and 0.5% of impurity B), the area of any peak corresponding to impurity C is not greater than the area of the peak in the chromatogram obtained with solution (3) (0.1%) and the area of any other *secondary peak* is not greater than the area of the peak in the chromatogram obtained with solution (4) (0.1%). The sum of the areas of any such peaks is not greater than 1%.

Primary amines

Dissolve 50 mg of the substance being examined in 10 mL of *ethanol (96%)*, add 0.1 mL of *hydrochloric acid* and 2 mL of a 5% w/v solution of *4-dimethylaminobenzaldehyde* in *ethanol (96%)*. The yellow colour obtained is not more intense than that obtained by treating 10 mL of a 0.00005% w/v solution of *2-aminobenzoic acid* in *ethanol (96%)* in the same manner.

Loss on drying

When dried for 3 hours at 100° to 105° at a pressure not exceeding 0.7 kPa, loses not more than 0.5% of its weight. Use 1 g.

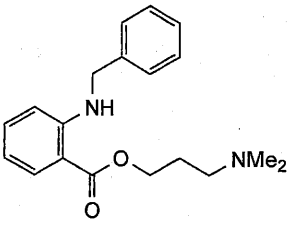
Sulfated ash

Not more than 0.1%, Appendix IX A. Use 1 g.

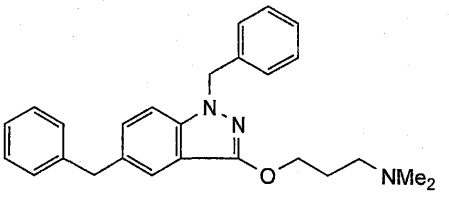
ASSAY

Dissolve 0.3 g in 100 mL of *anhydrous acetic acid* and carry out Method I for *non-aqueous titration*, Appendix VIII A, determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 34.59 mg of $C_{19}H_{23}N_3O$,HCl.

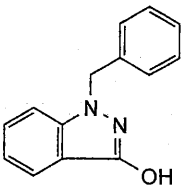
IMPURITIES



A. 3-dimethylaminopropyl 2-benzylaminobenzoate,



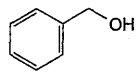
B. 3-(1,5-dibenzyl-1*H*-indazol-3-yl)oxypropyldimethylamine,



C. 1-benzyl-1*H*-indazol-3-ol.

Benzyl Alcohol[1]

(Ph. Eur. monograph 0256)



C_7H_8O 108.1 *100-51-6*

Action and use
Local anaesthetic; disinfectant.

Ph Eur

DEFINITION
Phenylmethanol.

Content
98.0 per cent to 100.5 per cent.

♦CHARACTERS
Appearance
Clear, colourless, oily liquid.

Solubility
Soluble in water, miscible with ethanol (96 per cent) and with fatty and essential oils.

Relative density
1.043 to 1.049.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzyl alcohol CRS*.♦

TESTS
♦Appearance of solution
Shake 2.0 mL with 60 mL of *water R*. It dissolves completely. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).♦

Acidity
To 10 mL add 10 mL of *ethanol (96 per cent) R* and 1 mL of *phenolphthalein solution R*. Not more than 1 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Refractive index (*2.2.6*)
1.538 to 1.541.

Peroxide value (*2.5.5, Method A*)
Maximum 5.

Related substances
Gas chromatography (*2.2.28*).

Test solution The substance to be examined.

Standard solution (a) Dissolve 0.100 g of *ethylbenzene R* in the test solution and dilute to 10.0 mL with the same solution. Dilute 2.0 mL of this solution to 20.0 mL with the test solution.

Standard solution (b) Dissolve 2.000 g of *dicyclohexyl R* in the test solution and dilute to 10.0 mL with the same solution. Dilute 2.0 mL of this solution to 20.0 mL with the test solution.

Reference solution (a) Dissolve 0.750 g of *benzaldehyde R* and 0.500 g of *cyclohexylmethanol R* in the test solution and dilute to 25.0 mL with the test solution. Add 1.0 mL of this solution to a mixture of 2.0 mL of standard solution (a) and 3.0 mL of standard solution (b) and dilute to 20.0 mL with the test solution.

Reference solution (b) Dissolve 0.250 g of *benzaldehyde R* and 0.500 g of *cyclohexylmethanol R* in the test solution and dilute to 25.0 mL with the test solution. Add 1.0 mL of this solution to a mixture of 2.0 mL of standard solution (a) and 2.0 mL of standard solution (b) and dilute to 20.0 mL with the test solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R*.

Linear velocity 25 cm/s at 50 °C.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 34	50 → 220
	34 - 69	220
Injection port		200
Detector		310

Detection Flame ionisation.

Benzyl alcohol not intended for parenteral administration

Injection Without air-plug, 0.1 µL of the test solution and reference solution (a).

Relative retention With reference to benzyl alcohol (retention time = about 26 min): ethylbenzene = about 0.28; dicyclohexyl = about 0.59; impurity A = about 0.68; impurity B = about 0.71.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurities A and B.

If any peaks in the chromatogram obtained with the test solution have the same retention time as the peaks due to ethyl benzene or dicyclohexyl, subtract the areas of any such peaks from the peak areas at these retention times in the chromatograms obtained with reference solutions (a) or (b) (corrected peak areas of ethyl benzene and dicyclohexyl). Any such peaks in the chromatogram obtained with the test solution are to be included in the assessments for the sum of other peaks.

Limits:
— *impurity A*: not more than the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (a) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.15 per cent);
— *impurity B*: not more than the difference between the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) and the area of the peak due to impurity B in the chromatogram obtained with the test solution (0.10 per cent);
— *sum of other peaks with a relative retention less than that of benzyl alcohol*: not more than 4 times the area of the peak due to ethylbenzene in the chromatogram obtained with reference solution (a) corrected if necessary as described above (0.04 per cent);
— *sum of peaks with a relative retention greater than that of benzyl alcohol*: not more than the area of the peak due

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8. Pharmacopoeial harmonisation.*

to dicyclohexyl in the chromatogram obtained with reference solution (a) corrected if necessary as described above (0.3 per cent);
— *disregard limit*: 0.01 times the area of the peak due to ethylbenzene in the chromatogram obtained with reference solution (a) corrected if necessary as described above (0.0001 per cent).

Benzyl alcohol intended for parenteral administration

Injection Without air-plug, 0.1 µL of the test solution and reference solution (b).

Relative retention With reference to benzyl alcohol (retention time = about 26 min): ethylbenzene = about 0.28; dicyclohexyl = about 0.59; impurity A = about 0.68; impurity B = about 0.71.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to impurities A and B.

If any peaks in the chromatogram obtained with the test solution have the same retention times as the peaks due to ethyl benzene or dicyclohexyl, subtract the areas of any such peaks from the peak areas at these retention times in the chromatograms obtained with reference solutions (a) or (b) (corrected peak areas of ethyl benzene and dicyclohexyl). Any such peaks in the chromatogram obtained with the test solution are to be included in the assessments for the sum of other peaks.

Limits:
— *impurity A*: not more than the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (b) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.05 per cent);
— *impurity B*: not more than the difference between the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) and the area of the peak due to impurity B in the chromatogram obtained with the test solution (0.10 per cent);
— *sum of other peaks with a relative retention less than that of benzyl alcohol*: not more than twice the area of the peak due to ethylbenzene in the chromatogram obtained with reference solution (b) corrected if necessary as described above (0.02 per cent);
— *sum of peaks with a relative retention greater than that of benzyl alcohol*: not more than the area of the peak due to dicyclohexyl in the chromatogram obtained with reference solution (b) corrected if necessary as described above (0.2 per cent);
— *disregard limit*: 0.01 times the area of the peak due to ethylbenzene in the chromatogram obtained with reference solution (b) corrected if necessary as described above (0.0001 per cent).

Residue on evaporation
Maximum 0.05 per cent.

After ensuring that the substance to be examined complies with the test for peroxide value, evaporate 10.0 g to dryness in a tared quartz or porcelain crucible or platinum dish on a hot plate at a temperature not exceeding 200 °C. Ensure that the substance to be examined does not boil during evaporation. Dry the residue on the hot plate for 1 h and allow to cool in a desiccator. The residue weighs a maximum of 5 mg.

ASSAY
To 0.900 g (*m* g) add 15.0 mL of a freshly prepared mixture of 1 volume of *acetic anhydride R* and 7 volumes of *anhydrous pyridine R* and heat under a reflux condenser on a boiling water-bath for 30 min. Cool and add 25 mL of *water R*. Using 0.25 mL of *phenolphthalein solution R* as indicator, titrate with *1 M sodium hydroxide* (n_1 mL). Carry out a blank titration (n_2 mL).

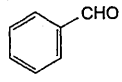
Calculate the percentage content of C_7H_8O using the following expression:

$$\frac{10.81(n_2 - n_1)}{m}$$

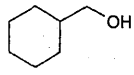
♦STORAGE
In an airtight container, under nitrogen, protected from light and at a temperature between 2 °C and 8 °C.

LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.♦

IMPURITIES
Specified impurities *A, B.*



A. benzaldehyde,

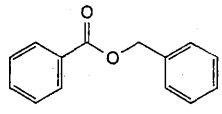


B. cyclohexylmethanol.

Ph Eur

Benzyl Benzoate

(Ph. Eur. monograph 0705)



$C_{14}H_{12}O_2$ 212.2 *120-51-4*

Action and use
Used topically in the treatment of scabies.

Preparation
Benzyl Benzoate Application

Ph Eur

DEFINITION
Phenylmethyl benzoate.

Content
99.0 per cent to 100.5 per cent.

CHARACTERS
Appearance
Colourless or almost colourless crystals or colourless or almost colourless, oily liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent), with methylene chloride and with fatty and essential oils.

Eb: about 320 °C.

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison Ph. Eur. reference spectrum of benzyl benzoate.

B. To 2 g add 25 mL of *alcoholic potassium hydroxide solution R* and boil under a reflux condenser for 2 h. Remove the ethanol on a water-bath, add 50 mL of *water R* and distill. Collect about 25 mL of distillate and use it for identification test C. Acidify the liquid remaining in the distillation flask with *dilute hydrochloric acid R*. A white precipitate is formed that, when washed with *water R* and dried *in vacuo* melts (*2.2.14*) at 121 °C to 124 °C.

C. To the distillate obtained in identification test B add 2.5 g of *potassium permanganate R* and 5 mL of *dilute sodium hydroxide solution R*. Boil under a reflux condenser for 15 min, cool and filter. Acidify the filtrate with *dilute hydrochloric acid R*. A white precipitate is formed that, when washed with *water R* and dried *in vacuo*, melts (*2.2.14*) at 121 °C to 124 °C.

TESTS
Acidity
Dissolve 2.0 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent. Titrate with *0.1 M sodium hydroxide* using *phenolphthalein solution R* as indicator. Not more than 0.2 mL is required to change the colour of the indicator to pink.

Relative density (*2.2.5*)
1.118 to 1.122.

Refractive index (*2.2.6*)
1.568 to 1.570.

Freezing point (*2.2.18*)
Minimum 17.0 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
To 2.000 g add 50.0 mL of 0.5 M alcoholic potassium hydroxide and boil gently under a reflux condenser for 1 h. Titrate the hot solution with *0.5 M hydrochloric acid* using 1 mL of *phenolphthalein solution R* as indicator. Carry out a blank determination.

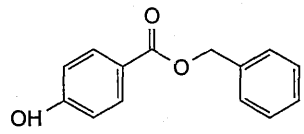
1 mL of 0.5 M alcoholic potassium hydroxide is equivalent to 106.1 mg of $C_{14}H_{12}O_2$.

STORAGE
In an airtight, well-filled container, protected from light.

Ph Eur

Benzyl Hydroxybenzoate

Benzylparaben



$C_{14}H_{12}O_3$ 228.3 *94-18-8*

Action and use
Antimicrobial preservative.

DEFINITION
Benzyl Hydroxybenzoate is benzyl 4-hydroxybenzoate. It contains not less than 99.0% and not more than 101.0% of $C_{14}H_{12}O_3$.

CHARACTERISTICS
A white to creamy white, crystalline powder.

Practically insoluble in *water*; freely soluble in *ethanol (96%)* and in *ether*. It dissolves in solutions of alkali hydroxides.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of benzyl hydroxybenzoate *(RS 028)*.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.001% w/v solution in *ethanol (96%)* exhibits a maximum only at 260 nm. The *absorbance* at the maximum at 260 nm is about 0.76.

C. Dissolve 0.1 g in 2 mL of *ethanol (96%)*, boil and add 0.5 mL of *nitric acid solution of mercury*. A precipitate is produced slowly and the supernatant liquid becomes red.

D. *Melting point*, about 112°, Appendix V A.

TESTS
Acidity
Dissolve 0.2 g in 10 mL of *ethanol (50%)* previously neutralised to *methyl red solution* and titrate with *0.1M sodium hydroxide VS* using *methyl red solution* as indicator. Not more than 0.1 mL of *0.1M sodium hydroxide VS* is required to change the colour of the solution.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using a plate precoated with silica gel F_{254}, the surface of which has been modified with chemically-bonded octadecylsilyl groups (Whatman KC18F plates are suitable) and a mixture of 70 volumes of *methanol*, 30 volumes of *water* and 1 volume of *glacial acetic acid* as the mobile phase. Apply separately to the plate 2 µL of each of two solutions of the substance being examined in *acetone* containing (1) 1.0% w/v and (2) 0.010% w/v. After removal of the plate, allow it to dry in air and examine under *ultraviolet light (254 nm)*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

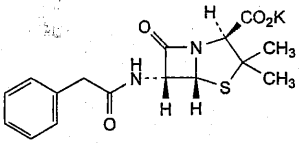
Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Gently boil 0.12 g under a reflux condenser with 20 mL of 2M *sodium hydroxide* for 30 minutes. Cool and extract with three 20-mL quantities of *1,2-dichloroethane*. Wash the combined extracts with 20 mL of 0.1M *sodium hydroxide* and

add the washings to the main aqueous phase, discarding the organic layer. To the aqueous solution add 25 mL of *0.0333M potassium bromate VS*, 5 mL of a 12.5% w/v solution of *potassium bromide* and 10 mL of *hydrochloric acid* and immediately stopper the flask. Shake for 15 minutes and allow to stand for 15 minutes. Add 25 mL of *dilute potassium iodide solution* and shake vigorously. Titrate the liberated iodine with 0.1M *sodium thiosulfate VS* using *starch mucilage*, added towards the end of the titration, as indicator. Repeat the operation without the substance being examined.
The difference between the titrations represents the amount of potassium bromate required. The volume of *0.0333M potassium bromate VS* used is equivalent to half of the volume of 0.1M *sodium thiosulfate VS* required for the titration.
Each mL of *0.0333M potassium bromate VS* is equivalent to 7.608 mg of $C_{14}H_{12}O_3$.

Benzylpenicillin Potassium

(Ph. Eur. monograph 0113)



$C_{16}H_{17}KN_2O_4S$ 372.5 *113-98-4*

Action and use
Penicillin antibacterial.

Preparation
Benzylpenicillin for Injection

Ph Eur

DEFINITION

Potassium (2*S*,5*R*,6*R*)-3,3-dimethyl-7-oxo-6-[(phenylacetyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Substance produced by the growth of certain strains of *Penicillium notatum* or related organisms.

Content
95.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, slightly hygroscopic, crystalline powder.

Solubility
Very soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzylpenicillin potassium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *benzylpenicillin potassium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *benzylpenicillin potassium CRS* and 25 mg of *phenoxymethylpenicillin potassium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube on a water-bath for 1 min; a reddish-brown colour develops.

D. It gives reaction (a) of potassium (*2.3.1*).

TESTS

pH (*2.2.3*)
5.5 to 7.5.

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*).

Dissolve 3.0 g in *water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Test solution (b) Dissolve 80.0 mg of the substance to be examined in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *benzylpenicillin sodium CRS* in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *benzylpenicillin for system suitability CRS* (containing impurities A, B, C, D, E, F, G and H) in 0.35 mL of *methanol R1* and add 0.65 mL of *water R.*

Reference solution (c) Dilute 1.0 mL of test solution (b) to 100.0 mL with *water R.*

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 20.0 mL with *water R.*

Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature:* 50 °C.

Mobile phase:
— *mobile phase A*: mix 10 volumes of a 68 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.4 with a 500 g/L solution of *phosphoric acid R*, 30 volumes of *methanol R1* and 60 volumes of *water for chromatography R*;
— *mobile phase B*: mix 10 volumes of a 68 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.4 with a 500 g/L solution of *phosphoric acid R*, 35 volumes of *water for chromatography R* and 55 volumes of *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	70	30
7 - 17	70 → 0	30 → 100
17 - 22	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (b) and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *benzylpenicillin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G and H.

Relative retention With reference to benzylpenicillin (retention time = about 7 min): impurity A = about 0.22; impurity D = about 0.33; impurity C = about 0.35; impurity E = about 0.48 and 0.55; impurity B = about 0.62; impurity F = about 0.81 and 0.83; impurity G = about 1.47; impurity H = about 1.90.

System suitability:
— *resolution*: minimum 1.2 between the peaks due to the epimers of impurity F and minimum 1.5 between the peaks due to impurities D and C in the chromatogram obtained with reference solution (b);
— *signal-to-noise ratio*: minimum 20 for the principal peak in the chromatogram obtained with reference solution (d).

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.4; impurity D = 0.6; impurity E = 2.0; impurity F = 1.7;
— for each impurity, use the concentration of benzylpenicillin potassium in reference solution (c).

Limits:
— *impurity F*: maximum 2.0 per cent for the sum of the 2 epimers;
— *impurity E*: maximum 1.0 per cent for the sum of the isomers;
— *impurity B*: maximum 0.5 per cent;
— *impurities A, C, D, G, H*: for each impurity, maximum 0.2 per cent;
— *any other impurity*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 3.0 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A, mobile phase B (70:30 *V/V*).

Flow rate 1.2 mL/min.

Injection 10 µL of test solution (a) and reference solution (a).

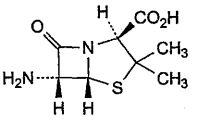
Calculate the percentage content of $C_{16}H_{17}KN_2O_4S$ taking into account the assigned content of *benzylpenicillin sodium CRS* and a conversion factor of 1.045.

STORAGE

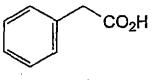
In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

IMPURITIES

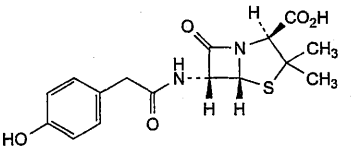
Specified impurities *A, B, C, D, E, F, G, H.*



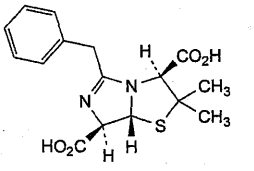
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



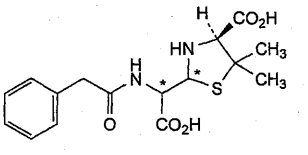
B. phenylacetic acid,



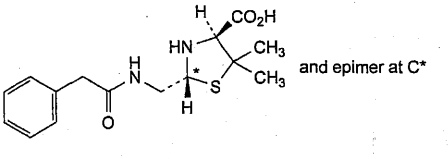
C. (2*S*,5*R*,6*R*)-6-[[(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,



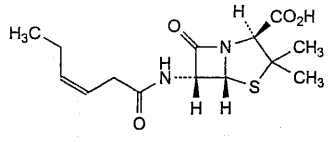
D. (3*S*,7*R*,7a*R*)-5-benzyl-2,2-dimethyl-2,3,7,7a-tetrahydroimidazo[5,1-*b*]thiazole-3,7-dicarboxylic acid (penillic acid of benzylpenicillin),



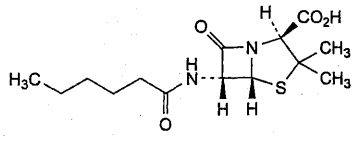
E. (4*S*)-2-[carboxy[(phenylacetyl)amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of benzylpenicillin),



F. (2*RS*,4*S*)-5,5-dimethyl-2-[[(phenylacetyl)amino]methyl]thiazolidine-4-carboxylic acid (penilloic acids of benzylpenicillin),



G. (2*S*,5*R*,6*R*)-6-[(3*Z*)-hex-3-enoylamino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,

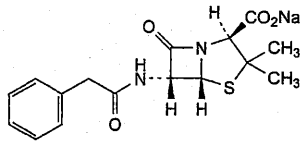


H. (2*S*,5*R*,6*R*)-6-(hexanoylamino)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (dihydropenicillin F).

Ph Eur

Benzylpenicillin Sodium

(Ph. Eur. monograph 0114)



$C_{16}H_{17}N_2NaO_4S$ 356.4 *69-57-8*

Action and use
Penicillin antibacterial.

Preparation
Benzylpenicillin for Injection

Ph Eur

DEFINITION
Sodium (2*S*,5*R*,6*R*)-3,3-dimethyl-7-oxo-6-[(phenylacetyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Substance produced by the growth of certain strains of *Penicillium notatum* or related organisms.

Content
95.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Very soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *benzylpenicillin sodium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *benzylpenicillin sodium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *benzylpenicillin sodium CRS* and 25 mg of *phenoxymethylpenicillin potassium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube on a water-bath for 1 min; a reddish-brown colour develops.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS
pH (*2.2.3*)
5.5 to 7.5.

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*).

Dissolve 3.0 g in *water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Test solution (b) Dissolve 80.0 mg of the substance to be examined in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *benzylpenicillin sodium CRS* in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *benzylpenicillin for system suitability CRS* (containing impurities A, B, C, D, E, F, G and H) in 0.35 mL of *methanol R1* and add 0.65 mL of *water R.*

Reference solution (c) Dilute 1.0 mL of test solution (b) to 100.0 mL with *water R.*

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 20.0 mL with *water R.*

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: mix 10 volumes of a 68 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.4 with a 500 g/L solution of *phosphoric acid R*, 30 volumes of *methanol R1* and 60 volumes of *water for chromatography R*;
— *mobile phase B*: mix 10 volumes of a 68 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.4 with a 500 g/L solution of *phosphoric acid R*, 35 volumes of *water for chromatography R* and 55 volumes of *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	70	30
7 - 17	70 → 0	30 → 100
17 - 22	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (b) and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *benzylpenicillin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G and H.

Relative retention With reference to benzylpenicillin (retention time = about 7 min): impurity A = about 0.22; impurity D = about 0.33; impurity C = about 0.35; impurity E = about 0.48 and 0.55; impurity B = about 0.62; impurity F = about 0.81 and 0.83; impurity G = about 1.47; impurity H = about 1.90.

System suitability:
— *resolution*: minimum 1.2 between the peaks due to the epimers of impurity F and minimum 1.5 between the peaks due to impurities D and C in the chromatogram obtained with reference solution (b);
— *signal-to-noise ratio*: minimum 20 for the principal peak in the chromatogram obtained with reference solution (d).

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.3; impurity D = 0.6; impurity E = 2.0; impurity F = 1.7;
— for each impurity, use the concentration of benzylpenicillin sodium in reference solution (c).

Limits:
— *impurity E*: maximum 2.0 per cent for the sum of the isomers;
— *impurity F*: maximum 1.0 per cent for the sum of the 2 epimers;
— *impurity B*: maximum 0.5 per cent;
— *impurities A, C, D, G, H*: for each impurity, maximum 0.2 per cent;
— *any other impurity*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 3.0 per cent;
— *reporting threshold*: 0.05 per cent.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.5 per cent *m/m*.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A, mobile phase B (70:30 *V/V*).

Flow rate 1.2 mL/min.

Injection 10 µL of test solution (a) and reference solution (a).

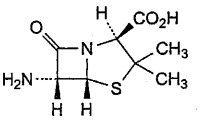
Calculate the percentage content of $C_{16}H_{17}N_2NaO_4S$ taking into account the assigned content of *benzylpenicillin sodium CRS*.

STORAGE

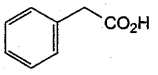
In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

IMPURITIES

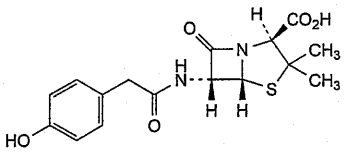
Specified impurities *A, B, C, D, E, F, G, H.*



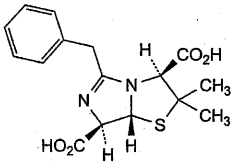
A. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



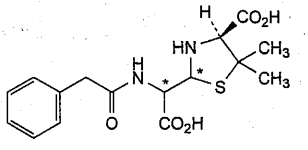
B. phenylacetic acid,



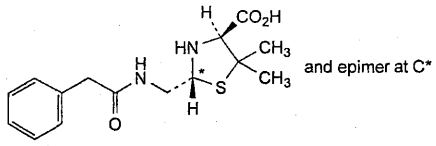
C. (2*S*,5*R*,6*R*)-6-[[(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid,



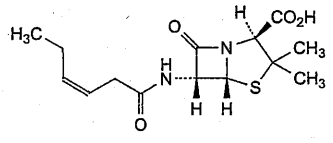
D. (3*S*,7*R*,7a*R*)-5-benzyl-2,2-dimethyl-2,3,7,7a-tetrahydroimidazo[5,1-*b*]thiazole-3,7-dicarboxylic acid (penillic acid of benzylpenicillin),



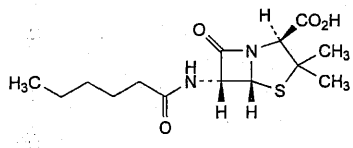
E. (4*S*)-2-[carboxy[(phenylacetyl)amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of benzylpenicillin),



F. (2*RS*,4*S*)-5,5-dimethyl-2-[[(phenylacetyl)amino] methyl]thiazolidine-4-carboxylic acid (penilloic acids of benzylpenicillin),



G. (2*S*,5*R*,6*R*)-6-[(3*Z*)-hex-3-enoylamino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (iso-penicillin F),

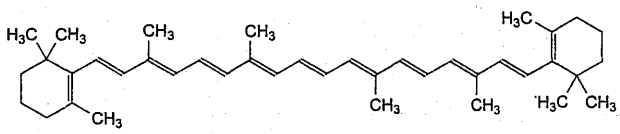


H. (2*S*,5*R*,6*R*)-6-(hexanoylamino)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (dihydropenicillin F).

Ph Eur

Betacarotene

(Ph. Eur. monograph 1069)



$C_{40}H_{56}$ 536.9 *7235-40-7*

Action and use
Precursor of vitamin A.

Ph Eur

DEFINITION

Main component 1,1′-[(1*E*,3*E*,5*E*,7*E*,9*E*,11*E*,13*E*,15*E*,17*E*)-3,7,12,16-tetramethyloctadeca-1,3,5,7,9,11,13,15,17-nonaene-1,18-diyl]bis(2,6,6-trimethylcyclohexene) (β,β-carotene, betacarotene, *all-trans*-betacarotene, *all-E*-betacarotene).

Content
95.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
Brown-red or brownish-red, crystalline powder.

Solubility
Practically insoluble in water, soluble in tetrahydrofuran, practically insoluble in anhydrous ethanol.

It is sensitive to air, heat and light, especially in solution.

IDENTIFICATION

Carry out all operations as rapidly as possible. Prepare the solutions immediately before use.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *tetrahydrofuran R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 50.0 mL with *tetrahydrofuran R*. Dilute 3.0 mL of this solution to 100.0 mL with *cyclohexane R*.

Absorption maximum 455 nm.

Shoulder About 427 nm.

Absorbance ratio A_{455}/A_{483} = 1.14 to 1.18.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Carry out all operations as rapidly as possible. Prepare the solutions immediately before use.

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Stabilised tetrahydrofuran *tetrahydrofuran R* containing 0.25 g/L of *butylhydroxytoluene R*.

Test solution Dissolve 50.0 mg of the substance to be examined in stabilised tetrahydrofuran and dilute to 200.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with *anhydrous ethanol R*.

Reference solution (a) Dissolve 50.0 mg of *betacarotene CRS* in stabilised tetrahydrofuran and dilute to 200.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with *anhydrous ethanol R*.

Reference solution (b) Dissolve 5 mg of *betacarotene for system suitability CRS* (containing impurities C, D, E, G and H) in stabilised tetrahydrofuran and dilute to 20 mL with the same solvent. Dilute 1 mL of the solution to 20 mL with *anhydrous ethanol R*.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with stabilised tetrahydrofuran. Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *amidoalkylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 50 mg of *butylhydroxytoluene R* in 20 mL of *2-propanol R*, then add 0.2 mL of *N,N-diisopropylethylamine R*, 25 mL of a 2.0 g/L solution of *ammonium acetate R*, 455 mL of *acetonitrile R* and 450 mL of *methanol R*; warm to room temperature and dilute to 1000 mL with *methanol R*.

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 448 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 2.7 times the retention time of betacarotene.

Identification of impurities Use the chromatogram supplied with *betacarotene for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C, D, E, G and H.

Relative retention With reference to betacarotene (retention time = about 28 min): impurity G = about 0.6; impurity C = about 0.95; impurity D = about 1.05; impurity E = about 1.15; impurity H = about 2.4.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurity C and betacarotene;
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to betacarotene.

Limits:
- *impurity E*: maximum 4.0 per cent;
- *impurity D*: maximum 2.0 per cent;
- *impurity C*: maximum 0.6 per cent;
- *impurity G*: maximum 0.4 per cent;
- *impurity H*: maximum 0.4 per cent;
- *unspecified impurities*: for each impurity, maximum 0.15 per cent;
- *sum of impurities other than D and E*: maximum 5.0 per cent;
- *reporting threshold*: 0.05 per cent (0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c)).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

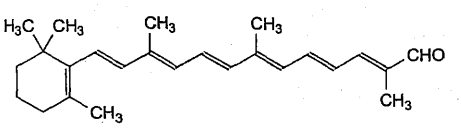
Loss on drying (*2.2.32*)
Maximum 0.2 per cent, determined on 1.000 g by drying *in vacuo* at 40 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 2.0 g.

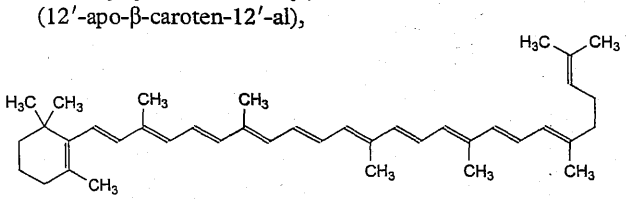
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification:
Injection Test solution and reference solution (a).
Calculate the percentage content of $C_{40}H_{56}$ taking into account the assigned content of *betacarotene CRS*.

STORAGE
In an airtight container, protected from light.

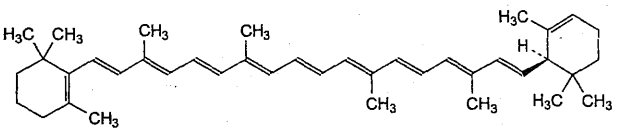
IMPURITIES
Specified impurities *C, D, E, G, H.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, F.*



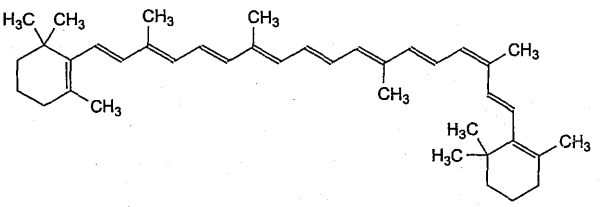
A. (2*E*,4*E*,6*E*,8*E*,10*E*,12*E*)-2,7,11-trimethyl-13-(2,6,6-trimethylcyclohex-1-en-1-yl)trideca-2,4,6,8,10,12-hexaenal (12′-apo-β-caroten-12′-al),



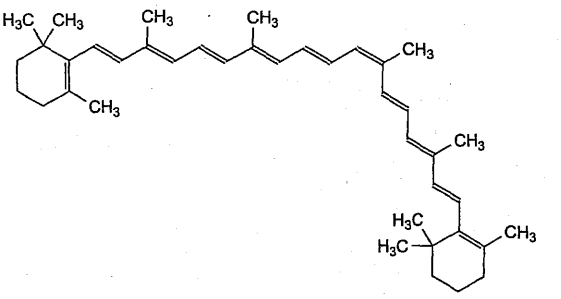
B. 1-[(1*E*,3*E*,5*E*,7*E*,9*E*,11*E*,13*E*,15*E*,17*E*,19*E*)-3,7,12,16,20,24-hexamethylpentacosa-1,3,5,7,9,11,13,15,17,19,23-undecaen-1-yl]-2,6,6-trimethylcyclohexene (β,Ψ-carotene, γ-carotene),



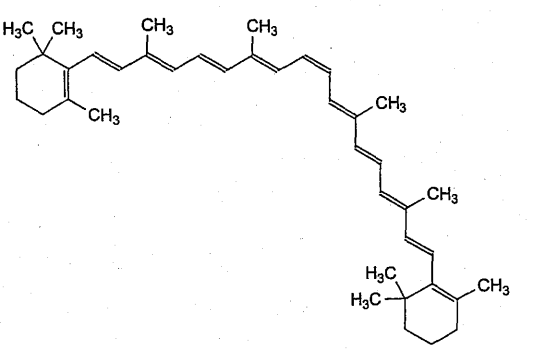
C. 1,3,3-trimethyl-2-[(1*E*,3*E*,5*E*,7*E*,9*E*,11*E*,13*E*,15*E*,17*E*)-3,7,12,16-tetramethyl-18-[(1*R*)-2,6,6-trimethylcyclohex-2-en-1-yl]octadeca-1,3,5,7,9,11,13,15,17-nonaen-1-yl] cyclohexene ((6′*R*)-β,ε-carotene, α-carotene),



D. 1,1′-[(1*E*,3*E*,5*E*,7*E*,9*E*,11*E*,13*E*,15*Z*,17*E*)-3,7,12,16-tetramethyloctadeca-1,3,5,7,9,11,13,15,17-nonaene-1,18-diyl]bis(2,6,6-trimethylcyclohexene) (9-*cis*-β,β-carotene, 9-*cis*-betacarotene),



E. 1,1′-[(1*E*,3*E*,5*E*,7*E*,9*E*,11*Z*,13*E*,15*E*,17*E*)-3,7,12,16-tetramethyloctadeca-1,3,5,7,9,11,13,15,17-nonaene-1,18-diyl]bis(2,6,6-trimethylcyclohexene) (13-*cis*-β,β-carotene, 13-*cis*-betacarotene),



F. 1,1′-[(1*E*,3*E*,5*E*,7*E*,9*Z*,11*E*,13*E*,15*E*,17*E*)-3,7,12,16-tetramethyloctadeca-1,3,5,7,9,11,13,15,17-nonaene-1,18-diyl]bis(2,6,6-trimethylcyclohexene) (15-cis-β,β-carotene, 15-*cis*-betacarotene),

G. unknown structure (oxidation products),

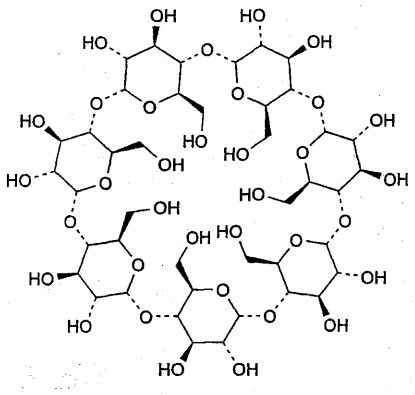
H. unknown structure.

Ph Eur

Betadex

Betacyclodextrin

(Ph. Eur. monograph 1070)



$[C_6H_{10}O_5]_7$ 1135 *7585-39-9*

Action and use
Carrier molecule for drug delivery systems.

Ph Eur

DEFINITION
Cycloheptakis-(1→4)-(α-D-glucopyranosyl) (cyclomaltoheptaose or β-cyclodextrin).

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, amorphous or crystalline, hygroscopic powder.

Solubility
Sparingly soluble in water and in propylene glycol, practically insoluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

C. Dissolve 0.2 g in 2 mL of *iodine solution R4* by warming on a water-bath, and allow to stand at room temperature. A yellowish-brown precipitate is formed.

TESTS
Solution S
Dissolve 1.000 g in *carbon dioxide-free water R* with heating, allow to cool and dilute to 100.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

pH (*2.2.3*)
5.0 to 8.0.

To 10 mL of solution S add 0.1 mL of a saturated solution of *potassium chloride R*.

Specific optical rotation (*2.2.7*)
+ 160 to + 164 (dried substance), determined on solution S.

Reducing sugars
Maximum 0.2 per cent.

Test solution To 1 mL of solution S add 1 mL of *cupri-tartaric solution R4*. Heat on a water-bath for 10 min, cool to room temperature. Add 10 mL of *ammonium molybdate reagent R1* and allow to stand for 15 min.

Reference solution Prepare a reference solution at the same time and in the same manner as the test solution, using 1 mL of a 0.02 g/L solution of *glucose R*.

Measure the absorbance (*2.2.25*) of the test solution and the reference solution at the absorption maximum at 740 nm using *water R* as the compensation liquid. The absorbance of the test solution is not greater than that of the reference solution.

Light-absorbing impurities
Examine solution S between 230 nm and 750 nm. Between 230 nm and 350 nm, the absorbance (*2.2.25*) is not greater than 0.10. Between 350 nm and 750 nm, the absorbance (*2.2.25*) is not greater than 0.05.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.250 g of the substance to be examined in *water R* with heating, cool and dilute to 25.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with *water R*.

Reference solution (a) Dissolve 25.0 mg of *alfadex CRS* (impurity A), 25.0 mg of *gammacyclodextrin CRS* (impurity B) and 50.0 mg of *betadex CRS* in *water R*, then dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with *water R*.

Reference solution (c) Dissolve 25.0 mg of *betadex CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase *methanol R, water for chromatography R* (10:90 *V/V*).

Flow rate 1.5 mL/min.

Detection Differential refractometer.

Equilibration With the mobile phase for about 3 h.

Injection 50 µL of test solution (a) and reference solutions (a) and (b).

Run time 1.5 times the retention time of betadex.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B.

Relative retention With reference to betadex (retention time = about 10 min): impurity B = about 0.3; impurity A = about 0.45.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurities B and A; if necessary, adjust the concentration of methanol in the mobile phase.

Calculation of percentage contents:
— for impurities A and B, use the concentration of the corresponding impurity in reference solution (b);
— for impurities other than A and B, use the concentration of betadex in reference solution (b).

Limits:
— *impurities A, B*: for each impurity, maximum 0.25 per cent;

— *sum of impurities other than A and B*: maximum 0.5 per cent;
— *reporting threshold*: 0.15 per cent.

Residual solvents
Head-space gas chromatography (*2.2.28*): use the standard additions method.

Internal standard *ethylene chloride R.*

Stock solution A To 20 µL of *ethylene chloride R* add 0.5 mL of *dimethyl sulfoxide R* and dilute to 25.0 mL with *water R.*

Stock solution B To 25 µL of *trichloroethylene R* add 25 µL of *toluene R* and 0.5 mL of *dimethyl sulfoxide R*, then dilute to 50.0 mL with *water R.*

Test solutions (a), (b), (c) and (d) In each of 4 identical vials, introduce 0.5 g of the substance to be examined, 0.10 g of *calcium chloride R*, 30 µL of *α-amylase solution R* and 1 mL of reference solutions (a), (b), (c) and (d), respectively, then add 9.0 mL of *water R*. Prepare test solutions (b), (c) and (d) in triplicate.

Reference solution (a) Dilute 250 µL of stock solution A to 10.0 mL with *water R.*

Reference solution (b) To 100 µL of stock solution B add 250 µL of stock solution A and dilute to 10.0 mL with *water R.*

Reference solution (c) To 200 µL of stock solution B add 250 µL of stock solution A and dilute to 10.0 mL with *water R.*

Reference solution (d) To 300 µL of stock solution B add 250 µL of stock solution A and dilute to 10.0 mL with *water R.*

Blank solution In a vial identical to those used for the test solutions, introduce 0.10 g of *calcium chloride R*, 30 µL of *α-amylase solution R*, 0.5 mL of *dimethyl sulfoxide R* and 10.0 mL of *water R.*

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.32 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1.7 mL/min.

Static head-space conditions that may be used: if the equipment has different setting parameters, adjust the equipment settings so as to comply with the system suitability criterion:
— *equilibration temperature*: 45 °C;
— *equilibration time*: 2 h;
— *syringe temperature*: 50 °C;
— *injection speed*: 500 µL/s.

Temperature:
— *column*: 50 °C;
— *injection port*: 140 °C;
— *detector*: 280 °C.

Detection Flame ionisation.

Injection 200 µL.

Relative retention With reference to the internal standard (retention time = about 13 min): trichloroethylene = about 0.6; toluene = about 0.8.

System suitability Test solutions (b), (c) and (d):
— *repeatability*: maximum relative standard deviations of the ratios of the areas of the peaks due to trichloroethylene and toluene to that of the peak due to ethylene chloride of 10.0 per cent, for each set of triplicate test solutions and each residual solvent.

Calculate the content of trichloroethylene and of toluene taking their relative densities to be 1.46 and 0.87, respectively.

Limits:
— *trichloroethylene*: maximum 10 ppm;
— *toluene*: maximum 10 ppm.

Loss on drying (*2.2.32*)
Maximum 16.0 per cent, determined on 1.000 g by drying in an oven at 120 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

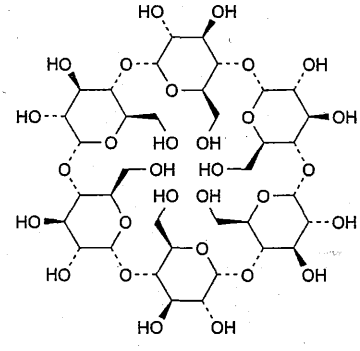
Injection Test solution (b) and reference solutions (a) and (c).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the area of the peak due to betadex, determined on 5 injections.

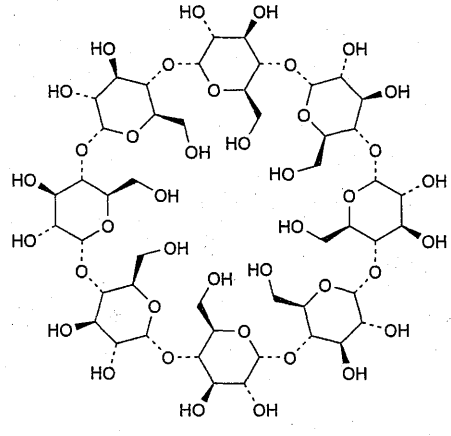
Calculate the percentage content of $[C_6H_{10}O_5]_7$ using the chromatogram obtained with reference solution (c) and taking into account the assigned content of *betadex CRS.*

STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B.*



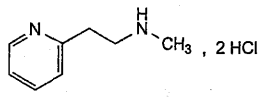
A. cyclohexakis-(1→4)-(α-D-glucopyranosyl) (alfadex or cyclomaltohexaose or α-cyclodextrin),



B. cyclooctakis-(1→4)-(α-D-glucopyranosyl) (cyclomaltooctaose or γ-cyclodextrin).

Betahistine Dihydrochloride

(Ph. Eur. monograph 1665)



$C_8H_{14}Cl_2N_2$ 209.1 *5579-84-0*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparation
Betahistine Dihydrochloride Tablets

Ph Eur

DEFINITION

N-Methyl-2-(pyridin-2-yl)ethanamine dihydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or slightly yellow powder, very hygroscopic.

Solubility
Very soluble in water, soluble in ethanol (96 per cent), practically insoluble in 2-propanol.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 150 °C to 154 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *betahistine dihydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 2 mL of *ethanol (96 per cent) R.*

Reference solution Dissolve 10 mg of *betahistine dihydrochloride CRS* in 2 mL of *ethanol (96 per cent) R.*

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *concentrated ammonia R, ethyl acetate R, methanol R* (0.75:15:30 *V/V/V*).

Application 2 µL.

Development Over 2/3 of the plate.

Drying At 110 °C for 10 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *carbon dioxide-free water R*, and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_8 (*2.2.2, Method II*).

pH (*2.2.3*)
2.0 to 3.0 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 10 mg of *betahistine dihydrochloride CRS* and 10 mg of *2-vinylpyridine R* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 2.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 2.0 mL of reference solution (b) to 10.0 mL with the mobile phase.

Column:
— *size: l* = 0.15 m, Ø = 3.0 mm;
— *stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 2.0 g of *sodium dodecyl sulfate R* in a mixture of 15 mL of a 10 per cent *V/V* solution of *sulfuric acid R*, 35 mL of a 17 g/L solution of *tetrabutylammonium hydrogen sulfate R* and 650 mL of *water R*; adjust to pH 3.3 using *dilute sodium hydroxide solution R* and mix with 300 mL of *acetonitrile R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 µL.

Run time 4 times the retention time of betahistine.

Relative retention With reference to betahistine (retention time = about 7 min): impurity B = about 0.2; impurity A = about 0.3; impurity C = about 3.

System suitability Reference solution (a):
— *resolution*: minimum 3.5 between the peaks due to 2-vinylpyridine and betahistine.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 0.4;
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times of the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 80.0 mg in 50 mL of *ethanol (96 per cent) R.* Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added to reach the second point of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 10.46 mg of $C_8H_{14}Cl_2N_2$.

STORAGE

In an airtight container.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
- the chromatogram shows 2 spots which may, however, not be completely separated.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. Add 1.0 mL of the filtrate to a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

E. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a deep reddish-brown colour develops. Add this solution to 10 mL of *water R* and mix. The colour is discharged and a clear solution remains.

TESTS

Specific optical rotation (*2.2.7*)

+ 118 to + 126 (dried substance).

Dissolve 0.125 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R* and *methanol R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 2 mg of *betamethasone CRS* and 2 mg of *methylprednisolone CRS* in mobile phase A, then dilute to 100.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 45 °C.

Mobile phase:
- *mobile phase A*: in a 1000 mL volumetric flask mix 250 mL of *acetonitrile R* with 700 mL of *water R* and allow to equilibrate; dilute to 1000 mL with *water R* and mix again;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100	0
15 - 40	100 → 0	0 → 100
40 - 41	0 → 100	100 → 0
41 - 46	100	0

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With mobile phase B for at least 30 min and then with mobile phase A for 5 min. For subsequent chromatograms, use the conditions described from 40 min to 46 min.

Injection 20 µL; inject the mixture of equal volumes of *acetonitrile R* and *methanol R* as a blank.

Retention time Methylprednisolone = about 11.5 min; betamethasone = about 12.5 min.

System suitability Reference solution (a):
- *resolution*: minimum 1.5 between the peaks due to methylprednisolone and betamethasone; if necessary, adjust the concentration of acetonitrile in mobile phase A.

Limits:
- *impurities A, B, C, D, E, F, G, H, I, J*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent), and not more than 1 such peak has an area greater than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 238.5 nm.

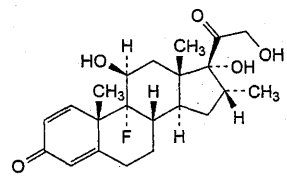
Calculate the content of $C_{22}H_{29}FO_5$ taking the specific absorbance to be 395.

STORAGE

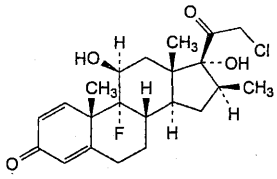
Protected from light.

IMPURITIES

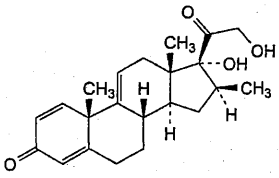
Specified impurities *A, B, C, D, E, F, G, H, I, J.*



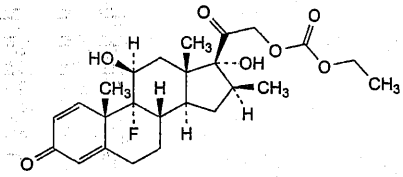
A. 9-fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione (dexamethasone),



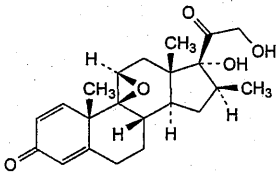
B. 21-chloro-9-fluoro-11β,17-dihydroxy-16β-methylpregna-1,4-diene-3,20-dione,



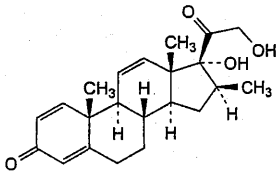
C. 17,21-dihydroxy-16β-methylpregna-1,4,9(11)-triene-3,20-dione,



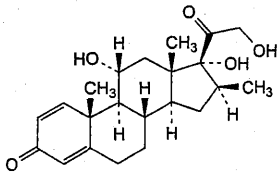
D. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl ethoxycarboxylate,



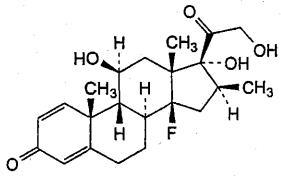
E. 9,11β-epoxy-17,21-dihydroxy-16β-methyl-9β-pregna-1,4-diene-3,20-dione,



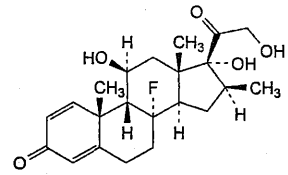
F. 17,21-dihydroxy-16β-methylpregna-1,4,11-triene-3,20-dione,



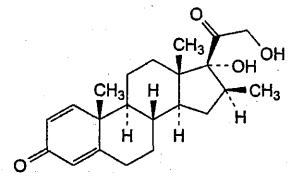
G. 11α,17,21-trihydroxy-16β-methylpregna-1,4-diene-3,20-dione,



H. 14-fluoro-11β,17,21-trihydroxy-16β-methyl-8α,9β,14β-pregna-1,4-diene-3,20-dione,



I. 8-fluoro-11β,17,21-trihydroxy-16β-methyl-8α,9β-pregna-1,4-diene-3,20-dione,

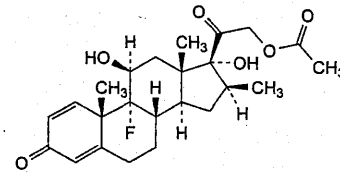


J. 17,21-dihydroxy-16β-methylpregna-1,4-diene-3,20-dione.

Ph Eur

Betamethasone Acetate

(Ph. Eur. monograph 0975)



$C_{24}H_{31}FO_6$ 434.5 *987-24-6*

Action and use
Glucocorticoid.

Ph Eur

DEFINITION
9-Fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-21-yl acetate.

Content
97.0 per cent to 103.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B, C.

Second identification: A, C, D, E, F.

A. Dissolve 10.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Place 2.0 mL of this solution in a ground-glass-stoppered tube, add 10.0 mL of *phenylhydrazine-sulfuric acid solution R*, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (*2.2.25*) measured at 419 nm is not greater than 0.10.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison betamethasone acetate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference

substance separately in the minimum volume of *methanol R*, evaporate to dryness on a water-bath and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *betamethasone acetate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *prednisolone acetate CRS* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R.* Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a deep reddish-brown colour develops. Add this solution to 10 mL of *water R* and mix. The colour is discharged and a clear solution remains.

E. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

F. About 10 mg gives the reaction of acetyl (*2.3.1*).

TESTS

Specific optical rotation (*2.2.7*)

+ 120 to + 128 (anhydrous substance).

Dissolve 0.250 g in *dioxan R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in 4 mL of *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of *betamethasone acetate CRS* and 2 mg of *dexamethasone acetate CRS* (impurity B) in the mobile phase, then dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase In a 1000 mL volumetric flask mix 380 mL of *acetonitrile R* with 550 mL of *water R* and allow to equilibrate; dilute to 1000 mL with *water R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 30 min.

Injection 20 µL.

Run time 2.5 times the retention time of betamethasone acetate.

Retention time Betamethasone acetate = about 19 min; impurity B = about 22 min.

System suitability Reference solution (a):
— *resolution*: minimum 3.3 between the peaks due to betamethasone acetate and impurity B; if necessary, adjust slightly the concentration of acetonitrile in the mobile phase.

Limits:
— *impurities A, B, C, D*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *total*: not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.25 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)

Maximum 4.0 per cent, determined on 0.100 g.

ASSAY

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 240 nm.

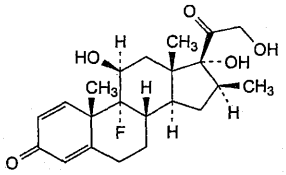
Calculate the content of $C_{24}H_{31}FO_6$ taking the specific absorbance to be 350.

STORAGE

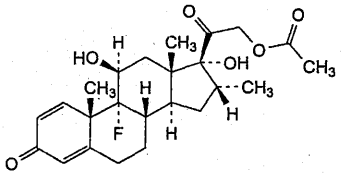
Protected from light.

IMPURITIES

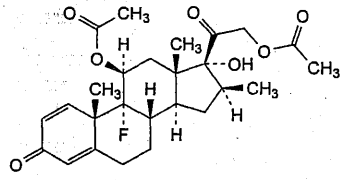
Specified impurities *A, B, C, D.*



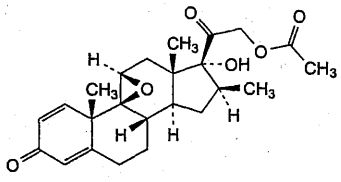
A. 9-fluoro-11β,17,21-trihydroxy-16β-methylpregna-1,4-diene-3,20-dione (betamethasone),



B. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (dexamethasone acetate),



C. 9-fluoro-17-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-11β,21-diyl diacetate (betamethasone 11,21-diacetate),

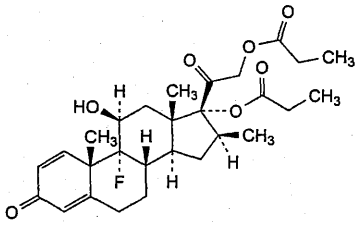


D. 9,11β-epoxy-17-hydroxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-diene-21-yl acetate.

Ph Eur

Betamethasone Dipropionate

(Ph. Eur. monograph 0809)



$C_{28}H_{37}FO_7$ 504.6 *5593-20-4*

Action and use
Glucocorticoid.

Ph Eur

DEFINITION
9-Fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in methylene chloride, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B.

Second identification: A, C, D, E.

A. Dissolve 10.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Place 2.0 mL of the solution in a ground-glass-stoppered tube, add 10.0 mL of *phenylhydrazine-sulfuric acid solution R*, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (*2.2.25*) measured at 419 nm is not more than 0.10.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *betamethasone dipropionate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 25 mg of the substance to be examined in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of solution A to 10 mL with *methylene chloride R.*

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a current of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 2 h. Allow to cool.

Reference solution (a) Dissolve 25 mg of *betamethasone dipropionate CRS* in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of solution B to 10 mL with *methylene chloride R.*

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a current of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 2 h. Allow to cool.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R*, heat at 120 °C for 10 min or until the spots appear, and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with

the corresponding reference solution; the principal spot in each of the chromatograms obtained with test solution (b) and reference solution (b) has an R_F value distinctly lower than that of the principal spot in each of the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a deep reddish-brown colour develops. Add this solution to 10 mL of *water R* and mix. The colour is discharged and a clear solution remains.

E. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. Add 1.0 mL of the filtrate to a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Specific optical rotation (*2.2.7*)

+ 84 to + 88 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 60.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *betamethasone dipropionate for system suitability A CRS* (containing impurities B, C, D, E, G and I) in the mobile phase and dilute to 2.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 60.0 mg of *betamethasone dipropionate CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 5 mg of *betamethasone dipropionate for peak identification CRS* (containing impurity H) in the mobile phase and dilute to 2.0 mL with the mobile phase.

Column:
- *size*: l = 0.10 m, Ø = 2.0 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (2.5 µm);
- *temperature*: 20 ± 2 °C.

Mobile phase Mix 35 mL of *water for chromatography R* and 56 mL of *acetonitrile R* and allow to equilibrate; dilute to 100 mL with *water for chromatography R* and mix.

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL of test solution (a) and reference solutions (a), (b) and (d).

Run time 3 times the retention time of betamethasone dipropionate.

Identification of impurities Use the chromatogram supplied with *betamethasone dipropionate for system suitability A CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C, D, E, G and I; use the chromatogram supplied with *betamethasone dipropionate for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity H.

Relative retention With reference to betamethasone dipropionate (retention time = about 10 min): impurity B = about 0.4; impurity C = about 0.5; impurity D = about 0.7; impurity I = about 1.16; impurity E = about 1.22; impurity H = about 1.7; impurity G = about 2.1.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to betamethasone dipropionate; minimum 4.0, where H_p = height above the baseline of the peak due to impurity I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity E.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity G = 1.3; impurity H = 1.4;
- *impurity C*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurities B, H*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurities D, E, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *impurity I*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{28}H_{37}FO_7$ taking into account the assigned content of *betamethasone dipropionate CRS*.

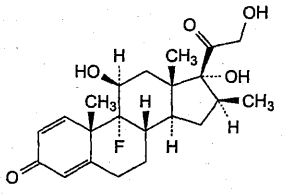
STORAGE

Protected from light.

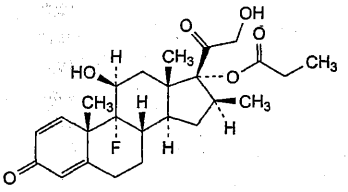
IMPURITIES

Specified impurities *B, C, D, E, G, H, I.*

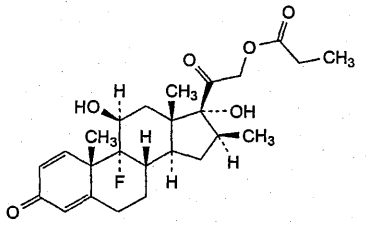
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, F.*



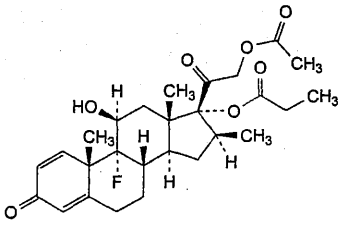
A. 9-fluoro-11β,17,21-trihydroxy-16β-methylpregna-1,4-diene-3,20-dione (betamethasone),



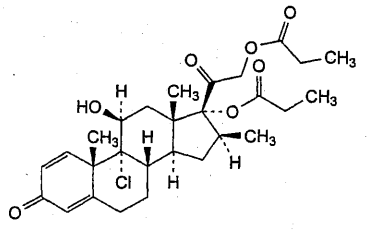
B. 9-fluoro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (betamethasone 17-propionate),



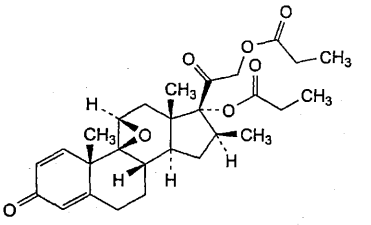
C. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl propanoate (betamethasone 21-propionate),



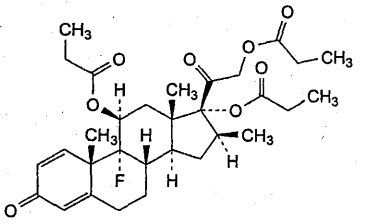
D. 21-(acetyloxy)-9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (betamethasone 21-acetate 17-propionate),



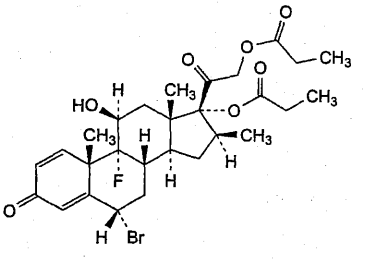
E. 9-chloro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate (beclometasone dipropionate),



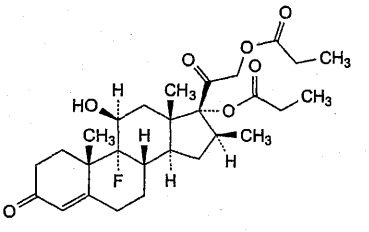
F. 9,11β-epoxy-16β-methyl-3,20-dioxo-9β-pregna-1,4-diene-17,21-diyl dipropanoate (9β,11β-epoxybetamethasone dipropionate),



G. 9-fluoro-16β-methyl-3,20-dioxopregna-1,4-diene-11β,17,21-triyl tripropanoate (betamethasone tripropionate),



H. 6α-bromo-9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-diene-17,21-diyl dipropanoate (6α-bromobetamethasone dipropionate),



I. 9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregn-4-ene-17,21-diyl dipropanoate (1,2-dihydrobetamethasone dipropionate).

Ph Eur

TESTS

Specific optical rotation (*2.2.7*)

+ 77 to + 83 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light. Prepare the solutions immediately before use.*

Solvent mixture *glacial acetic acid R*, mobile phase (1:1000 *V/V*).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 12.5 mg of *betamethasone valerate for system suitability CRS* (containing impurities D and G) in 5.0 mL of the solvent mixture. Use 1.0 mL of this solution to dissolve the contents of a vial of *betamethasone valerate impurity mixture CRS* (containing impurities C, H and I).

Reference solution (c) Dissolve 6 mg of *betamethasone CRS* (impurity A) and 3 mg of *betamethasone 21-valerate CRS* (impurity E) in 30.0 mL of the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 20 °C.

Mobile phase *acetonitrile R, water R* (50:50 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 239 nm.

Injection 20 µL.

Run time 2.5 times the retention time of betamethasone valerate.

Identification of impurities Use the chromatogram supplied with *betamethasone valerate for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C, D, G, H and I; use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and E.

Relative retention With reference to betamethasone valerate (retention time = about 20 min): impurity A = about 0.3; impurity I = about 0.6; impurity C = about 0.8; impurity H = about 1.3; impurity D = about 1.4; impurity E = about 1.6; impurity G = about 2.0.

System suitability Reference solution (b):

— *resolution*: minimum 1.7 between the peaks due to impurities H and D.

Limits:

— *impurity A*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);

— *impurities E, G*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— *impurities C, H, I*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 50.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 50.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 240 nm.

Calculate the content of $C_{27}H_{37}FO_6$ taking the specific absorbance to be 325.

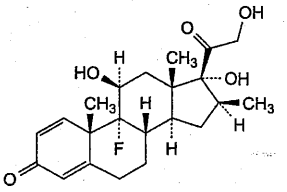
STORAGE

Protected from light.

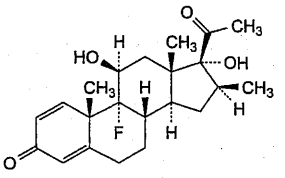
IMPURITIES

Specified impurities *A, C, E, G, H, I.*

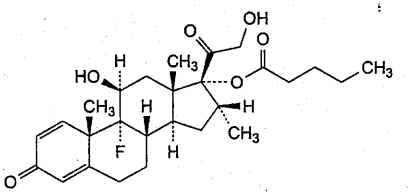
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, F.*



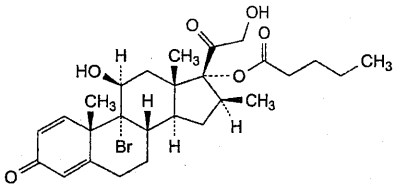
A. 9-fluoro-11β,17,21-trihydroxy-16β-methylpregna-1,4-diene-3,20-dione (betamethasone),



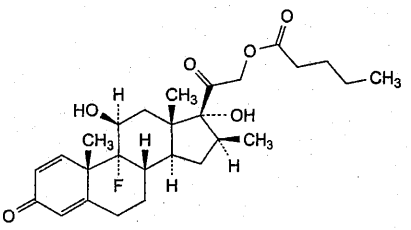
B. 9-fluoro-11β,17-dihydroxy-16β-methylpregna-1,4-diene-3,20-dione (21-deoxy-betamethasone),



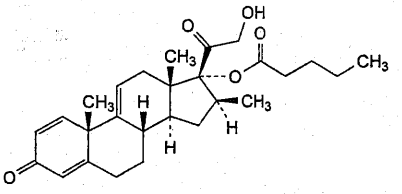
C. 9-fluoro-11β,21-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-17-yl pentanoate (dexamethasone 17-valerate),



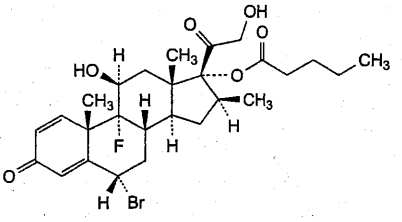
D. 9-bromo-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl pentanoate (9-bromo-betamethasone valerate),



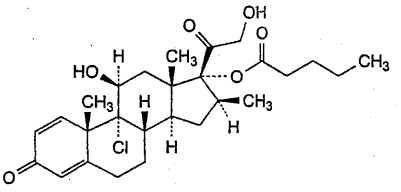
E. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl pentanoate (betamethasone 21-valerate),



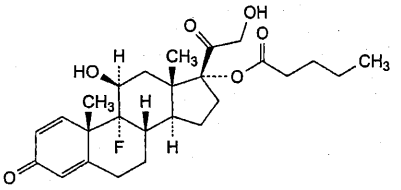
F. 21-hydroxy-16β-methyl-3,20-dioxopregna-1,4,9(11)-trien-17-yl pentanoate (betamethasone valerate δ-9(11)),



G. 6α-bromo-9-fluoro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl pentanoate (6α-bromo-betamethasone valerate),



H. 9-chloro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl pentanoate (beclomethasone 17-valerate),

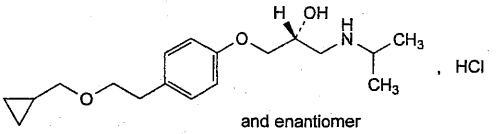


I. 9-fluoro-11β,21-dihydroxy-3,20-dioxopregna-1,4-dien-17-yl pentanoate (9-fluoro-prednisolone 17-valerate).

Ph Eur

Betaxolol Hydrochloride

(Ph. Eur. monograph 1072)



and enantiomer

$C_{18}H_{30}ClNO_3$ 343.9 *63659-19-8*

Action and use
Beta-adrenoceptor antagonist.

Preparations
Betaxolol Eye Drops, Solution
Betaxolol Eye Drops, Suspension

Ph Eur

DEFINITION

(2*RS*)-1-[4-[2-(Cyclopropylmethoxy)ethyl]phenoxy]-3-[(1-methylethyl)amino]propan-2-ol hydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent), soluble in methylene chloride.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 113 °C to 117 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *betaxolol hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 1 mL of *methanol R.*

Reference solution (a) Dissolve 20 mg of *betaxolol hydrochloride CRS* in 2 mL of *methanol R.*

Reference solution (b) Dissolve 10 mg of *oxprenolol hydrochloride CRS* in 1 mL of reference solution (a).

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *perchloric acid R, methanol R, water R* (0.5:50:50 *V/V/V*).

Application 2 μL.

Development Over a path of 10 cm.

Drying In air.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with a 50 g/L solution of *vanillin R* in a mixture of 5 volumes of *sulfuric acid R*, 10 volumes of *glacial acetic acid R* and 85 volumes of *methanol R*, heat at 100-105 °C until the colour of the spots reaches maximum intensity (10-15 min), and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.5 g in *water R* and dilute to 25 mL with the same solvent.

Acidity or alkalinity

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent. Add 0.2 mL of *methyl red solution R* and 0.2 mL of *0.01 M hydrochloric acid.* The solution is red. Add 0.4 mL of *0.01 M sodium hydroxide.* The solution is yellow.

Related substances

Liquid chromatography (*2.2.29*). *Prepare reference solutions (c) and (d) immediately before use.*

Test solution Dissolve 10 mg of the substance to be examined in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (a) Dissolve 8 mg of the substance to be examined and 4 mg of *betaxolol impurity A CRS* in 20.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 2 mg of *betaxolol impurity C CRS* in 50 mL of the mobile phase. Dilute 5 mL of the solution to 20 mL with the mobile phase.

Reference solution (d) Dissolve 10 mg of *betaxolol for peak identification CRS* (containing impurities B, D and E) in 5 mL of reference solution (c).

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 175 mL of *acetonitrile R* and 175 mL of *methanol R* and dilute to 1 L with a 3.4 g/L solution of *potassium dihydrogen phosphate R*, previously adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 µL of the test solution and reference solutions (a), (b) and (d).

Run time 4.5 the retention time of betaxolol.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *betaxolol for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B, C, D and E.

Relative retention With reference to betaxolol (retention time = about 8 min): impurity B = about 0.3; impurity A = about 0.8; impurity D = about 1.5; impurity E = about 2.2; impurity C = about 4.1.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity A and betaxolol.

Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 10.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

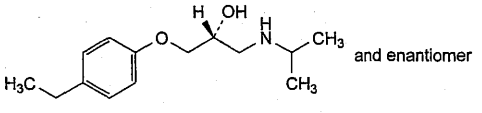
1 mL of *0.1 M sodium hydroxide* is equivalent to 34.39 mg of $C_{18}H_{30}ClNO_3$.

STORAGE

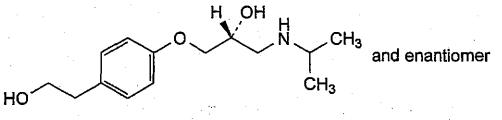
Protected from light.

IMPURITIES

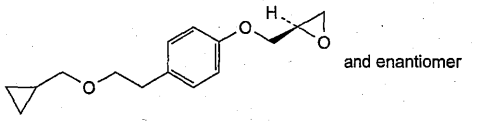
Specified impurities *A, B, C, D, E.*



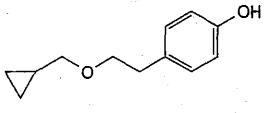
A. (2*RS*)-1-(4-ethylphenoxy)-3-[(1-methylethyl) amino]propan-2-ol,



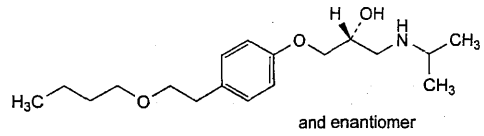
B. (2*RS*)-1-[4-(2-hydroxyethyl)phenoxy]-3-[(1-methylethyl)amino]propan-2-ol,



C. (2*RS*)-2-[[4-[2-(cyclopropylmethoxy)ethyl] phenoxy]methyl]oxirane,



D. 4-[2-(cyclopropylmethoxy)ethyl]phenol,



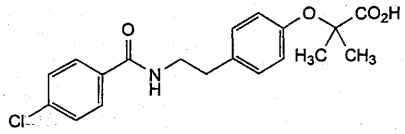
and enantiomer

E. (2*RS*)-1-[4-(2-butoxyethyl)phenoxy]-3-[(1-methylethyl)amino]propan-2-ol.

Ph Eur

Bezafibrate

(Ph. Eur. monograph 1394)



$C_{19}H_{20}ClNO_4$ 361.8 *41859-67-0*

Action and use
Fibrate; lipid-regulating drug.

Preparations
Bezafibrate Tablets

Bezafibrate Prolonged-release Tablets

Ph Eur

DEFINITION
2-[4-[2-[(4-Chlorobenzoyl)amino]ethyl]phenoxy]-2-methylpropanoic acid.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in dimethylformamide, sparingly soluble in acetone and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A, B.

Second identification: A, C.

A. Melting point (*2.2.14*): 181 °C to 185 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bezafibrate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *methanol R* and evaporate to dryness. Dry the residues *in vacuo* at 80 °C for 1 h and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 10 mg of *bezafibrate CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, methyl ethyl ketone R, xylene R* (2.7:30:60 *V/V/V*).

Application 5 µL.

Development Over half of the plate.

Drying At 120 °C for at least 15 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Solution S
Dissolve 1.0 g in *dimethylformamide R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 10.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the mobile phase.

Reference solution (c) To 1 mL of the test solution, add 1 mL of *0.1 M hydrochloric acid* and evaporate to dryness on a hot plate. Dissolve the residue in 20 mL of the mobile phase.

Column:
— *size: l* = 0.125 m, Ø = 4 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 40 volumes of a 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.3 with *phosphoric acid R*, and 60 volumes of *methanol R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 228 nm.

Injection 20 µL.

Run time The time necessary to detect the ester, which, depending on the route of synthesis, may be impurity C, D or E.

Relative retention With reference to bezafibrate (retention time = about 6.0 min): impurity A = about 0.5; impurity B = about 0.6; impurity C = about 1.5; impurity D = about 2.3; impurity E = about 6.2.

System suitability:
— *resolution*: minimum 5.0 between the 2 principal peaks in the chromatogram obtained with reference solution (c);
— *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.75 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 300 ppm.

Dilute 10 mL of solution S to 50 mL with *water R*. Filter the resultant suspension through a wet filter previously washed with *water R* until free from chlorides. Prepare the standard using 9 mL of *chloride standard solution (5 ppm Cl) R* and 6 mL of *water R*.

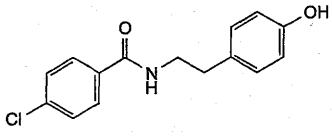
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

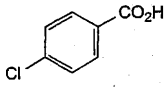
ASSAY
Dissolve 0.300 g in 50 mL of a mixture of 25 volumes of *water R* and 75 volumes of *ethanol (96 per cent) R*. Using 0.1 mL of *phenolphthalein solution R* as indicator, titrate with *0.1 M sodium hydroxide* until a pink colour is obtained. Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 36.18 mg of $C_{19}H_{20}ClNO_4$.

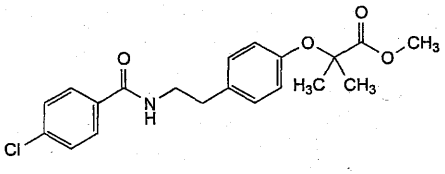
IMPURITIES
Specified impurities *A, B, C, D, E.*



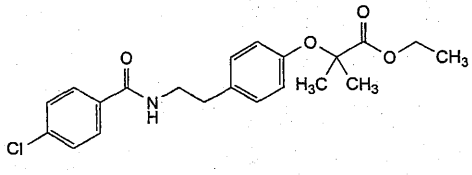
A. 4-chloro-*N*-[2-(4-hydroxyphenyl)ethyl]benzamide (chlorobenzoyltyramine),



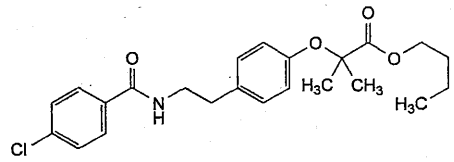
B. 4-chlorobenzoic acid,



C. methyl 2-[4-[2-[(4-chlorobenzoyl)amino]ethyl]phenoxy]-2-methylpropanoate,



D. ethyl 2-[4-[2-[(4-chlorobenzoyl)amino]ethyl]phenoxy]-2-methylpropanoate,

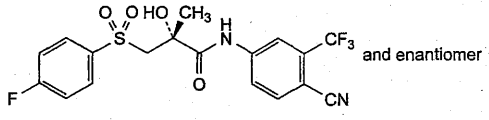


E. butyl 2-[4-[2-[(4-chlorobenzoyl)amino]ethyl]phenoxy]-2-methylpropanoate.

Ph Eur

Bicalutamide

(Ph. Eur. monograph 2196)



$C_{18}H_{14}F_4N_2O_4S$ 430.4 *90357-06-5*

Action and use
Antiandrogen; treatment of prostate cancer.

Preparation
Bicalutamide Tablets

Ph Eur

DEFINITION
(2*RS*)-*N*-[4-Cyano-3-(trifluoromethyl)phenyl]-3-[(4-fluorophenyl)sulfonyl]-2-hydroxy-2-methylpropanamide.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in acetone, slightly soluble in anhydrous ethanol and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bicalutamide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *phosphoric acid R, acetonitrile R1, water R* (0.05:50:50 *V/V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *bicalutamide for system suitability CRS* (containing impurities B and C) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of *bicalutamide CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 25.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: *phosphoric acid R, acetonitrile R1, water R* (1.9:100:1900 *V/V/V*);
- *mobile phase B*: *phosphoric acid R, water R, acetonitrile R1* (1.9:100:1900 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	92	8
3 - 23	92 → 67	8 → 33
23 - 43	67 → 50	33 → 50
43 - 50	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *bicalutamide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

Relative retention With reference to bicalutamide (retention time = about 38 min): impurity B = about 0.98; impurity C = about 1.1.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bicalutamide.

Limits:
- *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

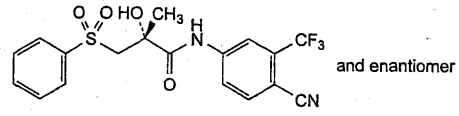
Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{18}H_{14}F_4N_2O_4S$ taking into account the assigned content of *bicalutamide CRS*.

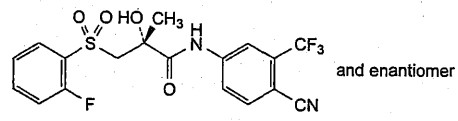
IMPURITIES

Specified impurities C.

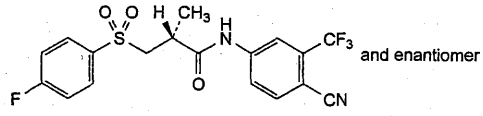
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, D, E, F, H, J, K, L, M.



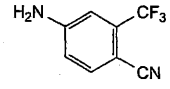
A. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-2-hydroxy-2-methyl-3-(phenylsulfonyl)propanamide,



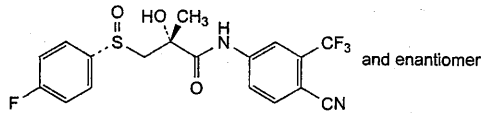
B. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-3-[(2-fluorophenyl)sulfonyl]-2-hydroxy-2-methylpropanamide,



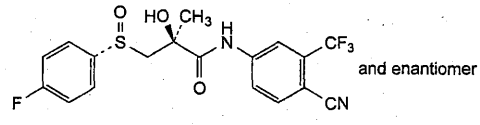
C. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-3-[(4-fluorophenyl)sulfonyl]-2-methylpropanamide,



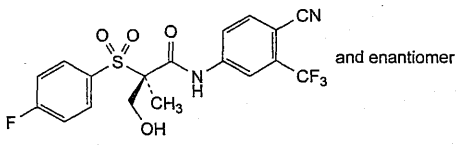
D. 4-amino-2-(trifluoromethyl)benzonitrile,



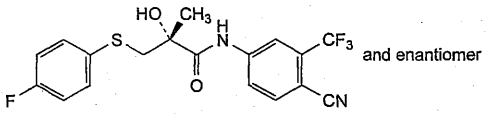
E. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-3-[(*RS*)-(4-fluorophenyl)sulfinyl]-2-hydroxy-2-methylpropanamide,



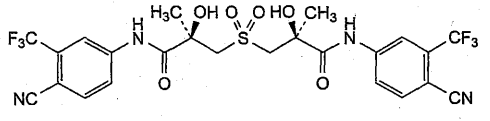
F. (2*SR*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-3-[(*RS*)-(4-fluorophenyl)sulfinyl]-2-hydroxy-2-methylpropanamide,


and enantiomer

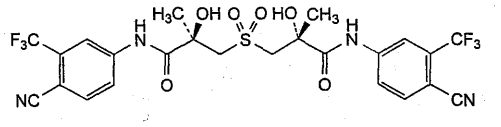
H. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-2-[(4-fluorophenyl)sulfonyl]-3-hydroxy-2-methylpropanamide,


and enantiomer

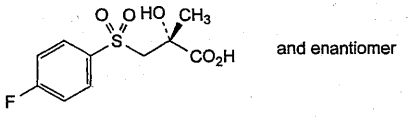
J. (2*RS*)-*N*-[4-cyano-3-(trifluoromethyl)phenyl]-3-[(4-fluorophenyl)sulfanyl]-2-hydroxy-2-methylpropanamide,



K. (2*R*,2′*S*)-3,3′-sulfonylbis[*N*-[4-cyano-3-(trifluoromethyl)phenyl]-2-hydroxy-2-methylpropanamide],


and enantiomer

L. (2*RS*,2′*RS*)-3,3′-sulfonylbis[*N*-[4-cyano-3-(trifluoromethyl)phenyl]-2-hydroxy-2-methylpropanamide],

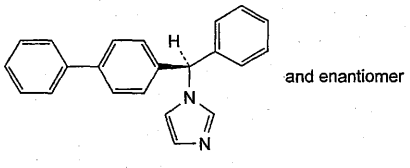

and enantiomer

M. (2*RS*)-3-[(4-fluorophenyl)sulfonyl]-2-hydroxy-2-methylpropanoic acid.

Ph Eur

Bifonazole

(Ph. Eur. monograph 1395)


and enantiomer

$C_{22}H_{18}N_2$ 310.4 *60628-96-8*

Action and use
Antifungal.

Ph Eur

DEFINITION
1-[(*RS*)-(Biphenyl-4-yl)phenylmethyl]-1*H*-imidazole.

Content
98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in anhydrous ethanol.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bifonazole CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *2-propanol R*, evaporate to dryness and record new spectra using the residues.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Buffer solution pH 3.2 Mix 2.0 mL of *phosphoric acid R* with 980 mL of *water R*, adjust to pH 3.2 (*2.2.3*) with *triethylamine R* and dilute to 1000.0 mL with *water R.*

Test solution Dissolve 50.0 mg of the substance to be examined in 25 mL of *acetonitrile R* and dilute to 50.0 mL with buffer solution pH 3.2.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with buffer solution pH 3.2. Dilute 1.0 mL of this solution to 10.0 mL with buffer solution pH 3.2.

Reference solution (b) Dissolve 2 mg of *bifonazole for system suitability CRS* (containing impurities A, B, C, D and E) in 2 mL of *acetonitrile R* and dilute to 10.0 mL with buffer solution pH 3.2.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, buffer solution pH 3.2 (20:80 *V/V*);
— *mobile phase B*: buffer solution pH 3.2, *acetonitrile R1* (20:80 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	60	40
8 - 12	60 → 10	40 → 90
12 - 30	10	90

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram supplied with *bifonazole for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to bifonazole (retention time = about 4 min): impurity C = about 0.2; impurity B = about 0.7; impurity A = about 3.2; impurity D = about 3.6; impurity E = about 5.8.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurity B and bifonazole.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 2;

— *impurities B, D*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurities A, C*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity E*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

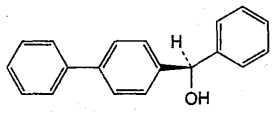
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 80 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

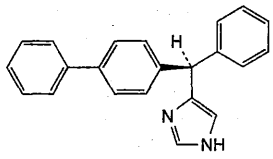
1 mL of *0.1 M perchloric acid* is equivalent to 31.04 mg of $C_{22}H_{18}N_2$.

IMPURITIES
Specified impurities *A, B, C, D, E.*



and enantiomer

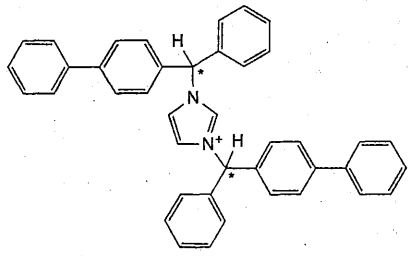
A. (*RS*)-(biphenyl-4-yl)phenylmethanol,



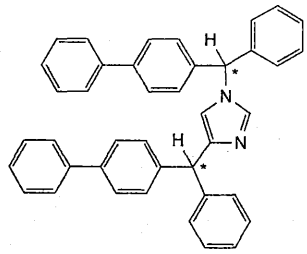
and enantiomer

B. 4-[(*RS*)-(biphenyl-4-yl)phenylmethyl]-1*H*-imidazole,

C. 1*H*-imidazole,



D. 1,3-bis[(biphenyl-4-yl)phenylmethyl]-1*H*-imidazolium ion,

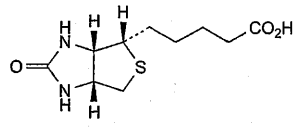


E. 1,4-bis[(biphenyl-4-yl)phenylmethyl]-1*H*-imidazole.

Ph Eur

Biotin

(Ph. Eur. monograph 1073)



$C_{10}H_{16}N_2O_3S$ 244.3 *58-85-5*

Action and use
Vitamin.

Ph Eur

DEFINITION
5-[(3a*S*,4*S*,6a*R*)-2-Oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]pentanoic acid.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Very slightly soluble in water and in ethanol (96 per cent), practically insoluble in acetone. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: A.
Second identification: B.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *biotin CRS.*

B. Thin-layer chromatography (*2.2.27*).
Prepare the solutions immediately before use and keep protected from bright light.
Test solution Dissolve 5 mg of the substance to be examined in *glacial acetic acid R* and dilute to 10 mL with the same solvent.
Reference solution Dissolve 5 mg of *biotin CRS* in *glacial acetic acid R* and dilute to 10 mL with the same solvent.
Plate *TLC silica gel plate R* (5 µm).
Mobile phase *methanol R, glacial acetic acid R, toluene R* (5:25:75 *V/V/V*).
Application 10 µL.
Development Over 3/4 of the plate.
Drying In a current of warm air.

Detection Allow to cool and spray with *4-dimethylaminocinnamaldehyde solution R*; examine immediately in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Solution S

Dissolve 0.250 g in a 4 g/L solution of *sodium hydroxide R* and dilute to 25.0 mL with the same alkaline solution.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 89 to + 93 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and keep protected from bright light.*

Solvent mixture *water R, acetonitrile R* (50:50 *V/V*).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture using sonication and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *biotin for system suitability CRS* (containing impurities A, C and E) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *methanesulfonic acid R, acetonitrile R1, water for chromatography R* (1:25:1000 *V/V/V*);
- *mobile phase B*: *methanesulfonic acid R, water for chromatography R, acetonitrile R1* (1:25:1000 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	95	5
5 - 20	95 → 0	5 → 100
20 - 28	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 200 nm from 0 to 5 min and at 210 nm from 5 to 28 min.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *biotin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, C and E.

Relative retention With reference to biotin (retention time = about 12 min): impurity C = about 0.2; impurity A = about 1.1; impurity E = about 1.3.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to biotin and impurity A.

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity E by 0.2;
- for each impurity, use the concentration of biotin in reference solution (a).

Limits:
- *impurities A, E*: for each impurity, maximum 0.5 per cent;
- *impurity C*: maximum 0.2 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 2.0 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Suspend 0.200 g in 5 mL of *dimethylformamide R*. Heat until the substance has dissolved completely. Add 50 mL of *ethanol R* and titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 24.43 mg of $C_{10}H_{16}N_2O_3S$.

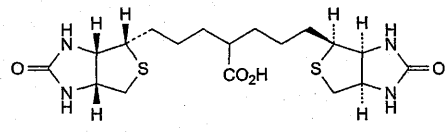
STORAGE

Store protected from light.

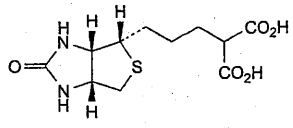
IMPURITIES

Specified impurities *A, C, E.*

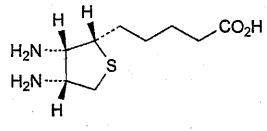
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, F, G, H.*



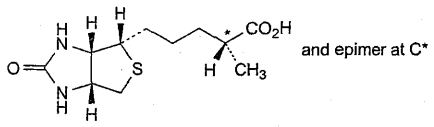
A. 5-[(3a*S*,4*S*,6a*R*)-2-oxohexahydro-1*H*-thieno[3,4-*d*] imidazol-4-yl]-2-[[(3a*S*,4*S*,6a*R*)-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]propyl]pentanoic acid,



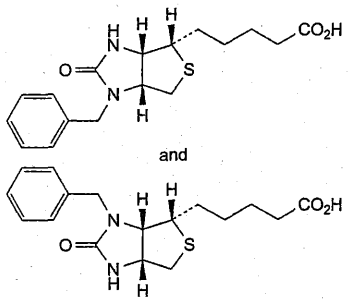
B. 4-[(3a*S*,4*S*,6a*R*)-2-oxohexahydro-1*H*-thieno[3,4-*d*] imidazol-4-yl]butane-1,1-dicarboxylic acid,



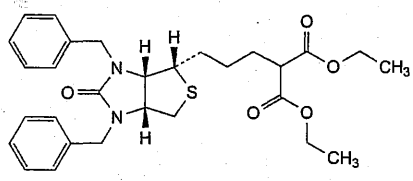
C. 5-[(2*S*,3*S*,4*R*)-3,4-diaminothiolan-2-yl]pentanoic acid,



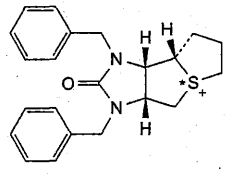
D. (2*RS*)-2-methyl-5-[(3a*S*,4*S*,6a*R*)-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]pentanoic acid,



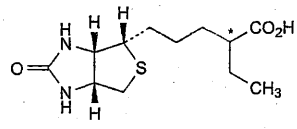
E. 5-[(3a*S*,4*S*,6a*R*)-1-benzyl-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]pentanoic acid and 5-[(3a*S*,4*S*,6a*R*)-3-benzyl-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]pentanoic acid,



F. diethyl 4-[(3a*S*,4*S*,6a*R*)-1,3-dibenzyl-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]butane-1,1-dicarboxylate,



G. (3a*R*,8a*S*,8b*S*)-1,3-dibenzyl-2-oxodecahydrothieno[1′,2′:1,2]thieno[3,4-*d*]imidazol-5-ium,

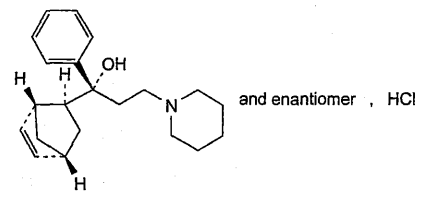


H. 2-ethyl-5-[(3a*S*,4*S*,6a*R*)-2-oxohexahydro-1*H*-thieno[3,4-*d*]imidazol-4-yl]pentanoic acid.

Ph Eur

Biperiden Hydrochloride

(Ph. Eur. monograph 1074)



$C_{21}H_{30}ClNO$ 347.9 *1235-82-1*

Action and use
Anticholinergic.

Ph Eur

DEFINITION

(1*RS*)-1-[(1*RS*,2*SR*,4*RS*)-Bicyclo[2.2.1]hept-5-en-2-yl]-1-phenyl-3-(piperidin-1-yl)propan-1-ol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water and in alcohol, very slightly soluble in methylene chloride.

mp
About 280 °C, with decomposition.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *biperiden hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of *biperiden hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *biperiden impurity A CRS* in reference solution (a) and dilute to 2 mL with the same solution.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *diethylamine R, methanol R, toluene R* (1:1:20 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *dilute potassium iodobismuthate solution R* and then with *sodium nitrite solution R* and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to

the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

C. To about 20 mg add 5 mL of *phosphoric acid R*. A green colour develops.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 0.10 g in *carbon dioxide-free water R*, heating gently if necessary, and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*) and is colourless (*2.2.2, Method II*).

pH (*2.2.3*)

5.0 to 6.5 for solution S.

Related substances

Gas chromatography (*2.2.28*).

Test solution Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 0.5 mL of the test solution to 100 mL with *methanol R*. Dilute 10 mL of this solution to 50 mL with *methanol R*.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *biperiden impurity A CRS* in *methanol R* and dilute to 5 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with *methanol R*.

Column:
— *material*: fused silica,
— *size*: *l* = 50 m, Ø = 0.25 mm,
— *stationary phase*: *poly(dimethyl) (diphenyl) (divinyl)siloxane R* (film thickness 0.25 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 0.4 mL/min.

Split ratio 1:250.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 5	200
	5 - 40	200 → 270
Injection port		250
Detector		300

Detection Flame ionisation.

Injection 2 µL.

Run time Twice the retention time of biperiden.

Relative retention With reference to biperiden: impurities A, B and C = between 0.95 and 1.05.

System suitability:
— *resolution*: minimum 2.5 between the peak due to biperiden (1st peak) and the peak due to impurity A (2nd peak) in the chromatogram obtained with reference solution (b),
— *signal-to-noise ratio*: minimum 6 for the principal peak in the chromatogram obtained with reference solution (a).

Limits:
— *impurities A, B, C*: for each impurity, maximum 0.50 per cent of the area of the principal peak,
— *any other impurity*: for each impurity, maximum 0.10 per cent of the area of the principal peak,
— *total of impurities A, B and C*: maximum 1.0 per cent of the area of the principal peak,
— *total of impurities other than A, B and C*: maximum 0.50 per cent of the area of the principal peak,
— *disregard limit*: 0.05 per cent of the area of the principal peak.

Impurity F (*2.4.24*)

Maximum 2 ppm.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 60 mL of *alcohol R*. In a closed vessel, titrate with *0.1 M alcoholic potassium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M alcoholic potassium hydroxide* is equivalent to 34.79 mg of $C_{21}H_{30}ClNO$.

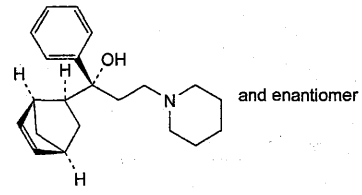
STORAGE

In an airtight container, protected from light.

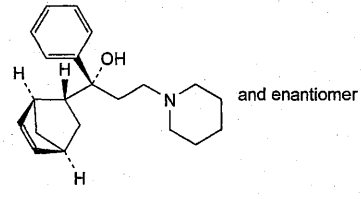
IMPURITIES

Specified impurities *A, B, C, F.*

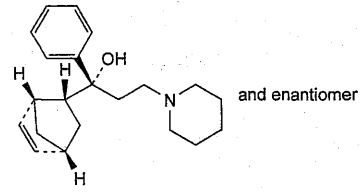
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E.*



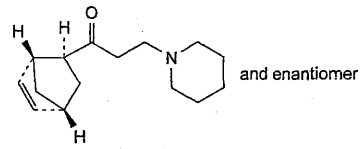
A. (1*RS*)-1-[(1*SR*,2*SR*,4*SR*)-bicyclo[2.2.1]hept-5-en-2-yl]-1-phenyl-3-(piperidin-1-yl)propan-1-ol (*endo* form),



B. (1*RS*)-1-[(1*SR*,2*RS*,4*SR*)-bicyclo[2.2.1]hept-5-en-2-yl]-1-phenyl-3-(piperidin-1-yl)propan-1-ol,

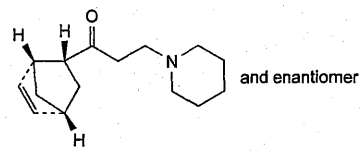


C. (1*RS*)-1-[(1*RS*,2*RS*,4*RS*)-bicyclo[2.2.1]hept-5-en-2-yl]-1-phenyl-3-(piperidin-1-yl)propan-1-ol,



and enantiomer

D. 1-[(1*RS*,2*SR*,4*RS*)-bicyclo[2.2.1]hept-5-en-2-yl]-3-(piperidin-1-yl)propan-1-one,



and enantiomer

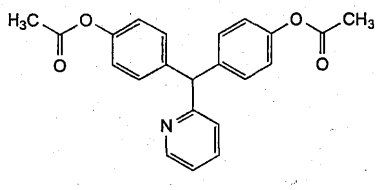
E. 1-[(1*RS*,2*RS*,4*RS*)-bicyclo[2.2.1]hept-5-en-2-yl]-3-(piperidin-1-yl)propan-1-one,

F. benzene.

Ph Eur

Bisacodyl

(Ph. Eur. monograph 0595)



$C_{22}H_{19}NO_4$ 361.4 *603-50-9*

Action and use
Stimulant laxative.

Preparations
Bisacodyl Suppositories
Bisacodyl Gastro-resistant Tablets

Ph Eur

DEFINITION
4,4′-(Pyridin-2-ylmethylene)diphenyl diacetate.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetone, sparingly soluble in ethanol (96 per cent). It dissolves in dilute mineral acids.

IDENTIFICATION
First identification: C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 131 °C to 135 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 10.0 mg in a 6 g/L solution of *potassium hydroxide R* in *methanol R* and dilute to 100.0 mL with the same solution. Dilute 10.0 mL of this solution to 100.0 mL with a 6 g/L solution of *potassium hydroxide R* in *methanol R*.

Spectral range 220-350 nm.

Absorption maximum At 248 nm.

Shoulder At 290 nm.

Specific absorbance at the absorption maximum 632 to 672.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bisacodyl CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *chloroform R*, evaporate to dryness and record new spectra using the residues.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *bisacodyl CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *methyl ethyl ketone R, xylene R* (50:50 *V/V*).

Application 10 µL.

Development Over a path of 10 cm.

Drying In air, if necessary heating at 100-105 °C.

Detection Spray with a mixture of equal volumes of *0.05 M iodine* and *dilute sulfuric acid R*.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Acidity or alkalinity
To 1.0 g add 20 mL of *carbon dioxide-free water R*, shake, heat to boiling, cool and filter. Add 0.2 mL of *0.01 M sodium hydroxide* and 0.1 mL of *methyl red solution R*. The solution is yellow. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *glacial acetic acid R, acetonitrile R, water R* (4:30:66 *V/V/V*).

Test solution Dissolve 50 mg of the substance to be examined in 25 mL of *acetonitrile R* and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.0 mg of *bisacodyl for system suitability CRS* (containing impurities A, B, C, D and E) in 1.0 mL of *acetonitrile R* and dilute to 2.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of *bisacodyl for peak identification CRS* (containing impurity F) in 2.5 mL of *acetonitrile R* and dilute to 5.0 mL with the solvent mixture.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 45 volumes of *acetonitrile R* and 55 volumes of a 1.58 g/L solution of *ammonium formate R* previously adjusted to pH 5.0 with *anhydrous formic acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 20 µL.

Run time 3.5 times the retention time of bisacodyl.

Identification of impurities Use the chromatogram supplied with *bisacodyl for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to bisacodyl (retention time = about 13 min): impurity A = about 0.2; impurity B = about 0.4; impurity C = about 0.45; impurity D = about 0.8; impurity E = about 0.9; impurity F = about 2.6.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bisacodyl.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.7;
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *impurities C, E*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity F*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

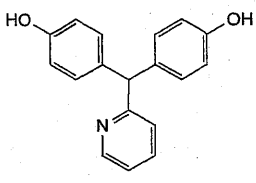
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

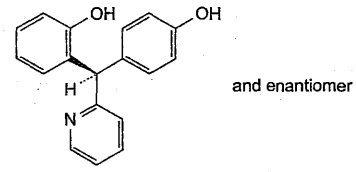
1 mL of *0.1 M perchloric acid* is equivalent to 36.14 mg of $C_{22}H_{19}NO_4$.

STORAGE
Protected from light.

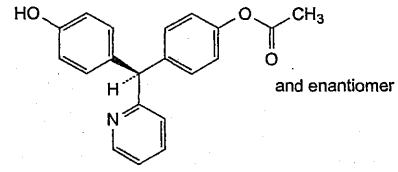
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



A. 4,4′-(pyridin-2-ylmethylene)diphenol,

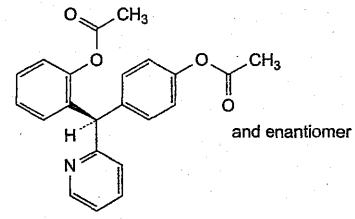


B. 2-[(*RS*)-(4-hydroxyphenyl)(pyridin-2-yl)methyl]phenol,



C. 4-[(*RS*)-(4-hydroxyphenyl)(pyridin-2-yl)methyl]phenyl acetate,

D. unknown structure,



E. 2-[(*RS*)-[4-(acetyloxy)phenyl](pyridin-2-yl)methyl]phenyl acetate,

F. unknown structure.

Ph Eur

Bismuth Subcarbonate

Bismuth Carbonate

(Ph. Eur. monograph 0012)

Ph Eur

DEFINITION

Content
80.0 per cent to 82.5 per cent of Bi (A_r 209.0) (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves with effervescence in mineral acids.

IDENTIFICATION
A. It gives the reaction of carbonates (*2.3.1*).

B. It gives the reactions of bismuth (*2.3.1*).

TESTS

Solution S
Shake 5.0 g with 10 mL of *water R* and add 20 mL of *nitric acid R*. Heat to dissolve, cool and dilute to 100 mL with *water R*.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and is colourless (*2.2.2, Method II*).

Chlorides (*2.4.4*)
Maximum 500 ppm.

To 6.6 mL of solution S add 4 mL of *nitric acid R* and dilute to 50 mL with *water R*.

Nitrates
Maximum 0.4 per cent.

To 0.25 g in a 125 mL conical flask, add 20 mL of *water R*, 0.05 mL of *indigo carmine solution R1* and then, as a single addition but with caution, 30 mL of *sulfuric acid R*. Titrate immediately with *indigo carmine solution R1* until a stable blue colour is obtained. Not more than *n* mL of the titrant is required, *n* being the volume corresponding to 1 mg of NO_3.

Alkali and alkaline-earth metals
Maximum 1.0 per cent.

To 1.0 g add 10 mL of *water R* and 10 mL of *acetic acid R*. Boil for 2 min, cool and filter. Wash the residue with 20 mL of *water R*. To the combined filtrate and washings add 2 mL of *dilute hydrochloric acid R* and 20 mL of *water R*. Boil and pass *hydrogen sulfide R* through the boiling solution until no further precipitate is formed. Filter, wash the residue with *water R*, evaporate the combined filtrate and washings to dryness on a water-bath and add 0.5 mL of *sulfuric acid R*. Ignite gently and allow to cool. The residue weighs a maximum of 10 mg.

Arsenic (*2.4.2, Method A*)
Maximum 5 ppm.

To 0.5 g in a distillation flask add 5 mL of *water R* and 7 mL of *sulfuric acid R*, allow to cool and add 5 g of *reducing mixture R* and 10 mL of *hydrochloric acid R*. Heat the contents of the flask to boiling gradually over 15-30 min and continue heating at such a rate that the distillation proceeds steadily until the volume in the flask is reduced by half or until 5 min after the air-condenser has become full of steam. It is important that distillation be discontinued before fumes of sulfur trioxide appear. Collect the distillate in a tube containing 15 mL of *water R* cooled in ice-water. Wash down the condenser with *water R* and dilute the distillate to 25 mL with the same solvent. Prepare the standard using a mixture of 2.5 mL of *arsenic standard solution (1 ppm As) R* and 22.5 mL of *water R*.

Copper
Maximum 50 ppm.

To 5 mL of solution S, add 2 mL of *ammonia R* and dilute to 50 mL with *water R*. Filter. To 10 mL of the filtrate add 1 mL of a 1 g/L solution of *sodium diethyldithiocarbamate R*. The solution is not more intensely coloured than a standard prepared at the same time in the same manner using a mixture of 0.25 mL of *copper standard solution (10 ppm Cu) R* and 9.75 mL of *water R* instead of 10 mL of the filtrate.

Lead
Maximum 20 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Dissolve 12.5 g in 75 mL of a mixture of equal volumes of *lead-free nitric acid R* and *water R*. Boil for 1 min, cool and dilute to 100.0 mL with *water R*.

Reference solutions Prepare the reference solutions using appropriate quantities of lead standard solution and a 37 per cent *V/V* solution of *lead-free nitric acid R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm (depending on the apparatus, the line at 217.0 nm may be used).

Atomisation device Air-acetylene flame.

Silver
Maximum 25 ppm.

To 2.0 g add 1 mL of *water R* and 4 mL of *nitric acid R*. Heat gently until dissolved and dilute to 11 mL with *water R*. Cool and add 2 mL of *1 M hydrochloric acid*. Allow to stand protected from light for 5 min. Any opalescence in the solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 10 mL of *silver standard solution (5 ppm Ag) R*, 1 mL of *nitric acid R* and 2 mL of *1 M hydrochloric acid*.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
Dissolve 0.500 g in 3 mL of *nitric acid R* and dilute to 250 mL with *water R*. Carry out the complexometric titration of bismuth (*2.5.11*).

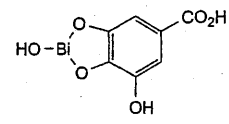
1 mL of *0.1 M sodium edetate* is equivalent to 20.90 mg of Bi.

STORAGE
Protected from light.

Ph Eur

Bismuth Subgallate

(Ph. Eur. monograph 1493)



$C_7H_5BiO_6$ 394.1 *99-26-3*

Ph Eur

DEFINITION
Complex of bismuth and gallic acid.

Content
48.0 per cent to 51.0 per cent of Bi (A_r 209.0) (dried substance).

CHARACTERS
Appearance
Yellow powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in mineral acids with decomposition and in solutions of alkali hydroxides, producing a reddish-brown liquid.

IDENTIFICATION
A. Mix 0.1 g with 5 mL of *water R* and 0.1 mL of *phosphoric acid R*. Heat to boiling and maintain boiling for 2 min. Cool and filter. To the filtrate, add 1.5 mL of *ferric chloride solution R1*; a blackish-blue colour develops.

B. It gives reaction (b) of bismuth (*2.3.1*).

TESTS
Solution S
In a porcelain or quartz dish, ignite 1.0 g, increasing the temperature very gradually. Heat in a muffle furnace at 600 ± 50 °C for 2 h. Cool and dissolve the residue with warming in 4 mL of a mixture of equal volumes of *lead-free nitric acid R* and *water R* and dilute to 20 mL with *water R*.

Acidity
Shake 1.0 g with 20 mL of *water R* for 1 min and filter. To the filtrate add 0.1 mL of *methyl red solution R*. Not more

than 0.15 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to yellow.

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 0.5 g add 10 mL of *dilute nitric acid R*. Heat on a water-bath for 5 min and filter. Dilute 5 mL of the filtrate to 15 mL with *water R*.

Nitrates
Maximum 0.2 per cent.

To 1.0 g add 25 mL of *water R* then 25 mL of a mixture of 2 volumes of *sulfuric acid R* and 9 volumes of *water R*. Heat at about 50 °C for 1 min with stirring and filter. To 10 mL of the filtrate, carefully add 30 mL of *sulfuric acid R*.
The solution is not more intensely brownish-yellow than a reference solution prepared at the same time as follows: to 0.4 g of *gallic acid R*, add 20 mL of *nitrate standard solution (100 ppm NO_3) R* and 30 mL of a mixture of 2 volumes of *sulfuric acid R* and 9 volumes of *water R*, then filter; to 10 mL of the filtrate, carefully add 30 mL of *sulfuric acid R*.

Copper
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *copper standard solution (10 ppm Cu) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Copper hollow-cathode lamp.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Lead
Maximum 20 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm (depending on the apparatus, the line at 217.0 nm may be used).

Atomisation device Air-acetylene flame.

Silver
Maximum 25 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *silver standard solution (5 ppm Ag) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Silver hollow-cathode lamp.

Wavelength 328.1 nm.

Atomisation device Air-acetylene flame.

Substances not precipitated by ammonia
Maximum 1.0 per cent.

In a porcelain or quartz dish, ignite 2.0 g, increasing the temperature very gradually to 600 ± 50 °C; allow to cool. Moisten the residue with 2 mL of *nitric acid R*, evaporate to dryness on a water-bath and carefully heat and ignite once more at 600 ± 50 °C. After cooling, dissolve the residue in 5 mL of *nitric acid R* and dilute to 20 mL with *water R*.
To 10 mL of this solution, add *concentrated ammonia R* until alkaline and filter. Wash the residue with *water R* and evaporate the combined filtrate and washings to dryness on a water-bath. Add 0.3 mL of *dilute sulfuric acid R* and ignite. The residue weighs a maximum of 10 mg.

Loss on drying (*2.2.32*)
Maximum 7.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY
To 0.300 g add 10 mL of a mixture of equal volumes of *nitric acid R* and *water R*, heat to boiling and maintain boiling for 2 min. Add 0.1 g of *potassium chlorate R*, heat to boiling and maintain boiling for 1 min. Add 10 mL of *water R* and heat until the solution becomes colourless. To the hot solution, add 200 mL of *water R* and 50 mg of *xylenol orange triturate R*. Titrate with *0.1 M sodium edetate* until a yellow colour is obtained.

1 mL of *0.1 M sodium edetate* is equivalent to 20.90 mg of Bi.

STORAGE
Protected from light.

Ph Eur

Heavy Bismuth Subnitrate

(Ph. Eur. monograph 1494)

$4[BiNO_3(OH)_2],BiO(OH)$ 1462 *1304-85-4*

Ph Eur

DEFINITION
Content
71.0 per cent to 74.0 per cent of Bi (A_r 209.0) (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in mineral acids with decomposition.

IDENTIFICATION
A. Dilute 1 mL of solution S1 (see Tests) to 5 mL with *water R* and add 0.3 mL of *potassium iodide solution R*.
A black precipitate is formed which dissolves into an orange solution with the addition of 2 mL of *potassium iodide solution R*.

B. It gives reaction (b) of bismuth (*2.3.1*).

C. It gives the reaction of nitrates (*2.3.1*).

D. pH (*2.2.3*): maximum 2.0 for solution S2 (see Tests).

TESTS
Solution S1
Shake 5.0 g by gently heating in 10 mL of *water R* and add 20 mL of *nitric acid R*. Heat until dissolution, cool and dilute to 100 mL with *water R*.

Solution S2
Place 1.00 g in a 20 mL volumetric flask and add 2.0 mL of *lead-free nitric acid R*. Allow acid attack to take place without heating and if necessary warm slightly at the end to completely dissolve the test sample. Add 10 mL of *water R*, shake and add, in small fractions, 4.5 mL of lead-free ammonia R; shake and allow to cool. Dilute to 20.0 mL with *water R*, shake again and allow the solids to settle. The clear supernatant solution is solution S2.

Acidity
Suspend 1.0 g in 15 mL of *water R* and shake several times. Allow to stand for 5 min and filter. To 10 mL of the filtrate, add 0.5 mL of *phenolphthalein solution R1*. Not more than 0.5 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 5.0 mL of solution S1, add 3 mL of *nitric acid R* and dilute to 15 mL with *water R*.

Copper
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S2.

Reference solutions Prepare the reference solutions using *copper standard solution (10 ppm Cu) R* and diluting with a 37 per cent *V/V* solution of *lead-free nitric acid R*.

Source Copper hollow-cathode lamp.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Lead
Maximum 20 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S2.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R* and diluting with a 37 per cent *V/V* solution of *lead-free nitric acid R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm (depending on the apparatus, the line at 217.0 nm may be used).

Atomisation device Air-acetylene flame.

Silver
Maximum 25 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S2.

Reference solutions Prepare the reference solutions using *silver standard solution (5 ppm Ag) R* and diluting with a 37 per cent *V/V* solution of *lead-free nitric acid R*.

Source Silver hollow-cathode lamp.

Wavelength 328.1 nm.

Atomisation device Air-acetylene flame.

Substances not precipitated by ammonia
Maximum 1.0 per cent.

To 20 mL of solution S1, add *concentrated ammonia R* until an alkaline reaction is produced and filter. Wash the residue with *water R*, and evaporate the combined filtrate and washings to dryness on a water-bath. To the residue, add 0.3 mL of *dilute sulfuric acid R* and ignite. The residue weighs a maximum of 10 mg.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
Dissolve with heating 0.250 g in 10 mL of a mixture of 2 volumes of *perchloric acid R* and 5 volumes of *water R*. To the hot solution, add 200 mL of *water R* and 50 mg of *xylenol orange triturate R*. Titrate with *0.1 M sodium edetate* until a yellow colour is obtained.

1 mL of *0.1 M sodium edetate* is equivalent to 20.90 mg of Bi.

Ph Eur

Bismuth Subsalicylate

(Ph. Eur. monograph 1495)

$C_7H_5BiO_4$ 362.1 *14882-18-9*

Ph Eur

DEFINITION
Complex of bismuth and salicylic acid.

Content
56.0 per cent to 59.4 per cent of Bi (A_r 209.0) (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water and in alcohol. It dissolves in mineral acids with decomposition.

IDENTIFICATION
A. To 0.5 g add 10 mL of *hydrochloric acid R1*. Heat on a boiling water-bath for 5 min. Cool and filter. Retain the filtrate for identification test B. Wash the residue with *dilute hydrochloric acid R* and then with *water R*. Dissolve the residue in 0.5-1 mL of *dilute sodium hydroxide solution R*. Add 15 mL of *water R*. Neutralise with *dilute hydrochloric acid R*. The solution gives reaction (a) of salicylates (*2.3.1*).

B. The filtrate obtained in identification test A gives reaction (b) of bismuth (*2.3.1*).

TESTS

Solution S
In a porcelain or quartz dish, ignite 1.0 g, increasing the temperature very gradually. Heat in a muffle furnace at 600 ± 25 °C for 2 h. Cool and dissolve the residue with warming in 4 mL of a mixture of equal volumes of *lead-free nitric acid R* and *water R* and dilute to 20 mL with *water R*.

Acidity
Shake 2.0 g with 30 mL of *ether R* for 1 min and filter. To the filtrate add 30 mL of *alcohol R* and 0.1 mL of *thymol blue solution R*. Not more than 0.35 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to blue.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dissolve 0.250 g in a mixture of 2 mL of *nitric acid R*, 5 mL of *water R* and 8 mL of *methanol R*.

Nitrates
Maximum 0.4 per cent.

To 0.1 g add 10 mL of *water R* and, with caution, 20 mL of *sulfuric acid R* and stir. The solution is not more intensely yellow coloured than a reference solution prepared at the same time using 0.1 g of *salicylic acid R*, 6 mL of *water R*, 4 mL of *nitrate standard solution (100 ppm NO_3) R* and 20 mL of *sulfuric acid R*.

Copper
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *copper standard solution (10 ppm Cu) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Copper hollow-cathode lamp.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Lead
Maximum 20 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm (depending on the apparatus, the line at 217.0 nm may be used).

Atomisation device Air-acetylene flame.

Silver
Maximum 25 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *silver standard solution (5 ppm Ag) R* and diluting with a 6.5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Silver hollow-cathode lamp.

Wavelength 328.1 nm.

Atomisation device Air-acetylene flame.

Soluble bismuth
Maximum 40 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Suspend 5.0 g in 100 mL of *water R*. Stir constantly for 2 h at 20-23 °C. Filter through filter paper (slow filtration) then through a cellulose micropore membrane filter (0.1 µm). To 10.0 mL of clear filtrate, add 0.1 mL of *nitric acid R*.

Reference solutions Prepare the reference solutions using *bismuth standard solution (100 ppm Bi) R* and diluting with a mixture of equal volumes of *dilute nitric acid R* and *water R*.

Source Bismuth hollow-cathode lamp.

Wavelength 223.06 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve with heating 0.300 g in 10 mL of a mixture of 2 volumes of *perchloric acid R* and 5 volumes of *water R*. To the hot solution, add 200 mL of *water R* and 50 mg of *xylenol orange triturate R*. Titrate with *0.1 M sodium edetate* until a yellow colour is obtained.

1 mL of *0.1 M sodium edetate* is equivalent to 20.90 mg of Bi.

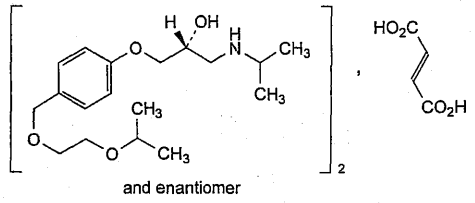
STORAGE

Protected from light.

Ph Eur

Bisoprolol Fumarate

(Ph. Eur. monograph 1710)



and enantiomer

$C_{40}H_{66}N_2O_{12}$ 767 *104344-23-2*

Action and use
Beta-adrenoceptor antagonist.

Preparation
Bisoprolol Tablets

Ph Eur

DEFINITION

(2*RS*)-1-[4-[[2-(1-Methylethoxy)ethoxy]methyl]phenoxy]-3-[(1-methylethyl)amino]propan-2-ol fumarate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, slightly hygroscopic powder.

Solubility
Very soluble in water, freely soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bisoprolol fumarate CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate and dry the residues at 60 °C at a pressure not exceeding 0.7 kPa and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water for chromatography R* (20:80 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *bisoprolol for peak identification CRS* (containing impurities A and E) in 1.0 mL of the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *bisoprolol for system suitability CRS* (containing impurity G) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 20 ± 2 °C.

Mobile phase:
— *mobile phase A*: 10 g/L solution of *phosphoric acid R*;
— *mobile phase B*: 10 g/L solution of *phosphoric acid R* in *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	95	5
4 - 8	95 → 80	5 → 20
8 - 15	80	20
15 - 34	80 → 20	20 → 80
34 - 36	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *bisoprolol for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to fumaric acid and impurities A and E; use the chromatogram supplied with *bisoprolol for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.

Relative retention With reference to bisoprolol (retention time = about 18 min): impurity A = about 0.5; impurity G = about 1.1; impurity E = about 1.2.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to bisoprolol.

Limits:
— *impurity G*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity E*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to fumaric acid.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 38.35 mg of $C_{40}H_{66}N_2O_{12}$.

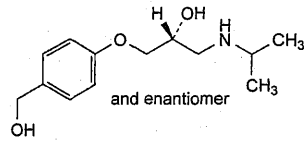
STORAGE

In an airtight container, protected from light.

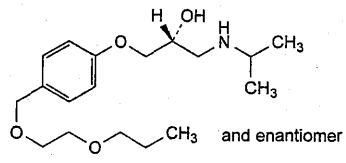
IMPURITIES

Specified impurities *A, E, G.*

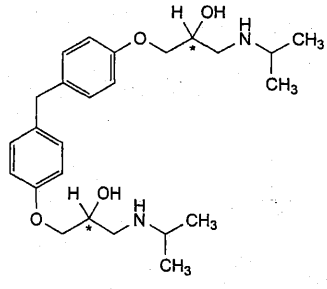
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, F, K, L, N, Q, R, S, T, U.*



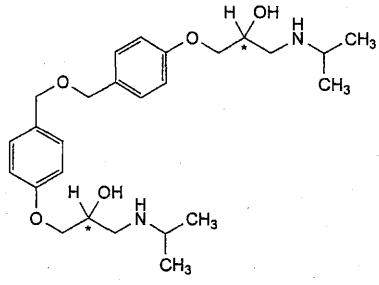
A. (2*RS*)-1-(4-hydroxymethyl-phenoxy)-3-isopropylaminopropan-2-ol,



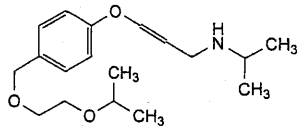
B. (2*RS*)-1-isopropylamino-3-[4-(2-propoxy-ethoxymethyl)phenoxy]propan-2-ol,



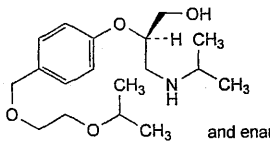
C. 1-[4-[4-(2-hydroxy-3-isopropylamino-propoxy)benzyl] phenoxy]-3-isopropylaminopropan-2-ol,



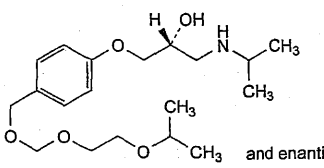
D. 1-[4-[4-(2-hydroxy-3-isopropylaminopropoxy) benzyloxylmethyl]phenoxy]-3-isopropylaminopropan-2-ol,



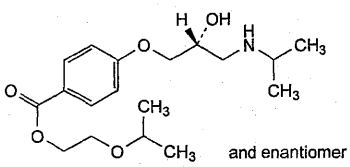
E. (*EZ*)-[3-[4-(2-isopropoxy-ethoxymethyl)phenoxy] allyl]isopropylamine,


and enantiomer

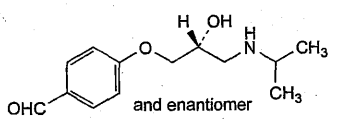
F. (2*RS*)-2-[4-(2-isopropoxy-ethoxymethyl)phenoxy]-3-isopropylaminopropan-2-ol,


and enantiomer

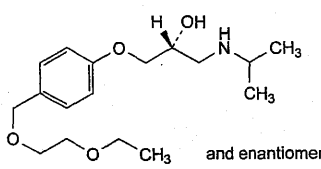
G. (2*RS*)-1-[4-[[(2-isopropoxyethoxy)methoxy]methyl]phenoxy]-3-isopropylaminopropan-2-ol,


and enantiomer

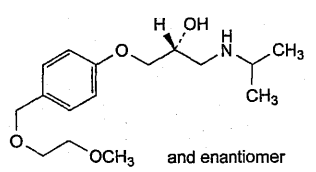
K. 2-isopropoxyethyl 4-[[(2*RS*)-2-hydroxy-3-(isopropylamino)propyl]oxy]benzoate,


and enantiomer

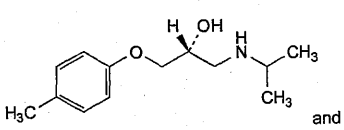
L. 4-[[(2*RS*)-2-hydroxy-3-(isopropylamino)propyl]oxy]benzaldehyde,


and enantiomer

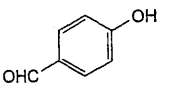
N. (2*RS*)-1-[4-[(2-ethoxyethoxy)methyl]phenoxy]-3-isopropylaminopropan-2-ol,


and enantiomer

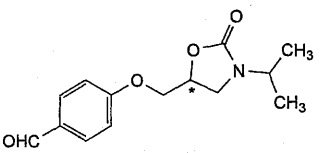
Q. (2*RS*)-1-(isopropylamino)-3-[4-(2-methoxyethoxy)methyl]phenoxypropan-2-ol,


and enantiomer

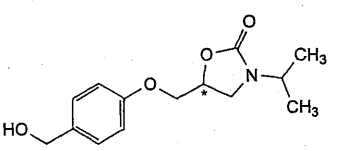
R. (2*RS*)-1-(isopropylamino)-3-(4-methylphenoxy)propan-2-ol,



S. 4-hydroxybenzaldehyde,



T. 4-[(3-isopropyl-2-oxo-1,3-oxazolidin-5-yl)methoxy]benzaldehyde,



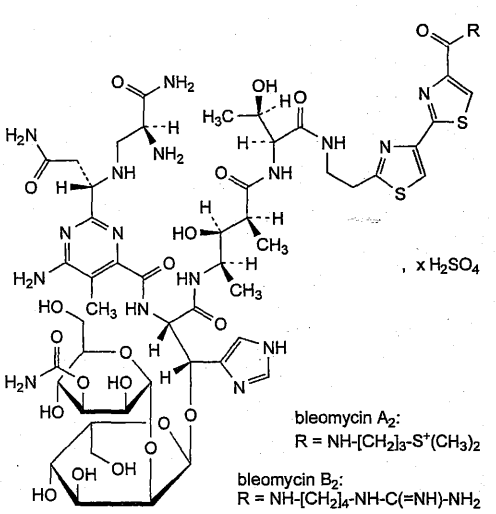
U. 5-[[4-(hydroxymethyl)phenoxy]methyl]-3-isopropyl-1,3-oxazolidin-2-one.

Ph Eur

Bleomycin Sulfate

Bleomycin Sulphate

(Ph. Eur. monograph 0976)



9041-93-4

Action and use
Cytotoxic antibacterial.

Preparation
Bleomycin Injection

Ph Eur

DEFINITION
Sulfate of a mixture of glycopeptides produced by *Streptomyces verticillus* or by any other means; the 2 principal components of the mixture are *N*-[3-(dimethylsulfonio)propyl]bleomycinamide (bleomycin A_2) and *N*-[4-(carbamimidoylamino)butyl]bleomycinamide (bleomycin B_2).

Potency
Minimum 1500 IU/mg (dried substance).

CHARACTERS

Appearance
White or yellowish-white, very hygroscopic powder.

Solubility
Very soluble in water, slightly soluble in anhydrous ethanol, practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for composition.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the 2 principal peaks in the chromatogram obtained with reference solution (a).

B. It gives the reactions of sulfates (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.10.

Dissolve 0.200 g in *water R* and dilute to 10.0 mL with the same solvent.

pH (*2.2.3*)
4.5 to 6.0.

Dissolve 50 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Composition
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of *bleomycin sulfate CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.5 mL of reference solution (a) to 100.0 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase:
— *mobile phase A*: *methanol R*;
— *mobile phase B*: dissolve 0.960 g of *sodium pentanesulfonate R* in 900 mL of acetic acid (4.8 g/L $C_2H_4O_2$), add 1.86 g of *sodium edetate R*, dilute to 1000 mL with the same solvent and adjust to pH 4.3 with *ammonia R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	10 → 40	90 → 60
60 - end	40	60

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time Until impurity D is eluted (about 80 min).

Relative retention With reference to bleomycin A_2: impurity D = 1.5 to 2.5.

System suitability:
— *resolution*: minimum 5 between the peaks due to bleomycin A_2 (1st principal peak) and bleomycin B_2 (2nd principal peak) in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 20 for the principal peak in the chromatogram obtained with reference solution (b);
— *repeatability*: maximum relative standard deviation of 2 per cent for the principal peak after 6 injections of reference solution (a).

Limits:
— *bleomycin A_2*: 55 per cent to 70 per cent;
— *bleomycin B_2*: 25 per cent to 32 per cent;
— *sum of bleomycin A_2 and B_2*: minimum 85 per cent;
— *impurity D*: maximum 5.5 per cent;
— *sum of impurities other than D*: maximum 9.5 per cent;
— *disregard limit*: 0.1 per cent of the total.

Copper
Maximum 200 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 50 mg in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution Dilute 1.0 mL of *copper standard solution (10 ppm Cu) R* to 10.0 mL with *water R*.

Source Copper hollow-cathode lamp.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 50 mg by drying at 60 °C at a pressure not exceeding 0.67 kPa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Carry out the microbiological assay of antibiotics (*2.7.2*), using the diffusion method. Use *bleomycin sulfate CRS* as the chemical reference substance.

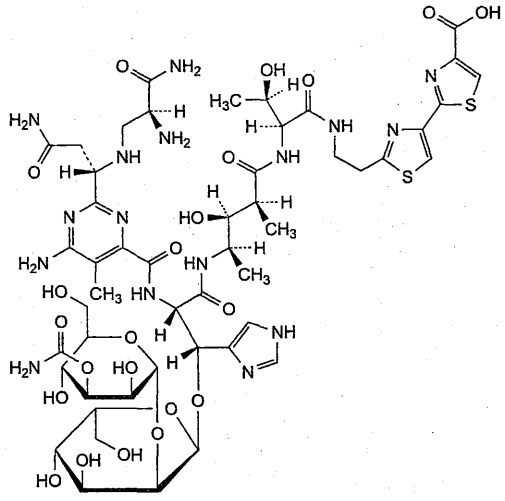
STORAGE

In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

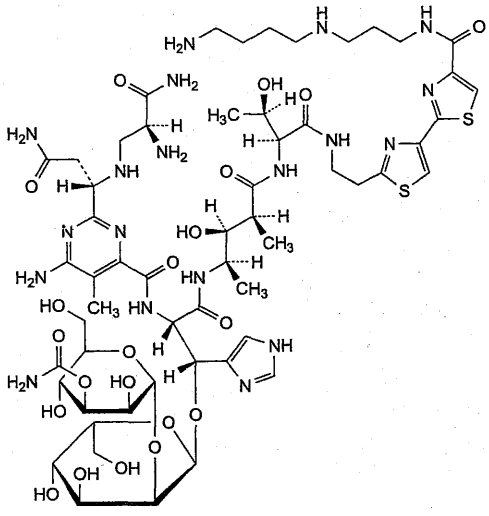
IMPURITIES

Specified impurities *D.*

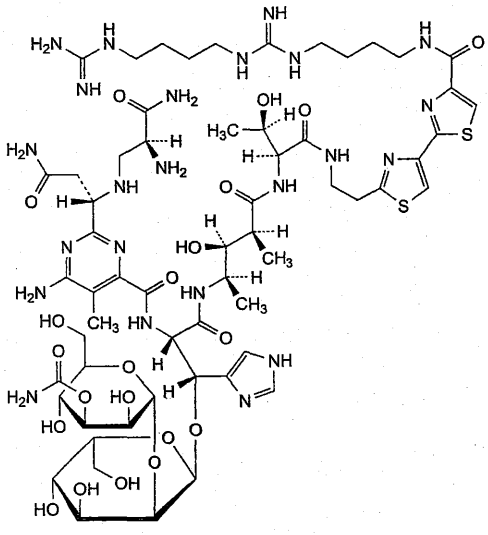
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



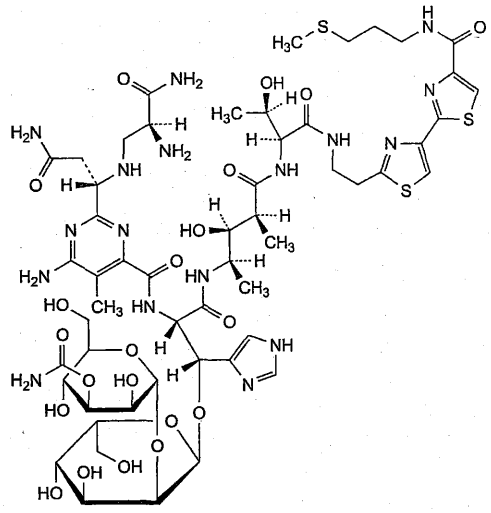
A. bleomycinic acid,



B. bleomycin A_5,



C. bleomycin B_4,

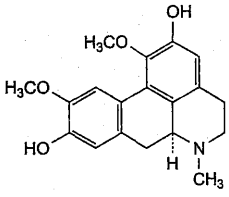


D. demethylbleomycin A_2.

Ph Eur

Boldine

(Ph. Eur. monograph 2971)



$C_{19}H_{21}NO_4$ 327.4 *476-70-0*

Action and use
Antioxidant.

Ph Eur

DEFINITION

(6a*S*)-1,10-Dimethoxy-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline-2,9-diol, of vegetable origin.

Content
98.5 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white or yellowish, crystalline powder.

Solubility
Very slightly soluble in water, soluble in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute acid solutions.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *boldine CRS*.

TESTS

Specific optical rotation (*2.2.7*)
+ 121.0 to + 127.0 (anhydrous substance).
Dissolve 0.500 g in *ethanol (96 per cent) R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 12.0 mg of the substance to be examined in 8 mL of *methanol R* using sonication and dilute to 10.0 mL with *methanol R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (b) Dissolve 12 mg of *boldine for system suitability CRS* (containing impurities C and E) in 8 mL of *methanol R* using sonication and dilute to 10 mL with *methanol R*.

Reference solution (c) In order to prepare impurity A *in situ*, add 0.5 mL of *strong hydrogen peroxide solution R* to 5 mL of reference solution (b) and stir for 1 h.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped extra-dense bonded octylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: dissolve 1.54 g of *ammonium acetate R* in 950 mL of *water for chromatography R*, adjust to pH 8.50 ± 0.05 with *diethylamine R* and dilute to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	85	15
2 - 38	85 → 64	15 → 36
38 - 50	64 → 20	36 → 80
50 - 53	20	80

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 302 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *boldine for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, C and E.

Relative retention With reference to boldine (retention time = about 23 min): impurity A = about 0.25; impurity C = about 0.6; impurity E = about 1.6.

System suitability Reference solution (b):
- *resolution*: minimum 10.0 between the peaks due to impurity C and boldine.

Calculation of percentage contents:
- for each impurity, use the concentration of boldine in reference solution (a).

Limits:
- *impurity C*: maximum 1.0 per cent;
- *impurity A*: maximum 0.3 per cent;
- *impurity E*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 1.5 per cent;
- *reporting threshold*: 0.05 per cent.

2-Propanol (*2.4.24*)
Maximum 1.0 per cent.

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 0.100 g using the evaporation technique at 120 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

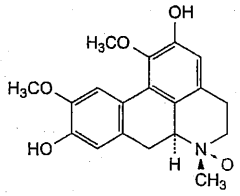
Dissolve 0.200 g in 50 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 32.74 mg of $C_{19}H_{21}NO_4$.

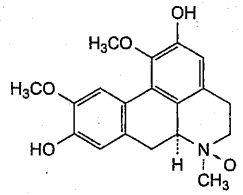
IMPURITIES

Specified impurities *A, C, E.*

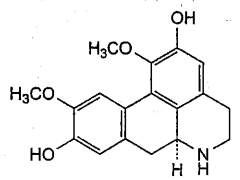
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D.*



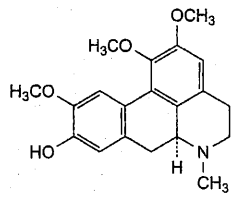
A. (6*S*,6a*S*)-2,9-dihydroxy-1,10-dimethoxy-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline *N*-oxide,



B. (6*R*,6a*S*)-2,9-dihydroxy-1,10-dimethoxy-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline *N*-oxide,



C. (6a*S*)-1,10-dimethoxy-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinoline-2,9-diol,



D. (6a*S*)-1,2,10-trimethoxy-6-methyl-5,6,6a,7-tetrahydro-4*H*-dibenzo[*de*,*g*]quinolin-9-ol,

E. unknown structure.

Refined Borage Oil

Refined Starflower Oil

(Refined Borage (Starflower) Oil, Ph. Eur. monograph 2105)

Ph Eur

DEFINITION

Fatty oil obtained from seeds of *Borago officinalis* L. by extraction and/or expression. It is then refined. A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, light yellow or yellow liquid.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum.

Relative density

About 0.921.

Refractive index

About 1.476.

IDENTIFICATION

First identification: B.

Second identification: A.

A. Identification of fatty oils by thin-layer chromatography (*2.3.2*).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

B. Composition of fatty acids (see Tests).

TESTS

Acid value (*2.5.1*)

Maximum 0.5, or maximum 0.3 if intended for use in the manufacture of parenteral preparations.

Peroxide value (*2.5.5, Method A*)

Maximum 10.0, or maximum 5.0 if intended for use in the manufacture of parenteral preparations.

Unsaponifiable matter (*2.5.7*)

Maximum 2.0 per cent, determined on 5.0 g.

Alkaline impurities (*2.4.19*)

It complies with the test.

Composition of fatty acids (*2.4.22, Method A*)

Use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty-acid fraction of the oil:

— *saturated fatty acids of chain length less than C_{16}*: maximum 0.3 per cent,
— *palmitic acid*: 9.0 per cent to 12.0 per cent,
— *palmitoleic acid*: maximum 0.6 per cent,
— *stearic acid*: 2.0 per cent to 6.0 per cent,
— *oleic acid*: 12.0 per cent to 22.0 per cent,
— *linoleic acid*: 30.0 per cent to 41.0 per cent,
— *gamma-linolenic acid*: 17.0 per cent to 27.0 per cent,
— *alpha-linolenic acid*: maximum 0.5 per cent,
— *arachidic acid*: maximum 0.5 per cent,
— *eicosenoic acid*: 2.8 per cent to 4.4 per cent,
— *erucic acid*: maximum 3.0 per cent,
— *nervonic acid*: maximum 4.5 per cent.

Brassicasterol (*2.4.23*)

Maximum 0.3 per cent in the sterol fraction of the oil.

Water (*2.5.32*)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

Under an inert gas, in a well-filled, airtight container, protected from light.

LABELLING

The label states, where applicable, that the oil is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Borax

Sodium Borate

Sodium Tetraborate

(Ph. Eur. monograph 0013)

$Na_2B_4O_7,10H_2O$ 381.4 *1303-96-4*

Ph Eur

DEFINITION

Disodium tetraborate decahydrate.

Content

99.0 per cent to 103.0 per cent of $Na_2B_4O_7,10H_2O$.

CHARACTERS

Appearance

White or almost white, crystalline powder, colourless crystals or crystalline masses, efflorescent.

Solubility

Soluble in water, very soluble in boiling water, freely soluble in glycerol.

IDENTIFICATION

A. To 1 mL of solution S (see Tests) add 0.1 mL of *sulfuric acid R* and 5 mL of *methanol R* and ignite. The flame has a green border.

B. To 5 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution is red. On the addition of 5 mL of *glycerol (85 per cent) R* the colour disappears.

C. Solution S gives the reactions of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 4.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

9.0 to 9.6 for solution S.

Sulfates (*2.4.13*)

Maximum 50 ppm, determined on solution S.

Use in this test 1.0 mL of *acetic acid R*. Prepare the standard using a mixture of 3 mL of *sulfate standard solution (10 ppm SO_4) R* and 12 mL of *distilled water R*.

Ammonium (*2.4.1*)

Maximum 10 ppm.

Dilute 6 mL of solution S to 14 mL with *water R*. Prepare the standard using a mixture of 2.5 mL of *ammonium standard solution (1 ppm NH_4) R* and 7.5 mL of *water R*.

Arsenic (*2.4.2, Method A*)

Maximum 5 ppm, determined on 5 mL of solution S.

Calcium (*2.4.3*)

Maximum 100 ppm, determined on solution S.

Prepare the standard using a mixture of 6 mL of *calcium standard solution (10 ppm Ca) R* and 9 mL of *distilled water R*.

ASSAY

Dissolve 20 g of *mannitol R* in 100 mL of *water R*, heating if necessary, cool and add 0.5 mL of *phenolphthalein solution R* and neutralise with *0.1 M sodium hydroxide* until a pink colour is obtained. Add 3.00 g of the substance to be examined, heat until dissolution is complete, cool, and titrate with *1 M sodium hydroxide* until the pink colour reappears.

1 mL of *1 M sodium hydroxide* is equivalent to 0.1907 g of $Na_2B_4O_7,10H_2O$.

Ph Eur

Boric Acid

(Ph. Eur. monograph 0001)

H_3BO_3 61.8 *10043-35-3*

Ph Eur

DEFINITION

Content

99.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder, colourless, shiny plates greasy to the touch, or white or almost white crystals.

Solubility

Soluble in water and in ethanol (96 per cent), freely soluble in boiling water and in glycerol (85 per cent).

IDENTIFICATION

A. Dissolve 0.1 g by gently heating in 5 mL of *methanol R*, add 0.1 mL of *sulfuric acid R* and ignite the solution. The flame has a green border.

B. Solution S (see Tests) is acid (*2.2.4*).

TESTS

Solution S

Dissolve 3.3 g in 80 mL of boiling *distilled water R*, cool and dilute to 100 mL with *carbon dioxide-free water R* prepared from *distilled water R*.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

3.8 to 4.8 for solution S.

Solubility in ethanol (96 per cent)

The solution is not more opalescent than reference suspension II (*2.2.1*) and is colourless (*2.2.2, Method II*).

Dissolve 1.0 g in 10 mL of boiling *ethanol (96 per cent) R*.

Organic matter

It does not darken on progressive heating to dull redness.

Sulfates (*2.4.13*)

Maximum 450 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

ASSAY

Dissolve 1.000 g with heating in 100 mL of *water R* containing 15 g of *mannitol R*. Titrate with *1 M sodium hydroxide*, using 0.5 mL of *phenolphthalein solution R* as indicator, until a pink colour is obtained.

1 mL of *1 M sodium hydroxide* is equivalent to 61.8 mg of H_3BO_3.

Ph Eur

Botulinum Toxin Type A for Injection

(Ph. Eur. monograph 2113)

Ph Eur

DEFINITION

Botulinum toxin type A for injection is a dried preparation containing purified botulinum neurotoxin type A, which may be present in the form of a complex with haemagglutinins and non-toxic proteins. Botulinum neurotoxin type A or its haemagglutinin complex is prepared by a suitable purification process of the liquid supernatant from a broth-culture of a suitable strain of *Clostridium botulinum* type A.

The purified complexes consist of several proteins and can be of various sizes. The largest complex (relative molecular mass of about 900 000) consists of a 150 000 relative molecular mass neurotoxin, a 130 000 relative molecular mass non-toxic protein and various haemagglutinins ranging between relative molecular mass 14 000 and 43 000. The purified toxin moiety is composed of only the same 150 000 relative molecular mass neurotoxin as is found in the 900 000 relative molecular mass neurotoxin complex, which is initially produced as a single chain and further cleaved (nicked) by endogenous proteases into a fully active, disulfide-linked, 54 000 relative molecular mass light chain and a 97 000 relative molecular mass heavy chain.

The preparation is reconstituted before use, as stated on the label.

PRODUCTION

GENERAL PROVISIONS

Production of the toxin is based on seed cultures, managed in a defined seed-lot system in which the ability to produce toxin is conserved. The production method must be shown to yield consistently product of activity and profile comparable to that of lots shown in clinical studies to be of adequate safety and efficacy.

The production method and stability of the finished product and relevant intermediates are evaluated using the tests below. Such tests include the specific toxin activity per milligram of protein of purified toxin in an appropriate functional model of toxin activity and may be supported by tests confirming the presence of botulinum toxin type A, and, if appropriate, associated non-toxic proteins.

BACTERIAL SEED LOTS

A highly toxigenic strain of *C. botulinum* of known toxin type A and confirmed absence of genes encoding other botulinum toxins (particularly botulinum toxin types B and F), with known origin and history, is grown using suitable media. The bacterial strain, used for the master seed lot, shall be identified by historical records that include information on its origin and the tests used to characterise the strain. These will include morphological, cultural, biochemical, genetic and serological properties of the strain. The master seed lot and the working seed lot, where applicable, must be demonstrated to have identical profiles. Only a seed lot that complies with the following requirements may be used.

Identification

Each seed lot is identified as containing pure cultures of *C. botulinum* type A bacteria with no extraneous bacterial or fungal contamination.

Microbial purity
Each seed lot complies with the requirements for absence of contaminating micro-organisms. The purity of bacterial cultures is verified by methods of suitable sensitivity. These may include inoculation into suitable media and examination of colony morphology.

Phenotypic parameters
Each seed lot must have a known fatty acid profile, sugar fermentation profile (glucose, lactose, mannose, etc.) and proteolytic activity and must demonstrate relevant lipase, lecithinase and gelatinase activity.

Genetic purity
Each seed lot must have information on the toxin gene sequence and comply with requirements for the absence of other genes encoding other toxin serotypes.

Production of active toxin
A bacterial strain producing a high yield of active toxin, as determined by an acute toxicity assay, is suitable. Seed lots demonstrate a capability of producing at least a minimum toxicity level appropriate for the manufacturing process and scale.

MANUFACTURER'S REFERENCE PREPARATIONS
During development, reference preparations are established for subsequent verification of batch consistency during production and for control of the bulk purified toxin and finished product. They are derived from representative batches of botulinum toxin type A that are characterised as described under Bulk Purified Toxin.

The reference preparations are suitably characterised for their intended purpose and are stored in suitably sized aliquots under conditions ensuring their suitability.

BULK PURIFIED TOXIN
C. botulinum Type A strain is grown anaerobically, in suitable media, from which cultures are selected for step-up incubations under a suitably controlled anaerobic atmosphere through the seed culture and bulk fermentation stages to allow maximum production of toxin. The toxin is purified by suitable methods to remove nucleic acids and components likely to cause adverse reactions.

Only a purified toxin that complies with the following requirements may be used in the preparation of the final bulk. For each test and for each product, limits of acceptance are established and each new purified toxin must comply with these limits.

Residual reagents
Removal of residual reagents used in purification steps is confirmed by suitable limit tests or by validation of the process.

Nucleic acids
Removal of nucleic acids is confirmed by suitable limit tests or by validation of the process.

Immunological identity
The presence of specific type A toxin is confirmed by a suitable immunochemical method (*2.7.1*).

Specific activity
The specific activity is confirmed in a mouse model of toxicity or by *in vivo*/*ex vivo* methods validated with respect to the LD_{50} assay and expressed in mouse LD_{50} units per milligram of protein. Specific activity must not be less than 1×10^8 mouse LD_{50} units per milligram of protein for the 150 000 relative molecular mass neurotoxin and must not be less than 1×10^7 mouse LD_{50} units per milligram of protein for the 900 000 relative molecular mass neurotoxin complex.

Protein
The total protein concentration is determined by a suitable method. An acceptable value is established for the product and each batch must be shown to comply with the limits.

Protein profile
Identity and protein composition are determined by polyacrylamide gel electrophoresis (*2.2.31*) under reducing or non-reducing conditions or by other suitable physicochemical methods such as size-exclusion chromatography (*2.2.30*), comparing with suitable reference standards.

Total viable count
It complies with the limits approved for the particular product.

FINAL BULK
The final bulk is prepared by adding approved excipients to the bulk purified toxin. The solution is filtered through a bacteria-retentive filter. If human albumin is added, it complies with the monograph *Human albumin solution (0255)*.

FINAL LOT
The final bulk is distributed aseptically into sterile, tamper-proof containers. Uniformity of fill is verified during filling and the test for uniformity of content (*2.9.6*) is not required. The containers are closed so as to prevent contamination.

Only a final lot that is within the limits approved for the particular product and is satisfactory with respect to each of the requirements given below under Identification, Tests and Assay may be released for use.

pH (*2.2.3*)
The pH of the reconstituted product is within ± 0.5 pH units of the limit approved for the particular product.

Water
Not more than the limit approved for the particular product.

IDENTIFICATION
The presence of botulinum toxin type A is confirmed by a suitable immunochemical method (*2.7.1*).

TESTS
Sterility (*2.6.1*)
It complies with the test for sterility.

Bacterial endotoxins (*2.6.14*)
Less than 10 IU per vial.

ASSAY
In accordance with the provisions of the European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes, tests must be carried out in such a way as to use the minimum number of animals and to cause the least pain, suffering, distress or lasting harm. The LD_{50} assay is associated with severe suffering of animals and manufacturers are strongly encouraged to develop and validate assays that will reduce the number of animals used, or refine or replace the test procedure with the goal of promoting animal welfare.

The potency of the reconstituted product is determined by an LD_{50} assay in mice or by a method validated with respect to the LD_{50} assay. The potency is expressed in terms of the LD_{50} for mice or relative to the reference preparation.

For determination of the LD_{50}, graded doses of the product are injected intraperitoneally into groups of mice and the LD_{50} is calculated by the usual statistical methods (*5.3*) from the mouse lethality in each group. A suitable reference preparation is assayed in parallel; the potency of the toxin is expressed relative to the reference or the value found for the

reference is within suitable limits defined in terms of the assigned potency.

After validation with respect to the LD_{50} assay (reference method), the product may also be assayed by other methods that are preferable in terms of animal welfare, for example mouse bioassays using paralysis as the end-point, *ex vivo* assays using mouse phrenic nerve diaphragm, endopeptidase assays *in vitro* and cell-based assays.

For alternative replacement methods the potency is calculated with respect to a suitable reference preparation calibrated in mouse LD_{50} units.

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits ($P = 0.95$) are not less than 80 per cent and not more than 125 per cent of the estimated potency.

The test may be repeated but when more than 1 test is performed, the results of all valid tests must be combined in the estimate of potency.

LABELLING

The label states:
— the number of units of toxin per vial with a statement that units are product specific and not applicable to other preparations containing botulinum toxin type A;
— the name and the volume of the diluent to be added for reconstitution of the dried product.

Ph Eur

Botulinum Toxin Type B for Injection

(Ph. Eur. monograph 2581)

Ph Eur

DEFINITION

Botulinum toxin type B for injection is a liquid preparation containing purified botulinum neurotoxin type B, which may be present in the form of a complex with haemagglutinins and non-toxic proteins. Botulinum neurotoxin type B or its haemagglutinin complex is prepared by a suitable purification process of the liquid supernatant from a broth-culture of a suitable strain of *Clostridium botulinum* type B. Suitable stabilisers may be added.

The toxin is present in its native form as a complex of neurotoxin and non-toxin proteins and haemagglutinins with a total relative molecular mass of approximately 700 000. The neurotoxin is synthesised by the bacterium as a single-chain polypeptide of approximately 150 000 relative molecular mass that is activated during the fermentation process via a proteolytic cleavage (nicking) by endogenous proteases. The nicked protein is a fully active double-chain polypeptide consisting of a heavy chain (100 000 relative molecular mass) and a light chain (50 000 relative molecular mass), connected by a disulfide bond.

PRODUCTION

GENERAL PROVISIONS

Production of the toxin is based on seed cultures, managed in a defined seed-lot system in which the ability to produce toxin is conserved. The production method must be shown to yield consistently product of activity and profile comparable to that of lots shown in clinical studies to be of adequate safety and efficacy.

The production method and stability of the finished product and relevant intermediates are evaluated using the tests below. Such tests include the specific toxin activity per milligram of protein of purified toxin in an appropriate functional model of toxin activity and may be supported by tests confirming the presence of botulinum toxin type B, and, if appropriate, associated non-toxic proteins.

BACTERIAL SEED LOTS

A highly toxigenic strain of *C. botulinum* of known toxin type B and confirmed absence of genes encoding other botulinum toxins (particularly botulinum toxin types A and F), with known origin and history, is grown using suitable media. The bacterial strain, used for the master seed lot, shall be identified by historical records that include information on its origin and the tests used to characterise the strain. These will include morphological, cultural, biochemical, genetic and serological properties of the strain. The master seed lot and the working seed lot, where applicable, must be demonstrated to have identical profiles. Only a seed lot that complies with the following requirements may be used.

Identification

Each seed lot is identified as containing pure cultures of *C. botulinum* type B bacteria with no extraneous bacterial or fungal contamination.

Microbial purity

Each seed lot complies with the requirements for absence of contaminating micro-organisms. The purity of bacterial cultures is verified by methods of suitable sensitivity. These may include inoculation into suitable media and examination of colony morphology.

Phenotypic parameters

Each seed lot must have a known fatty acid profile, sugar fermentation profile (glucose, lactose, mannose, etc.) and proteolytic activity and must demonstrate relevant lipase, lecithinase and gelatinase activity.

Genetic purity

Each seed lot must have information on the toxin gene genomic location and on the toxin gene sequence, and comply with requirements for the absence of other genes encoding other toxin serotypes.

Production of active toxin

A bacterial strain producing a high yield of active toxin, as determined by an acute toxicity assay, is suitable. Seed lots demonstrate a capability of producing at least a minimum toxicity level appropriate for the manufacturing process and scale.

MANUFACTURER'S REFERENCE PREPARATIONS

During development, reference preparations are established for subsequent verification of batch consistency during production and for control of the bulk purified toxin and finished product. They are derived from representative batches of botulinum toxin type B that are characterised as described under Bulk Purified Toxin.

The reference preparations are suitably characterised for their intended purpose and are stored in suitably sized aliquots under conditions ensuring their suitability.

BULK PURIFIED TOXIN

C. botulinum type B strain is grown anaerobically, in suitable media, from which cultures are selected for step-up incubations under a suitably controlled anaerobic atmosphere through the seed culture and bulk fermentation stages to allow maximum production of toxin. The toxin is purified by

suitable methods to remove nucleic acids and components likely to cause adverse reactions.

Only a purified toxin that complies with the following requirements may be used in the preparation of the final bulk. For each test and for each product, limits of acceptance are established and each new purified toxin must comply with these limits.

Residual reagents
Removal of residual reagents used in purification steps is confirmed by suitable limit tests or by validation of the process.

Nucleic acids
Removal of nucleic acids is confirmed by suitable limit tests or by validation of the process.

Immunological identity
The presence of specific type B toxin is confirmed by a suitable immunochemical method (*2.7.1*).

Specific activity
The specific activity is confirmed in a mouse model of toxicity or by *in vivo*/*ex vivo* methods validated with respect to the LD_{50} assay and expressed in mouse LD_{50} units per milligram of protein. Specific activity must not be less than 1×10^8 mouse LD_{50} units per milligram of protein.

Protein
The total protein concentration is determined by a suitable method. An acceptable value is established for the product and each batch must be shown to comply with the limits.

Protein profile
Identity and protein composition are determined by polyacrylamide gel electrophoresis (*2.2.31*) under reducing or non-reducing conditions or by other suitable physicochemical methods such as size-exclusion chromatography (*2.2.30*), comparing with suitable reference standards.

Total viable count
It complies with the limits approved for the particular product.

FINAL BULK
The final bulk is prepared by adding approved excipients to the bulk purified toxin. The solution is filtered through a bacteria-retentive filter. If human albumin is added, it complies with the monograph *Human albumin solution (0255)*.

FINAL LOT
The final bulk is distributed aseptically into sterile, tamper-proof containers. Uniformity of fill is verified during filling and the test for uniformity of content (*2.9.6*) is not required. The containers are closed so as to prevent contamination.

Only a final lot that is within the limits approved for the particular product and is satisfactory with respect to each of the requirements given below under Identification, Tests and Assay may be released for use.

pH (*2.2.3*)
The pH of the product is within ± 0.5 pH units of the limit approved for the particular product.

IDENTIFICATION
The presence of botulinum toxin type B is confirmed by a suitable immunochemical method (*2.7.1*).

TESTS
Sterility (*2.6.1*)
It complies with the test for sterility.

Bacterial endotoxins (*2.6.14*)
Less than 10 IU per vial.

ASSAY
In accordance with the provisions of the European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes, tests must be carried out in such a way as to use the minimum number of animals and to cause the least pain, suffering, distress or lasting harm. The LD_{50} assay is associated with severe suffering of animals and manufacturers are strongly encouraged to develop and validate assays that will reduce the number of animals used, or refine or replace the test procedure with the goal of promoting animal welfare.

The potency of the product is determined by an LD_{50} assay in mice or by a method validated with respect to the LD_{50} assay. The potency is expressed in terms of the LD_{50} for mice or relative to the reference preparation.

For determination of the LD_{50}, graded doses of the product are injected intraperitoneally into groups of mice and the LD_{50} is calculated by the usual statistical methods (*5.3*) from the mouse lethality in each group. A suitable reference preparation is assayed in parallel; the potency of the toxin is expressed relative to the reference or the value found for the reference is within suitable limits defined in terms of the assigned potency.

After validation with respect to the LD_{50} assay (reference method), the product may also be assayed by other methods that are preferable in terms of animal welfare, for example mouse bioassays using paralysis as the end-point, *ex vivo* assays using mouse phrenic nerve diaphragm, endopeptidase assays *in vitro* and cell-based assays.

For alternative replacement methods the potency is calculated with respect to a suitable reference preparation calibrated in mouse LD_{50} units.

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits ($P = 0.95$) are not less than 80 per cent and not more than 125 per cent of the estimated potency.

The test may be repeated but when more than 1 test is performed, the results of all valid tests must be combined in the estimate of potency.

LABELLING
The label states the number of units of toxin per vial with a statement that units are product specific and not applicable to other preparations containing botulinum toxin type B.

Ph Eur

Bovine Serum

(Ph. Eur. monograph 2262)

Ph Eur

DEFINITION
Liquid fraction of blood obtained from the ox (*Bos taurus* L.) and from which cells, fibrin and clotting factors have been removed.

Different types of bovine serum are used:
- *adult bovine serum* obtained at slaughter from cattle that are declared fit for human consumption;
- *calf serum* obtained at slaughter from animals, fit for human consumption, before the age of 12 months;
- *new-born calf serum* obtained at slaughter from animals before the age of 20 days;

— *foetal bovine serum* obtained from normal foetuses from dams fit for human consumption;
— *donor bovine serum* obtained by repeated bleeding of donor animals from controlled donor herds.

This monograph provides a general quality specification for bovine serum. Various measures are applied during the production of bovine serum aimed at obtaining a product that is acceptable as regards viral safety. No single measure, nor the combination of measures outlined below can guarantee complete viral safety but they rather reduce the risk involved in the use of serum in the manufacture of medicinal products. It is therefore necessary for the manufacturer of a medicinal product to take account of this when choosing the serum for a particular use by making a risk assessment.

PRODUCTION

All stages of serum production are submitted to a suitable quality management system.

Traceability of serum is maintained from the final container to the abattoir of origin (for blood collected from slaughtered animals) or to the herd of origin (for blood collected from donor animals).

Further guarantee of the safety and quality of serum may be ensured by the use of a controlled donor herd. Where serum is obtained from such a herd, the animals are subjected to regular veterinary examination to ascertain their health status. Animals introduced into the herd are traceable as regards source, breeding and rearing history. The introduction of animals into the herd follows specified procedures, including defined quarantine measures. During the quarantine period the animals are observed and tested to establish that they are free from all agents and antibodies from which the donor herd is claimed to be free. It may be necessary to test the animals in quarantine for freedom from additional agents, depending on factors such as information available on their breeding and rearing history. It is recommended that animals in the herd should not be vaccinated against bovine viral diarrhoea virus. Tests are carried out for any agent and/or antibody from which the herd is claimed to be free.

Serum is obtained by separation of the serum from blood cells and clot under conditions designed to minimise microbial contamination. Serum from a number of animals is pooled and a batch number is allocated to the pool. Appropriate steps are taken to ensure homogeneity of the harvested material, intermediate pools and the final batch. Suitable measures (for example filtration) are taken to ensure sterility or a low bioburden. Before further processing, the serum is tested for sterility or bioburden. General and specific tests for viral contaminants are carried out as described below.

A step or steps for virus inactivation/removal are applied to serum intended for production of immunological veterinary medicinal products. Unless otherwise justified and authorised for a particular medicinal product, a step or steps for virus inactivation/removal are applied to serum intended for production of human and non-immunological veterinary medicinal products.

INACTIVATION

The inactivation procedure applied is validated with respect to a suitable representative range of viruses covering different types (enveloped, non-enveloped, DNA, RNA viruses). The optimal choice of relevant and model viruses depends strongly on the specific inactivation/removal procedure; representative viruses with different degrees of resistance to the type of treatment must be included. Bovine viral diarrhoea virus must be included in the viruses used for validation. Serum free from antibodies against bovine viral diarrhoea virus is used in part or all of the validation studies.

For bovine serum intended for use in immunological veterinary medicinal products, for inactivation by gamma irradiation a minimum dose of 30 kGy is applied, unless otherwise justified and authorised.

Critical parameters for the method of virus inactivation/removal are established and the parameters used in the validation study are strictly adhered to during subsequent application of the procedures to each batch of serum.

For inactivation by gamma irradiation, critical parameters include:
— the temperature;
— packaging configuration;
— distribution of dosimeters to assess the effective dose received by the product whatever its position;
— the minimum and maximum dose received.

QUALITY CONTROL TESTS APPLIED TO EACH BATCH

A suitable sample size for each batch is established. Specific tests for viral contaminants are validated with respect to sensitivity and specificity. The cell cultures used for general tests for viral contaminants are shown to be sensitive to a suitable range of potential contaminants. Control cells used in the tests are cultivated, where relevant, with a bovine serum controlled and inactivated as described in this monograph. Serum free from antibodies to bovine viral diarrhoea virus is required for validation of the effect of antibodies on the detection limits for bovine viral diarrhoea virus.

Tests carried out on the batch prior to treatment

The following tests are carried out on the serum (before any virus inactivation/removal steps, where applicable).

Tests for viral contaminants General tests supplemented by specific tests are carried out.

General tests Validated tests are carried out by inoculation of the serum on at least 2 distinct cell lines, one of which is of bovine origin. The cell lines used are suitable for detecting haemadsorbing viruses such as bovine parainfluenza virus 3 and cytopathic agents such as bovine herpesvirus 1.

Specific tests for viral contaminants (if not detected by general tests), where relevant in view of the country of origin of the serum Bluetongue virus, bovine adenovirus, bovine parvovirus, bovine respiratory syncytial virus, bovine viral diarrhoea virus, rabies virus and reovirus. Depending on the country of origin, specific tests for other viruses may be needed. The animal health status of countries is defined by the 'Office International des Epizooties' (OIE).

For serum to be subjected to a virus inactivation/removal procedure, if evidence of viral contamination is found in any of the tests described above, the serum is acceptable only if the virus is identified and shown to be present in an amount that has been shown in a validation study to be effectively inactivated.

For serum that is not to be subjected to a virus inactivation/removal procedure, if evidence of viral contamination is found in any of the tests described above, the serum is not acceptable.

A test for bovine viral diarrhoea virus antibodies is carried out; an acceptance criterion for the titre is established taking account of the risk assessment.

Composition The content of a suitable selection of the following components is determined and shown to be within

the expected range for the type of serum: cholesterol, α-, β- and γ-globulin, albumin, creatinine, bilirubin, glucose, serum aspartate transaminase (SAST, formerly SGOT - serum glutamic-oxaloacetic transaminase), serum alanine transaminase (SALT, formerly SGPT - glutamic-pyruvic transaminase), phosphorus, potassium, calcium, sodium and pH.

Tests carried out on the batch post-treatment

If bovine viral diarrhoea virus was detected before virus inactivation/removal, the following test for bovine viral diarrhoea virus is carried out after virus inactivation/removal.

Test for bovine viral diarrhoea virus A validated test for bovine viral diarrhoea virus is carried out, for example by inoculation into susceptible cell cultures, followed by not fewer than 3 subcultures and detection by immunostaining. No evidence of the presence of bovine viral diarrhoea virus is found.

IDENTIFICATION

A. The electrophoretic pattern corresponds to that for serum and is consistent with the type (foetal or other) of bovine serum.

B. Bovine origin is confirmed by a suitable immunochemical method (*2.7.1*).

TESTS

Osmolality (*2.2.35*)

280 mosmol/kg to 365 mosmol/kg for foetal bovine serum and 240 mosmol/kg to 340 mosmol/kg for other types.

Total protein (*2.5.33*)

30 mg/mL to 45 mg/mL for foetal bovine serum and minimum 35 mg/mL for other types.

Haemoglobin

Maximum 4 mg/mL, determined by a validated method, such as spectrophotometry.

Bacterial endotoxins (*2.6.14*)

Less than 10 IU/mL for donor bovine serum, less than 25 IU/mL for foetal bovine serum, less than 100 IU/mL for other types.

Sterility (*2.6.1*)

It complies with the test. Use 10 mL for each medium.

Mycoplasmas (*2.6.7*)

It complies with the test.

STORAGE

Frozen at −10 °C or below.

LABELLING

The label states:

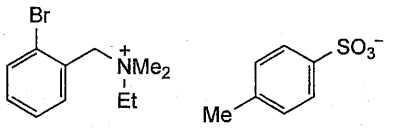
— the type of serum;

— where applicable, that the serum has been inactivated and the inactivation method;

— where the serum has been inactivated by gamma irradiation, the target minimum dose of the irradiation procedure.

Ph Eur

Bretylium Tosilate



$C_{18}H_{24}BrNO_3S$ 414.4 *61-75-6*

Action and use

Antiarrhythmic.

Preparation

Bretylium Injection

DEFINITION

Bretylium Tosilate is 2-bromobenzyl-*N*-ethyldimethylammonium-*p*-toluenesulfonate. It contains not less than 99.0% and not more than 101.0% of $C_{18}H_{24}BrNO_3S$, calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder. It melts at about 98°. It exhibits polymorphism.

Freely soluble in *water*, in *ethanol (96%)* and in *methanol*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of bretylium tosilate *(RS 030)*. If the spectra are not concordant, dissolve a quantity of the substance being examined in the minimum volume of *acetone* by heating on a water bath at 50°, evaporate to dryness at room temperature under a current of nitrogen and prepare a new spectrum of the residue.

B. Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in *water*.

(1) 0.5% w/v of the substance being examined.

(2) 0.5% w/v of *bretylium tosilate BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a *silica gel F_{254} precoated plate* (Merck plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry it in a current of air and examine under *ultraviolet light (254 nm)*.

MOBILE PHASE

15 volumes of *glacial acetic acid*, 30 volumes of *water* and 75 volumes of *butan-1-ol*.

CONFIRMATION

The two principal spots in the chromatogram obtained with solution (1) correspond to those in the chromatogram obtained with solution (2).

TESTS

Acidity

pH of a 5.0% w/v solution, 5.0 to 6.5, Appendix V L.

Clarity and colour of solution

A 5.0% w/v solution is *clear*, Appendix IV A, and *colourless*, Appendix IV B, Method II.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in mobile phase.

(1) 0.20% w/v of the substance being examined.

(2) 0.002% w/v of the substance being examined.

(3) 0.05% w/v of *bretylium tosilate BPCRS* and 0.05% w/v of *2-bromobenzyldimethylamine hydrochloride BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with particles of silica the surface of which has been modified by chemically bonded phenyl groups (5 µm) (Spherisorb Phenyl is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 265 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

0.5 volume of *triethylamine*, 2 volumes of *glacial acetic acid*, 19 volumes of *acetonitrile* and 81 volumes of *0.01M sodium octanesulfonate*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution factor* between the two principal peaks is at least 6.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any *secondary peak* is not greater than half the area of the peak in the chromatogram obtained with solution (2) (0.5%);

the sum of the areas of all such peaks is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%).

Disregard the peak due to tosilate (retention time, about 2 minutes) and any peak with an area less than 0.05 times the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

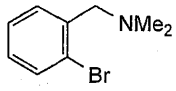
Loss on drying
When dried to constant weight over *phosphorus pentoxide* at 60° at a pressure not exceeding 0.7 kPa, loses not more than 3.0% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A. Use 1 g.

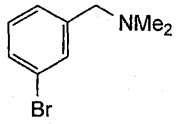
ASSAY
Dissolve 0.2 g in 50 mL of *1,4-dioxan* and carry out Method I for *non-aqueous titration*, Appendix VIII A, using *0.025M perchloric acid VS* as titrant and determining the end-point potentiometrically. Each mL of *0.025M perchloric acid VS* is equivalent to 10.36 mg of $C_{18}H_{24}BrNO_3S$.

STORAGE
Bretylium Tosilate should be kept in an airtight container and protected from light.

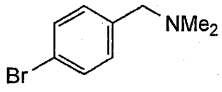
IMPURITIES



A. 2-bromobenzyldimethylamine,



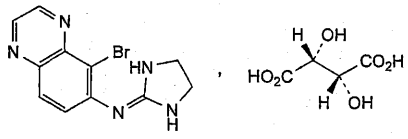
B. 3-bromobenzyldimethylamine,



C. 4-bromobenzyldimethylamine.

Brimonidine Tartrate

(Ph. Eur. monograph 2760)



$C_{15}H_{16}BrN_5O_6$ 442.2 *70359-46-5*

Action and use
Alpha$_2$-adrenoceptor agonist; treatment of hypertension.

Ph Eur

DEFINITION
5-Bromo-*N*-(imidazolidin-2-ylidene)quinoxalin-6-amine (2*R*,3*R*)-2,3-dihydroxybutanedioate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or slightly yellowish or slightly brownish powder.

Solubility
Soluble in water, practically insoluble in anhydrous ethanol and in toluene.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *brimonidine tartrate CRS*.

TESTS

Specific optical rotation (*2.2.7*)
+ 9.0 to + 10.5 (dried substance).

Dissolve 0.50 g in *water R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 65.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve the contents of a vial of *brimonidine for system suitability CRS* (containing impurity E) in 1.0 mL of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 2.6 g of *sodium heptanesulfonate R* in 310 mL of *methanol R*, add 2.5 mL of *triethylamine R* and 7.5 mL of *glacial acetic acid R*, and dilute to 1000 mL with *water R*. Use a freshly prepared mixture.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 264 nm.

Injection 20 µL.

Run time 3 times the retention time of brimonidine.

Identification of impurities Use the chromatogram supplied with *brimonidine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity E.

Relative retention With reference to brimonidine (retention time = about 19 min): impurity E = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity E and brimonidine.

Calculation of percentage contents:
— for each impurity, use the concentration of brimonidine in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

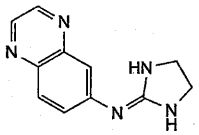
ASSAY

Dissolve 0.350 g in 70 mL of *anhydrous acetic acid R* using sonication until complete dissolution. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

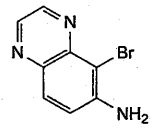
1 mL of *0.1 M perchloric acid* is equivalent to 44.22 mg of $C_{15}H_{16}BrN_5O_6$.

IMPURITIES

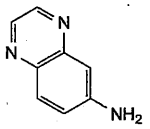
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



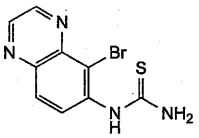
A. *N*-(imidazolidin-2-ylidene)quinoxalin-6-amine,



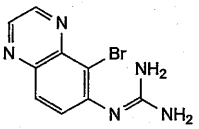
B. 5-bromoquinoxalin-6-amine,



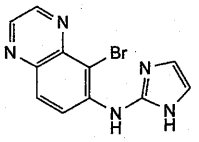
C. quinoxalin-6-amine,



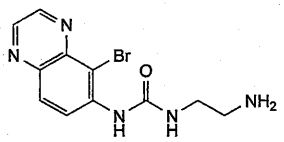
D. 1-(5-bromoquinoxalin-6-yl)thiourea,



E. 2-(5-bromoquinoxalin-6-yl)guanidine,



F. 5-bromo-*N*-(1*H*-imidazol-2-yl)quinoxalin-6-amine,

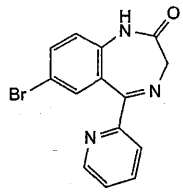


G. 1-(2-aminoethyl)-3-(5-bromoquinoxalin-6-yl)urea.

Ph Eur

Bromazepam

(Ph. Eur. monograph 0879)



$C_{14}H_{10}BrN_3O$ 316.2 *1812-30-2*

Action and use
Benzodiazepine.

Ph Eur

DEFINITION

7-Bromo-5-(pyridin-2-yl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or yellowish, crystalline powder.

Solubility
Practically insoluble in water, slightly soluble or sparingly soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bromazepam CRS.*

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in 9 mL of a mixture of 1 volume of *acetonitrile R* and 8 volumes of *methanol R*. Dilute to 20.0 mL with an 11.33 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.0 with a 100 g/L solution of *potassium hydroxide R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *bromazepam for system suitability CRS* (containing impurities A, B, C, D and E) in 5 mL of a mixture of 1 volume of *acetonitrile R* and 8 volumes of *methanol R*. Dilute to 10.0 mL with an 11.33 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.0 with a 100 g/L solution of *potassium hydroxide R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 50 °C.

Mobile phase Mix 5 volumes of *acetonitrile R*, 45 volumes of *methanol R* and 50 volumes of an 11.33 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.0 with a 100 g/L solution of *potassium hydroxide R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 µL.

Run time 4 times the retention time of bromazepam.

Identification of impurities Use the chromatogram supplied with *bromazepam for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to bromazepam (retention time = about 5 min): impurity D = about 1.4; impurity A = about 1.5; impurity C = about 1.6; impurity E = about 2.1; impurity B = about 2.2.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to bromazepam and impurity D and minimum 1.2 between the peaks due to impurities A and C.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.3; impurity B = 1.8; impurity E = 2.1;
— *impurities A, B, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.2 per cent, determined on 1.000 g by drying at 80 °C at a pressure not exceeding 2.7 kPa for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 20 mL of *anhydrous acetic acid R*. Add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 31.62 mg of $C_{14}H_{10}BrN_3O$.

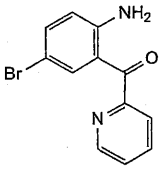
STORAGE

Protected from light.

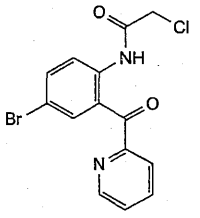
IMPURITIES

Specified impurities *A, B, E.*

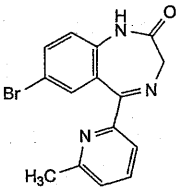
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D.*



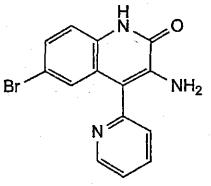
A. (2-amino-5-bromophenyl)(pyridin-2-yl)methanone,



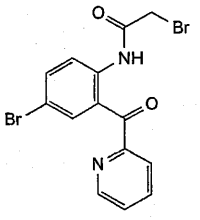
B. *N*-[4-bromo-2-(pyridin-2-ylcarbonyl)phenyl]-2-chloroacetamide,



C. 7-bromo-5-(6-methylpyridin-2-yl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one,



D. 3-amino-6-bromo-4-(pyridin-2-yl)quinolin-2(1*H*)-one,

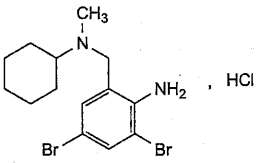


E. 2-bromo-*N*-[4-bromo-2-(pyridin-2-ylcarbonyl)phenyl]acetamide.

Ph Eur

Bromhexine Hydrochloride

(Ph. Eur. monograph 0706)



$C_{14}H_{21}Br_2ClN_2$ 412.6 *611-75-6*

Action and use
Mucolytic.

Ph Eur

DEFINITION

2,4-Dibromo-6-[[cyclohexyl(methyl)amino]methyl]aniline hydrochloride.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, slightly soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bromhexine hydrochloride CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution Dissolve 10 mg of *bromhexine hydrochloride CRS* in *methanol R* and dilute to 5.0 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R, 2-propanol R, acetone R* (1:29:70 *V/V/V*).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve about 20 mg in 1 mL of *methanol R* and add 1 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 0.6 g in *methanol R* and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 1.26 g of *ammonium formate R* in 950 mL of *water for chromatography R* and adjust to pH 4.4 with *anhydrous formic acid R*. Dilute to 1000.0 mL with *water for chromatography R*.

Solvent mixture *acetonitrile for chromatography R, water for chromatography R* (50:50 *V/V*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of *bromhexine for system suitability CRS* (containing impurities C and D) in the solvent mixture and dilute to 2.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *bromhexine hydrochloride CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped solid core octadecylsilyl silica gel for chromatography R* (2.6 µm);
- *temperature*: 30 °C.

Mobile phase *acetonitrile for chromatography R*, buffer solution (40:60 *V/V*).

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 248 nm.

Injection 3.0 µL of test solution (a) and reference solutions (a) and (b).

Run time Twice the retention time of bromhexine.

Identification of impurities Use the chromatogram supplied with *bromhexine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities C and D.

Relative retention With reference to bromhexine (retention time = about 10 min): impurity C = about 0.2; impurity D = about 0.3.

System suitability Reference solution (a):
- *resolution*: minimum 2.0 between the peaks due to impurities C and D.

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity C by 1.6;
- for each impurity, use the concentration of bromhexine in reference solution (b).

Limits:
- *impurity C*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.2 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{14}H_{21}Br_2ClN_2$ taking into account the assigned content of *bromhexine hydrochloride CRS*.

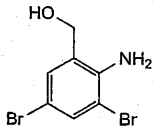
STORAGE

Protected from light.

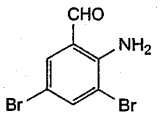
IMPURITIES

Specified impurities *C.*

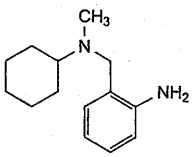
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, D, E.*



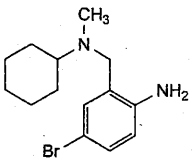
A. (2-amino-3,5-dibromophenyl)methanol,



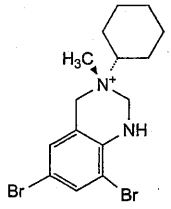
B. 2-amino-3,5-dibromobenzaldehyde,



C. 2-[[cyclohexyl(methyl)amino]methyl]aniline,



D. 4-bromo-2-[[cyclohexyl(methyl)amino]methyl]aniline,



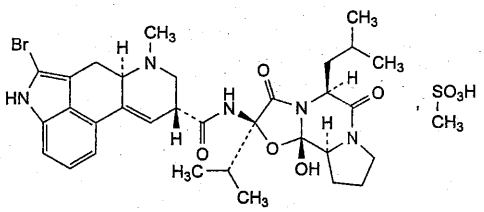
and enantiomer

E. (3*RS*)-6,8-dibromo-3-cyclohexyl-3-methyl-1,2,3,4-tetrahydroquinazolin-3-ium.

Ph Eur

Bromocriptine Mesilate

(Ph. Eur. monograph 0596)



$C_{33}H_{44}BrN_5O_8S$ 751 *22260-51-1*

Action and use
Dopamine receptor agonist.

Preparations
Bromocriptine Capsules
Bromocriptine Tablets

Ph Eur

DEFINITION

(6a*R*,9*R*)-5-Bromo-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide monomethanesulfonate.

Content
98.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in bromocriptine mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation.
The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance
White or slightly coloured, fine crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methanol, soluble in ethanol (96 per cent), sparingly soluble in methylene chloride.

It is very sensitive to light.

The identification, tests and assay are to be carried out as rapidly as possible, protected from light.

IDENTIFICATION

First identification: B.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 10.0 mg in 10 mL of *methanol R* and dilute to 200.0 mL with *0.01 M hydrochloric acid.*

Spectral range 250-380 nm.

Absorption maximum At 305 nm.

Absorption minimum At 270 nm.

Specific absorbance at the absorption maximum 120 to 135 (dried substance).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bromocriptine mesilate CRS.*

C. Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Solvent mixture *ethanol (96 per cent) R, methanol R, methylene chloride R* (30:30:40 *V/V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution Dissolve 10 mg of *bromocriptine mesilate CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture.

Plate *TLC silica gel G plate R.*

Mobile phase *concentrated ammonia R, water R, 2-propanol R, methylene chloride R, ether R* (0.1:1.5:3:88:100 *V/V/V/V/V*).

Application 10 µL.

Development Immediately in an unsaturated tank, over a path of 15 cm.

Drying In a current of cold air for 2 min.

Detection Spray with *ammonium molybdate solution R3* and dry at 100 °C until the spots appear (about 10 min).

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 g add 5 mL of *dilute hydrochloric acid R* and shake for about 5 min. Filter and add 1 mL of *barium chloride solution R1*. The filtrate remains clear. To a further 0.1 g add 0.5 g of *anhydrous sodium carbonate R*, mix and ignite until a white residue is obtained. Allow to cool and dissolve the residue in 7 mL of *water R* (solution A). Solution A gives reaction (a) of sulfates (*2.3.1*).

E. Solution A obtained in identification test D gives reaction (a) of bromides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_5, BY_5 or Y_5 (*2.2.2, Method II*).

Dissolve 0.25 g in *methanol R* and dilute to 25 mL with the same solvent.

pH (*2.2.3*)
3.1 to 3.8.

Dissolve 0.2 g in a mixture of 2 volumes of *methanol R* and 8 volumes of *carbon dioxide-free water R* and dilute to 20 mL with the same mixture of solvents.

Specific optical rotation (*2.2.7*)
+ 95 to + 105 (dried substance).

Dissolve 0.100 g in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10.0 mL with the same mixture of solvents.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *buffer solution pH 2.0 R, methanol R* (50:50 *V/V*).

Test solution Dissolve 0.500 g of the substance to be examined in 5.0 mL of *methanol R* and dilute to 10.0 mL with *buffer solution pH 2.0 R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *bromocriptine mesilate for system suitability CRS* (containing impurities A and B) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.12 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 0.791 g/L solution of *ammonium carbonate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	90 → 40	10 → 60
30 - 45	40	60

Flow rate 2 mL/min.

Detection Spectrophotometer at 300 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *bromocriptine mesilate for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to bromocriptine: impurity C = about 1.2.

System suitability Reference solution (c):
— *resolution*: minimum 1.1 between the peaks due to impurities A and B.

Limits:
— *impurity A*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent);
— *impurity C*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
— *impurities B, D, E, F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent) and not more than 1 such peak has an area greater than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent), apart from the peak due to impurity A.

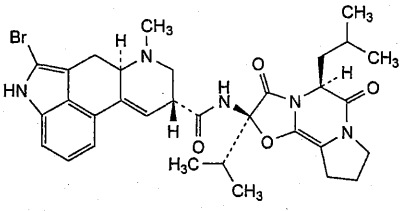
Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 0.500 g by drying *in vacuo* at 80 °C for 5 h.

ASSAY
Dissolve 0.500 g in 80 mL of a mixture of 10 volumes of *anhydrous acetic acid R* and 70 volumes of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

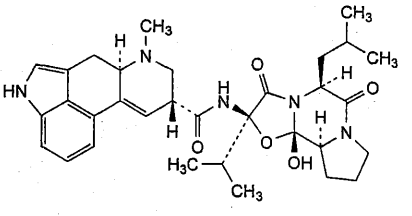
1 mL of *0.1 M perchloric acid* is equivalent to 75.1 mg of $C_{33}H_{44}BrN_5O_8S$.

STORAGE
In an airtight container, protected from light, at a temperature not exceeding −15 °C.

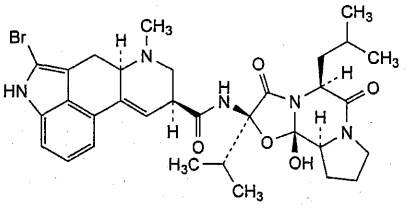
IMPURITIES
Specified impurities *A, B, C, D, E, F, G.*



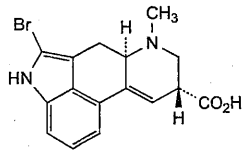
A. (6a*R*,9*R*)-5-bromo-7-methyl-*N*-[(2*R*,5*S*)-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxo-2,3,5,6,9,10-hexahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (2-bromoanhydro-α-ergocryptine),



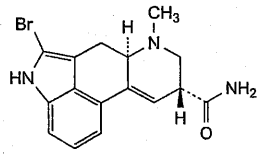
B. (6a*R*,9*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (α-ergocryptine),



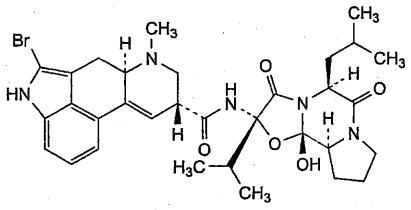
C. (6a*R*,9*S*)-5-bromo-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide ((8*S*)-2-bromo-α-ergocryptine),



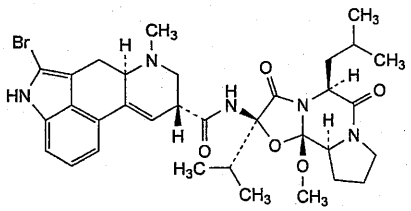
D. (6a*R*,9*R*)-5-bromo-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxylic acid,



E. (6a*R*,9*R*)-5-bromo-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide,



F. (6a*R*,9*R*)-5-bromo-*N*-[(2*S*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide ((2′*S*)-2-bromo-α-ergocryptine),

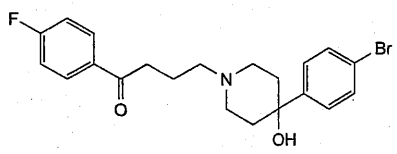


G. (6a*R*,9*R*)-5-bromo-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-methoxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (2-bromo-10′b-*O*-methyl-α-ergocryptine).

Ph Eur

Bromperidol

(Ph. Eur. monograph 1178)



$C_{21}H_{23}BrFNO_2$ 420.3 *10457-90-6*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
4-[4-(4-Bromophenyl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, sparingly soluble in methanol and in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Melting point (*2.2.14*): 156 °C to 159 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bromperidol CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *bromperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *bromperidol CRS* and 10 mg of *haloperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *tetrahydrofuran R, methanol R,* 58 g/L solution of *sodium chloride R* (10:45:45 *V/V/V*).

Application 1 µL.

Development In an unsaturated tank, over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 spots which may, however, not be completely separated.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 5 mL of *anhydrous ethanol R.* Add 0.5 mL of *dinitrobenzene solution R* and 0.5 mL of *2 M alcoholic potassium hydroxide R.* A violet colour is produced that becomes brownish-red after 20 min.

E. To 0.1 g in a porcelain crucible add 0.5 g of *anhydrous sodium carbonate R.* Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R.* The solution gives reaction (a) of bromides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 0.2 g in 20 mL of a 1 per cent *V/V* solution of *lactic acid R.*

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of *bromperidol CRS* and 5.0 mg of *haloperidol CRS* in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.1 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 17 g/L solution of *tetrabutylammonium hydrogen sulfate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	90 → 50	10 → 50
15 - 20	50	50
20 - 25	90	10

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Relative retention With reference to bromperidol (retention time = about 6 min): impurity A = about 0.5; impurity B = about 0.8; haloperidol = about 0.9; impurity C = about 1.4; impurity D = about 1.5; impurity E = about 1.8; impurity F = about 1.85.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to haloperidol and bromperidol.

Limits:
— *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

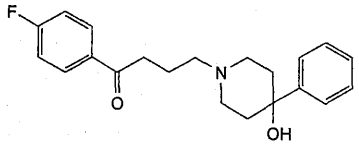
1 mL of *0.1 M perchloric acid* is equivalent to 42.03 mg of $C_{21}H_{23}BrFNO_2$.

STORAGE

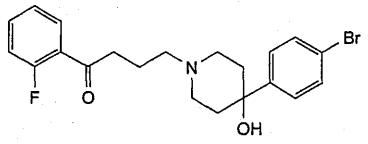
Protected from light.

IMPURITIES

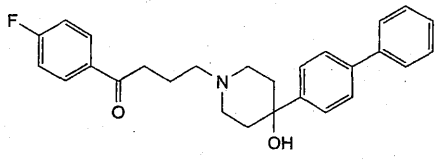
Specified impurities *A, B, C, D, E, F.*



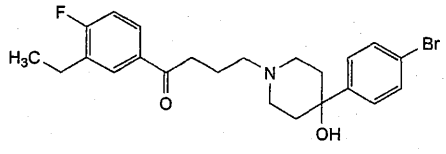
A. 1-(4-fluorophenyl)-4-(4-hydroxy-4-phenylpiperidin-1-yl) butan-1-one,



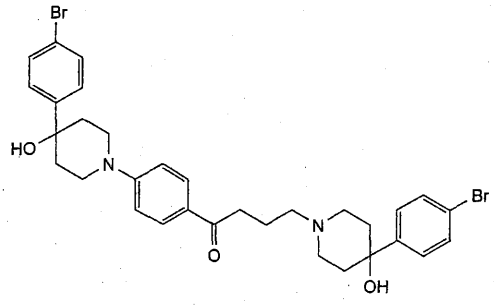
B. 4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]-1-(2-fluorophenyl)butan-1-one,



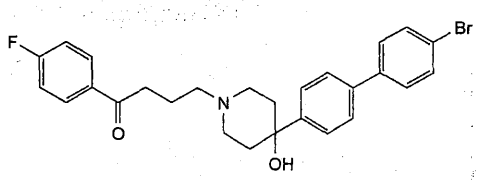
C. 4-[4-(biphenyl-4-yl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one,



D. 4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]-1-(3-ethyl-4-fluorophenyl)butan-1-one,



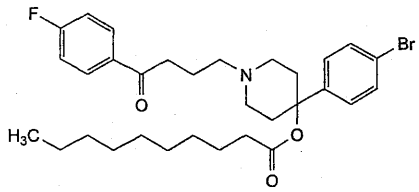
E. 4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]-1-[4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]phenyl]butan-1-one,



F. 4-[4-(4′-bromobiphenyl-4-yl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one.

Bromperidol Decanoate

(Ph. Eur. monograph 1397)



$C_{31}H_{41}BrFNO_3$ 574.6 *75067-66-2*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
4-(4-Bromophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, very soluble in methylene chloride, soluble in ethanol (96 per cent).

mp
About 60 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bromperidol decanoate CRS.*

B. To 0.1 g in a porcelain crucible add 0.5 g of *anhydrous sodium carbonate R*. Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R*. The solution gives reaction (a) of bromides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_5 (*2.2.2, Method II*).

Dissolve 2.0 g in *methylene chloride R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of *bromperidol decanoate CRS* and 2.5 mg of *haloperidol decanoate CRS* in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.1 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 27 g/L solution of *tetrabutylammonium hydrogen sulfate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	80 → 40	20 → 60
30 - 35	40	60
35 - 40	40 → 80	60 → 20

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Relative retention With reference to bromperidol decanoate (retention time = about 24 min): impurity G = about 0.10; impurity L = about 0.15; impurity H = about 0.8; impurity A = about 0.89; impurity I = about 0.91; impurity B = about 0.96; haloperidol decanoate = about 0.98; impurity F = about 1.10; impurity C = about 1.15; impurity K = about 1.2; impurity E = about 1.23; impurity D = about 1.25.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to haloperidol decanoate and bromperidol decanoate.

Limits:
— *impurities A, B, C, D, E, F, G, H, I, J, K*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 30 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.450 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

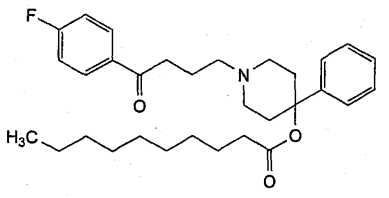
1 mL of *0.1 M perchloric acid* is equivalent to 57.46 mg of $C_{31}H_{41}BrFNO_3$.

STORAGE
Protected from light, at a temperature below 25 °C.

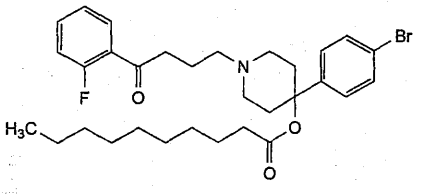
IMPURITIES
Specified impurities *A, B, C, D, E, F, G, H, I, J, K.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

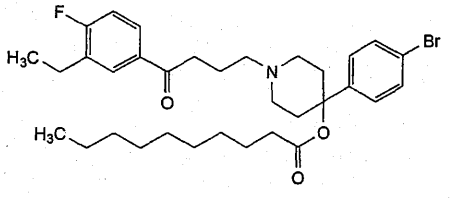
demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) L.



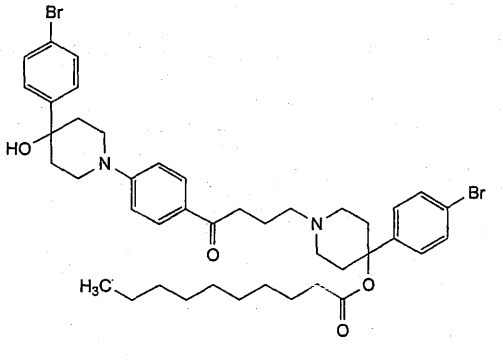
A. 1-[4-(4-fluorophenyl)-4-oxobutyl]-4-phenylpiperidin-4-yl decanoate,



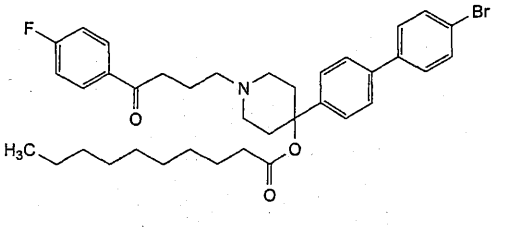
B. 4-(4-bromophenyl)-1-[4-(2-fluorophenyl)-4-oxobutyl]-piperidin-4-yl decanoate,



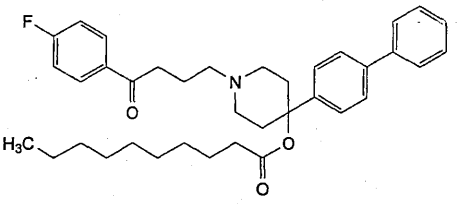
C. 4-(4-bromophenyl)-1-[4-(3-ethyl-4-fluorophenyl)-4-oxobutyl]-piperidin-4-yl decanoate,



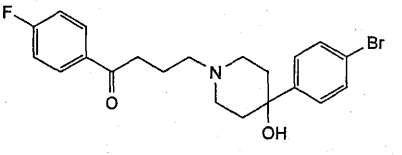
D. 4-(4-bromophenyl)-1-[4-[4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]phenyl]-4-oxobutyl]piperidin-4-yl decanoate,



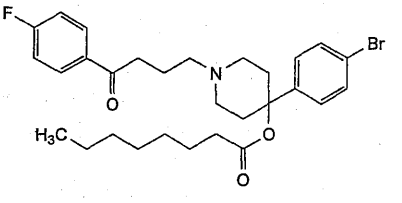
E. 4-(4′-bromobiphenyl-4-yl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate,



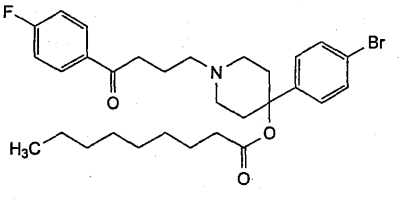
F. 4-(biphenyl-4-yl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl decanoate,



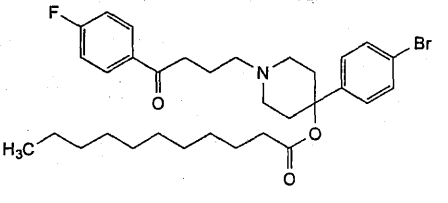
G. 4-[4-(4-bromophenyl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one (bromperidol),



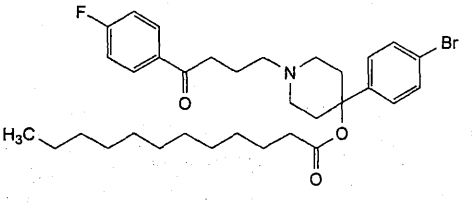
H. 4-(4-bromophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl octanoate,



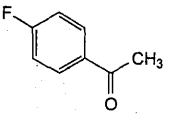
I. 4-(4-bromophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl nonanoate,



J. 4-(4-bromophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl undecanoate,



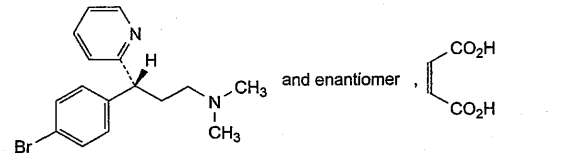
K. 4-(4-bromophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl dodecanoate,



L. 1-(4-fluorophenyl)ethanone.

Brompheniramine Maleate

(Ph. Eur. monograph 0977)



$C_{20}H_{23}BrN_2O_4$ 435.3 *980-71-2*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION

(3*RS*)-3-(4-Bromophenyl)-*N*,*N*-dimethyl-3-(pyridin-2-yl) propan-1-amine (*Z*)-butenedioate.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, freely soluble in ethanol (96 per cent), in methanol and in methylene chloride.

IDENTIFICATION

First identification: C, F.

Second identification: A, B, D, E, F.

A. Melting point (*2.2.14*): 130 °C to 135 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 65 mg in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with the same solution. Dilute 5.0 mL of this solution to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*.

Spectral range 220-320 nm.

Absorption maximum At 265 nm.

Specific absorbance at the absorption maximum 190 to 210.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *brompheniramine maleate CRS*.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution Dissolve 56 mg of *maleic acid R* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *water R, anhydrous formic acid R, methanol R, di-isopropyl ether R* (3:7:20:70 *V/V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In a current of air for 5 min.

Detection Examine in ultraviolet light at 254 nm.

Results The chromatogram obtained with the test solution shows 2 clearly separated spots; the upper spot is similar in position and size to the spot in the chromatogram obtained with the reference solution.

E. To 0.15 g in a porcelain crucible add 0.5 g of *anhydrous sodium carbonate R*. Heat over an open flame for 10 min. Allow to cool. Take up the residue in 10 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R*. The solution gives reaction (a) of bromides (*2.3.1*).

F. Optical rotation (see Tests).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 2.0 g in *methanol R* and dilute to 20 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.0.

Dissolve 0.20 g in 20 mL of *carbon dioxide-free water R*.

Optical rotation (*2.2.7*)
−0.20° to + 0.20° (measured in a 2 dm tube).

Dissolve 2.5 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Gas chromatography (*2.2.28*).

Test solution Dissolve 0.100 g of the substance to be examined in 10.0 mL of *methylene chloride R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methylene chloride R*. Dilute 1.0 mL of this solution to 10.0 mL with *methylene chloride R*.

Reference solution (b) Dissolve 10 mg of *chlorphenamine maleate CRS* (impurity A) and 10 mg of *pheniramine maleate CRS* (impurity C) in *methylene chloride R* and dilute to 5 mL with the same solvent. To 2.5 mL of the solution add 2.5 mL of the test solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *polymethylphenylsiloxane R* (film thickness 0.5 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 1.0 mL/min.

Split ratio 1:5.

Temperature:
— *column*: 205 °C;
— *injection port and detector*: 250 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 1.2 times the retention time of brompheniramine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C.

Relative retention With reference to brompheniramine (retention time = about 34 min): impurity C = about 0.4; impurity A = about 0.7.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity A and brompheniramine.

Limits:
— *impurities A, C*: for each impurity, not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

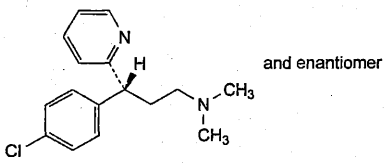
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.260 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

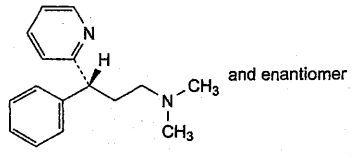
1 mL of *0.1 M perchloric acid* is equivalent to 21.77 mg of $C_{20}H_{23}BrN_2O_4$.

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, C.*



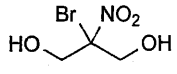
A. (3*RS*)-3-(4-chlorophenyl)-*N*,*N*-dimethyl-3-(pyridin-2-yl) propan-1-amine (chlorphenamine),



C. (3*RS*)-*N*,*N*-dimethyl-3-phenyl-3-(pyridin-2-yl)propan-1-amine (pheniramine).

Ph Eur

Bronopol



$C_3H_6BrNO_4$ 200.0 *52-51-7*

Action and use
Antibacterial preservative.

DEFINITION
Bronopol is 2-bromo-2-nitropropane-1,3-diol. It contains not less than 99.0% and not more than 101.0% of $C_3H_6BrNO_4$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
White or almost white crystals or crystalline powder.

Freely soluble in *water* and in *ethanol (96%)*; slightly soluble in *glycerol* and in *liquid paraffin*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of bronopol *(RS 031)*.

B. Dissolve 0.1 g in 10 mL of *water*, add 10 mL of 7.5M *sodium hydroxide* and, carefully with constant stirring and cooling, 0.5 g of *nickel-aluminium alloy*. Allow the reaction to subside, filter and carefully neutralise with *nitric acid*. The resulting solution yields reaction A characteristic of *bromides*, Appendix VI.

C. *Melting point*, after drying over *phosphorus pentoxide* at a pressure not exceeding 0.7 kPa, about 130°, Appendix V A.

TESTS
Acidity or alkalinity
pH of a 1% w/v solution, 5.0 to 7.0, Appendix V L.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in the mobile phase.

(1) 0.2% w/v of the substance being examined.

(2) Dilute a volume of solution (1) to produce a solution containing 0.0002% w/v of the substance being examined.

(3) 0.001% w/v each of *2-methyl-2-nitropropan-1,3-diol* and *tris(hydroxymethyl)nitromethane*.

(4) 0.0002% w/v each of 2-methyl-2-nitropropane-1,3-diol, 2-nitroethanol, sodium bromide and tris(hydroxymethyl) nitromethane and 0.2% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Phenomenex Luna C18 (2) is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use a column temperature of 35°.

(e) Use a detection wavelength of 214 nm.

(f) Inject 20 µL of each solution.

(g) For solution (1) allow the chromatography to proceed for at least 3 times the retention time of the principal peak.

MOBILE PHASE

1 volume of a 10% v/v solution of *orthophosphoric acid*, 10 volumes of *acetonitrile* and 189 volumes of *water*, adjust the pH to 3.0 using 2M *sodium hydroxide*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (4):

the *resolution factor* between the peaks due to sodium bromide and tris(hydroxymethyl)nitromethane is at least 1.0;

the *resolution factor* between the peaks due to tris (hydroxymethyl)nitromethane and 2-nitroethanol is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to 2-methyl-2-nitropropane-1,3-diol and tris(hydroxymethyl)nitromethane are not greater than the area of the corresponding peaks in the chromatogram obtained with solution (3) (0.5% of each);

the area of any other *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.1%).

Sulfated ash
Not more than 0.1%, Appendix IX A.

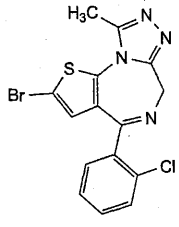
Water
Not more than 0.5% w/w, Appendix IX C, Method I B. Use 5 g.

ASSAY
In a flask fitted with a reflux condenser dissolve 0.4 g in 15 mL of *water* and add 15 mL of *7.5M sodium hydroxide*. Slowly, with caution, add 2 g of *nickel-aluminium alloy* through the reflux condenser, agitating the flask whilst cooling under running water. Allow the mixture to stand for 10 minutes and boil for 1 hour. Cool and filter under reduced pressure, washing the condenser, flask and residue with 150 mL of *water*. Combine the filtrate and washings, add 25 mL of *nitric acid* and 40 mL of *0.1M silver nitrate VS*, shake vigorously and titrate with *0.1M ammonium thiocyanate VS* using *ammonium iron(III) sulfate solution R2* as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of silver nitrate required. Each mL of *0.1M silver nitrate VS* is equivalent to 20.00 mg of $C_3H_6BrNO_4$.

STORAGE
Bronopol should be protected from light.

Brotizolam

(Ph. Eur. monograph 2197)



$C_{15}H_{10}BrClN_4S$ 393.7 *57801-81-7*

Action and use
Benzodiazepine.

Ph Eur

DEFINITION
2-Bromo-4-(2-chlorophenyl)-9-methyl-6*H*-thieno-[3,2-*f*][1,2,4]-triazolo[4,3-*a*][1,4]diazepine.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish powder.

Solubility
Practically insoluble in water, sparingly soluble or slightly soluble in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *brotizolam CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL of *acetonitrile R*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R*.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *brotizolam impurity B CRS* in 50 mL of *acetonitrile R*. Dilute 2 mL of this solution to 20 mL with *acetonitrile R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: 2 g/L solution of *sodium heptanesulfonate monohydrate R*;
— *mobile phase B*: mix 25 volumes of a 2 g/L solution of *sodium heptanesulfonate R* and 75 volumes of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	63	37
4 - 15	63 → 12	37 → 88

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 242 nm.

Injection 5 µL.

Relative retention With reference to brotizolam (retention time = about 7.4 min): impurity A = about 0.5; impurity B = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity B and brotizolam.

Limits:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 100 ppm.
Dissolve 0.67 g in 20.0 mL of *methanol R*, mix and filter.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

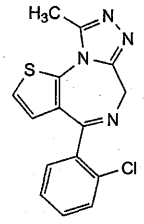
ASSAY
Dissolve 0.150 g in a mixture of 25 mL of *glacial acetic acid R* and 50 mL of *acetic anhydride R*. Titrate to the second point of inflexion with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 19.68 mg of $C_{15}H_{10}BrClN_4S$.

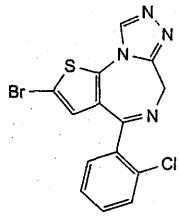
IMPURITIES

Specified impurities B.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A.



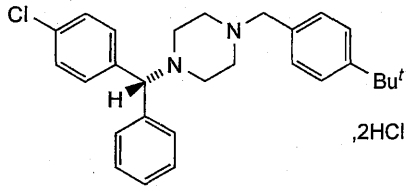
A. 4-(2-chlorophenyl)-9-methyl-6*H*-thieno[3,2-*f*][1,2,4]triazolo[4,3-*a*][1,4]diazepine (desbromobrotizolam),



B. 2-bromo-4-(2-chlorophenyl)-6*H*-thieno[3,2-*f*][1,2,4]triazolo[4,3-*a*][1,4]diazepine (desmethylbrotizolam).

Ph Eur

Buclizine Hydrochloride



$C_{28}H_{33}ClN_2,2HCl$ 506.0 *129-74-8*

Action and use

Histamine H_1 receptor antagonist; antiemetic.

DEFINITION

Buclizine Hydrochloride is (*RS*)-1-(4-*tert*-butylbenzyl)-4-(4-chlorobenzhydryl)piperazine dihydrochloride. It contains not less than 99.0% and not more than 100.5% of $C_{28}H_{33}ClN_2,2HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or slightly yellowish, crystalline powder.

Practically insoluble in *water*; sparingly soluble in *propane-1,2-diol*; very slightly soluble in *ethanol (96%)*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of buclizine hydrochloride *(RS 032)*.

B. A 0.25% w/v solution in *ethanol (50%)* yields reaction A characteristic of *chlorides*, Appendix VI.

TESTS

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using four solutions in the initial mobile phase containing (1) 0.0010% w/v of the substance being examined, (2) 0.50% w/v of the substance being examined, (3) 0.0010% w/v of *1,4-bis(4-chlorobenzhydryl) piperazine BPCRS* and (4) 0.50% w/v of *buclizine hydrochloride impurity standard BPCRS*.

The chromatographic procedure may be carried out using a stainless steel column (20 cm × 4 mm) packed with *octadecylsilyl silica gel for chromatography* (10 µm) (Nucleosil C18 is suitable). Use as the initial mobile phase *0.01M sodium heptanesulfonate* in a mixture of 55 volumes of *water* and 45 volumes of *acetonitrile* and as the final mobile phase *0.01M sodium heptanesulfonate* in a mixture of 20 volumes of *water* and 80 volumes of *acetonitrile*. Before use, adjust the pH of both the initial and final mobile phases to 4.0 with *1M orthophosphoric acid*. Carry out a linear gradient elution with a flow rate of 2 mL per minute for 30 minutes and maintain the final mobile phase for 10 minutes with the same flow rate. Use a detection wavelength of 230 nm.

The test is not valid unless the chromatogram obtained with solution (4) closely resembles the chromatogram supplied with *buclizine hydrochloride impurity standard BPCRS*.

In the chromatogram obtained with solution (2) the area of any peak corresponding to 1,4-bis(4-chlorobenzhydryl)-piperazine is not greater than the area of the peak obtained in the chromatogram with solution (3) and the area of any other *secondary peak* is not greater than the area of the peak in the chromatogram obtained with solution (1).

Loss on drying

When dried to constant weight at 100° to 105°, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.4 g and determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 25.30 mg of $C_{28}H_{33}ClN_2,2HCl$.

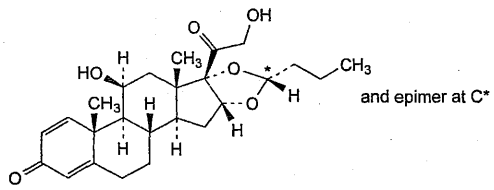
IMPURITIES

A. 1,4-bis(4-chlorobenzhydryl)piperazine,

B. 4-chlorobenzhydrol, 1-(4-chlorobenzhydryl)piperazine, 4-chlorobenzophenone.

Budesonide

(Ph. Eur. monograph 1075)



$C_{25}H_{34}O_6$ 430.5 *51333-22-3*

Action and use
Glucocorticoid.

Preparations
Budesonide Aqueous Nasal Spray
Budesonide Inhalation Powder
Budesonide Inhalation Powder, pre-metered
Budesonide Nebuliser Suspension
Budesonide Pressurised Inhalation

Ph Eur

DEFINITION

Mixture of the C-22*S* (epimer A) and the C-22*R* (epimer B) epimers of 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β,21-dihydroxypregna-1,4-diene-3,20-dione.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *budesonide CRS.*

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (10:90 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 25 mg of *budesonide CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 12.5 mg of *triamcinolone acetonide CRS* in reference solution (a) and dilute to 5 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*; heat at 120 °C for 10 min or until the spots appear and allow to cool; examine the chromatograms in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

C. Dissolve about 2 mg in 2 mL of *sulfuric acid R.* Within 5 min a yellow colour develops. Within 30 min the colour changes to brown or reddish-brown. Cautiously add the solution to 10 mL of *water R* and mix. The colour fades and a clear solution remains.

D. Dissolve about 1 mg in 2 mL of a solution containing 2 g of *phosphomolybdic acid R* dissolved in a mixture of 10 mL of *dilute sodium hydroxide solution R*, 15 mL of *water R* and 25 mL of *glacial acetic acid R.* Heat for 5 min on a water-bath. Cool in iced water for 10 min and add 3 mL of *dilute sodium hydroxide solution R.* The solution is blue.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture *acetonitrile R, phosphate buffer solution pH 3.2 R* (32:68 *V/V*).

Test solution (a) Dissolve 50 mg of the substance to be examined in 15 mL of *acetonitrile R* and dilute to 50 mL with *phosphate buffer solution pH 3.2 R.*

Test solution (b) Dissolve 25.0 mg of the substance to be examined in 15 mL of *acetonitrile R* and dilute to 50.0 mL with *phosphate buffer solution pH 3.2 R.*

Reference solution (a) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *budesonide for system suitability CRS* (containing impurities A, D, G, K and L) in 1.5 mL of *acetonitrile R* and dilute to 5 mL with *phosphate buffer solution pH 3.2 R.*

Reference solution (c) Dissolve 25.0 mg of *budesonide CRS* in 15 mL of *acetonitrile R* and dilute to 50.0 mL with *phosphate buffer solution pH 3.2 R.*

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: *anhydrous ethanol R, acetonitrile R, phosphate buffer solution pH 3.2 R* (2:32:68 *V/V/V*);
— *mobile phase B*: *acetonitrile R, phosphate buffer solution pH 3.2 R* (50:50 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 38	100	0
38 - 50	100 → 0	0 → 100
50 - 60	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *budesonide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, D, G, K and L.

Relative retention With reference to budesonide epimer B (retention time = about 17 min): impurity A = about 0.1; epimers of impurity D = about 0.63 and 0.67; impurity L = about 0.95; epimers of impurity G = about 1.2 and 1.3; epimers of impurity K = about 2.9 and 3.0.

System suitability Reference solution (b):

— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the 1st of the 2 peaks due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to budesonide epimer A (the 2nd of the 2 principal peaks); and minimum 3, where H_p = height above the baseline of the peak due to impurity L and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to budesonide epimer B (the 1st of the 2 principal peaks).

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 1.8; impurity K = 1.3;

— *impurities A, L*: for each impurity, not more than twice the sum of the areas of the 2 peaks due to the budesonide epimers in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurities D, K*: for each impurity, for the sum of the areas of the 2 epimer peaks, not more than twice the sum of the areas of the 2 peaks due to the budesonide epimers in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *unspecified impurities*: for each individual peak, not more than the sum of the areas of the 2 peaks due to the budesonide epimers in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the sum of the areas of the 2 peaks due to the budesonide epimers in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.5 times the sum of the areas of the 2 peaks due to the budesonide epimers in the chromatogram obtained with reference solution (a) (0.05 per cent).

Epimer A

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 21	100	0
21 - 22	100 → 0	0 → 100
22 - 31	0	100

Injection 20 µL of test solution (b) and reference solutions (b) and (c).

Retention time Budesonide epimer B = about 17 min; budesonide epimer A = about 19 min.

System suitability:

— *resolution*: minimum 1.5 between the 2 principal peaks (budesonide epimers A and B) in the chromatogram obtained with reference solution (c);

— *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to impurity L and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to budesonide epimer B (the 1st of the 2 principal peaks) in the chromatogram obtained with reference solution (b).

Limit:

— *epimer A*: 40.0 per cent to 51.0 per cent of the sum of the areas of the 2 peaks due to the budesonide epimers.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

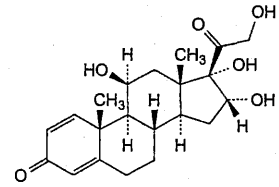
Liquid chromatography (*2.2.29*). Examine the chromatograms obtained in the test for epimer A.

Calculate the percentage content of $C_{25}H_{34}O_6$ from the sum of the areas of the 2 peaks due to the budesonide epimers and the declared content of *budesonide CRS*.

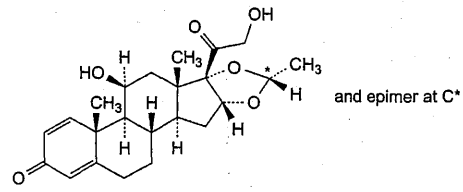
IMPURITIES

Specified impurities *A, D, K, L.*

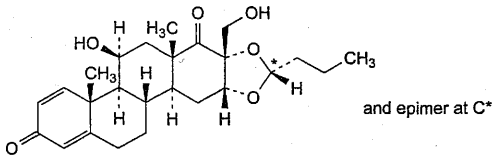
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, E, F, G, H, I, J.*



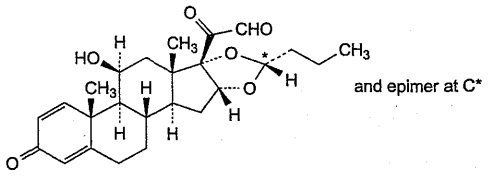
A. 11β,16α,17,21-tetrahydroxypregna-1,4-diene-3,20-dione,



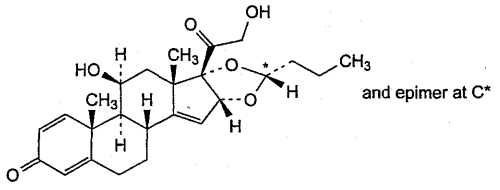
B. 16α,17-[(1*RS*)-ethylidenebis(oxy)]-11β,21-dihydroxypregna-1,4-diene-3,20-dione,



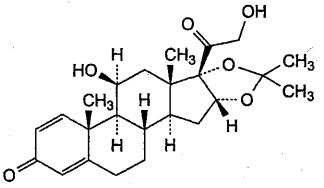
C. 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β-hydroxy-17-(hydroxymethyl)-*D*-homoandrosta-1,4-diene-3,17a-dione,



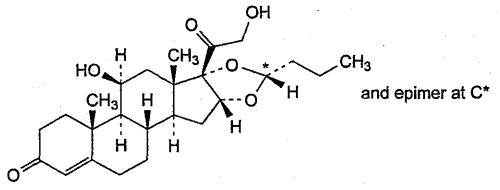
D. 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β-hydroxy-3,20-dioxopregna-1,4-dien-21-al,



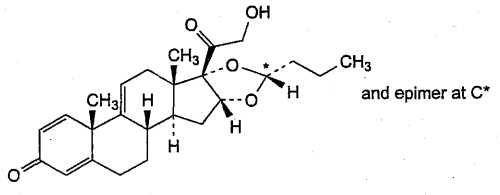
E. 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β,21-dihydroxypregna-1,4,14-triene-3,20-dione,



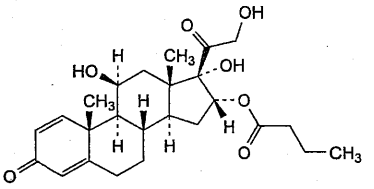
F. 16α,17-[1-methylethylidenebis(oxy)]-11β,21-dihydroxypregna-1,4-diene-3,20-dione,



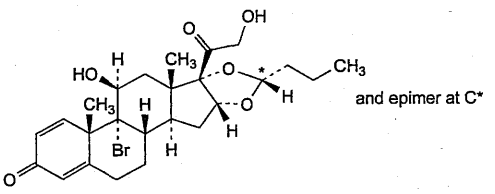
G. 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β,21-dihydroxypregn-4-ene-3,20-dione.



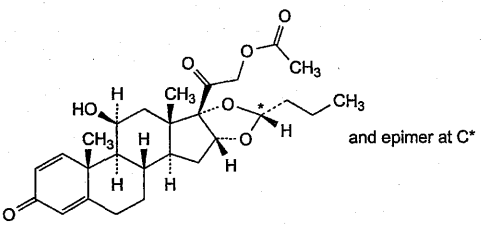
H. 16α,17-[(1*RS*)-butylidenebis(oxy)]-21-hydroxypregna1,4,9(11)-triene-3,20-dione,



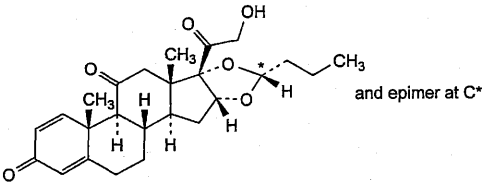
I. 11β,17,21-trihydroxy-3,20-dioxopregna-1,4-dien-16α-yl butanoate,



J. 16α,17-[(1*RS*)-butylidenebis(oxy)]-9α-bromo-11β,21-dihydroxypregna-1,4-diene-3,20-dione,



K. 16α,17-[(1*RS*)-butylidenebis(oxy)]-11β,21-dihydroxypregna-1,4-diene-3,20-dione-21-acetate,

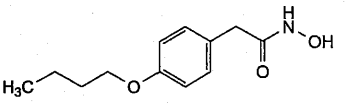


L. 16α,17-[(1*RS*)-butylidenebis(oxy)]-21-hydroxypregna-1,4-diene-3,11,20-trione.

Ph Eur

Bufexamac

(Ph. Eur. monograph 1179)



$C_{12}H_{17}NO_3$ 223.3 *2438-72-4*

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION

2-(4-Butoxyphenyl)-*N*-hydroxyacetamide.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in dimethylformamide, slightly soluble in ethyl acetate and in methanol.

IDENTIFICATION

First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 20 mg in *methanol R* and dilute to 20 mL with the same solvent. Dilute 1 mL of the solution to 50 mL with *methanol R*.

Spectral range 210-360 nm.

Absorption maxima At 228 nm, 277 nm and 284 nm.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bufexamac CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *bufexamac CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *salicylic acid R* in reference solution (a) and dilute to 5 mL with the same solution.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *glacial acetic acid R, dioxan R, toluene R* (4:20:90 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dilute 5.0 mL of the test solution to 25.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *bufexamac CRS*, 5 mg of *bufexamac impurity A CRS*, 5 mg of *bufexamac impurity B CRS*, 5 mg of *bufexamac impurity C CRS* and 5 mg of *bufexamac impurity D CRS* in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 10.0 mL with mobile phase A.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: mix 30 volumes of a 1.4 g/L solution of *dipotassium hydrogen phosphate R* and 70 volumes of *methanol R*, then adjust to pH 3.6 with *dilute phosphoric acid R*;

— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100	0
10 - 13	100 → 70	0 → 30
13 - 40	70	30

Flow rate 1 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to bufexamac (retention time = about 5.7 min): impurity D = about 1.3; impurity A = about 1.8; impurity B = about 3.0; impurity C = about 5.4.

System suitability Reference solution (b):

— *resolution*: minimum 2.0 between the peaks due to bufexamac and impurity D.

Limits:

— *impurities A, B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of *dimethylformamide R*. Titrate with *0.1 M lithium methoxide*, determining the end-point potentiometrically (*2.2.20*).

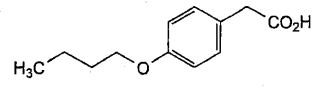
1 mL of *0.1 M lithium methoxide* is equivalent to 22.33 mg of $C_{12}H_{17}NO_3$.

STORAGE

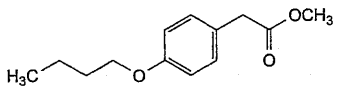
Protected from light.

IMPURITIES

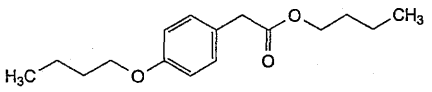
Specified impurities *A, B, C, D.*



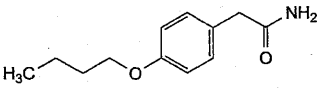
A. 2-(4-butoxyphenyl)acetic acid,



B. methyl 2-(4-butoxyphenyl)acetate,



C. butyl 2-(4-butoxyphenyl)acetate,

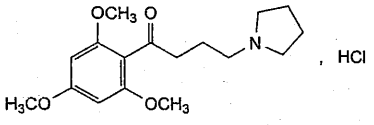


D. 2-(4-butoxyphenyl)acetamide.

Ph Eur

Buflomedil Hydrochloride

(Ph. Eur. monograph 1398)



$C_{17}H_{26}ClNO_4$ 343.9 *35543-24-9*

Action and use
Vasodilator.

Ph Eur

DEFINITION
4-(Pyrrolidin-1-yl)-1-(2,4,6-trimethoxyphenyl)butan-1-one hydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, microcrystalline powder.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent), very slightly soluble in acetone.

mp
About 195 °C, with decomposition.

IDENTIFICATION
First identification: B, D.
Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 25.0 mg in *ethanol (96 per cent) R* and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 20.0 mL with *ethanol (96 per cent) R*.

Spectral range 220-350 nm.

Absorption maximum At 275 nm.

Specific absorbance at the absorption maximum 143 to 149.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *buflomedil hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 40 mg of the substance to be examined in *methanol R* and dilute to 2 mL with the same solvent.

Reference solution Dissolve 40 mg of *buflomedil hydrochloride CRS* in *methanol R* and dilute to 2 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *triethylamine R, 2-propanol R, toluene R* (5:50:50 *V/V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
5.0 to 6.5 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.10 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of *buflomedil impurity B CRS* in the mobile phase, add 0.5 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *buflomedil for peak identification CRS* (containing impurities A and C) in 1.0 mL of reference solution (b).

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 45 volumes of *acetonitrile R1* and 55 volumes of a 9.25 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.5 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Run time Twice the retention time of buflomedil.

Identification of impurities Use the chromatogram supplied with *buflomedil for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to buflomedil (retention time = about 5 min): impurity B = about 0.6; impurity C = about 0.7; impurity A = about 1.5.

System suitability Reference solution (c):

— *resolution*: minimum 1.5 between the peaks due to impurity B and impurity C.

Limits:

— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);

— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

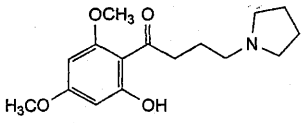
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

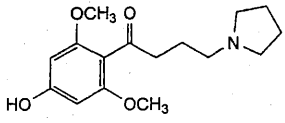
ASSAY
Dissolve 0.300 g in 15 mL of *anhydrous acetic acid R* and add 35 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 34.39 mg of $C_{17}H_{26}ClNO_4$.

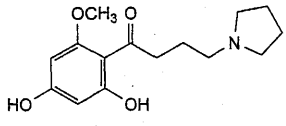
IMPURITIES
Specified impurities *A, B, C.*



A. 1-(2-hydroxy-4,6-dimethoxyphenyl)-4-(pyrrolidin-1-yl) butan-1-one,



B. 1-(4-hydroxy-2,6-dimethoxyphenyl)-4-(pyrrolidin-1-yl) butan-1-one,

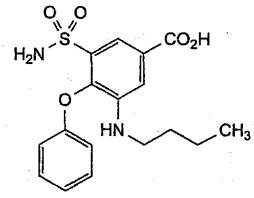


C. 1-(2,4-dihydroxy-6-methoxyphenyl)-4-(pyrrolidin-1-yl) butan-1-one.

Ph Eur

Bumetanide

(Ph. Eur. monograph 1076)



$C_{17}H_{20}N_2O_5S$ 364.4 *28395-03-1*

Action and use
Loop diuretic.

Preparations
Bumetanide Injection
Bumetanide Oral Solution
Bumetanide and Prolonged-release Potassium Tablets
Bumetanide Tablets

Ph Eur

DEFINITION
3-(Butylamino)-4-phenoxy-5-sulfamoylbenzoic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetone and in ethanol (96 per cent), slightly soluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

mp
About 233 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *bumetanide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.1 g in a 6 g/L solution of *potassium hydroxide R* and dilute to 20 mL with the same solution.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of *bumetanide impurity A CRS* and 2 mg of *bumetanide impurity B CRS* in the mobile phase and dilute to 10.0 mL with the mobile

phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (3.5 µm).

Mobile phase Mix 70 volumes of *methanol R*, 25 volumes of *water for chromatography R* and 5 volumes of a 27.2 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.0 with a 280 g/L solution of *potassium hydroxide R*; add *tetrahexylammonium bromide R* to this mixture to obtain a concentration of 2.17 g/L.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 5 times the retention time of bumetanide.

Relative retention With reference to bumetanide (retention time = about 6 min): impurity B = about 0.4; impurity A = about 0.6; impurity C = about 2.5; impurity D = about 4.4.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity A and impurity B.

Limits:
— *impurities A, B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

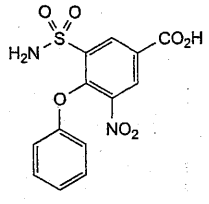
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 50 mL of *alcohol R*. Add 0.1 mL of *phenol red solution R*. Titrate with *0.1 M sodium hydroxide* until a violet-red colour is obtained. Carry out a blank titration.

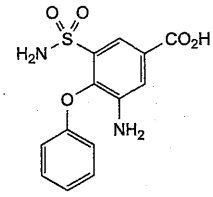
1 mL of *0.1 M sodium hydroxide* is equivalent to 36.44 mg of $C_{17}H_{20}N_2O_5S$.

STORAGE
Protected from light.

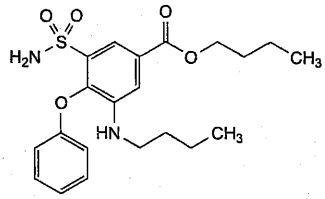
IMPURITIES
Specified impurities *A, B, C, D.*



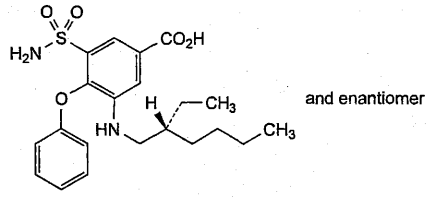
A. 3-nitro-4-phenoxy-5-sulfamoylbenzoic acid,



B. 3-amino-4-phenoxy-5-sulfamoylbenzoic acid,



C. butyl 3-(butylamino)-4-phenoxy-5-sulfamoylbenzoate,

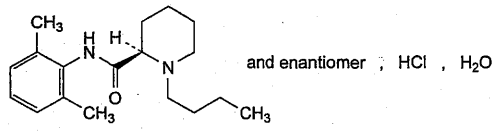


D. 3-[[(2*RS*)-2-ethylhexyl]amino]-4-phenoxy-5-sulfamoylbenzoic acid.

Ph Eur

Bupivacaine Hydrochloride

(Ph. Eur. monograph 0541)



$C_{18}H_{29}ClN_2O,H_2O$ 342.9 *73360-54-0*

Action and use
Local anaesthetic.

Preparations
Bupivacaine Injection
Bupivacaine Heavy Injection
Bupivacaine and Adrenaline Injection/Bupivacaine and Epinephrine Injection
Bupivacaine and Diamorphine Injection
Bupivacaine and Fentanyl Injection

Ph Eur

DEFINITION
(2*RS*)-1-Butyl-*N*-(2,6-dimethylphenyl)piperidine-2-carboxamide hydrochloride monohydrate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, D, E.
Second identification: B, C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *bupivacaine hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.
Reference solution Dissolve 25 mg of *bupivacaine hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.
Plate *TLC silica gel G plate R.*
Mobile phase *concentrated ammonia R, methanol R* (0.1:100 *V/V*).
Application 5 µL.
Development Over a path of 10 cm.
Drying In air.
Detection Spray with *dilute potassium iodobismuthate solution R.*
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 0.1 g in 10 mL of *water R*, add 2 mL of *dilute sodium hydroxide solution R* and shake with 2 quantities, each of 15 mL, of *1,1-dimethylethyl methyl ether R*. Dry the combined upper layers over *anhydrous sodium sulfate R* and filter. Evaporate the filtrate, recrystallise the residue from *ethanol (90 per cent V/V) R* and dry under reduced pressure. The crystals melt (*2.2.14*) at 105 °C to 108 °C.

D. It gives reaction (a) of chlorides (*2.3.1*).

E. Optical rotation (see Tests).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.2 mL of *0.01 M sodium hydroxide*; the pH (*2.2.3*) is not less than 4.7. Add 0.4 mL of *0.01 M hydrochloric acid*; the pH is not greater than 4.7.

Optical rotation (*2.2.7*)
−0.10° to + 0.10°.
Dissolve 1.0 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Related substances
Gas chromatography (*2.2.28*).
Internal standard solution Dissolve 25 mg of *methyl behenate R* in *methylene chloride R* and dilute to 500 mL with the same solvent.
Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *water R*, add 2.5 mL of *dilute sodium hydroxide solution R* and extract with 2 quantities, each of 5 mL, of the internal standard solution. Filter the lower layer.
Reference solution (a) Dissolve 10 mg of the substance to be examined, 10 mg of *bupivacaine impurity B CRS* and 10 mg of *bupivacaine impurity E CRS* in 2.5 mL of *water R*, add 2.5 mL of *dilute sodium hydroxide solution R* and extract with 2 quantities, each of 5 mL, of the internal standard solution. Filter the lower layer and dilute to 20 mL with the internal standard solution.
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the internal standard solution.
Reference solution (c) Dilute 5.0 mL of reference solution (b) to 10.0 mL with the internal standard solution.
Reference solution (d) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the internal standard solution.
Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.25 µm).
Carrier gas *helium for chromatography R.*
Flow rate 2.5 mL/min.
Split ratio 1:12.
Temperature:

	Time (min)	Temperature (°C)
	0	180
Column	0 - 10	180 → 230
	10 - 15	230
Injection port		250
Detector		250

Detection Flame ionisation.
Injection 1 µL.
Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and E.
Relative retention With reference to bupivacaine (retention time = about 10 min): impurity B = about 0.7; impurity E = about 1.1; internal standard = about 1.4.
System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to bupivacaine and impurity E.
Limits:
— *impurity B*: calculate the ratio (R_1) of the area of the principal peak to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (c); from the chromatogram obtained with the test solution, calculate the ratio of the area of the peak due to impurity B to the area of the peak due to the internal standard: this ratio is not greater than R_1 (0.5 per cent);
— *unspecified impurities*: calculate the ratio (R_2) of the area of the principal peak to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (d); from the chromatogram obtained with the test solution, calculate for each impurity the ratio of the area of any peak, apart from the principal peak, the peak due to impurity B and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than R_2 (0.10 per cent);
— *total*: calculate the ratio (R_3) of the area of the principal peak to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (b); from the chromatogram obtained with the

test solution, calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than R_3 (1.0 per cent);
— *disregard limit*: ratio less than 0.05 times R_3 (0.05 per cent).

Impurity F

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 5.0 mg of *bupivacaine impurity F CRS* in mobile phase A and dilute to 100.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 20 mg of *methyl benzoate R* and 25 mg of *bupivacaine impurity F CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A. Dilute 3.0 mL of the solution to 50.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 0.23 g of *sodium dihydrogen phosphate monohydrate R* and 3.626 g of *disodium hydrogen phosphate dihydrate R* in *water R* and dilute to 1000 mL with the same solvent; mix equal volumes of this solution (pH 8.0) and *acetonitrile R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100	0
10 - 15	100 → 80	0 → 20
15 - 25	80	20

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity F.

Relative retention with reference to bupivacaine (retention time = about 20 min): impurity F = about 0.3; methyl benzoate = about 0.4.

System suitability:
— *resolution*: minimum 4.0 between the peaks due to impurity F and methyl benzoate in the chromatogram obtained with reference solution (b);
— *signal-to-noise ratio*: minimum 40 for the principal peak in the chromatogram obtained with reference solution (a).

Limit:
— *impurity F*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (10 ppm).

Loss on drying (*2.2.32*)

4.5 per cent to 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 20 mL of *water R* and 25 mL of *ethanol (96 per cent) R*. Add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M ethanolic sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 32.49 mg of $C_{18}H_{29}ClN_2O$.

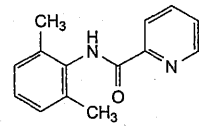
STORAGE

Protected from light.

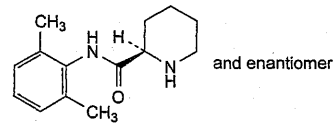
IMPURITIES

Specified impurities *B, F.*

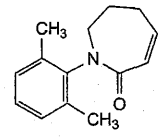
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E.*



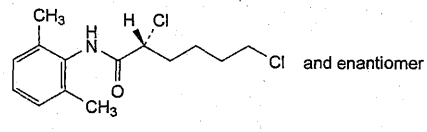
A. *N*-(2,6-dimethylphenyl)pyridine-2-carboxamide,



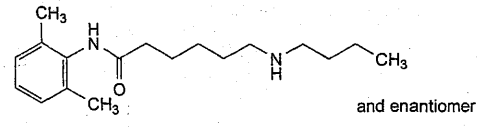
B. (2*RS*)-*N*-(2,6-dimethylphenyl)piperidine-2-carboxamide,



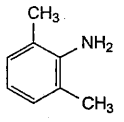
C. 1-(2,6-dimethylphenyl)-1,5,6,7-tetrahydro-2*H*-azepin-2-one,



D. (2*RS*)-2,6-dichloro-*N*-(2,6-dimethylphenyl)hexanamide,



E. 6-(butylamino)-*N*-(2,6-dimethylphenyl)hexanamide,

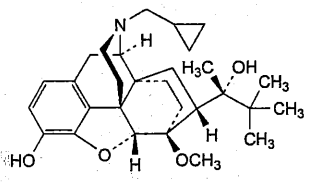


F. 2,6-dimethylaniline.

Ph Eur

Buprenorphine

(Ph. Eur. monograph 1180)



$C_{29}H_{41}NO_4$ 467.6 *52485-79-7*

Action and use
Opioid receptor partial agonist; analgesic.

Preparation
Buprenorphine Transdermal Patches

Ph Eur

DEFINITION
(2*S*)-2-[17-(Cyclopropylmethyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, freely soluble in acetone, soluble in methanol, slightly soluble in cyclohexane. It dissolves in dilute solutions of acids.

mp
About 217 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *buprenorphine CRS*.

TESTS

Solution S
Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
−103 to −107 (dried substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 50.0 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.
Reference solution (b) Dissolve 5 mg of *buprenorphine for system suitability CRS* (containing impurities A, B, F, G, H and J) in 1.0 mL of *methanol R*.
Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 30 °C.
Mobile phase:
— *mobile phase A*: mix 10 volumes of *acetonitrile R* and 90 volumes of the following solution: dissolve 5.44 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 4.5 with a 5 per cent *V/V* solution of *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	89	11
2 - 12	89 → 64	11 → 36
12 - 15	64 → 41	36 → 59
15 - 20	41 → 39	59 → 61

Flow rate 1.3 mL/min.
Detection Spectrophotometer at 240 nm.
Injection 5 µL.
Identification of impurities Use the chromatogram supplied with *buprenorphine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, F, G, H and J.
Relative retention With reference to buprenorphine (retention time = about 8.5 min): impurity B = about 0.4; impurity J = about 1.1; impurity F = about 1.27; impurity H = about 1.33; impurity A = about 1.40; impurity G = about 1.8.
System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to buprenorphine and impurity J.
Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity G by 0.3;
— *impurity H*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);
— *impurities A, B, F, J*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity G*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.400 g in 40 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 46.76 mg of $C_{29}H_{41}NO_4$.

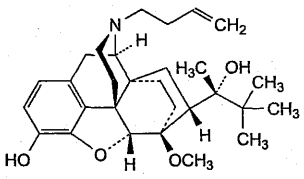
STORAGE

Protected from light.

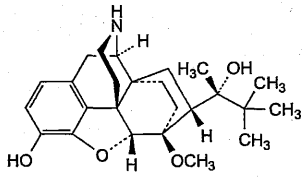
IMPURITIES

Specified impurities *A, B, F, G, H, J.*

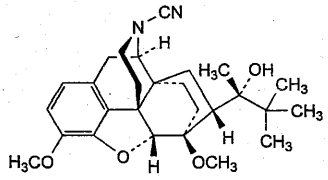
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, I.*



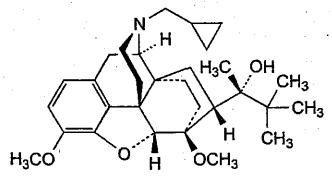
A. (2*S*)-2-[17-(but-3-enyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol,



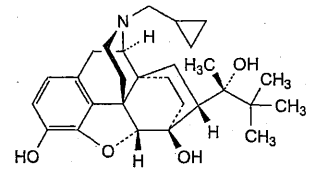
B. (2*S*)-2-(4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl)-3,3-dimethylbutan-2-ol (norbuprenorphine),



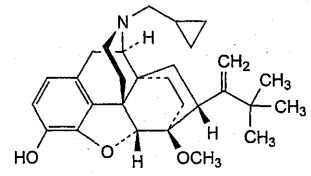
C. 4,5α-epoxy-7α-[(1*S*)-1-hydroxy-1,2,2-trimethylpropyl]-3,6-dimethoxy-6α,14-ethano-14α-morphinan-17-carbonitrile,



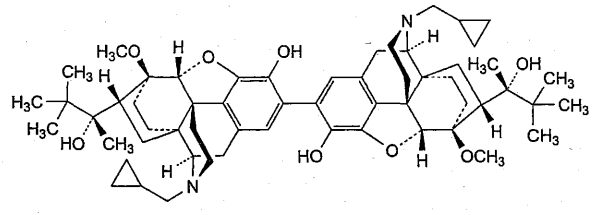
D. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3,6-dimethoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol (3-*O*-methylbuprenorphine),



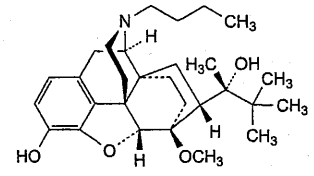
E. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3,6-dihydroxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol (6-*O*-desmethylbuprenorphine),



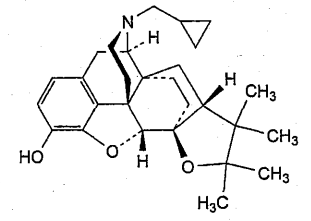
F. 17-(cyclopropylmethyl)-4,5α-epoxy-6-methoxy-7α-[1-(1,1-dimethylethyl)ethenyl]-6α,14-ethano-14α-morphinan-3-ol,



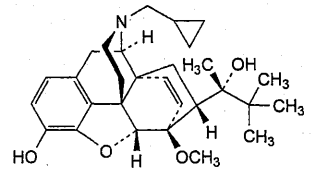
G. 17,17′-di(cyclopropylmethyl)-4,5α;4′,5α′-diepoxy-7α,7α′-di[(1*S*)-1-hydroxy-1,2,2-trimethylpropyl]-6,6′-dimethoxy-2,2′-bi(6α,14-ethano-14α-morphinan)-3,3′-diol (2,2′-bibuprenorphine),



H. (2*S*)-2-[17-butyl-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]3,3-dimethylbutan-2-ol,



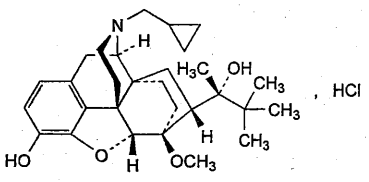
I. 17-(cyclopropylmethyl)-4″,4″,5″,5″-tetramethyl-4″,5″dihydro-(7β*H*)-6α,14-ethano-(5β*H*)-difurano[2′,3′,4′,5′:4,12,13,5;2″,3″:6,7]-14α-morphinan-3-ol,



J. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-etheno-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol.

Buprenorphine Hydrochloride

(Ph. Eur. monograph 1181)



$C_{29}H_{42}ClNO_4$ 504.1 *53152-21-9*

Action and use
Opioid receptor partial agonist; analgesic.

Preparations
Buprenorphine Injection
Buprenorphine Sublingual Tablets

Ph Eur

DEFINITION

(2*S*)-2-[17-(Cyclopropylmethyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol hydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *buprenorphine hydrochloride CRS*.

B. 3 mL of solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.250 g in 5.0 mL of *methanol R* and, while stirring, dilute to 25.0 mL with *carbon dioxide-free water R*.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10.0 mL of solution S add 0.05 mL of *methyl red solution R*. Not more than 0.2 mL of *0.02 M sodium hydroxide* or *0.02 M hydrochloric acid* is required to change the colour of the indicator.

Specific optical rotation (*2.2.7*)
−92 to −98 (dried substance).

Dissolve 0.200 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (b) Dissolve 5 mg of *buprenorphine for system suitability CRS* (containing impurities A, B, F, G, H and J) in 1.0 mL of *methanol R*.

Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 10 volumes of *acetonitrile R* and 90 volumes of the following solution: dissolve 5.44 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 4.5 with a 5 per cent *V/V* solution of *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	89	11
2 - 12	89 → 64	11 → 36
12 - 15	64 → 41	36 → 59
15 - 20	41 → 39	59 → 61

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *buprenorphine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, F, G, H and J.

Relative retention With reference to buprenorphine (retention time = about 8.5 min): impurity B = about 0.4; impurity J = about 1.1; impurity F = about 1.27; impurity H = about 1.33; impurity A = about 1.40; impurity G = about 1.8.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to buprenorphine and impurity J.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity G by 0.3;
— *impurity H*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);
— *impurities A, B, F, J*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity G*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by heating in an oven at 115-120 °C.

ASSAY

Dissolve 0.400 g in a mixture of 5 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R.* Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion. Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 50.41 mg of $C_{29}H_{42}ClNO_4$.

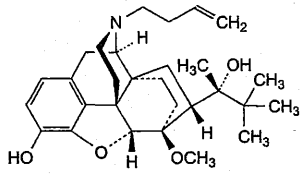
STORAGE

Protected from light.

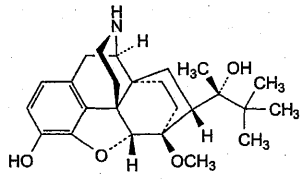
IMPURITIES

Specified impurities *A, B, F, G, H, J.*

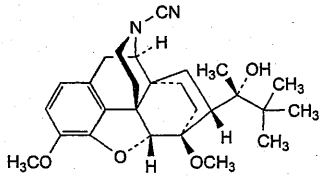
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, I.*



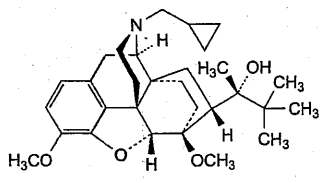
A. (2*S*)-2-[17-(but-3-enyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol,



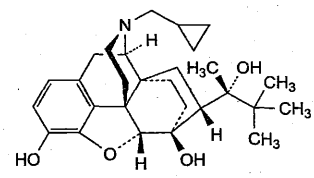
B. (2*S*)-2-(4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl)-3,3-dimethylbutan-2-ol (norbuprenorphine),



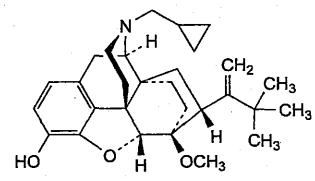
C. 4,5α-epoxy-7α-[(1*S*)-1-hydroxy-1,2,2-trimethylpropyl]-3,6-dimethoxy-6α,14-ethano-14α-morphinan-17-carbonitrile,



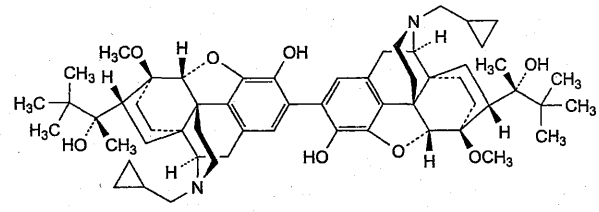
D. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3,6-dimethoxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol (3-*O*-methylbuprenorphine),



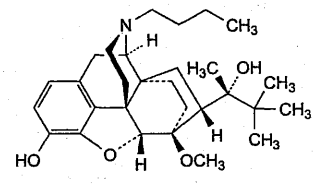
E. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3,6-dihydroxy-6α,14-ethano-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol (6-*O*-desmethylbuprenorphine),



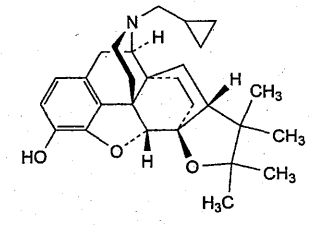
F. 17-(cyclopropylmethyl)-4,5α-epoxy-6-methoxy-7α-[1-(1,1-dimethylethyl)ethenyl]-6α,14-ethano-14α-morphinan-3-ol,



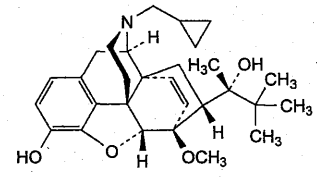
G. 17,17′-di(cyclopropylmethyl)-4,5α;4′,5α′-diepoxy-7α,7α′-di[(1*S*)-1-hydroxy-1,2,2-trimethylpropyl]-6,6′-dimethoxy-2,2′-bi(6α,14-ethano-14α-morphinan)-3,3′-diol (2,2′-bibuprenorphine),



H. (2*S*)-2-[17-butyl-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-ethano-14α-morphinan-7α-yl]3,3-dimethylbutan-2-ol,



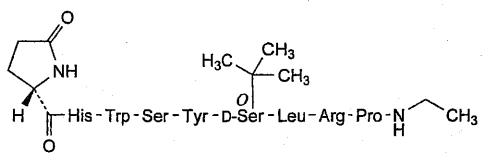
I. 17-(cyclopropylmethyl)-4″,4″,5″,5″-tetramethyl-4″,5″dihydro-(7β*H*)-6α,14-ethano-(5β*H*)-difurano[2′,3′,4′,5′:4,12,13,5;2″,3″:6,7]-14α-morphinan-3-ol,



J. (2*S*)-2-[17-(cyclopropylmethyl)-4,5α-epoxy-3-hydroxy-6-methoxy-6α,14-etheno-14α-morphinan-7α-yl]-3,3-dimethylbutan-2-ol.

Buserelin

(Ph. Eur. monograph 1077)



$C_{60}H_{86}N_{16}O_{13}$ 1239 *57982-77-1*

Action and use
Gonadotrophin releasing hormone (gonadorelin) analogue; treatment of prostate cancer.

Ph Eur

DEFINITION

5-Oxo-L-prolyl-L-histidyl-L-tryptophyl-L-seryl-L-tyrosyl-*O*-(1,1-dimethylethyl)-D-seryl-L-leucyl-L-arginyl-*N*-ethyl-L-prolinamide.

Synthetic nonapeptide analogue of human gonadotrophin-releasing hormone GnRH with agonistic activity to gonadorelin. It is obtained by chemical synthesis and is available as an acetate.

Content
95.0 per cent to 102.0 per cent (anhydrous, acetic acid-free substance).

CHARACTERS

Appearance
White or slightly yellowish powder, hygroscopic.

Solubility
Sparingly soluble in water and in dilute acids.

IDENTIFICATION

Carry out either tests A and B or tests A and C.

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (b).

B. Nuclear magnetic resonance spectrometry (*2.2.64*).

Preparation 4 mg/mL solution in a mixture of 20 volumes of *deuterated acetic acid R* and 80 volumes of *deuterium oxide R*.

Comparison Dissolve the contents of a vial of *buserelin for NMR identification CRS* in a mixture of 20 volumes of *deuterated acetic acid R* and 80 volumes of *deuterium oxide R* to obtain a concentration of 4 mg/mL.

Operating conditions:
— *field strength*: minimum 300 MHz;
— *temperature*: 27 °C.

Results Examine the ^{1}H NMR spectrum from 0 to 9 ppm. The ^{1}H NMR spectrum obtained is qualitatively similar to the ^{1}H NMR spectrum obtained with *buserelin for NMR identification CRS*.

C. Amino acid analysis (*2.2.56*). Method 1 for hydrolysis and method 1 for analysis are suitable.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids, taking 1/6 of the sum of the number of moles of glutamic acid, histidine, tyrosine, leucine, arginine and proline as equal to 1. The values fall within the following limits: serine 1.4 to 2.0; proline 0.8 to 1.2; glutamic acid 0.9 to 1.1; leucine 0.9 to 1.1; tyrosine 0.9 to 1.1; histidine 0.9 to 1.1; arginine 0.9 to 1.1. Not more than traces of other amino acids are present.

TESTS

Appearance of solution
A 10 g/L solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
−58 to −49 (anhydrous, acetic acid-free substance), determined on a 10 g/L solution.

Specific absorbance (*2.2.25*)
49 to 56, measured at the absorption maximum at 278 nm (anhydrous, acetic acid-free substance).

Dissolve 10.0 mg in 100.0 mL of *0.01 M hydrochloric acid*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 5.0 mg of the substance to be examined in 5.0 mL of the mobile phase.

Reference solution (a) Dissolve the contents of a vial of *D-His-buserelin CRS* (impurity A) in the mobile phase. Dilute an appropriate volume of this solution in the mobile phase to obtain a final concentration of 1 mg/mL. Add 1.0 mL of the test solution to 1.0 mL of this solution.

Reference solution (b) Dissolve the contents of a vial of *buserelin CRS* in the mobile phase. Dilute an appropriate volume of this solution in the mobile phase to obtain a final concentration of 1.0 mg/mL.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve the contents of a vial of *buserelin for peak identification CRS* (containing impurities F and G) in the mobile phase. Dilute an appropriate volume of this solution in the mobile phase to obtain a final concentration of 1 mg/mL.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 42 °C.

Mobile phase Mix 200 mL of *acetonitrile R* and 700 mL of an 11.2 g/L solution of *phosphoric acid R* previously adjusted to pH 2.5 with *triethylamine R*.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (a), (c) and (d).

Run time 60 min.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities F and G.

Relative retention With reference to buserelin (retention time = about 23 min): impurity F = about 0.83; impurity A = about 0.91; impurity G = about 1.26.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurity A and buserelin.

Calculation of percentage contents:
— for each impurity, use the concentration of buserelin in reference solution (c).

Limits:
— *impurity A*: maximum 2.5 per cent;
— *impurity F*: maximum 1.0 per cent;

— *impurity G*: maximum 1.0 per cent;
— *unspecified impurities*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 4.0 per cent;
— *reporting threshold*: 0.1 per cent.

Acetic acid (*2.5.34*)
3.0 per cent to 7.0 per cent.

Test solution Dissolve 20.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of solvents.

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 80.0 mg.

Bacterial endotoxins (*2.6.14*)
Less than 55.5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of buserelin ($C_{60}H_{86}N_{16}O_{13}$) taking into account the assigned content of $C_{60}H_{86}N_{16}O_{13}$ in *buserelin CRS*.

STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, the container is also sterile and tamper-proof.

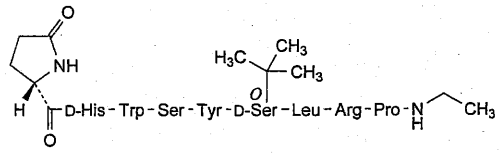
LABELLING

The label states:
— the mass of peptide in the container;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

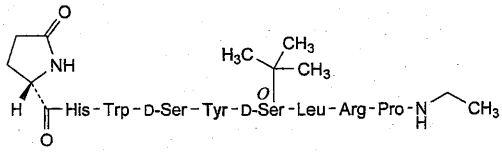
IMPURITIES

Specified impurities *A, F, G.*

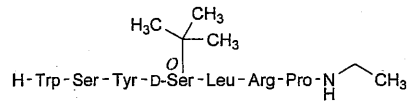
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E.*



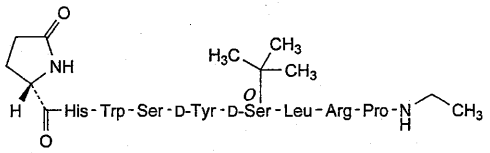
A. [2-D-histidine]buserelin,



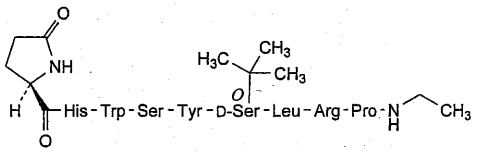
B. [4-D-serine]buserelin,



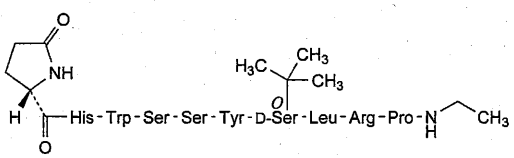
C. buserelin-(3-9)-peptide,



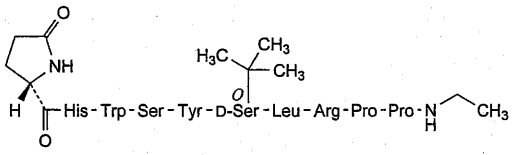
D. [5-D-tyrosine]buserelin,



E. [1-(5-oxo-D-proline)]buserelin,



F. endo-3a-serine-buserelin,

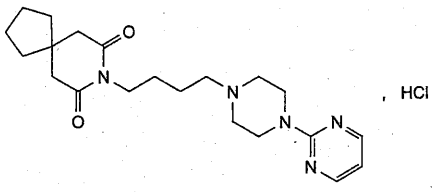


G. endo-8a-proline-buserelin.

Ph Eur

Buspirone Hydrochloride

(Ph. Eur. monograph 1711)



$C_{21}H_{32}ClN_5O_2$ 422.0 *33386-08-2*

Action and use
Non-benzodiazepine hypnotic; treatment of anxiety.

Ph Eur

DEFINITION

8-[4-[4-(Pyrimidin-2-yl)piperazin-1-yl]butyl]-8-azaspiro[4.5]decane-7,9-dione hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in methanol, practically insoluble in acetone.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *buspirone hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 25.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *buspirone for system suitability CRS* (containing impurities E, G, J, L and N) in 2.0 mL of mobile phase A and sonicate for 10 min.

Column:
- *size*: l = 0.15 m, Ø = 4.6 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm),
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: mix 950 volumes of a solution containing 6.8 g/L of *potassium dihydrogen phosphate R* and 0.93 g/L of *sodium hexanesulfonate monohydrate R*, previously adjusted to pH 3.4 with *phosphoric acid R* and 50 volumes of *acetonitrile R1*;
- *mobile phase B*: mix 250 volumes of a solution containing 3.4 g/L of *potassium dihydrogen phosphate R* and 3.52 g/L of *sodium hexanesulfonate monohydrate R*, previously adjusted to pH 2.2 with *phosphoric acid R* and 750 volumes of *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 6	90	10
6 - 34	90 → 42	10 → 58
34 - 45	42	58
45 - 55	42 → 0	58 → 100
55 - 56	0 → 100	100 → 0
56 - 60	100	0
60 - 61	100 → 90	0 → 10

Flow rate 1 mL/min.

Detection Variable wavelength spectrophotometer capable of operating at 240 nm and at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *buspirone for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities E, G, J, L and N.

Relative retention at 240 nm With reference to buspirone (retention time = about 25 min): impurity A = about 0.2; impurity B = about 0.3; impurity C = about 0.6; impurity D = about 0.7; impurity E = about 0.8; impurity F = about 0.9; impurity G = about 1.05; impurity H = about 1.1; impurity I = about 1.2; impurity J = about 1.5.

Relative retention at 210 nm With reference to buspirone (retention time = about 25 min): impurity K = about 0.6; impurity L = about 1.7; impurity M = about 1.8; impurity N = about 1.9.

System suitability Reference solution (b):
- *peak-to-valley ratio at 240 nm*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to buspirone;
- *resolution at 210 nm*: minimum 4.0 between the peaks due to impurity L and impurity N;
- the chromatograms obtained are similar to the chromatograms supplied with *buspirone for system suitability CRS*.

Limits Spectrophotometer at 240 nm:
- *correction factor*: for the calculation of content, multiply the peak area of impurity J by 2,
- *impurity E*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
- *impurity J*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
- *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent),
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Limits Spectrophotometer at 210 nm:
- *impurity K*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *any other impurity eluting with a relative retention greater than 1.6*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 10 mL of *glacial acetic acid R* and add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 21.10 mg of $C_{21}H_{32}ClN_5O_2$.

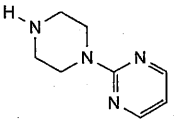
STORAGE

Protected from light.

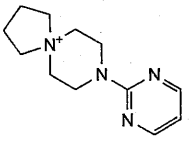
IMPURITIES

Specified impurities E, J, K.

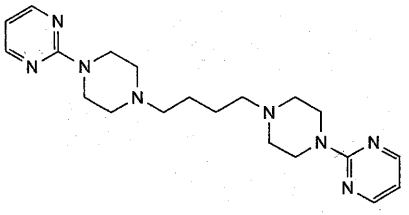
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, C, D, F, G, H, I, L, M, N.



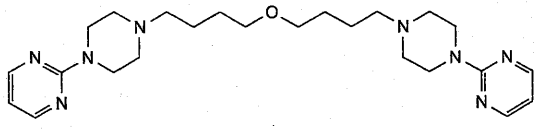
A. 2-(piperazin-1-yl)pyrimidine,



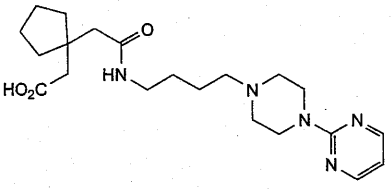
B. 8-(pyrimidin-2-yl)-8-aza-5-azoniaspiro[4.5]decane,



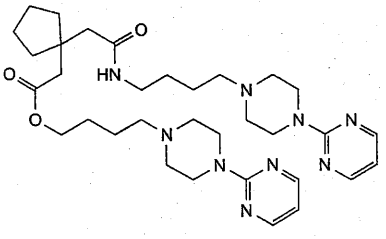
C. 2,2′-[butane-1,4-diylbis(piperazine-1,4-diyl)]dipyrimidine,



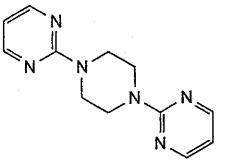
D. 2,2′-[oxybis[butane-1,4-diyl(piperazine-1,4-diyl)]] dipyrimidine,



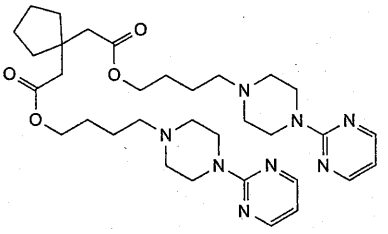
E. [1-[2-oxo-2-[[4-[4-(pyrimidin-2-yl)piperazin-1-yl]butyl] amino]ethyl]cyclopentyl]acetic acid,



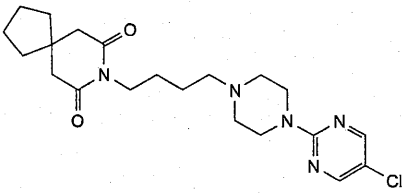
F. 4-[4-(pyrimidin-2-yl)piperazin-1-yl]butyl [1-[2-oxo-2-[[4-[4-(pyrimidin-2-yl)piperazin-1-yl]butyl]amino]ethyl] cyclopentyl]acetate,



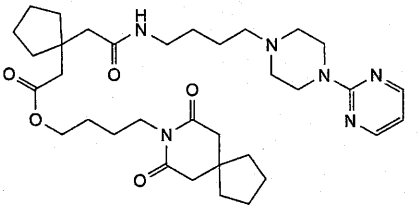
G. 2,2′-(piperazine-1,4-diyl)dipyrimidine,



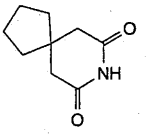
H. bis[4-[4-(pyrimidin-2-yl)piperazin-1-yl]butyl] (cyclopentane-1,1-diyl)diacetate,



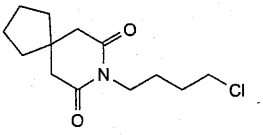
I. 8-[4-[4-(5-chloropyrimidin-2-yl)piperazin-1-yl]butyl]-8-azaspiro[4.5]decane-7,9-dione,



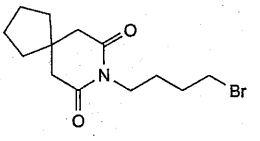
J. 4-(7,9-dioxo-8-azaspiro[4.5]dec-8-yl)butyl [1-[2-oxo-2-[[4-[4-(pyrimidin-2-yl)piperazin-1-yl]butyl]amino] ethyl]cyclopentyl]acetate,



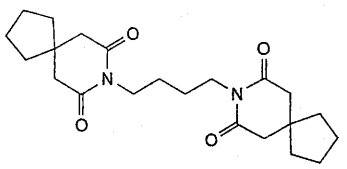
K. 8-azaspiro[4.5]decane-7,9-dione,



L. 8-(4-chlorobutyl)-8-azaspiro[4.5]decane-7,9-dione,



M. 8-(4-bromobutyl)-8-azaspiro[4.5]decane-7,9-dione,

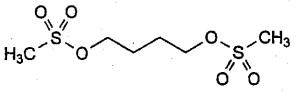


N. 8,8′-(butane-1,4-diyl)bis(8-azaspiro[4.5]decane-7,9-dione).

Ph Eur

Busulfan

(Ph. Eur. monograph 0542)



$C_6H_{14}O_6S_2$ 246.3 *55-98-1*

Action and use
Cytotoxic alkylating agent.

Preparation
Busulfan Tablets

Ph Eur

DEFINITION
Butane-1,4-diyl di(methanesulfonate).

Content
99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, freely soluble in acetone and in acetonitrile, very slightly soluble in ethanol (96 per cent).

mp
About 116 °C.

IDENTIFICATION
First identification: A.
Second identification: B, C, D.
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *busulfan CRS.*
B. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 20 mg of the substance to be examined in 2 mL of *acetone R.*
Reference solution Dissolve 20 mg of *busulfan CRS* in 2 mL of *acetone R.*
Plate *TLC silica gel G plate R.*
Mobile phase *acetone R, toluene R* (50:50 *V/V*).
Application 5 µL.
Development Over a path of 15 cm.
Drying In a current of warm air.
Detection Spray with *anisaldehyde solution R* and heat at 120 °C.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 0.1 g add 5 mL of *1 M sodium hydroxide.* Heat until a clear solution is obtained. Allow to cool. To 2 mL of the solution add 0.1 mL of *potassium permanganate solution R.* The colour changes from purple through violet to blue and finally to green. Filter and add 1 mL of *ammoniacal silver nitrate solution R.* A precipitate is formed.

D. To 0.1 g add 0.1 g of *potassium nitrate R* and 0.25 g of *sodium hydroxide R,* mix and heat to fusion. Allow to cool and dissolve the residue in 5 mL of *water R.* Adjust to pH 1-2 using *dilute hydrochloric acid R.* The solution gives reaction (a) of sulfates (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).
Dissolve 0.25 g in 20 mL of *acetonitrile R,* dilute to 25 mL with *water R* and examine immediately.

Acidity
Dissolve 0.20 g with heating in 50 mL of *anhydrous ethanol R.* Add 0.1 mL of *methyl red solution R.* Not more than 0.05 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

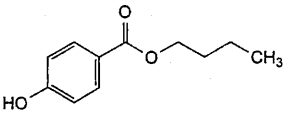
ASSAY
To 0.250 g add 50 mL of *water R.* Shake. Boil under a reflux condenser for 30 min and, if necessary, make up to the initial volume with *water R.* Allow to cool. Using 0.3 mL of *phenolphthalein solution R* as indicator, titrate with *0.1 M sodium hydroxide* until a pink colour is obtained.
1 mL of *0.1 M sodium hydroxide* is equivalent to 12.32 mg of $C_6H_{14}O_6S_2$.

STORAGE
In an airtight container, protected from light.

Ph Eur

Butyl Hydroxybenzoate

Butylparaben
(Butyl Parahydroxybenzoate, Ph. Eur. monograph 0881)



$C_{11}H_{14}O_3$ 194.2 *94-26-8*

Action and use
Excipient.

Ph Eur

DEFINITION
Butyl 4-hydroxybenzoate.

Content
98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification: A, B.

Second identification: A, C.

A. Melting point (*2.2.14*): 68 °C to 71 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *butyl parahydroxybenzoate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.10 g of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *acetone R.*

Reference solution (a) Dissolve 10 mg of *butyl parahydroxybenzoate CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *propyl parahydroxybenzoate R* in 1 mL of test solution (a) and dilute to 10 mL with *acetone R.*

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, methanol R* (1:30:70 *V/V/V*).

Application 2 µL of test solution (b) and reference solutions (a) and (b).

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 1.0 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity

To 2 mL of solution S add 3 mL of *ethanol (96 per cent) R*, 5 mL of *carbon dioxide-free water R* and 0.1 mL of *bromocresol green solution R.* Not more than 0.1 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *4-hydroxybenzoic acid R* (impurity A), 5 mg of *propyl parahydroxybenzoate R* (impurity D) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of *butyl parahydroxybenzoate CRS* in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 5 mg of *butyl parahydroxybenzoate impurity E CRS* (iso-butyl parahydroxybenzoate) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (e) Dilute 0.5 mL of reference solution (d) to 50.0 mL with reference solution (b).

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 35 °C.

Mobile phase 6.8 g/L solution of *potassium dihydrogen phosphate R, methanol R* (50:50 *V/V*).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 µL of the test solution and reference solutions (a), (c) and (e).

Run time 1.5 times the retention time of butyl parahydroxybenzoate.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and D; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity E.

Relative retention With reference to butyl parahydroxybenzoate (retention time = about 22 min): impurity A = about 0.1; impurity D = about 0.5; impurity E = about 0.9.

System suitability:

— *resolution*:

  — minimum 5.0 between the peaks due to impurity D and butyl parahydroxybenzoate in the chromatogram obtained with reference solution (a);

  — minimum 1.5 between the peaks due to impurity E and butyl parahydroxybenzoate in the chromatogram obtained with reference solution (e).

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

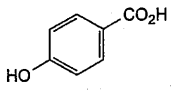
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

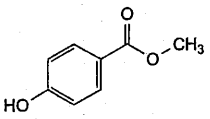
Calculate the percentage content of $C_{11}H_{14}O_3$ from the declared content of *butyl parahydroxybenzoate CRS*.

IMPURITIES
Specified impurities *A.*

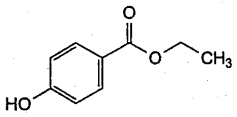
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E.*



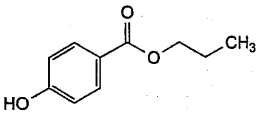
A. 4-hydroxybenzoic acid,



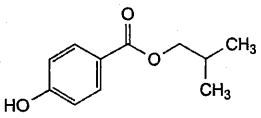
B. methyl 4-hydroxybenzoate (methyl parahydroxybenzoate),



C. ethyl 4-hydroxybenzoate (ethyl parahydroxybenzoate),



D. propyl 4-hydroxybenzoate (propyl parahydroxybenzoate),

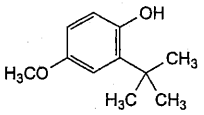


E. 2-methylpropyl 4-hydroxybenzoate (iso-butyl parahydroxybenzoate).

Ph Eur

Butylated Hydroxyanisole

(Butylhydroxanisole, Ph. Eur. monograph 0880)



$C_{11}H_{16}O_2$ 180.3 *25013-16-5*

Action and use
Antioxidant.

Ph Eur

DEFINITION
Butylhydroxyanisole is 2-(1,1-dimethylethyl)-4-methoxyphenol containing not more than 10 per cent of 3-(1,1-dimethylethyl)-4-methoxyphenol.

CHARACTERS
A white, yellowish or slightly pinkish, crystalline powder, practically insoluble in water, very soluble in methylene chloride, freely soluble in alcohol and in fatty oils. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
A. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To 0.5 mL of solution S (see Tests) add 10 mL of *aminopyrazolone solution R* and 1 mL of *potassium ferricyanide solution R*. Mix and add 10 mL of *methylene chloride R*. Shake vigorously. After separation, the organic layer is red.

C. Dissolve about 10 mg in 2 mL of *alcohol R*. Add 1 mL of a 1 g/L solution of *testosterone propionate R* in *alcohol R* and 2 mL of *dilute sodium hydroxide solution R*. Heat in a water-bath at 80 °C for 10 min and allow to cool. A red colour develops.

TESTS
Solution S
Dissolve 2.5 g in *alcohol R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.25 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methylene chloride R*.

Reference solution (a) Dissolve 25 mg of *butylhydroxyanisole CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of reference solution (a) to 20 mL with *methylene chloride R*.

Reference solution (c) Dissolve 50 mg of *hydroquinone R* in 5 mL of *alcohol R* and dilute to 100 mL with *methylene*

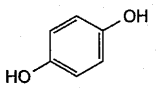
chloride R. Dilute 1 mL of this solution to 10 mL with *methylene chloride R*.

Apply separately to the plate 5 µL of each solution. Develop over a path of 10 cm using *methylene chloride R*. Allow the plate to dry in air and spray with a freshly prepared mixture of 10 volumes of *potassium ferricyanide solution R*, 20 volumes of *ferric chloride solution R1* and 70 volumes of *water R*. In the chromatogram obtained with test solution (a): any violet-blue spot with an R_F value of about 0.35 (corresponding to 3-(1,1-dimethylethyl)-4-methoxyphenol) is not more intense than the principal spot in the chromatogram obtained with reference solution (a) (10 per cent); any spot corresponding to hydroquinone is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.2 per cent); any spot, apart from the principal spot and any spots corresponding to 3-(1,1-dimethylethyl)-4-methoxyphenol and hydroquinone, is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

STORAGE
Store protected from light.

IMPURITIES

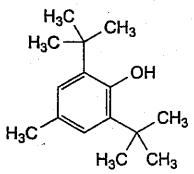


A. benzene-1,4-diol (hydroquinone).

Ph Eur

Butylated Hydroxytoluene

(Butylhydroxytoluene, Ph. Eur. monograph 0581)



$C_{15}H_{24}O$ 220.4 *128-37-0*

Action and use
Antioxidant.

Ph Eur

DEFINITION
Butylhydroxytoluene is 2,6-bis(1,1-dimethylethyl)-4-methylphenol.

CHARACTERS
A white or yellowish-white, crystalline powder, practically insoluble in water, very soluble in acetone, freely soluble in alcohol and in vegetable oils.

IDENTIFICATION
First identification: A, C.
Second identification: A, B, D.

A. Freezing-point (see Tests).

B. Dissolve 0.500 g in *ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *ethanol R*. Examined between 230 nm and 300 nm (*2.2.25*), the solution shows an absorption maximum at 278 nm. The specific absorbance at the maximum is 80 to 90.

C. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *butylhydroxytoluene CRS*.

D. Dissolve about 10 mg in 2 mL of *alcohol R*. Add 1 mL of a 1 g/L solution of *testosterone propionate R* in *alcohol R* and 2 mL of *dilute sodium hydroxide solution R*. Heat in a water-bath at 80 °C for 10 min and allow to cool. A blue colour develops.

TESTS

Appearance of solution
Dissolve 1.0 g in *methanol R* and dilute to 10 mL with the same solvent. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 or BY_5 (*2.2.2, Method II*).

Freezing-point (*2.2.18*)
69 °C to 70 °C.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution Dissolve 0.2 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution Dilute 1 mL of the test solution to 200 mL with *methanol R*.

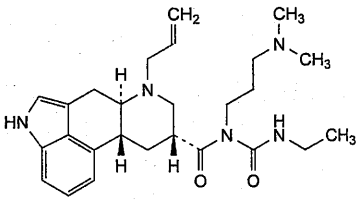
Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using *methylene chloride R*. Dry the plate in air and spray with a freshly prepared mixture of 10 volumes of *potassium ferricyanide solution R*, 20 volumes of *ferric chloride solution R1* and 70 volumes of *water R*. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

Ph Eur

Cabergoline

(Ph. Eur. monograph 1773)



$C_{26}H_{37}N_5O_2$ 451.6 *81409-90-7*

Action and use
Dopamine D2 receptor agonist.

Preparation
Cabergoline Tablets

Ph Eur

DEFINITION
1-Ethyl-3-[3-(dimethylamino)propyl]-3-[[(6a*R*,9*R*,10a*R*)-7-(prop-2-enyl)-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinolin-9-yl]carbonyl]urea.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), very slightly soluble in hexane. It is slightly soluble in 0.1 M hydrochloric acid.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cabergoline CRS*.

If the spectra obtained in the solid state show differences, dissolve 50 mg of the substance to be examined and 50 mg of the reference substance separately in 1 mL of *ethanol (96 per cent) R*, evaporate to dryness and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)
−77 to −83 (anhydrous substance).

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protected from light.*

Test solution Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 30.0 mg of *cabergoline CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Suspend 50 mg of the substance to be examined in 10 mL of *0.1 M sodium hydroxide*. Stir for about 15 min. To 1 mL of the suspension add 1 mL of *0.1 M hydrochloric acid* and dilute to 10 mL with the mobile phase. Sonicate until dissolution is complete. The main degradation product obtained is impurity A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase Mix 16 volumes of *acetonitrile R* and 84 volumes of a freshly prepared 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 2.0 with *phosphoric acid R*. Add 0.2 volumes of *triethylamine R*.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 4 times the retention time of cabergoline.

Relative retention With reference to cabergoline (retention time = about 12 min): impurity D = about 0.3; impurity B = about 0.6; impurity A = about 0.8; impurity C = about 2.9.

System suitability Reference solution (c):
— *resolution*: minimum 3.0 between the peaks due to cabergoline and impurity A.

Limits:
— *impurities A, C*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurities B, D*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

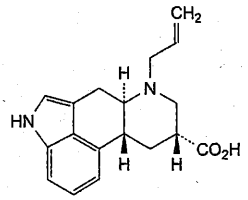
Calculate the percentage content of $C_{26}H_{37}N_5O_2$ from the areas of the peaks and the declared content of *cabergoline CRS*.

STORAGE

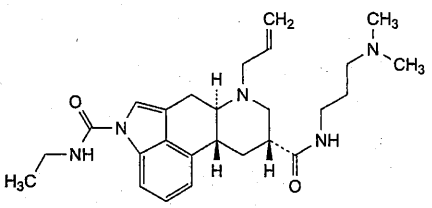
Protected from light.

IMPURITIES

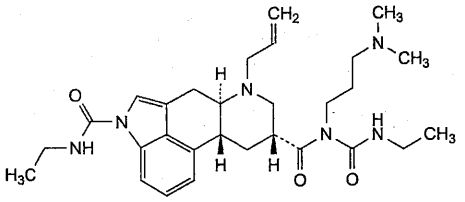
Specified impurities *A, B, C, D.*



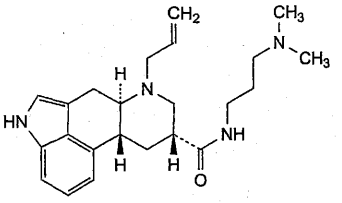
A. (6a*R*,9*R*,10a*R*)-7-(prop-2-enyl)-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxylic acid,



B. (6a*R*,9*R*,10a*R*)-N^9-[3-(dimethylamino)propyl]-N^4-ethyl-7-(prop-2-enyl)-6a,7,8,9,10,10a-hexahydroindolo[4,3-*fg*]quinoline-4,9(6*H*)-dicarboxamide,



C. (6a*R*,9*R*,10a*R*)-N^9-[3-(dimethylamino)propyl]-N^4-ethyl-N^9-(ethylcarbamoyl)-7-(prop-2-enyl)-6a,7,8,9,10,10a-hexahydroindolo[4,3-*fg*]quinoline-4,9(6*H*)-dicarboxamide,



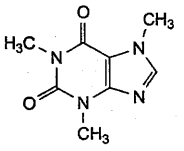
D. (6a*R*,9*R*,10a*R*)-*N*-[3-(dimethylamino)propyl]-7-(prop-2-enyl)-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide.

Ph Eur

Caffeine

Anhydrous Caffeine

(Ph. Eur. monograph 0267)



$C_8H_{10}N_4O_2$ 194.2 *58-08-2*

Action and use
Central nervous system stimulant.

Preparations
Aspirin and Caffeine Tablets
Caffeine Citrate Injection
Caffeine Citrate Oral Solution
Paracetamol and Caffeine Capsules
Paracetamol and Caffeine Tablets
Paracetamol, Codeine Phosphate and Caffeine Capsules
Paracetamol, Codeine Phosphate and Caffeine Tablets

Ph Eur

DEFINITION
1,3,7-Trimethyl-3,7-dihydro-1*H*-purine-2,6-dione.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or silky, white or almost white, crystals.

Solubility
Sparingly soluble in water, freely soluble in boiling water, slightly soluble in ethanol (96 per cent). It dissolves in concentrated solutions of alkali benzoates or salicylates. It sublimes readily.

IDENTIFICATION
First identification: A, B, E.

Second identification: A, C, D, E, F.

A. Melting point (*2.2.14*): 234 °C to 239 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *caffeine CRS.*

C. To 2 mL of a saturated solution add 0.05 mL of *iodinated potassium iodide solution R*. The solution remains clear. Add 0.1 mL of *dilute hydrochloric acid R*; a brown precipitate is formed. Neutralise with *dilute sodium hydroxide solution R*; the precipitate dissolves.

D. In a ground-glass-stoppered tube, dissolve about 10 mg in 0.25 mL of a mixture of 0.5 mL of *acetylacetone R* and 5 mL of *dilute sodium hydroxide solution R*. Heat in a water-bath at 80 °C for 7 min. Cool and add 0.5 mL of *dimethylaminobenzaldehyde solution R2*. Heat again in a water-bath at 80 °C for 7 min. Allow to cool and add 10 mL of *water R*; an intense blue colour develops.

E. Loss on drying (see Tests).

F. It gives the reaction of xanthines (*2.3.1*).

TESTS
Solution S
Dissolve 0.5 g with heating in 30 mL of *carbon dioxide-free water R* prepared from *distilled water R*, cool and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity
To 10 mL of solution S add 0.05 mL of *bromothymol blue solution R1*; the solution is green or yellow. Not more than 0.2 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *caffeine for system suitability CRS* (containing impurities A, C, D and F) in the mobile phase and dilute to 5 mL with the mobile phase. Dilute 2 mL of this solution to 10 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 20 volumes of *tetrahydrofuran R*, 25 volumes of *acetonitrile R* and 955 volumes of a solution containing 0.82 g/L of *anhydrous sodium acetate R* previously adjusted to pH 4.5 with *glacial acetic acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL.

Run time 1.5 times the retention time of caffeine.

Identification of impurities Use the chromatogram supplied with *caffeine for system suitability CRS* and the chromatogram

obtained with reference solution (b) to identify the peaks due to impurities A, C, D and F.

Retention time Caffeine = about 8 min.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities C and D and minimum 2.5 between the peaks due to impurities F and A.

Limits:
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

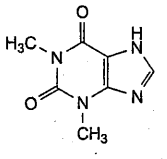
Sulfates (*2.4.13*)
Maximum 500 ppm, determined on 15 mL of solution S.
Prepare the standard using a mixture of 7.5 mL of *sulfate standard solution (10 ppm SO_4) R* and 7.5 mL of *distilled water R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

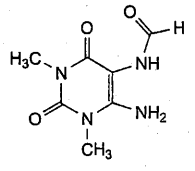
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.170 g with heating in 5 mL of *anhydrous acetic acid R*. Allow to cool, add 10 mL of *acetic anhydride R* and 20 mL of *toluene R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M perchloric acid* is equivalent to 19.42 mg of $C_8H_{10}N_4O_2$.

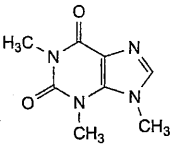
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F.*



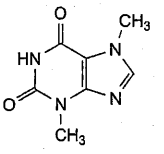
A. 1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theophylline),



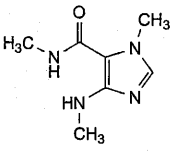
B. *N*-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide,



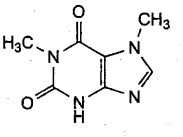
C. 1,3,9-trimethyl-3,9-dihydro-1*H*-purine-2,6-dione (isocaffeine),



D. 3,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theobromine),



E. *N*,1-dimethyl-4-(methylamino)-1*H*-imidazole-5-carboxamide (caffeidine),

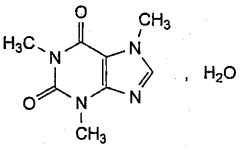


F. 1,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione.

Ph Eur

Caffeine Hydrate

(Caffeine Monohydrate, Ph. Eur. monograph 0268)



$C_8H_{10}N_4O_2,H_2O$ 212.2 *5743-12-4*

Action and use
Central nervous system stimulant.

Ph Eur

DEFINITION
1,3,7-Trimethyl-3,7-dihydro-1*H*-purine-2,6-dione monohydrate.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or silky, white or almost white crystals.

Solubility
Sparingly soluble in water, freely soluble in boiling water, slightly soluble in ethanol (96 per cent). It dissolves in concentrated solutions of alkali benzoates or salicylates.

It sublimes readily.

IDENTIFICATION

First identification: A, B, E.

Second identification: A, C, D, E, F.

A. Melting point (*2.2.14*): 234 °C to 239 °C, determined after drying at 100-105 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dry the substance to be examined at 100-105 °C before use.

Comparison *caffeine CRS.*

C. To 2 mL of a saturated solution add 0.05 mL of *iodinated potassium iodide solution R*; the solution remains clear. Add 0.1 mL of *dilute hydrochloric acid R*; a brown precipitate is formed. Neutralise with *dilute sodium hydroxide solution R*; the precipitate dissolves.

D. In a glass-stoppered tube, dissolve about 10 mg in 0.25 mL of a mixture of 0.5 mL of *acetylacetone R* and 5 mL of *dilute sodium hydroxide solution R*. Heat in a water-bath at 80 °C for 7 min. Cool and add 0.5 mL of *dimethylaminobenzaldehyde solution R2*. Heat again in a water-bath at 80 °C for 7 min. Allow to cool and add 10 mL of *water R*; an intense blue colour develops.

E. Loss on drying (see Tests).

F. It gives the reaction of xanthines (*2.3.1*).

TESTS

Solution S
Dissolve 0.5 g with heating in 30 mL of *carbon dioxide-free water R* prepared from *distilled water R*, cool, and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity
To 10 mL of solution S add 0.05 mL of *bromothymol blue solution R1*; the solution is green or yellow. Not more than 0.2 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.110 g of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *caffeine for system suitability CRS* (containing impurities A, C, D and F) in the mobile phase and dilute to 5.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 20 volumes of *tetrahydrofuran R*, 25 volumes of *acetonitrile R* and 955 volumes of a solution containing 0.82 g/L of *anhydrous sodium acetate R* previously adjusted to pH 4.5 with *glacial acetic acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL.

Run time 1.5 times the retention time of caffeine.

Identification of impurities Use the chromatogram supplied with *caffeine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, C, D and F.

Retention time Caffeine = about 8 min.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities C and D; minimum 2.5 between the peaks due to impurities F and A.

Limits:
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfates (*2.4.13*)
Maximum 500 ppm, determined on 15 mL of solution S.

Prepare the standard using a mixture of 7.5 mL of *sulfate standard solution (10 ppm SO_4) R* and 7.5 mL of *distilled water R*.

Loss on drying (*2.2.32*)
5.0 per cent to 9.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

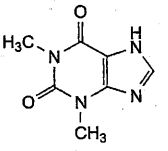
ASSAY

Dissolve 0.170 g, previously dried at 100-105 °C, with heating in 5 mL of *anhydrous acetic acid R*. Allow to cool, and add 10 mL of *acetic anhydride R* and 20 mL of *toluene R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

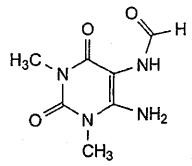
1 mL of *0.1 M perchloric acid* is equivalent to 19.42 mg of $C_8H_{10}N_4O_2$.

IMPURITIES

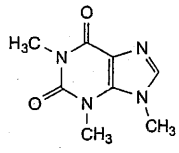
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F.*



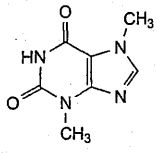
A. 1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theophylline),



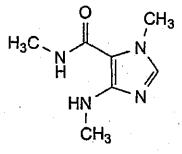
B. *N*-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide,



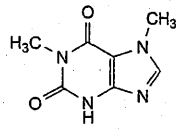
C. 1,3,9-trimethyl-3,9-dihydro-1*H*-purine-2,6-dione (isocaffeine),



D. 3,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theobromine),



E. *N*,1-dimethyl-4-(methylamino)-1*H*-imidazole-5-carboxamide (caffeidine),



F. 1,7-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione.

Ph Eur

Calamine

Prepared Calamine

Action and use
Antipruritic.

Preparations
Aqueous Calamine Cream
Calamine Lotion
Calamine Ointment
Calamine and Coal Tar Ointment

DEFINITION
Calamine is a basic zinc carbonate suitably coloured with iron(III) oxide.

CHARACTERISTICS
An amorphous, impalpable, pink or reddish brown powder, the colour depending on the variety and amount of iron(III) oxide present and the process by which it is incorporated.

Practically insoluble in *water*. It dissolves with effervescence in *hydrochloric acid*.

IDENTIFICATION
A. Yields the reactions characteristic of *carbonates*, Appendix VI.

B. To 2 g add 5 mL of *hydrochloric acid* and heat to boiling; if necessary, add *hydrochloric acid* drop wise until a bright yellow solution is obtained. Cool and add *13.5M ammonia* until the first sign of precipitate (solution A). The solution yields reaction B characteristic of *iron salts*, Appendix VI. Dilute 1 mL of solution A to 5 mL with *water*; the solution yields the reaction characteristic of *zinc salts*, Appendix VI.

TESTS

Calcium
Dissolve 0.50 g in a mixture of 10 mL of *water* and 2.5 mL of *glacial acetic acid* and filter. To 0.5 mL of the filtrate add 15 mL of *5M ammonia* and 2 mL of a 2.5% w/v solution of *ammonium oxalate* and allow to stand for 2 minutes. The solution remains clear.

Soluble barium salts
To the remainder of the filtrate obtained in the test for Calcium add 2 mL of *1M sulfuric acid* and allow to stand for 5 minutes. The solution remains clear.

Lead
Not more than 150 ppm when determined by *atomic absorption spectrophotometry*, Appendix II D, Method II, measuring at 283.3 nm or 217 nm and using an air-acetylene flame. Carefully add 5 g of the substance being examined to 25 mL of *hydrochloric acid* and allow to stand for 18 hours. Add 5 mL of *nitric acid* and sufficient *water* to produce 200 mL. Use *lead standard solution (100 ppm Pb)* suitably diluted with a 3.5% v/v solution of *nitric acid* to prepare the standard solution.

Chloride
Dissolve 0.15 g in *water* with the addition of 1 mL of *nitric acid*, filter and dilute to 30 mL with *water*. The resulting solution complies with the *limit test for chlorides*, Appendix VII (0.07%).

Sulfate
Dissolve 0.1 g in *water* with the addition of 3 mL of *2M hydrochloric acid*, filter and dilute to 60 mL with *water*. The resulting solution complies with the *limit test for sulfates*, Appendix VII (0.6%).

Ethanol-soluble dyes
Shake 1.0 g with 10 mL of *ethanol (90%)* and filter. The filtrate is *colourless*, Appendix IV B, Method II.

Matter insoluble in hydrochloric acid
Dissolve 1 g in 20 mL of warm *2M hydrochloric acid* and filter. The residue, when washed with *water* and dried to constant weight at 105°, weighs not more than 10 mg.

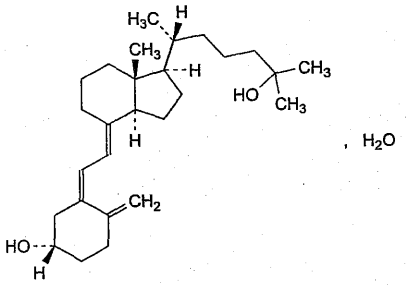
Water-soluble dyes
Shake 1.0 g with 10 mL of *water* and filter. The filtrate is *colourless*, Appendix IV B, Method II.

Residue on ignition
68.0 to 74.0%, when ignited at a temperature not lower than 900° until, after further ignition, two successive weighings do not differ by more than 0.2% of the weight of the residue. Use 1 g.

Calcifediol Monohydrate

Calcifediol

(Ph. Eur. monograph 1295)



$C_{27}H_{44}O_2,H_2O$ 418.7 *63283-36-3*

Action and use
Vitamin D analogue.

Ph Eur

DEFINITION

(3*S*,5*Z*,7*E*)-9,10-Secocholesta-5,7,10(19)-triene-3,25-diol monohydrate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

A reversible isomerisation to pre-calcifediol takes place in solution, depending on temperature and time. The activity is due to both compounds (see Assay).

CHARACTERS

Appearance
White or almost white crystals.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in fatty oils.

It is sensitive to air, heat and light.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of calcifediol.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Carry out the test as rapidly as possible, avoiding exposure to actinic light and air.*

Test solution Dissolve 5.0 mg of the substance to be examined without heating in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *calcifediol CRS* without heating in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Heat 2 mL of reference solution (a) in a water-bath at 80 °C under a reflux condenser for 2 h and cool.

Reference solution (d) Dissolve 1 mg of *calcifediol for impurity D identification CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water for chromatography R, methanol R* (20:80 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 50 µL of the test solution and reference solutions (b), (c) and (d).

Run time Twice the retention time of calcifediol.

Identification of impurities Use the chromatogram supplied with *calcifediol for impurity D identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity D.

Relative retention With reference to calcifediol (retention time = about 11 min): pre-calcifediol = about 1.3; impurity D = about 1.7.

System suitability Reference solution (c):
— *resolution*: minimum 5.0 between the peaks due to calcifediol and pre-calcifediol.

Limits:
— *impurity D*: maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent (0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)); disregard the peak due to pre-calcifediol.

Water (*2.5.32*)
3.8 per cent to 5.0 per cent, determined on 10.0 mg by direct introduction of the sample.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{27}H_{44}O_2$ taking into account the assigned content of *calcifediol CRS* and, if necessary, the peak due to pre-calcifediol.

STORAGE

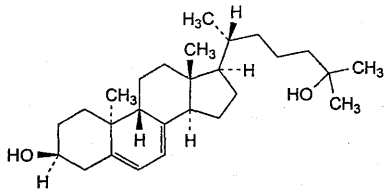
Under nitrogen, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

The contents of an opened container are to be used immediately.

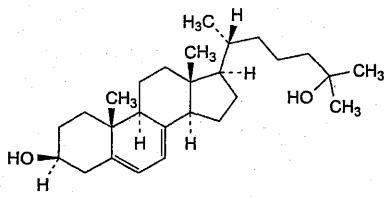
IMPURITIES

Specified impurities *D.*

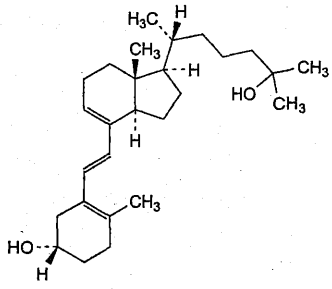
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



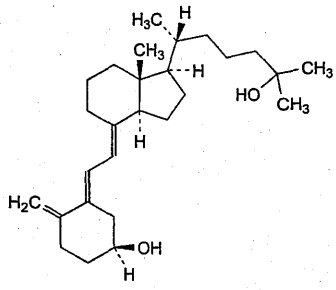
A. 9β,10α-cholesta-5,7-diene-3β,25-diol,



B. cholesta-5,7-diene-3β,25-diol,



C. (3*S*,6*E*)-9,10-secocholesta-5(10),6,8-triene-3,25-diol,



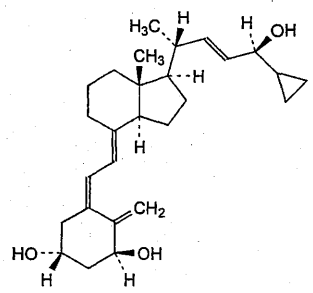
D. (3*S*,5*E*,7*E*)-9,10-secocholesta-5,7,10(19)-triene-3,25-diol.

Ph Eur

Calcipotriol

Anhydrous Calcipotriol

(Ph. Eur. monograph 2011)



$C_{27}H_{40}O_3$ 412.6 *112965-21-6*

Action and use
Vitamin D analogue.

Preparations
Calcipotriol Cream
Calcipotriol Ointment
Calcipotriol Scalp Application

Ph Eur

DEFINITION
(5*Z*,7*E*,22*E*,24*S*)-24-Cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol.

Content
95.5 per cent to 102.0 per cent (dried substance).

A reversible isomerisation to pre-calcipotriol takes place in solution, depending on temperature and time. The activity is due to both compounds.

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It is sensitive to heat and light.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of anhydrous calcipotriol.*

B. Loss on drying (see Tests).

TESTS
Carry out the tests for related substances and the assay as rapidly as possible and protected from actinic light and air.

Related substances
A. Thin-layer chromatography (*2.2.27*).

Solution A To 1 mL of *triethylamine R* add 9 mL of *chloroform R*.

Test solution Dissolve 1 mg of the substance to be examined in 100 µL of solution A.

Reference solution (a) To 10 µL of the test solution add 990 µL of solution A.

Reference solution (b) To 250 µL of reference solution (a) add 750 µL of solution A.

Reference solution (c) To 100 µL of reference solution (a) add 900 µL of solution A.

Reference solution (d) Place 2 mg of the substance to be examined in a vial and dissolve in 200 µL of solution A. Close the vial and keep it in a water-bath at 60 °C for 2 h.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *2-methylpropanol R, methylene chloride R* (20:80 *V/V*).

Application 10 µL of the test solution and reference solutions (b), (c) and (d).

Development Over 2/3 of the plate.

Drying In air, then at 140 °C for 10 min.

Detection Spray the hot plate with an *alcoholic solution of sulfuric acid R*, dry at 140 °C for not more than 1 min and examine in ultraviolet light at 366 nm.

Relative retention With reference to calcipotriol (R_F = about 0.4): impurity G = about 0.4; impurity H = about 0.4; pre-calcipotriol = about 0.9; impurity A = about 1.2.

System suitability Reference solution (d):

— the chromatogram shows a secondary spot due to pre-calcipotriol.

Limits:

— *impurity A*: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent);

— *impurities G, H*: any spot due to impurity G or H is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent for the sum);

— *unspecified impurities*: any other spot is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.1 per cent).

B. Liquid chromatography (*2.2.29*).

Solution A Dissolve 1.32 g of *ammonium phosphate R* in *water R* and dilute to 10.0 mL with the same solvent.

Solvent mixture Solution A, *water R*, *methanol R* (0.3:29.7:70 *V/V/V*).

Test solution (a) Dissolve 2 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Test solution (b) Dissolve 2.00 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 1 mg of *calcipotriol for system suitability CRS* (containing impurities B, C and D) in the solvent mixture and dilute to 2.5 mL with the solvent mixture.

Reference solution (d) Dissolve 2.00 mg of *calcipotriol monohydrate CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.10 m, Ø = 4.0 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase *water for chromatography R*, *methanol R* (30:70 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 264 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Run time Twice the retention time of calcipotriol.

Identification of impurities Use the chromatogram supplied with *calcipotriol for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C and D.

Relative retention With reference to calcipotriol (retention time = about 13.5 min): impurity B = about 0.86; impurity C = about 0.92; impurity D = about 1.3.

System suitability Reference solution (c):

— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to calcipotriol.

Limits:

— *impurity B*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *impurities C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (2.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying

Maximum 1.0 per cent, determined on 5 mg by thermogravimetry (*2.2.34*). Heat to 105 °C at a rate of 10 °C/min and maintain at 105 °C for 60 min.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{27}H_{40}O_3$ taking into account the assigned content of *calcipotriol monohydrate CRS* and, if necessary, the peak due to pre-calcipotriol.

STORAGE

In an airtight container, protected from light, at −20 °C or below.

IMPURITIES

Test A for related substances

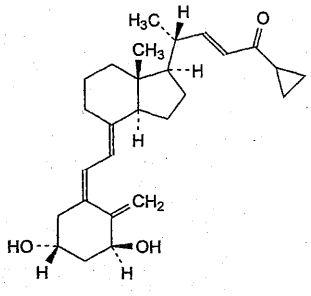
A, G, H, I.

Test B for related substances

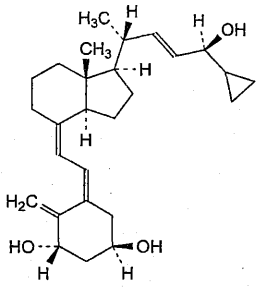
B, C, D, E, F.

Specified impurities *A, B, C, D, G, H.*

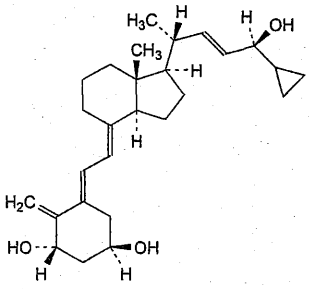
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F, I.*



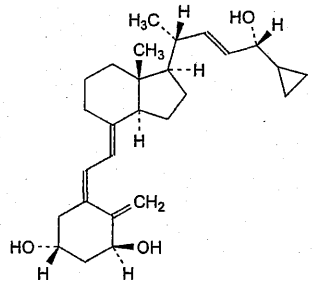
A. (5*Z*,7*E*,22*E*)-24-cyclopropyl-1α,3β-dihydroxy-9,10-secochola-5,7,10(19),22-tetraen-24-one,



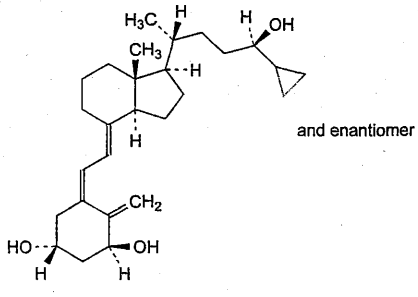
B. (5*Z*,7*Z*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol ((7*Z*)-calcipotriol),



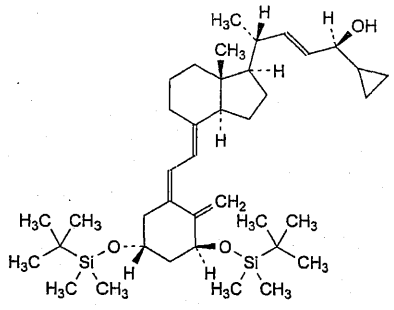
C. (5*E*,7*E*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol ((5*E*)-calcipotriol),



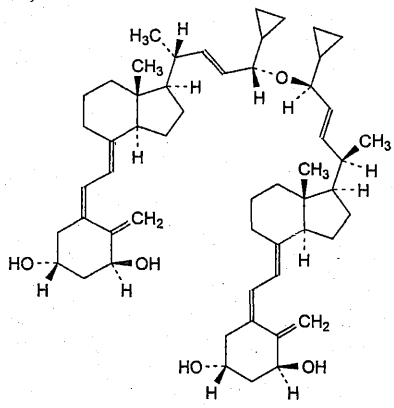
D. (5*Z*,7*E*,22*E*,24*R*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol (24-*epi*-calcipotriol),



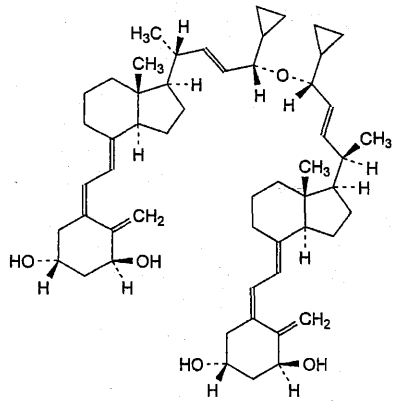
E. *rac*-(5*Z*,7*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19)-triene-1α,3β,24-triol,



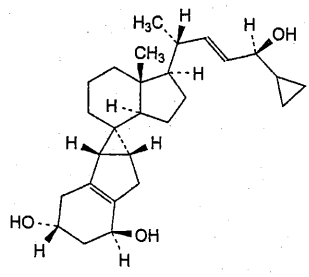
F. (5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-1α,3β-bis[[(1,1-dimethylethyl)dimethylsilyl]oxy]-9,10-secochola-5,7,10(19),22-tetraen-24-ol,



G. 24,24′-oxybis[(5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β-diol],



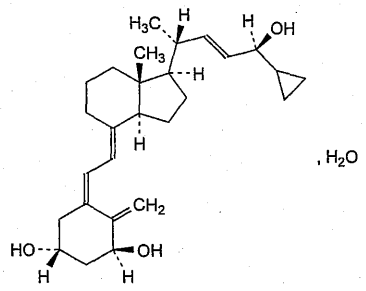
H. (5*Z*,7*E*,22*E*,24*R*)-24-cyclopropyl-24-[[(5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-1α,3β-dihydroxy-9,10-secochola-5,7,10(19),22-tetraen-24-yl]oxy]-9,10-secochola-5,7,10(19),22-tetraene-1α,3β-diol,



I. (6*S*,7*R*,8*R*,22*E*,24*S*)-24-cyclopropyl-6,8:7,19-dicyclo-9,10-secochola-5(10),22-diene-1α,3β,24-triol (suprasterol of calcipotriol).

Calcipotriol Monohydrate

(Ph. Eur. monograph 2284)



$C_{27}H_{40}O_3,H_2O$ 430.6 *147657-22-5*

Action and use
Vitamin D analogue.

Preparations
Calcipotriol Cream
Calcipotriol Ointment
Calcipotriol Scalp Application

Ph Eur

DEFINITION

(5*Z*,7*E*,22*E*,24*S*)-24-Cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol monohydrate.

Content
95.5 per cent to 102.0 per cent (anhydrous substance).

A reversible isomerisation to pre-calcipotriol takes place in solution, depending on temperature and time. The activity is due to both compounds.

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It is sensitive to light.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of calcipotriol monohydrate.*

B. Water (see Tests).

TESTS

Carry out the tests for related substances and the assay as rapidly as possible and protected from actinic light and air.

Related substances
A. Thin-layer chromatography (*2.2.27*).

Solution A To 1 mL of *triethylamine R* add 9 mL of *chloroform R*.

Test solution Dissolve 1 mg of the substance to be examined in 100 µL of solution A.

Reference solution (a) To 10 µL of the test solution add 990 µL of solution A.

Reference solution (b) To 250 µL of reference solution (a) add 750 µL of solution A.

Reference solution (c) To 100 µL of reference solution (a) add 900 µL of solution A.

Reference solution (d) Place 2 mg of the substance to be examined in a vial and dissolve in 200 µL of solution A. Close the vial and keep it in a water bath at 60 °C for 2 h.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *2-methylpropanol R, methylene chloride R* (20:80 *V/V*).

Application 10 µL of the test solution and reference solutions (b), (c) and (d).

Development Over 2/3 of the plate.

Drying In air, then at 140 °C for 10 min.

Detection Spray the hot plate with an *alcoholic solution of sulfuric acid R*, dry at 140 °C for not more than 1 min and examine in ultraviolet light at 366 nm.

Relative retention With reference to calcipotriol (R_F = about 0.4): impurity G = about 0.4; impurity H = about 0.4; pre-calcipotriol = about 0.9; impurity A = about 1.2.

System suitability Reference solution (d):
- — the chromatogram shows a secondary spot due to pre-calcipotriol.

Limits:
- — *impurity A*: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent);
- — *impurities G, H*: any spot due to impurity G or H is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent for the sum);
- — *unspecified impurities*: any other spot is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.1 per cent).

B. Liquid chromatography (*2.2.29*).

Solution A Dissolve 1.32 g of *ammonium phosphate R* in *water R* and dilute to 10.0 mL with the same solvent.

Solvent mixture Solution A, *water R, methanol R* (0.3:29.7:70 *V/V/V*).

Test solution (a) Dissolve 2 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Test solution (b) Dissolve 2.00 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 1 mg of *calcipotriol for system suitability CRS* (containing impurities B, C and D) in the solvent mixture and dilute to 2.5 mL with the solvent mixture.

Reference solution (d) Dissolve 2.00 mg of *calcipotriol monohydrate CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Column:
- — *size*: *l* = 0.10 m, Ø = 4.0 mm;
- — *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase *water for chromatography R, methanol R* (30:70 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 264 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Run time Twice the retention time of calcipotriol.

Identification of impurities Use the chromatogram supplied with *calcipotriol for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C and D.

Relative retention With reference to calcipotriol (retention time = about 13.5 min): impurity B = about 0.86; impurity C = about 0.92; impurity D = about 1.3.

System suitability Reference solution (c):

— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to calcipotriol.

Limits:

— *impurity B*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *impurities C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (2.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)

3.3 per cent to 5.0 per cent, determined on 0.100 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{27}H_{40}O_3$ taking into account the assigned content of *calcipotriol monohydrate CRS* and, if necessary, the peak due to pre-calcipotriol.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Test A for related substances

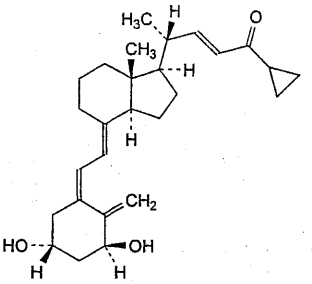
A, G, H, I.

Test B for related substances

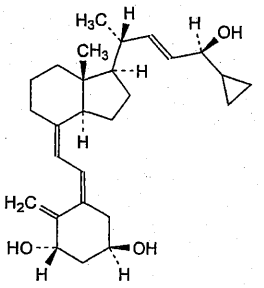
B, C, D, E, F.

Specified impurities *A, B, C, D, G, H.*

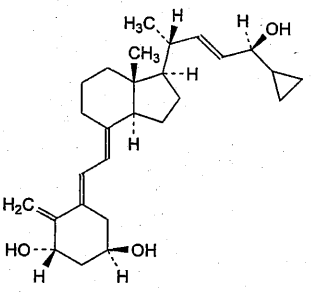
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F, I.*



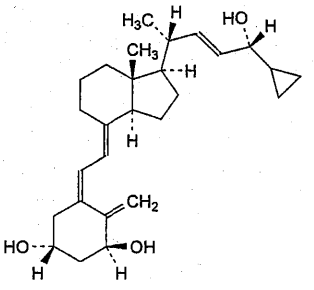
A. (5*Z*,7*E*,22*E*)-24-cyclopropyl-1α,3β-dihydroxy-9,10-secochola-5,7,10(19),22-tetraen-24-one,



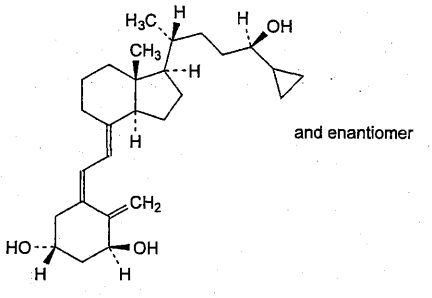
B. (5*Z*,7*Z*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol ((7*Z*)-calcipotriol),



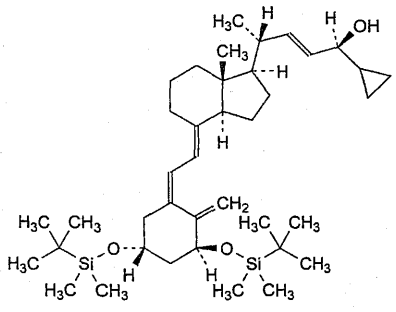
C. (5*E*,7*E*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol ((5*E*)-calcipotriol),



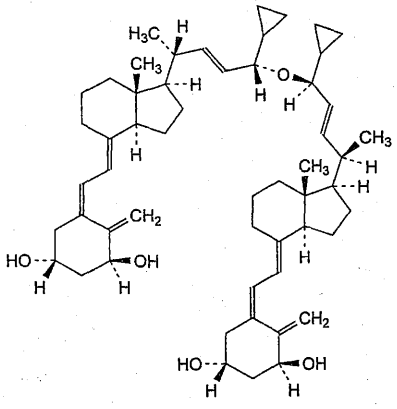
D. (5*Z*,7*E*,22*E*,24*R*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β,24-triol (24-*epi*-calcipotriol),



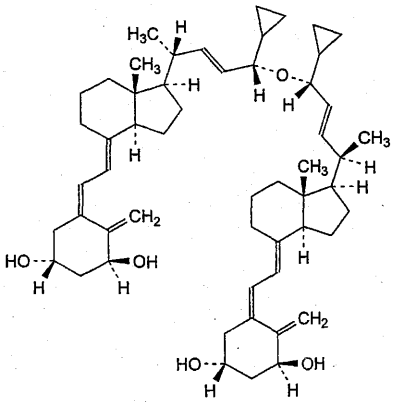
E. *rac*-(5*Z*,7*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19)-triene-1α,3β,24-triol,



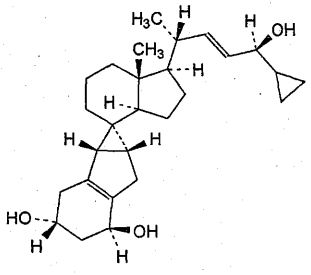
F. (5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-1α,3β-bis[[(1,1-dimethylethyl)dimethylsilyl]oxy]-9,10-secochola-5,7,10(19),22-tetraen-24-ol,



G. 24,24′-oxybis[(5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-9,10-secochola-5,7,10(19),22-tetraene-1α,3β-diol],



H. (5*Z*,7*E*,22*E*,24*R*)-24-cyclopropyl-24-[[(5*Z*,7*E*,22*E*,24*S*)-24-cyclopropyl-1α,3β-dihydroxy-9,10-secochola-5,7,10(19),22-tetraen-24-yl]oxy]-9,10-secochola-5,7,10(19),22-tetraene-1α,3β-diol,

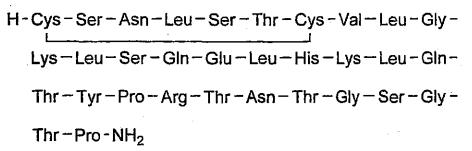


I. (6*S*,7*R*,8*R*,22*E*,24*S*)-24-cyclopropyl-6,8:7,19-dicyclo-9,10-secochola-5(10),22-diene-1α,3β,24-triol (suprasterol of calcipotriol).

Ph Eur

Calcitonin (Salmon)

(Ph. Eur. monograph 0471)



$C_{145}H_{240}N_{44}O_{48}S_2$ 3432

Action and use
Hormone.

Preparation
Calcitonin (Salmon) Injection

Ph Eur

DEFINITION

Polypeptide having the structure determined for salmon calcitonin I. It lowers the calcium concentration in plasma of mammals by diminishing the rate of bone resorption. It is obtained by chemical synthesis or by a method based on recombinant DNA (rDNA) technology. It is available as an acetate.

Content
90.0 per cent to 105.0 per cent of the peptide $C_{145}H_{240}N_{44}O_{48}S_2$ (anhydrous and acetic acid-free substance).

By convention, for the purpose of labelling calcitonin (salmon) preparations, 1 mg of calcitonin (salmon) ($C_{145}H_{240}N_{44}O_{48}S_2$) is equivalent to 6000 IU of biological activity.

PRODUCTION

The following requirements apply only to calcitonin (salmon) produced by a method based on rDNA technology.

Prior to release the following tests are carried out on each batch of final bulk product unless exemption has been granted by the competent authority.

Host-cell-derived proteins
The limit is approved by the competent authority.

Host-cell or vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

The following requirement applies only to calcitonin (salmon) obtained by chemical synthesis.

B. Amino acid analysis (*2.2.56*).

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids taking as equivalent to 1 the sum, divided by 20, of the number of moles of aspartic acid, glutamic acid, proline, glycine, valine, leucine, histidine, arginine and lysine. The values fall within

the following limits: aspartic acid: 1.8 to 2.2; glutamic acid: 2.7 to 3.3; proline: 1.7 to 2.3; glycine: 2.7 to 3.3; valine: 0.9 to 1.1; leucine: 4.5 to 5.3; histidine: 0.9 to 1.1; arginine: 0.9 to 1.1; lysine: 1.8 to 2.2; serine: 3.2 to 4.2; threonine: 4.2 to 5.2; tyrosine: 0.7 to 1.1; half-cystine: 1.4 to 2.1.

The following requirement applies only to calcitonin (salmon) produced by a method based on rDNA technology.

C. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a 1 mg/mL solution of the substance to be examined. Transfer 1.0 mL to a clean tube. Add 100 µL of *1 M tris-hydrochloride buffer solution pH 8.0 R* and 20 µL of a freshly prepared 1.0 mg/mL solution of *trypsin for peptide mapping R*. Allow to stand at 2-8 °C for 16-20 h. Stop the reaction by adding 10 µL of a 50 per cent *V/V* solution of *trifluoroacetic acid R*. Cap the vial and mix. Centrifuge the vials to remove air bubbles.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *calcitonin (salmon) CRS* instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION Liquid chromatography (*2.2.29*).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- *mobile phase A*: mix 1 mL of *trifluoroacetic acid R* and 1000 mL of *water R*; filter and degas;
- *mobile phase B*: mix 0.850 mL of *trifluoroacetic acid R*, 200 mL of *water R* and 800 mL of *acetonitrile for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 50	100 → 65	0 → 35
50 - 60	65 → 40	35 → 60
60 - 60.1	40 → 0	60 → 100
60.1 - 65.1	0	100
65.1 - 65.2	0 → 100	100 → 0
65.2 - 80.2	100	0

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 20 µL.

System suitability The chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of calcitonin (salmon) digest supplied with *calcitonin (salmon) CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution: the retention times of the fragment peaks in the chromatogram obtained with the test solution are within 5 per cent of the retention times of the fragments obtained with the reference solution; the peak area ratios of the fragment peaks in the chromatogram obtained with the test solution, normalised to the area of peak T_2, are within 5 per cent of the corresponding peak ratios in the chromatogram obtained with the reference solution.

TESTS

Acetic acid (*2.5.34*)

4.0 per cent to 15.0 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure.

The following requirement applies to calcitonin (salmon), whether obtained by chemical synthesis or by a method based on rDNA technology.

A. *Test solution.* Prepare a 1.0 mg/mL solution of the substance to be examined in mobile phase A.

Reference solution Dissolve the contents of a vial of *calcitonin (salmon) CRS* in mobile phase A to obtain a concentration of 1.0 mg/mL.

Resolution solution Dissolve the contents of a vial of *N-acetyl-Cys[1]-calcitonin CRS* in 400 µL of mobile phase A and add 100 µL of the test solution.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 65 °C.

Mobile phase:
- *mobile phase A*: dissolve 3.26 g of *tetramethylammonium hydroxide R* in 900 mL of *water R*, adjust to pH 2.5 with *phosphoric acid R* and mix with 100 mL of *acetonitrile for chromatography R*; filter and degas;
- *mobile phase B*: dissolve 1.45 g of *tetramethylammonium hydroxide R* in 400 mL of *water R*, adjust to pH 2.5 with *phosphoric acid R* and mix with 600 mL of *acetonitrile for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	72 → 48	28 → 52
30 - 32	48 → 72	52 → 28
32 - 55	72	28

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Relative retention With reference to calcitonin (salmon) (retention time = about 20 min): impurity B = about 0.8; impurity C = about 0.9; impurity D = about 1.05; impurity A = about 1.15.

System suitability Resolution solution:
- *resolution*: minimum 5.0 between the peaks due to calcitonin (salmon) and impurity A,
- *symmetry factor*: maximum 2.5 for the peak due to impurity A.

Limits:
- *impurities A, B, C, D*: for each impurity, maximum 3.0 per cent; other unidentified, specified impurities may occur that co-elute with impurities A, B, C and D; the acceptance criterion applies irrespective of whether these impurities co-elute;
- *total*: maximum 5.0 per cent;
- *disregard limit*: 0.1 per cent.

The following requirement applies only to calcitonin (salmon) produced by a method based on rDNA technology.

B. *Test solution.* Prepare a 0.5 mg/mL solution of the substance to be examined. To 1.0 mL of this solution add 100 µL of *0.25 M citrate buffer solution pH 3.0 R.*

Resolution solution Prepare a 1 mg/mL solution of the substance to be examined. Mix 1 volume of the solution and 1 volume of *calcitonin-Gly CRS*. To 1.0 mL of this mixture add 100 µL of *0.25 M citrate buffer solution pH 3.0 R.*

Column:
- — *size*: *l* = 0.20 m, Ø = 4.6 mm;
- — *stationary phase*: a suitable polysulfoethylaspartamide ion-exchange gel (5 µm).

Mobile phase:
- — *mobile phase A*: mix 15 volumes of *acetonitrile for chromatography R* and 85 volumes of a 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 5.0 with a 600 g/L solution of *potassium hydroxide R*;
- — *mobile phase B*: mix 15 volumes of *acetonitrile for chromatography R* and 85 volumes of a solution containing 2.72 g/L of *potassium dihydrogen phosphate R* and 29.22 g/L of *sodium chloride R* adjusted to pH 4.6 with a 600 g/L solution of *potassium hydroxide R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100 → 0	0 → 100
10 - 15	0	100
15 - 15.1	0 → 100	100 → 0
15.1 - 22.1	100	0

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 µL; rinse the injector with a 40 per cent *V/V* solution of *acetonitrile for chromatography R.*

Relative retention With reference to calcitonin (salmon) (retention time = about 9 min): impurity G = about 0.4; impurity F = about 0.6; impurity E = about 0.9.

System suitability Resolution solution:
- — *resolution*: minimum 3.0 between the peaks due to impurity E and calcitonin (salmon).

Limits:
- — *impurity E*: maximum 0.6 per cent;
- — *impurities F, G*: for each impurity, maximum 0.2 per cent.

Water (*2.5.32*)
Maximum 10.0 per cent.

Acetic acid and water
Maximum 20 per cent, calculated by adding together the percentage contents of acetic acid and water determined by the methods described above.

Bacterial endotoxins (*2.6.14*)
Less than 25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

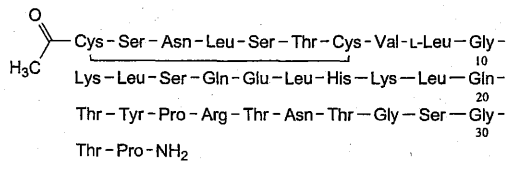
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances. Use method A for calcitonin (salmon) obtained by chemical synthesis and method B for calcitonin (salmon) obtained by a method based on rDNA technology.

Calculate the content of calcitonin (salmon) ($C_{145}H_{240}N_{44}O_{48}S_2$) from the area of the principal peak in each of the chromatograms obtained with the test solution and the reference solution and the declared content of $C_{145}H_{240}N_{44}O_{48}S_2$ in *calcitonin (salmon) CRS*. Proceed with tangential integration of the peak areas.

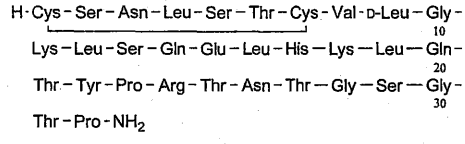
STORAGE
Protected from light at a temperature between 2 °C and 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING
The label states:
— the calcitonin peptide content ($C_{145}H_{240}N_{44}O_{48}S_2$);
— the origin: synthetic or rDNA technology.

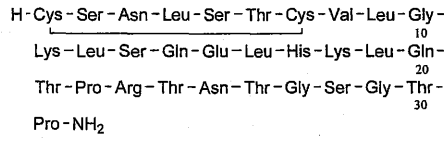
IMPURITIES
Specified impurities *A, B, C, D, E, F, G.*



A. acetylcalcitonin (salmon),

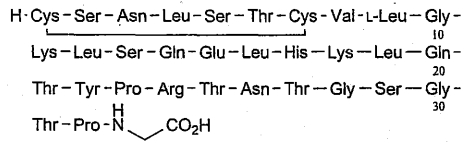


B. [9-D-leucine]calcitonin (salmon),

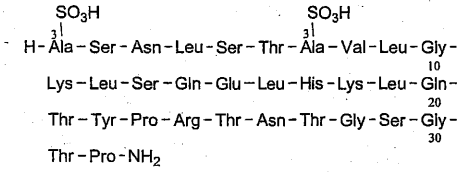


C. des-22-tyrosine-calcitonin (salmon),

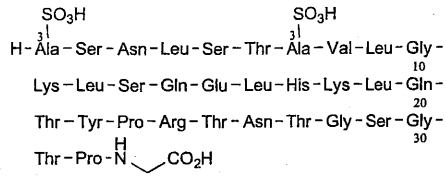
D. *O*-acetylated calcitonin (salmon),



E. salmon calcitoninylglycine,



F. [1,7-bis(3-sulfo-L-alanine)]calcitonin (salmon),

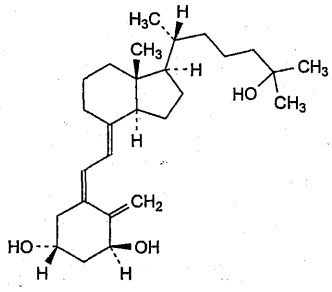


G. [1,7-bis(3-sulfo-L-alanine)]calcitoninylglycine (salmon).

Ph Eur

Calcitriol

(Ph. Eur. monograph 0883)



$C_{27}H_{44}O_3$ 416.6 *32222-06-3*

Action and use
Vitamin D analogue.

Preparation
Calcitriol Capsules

Ph Eur

DEFINITION
(5*Z*,7*E*)-9,10-Secocholesta-5,7,10(19)-triene-1α,3β,25-triol.

Content
97.0 per cent to 103.0 per cent.

A reversible isomerisation to pre-calcitriol takes place in solution, depending on temperature and time. The activity is due to both compounds (see Assay).

CHARACTERS
Appearance
White or almost white crystals.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in fatty oils.

It is sensitive to air, heat and light.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of calcitriol.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Carry out the test as rapidly as possible, avoiding exposure to actinic light and air.*

Test solution Dissolve 1.00 mg of the substance to be examined without heating in 10.0 mL of the mobile phase.

Reference solution (a) Dissolve 1.00 mg of *calcitriol CRS* without heating in 10.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Heat 2 mL of reference solution (a) at 80 °C for 30 min.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R1* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 450 volumes of a 1.0 g/L solution of *tris(hydroxymethyl)aminomethane R* adjusted to pH 7.0-7.5 with *phosphoric acid R*, and 550 volumes of *acetonitrile R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 50 µL.

Run time Twice the retention time of calcitriol.

Relative retention With reference to calcitriol (retention time = about 14 min): impurity C = about 0.4; pre-calcitriol = about 0.88; impurity A = about 0.95; impurity B = about 1.1.

System suitability:
— *resolution*: minimum 3.5 between the peaks due to pre-calcitriol and calcitriol in the chromatogram obtained with reference solution (c);
— *number of theoretical plates*: minimum 10 000, calculated for the peak due to calcitriol in the chromatogram obtained with reference solution (a).

Limits:
— *impurities A, B, C*: for each impurity, maximum 0.5 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to pre-calcitriol.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

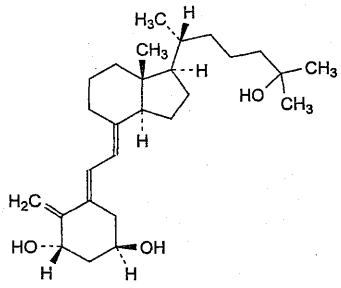
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1 per cent for the peak due to calcitriol after 6 injections.

Calculate the percentage content of $C_{27}H_{44}O_3$ taking into account the assigned content of *calcitriol CRS* and, if necessary, the peak due to pre-calcitriol.

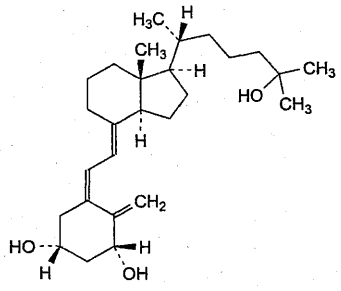
STORAGE
Under nitrogen, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

The contents of an opened container are to be used immediately.

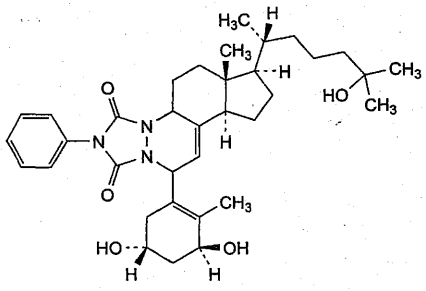
IMPURITIES
Specified impurities *A, B, C.*



A. (5*E*,7*E*)-9,10-secocholesta-5,7,10(19)-triene-1α,3β,25-triol (*trans*-calcitriol),



B. (5*Z*,7*E*)-9,10-secocholesta-5,7,10(19)-triene-1β,3β,25-triol (1β-calcitriol),

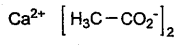


C. (6a*R*,7*R*,9a*R*)-11-[(3*S*,5*R*)-3,5-dihydroxy-2-methylcyclohex-1-enyl]-7-[(1*R*)-5-hydroxy-1,5-dimethylhexyl]-6a-methyl-2-phenyl-5,6,6a,7,8,9,9a,11-octahydro-1*H*,4a*H*-cyclopenta[*f*][1,2,4]triazolo[1,2-*a*]cinnoline-1,3(2*H*)-dione (triazoline adduct of pre-calcitriol).

Ph Eur

Calcium Acetate

(Ph. Eur. monograph 2128)



$C_4H_6CaO_4$ 158.2 *62-54-4*

Action and use
Used in solutions for haemodialysis and peritoneal dialysis.

Ph Eur

DEFINITION
Calcium diacetate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. It gives reaction (b) of calcium (*2.3.1*).
B. It gives reaction (b) of acetates (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
7.2 to 8.2.

Dilute 5.0 mL of solution S to 10.0 mL with *carbon dioxide-free water R*.

Readily oxidisable substances
Dissolve 2.0 g in boiling *water R* and dilute to 100 mL with boiling *water R*, add a few glass beads, 6 mL of *5 M sulfuric acid R* and 0.3 mL of *0.02 M potassium permanganate*, mix, boil gently for 5 min and allow the precipitate to settle. The pink colour in the supernatant is not completely discharged.

Chlorides (*2.4.4*)
Maximum 330 ppm.

Dissolve 0.15 g in *water R* and dilute to 15 mL with the same solvent.

Fluorides
Maximum 50 ppm.

Potentiometry (*2.2.36, Method I*).

Test solution In a 50 mL volumetric flask, dissolve 0.200 g in a 10.3 g/L solution of *hydrochloric acid R*, add 5.0 mL of *fluoride standard solution (1 ppm F) R* and dilute to 50.0 mL with a 10.3 g/L solution of *hydrochloric acid R*. To 20.0 mL of the solution add 20.0 mL of *total-ionic-strength-adjustment buffer R* and 3 mL of an 82 g/L solution of *anhydrous sodium acetate R*. Adjust to pH 5.2 with *ammonia R* and dilute to 50.0 mL with *distilled water R*.

Reference solutions To 0.25 mL, 0.5 mL, 0.75 mL and 1.0 mL of *fluoride standard solution (10 ppm F) R* add 20.0 mL of *total-ionic-strength-adjustment buffer R* and dilute to 50.0 mL with *distilled water R*.

Indicator electrode Fluoride selective.

Reference electrode Silver-silver chloride.

Take into account the addition of fluoride to the test solution for the calculation.

Nitrates
To 10.0 mL of solution S add 5 mg of *sodium chloride R*, 0.05 mL of *indigo carmine solution R* and add with stirring, 10 mL of *nitrogen-free sulfuric acid R*. The blue colour remains for at least 10 min.

Sulfates (*2.4.13*)
Maximum 600 ppm.

Dissolve 0.25 g in *distilled water R* and dilute to 15 mL with the same solvent.

Aluminium (*2.4.17*)
Maximum 1 ppm, if intended for use in the manufacture of peritoneal dialysis solutions, haemofiltration solutions or haemodialysis solutions.

Test solution Dissolve 4.0 g of the substance to be examined in 100 mL of *water R* and add 10 mL of *acetate buffer solution pH 6.0 R*.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al) R*, 10 mL of *acetate buffer solution pH 6.0 R* and 98 mL of *water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *water R*.

Arsenic (*2.4.2*)
Maximum 3 ppm.

3.3 mL of solution S complies with test A.

Barium
Maximum 50 ppm.

Inductively coupled plasma-atomic emission spectrometry (*2.2.57*).

Test solution Dissolve 5.00 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using barium standard solution (0.1 per cent Ba) R, diluted as necessary with *water R*.

Wavelength 455.4 nm.

Iron (*2.4.9*)
Maximum 20 ppm, if intended for use in the manufacture of parenteral preparations or haemodialysis solutions.

Dilute 5 mL of solution S to 10 mL of *water R*.

Magnesium
Maximum 500 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using magnesium standard solution (0.1 per cent Mg) R, diluted as necessary with *water R*.

Source Magnesium hollow-cathode lamp.

Wavelength 285.2 nm.

Atomisation device Air-acetylene flame.

Potassium
Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations or haemodialysis solutions.

Atomic emission spectrometry (*2.2.22, Method II*).

Test solution Dissolve 1.00 g of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using *potassium standard solution (0.2 per cent K) R*, diluted as necessary with *water R*.

Wavelength 766.5 nm.

Sodium
Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations or haemodialysis solutions.

Atomic emission spectrometry (*2.2.22, Method II*).

Test solution Dissolve 1.00 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using *sodium standard solution (200 ppm Na) R*, diluted as necessary with *water R*.

Wavelength 589 nm.

Strontium
Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations or haemodialysis solutions.

Atomic emission spectrometry (*2.2.22, Method II*).

Test solution Dissolve 2.00 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using *strontium standard solution (1.0 per cent Sr) R*, diluted as necessary with *water R*.

Wavelength 460.7 nm.

Water (*2.5.12*)
Maximum 7.0 per cent, determined on 0.100 g. Add 2 mL of *anhydrous acetic acid R* to the titration vessel in addition to the methanol. Clean the titration vessel after each determination.

ASSAY
Dissolve 0.150 g in 100 mL of *water R* and carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 15.82 mg of $C_4H_6CaO_4$.

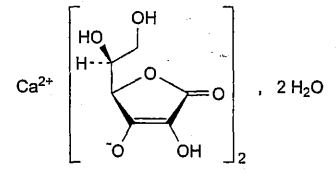
STORAGE
In an airtight container.

LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations, peritoneal dialysis solutions, haemofiltration solutions or haemodialysis solutions.

Ph Eur

Calcium Ascorbate

(Ph. Eur. monograph 1182)



$C_{12}H_{14}CaO_{12},2H_2O$ 426.3 *5743-28-2*

Action and use
Excipient.

Ph Eur

DEFINITION
Calcium di[(*R*)-2-[(*S*)-1,2-dihydroxyethyl]-4-hydroxy-5-oxo-2*H*-furan-3-olate] dihydrate.

Content
99.0 per cent to 100.5 per cent of $C_{12}H_{14}CaO_{12},2H_2O$.

CHARACTERS
Appearance
White or slightly yellowish, crystalline powder.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B, E.
Second identification: A, C, D, E.
A. Specific optical rotation (see Tests).
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of calcium ascorbate.*
C. Dilute 1 mL of solution S (see Tests) to 10 mL with *water R*. To 2 mL of the solution add 0.2 mL of a 100 g/L solution of *ferrous sulfate R*. A deep violet colour develops.
D. To 1 mL of solution S add 0.2 mL of *dilute nitric acid R* and 0.2 mL of *silver nitrate solution R2*. A grey precipitate is formed.
E. The substance gives reaction (b) of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 5.00 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*). Examine the colour of the solution immediately after preparation of the solution.

pH (*2.2.3*)
6.8 to 7.4 for solution S.

Specific optical rotation (*2.2.7*)
+ 95 to + 97 (dried substance), determined using freshly prepared solution S.

Related substances
The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Fluorides
Maximum 10 ppm.
Potentiometry (*2.2.36, Method I*).
Test solution In a 50 mL volumetric flask, dissolve 1.000 g in a 10.3 g/L solution of *hydrochloric acid R*, add 5.0 mL of *fluoride standard solution (1 ppm F) R* and dilute to 50.0 mL with a 10.3 g/L solution of *hydrochloric acid R*. To 20.0 mL of the solution add 20.0 mL of *total-ionic-strength-adjustment buffer R* and 3 mL of an 82 g/L solution of *anhydrous sodium acetate R*. Adjust to pH 5.2 with *ammonia R* and dilute to 50.0 mL with *distilled water R*.
Reference solutions To 0.25 mL, 0.5 mL, 1.0 mL, 2.0 mL and 5.0 mL of *fluoride standard solution (10 ppm F) R* add 20.0 mL of *total-ionic-strength-adjustment buffer R* and dilute to 50.0 mL with *distilled water R*.
Indicator electrode Fluoride selective.
Reference electrode Silver-silver chloride.
Take into account the addition of fluoride to the test solution for the calculation.

Copper
Maximum 5 ppm.
Atomic absorption spectrometry (*2.2.23, Method I*).
Test solution Dissolve 2.0 g in a 9.7 g/L solution of *nitric acid R* and dilute to 25.0 mL with the same acid solution.
Reference solutions Prepare the reference solutions using *copper standard solution (10 ppm Cu) R*, diluting with a 9.7 g/L solution of *nitric acid R*.
Source Copper hollow-cathode lamp.
Wavelength 324.8 nm.
Atomisation device Air-acetylene flame.

Iron
Maximum 2 ppm.
Atomic absorption spectrometry (*2.2.23, Method I*).
Test solution Dissolve 5.0 g in a 9.7 g/L solution of *nitric acid R* and dilute to 25.0 mL with the same acid solution.
Reference solutions Prepare the reference solutions using *iron standard solution (10 ppm Fe) R*, diluting with a 9.7 g/L solution of *nitric acid R*.
Source Iron hollow-cathode lamp.
Wavelength 248.3 nm.
Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)
Maximum 0.1 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

ASSAY
Dissolve 80.0 mg in a mixture of 10 mL of *dilute sulfuric acid R* and 80 mL of *carbon dioxide-free water R*. Add 1 mL of *starch solution R*. Titrate with *0.05 M iodine* until a persistent violet-blue colour is obtained.
1 mL of *0.05 M iodine* is equivalent to 10.66 mg of $C_{12}H_{14}CaO_{12},2H_2O$.

STORAGE
In a non-metallic container, protected from light.

Ph Eur

Calcium Carbonate

(Ph. Eur. monograph 0014)

$CaCO_3$ 100.1 *471-34-1*

Action and use
Antacid.

Preparations
Calcium Carbonate Chewable Tablets
Calcium Carbonate Oral Suspension
Calcium and Colecalciferol Tablets
Calcium and Colecalciferol Chewable Tablets

Ph Eur

DEFINITION
Content
98.5 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water.

IDENTIFICATION
A. It gives the reaction of carbonates (*2.3.1*).
B. 0.2 mL of solution S (see Tests) gives the reactions of calcium (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in 80 mL of *dilute acetic acid R*. When the effervescence ceases, boil for 2 min. Allow to cool, dilute to 100 mL with *dilute acetic acid R* and filter, if necessary, through a sintered-glass filter (*2.1.2*).

Substances insoluble in acetic acid
Maximum 0.2 per cent.

Wash any residue obtained during the preparation of solution S with 4 quantities, each of 5 mL, of hot *water R* and dry at 100-105 °C for 1 h. The residue weighs a maximum of 10 mg.

Chlorides (*2.4.4*)
Maximum 330 ppm.

Dilute 3 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 0.25 per cent.

Dilute 1.2 mL of solution S to 15 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 4 ppm, determined on 5 mL of solution S.

Barium
To 10 mL of solution S add 10 mL of *calcium sulfate solution R*. After at least 15 min, any opalescence in the solution is not more intense than that in a mixture of 10 mL of solution S and 10 mL of *distilled water R*.

Iron (*2.4.9*)
Maximum 200 ppm.

Dissolve 50 mg in 5 mL of *dilute hydrochloric acid R* and dilute to 10 mL with *water R*.

Magnesium and alkali metals
Maximum 1.5 per cent.

Dissolve 1.0 g in 12 mL of *dilute hydrochloric acid R*. Boil the solution for about 2 min and add 20 mL of *water R*, 1 g of *ammonium chloride R* and 0.1 mL of *methyl red solution R*. Add *dilute ammonia R1* until the colour of the indicator changes and then add 2 mL in excess. Heat to boiling and add 50 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 100 mL with *water R* and filter through a suitable filter. To 50 mL of the filtrate add 0.25 mL of *sulfuric acid R*. Evaporate to dryness on a water-bath and ignite to constant mass at 600 ± 50 °C.
The residue weighs a maximum of 7.5 mg.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 200 ± 10 °C.

ASSAY

Dissolve 0.150 g in a mixture of 3 mL of *dilute hydrochloric acid R* and 20 mL of *water R*. Boil for 2 min, allow to cool and dilute to 50 mL with *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 10.01 mg of $CaCO_3$.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium carbonate used as filler in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Calcium Chloride Dihydrate

(Ph. Eur. monograph 0015)

$CaCl_2,2H_2O$ 147.0 *10035-04-8*

Preparations
Calcium Chloride Injection
Compound Sodium Lactate Infusion

Ph Eur

DEFINITION

Content
97.0 per cent to 103.0 per cent of $CaCl_2,2H_2O$.

CHARACTERS

Appearance
White or almost white, crystalline powder, hygroscopic.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

B. It gives the reactions of calcium (*2.3.1*).

C. It complies with the limits of the assay.

TESTS

Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of freshly prepared solution S add 0.1 mL of *phenolphthalein solution R*. If the solution is red, not more than 0.2 mL of *0.01 M hydrochloric acid* is required to discharge the colour and if the solution is colourless, not more than 0.2 mL of *0.01 M sodium hydroxide* is required to turn it red.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dilute 5 mL of solution S to 15 mL with *distilled water R*.

Aluminium
To 10 mL of solution S add 2 mL of *ammonium chloride solution R* and 1 mL of *dilute ammonia R1* and boil the solution. No turbidity or precipitate is formed.

If intended for use in the manufacture of dialysis solutions, the above test is replaced by the following test for aluminium (*2.4.17*): maximum 1 ppm.

Prescribed solution Dissolve 4 g in 100 mL of *water R* and add 10 mL of *acetate buffer solution pH 6.0 R*.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al) R*, 10 mL of *acetate buffer solution pH 6.0 R* and 98 mL of *water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *water R*.

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R*. After at least 15 min, any opalescence in the solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S.

Magnesium and alkali metals
Maximum 0.5 per cent.

To a mixture of 20 mL of solution S and 80 mL of *water R* add 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1*, heat to boiling and pour into the boiling solution a hot solution of 5 g of *ammonium oxalate R* in 75 mL of *water R*. Allow to stand for 4 h, dilute to 200 mL with *water R* and filter through a suitable filter. To 100 mL of the filtrate add 0.5 mL of *sulfuric acid R*. Evaporate to dryness on a water-bath and ignite to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

ASSAY
Dissolve 0.280 g in 100 mL of *water R* and carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 14.70 mg of $CaCl_2,2H_2O$.

LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of dialysis solutions.

STORAGE
In an airtight container.

Ph Eur

Calcium Chloride Hexahydrate

(Ph. Eur. monograph 0707)

$CaCl_2,6H_2O$ 219.1 *7774-34-7*

Ph Eur

DEFINITION
Content
97.0 per cent to 103.0 per cent of $CaCl_2,6H_2O$.

CHARACTERS
Appearance
White or almost white, crystalline mass or colourless crystals.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

It solidifies at about 29 °C.

IDENTIFICATION
A. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

B. It gives the reactions of calcium (*2.3.1*).

C. It complies with the limits of the assay.

TESTS
Solution S
Dissolve 15.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of freshly prepared solution S add 0.1 mL of *phenolphthalein solution R*. If the solution is red, not more than 0.2 mL of *0.01 M hydrochloric acid* is required to discharge the colour and if the solution is colourless, not more than 0.2 mL of *0.01 M sodium hydroxide* is required to turn it red.

Sulfates (*2.4.13*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *distilled water R*.

Aluminium
To 10 mL of solution S add 2 mL of *ammonium chloride solution R* and 1 mL of *dilute ammonia R1*. Heat to boiling. No turbidity or precipitate is formed.

If intended for use in the manufacture of dialysis solutions, the above test is replaced by the following test for aluminium (*2.4.17*): maximum 1 ppm.

Prescribed solution Dissolve 6 g in 100 mL of *water R* and add 10 mL of *acetate buffer solution pH 6.0 R*.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al) R*, 10 mL of *acetate buffer solution pH 6.0 R* and 98 mL of *water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *water R*.

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R*. After at least 15 min, any opalescence in the solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 7 ppm, determined on solution S.

Magnesium and alkali metals
Maximum 0.3 per cent.

To a mixture of 20 mL of solution S and 80 mL of *water R* add 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1*, heat to boiling and pour into the boiling solution a hot solution of 5 g of *ammonium oxalate R* in 75 mL of *water R*. Allow to stand for 4 h, dilute to 200 mL with *water R* and filter through a suitable filter. To 100 mL of the filtrate add 0.5 mL of *sulfuric acid R*. Evaporate to dryness on a water-bath and ignite to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

ASSAY
Dissolve 0.200 g in 100 mL of *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

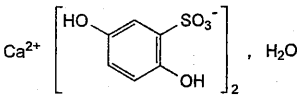
1 mL of *0.1 M sodium edetate* is equivalent to 21.91 mg of $CaCl_2,6H_2O$.

LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of dialysis solutions.

Ph Eur

Calcium Dobesilate Monohydrate

(Ph. Eur. monograph 1183)



$C_{12}H_{10}CaO_{10}S_2,H_2O$ 436.4 *20123-80-2*

Ph Eur

DEFINITION

Calcium di(2,5-dihydroxybenzenesulfonate) monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Very soluble in water, freely soluble in anhydrous ethanol, very slightly soluble in 2-propanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.100 g in *water R* and dilute to 200.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *water R*.

Spectral range 210-350 nm.

Absorption maxima At 221 nm and 301 nm.

Specific absorbance at the absorption maximum at 301 nm 174 to 181.

B. Mix 1 mL of *ferric chloride solution R2*, 1 mL of a freshly prepared 10 g/L solution of *potassium ferricyanide R* and 0.1 mL of *nitric acid R*. To this mixture add 5 mL of freshly prepared solution S (see Tests): a blue colour and a precipitate are immediately produced.

C. 2 mL of freshly prepared solution S gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S

Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S, when freshly prepared, is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

4.5 to 6.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*). *Keep all solutions at 2-8 °C.*

Buffer solution Dissolve 1.2 g of *anhydrous sodium dihydrogen phosphate R* in 900 mL of *water for chromatography R*, adjust to pH 6.5 with *disodium hydrogen phosphate solution R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of *hydroquinone R* (impurity A) in *water R* and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 100 mL with *water R*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R1*, buffer solution (10:90 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 2.5 times the retention time of dobesilate.

Relative retention With reference to dobesilate (retention time = about 6 min): impurity A = about 1.7.

System suitability Reference solution (b):

— *resolution*: minimum 8.0 between the peaks due to dobesilate and impurity A.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.6;

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Iron (*2.4.9*)

Maximum 10 ppm, determined on 10 mL of solution S.

Water (*2.5.12*)

4.0 per cent to 6.0 per cent, determined on 0.500 g.

ASSAY

Dissolve 0.200 g in a mixture of 10 mL of *water R* and 40 mL of *dilute sulfuric acid R*. Titrate with *0.1 M cerium sulfate*, determining the end-point potentiometrically (*2.2.20*).

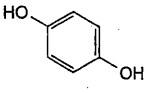
1 mL of *0.1 M cerium sulfate* is equivalent to 10.45 mg of $C_{12}H_{10}CaO_{10}S_2$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities *A*.



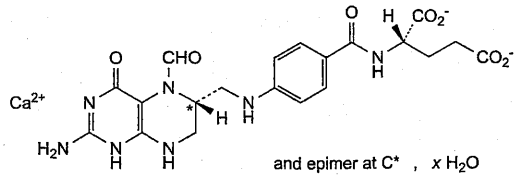
A. benzene-1,4-diol (hydroquinone).

Ph Eur

Calcium Folinate Hydrate

Calcium Folinate

(Ph. Eur. monograph 0978)



$C_{20}H_{21}CaN_7O_7,xH_2O$ 511.5 (anhydrous substance)

2060570-47-8

Action and use
Antidote to folic acid antagonists.

Preparations
Calcium Folinate Injection
Calcium Folinate Tablets

Ph Eur

DEFINITION

Calcium (2*S*)-2-[4-[[[(6*RS*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl]amino]benzamido]pentanedioate hydrate.

Content
— *calcium folinate* ($C_{20}H_{21}CaN_7O_7$): 97.0 per cent to 102.0 per cent (anhydrous substance);
— *calcium* (Ca; A_r 40.08): 7.54 per cent to 8.14 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS

Appearance
White or light yellow, amorphous or crystalline, hygroscopic powder.

Solubility
Sparingly soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

The amorphous form may produce supersaturated solutions in water.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, B, D.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *calcium folinate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *water R* and add dropwise sufficient *acetone R* to produce a precipitate. Allow to stand for 15 min, collect the precipitate by centrifugation, wash the precipitate with 2 small quantities of *acetone R* and dry. Record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 15 mg of the substance to be examined in a 3 per cent *V/V* solution of *ammonia R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 15 mg of *calcium folinate CRS* in a 3 per cent *V/V* solution of *ammonia R* and dilute to 5 mL with the same solvent.

Plate *cellulose for chromatography F_{254} R* as the coating substance.

Mobile phase The lower layer of a mixture of 1 volume of *isoamyl alcohol R* and 10 volumes of a 50 g/L solution of *citric acid monohydrate R* previously adjusted to pH 8 with *ammonia R*.

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (b) of calcium (*2.3.1*).

TESTS

Carry out the tests as rapidly as possible, protected from actinic light.

Solution S
Dissolve 1.25 g in *carbon dioxide-free water R*, heating at 40 °C if necessary, and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

pH (*2.2.3*)
6.8 to 8.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 14.4 to + 18.0 (anhydrous substance), determined on solution S.

Absorbance (*2.2.25*)
Maximum 0.60, determined at 420 nm on solution S.

Ethanol
Head-space gas chromatography (*2.2.28*): use the standard additions method.

Test solution Dissolve 0.25 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution Dilute 0.750 g of *anhydrous ethanol R* to 1000.0 mL with *water R*.

Column:
— *material*: fused silica;
— *size*: *l* = 10 m, Ø = 0.32 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R*.

Carrier gas *nitrogen for chromatography R*.

Flow rate 4 mL/min.

Static head-space conditions that may be used:
— *equilibration temperature*: 80 °C;
— *equilibration time*: 20 min;
— *pressurisation time*: 30 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 6	125 → 185
	6 - 15	185
Injection port		250
Detector		250

Detection Flame ionisation.

Injection At least 3 times.

Limit:
— *ethanol*: maximum 3.0 per cent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *calcium folinate CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (c) Dissolve 5 mg of *calcium folinate for system suitability CRS* (containing impurities A, E and F) in 5 mL of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 165 mL of *methanol R* and 835 mL of a solution containing 4.0 mL of *tetrabutylammonium dihydrogen phosphate solution R* and 1.42 g of *disodium hydrogen phosphate dihydrate R* in *water for chromatography R*, previously adjusted to pH 7.7 with *phosphoric acid R* or *dilute sodium hydroxide solution R*.

Flow rate 1.25 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 3.5 times the retention time of folinic acid.

Identification of impurities Use the chromatogram supplied with *calcium folinate for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, E and F.

Relative retention With reference to folinic acid (retention time = about 13.9 min): impurity E = about 0.4; impurity A = about 0.6; impurity F = about 0.7.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to impurities A and F.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity E = 0.6; impurity F = 0.6;
— for each impurity, use the concentration of calcium folinate hydrate in reference solution (b).

Limits:
— *impurities A, E, F*: for each impurity, maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.20 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides

Maximum 0.5 per cent.

Dissolve 0.300 g in 50 mL of *water R* heating at 40 °C if necessary. Add 10 mL of *2 M nitric acid* and titrate with *0.005 M silver nitrate* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.005 M silver nitrate* is equivalent to 0.177 mg of Cl.

Water

(*2.5.12*): 10.0 per cent to 17.0 per cent.

Dissolve 0.100 g in a mixture of 15 mL of *formamide R* and 50 mL of the titration solvent. Stir for about 6 min before titrating and use a suitable titrant that does not contain pyridine.

ASSAY

Carry out the assays as rapidly as possible, protected from actinic light.

Calcium

Dissolve 0.400 g in 150 mL of *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 4.008 mg of Ca.

Calcium folinate

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{20}H_{21}CaN_7O_7$ taking into account the assigned content of *calcium folinate CRS*.

STORAGE

In an airtight container, protected from light. If the substance is sterile, the container is also sterile and tamper-proof.

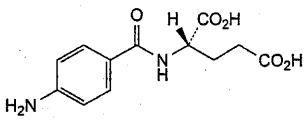
LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

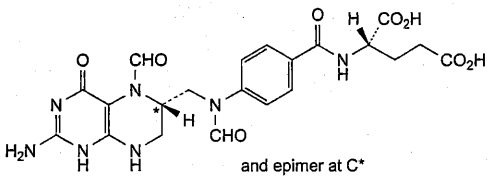
IMPURITIES

Specified impurities *A, E, F.*

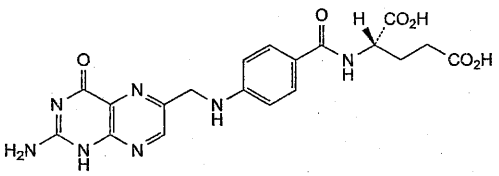
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, G, I.*



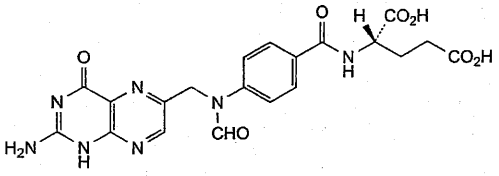
A. (2*S*)-2-(4-aminobenzamido)pentanedioic acid,



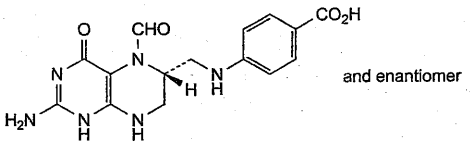
B. (2*S*)-2-[4-[[[(6*RS*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl](formyl)amino]benzamido] pentanedioic acid (5,10-diformyltetrahydrofolic acid),



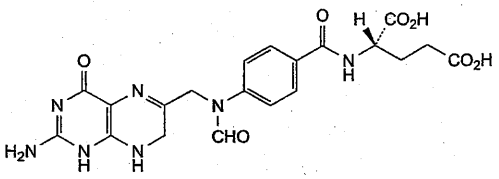
C. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid (folic acid),



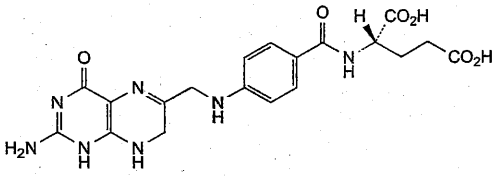
D. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl](formyl)amino]benzamido]pentanedioic acid (10-formylfolic acid),



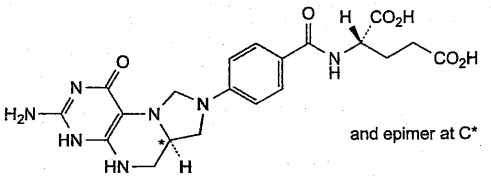
E. 4-[[[(6*RS*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl]amino]benzoic acid (5-formyltetrahydropteroic acid),



F. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4,7,8-tetrahydropteridin-6-yl)methyl](formyl)amino]benzamido]pentanedioic acid (10-formyldihydrofolic acid),



G. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4,7,8-tetrahydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid (dihydrofolic acid),

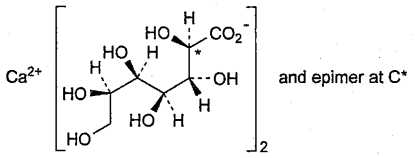


I. (2*S*)-2-[4-[(6a*RS*)-3-amino-1-oxo-1,2,5,6,6a,7-hexahydroimidazo[1,5-*f*]pteridin-8(9*H*)-yl]benzamido]pentanedioic acid ((6a*RS*)-5,10-methylenetetrahydrofolic acid).

Ph Eur

Calcium Glucoheptonate

(Ph. Eur. monograph 1399)



$C_{14}H_{26}CaO_{16}$ 490.4

Action and use
Used in treatment of calcium deficiency.

Ph Eur

DEFINITION
Mixture in variable proportions, of calcium di(D-*glycero*-D-*gulo*-heptonate) and calcium di(D-*glycero*-D-*ido*-heptonate).

Content
98.0 per cent to 102.0 per cent of calcium 2,3,4,5,6,7-hexahydroxyheptanoate (dried substance).

CHARACTERS
Appearance
White or very slightly yellow, amorphous powder, hygroscopic.

Solubility
Very soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *water R*.

Reference solution (a) Dissolve 20 mg of *calcium glucoheptonate CRS* in 1 mL of *water R*.

Reference solution (b) Dissolve 10 mg of *calcium gluconate CRS* in 0.5 mL of the test solution and dilute to 1 mL with *water R*.

Plate *cellulose for chromatography R1* as the coating substance.

Mobile phase *anhydrous formic acid R, water R, acetone R, butanol R* (20:20:30:30 *V/V/V/V*); use a freshly prepared mixture.

Application 10 µL as bands of 20 mm by 2 mm.

Development In a tank previously allowed to saturate for 10 min, over a path of 12 cm.

Drying In air.

Detection Spray with *0.02 M potassium permanganate*.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. 0.2 mL of solution S (see Tests) gives reaction (b) of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
6.0 to 8.0 for solution S.

Reducing sugars
Maximum 1 per cent, expressed as glucose.

Dissolve 1.0 g in 5 mL of *water R* with the aid of gentle heat. Cool and add 20 mL of *cupri-citric solution R* and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent *V/V* solution of *glacial acetic acid R* and 20.0 mL of *0.025 M iodine*. With continuous shaking, add 25 mL of a mixture of 6 volumes of *hydrochloric acid R* and 94 volumes of *water R* until the precipitate dissolves, titrate the excess of iodine with *0.05 M sodium thiosulfate* using 1 mL of *starch solution R* added towards the end of the titration, as indicator. Not less than 12.6 mL of *0.05 M sodium thiosulfate* is required.

Cyanide
Dissolve 5.0 g in 50 mL of *water R* and add 2.0 g of *tartaric acid R*. Place this solution in a distillation apparatus (*2.2.11*). The plain bend adapter attached to the end of the condenser has a vertical part that is long enough to extend to 1 cm from the bottom of a 50 mL test-tube used as a receiver. Place 10 mL of *water R* and 2 mL of *0.1 M sodium hydroxide* into the receiver. Distil, collect 25 mL of distillate and dilute to 50 mL with *water R*. To 25 mL of this solution add 25 mg of *ferrous sulfate R* and boil for a short time. After cooling to about 70 °C add 10 mL of *hydrochloric acid R1*. After 30 min, filter the solution and wash the filter. A yellow spot appears on the filter; there is no blue or green spot.

Chlorides (*2.4.4*)
Maximum 100 ppm.

To 5 mL of solution S, add 10 mL of *water R*.

Sulfates (*2.4.13*)
Maximum 100 ppm, determined on solution S.

Iron (*2.4.9*)
Maximum 40 ppm.

Dilute 2.5 mL of solution S to 10 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 167 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Dissolve 0.800 g in a mixture of 2 mL of *3 M hydrochloric acid* and 150 mL of *water R*. While stirring, add 12.5 mL of *0.1 M sodium edetate*, 15 mL of *1 M sodium hydroxide* and 0.3 g of *hydroxynaphthol blue, sodium salt R*. Titrate with *0.1 M sodium edetate* until the colour changes from violet to pure blue.

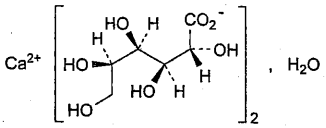
1 mL of *0.1 M sodium edetate* is equivalent to 49.04 mg of $C_{14}H_{26}CaO_{16}$.

STORAGE
In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

Ph Eur

Calcium Gluconate

(Ph. Eur. monograph 0172)



$C_{12}H_{22}CaO_{14},H_2O$ 448.4 *18016-24-5*

Action and use
Used in treatment of calcium deficiency.

Preparations
Calcium Gluconate Tablets
Calcium Gluconate Chewable Tablets
Calcium Gluconate Effervescent Tablets

Ph Eur

DEFINITION
Calcium bis[(2*R*,3*S*,4*R*,5*R*)-2,3,4,5,6-pentahydroxyhexanoate] monohydrate (calcium di(D-gluconate) monohydrate).

Content
98.5 per cent to 102.0 per cent of $C_{12}H_{22}CaO_{14},H_2O$.

CHARACTERS
Appearance
White or almost white, crystalline or granular powder.

Solubility
Sparingly soluble in water, freely soluble in boiling water.

IDENTIFICATION
A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Reference solution Dissolve 20 mg of *calcium gluconate CRS* in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Plate *TLC silica gel plate R* (5-40 µm) [or *TLC silica gel plate R* (2-10 µm)].

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).

Application 1 µL.

Development Over 2/3 of the plate.

Drying At 100 °C for 20 min; allow to cool.

Detection Spray with a solution containing 10 g/L of *cerium sulfate R* and 25 g/L of *ammonium molybdate R* in *dilute sulfuric acid R* and heat at 105 °C for about 10 min.

Results After 5 min, the principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Solution S (see Tests) gives the reactions of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *water R* heated to 60 °C and dilute to 50 mL with the same solvent.

Appearance of solution
At 60 °C, solution S is not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*). After cooling, it is not more opalescent than reference suspension II (*2.2.1*).

Organic impurities and boric acid
Introduce 0.5 g into a porcelain dish previously rinsed with *sulfuric acid R* and placed in a bath of iced water. Add 2 mL of cooled *sulfuric acid R* and mix. No yellow or brown colour develops. Add 1 mL of *chromotrope II B solution R*. A violet colour develops and does not become dark blue.
The solution is not more intensely coloured than that of a mixture of 1 mL of *chromotrope II B solution R* and 2 mL of cooled *sulfuric acid R*.

Sucrose and reducing sugars
Dissolve 0.5 g in a mixture of 2 mL of *hydrochloric acid R1* and 10 mL of *water R*. Boil for 5 min, allow to cool, add 10 mL of *sodium carbonate solution R* and allow to stand. Dilute to 25 mL with *water R* and filter. To 5 mL of the filtrate add 2 mL of *cupri-tartaric solution R* and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 12.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 100 ppm.

Dissolve 10.0 g with heating in a mixture of 10 mL of *acetic acid R* and 90 mL of *distilled water R*.

Magnesium and alkali metals
Maximum 0.4 per cent.

Dissolve 1.00 g in 100 mL of boiling *water R*, add 10 mL of *ammonium chloride solution R*, 1 mL of *ammonia R* and, dropwise, 50 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 200 mL with *water R* and filter. Evaporate 100 mL of the filtrate to dryness and ignite. The residue weighs a maximum of 2 mg.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

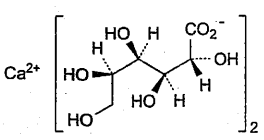
ASSAY
Dissolve 0.8000 g in 20 mL of hot *water R*, allow to cool and dilute to 300 mL with *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 44.84 mg of $C_{12}H_{22}CaO_{14},H_2O$.

Ph Eur

Anhydrous Calcium Gluconate

(Ph. Eur. monograph 2364)



$C_{12}H_{22}CaO_{14}$ 430.4

Action and use
Used in treatment of calcium deficiency.

Ph Eur

DEFINITION
Anhydrous calcium D-gluconate.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline or granular powder.

Solubility
Sparingly soluble in water, freely soluble in boiling water.

IDENTIFICATION
A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Reference solution Dissolve 20 mg of *calcium gluconate CRS* in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Plate *TLC silica gel plate R* (5-40 µm) [or *TLC silica gel plate R* (2-10 µm)].

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).

Application 1 µL.

Development Over 2/3 of the plate.

Drying At 100 °C for 20 min, then allow to cool.

Detection Spray with a solution containing 25 g/L of *ammonium molybdate R* and 10 g/L of *cerium sulfate R* in *dilute sulfuric acid R*, and heat at 100-105 °C for about 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Solution S (see Tests) gives the reactions of calcium (*2.3.1*).

C. Loss on drying (see Tests).

TESTS
Solution S
Dissolve 1.0 g in *water R* heated to 60 °C and dilute to 50 mL with the same solvent.

Appearance of solution
At 60 °C, solution S is not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*). After cooling, it is not more opalescent than reference suspension II (*2.2.1*).

Organic impurities and boric acid
Place 0.5 g in a porcelain dish previously rinsed with *sulfuric acid R* and placed in a bath of iced water. Add 2 mL of cooled *sulfuric acid R* and mix. No yellow or brown colour develops. Add 1 mL of *chromotrope II B solution R*. A violet colour develops and does not become dark blue. Compare the colour obtained with that of a mixture of 1 mL of *chromotrope II B solution R* and 2 mL of cooled *sulfuric acid R*.

Sucrose and reducing sugars
Dissolve 0.5 g in a mixture of 2 mL of *hydrochloric acid R1* and 10 mL of *water R*. Boil for 5 min, allow to cool, add 10 mL of *sodium carbonate solution R* and allow to stand for 10 min. Dilute to 25 mL with *water R* and filter. To 5 mL of the filtrate add 2 mL of *cupri-tartaric solution R* and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 12.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 100 ppm.

Dissolve 10.0 g with heating in a mixture of 10 mL of *acetic acid R* and 90 mL of *distilled water R*.

Magnesium and alkali metals
Maximum 0.4 per cent (expressed as MgO).

Dissolve 1.00 g in 100 mL of boiling *water R*, add 10 mL of *ammonium chloride solution R*, 1 mL of *ammonia R* and, dropwise, 50 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 200 mL with *water R* and filter. Evaporate 100 mL of the filtrate to dryness and ignite. The residue weighs a maximum of 2 mg.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 16 h.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

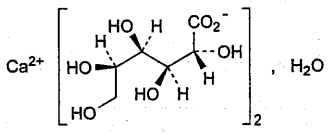
ASSAY
Dissolve 0.350 g in 20 mL of hot *water R*, allow to cool and dilute to 300 mL with *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 43.04 mg of $C_{12}H_{22}CaO_{14}$.

Ph Eur

Calcium Gluconate for Injection

(Ph. Eur. monograph 0979)



$C_{12}H_{22}CaO_{14},H_2O$ 448.4 *18016-24-5*

Action and use
Used in treatment of calcium deficiency.

Preparation
Calcium Gluconate Injection

Ph Eur

DEFINITION
Calcium bis[(2*R*,3*S*,4*R*,5*R*)-2,3,4,5,6-pentahydroxyhexanoate] monohydrate (calcium di(D-gluconate) monohydrate).

Content
99.0 per cent to 101.0 per cent of $C_{12}H_{22}CaO_{14},H_2O$.

CHARACTERS
Appearance
White or almost white, crystalline or granular powder.

Solubility
Sparingly soluble in water, freely soluble in boiling water.

IDENTIFICATION
A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Reference solution Dissolve 20 mg of *calcium gluconate CRS* in 1 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Plate *TLC silica gel plate R* (5-40 µm) [or *TLC silica gel plate R* (2-10 µm)].

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).

Application 1 µL.

Development Over 2/3 of the plate.

Drying At 100 °C for 20 min; allow to cool.

Detection Spray with a solution containing 10 g/L of *cerium sulfate R* and 25 g/L of *ammonium molybdate R* in *dilute sulfuric acid R* and heat at 105 °C for about 10 min.

Results After 5 min, the principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. About 20 mg gives reaction (b) of calcium (*2.3.1*).

TESTS
Solution S
To 10.0 g add 90 mL of boiling *distilled water R* and boil with stirring, for not more than 10 s, until completely dissolved, then dilute to 100.0 mL with the same solvent.

Appearance of solution
At 60 °C, solution S is not more intensely coloured than reference solution B_7 (*2.2.2, Method II*). After cooling to 20 °C, it is not more opalescent than reference suspension II (*2.2.1*).

pH (*2.2.3*)
6.4 to 8.3.

Dissolve 1.0 g in 20 mL of *carbon dioxide-free water R*, heating on a water-bath.

Organic impurities and boric acid
Introduce 0.5 g into a porcelain dish previously rinsed with *sulfuric acid R* and placed in a bath of iced water. Add 2 mL of cooled *sulfuric acid R* and mix. No yellow or brown colour develops. Add 1 mL of *chromotrope II B solution R*. A violet colour develops and does not become dark blue.
The solution is not more intensely coloured than that of a mixture of 1 mL of *chromotrope II B solution R* and 2 mL of cooled *sulfuric acid R*.

Oxalates
Liquid chromatography (*2.2.29*).

Test solution Dissolve 1.00 g of the substance to be examined in *water for chromatography R* and dilute to 100.0 mL with the same solvent.

Reference solution Dissolve 1.00 g of the substance to be examined in *water for chromatography R*, add 0.5 mL of a 0.152 g/L solution of *sodium oxalate R* in *water for chromatography R* and dilute to 100.0 mL with the same solvent.

Precolumn:
— *size*: *l* = 30 mm, Ø = 4 mm;
— *stationary phase*: suitable strong anion-exchange resin (30-50 µm).

Columns 1 and 2:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: suitable strong anion-exchange resin (30-50 µm).

Anion-suppresser column Connected in series with the precolumn and analytical columns and equipped with a micromembrane that separates the mobile phase from the suppressor regeneration solution, flowing countercurrent to the mobile phase.

Mobile phase Dissolve 0.212 g of *anhydrous sodium carbonate R* and 63 mg of *sodium hydrogen carbonate R* in *water for chromatography R* and dilute to 1000.0 mL with the same solvent.

Flow rate of the mobile phase 2 mL/min.

Suppressor regeneration solution 1.23 g/L solution of *sulfuric acid R* in *water for chromatography R*.

Flow rate of the suppressor regeneration solution 4 mL/min.

Detection Conductance.

Injection 50 µL.

System suitability Reference solution:
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the area of the peak due to oxalate after 5 injections.

Inject 50 µL of each solution 3 times. Calculate the content of oxalates in parts per million using the following expression:

$$\frac{S_T \times 50}{S_R - S_T}$$

S_T = area of the peak due to oxalate in the chromatogram obtained with the test solution;
S_R = area of the peak due to oxalate in the chromatogram obtained with the reference solution.

Limit:
— *oxalates*: maximum 100 ppm.

Sucrose and reducing sugars
Dissolve 0.5 g in a mixture of 2 mL of *hydrochloric acid R1* and 10 mL of *water R*. Boil for 5 min, allow to cool, add 10 mL of *sodium carbonate solution R* and allow to stand for 10 min. Dilute to 25 mL with *water R* and filter. To 5 mL of the filtrate add 2 mL of *cupri-tartaric solution R* and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (*2.4.4*)
Maximum 50 ppm.

To 10 mL of previously filtered solution S add 5 mL of *water R*.

Phosphates (*2.4.11*)
Maximum 100 ppm.

Dilute 1 mL of solution S to 100 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 50 ppm, determined on previously filtered solution S.

Prepare the standard using a mixture of 7.5 mL of *sulfate standard solution (10 ppm SO_4) R* and 7.5 mL of *distilled water R*.

Iron
Maximum 5 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Introduce 2.0 g into a 100 mL polytetrafluoroethylene beaker and add 5 mL of *nitric acid R*. Boil, evaporating almost to dryness. Add 1 mL of *strong hydrogen peroxide solution R* and evaporate again almost to dryness. Repeat the hydrogen peroxide treatment until a clear solution is obtained. Using 2 mL of *nitric acid R*, transfer the solution into a 25 mL volumetric flask. Dilute to 25.0 mL with *dilute hydrochloric acid R*. In the same manner, prepare a compensation solution using 0.65 g of *calcium chloride R1* instead of the substance to be examined.

Reference solutions Prepare the reference solutions from *iron standard solution (20 ppm Fe) R*, diluting with *dilute hydrochloric acid R*.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Carry out a basic correction using a deuterium lamp.

Magnesium and alkali metals
Maximum 0.4 per cent.

To 0.50 g add a mixture of 1.0 mL of *dilute acetic acid R* and 10.0 mL of *water R* and rapidly boil, whilst shaking, until completely dissolved. To the boiling solution add 5.0 mL of *ammonium oxalate solution R* and allow to stand for at least 6 h. Filter through a sintered-glass filter (1.6) (*2.1.2*) into a porcelain crucible. Carefully evaporate the filtrate to dryness and ignite. The residue weighs not more than 2 mg.

Bacterial endotoxins (*2.6.14*)
Less than 167 IU/g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

ASSAY
Dissolve 0.350 g in 20 mL of hot *water R*, allow to cool and dilute to 300 mL with *water R*. Carry out the complexometric titration of calcium (*2.5.11*). Use 50 mg of *calconecarboxylic acid triturate R*.

1 mL of *0.1 M sodium edetate* is equivalent to 44.84 mg of $C_{12}H_{22}CaO_{14},H_2O$.

Ph Eur

Calcium Glycerophosphate

(Ph. Eur. monograph 0980)

$C_3H_7CaO_6P$ 210.1

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture in variable proportions of the calcium salt of (*RS*)-2,3-dihydroxypropyl phosphate and of 2-hydroxy-1-(hydroxymethyl)ethyl phosphate which may be hydrated.

Content
18.6 per cent to 19.4 per cent of Ca (dried substance).

CHARACTERS
Appearance
White or almost white powder, hygroscopic.

Solubility
Sparingly soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Mix 1 g with 1 g of *potassium hydrogen sulfate R* in a test tube fitted with a glass tube. Heat strongly and direct the white vapour towards a piece of filter paper impregnated with a freshly prepared 10 g/L solution of *sodium nitroprusside R*.

The filter paper develops a blue colour in contact with *piperidine R*.

B. Ignite 0.1 g in a crucible. Take up the residue with 5 mL of *nitric acid R* and heat on a water-bath for 1 min. Filter. The filtrate gives reaction (b) of phosphates (*2.3.1*).

C. It gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S

Dissolve 1.5 g at room temperature in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 150 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension III (*2.2.1*).

Acidity or alkalinity

To 100 mL of solution S add 0.1 mL of *phenolphthalein solution R*. Not more than 1.5 mL of *0.1 M hydrochloric acid* or 0.5 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Citric acid

Shake 5.0 g with 20 mL of *carbon dioxide-free water R* and filter. To the filtrate add 0.15 mL of *sulfuric acid R* and filter again. To the filtrate add 5 mL of mercuric sulfate solution R and heat to boiling. Add 0.5 mL of a 3.2 g/L solution of *potassium permanganate R* and again heat to boiling. No precipitate is formed.

Glycerol and ethanol (96 per cent)-soluble substances

Maximum 0.5 per cent.

Shake 1.000 g with 25 mL of *ethanol (96 per cent) R* for 1 min. Filter. Evaporate the filtrate on a water-bath and dry the residue at 70 °C for 1 h. The residue weighs a maximum of 5 mg.

Chlorides (*2.4.4*)

Maximum 500 ppm.

Dissolve 0.1 g in a mixture of 2 mL of *acetic acid R* and 8 mL of *water R* and dilute to 15 mL with *water R*.

Phosphates (*2.4.11*)

Maximum 400 ppm.

Dilute 2.5 mL of solution S to 100 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 0.1 per cent, determined on solution S.

Arsenic (*2.4.2, Method A*)

Maximum 3 ppm.

Dissolve 0.33 g in *water R* and dilute to 25 mL with the same solvent.

Iron (*2.4.9*)

Maximum 50 ppm, detemined on 0.20 g.

Loss on drying (*2.2.32*)

Maximum 12.0 per cent, determined on 1.000 g by drying in an oven at 150 °C for 4 h.

ASSAY

Dissolve 0.200 g in *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 4.008 mg of Ca.

Ph Eur

Calcium Hydrogen Phosphate[1]

Anhydrous Calcium Hydrogen Phosphate

(Ph. Eur. monograph 0981)

$CaHPO_4$ 136.1 *7757-93-9*

Ph Eur

DEFINITION

Content

97.5 per cent to 102.5 per cent.

♦CHARACTERS

Appearance

White or almost white, crystalline powder, or colourless crystals.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute hydrochloric acid and in dilute nitric acid.♦

IDENTIFICATION

A. Dissolve with heating 0.1 g in 10 mL of *dilute hydrochloric acid R*. Add 2.5 mL of *dilute ammonia R1*, shake, and add 5 mL of a 35 g/L solution of *ammonium oxalate R*. A white precipitate is produced.

B. Dissolve 0.1 g in 5 mL of *dilute nitric acid R*, add 2 mL of *ammonium molybdate solution R* and heat at 70 °C for 1-2 min. A yellow precipitate is produced.

◊C. It complies with the limits of the assay.◊

TESTS

♦Solution S

Dissolve 2.5 g in 20 mL of *dilute hydrochloric acid R*, filter if necessary and add *dilute ammonia R1* until a precipitate is formed. Add just sufficient *dilute hydrochloric acid R* to dissolve the precipitate and dilute to 50 mL with *distilled water R*.♦

Acid-insoluble substances

Maximum 0.2 per cent.

Dissolve 5.0 g in 40 mL of *water R*, add 10 mL of *hydrochloric acid R* and heat to boiling for 5 min. Cool, then collect the insoluble substances using ashless filter paper. Wash with *water R* until turbidity is no longer produced when *silver nitrate solution R2* is added. Ignite the residue and the filter paper at 600 ± 50 °C. The residue weighs not more than 10 mg.

Carbonates

Shake 1.0 g with 5 mL of *carbon dioxide-free water R* and add 2 mL of *hydrochloric acid R*. No effervescence is produced.

Chlorides

Maximum 0.25 per cent.

Test solution Dissolve 0.20 g in a mixture of 20 mL of *water R* and 13 mL of *dilute nitric acid R* by warming if necessary, dilute to 100 mL with *water R* and filter if necessary. Use 50 mL of this solution.

Reference solution To 0.70 mL of *0.01 M hydrochloric acid*, add 6 mL of *dilute nitric acid R* and dilute to 50 mL with *water R*.

Add 1 mL of *silver nitrate solution R2* to the test solution and to the reference solution and mix. After standing for 5 min protected from light, any opalescence in the test solution, by

[1] *This monograph has undergone the process of Pharmacopoeial harmonisation. See chapter 5.8. Pharmacopoeial harmonisation.*

viewing vertically or horizontally against a black background, is not more intense than that in the reference solution.

♦Fluorides
Maximum 100 ppm.

Potentiometry (*2.2.36, Method II*).

Chelating solution Dissolve 45 g of *cyclohexylenedinitrilotetra-acetic acid R* in 75 mL of *sodium hydroxide solution R* and dilute to 250 mL with *water R*.

Test solution Dissolve 1.000 g in 4 mL of *hydrochloric acid R1*, add 20 mL of chelating solution, 2.7 mL of *glacial acetic acid R* and 2.8 g of *sodium chloride R*, adjust to pH 5-6 with *sodium hydroxide solution R* and dilute to 50.0 mL with *water R*.

Reference solution Dissolve 4.42 g of *sodium fluoride R*, previously dried at 300 °C for 12 h, in *water R* and dilute to 1000.0 mL with the same solvent. Dilute 50.0 mL of this solution to 500.0 mL with *total-ionic-strength-adjustment buffer R* (200 ppm F).

Indicator electrode Fluoride-selective.

Reference electrode Silver-silver chloride.

Carry out the measurement on 20.0 mL of the test solution. Add at least 3 times 0.10 mL of the reference solution and carry out the measurement after each addition. Calculate the concentration of fluorides using the calibration curve.♦

Sulfates
Maximum 0.5 per cent.

Test solution Dissolve 0.5 g in a mixture of 5 mL of *water R* and 5 mL of *dilute hydrochloric acid R* and dilute to 100 mL with *water R*. Filter if necessary. To 20 mL of this solution, add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R*.

Reference solution To 1.0 mL of *0.005 M sulfuric acid*, add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R*. Filter if necessary.

To the test solution and to the reference solution, add 2 mL of a 120 g/L solution of *barium chloride R* and allow to stand for 10 min. Any opalescence in the test solution, by viewing vertically or horizontally against a black background, is not more intense than that in the reference solution.

♦Arsenic (*2.4.2, Method A*)
Maximum 10 ppm, determined on 2 mL of solution S.♦

Barium
To 0.5 g, add 10 mL of *water R* and heat to boiling. While stirring, add 1 mL of *hydrochloric acid R* dropwise. Allow to cool and filter if necessary. Add 2 mL of a 10 g/L solution of *dipotassium sulfate R* and allow to stand for 10 min.
No turbidity is produced.

♦Iron (*2.4.9*)
Maximum 400 ppm.

Dilute 0.5 mL of solution S to 10 mL with *water R*.♦

Loss on ignition
6.6 per cent to 8.7 per cent, determined on 1.000 g to constant mass at 800-825 °C.

ASSAY
Dissolve 0.4 g in 12 mL of *dilute hydrochloric acid R* by heating on a water bath if necessary and dilute to 200.0 mL with *water R*. To 20.0 mL of this solution add 25.0 mL of *0.02 M sodium edetate*, 50 mL of *water R*, 5 mL of *ammonium chloride buffer solution pH 10.7 R* and about 25 mg of *mordant black 11 triturate R*. Titrate the excess of sodium edetate with *0.02 M zinc sulfate*. Carry out a blank titration.

1 mL of *0.02 M sodium edetate* is equivalent to 2.721 mg of $CaHPO_4$.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium hydrogen phosphate used as filler in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

Bulk and tapped density (*2.9.34*)

Powder flow (*2.9.36*)

Ph Eur

Calcium Hydrogen Phosphate Dihydrate

Dibasic Calcium Phosphate

(Ph. Eur. monograph 0116)

NOTE: The name Calcium Hydrogen Phosphate was formerly used in the United Kingdom.

$CaHPO_4,2H_2O$ 172.1 *7789-77-7*

Ph Eur

DEFINITION
Content
98.0 per cent to 105.0 per cent.

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute hydrochloric acid and in dilute nitric acid.

IDENTIFICATION
A. Dissolve with heating 0.1 g in 10 mL of *dilute hydrochloric acid R*. Add 2.5 mL of *dilute ammonia R1*, shake and add 5 mL of a 35 g/L solution of *ammonium oxalate R*. A white precipitate is produced.

B. Dissolve 0.1 g in 5 mL of *dilute nitric acid R*, add 2 mL of *ammonium molybdate solution R* and heat at 70 °C for 2 min. A yellow precipitate is produced.

C. It complies with the limits of the assay.

TESTS
Solution S
Dissolve 2.5 g in 20 mL of *dilute hydrochloric acid R*, filter if necessary and add *dilute ammonia R1* until a precipitate is formed. Add just sufficient *dilute hydrochloric acid R* to

dissolve the precipitate and dilute to 50 mL with *distilled water R*.

Acid-insoluble substances
Maximum 0.2 per cent.

Dissolve 5.0 g in 40 mL of *water R*, add 10 mL of *hydrochloric acid R* and heat to boiling for 5 min. Cool, then collect the insoluble substances using ashless filter paper. Wash with *water R* until turbidity is no longer produced when *silver nitrate solution R2* is added to the filtrate. Ignite at 600 ± 50 °C. The residue weighs not more than 10 mg.

Carbonates
Shake 0.5 g with 5 mL of *carbon dioxide-free water R* and add 1 mL of *hydrochloric acid R*. No effervescence is produced.

Chlorides
Maximum 0.25 per cent.

Test solution Dissolve 0.20 g in a mixture of 20 mL of *water R* and 13 mL of *dilute nitric acid R* by warming if necessary, dilute to 100 mL with *water R* and filter if necessary. Use 50 mL of this solution.

Reference solution To 0.70 mL of *0.01 M hydrochloric acid*, add 6 mL of *dilute nitric acid R* and dilute to 50 mL with *water R*.

Add 1 mL of *silver nitrate solution R2* to the test solution and to the reference solution and mix. After standing for 5 min protected from light, any opalescence in the test solution is not more intense than that in the reference solution.

Fluorides
Maximum 100 ppm.

Potentiometry (*2.2.36, Method II*).

Chelating solution Dissolve 45 g of *cyclohexylenedinitrilotetra-acetic acid R* in 75 mL of *sodium hydroxide solution R* and dilute to 250 mL with *water R*.

Test solution Dissolve 1.000 g in 4 mL of *hydrochloric acid R1*, add 20 mL of chelating solution, 2.7 mL of *glacial acetic acid R* and 2.8 g of *sodium chloride R*, adjust to pH 5-6 with *sodium hydroxide solution R* and dilute to 50.0 mL with *water R*.

Reference solution Dissolve 4.42 g of *sodium fluoride R*, previously dried at 300 °C for 12 h, in *water R* and dilute to 1000.0 mL with the same solvent. Dilute 50.0 mL of this solution to 500.0 mL with *total-ionic-strength-adjustment buffer R* (200 ppm F).

Indicator electrode Fluoride-selective.

Reference electrode Silver-silver chloride.

Carry out the measurement on 20.0 mL of the test solution. Add at least 3 times 0.10 mL of the reference solution and carry out the measurement after each addition. Calculate the concentration of fluorides using the calibration curve.

Sulfates
Maximum 0.5 per cent.

Test solution Dissolve 0.5 g in a mixture of 5 mL of *water R* and 5 mL of *dilute hydrochloric acid R* and dilute to 100 mL with *water R*. Filter if necessary. To 20 mL of this solution, add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R*.

Reference solution To 1.0 mL of *0.005 M sulfuric acid*, add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R*. Filter if necessary.

To the test solution and to the reference solution, add 2 mL of a 120 g/L solution of *barium chloride R* and allow to stand for 10 min. Any opalescence in the test solution is not more intense than that in the reference solution.

Arsenic (*2.4.2, Method A*)
Maximum 10 ppm, determined on 2 mL of solution S.

Barium
To 0.5 g, add 10 mL of *water R* and heat to boiling. While stirring, add 1 mL of *hydrochloric acid R* dropwise. Allow to cool and filter if necessary. Add 2 mL of a 10 g/L solution of *dipotassium sulfate R* and allow to stand for 10 min.
No turbidity is produced.

Iron (*2.4.9*)
Maximum 400 ppm.

Dilute 0.5 mL of solution S to 10 mL with *water R*.

Loss on ignition
24.5 per cent to 26.5 per cent, determined on 1.000 g by ignition to constant mass at 800-825 °C.

ASSAY
Dissolve 0.4 g in 12 mL of *dilute hydrochloric acid R* by heating on a water bath if necessary and dilute to 200 mL with *water R*. To 20.0 mL of this solution add 25.0 mL of *0.02 M sodium edetate*, 50 mL of *water R*, 5 mL of *ammonium chloride buffer solution pH 10.7 R* and about 25 mg of *mordant black 11 triturate R*. Titrate the excess of sodium edetate with *0.02 M zinc sulfate*. Carry out a blank titration.

1 mL of *0.02 M sodium edetate* is equivalent to 3.44 mg of $CaHPO_4,2H_2O$.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium hydrogen phosphate dihydrate used as filler in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

Bulk and tapped density (*2.9.34*)

Powder flow (*2.9.36*)

Ph Eur

Calcium Hydroxide

(Ph. Eur. monograph 1078)

$Ca(OH)_2$ 74.1 *1305-62-0*

Preparation
Calcium Hydroxide Solution

Ph Eur

DEFINITION
Content
95.0 per cent to 100.5 per cent.

CHARACTERS
Appearance
White or almost white, fine powder.

Solubility
Practically insoluble in water.

IDENTIFICATION
A. To 0.80 g in a mortar, add 10 mL of *water R* and 0.5 mL of *phenolphthalein solution R* and mix. The suspension turns red. On addition of 17.5 mL of *1 M hydrochloric acid*, the suspension becomes colourless without effervescing. The red colour occurs again when the mixture is triturated for 1 min. On addition of a further 6 mL of *1 M hydrochloric acid* and triturating, the solution becomes colourless.

B. Dissolve about 0.1 g in *dilute hydrochloric acid R* and dilute to 10 mL with *water R*. 5 mL of the solution give reaction (b) of calcium (*2.3.1*).

TESTS
Matter insoluble in hydrochloric acid
Maximum 0.5 per cent.

Dissolve 2.0 g in 30 mL of *hydrochloric acid R*. Boil the solution and filter. Wash the residue with hot *water R*. The residue weighs a maximum of 10 mg.

Carbonates
Maximum 5.0 per cent of $CaCO_3$.

Add 5.0 mL of *1 M hydrochloric acid* to the titrated solution obtained under Assay and titrate with *1 M sodium hydroxide* using 0.5 mL of *methyl orange solution R* as indicator.

1 mL of *1 M hydrochloric acid* is equivalent to 50.05 mg of $CaCO_3$.

Chlorides (*2.4.4*)
Maximum 330 ppm.

Dissolve 0.30 g in a mixture of 2 mL of *nitric acid R* and 10 mL of *water R* and dilute to 30 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 0.4 per cent.

Dissolve 0.15 g in a mixture of 5 mL of *dilute hydrochloric acid R* and 10 mL of *distilled water R* and dilute to 60 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 4 ppm.

Dissolve 0.50 g in 5 mL of *brominated hydrochloric acid R* and dilute to 50 mL with *water R*. Use 25 mL of this solution.

Magnesium and alkali metals
Maximum 4.0 per cent calculated as sulfates.

Dissolve 1.0 g in a mixture of 10 mL of *hydrochloric acid R* and 40 mL of *water R*. Boil and add 50 mL of a 63 g/L solution of *oxalic acid R*. Neutralise with *ammonia R* and dilute to 200 mL with *water R*. Allow to stand for 1 h and filter through a suitable filter. To 100 mL of the filtrate, add 0.5 mL of *sulfuric acid R*. Cautiously evaporate to dryness and ignite. The residue weighs a maximum of 20 mg.

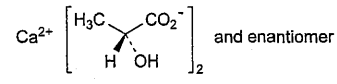
ASSAY
To 1.500 g in a mortar, add 20-30 mL of *water R* and 0.5 mL of *phenolphthalein solution R*. Titrate with *1 M hydrochloric acid* by triturating the substance until the red colour disappears. The final solution is used in the tests for carbonates.

1 mL of *1 M hydrochloric acid* is equivalent to 37.05 mg of $Ca(OH)_2$.

Ph Eur

Calcium Lactate

Anhydrous Calcium Lactate
(Ph. Eur. monograph 2118)



$C_6H_{10}CaO_6$ 218.2

Action and use
Used in treatment of calcium deficiency.

Ph Eur

DEFINITION
Calcium bis(2-hydroxypropanoate) or mixture of calcium (2*R*)-, (2*S*)- and (2*RS*)-2-hydroxypropanoates.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline or granular powder.

Solubility
Soluble in water, freely soluble in boiling water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Loss on drying (see Tests).

B. It gives the reaction of lactates (*2.3.1*).

C. It gives reaction (b) of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g with heating in *carbon dioxide-free water R* prepared from *distilled water R*, allow to cool and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M hydrochloric acid*.
The solution is colourless. Not more than 2.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 400 ppm.

Dilute 7.5 mL of solution S to 15 mL with *distilled water R*.

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R*. Allow to stand for 15 min. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 50 ppm.

Dilute 4 mL of solution S to 10 mL with *water R*.

Magnesium and alkali salts
Maximum 1 per cent.

To 20 mL of solution S add 20 mL of *water R*, 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1*. Heat to boiling and rapidly add 40 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 100.0 mL with *water R* and filter. To 50.0 mL of the filtrate add 0.5 mL of *sulfuric acid R*. Evaporate to dryness and ignite the residue to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 0.500 g by drying in an oven at 125 °C.

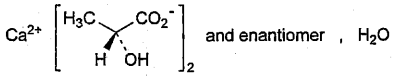
ASSAY
Dissolve 0.200 g in *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 21.82 mg of $C_6H_{10}CaO_6$.

Ph Eur

Calcium Lactate Monohydrate

(Ph. Eur. monograph 2117)



$C_6H_{10}CaO_6,H_2O$ 236.0

Action and use
Used in treatment of calcium deficiency.

Ph Eur

DEFINITION
Calcium bis(2-hydroxypropanoate) or mixture of calcium (2*R*)-, (2*S*)- and (2*RS*)-2-hydroxypropanoates monohydrates.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline or granular powder.

Solubility
Soluble in water, freely soluble in boiling water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Loss on drying (see Tests).
B. It gives the reaction of lactates (*2.3.1*).
C. It gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S
Dissolve 5.4 g (equivalent to 5.0 g of the dried substance) with heating in *carbon dioxide-free water R* prepared from *distilled water R*, allow to cool and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M hydrochloric acid*. The solution is colourless. Not more than 2.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Chlorides (*2.4.4*)
Maximum 200 ppm.
Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 400 ppm.
Dilute 7.5 mL of solution S to 15 mL with *distilled water R*.

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R*. Allow to stand for 15 min. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 50 ppm.
Dilute 4 mL of solution S to 10 mL with *water R*.

Magnesium and alkali salts
Maximum 1 per cent.
To 20 mL of solution S add 20 mL of *water R*, 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1*. Heat to boiling and rapidly add 40 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 100.0 mL with *water R* and filter. To 50.0 mL of the filtrate add 0.5 mL of *sulfuric acid R*. Evaporate to dryness and ignite the residue to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

Loss on drying (*2.2.32*)
5.0 per cent to 8.0 per cent, determined on 0.500 g by drying in an oven at 125 °C.

ASSAY
Dissolve a quantity equivalent to 0.200 g of the dried substance in *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).

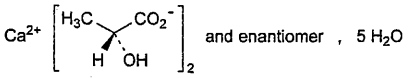
1 mL of *0.1 M sodium edetate* is equivalent to 21.82 mg of $C_6H_{10}CaO_6$.

Ph Eur

Calcium Lactate Pentahydrate

Calcium Lactate

(Ph. Eur. monograph 0468)



$C_6H_{10}CaO_6,5H_2O$ 308.3

Action and use
Used in treatment of calcium deficiency.

Preparations
Calcium and Ergocalciferol Tablets
Calcium Lactate Tablets
Calcium and Ergocalciferol Chewable Tablets

Ph Eur

DEFINITION
Calcium bis(2-hydroxypropanoate) or mixture of calcium (2*R*)-, (2*S*)- and (2*RS*)-2-hydroxypropanoates pentahydrates.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline or granular powder, slightly efflorescent.

Solubility
Soluble in water, freely soluble in boiling water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Loss on drying (see Tests).
B. It gives the reaction of lactates (*2.3.1*).
C. It gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S
Dissolve 7.1 g (equivalent to 5.0 g of the dried substance) with heating in *carbon dioxide-free water R* prepared from *distilled water R*, allow to cool and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M hydrochloric acid.*
The solution is colourless. Not more than 2.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Chlorides (*2.4.4*)
Maximum 200 ppm.
Dilute 5 mL of solution S to 15 mL with *water R.*

Sulfates (*2.4.13*)
Maximum 400 ppm.
Dilute 7.5 mL of solution S to 15 mL with *distilled water R.*

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R.* Allow to stand for 15 min. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 50 ppm.
Dilute 4 mL of solution S to 10 mL with *water R.*

Magnesium and alkali salts
Maximum 1 per cent.
To 20 mL of solution S add 20 mL of *water R*, 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1.* Heat to boiling and rapidly add 40 mL of hot *ammonium oxalate solution R.* Allow to stand for 4 h, dilute to 100.0 mL with *water R* and filter. To 50.0 mL of the filtrate add 0.5 mL of *sulfuric acid R.* Evaporate to dryness and ignite the residue to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

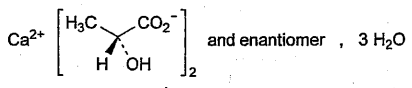
Loss on drying (*2.2.32*)
22.0 per cent to 27.0 per cent, determined on 0.500 g by drying in an oven at 125 °C.

ASSAY
Dissolve a quantity equivalent to 0.200 g of the dried substance in *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).
1 mL of *0.1 M sodium edetate* is equivalent to 21.82 mg of $C_6H_{10}CaO_6$.

Ph Eur

Calcium Lactate Trihydrate

(Ph. Eur. monograph 0469)



$C_6H_{10}CaO_6,3H_2O$ 272.3

Action and use
Used in treatment of calcium deficiency.

Preparation
Calcium Lactate Tablets

Ph Eur

DEFINITION
Calcium bis(2-hydroxypropanoate) or mixture of calcium (2*R*)-, (2*S*)- and (2*RS*)-2-hydroxypropanoates trihydrates.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline or granular powder.

Solubility
Soluble in water, freely soluble in boiling water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Loss on drying (see Tests).
B. It gives the reaction of lactates (*2.3.1*).
C. It gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S
Dissolve 6.2 g (equivalent to 5.0 g of the dried substance) with heating in *carbon dioxide-free water R* prepared from *distilled water R*, allow to cool and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M hydrochloric acid.*
The solution is colourless. Not more than 2.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Chlorides (*2.4.4*)
Maximum 200 ppm.
Dilute 5 mL of solution S to 15 mL with *water R.*

Sulfates (*2.4.13*)
Maximum 400 ppm.
Dilute 7.5 mL of solution S to 15 mL with *distilled water R.*

Barium
To 10 mL of solution S add 1 mL of *calcium sulfate solution R.* Allow to stand for 15 min. Any opalescence in the

solution is not more intense than that in a mixture of 1 mL of *distilled water R* and 10 mL of solution S.

Iron (*2.4.9*)
Maximum 50 ppm.

Dilute 4 mL of solution S to 10 mL with *water R*.

Magnesium and alkali salts
Maximum 1 per cent.

To 20 mL of solution S add 20 mL of *water R*, 2 g of *ammonium chloride R* and 2 mL of *dilute ammonia R1*. Heat to boiling and rapidly add 40 mL of hot *ammonium oxalate solution R*. Allow to stand for 4 h, dilute to 100.0 mL with *water R* and filter. To 50.0 mL of the filtrate add 0.5 mL of *sulfuric acid R*. Evaporate to dryness and ignite the residue to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5 mg.

Loss on drying (*2.2.32*)
15.0 per cent to 20.0 per cent, determined on 0.500 g by drying in an oven at 125 °C.

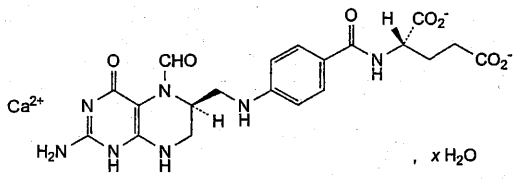
ASSAY
Dissolve a quantity equivalent to 0.200 g of the dried substance in *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 21.82 mg of $C_6H_{10}CaO_6$.

Ph Eur

Calcium Levofolinate Hydrate

Calcium Levofolinate Pentahydrate
(Ph. Eur. monograph 1606)



$C_{20}H_{21}CaN_7O_7,xH_2O$ 511.5
(anhydrous substance)

Action and use
Antidote to folic acid antagonists.

Ph Eur

DEFINITION
Calcium (2*S*)-2-[4-[[[(6*S*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl]amino]benzamido] pentanedioate hydrate.

Content
— *calcium levofolinate* ($C_{20}H_{21}CaN_7O_7$; M_r 511.5): 97.0 per cent to 102.0 per cent (anhydrous substance);
— *calcium* (Ca; A_r 40.08): 7.54 per cent to 8.14 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS
Appearance
White or light yellow, amorphous or crystalline, hygroscopic powder.

Solubility
Slightly soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A, B, D.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison: calcium folinate CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum quantity of *water R* and add dropwise sufficient *acetone R* to produce a precipitate. Allow to stand for 15 min, collect the precipitate by centrifugation, wash the precipitate twice with a minimum quantity of *acetone R* and dry. Record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 15 mg of the substance to be examined in a 3 per cent *V/V* solution of *ammonia R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 15 mg of *calcium folinate CRS* in a 3 per cent *V/V* solution of *ammonia R* and dilute to 5 mL with the same solvent.

Plate *cellulose for chromatography F_{254} R* as the coating substance.

Mobile phase The lower layer of a mixture of 1 volume of *isoamyl alcohol R* and 10 volumes of a 50 g/L solution of *citric acid monohydrate R* previously adjusted to pH 8 with *ammonia R*.

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (b) of calcium (*2.3.1*).

TESTS
Carry out the tests and the assay as rapidly as possible, protected from actinic light.

Solution S
Dissolve 0.40 g in *carbon dioxide-free water R*, heating at 40 °C if necessary, and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

pH (*2.2.3*)
7.5 to 8.5 for solution S.

Specific optical rotation (*2.2.7*)
−15.0 to −10.0 (anhydrous substance), measured at 25 °C.

Dissolve 0.200 g in *tris(hydroxymethyl)aminomethane solution R* previously adjusted to pH 8.1 with *sodium hydroxide solution R* or *hydrochloric acid R1* and dilute to 20.0 mL with the same solvent.

Absorbance (*2.2.25*)
Maximum 0.25, determined at 420 nm on solution S.

Ethanol

Head-space gas chromatography (*2.2.28*): use the standard additions method.

Test solution Dissolve 0.25 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution Dilute 0.750 g of *anhydrous ethanol R* to 1000.0 mL with *water R*.

Column:
— *material*: fused silica;
— *size*: *l* = 10 m, Ø = 0.32 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R*.

Carrier gas *nitrogen for chromatography R*.

Flow rate 4 mL/min.

Static head-space conditions that may be used:
— *equilibration temperature*: 80 °C;
— *equilibration time*: 20 min;
— *pressurisation time*: 30 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 14	80 → 220
Injection port		110
Detector		270

Detection Flame ionisation.

Injection At least 3 times.

Limit:
— *ethanol*: maximum 3.0 per cent.

Related substances

Liquid chromatography (*2.2.29*).

Prepare the solutions immediately before use.

Test solution Dissolve 10.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *calcium folinate CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (c) Dissolve 5 mg of *calcium folinate for system suitability CRS* (containing impurities A, E, F and I) in 5 mL of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 165 mL of *methanol R* and 835 mL of a solution containing 4.0 mL of *tetrabutylammonium dihydrogen phosphate solution R* and 1.42 g of *disodium hydrogen phosphate dihydrate R* in *water for chromatography R*, previously adjusted to pH 7.7 with *phosphoric acid R* or *dilute sodium hydroxide solution R*.

Flow rate 1.25 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 4 times the retention time of folinic acid.

Identification of impurities Use the chromatogram supplied with *calcium folinate for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, E, F and I.

Relative retention With reference to folinic acid (retention time = about 13.9 min): impurity E = about 0.4; impurity A = about 0.6; impurity F = about 0.7; impurity I = about 1.2.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to impurities A and F.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity E = 0.6; impurity F = 0.6; impurity I = 0.6;
— for each impurity, use the concentration of calcium levofolinate hydrate in reference solution (b).

Limits:
— *impurities A, E*: for each impurity, maximum 0.3 per cent;
— *impurities F, I*: for each impurity, maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.20 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Impurity H

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *calcium folinate CRS* in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *water R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4 mm;
— *stationary phase*: *human albumin coated silica gel for chiral separation R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Dissolve 9.72 g of *sodium dihydrogen phosphate R* in 890 mL of *water for chromatography R* and adjust to pH 5.0 with *sodium hydroxide solution R*; add 100 mL of *2-propanol R* and 10 mL of *acetonitrile R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 286 nm.

Injection 10 µL.

Retention times Levofolinic acid = about 9 min; impurity H = about 19 min.

System suitability:
— *resolution*: minimum 5.0 between the peaks due to levofolinic acid and impurity H in the chromatogram obtained with reference solution (a). The sum of the areas of the 2 peaks is 100 per cent. The peak area of impurity H is 48 per cent to 52 per cent. In the chromatogram obtained with reference solution (b) 2 clearly visible peaks are obtained.

Limit:
— *impurity H*: maximum 0.5 per cent.

Chlorides
Maximum 0.5 per cent.

Dissolve 0.300 g in 50 mL of *water R* heating at 40 °C if necessary. Add 10 mL of *2 M nitric acid* and titrate with *0.005 M silver nitrate* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.005 M silver nitrate* is equivalent to 0.177 mg of Cl.

Water
(*2.5.12*): 10.0 per cent to 17.0 per cent.

Dissolve 0.100 g in a mixture of 15 mL of *formamide R* and 50 mL of the titration solvent. Stir for about 6 min before titrating and use a suitable titrant that does not contain pyridine.

ASSAY

Carry out the assays as rapidly as possible, protected from actinic light.

Calcium
Dissolve 0.400 g in 150 mL of *water R* and dilute to 300 mL with the same solvent. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 4.008 mg of Ca.

Calcium folinate
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{20}H_{21}CaN_7O_7$ taking into account the assigned content of *calcium folinate CRS*.

STORAGE

In an airtight container, protected from light. If the substance is sterile, the container is also sterile and tamper-proof.

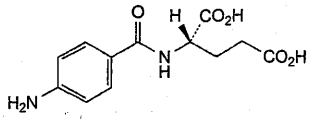
LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

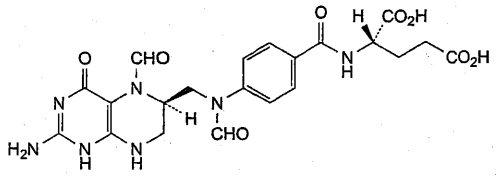
IMPURITIES

Specified impurities *A, E, F, H, I.*

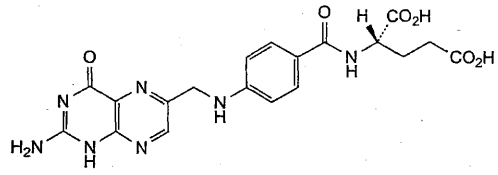
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, G.*



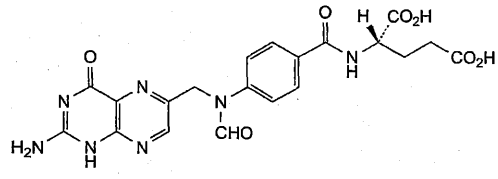
A. (2*S*)-2-(4-aminobenzamido)pentanedioic acid,



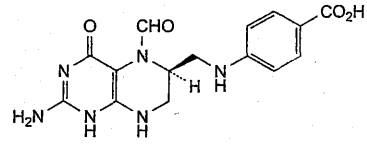
B. (2*S*)-2-[4-[[[(6*R*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl](formyl)amino]benzamido]pentanedioic acid (5,10-diformyltetrahydrolevofolic acid),



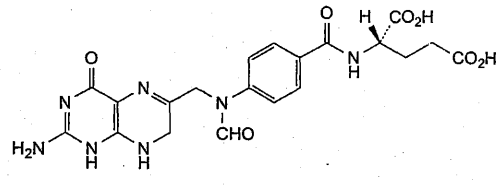
C. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid (folic acid),



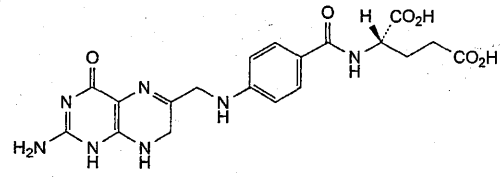
D. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl](formyl)amino]benzamido]pentanedioic acid (10-formylfolic acid),



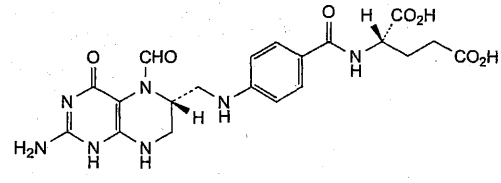
E. 4-[[[(6*S*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl]amino]benzoic acid ((6*S*)-5-formyltetrahydropteroic acid),



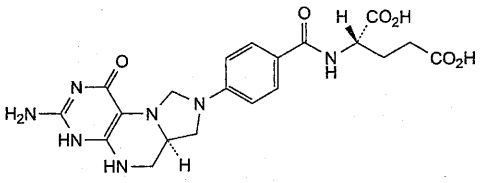
F. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4,7,8-tetrahydropteridin-6-yl)methyl](formyl)amino]benzamido]pentanedioic acid (10-formyldihydrofolic acid),



G. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4,7,8-tetrahydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid (dihydrofolic acid),



H. (2*S*)-2-[4-[[[(6*R*)-2-amino-5-formyl-4-oxo-1,4,5,6,7,8-hexahydropteridin-6-yl]methyl]amino]benzamido]pentanedioic acid (dextrofolinic acid),

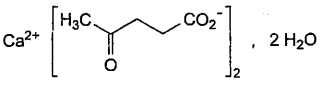


I. (2*S*)-2-[4-[(6a*R*)-3-amino-1-oxo-1,2,5,6,6a,7-hexahydroimidazo[1,5-*f*]pteridin-8(9*H*)-yl]benzamido] pentanedioic acid ((6a*R*)-5,10-methylenetetrahydrofolic acid).

Ph Eur

Calcium Levulinate Dihydrate

(Ph. Eur. monograph 1296)



$C_{10}H_{14}CaO_6,2H_2O$ 306.3 *5743-49-7*

Action and use
Source of calcium.

Ph Eur

DEFINITION
Calcium di(4-oxopentanoate) dihydrate.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION
First identification: A, D, E.
Second identification: B, C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *calcium levulinate dihydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 60 mg of the substance to be examined in *water R* and dilute to 1 mL with the same solvent.
Reference solution Dissolve 60 mg of *calcium levulinate dihydrate CRS* in *water R* and dilute to 1 mL with the same solvent.
Plate *TLC silica gel plate R.*
Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).
Application 10 µL.
Development Over a path of 10 cm.
Drying At 100-105 °C for 20 min and allow to cool.
Detection Spray with a 30 g/L solution of *potassium permanganate R*. Dry in a current of warm air for about 5 min or until the spots become yellow. Examine in daylight.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 1 mL of solution S (see Tests), add 20 mL of a 2.5 g/L solution of *dinitrophenylhydrazine R* in *dilute hydrochloric acid R*. Allow to stand for 15 min. Filter, wash the precipitate with *water R*. Dry the precipitate in an oven at 100-105 °C. The melting point (*2.2.14*) is 203 °C to 210 °C.

D. It gives reaction (b) of calcium (*2.3.1*).

E. Loss on drying (see Tests).

TESTS

Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
6.8 to 7.8 for solution S.

Oxidisable substances
To 1 mL of solution S, add 10 mL of *water R*, 1 mL of *dilute sulfuric acid R* and 0.25 mL of a 3.0 g/L solution of *potassium permanganate R*. Mix. After 5 min, the violet colour of the mixture is still visible.

Sucrose and reducing sugars
To 5 mL of solution S add 2 mL of *hydrochloric acid R1* and dilute to 10 mL with *water R*. Heat to boiling for 5 min and allow to cool. Add 10 mL of *sodium carbonate solution R*. Allow to stand for 5 min, dilute to 25 mL with *water R* and filter. To 5 mL of the filtrate add 2 mL of *cupri-tartaric solution R* and heat to boiling for 1 min. No red precipitate is formed.

Chlorides (*2.4.4*)
Maximum 50 ppm.
Dilute 10 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 200 ppm.
Dilute 7.5 mL of solution S to 15 mL with *distilled water R*.

Magnesium and alkali metals
Maximum 1.0 per cent.
To 10 mL of solution S, add 80 mL of *water R*, 10 mL of *ammonium chloride solution R* and 1 mL of *ammonia R*. Heat to boiling. To the boiling solution, add dropwise 50 mL of warm *ammonium oxalate solution R*. Allow to stand for 4 h, then dilute to 200 mL with *water R* and filter. To 100 mL of the filtrate, add 0.5 mL of *sulfuric acid R*. Evaporate to dryness on a water-bath and ignite to constant mass at 600 ± 50 °C. The residue weighs a maximum of 5.0 mg.

Loss on drying (*2.2.32*)
11.0 per cent to 12.5 per cent, determined on 0.200 g by drying at 105 °C.

Pyrogens (*2.6.8*)
If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of pyrogens, it complies with the test for pyrogens. Inject per kilogram of the rabbit's mass 4 mL of a solution containing per millilitre 50 mg of the substance to be examined.

ASSAY
Dissolve 0.240 g in 50 mL of *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

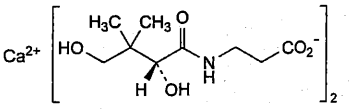
1 mL of *0.1 M sodium edetate* is equivalent to 27.03 mg of $C_{10}H_{14}CaO_6$.

STORAGE
Protected from light.

Ph Eur

Calcium Pantothenate

(Ph. Eur. monograph 0470)



$C_{18}H_{32}CaN_2O_{10}$ 476.5 *137-08-6*

Action and use
Component of vitamin B.

Ph Eur

DEFINITION
Calcium bis[3-[(2*R*)-2,4-dihydroxy-3,3-dimethylbutanamido] propanoate].

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder, slightly hygroscopic.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

C. To 1 mL of solution S (see Tests) add 1 mL of *dilute sodium hydroxide solution R* and 0.1 mL of *copper sulfate solution R*. A blue colour develops.

D. It gives reaction (a) of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
6.8 to 8.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 25.5 to + 27.5 (dried substance), determined on solution S.

Impurity A and other aminocarboxylic acid impurities
Maximum 0.50 per cent.

Dissolve 8.000 g in 40 mL of *water R* and dilute to 100.0 mL with the same solvent. Add 25 mL of *formaldehyde solution R* and titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 8.91 mg of $C_3H_7NO_2$.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.600 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in *water R* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 20.0 mg of *calcium pantothenate CRS* in *water R* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (c) Dissolve 3.0 mg of *pantolactone CRS* (impurity C) in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (d) Mix 1 mL of reference solution (a) and 1.0 mL of reference solution (c) and dilute to 10.0 mL with *water R*.

Reference solution (e) Dissolve 3 mg of *pantothenate for peak identification CRS* (containing impurities B, E and H) in 0.5 mL of *water R*.

Column:
— *size*: *l* = 0.15 m, Ø = 3.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 35 °C.

Mobile phase *acetonitrile R1*, 1.56 g/L solution of *sodium dihydrogen phosphate R* in *water for chromatography R*, adjusted to pH 2.5 with *phosphoric acid R* (1:99 *V/V*).

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 5 µL of test solution (a) and reference solutions (b), (c), (d) and (e).

Run time 7 times the retention time of calcium pantothenate.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C; use the chromatogram supplied with *pantothenate for peak identification CRS* and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities B, E and H.

Relative retention With reference to calcium pantothenate (retention time = about 4 min): impurity B = about 0.5; impurity C = about 0.8; impurity E = about 1.7; impurity H = about 3.5.

System suitability Reference solution (d):
— *resolution*: minimum 3.0 between the peaks due to impurity C and calcium pantothenate;
— *signal-to-noise ratio*: minimum 10 for the peak due to impurity C.

Calculation of percentage contents:
— for impurities B and C, use the concentration of impurity C in reference solution (c);
— for impurities other than B and C, use the concentration of calcium pantothenate in reference solution (b).

Limits:
— *impurity B*: maximum 0.8 per cent;
— *impurity C*: maximum 0.5 per cent;
— *impurity E*: maximum 0.25 per cent;
— *impurity H*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent; disregard any peak due to impurity A eluting before 1 min.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

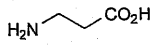
ASSAY
Dissolve 0.180 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 23.83 mg of $C_{18}H_{32}CaN_2O_{10}$.

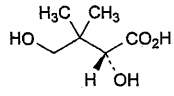
STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B, C, E, H.*

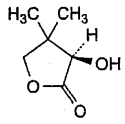
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, F, G.*



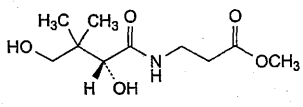
A. 3-aminopropanoic acid (β-alanine),



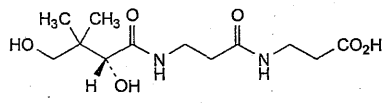
B. (2*R*)-2,4-dihydroxy-3,3-dimethylbutanoic acid (pantoic acid),



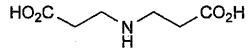
C. (3*R*)-3-hydroxy-4,4-dimethyldihydrofuran-2(3*H*)-one (pantolactone),



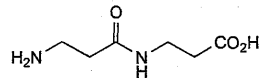
D. methyl 3-[(2*R*)-2,4-dihydroxy-3,3-dimethylbutanamido] propanoate (methyl pantothenate),



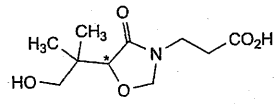
E. 3-[3-[(2*R*)-2,4-dihydroxy-3,3-dimethylbutanamido]propanamido]propanoic acid (β-alanyl pantothenamide),



F. 3,3′-azanediyldipropanoic acid,



G. 3-(3-aminopropanamido)propanoic acid (β-alanyl-β-alanine),



H. 3-[(5Ξ)-5-(1-hydroxy-2-methylpropan-2-yl)-4-oxo-1,3-oxazolidin-3-yl]propanoic acid.

Ph Eur

Calcium Phosphate

Tribasic Calcium Phosphate
(Ph. Eur. monograph 1052)

Action and use
Excipient.

Preparations
Calcium and Ergocalciferol Tablets
Calcium Phosphate for Homoeopathic Preparations
Calcium and Ergocalciferol Chewable Tablets

Ph Eur

DEFINITION
Mixture of calcium phosphates.

Content
35.0 per cent to 40.0 per cent of Ca (A_r 40.08).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water. It dissolves in dilute hydrochloric acid and in dilute nitric acid.

IDENTIFICATION
A. Dissolve 0.1 g in 5 mL of a 25 per cent *V/V* solution of *nitric acid R*. The solution gives reaction (b) of phosphates (*2.3.1*).

B. It gives reaction (b) of calcium (*2.3.1*). Filter before adding *potassium ferrocyanide solution R*.

C. It complies with the limits of the assay.

TESTS
Solution S
Dissolve 2.50 g in 20 mL of *dilute hydrochloric acid R*. If the solution is not clear, filter it. Add *dilute ammonia R1* dropwise until a precipitate is formed. Dissolve the precipitate by adding *dilute hydrochloric acid R* and dilute to 50 mL with *distilled water R*.

Chlorides (*2.4.4*)
Maximum 0.15 per cent.

Dissolve 0.22 g in a mixture of 1 mL of *nitric acid R* and 10 mL of *water R* and dilute to 100 mL with *water R*.

Fluorides
Maximum 75 ppm.

Potentiometry (*2.2.36, Method II*).

Test solution Dissolve 0.250 g in *0.1 M hydrochloric acid*, add 5.0 mL of *fluoride standard solution (1 ppm F) R* and dilute to 50.0 mL with *0.1 M hydrochloric acid*. To 20.0 mL of this solution add 20.0 mL of *total-ionic-strength-adjustment buffer R* and 3 mL of an 82 g/L solution of *anhydrous sodium acetate R*. Adjust to pH 5.2 with *ammonia R* and dilute to 50.0 mL with *distilled water R*.

Reference solution *Fluoride standard solution (10 ppm F) R*.

Indicator electrode Fluoride-selective.

Reference electrode Silver-silver chloride.

Carry out the measurements on the test solution, then add at least 3 quantities, each of 0.5 mL, of the reference solution, carrying out a measurement after each addition. Calculate the concentration of fluorides using the calibration curve, taking into account the addition of fluoride to the test solution.

Sulfates (*2.4.13*)
Maximum 0.5 per cent.

Dilute 1 mL of solution S to 25 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 4 ppm, determined on 5 mL of solution S.

Iron (*2.4.9*)
Maximum 400 ppm.

Dilute 0.5 mL of solution S to 10 mL with *water R*.

Acid-insoluble matter
Maximum 0.2 per cent.

Dissolve 5.0 g in a mixture of 10 mL of *hydrochloric acid R* and 30 mL of *water R*. Filter, wash the residue with *water R* and dry to constant mass at 100-105 °C. The residue weighs a maximum of 10 mg.

Loss on ignition
Maximum 8.0 per cent, determined on 1.000 g by ignition at 800 ± 50 °C for 30 min.

ASSAY
Dissolve 0.200 g in a mixture of 1 mL of *hydrochloric acid R1* and 5 mL of *water R*. Add 25.0 mL of *0.1 M sodium edetate* and dilute to 200 mL with *water R*. Adjust to about pH 10 with *concentrated ammonia R*. Add 10 mL of *ammonium chloride buffer solution pH 10.0 R* and a few milligrams of *mordant black 11 triturate R*. Titrate the excess sodium edetate with *0.1 M zinc sulfate* until the colour changes from blue to violet.

1 mL of *0.1 M sodium edetate* is equivalent to 4.008 mg of Ca.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium phosphate is used as a filler in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

Bulk and tapped density (*2.9.34*)

Powder flow (*2.9.36*)

Ph Eur

Calcium Polystyrene Sulfonate

Calcium Polystyrene Sulphonate

Action and use
Used in the treatment of hyperkalaemia.

DEFINITION
Calcium Polystyrene Sulfonate is a cation-exchange resin prepared in the calcium form containing not less than 6.5% w/w and not more than 9.5% w/w of calcium, calculated with reference to the dried substance. Each g exchanges not less than 1.3 mEq and not more than 2.0 mEq of potassium, calculated with reference to the dried substance.

CHARACTERISTICS
A cream to light brown, fine powder.

Practically insoluble in *water* and in *ethanol (96%)*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of calcium polystyrene sulfonate *(RS 037)*.

B. Yields reaction C characteristic of *calcium salts*, Appendix VI.

TESTS
Particle size
Not more than 1% w/w is retained on a 150-µm sieve, Appendix XVII B. Use 20 g and sieve for 5 minutes.

Potassium
Not more than 0.1% of K when determined by *atomic emission spectrophotometry*, Appendix II D, measuring at 766.5 nm and using a solution prepared in the following manner. To 1.1 g of the substance being examined add 5 mL of *hydrochloric acid*, heat to boiling, cool and add 10 mL of *water*. Filter, wash the filter and residue with *water* and dilute the filtrate and washings to 25 mL with *water*. Use *potassium standard solution (100 ppm K)*, suitably diluted with *water*, to prepare the standard solutions.

Sodium
Not more than 0.1% of Na when determined by *atomic emission spectrophotometry*, Appendix II D, measuring at 589.0 nm and using a solution prepared in the following manner. To 1.1 g of the substance being examined add 5 mL of *hydrochloric acid*, heat to boiling, cool and add 10 mL of *water*. Filter, wash the filter and residue with *water* and dilute the filtrate and washings to 25 mL with *water*. Use *sodium solution (200 ppm Na)*, suitably diluted with *water*, to prepare the standard solutions.

Arsenic
1 g dispersed in 25 mL of *water* complies with the *limit test for arsenic*, Appendix VII (1 ppm).

Styrene
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) Shake 10 g of the substance being examined with 10 mL of *acetone* for 30 minutes, centrifuge and use the supernatant liquid.

(2) 0.0001% w/v of *styrene* in *acetone*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (30 cm × 4 mm) packed with *octadecylsilyl silica gel for chromatography* (μBondapak C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 μL of each solution.

MOBILE PHASE

Equal volumes of *acetonitrile* and *water*.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to styrene is not greater than the area of the peak in the chromatogram obtained with solution (2) (1 ppm).

Potassium exchange capacity
To 3 g of the substance being examined in a dry 250 mL glass-stoppered flask add 100 mL of a solution containing 0.7455% w/v of *potassium chloride* and 0.4401% w/v of *potassium hydrogen carbonate* in *water* (solution A), stopper and shake for 15 minutes. Filter and dilute 2 mL of the filtrate to 1000 mL with *water*. Determine the concentration of unbound potassium in this solution by *atomic emission spectrophotometry*, Appendix II D, measuring at 766.5 nm and using solution A suitably diluted with *water*, to prepare the standard solutions. Calculate the potassium exchange capacity of the substance being examined in milliequivalents taking the concentration of potassium in solution A as 144 milliequivalents of K per litre.

Loss on drying
When dried at 70° at a pressure not exceeding 0.7 kPa for 16 hours, loses not more than 8.0% of its weight. Use 2 g.

Microbial contamination
Carry out a quantitative evaluation for Enterobacteria and certain other Gram-negative bacteria, Appendix XVI B1. 0.01 g of the substance being examined gives a negative result, Table I (most probable number of bacteria per gram fewer than 10^2).

ASSAY

For calcium
Carefully heat 1 g in a platinum crucible until a white ash is obtained and dissolve in 10 mL of *2M hydrochloric acid* with the aid of heat. Transfer the resulting solution to a conical flask using 20 mL of *water*. Add 50 mL of *0.05M disodium edetate VS*, 20 mL of *ammonia buffer pH 10.9* and titrate the excess of disodium edetate with *0.02M zinc sulfate VS*, using a 0.5% w/v solution of *mordant black 11* in *ethanol (96%)* as indicator to a red purple end point. Each mL of *0.05M disodium edetate VS* is equivalent to 2.004 mg of Ca.

STORAGE

Calcium Polystyrene Sulfonate should be kept in an airtight container.

Calcium Stearate

(Ph. Eur. monograph 0882)

1592-23-0

Action and use
Excipient.

Ph Eur

DEFINITION

Mixture of calcium salts of different fatty acids consisting mainly of stearic (octadecanoic) acid [$(C_{17}H_{35}COO)_2Ca$; M_r 607] and palmitic (hexadecanoic) acid [$(C_{15}H_{31}COO)_2Ca$; M_r 550.9] with minor proportions of other fatty acids.

Content
- *calcium*: 6.4 per cent to 7.4 per cent (A_r 40.08) (dried substance);
- *stearic acid in the fatty acid fraction*: minimum 40.0 per cent;
- *sum of stearic acid and palmitic acid in the fatty acid fraction*: minimum 90.0 per cent.

CHARACTERS

Appearance
Fine, white or almost white, crystalline powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification: C, D.

Second identification: A, B, D.

A. Freezing point (*2.2.18*): minimum 53 °C, for the residue obtained in the preparation of solution S (see Tests).

B. Acid value (*2.5.1*): 195 to 210.

Dissolve 0.200 g of the residue obtained in the preparation of solution S in 25 mL of the prescribed mixture of solvents.

C. Examine the chromatograms obtained in the test for fatty acid composition.

Results The retention times of the principal peaks in the chromatogram obtained with the test solution are approximately the same as those of the principal peaks in the chromatogram obtained with the reference solution.

D. Neutralise 5 mL of solution S to *red litmus paper R* using *strong sodium hydroxide solution R*. The solution gives reaction (b) of calcium (*2.3.1*).

TESTS

Solution S
To 5.0 g add 50 mL of *peroxide-free ether R*, 20 mL of *dilute nitric acid R* and 20 mL of *distilled water R*. Boil under a reflux condenser until dissolution is complete. Allow to cool. In a separating funnel, separate the aqueous layer and shake the ether layer with 2 quantities, each of 5 mL, of *distilled water R*. Combine the aqueous layers, wash with 15 mL of *peroxide-free ether R* and dilute the aqueous layer to 50 mL with *distilled water R* (solution S). Evaporate the ether layer to dryness and dry the residue at 100-105 °C. Keep the residue for identification tests A and B.

Acidity or alkalinity
To 1.0 g add 20 mL of *carbon dioxide-free water R* and boil for 1 min with continuous shaking. Cool and filter.
To 10 mL of the filtrate add 0.05 mL of *bromothymol blue solution R1*. Not more than 0.5 mL of *0.01 M hydrochloric*

acid or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Chlorides (*2.4.4*)
Maximum 0.1 per cent.

Dilute 0.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 0.3 per cent.

Dilute 0.5 mL of solution S to 15 mL with *distilled water R*.

Cadmium
Maximum 3 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Place 50.0 mg in a polytetrafluoroethylene digestion bomb and add 0.5 mL of a mixture of 1 volume of *hydrochloric acid R* and 5 volumes of *cadmium- and lead-free nitric acid R*. Allow to digest at 170 °C for 5 h. Allow to cool. Dissolve the residue in *water R* and dilute to 5.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using *cadmium standard solution (10 ppm Cd) R*, diluted if necessary with a 1 per cent *V/V* solution of *hydrochloric acid R*.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Graphite furnace.

Lead
Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Use the solution described in the test for cadmium.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R*, diluted if necessary with *water R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm; 217.0 nm may be used depending on the apparatus.

Atomisation device Graphite furnace.

Nickel
Maximum 5 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Use the solution described in the test for cadmium.

Reference solutions Prepare the reference solutions using *nickel standard solution (10 ppm Ni) R*, diluted if necessary with *water R*.

Source Nickel hollow-cathode lamp.

Wavelength 232.0 nm.

Atomisation device Graphite furnace.

Loss on drying (*2.2.32*)
Maximum 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

ASSAY

Calcium
To 0.500 g in a 250 mL conical flask add 50 mL of a mixture of equal volumes of *anhydrous ethanol R* and *butanol R*, 5 mL of *concentrated ammonia R*, 3 mL of *ammonium chloride buffer solution pH 10.0 R*, 30.0 mL of *0.1 M sodium edetate* and 15 mg of *mordant black 11 triturate R*. Heat to 45-50 °C until the solution is clear. Cool and titrate with *0.1 M zinc sulfate* until the colour changes from blue to violet. Carry out a blank titration.

1 mL of *0.1 M sodium edetate* is equivalent to 4.008 mg of Ca.

Composition of fatty acids
Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution In a conical flask fitted with a reflux condenser, dissolve 0.10 g of the substance to be examined in 5 mL of *boron trifluoride-methanol solution R*. Boil under a reflux condenser for 10 min. Add 4 mL of *heptane R* through the condenser. Boil under a reflux condenser for 10 min. Allow to cool. Add 20 mL of *saturated sodium chloride solution R*. Shake and allow the layers to separate. Remove about 2 mL of the organic layer and dry over 0.2 g of *anhydrous sodium sulfate R*. Dilute 1.0 mL of the solution to 10.0 mL with *heptane R*.

Reference solution Prepare the reference solution in the same manner as the test solution using 50.0 mg of *palmitic acid CRS* and 50.0 mg of *stearic acid CRS* instead of calcium stearate.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R*.

Flow rate 2.4 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	70
	2 - 36	70 → 240
	36 - 41	240
Injection port		220
Detector		260

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to methyl stearate: methyl palmitate = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 5.0 between the peaks due to methyl palmitate and methyl stearate.

Calculate the content of palmitic acid and stearic acid. Disregard the peak due to the solvent.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium stearate used as a lubricant in tablets and capsules.

Particle-size distribution (*2.9.31*)

Specific surface area (*2.9.26, Method I*)
Determine the specific surface area in the P/P_o range of 0.05 to 0.15.

Sample outgassing 2 h at 40 °C.

Ph Eur

Dried Calcium Sulfate

Exsiccated Calcium Sulfate; Plaster of Paris; Dried Calcium Sulphate

$CaSO_4,½H_2O$ 145.1 *26499-65-0*

DEFINITION
Dried Calcium Sulfate is prepared by heating powdered gypsum, $CaSO_4,2H_2O$, at about 150° in a controlled manner such that it is substantially converted into the hemihydrate, $CaSO_4,½H_2O$, with minimum production of the anhydrous phases of calcium sulfate. It may contain suitable setting accelerators or decelerators.

CHARACTERISTICS
A white or almost white powder; hygroscopic.

Slightly soluble in *water*; more soluble in dilute mineral acids; practically insoluble in *ethanol (96%)*.

IDENTIFICATION
Yields the reactions characteristic of *calcium salts* and of *sulfates*, Appendix VI.

TESTS
Setting properties
20 g mixed with 10 mL of *water* at 15° to 20° in a cylindrical mould about 2.4 cm in diameter sets in 4 to 11 minutes. The mass thus produced, after standing for 3 hours, possesses sufficient hardness to resist pressure of the fingers at the edges, which retain their sharpness of outline and do not crumble.

Loss on ignition
When ignited to constant weight at red heat, loses 4.5% to 8.0% of its weight. Use 1 g.

Calcium Sulfate Dihydrate

Calcium Sulphate

(Ph. Eur. monograph 0982)

$CaSO_4,2H_2O$ 172.2 *10101-41-4*

Action and use
Excipient.

Ph Eur

DEFINITION
Content
98.0 per cent to 102.0 per cent of $CaSO_4,2H_2O$.

CHARACTERS
Appearance
White or almost white fine powder.

Solubility
Very slightly soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Loss on ignition (see Tests).

B. Solution S (see Tests) gives reaction (a) of sulfates (*2.3.1*).

C. Solution S gives reaction (a) of calcium (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in 50 mL of a 10 per cent *V/V* solution of *hydrochloric acid R* by heating at 50 °C for 5 min. Allow to cool.

Acidity or alkalinity
Shake 1.5 g with 15 mL of *carbon dioxide-free water R* for 5 min. Allow to stand for 5 min and filter. To 10 mL of the filtrate add 0.1 mL of *phenolphthalein solution R* and 0.25 mL of *0.01 M sodium hydroxide*. The solution is red.
Add 0.30 mL of *0.01 M hydrochloric acid*. The solution is colourless. Add 0.2 mL of *methyl red solution R*. The solution is reddish-orange.

Chlorides (*2.4.4*)
Maximum 300 ppm.

Shake 0.5 g with 15 mL of *water R* for 5 min. Allow to stand for 15 min and filter. Dilute 5 mL of the filtrate to 15 mL with *water R*.

Arsenic (*2.4.2, Method A*)
Maximum 10 ppm, determined on 5 mL of solution S.

Iron (*2.4.9*)
Maximum 100 ppm.

To 0.25 g add a mixture of 5 mL of *hydrochloric acid R* and 20 mL of *water R*. Heat to boiling, cool and filter.

Loss on ignition
18.0 per cent to 22.0 per cent, determined on 1.000 g by ignition to constant mass at 800 ± 50 °C.

ASSAY
Dissolve 0.150 g in 120 mL of *water R*. Carry out the complexometric titration of calcium (*2.5.11*).

1 mL of *0.1 M sodium edetate* is equivalent to 17.22 mg of $CaSO_4,2H_2O$.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for calcium sulfate dihydrate used as filler in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

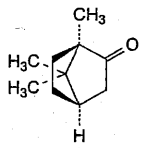
Bulk and tapped density (*2.9.34*)

Powder flow (*2.9.36*)

Ph Eur

Natural Camphor

(D-Camphor, Ph. Eur. monograph 1400)



$C_{10}H_{16}O$ 152.2 *464-49-3*

Ph Eur

DEFINITION

(1*R*,4*R*)-1,7,7-Trimethylbicyclo[2.2.1]heptan-2-one.

CHARACTERS

Appearance

White or almost white, crystalline powder or friable, crystalline masses.

Highly volatile even at room temperature.

Solubility

Slightly soluble in water, very soluble in alcohol and in light petroleum, freely soluble in fatty oils, very slightly soluble in glycerol.

IDENTIFICATION

First identification: A, C.

Second identification: A, B, D.

A. Specific optical rotation (see Tests).

B. Melting point (*2.2.14*): 175 °C to 179 °C.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *racemic camphor CRS.*

D. Dissolve 1.0 g in 30 mL of *methanol R*. Add 1.0 g of *hydroxylamine hydrochloride R* and 1.0 g of *anhydrous sodium acetate R*. Boil under a reflux condenser for 2 h. Allow to cool and add 100 mL of *water R*. Filter, wash the precipitate obtained with 10 mL of *water R* and recrystallise from 10 mL of a mixture of 4 volumes of *alcohol R* and 6 volumes of *water R*. The crystals, dried *in vacuo*, melt (*2.2.14*) at 118 °C to 121 °C.

TESTS

Carry out the weighings and dissolution rapidly.

Solution S

Dissolve 2.50 g in 10 mL of *alcohol R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R1*. The solution is colourless. Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Specific optical rotation (*2.2.7*)

+ 41.0 to + 44.0, determined on solution S.

Related substances

Gas chromatography (*2.2.28*).

Test solution Dissolve 2.50 g of the substance to be examined in *heptane R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *heptane R*.

Reference solution (b) Dilute 10.0 mL of reference solution (a) to 20.0 mL with *heptane R*.

Reference solution (c) Dissolve 0.50 g of *borneol R* in *heptane R* and dilute to 25.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *heptane R*.

Reference solution (d) Dissolve 50 mg of *linalol R* and 50 mg of *bornyl acetate R* in *heptane R* and dilute to 100.0 mL with the same solvent.

Column:

— *size*: *l* = 30 m, Ø = 0.25 mm,

— *stationary phase*: *macrogol 20 000 R* (0.25 µm).

Carrier gas *helium for chromatography R.*

Split ratio 1:70.

Flow rate 45 cm/s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 10	50
	10 - 35	50 → 100
	35 - 45	100 → 200
	45 - 55	200
Injection port		220
Detector		250

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution (d).

— *resolution*: minimum 3.0 between the peaks due to bornyl acetate and to linalol.

Limits:

— *borneol*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent),

— *any other impurity*: not more than half of the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),

— *total of other impurities*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (4.0 per cent),

— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Halogens

Maximum 100 ppm.

Dissolve 1.0 g in 10 mL of *2-propanol R* in a distillation flask. Add 1.5 mL of *dilute sodium hydroxide solution R* and 50 mg of *nickel-aluminium alloy R*. Heat on a water-bath until the *2-propanol R* has evaporated. Allow to cool and add 5 mL of *water R*. Mix and filter through a wet filter previously washed with *water R* until free from chlorides. Dilute the filtrate to 10.0 mL with *water R*. To 5.0 mL of the solution, add *nitric acid R* dropwise until the precipitate which forms is redissolved and dilute to 15 mL with *water R*. The solution complies with the limit test for chlorides (*2.4.4*).

Residue on evaporation (*2.8.9*)

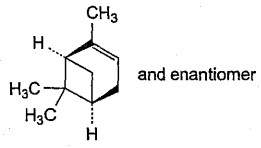
Maximum 0.05 per cent.

Evaporate 2.0 g on a water-bath and dry in an oven at 100-105 °C for 1 h. The residue weighs a maximum of 1 mg.

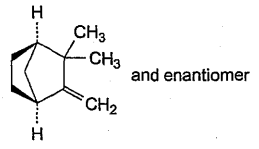
Water

Dissolve 1 g in 10 mL of *light petroleum R*. The solution is clear (*2.2.1*).

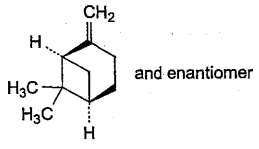
IMPURITIES



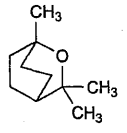
A. 2,6,6-trimethylbicyclo[3.1.1]hept-2-ene (α-pinene),



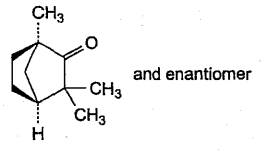
B. 2,2-dimethyl-3-methylenebicyclo[2.2.1]heptane (camphene),



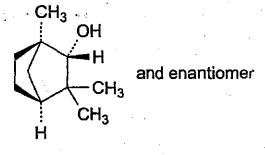
C. 6,6-dimethyl-2-methylenebicyclo[3.1.1]heptane (β-pinene),



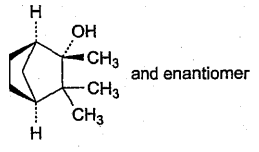
D. 1,3,3-trimethyl-2-oxabicyclo[2.2.2]octane (cineole),



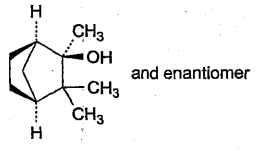
E. 1,3,3-trimethylbicyclo[2.2.1]heptan-2-one (fenchone),



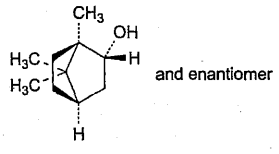
F. *exo*-1,3,3-trimethylbicyclo[2.2.1]heptan-2-ol (fenchol),



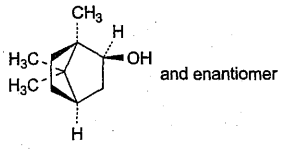
G. *exo*-2,3,3-trimethylbicyclo[2.2.1]heptan-2-ol (camphene hydrate),



H. *endo*-2,3,3-trimethylbicyclo[2.2.1]heptan-2-ol (methylcamphenilol),



I. *exo*-1,7,7-trimethylbicyclo[2.2.1]heptan-2-ol (*exo*-borneol),

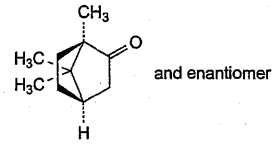


J. *endo*-1,7,7-trimethylbicyclo[2.2.1]heptan-2-ol (*endo*-borneol).

Ph Eur

Racemic Camphor

(Ph. Eur. monograph 0655)



$C_{10}H_{16}O$ 152.2 *76-22-2*

Action and use
Counter-irritant.

Preparations
Camphorated Opium Tincture
Concentrated Camphorated Opium Tincture
Concentrated Camphor Water

Ph Eur

DEFINITION
(1*RS*,4*RS*)-1,7,7-Trimethylbicyclo[2.2.1]heptan-2-one.

CHARACTERS

Appearance
White or almost white, crystalline powder or friable, crystalline masses, highly volatile even at room temperature.

Solubility
Slightly soluble in water, very soluble in ethanol (96 per cent) and in light petroleum, freely soluble in fatty oils, very slightly soluble in glycerol.

IDENTIFICATION
First identification: A, C.
Second identification: A, B, D.
A. Optical rotation (see Tests).
B. Melting point (*2.2.14*): 172 °C to 180 °C.
C. Infrared absorption spectrophotometry (*2.2.24*).
Preparation Mulls in *liquid paraffin R.*
Comparison *racemic camphor CRS.*
D. Dissolve 1.0 g in 30 mL of *methanol R.* Add 1.0 g of *hydroxylamine hydrochloride R* and 1.0 g of *anhydrous sodium acetate R.* Boil under a reflux condenser for 2 h. Allow to cool and add 100 mL of *water R.* A precipitate is formed. Filter, wash with 10 mL of *water R* and recrystallise from

10 mL of a mixture of 4 volumes of *ethanol (96 per cent) R* and 6 volumes of *water R*. The crystals, dried *in vacuo*, melt (*2.2.14*) at 118 °C to 121 °C.

TESTS

Carry out the weighings rapidly.

Solution S

Dissolve 2.50 g in 10 mL of *ethanol (96 per cent) R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

Dissolve 1.0 g in 10 mL of *ethanol (96 per cent) R* and add 0.1 mL of *phenolphthalein solution R1*. The solution is colourless. Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Optical rotation (*2.2.7*)

−0.15° to + 0.15°, determined on solution S.

Related substances

Gas chromatography (*2.2.28*).

Test solution Dissolve 50 mg of the substance to be examined in *hexane R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of the substance to be examined and 50 mg of *bornyl acetate R* in *hexane R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with *hexane R*.

Column:

— *size*: *l* = 2 m, Ø = 2 mm;

— *stationary phase*: diatomaceous earth for gas chromatography R impregnated with 10 per cent *m/m* of *macrogol 20 000 R*.

Carrier gas *nitrogen for chromatography R*.

Flow rate 30 mL/min.

Temperature:

— *column*: 130 °C;

— *injection port and detector*: 200 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 3 times the retention time of camphor.

System suitability:

— *resolution*: minimum 1.5 between the peaks due to camphor and bornyl acetate in the chromatogram obtained with reference solution (a);

— *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:

— *any impurity*: for each impurity, not more than 2 per cent of the area of the principal peak;

— *total*: not more than 4 per cent of the area of the principal peak;

— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b).

Halogens

Maximum 100 ppm.

Dissolve 1.0 g in 10 mL of *2-propanol R* in a distillation flask. Add 1.5 mL of *dilute sodium hydroxide solution R* and 50 mg of *nickel-aluminium alloy R*. Heat on a water-bath until the *2-propanol R* has evaporated. Allow to cool and add 5 mL of *water R*. Mix and filter through a wet filter previously washed with *water R* until free from chlorides. Dilute the filtrate to 10.0 mL with *water R*. To 5.0 mL of this solution, add *nitric acid R* dropwise until the precipitate which forms is redissolved and dilute to 15 mL with *water R*. The solution complies with the limit test for chlorides (*2.4.4*).

Water

Dissolve 1 g in 10 mL of *light petroleum R*. The solution is clear (*2.2.1*).

Residue on evaporation

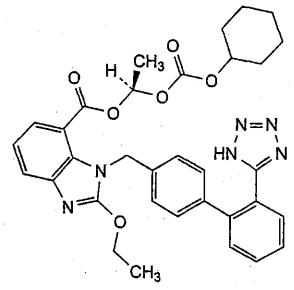
Maximum 0.05 per cent.

Evaporate 2.0 g on a water-bath and dry at 100-105 °C for 1 h. The residue weighs not more than 1 mg.

Ph Eur

Candesartan Cilexetil

(Ph. Eur. monograph 2573)



and enantiomer

$C_{33}H_{34}N_6O_6$ 611 *145040-37-5*

Action and use

Angiotensin.

Preparation

Candesartan Tablets

Ph Eur

DEFINITION

(1*RS*)-1-[[(Cyclohexyloxy)carbonyl]oxy]ethyl 2-ethoxy-1-[[2′-(1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride and slightly soluble in anhydrous ethanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *candesartan cilexetil CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *water R, acetonitrile R* (40:60 *V/V*).

Test solution Dissolve 20 mg of the substance to be examined in 50.0 mL of the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *candesartan cilexetil for system suitability CRS* (containing impurities A, B and F) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2.5 mg of *candesartan cilexetil for peak identification CRS* (containing impurities G and H) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (4 µm).

Mobile phase:
— *mobile phase A*: *glacial acetic acid R, water R, acetonitrile R* (1:43:57 *V/V/V*);
— *mobile phase B*: *glacial acetic acid R, water R, acetonitrile R* (1:10:90 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 33	100 → 0	0 → 100
33 - 40	0	100

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *candesartan cilexetil for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and F; use the chromatogram supplied with *candesartan cilexetil for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities G and H.

Relative retention With reference to candesartan cilexetil (retention time = about 11 min): impurity G = about 0.2; impurity A = about 0.4; impurity B = about 0.5; impurity F = about 2.0; impurity H = about 3.5.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to impurities A and B.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurities A and G = 0.7; impurity H = 1.6;
— *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurities A, H*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.32*)
Maximum 0.3 per cent, determined on 60.0 mg.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

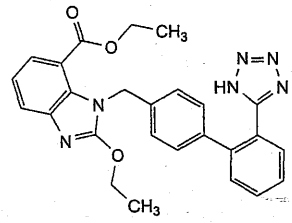
Dissolve 0.500 g in 60 mL of *glacial acetic acid R*. Titrate immediately with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*) at the 1st inflexion point.

1 mL of *0.1 M perchloric acid* is equivalent to 61.1 mg of $C_{33}H_{34}N_6O_6$.

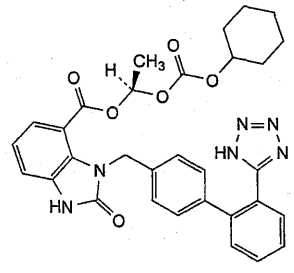
IMPURITIES

Specified impurities *A, B, F, G, H.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, I.*

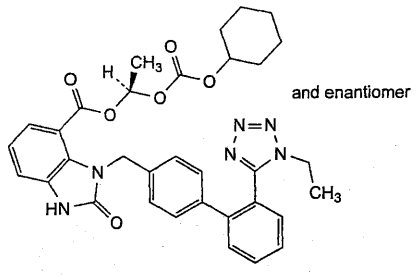


A. ethyl 2-ethoxy-1-[[2′-(1*H*-tetrazol-5-yl)biphenyl-4-yl] methyl]-1*H*-benzimidazole-7-carboxylate,

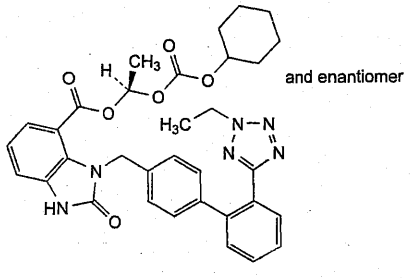


and enantiomer

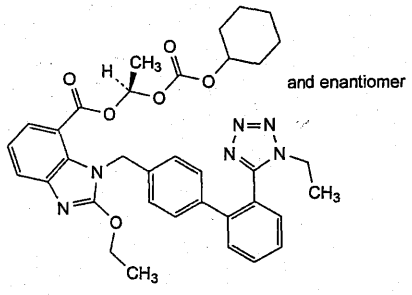
B. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 2-oxo-3-[[2′-(1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-2,3-dihydro-1*H*-benzimidazole-4-carboxylate,



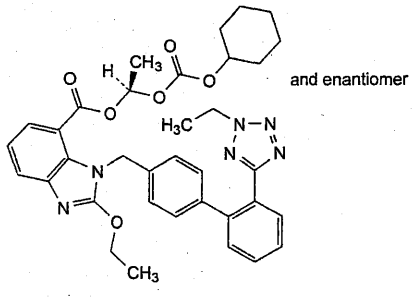
C. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 3-[[2′-(1-ethyl-1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-2-oxo-2,3-dihydro-1*H*-benzimidazole-4-carboxylate,



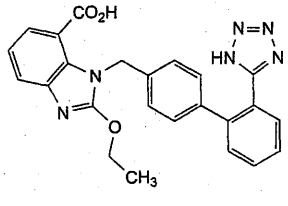
D. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 3-[[2′-(2-ethyl-2*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-2-oxo-2,3-dihydro-1*H*-benzimidazole-4-carboxylate,



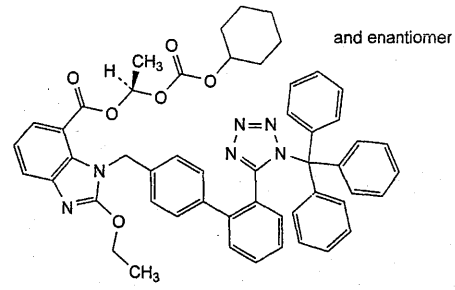
E. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 2-ethoxy-1-[[2′-(1-ethyl-1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylate,



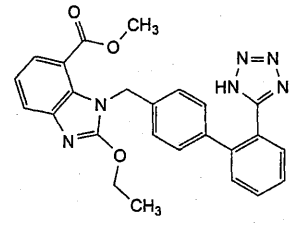
F. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 2-ethoxy-1-[[2′-(2-ethyl-2*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylate,



G. 2-ethoxy-1-[[2′-(1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylic acid (candesartan),



H. (1*RS*)-1-[[(cyclohexyloxy)carbonyl]oxy]ethyl 2-ethoxy-1-[[2′-[1-(triphenylmethyl)-1*H*-tetrazol-5-yl]biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylate (*N*-tritylcandesartan),

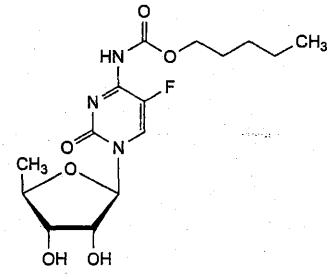


I. methyl 2-ethoxy-1-[[2′-(1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-benzimidazole-7-carboxylate.

Ph Eur

Capecitabine

(Ph. Eur. monograph 2762)



$C_{15}H_{22}FN_3O_6$ 359.3 *154361-50-9*

Action and use
Pyrimidine analogue; cytotoxic; treatment of colorectal cancer.

Preparation
Capecitabine Tablets

Ph Eur

DEFINITION
Pentyl [1-(5-deoxy-β-D-ribofuranosyl)-5-fluoro-2-oxo-1,2-dihydropyrimidin-4-yl]carbamate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Sparingly soluble in water, freely soluble in anhydrous ethanol, practically insoluble in heptane.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *capecitabine CRS*.

TESTS

Specific optical rotation (*2.2.7*)

+ 96.0 to + 100.0 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use or store them at 2-8 °C.*

Solvent mixture *acetonitrile R, methanol R, water R* (5:35:60 *V/V/V*).

Test solution Dissolve 60.0 mg of the substance to be examined in 80 mL of the solvent mixture, sonicate until dissolution is complete and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 60.0 mg of *capecitabine CRS* in 80 mL of the solvent mixture, sonicate until dissolution is complete and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 3 mg of *capecitabine impurity A CRS*, 3 mg of *capecitabine impurity B CRS* and 5 mg of *capecitabine impurity D CRS* in 80 mL of the solvent mixture, sonicate until dissolution is complete and dilute to 100.0 mL with the solvent mixture. Dilute 1 mL of the solution to 50 mL with the test solution.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R, methanol R*, 0.1 per cent *V/V* solution of *glacial acetic acid R* (5:35:60 *V/V/V*);
- *mobile phase B*: *acetonitrile R*, 0.1 per cent *V/V* solution of *glacial acetic acid R, methanol R* (5:15:80 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 20	100 → 49	0 → 51
20 - 30	49	51

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and D.

Relative retention With reference to capecitabine (retention time = about 17 min): impurity A = about 0.18; impurity B = about 0.19; impurities D and E = about 0.95.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to impurities A and B; minimum 2.0 between the peaks due to impurity D and capecitabine.

Calculation of percentage contents:
- for each impurity, use the concentration of capecitabine in reference solution (b);
- *correction factor*: multiply the peak area of impurity B by 1.3.

Limits:
- *impurities A, B*: for each impurity, maximum 0.3 per cent;
- *sum of impurities D and E*: maximum 0.2 per cent;
- *unspecified impurities*: for each impurity, maximum 0.05 per cent;
- *total*: maximum 0.5 per cent;
- *reporting threshold*: 0.03 per cent.

Water (*2.5.32*)

Maximum 0.3 per cent.

Inject 1.0 mL of a 0.200 g/mL solution of the substance to be examined in *methanol R*.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

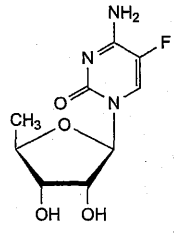
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{15}H_{22}FN_3O_6$ taking into account the assigned content of *capecitabine CRS*.

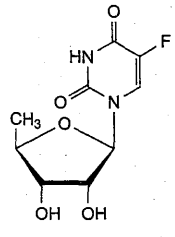
IMPURITIES

Specified impurities *A, B, D, E.*

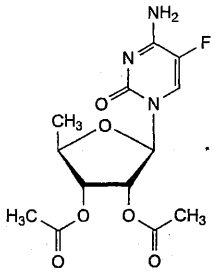
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, F, G.*



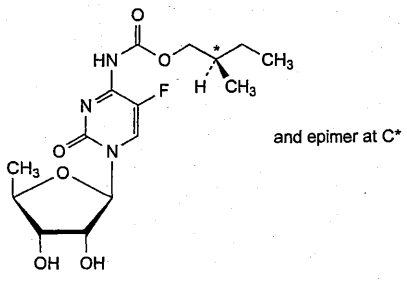
A. 4-amino-1-(5-deoxy-β-D-ribofuranosyl)-5-fluoropyrimidin-2(1*H*)-one (5′-deoxy-5-fluorocytidine),



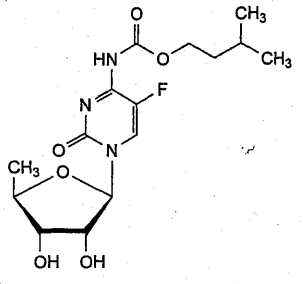
B. 1-(5-deoxy-β-D-ribofuranosyl)-5-fluoropyrimidine-2,4(1*H*,3*H*)-dione (5′-deoxy-5-fluorouridine),



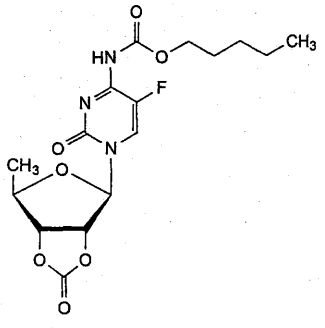
C. 1-(2,3-di-*O*-acetyl-5-deoxy-β-D-ribofuranosyl)-4-amino-5-fluoropyrimidin-2(1*H*)-one,



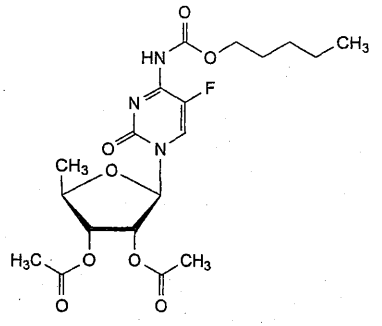
D. (2*RS*)-2-methylbutyl [1-(5-deoxy-β-D-ribofuranosyl)-5-fluoro-2-oxo-1,2-dihydropyrimidin-4-yl]carbamate,



E. 3-methylbutyl [1-(5-deoxy-β-D-ribofuranosyl)-5-fluoro-2-oxo-1,2-dihydropyrimidin-4-yl]carbamate,



F. pentyl [5-fluoro-1-[(3a*R*,4*R*,6*R*,6a*R*)-6-methyl-2-oxotetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl]-2-oxo-1,2-dihydropyrimidin-4-yl]carbamate,



G. pentyl [1-(2,3-di-*O*-acetyl-5-deoxy-β-D-ribofuranosyl)-5-fluoro-2-oxo-1,2-dihydropyrimidin-4-yl]carbamate.

Ph Eur

Caprylocaproyl Macrogolglycerides

(Ph. Eur. monograph 1184)

Action and use
Excipient.

Ph Eur

DEFINITION

Mixtures of monoesters, diesters and triesters of glycerol and monoesters and diesters of macrogols with a mean relative molecular mass between 200 and 400.

They are obtained by partial alcoholysis of medium-chain triglycerides using macrogol, or by esterification of glycerol and macrogol with caprylic (octanoic) acid and capric (decanoic) acid, or by mixing glycerol esters and condensates of ethylene oxide with caprylic acid and capric acid. They may contain free macrogols.

CHARACTERS

Appearance
Pale-yellow, oily liquid.

Solubility
Dispersible in hot water, freely soluble in methylene chloride.

Density: about 1.0 at 20 °C.

Refractive index
About 1.4 at 20 °C.

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 20 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *hexane R, ether R* (30:70 *V/V*).

Application 50 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of *rhodamine B R* in *ethanol (96 per cent) R* and examine in ultraviolet light at 365 nm.

Results The chromatogram shows a spot due to triglycerides with an R_F value of about 0.9 (R_{st} 1) and spots due to 1,3-diglycerides (R_{st} 0.7), to 1,2-diglycerides (R_{st} 0.6), to monoglycerides (R_{st} 0.1) and to esters of macrogol (R_{st} 0).

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Composition of fatty acids (see Tests).

TESTS

Viscosity (*2.2.9*)

Carry out the determination at 20 ± 0.5 °C.

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Viscosity (mPa·s)
4	200	30 to 50
6	300	60 to 80
8	400	80 to 110

Acid value (*2.5.1*)

Maximum 2.0, determined on 2.0 g.

Hydroxyl value (*2.5.3, Method A*)

Use 1.0 g.

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Hydroxyl value
4	200	80 to 120
6	300	140 to 180
8	400	170 to 205

Peroxide value (*2.5.5, Method A*)

Maximum 6.0, determined on 2.0 g.

Saponification value (*2.5.6*)

Use 2.0 g.

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Saponification value
4	200	265 to 285
6	300	170 to 190
8	400	85 to 105

Alkaline impurities

Introduce 5.0 g into a test-tube and carefully add a mixture, neutralised if necessary with *0.01 M hydrochloric acid* or with *0.01 M sodium hydroxide*, of 0.05 mL of a 0.4 g/L solution of *bromophenol blue R* in *ethanol (96 per cent) R*, 0.3 mL of *water R* and 10 mL of *ethanol (96 per cent) R*. Shake and allow to stand. Not more than 1.0 mL of *0.01 M hydrochloric acid* is required to change the colour of the upper layer to yellow.

Free glycerol

Maximum 5.0 per cent.

Dissolve 1.20 g in 25.0 mL of *methylene chloride R*. Heat if necessary. After cooling, add 100 mL of *water R*. Shake and add 25.0 mL of *periodic acetic acid solution R*. Shake and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of *potassium iodide R*. Allow to stand for 1 min. Add 1 mL of *starch solution R*. Titrate the iodine with *0.1 M sodium thiosulfate*. Carry out a blank titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 2.3 mg of glycerol.

Composition of fatty acids (*2.4.22, Method A*)

Composition of the fatty-acid fraction of the substance:

— *caproic acid*: maximum 2.0 per cent;

— *caprylic acid*: 50.0 per cent to 80.0 per cent;

— *capric acid*: 20.0 per cent to 50.0 per cent;

— *lauric acid*: maximum 3.0 per cent;

— *myristic acid*: maximum 1.0 per cent.

Ethylene oxide and dioxan (*2.4.25*)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 1.00 g. Use a mixture of 30 volumes of *anhydrous methanol R* and 70 volumes of *methylene chloride R* as solvent.

Total ash (*2.4.16*)

Maximum 0.1 per cent.

LABELLING

The label states the type of macrogol used (mean relative molecular mass) or the number of ethylene oxide units per molecule (nominal value).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for caprylocaproyl macrogolglycerides used as self-emulsifying agents and solubilisers.

Hydroxyl value

(see Tests).

Saponification value

(see Tests).

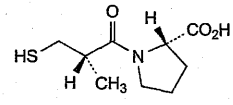
Composition of fatty acids

(see Tests).

Ph Eur

Captopril

(Ph. Eur. monograph 1079)



$C_9H_{15}NO_3S$ 217.3 *62571-86-2*

Action and use

Angiotensin converting enzyme inhibitor.

Preparations

Captopril Oral Solution

Captopril Tablets

Ph Eur

DEFINITION

(2*S*)-1-[(2*S*)-2-Methyl-3-sulfanylpropanoyl]pyrrolidine-2-carboxylic acid.

Content
98.0 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, freely soluble in methanol and in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *captopril CRS*.

TESTS

Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
2.0 to 2.6 for solution S.

Specific optical rotation (*2.2.7*)
−132 to −127 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Impurity F
Gas chromatography (*2.2.28*).

Reagent solution Add 2.8 mL of *acetyl chloride R* dropwise to 17.2 mL of *anhydrous methanol R* at 0 °C and mix. Allow to stand for 20 min at room temperature before use.

Test solution Introduce 20.0 mg of the substance to be examined into a vial and add 1.0 mL of the reagent solution. Mix and heat at 60 °C for 30 min. Evaporate to dryness under a stream of *nitrogen R*. Dissolve the residue in 0.5 mL of *ethyl acetate R*, add 0.5 mL of *pentafluoropropionic anhydride R*, mix and heat at 60 °C for 30 min. Evaporate to dryness under a stream of *nitrogen R*. Dissolve the residue in 1.0 mL of *butyl acetate R*.

Reference solution (a) Dissolve the contents of a vial of *captopril for system suitability CRS* (containing impurity F) in 1.0 mL of the reagent solution. Prepare as described for the test solution.

Reference solution (b) Mix 0.25 mL of reference solution (a) and 0.75 mL of *butyl acetate R*.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 1 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1.2 mL/min.

Split ratio 1:20.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 10	200
	10 - 14	200 → 240
	14 - 34	240
Injection port		270
Detector		300

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to captopril (retention time = about 6 min): impurity F = about 0.96.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to impurity F and captopril in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 10 for the peak due to impurity F in the chromatogram obtained with reference solution (b).

Calculate the percentage content of impurity F using the following expression:

$$\frac{A}{A+B} \times 100$$

A = area of the peak due to impurity F in the chromatogram obtained with the test solution;
B = area of the peak due to captopril in the chromatogram obtained with the test solution.

Limit:
— *impurity F*: maximum 0.2 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *phosphoric acid R, acetonitrile R1, water R* (0.08:10:90 *V/V/V*).

Test solution Dissolve 0.125 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 4.0 mg of *captopril impurity J CRS*, 5.0 mg of *captopril impurity B CRS*, 5.0 mg of *captopril impurity C CRS* and 5.0 mg of *captopril impurity D CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 20.0 mL with the solvent mixture. Prepare immediately before use.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *captopril impurity E CRS* in *acetonitrile R* and dilute to 25.0 mL with the same solvent. Dilute 4 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (c) In order to prepare impurity A *in situ*, introduce 1.0 mL of the test solution into a volumetric flask and add 230 µL of *0.05 M iodine*. If the solution is not colourless, add *0.1 M sodium thiosulfate* dropwise until it becomes colourless, and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.3 m, Ø = 3.9 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (10 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: *phosphoric acid R, water R* (0.08:100 *V/V*);
— *mobile phase B*: *phosphoric acid R, acetonitrile R1, water R* (0.08:50:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90	10
5 - 20	90 → 50	10 → 50
20 - 45	50	50

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 25 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C, D and J; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to captopril (retention time = about 15 min): impurity C = about 0.6; impurity D = about 0.8; impurity E = about 0.9; impurity B = about 1.17; impurity J = about 1.22; impurity A = about 1.7.

System suitability:

— *resolution*: minimum 1.5 between the peaks due to impurities B and J in the chromatogram obtained with reference solution (a);

— *resolution*: minimum 2.0 between the peaks due to impurity E and captopril in the chromatogram obtained with reference solution (b).

Limits:

— *impurity A*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (d) (1.0 per cent);

— *impurity J*: not more than 2.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurities B, C, D*: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— *impurity E*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.10 per cent);

— *total*: maximum 1.2 per cent;

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying under high vacuum at 60 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

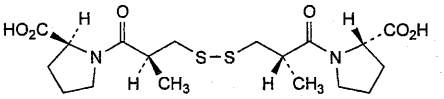
Dissolve 0.150 g in 30 mL of *water R*. Titrate with *0.05 M iodine*, determining the end-point potentiometrically (*2.2.20*). Use a combined platinum electrode.

1 mL of *0.05 M iodine* is equivalent to 21.73 mg of $C_9H_{15}NO_3S$.

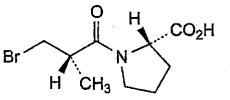
IMPURITIES

Specified impurities *A, B, C, D, E, F, J.*

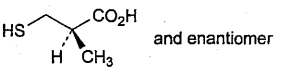
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, H, I, L, M, N, O.*



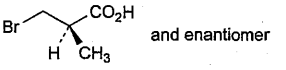
A. 1,1′-[disulfanediylbis[(2*S*)-2-methyl-1-oxopropane-3,1-diyl]]bis[(2*S*)-pyrrolidine-2-carboxylic] acid (captopril disulfide),



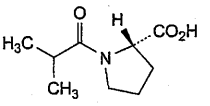
B. (2*S*)-1-[(2*S*)-3-bromo-2-methylpropanoyl]pyrrolidine-2-carboxylic acid,



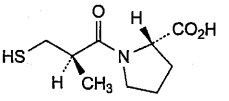
C. (2*RS*)-2-methyl-3-sulfanylpropanoic acid,



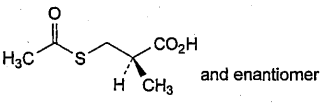
D. (2*RS*)-3-bromo-2-methylpropanoic acid,



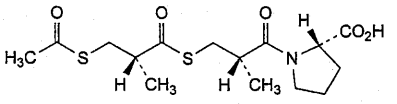
E. (2*S*)-1-(2-methylpropanoyl)pyrrolidine-2-carboxylic acid,



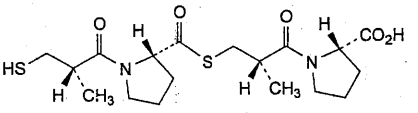
F. (2*S*)-1-[(2*R*)-2-methyl-3-sulfanylpropanoyl]pyrrolidine-2-carboxylic acid (*epi*-captopril),



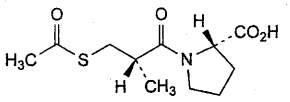
G. (2*RS*)-3-(acetylsulfanyl)-2-methylpropanoic acid,



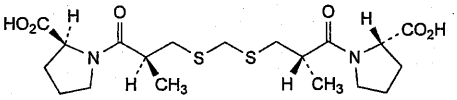
H. (2*S*)-1-[(2*S*)-3-[[(2*R*)-3-(acetylsulfanyl)-2-methylpropanoyl]sulfanyl]-2-methylpropanoyl]pyrrolidine-2-carboxylic acid,



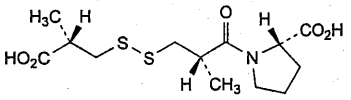
I. (2*S*)-1-[(2*S*)-3-[[[(2*S*)-1-[(2*S*)-2-methyl-3-sulfanylpropanoyl]pyrrolidin-2-yl]carbonyl]sulfanyl]-2-methylpropanoyl]pyrrolidine-2-carboxylic acid,



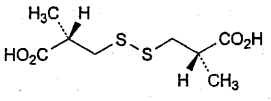
J. (2*S*)-1-[(2*S*)-3-(acetylsulfanyl)-2-methylpropanoyl] pyrrolidine-2-carboxylic acid (acetylcaptopril),



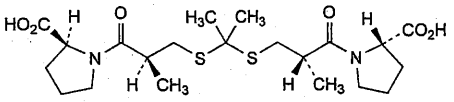
L. 1,1′-[methylenebis[sulfanediyl[(2*S*)-2-methyl-1-oxopropane-3,1-diyl]]]bis[(2*S*)-pyrrolidine-2-carboxylic] acid,



M. (2*S*)-1-[(2*S*)-3-[[(2*S*)-2-carboxypropyl]disulfanyl]-2-methylpropanoyl]pyrrolidine-2-carboxylic acid,



N. 3,3′-disulfanediylbis[(2*S*)-2-methylpropanoic] acid,

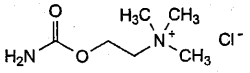


O. 1,1′-[propane-2,2-diylbis[sulfanediyl[(2*S*)-2-methyl-1-oxopropane-3,1-diyl]]]bis[(2*S*)-pyrrolidine-2-carboxylic] acid.

Ph Eur

Carbachol

(Ph. Eur. monograph 1971)



$C_6H_{15}ClN_2O_2$ 182.7 *51-83-2*

Action and use
Cholinoceptor agonist.

Ph Eur

DEFINITION
2-(Carbamoyloxy)-*N*,*N*,*N*-trimethylethanaminium chloride.

Content
99.0 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline, hygroscopic powder.

Solubility
Very soluble in water, sparingly soluble in alcohol, practically insoluble in acetone.

IDENTIFICATION
First identification: A, C.
Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *carbachol CRS.*

B. Examine the chromatograms obtained in the test for related substances.
Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. 0.5 mL of solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 2.0 mL of solution S, add 0.05 mL of *methyl red mixed solution R*. Not more than 0.2 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Related substances
Thin-layer chromatography (*2.2.27*).
Prepare the solutions immediately before use.
Test solution (a) Dissolve 0.20 g of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.
Test solution (b) Dilute 2.0 mL of test solution (a) to 20.0 mL with *methanol R*.
Reference solution (a) Dissolve 20 mg of *carbachol CRS* in *methanol R* and dilute to 5.0 mL with the same solvent.
Reference solution (b) Dissolve 8 mg of *choline chloride R* and 8 mg of *acetylcholine chloride CRS* in *methanol R* and dilute to 10.0 mL with the same solvent. Dilute 5.0 mL to 10.0 mL with *methanol R*.
Plate *cellulose for chromatography R* as the coating substance.
Mobile phase *water R, methanol R* (10:90 *V/V*).
Application 10 µL.
Development Over 2/3 of the plate.
Detection Spray with *potassium iodobismuthate solution R3.*
System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.
Limits In the chromatogram obtained with test solution (a):
— *any impurity*: any spot, apart from the principal spot, is not more intense than one or other of the 2 principal spots in the chromatogram obtained with reference solution (b) (1 per cent). Compare the spots with the spot of the most appropriate colour in the chromatogram obtained with reference solution (b).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g of the residue obtained in the test for loss on drying.

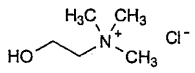
ASSAY
Dissolve 0.150 g in a mixture of 10 mL of *anhydrous acetic acid R* and 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*. Determine the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 18.27 mg of $C_6H_{15}ClN_2O_2$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

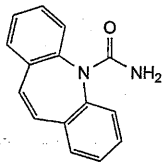


A. 2-hydroxy-*N*,*N*,*N*-trimethylethanaminium chloride (choline chloride).

Ph Eur

Carbamazepine

(Ph. Eur. monograph 0543)



$C_{15}H_{12}N_2O$ 236.3 *298-46-4*

Action and use

Antiepileptic.

Preparations

Carbamazepine Oral Suspension

Carbamazepine Tablets

Carbamazepine Chewable Tablets

Carbamazepine Prolonged-release Tablets

Ph Eur

DEFINITION

5*H*-Dibenzo[*b*,*f*]azepine-5-carboxamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in methylene chloride, sparingly soluble in acetone and in ethanol (96 per cent).

It shows polymorphism (*5.9*). The acceptable crystalline form corresponds to *carbamazepine CRS*.

IDENTIFICATION

A. Melting point (*2.2.14*): 189 °C to 193 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *carbamazepine CRS*.

Preparation Examine the substances as discs without prior treatment.

TESTS

Acidity or alkalinity

To 1.0 g add 20 mL of *carbon dioxide-free water R*, shake for 15 min and filter. To 10 mL of the filtrate add 0.05 mL of *phenolphthalein solution R1* and 0.5 mL of *0.01 M sodium hydroxide*; the solution is red. Add 1.0 mL of *0.01 M hydrochloric acid*; the solution is colourless. Add 0.15 mL of *methyl red solution R*; the solution is red.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 60.0 mg of the substance to be examined in *methanol R2* and dilute to 20.0 mL with the same solvent. Sonicate. Dilute 10.0 mL of this solution to 20.0 mL with *water R*.

Test solution (b) Dilute 10.0 mL of test solution (a) to 50.0 mL with a mixture of equal volumes of *methanol R2* and *water R*.

Reference solution (a) Dissolve 7.5 mg of *carbamazepine CRS*, 7.5 mg of *carbamazepine impurity A CRS* and 7.5 mg of *iminodibenzyl R* (impurity E) in *methanol R2* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with a mixture of equal volumes of *methanol R2* and *water R*.

Reference solution (b) Dissolve 60.0 mg of *carbamazepine CRS* in *methanol R2* and dilute to 20.0 mL with the same solvent. Sonicate. Dilute 5.0 mL of this solution to 50.0 mL with a mixture of equal volumes of *methanol R2* and *water R*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *nitrile silica gel for chromatography R1* (10 µm).

Mobile phase *tetrahydrofuran R*, *methanol R2*, *water R* (3:12:85 *V/V/V*); to 1000 mL of this solution add 0.2 mL of *anhydrous formic acid R* and 0.5 mL of *triethylamine R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL of test solution (a) and reference solution (a).

Run time 8 times the retention time of carbamazepine.

Relative retention With reference to carbamazepine (retention time = about 10 min): impurity A = about 0.9; impurity E = about 3.5.

System suitability:

— *resolution*: minimum 1.7 between the peaks due to impurity A and carbamazepine in the chromatogram obtained with reference solution (a).

Limits:

— *impurities A, E*: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— *unspecified impurities*: not more than the area of the peak due to carbamazepine in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the area of the peak due to carbamazepine in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the peak due to carbamazepine in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)

Maximum 140 ppm.

Suspend 0.715 g in 20 mL of *water R* and boil for 10 min. Cool and dilute to 20 mL with *water R*. Filter through a membrane filter (nominal pore size 0.8 µm). Dilute 10 mL of the filtrate to 15 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (b).

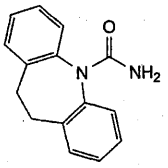
System suitability:
— *repeatability*: reference solution (b).

Calculate the percentage content of $C_{15}H_{12}N_2O$ from the declared content of *carbamazepine CRS*.

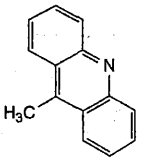
STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, E.*

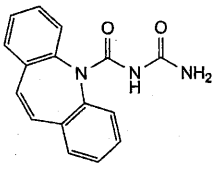
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, F, G.*



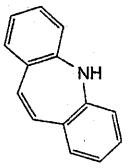
A. 10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepine-5-carboxamide (10,11-dihydrocarbamazepine),



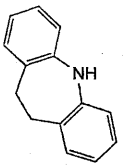
B. 9-methylacridine,



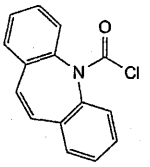
C. (5*H*-dibenzo[*b*,*f*]azepin-5-ylcarbonyl)urea (*N*-carbamoylcarbamazepine),



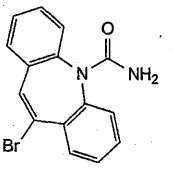
D. 5*H*-dibenzo[*b*,*f*]azepine (iminostilbene),



E. 10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepine (iminodibenzyl),



F. 5*H*-dibenzo[*b*,*f*]azepine-5-carbonyl chloride (5-chlorocarbonyliminostilbene),

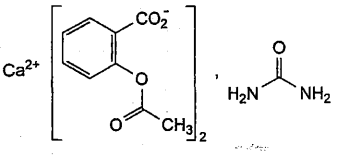


G. 10-bromo-5*H*-dibenzo[*b*,*f*]azepine-5-carboxamide (10-bromocarbamazepine).

Ph Eur

Carbasalate Calcium

(Ph. Eur. monograph 1185)



$C_{19}H_{18}CaN_2O_9$ 458.4 *5749-67-7*

Action and use
Salicylate; non-selective cyclo-oxygenase inhibitor; antipyretic; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
Equimolecular compound of calcium di[2-(acetyloxy) benzoate] and urea.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in dimethylformamide, practically insoluble in acetone and in anhydrous methanol.

Protect the substance from moisture during handling. Examination in aqueous solutions has to be performed immediately after preparation.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.250 g in *water R* and dilute to 100.0 mL with the same solvent. To 1.0 mL of the solution add 75 mL of *water R* and 5 mL of *dilute hydrochloric acid R*, mix and dilute to 100.0 mL with *water R*. Examine immediately.

Spectral range 220-350 nm.

Absorption maxima At 228 nm and 276 nm.

Specific absorbance at the absorption maxima:
— at 228 nm: 363 to 379,
— at 276 nm: 49 to 53.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of carbasalate calcium.*

C. Dissolve 0.1 g in 10 mL of *water R*, boil for 2 min and cool. The solution gives reaction (a) of salicylates (*2.3.1*).

D. Heat 0.2 g with 0.2 g of *sodium hydroxide R*; a yellow or yellowish-brown colour is produced and the vapour turns *red litmus paper R* blue.

E. It gives reaction (a) of calcium (*2.3.1*).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and is colourless (*2.2.2, Method II*).

Dissolve 2.5 g in 50 mL of *water R*.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *phosphoric acid R, methanol R, acetonitrile for chromatography R* (0.5:8:92 *V/V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in 5 mL of the solvent mixture, sonicate for 15 min and dilute to 10.0 mL with the solvent mixture. Filter the solution through a membrane filter (nominal pore size 0.45 µm).

Reference solution (a) Dissolve 10.0 mg of *salicylic acid CRS* (impurity C) in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of *carbasalate impurity B CRS* in 20.0 mL of the solvent mixture.

Reference solution (d) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Mix 1.0 mL of this solution with 5.0 mL of reference solution (a), add 1.0 mL of reference solution (c) and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase *phosphoric acid R, acetonitrile for chromatography R, water R* (0.5:40:60 *V/V/V*).

Flow rate 1.8 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL of the test solution and reference solutions (b) and (d).

Run time 8 times the retention time of acetylsalicylic acid.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B and C.

Relative retention With reference to acetylsalicylic acid (retention time = about 2 min): impurity C = about 1.3; impurity B = about 2.5.

System suitability Reference solution (d):
— *resolution*: minimum 5.0 between the peaks due to acetylsalicylic acid and impurity C.

Limits:
— *impurity C*: not more than 5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurity B*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

Sodium

Maximum 0.1 per cent.

Atomic emission spectrometry (*2.2.22, Method I*).

Test solution Dissolve 1.0 g in 500.0 mL of *water R*.

Water (*2.5.12*)

Maximum 0.1 per cent, determined on 1.000 g. Use a mixture of 15 mL of *anhydrous methanol R* and 15 mL of *dimethylformamide R* as the solvent.

ASSAY

In a flask with a ground-glass stopper, dissolve 0.400 g in 25 mL of *water R*. Add 25.0 mL of *0.1 M sodium hydroxide*. Close the flask and allow to stand for 2 h. Titrate with *0.1 M hydrochloric acid*, using 0.2 mL of *phenolphthalein solution R*. Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 22.92 mg of $C_{19}H_{18}CaN_2O_9$.

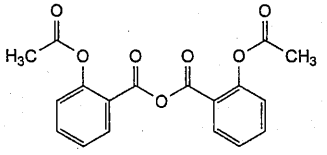
STORAGE

In an airtight container.

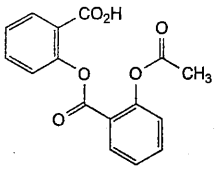
IMPURITIES

Specified impurities *B, C.*

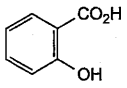
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D.*



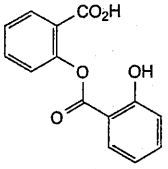
A. 2-(acetyloxy)benzoic anhydride,



B. 2-[[2-(acetyloxy)benzoyl]oxy]benzoic acid (acetylsalicylsalicylic acid),



C. 2-hydroxybenzenecarboxylic acid (salicylic acid),

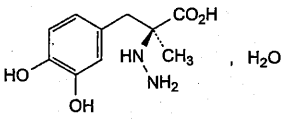


D. 2-[(2-hydroxybenzoyl)oxy]benzoic acid (salicylsalicylic acid).

Ph Eur

Carbidopa

(Ph. Eur. monograph 0755)



$C_{10}H_{14}N_2O_4,H_2O$ 244.2 *38821-49-7*

Action and use
Dopa decarboxylase inhibitor.

Preparation
Co-careldopa Tablets

Ph Eur

DEFINITION
(2*S*)-3-(3,4-Dihydroxyphenyl)-2-hydrazino-2-methylpropanoic acid monohydrate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish-white powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in dilute solutions of mineral acids.

IDENTIFICATION
First identification: A, C.

Second identification: A, B, D, E.

A. Specific optical rotation (see Tests).

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50 mg in an 8.5 g/L solution of *hydrochloric acid R* in *methanol R* and dilute to 100.0 mL with the same solution. Dilute 10.0 mL of this solution to 100.0 mL with an 8.5 g/L solution of *hydrochloric acid R* in *methanol R*.

Spectral range 230-350 nm.

Absorption maximum At 283 nm.

Specific absorbance at the absorption maximum 135 to 150 (dried substance).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *carbidopa CRS*.

D. Shake vigorously about 5 mg with 10 mL of *water R* for 1 min and add 0.3 mL of *ferric chloride solution R2*. An intense green colour is produced, which quickly changes to reddish-brown.

E. Suspend about 20 mg in 5 mL of *water R* and add 5 mL of *cupri-tartaric solution R*. On heating, the colour of the solution changes to dark brown and a red precipitate is formed.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 or B_6 (*2.2.2, Method II*).

Dissolve 0.25 g in 25 mL of *1 M hydrochloric acid*.

Specific optical rotation (*2.2.7*)
−26.5 to −22.5 (dried substance).

Using an ultrasonic bath, dissolve completely 0.250 g in *aluminium chloride solution R* and dilute to 25.0 mL with the same solution.

Hydrazine
Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.50 g in *dilute hydrochloric acid R* and dilute to 2.0 mL with the same acid.

Test solution (b) Place 25 g of *strongly basic anion-exchange resin R* into each of 2 conical flasks with ground-glass stoppers. To each, add 150 mL of *carbon dioxide-free water R* and shake from time to time during 30 min. Decant the liquid from both flasks and repeat the process with further quantities, each of 150 mL, of *carbon dioxide-free water R*.

Take two 100 mL measuring cylinders 3.5-4.5 cm in internal diameter and label these A and B. Into cylinder A, transfer as completely as possible the resin from 1 conical flask using 60 mL of *carbon dioxide-free water R*; into cylinder B, transfer the 2nd quantity of resin, this time using 20 mL of *carbon dioxide-free water R*.

Into each cylinder, insert a gas-inlet tube, the end of which has an internal diameter of 2-3 mm and which reaches almost to the bottom of the cylinder. Pass a rapid stream of *nitrogen for chromatography R* through each mixture so that homogeneous suspensions are formed. After 30 min, without interrupting the gas flow, add 1.0 mL of test solution (a) to cylinder A; after 1 min stop the gas flow into cylinder A and transfer the contents, through a moistened filter paper, into cylinder B. After 1 min, stop the gas flow to cylinder B and pour the solution immediately through a moistened filter paper into a freshly prepared mixture of 1 mL of a 200 g/L solution of *salicylaldehyde R* in *methanol R* and 20 mL of *phosphate buffer solution pH 5.5 R* in a conical flask; shake thoroughly for 1 min and heat in a water-bath at 60 °C for 15 min. The liquid becomes clear. Allow to cool, add 2.0 mL of *toluene R* and shake vigorously for 2 min. Transfer the mixture into a centrifuge tube and centrifuge.

Separate the toluene layer in a 100 mL separating funnel and shake vigorously with 2 quantities, each of 20 mL, of a 200 g/L solution of *sodium metabisulfite R* and finally with 2 quantities, each of 50 mL, of *water R*. Separate the toluene layer.

Reference solution (a) Dissolve 10 mg of *hydrazine sulfate R* in *dilute hydrochloric acid R* and dilute to 50 mL with the same acid. Dilute 1.0 mL of this solution to 10.0 mL with *dilute hydrochloric acid R*.

Reference solution (b) Prepare the solution at the same time and in the same manner as described for test solution (b) using 1.0 mL of reference solution (a) instead of 1.0 mL of test solution (a).

Plate *TLC silanised silica gel plate R.*

Mobile phase *water R, methanol R* (10:20 *V/V*).

Application 10 µL of test solution (b) and reference solution (b).

Development Over 1/2 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 365 nm.

Limit:

— *hydrazine*: any spot showing a yellow fluorescence is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (20 ppm).

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Buffer solution Dissolve 6.9 g of *sodium dihydrogen phosphate monohydrate R* in 900 mL of *water for chromatography R*, adjust to pH 2.2 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase, add 20 µL of *hydrochloric acid R* and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 4 mg of *carbidopa for system suitability CRS* (containing impurities A, D, E, I and J) in the mobile phase, add 4 µL of *hydrochloric acid R* and dilute to 2.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *carbidopa impurity mixture CRS* (impurities F and H) in 1.0 mL of reference solution (b).

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R* (3.5 µm);

— *temperature*: 25 °C.

Mobile phase *ethanol (96 per cent) R*, buffer solution (7:93 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 6 times the retention time of carbidopa.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, D + E, F, H, I and J.

Relative retention With reference to carbidopa (retention time = about 3 min): impurity A = about 0.9; impurities D and E = about 1.9; impurity H = about 2.5; impurity I = about 3.7; impurity J = about 4.0; impurity F = about 4.4.

System suitability:

— *resolution*: minimum 1.5 between the peaks due to impurity A and carbidopa in the chromatogram obtained with reference solution (b); minimum 1.5 between the peaks due to impurities I and J in the chromatogram obtained with reference solution (b);

— *signal-to-noise ratio*: minimum 30 for the principal peak in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:

— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurities D and E = 1.5; impurity I = 1.5; impurity J = 1.5;

— for each impurity, use the concentration of carbidopa in reference solution (a).

Limits:

— *impurity A*: maximum 0.5 per cent;

— *impurity J*: maximum 0.25 per cent;

— *sum of impurities D and E*: maximum 0.2 per cent;

— *impurities F, H, I*: for each impurity, maximum 0.15 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 1.0 per cent;

— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

6.9 per cent to 7.9 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g with gentle heating in 75 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 22.62 mg of $C_{10}H_{14}N_2O_4$.

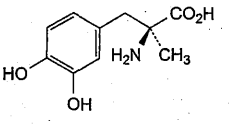
STORAGE

Protected from light.

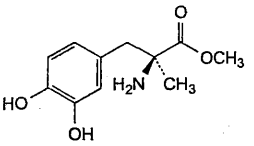
IMPURITIES

Specified impurities *A, D, E, F, H, I, J.*

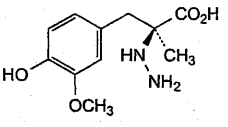
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, G.*



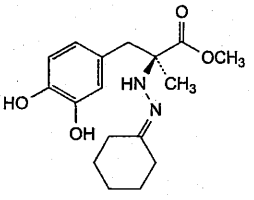
A. (2*S*)-2-amino-3-(3,4-dihydroxyphenyl)-2-methylpropanoic acid (L-methyldopa),



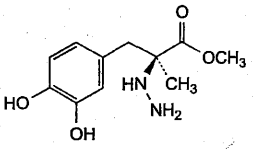
B. methyl (2*S*)-2-amino-3-(3,4-dihydroxyphenyl)-2-methylpropanoate,



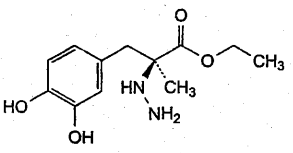
C. (2*S*)-2-hydrazino-3-(4-hydroxy-3-methoxyphenyl)-2-methylpropanoic acid (3-*O*-methylcarbidopa),



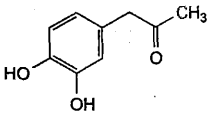
D. methyl (2*S*)-2-(2-cyclohexylidenehydrazino)-3-(3,4-dihydroxyphenyl)-2-methylpropanoate,



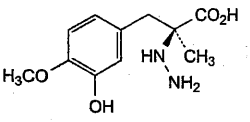
E. methyl (2*S*)-3-(3,4-dihydroxyphenyl)-2-hydrazino-2-methylpropanoate,



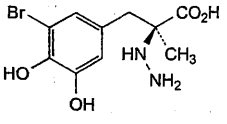
F. ethyl (2*S*)-3-(3,4-dihydroxyphenyl)-2-hydrazino-2-methylpropanoate,



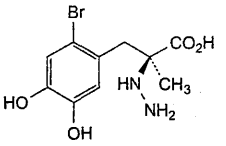
G. 1-(3,4-dihydroxyphenyl)propan-2-one,



H. (2*S*)-2-hydrazino-3-(3-hydroxy-4-methoxyphenyl)-2-methylpropanoic acid,



I. (2*S*)-3-(3-bromo-4,5-dihydroxyphenyl)-2-hydrazino-2-methylpropanoic acid,

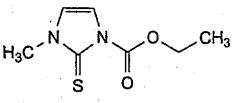


J. (2*S*)-3-(2-bromo-4,5-dihydroxyphenyl)-2-hydrazino-2-methylpropanoic acid.

Ph Eur

Carbimazole

(Ph. Eur. monograph 0884)



$C_7H_{10}N_2O_2S$ 186.2 *22232-54-8*

Action and use
Thionamide antithyroid drug.

Preparation
Carbimazole Tablets

Ph Eur

DEFINITION

Ethyl 3-methyl-2-thioxo-2,3-dihydro-1*H*-imidazole-1-carboxylate.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or yellowish-white, crystalline powder.

Solubility
Slightly soluble in water, soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 122 °C to 125 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *carbimazole CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methylene chloride R* and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 10 mg of *carbimazole CRS* in *methylene chloride R* and dilute to 10.0 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetone R, methylene chloride R* (20:80 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air for 30 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 10 mg in a mixture of 0.05 mL of *dilute hydrochloric acid R* and 50 mL of *water R*. Add 1 mL of *potassium iodobismuthate solution R*. A red precipitate is formed.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *acetonitrile R, water R* (20:80 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *thiamazole CRS* (impurity A) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Mix 1 mL of the solution with 2 mL of the test solution and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of *thiamazole CRS* (impurity A) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 25.0 mg of *carbimazole CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 3.9 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R, water R* (10:90 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a), (b) and (c).

Run time 1.5 times the retention time of carbimazole.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to carbimazole (retention time = about 6 min): impurity A = about 0.2.

System suitability Reference solution (a):
- *resolution*: minimum 5.0 between the peaks due to impurity A and carbimazole.

Limits:
- *impurity A*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
- *total*: maximum 0.6 per cent;
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 24 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

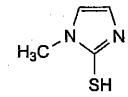
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (d).

Calculate the percentage content of $C_7H_{10}N_2O_2S$ taking into account the assigned content of *carbimazole CRS*.

IMPURITIES

Specified impurities *A*.

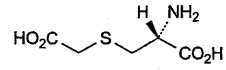


A. 1-methyl-1*H*-imidazole-2-thiol (thiamazole).

Ph Eur

Carbocisteine

(Ph. Eur. monograph 0885)



$C_5H_9NO_4S$ 179.2 *638-23-3*

Action and use

Mucolytic.

Ph Eur

DEFINITION

Carbocisteine contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of (2*R*)-2-amino-3-[(carboxymethyl)sulfanyl]propanoic acid, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, practically insoluble in water and in alcohol. It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *carbocisteine CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for ninhydrin-positive substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 4.5 mL of *dilute sodium hydroxide solution R*. Heat on a water-bath for 10 min. Cool and add 1 mL of a 25 g/L solution of *sodium nitroprusside R*. A dark red colour is produced, which changes to brown and then to yellow within a few minutes.

TESTS

Solution S

Disperse 5.00 g in 20 mL of *water R* and add dropwise with shaking 2.5 mL of *strong sodium hydroxide solution R*. Adjust to pH 6.3 with *1 M sodium hydroxide* and dilute to 50.0 mL with *water R*.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

Shake 0.2 g with 20 mL of *carbon dioxide-free water R*. The pH of the suspension is 2.8 to 3.0.

Specific optical rotation (*2.2.7*)

−32.5 to −35.5, determined on solution S and calculated with reference to the dried substance.

Ninhydrin-positive substances

Examine by thin-layer chromatography (*2.2.27*), using a suitable silica gel as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *dilute ammonia R2* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with *water R*.

Reference solution (a) Dissolve 10 mg of *carbocisteine CRS* in *dilute ammonia R2* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (b) to 20 mL with *water R*.

Reference solution (c) Dissolve 10 mg of *carbocisteine CRS* and 10 mg of *arginine hydrochloride CRS* in 5 mL of *dilute ammonia R2* and dilute to 25 mL with *water R*.

Apply separately to the plate 5 µL of each solution. Allow the plate to dry in air. Develop over a path of 15 cm using a mixture of 20 volumes of *glacial acetic acid R*, 20 volumes of *water R* and 60 volumes of *butanol R*. Dry the plate in a current of warm air. Spray with *ninhydrin solution R* and heat at 100 °C to 105 °C for 15 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated principal spots.

Chlorides (*2.4.4*)

Dissolve 33 mg in 5 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*. The solution, without further addition of nitric acid, complies with the limit test for chlorides (0.15 per cent).

Sulfates (*2.4.13*)

Dissolve 0.5 g in 5 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*. The solution complies with the limit test for sulfates (300 ppm).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Not more than 0.3 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 10 mL of *anhydrous formic acid R* with slight heating and shake until dissolution is complete. Add 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 17.92 mg of $C_5H_9NO_4S$.

STORAGE

Store protected from light.

Ph Eur

Carbomers

(Ph. Eur. monograph 1299)

Action and use

Stabilizer in pharmaceutical products.

Preparation

Carbomer Eye Drops

Ph Eur

DEFINITION

High-molecular-mass polymers of acrylic acid cross-linked with alkenyl ethers of sugars or polyalcohols.

Content

56.0 per cent to 68.0 per cent of carboxylic acid (-CO_2H) groups (dried substance).

CHARACTERS

Appearance

White or almost white, fluffy, hygroscopic powder.

Solubility

Swells in water and in other polar solvents after dispersion and neutralisation with sodium hydroxide solution.

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Main bands At 1710 ± 5 cm^{-1}, 1454 ± 5 cm^{-1}, 1414 ± 5 cm^{-1}, 1245 ± 5 cm^{-1}, 1172 ± 5 cm^{-1}, 1115 ± 5 cm^{-1} and 801 ± 5 cm^{-1}, with the strongest band at 1710 ± 5 cm^{-1}.

B. Adjust a 10 g/L dispersion to about pH 7.5 with *1 M sodium hydroxide*. A highly viscous gel is formed.

C. Add 2 mL of a 100 g/L solution of *calcium chloride R*, with continuous stirring, to 10 mL of the gel from identification test B. A white precipitate is immediately produced.

D. Add 0.5 mL of *thymol blue solution R* to 10 mL of a 10 g/L dispersion. An orange colour is produced. Add 0.5 mL of *cresol red solution R* to 10 mL of a 10 g/L dispersion. A yellow colour is produced.

TESTS

Free acrylic acid

Liquid chromatography (*2.2.29*).

Test solution Mix 0.125 g of the substance to be examined with a 25 g/L solution of *aluminium potassium sulfate R* and dilute to 25.0 mL with the same solution. Heat the suspension at 50 °C for 20 min with shaking, then shake the suspension at room temperature for 60 min. Centrifuge and use the clear supernatant solution as the test solution.

Reference solution Dissolve 62.5 mg of *acrylic acid R* in a 25 g/L solution of *aluminium potassium sulfate R* and dilute to 100.0 mL with the same solution. Dilute 1.0 mL of this solution to 50.0 mL with a 25 g/L solution of *aluminium potassium sulfate R*.

Column:
— *size*: *l* = 0.12 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 1.361 g/L solution of *potassium dihydrogen phosphate R*, adjusted to pH 2.5 using *dilute phosphoric acid R*;
— *mobile phase B*: mixture of equal volumes of a 1.361 g/L solution of *potassium dihydrogen phosphate R* and *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	100	0
8 - 9	100 → 0	0 → 100
9 - 20	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 µL.

Retention time Acrylic acid = about 6.0 min.

Limit:
— *acrylic acid*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.25 per cent).

Benzene

Gas chromatography (*2.4.24, System A*).

Solution A Dissolve 0.100 g of *benzene R* in *dimethyl sulfoxide R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 100.0 mL with *water R*.

Test solution Weigh 50.0 mg of the substance to be examined into an injection vial and add 5.0 mL of *water R* and 1.0 mL of *dimethyl sulfoxide R*.

Reference solution Weigh 50.0 mg of the substance to be examined into an injection vial and add 4.0 mL of *water R*, 1.0 mL of *dimethyl sulfoxide R* and 1.0 mL of solution A.

Close the vials with a tight rubber membrane stopper coated with polytetrafluoroethylene and secure with an aluminium crimped cap. Shake to obtain a homogeneous dispersion.

Static head-space conditions that may be used:
— *equilibration temperature*: 80 °C;
— *equilibration time*: 60 min;
— *transfer-line temperature*: 90 °C.

Injection 1 mL of the gaseous phase of the test solution and 1 mL of the gaseous phase of the reference solution; repeat these injections twice more.

System suitability:
— *repeatability*: maximum relative standard deviation of the differences in area between the analyte peaks obtained from the 3 replicate pair injections of the reference solution and the test solution is 15 per cent.

Limit:
— *benzene*: the mean area of the peak due to benzene in the chromatograms obtained with the test solution is not greater than 0.5 times the mean area of the peak due to benzene in the chromatograms obtained with the reference solution (2 ppm).

Loss on drying (*2.2.32*)

Maximum 3.0 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 60 min.

Sulfated ash (*2.4.14*)

Maximum 4.0 per cent, determined on 1.0 g.

ASSAY

Slowly add 50 mL of *water R* to 0.120 g whilst stirring and heating at 60 °C for 15 min. Stop heating, add 150 mL of *water R* and continue stirring for 30 min. Add 2 g of *potassium chloride R* and titrate with *0.2 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.2 M sodium hydroxide* is equivalent to 9.0 mg of carboxylic acid (-CO_2H) groups.

STORAGE

In an airtight container.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for carbomers used as viscosity-increasing agents and gelling agents.

Apparent viscosity (*2.2.10*)

The nominal apparent viscosity is typically between 300 mPa·s and 115 000 mPa·s. For a product with a nominal apparent viscosity of 20 000 mPa·s or greater, the apparent viscosity is typically 70.0 per cent to 130.0 per cent of the nominal value; for a product with a nominal apparent viscosity of less than 20 000 mPa·s, the apparent viscosity is typically 50.0 per cent to 150.0 per cent of the nominal value.

Dry the substance to be examined *in vacuo* at 80 °C for 1 h. Carefully add 2.50 g of the previously dried substance to be examined to 500 mL of *water R* in a 1000 mL beaker while stirring continuously at 1000 ± 50 r/min, with the stirrer shaft set at an angle of 60° to one side of the beaker. Add the previously dried substance over a period of 45-90 s, at a uniform rate, ensuring that loose agglomerates of powder are broken up, and continue stirring at 1000 ± 50 r/min for 15 min. Remove the stirrer and place the beaker containing the dispersion in a water-bath at 25 ± 1 °C for 30 min. Insert the stirrer to a depth necessary to ensure that air is not drawn into the dispersion and, while stirring at 300 ± 25 r/min, titrate to pH 7.3-7.8 by adding a 180 g/L solution of *sodium hydroxide R* below the surface, determining the end-point potentiometrically (*2.2.20*). The total volume of the 180 g/L solution of *sodium hydroxide R* used is about 6.2 mL. Allow 2-3 min before the final pH determination. If the final pH exceeds 7.8, discard the preparation and prepare another using a smaller amount of sodium hydroxide for titration. Return the neutralised preparation to the water-bath at 25 °C for 1 h, then perform the viscosity determination without delay to avoid slight viscosity changes that occur 75 min after neutralisation. Determine the viscosity using a rotating viscometer with a spindle rotating at

20 r/min, using a spindle suitable for the expected apparent viscosity.

Carboxylic acid groups
See Assay.

Ph Eur

Carbon Dioxide

(Ph. Eur. monograph 0375)

Carbon Dioxide should be kept in approved metal cylinders which are painted grey and carry a label stating 'Carbon Dioxide'. In addition, 'Carbon Dioxide' or the symbol 'CO2' should be stencilled in paint on the shoulder of the cylinder.

CO_2 44.01 *124-38-9*

Ph Eur

DEFINITION

Content
Minimum 99.5 per cent *V/V* of CO_2 in the gaseous phase.

This monograph applies to carbon dioxide for medicinal use.

CHARACTERS

Appearance
Colourless gas.

Solubility
At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 1 volume of water.

PRODUCTION

Examine the gaseous phase.

If the test is performed on a cylinder of gas, keep the cylinder of the substance to be examined at room temperature for not less than 6 h before carrying out the tests. Keep the cylinder in the vertical position with the outlet valve uppermost.

Carbon monoxide
Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas A mixture containing 5 ppm *V/V* of *carbon monoxide R* in *nitrogen R1*.

Column:
— *material*: stainless steel,
— *size*: *l* = 2 m, Ø = 4 mm,
— *stationary phase*: an appropriate molecular sieve for chromatography (0.5 nm).

Carrier gas *helium for chromatography R.*

Flow rate 60 mL/min.

Temperature:
— *column*: 50 °C,
— *injection port and detector*: 130 °C.

Detection Flame ionisation with methaniser.

Injection Loop injector.

Adjust the injected volumes and the operating conditions so that the height of the peak due to carbon monoxide in the chromatogram obtained with the reference gas is at least 35 per cent of the full scale of the recorder.

Limit:
— *carbon monoxide*: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (5 ppm *V/V*).

Nitrogen monoxide and nitrogen dioxide
Maximum 2 ppm *V/V* in total, determined using a chemiluminescence analyser (*2.5.26*).

Gas to be examined The substance to be examined.

Reference gas (a) *Carbon dioxide R1.*

Reference gas (b) A mixture containing 2 ppm *V/V* of *nitrogen monoxide R* in *carbon dioxide R1* or in *nitrogen R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of nitrogen monoxide and nitrogen dioxide in the gas to be examined.

If nitrogen is used instead of carbon dioxide in reference gas (b), multiply the result obtained by the quenching correction factor in order to correct the quenching effect on the analyser response caused by the carbon dioxide matrix effect.

The quenching correction factor is determined by applying a known reference mixture of nitrogen monoxide in carbon dioxide and comparing the actual content with the content indicated by the analyser which has been calibrated with a NO/N_2 reference mixture.

$$\text{Quenching correction factor} = \frac{\text{actual nitrogen monoxide content}}{\text{indicated nitrogen monoxide content}}$$

Total sulfur
Maximum 1 ppm *V/V*, determined using an ultraviolet fluorescence analyser after oxidation of the sulfur compounds by heating at 1000 °C (Figure 0375.-1).

The apparatus consists of the following:
— a system generating ultraviolet radiation with a wavelength of 210 nm, made up of an ultraviolet lamp, a collimator, and a selective filter; the beam is blocked periodically by a chopper rotating at high speed,
— a reaction chamber through which flows the previously filtered gas to be examined,
— a system that detects radiation emitted at a wavelength of 350 nm, made up of a selective filter, a photomultiplier tube and an amplifier.

Gas to be examined The substance to be examined.

Reference gas (a) *Carbon dioxide R1.*

Reference gas (b) A mixture containing between 0.5 ppm *V/V* and 2 ppm *V/V* of *hydrogen sulfide R1* in *carbon dioxide R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Pass the gas to be examined through a quartz oven heated to 1000 °C. *Oxygen R* is circulated in the oven at a tenth of the flow rate of the gas to be examined. Measure the sulfur dioxide content in the gaseous mixture leaving the oven.

Water
Maximum 67 ppm *V/V*, determined using an electrolytic hygrometer (*2.5.28*).

Assay
Infrared analyser (*2.5.24*).

Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena.

Reference gas (a) *Carbon dioxide R1.*

Reference gas (b) A mixture containing 95.0 per cent *V/V* of *carbon dioxide R1* and 5.0 per cent *V/V* of *nitrogen R1*.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon dioxide in the gas to be examined.

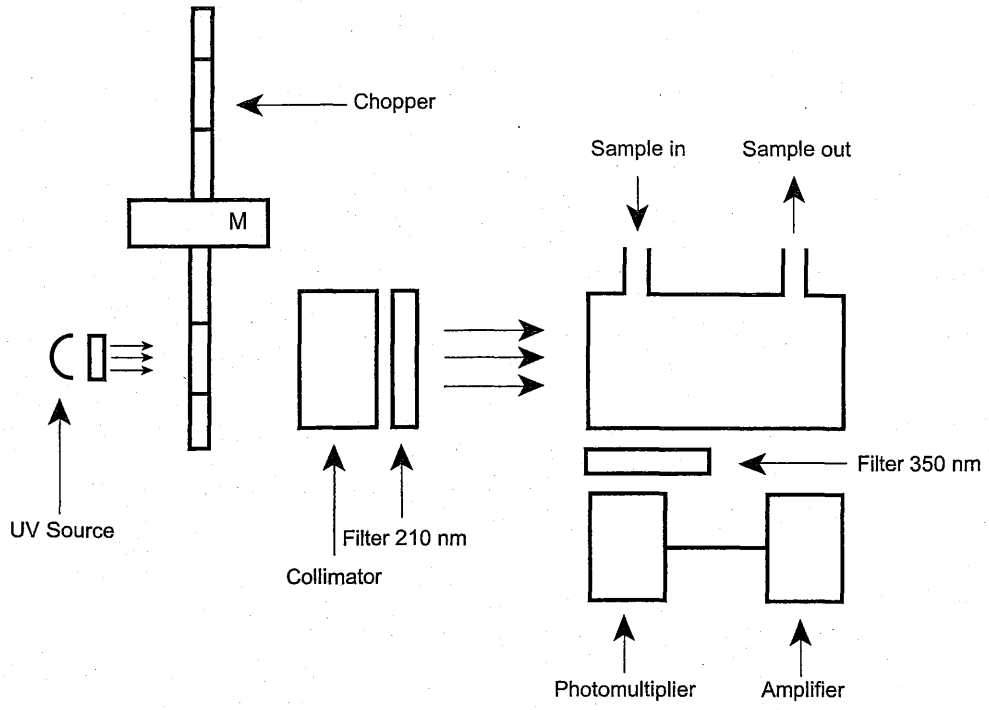


Figure 0375.-1.– *UV Fluorescence Analyser*

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison Ph. Eur. reference spectrum of carbon dioxide.

B. Place a glowing splinter of wood in an atmosphere of the substance to be examined. It is extinguished.

C. Pass a stream of the substance to be examined through *barium hydroxide solution R*. A white precipitate is formed which dissolves with effervescence in *dilute acetic acid R*.

TESTS

Examine the gaseous phase.

If the test is performed on a cylinder of gas, keep the cylinder of the substance to be examined at room temperature for not less than 6 h before carrying out the tests. Keep the cylinder in the vertical position with the outlet valve uppermost.

Carbon monoxide

Maximum 5 ppm *V/V*, determined using a carbon monoxide detector tube (*2.1.6*).

Hydrogen sulfide

Maximum 1 ppm *V/V*, determined using a hydrogen sulfide detector tube (*2.1.6*).

Nitrogen monoxide and nitrogen dioxide

Maximum 2 ppm *V/V* in total, determined using a nitrogen monoxide and nitrogen dioxide detector tube (*2.1.6*).

Sulfur dioxide

Maximum 2 ppm *V/V*, determined using a sulfur dioxide detector tube (*2.1.6*).

Water vapour

Maximum 67 ppm *V/V*, determined using a water vapour detector tube (*2.1.6*).

STORAGE

Store liquefied under pressure in suitable containers complying with the legal regulations.

IMPURITIES

A. NO: nitrogen monoxide,

B. NO_2: nitrogen dioxide,

C. CO: carbon monoxide,

D. total sulfur,

E. H_2O: water.

Ph Eur

Carbon Monoxide

(Ph. Eur. monograph 2408)

CO 28.00 *630-08-0*

Ph Eur

DEFINITION

Gas obtained by steam reforming (catalytic oxidation) of hydrocarbons.

Content

Minimum 99.5 per cent *V/V* of CO.

This monograph applies to carbon monoxide for medicinal use.

CHARACTERS

Appearance

Colourless, flammable gas.

Solubility

At 20 °C and at a pressure of 101 kPa, 2.266 volumes of carbon monoxide dissolve in 100 volumes of water.

IDENTIFICATION

Carry out either test A or B.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison Ph. Eur. reference spectrum of carbon monoxide.

B. It complies with the limits of the assay.

TESTS

Carbon dioxide

Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas A mixture containing 300 ppm *V/V* of *carbon dioxide R1* in *carbon monoxide R.*

Column:
— *material*: stainless steel;
— *size*: *l* = 2 m, Ø = 2 mm;
— *stationary phase*: an appropriate divinylbenzene porous polymer (149-177 µm).

Carrier gas *helium for chromatography R.*

Flow rate 30 mL/min.

Temperature:
— *column*: 50 °C;
— *detector*: 220 °C.

Detection Thermal conductivity.

Injection 1 mL.

Run time 3 min.

Relative retention With reference to carbon monoxide (retention time = about 0.4 min): carbon dioxide = about 3.5.

Limit:
— *carbon dioxide*: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (300 ppm *V/V*).

Methane

Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas A mixture containing 100 ppm *V/V* of *methane R* in *carbon monoxide R.*

Column:
— *material*: stainless steel;
— *size*: *l* = 2 m, Ø = 4 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (177-250 µm).

Carrier gas *nitrogen for chromatography R.*

Flow rate 10 mL/min.

Temperature:
— *column*: 95 °C;
— *detector*: 240 °C.

Detection Flame ionisation.

Injection 1 mL.

Run time 3 min.

Retention time Methane = about 1.8 min.

Limit:
— *methane*: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (100 ppm *V/V*).

Hydrogen

Gas chromatography.

Gas to be examined The substance to be examined.

Reference gas A mixture containing 300 ppm *V/V* of *hydrogen for chromatography R* in *carbon monoxide R.*

Column:
— *material*: stainless steel;
— *size*: *l* = 2 m, Ø = 2 mm;
— *stationary phase*: molecular sieve for chromatography (149-177 µm) with a nominal pore size of 0.5 nm.

Carrier gas *argon for chromatography R.*

Flow rate 30 mL/min.

Temperature:
— *column*: 100 °C;
— *detector*: 160 °C.

Detection Thermal conductivity.

Injection 1 mL.

Run time 4 min.

Relative retention With reference to carbon monoxide (retention time = about 2.3 min): hydrogen = about 0.4.

Limit:
— *hydrogen*: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (300 ppm *V/V*).

Nickel tetracarbonyl and iron pentacarbonyl

Not detectable, using a detector tube having a limit of detection of 0.1 ppm *V/V* (*2.1.6*).

Water

Maximum 10 ppm *V/V*, determined using an electrolytic hygrometer (*2.5.28*).

ASSAY

Infrared analyser (*2.5.25*).

Gas to be examined The substance to be examined, previously filtered to avoid stray light phenomena.

Reference gas (a) *Carbon monoxide R.*

Reference gas (b) A mixture containing 95.0 per cent *V/V* of *carbon monoxide R* and 5.0 per cent *V/V* of *nitrogen R1.*

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon monoxide in the gas to be examined.

STORAGE

Under pressure in suitable containers complying with the legal regulations.

IMPURITIES

Specified impurities *A, B, C, D, E, F.*

A. CO_2: carbon dioxide,

B. CH_4: methane,

C. H_2: hydrogen,

D. $Ni(CO)_4$: nickel tetracarbonyl,

E. $Fe(CO)_5$: iron pentacarbonyl,

F. H_2O: water.

Ph Eur

Carbon Monoxide Intermix (5 per cent) in Nitrogen

(Ph. Eur. monograph 2904)

Ph Eur

DEFINITION

A mixture containing 5 per cent *V/V* of *Carbon monoxide (2408)* in *Low-oxygen nitrogen (1685).*

Content

4.75 per cent *V/V* to 5.25 per cent *V/V* of carbon monoxide (CO) in nitrogen (N_2).

This monograph applies to carbon monoxide intermix (5 per cent) in nitrogen used in the preparation of lung function test gas mixtures for medicinal use.

CHARACTERS

Appearance
Colourless gas.

IDENTIFICATION

Carry out either tests A, C or tests B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of carbon monoxide.*

B. It complies with the limits of the assay.

C. Gas chromatography (*2.2.28*).

Gas to be examined The substance to be examined.

Reference gas *Nitrogen R1.*

Column:
- — *material*: stainless steel;
- — *size*: *l* = 2 m, Ø = 2 mm;
- — *stationary phase*: *molecular sieve for chromatography R* (0.5 nm).

Carrier gas *helium for chromatography R.*

Flow rate 20 mL/min.

Temperature:
- — *column*: 80 °C;
- — *detector*: 130 °C.

Detection Thermal conductivity.

Injection 10 µL.

Retention time Nitrogen = about 2 min.

Results The principal peak in the chromatogram obtained with the gas to be examined is similar in retention time to the principal peak in the chromatogram obtained with the reference gas.

TESTS

Water (*2.5.28*)
Maximum 10 ppm *V/V*.

ASSAY

Infrared analyser (*2.5.25*).

Gas to be examined The substance to be examined. It must be filtered to avoid stray-light phenomena.

Reference gas (a) Mixture containing 5.0 per cent *V/V* of *carbon monoxide R* in *nitrogen R1*.

Reference gas (b) *Nitrogen R1.*

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon monoxide in the gas to be examined.

STORAGE

As a compressed gas, in appropriate high-pressure cylinders complying with the legal regulations.

LABELLING

The label states the nominal content, in per cent *V/V*, of carbon monoxide in nitrogen.

IMPURITIES

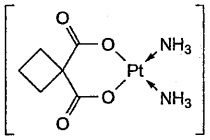
Specified impurities *A.*

A. water.

Ph Eur

Carboplatin

(Ph. Eur. monograph 1081)



$C_6H_{12}N_2O_4Pt$ 371.3 *41575-94-4*

Action and use
Platinum-containing cytotoxic.

Preparation
Carboplatin Injection

Ph Eur

DEFINITION

(*SP*-4-2)-Diammine[cyclobutan-1,1-di(carboxylato-κ*O*)(2-)] platinum.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
Colourless, crystalline powder.

Solubility
Sparingly soluble in water, very slightly soluble in acetone and in ethanol (96 per cent).

mp
About 200 °C, with decomposition.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of carboplatin.*

TESTS

Solution S
Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Impurity B and acidity
Maximum 0.5 per cent, calculated as impurity B.

To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R1*. The solution is colourless. Not more than 0.7 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R* and *water R* and dilute to 20.0 mL with the same mixture of solvents.

Reference solution Dilute 0.5 mL of the test solution to 200.0 mL with the mobile phase.

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *aminopropylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water for chromatography R, acetonitrile for chromatography R* (13:87 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Run time 2.5 times the retention time of carboplatin.

Relative retention With reference to carboplatin (retention time = about 7 min): impurity A = about 0.3.

System suitability Test solution:

— *number of theoretical plates*: minimum 5000; if necessary, adjust the concentration of acetonitrile in the mobile phase;

— *symmetry factor*: maximum 2.0; if necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (0.25 per cent);

— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with the reference solution (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with the reference solution (0.5 per cent);

— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with the reference solution (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 100 ppm.

Dissolve 0.5 g in *water R*, heating slightly if necessary, and dilute to 20 mL with the same solvent. Filter if necessary. Dilute 10 mL of this solution to 15 mL with *water R*. Prepare the standard using 5 mL of *chloride standard solution (5 ppm Cl) R*.

Ammonium (*2.4.1, Method B*)
Maximum 100 ppm, determined on 0.20 g.

Prepare the standard using 0.2 mL of *ammonium standard solution (100 ppm NH_4) R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Use the residue obtained in the test for loss on drying. Ignite 0.200 g of the residue to constant mass at 800 ± 50 °C.

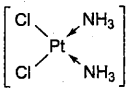
1 mg of the residue is equivalent to 1.903 mg of $C_6H_{12}N_2O_4Pt$.

STORAGE

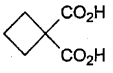
Protected from light.

IMPURITIES

Specified impurities *A, B.*



A. (*SP*-4-2)-diamminedichloridoplatinum(II) (cisplatin),

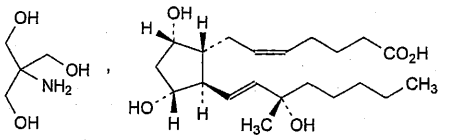


B. cyclobutane-1,1-dicarboxylic acid.

Ph Eur

Carboprost Trometamol

(Ph. Eur. monograph 1712)



$C_{25}H_{47}NO_8$ 489.7 *58551-69-2*

Action and use
Prostaglandin ($PGF_{2\alpha}$) analogue.

Ph Eur

DEFINITION

2-Amino-2-(hydroxymethyl)propane-1,3-diol (5*Z*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(1*E*,3*S*)-3-hydroxy-3-methyloct-1-enyl]cyclopentyl]hept-5-enoate ((15*S*)-15-methyl-PGF_2).

Content
94.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Soluble in water.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of carboprost trometamol.*

TESTS

Specific optical rotation (*2.2.7*)
+ 18 to + 24 (anhydrous substance).

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 15.0 mg of the substance to be examined in a mixture of 23 volumes of *acetonitrile R* and 77 volumes of *water for chromatography R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 15.0 mg of *carboprost trometamol CRS* (containing impurity A) in a mixture of 23 volumes of *acetonitrile R* and 77 volumes of *water for chromatography R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (b) Dilute 1.0 mL of reference solution (a) and 0.15 mL of *(15R)-15-methylprostaglandin $F_{2\alpha}$ R* (impurity B) to 100.0 mL with a mixture of 23 volumes of *acetonitrile R* and 77 volumes of *water for chromatography R*.

Reference solution (c) Dilute 2.0 mL of the test solution to 20.0 mL with a mixture of 23 volumes of *acetonitrile R* and 77 volumes of *water for chromatography R*. Dilute 2.0 mL of this solution to 20.0 mL with a mixture of 23 volumes of *acetonitrile R* and 77 volumes of *water for chromatography R*.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm,

— *stationary phase*: *octadecylsilyl silica gel for chromatography R1* (5 µm) with a pore size of 8-10 nm and a carbon loading of 12-19 per cent.

Mobile phase Mix 23 volumes of *acetonitrile R1* and 77 volumes of a 2.44 g/L solution of *sodium dihydrogen phosphate R* in *water for chromatography R* previously adjusted to pH 2.5 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

Run time 1.3 times the retention time of carboprost.

Relative retention With reference to carboprost (retention time = about 80 min): impurity B = about 0.85; impurity A = about 0.9.

Identification of impurities Use the chromatogram obtained with reference solution (a) and the chromatogram supplied with *carboprost trometamol CRS* to identify the peak due to impurity A.

System suitability:
- *resolution*: minimum 3.4 between the peaks due to impurity B and carboprost in the chromatogram obtained with reference solution (b);
- *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity B in the chromatogram obtained with reference solution (a).

Limits:
- *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent),
- *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent),
- *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent),
- *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (4.0 per cent),
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 50 mg.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mix 27 volumes of *acetonitrile R1* and 73 volumes of a 2.44 g/L solution of *sodium dihydrogen phosphate R* in *water for chromatography R* previously adjusted to pH 2.5 with *phosphoric acid R*.

Injection Test solution and reference solution (a).

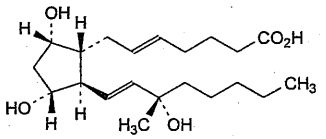
Run time 1.2 times the retention time of carboprost.

Retention time Carboprost = about 29 min.

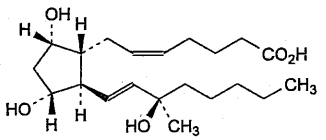
Calculate the percentage content of $C_{25}H_{47}NO_8$ using the declared content of *carboprost trometamol CRS*.

STORAGE
At a temperature below −15 ° C.

IMPURITIES
Specified impurities *A, B.*



A. (5*E*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(1*E*,3*S*)-3-hydroxy-3-methyloct-1-enyl]cyclopentyl]hept-5-enoic acid,

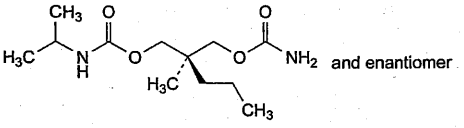


B. (5*Z*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(1*E*,3*R*)-3-hydroxy-3-methyloct-1-enyl]cyclopentyl]hept-5-enoic acid.

Ph Eur

Carisoprodol

(Ph. Eur. monograph 1689)



$C_{12}H_{24}N_2O_4$ 260.3 *78-44-4*

Action and use
Skeletal muscle relaxant.

Ph Eur

DEFINITION
(2*RS*)-2-[(Carbamoyloxy)methyl]-2-methylpentyl (1-methylethyl)carbamate.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, fine powder.

Solubility
Very slightly soluble in water, freely soluble in acetone, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 92 °C to 95 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *carisoprodol CRS.*

C. Examine the chromatograms obtained in the test for related substances.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (d).

D. Dissolve 0.2 g in 15 mL of a 28 g/L solution of *potassium hydroxide R* in *alcohol R* and boil under a reflux condenser for 15 min. Add 0.5 mL of *glacial acetic acid R* and 1 mL of a 50 g/L solution of *cobalt nitrate R* in *ethanol R*. An intense blue colour develops.

TESTS

Optical rotation (*2.2.7*)
−0.10° to + 0.10°.

Dissolve 2.5 g in *alcohol R* and dilute to 25.0 mL with the same solvent.

Related substances
Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.20 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methylene chloride R*.

Reference solution (a) Dissolve 5.0 mg of *meprobamate CRS* in *methylene chloride R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 50 mL with *methylene chloride R*.

Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with *methylene chloride R*.

Reference solution (d) Dissolve 20 mg of *carisoprodol CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (e) Dissolve the contents of a vial of *carisoprodol impurity A CRS* in 5 mL of reference solution (d) and dilute to 50 mL with *methylene chloride R*.

Plate *TLC silica gel plate R*.

Mobile phase *acetone R, methylene chloride R* (20:80 *V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air for 15 min.

Detection Spray with a solution prepared as follows: dissolve 5 g of *phosphomolybdic acid R* in a mixture of 50 mL of *glacial acetic acid R* and 10 mL of *sulfuric acid R*, and dilute to 100 mL with *glacial acetic acid R*. Heat the plate at 100-105 °C for 30 min.

System suitability:
— the chromatogram obtained with reference solution (c) shows 1 clearly visible spot;
— the chromatogram obtained with reference solution (e) shows 2 clearly separated spots.

Limits In the chromatogram obtained with test solution (a):
— *impurity D*: any spot due to impurity D is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *any other impurity*: any spot, apart from the principal spot and any spot due to impurity D, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g *in vacuo* at 60 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

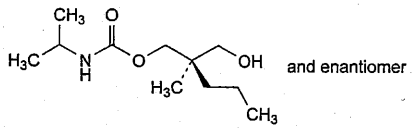
ASSAY

Dissolve 0.100 g in 15 mL of a 25 per cent *V/V* solution of *sulfuric acid R* and boil under a reflux condenser for 3 h. Cool, dissolve by cautiously adding 30 mL of *water R*, cool again and place in a steam-distillation apparatus. Add 40 mL of *strong sodium hydroxide solution R* and distil immediately by passing steam through the mixture. Collect the distillate into 40 mL of a 40 g/L solution of *boric acid R* until the total volume in the receiver reaches about 200 mL.

Add 0.25 mL of *methyl red mixed solution R*. Titrate with *0.1 M hydrochloric acid*, until the colour changes from green to violet. Carry out a blank titration.

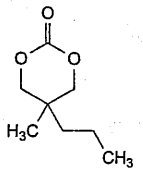
1 mL of *0.1 M hydrochloric acid* is equivalent to 13.02 mg of $C_{12}H_{24}N_2O_4$.

IMPURITIES

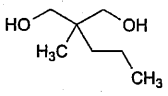


and enantiomer

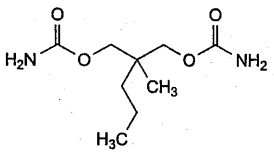
A. (2*RS*)-2-(hydroxymethyl)-2-methylpentyl (1-methylethyl)carbamate,



B. 5-methyl-5-propyl-1,3-dioxan-2-one,



C. 2-methyl-2-propylpropane-1,3-diol,



D. 2-methyl-2-propylpropane-1,3-diyl dicarbamate (meprobamate).

Ph Eur

Carmellose[1]

(Ph. Eur. monograph 2360)

9000-11-7

Action and use
Excipient; bulk laxative.

Ph Eur

DEFINITION

Carboxymethylether of cellulose.

Partly *O*-carboxymethylated cellulose.

♦CHARACTERS

Appearance
White or almost white powder, hygroscopic.

Solubility
Practically insoluble in anhydrous ethanol. It swells with water to form a suspension and becomes viscid in 1 M sodium hydroxide.♦

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

IDENTIFICATION

A. pH (*2.2.3*): 3.5 to 5.0.

Suspend 1.0 g in 100 mL of *carbon dioxide-free water R.*

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison carmellose CRS.

TESTS

Chlorides

Maximum 0.36 per cent.

Shake 0.8 g with 50 mL of *water R*, dissolve in 10 mL of *1 M sodium hydroxide* and dilute to 100 mL with *water R*. Heat on a water-bath a mixture of 10 mL of *dilute nitric acid R* and 20 mL of this solution until a flocculent precipitate is produced. Cool, centrifuge and take out the supernatant. Wash the precipitate with 3 quantities, each of 10 mL, of *water R*, centrifuging each time. Combine the supernatant and the washings and dilute to 100 mL with *water R*. To 25 mL of this solution add 6 mL of *dilute nitric acid R* and dilute to 50 mL with *water R* (test solution). Prepare the reference solution in the same manner, using 0.40 mL of *0.01 M hydrochloric acid*. Add 1 mL of *silver nitrate solution R2* to the test solution and the reference solution. Allow to stand protected from light for 5 min. Any opalescence in the test solution is not more intense than that in the reference solution.

Sulfates

Maximum 0.72 per cent.

Shake 0.40 g with 25 mL of *water R*, dissolve in 5 mL of *1 M sodium hydroxide* and add 20 mL of *water R*. Heat this solution with 2.5 mL of *hydrochloric acid R* in a water-bath until a flocculent precipitate is produced. Cool, centrifuge, and take out the supernatant. Wash the precipitate with 3 quantities, each of 10 mL, of *water R*, centrifuging each time. Combine the supernatant and the washings, and dilute to 100 mL with *water R*. Filter, and discard the first 5 mL of the filtrate. To 25 mL of the filtrate add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R* (test solution). Prepare the reference solution in the same manner, using 1.5 mL of *0.005 M sulfuric acid*. Add 2 mL of a 120 g/L solution of *barium chloride R* to the test solution and the reference solution. Mix and allow to stand for 10 min. The white turbidity produced in the test solution is not thicker than that in the reference solution.

Loss on drying (*2.2.32*)

Maximum 8.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 1.5 per cent (dried substance), determined on 1.0 g.

♦STORAGE

In an airtight container.♦

Ph Eur

Carmellose Calcium[1]

(Ph. Eur. monograph 0886)

9050-04-8

Action and use

Excipient in pharmaceutical products; bulk laxative.

Ph Eur

DEFINITION

Calcium salt of partly *O*-carboxymethylated cellulose.

Calcium salt of a polycarboxymethyl ether of cellulose.

♦CHARACTERS

Appearance

White or yellowish-white powder, hygroscopic after drying.

Solubility

Practically insoluble in acetone, in ethanol (96 per cent) and in toluene. It swells with water to form a suspension.♦

IDENTIFICATION

A. Shake 0.1 g thoroughly with 10 mL of *water R*. Add 2 mL of a 42 g/L solution of *sodium hydroxide R* and allow to stand for 10 min (solution A). Dilute 1 mL of solution A to 5 mL with *water R*. To 0.05 mL of this solution add 0.5 mL of a 0.5 g/L solution of *chromotropic acid, sodium salt R* in a 75 per cent *m/m* solution of *sulfuric acid R* and heat on a water-bath for 10 min. A reddish-violet colour develops.

B. Shake 5 mL of solution A obtained in identification test A with 10 mL of *acetone R*. A white, flocculent precipitate is produced.

C. Shake 5 mL of solution A obtained in identification test A with 1 mL of *ferric chloride solution R1*. A brown, flocculent precipitate is formed.

D. Ignite 1 g to ash and dissolve the residue in a mixture of 5 mL of *acetic acid R* and 10 mL of *water R*. Filter if necessary and heat the filtrate to boiling. Cool and neutralise with *dilute ammonia R1*. The solution gives reaction (a) of calcium (*2.3.1*).

TESTS

Solution S

Shake 1.0 g with 50 mL of *distilled water R*, add 5 mL of *dilute sodium hydroxide solution R* and dilute to 100 mL with *distilled water R*.

Alkalinity

Shake 1.0 g thoroughly with 50 mL of *carbon dioxide-free water R* and add 0.1 mL of *phenolphthalein solution R*. No red colour develops.

Chlorides (*2.4.4*)

Maximum 0.36 per cent.

Heat 28 mL of solution S with 10 mL of *dilute nitric acid R* on a water-bath until a flocculent precipitate is produced. Cool, centrifuge and separate the supernatant. Wash the precipitate with 3 quantities, each of 10 mL, of *water R*, centrifuging each time. Combine the supernatant and the washings and dilute to 100 mL with *water R*. To 25 mL add 6 mL of *dilute nitric acid R* and dilute to 50 mL with *water R*. Dilute 10 mL of the solution to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 1 per cent.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Heat 20 mL of solution S with 1 mL of *hydrochloric acid R* on a water-bath until a flocculent precipitate is produced. Cool, centrifuge and separate the supernatant. Wash the precipitate with 3 quantities, each of 10 mL, of *distilled water R*, centrifuging each time. Combine the supernatant and the washings and dilute to 100 mL with *distilled water R*. To 25 mL add 1 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *distilled water R*.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
10.0 per cent to 20.0 per cent, determined on 1.0 g of the dried substance.

♦STORAGE
In an airtight container.♦

Ph Eur

Carmellose Sodium

(Ph. Eur. monograph 0472)

9004-32-4

Action and use
Excipient; bulk laxative.

Preparation
Carmellose Sodium Eye Drops

Ph Eur

DEFINITION
Carmellose sodium (carboxymethylcellulose sodium) is the sodium salt of a partly *O*-carboxymethylated cellulose. It contains not less than 6.5 per cent and not more than 10.8 per cent of sodium (Na), calculated with reference to the dried substance.

CHARACTERS
A white or almost white, granular powder, hygroscopic after drying, practically insoluble in acetone, in ethanol and in toluene. It is easily dispersed in water giving colloidal solutions.

IDENTIFICATION
A. To 10 mL of solution S (see Tests) add 1 mL of *copper sulfate solution R*. A blue, cotton-like precipitate is formed.

B. Boil 5 mL of solution S for a few minutes. No precipitate is formed.

C. To the residue obtained in the determination of the sulfated ash, add 1 mL of *hydrochloric acid R* and evaporate on a water-bath. Take up the residue in 20 mL of *water R*. The solution gives the reactions of sodium (*2.3.1*).

TESTS
Solution S
Sprinkle a quantity of the substance to be examined equivalent to 1.0 g of the dried substance onto 90 mL of *carbon dioxide-free water R* at 40 °C to 50 °C stirring vigorously. Continue stirring until a colloidal solution is obtained, cool and dilute to 100 mL with *carbon dioxide-free water R*.

Appearance of solution
Solution S is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
The pH of solution S is 6.0 to 8.0.

Apparent viscosity
While stirring, introduce a quantity of the substance to be examined equivalent to 2.00 g of the dried substance into 50 mL of *water R* heated to 90 °C. For a product of low viscosity, use if necessary, the quantity required to give the concentration indicated on the label. Allow to cool, dilute to 100.0 mL with *water R* and stir until dissolution is complete. Determine the viscosity (*2.2.10*) using a rotating viscometer at 20 °C and a shear rate of 10 s^{-1}. If it is impossible to obtain a shear rate of exactly 10 s^{-1}, use a shear rate slightly higher and a rate slightly lower and interpolate. The apparent viscosity is not less than 75 per cent and not more than 140 per cent of the value stated on the label.

Sodium glycollate
Place a quantity of the substance to be examined equivalent to 0.500 g of dried substance in a beaker. Add 5 mL of *acetic acid R* and 5 mL of *water R*. Stir until dissolution is complete (about 30 min). Add 80 mL of *acetone R* and 2 g of *sodium chloride R*. Filter through a fast filter paper impregnated with *acetone R* into a volumetric flask, rinse the beaker and filter with *acetone R* and dilute the filtrate to 100.0 mL with the same solvent. Allow to stand for 24 h without shaking. Use the clear supernatant to prepare the test solution.

In a volumetric flask, dissolve 0.310 g of *glycollic acid R*, previously dried *in vacuo* over *diphosphorus pentoxide R*, in *water R* and dilute to 1000.0 mL with the same solvent. Place 5.0 mL of this solution in a volumetric flask, add 5 mL of *acetic acid R* and allow to stand for about 30 min. Add 80 mL of *acetone R* and 2 g of *sodium chloride R* and dilute to 100.0 mL with *acetone R*. Use this solution to prepare the reference solution.

Place 2.0 mL of each solution in a separate 25 mL volumetric flask. Heat on a water-bath to eliminate acetone. Cool to room temperature and add 5.0 mL of *2,7-dihydroxynaphthalene solution R* to each flask. Shake and add 15.0 mL of *2,7-dihydroxynaphthalene solution R*. Close the flasks with aluminium foil and heat on a water-bath for 20 min. Cool under running water and dilute to 25.0 mL with *sulfuric acid R*. Within 10 min, transfer 10.0 mL of each solution to a flat-bottomed tube. Examine the solutions viewing vertically. The test solution is not more intensely coloured than the reference solution (0.4 per cent).

Chlorides (*2.4.4*)
Dilute 2 mL of solution S to 15 mL with *water R*. The solution complies with the limit test for chlorides (0.25 per cent).

Loss on drying (*2.2.32*)
Not more than 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
20.0 per cent to 33.3 per cent, determined on 1.0 g using a mixture of equal volumes of *sulfuric acid R* and *water R* and calculated with reference to the dried substance. These limits correspond to a content of 6.5 per cent to 10.8 per cent of sodium (Na).

LABELLING
The label states the apparent viscosity in millipascal seconds for a 20 g/L solution; for a product of low viscosity, the label states the concentration of the solution to be used and the apparent viscosity in millipascal seconds.

Ph Eur

Low-substituted Carmellose Sodium

(Ph. Eur. monograph 1186)

9050-32-4

Action and use
Excipient in pharmaceutical products; bulk laxative.

Ph Eur

DEFINITION

Low-substituted sodium carboxymethylcellulose. Sodium salt of a partly *O*-(carboxymethylated) cellulose.

Content
2.0 per cent to 4.5 per cent of sodium (Na) (dried substance).

CHARACTERS

Appearance
White or almost white powder or short fibres.

Solubility
Practically insoluble in acetone, in anhydrous ethanol and in toluene. It swells in water to form a gel.

IDENTIFICATION

A. Shake 1 g with 100 mL of a 100 g/L solution of *sodium hydroxide R*. A suspension is produced.

B. Shake 1 g with 50 mL of *water R*. Transfer 1 mL of the mixture to a test tube, add 1 mL of *water R* and 0.05 mL of a freshly prepared 40 g/L solution of *α-naphthol R* in *methanol R*. Incline the test tube and add carefully 2 mL of *sulfuric acid R* down the side so that it forms a lower layer. A reddish-purple colour develops at the interface.

C. Sulfated ash (*2.4.14*) (see Tests).

D. To the residue obtained in the determination of the sulfated ash, add 1 mL of *hydrochloric acid R* and evaporate on a water-bath. Take up the residue in 20 mL of *water R*. The solution gives reaction (a) of sodium (*2.3.1*).

TESTS

pH (*2.2.3*)
6.0 to 8.5.

Shake 1 g with 100 mL of *carbon dioxide-free water R* for 5 min. Centrifuge.

Sodium chloride and sodium glycollate
Maximum 0.5 per cent (dried substance) for the sum of the percentage contents.

Sodium chloride Place 5.00 g in a 250 mL conical flask, add 50 mL of *water R* and 5 mL of *strong hydrogen peroxide solution R* and heat on a water bath for 20 min, stirring occasionally to ensure total hydration. Cool, add 100 mL of *water R* and 10 mL of *nitric acid R*. Titrate with *0.05 M silver nitrate* determining the end-point potentiometrically (*2.2.20*) using a silver-based indicator electrode and a double-junction reference electrode containing a 100 g/L solution of *potassium nitrate R* in the outer jacket and a standard filling solution in the inner jacket.

1 mL of *0.05 M silver nitrate* is equivalent to 2.922 mg of NaCl.

Sodium glycollate Place a quantity of the substance to be examined equivalent to 0.500 g of the dried substance in a beaker. Add 5 mL of *glacial acetic acid R* and 5 mL of *water R* and stir to ensure total hydration (about 30 min). Add 80 mL of *acetone R* and 2 g of *sodium chloride R*. Stir for several minutes to ensure complete precipitation of the carboxymethylcellulose. Filter through a fast filter paper impregnated with *acetone R* into a volumetric flask, rinse the beaker and filter with *acetone R* and dilute the filtrate to 100.0 mL with the same solvent. Allow to stand for 24 h without shaking. Use the clear supernatant as the test solution.

Prepare the reference solutions as follows: in a 100 mL volumetric flask, dissolve 0.100 g of *glycollic acid R*, previously dried *in vacuo* over *diphosphorus pentoxide R*, in *water R* and dilute to 100.0 mL with the same solvent. Transfer 0.5 mL, 1.0 mL, 1.5 mL and 2.0 mL of the solution to separate volumetric flasks; dilute the contents of each flask to 5.0 mL with *water R*, add 5 mL of *glacial acetic acid R*, dilute to 100.0 mL with *acetone R* and mix.

Transfer 2.0 mL of the test solution and 2.0 mL of each of the reference solutions to separate 25 mL volumetric flasks. Heat the uncovered flasks in a water-bath to eliminate the acetone. Allow to cool and add 5.0 mL of *2,7-dihydroxynaphthalene solution R* to each flask. Mix, add a further 15.0 mL of *2,7-dihydroxynaphthalene solution R* and mix again. Close the flasks with aluminium foil and heat in a water-bath for 20 min. Cool and dilute to 25.0 mL with *sulfuric acid R*.

Measure the absorbance (*2.2.25*) of each solution at 540 nm. Prepare a blank using 2.0 mL of a solution containing 5 per cent *V/V* each of *glacial acetic acid R* and *water R* in *acetone R*. Prepare a standard curve using the absorbances obtained with the reference solutions. From the standard curve and the absorbance of the test solution, determine the mass *a*, in milligrams, of glycollic acid in the substance to be examined and calculate the content of sodium glycollate from the following expression:

$$\frac{10 \times 1.29 \times a}{(100 - b)m}$$

1.29 = the factor converting glycollic acid to sodium glycollate,
b = the loss on drying as a percentage,
m = the mass of the substance to be examined, in grams.

Water-soluble substances
Maximum 70.0 per cent.

Disperse 5.00 g in 400.0 mL of *water R* and stir for 1 min every 10 min during the first 30 min. Allow to stand for 1 h and centrifuge, if necessary. Decant 100.0 mL of the supernatant onto a fast filter paper in a vacuum filtration funnel, apply vacuum and collect 75.0 mL of the filtrate. Evaporate to dryness and dry the residue at 100-105 °C for 4 h.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
6.5 per cent to 13.5 per cent (dried substance), corresponding to a content of 2.0 per cent to 4.5 per cent of Na.

Use 1.0 g with a mixture of equal volumes of *sulfuric acid R* and *water R*.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the

mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for low-substituted carmellose sodium used as disintegrant.

Settling volume

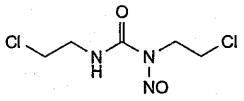
15.0 mL to 35.0 mL.

In a 100 mL graduated cylinder, place 20 mL of *2-propanol R*, add 5.0 g of the substance to be examined and shake vigorously. Dilute to 30 mL with *2-propanol R* then to 50 mL with *water R* and shake vigorously. Within 15 min, repeat the shaking 3 times. Allow to stand for 4 h and determine the volume of the settled mass.

Ph Eur

Carmustine

(Ph. Eur. monograph 1187)



$C_5H_9Cl_2N_3O_2$ 214.1 *154-93-8*

Action and use

Cytotoxic alkylating agent.

Ph Eur

DEFINITION

Carmustine contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of 1,3-bis(2-chloroethyl)-1-nitrosourea, calculated with reference to the anhydrous substance.

CHARACTERS

A yellowish, granular powder, very slightly soluble in water, very soluble in methylene chloride, freely soluble in ethanol.

It melts at about 31 °C with decomposition.

IDENTIFICATION

Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the *Ph. Eur. reference spectrum of carmustine*. Examine the melted substances prepared as films.

TESTS

1,3-Bis(2-chloroethyl)urea (impurity A)

Examine by thin-layer chromatography (*2.2.27*), using a suitable silica gel as the coating substance.

Test solution Dissolve 0.10 g of the substance to be examined in *methylene chloride R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of *carmustine impurity A CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of the test solution to 10 mL with *methylene chloride R*. To 5 mL of this solution, add 5 mL of reference solution (a).

Apply separately to the plate 2 µL of each solution. Develop over a path of 10 cm using a mixture of 10 volumes of *methanol R* and 90 volumes of *methylene chloride R*. Allow the plate to dry in air. Spray with *diethylamine R* and heat at 125 °C for 10 min. Allow to cool and spray with *silver nitrate solution R2*. Expose to ultraviolet light at 365 nm until brown to black spots appear. Any spot corresponding to carmustine impurity A in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with reference solution (a) (1 per cent). The test is not valid unless the chromatogram obtained with reference solution (b) shows two clearly separated spots.

Water (*2.5.12*)

Not more than 1.0 per cent, determined on 0.50 g by the semi-micro determination of water.

ASSAY

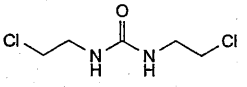
Dissolve 0.100 g in 30 mL of *ethanol R* and dilute to 100.0 mL with *water R*. Dilute 3.0 mL of the solution to 100.0 mL with *water R*. Measure the absorbance (*2.2.25*) at the maximum at 230 nm.

Calculate the content of $C_5H_9Cl_2N_3O_2$ taking the specific absorbance to be 270.

STORAGE

Store in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

IMPURITIES



A. 1,3-bis(2-chloroethyl)urea.

Ph Eur

Carnauba Wax

(Ph. Eur. monograph 0597)

8015-86-9

Action and use

Excipient.

Ph Eur

DEFINITION

Purified wax obtained from the leaves of *Copernicia cerifera* Mart.

CHARACTERS

Appearance

Pale yellow or yellow powder, flakes or hard masses.

Solubility

Practically insoluble in water, soluble on heating in ethyl acetate and in xylene, practically insoluble in ethanol (96 per cent).

Relative density

About 0.97.

IDENTIFICATION

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined with heating in 5 mL of *chloroform R*. Use the warm solution.

Reference solution Dissolve 5 mg of *menthol R*, 5 µL of *menthyl acetate R* and 5 mg of *thymol R* in 10 mL of *toluene R*.

Plate *TLC silica gel plate R.*

Mobile phase *ethyl acetate R, chloroform R* (2:98 *V/V*).

Application 30 µL of the test solution and 10 µL of the reference solution as bands 20 mm by 3 mm.

Development Over 1/2 of the plate.

Drying In air.

Detection Spray with a freshly prepared 200 g/L solution of *phosphomolybdic acid R* in *ethanol (96 per cent) R* (about 10 mL for a 20 cm plate). Heat at 100-105 °C for 10-15 min.

Results The chromatogram obtained with the reference solution shows in the lower part a dark blue zone (menthol), above this zone a reddish zone (thymol) and in the upper part a dark blue zone (menthyl acetate). The chromatogram obtained with the test solution shows a large blue zone (triacontanol = melissyl alcohol) at a level between the thymol and menthol zones in the chromatogram obtained with the reference solution. Further blue zones are visible in the upper part of the chromatogram obtained with the test solution, at levels between those of the menthyl acetate and thymol zones in the chromatogram obtained with the reference solution; above these zones, further zones are visible in the chromatogram obtained with the test solution; the zone with the highest R_F value is very pronounced. A number of faint zones are visible below the triacontanol zone and the point of application is coloured blue.

TESTS

Melting point (*2.2.15*)
80 °C to 88 °C.

Melt the substance to be examined carefully on a water-bath before introduction into the capillary tubes. Allow the tubes to stand in the refrigerator for 24 h or at 0 °C for 2 h.

Acid value
2 to 7.

To 2.000 g (*m* g) in a 250 mL conical flask fitted with a reflux condenser add 40 mL of *xylene R* and a few glass beads. Heat with stirring until the substance is completely dissolved. Add 20 mL of *ethanol (96 per cent) R* and 1 mL of *bromothymol blue solution R3* and titrate the hot solution with 0.5 M alcoholic potassium hydroxide until a green colour persisting for at least 10 s is obtained (n_1 mL). Carry out a blank test (n_2 mL). Calculate the acid value using the following expression:

$$\frac{28.05(n_1 - n_2)}{m}$$

Saponification value
78 to 95.

To 2.000 g (*m* g) in a 250 mL conical flask fitted with a reflux condenser add 40 mL of *xylene R* and a few glass beads. Heat with stirring until the substance is completely dissolved. Add 20 mL of *ethanol (96 per cent) R* and 20.0 mL of 0.5 M alcoholic potassium hydroxide. Boil under a reflux condenser for 3 h. Add 1 mL of *phenolphthalein solution R1* and titrate the hot solution immediately with *0.5 M hydrochloric acid* until the red colour disappears. Repeat the heating and titration until the colour no longer reappears on heating (n_3 mL). Carry out a blank test (n_4 mL). Calculate the saponification value using the following expression:

$$\frac{28.05(n_4 - n_3)}{m}$$

Total ash (*2.4.16*)
Maximum 0.25 per cent, determined on 2.0 g.

STORAGE

Protected from light.

Ph Eur

Carrageenan

(Ph. Eur. monograph 2138)

Ph Eur

DEFINITION

Polysaccharides extracted from different Rhodophyceae with boiling water or aqueous alkali solutions. Carrageenan is separated by alcohol precipitation, potassium chloride precipitation, gel pressing, drum drying or freezing.
The alcohol used during separation and purification is generally 2-propanol. The main components are potassium, sodium, calcium or magnesium salts of the sulfate esters of D-galactose and 3,6-anhydro-D-galactose copolymers. They exist in different proportions depending on the biological origin of the polymer.

The prevalent copolymers are designated as κ-, ι- and λ-carrageenan.

CHARACTERS

Appearance
Yellowish, brownish, or white or almost white powder.

Solubility
Soluble in water giving a viscous or colloidal solution, insoluble in organic solvents.

IDENTIFICATION

A. Prepare a 20 g/L dispersion and heat in a water-bath at 80 °C. Mix 1 volume of this solution and about 4 volumes of *water R* and add 2-3 drops of a 0.5 g/L solution of *methylene blue R* in *ethanol (96 per cent) R*. A blue precipitate is formed.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Prepare a 2 g/L solution of the substance to be examined; cast 4.5 mL of the solution into a plastic flat-bottomed weighing boat about 35 mm in diameter and allow to dry completely in an air-flow oven at 60 °C until a film, about 10 µm thick, is obtained (about 4 h).

Carrageenan has strong, broad absorption bands, typical of all polysaccharides, in the 1000-1100 cm^{-1} region. Absorption maxima are 1065 cm^{-1} and 1020 cm^{-1} for gelling and non-gelling types, respectively. Other characteristic absorption bands and their intensities relative to the absorbance at 1050 cm^{-1} are shown in Table 2138.-1.

Table 2138.-1 shows absorbance ratios corresponding to copolymer types isolated from carrageenans. Native extracts of carrageenan from most cold-water seaweed species comprise κ-, ι- and λ-structures from the natural mix of haploid and diploid plants. Such extracts exhibit, in practice, the characteristic absorbance peaks of κ-, ι- and λ-structures and yield absorbance ratios somewhere between the above-mentioned individual ranges.

Table 2138.-1. – *Characteristic absorption bands for carrageenan identification by infrared absorption spectrophotometry*

Wavenumber (cm^{-1})	Molecular structure	Absorbance relative to the absorbance at 1050 cm^{-1}		
		κ	ι	λ
1220 - 1260	Ester sulfate	0.7 - 1.2	1.2 -2.0	1.4 - 2.0
928 - 933	3,6-Anhydro-D-galactose	0.3 - 0.6	0.2 - 0.5	≤ 0.2
840 - 850	Galactose-4-sulfate	0.3 - 0.5	0.2 - 0.4	-
825 - 830	Galactose-2-sulfate	-	-	0.2 - 0.4
810 - 820	Galactose-6-sulfate	-	-	0.1 - 0.3
800 - 805	3,6-Anhydro-D-galactose-2-sulfate	≤ 0.2	0.2 - 0.4	-

TESTS

Viscosity (*2.2.10*)
Minimum 5 mPa·s. Heat a 15 g/L dispersion (dried substance) at 80 °C for at least 15 min to dissolve. Compensate for any loss of water by evaporation, allow to cool to 75 °C and carry out the test at this temperature.

Arsenic (*2.4.27*)
Maximum 3.0 ppm.

Cadmium (*2.4.27*)
Maximum 2.0 ppm.

Lead (*2.4.27*)
Maximum 5.0 ppm.

Mercury (*2.4.27*)
Maximum 1.0 ppm.

Loss on drying (*2.2.32*)
Maximum 12.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Total ash (*2.4.16*)
Maximum 40.0 per cent.

Ash insoluble in hydrochloric acid (*2.8.1*)
Maximum 2.0 per cent.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for carrageenan used as viscosity-increasing agent.

Gel formation
Prepare a 20 g/L dispersion and heat in a water-bath at 80 °C (solution A). Allow to cool; it becomes more viscous upon cooling and may form a gel.

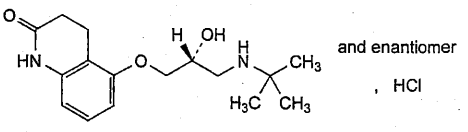
To 10 mL of solution A, while still hot, add 4 drops of a 100 g/L solution of *potassium chloride R*, mix and allow to cool. A 'brittle' gel indicates a carrageenan of a predominantly κ-type; an 'elastic' gel indicates a predominantly ι-type; if the solution does not form a gel, the carrageenan is of a predominantly λ-type; a weak or pourable gel obtained on cooling indicates a carrageenan comprising a mixture of κ- and λ-types, which can be confirmed by infrared absorption spectrophotometry.

Viscosity
(see Tests).

Ph Eur

Carteolol Hydrochloride

(Ph. Eur. monograph 1972)



$C_{16}H_{25}N_2O_3Cl$ 328.8 *51781-21-6*

Action and use
Beta-adrenoceptor antagonist.

Preparation
Carteolol Eye Drops

Ph Eur

DEFINITION
5-[(2*RS*)-3-[(1,1-Dimethylethyl)amino]-2-hydroxypropoxy]-3,4-dihydroquinolin-2(1*H*)-one hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white crystals or crystalline powder.

Solubility
Soluble in water, sparingly soluble in methanol, slightly soluble in ethanol 96 per cent, practically insoluble in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison Ph. Eur. reference spectrum of carteolol hydrochloride.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.300 g in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)
5.0 to 6.0.

Dissolve 0.250 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 10 mg of *carteolol for system suitability CRS* in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (d) Dilute 5.0 mL of reference solution (b) to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 1 volume of *methanol R2*, 20 volumes of *acetonitrile R* and 79 volumes of a 2.82 g/L solution of *sodium hexanesulfonate R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 252 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *carteolol for system suitability CRS* to identify the peak due to impurity H.

System suitability:
— the chromatogram obtained with reference solution (c) is similar to the chromatogram provided with *carteolol for system suitability CRS*; the peaks due to impurity H and carteolol show base-line separation;
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (d);
— *number of theoretical plates*: minimum 6000, calculated for the principal peak in the chromatogram obtained with reference solution (a).

Limits:
— *impurity H*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than half the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 60 mL of *ethanol (96 per cent) R*. Add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 32.88 mg of $C_{16}H_{25}N_2O_3Cl$.

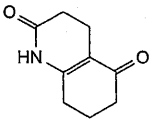
STORAGE

In an airtight container.

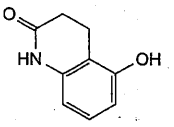
IMPURITIES

Specified impurities *H.*

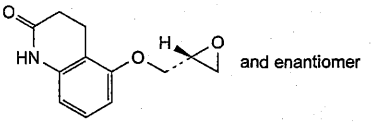
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, I.*



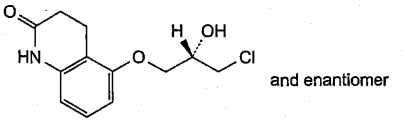
A. 4,6,7,8-tetrahydroquinoline-2,5(1*H*,3*H*)-dione,



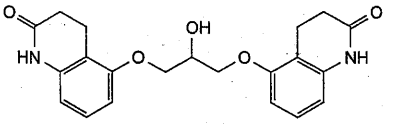
B. 5-hydroxy-3,4-dihydroquinolin-2(1*H*)-one,



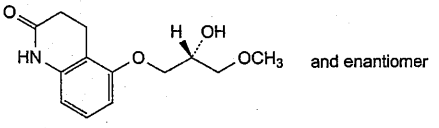
C. 5-[[(2*RS*)-oxiran-2-yl]methoxy]-3,4-dihydroquinolin-2(1*H*)-one,



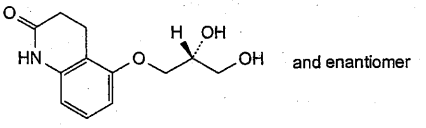
D. 5-[(2*RS*)-3-chloro-2-hydroxypropoxy]-3,4-dihydroquinolin-2(1*H*)-one,



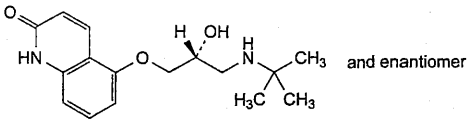
E. 5,5′-[(2-hydroxypropan-1,3-diyl)bis(oxy)]bis(3,4-dihydroquinolin-2(1*H*)-one),



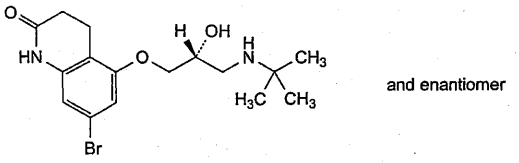
F. 5-[(2*RS*)-2-hydroxy-3-methoxypropoxy]-3,4-dihydroquinolin-2(1*H*)-one,



G. 5-[(2*RS*)-2,3-dihydroxypropoxy]-3,4-dihydroquinolin-2(1*H*)-one,



H. 5-[(2*RS*)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]quinolin-2(1*H*)-one,

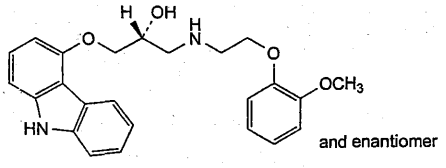


I. 7-bromo-5-[(2*RS*)-3-[(1,1-(dimethylethyl)amino]-2-hydroxypropoxy]-3,4-dihydroquinolin-2(1*H*)-one.

Ph Eur

Carvedilol

(Ph. Eur. monograph 1745)



$C_{24}H_{26}N_2O_4$ 406.5 *72956-09-3*

Action and use
Beta-adrenoceptor antagonist; arteriolar vasodilator.

Preparation
Carvedilol Tablets

Ph Eur

DEFINITION
(2*RS*)-1-(9*H*-Carbazol-4-yloxy)-3-[[2-(2-methoxyphenoxy)ethyl]amino]propan-2-ol.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent). It is practically insoluble in dilute acids.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *carvedilol CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *2-propanol R*, evaporate to dryness and record new spectra using the residues.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *carvedilol impurity C CRS* in 5.0 mL of the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 4.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *carvedilol for system suitability CRS* (containing impurities A and D) in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.150 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 55 °C.

Mobile phase Dissolve 1.77 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 650 mL with the same solvent; adjust to pH 2.0 with *phosphoric acid R* and add 350 mL of *acetonitrile R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL.

Run time 6 times the retention time of carvedilol.

Identification of impurities Use the chromatogram supplied with *carvedilol for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and D; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to carvedilol (retention time = about 4 min): impurity A = about 0.5; impurity C = about 2.9; impurity D = about 3.8.

System suitability:
— *resolution*: minimum 3.5 between the peaks due to impurity A and carvedilol in the chromatogram obtained with reference solution (c);
— *signal-to-noise ratio*: minimum 10 for the peak due to impurity C in the chromatogram obtained with reference solution (b).

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 2.0;
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity D*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *impurity C*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.02 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than C*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

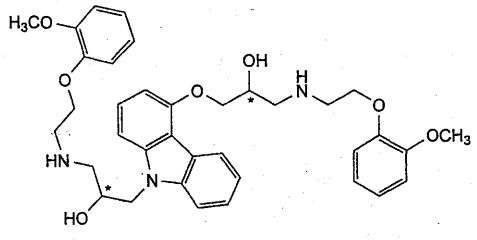
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.350 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

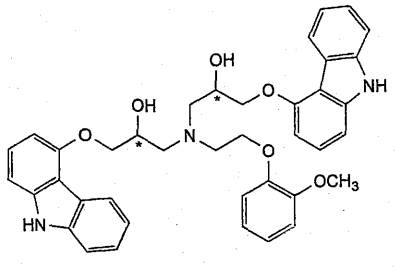
1 mL of *0.1 M perchloric acid* is equivalent to 40.65 mg of $C_{24}H_{26}N_2O_4$.

IMPURITIES
Specified impurities *A, C, D.*

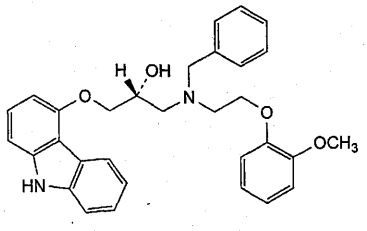
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. 1-[[9-[2-hydroxy-3-[[2-(2-methoxyphenoxy)ethyl]amino]propyl]-9*H*-carbazol-4-yl]oxy]-3-[[2-(2-methoxyphenoxy)ethyl]amino]propan-2-ol,

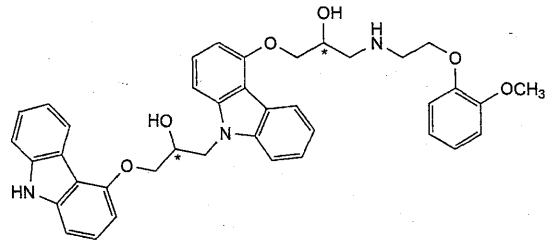


B. 1,1′-[[2-(2-methoxyphenoxy)ethyl]nitrilo]bis[3-(9*H*-carbazol-4-yloxy)propan-2-ol],



and enantiomer

C. (2*RS*)-1-[benzyl[2-(2-methoxyphenoxy)ethyl]amino]-3-(9*H*-carbazol-4-yloxy)propan-2-ol,



D. 1-(9*H*-carbazol-4-yloxy)-3-[4-[2-hydroxy-3-[[2-(2-methoxyphenoxy)ethyl]amino]propoxy]-9*H*-carbazol-9-yl]propan-2-ol.

Ph Eur

Hydrogenated Castor Oil

(Ph. Eur. monograph 1497)

Action and use
Excipient.

Ph Eur

DEFINITION
Fatty oil obtained by hydrogenation of *Virgin Castor oil (0051)*. It consists mainly of the triglyceride of 12-hydroxystearic (12-hydroxyoctadecanoic) acid.

CHARACTERS
Appearance
Fine, almost white or pale yellow powder or almost white or pale yellow masses or flakes.

Solubility
Practically insoluble in water, slightly soluble in methylene chloride, very slightly soluble in anhydrous ethanol, practically insoluble in light petroleum.

IDENTIFICATION
A. Melting point (*2.2.14*): 83 °C to 88 °C.

B. Hydroxyl value (see Tests).

C. Composition of fatty acids (see Tests).

TESTS
Acid value (*2.5.1*)
Maximum 4.0, determined on 10.0 g dissolved in 75 mL of hot *ethanol (96 per cent) R*.

Hydroxyl value (*2.5.3, Method A*)
145 to 165, determined on a warm solution.

Iodine value (*2.5.4, Method A*)
Maximum 5.0.

Alkaline impurities
Dissolve 1.0 g by gentle heating in a mixture of 1.5 mL of *ethanol (96 per cent) R* and 3 mL of *toluene R*. Add 0.05 mL of a 0.4 g/L solution of *bromophenol blue R* in *ethanol (96 per cent) R*. Not more than 0.2 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to yellow.

Composition of fatty acids (*2.4.22*)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Test solution Introduce 75 mg of the substance to be examined into a 10 mL centrifuge tube with a screw cap. Dissolve in 2 mL of *1,1-dimethylethyl methyl ether R1* by shaking and heat gently (50-60 °C). Add, when still warm, 1 mL of a 12 g/L solution of *sodium R* in *anhydrous methanol R*, prepared with the necessary precautions, and mix

vigorously for at least 5 min. Add 5 mL of *distilled water R* and mix vigorously for about 30 s. Centrifuge for 15 min at 1500 *g*. Use the upper layer.

Reference solution Dissolve 50 mg of *methyl 12-hydroxystearate CRS* and 50 mg of *methyl stearate CRS* in 10.0 mL of *1,1-dimethylethyl methyl ether R1*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m; Ø = 0.25 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Flow rate 0.9 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 55	215
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Calculate the fraction of each fatty-acid using the following expression:

$$A_{x,s,c} / \sum A_{x,s,c} \times 100 \text{ per cent } m/m$$

$A_{x,s,c}$ = corrected peak area of the fatty acid in the test solution:

$$A_{x,s,c} = A_{x,s} \times R_c$$

R_c = relative correction factor for the peak due to methyl 12-hydroxystearate:

$$R_c = \frac{m_{1,r} \times A_{2,r}}{A_{1,r} \times m_{2,r}}$$

R_c = 1 for peaks corresponding to each of the other specified fatty acids or any unspecified fatty acid;
$m_{1,r}$ = mass of methyl 12-hydroxystearate in the reference solution;
$m_{2,r}$ = mass of methyl stearate in the reference solution;
$A_{1,r}$ = area of any peak due to methyl 12-hydroxystearate in the chromatogram obtained with the reference solution;
$A_{2,r}$ = area of any peak due to methyl stearate in the chromatogram obtained with the reference solution;
$A_{x,s}$ = area of the peaks due to any specified or unspecified fatty acid methyl esters.

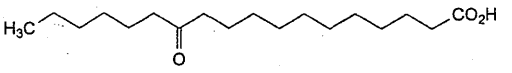
Composition of the fatty acid fraction of the oil:
— *palmitic acid*: not more than 2.0 per cent;
— *stearic acid*: 7.0 per cent to 14.0 per cent;
— *arachidic acid*: not more than 1.0 per cent;
— *12-oxostearic acid*: not more than 5.0 per cent;
— *12-hydroxystearic acid*: 78.0 per cent to 91.0 per cent;
— *any other fatty acid*: not more than 3.0 per cent.

Nickel (*2.4.31*)
Maximum 1 ppm.

STORAGE

In a well-filled container.

IMPURITIES



A. 12-oxostearic acid.

Ph Eur

Polyoxyl Castor Oil

(Macrogolglycerol Ricinoleate, Ph. Eur. monograph 1082)

Action and use
Excipient.

Ph Eur

DEFINITION

Contains mainly ricinoleyl glycerol ethoxylated with 30-50 molecules of ethylene oxide (nominal value), with small amounts of macrogol ricinoleate and of the corresponding free glycols. It results from the reaction of castor oil with ethylene oxide.

CHARACTERS

Appearance
Clear, yellow viscous liquid or semi-solid.

Solubility
Freely soluble in water, very soluble in methylene chloride, freely soluble in ethanol (96 per cent).

Relative density
About 1.05.

Viscosity
500 mPa·s to 800 mPa·s at 25 °C.

IDENTIFICATION

A. Iodine value (see Tests).

B. Saponification value (see Tests).

C. Thin-layer chromatography (*2.2.27*).

Test solution To 1 g of the substance to be examined add 100 mL of a 100 g/L solution of *potassium hydroxide R* and boil under a reflux condenser for 30 min. Allow to cool. Acidify the solution with 20 mL of *hydrochloric acid R*. Shake the mixture with 50 mL of *ether R* and allow to stand until separation of the layers is obtained. Transfer the clear upper layer to a suitable tube, add 5 g of *anhydrous sodium sulfate R*, close the tube and allow to stand for 30 min. Filter and evaporate the filtrate to dryness on a water-bath. Dissolve 50 mg of the residue in 25 mL of *ether R*.

Reference solution Dissolve 50 mg of *ricinoleic acid R* in *methylene chloride R* and dilute to 25 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R*.

Mobile phase *methylene chloride R, glacial acetic acid R, acetone R* (10:40:50 *V/V/V*).

Application 2 µL.

Development Over a path of 8 cm.

Drying In a current of cold air.

Detection Spray with an 80 g/L solution of *phosphomolybdic acid R* in *2-propanol R* and heat at 120 °C for 1-2 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the

principal spot in the chromatogram obtained with the reference solution.

D. Place about 2 g of the substance to be examined in a test-tube and add 0.2 mL of *sulfuric acid R*. Close the tube using a stopper fitted with a glass tube bent twice at right angles. Heat the tube until white fumes appear. Collect the fumes in 1 mL of *mercuric chloride solution R*. A white precipitate is formed and the fumes turn a filter paper impregnated with *alkaline potassium tetraiodomercurate solution R* black.

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*). If intended for use in the manufacture of parenteral preparations, solution S is not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of *water R* and 10 mL of *ethanol (96 per cent) R*. Add 0.1 mL of *bromothymol blue solution R1*. Not more than 0.5 mL of *0.1 M hydrochloric acid* is required to change the colour of the indicator to yellow.

Acid value (*2.5.1*)
Maximum 2.0, determined on 5.0 g.

Hydroxyl value (*2.5.3, Method A*)
See Table 1082.-1.

Iodine value (*2.5.4*)
25 to 35.

Saponification value (*2.5.6*)
See Table 1082.-1.

Table 1082.-1

Ethylene oxide units per molecule (nominal value)	Hydroxyl value	Saponification value
30 - 35	65 - 82	60 - 75
50	48 - 68	38 - 52

Residual ethylene oxide and dioxan (*2.4.25*)
Maximum 1 ppm of residual ethylene oxide and 10 ppm of residual dioxan.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 2.000 g.

Total ash (*2.4.16*)
Maximum 0.3 per cent, determined on 2.0 g.

STORAGE

Protected from light.

LABELLING

The label states:
— the amount of ethylene oxide reacted with castor oil (nominal value),
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Hydrogenated Polyoxyl Castor Oil

(Macrogolglycerol Hydroxystearate, Ph. Eur. monograph 1083)

Ph Eur

DEFINITION

Contains mainly tris(12-hydroxystearyl) glycerol ethoxylated with 7 to 60 molecules of ethylene oxide (nominal value), with small amounts of macrogol hydroxystearate and of the corresponding free glycols. It results from the reaction of hydrogenated castor oil with ethylene oxide.

CHARACTERS

Appearance

— if less than 10 units of ethylene oxide per molecule: yellowish, turbid, viscous liquid;
— if more than 20 units of ethylene oxide per molecule: white or yellowish semi-liquid or pasty mass.

Solubility

— if less than 10 units of ethylene oxide per molecule: practically insoluble in water, soluble in acetone, dispersible in ethanol (96 per cent);
— if more than 20 units of ethylene oxide per molecule: freely soluble in water, in acetone and in ethanol (96 per cent), practically insoluble in light petroleum.

IDENTIFICATION

A. Iodine value (see Tests).

B. Saponification value (see Tests).

C. Thin-layer chromatography (*2.2.27*).

Test solution To 1 g of the substance to be examined, add 100 mL of a 100 g/L solution of *potassium hydroxide R* and boil under a reflux condenser for 30 min. Allow to cool. Acidify the solution with 20 mL of *hydrochloric acid R*. Shake the mixture with 50 mL of *ether R* and allow to stand until separation of the layers is obtained. Transfer the clear upper layer to a suitable tube, add 5 g of *anhydrous sodium sulfate R*, close the tube and allow to stand for 30 min. Filter and evaporate the filtrate to dryness on a water-bath. Dissolve 50 mg of the residue in 25 mL of *ether R*.

Reference solution Dissolve 50 mg of *12-hydroxystearic acid R* in *methylene chloride R* and dilute to 25 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *methylene chloride R, glacial acetic acid R, acetone R* (10:40:50 *V/V/V*).

Application 2 µL.

Development Over a path of 8 cm.

Drying In a current of cold air.

Detection Spray with a 80 g/L solution of *phosphomolybdic acid R* in *2-propanol R* and heat at 120 °C for about 1-2 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the principal spot in the chromatogram obtained with the reference solution.

D. Place about 2 g in a test-tube and add 0.2 mL of *sulfuric acid R*. Close the tube using a stopper fitted with a glass tube bent twice at right angles. Heat the tube until white fumes appear. Collect the fumes in 1 mL of *mercuric chloride solution R*. A white precipitate is formed and the fumes turn a filter paper impregnated with *alkaline potassium tetraiodomercurate solution R* black.

TESTS

Solution S

Dissolve 5.0 g of macrogolglycerol hydroxystearate with less than 40 units of ethylene oxide per molecule in a mixture of 50 volumes of *acetone R* and 50 volumes of *anhydrous ethanol R* and dilute to 50 mL with the same mixture of solvents.

Dissolve 5.0 g of macrogolglycerol hydroxystearate with 40 units or more of ethylene oxide per molecule in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Alkalinity
To 2 mL of solution S add 0.5 mL of *bromothymol blue solution R1*. The solution is not blue.

Acid value (*2.5.1*)
Maximum 2.0, determined on 5.0 g.

Hydroxyl value (*2.5.3, Method A*)
See Table 1083.-1.

Iodine value (*2.5.4*)
Maximum 5.0.

Saponification value (*2.5.6*)
See Table 1083.-1.

Table 1083.-1

Ethylene oxide units per molecule (nominal value)	Hydroxyl value	Saponification value
7	115 - 135	125 - 140
25	70 - 90	70 - 90
40	57 - 80	45 - 69
60	45 - 67	40 - 51

Residual ethylene oxide and dioxan (*2.4.25*)
Maximum 1 ppm of residual ethylene oxide and 10 ppm of residual dioxan.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 2.000 g.

Total ash (*2.4.16*)
Maximum 0.3 per cent, determined on 2.0 g.

LABELLING

The label states the number of ethylene oxide units per molecule (nominal value).

Ph Eur

Refined Castor Oil

(Ph. Eur. monograph 2367)

Ph Eur

DEFINITION

Fatty oil obtained from the seeds of *Ricinus communis* L. by cold expression. It is then refined. A suitable antioxidant may be added.

PRODUCTION

During the expression step, the temperature of the oil must not exceed 50 °C.

CHARACTERS

Appearance

Clear, almost colourless or slightly yellow, viscous, hygroscopic liquid.

Solubility

Slightly soluble in light petroleum, miscible with ethanol (96 per cent) and with glacial acetic acid.

Relative density

About 0.958.

Refractive index

About 1.479.

Viscosity

About 1000 mPa·s.

IDENTIFICATION

First identification: B, C.

Second identification: A, B.

A. A mixture of 2 mL of the substance to be examined and 8 mL of *ethanol (96 per cent) R* is clear (*2.2.1*).

B. Specific absorbance (see Tests).

C. Composition of fatty acids (see Tests).

TESTS

Appearance

The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution BY_3 or Y_3 (*2.2.2, Method I*).

Optical rotation (*2.2.7*)
+ 3.5° to + 6.0°.

Specific absorbance (*2.2.25*)
Greater than 0.7 and maximum 1.5, determined at the absorption maximum at 270 nm.

To 1.00 g add *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent.

Acid value (*2.5.1*)
Maximum 0.8.

Dissolve 5.00 g in 25 mL of the prescribed mixture of solvents.

Hydroxyl value (*2.5.3, Method A*)
Minimum 160.

Peroxide value (*2.5.5, Method A*)
Maximum 5.0.

Unsaponifiable matter (*2.5.7*)
Maximum 0.8 per cent, determined on 5.0 g.

Oil obtained by extraction and adulteration
In a ground-glass-stoppered tube about 125 mm long and 18 mm in internal diameter, thoroughly mix 3 mL of the substance to be examined with 3 mL of *carbon disulfide R*. Shake for 3 min with 1 mL of *sulfuric acid R*. The mixture is less intensely coloured than a freshly prepared mixture of 3.2 mL of *ferric chloride solution R1*, 2.3 mL of *water R* and 0.5 mL of *dilute ammonia R1*.

Composition of fatty acids
Gas chromatography (*2.4.22*) with the following modifications.

Use the mixture of calibrating substances in Table 2.4.22.-3.

Test solution Introduce 75 mg of the substance to be examined into a 10 mL centrifuge tube with a screw cap. Dissolve in 2 mL of *1,1-dimethylethyl methyl ether R1* with shaking and heat gently (50-60 °C). To the still-warm solution, add 1 mL of a 12 g/L solution of *sodium R* in *anhydrous methanol R*, prepared with the necessary

precautions, and shake vigorously for at least 5 min. Add 5 mL of *distilled water R* and shake vigorously for about 30 s. Centrifuge for 15 min at 1500 *g*. Use the upper layer.

Reference solution Dissolve 50 mg of *methyl ricinoleate CRS* and 50 mg of *methyl stearate CRS* in 10.0 mL of *1,1-dimethylethyl methyl ether R1*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Flow rate 0.9 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 55	215
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Calculate the percentage content of each fatty acid by the normalisation procedure.

Correct the area of the peak due to methyl ricinoleate, by multiplying by a factor *R* calculated using the following expression:

$$\frac{m_1 \times A_2}{A_1 \times m_2}$$

m_1 = mass of methyl ricinoleate in the reference solution;
m_2 = mass of methyl stearate in the reference solution;
A_1 = area of the peak due to methyl ricinoleate in the chromatogram obtained with the reference solution;
A_2 = area of the peak due to methyl stearate in the chromatogram obtained with the reference solution.

Composition of the fatty-acid fraction of the oil:
— *palmitic acid*: maximum 2.0 per cent;
— *stearic acid*: maximum 2.5 per cent;
— *oleic acid*: 2.5 per cent to 6.0 per cent;
— *linoleic acid*: 2.5 per cent to 7.0 per cent;
— *linolenic acid*: maximum 1.0 per cent;
— *eicosenoic acid*: maximum 1.0 per cent;
— *ricinoleic acid*: 85.0 per cent to 92.0 per cent;
— *any other fatty acid*: maximum 1.0 per cent.

Water (*2.5.32*)
Maximum 0.3 per cent, or maximum 0.2 per cent if intended for use in the manufacture of parenteral preparations, determined on 1.00 g.

STORAGE

In an airtight, well-filled container, protected from light. If intended for use in the manufacture of parenteral preparations, store under an inert gas.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations and the name of the inert gas.

Ph Eur

Virgin Castor Oil

Castor Oil

(Ph. Eur. monograph 0051)

Action and use
Stimulant laxative; emollient.

Preparation
Zinc and Castor Oil Ointment

Ph Eur

DEFINITION

Fatty oil obtained by cold expression from the seeds of *Ricinus communis* L. A suitable antioxidant may be added.

PRODUCTION

During the expression step, the temperature of the oil must not exceed 50 °C.

CHARACTERS

Appearance
Clear at 40 °C, slightly yellow, viscous, hygroscopic liquid.

Solubility
Slightly soluble in light petroleum, miscible with ethanol (96 per cent) and with glacial acetic acid.

Relative density
About 0.958.

Refractive index
About 1.479.

IDENTIFICATION

First identification: B, C.

Second identification: A, B.

A. A mixture of 2 mL of the substance to be examined and 8 mL of *ethanol (96 per cent) R* is clear (*2.2.1*).

B. Specific absorbance (see Tests).

C. Composition of fatty acids (see Tests).

TESTS

Optical rotation (*2.2.7*)
+ 3.5° to + 6.0°.

Specific absorbance (*2.2.25*)
Maximum 0.7, determined at the absorption maximum at 270 nm.

To 1.00 g add *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent.

Acid value (*2.5.1*)
Maximum 1.5.

Dissolve 5.00 g in 25 mL of the prescribed mixture of solvents.

Hydroxyl value (*2.5.3, Method A*)
Minimum 160.

Peroxide value (*2.5.5, Method A*)
Maximum 10.0.

Unsaponifiable matter (*2.5.7*)
Maximum 0.8 per cent, determined on 5.0 g.

Composition of fatty acids
Gas chromatography (*2.4.22*) with the following modifications.

Use the mixture of calibrating substances in Table 2.4.22.-3.

Test solution Introduce 75 mg of the substance to be examined into a 10 mL centrifuge tube with a screw cap. Dissolve in 2 mL of *1,1-dimethylethyl methyl ether R1* with

shaking and heat gently (50-60 °C). Add, while still warm, 1 mL of a 12 g/L solution of *sodium R* in *anhydrous methanol R*, prepared with the necessary precautions, and mix vigorously for at least 5 min. Add 5 mL of *distilled water R* and mix vigorously for about 30 s. Centrifuge for 15 min at 1500 *g*. Use the upper layer.

Reference solution Dissolve 50 mg of *methyl ricinoleate CRS* and 50 mg of *methyl stearate CRS* in 10.0 mL of *1,1-dimethylethyl methyl ether R1*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Flow rate 0.9 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 55	215
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Calculate the percentage content of each fatty acid by the normalisation procedure.

Correct the area of the peak due to methyl ricinoleate, by multiplying by a factor *R* calculated using the following expression:

$$\frac{m_1 \times A_2}{A_1 \times m_2}$$

m_1 = mass of methyl ricinoleate in the reference solution;
m_2 = mass of methyl stearate in the reference solution;
A_1 = area of the peak due to methyl ricinoleate in the chromatogram obtained with the reference solution;
A_2 = area of the peak due to methyl stearate in the chromatogram obtained with the reference solution.

Composition of the fatty-acid fraction of the oil:
— *palmitic acid*: maximum 2.0 per cent;
— *stearic acid*: maximum 2.5 per cent;
— *oleic acid* : 2.5 per cent to 6.0 per cent;
— *linoleic acid* : 2.5 per cent to 7.0 per cent;
— *linolenic acid* : maximum 1.0 per cent;
— *eicosenoic acid* : maximum 1.0 per cent;
— *ricinoleic acid* : 85.0 per cent to 92.0 per cent;
— *any other fatty acid*: maximum 1.0 per cent.

Water (*2.5.32*)
Maximum 0.3 per cent, determined on 1.00 g.

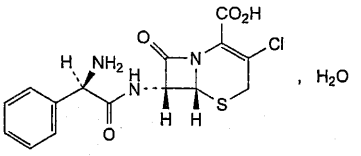
STORAGE

In an airtight, well-filled container, protected from light.

Ph Eur

Cefaclor

(Ph. Eur. monograph 0986)



$C_{15}H_{14}ClN_3O_4S,H_2O$ 385.8 *70356-03-5*

Action and use
Cephalosporin antibacterial.

Preparations
Cefaclor Capsules
Cefaclor Oral Suspension
Cefaclor Prolonged-release Tablets

Ph Eur

DEFINITION

(6*R*,7*R*)-7-[[(2*R*)-2-Amino-2-phenylacetyl]amino]-3-chloro-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent of $C_{15}H_{14}ClN_3O_4S$ (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow powder.

Solubility
Slightly soluble in water, practically insoluble in methanol and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefaclor CRS*.

TESTS

pH (*2.2.3*)
3.0 to 4.5.

Suspend 0.250 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 101 to + 111 (anhydrous substance).

Dissolve 0.250 g in a 10 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same solution.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in 10.0 mL of a 2.7 g/L solution of *sodium dihydrogen phosphate R* adjusted to pH 2.5 with *phosphoric acid R*.

Reference solution (a) Dissolve 2.5 mg of *cefaclor CRS* and 5.0 mg of *delta-3-cefaclor CRS* (impurity D) in 100.0 mL of a 2.7 g/L solution of *sodium dihydrogen phosphate R* adjusted to pH 2.5 with *phosphoric acid R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a 2.7 g/L solution of *sodium dihydrogen phosphate R* adjusted to pH 2.5 with *phosphoric acid R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 7.8 g/L solution of *sodium dihydrogen phosphate R* adjusted to pH 4.0 with *phosphoric acid R*;
— *mobile phase B*: mix 450 mL of *acetonitrile R* with 550 mL of mobile phase A;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	95 → 75	5 → 25
30 - 45	75 → 0	25 → 100
45 - 55	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

System suitability Reference solution (a):
— *resolution*: minimum 2 between the peaks due to cefaclor and impurity D; if necessary, adjust the acetonitrile content in the mobile phase;
— *symmetry factor*: maximum 1.2 for the peak due to cefaclor; if necessary, adjust the acetonitrile content in the mobile phase.

Limits:
— *any impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Water (*2.5.12*)
3.0 per cent to 6.5 per cent, determined on 0.200 g.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 15.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 15.0 mg of *cefaclor CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 3.0 mg of *cefaclor CRS* and 3.0 mg of *delta-3-cefaclor CRS* (impurity D) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Add 220 mL of *methanol R* to a mixture of 780 mL of *water R*, 10 mL of *triethylamine R* and 1 g of *sodium pentanesulfonate R*, then adjust to pH 2.5 with *phosphoric acid R*.

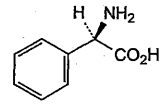
Flow rate 1.5 mL/min.

Detection Spectrophotometer at 265 nm.

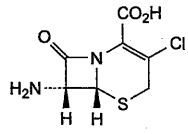
Injection 20 µL.

System suitability:
— *resolution*: minimum 2.5 between the peaks due to cefaclor and impurity D in the chromatogram obtained with reference solution (b); if necessary, adjust the concentration of methanol in the mobile phase;
— *symmetry factor*: maximum 1.5 for the peak due to cefaclor in the chromatogram obtained with reference solution (b);
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections of reference solution (a).

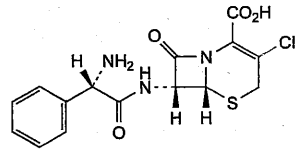
IMPURITIES



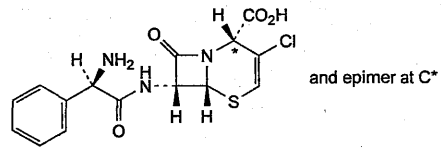
A. (2*R*)-2-amino-2-phenylacetic acid (phenylglycine),



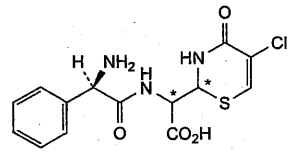
B. (6*R*,7*R*)-7-amino-3-chloro-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



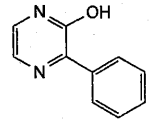
C. (6*R*,7*R*)-7-[[(2*S*)-2-amino-2-phenylacetyl]amino]-3-chloro-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



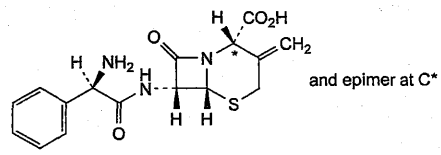
D. (2*R*,6*R*,7*R*)- and (2*S*,6*R*,7*R*)-7-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3-chloro-8-oxo-5-thia- 1-azabicyclo [4.2.0]oct-3-ene-2-carboxylic acid (delta-3-cefaclor),



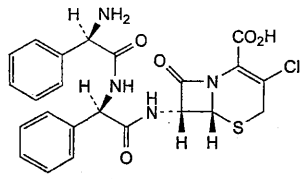
E. 2-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-(5-chloro-4-oxo-3,4-dihydro-2*H*-1,3-thiazin-2-yl)acetic acid,



F. 3-phenylpyrazin-2-ol,



G. (2*R*,6*R*,7*R*)- and (2*S*,6*R*,7*R*)-7-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3-methylene-8-oxo-5-thia-1-azabicyclo[4.2.0]octane-2-carboxylic acid (isocefalexine),

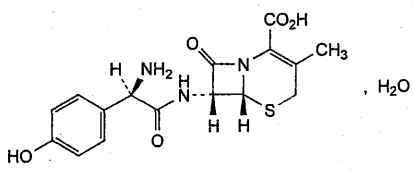


H. (6*R*,7*R*)-7-[[(2*R*)-2-[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-phenylacetyl]amino]-3-chloro-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (*N*-phenylglycyl cefaclor).

Ph Eur

Cefadroxil Monohydrate

(Ph. Eur. monograph 0813)



$C_{16}H_{17}N_3O_5S,H_2O$ 381.4 *66592-87-8*

Action and use
Cephalosporin antibacterial.

Preparations
Cefadroxil Capsules
Cefadroxil Oral Suspension

Ph Eur

DEFINITION

(6*R*,7*R*)-7-[[(2*R*)-2-Amino-2-(4-hydroxyphenyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefadroxil CRS.*

TESTS

pH (*2.2.3*)
4.0 to 6.0.

Suspend 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 165 to + 178 (anhydrous substance).

Dissolve 0.500 g in *water R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 10.0 mg of *D-α-(4-hydroxyphenyl)glycine CRS* (impurity A) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 10.0 mg of *7-aminodesacetoxycephalosporanic acid CRS* (impurity B) in *phosphate buffer solution pH 7.0 R5* and dilute to 10.0 mL with the same buffer solution.

Reference solution (c) Dilute 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b) to 100.0 mL with mobile phase A.

Reference solution (d) Dissolve 10 mg of *dimethylformamide R* and 10 mg of *dimethylacetamide R* in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (e) Dilute 1.0 mL of reference solution (c) to 25.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *phosphate buffer solution pH 5.0 R,*
— *mobile phase B*: *methanol R2,*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	98	2
1 - 20	98 → 70	2 → 30

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (c), (d) and (e).

Relative retention With reference to cefadroxil (retention time = about 6 min): dimethylformamide = about 0.4; dimethylacetamide = about 0.75.

System suitability:
— *resolution*: minimum 5.0 between the peaks due to impurities A and B in the chromatogram obtained with reference solution (c),
— *signal-to-noise ratio*: minimum 10 for the 2nd peak in the chromatogram obtained with reference solution (e).

Limits:
— *impurity A*: not more than the area of the 1st peak in the chromatogram obtained with reference solution (c) (1.0 per cent),
— *any other impurity*: for each impurity, not more than the area of the 2nd peak in the chromatogram obtained with reference solution (c) (1.0 per cent),
— *total*: not more than 3 times the area of the 2nd peak in the chromatogram obtained with reference solution (c) (3.0 per cent),
— *disregard limit*: 0.05 times the area of the 2nd peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard the peaks due to dimethylformamide and dimethylacetamide.

***N,N*-Dimethylaniline** (*2.4.26, Method B*)
Maximum 20 ppm.

Water (*2.5.12*)
4.0 per cent to 6.0 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *cefadroxil CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *cefadroxil CRS* and 50 mg of *amoxicillin trihydrate CRS* in the mobile phase and dilute to 100 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R*, a 2.72 g/L solution of *potassium dihydrogen phosphate R* (4:96 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

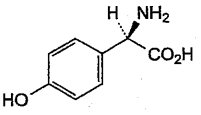
Injection 20 µL.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to cefadroxil and to amoxicillin.

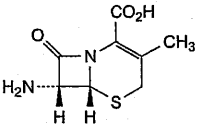
Calculate the percentage content of cefadroxil.

STORAGE
Protected from light.

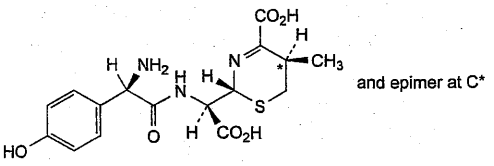
IMPURITIES



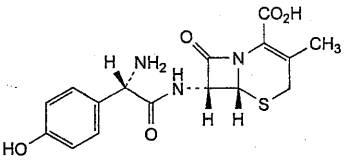
A. (2*R*)-2-amino-2-(4-hydroxyphenyl)acetic acid,



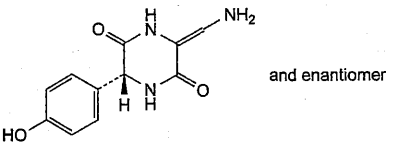
B. (6*R*,7*R*)-7-amino-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ADCA),



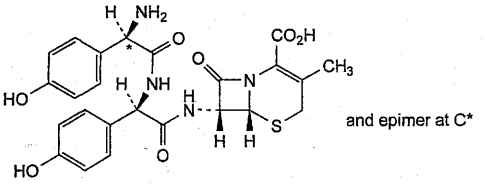
C. (2*R*,5*RS*)-2-[(*R*)-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]carboxymethyl]-5-methyl-5,6-dihydro-2*H*-1,3-thiazine-4-carboxylic acid,



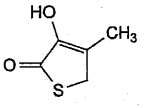
D. (6*R*,7*R*)-7-[[(2*S*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (L-cefadroxil),



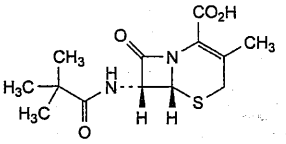
E. (6*RS*)-3-(aminomethylene)-6-(4-hydroxyphenyl)piperazine-2,5-dione,



F. (6*R*,7*R*)-7-[[(2*R*)-2-[[(2*RS*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



G. 3-hydroxy-4-methylthiophen-2(5*H*)-one,

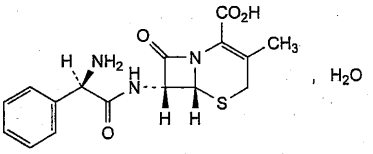


H. (6*R*,7*R*)-7-[(2,2-dimethylpropanoyl)amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ADCA pivalamide).

Ph Eur

Cefalexin Monohydrate

(Ph. Eur. monograph 0708)



$C_{16}H_{17}N_3O_4S,H_2O$ 365.4 *23325-78-2*

Action and use
Cephalosporin antibacterial.

Preparations
Cefalexin Capsules
Cefalexin Oral Suspension
Cefalexin Tablets

Ph Eur

DEFINITION

(6*R*,7*R*)-7-[[(2*R*)-2-Amino-2-phenylacetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid monohydrate.

Semi-synthetic product derived from a fermentation product.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefalexin monohydrate CRS.*

TESTS

pH (*2.2.3*)

4.0 to 5.5.

Dissolve 50 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)

+ 149 to + 158 (anhydrous substance).

Dissolve 0.125 g in *phthalate buffer solution pH 4.4 R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 10.0 mg of *D-phenylglycine R* in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 10.0 mg of *7-aminodesacetoxycephalosporanic acid CRS* in *phosphate buffer solution pH 7.0 R5* and dilute to 10.0 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b) to 100.0 mL with mobile phase A.

Reference solution (d) Dissolve 10 mg of *dimethylformamide R* and 10 mg of *dimethylacetamide R* in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (e) Dilute 1.0 mL of reference solution (c) to 20.0 mL with mobile phase A.

Reference solution (f) Dissolve 10 mg of *cefotaxime sodium CRS* in mobile phase A and dilute to 10.0 mL with mobile phase A. To 1.0 mL of this solution add 1.0 mL of the test solution and dilute to 100 mL with mobile phase A.

Column:

— *size*: *l* = 0.10 m, Ø = 4.6 mm;

— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: *phosphate buffer solution pH 5.0 R*;

— *mobile phase B*: *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	98	2
1 - 20	98 → 70	2 → 30

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (c), (d), (e) and (f).

System suitability:

— *resolution*: minimum 2.0 between the peaks due to impurities A and B in the chromatogram obtained with reference solution (c) and minimum 1.5 between the peaks due to cefalexin and cefotaxime in the chromatogram obtained with reference solution (f).

Limits:

— *impurity B*: not more than the area of the 2nd peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— *any other impurity*: not more than the area of the 1st peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— *total*: not more than 3 times the area of the 1st peak in the chromatogram obtained with reference solution (c) (3.0 per cent);

— *disregard limit*: the area of the 2nd peak in the chromatogram obtained with reference solution (e) (0.05 per cent); disregard any peaks due to dimethylformamide or dimethylacetamide.

***N,N*-Dimethylaniline** (*2.4.26, Method B*)

Maximum 20 ppm.

Water (*2.5.12*)

4.0 per cent to 8.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *cefalexin monohydrate CRS* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *cefradine CRS* in 20 mL of reference solution (a) and dilute to 100 mL with *water R*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, acetonitrile R*, 13.6 g/L solution of *potassium dihydrogen phosphate R, water R* (2:5:10:83 *V/V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

System suitability Reference solution (b):

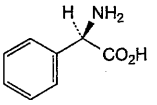
— *resolution*: minimum 4.0 between the peaks due to cefalexin and cefradine.

Calculate the percentage content of cefalexin monohydrate.

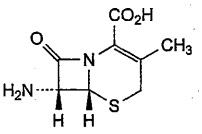
STORAGE

Protected from light.

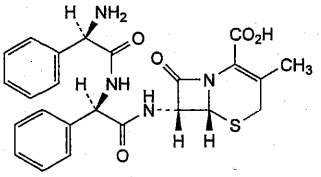
IMPURITIES



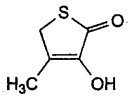
A. (2*R*)-2-amino-2-phenylacetic acid (D-phenylglycine),



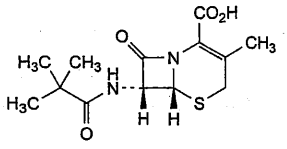
B. (6*R*,7*R*)-7-amino-3-methyl-8-oxo-5-thia-1-azabicyclo [4.2.0]oct-2-ene-2-carboxylic acid (7-aminodesacetoxycephalosporanic acid, 7-ADCA),



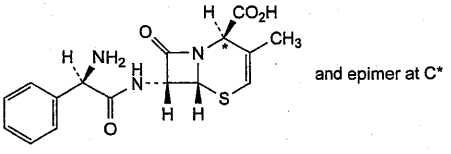
C. (6*R*,7*R*)-7-[[(2*R*)-2-[[(2*R*)-2-amino-2-phenylacetyl] amino]-2-phenylacetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



D. 3-hydroxy-4-methylthiophen-2(5*H*)-one,



E. (6*R*,7*R*)-7-[(2,2-dimethylpropanoyl)amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ADCA pivalamide),

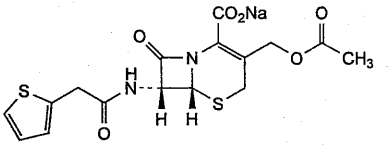


F. (2*RS*,6*R*,7*R*)-7-[[(2*R*)-2-amino-2-phenylacetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylic acid (delta-2-cefalexin).

Ph Eur

Cefalotin Sodium

(Ph. Eur. monograph 0987)



$C_{16}H_{15}N_2NaO_6S_2$ 418.4 *58-71-9*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION

Sodium (6*R*,7*R*)-3-[(acetyloxy)methyl]-8-oxo-7-[(thiophen-2-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, slightly soluble in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cefalotin sodium CRS*.

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 450 nm is not greater than 0.20.

pH (*2.2.3*)
4.5 to 7.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 124 to + 134 (anhydrous substance).

Dissolve 1.25 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 75.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with *water R*.

Reference solution (a) Dissolve 75.0 mg of *cefalotin sodium CRS* in *water R* and dilute to 25.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *water R*.

Reference solution (c) Mix 1 mL of test solution (a), 1 mL of *hydrochloric acid R1* and 8 mL of *water R*. Heat at 60 °C for

12 min and cool to room temperature in iced water. Inject immediately.

Reference solution (d) Dissolve 5 mg of *cefalotin for impurity B identification CRS* in *water R* and dilute to 5 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: mix 3 volumes of *acetonitrile R1* and 97 volumes of a 1.742 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 2.5 with *phosphoric acid R*;
— *mobile phase B*: mix 40 volumes of *acetonitrile R1* and 60 volumes of a 1.742 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 2.5 with *phosphoric acid R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	100 → 0	0 → 100
30 - 35	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (c) and (d).

Relative retention With reference to cefalotin (retention time = about 26 min): impurity C = about 0.2; impurity B = about 0.7; impurity D = about 0.88; impurity A = about 0.96.

System suitability Reference solution (c):
— *resolution*: minimum 7.0 between the peaks due to impurity D and cefalotin.

Limits:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *impurity D*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *any other impurity*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.5 per cent.

Water (*2.5.12*)
Maximum 1.5 per cent, determined on 0.500 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.13 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mix 14 volumes of *acetonitrile R* and 86 volumes of a 6.967 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 6.0 with *phosphoric acid R*.

Detection Spectrophotometer at 260 nm.

Injection 5 µL of test solution (b) and reference solution (a).

Run time 1.5 times the retention time of cefalotin (retention time = about 10 min).

Calculate the percentage content of $C_{16}H_{15}N_2NaO_6S_2$ using the chromatogram obtained with reference solution (a) and taking into account the assigned content of *cefalotin sodium CRS*.

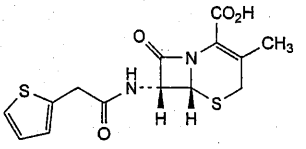
STORAGE

Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

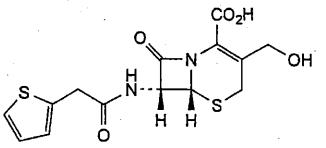
IMPURITIES

Specified impurities *B, D.*

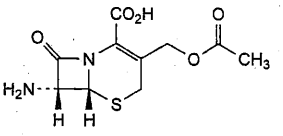
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C.*



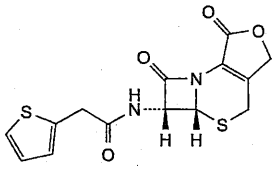
A. (6*R*,7*R*)-3-methyl-8-oxo-7-[(thiophen-2-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetoxycefalotin),



B. (6*R*,7*R*)-3-(hydroxymethyl)-8-oxo-7-[(thiophen-2-ylacetyl) amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetylcefalotin),



C. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-amino-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ACA),

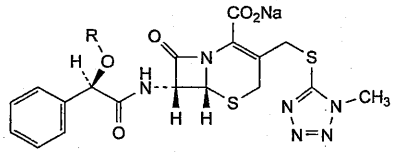


D. (5a*R*,6*R*)-6-[(thiophen-2-ylacetyl)amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (cefalotin lactone).

Ph Eur

Cefamandole Nafate

(Ph. Eur. monograph 1402)



Compound	R	Molecular Formula	M_r
Cefamandole nafate	CHO	$C_{19}H_{17}N_6NaO_6S_2$	512.5
Cefamandole sodium	H	$C_{18}H_{17}N_6NaO_5S_2$	484.5

Cefamandole nafate *42540-40-9*

Cefamandole sodium *30034-03-8*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION
Mixture of sodium (6*R*,7*R*)-7-[[(2*R*)-2-(formyloxy)-2-phenylacetyl]amino]-3-[[(1-methyl-1*H*-tetrazol-5-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate and sodium (6*R*,7*R*)-7-[[(2*R*)-2-hydroxy-2-phenylacetyl]amino]-3-[[(1-methyl-1*H*-tetrazol-5-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (cefamandole sodium), with sodium carbonate.

Semi-synthetic product derived from a fermentation product.

Content
— *cefamandole nafate* ($C_{19}H_{17}N_6NaO_6S_2$): 93.0 per cent to 102.0 per cent (anhydrous and sodium carbonate-free substance), for the sum of the content of cefamandole nafate and cefamandole sodium expressed as cefamandole nafate;
— *cefamandole sodium* ($C_{18}H_{17}N_6NaO_5S_2$): maximum 10.0 per cent (anhydrous and sodium carbonate-free substance);
— *sodium carbonate* (Na_2CO_3): 4.8 per cent to 6.4 per cent.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Freely soluble in water, sparingly soluble in methanol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cefamandole nafate CRS*.
B. It gives reaction (a) of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 475 nm is not greater than 0.03.

pH
6.0 to 8.0 for solution S, measured after 30 min.

Specific optical rotation (*2.2.7*)
−35.0 to −45.0 (anhydrous and sodium carbonate-free substance).

Dissolve 1.00 g in *acetate buffer solution pH 4.7 R1* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mix 18 volumes of *acetonitrile R* and 75 volumes of a 10 per cent *V/V* solution of *triethylamine R* previously adjusted to pH 2.5 with *phosphoric acid R*.

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1 mL of the test solution to 10 mL with the solvent mixture, then heat at 60 °C for 30 min.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *triethylamine phosphate solution*: dissolve 2.0 g of *sodium pentanesulfonate R* in 350 mL of *water R*, add 40 mL of *triethylamine R*, adjust to pH 2.5 with *phosphoric acid R* and dilute to 700 mL with *water R*;
— *mobile phase A:* mix 1 volume of the triethylamine phosphate solution and 2 volumes of *water R*;
— *mobile phase B*: mix equal volumes of the triethylamine phosphate solution, *methanol R* and *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	100	0
1 - 35	100 → 0	0 → 100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL loop injector.

Relative retention With reference to cefamandole nafate (retention time = about 24 min): cefamandole = about 0.8.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to cefamandole and cefamandole nafate.

Limits:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5.0 per cent);

— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.3 per cent *m/m*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.15 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Cefamandole nafate
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *cefamandole nafate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1 mL of the test solution to 10 mL with the mobile phase, then heat at 60 °C for 30 min.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 25 volumes of *acetonitrile R* and 75 volumes of a 10 per cent *V/V* solution of *triethylamine R* previously adjusted to pH 2.5 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL loop injector.

System suitability:
— *resolution*: minimum 7.0 between the 2 principal peaks in the chromatogram obtained with reference solution (b);
— *repeatability*: maximum relative standard deviation of 0.8 per cent after a series of single injections of not less than 3 freshly prepared reference solutions (a).

Calculate the percentage content of cefamandole nafate ($C_{19}H_{17}N_6NaO_6S_2$) from the sum of the contents of cefamandole nafate and cefamandole sodium expressed as cefamandole nafate, using the declared content of *cefamandole nafate CRS*.

1 mg of cefamandole sodium is equivalent to 1.0578 mg of cefamandole nafate.

Sodium carbonate
Dissolve 0.500 g in 50 mL of *water R*. Titrate with *0.1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M hydrochloric acid* is equivalent to 5.3 mg of Na_2CO_3.

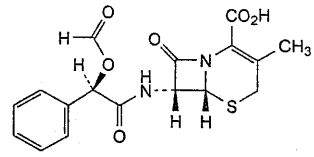
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

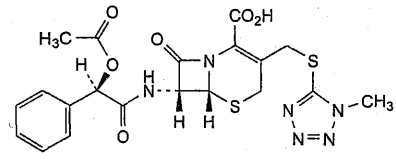
LABELLING

The label states that the substance contains sodium carbonate.

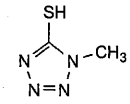
IMPURITIES



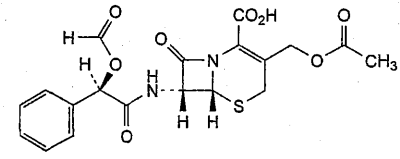
A. (6*R*,7*R*)-7-[[(2*R*)-2-(formyloxy)-2-phenylacetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (formylmandeloyl-7-amino-desacetoxy-cephalosporanic acid),



C. (6*R*,7*R*)-7-[[(2*R*)-2-(acetyloxy)-2-phenylacetyl]amino]-3-[[(1-methyl-1*H*-tetrazol-5-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (*O*-acetylcefamandole),



D. 1-methyl-1*H*-tetrazole-5-thiol,

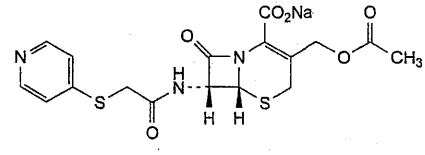


E. (6*R*,7*R*)-7-[[(2*R*)-2-(formyloxy)-2-phenylacetyl]amino]-3-[(acetyloxy)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (formylmandeloyl-7-ACA).

Ph Eur

Cefapirin Sodium

(Ph. Eur. monograph 1650)



$C_{17}H_{16}N_3NaO_6S_2$ 445.5 *24356-60-3*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION

Sodium (6*R*,7*R*)-3-[(acetyloxy)methyl]-8-oxo-7-[[[(pyridin-4-yl)sulfanyl]acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or pale yellow powder.

Solubility
Soluble in water, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefapirin sodium CRS*.

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Appearance of solution
Dissolve 2.0 g in *water R* and dilute to 10.0 mL with the same solvent. The solution is clear (*2.2.1*). Dilute 5.0 mL to 10.0 mL with *water R*. The absorbance (*2.2.25*) of this solution at 450 nm is not greater than 0.25.

pH (*2.2.3*)
6.5 to 8.5.

Dissolve 0.100 g in *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 150 to + 165 (anhydrous substance).

Dissolve 0.500 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 42 mg of the substance to be examined in the mobile phase and dilute to 200.0 mL with the mobile phase.

Reference solution (a) Dissolve 42 mg of *cefapirin sodium CRS* in the mobile phase and dilute to 200.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 20.0 mL with the mobile phase.

Reference solution (d) Mix 1 mL of the test solution, 8 mL of the mobile phase and 1 mL of *hydrochloric acid R1*. Heat at 60 °C for 10 min.

Column:
— *size*: *l* = 0.30 m, Ø = 4 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase Mix 80 mL of *dimethylformamide R*, 4.0 mL of *glacial acetic acid R* and 20 mL of a 4.5 per cent *m/m* solution of *potassium hydroxide R*. Dilute to 2 L with *water R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time Twice the retention time of cefapirin.

Relative retention With reference to cefapirin (retention time = about 13 min): impurity B = about 0.3; impurity C = about 0.5; impurity A = about 0.75.

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to cefapirin and impurity A.

Limits:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent), and not more than 1 such peak has an area greater than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent),
— *disregard limit*: area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

***N,N*-Dimethylaniline** (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.5 per cent.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.300 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.17 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

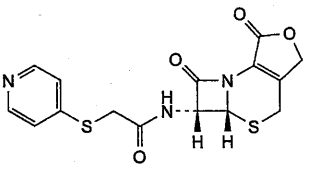
Calculate the percentage content of $C_{17}H_{16}N_3NaO_6S_2$.

STORAGE

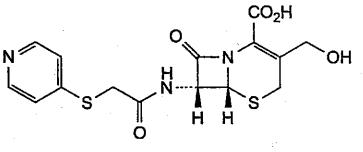
Protected from light. If the substance is sterile, store in a sterile, tamper-proof container.

IMPURITIES

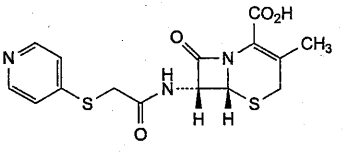
Specified impurities *A, B, C.*



A. (5a*R*,6*R*)-6-[[[(pyridin-4-yl)sulfanyl]acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (deacetylcefapirin lactone),



B. (6*R*,7*R*)-3-(hydroxymethyl)-8-oxo-7-[[[(pyridin-4-yl)sulfanyl]acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetylcefapirin),

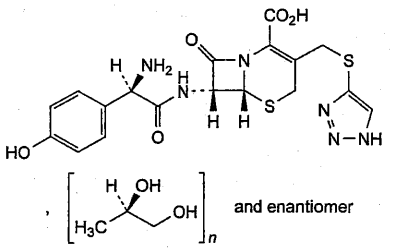


C. (6*R*,7*R*)-3-methyl-8-oxo-7-[[[(pyridin-4-yl)sulfanyl]acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetoxycefapirin).

Ph Eur

Cefatrizine Propylene Glycol

(Ph. Eur. monograph 1403)



$C_{18}H_{18}N_6O_5S_2,(C_3H_8O_2)_n$ 462.5 (base)

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION

Mixture of (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-8-oxo-3-[[(1*H*-1,2,3-triazol-4-yl)sulfanyl]methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid and propane-1,2-diol in molecular proportions of about 1:1.

Content
95.0 per cent to 102.0 per cent of $C_{18}H_{18}N_6O_5S_2$ (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Slightly soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefatrizine propylene glycol CRS*.

B. Examine the chromatograms obtained in the test for propylene glycol.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (b).

TESTS

Specific optical rotation (*2.2.7*)
+ 63 to + 69 (anhydrous substance).

Dissolve 0.400 g in *1 M hydrochloric acid* and dilute to 20.0 mL with the same acid.

Propylene glycol
Gas chromatography (*2.2.28*).

Solvent mixture *acetone R, water R* (20:80 *V/V*).

Internal standard solution Dissolve 1.0 g of *dimethylacetamide R* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution Introduce 0.40 g of the substance to be examined into a ground-glass-stoppered test-tube. Add 3.0 mL of the internal standard solution, 1.0 mL of the solvent mixture and 2.0 mL of *hydrochloric acid R*. Seal the test-tube and shake.

Reference solution (a) Dissolve 2.0 g of *propylene glycol R* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Introduce into a ground-glass-stoppered test-tube 1.0 mL of reference solution (a) and 1.0 mL of the internal standard solution.

Column:
— *material*: stainless steel;
— *size*: *l* = 2 m, Ø = 2 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (150-180 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 30 mL/min.

Temperature:
— *column*: 200 °C;
— *injection port and detector*: 250 °C.

Detection Flame ionisation.

Injection 1 µL of the test solution and reference solution (b).

Limit:
— *propylene glycol*: 13.0 per cent to 18.0 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 60.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 60.0 mg of *cefatrizine propylene glycol CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 30.0 mg of *cefatrizine impurity A CRS* in *buffer solution pH 7.0 R* and dilute to 100.0 mL with the same buffer solution.

Reference solution (c) Dilute 0.6 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (b) to 100.0 mL with *buffer solution pH 7.0 R*.

Reference solution (e) To 1.0 mL of reference solution (a) add 1.0 mL of reference solution (b) and dilute to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 5 volumes of *acetonitrile R* and 95 volumes of a 2.72 g/L solution of *potassium dihydrogen phosphate R* in *water R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 µL of the test solution and reference solutions (c), (d) and (e).

Run time At least twice the retention time of cefatrizine.

System suitability Reference solution (e):
— *resolution*: minimum 5.0 between the peaks due to cefatrizine and impurity A.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.6 per cent);

— *sum of impurities other than A*: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (2.1 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.03 per cent).

Water (*2.5.12*)
Maximum 1.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

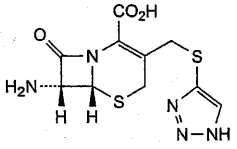
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of $C_{18}H_{18}N_6O_5S_2$ from the declared content of $C_{18}H_{18}N_6O_5S_2$ in *cefatrizine propylene glycol CRS*.

IMPURITIES
Specified impurities *A*.

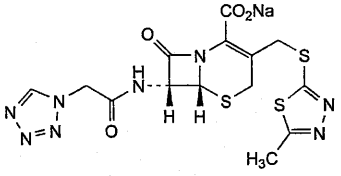


A. (6*R*,7*R*)-7-amino-8-oxo-3-[[(1*H*-1,2,3-triazol-4-yl)sulfanyl]methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ACA triazole).

Ph Eur

Cefazolin Sodium

(Ph. Eur. monograph 0988)



$C_{14}H_{13}N_8NaO_4S_3$ 476.5 *27164-46-1*

Action and use
Cephalosporin antibacterial.

Preparation
Cefazolin Injection

Ph Eur

DEFINITION
Sodium (6*R*,7*R*)-3-[[(5-methyl-1,3,4-thiadiazol-2-yl)sulfanyl]methyl]-8-oxo-7-[(1*H*-tetrazol-1-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder, very hygroscopic.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 0.150 g in 5 mL of *water R*, add 0.5 mL of *dilute acetic acid R*, swirl and allow to stand for 10 min in iced water. Filter the precipitate and rinse with 1-2 mL of *water R*. Dissolve in a mixture of 1 volume of *water R* and 9 volumes of *acetone R*. Evaporate the solvent almost to dryness, then dry in an oven at 60 °C for 30 min.

Comparison *cefazolin CRS*.

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.15.

pH (*2.2.3*)
4.0 to 6.0 for solution S.

Specific optical rotation (*2.2.7*)
−24 to −15 (anhydrous substance).

Dissolve 1.25 g in *water R* and dilute to 25.0 mL with the same solvent.

Absorbance (*2.2.25*)
Dissolve 0.100 g in *water R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with *sodium hydrogen carbonate solution R*. Examined between 220 nm and 350 nm, the solution shows an absorption maximum at 272 nm. The specific absorbance at the maximum is 260 to 300 (anhydrous substance).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 20 mg of the substance to be examined in 10 mL of a 2 g/L solution of *sodium hydroxide R*. Allow to stand for 15-30 min. Dilute 1.0 mL of the solution to 20 mL with mobile phase A.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 45 °C.

Mobile phase:
— *mobile phase A*: solution containing 14.54 g/L of *disodium hydrogen phosphate dodecahydrate R* and 3.53 g/L of *potassium dihydrogen phosphate R*;
— *mobile phase B*: *acetonitrile R*;

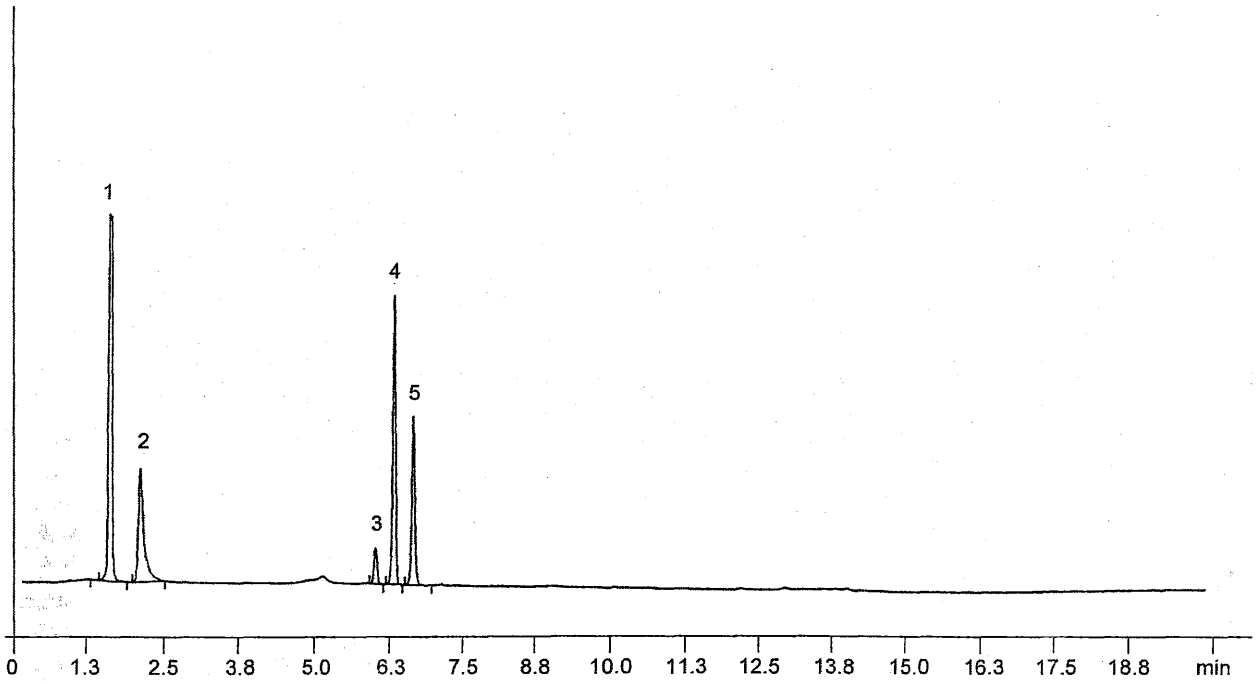


Figure 0988.-1. – *Chromatogram for the test for related substances of cefazolin sodium: reference solution (b) (in situ degradation)*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	98	2
2 - 4	98 → 85	2 → 15
4 - 10	85 → 60	15 → 40
10 - 11.5	60 → 35	40 → 65
11.5 - 12	35	65
12 - 15	35 → 98	65 → 2
15 - 21	98	2

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to cefazolin and impurity L (see Figure 0988.-1).

Limits:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *total*: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

_N,N_-Dimethylaniline (*2.4.26, Method B*)
Maximum 20 ppm.

Water (*2.5.12*)
Maximum 6.0 per cent, determined on 0.300 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.15 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *cefazolin CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *cefuroxime sodium CRS* in 10.0 mL of reference solution (a) and dilute to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 10 volumes of *acetonitrile R* and 90 volumes of a solution containing 2.77 g/L of *disodium hydrogen phosphate dodecahydrate R* and 1.86 g/L of *citric acid monohydrate R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 µL.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to cefazolin and cefuroxime in the chromatogram obtained with reference solution (b);
— *symmetry factor*: 0.8 to 3.0 for the peak due to cefazolin in the chromatogram obtained with reference solution (a).

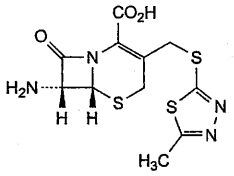
Calculate the percentage content of $C_{14}H_{13}N_8NaO_4S_3$ taking into account the assigned content of *cefazolin CRS* and a conversion factor of 1.048.

STORAGE

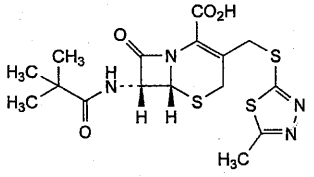
In an airtight container, protected from light. If the substance is sterile, the container is also sterile and tamper-proof.

IMPURITIES

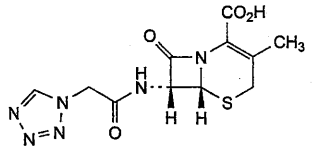
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, G, H, I, J, K, L.*



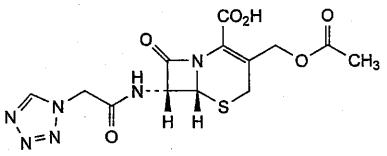
A. (6*R*,7*R*)-7-amino-3-[[(5-methyl-1,3,4-thiadiazol-2-yl) sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



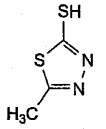
B. (6*R*,7*R*)-7-[(2,2-dimethylpropanoyl)amino]-3-[[(5-methyl-1,3,4-thiadiazol-2-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



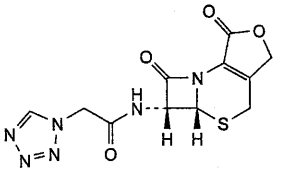
C. (6*R*,7*R*)-3-methyl-8-oxo-7-[(1*H*-tetrazol-1-ylacetyl) amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



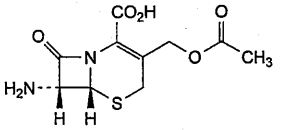
D. (6*R*,7*R*)-3-[(acetyloxy)methyl]-8-oxo-7-[(1*H*-tetrazol-1-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



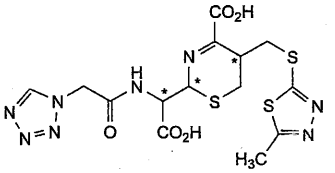
E. 5-methyl-1,3,4-thiadiazol-2-thiol (MMTD),



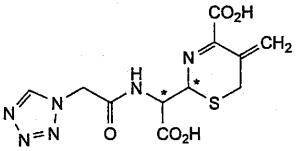
G. (5a*R*,6*R*)-6-[(1*H*-tetrazol-1-ylacetyl)amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione,



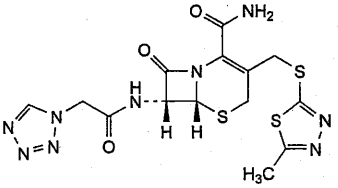
H. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-amino-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ACA),



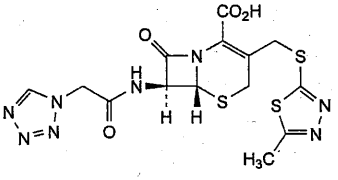
I. 2-[carboxy[(1*H*-tetrazol-1-ylacetyl)amino]methyl]-5-[[(5-methyl-1,3,4-thiadiazol-2-yl)sulfanyl]methyl]-5,6-dihydro-2*H*-1,3-thiazine-4-carboxylic acid (cefazoloic acid),



J. 2-[carboxy[(1*H*-tetrazol-1-ylacetyl)amino]methyl]-5-methylidene-5,6-dihydro-2*H*-1,3-thiazine-4-carboxylic acid,



K. (6*R*,7*R*)-3-[[(5-methyl-1,3,4-thiadiazol-2-yl)sulfanyl] methyl]-8-oxo-7-[(1*H*-tetrazol-1-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxamide (cefazolinamide),

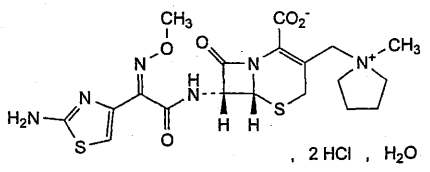


L. (6*R*,7*S*)-3-[[(5-methyl-1,3,4-thiadiazol-2-yl)sulfanyl] methyl]-8-oxo-7-[(1*H*-tetrazol-1-ylacetyl)amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid.

Ph Eur

Cefepime Hydrochloride Monohydrate

(Cefepime Dihydrochloride Monohydrate, Ph. Eur. monograph 2126)



$C_{19}H_{26}Cl_2N_6O_5S_2,H_2O$ 571.5 *123171-59-5*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION

(6*R*,7*R*)-7-[[(2*Z*)-(2-Aminothiazol-4-yl)(methoxyimino) acetyl]amino]-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate dihydrochloride monohydrate. Semi-synthetic product derived from a fermentation product.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefepime dihydrochloride monohydrate CRS.*

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_3 (*2.2.2, Method II*).

Dissolve 2.0 g in *water R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 40 to + 45 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Impurity G
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in *0.01 M nitric acid* and dilute to 10.0 mL with the same acid.

Reference solution (a) Dilute 0.250 g of *N-methylpyrrolidine R* (impurity G) to 100.0 mL with *water R*. Dilute 2.0 mL of this solution to 100.0 mL with *0.01 M nitric acid.*

Reference solution (b) Dilute 0.250 g of *pyrrolidine R* to 100 mL with *0.01 M nitric acid.* Dilute 2 mL of the solution to 100 mL with *0.01 M nitric acid.* Mix 5 mL of this solution with 5 mL of reference solution (a).

Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *strong cation-exchange resin R* (5 µm).

Mobile phase Mix 1 volume of *acetonitrile R* and 100 volumes of *0.01 M nitric acid*; filter through a 0.2 µm filter.

Flow rate 1 mL/min.

Detection Conductivity detector.

Injection 100 µL.

Run time 1.1 times the retention time of cefepime.

Retention time Cefepime = about 50 min, eluting as a broadened peak.

System suitability:
— *symmetry factor*: maximum 2.5 for the peak due to impurity G in the chromatogram obtained with reference solution (a);
— *repeatability*: maximum relative standard deviation of 5.0 per cent after 6 injections of reference solution (a);
— *peak-to-valley ratio*: minimum 3 between the peaks due to pyrrolidine and impurity G in the chromatogram obtained with reference solution (b).

Calculate the percentage content of impurity G in the test solution using reference solution (a).

Limit:
— *impurity G*: maximum 0.5 per cent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use or keep refrigerated at 4-8 °C for not more than 12 h.*

Test solution Dissolve 70.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A. Sonicate for 30 s and stir for about 5 min.

Reference solution (a) Dissolve 70.0 mg of *cefepime dihydrochloride monohydrate CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A. Sonicate for 30 s and stir for about 5 min.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with mobile phase A. Dilute 2.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (c) Dissolve 7 mg of *cefepime dihydrochloride monohydrate for system suitability CRS* (containing impurities A, B and F) in mobile phase A and dilute to 5 mL with mobile phase A.

Reference solution (d) Dissolve 2 mg of *cefepime impurity E CRS* in mobile phase A and dilute to 25.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 10 volumes of *acetonitrile R* and 90 volumes of a 0.68 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 5.0 with a 0.5 M potassium hydroxide solution prepared from *potassium hydroxide R*;
— *mobile phase B*: mix equal volumes of *acetonitrile R* and a 0.68 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 5.0 with a 0.5 M potassium hydroxide solution prepared from *potassium hydroxide R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100	0
10 - 30	100 → 50	0 → 50
30 - 35	50	50
35 - 36	50 → 100	50 → 0
36 - 45	100	0

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *cefepime dihydrochloride monohydrate for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and F; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity E.

Relative retention With reference to cefepime (retention time = about 7 min): impurity E = about 0.4; impurity F = about 0.8; impurity A = about 2.5; impurity B = about 4.1.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity F and cefepime.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.4; impurity B = 1.4; impurity E = 1.8;
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurities B, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurity E*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
3.0 per cent to 4.5 per cent, determined on 0.400 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.04 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A.

Injection Test solution and reference solution (a).

Run time 1.4 times the retention time of cefepime.

Calculate the percentage content of $C_{19}H_{26}Cl_2N_6O_5S_2$ from the declared content of *cefepime dihydrochloride monohydrate CRS*.

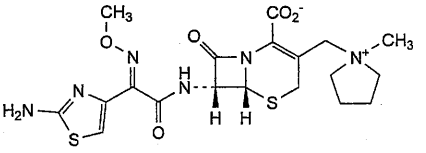
STORAGE

Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

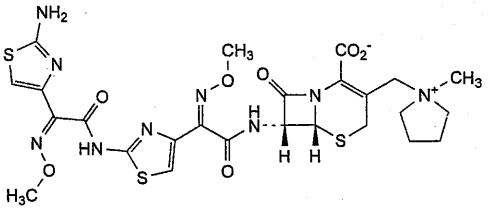
IMPURITIES

Specified impurities *A, B, E, F, G.*

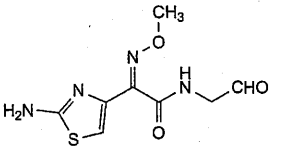
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D.*



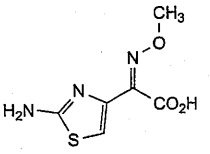
A. (6*R*,7*R*)-7-[[(2*E*)-(2-aminothiazol-4-yl)(methoxyimino) acetyl]amino]-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (*anti*-cefepime),



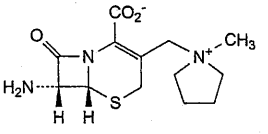
B. (6*R*,7*R*)-7-[[(2*Z*)-[2-[[(2*Z*)-(2-aminothiazol-4-yl) (methoxyimino)acetyl]amino]thiazol-4-yl](methoxyimino) acetyl]amino]-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,



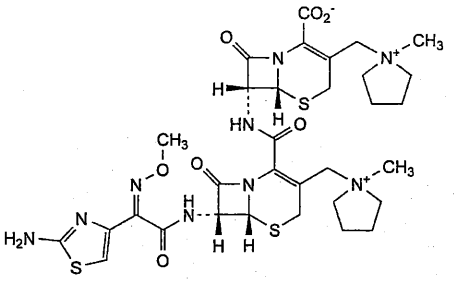
C. (2*Z*)-2-(2-aminothiazol-4-yl)-*N*-(formylmethyl)-2-(methoxyimino)acetamide,



D. (2*Z*)-(2-aminothiazol-4-yl)(methoxyimino)acetic acid,



E. (6*R*,7*R*)-7-amino-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,



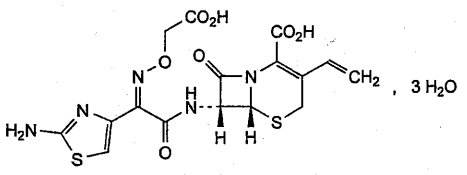
F. (6*R*,7*R*)-7-[[[(6*R*,7*R*)-7-[[(2*Z*)-(2-aminothiazol-4-yl)(methoxyimino)acetyl]amino]-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-en-2-yl]carbonyl]amino]-3-[(1-methylpyrrolidinio)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

G. 1-methylpyrrolidine (*N*-methylpyrrolidine).

Ph Eur

Cefixime

(Ph. Eur. monograph 1188)



$C_{16}H_{15}N_5O_7S_2,3H_2O$ 507.5

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION
(6*R*,7*R*)-7-[[(*Z*)-2-(2-Aminothiazol-4-yl)-2-[(carboxymethoxy)imino]acetyl]amino]-3-ethenyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid trihydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, slightly hygroscopic powder.

Solubility
Slightly soluble in water, soluble in methanol, sparingly soluble in anhydrous ethanol, practically insoluble in ethyl acetate.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefixime CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS
pH (*2.2.3*)
2.6 to 4.1.

Suspend 0.5 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *cefixime CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) In order to prepare impurity D *in situ*, dissolve 10 mg of *cefixime CRS* in 10 mL of *water R*, heat on a water-bath for 45 min and cool. Inject immediately.

Column:
— *size*: *l* = 0.125 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.
— *Mobile phase*: mix 250 volumes of *acetonitrile R* and 750 volumes of a tetrabutylammonium hydroxide solution prepared as follows: dissolve 8.2 g of *tetrabutylammonium hydroxide R* in *water for chromatography R* and dilute to 800 mL with the same solvent; adjust to pH 6.5 with *dilute phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Autosampler Set at 4 °C.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 3 times the retention time of cefixime.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to cefixime and impurity D; if necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:
— *any impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Ethanol (*2.4.24*)
Head-space gas chromatography (*2.2.28*): use the standard additions method.

Sample solution Dissolve 0.250 g of the substance to be examined in *dimethylformamide R* and dilute to 25.0 mL with the same solvent.

Limit:
— *ethanol*: maximum 1.0 per cent.

Water (*2.5.12*)
9.0 per cent to 12.0 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

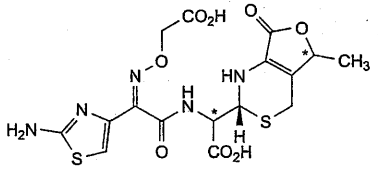
Injection Test solution and reference solution (a).

System suitability Reference solution (a):
— *symmetry factor*: maximum 3.0 for the peak due to cefixime;
— *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

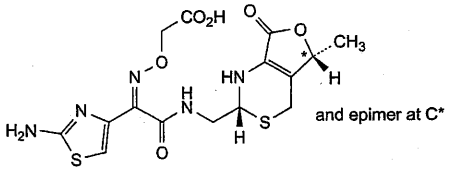
Calculate the percentage content of $C_{16}H_{15}N_5O_7S_2$ taking into account the assigned content of *cefixime CRS*.

STORAGE
In an airtight container, protected from light.

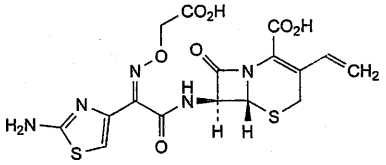
IMPURITIES



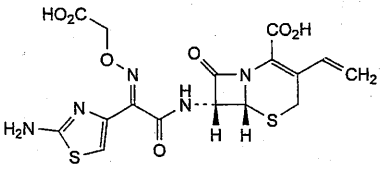
A. 2-[[(*Z*)-2-(2-aminothiazol-4-yl)-2-[(carboxymethoxy)imino]acetyl]amino]-2-[(2*R*)-5-methyl-7-oxo-1,2,5,7-tetrahydro-4*H*-furo[3,4-*d*][1,3]thiazin-2-yl] acetic acid,



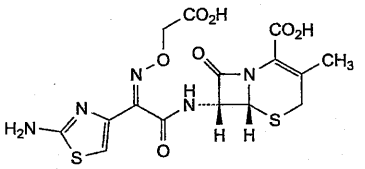
B. 2-[[[(*Z*)-1-(2-aminothiazol-4-yl)-2-[[[(2*R*,5*RS*)-5-methyl-7-oxo-1,2,5,7-tetrahydro-4*H*-furo[3,4-*d*][1,3]thiazin-2-yl] methyl]amino]-2-oxoethylidene]amino]oxy]acetic acid,



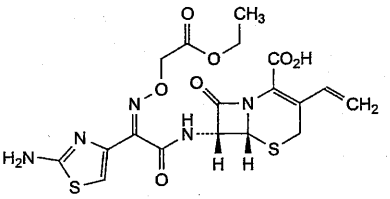
C. (6*R*,7*S*)-7-[[(*Z*)-2-(2-aminothiazol-4-yl)-2-[(carboxymethoxy)imino]acetyl]amino]-3-ethenyl-8-oxo5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefixime 7-epimer),



D. (6*R*,7*R*)-7-[[(*E*)-2-(2-aminothiazol-4-yl)-2-[(carboxymethoxy)imino]acetyl]amino]-3-ethenyl-8-oxo5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefixime (*E*)-isomer),



E. (6*R*,7*R*)-7-[[(*Z*)-2-(2-aminothiazol-4-yl)-2-[(carboxymethoxy)imino]acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,

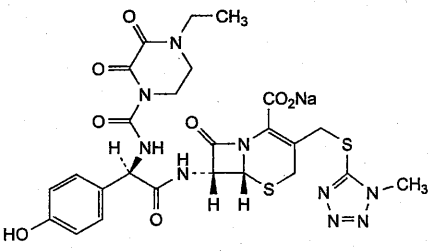


F. (6*R*,7*R*)-7-[[(*Z*)-2-(2-aminothiazol-4-yl)-2-[(2-ethoxy-2-oxoethoxy)imino]acetyl]amino]-3-ethenyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid.

Ph Eur

Cefoperazone Sodium

(Ph. Eur. monograph 1404)



$C_{25}H_{26}N_9NaO_8S_2$ 668 *62893-20-3*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION
Sodium (6*R*,7*R*)-7-[[(2*R*)-2-[[(4-ethyl-2,3-dioxopiperazin-1-yl)carbonyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-3-[[(1-methyl-1*H*-tetrazol-5-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow, hygroscopic powder.

Solubility
Freely soluble in water, soluble in methanol, slightly soluble in ethanol (96 per cent).

If crystalline, it shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve the substance to be examined in *methanol R* and evaporate to dryness; examine the residue.

Comparison *Ph. Eur. reference spectrum of cefoperazone sodium.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (a) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

C. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.15.

Dissolve 2.5 g in *water R* and dilute to 25.0 mL with the same solvent.

pH (*2.2.3*)

4.5 to 6.5.

Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 250.0 mL with the mobile phase.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *cefoperazone dihydrate CRS* in the mobile phase and dilute to 250.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 884 volumes of *water R*, 110 volumes of *acetonitrile R*, 3.5 volumes of a 60 g/L solution of *acetic acid R* and 2.5 volumes of a triethylammonium acetate solution prepared as follows: dilute 14 mL of *triethylamine R* and 5.7 mL of *glacial acetic acid R* to 100 mL with *water R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of test solution (b) and reference solutions (a) and (b).

Run time 2.5 times the retention time of cefoperazone.

Retention time Cefoperazone = about 15 min.

System suitability Reference solution (a):
- *number of theoretical plates*: minimum 5000, calculated for the principal peak; if necessary, adjust the content of *acetonitrile R* in the mobile phase;
- *symmetry factor*: maximum 1.6 for the principal peak; if necessary, adjust the content of *acetonitrile R* in the mobile phase.

Limits:
- *any impurity*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
- *total*: not more than 4.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (4.5 per cent);
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Acetone (*2.4.24, System B*)

Maximum 2.0 per cent.

Sample solution Dissolve 0.500 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Solvent solution Dissolve 0.350 g of *acetone R* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with *water R*.

Prepare each of 4 injection vials as shown in the table below:

Vial No.	Sample solution (mL)	Solvent solution (mL)	*Water R* (mL)
1	1.0	0	4.0
2	1.0	1.0	3.0
3	1.0	2.0	2.0
4	1.0	3.0	1.0

Static head-space conditions that may be used:
- *equilibration time*: 15 min;
- *transfer-line temperature*: 110 °C.

Temperature:
- *Column*: 40 °C for 10 min.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.200 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.20 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (a) and reference solution (a).

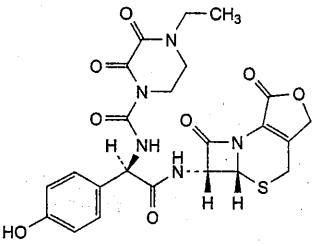
System suitability Reference solution (a):
- *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of cefoperazone sodium by multiplying the percentage content of cefoperazone by 1.034.

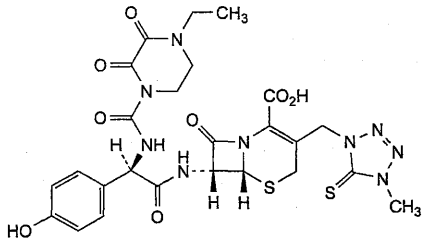
STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

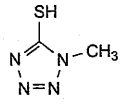
IMPURITIES



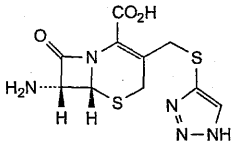
A. (5a*R*,6*R*)-6-[[(2*R*)-2-[[(4-ethyl-2,3-dioxopiperazin-1-yl)carbonyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione,



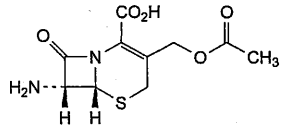
B. (6*R*,7*R*)-7-[[(2*R*)-2-[[(4-ethyl-2,3-dioxopiperazin-1-yl)carbonyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-3-[(4-methyl-5-thioxo-4,5-dihydro-1*H*-tetrazol-1-yl)methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



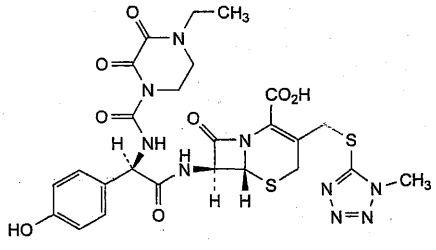
C. 1-methyl-1*H*-tetrazole-5-thiol,



D. (6*R*,7*R*)-7-amino-8-oxo-3-[(1*H*-1,2,3-triazol-4-ylsulfanyl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-TACA),



E. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-amino-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ACA),

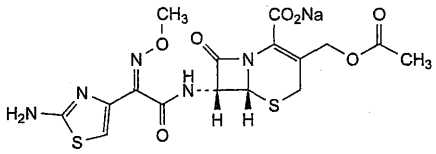


F. (6*R*,7*S*)-7-[[(2*R*)-2-[[(4-ethyl-2,3-dioxopiperazine-1-yl)carbonyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-3-[[(1-methyl-1*H*-tetrazol-5-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid.

Ph Eur

Cefotaxime Sodium

(Ph. Eur. monograph 0989)



$C_{16}H_{16}N_5NaO_7S_2$ 477.4 *64485-93-4*

Action and use
Cephalosporin antibacterial.

Preparation
Cefotaxime Injection

Ph Eur

DEFINITION

Sodium (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow powder, hygroscopic.

Solubility
Freely soluble in water, sparingly soluble in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefotaxime sodium CRS*.

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*). Add 1 mL of *glacial acetic acid R* to 10 mL of solution S. The solution, examined immediately, is clear.

pH (*2.2.3*)
4.5 to 6.5 for solution S.

Specific optical rotation (*2.2.7*)
+ 58.0 to + 64.0 (anhydrous substance).

Dissolve 0.100 g in *water R* and dilute to 10.0 mL with the same solvent.

Absorbance (*2.2.25*)
Maximum 0.40 at 430 nm for solution S.

Specific absorbance (*2.2.25*)
360 to 390, determined at the absorption maximum at 235 nm (anhydrous substance).

Dissolve 20.0 mg in *water R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Mobile phase B, mobile phase A (14:86 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with the same solution.

Reference solution (a) Dissolve 8.0 mg of *cefotaxime acid CRS* in solution A and dilute to 10.0 mL with the same solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with solution A.

Reference solution (c) Add 1.0 mL of *dilute hydrochloric acid R* to 4.0 mL of the test solution. Heat the solution at 40 °C for 2 h. Add 5.0 mL of *buffer solution pH 6.6 R* and 1.0 mL of *dilute sodium hydroxide solution R*.

Reference solution (d) Dissolve 4 mg of *cefotaxime for peak identification CRS* (containing impurities A, B, C, E and F) in 5 mL of solution A.

Column:
- *size*: *l* = 0.15 m, Ø = 3.9 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm),
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: 7.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R* adjusted to pH 6.25 using *phosphoric acid R*;
- *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	86	14
7 - 9	86 → 82	14 → 18
9 - 16	82	18
16 - 45	82 → 60	18 → 40
45 - 50	60	40
50 - 55	60 → 86	40 → 14
55 - 60	86	14

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *cefotaxime for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, E and F.

Relative retention With reference to cefotaxime (retention time = about 13 min): impurity B = about 0.3; impurity A = about 0.5; impurity E = about 0.6; impurity C = about 1.9; impurity D = about 2.3; impurity F = about 2.4; impurity G = about 3.1.

System suitability Reference solution (c):
- *resolution*: minimum 3.5 between the peaks due to impurity E and cefotaxime;
- *symmetry factor*: maximum 2.0 for the peak due to cefotaxime.

Limits:
- *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Ethanol (*2.4.24, System A*)
Maximum 1.0 per cent.

***N,N*-Dimethylaniline** (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.5 per cent *m/m*.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 0.300 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.05 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{16}H_{16}N_5NaO_7S_2$ by multiplying the percentage content of cefotaxime by 1.048.

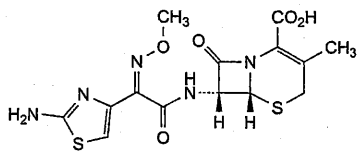
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

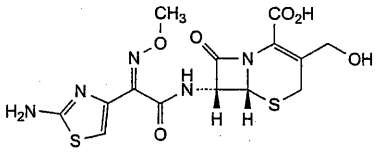
IMPURITIES

Specified impurities *A, B, C, D, E, F.*

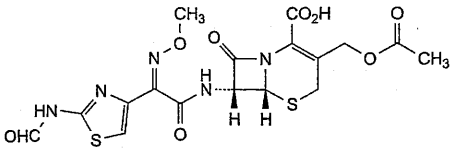
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G.*



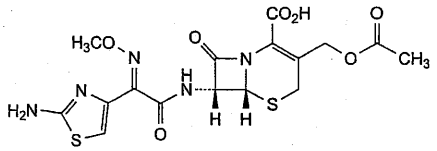
A. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetoxycefotaxime),



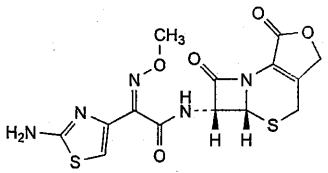
B. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(hydroxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (deacetylcefotaxime),



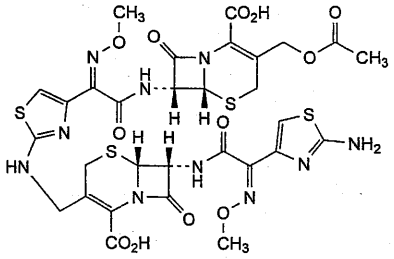
C. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(2*Z*)-2-[2-(formylamino)thiazol-4-yl]-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (*N*-formylcefotaxime),



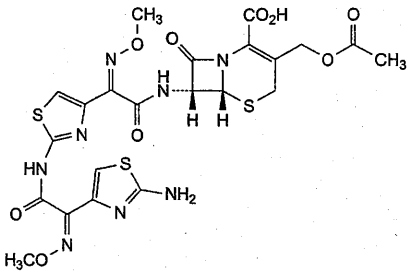
D. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(2*E*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid ((*E*)-cefotaxime),



E. (5a*R*,6*R*)-6-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (deacetylcefotaxime lactone),



F. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(2*Z*)-2-[2-[[[(6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-2-carboxy-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-en-2-yl]methyl]amino]thiazol-4-yl]-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefotaxime dimer),

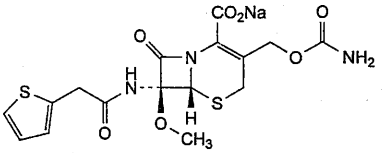


G. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(2*Z*)-2-[2-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]thiazol-4-yl]-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (ATA cefotaxime).

Ph Eur

Cefoxitin Sodium

(Ph. Eur. monograph 0990)



$C_{16}H_{16}N_3NaO_7S_2$ 449.4 *33564-30-6*

Action and use
Cephalosporin antibacterial.

Preparation
Cefoxitin Injection

Ph Eur

DEFINITION

Sodium (6*R*,7*S*)-3-[(carbamoyloxy)methyl]-7-methoxy-8-oxo-7-[[(thiophen-2-yl)acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, very hygroscopic powder.

Solubility
Very soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefoxitin sodium CRS.*

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

pH (*2.2.3*)
4.2 to 7.0.

Dilute 2 mL of solution S to 20 mL with *carbon dioxide-free water R.*

Specific optical rotation (*2.2.7*)
+ 206 to + 214 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 1.0 g of *potassium dihydrogen phosphate R* and 1.8 g of *anhydrous disodium hydrogen phosphate R* in 1000 mL of *water R.* To 100 mL of the solution add 800 mL of *water R*, adjust to pH 7.0 with *phosphoric acid R* or a 40 g/L solution of *sodium hydroxide R* and dilute to 1000 mL with *water R.*

Test solution Dissolve 50 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 20.0 mL with solution A.

Reference solution (c) Dissolve 5 mg of *cefoxitin for peak identification CRS* (containing impurities A, B, E, H, I and J) in solution A and dilute to 5 mL with solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *phenylsilyl silica gel for chromatography R* (3.0 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: 1.0 g/L solution of *ammonium formate R* adjusted to pH 2.7 with *anhydrous formic acid R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	92	8
5 - 50	92 → 74	8 → 26
50 - 85	74	26

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *cefoxitin for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, E, H, I and J.

Relative retention With reference to cefoxitin (retention time = about 30 min): impurity A = about 0.83; impurity I = about 0.98; impurity H = about 1.06; impurity E = about 1.11; impurity B = about 1.18; impurity J = about 1.66.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to impurities H and E;
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cefoxitin.

Limits:
— *impurity I*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *impurities E, H*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity J*: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities A, B*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3.0 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 0.500 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.13 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 25.0 mg of *cefoxitin sodium CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dissolve 20.0 mg of *2-(2-thienyl)acetic acid R* in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (c) Mix 1.0 mL of reference solution (a) and 5.0 mL of reference solution (b).

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetic acid R, acetonitrile R, water R* (1:19:81 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (a) and (c).

Run time 12 min.

System suitability Reference solution (c):
— *resolution*: minimum 3.5 between the 2 principal peaks.

Calculate the percentage content of $C_{16}H_{16}N_3NaO_7S_2$ taking into account the assigned content of *cefoxitin sodium CRS*.

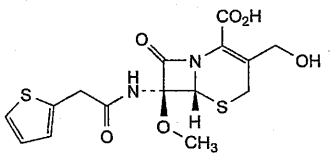
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

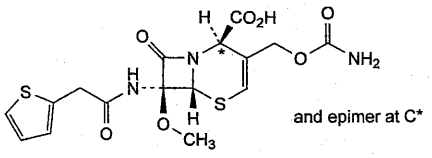
IMPURITIES

Specified impurities *A, B, E, H, I, J.*

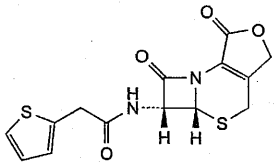
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, F, G.*



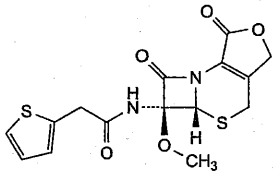
A. (6*R*,7*S*)-3-(hydroxymethyl)-7-methoxy-8-oxo-7-[[(thiophen-2-yl)acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (decarbamoylcefoxitin),



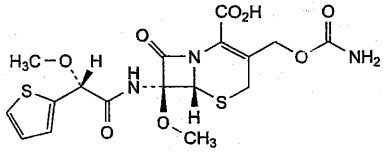
B. (2*RS*,6*R*,7*S*)-3-[(carbamoyloxy)methyl]-7-methoxy-8-oxo-7-[[(thiophen-2-yl)acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylic acid (delta-3-cefoxitin),



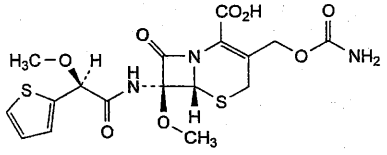
C. (5a*R*,6*R*)-6-[[(thiophen-2-yl)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (cefalotin lactone),



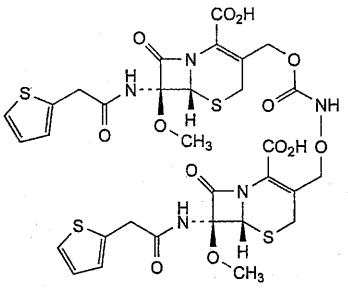
D. (5a*R*,6*S*)-6-methoxy-6-[[(thiophen-2-yl)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (cefoxitin lactone),



E. (6*R*,7*S*)-3-[(carbamoyloxy)methyl]-7-methoxy-7-[[(2*R*)-2-methoxy-2-(thiophen-2-yl)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid ((*R*)-methoxy cefoxitin),



F. (6*R*,7*S*)-3-[(carbamoyloxy)methyl]-7-methoxy-7-[[(2*S*)-2-methoxy-2-(thiophen-2-yl)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid ((*S*)-methoxy cefoxitin),



G. (6*R*,7*S*)-3-[[[[[[(6*R*,7*S*)-2-carboxy-7-methoxy-8-oxo-7-[[2-(thiophen-2-yl)acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-en-3-yl]methyl]oxy]carbamoyl]oxy]methyl]-7-methoxy-8-oxo-7-[[2-(thiophen-2-yl)acetyl]amino]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefoxitin dimer),

H. unknown structure,

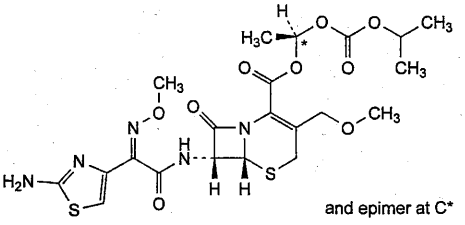
I. unknown structure,

J. unknown structure.

Ph Eur

Cefpodoxime Proxetil

(Ph. Eur. monograph 2341)



$C_{21}H_{27}N_5O_9S_2$ 557.6 *87239-81-4*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION
(1*RS*)-1-[[(1-Methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
94.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or pale yellow or light brown, amorphous powder.

Solubility
Very slightly soluble or practically insoluble in water, very soluble in acetonitrile and in methanol, freely soluble in anhydrous ethanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cefpodoxime proxetil CRS.*

TESTS

Diastereoisomer ratio

Liquid chromatography (*2.2.29*) as described under Assay. Use the normalisation procedure.

Limit Test solution:

— the ratio of the area of the peak due to cefpodoxime proxetil diastereoisomer II to the sum of the areas of the peaks due to cefpodoxime proxetil diastereoisomers I and II is between 0.5 and 0.6.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use or store them at 2-8 °C.*

Solvent mixture *glacial acetic acid R, acetonitrile R, water R* (2:99:99 *V/V/V*).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *cefpodoxime proxetil for peak identification CRS* (containing impurities B, C and D) in 5.0 mL of the solvent mixture.

Reference solution (c) Dissolve 5 mg of *cefpodoxime proxetil for impurity H identification CRS* in 5.0 mL of the solvent mixture.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: maintain at a constant temperature of 20 °C.

Mobile phase:

— *mobile phase A*: *anhydrous formic acid R, methanol R, water R* (1:400:600 *V/V/V*);

— *mobile phase B*: *anhydrous formic acid R, water R, methanol R* (1:50:950 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 65	95	5
65 - 145	95 → 15	5 → 85
145 - 155	15	85

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *cefpodoxime proxetil for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C and D; use the chromatogram supplied with *cefpodoxime proxetil for impurity H identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurity H.

Relative retention With reference to cefpodoxime proxetil diastereoisomer II (retention time = about 58 min): diastereoisomer I of impurity B = about 0.68; diastereoisomer I of cefpodoxime proxetil = about 0.74; impurity C = about 0.82; diastereoisomer II of impurity B = about 0.85; impurity D (2 peaks) = about 0.88 and 1.13; peaks due to diastereoisomers of impurity H: between about 1.9 and 2.3.

System suitability:

— the chromatogram obtained with reference solution (b) is similar to the chromatogram supplied with *cefpodoxime proxetil for peak identification CRS*;

— *resolution*: minimum 6.0 between the peaks due to cefpodoxime proxetil diastereoisomers I and II in the chromatogram obtained with reference solution (a);

— *peak-to-valley ratio*: minimum 1.1, where H_p = height above the baseline of the peak due to diastereoisomer II of impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity C in the chromatogram obtained with reference solution (b).

Limits:

— *impurity C*: not more than twice the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (2.0 per cent);

— *impurity D (sum of the 2 diastereoisomers)*: not more than the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *impurity H (sum of the diastereoisomers)*: not more than the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *impurity B (sum of the 2 diastereoisomers)*: not more than 0.5 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *any other impurity*: for each impurity, not more than 0.2 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *total*: not more than 4 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (4.0 per cent);

— *disregard limit*: 0.05 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

Maximum 2.5 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (*2.2.29*).

Solution A 20 mg/L solution of *anhydrous citric acid R* in *acetonitrile R*.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution Dissolve 30.0 mg of *cefpodoxime proxetil CRS* in solution A and dilute to 50.0 mL with solution A.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 40 °C.

Mobile phase *methanol R, water R* (9:11 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 1.2 times the retention time of cefpodoxime proxetil diastereoisomer II.

Retention time Cefpodoxime proxetil diastereoisomer II = about 30 min.

System suitability Reference solution:

— *resolution*: minimum 4.0 between the peaks due to cefpodoxime proxetil diastereoisomers I and II.

Calculate the percentage content of $C_{21}H_{27}N_5O_9S_2$ from the sum of the areas of the 2 peaks due to the diastereoisomers and using the declared content of *cefpodoxime proxetil CRS*.

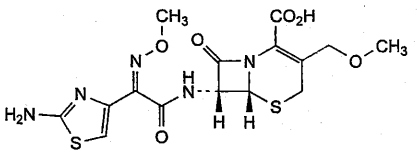
STORAGE

Protected from light.

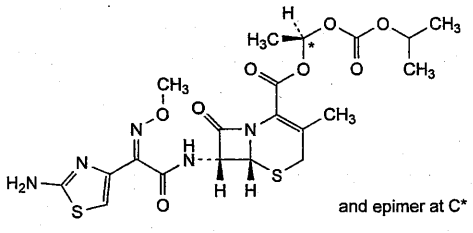
IMPURITIES

Specified impurities *B, C, D, H.*

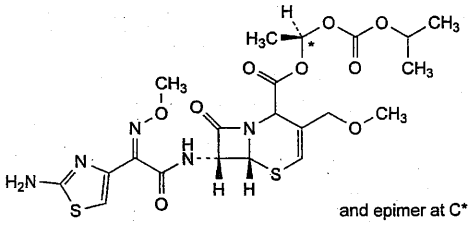
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, E, F, G.*



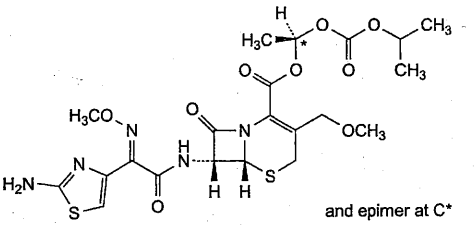
A. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefpodoxime),



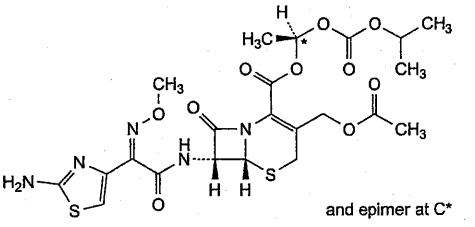
B. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (ADCA-analogue of cefpodoxime proxetil),



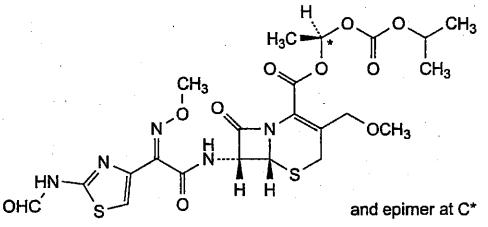
C. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylate (delta-2-cefpodoxime proxetil),



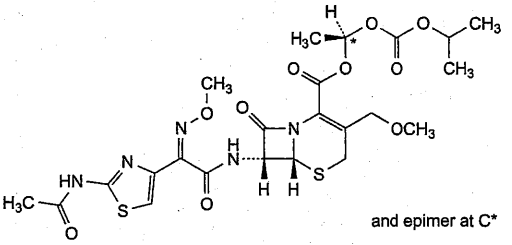
D. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*E*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (anti-cefpodoxime proxetil),



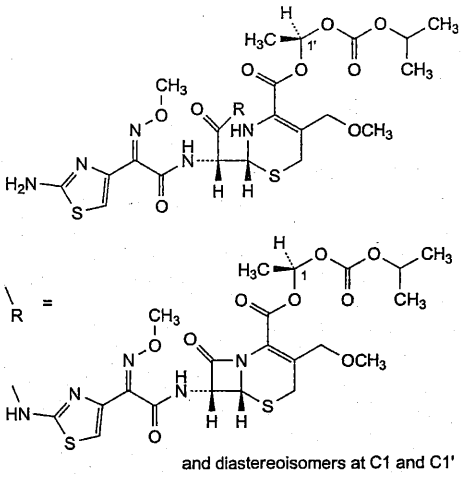
E. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-3-(acetoxymethyl)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (ACA-analogue of cefpodoxime proxetil),



F. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-2-[(2-formylamino)thiazol-4-yl)-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (*N*-formyl cefpodoxime proxetil),



G. (1*RS*)-1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-[2-(2-acetylamino)thiazol-4-yl]-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (*N*-acetyl-cefpodoxime proxetil),

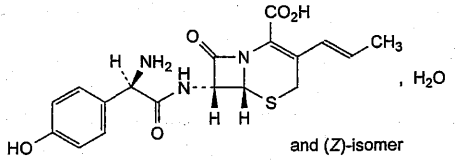


H. mixture of the diastereoisomers of 1-[[(1-methylethoxy)carbonyl]oxy]ethyl (6*R*,7*R*)-7-[[(2*Z*)-2-[2-[[(2*R*)-2-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-(methoxyimino)acetyl]amino]-2-[(2*R*)-5-(methoxymethyl)-4-[[1-[[(1-methylethoxy)carbonyl]oxy]ethoxy]carbonyl]-3,6-dihydro-2*H*-1,3-thiazin-2-yl]acetyl]amino]thiazol-4-yl]-2-(methoxyimino)acetyl]amino]-3-(methoxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (cefpodoxime proxetil dimer).

Ph Eur

Cefprozil Monohydrate

(Ph. Eur. monograph 2342)



$C_{18}H_{19}N_3O_5S, H_2O$ 407.4 *121123-17-9*

Action and use
Cephalosporin antibacterial.

Ph Eur

DEFINITION
Mixture of the 2 diastereoisomers of (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-8-oxo-3-[(1*EZ*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or yellow, crystalline powder, slightly hygroscopic.

Solubility
Slightly soluble in water and in methanol, practically insoluble in acetone.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefprozil CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 0.125 g of the substance to be examined in 1 mL of a 103 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with mobile phase A.

Test solution (b) Dissolve 30.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *cefprozil for peak identification CRS* (containing impurities B, H and M) in 0.05 mL of a 103 g/L solution of *hydrochloric acid R* and add 1 mL of mobile phase A.

Reference solution (c) Dissolve 3 mg of *cefprozil CRS* and 6 mg of *cefprozil impurity mixture CRS* (containing impurities D and F) in 2 mL of a 103 g/L solution of *hydrochloric acid R* and dilute to 50 mL with mobile phase A.

Reference solution (d) Dissolve 30.0 mg of *cefprozil CRS* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (e) Dissolve 10.0 mg of *cefadroxil CRS* (impurity B) in *water R* and dilute to 20.0 mL with the same solvent. Dilute 1.0 mL of the solution to 20.0 mL with *water R.*

Reference solution (f) Dissolve 10.0 mg of *cefprozil impurity A CRS* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *water R.*

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm);
— *temperature:* 40 °C.

Mobile phase:
— *mobile phase A:* dissolve 11.5 g of *ammonium dihydrogen phosphate R* in *water for chromatography R*, adjust to pH 4.4 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R;*
— *mobile phase B: acetonitrile for chromatography R*, mobile phase A (50:50 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	81	19
8 - 20	81 → 36	19 → 64
20 - 25	36	64

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b), (c), (e) and (f).

Identification of impurities Use the chromatogram supplied with *cefprozil for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, H and M; use the chromatogram supplied with *cefprozil impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D and F; impurities G and I are identified by their relative retention.

Relative retention With reference to cefprozil (*Z*)-isomer (retention time = about 7 min): impurity A = about 0.4; impurity B = about 0.5; impurity D = about 0.7; impurity F = about 0.9; cefprozil (*E*)-isomer = about 1.4; impurity G = about 1.7; impurity H = about 2.0; impurity I = about 2.1; impurity M = about 2.9.

System suitability Reference solution (c):
— *resolution*: minimum 1.4 between the peaks due to impurity F and cefprozil (*Z*)-isomer.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 2.3;
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.5 per cent);
— *impurities D, G, H, I, M*: for each impurity, not more than 0.3 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (f) (0.2 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *total*: maximum 2.0 per cent;
— *disregard limit*: 0.05 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.05 per cent).

(*E*)-isomer ratio

Liquid chromatography (*2.2.29*) as described under Assay.

Determine the area of the peak due to the (*E*)-isomer in the chromatogram obtained with test solution (b) and reference solution (d). Calculate the ratio of the (*E*)-isomer to the sum of both cefprozil isomers, as determined under Assay.

Limit:
— *(E)-isomer ratio*: 0.06 to 0.11.

Water (*2.5.12*)

3.5 per cent to 6.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase B, mobile phase A (18:82 *V/V*).

Detection Spectrophotometer at 280 nm.

Injection 10 µL of test solution (b) and reference solution (d).

Run time Twice the retention time of cefprozil (*Z*)-isomer.

Elution order (*Z*)-isomer, (*E*)-isomer.

Retention time Cefprozil (*Z*)-isomer = about 8 min.

System suitability Reference solution (d):
— *resolution*: minimum 2.5 between the peaks due to cefprozil (*Z*)-isomer and the (*E*)-isomer.

Calculate the percentage content of the sum of both isomers of cefprozil ($C_{18}H_{19}N_3O_5S$) taking into account the assigned contents of both (*E*)-isomer and (*Z*)-isomer of *cefprozil CRS*.

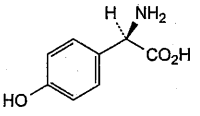
STORAGE

In an airtight container.

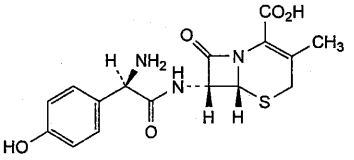
IMPURITIES

Specified impurities *A, B, D, G, H, I, M.*

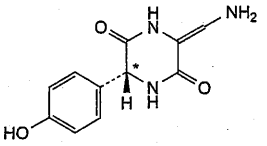
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, F, J, K, L, N.*



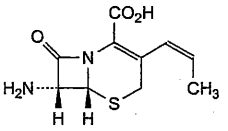
A. (2*R*)-2-amino-2-(4-hydroxyphenyl)acetic acid (*p*-hydroxyphenylglycine),



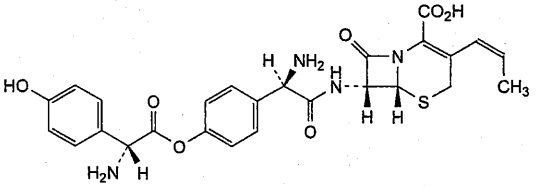
B. (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-(4-hydroxyphenyl) acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0] oct-2-ene-2-carboxylic acid (cefadroxil),



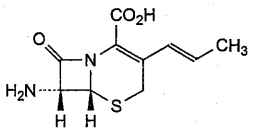
C. (6*RS*)-3-(aminomethylene)-6-(4-hydroxyphenyl)piperazine-2,5-dione,



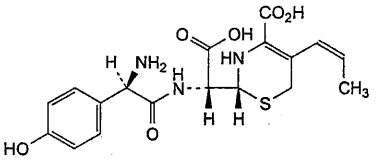
D. (6*R*,7*R*)-7-amino-8-oxo-3-[(1*Z*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



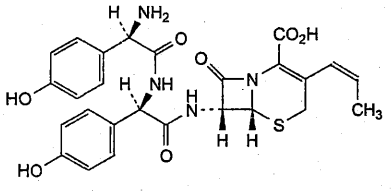
E. (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-[4-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]oxy]phenyl]acetyl]amino]-8-oxo-3-[(1*Z*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



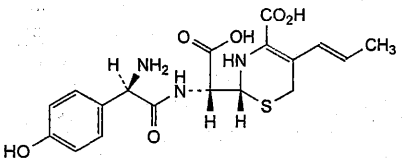
F. (6*R*,7*R*)-7-amino-8-oxo-3-[(1*E*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



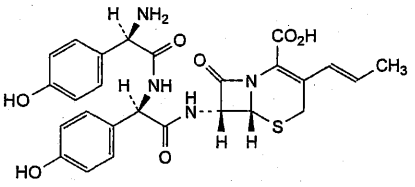
G. (2*R*)-2-[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino-2-[(2*R*)-4-carboxy-5-[(1*Z*)-prop-1-enyl]-3,6-dihydro-2*H*-1,3-thiazin-2-yl]-acetic acid,



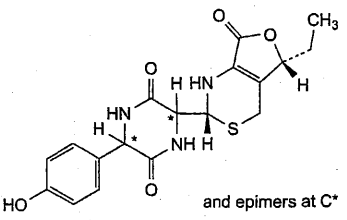
H. (6*R*,7*R*)-7-[[(2*R*)-2-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-8-oxo-3-[(1*Z*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



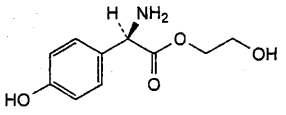
I. (2*R*)-2-[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino-2-[(2*R*)-4-carboxy-5-[(1*E*)-prop-1-enyl]-3,6-dihydro-2*H*-1,3-thiazin-2-yl]-acetic acid,



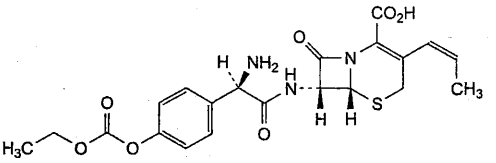
J. (6*R*,7*R*)-7-[[(2*R*)-2-[[(2*R*)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-2-(4-hydroxyphenyl)acetyl]amino]-8-oxo-3-[(1*E*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



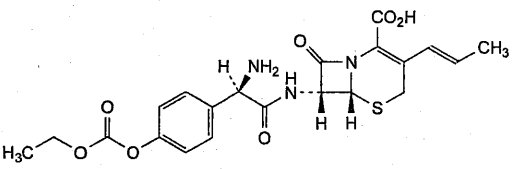
K. mixture of 4 diastereoisomers of (3*RS*,6*RS*)-3-[(2*R*,5*R*)-5-ethyl-7-oxo-1,2,5,7-tetrahydro-4*H*-furo[3,4-*d*][1,3]thiazin-2-yl]-6-(4-hydroxyphenyl)piperazine-2,5-dione,



L. 2-hydroxyethyl (2*R*)-2-amino-2-(4-hydroxyphenyl)acetate,



M. (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-[4-[(ethoxycarbonyl)oxy]phenyl]acetyl]amino]-8-oxo-3-[(1*Z*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,

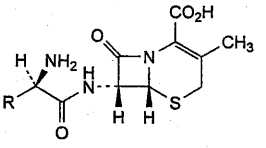


N. (6*R*,7*R*)-7-[[(2*R*)-2-amino-2-[4-(ethoxycarbonyl)oxy]phenyl]acetyl]amino]-8-oxo-3-[(1*E*)-prop-1-enyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid.

Ph Eur

Cefradine

(Ph. Eur. monograph 0814)



Compound	R	Mol. Formula	M_r
cefradine		$C_{16}H_{19}N_3O_4S$	349.4
cefalexin		$C_{16}H_{17}N_3O_4S$	347.4
4',5'-dihydrocefradine		$C_{16}H_{21}N_3O_4S$	351.4

Cefradine *38821-53-3*

Action and use
Cephalosporin antibacterial.

Preparations
Cefradine Capsules
Cefradine Injection
Cefradine Oral Suspension

Ph Eur

DEFINITION

Main component (6*R*,7*R*)-7-[[(2*R*)-amino(cyclohexa-1,4-dienyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (cefradine).

Semi-synthetic product derived from a fermentation product.

Content
— *cefradine*: minimum 90.0 per cent (anhydrous substance);
— *cefalexin*: maximum 5.0 per cent (anhydrous substance);
— *4',5'-dihydrocefradine*: maximum 2.0 per cent (anhydrous substance);

— *sum of the percentage contents of cefradine, cefalexin and 4′,5′-dihydrocefradine*: 96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly yellow, hygroscopic powder.

Solubility

Sparingly soluble in water, practically insoluble in ethanol (96 per cent) and in hexane.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefradine CRS.*

If the spectra obtained in the solid state show differences, dissolve 30 mg of the substance to be examined and 30 mg of the reference substance separately in 10 mL of *methanol R*, evaporate to dryness at 40 °C at a pressure less than 2 kPa and record new spectra using the residues.

TESTS

Solution S

Dissolve 2.50 g in *sodium carbonate solution R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*). Allow solution S to stand for 5 min. The absorbance (*2.2.25*) of solution S measured at 450 nm is not greater than 0.60.

pH (*2.2.3*)

3.5 to 6.0.

Dissolve 0.100 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)

+ 80.0 to + 90.0 (anhydrous substance).

Dissolve 0.250 g in *acetate buffer solution pH 4.6 R* and dilute to 25.0 mL with the same solution.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.300 g of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 3.0 mg of *cyclohexa-1,4-dienylglycine CRS* (impurity B) in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 3 mg of the substance to be examined and 3 mg of *cefalexin monohydrate CRS* in mobile phase A and dilute to 25 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (d) Dissolve 6 mg of *cefradine for peak identification CRS* (containing impurities C, D and E) in 1.0 mL of mobile phase A.

Reference solution (e) Dissolve the contents of a vial of *cefradine impurity mixture CRS* (impurities A and G) in 1.0 mL of mobile phase A.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase:

— *mobile phase A*: 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.0 with *dilute phosphoric acid R*;

— *mobile phase B*: *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2.5	99.5 → 97	0.5 → 3
2.5 - 11	97 → 75	3 → 25
11 - 13	75 → 60	25 → 40
13 - 16	60	40
16 - 19	60 → 20	40 → 80
19 - 19.1	20 → 99.5	80 → 0.5
19.1 - 25	99.5	0.5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 25 µL.

Identification of impurities Use the chromatogram supplied with *cefradine for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities C, D and E; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity B; use the chromatogram supplied with *cefradine impurity mixture CRS* and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities A and G.

Relative retention With reference to cefradine (retention time = about 15 min): impurity A = about 0.27; impurity B = about 0.32; impurity C = about 0.53; impurity D = about 0.63; impurity E = about 0.80; impurity F = about 0.92; cefalexin = about 0.95; 4′,5′-dihydrocefradine = about 1.06; impurity G = about 1.32.

System suitability Reference solution (b):

— *resolution*: minimum 4.0 between the peaks due to cefalexin and cefradine.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 3.4;

— *impurities A, B, C, D, E, F, G*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.25 per cent);

— *any other impurity*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.25 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent);

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard the peaks due to cefalexin and 4′,5′-dihydrocefradine.

***N,N*-Dimethylaniline** (*2.4.26, Method B*)

Maximum 20 ppm.

Water (*2.5.12*)

Maximum 6.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *phosphate buffer solution pH 5.0 R* and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 50.0 mg of *cefradine CRS* (containing 4′,5′-dihydrocefradine) in *phosphate buffer solution pH 5.0 R* and dilute to 100.0 mL with the same solution.

Reference solution (b) Dissolve 5.0 mg of *cefalexin monohydrate CRS* in *phosphate buffer solution pH 5.0 R* and dilute to 100.0 mL with the same solution.

Reference solution (c) Dilute 1 mL of reference solution (a) to 10 mL with *phosphate buffer solution pH 5.0 R*. Mix 5 mL of this solution and 5 mL of reference solution (b).

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, phosphate buffer solution pH 5.0 R* (25:75 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL.

Run time Twice the retention time of cefradine.

Relative retention With reference to cefradine (retention time = about 3 min): cefalexin = about 0.7; 4′,5′-dihydrocefradine = about 1.5.

System suitability Reference solution (c):
— *resolution*: minimum 4.0 between the peaks due to cefalexin and cefradine.

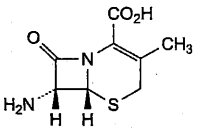
Calculate the percentage content of cefradine using the chromatogram obtained with reference solution (a) and taking into account the assigned content of *cefradine CRS*. Calculate the percentage content of cefalexin using the chromatogram obtained with reference solution (b) and taking into account the assigned content of *cefalexin monohydrate CRS*. Calculate the percentage content of 4′,5′-dihydrocefradine using the chromatogram obtained with reference solution (b), taking into account the assigned content of *cefalexin monohydrate CRS* and multiplying the area of the peak due to 4′,5′-dihydrocefradine by a correction factor of 1.6.

STORAGE

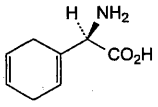
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

IMPURITIES

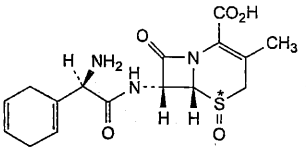
Specified impurities *A, B, C, D, E, F, G.*



A. (6*R*,7*R*)-7-amino-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-aminodeacetoxycephalosporanic acid, 7-ADCA),

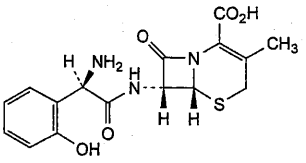


B. (2*R*)-amino(cyclohexa-1,4-dienyl)acetic acid (D-dihydrophenylglycine, cyclohexa-1,4-dienylglycine),

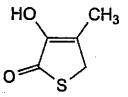


C. (6*R*,7*R*)-7-[[(2*R*)-amino(cyclohexa-1,4-dienyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid 5-oxide (isomer 1),

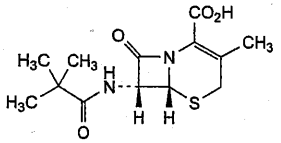
D. (6*R*,7*R*)-7-[[(2*R*)-amino(cyclohexa-1,4-dienyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid 5-oxide (isomer 2),



E. (6*R*,7*R*)-7-[[(2*R*)-amino(2-hydroxyphenyl)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



F. 3-hydroxy-4-methylthiophen-2(5*H*)-one,



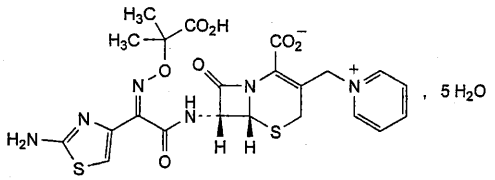
G. (6*R*,7*R*)-7-[(2,2-dimethylpropanoyl)amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (7-ADCA pivalamide).

Ph Eur

Ceftazidime Pentahydrate

Ceftazidime

(Ph. Eur. monograph 1405)



$C_{22}H_{22}N_6O_7S_2,5H_2O$ 637 *78439-06-2*

Action and use

Cephalosporin antibacterial.

Preparations

Ceftazidime Eye Drops

Ceftazidime for Injection

Ceftazidime Injection

Ph Eur

DEFINITION

(6*R*,7*R*)-7-[[(2*Z*)-2-(2-Aminothiazol-4-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate pentahydrate.

Semi-synthetic product derived from a fermentation product.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water and in methanol, practically insoluble in acetone and in ethanol (96 per cent). It dissolves in acid and alkali solutions.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ceftazidime CRS.*

TESTS

Solution S

Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

3.0 to 4.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Suspend 0.150 g of the substance to be examined in 5 mL of *acetonitrile R*, dissolve by adding *water R* and dilute to 100 mL with *water R*.

Reference solution (a) To 1.0 mL of the test solution add 5.0 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 5.0 mL with *water R*.

Reference solution (b) In order to prepare impurity B *in situ*, expose 5 mL of the test solution to ultraviolet light at 254 nm for about 24 h.

Reference solution (c) Dissolve the contents of a vial of *ceftazidime for peak identification CRS* (containing impurities A and G) in 2.0 mL of *water R*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: solution containing 3.6 g/L of *disodium hydrogen phosphate dodecahydrate R* and 1.4 g/L of *potassium dihydrogen phosphate R*, adjusted to pH 3.4 with a 10 per cent *V/V* solution of *phosphoric acid R*;

— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	96 → 89	4 → 11
4 - 5	89	11
5 - 8	89 → 84	11 → 16
8 - 11	84 → 80	16 → 20
11 - 15	80 → 50	20 → 50
15 - 18	50 → 20	50 → 80
18 - 22	20	80

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Relative retention With reference to ceftazidime (retention time = about 8 min): impurity F = about 0.4; impurity G = about 0.8; impurity A = about 0.9; impurity B = about 1.4.

Identification of impurities Use the chromatogram supplied with *ceftazidime for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and G; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

System suitability Reference solution (c):

— *resolution*: minimum 4.0 between the peaks due to impurity A and ceftazidime.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity G by 3.0;

— *impurities A, B, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to impurity F.

Impurity F

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Phosphate buffer solution Prepare a 10 per cent *V/V* solution of *phosphate buffer solution pH 7.0 R4*.

Test solution Dissolve 0.500 g of the substance to be examined in phosphate buffer solution and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 1.00 g of *pyridine R* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 200.0 mL with *water R*. Dilute 1.0 mL of this solution to 100.0 mL with phosphate buffer solution.

Reference solution (b) Dilute 1 mL of the test solution to 200 mL with phosphate buffer solution. To 1 mL of this solution add 20 mL of reference solution (a) and dilute to 200 mL with phosphate buffer solution.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 8 volumes of a 28.8 g/L solution of *ammonium dihydrogen phosphate R* previously adjusted to pH 7.0 with *ammonia R*, 24 volumes of *acetonitrile R* and 68 volumes of *water R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 255 nm.

Injection 20 µL.

Run time 10 min.

System suitability Reference solution (b):

— *resolution*: minimum 7.0 between the peaks due to ceftazidime and impurity F.

Limit:

— *impurity F*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (500 ppm).

Water (*2.5.12*)

13.0 per cent to 15.0 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *ceftazidime CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *ceftazidime for peak identification CRS* (containing impurities A and G) in 3.0 mL of the mobile phase.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *hexylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 4.3 g of *disodium hydrogen phosphate dodecahydrate R* and 2.7 g of *potassium dihydrogen phosphate R* in 980 mL of *water R*, then add 20 mL of *acetonitrile R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 20 µL.

Run time 6 min.

Relative retention With reference to ceftazidime (retention time = about 4.5 min): impurity A = about 0.7.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the peaks due to impurity A and ceftazidime.

Calculate the content of ceftazidime ($C_{22}H_{22}N_6O_7S_2$) taking into account the assigned content of $C_{22}H_{22}N_6O_7S_2$ in *ceftazidime CRS*.

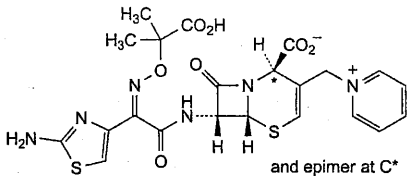
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

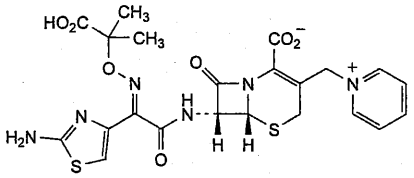
IMPURITIES

Specified impurities *A, B, F, G.*

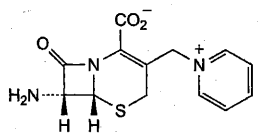
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, H.*



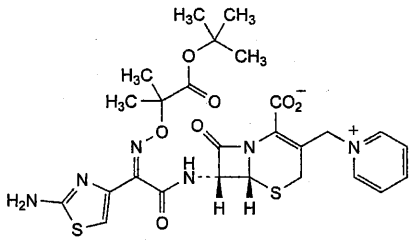
A. (2*RS*,6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylate (Δ-2-ceftazidime),



B. (6*R*,7*R*)-7-[[(2*E*)-2-(2-aminothiazol-4-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

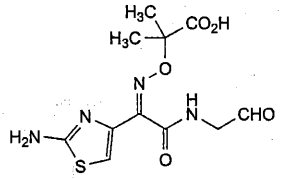


C. (6*R*,7*R*)-7-amino-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

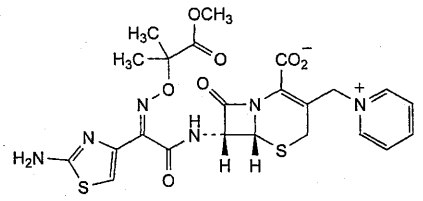


E. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[[2-(1,1-dimethylethoxy)-1,1-dimethyl-2-oxoethoxy]imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

F. pyridine,



G. 2-[[[(1*Z*)-1-(2-aminothiazol-4-yl)-2-[(oxoethyl)amino]-2-oxoethylidene]amino]oxy]-2-methylpropanoic acid,



H. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[(2-methoxy-1,1-dimethyl-2-oxoethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Ph Eur

Ceftazidime Pentahydrate with Sodium Carbonate for Injection

(Ph. Eur. monograph 2344)

Action and use
Cephalosporin antibacterial.

Preparations
Ceftazidime Eye Drops
Ceftazidime for Injection
Ceftazidime Injection

Ph Eur

DEFINITION

Sterile mixture of *Ceftazidime pentahydrate (1405)* and *Anhydrous sodium carbonate (0773)*.

Semi-synthetic product derived from a fermentation product.

Content
— *ceftazidime*: 93.0 per cent to 105.0 per cent (dried and carbonate-free substance);
— *sodium carbonate*: 8.0 per cent to 10.0 per cent.

CHARACTERS

Appearance
White or pale yellow powder.

Solubility
Freely soluble in water and in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. It gives the reaction of carbonates (*2.3.1*).

TESTS

Solution S
Dissolve 2.60 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 425 nm is not greater than 0.50.

pH (*2.2.3*)
5.0 to 7.5 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Suspend 0.150 g of the substance to be examined in 5 mL of *acetonitrile R*, dissolve by adding *water R* and dilute to 100 mL with *water R*.

Reference solution (a) To 1.0 mL of the test solution add 5.0 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 5.0 mL with *water R*.

Reference solution (b) In order to prepare impurity B *in situ*, expose 5 mL of the test solution to ultraviolet light at 254 nm for about 24 h.

Reference solution (c) Dissolve the contents of a vial of *ceftazidime for peak identification CRS* (containing impurities A and G) in 2.0 mL of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: solution containing 3.6 g/L of *disodium hydrogen phosphate dodecahydrate R* and 1.4 g/L of *potassium dihydrogen phosphate R*, adjusted to pH 3.4 with a 10 per cent *V/V* solution of *phosphoric acid R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	96 → 89	4 → 11
4 - 5	89	11
5 - 8	89 → 84	11 → 16
8 - 11	84 → 80	16 → 20
11 - 15	80 → 50	20 → 50
15 - 18	50 → 20	50 → 80
18 - 22	20	80

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Relative retention With reference to ceftazidime (retention time = about 8 min): impurity F = about 0.4; impurity G = about 0.8; impurity A = about 0.9; impurity B = about 1.4.

Identification of impurities Use the chromatogram supplied with *ceftazidime for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and G; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

System suitability Reference solution (c):
— *resolution*: minimum 4.0 between the peaks due to impurity A and ceftazidime.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity G by 3.0;
— *impurities A, B, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to impurity F.

Impurity F

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Phosphate buffer solution Prepare a 10 per cent *V/V* solution of *phosphate buffer solution pH 7.0 R4.*

Test solution Dissolve 0.500 g of the substance to be examined in phosphate buffer solution and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 1.00 g of *pyridine R* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 200.0 mL with *water R*. Dilute 1.0 mL of this solution to 100.0 mL with phosphate buffer solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with phosphate buffer solution. To 1.0 mL of this solution add 20.0 mL of reference solution (a) and dilute to 200.0 mL with phosphate buffer solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 8 volumes of a 28.8 g/L solution of *ammonium dihydrogen phosphate R* previously adjusted to pH 7.0 with *ammonia R*, 24 volumes of *acetonitrile R* and 68 volumes of *water R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 255 nm.

Injection 20 µL.

Run time 10 min.

System suitability Reference solution (b):
— *resolution*: minimum 7.0 between the peaks due to ceftazidime and impurity F.

Limit:
— *impurity F*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent).

Loss on drying (*2.2.32*)

Maximum 13.5 per cent, determined on 0.300 g. Dry at 25 °C at a pressure not exceeding 0.67 kPa for 4 h then heat the residue at 100 °C at a pressure not exceeding 0.67 kPa for 3 h.

Bacterial endotoxins (*2.6.14*)

Less than 0.10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Ceftazidime

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *ceftazidime CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *ceftazidime for peak identification CRS* (containing impurities A and G) in 3.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *hexylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 4.3 g of *disodium hydrogen phosphate dodecahydrate R* and 2.7 g of *potassium dihydrogen phosphate R* in 980 mL of *water R*, then add 20 mL of *acetonitrile R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 20 µL.

Run time 6 min.

Relative retention With reference to ceftazidime (retention time = about 4.5 min): impurity A = about 0.7.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity A and ceftazidime.

Calculate the content of ceftazidime ($C_{22}H_{22}N_6O_7S_2$) taking into account the assigned content of $C_{22}H_{22}N_6O_7S_2$ in *ceftazidime CRS*.

Sodium carbonate

Atomic absorption spectrometry (*2.2.23, Method I*).

Caesium chloride buffer solution To 12.7 g of *caesium chloride R* add 500 mL of *water R* and 86 mL of *hydrochloric acid R* and dilute to 1000.0 mL with *water R*.

Sodium standard solution (1000 mg/L) Dissolve 3.70 g of *sodium nitrate R* in *water R* and dilute to 500 mL with the same solvent, add 48.5 g of *nitric acid R* and dilute to 1000 mL with *water R*.

Test solution Dissolve 650.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent. To 10.0 mL of this solution add 5.0 mL of caesium chloride buffer solution and dilute to 50.0 mL with *water R*.

Reference solution Into 4 identical flasks, each containing 20.0 mL of caesium chloride buffer solution, introduce respectively 0 mL, 5.00 mL, 10.00 mL and 15.00 mL of sodium standard solution (1000 mg/L) and dilute to 200.0 mL with *water R*.

Source Sodium hollow-cathode lamp.

Wavelength 330.2 nm to 330.3 nm.

Atomisation device Air-acetylene flame.

Calculate the percentage content of sodium carbonate.

STORAGE

In a sterile, airtight, tamper-proof container, protected from light and humidity.

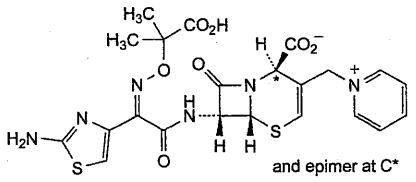
LABELLING

The label states the percentage content *m/m* of ceftazidime.

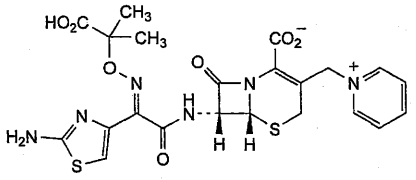
IMPURITIES

Specified impurities *A, B, F, G.*

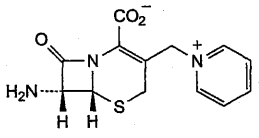
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, H.*



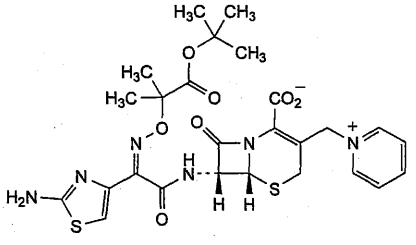
A. (2*RS*,6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylate (Δ-2-ceftazidime),



B. (6*R*,7*R*)-7-[[(2*E*)-2-(2-aminothiazol-4-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

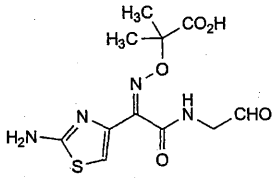


C. (6*R*,7*R*)-7-amino-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

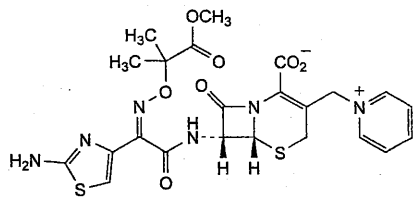


E. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[[2-(1,1-dimethylethoxy)-1,1-dimethyl-2-oxoethoxy]imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate,

F. pyridine,



G. 2-[[[(1*Z*)-1-(2-aminothiazol-4-yl)-2-[(oxoethyl)amino]-2-oxoethylidene]amino]oxy]-2-methylpropanoic acid,

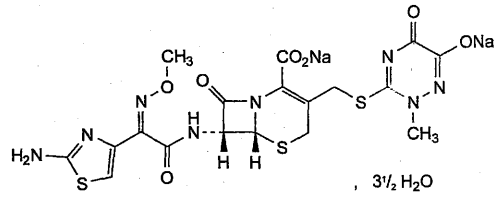


H. (6*R*,7*R*)-7-[[(2*Z*)-2-(2-aminothiazol-4-yl)-2-[(2-methoxy-1,1-dimethyl-2-oxoethoxy)imino]acetyl]amino]-8-oxo-3-[(pyridin-1-ium-1-yl)methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Ph Eur

Ceftriaxone Sodium

(Ph. Eur. monograph 0991)



$C_{18}H_{16}N_8Na_2O_7S_3,3½H_2O$ 662 *104376-79-6*

Action and use
Cephalosporin antibacterial.

Preparation
Ceftriaxone Injection

Ph Eur

DEFINITION
Disodium (6*R*,7*R*)-7-[[(2*Z*)-(2-aminothiazol-4-yl)(methoxyimino)acetyl]amino]-3-[[(2-methyl-6-oxido-5-oxo-2,5-dihydro-1,2,4-triazin-3-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate 3.5 hydrate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Almost white or yellowish, slightly hygroscopic, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in methanol, very slightly soluble in anhydrous ethanol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *ceftriaxone sodium CRS.*
B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.40 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 or BY_5 (*2.2.2*).
Dilute 2 mL of solution S to 20 mL with *water R.*

pH (*2.2.3*)
6.0 to 8.0 for solution S.

Specific optical rotation (*2.2.7*)
−155 to −170 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 30.0 mg of *ceftriaxone sodium CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *ceftriaxone sodium CRS* and 5.0 mg of *ceftriaxone impurity A CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 2.0 g of *tetradecylammonium bromide R* and 2.0 g of *tetraheptylammonium bromide R* in a mixture of 440 mL of *water R*, 55 mL of *0.067 M phosphate buffer solution pH 7.0 R*, 5.0 mL of citrate buffer solution pH 5.0 prepared by dissolving 20.17 g of *citric acid monohydrate R* in 800 mL of *water R*, adjusting to pH 5.0 with *strong sodium hydroxide solution R* and diluting to 1000.0 mL with *water R*, and 500 mL of *acetonitrile R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of ceftriaxone.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to ceftriaxone and impurity A.

Limits:
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (4.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

***N,N*-Dimethylaniline** (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.8 per cent *m/m*.

Water (*2.5.12*)
8.0 per cent to 11.0 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.08 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

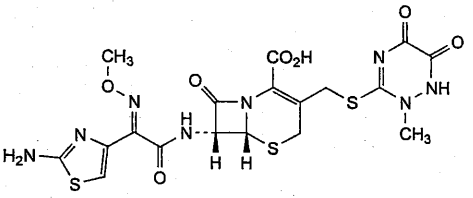
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

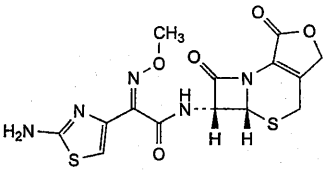
Calculate the percentage content of $C_{18}H_{16}N_8Na_2O_7S_3$ from the declared content of *ceftriaxone sodium CRS*.

STORAGE
In an airtight container protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

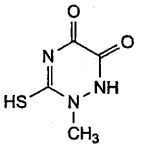
IMPURITIES



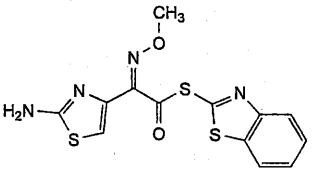
A. (6*R*,7*R*)-7-[[(2*E*)-(2-aminothiazol-4-yl)(methoxyimino)acetyl]amino]-3-[[(2-methyl-5,6-dioxo-1,2,5,6-tetrahydro-1,2,4-triazin-3-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (*(E)*-isomer),



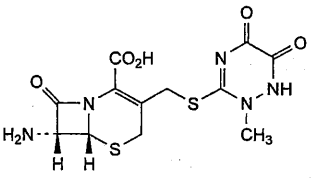
B. (5a*R*,6*R*)-6-[[(2*Z*)-(2-aminothiazol-4-yl)(methoxyimino)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione,



C. 2-methyl-3-sulfanyl-1,2-dihydro-1,2,4-triazine-5,6-dione,



D. *S*-benzothiazol-2-yl (2*Z*)-(2-aminothiazol-4-yl)(methoxyimino)thioacetate,

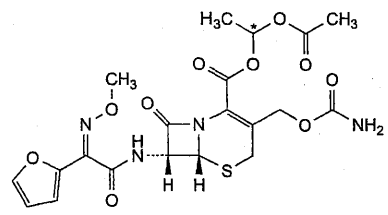


E. (6*R*,7*R*)-7-amino-3-[[(2-methyl-5,6-dioxo-1,2,5,6-tetrahydro-1,2,4-triazin-3-yl)sulfanyl]methyl]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid.

Ph Eur.

Cefuroxime Axetil

(Ph. Eur. monograph 1300)



$C_{20}H_{22}N_4O_{10}S$ 510.5 *64544-07-6*

Action and use
Cephalosporin antibacterial.

Preparations
Cefuroxime Axetil Oral Suspension
Cefuroxime Axetil Tablets

Ph Eur

DEFINITION

Mixture of the 2 diastereoisomers of (1*RS*)-1-(acetyloxy)ethyl (6*R*,7*R*)-3-[(carbamoyloxy)methyl]-7-[[(*Z*)-2-(furan-2-yl)-2(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Slightly soluble in water, soluble in acetone, in ethyl acetate and in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cefuroxime axetil CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the peaks due to cefuroxime axetil diastereoisomers A and B in the chromatogram obtained with reference solution (d).

TESTS

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the test solution and reference solution (d) immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) In order to prepare *in situ* impurity A, heat 5 mL of the test solution at 60 °C for 1 h.

Reference solution (c) In order to prepare *in situ* impurity B, expose 5 mL of the test solution to ultraviolet light at 254 nm for 24 h.

Reference solution (d) Dissolve 10.0 mg of *cefuroxime axetil CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *trimethylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R*, 23 g/L solution of *ammonium dihydrogen phosphate R* (38:62 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 278 nm.

Injection 20 µL of the test solution and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the pair of peaks due to impurity A and use the chromatogram obtained with reference solution (c) to identify the pair of peaks due to impurity B.

Relative retention With reference to cefuroxime axetil diastereoisomer A: cefuroxime axetil diastereoisomer B = about 0.9, impurity A = about 1.2; impurity B = 1.7 and 2.1.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to cefuroxime axetil diastereoisomer A and impurity A.

Limits:
— *impurity A*: maximum 1.5 per cent for the sum of the pair of peaks;
— *impurity B*: maximum 1.0 per cent for the sum of the pair of peaks;
— *impurity E*: maximum 0.5 per cent;
— *any other impurity*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 3.0 per cent;
— *disregard limit*: 0.05 times the area of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.05 per cent).

Diastereoisomer ratio
Liquid chromatography (*2.2.29*) as described in the test for related substances.

Limit Test solution:
— the ratio of the area of the peak due to cefuroxime axetil diastereoisomer A to the sum of the areas of the peaks due to cefuroxime axetil diastereoisomers A and B is between 0.48 and 0.55.

Acetone (*2.4.24*)
Maximum 1.1 per cent.

Water (*2.5.12*)
Maximum 1.5 per cent, determined on 0.400 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (d).

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to cefuroxime axetil diastereoisomers A and B;
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the sum of the peaks due to cefuroxime axetil diastereoisomers A and B after 6 injections.

Calculate the percentage content of $C_{20}H_{22}N_4O_{10}S$ from the sum of the areas of the 2 diastereoisomer peaks and the declared content of $C_{20}H_{22}N_4O_{10}S$ in *cefuroxime axetil CRS*.

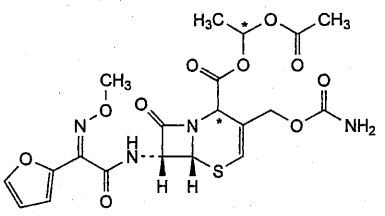
STORAGE

In an airtight container, protected from light.

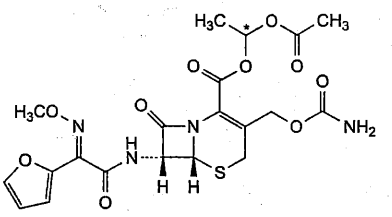
IMPURITIES

Specified impurities *A, B, E.*

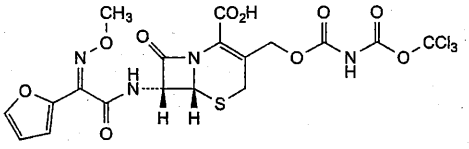
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D.*



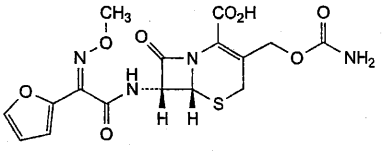
A. 1-(acetyloxy)ethyl (6*R*,7*R*)-3-[(carbamoyloxy)methyl]-7-[[(*Z*)-2-(furan-2-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylate (Δ^3-isomers),



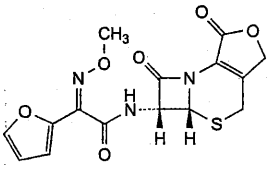
B. (1*RS*)-1-(acetyloxy)ethyl (6*R*,7*R*)-3-[(carbamoyloxy)methyl]-7-[[(*E*)-2-(furan-2-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2carboxylate ((*E*)-isomers),



C. (6*R*,7*R*)-7-[[(*Z*)-2-(furan-2-yl)-2-(methoxyimino)acetyl]amino]-8-oxo-3-[[[(trichloroacetyl)carbamoyl]oxy]methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



D. cefuroxime.

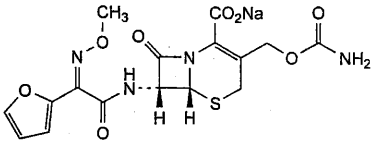


E. (5a*R*,6*R*)-6-[[(2*Z*)-2-(furan-2-yl)-2-(methoxyimino)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,4-*d*][1,3]thiazine-1,7(4*H*)-dione (descarbamoylcefuroxime lactone).

Ph Eur

Cefuroxime Sodium

(Ph. Eur. monograph 0992)



$C_{16}H_{15}N_4NaO_8S$ 446.4 *56238-63-2*

Action and use
Cephalosporin antibacterial.

Preparations
Cefuroxime Eye Drops
Cefuroxime Injection
Cefuroxime Intracameral Injection

Ph Eur

DEFINITION

Sodium (6*R*,7*R*)-3-[(carbamoyloxy)methyl]-7-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, slightly hygroscopic powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cefuroxime sodium CRS.*

B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*). The absorbance (*2.2.25*) of solution S measured at 450 nm is not greater than 0.25.

pH (*2.2.3*)
5.5 to 8.5.

Dilute 2 mL of solution S to 20 mL with *carbon dioxide-free water R.*

Specific optical rotation (*2.2.7*)
+ 59 to + 66 (anhydrous substance).

Dissolve 0.500 g in *acetate buffer solution pH 4.6 R* and dilute to 25.0 mL with the same buffer solution.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use or keep at 2-8 °C.*

Test solution (a) Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with *water R.*

Reference solution (a) Dissolve 25.0 mg of *cefuroxime sodium CRS* in *water R* and dilute to 25.0 mL with the same solvent. Dilute 5.0 mL to 50.0 mL with *water R*.

Reference solution (b) Place 20 mL of reference solution (a) in a water-bath at 80 °C for 15 min. Cool and inject immediately.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with *water R*.

Column:
— *size*: *l* = 0.125 m, Ø = 4.6 mm;
— *stationary phase*: *hexylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 1 volume of *acetonitrile R* and 99 volumes of an acetate buffer solution pH 3.4, prepared by dissolving 6.01 g of *glacial acetic acid R* and 0.68 g of *sodium acetate R* in *water R* and diluting to 1000 mL with the same solvent.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 µL loop injector; inject test solution (a) and reference solutions (b) and (c).

Run time 4 times the retention time of cefuroxime.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to cefuroxime and impurity A.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *any other impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.5 per cent *m/m*.

Water (*2.5.12*)
Maximum 3.5 per cent, determined on 0.400 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

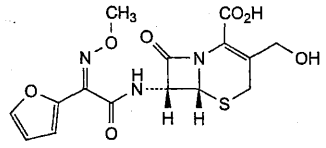
Injection Test solution (b) and reference solution (a).

Calculate the percentage content of cefuroxime sodium.

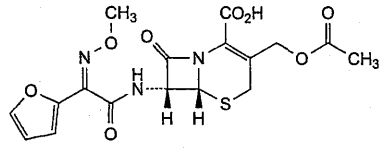
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

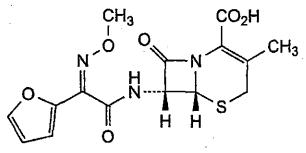
IMPURITIES



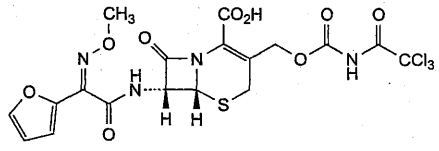
A. (6*R*,7*R*)-7-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-3-(hydroxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (descarbamoylcefuroxime),



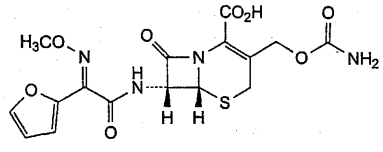
B. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



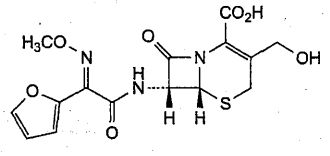
C. (6*R*,7*R*)-7-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



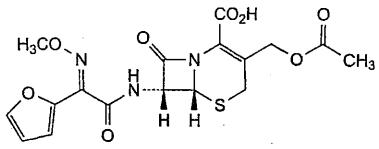
D. (6*R*,7*R*)-7-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-8-oxo-3-[[[(trichloroacetyl)carbamoyl]oxy]methyl]-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



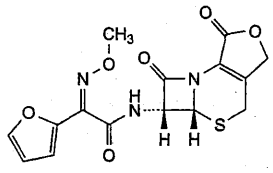
E. (6*R*,7*R*)-3-[(carbamoyloxy)methyl]-7-[[(*E*)-(furan-2-yl)(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (*trans*-cefuroxime),



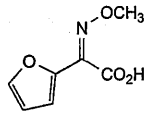
F. (6*R*,7*R*)-7-[[(*E*)-(furan-2-yl)(methoxyimino)acetyl]amino]-3-(hydroxymethyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



G. (6*R*,7*R*)-3-[(acetyloxy)methyl]-7-[[(*E*)-(furan-2-yl)(methoxyimino)acetyl]amino]-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid,



H. (5a*R*,6*R*)-6-[[(*Z*)-(furan-2-yl)(methoxyimino)acetyl]amino]-5a,6-dihydro-3*H*,7*H*-azeto[2,1-*b*]furo[3,3,4-*d*][1,3]thiazine-1,7(4*H*)-dione,

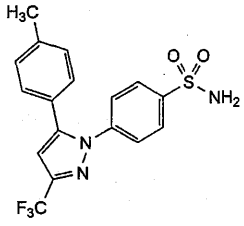


I. (*Z*)-(furan-2-yl)(methoxyimino)acetic acid.

Ph Eur

Celecoxib

(Ph. Eur. monograph 2591)



$C_{17}H_{14}F_3N_3O_2S$ 381.4 *169590-42-5*

Action and use
Cyclo-oxygenase (COX-2) inhibitor; analgesic; anti-inflammatory.

Preparation
Celecoxib Capsules

Ph Eur

DEFINITION
4-[5-(4-Methylphenyl)-3-(trifluoromethyl)-1*H*-pyrazol-1-yl]benzenesulfonamide.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline or amorphous powder.

Solubility
Practically insoluble in water, freely soluble to soluble in anhydrous ethanol, soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *celecoxib CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *2-propanol R*, evaporate to dryness and record new spectra using the residues.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *water R, methanol R2* (25:75 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of *celecoxib CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 3 mg of *celecoxib impurity A CRS* and 3 mg of *celecoxib impurity B CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 25.0 mL with reference solution (a).

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped phenylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 60 °C.

Mobile phase Mix 10 volumes of *acetonitrile R1*, 30 volumes of *methanol R2* and 60 volumes of a 2.7 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 25 µL of the test solution and reference solutions (b) and (c).

Run time 1.5 times the retention time of celecoxib.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to celecoxib (retention time = about 27 min): impurity A = about 0.9; impurity B = about 1.1.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to impurity A and celecoxib and minimum 1.8 between the peaks due to celecoxib and impurity B in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
— for all impurities, use the concentration of celecoxib in reference solution (c).

Limits:
— *impurity A*: maximum 0.4 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 0.400 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g in a platinum crucible.

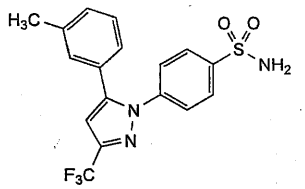
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

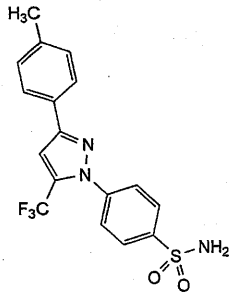
Calculate the percentage content of $C_{17}H_{14}F_3N_3O_2S$ taking into account the assigned content of *celecoxib CRS*.

IMPURITIES
Specified impurities *A*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B*.



A. 4-[5-(3-methylphenyl)-3-(trifluoromethyl)-1*H*-pyrazol-1-yl]benzenesulfonamide,

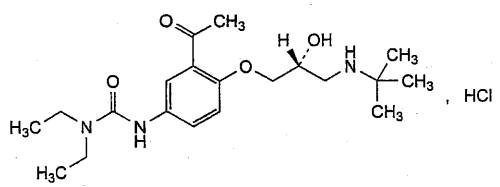


B. 4-[3-(4-methylphenyl)-5-(trifluoromethyl)-1*H*-pyrazol-1-yl]benzenesulfonamide.

Ph Eur

Celiprolol Hydrochloride

(Ph. Eur. monograph 1632)



and enantiomer

$C_{20}H_{34}ClN_3O_4$ 416.0 *57470-78-7*

Action and use
Beta-adrenoceptor antagonist.

Preparation
Celiprolol Tablets

Ph Eur

DEFINITION
3-[3-Acetyl-4-[(2*RS*)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]phenyl]-1,1-diethylurea hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or very slightly yellow, crystalline powder.

Solubility
Freely soluble in water and in methanol, soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *celiprolol hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Optical rotation (*2.2.7*)
−0.10° to + 0.10°.

Dissolve 1.0 g in *water R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dissolve 2 mg of the substance to be examined and 2 mg of *acebutolol hydrochloride R* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 10 mg of the substance to be examined in 2 mL of mobile phase A and allow to stand for 24 h (for identification of impurity A).

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (d) Dissolve 10 mg of *celiprolol for peak identification CRS* in mobile phase A and dilute to 2 mL with mobile phase A.

Reference solution (e) *This solution is only prepared if required (see below) and is used to determine the identity of impurity I which co-elutes with impurity H (the 2 impurities originate from different routes of synthesis).* Dissolve the contents of a vial of *celiprolol impurity I CRS* in mobile phase A and dilute to 2.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 91 mL of *tetrahydrofuran R*, 63 mL of *acetonitrile R1*, 0.6 mL of *pentafluoropropanoic acid R* and

0.2 mL of *trifluoroacetic acid R*; dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 50	100 → 80	0 → 20
50 - 51	80 → 100	20 → 0
51 - 65	100	0

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *celiprolol for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B, E and F.

Relative retention With reference to celiprolol (retention time = about 10 min): impurity A = about 0.3; impurity D = about 0.7; impurity G = about 1.2; impurity B = about 1.4; impurity F = about 1.6; impurity C = about 2.2; impurity H or I = about 2.5; impurity E = about 3.9.

System suitability Reference solution (a):
— *resolution*: minimum 4.0 between the peaks due to celiprolol and acebutolol.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 4.0; impurity B = 1.5; impurity E = 2.3; impurity F = 0.5; impurity I = 1.7;
— *any impurity*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent), and not more than 1 such peak has an area greater than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— if any of the above limits are exceeded and if a peak occurs with a relative retention of about 2.5 (impurity H or I), the identity of this peak has to be clarified by use of a UV spectrum recorded with a diode array detector; if this spectrum is different from the one obtained with reference solution (e), no correction factor is applied;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Dissolve 0.350 g under an atmosphere of nitrogen in 50 mL of *ethanol (96 per cent) R* and add 1.0 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

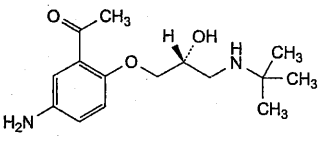
1 mL of *0.1 M sodium hydroxide* is equivalent to 41.60 mg of $C_{20}H_{34}ClN_3O_4$.

STORAGE

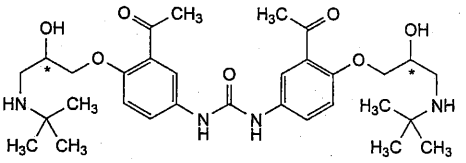
Protected from light.

IMPURITIES

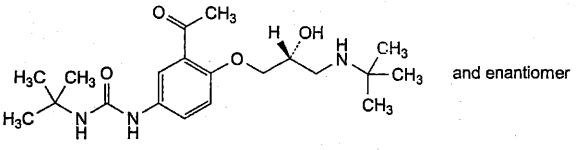
Specified impurities *A, B, C, D, E, F, G, H, I.*



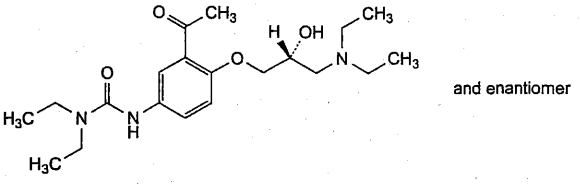
A. 1-[5-amino-2-[(2*RS*)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]phenyl]ethanone,



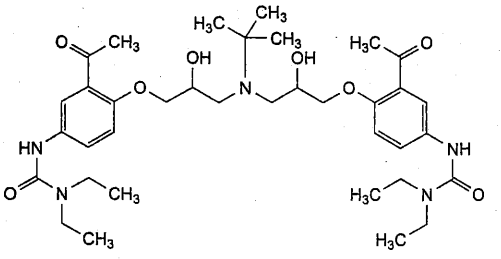
B. 1,3-bis[3-acetyl-4-[3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]phenyl]urea,



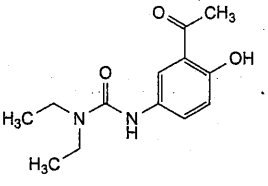
C. 1-[3-acetyl-4-[(2*RS*)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]phenyl]-3-(1,1-dimethylethyl)urea,



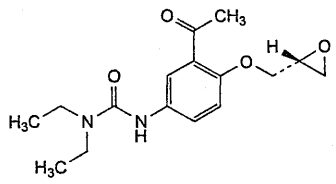
D. 3-[3-acetyl-4-[(2*RS*)-3-(diethylamino)-2-hydroxypropoxy]phenyl]-1,1-diethylurea,



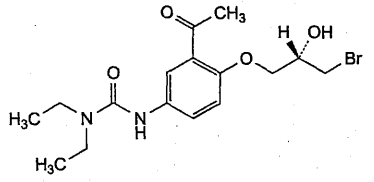
E. 1,1′-[[(1,1-dimethylethyl)imino]bis[(2-hydroxypropane-1,3-diyl)oxy(3-acetyl-1,4-phenylene)]]bis(3,3-diethylurea),



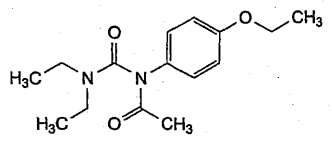
F. 3-(3-acetyl-4-hydroxyphenyl)-1,1-diethylurea,



G. 3-[3-acetyl-4-[[(*RS*)-oxiranyl]methoxy]phenyl]-1,1-diethylurea,



H. 3-[3-acetyl-4-[(2*RS*)-3-bromo-2-hydroxypropoxy]phenyl]-1,1-diethylurea (bromhydrin compound),



I. 1-acetyl-1-(4-ethoxyphenyl)-3,3-diethylurea.

Ph Eur

Cellacefate[1]

(Cellulose Acetate Phthalate, Ph. Eur. monograph 0314)

9004-38-0

Action and use
Enteric coating in pharmaceutical products.

Ph Eur

DEFINITION
Partly *O*-acetylated and *O*-phthalylated cellulose.

Content
— *phthaloyl groups* ($C_8H_5O_3$; M_r 149.1): 30.0 per cent to 36.0 per cent (anhydrous and acid-free substance);
— *acetyl groups* (C_2H_3O; M_r 43.04): 21.5 per cent to 26.0 per cent (anhydrous and acid-free substance).

♦CHARACTERS
Appearance
White or almost white, free-flowing powder or colourless flakes, hygroscopic.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in diethylene glycol, practically insoluble in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.♦

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cellulose acetate phthalate CRS.*

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

TESTS
Viscosity (*2.2.9*)
45 mPa·s to 90 mPa·s, determined at 25 ± 0.2 °C.
Dissolve 15 g, calculated with reference to the anhydrous substance, in 85 g of a mixture of 1 part by mass of *water R* and 249 parts by mass of *acetone R.*

Free acid
Maximum 3.0 per cent, calculated as phthalic acid (anhydrous substance).
Shake 3.0 g for 2 h with 100 mL of a 50 per cent *V/V* solution of *methanol R* and filter. Wash the flask and the filter with 2 quantities, each of 10 mL, of a 50 per cent *V/V* solution of *methanol R.* Combine the filtrate and washings, add 0.1 mL of *phenolphthalein solution R1* and titrate with *0.1 M sodium hydroxide* until a faint pink colour is obtained. Carry out a blank titration using 120 mL of a 50 per cent *V/V* solution of *methanol R.*
1 mL of *0.1 M sodium hydroxide* is equivalent to 8.3 mg of free acid, calculated as phthalic acid.

Water (*2.5.12*)
Maximum 5.0 per cent, determined on 0.500 g.
Carry out the test using a mixture of 2 volumes of *methylene chloride R* and 3 volumes of *anhydrous ethanol R.*

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Phthaloyl groups
Dissolve 1.000 g in 50 mL of a mixture of 2 volumes of *acetone R* and 3 volumes of *ethanol (96 per cent) R.*
Add about 0.1 mL of *phenolphthalein solution R1* and titrate with *0.1 M sodium hydroxide.* Carry out a blank titration.
Calculate the percentage content of phthaloyl groups (*P*) using the following expression:

$$\frac{14910(n_1 - n_2)}{(100 - a)(100 - S)m} - \frac{179.5S}{(100 - S)}$$

a = percentage content of water (see Tests);
m = mass of the substance to be examined, in grams;
n_1 = volume of *0.1 M sodium hydroxide* used in the titration, in millilitres;
n_2 = volume of *0.1 M sodium hydroxide* used in the blank titration, in millilitres;
S = percentage content of free acid (see Tests).

Acetyl groups
To 0.100 g add 25.0 mL of *0.1 M sodium hydroxide* and heat on a water-bath under a reflux condenser for 30 min. Cool, add about 0.1 mL of *phenolphthalein solution R1* and titrate with *0.1 M hydrochloric acid.* Carry out a blank titration.
Calculate the percentage content of acetyl groups using the following expression:

$$\left[\frac{4305(n_2 - n_1)}{(100 - a)(100 - S)m} - \frac{51.82S}{(100 - S)}\right] - 0.5772P$$

a = percentage content of water (see Tests);
m = mass of the substance to be examined, in grams;
n_1 = volume of *0.1 M hydrochloric acid* used in the titration, in millilitres;
n_2 = volume of *0.1 M hydrochloric acid* used in the blank titration, in millilitres;
P = percentage content of phthaloyl groups;
S = percentage content of free acid (see Tests).

STORAGE
In an airtight container.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for cellulose acetate phthalate used as film former in gastro-resistant tablets and capsules.

Viscosity
See Tests.

Solubility of a film
Dissolve about 0.15 g in 1 mL of *acetone R* and pour onto a clear glass plate. A film is formed. Take a piece of the film and place it in a flask containing *0.1 M hydrochloric acid.* It does not dissolve. Then place the piece of film in a flask containing phosphate buffer solution pH 6.8 R. It dissolves.

Phthaloyl groups
See Assay.

Acetyl groups
See Assay.

Ph Eur

Dispersible Cellulose

Action and use
Pharmaceutical excipient.

DEFINITION
Dispersible Cellulose is a mixture of Microcrystalline Cellulose co-processed with Carmellose Sodium that readily forms a colloidal dispersion.

Content of carmellose sodium
75.0 to 125.0% w/w of the stated amount.

CHARACTERISTICS
A white or off-white, coarse or fine powder; hygroscopic.

Disperses in *water* producing a white, opaque dispersion or gel; practically insoluble in organic solvents and in dilute acids.

IDENTIFICATION
A. Mix 6 g with 300 mL of *water* stirring at 18,000 revolutions per minute for 5 minutes. A white, opaque dispersion is obtained which does not produce a supernatant liquid.

B. Add several drops of the dispersion obtained in test A to a 10% w/v solution of *aluminium chloride.* Each drop forms a white, opaque globule which does not disperse on standing.

C. Add 2 mL of *iodinated potassium iodide solution* to the dispersion obtained in test A. No blue or purplish colour is produced.

D. To the residue obtained in the test for Sulfated ash add 1 mL of *hydrochloric acid,* evaporate to dryness on a water bath and dissolve the residue in 20 mL of *water.* The resulting solution yields the reactions characteristic of *sodium salts,* Appendix VI, except that in test A the white precipitate produced may not be dense.

TESTS

Acidity or alkalinity
pH of the dispersion obtained in the test for Apparent viscosity, 6.0 to 8.0, Appendix V L.

Solubility
Add 50 mg to 10 mL of *ammoniacal solution of copper tetrammine* and shake. It dissolves completely leaving no residue.

Apparent viscosity
60 to 140% of the declared value when determined by the following method. Calculate the quantity (*x* g) needed to prepare exactly 600 g of a dispersion of the stated percentage w/w, with reference to the dried substance. To (600-*x*) g of *water* at 23° to 25° contained in a 1000 mL high-speed blender bowl add *x* g of the substance being examined, stirring at reduced speed, taking care to avoid contacting the sides of the bowl with the powder. Continue stirring at low speed for 15 seconds after the addition and then stir at 18,000 revolutions per minute for exactly 2 minutes. Immerse the appropriate spindle of a rotational viscometer, switch on after 30 seconds and after a further 30 seconds determine the *viscosity,* Appendix V H, Method III, using a speed of 20 revolutions per minute (2.09 radians per second).

Loss on drying
When dried to constant weight at 105°, loses not more than 8.0% of its weight. Use 1 g.

Sulfated ash
Not more than 5.0%, Appendix IX A. Use 2 g.

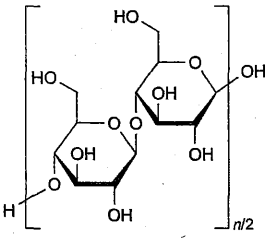
ASSAY
Heat 2 g with 75 mL of *anhydrous acetic acid* under a reflux condenser for 2 hours, cool and carry out Method I for *non-aqueous titration,* Appendix VIII A, determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 29.6 mg of carmellose sodium.

STORAGE
Dispersible Cellulose should be protected from moisture.

LABELLING
The label states (1) the percentage w/w of Carmellose Sodium; (2) the viscosity of a dispersion in water of a stated percentage w/w of Carmellose Sodium.

Microcrystalline Cellulose[1]

(Ph. Eur. monograph 0316)



$C_{6n}H_{10n+2}O_{5n+1}$ *9004-34-6*

Action and use
Excipient.

Ph Eur

DEFINITION
Purified, partly depolymerised cellulose prepared by treating alpha-cellulose, obtained as a pulp from fibrous plant material, with mineral acids.

♦CHARACTERS
Appearance
White or almost white, fine or granular, slightly hygroscopic powder.

Solubility
Practically insoluble in water, in acetone, in anhydrous ethanol, in toluene, in dilute acids and in a 50 g/L solution of sodium hydroxide.♦

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison microcrystalline cellulose CRS.

Disregard any band between 800 cm^{-1} and 825 cm^{-1} or between 950 cm^{-1} and 1000 cm^{-1}.

B. Place about 10 mg on a watch-glass and disperse in 2 mL of *iodinated zinc chloride solution R*. The substance becomes violet-blue.

C. The degree of polymerisation is not more than 350.

Transfer 1.300 g to a 125 mL conical flask. Add 25.0 mL of *water R* and 25.0 mL of *cupriethylenediamine hydroxide solution R*. Immediately purge the solution with *nitrogen R*, insert the stopper and shake until completely dissolved. Transfer an appropriate volume of the solution to a suitable capillary viscometer (*2.2.9*). Equilibrate the solution at 25 ± 0.1 °C for at least 5 min. Record the flow time (t_1) in seconds between the 2 marks on the viscometer. Calculate the kinematic viscosity (v_1) of the solution using the following expression:

$$t_1(k_1)$$

k_1 = viscometer constant.

Dilute a suitable volume of *cupriethylenediamine hydroxide solution R* with an equal volume of *water R* and measure the flow time (t_2) using a suitable capillary viscometer. Calculate the kinematic viscosity (v_2) of the solvent using the following expression:

$$t_2(k_2)$$

k_2 = viscometer constant.

Determine the relative viscosity (η_{rel}) of the substance to be examined using the following expression:

$$v_1/v_2$$

Determine the intrinsic viscosity ($[\eta]_c$) by interpolation, using the intrinsic viscosity table (Table 0316.-1).

Calculate the degree of polymerisation (P) using the following expression:

$$\frac{95[\eta]_c}{m[(100-b)/100]}$$

m = mass of the substance to be examined, in grams;

b = loss on drying, in per cent.

TESTS
◊Solubility
Dissolve 50 mg in 10 mL of *ammoniacal solution of copper tetrammine R*. It dissolves completely, leaving no residue.◊

pH (*2.2.3*)
5.0 to 7.5 for the supernatant.

Shake 5 g with 40 mL of *carbon dioxide-free water R* for 20 min and centrifuge.

Conductivity (*2.2.38*)
The conductivity of the test solution does not exceed the conductivity of the water by more than 75 $\mu S \cdot cm^{-1}$.

Use as test solution the supernatant obtained in the test for pH. Measure the conductivity of the supernatant after a stable reading has been obtained and measure the conductivity of the water used to prepare the test solution.

Ether-soluble substances
Maximum 0.05 per cent (5.0 mg) for the difference between the mass of the residue and the mass obtained from a blank determination.

Place 10.0 g in a chromatography column about 20 mm in internal diameter and pass 50 mL of *peroxide-free ether R* through the column. Evaporate the eluate to dryness in a previously dried and tared evaporating dish, with the aid of a current of air in a fume cupboard. After all ether has evaporated, dry the residue at 105 °C for 30 min, allow to cool in a desiccator and weigh. Carry out a blank determination.

Water-soluble substances
Maximum 0.25 per cent (12.5 mg) for the difference between the mass of the residue and the mass obtained from a blank determination.

Shake 5.0 g with 80 mL of *water R* for 10 min. Filter through a filter paper with the aid of vacuum into a tared flask. Evaporate to dryness on a water-bath avoiding charring. Dry at 105 °C for 1 h, allow to cool in a desiccator and weigh. Carry out a blank determination.

Loss on drying (*2.2.32*)
Maximum 7.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Table 0316.-1. – *Intrinsic viscosity table*

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}										
η_{rel}	0.00	0.01	0.02	0.03	$[\eta]_c$ 0.04	0.05	0.06	0.07	0.08	0.09
1.1	0.098	0.106	0.115	0.125	0.134	0.143	0.152	0.161	0.170	0.180
1.2	0.189	0.198	0.207	0.216	0.225	0.233	0.242	0.250	0.259	0.268
1.3	0.276	0.285	0.293	0.302	0.310	0.318	0.326	0.334	0.342	0.350
1.4	0.358	0.367	0.375	0.383	0.391	0.399	0.407	0.414	0.422	0.430
1.5	0.437	0.445	0.453	0.460	0.468	0.476	0.484	0.491	0.499	0.507
1.6	0.515	0.522	0.529	0.536	0.544	0.551	0.558	0.566	0.573	0.580
1.7	0.587	0.595	0.602	0.608	0.615	0.622	0.629	0.636	0.642	0.649
1.8	0.656	0.663	0.670	0.677	0.683	0.690	0.697	0.704	0.710	0.717
1.9	0.723	0.730	0.736	0.743	0.749	0.756	0.762	0.769	0.775	0.782
2.0	0.788	0.795	0.802	0.809	0.815	0.821	0.827	0.833	0.840	0.846
2.1	0.852	0.858	0.864	0.870	0.876	0.882	0.888	0.894	0.900	0.906
2.2	0.912	0.918	0.924	0.929	0.935	0.941	0.948	0.953	0.959	0.965
2.3	0.971	0.976	0.983	0.988	0.994	1.000	1.006	1.011	1.017	1.022
2.4	1.028	1.033	1.039	1.044	1.050	1.056	1.061	1.067	1.072	1.078
2.5	1.083	1.089	1.094	1.100	1.105	1.111	1.116	1.121	1.126	1.131
2.6	1.137	1.142	1.147	1.153	1.158	1.163	1.169	1.174	1.179	1.184
2.7	1.190	1.195	1.200	1.205	1.210	1.215	1.220	1.225	1.230	1.235
2.8	1.240	1.245	1.250	1.255	1.260	1.265	1.270	1.275	1.280	1.285
2.9	1.290	1.295	1.300	1.305	1.310	1.314	1.319	1.324	1.329	1.333
3.0	1.338	1.343	1.348	1.352	1.357	1.362	1.367	1.371	1.376	1.381
3.1	1.386	1.390	1.395	1.400	1.405	1.409	1.414	1.418	1.423	1.427
3.2	1.432	1.436	1.441	1.446	1.450	1.455	1.459	1.464	1.468	1.473
3.3	1.477	1.482	1.486	1.491	1.496	1.500	1.504	1.508	1.513	1.517
3.4	1.521	1.525	1.529	1.533	1.537	1.542	1.546	1.550	1.554	1.558
3.5	1.562	1.566	1.570	1.575	1.579	1.583	1.587	1.591	1.595	1.600
3.6	1.604	1.608	1.612	1.617	1.621	1.625	1.629	1.633	1.637	1.642
3.7	1.646	1.650	1.654	1.658	1.662	1.666	1.671	1.675	1.679	1.683
3.8	1.687	1.691	1.695	1.700	1.704	1.708	1.712	1.715	1.719	1.723
3.9	1.727	1.731	1.735	1.739	1.742	1.746	1.750	1.754	1.758	1.762
4.0	1.765	1.769	1.773	1.777	1.781	1.785	1.789	1.792	1.796	1.800
4.1	1.804	1.808	1.811	1.815	1.819	1.822	1.826	1.830	1.833	1.837
4.2	1.841	1.845	1.848	1.852	1.856	1.859	1.863	1.867	1.870	1.874
4.3	1.878	1.882	1.885	1.889	1.893	1.896	1.900	1.904	1.907	1.911
4.4	1.914	1.918	1.921	1.925	1.929	1.932	1.936	1.939	1.943	1.946
4.5	1.950	1.954	1.957	1.961	1.964	1.968	1.971	1.975	1.979	1.982
4.6	1.986	1.989	1.993	1.996	2.000	2.003	2.007	2.010	2.013	2.017
4.7	2.020	2.023	2.027	2.030	2.033	2.037	2.040	2.043	2.047	2.050
4.8	2.053	2.057	2.060	2.063	2.067	2.070	2.073	2.077	2.080	2.083
4.9	2.087	2.090	2.093	2.097	2.100	2.103	2.107	2.110	2.113	2.116
5.0	2.119	2.122	2.125	2.129	2.132	2.135	2.139	2.142	2.145	2.148
5.1	2.151	2.154	2.158	2.160	2.164	2.167	2.170	2.173	2.176	2.180
5.2	2.183	2.186	2.190	2.192	2.195	2.197	2.200	2.203	2.206	2.209
5.3	2.212	2.215	2.218	2.221	2.224	2.227	2.230	2.233	2.236	2.240
5.4	2.243	2.246	2.249	2.252	2.255	2.258	2.261	2.264	2.267	2.270
5.5	2.273	2.276	2.279	2.282	2.285	2.288	2.291	2.294	2.297	2.300
5.6	2.303	2.306	2.309	2.312	2.315	2.318	2.320	2.324	2.326	2.329
5.7	2.332	2.335	2.338	2.341	2.344	2.347	2.350	2.353	2.355	2.358
5.8	2.361	2.364	2.367	2.370	2.373	2.376	2.379	2.382	2.384	2.387
5.9	2.390	2.393	2.396	2.400	2.403	2.405	2.408	2.411	2.414	2.417
6.0	2.419	2.422	2.425	2.428	2.431	2.433	2.436	2.439	2.442	2.444
6.1	2.447	2.450	2.453	2.456	2.458	2.461	2.464	2.467	2.470	2.472
6.2	2.475	2.478	2.481	2.483	2.486	2.489	2.492	2.494	2.497	2.500
6.3	2.503	2.505	2.508	2.511	2.513	2.516	2.518	2.521	2.524	2.526
6.4	2.529	2.532	2.534	2.537	2.540	2.542	2.545	2.547	2.550	2.553
6.5	2.555	2.558	2.561	2.563	2.566	2.568	2.571	2.574	2.576	2.579
6.6	2.581	2.584	2.587	2.590	2.592	2.595	2.597	2.600	2.603	2.605

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}

η_{rel}	0.00	0.01	0.02	0.03	$[\eta]_c$ 0.04	0.05	0.06	0.07	0.08	0.09
6.7	2.608	2.610	2.613	2.615	2.618	2.620	2.623	2.625	2.627	2.630
6.8	2.633	2.635	2.637	2.640	2.643	2.645	2.648	2.650	2.653	2.655
6.9	2.658	2.660	2.663	2.665	2.668	2.670	2.673	2.675	2.678	2.680
7.0	2.683	2.685	2.687	2.690	2.693	2.695	2.698	2.700	2.702	2.705
7.1	2.707	2.710	2.712	2.714	2.717	2.719	2.721	2.724	2.726	2.729
7.2	2.731	2.733	2.736	2.738	2.740	2.743	2.745	2.748	2.750	2.752
7.3	2.755	2.757	2.760	2.762	2.764	2.767	2.769	2.771	2.774	2.776
7.4	2.779	2.781	2.783	2.786	2.788	2.790	2.793	2.795	2.798	2.800
7.5	2.802	2.805	2.807	2.809	2.812	2.814	2.816	2.819	2.821	2.823
7.6	2.826	2.828	2.830	2.833	2.835	2.837	2.840	2.842	2.844	2.847
7.7	2.849	2.851	2.854	2.856	2.858	2.860	2.863	2.865	2.868	2.870
7.8	2.873	2.875	2.877	2.879	2.881	2.884	2.887	2.889	2.891	2.893
7.9	2.895	2.898	2.900	2.902	2.905	2.907	2.909	2.911	2.913	2.915
8.0	2.918	2.920	2.922	2.924	2.926	2.928	2.931	2.933	2.935	2.937
8.1	2.939	2.942	2.944	2.946	2.948	2.950	2.952	2.955	2.957	2.959
8.2	2.961	2.963	2.966	2.968	2.970	2.972	2.974	2.976	2.979	2.981
8.3	2.983	2.985	2.987	2.990	2.992	2.994	2.996	2.998	3.000	3.002
8.4	3.004	3.006	3.008	3.010	3.012	3.015	3.017	3.019	3.021	3.023
8.5	3.025	3.027	3.029	3.031	3.033	3.035	3.037	3.040	3.042	3.044
8.6	3.046	3.048	3.050	3.052	3.054	3.056	3.058	3.060	3.062	3.064
8.7	3.067	3.069	3.071	3.073	3.075	3.077	3.079	3.081	3.083	3.085
8.8	3.087	3.089	3.092	3.094	3.096	3.098	3.100	3.102	3.104	3.106
8.9	3.108	3.110	3.112	3.114	3.116	3.118	3.120	3.122	3.124	3.126
9.0	3.128	3.130	3.132	3.134	3.136	3.138	3.140	3.142	3.144	3.146
9.1	3.148	3.150	3.152	3.154	3.156	3.158	3.160	3.162	3.164	3.166
9.2	3.168	3.170	3.172	3.174	3.176	3.178	3.180	3.182	3.184	3.186
9.3	3.188	3.190	3.192	3.194	3.196	3.198	3.200	3.202	3.204	3.206
9.4	3.208	3.210	3.212	3.214	3.215	3.217	3.219	3.221	3.223	3.225
9.5	3.227	3.229	3.231	3.233	3.235	3.237	3.239	3.241	3.242	3.244
9.6	3.246	3.248	3.250	3.252	3.254	3.256	3.258	3.260	3.262	3.264
9.7	3.266	3.268	3.269	3.271	3.273	3.275	3.277	3.279	3.281	3.283
9.8	3.285	3.287	3.289	3.291	3.293	3.295	3.297	3.298	3.300	3.302
9.9	3.304	3.305	3.307	3.309	3.311	3.313	3.316	3.318	3.320	3.321

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}

η_{rel}	0.0	0.1	0.2	0.3	$[\eta]_c$ 0.4	0.5	0.6	0.7	0.8	0.9
10	3.32	3.34	3.36	3.37	3.39	3.41	3.43	3.45	3.46	3.48
11	3.50	3.52	3.53	3.55	3.56	3.58	3.60	3.61	3.63	3.64
12	3.66	3.68	3.69	3.71	3.72	3.74	3.76	3.77	3.79	3.80
13	3.80	3.83	3.85	3.86	3.88	3.89	3.90	3.92	3.93	3.95
14	3.96	3.97	3.99	4.00	4.02	4.03	4.04	4.06	4.07	4.09
15	4.10	4.11	4.13	4.14	4.15	4.17	4.18	4.19	4.20	4.22
16	4.23	4.24	4.25	4.27	4.28	4.29	4.30	4.31	4.33	4.34
17	4.35	4.36	4.37	4.38	4.39	4.41	4.42	4.43	4.44	4.45
18	4.46	4.47	4.48	4.49	4.50	4.52	4.53	4.54	4.55	4.56
19	4.57	4.58	4.59	4.60	4.61	4.62	4.63	4.64	4.65	4.66

♦Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Pseudomonas aeruginosa* (*2.6.13*).

Absence of *Staphylococcus aureus* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).♦

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for microcrystalline cellulose used as binder, diluent or disintegrant.

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Microcrystalline Cellulose and Carmellose Sodium

(Ph. Eur. monograph 2050)

Action and use
Excipient.

Ph Eur

DEFINITION
Colloid-forming, powdered mixture of *Microcrystalline cellulose (0316)* with 5 per cent to 22 per cent of *Carmellose sodium (0472)*.

Content
75.0 per cent to 125.0 per cent of the nominal content of carmellose sodium (dried substance).

CHARACTERS
Appearance
White or off-white, coarse or fine, hygroscopic powder.

Solubility
Dispersible in water producing a white, opaque colloidal dispersion; practically insoluble in organic solvents and in dilute acids.

IDENTIFICATION
A. Mix 6 g with 300 mL of *water R* and stir at 18 000 r/min for 5 min. A white opaque dispersion is obtained which does not produce a supernatant.

B. Add several drops of the dispersion obtained in identification A to a 100 g/L solution of *aluminium chloride R*. Each drop forms a white, opaque globule which does not disperse on standing.

C. Add 2 mL of *iodinated potassium iodide solution R* to the dispersion obtained in test A. No blue or purplish colour is produced.

D. It complies with the limits of the assay.

TESTS
Solubility
Add 50 mg to 10 mL of *ammoniacal solution of copper tetrammine R* and shake. It dissolves completely, leaving no residue.

pH (*2.2.3*)
6.0 to 8.0 for the dispersion obtained in identification A.

Loss on drying (*2.2.32*)
Maximum 8.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 7.4 per cent, determined on 2.0 g.

ASSAY
Heat 2.00 g with 75 mL of *anhydrous acetic acid R* under a reflux condenser for 2 h, cool and titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 29.6 mg of carmellose sodium.

LABELLING
The label states the nominal content of carmellose sodium in per cent *m/m*.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for microcrystalline cellulose and carmellose sodium used as a suspending agent.

Viscosity (*2.2.10*)
60 per cent to 140 per cent of the nominal value.

Calculate the quantity (*x* g) needed to prepare exactly 600 g of a dispersion of the stated percentage *m/m* (dried substance). To (600 - *x*) g of *water R* at 23-25 °C contained in a 1000 mL high-speed blender bowl, add *x* g of the substance to be examined and stir at reduced speed, taking care to avoid contacting the sides of the bowl with the powder. Continue stirring at low speed for 15 s after the addition of the powder and then stir at 18 000 r/min for exactly 2 min.

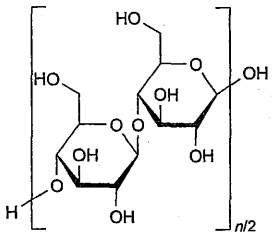
Determine the viscosity with a suitable relative rotational viscometer under the following conditions:
— spindle: as appropriate;
— speed: 20 r/min.

Immerse the spindle into the suspension immediately after preparation, switch on the rotation spindle after 30 s; after a further 30 s, take scale readings and calculate the viscosity according to the viscometer manual.

Ph Eur

Powdered Cellulose[1]

(Ph. Eur. monograph 0315)



$C_{6n}H_{10n+2}O_{5n+1}$

Action and use
Excipient.

Ph Eur

DEFINITION
Purified, mechanically disintegrated cellulose prepared by processing alpha-cellulose obtained as a pulp from fibrous plant material.

♦CHARACTERS
Appearance
White or almost white, fine or granular powder.

Solubility
Practically insoluble in water, slightly soluble in a 50 g/L solution of sodium hydroxide, practically insoluble in acetone, in anhydrous ethanol, in toluene, in dilute acids and in most organic solvents.♦

IDENTIFICATION
A. Place about 10 mg on a watch-glass and disperse in 2 mL of *iodinated zinc chloride solution R*. The substance becomes violet-blue.

B. The degree of polymerisation is greater than 440.

Transfer 0.250 g to a 125 mL conical flask. Add 25.0 mL of *water R* and 25.0 mL of *cupriethylenediamine hydroxide solution R*. Immediately purge the solution with *nitrogen R*, insert the stopper and shake until completely dissolved. Transfer an appropriate volume of the solution to a suitable capillary viscometer (*2.2.9*). Equilibrate the solution at 25 ± 0.1 °C for at least 5 min. Record the flow time (t_1) in seconds between the 2 marks on the viscometer. Calculate the kinematic viscosity (v_1) of the solution using the following expression:

$$t_1(k_1)$$

k_1 = viscometer constant.

Dilute a suitable volume of *cupriethylenediamine hydroxide solution R* with an equal volume of *water R* and measure the flow time (t_2) using a suitable capillary viscometer. Calculate the kinematic viscosity (v_2) of the solvent using the following expression:

$$t_2(k_2)$$

k_2 = viscometer constant.

Determine the relative viscosity (η_{rel}) of the substance to be examined using the following expression:

$$v_1/v_2$$

Determine the intrinsic viscosity ($[\eta]_c$) by interpolation, using the intrinsic viscosity table (Table 0315.-1).

Calculate the degree of polymerisation (P) using the following expression:

$$\frac{95[\eta]_c}{m[(100-b)/100]}$$

m = mass of the substance to be examined, in grams;
b = loss on drying, in per cent.

TESTS
◊Solubility
Dissolve 50 mg in 10 mL of *ammoniacal solution of copper tetrammine R*. It dissolves completely, leaving no residue.◊

pH (*2.2.3*)
5.0 to 7.5 for the supernatant.

Mix 10 g with 90 mL of *carbon dioxide-free water R* and allow to stand with occasional stirring for 1 h.

Ether-soluble substances
Maximum 0.15 per cent (15.0 mg) for the difference between the mass of the residue and the mass obtained from a blank determination.

Place 10.0 g in a chromatography column about 20 mm in internal diameter and pass 50 mL of *peroxide-free ether R* through the column. Evaporate the eluate to dryness in a previously dried and tared evaporating dish, with the aid of a current of air in a fume cupboard. After all the ether has evaporated, dry the residue at 105 °C for 30 min, allow to cool in a desiccator and weigh. Carry out a blank determination.

Water-soluble substances
Maximum 1.5 per cent (15.0 mg) for the difference between the mass of the residue and the mass obtained from a blank determination.

Shake 6.0 g with 90 mL of *carbon dioxide-free water R* for 10 min. Filter with the aid of vacuum into a tared flask. Discard the first 10 mL of the filtrate and pass the filtrate through the same filter a second time, if necessary, to obtain a clear filtrate. Evaporate a 15.0 mL portion of the filtrate to dryness in a tared evaporating dish without charring. Dry at 105 °C for 1 h, allow to cool in a desiccator and weigh. Carry out a blank determination.

Loss on drying (*2.2.32*)
Maximum 6.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.3 per cent (dried substance), determined on 1.0 g.

♦Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Pseudomonas aeruginosa* (*2.6.13*).

Absence of *Staphylococcus aureus* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).♦

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Table 0315.-1. – *Intrinsic viscosity table*

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}										
η_{rel}	0.00	0.01	0.02	0.03	$[\eta]_c$ 0.04	0.05	0.06	0.07	0.08	0.09
1.1	0.098	0.106	0.115	0.125	0.134	0.143	0.152	0.161	0.170	0.180
1.2	0.189	0.198	0.207	0.216	0.225	0.233	0.242	0.250	0.259	0.268
1.3	0.276	0.285	0.293	0.302	0.310	0.318	0.326	0.334	0.342	0.350
1.4	0.358	0.367	0.375	0.383	0.391	0.399	0.407	0.414	0.422	0.430
1.5	0.437	0.445	0.453	0.460	0.468	0.476	0.484	0.491	0.499	0.507
1.6	0.515	0.522	0.529	0.536	0.544	0.551	0.558	0.566	0.573	0.580
1.7	0.587	0.595	0.602	0.608	0.615	0.622	0.629	0.636	0.642	0.649
1.8	0.656	0.663	0.670	0.677	0.683	0.690	0.697	0.704	0.710	0.717
1.9	0.723	0.730	0.736	0.743	0.749	0.756	0.762	0.769	0.775	0.782
2.0	0.788	0.795	0.802	0.809	0.815	0.821	0.827	0.833	0.840	0.846
2.1	0.852	0.858	0.864	0.870	0.876	0.882	0.888	0.894	0.900	0.906
2.2	0.912	0.918	0.924	0.929	0.935	0.941	0.948	0.953	0.959	0.965
2.3	0.971	0.976	0.983	0.988	0.994	1.000	1.006	1.011	1.017	1.022
2.4	1.028	1.033	1.039	1.044	1.050	1.056	1.061	1.067	1.072	1.078
2.5	1.083	1.089	1.094	1.100	1.105	1.111	1.116	1.121	1.126	1.131
2.6	1.137	1.142	1.147	1.153	1.158	1.163	1.169	1.174	1.179	1.184
2.7	1.190	1.195	1.200	1.205	1.210	1.215	1.220	1.225	1.230	1.235
2.8	1.240	1.245	1.250	1.255	1.260	1.265	1.270	1.275	1.280	1.285
2.9	1.290	1.295	1.300	1.305	1.310	1.314	1.319	1.324	1.329	1.333
3.0	1.338	1.343	1.348	1.352	1.357	1.362	1.367	1.371	1.376	1.381
3.1	1.386	1.390	1.395	1.400	1.405	1.409	1.414	1.418	1.423	1.427
3.2	1.432	1.436	1.441	1.446	1.450	1.455	1.459	1.464	1.468	1.473
3.3	1.477	1.482	1.486	1.491	1.496	1.500	1.504	1.508	1.513	1.517
3.4	1.521	1.525	1.529	1.533	1.537	1.542	1.546	1.550	1.554	1.558
3.5	1.562	1.566	1.570	1.575	1.579	1.583	1.587	1.591	1.595	1.600
3.6	1.604	1.608	1.612	1.617	1.621	1.625	1.629	1.633	1.637	1.642
3.7	1.646	1.650	1.654	1.658	1.662	1.666	1.671	1.675	1.679	1.683
3.8	1.687	1.691	1.695	1.700	1.704	1.708	1.712	1.715	1.719	1.723
3.9	1.727	1.731	1.735	1.739	1.742	1.746	1.750	1.754	1.758	1.762
4.0	1.765	1.769	1.773	1.777	1.781	1.785	1.789	1.792	1.796	1.800
4.1	1.804	1.808	1.811	1.815	1.819	1.822	1.826	1.830	1.833	1.837
4.2	1.841	1.845	1.848	1.852	1.856	1.859	1.863	1.867	1.870	1.874
4.3	1.878	1.882	1.885	1.889	1.893	1.896	1.900	1.904	1.907	1.911
4.4	1.914	1.918	1.921	1.925	1.929	1.932	1.936	1.939	1.943	1.946
4.5	1.950	1.954	1.957	1.961	1.964	1.968	1.971	1.975	1.979	1.982
4.6	1.986	1.989	1.993	1.996	2.000	2.003	2.007	2.010	2.013	2.017
4.7	2.020	2.023	2.027	2.030	2.033	2.037	2.040	2.043	2.047	2.050
4.8	2.053	2.057	2.060	2.063	2.067	2.070	2.073	2.077	2.080	2.083
4.9	2.087	2.090	2.093	2.097	2.100	2.103	2.107	2.110	2.113	2.116
5.0	2.119	2.122	2.125	2.129	2.132	2.135	2.139	2.142	2.145	2.148
5.1	2.151	2.154	2.158	2.160	2.164	2.167	2.170	2.173	2.176	2.180
5.2	2.183	2.186	2.190	2.192	2.195	2.197	2.200	2.203	2.206	2.209
5.3	2.212	2.215	2.218	2.221	2.224	2.227	2.230	2.233	2.236	2.240
5.4	2.243	2.246	2.249	2.252	2.255	2.258	2.261	2.264	2.267	2.270
5.5	2.273	2.276	2.279	2.282	2.285	2.288	2.291	2.294	2.297	2.300
5.6	2.303	2.306	2.309	2.312	2.315	2.318	2.320	2.324	2.326	2.329
5.7	2.332	2.335	2.338	2.341	2.344	2.347	2.350	2.353	2.355	2.358
5.8	2.361	2.364	2.367	2.370	2.373	2.376	2.379	2.382	2.384	2.387
5.9	2.390	2.393	2.396	2.400	2.403	2.405	2.408	2.411	2.414	2.417
6.0	2.419	2.422	2.425	2.428	2.431	2.433	2.436	2.439	2.442	2.444
6.1	2.447	2.450	2.453	2.456	2.458	2.461	2.464	2.467	2.470	2.472
6.2	2.475	2.478	2.481	2.483	2.486	2.489	2.492	2.494	2.497	2.500
6.3	2.503	2.505	2.508	2.511	2.513	2.516	2.518	2.521	2.524	2.526
6.4	2.529	2.532	2.534	2.537	2.540	2.542	2.545	2.547	2.550	2.553
6.5	2.555	2.558	2.561	2.563	2.566	2.568	2.571	2.574	2.576	2.579
6.6	2.581	2.584	2.587	2.590	2.592	2.595	2.597	2.600	2.603	2.605

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}

η_{rel}	0.00	0.01	0.02	0.03	$[\eta]_c$ 0.04	0.05	0.06	0.07	0.08	0.09
6.7	2.608	2.610	2.613	2.615	2.618	2.620	2.623	2.625	2.627	2.630
6.8	2.633	2.635	2.637	2.640	2.643	2.645	2.648	2.650	2.653	2.655
6.9	2.658	2.660	2.663	2.665	2.668	2.670	2.673	2.675	2.678	2.680
7.0	2.683	2.685	2.687	2.690	2.693	2.695	2.698	2.700	2.702	2.705
7.1	2.707	2.710	2.712	2.714	2.717	2.719	2.721	2.724	2.726	2.729
7.2	2.731	2.733	2.736	2.738	2.740	2.743	2.745	2.748	2.750	2.752
7.3	2.755	2.757	2.760	2.762	2.764	2.767	2.769	2.771	2.774	2.776
7.4	2.779	2.781	2.783	2.786	2.788	2.790	2.793	2.795	2.798	2.800
7.5	2.802	2.805	2.807	2.809	2.812	2.814	2.816	2.819	2.821	2.823
7.6	2.826	2.828	2.830	2.833	2.835	2.837	2.840	2.842	2.844	2.847
7.7	2.849	2.851	2.854	2.856	2.858	2.860	2.863	2.865	2.868	2.870
7.8	2.873	2.875	2.877	2.879	2.881	2.884	2.887	2.889	2.891	2.893
7.9	2.895	2.898	2.900	2.902	2.905	2.907	2.909	2.911	2.913	2.915
8.0	2.918	2.920	2.922	2.924	2.926	2.928	2.931	2.933	2.935	2.937
8.1	2.939	2.942	2.944	2.946	2.948	2.950	2.952	2.955	2.957	2.959
8.2	2.961	2.963	2.966	2.968	2.970	2.972	2.974	2.976	2.979	2.981
8.3	2.983	2.985	2.987	2.990	2.992	2.994	2.996	2.998	3.000	3.002
8.4	3.004	3.006	3.008	3.010	3.012	3.015	3.017	3.019	3.021	3.023
8.5	3.025	3.027	3.029	3.031	3.033	3.035	3.037	3.040	3.042	3.044
8.6	3.046	3.048	3.050	3.052	3.054	3.056	3.058	3.060	3.062	3.064
8.7	3.067	3.069	3.071	3.073	3.075	3.077	3.079	3.081	3.083	3.085
8.8	3.087	3.089	3.092	3.094	3.096	3.098	3.100	3.102	3.104	3.106
8.9	3.108	3.110	3.112	3.114	3.116	3.118	3.120	3.122	3.124	3.126
9.0	3.128	3.130	3.132	3.134	3.136	3.138	3.140	3.142	3.144	3.146
9.1	3.148	3.150	3.152	3.154	3.156	3.158	3.160	3.162	3.164	3.166
9.2	3.168	3.170	3.172	3.174	3.176	3.178	3.180	3.182	3.184	3.186
9.3	3.188	3.190	3.192	3.194	3.196	3.198	3.200	3.202	3.204	3.206
9.4	3.208	3.210	3.212	3.214	3.215	3.217	3.219	3.221	3.223	3.225
9.5	3.227	3.229	3.231	3.233	3.235	3.237	3.239	3.241	3.242	3.244
9.6	3.246	3.248	3.250	3.252	3.254	3.256	3.258	3.260	3.262	3.264
9.7	3.266	3.268	3.269	3.271	3.273	3.275	3.277	3.279	3.281	3.283
9.8	3.285	3.287	3.289	3.291	3.293	3.295	3.297	3.298	3.300	3.302
9.9	3.304	3.305	3.307	3.309	3.311	3.313	3.316	3.318	3.320	3.321

Intrinsic viscosity $[\eta]_c$ at different values of relative viscosity η_{rel}

η_{rel}	0.0	0.1	0.2	0.3	$[\eta]_c$ 0.4	0.5	0.6	0.7	0.8	0.9
10	3.32	3.34	3.36	3.37	3.39	3.41	3.43	3.45	3.46	3.48
11	3.50	3.52	3.53	3.55	3.56	3.58	3.60	3.61	3.63	3.64
12	3.66	3.68	3.69	3.71	3.72	3.74	3.76	3.77	3.79	3.80
13	3.80	3.83	3.85	3.86	3.88	3.89	3.90	3.92	3.93	3.95
14	3.96	3.97	3.99	4.00	4.02	4.03	4.04	4.06	4.07	4.09
15	4.10	4.11	4.13	4.14	4.15	4.17	4.18	4.19	4.20	4.22
16	4.23	4.24	4.25	4.27	4.28	4.29	4.30	4.31	4.33	4.34
17	4.35	4.36	4.37	4.38	4.39	4.41	4.42	4.43	4.44	4.45
18	4.46	4.47	4.48	4.49	4.50	4.52	4.53	4.54	4.55	4.56
19	4.57	4.58	4.59	4.60	4.61	4.62	4.63	4.64	4.65	4.66

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for powdered cellulose used as diluent or disintegrant.

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Cellulose Acetate[1]

(Ph. Eur. monograph 0887)

Action and use
Excipient.

Ph Eur

DEFINITION
Partly or completely *O*-acetylated cellulose.

Content
— *acetyl groups* (C_2H_3O; M_r 43.04): 29.0 per cent to 44.8 per cent (dried substance); 90.0 per cent to 110.0 per cent of the nominal content (dried substance).

CHARACTERS
Appearance
White, yellowish-white or greyish-white, hygroscopic powder or granules.

Solubility
Practically insoluble in water, soluble in acetone, in formic acid and in a mixture of equal volumes of methanol and methylene chloride, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cellulose acetate CRS*.

Preparation Prepare a 20 g/L solution of cellulose acetate, previously dried, in *acetone R* (mono- or diester) or in *methylene chloride R* (di- or triester), and spread 1 drop of the solution between 2 sodium chloride plates; separate the plates, heat them both at 105 °C for 1 h, and reassemble the dried plates.

TESTS
Free acid
Maximum 0.1 per cent, calculated as acetic acid (dried substance).

To 5.00 g in a 250 mL conical flask, add 150 mL of *carbon dioxide-free water R*, insert the stopper, swirl the suspension gently and allow to stand for 3 h. Filter, then wash the flask and the filter with *carbon dioxide-free water R*, adding the washings to the filtrate. Add 0.1 mL of *phenolphthalein solution R1* and titrate the combined filtrate and washings with *0.01 M sodium hydroxide* until a pale pink colour is obtained.

1 mL of *0.01 M sodium hydroxide* is equivalent to 0.6005 mg of free acid, calculated as acetic acid.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

ASSAY
A. Cellulose acetate containing not more than 42.0 per cent of acetyl groups.

To 2.000 g in a 500 mL conical flask, add 100 mL of *acetone R* and 5-10 mL of *water R*. Close the flask and stir with a magnetic stirrer until dissolution is complete. Add 30.0 mL of *1 M sodium hydroxide* with constant stirring. A finely divided precipitate of regenerated cellulose, free from lumps, is obtained. Close the flask and stir with a magnetic stirrer for 30 min. Add 100 mL of *water R* at 80 °C, washing down the sides of the flask, stir for 2 min and cool to room temperature. Titrate with *0.5 M sulfuric acid*, using 0.1 mL of *phenolphthalein solution R1* as indicator. Carry out a blank titration.

Calculate the percentage content of acetyl groups using the following expression:

$$\frac{4.305(n_2 - n_1)}{(100 - d) \times m} \times 100$$

d = loss on drying as a percentage;
m = mass of the substance to be examined, in grams;
n_1 = volume of *0.5 M sulfuric acid* used in the titration, in millilitres;
n_2 = volume of *0.5 M sulfuric acid* used in the blank titration, in millilitres.

B. Cellulose acetate containing more than 42.0 per cent of acetyl groups.

To 2.000 g in a 500 mL conical flask, add 30 mL of *dimethyl sulfoxide R* and 100 mL of *acetone R*. Close the flask and stir with a magnetic stirrer for 16 h. Add 30.0 mL of *1 M sodium hydroxide* with constant stirring. Close the flask and stir with a magnetic stirrer for 6 min. Allow to stand without stirring for 60 min. Resume stirring and add 100 mL of *water R* at 80 °C, washing down the sides of the flask, stir for 2 min and cool to room temperature. Titrate with *0.5 M hydrochloric acid*, using 0.1 mL of *phenolphthalein solution R1* as indicator. Add 0.5 mL of *0.5 M hydrochloric acid* in excess, stir for 5 min and allow to stand for 30 min. Titrate with *0.5 M sodium hydroxide* until a persistent pink colour is obtained, stirring with a magnetic stirrer. Calculate the net amount of *0.5 M sodium hydroxide* consumed, in millimoles, taking the mean of 2 blank titrations into consideration.

Calculate the percentage content of acetyl groups using the following expression:

$$\frac{4.305 \times n}{(100 - d) \times m} \times 100$$

d = loss on drying as a percentage;
m = mass of the substance to be examined, in grams;
n = net amount of *0.5 M sodium hydroxide* consumed, in millimoles.

STORAGE
In an airtight container.

LABELLING
The label states the nominal percentage content of acetyl groups.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for cellulose acetate used as film former.

Viscosity
Dissolve 10 g in a mixture of 50 mL of *methanol R* and 50 mL of *methylene chloride R* by shaking. Determine the viscosity of this solution at 20 ± 0.1 °C using a rotating viscometer (*2.2.10*).

Acetyl groups
See Assay.

The following characteristics may be relevant for cellulose acetate used as matrix former in prolonged-release tablets.

Viscosity
See test above.

Acetyl groups
See Assay.

Molecular mass distribution (*2.2.30*)

Particle-size distribution (*2.9.31*)

Powder flow (*2.9.36*)

Ph Eur

Cellulose Acetate Butyrate

(Ph. Eur. monograph 1406)

Action and use
Excipient.

Ph Eur

DEFINITION
Partly or completely *O*-acetylated and *O*-butyrated cellulose.

Content
— *acetyl groups* (C_2H_3O): 2.0 per cent to 30.0 per cent (dried substance); 90.0 per cent to 110.0 per cent of that stated on the label (dried substance);
— *butyryl groups* (C_4H_7O): 16.0 per cent to 53.0 per cent (dried substance); 90.0 per cent to 110.0 per cent of that stated on the label (dried substance).

CHARACTERS
Appearance
White, yellowish-white or greyish-white powder or granules, slightly hygroscopic.

Solubility
Practically insoluble in water, soluble in acetone, in formic acid and in a mixture of equal volumes of methanol and methylene chloride, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cellulose acetate butyrate CRS.*

The intensity of the bands may vary according to the degree of substitution.

B. It complies with the limits of the assay.

TESTS
Acidity
To 5.00 g in a 250 mL conical flask, add 150 mL of *carbon dioxide-free water R*, insert the stopper, swirl the suspension gently and allow to stand for 3 h. Filter, wash the flask and the filter with *carbon dioxide-free water R*. Combine the filtrate and washings. Add 0.1 mL of *phenolphthalein solution R1*. Not more than 3.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution To 1.000 g of the substance to be examined in a 500 mL conical flask, add 100 mL of *acetone R* and 10 mL of *water R*. Close the flask and stir with a magnetic stirrer until dissolution is complete. Add 30.0 mL of *1 M sodium hydroxide* with constant stirring. Close the flask and stir with a magnetic stirrer for 30 min. Add 100 mL of hot *water R* at 80 °C, washing down the sides of the flask and stir for 2 min. Cool, centrifuge or filter the suspension and wash the residue with *water R*. Combine the filtrate and washings, adjust to pH 3 with *dilute phosphoric acid R* and dilute to 500.0 mL with *water R*.

Reference solution Dissolve 0.200 g of *glacial acetic acid R* and 0.400 g of *butyric acid R* in *water R*, adjust to pH 3 with *dilute phosphoric acid R* and dilute to 500.0 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *methanol R*;
— *mobile phase B*: *phosphate buffer solution pH 3.0 R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	5	95
30 - 35	5 → 20	95 → 80
35 - 60	20	80
60 - 61	5	95

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Calculate the percentage content of acetic acid and butyric acid using the chromatograms obtained with the 2 solutions. To calculate the percentage content of acetyl (C_2H_3O) and of butyryl (C_4H_7O) groups, multiply the percentage content of acetic acid and butyric acid by 0.717 and 0.807, respectively.

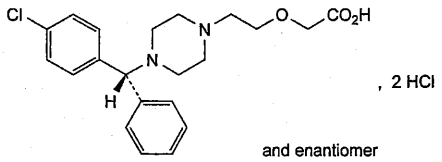
STORAGE
In an airtight container.

LABELLING
The label states the nominal percentage content of acetyl and butyryl groups.

Ph Eur

Cetirizine Hydrochloride

(Cetirizine Dihydrochloride, Ph. Eur. monograph 1084)



$C_{21}H_{27}Cl_3N_2O_3$ 461.8 *83881-52-1*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparations
Cetirizine Capsules
Cetirizine Oral Solution
Cetirizine Tablets

Ph Eur

DEFINITION
(*RS*)-2-[2-[4-[(4-Chlorophenyl)phenylmethyl]piperazin-1-yl]ethoxy]acetic acid dihydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Freely soluble in water, practically insoluble in acetone and in methylene chloride.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 20.0 mg in 50 mL of a 10.3 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of this solution to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*.

Spectral range 210-350 nm.

Absorption maximum At 231 nm.

Specific absorbance at the absorption maximum 359 to 381.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cetirizine dihydrochloride CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *cetirizine dihydrochloride CRS* in *water R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *chlorphenamine maleate CRS* in *water R* and dilute to 5 mL with the same solvent. Mix 1 mL of the solution and 1 mL of reference solution (a).

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *ammonia R, methanol R, methylene chloride R* (1:10:90 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In a current of cold air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

pH (*2.2.3*)
1.2 to 1.8 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *cetirizine dihydrochloride CRS* and 2 mg of *cetirizine impurity A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *cetirizine for peak identification CRS* (containing impurities B, C, D, E and F) in 5.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *dilute sulfuric acid R, water R, acetonitrile R* (0.4:6.6:93 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 3 times the retention time of cetirizine.

Identification of impurities Use the chromatogram supplied with *cetirizine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C, D, E and F; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to cetirizine (retention time = about 9 min): impurity D = about 0.6; impurity B = about 0.8; impurity C = about 0.9; impurity E = about 1.2; impurity F = about 1.37; impurity A = about 1.42.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cetirizine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7; impurity C = 1.9; impurity D = 0.6; impurity E = 1.3; impurity F = 1.9;
— *impurities A, B, C, D, E, F*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

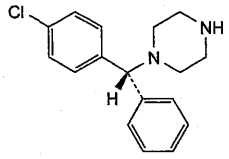
ASSAY
Dissolve 0.100 g in 70 mL of a mixture of 30 volumes of *water R* and 70 volumes of *acetone R*. Titrate with *0.1 M sodium hydroxide* to the 2nd point of inflexion. Determine the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 15.39 mg of $C_{21}H_{27}Cl_3N_2O_3$.

STORAGE
Protected from light.

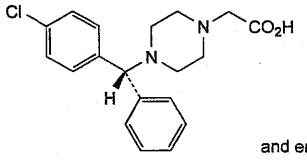
IMPURITIES
Specified impurities *A, B, C, D, E, F.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) G.



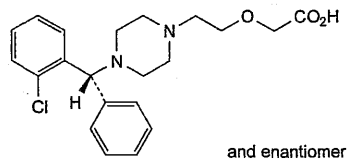
and enantiomer

A. (*RS*)-1-[(4-chlorophenyl)phenylmethyl]piperazine,



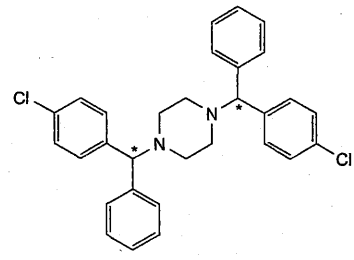
and enantiomer

B. (*RS*)-2-[4-[(4-chlorophenyl)phenylmethyl]piperazin-1-yl]acetic acid,

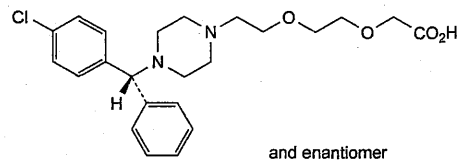


and enantiomer

C. (*RS*)-2-[2-[4-[(2-chlorophenyl)phenylmethyl]piperazin-1-yl]ethoxy]acetic acid,

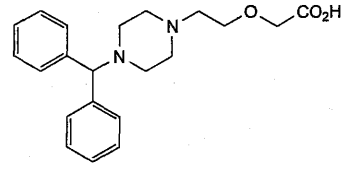


D. 1,4-bis[(4-chlorophenyl)phenylmethyl]piperazine,

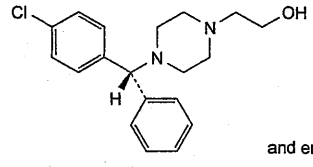


and enantiomer

E. (*RS*)-2-[2-[2-[4-[(4-chlorophenyl)phenylmethyl]piperazin-1-yl]ethoxy]ethoxy]acetic acid (ethoxycetirizine),



F. 2-[2-[4-(diphenylmethyl)piperazin-1-yl]ethoxy]acetic acid,



and enantiomer

G. (*RS*)-2-[4-[(4-chlorophenyl)phenylmethyl]piperazin-1-yl]ethan-1-ol.

Ph Eur

Cetostearyl Alcohol

(Ph. Eur. monograph 0702)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of solid aliphatic alcohols, mainly octadecan-1-ol (stearyl alcohol; $C_{18}H_{38}O$; M_r 270.5) and hexadecan-1-ol (cetyl alcohol; $C_{16}H_{34}O$; M_r 242.4), of animal or vegetable origin.

Content
— *stearyl alcohol*: minimum 40.0 per cent,

— *sum of the contents of stearyl alcohol and cetyl alcohol*: minimum 90.0 per cent.

CHARACTERS

Appearance

White or pale yellow, wax-like mass, plates, flakes or granules.

Solubility

Practically insoluble in water, soluble in ethanol (96 per cent) and in light petroleum. When melted, it is miscible with fatty oils, with liquid paraffin and with melted wool fat.

IDENTIFICATION

Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the principal peaks in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_6 (*2.2.2, Method II*).

Dissolve 0.50 g in 20 mL of boiling *ethanol (96 per cent) R*. Allow to cool.

Melting point (*2.2.14*)

49 °C to 56 °C.

Acid value (*2.5.1*)

Maximum 1.0.

Hydroxyl value (*2.5.3, Method A*)

208 to 228.

Iodine value (*2.5.4, Method A*)

Maximum 2.0.

Dissolve 2.00 g in *methylene chloride R* and dilute to 25 mL with the same solvent.

Saponification value (*2.5.6*)

Maximum 2.0.

ASSAY

Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 0.100 g of the substance to be examined in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 60 mg of *cetyl alcohol CRS* and 40 mg of *stearyl alcohol CRS* in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with *ethanol (96 per cent) R*.

Column:

— *size*: l = 30 m, Ø = 0.32 mm,

— *stationary phase*: *poly(dimethyl)siloxane R* (1 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 20	150 → 250
	20 - 40	250
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution:

— *resolution*: minimum 5.0 between the peaks due to cetyl alcohol and stearyl alcohol.

Calculate the percentage contents of $C_{16}H_{34}O$ and $C_{18}H_{38}O$.

Ph Eur

Emulsifying Cetostearyl Alcohol (Type A)

(Ph. Eur. monograph 0801)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of cetostearyl alcohol and sodium cetostearyl sulfate. A suitable buffer may be added.

Content

— *cetostearyl alcohol*: minimum 80.0 per cent (anhydrous substance);

— *sodium cetostearyl sulfate*: minimum 7.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or pale yellow, waxy mass, plates, flakes or granules.

Solubility

Soluble in hot water giving an opalescent solution, practically insoluble in cold water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: B, C, D.

Second identification: A, C, D.

A. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.1 g of the substance to be examined in 10 mL of *trimethylpentane R*, heating on a water-bath. Shake with 2 mL of *ethanol (70 per cent V/V) R* and allow to separate. Use the lower layer as test solution (b). Dilute 1 mL of the upper layer to 8 mL with *trimethylpentane R*.

Test solution (b) Use the lower layer obtained in the preparation of test solution (a).

Reference solution (a) Dissolve 24 mg of *cetyl alcohol CRS* and 16 mg of *stearyl alcohol CRS* in 10 mL of *trimethylpentane R*.

Reference solution (b) Dissolve 20 mg of *sodium cetostearyl sulfate R* in 10 mL of *ethanol (70 per cent V/V) R*, heating on a water-bath.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *water R, acetone R, methanol R* (20:40:40 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 50 g/L solution of *phosphomolybdic acid R* in *ethanol (96 per cent) R*; heat at 120 °C until spots appear (about 5 min).

Results:
- — the 2 principal spots in the chromatogram obtained with test solution (a) are similar in position and colour to the principal spots in the chromatogram obtained with reference solution (a);
- — 2 of the spots in the chromatogram obtained with test solution (b) are similar in position and colour to the principal spots in the chromatogram obtained with reference solution (b).

B. Examine the chromatograms obtained in the assay of cetostearyl alcohol.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 2 principal peaks in the chromatograms obtained with reference solutions (a) and (b).

C. It gives a yellow colour to a non-luminous flame.

D. To 0.3 g add 20 mL of *anhydrous ethanol R* and heat to boiling on a water-bath with shaking. Filter the mixture immediately, evaporate to dryness and take up the residue in 7 mL of *water R*. To 1 mL of the solution add 0.1 mL of a 1 g/L solution of *methylene blue R*, 2 mL of *dilute sulfuric acid R* and 2 mL of *methylene chloride R* and shake. A blue colour develops in the lower layer.

TESTS

Acid value (*2.5.1*)
Maximum 2.0.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Dissolve 2.00 g in 25 mL of *methylene chloride R*.

Saponification value (*2.5.6*)
Maximum 2.0.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 2.50 g.

ASSAY

Cetostearyl alcohol

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.200 g of *1-nonadecanol CRS* in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 0.200 g of the substance to be examined in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Reference solution (a) Dissolve 0.100 g of *cetyl alcohol CRS* in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Reference solution (b) Dissolve 0.100 g of *stearyl alcohol CRS* in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Column:
- — *material*: fused silica;
- — *size*: l = 25 m, Ø = 0.25 mm;
- — *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 20	150 → 250
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Elution order Cetyl alcohol, stearyl alcohol, 1-nonadecanol.

Calculate the percentage content of cetyl alcohol in the substance to be examined using the following expression and taking into account the assigned content of *cetyl alcohol CRS*:

$$A_x \times \frac{A_2}{A_1} \times \frac{m_{x,y}}{A_{x,y}} \times \frac{1}{m} \times 100$$

A_x = area of the peak due to cetyl alcohol in the chromatogram obtained with the test solution;
$A_{x,y}$ = area of the peak due to *cetyl alcohol CRS* in the chromatogram obtained with reference solution (a);
A_1 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_2 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (a);
m = mass of the substance to be examined in the test solution, in milligrams;
$m_{x,y}$ = mass of *cetyl alcohol CRS* in reference solution (a), in milligrams.

Calculate the percentage content of stearyl alcohol in the substance to be examined using the following expression and taking into account the assigned content of *stearyl alcohol CRS*:

$$A_z \times \frac{A_3}{A_1} \times \frac{m_{z,y}}{A_{z,y}} \times \frac{1}{m} \times 100$$

A_z = area of the peak due to stearyl alcohol in the chromatogram obtained with the test solution;
$A_{z,y}$ = area of the peak due to *stearyl alcohol CRS* in the chromatogram obtained with reference solution (b);
A_1 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_3 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (b);
m = mass of the substance to be examined in the test solution, in milligrams;
$m_{z,y}$ = mass of *stearyl alcohol CRS* in reference solution (b), in milligrams.

The percentage content of cetostearyl alcohol corresponds to the sum of the percentage contents of cetyl alcohol and stearyl alcohol.

Sodium cetostearyl sulfate

Disperse 0.300 g in 25 mL of *methylene chloride R*. Add 50 mL of *water R* and 10 mL of *dimidium bromide-sulfan blue mixed solution R*. Titrate with 0.004 M benzethonium chloride, using sonication, heating, and allowing the layers to separate before each addition, until the colour of the lower layer changes from pink to grey.

1 mL of 0.004 M benzethonium chloride is equivalent to 1.434 mg of sodium cetostearyl sulfate.

LABELLING
The label states, where applicable, the name and concentration of any added buffer.

Ph Eur

Emulsifying Cetostearyl Alcohol (Type B)

(Ph. Eur. monograph 0802)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of cetostearyl alcohol and sodium laurilsulfate.
A suitable buffer may be added.

Content
— *cetostearyl alcohol*: minimum 80.0 per cent (anhydrous substance);
— *sodium laurilsulfate*: minimum 7.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or pale yellow, waxy mass, plates, flakes or granules.

Solubility
Soluble in hot water giving an opalescent solution, practically insoluble in cold water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B, C, D.

Second identification: A, C, D.

A. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.1 g of the substance to be examined in 10 mL of *trimethylpentane R*, heating on a water-bath. Shake with 2 mL of *ethanol (70 per cent V/V) R* and allow to separate. Use the lower layer as test solution (b). Dilute 1 mL of the upper layer to 8 mL with *trimethylpentane R*.

Test solution (b) Use the lower layer obtained in the preparation of test solution (a).

Reference solution (a) Dissolve 24 mg of *cetyl alcohol CRS* and 16 mg of *stearyl alcohol CRS* in 10 mL of *trimethylpentane R*.

Reference solution (b) Dissolve 20 mg of *sodium laurilsulfate CRS* in 10 mL of *ethanol (70 per cent V/V) R*, heating on a water-bath.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *water R, acetone R, methanol R* (20:40:40 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 50 g/L solution of *phosphomolybdic acid R* in *ethanol (96 per cent) R*; heat at 120 °C until spots appear (about 5 min).

Results:
— the 2 principal spots in the chromatogram obtained with test solution (a) are similar in position and colour to the principal spots in the chromatogram obtained with reference solution (a);
— 1 of the spots in the chromatogram obtained with test solution (b) is similar in position and colour to the principal spot in the chromatogram obtained with reference solution (b).

B. Examine the chromatograms obtained in the assay of cetostearyl alcohol.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 2 principal peaks in the chromatograms obtained with reference solutions (a) and (b).

C. It gives a yellow colour to a non-luminous flame.

D. To 0.3 g add 20 mL of *anhydrous ethanol R* and heat to boiling on a water-bath with shaking. Filter the mixture immediately, evaporate to dryness and take up the residue in 7 mL of *water R*. To 1 mL of the solution add 0.1 mL of a 1 g/L solution of *methylene blue R*, 2 mL of *dilute sulfuric acid R* and 2 mL of *methylene chloride R* and shake. A blue colour develops in the lower layer.

TESTS
Acid value (*2.5.1*)
Maximum 2.0.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Dissolve 2.00 g in 25 mL of *methylene chloride R*.

Saponification value (*2.5.6*)
Maximum 2.0.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 2.50 g.

ASSAY
Cetostearyl alcohol
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.200 g of *1-nonadecanol CRS* in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 0.200 g of the substance to be examined in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Reference solution (a) Dissolve 0.100 g of *cetyl alcohol CRS* in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Reference solution (b) Dissolve 0.100 g of *stearyl alcohol CRS* in 25.0 mL of the internal standard solution. Add 25 mL of *water R* and shake with 4 quantities, each of 25 mL, of *pentane R*, adding *sodium chloride R*, if necessary, to facilitate the separation of the layers. Combine the upper layers, wash with 2 quantities, each of 30 mL, of *water R*, dry over *anhydrous sodium sulfate R* and filter.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.25 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R.*
Flow rate 1 mL/min.
Split ratio 1:100.
Temperature:

	Time (min)	Temperature (°C)
Column	0 - 20	150 → 250
Injection port		250
Detector		250

Detection Flame ionisation.
Injection 1 µL.
Elution order Cetyl alcohol, stearyl alcohol, 1-nonadecanol.

Calculate the percentage content of cetyl alcohol in the substance to be examined using the following expression and taking into account the assigned content of *cetyl alcohol CRS*:

$$A_x \times \frac{A_2}{A_1} \times \frac{m_{x,y}}{A_{x,y}} \times \frac{1}{m} \times 100$$

A_x = area of the peak due to cetyl alcohol in the chromatogram obtained with the test solution;
$A_{x,y}$ = area of the peak due to *cetyl alcohol CRS* in the chromatogram obtained with reference solution (a);
A_1 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_2 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (a);
m = mass of the substance to be examined in the test solution, in milligrams;
$m_{x,y}$ = mass of *cetyl alcohol CRS* in reference solution (a), in milligrams.

Calculate the percentage content of stearyl alcohol in the substance to be examined using the following expression and taking into account the assigned content of *stearyl alcohol CRS*:

$$A_z \times \frac{A_3}{A_1} \times \frac{m_{z,y}}{A_{z,y}} \times \frac{1}{m} \times 100$$

A_z = area of the peak due to stearyl alcohol in the chromatogram obtained with the test solution;
$A_{z,y}$ = area of the peak due to *stearyl alcohol CRS* in the chromatogram obtained with reference solution (b);
A_1 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_3 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (b);
m = mass of the substance to be examined in the test solution, in milligrams;
$m_{z,y}$ = mass of *stearyl alcohol CRS* in reference solution (b), in milligrams.

The percentage content of cetostearyl alcohol corresponds to the sum of the percentage contents of cetyl alcohol and stearyl alcohol.

Sodium laurilsulfate
Disperse 0.300 g in 25 mL of *methylene chloride R.* Add 50 mL of *water R* and 10 mL of *dimidium bromide-sulfan blue mixed solution R.* Titrate with 0.004 M benzethonium chloride, using sonication, heating, and allowing the layers to separate before each addition, until the colour of the lower layer changes from pink to grey.
1 mL of 0.004 M benzethonium chloride is equivalent to 1.154 mg of sodium laurilsulfate.

LABELLING
The label states, where applicable, the name and concentration of any added buffer.

Ph Eur

Cetostearyl Isononanoate

(Ph. Eur. monograph 1085)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of esters of cetostearyl alcohol with isononanoic acid, mainly 3,5,5-trimethylhexanoic acid.

CHARACTERS
Appearance
Clear, colourless or slightly yellowish liquid.

Solubility
Practically insoluble in water, soluble in ethanol (96 per cent) and in light petroleum, miscible with fatty oils and with liquid paraffins.

Viscosity
15 mPa·s to 30 mPa·s.

Relative density
0.85 to 0.86.

Refractive index
1.44 to 1.45.

IDENTIFICATION
A. On cooling, turbidity occurs below 15 °C.
B. Saponification value (see Tests).
C. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of cetostearyl isononanoate.*

TESTS
Appearance
The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method I*).

Acid value (*2.5.1*)
Maximum 1.0, determined on 5.0 g.

Hydroxyl value (*2.5.3, Method A*)
Maximum 5.0, determined on 4.0 g. Add 5.0 mL of acetylating reagent.

Iodine value (*2.5.4, Method A*)
Maximum 1.0.

Saponification value (*2.5.6*)
135 to 148, determined on 1.0 g.

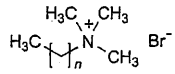
Water (*2.5.12*)
Maximum 0.2 per cent, determined on 10.0 g.

Total ash (*2.4.16*)
Maximum 0.2 per cent, determined on 2.0 g.

Ph Eur

Cetrimide

(Ph. Eur. monograph 0378)



Action and use
Antiseptic.

Preparations
Cetrimide Cream
Cetrimide Emulsifying Ointment
Strong Cetrimide Solution

Ph Eur

DEFINITION
Cetrimide consists of trimethyltetradecylammonium bromide and may contain smaller amounts of dodecyl- and hexadecyltrimethylammonium bromides.

Content
96.0 per cent to 101.0 per cent of alkyltrimethylammonium bromides, calculated as $C_{17}H_{38}BrN$ (M_r 336.4) (dried substance).

CHARACTERS
Appearance
White or almost white, voluminous, free-flowing powder.

Solubility
Freely soluble in water and in alcohol.

IDENTIFICATION
A. Dissolve 0.25 g in *alcohol R* and dilute to 25.0 mL with the same solvent. At wavelengths from 260 nm to 280 nm, the absorbance (*2.2.25*) of the solution has a maximum of 0.05.

B. Dissolve about 5 mg in 5 mL of *buffer solution pH 8.0 R*. Add about 10 mg of *potassium ferricyanide R*. A yellow precipitate is formed. Prepare a blank in the same manner but omitting the substance to be examined: a yellow solution is observed but no precipitate is formed.

C. Solution S (see Tests) froths copiously when shaken.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 0.10 g of *trimethyltetradecylammonium bromide CRS* in *water R* and dilute to 5 mL with the same solvent.

Plate *TLC silanised silica gel F_{254} plate R.*

Mobile phase *acetone R*, 270 g/L solution of *sodium acetate R*, *methanol R* (20:35:45 *V/V/V*).

Application 1 µL.

Development Over a path of 12 cm.

Drying In a current of hot air.

Detection Allow to cool; expose the plate to iodine vapour and examine in daylight.

Result The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

E. It gives reaction (a) of bromides (*2.3.1*).

TESTS
Solution S
Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 50 mL of solution S add 0.1 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.1 M hydrochloric acid* or *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Amines and amine salts
Dissolve 5.0 g in 30 mL of a mixture of 1 volume of *1 M hydrochloric acid* and 99 volumes of *methanol R* and add 100 mL of *2-propanol R*. Pass a stream of *nitrogen R* slowly through the solution. Gradually add 15.0 mL of 0.1 M tetrabutylammonium hydroxide and record the potentiometric titration curve (*2.2.20*). If the curve shows 2 points of inflexion, the volume of titrant added between the 2 points is not greater than 2.0 mL.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY
Dissolve 2.000 g in *water R* and dilute to 100.0 mL with the same solvent. Transfer 25.0 mL of the solution to a separating funnel, add 25 mL of *chloroform R*, 10 mL of *0.1 M sodium hydroxide* and 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R*. Shake, allow to separate and discard the chloroform layer. Shake the aqueous layer with 3 quantities, each of 10 mL, of *chloroform R* and discard the chloroform layers. Add 40 mL of *hydrochloric acid R*, allow to cool and titrate with *0.05 M potassium iodate* until the deep brown colour is almost discharged. Add 2 mL of *chloroform R* and continue the titration, shaking vigorously, until the colour of the chloroform layer no longer changes. Carry out a blank titration on a mixture of 10.0 mL of the freshly prepared 50 g/L solution of *potassium iodide R*, 20 mL of *water R* and 40 mL of *hydrochloric acid R*.

1 mL of *0.05 M potassium iodate* is equivalent to 33.64 mg of $C_{17}H_{38}BrN$.

Ph Eur

Strong Cetrimide Solution

Action and use
Antiseptic.

Preparation
Cetrimide Solution

DEFINITION
Strong Cetrimide Solution is an aqueous solution of cetrimide. It contains 20 to 40% w/v of cetrimide, calculated as $C_{17}H_{38}BrN$. It contains Ethanol (96 per cent) or Isopropyl Alcohol or both. It may be perfumed and may contain colouring matter.

PRODUCTION
In making Strong Cetrimide Solution, Ethanol (96 per cent) may be replaced by Industrial Methylated Spirit, provided

that the law and the statutory regulations governing the use of Industrial Methylated Spirit are observed.

Content of cetrimide $C_{17}H_{38}BrN$
95.0 to 105.0% of the stated amount.

IDENTIFICATION

A. Dilute a volume of the solution containing 0.1 g of cetrimide to 5 mL with *water* and add 2 mL of a 5% w/v solution of *potassium hexacyanoferrate(III)*. A yellow precipitate is produced.

B. Shake together 5 mL of *water*, 1 mL of 2M *sulfuric acid*, 2 mL of *chloroform* and 0.05 mL of *methyl orange solution*; the chloroform layer is colourless. Add 0.1 mL of the solution being examined and shake; a yellow colour is produced slowly in the chloroform layer.

C. Yields reaction A characteristic of *bromides*, Appendix VI.

TESTS

Acidity or alkalinity
Dilute a volume of the solution containing 10 g of cetrimide to 100 mL and add 0.1 mL of *bromocresol purple solution*. Not more than 1.0 mL of either *0.1M hydrochloric acid VS* or *0.1M sodium hydroxide VS* is required to change the colour of the solution.

Miscibility with ethanol
Mix a volume of the solution containing 1.6 g of cetrimide with a mixture of 2 mL of *water* and 16 mL of *ethanol (96%)*. The solution remains *clear*, Appendix IV A.

Neutral substances
To a volume of the solution containing 10 g of cetrimide add 25 mL of *ethanol (50%)*, acidify to *bromophenol blue solution* by the drop wise addition of *hydrochloric acid* and add 0.05 mL in excess. Transfer quantitatively to the extraction compartment of an apparatus designed for continuous liquid-liquid extraction by fluids of a lesser density than water, washing out the beaker with 10 mL *ethanol (50%)* and adding the washings to the bulk of the solution in the extractor. Add sufficient *ethanol (50%)*, if necessary, to half-fill the extraction chamber to the level of the overflow limb. Add sufficient *purified hexane* to fill the extraction chamber, secure an overflow volume of about 30 mL in the ebullition flask and heat using an electrically heated mantle. Ensure that a continuous flow of hexane through the aqueous ethanol layer is observed and continue the extraction for 16 hours. Transfer the hexane extract to a separating funnel, washing out the flask with 10 mL of *purified hexane*. Shake the combined extract and washings with 25 mL of *ethanol (50%)* and discard the aqueous ethanol layer. Filter the hexane layer through a dry filter paper (Whatman No. 1 is suitable) into a tared flask and remove the solvent using a rotary evaporator at 40° and then at room temperature at a pressure not exceeding 0.7 kPa for 2 hours. The residue weighs not more than 0.4 g.

Non-quaternised amines
To a volume of the solution containing 10 g of cetrimide add a mixture of 100 mL of *propan-2-ol*, 0.1 mL of *hydrochloric acid* and 20 mL of *methanol*. Titrate with 0.1M *tetrabutylammonium hydroxide VS* passing a slow current of *nitrogen* through the solution and determining the end point potentiometrically using a platinum-glass electrode system. Inflections in the titration curve indicate (A) neutralisation of excess hydrochloric acid and (B) neutralisation of non-quaternised amine salts. The difference between the volumes corresponding to A and B is not more than 10 mL (2.4%, calculated as $C_{16}H_{35}N$).

Ethanol; Isopropyl alcohol
Carry out one or both of the following methods according to the declared alcohol content of the solution being examined.

Ethanol Not more than 10.0% v/v, by the method for the *determination of ethanol*, Appendix VIII F. Use on-column injection and do not heat the injection port.

Isopropyl alcohol Not more than 10.0% v/v, by the method for the *determination of ethanol*, Appendix VIII F, with the following modifications. For solution (1) use a solution containing 5.0% v/v of *propan-2-ol* and 5.0% v/v of *propan-1-ol* (internal standard). For solution (2) use the solution being examined, diluted with *water*, if necessary, to contain about 5.0% v/v of isopropyl alcohol. Maintain the column temperature at 170°, use on-column injection and do not heat the injection port.

ASSAY

Dilute a volume containing 4 g of cetrimide with sufficient *water* to produce 100 mL. Transfer 25 mL of the solution to a separating funnel and add 25 mL of *chloroform*, 10 mL of 0.1M *sodium hydroxide* and 10 mL of a freshly prepared 8.0% w/v solution of *potassium iodide*. Shake well, allow to separate and discard the chloroform layer. Wash the aqueous layer with three 10 mL quantities of *chloroform* and discard the washings. Add 40 mL of *hydrochloric acid*, cool and titrate with *0.05M potassium iodate VS* until the deep brown colour is almost discharged. Add 2 mL of *chloroform* and continue the titration, with shaking, until the chloroform layer becomes colourless. Carry out a blank titration on a mixture of 10 mL of the freshly prepared potassium iodide solution, 20 mL of *water* and 40 mL of *hydrochloric acid*. The difference between the titrations represents the amount of potassium iodate required. Each mL of *0.05M potassium iodate VS* is equivalent to 33.64 mg of $C_{17}H_{38}BrN$.

STORAGE

Strong Cetrimide Solution should be stored at a temperature above 15°.

LABELLING

The label states whether Ethanol, Isopropyl Alcohol or both are present and the percentage of cetrimide, weight in volume.

Cetyl Alcohol

(Ph. Eur. monograph 0540)

Action and use
Excipient.

Ph Eur

DEFINITION

Mixture of solid alcohols, mainly hexadecan-1-ol ($C_{16}H_{34}O$; M_r 242.4), of animal or vegetable origin.

Content
Minimum 95.0 per cent of $C_{16}H_{34}O$.

CHARACTERS

Appearance
White or almost white, unctuous mass, powder, flakes or granules.

Solubility
Practically insoluble in water, freely soluble or sparingly soluble in ethanol (96 per cent). When melted, it is miscible

with vegetable and animal oils, with liquid paraffin and with melted wool fat.

IDENTIFICATION

Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_6 (*2.2.2, Method II*).

Dissolve 0.50 g in 20 mL of boiling *ethanol (96 per cent) R*. Allow to cool.

Melting point (*2.2.14*)

46 °C to 52 °C.

Acid value (*2.5.1*)

Maximum 1.0.

Hydroxyl value (*2.5.3, Method A*)

218 to 238.

Iodine value (*2.5.4, Method A*)

Maximum 2.0.

Dissolve 2.00 g in *methylene chloride R* and dilute to 25 mL with the same solvent.

Saponification value (*2.5.6*)

Maximum 2.0.

ASSAY

Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 0.100 g of the substance to be examined in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *cetyl alcohol CRS* in *ethanol (96 per cent) R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 50 mg of *stearyl alcohol R* in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Reference solution (c) Mix 1 mL of reference solution (a) and 1 mL of reference solution (b) and dilute to 10 mL with *ethanol (96 per cent) R*.

Column:

— *size*: *l* = 30 m, Ø = 0.32 mm,

— *stationary phase*: *poly(dimethyl)siloxane R* (1 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 20	150 → 250
	20 - 40	250
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL of the test solution and reference solutions (a) and (c).

System suitability Reference solution (c):

— *resolution*: minimum 5.0 between the peaks due to cetyl alcohol and stearyl alcohol.

Calculate the percentage content of $C_{16}H_{34}O$.

Ph Eur

Cetyl Palmitate

(Ph. Eur. monograph 1906)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of esters of C_{14}-C_{18} alcohols with lauric (dodecanoic), myristic (tetradecanoic), palmitic (hexadecanoic) and stearic (octadecanoic) acids ('Cetyl esters wax').

Content

(expressed as hexadecyl hexadecanoate):

10.0 per cent to 20.0 per cent for Cetyl palmitate 15, 60.0 per cent to 70.0 per cent for Cetyl palmitate 65 and minimum 90.0 per cent for Cetyl palmitate 95.

CHARACTERS

Appearance

White or almost white, waxy plates, flakes or powder.

Solubility

Practically insoluble in water, soluble in boiling anhydrous ethanol and in methylene chloride, slightly soluble in light petroleum, practically insoluble in anhydrous ethanol.

mp

About 45 °C for Cetyl palmitate 15 and Cetyl palmitate 65 and about 52 °C for Cetyl palmitate 95.

IDENTIFICATION

A. It complies with the limits of the assay and the chromatogram obtained with the test solution shows the typical main peak(s).

B. Saponification value (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 4.0 g in *methylene chloride R* and dilute to 20 mL with the same solvent.

Acid value (*2.5.1*)

Maximum 4.0.

Dissolve 10.0 g in 50 mL of the solvent mixture described by heating under reflux on a water-bath for 5 min.

Hydroxyl value (*2.5.3, Method A*)

Maximum 20.0.

Carry out the titration at a temperature between 55 °C and 70 °C, shaking the flask towards the end of the titration.

Iodine value (*2.5.4, Method A*)

Maximum 2.0.

Saponification value (*2.5.6*)

105 to 120.

Heat under reflux for 2 h.

Alkaline impurities
Dissolve 2.0 g with gentle heating in a mixture of 1.5 mL of *ethanol (96 per cent) R* and 3 mL of *toluene R*. Add 0.05 mL of a 0.4 g/L solution of *bromophenol blue R* in *ethanol (96 per cent) R*. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the solution to yellow.

Nickel (*2.4.31*)
Maximum 1 ppm.

Water (*2.5.12*)
Maximum 0.3 per cent, determined on 1.0 g using a mixture of equal volumes of *anhydrous methanol R* and *methylene chloride R* as solvent.

Total ash (*2.4.16*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 20.0 mg of the substance to be examined in *hexane R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 20.0 mg of *cetyl palmitate 95 CRS* in *hexane R* and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dissolve 20.0 mg of *cetyl palmitate 15 CRS* in *hexane R* and dilute to 20.0 mL with the same solvent.

Column:
— *material*: stainless steel;
— *size*: *l* = 10 m, Ø = 0.53 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 2.65 µm).

Carrier gas *helium for chromatography R.*

Flow rate 6.5 mL/min.

Split ratio 1:10.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 10	100 → 300
	10 - 15	300
Injection port		350
Detector		350

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to cetyl palmitate (retention time = about 9 min): cetyl alcohol = about 0.3; palmitic acid = about 0.4; lauric ester = about 0.8; myristic ester = about 0.9; stearic ester = about 1.1.

System suitability Reference solution (b):
— *resolution*: minimum of 1.5 between the peaks due to cetyl palmitate and cetyl stearate.

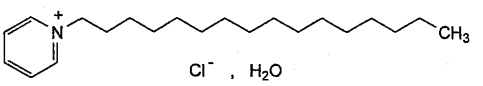
STORAGE
At a temperature not exceeding 25 °C.

LABELLING
The label states the type of cetyl palmitate.

Ph Eur

Cetylpyridinium Chloride

(Ph. Eur. monograph 0379)



$C_{21}H_{38}ClN,H_2O$ 358.0 *6004-24-6*

Action and use
Antiseptic.

Ph Eur

DEFINITION
Cetylpyridinium chloride contains not less than 96.0 per cent and not more than the equivalent of 101.0 per cent of 1-hexadecylpyridinium chloride, calculated with reference to the anhydrous substance.

CHARACTERS
A white or almost white powder, slightly soapy to the touch, soluble in water and in alcohol. An aqueous solution froths copiously when shaken.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Dissolve 0.10 g in *water R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *water R*. Examined between 240 nm and 300 nm (*2.2.25*), the solution shows an absorption maximum at 259 nm and 2 shoulders at about 254 nm and at about 265 nm. The specific absorbance at the maximum is 126 to 134, calculated with reference to the anhydrous substance.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *cetylpyridinium chloride CRS*. Examine the substances in the solid state.

C. To 5 mL of *dilute sodium hydroxide solution R* add 0.1 mL of *bromophenol blue solution R1* and 5 mL of *chloroform R* and shake. The chloroform layer is colourless. Add 0.1 mL of solution S (see Tests) and shake. The chloroform layer becomes blue.

D. Solution S gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and is colourless (*2.2.2, Method II*).

Acidity
To 50 mL of solution S add 0.1 mL of *phenolphthalein solution R*. Not more than 2.5 mL of *0.02 M sodium hydroxide* is required to change the colour of the indicator.

Amines and amine salts
Dissolve 5.0 g with heating in 20 mL of a mixture of 3 volumes of *1 M hydrochloric acid* and 97 volumes of *methanol R* and add 100 mL of *2-propanol R*. Pass a stream of *nitrogen R* slowly through the solution. Gradually add 12.0 mL of 0.1 M tetrabutylammonium hydroxide and record the potentiometric titration curve (*2.2.20*). If the curve shows 2 points of inflexion, the volume of titrant added between the two points is not greater than 5.0 mL. If the

curve shows no point of inflexion, the substance to be examined does not comply with the test. If the curve shows one point of inflexion, repeat the test but add 3.0 mL of a 25.0 g/L solution of *dimethyldecylamine R* in *2-propanol R* before the titration. If the titration curve after the addition of 12.0 mL of the titrant shows only one point of inflexion, the substance to be examined does not comply with the test.

Water (*2.5.12*)
4.5 per cent to 5.5 per cent, determined on 0.300 g by the semi-micro determination of water.

Sulfated ash (*2.4.14*)
Not more than 0.2 per cent, determined on 1.0 g.

ASSAY
Dissolve 2.00 g in *water R* and dilute to 100.0 mL with the same solvent. Transfer 25.0 mL of the solution to a separating funnel, add 25 mL of *chloroform R*, 10 mL of *0.1 M sodium hydroxide* and 10.0 mL of a freshly prepared 50 g/L solution of *potassium iodide R*. Shake well, allow to separate and discard the chloroform layer. Shake the aqueous layer with three quantities, each of 10 mL, of *chloroform R* and discard the chloroform layers. To the aqueous layer add 40 mL of *hydrochloric acid R*, allow to cool and titrate with *0.05 M potassium iodate* until the deep-brown colour is almost discharged. Add 2 mL of *chloroform R* and continue the titration, shaking vigorously, until the chloroform layer no longer changes colour. Carry out a blank titration on a mixture of 10.0 mL of the freshly prepared 50 g/L solution of *potassium iodide R*, 20 mL of *water R* and 40 mL of *hydrochloric acid R*.

1 mL of *0.05 M potassium iodate* is equivalent to 34.0 mg of $C_{21}H_{38}ClN$.

Ph Eur

Chalk

Prepared Chalk

$CaCO_3$ 100.1

Action and use
Antacid.

DEFINITION
Chalk is a native form of calcium carbonate freed from most of its impurities by elutriation and dried. It contains not less than 97.0% and not more than 100.5% of $CaCO_3$, calculated with reference to the dried substance.

CHARACTERISTICS
Chalk absorbs water readily.

Practically insoluble in *water*; slightly soluble in *water* containing carbon dioxide.

Macroscopical White or greyish white, small friable masses, usually conical in form, or in powder; amorphous; earthy; soft to the touch.

Microscopical Consists of the calcareous shells and detritus of various foraminifera; the calcareous shells vary from about 35 to 100 µm in breadth and from about 50 to 180 µm in length; among the detritus are numerous small rings and discs about 5 to 10 µm in diameter.

IDENTIFICATION
A. A solution in 6M *acetic acid* yields reaction C characteristic of *calcium salts*, Appendix VI.

B. Yields reaction A characteristic of *carbonates*, Appendix VI.

TESTS
Acidity or alkalinity
1 g, boiled with 50 mL of *water* and filtered, yields a filtrate which is neutral to *bromothymol blue solution R3* or requires not more than 0.05 mL of *0.1M hydrochloric acid VS* to make it so.

Aluminium, iron, phosphate and matter insoluble in hydrochloric acid
Dissolve 2 g in a mixture of 5 mL of *hydrochloric acid* and 75 mL of *water*, boil to remove carbon dioxide and make alkaline with 5M *ammonia* using *methyl red solution* as indicator. Boil for 1 minute, filter and wash the precipitate with a hot 2% w/v solution of *ammonium chloride*. Dissolve the precipitate as completely as possible by passing 20 mL of hot 2M *hydrochloric acid* through the filter and wash the filter with sufficient hot *water* to adjust the volume of the solution to 50 mL. Boil the solution and make alkaline with 5M *ammonia* using *methyl red solution* as indicator. Boil for 1 minute, filter through the same filter, wash the precipitate with a hot 2% w/v solution of *ammonium nitrate*, dry and ignite at a temperature not lower than 1000°. The residue weighs not more than 40 mg.

Arsenic
Dissolve 0.5 g in 5 mL of *brominated hydrochloric acid* and dilute to 50 mL with *water*. 25 mL of the resulting solution complies with the *limit test for arsenic*, Appendix VII (4 ppm).

Chloride
Dissolve 0.3 g in 2 mL of *nitric acid* and 10 mL of *water*, filter and dilute the filtrate to 30 mL with *water*. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (330 ppm).

Sulfate
Dissolve 0.25 g in 5.5 mL of 2M *hydrochloric acid*, dilute to 30 mL with *water* and filter. 15 mL of the resulting solution complies with the *limit test for sulfates*, Appendix VII (0.12%).

Loss on drying
When dried to constant weight at 105°, loses not more than 1.0% of its weight. Use 1 g.

ASSAY
To 2 g in 100 mL of *water* add 50 mL of *1M hydrochloric acid VS*, boil to remove carbon dioxide, cool and titrate the excess of acid with *1M sodium hydroxide VS* using *methyl orange solution* as indicator. Each mL of *1M hydrochloric acid VS* is equivalent to 50.04 mg of $CaCO_3$.

Activated Charcoal

Decolourising Charcoal

(Ph. Eur. monograph 0313)

Action and use
Adsorbent.

Ph Eur

DEFINITION
Obtained from vegetable matter by suitable carbonisation processes intended to confer a high adsorption power.

CHARACTERS
Appearance
Black, light powder free from grittiness.

Solubility
Practically insoluble in all usual solvents.

IDENTIFICATION

A. When heated to redness it burns slowly without a flame.

B. Adsorption power (see Tests).

TESTS

Solution S

To 2.0 g in a conical flask with a ground-glass neck add 50 mL of *dilute hydrochloric acid R*. Boil gently under a reflux condenser for 1 h, filter and wash the filter with *dilute hydrochloric acid R*. Evaporate the combined filtrate and washings to dryness on a water-bath, dissolve the residue in *0.1 M hydrochloric acid* and dilute to 50.0 mL with the same acid.

Acidity or alkalinity

To 2.0 g add 40 mL of *water R* and boil for 5 min. Cool, restore to the original mass with *carbon dioxide-free water R* and filter. Reject the first 20 mL of the filtrate. To 10 mL of the filtrate add 0.25 mL of *bromothymol blue solution R1* and 0.25 mL of *0.02 M sodium hydroxide*. The solution is blue. Not more than 0.75 mL of *0.02 M hydrochloric acid* is required to change the colour of the indicator to yellow.

Acid-soluble substances

Maximum 3 per cent.

To 1.0 g add 25 mL of *dilute nitric acid R* and boil for 5 min. Filter whilst hot through a sintered-glass filter (10) (*2.1.2*) and wash with 10 mL of hot *water R*. Evaporate the combined filtrate and washings to dryness on a water-bath, add to the residue 1 mL of *hydrochloric acid R*, evaporate to dryness again and dry the residue to constant mass at 100-105 °C. The residue weighs a maximum of 30 mg.

Alkali-soluble coloured substances

To 0.25 g add 10 mL of *dilute sodium hydroxide solution R* and boil for 1 min. Cool, filter and dilute the filtrate to 10 mL with *water R*. The solution is not more intensely coloured than reference solution GY_4 (*2.2.2, Method II*).

Ethanol (96 per cent) soluble substances

Maximum 0.5 per cent.

To 2.0 g add 50 mL of *ethanol (96 per cent) R* and boil under a reflux condenser for 10 min. Filter immediately, cool, and dilute to 50 mL with *ethanol (96 per cent) R*. The filtrate is not more intensely coloured than reference solution Y_6 or BY_6 (*2.2.2, Method II*). Evaporate 40 mL of the filtrate to dryness and dry to constant mass at 100-105 °C. The residue weighs a maximum of 8 mg.

Fluorescent substances

In an intermittent-extraction apparatus, treat 10.0 g with 100 mL of *cyclohexane R1* for 2 h. Collect the liquid and dilute to 100 mL with *cyclohexane R1*. Examine in ultraviolet light at 365 nm. The fluorescence of the solution is not more intense than that of a solution of 83 µg of *quinine R* in 1000 mL of *0.005 M sulfuric acid* examined under the same conditions.

Sulfides

To 1.0 g in a conical flask add 5 mL of *hydrochloric acid R1* and 20 mL of *water R*. Heat to boiling. The fumes released do not turn *lead acetate paper R* brown.

Copper

Maximum 25 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Use solution S.

Reference solutions Prepare the reference solutions using *copper standard solution (0.1 per cent Cu) R* and diluting with *0.1 M hydrochloric acid.*

Source Copper hollow-cathode lamp.

Wavelength 325.0 nm.

Atomisation device Air-acetylene flame.

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Use solution S.

Reference solutions Prepare the reference solutions using *lead standard solution (100 ppm Pb) R* and diluting with *0.1 M hydrochloric acid.*

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm; 217.0 nm may be used depending on the apparatus.

Atomisation device Air-acetylene flame.

Zinc

Maximum 25 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Use solution S.

Reference solutions Prepare the reference solutions using *zinc standard solution (100 ppm Zn) R* and diluting with *0.1 M hydrochloric acid.*

Source Zinc hollow-cathode lamp.

Wavelength 214.0 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)

Maximum 15 per cent, determined on 1.00 g by drying in an oven at 120 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 5.0 per cent, determined on 1.0 g.

Adsorption power

To 0.300 g in a 100 mL ground-glass-stoppered conical flask add 25.0 mL of a freshly prepared solution of 0.5 g of *phenazone R* in 50 mL of *water R*. Shake thoroughly for 15 min. Filter and reject the first 5 mL of filtrate. To 10.0 mL of the filtrate add 1.0 g of *potassium bromide R* and 20 mL of *dilute hydrochloric acid R*. Using 0.1 mL of *methyl red solution R* as indicator, titrate with *0.0167 M potassium bromate* until the red colour is discharged. Titrate slowly (1 drop every 15 s) towards the end of the titration. Carry out a blank titration using 10.0 mL of the phenazone solution.

Calculate the quantity of phenazone adsorbed per 100 g of activated charcoal from the following expression:

$$\frac{2.353(a-b)}{m}$$

a = number of millilitres of *0.0167 M potassium bromate* used for the blank;

b = number of millilitres of *0.0167 M potassium bromate* used for the test;

m = mass in grams of the substance to be examined.

Minimum 40 g of phenazone is adsorbed per 100 g of activated charcoal, calculated with reference to the dried substance.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

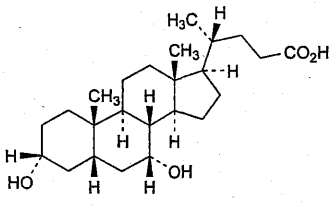
STORAGE

In an airtight container.

Ph Eur

Chenodeoxycholic Acid

(Ph. Eur. monograph 1189)



$C_{24}H_{40}O_4$ 392.6 *474-25-9*

Action and use
Bile acid; treatment of gallstones.

Ph Eur

DEFINITION
Chenodeoxycholic acid contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 3α,7α-dihydroxy-5β-cholan-24-oic acid, calculated with reference to the dried substance.

CHARACTERS
A white or almost white powder, very slightly soluble in water, freely soluble in alcohol, soluble in acetone, slightly soluble in methylene chloride.

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *chenodeoxycholic acid CRS*. Examine the substances prepared as discs using *potassium bromide R*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 1 mL of *sulfuric acid R*. Add 0.1 mL of *formaldehyde solution R* and allow to stand for 5 min. Add 5 mL of *water R*. The suspension obtained is greenish-blue.

TESTS
Specific optical rotation (*2.2.7*)
Dissolve 0.500 g in *methanol R* and dilute to 25.0 mL with the same solvent. The specific optical rotation is + 11.0 to + 13.0, calculated with reference to the dried substance.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using a suitable silica gel as the coating substance.

Test solution (a) Dissolve 0.40 g of the substance to be examined in a mixture of 1 volume of *water R* and 9 volumes of *acetone R* and dilute to 10 mL with the same mixture of solvents.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of *water R* and 9 volumes of *acetone R*.

Reference solution (a) Dissolve 40 mg of *chenodeoxycholic acid CRS* in a mixture of 1 volume of *water R* and 9 volumes of *acetone R* and dilute to 10 mL with the same mixture of solvents.

Reference solution (b) Dissolve 20 mg of *lithocholic acid CRS* in a mixture of 1 volume of *water R* and 9 volumes of *acetone R* and dilute to 10 mL with the same mixture of solvents. Dilute 2 mL of the solution to 100 mL with a mixture of 1 volume of *water R* and 9 volumes of *acetone R*.

Reference solution (c) Dissolve 20 mg of *ursodeoxycholic acid CRS* in a mixture of 1 volume of *water R* and 9 volumes of *acetone R* and dilute to 50 mL with the same mixture of solvents.

Reference solution (d) Dissolve 20 mg of *cholic acid CRS* in a mixture of 1 volume of *water R* and 9 volumes of *acetone R* and dilute to 100 mL with the same mixture of solvents.

Reference solution (e) Dilute 0.5 mL of test solution (a) to 20 mL with a mixture of 1 volume of *water R* and 9 volumes of *acetone R*. Dilute 1 mL of the solution to 10 mL with a mixture of 1 volume of *water R* and 9 volumes of *acetone R*.

Reference solution (f) Dissolve 10 mg of *chenodeoxycholic acid CRS* in reference solution (c) and dilute to 25 mL with the same solution.

Apply separately to the plate 5 µL of each solution. Develop in an unsaturated tank over a path of 15 cm using a mixture of 1 volume of *glacial acetic acid R*, 30 volumes of *acetone R* and 60 volumes of *methylene chloride R*. Dry the plate at 120 °C for 10 min. Spray the plate immediately with a 47.6 g/L solution of *phosphomolybdic acid R* in a mixture of 1 volume of *sulfuric acid R* and 20 volumes of *glacial acetic acid R* and heat again at 120 °C until blue spots appear on a lighter background. In the chromatogram obtained with test solution (a): any spot corresponding to lithocholic acid is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.1 per cent); any spot corresponding to ursodeoxycholic acid is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (1 per cent); any spot corresponding to cholic acid is not more intense than the principal spot in the chromatogram obtained with reference solution (d) (0.5 per cent); any spot apart from the principal spot and any spots corresponding to lithocholic acid, ursodeoxycholic acid and cholic acid, is not more intense than the principal spot in the chromatogram obtained with reference solution (e) (0.25 per cent). The test is not valid unless the chromatogram obtained with reference solution (f) shows two clearly separated principal spots.

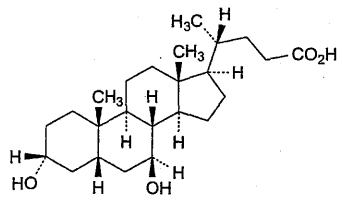
Loss on drying (*2.2.32*)
Not more than 1.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

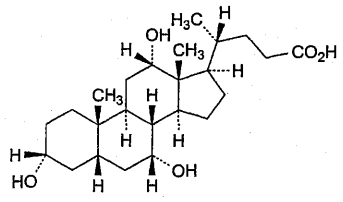
ASSAY
Dissolve 0.350 g in 50 mL of *alcohol R*, previously neutralised to 0.2 mL of *phenolphthalein solution R*. Add 50 mL of *water R* and titrate with *0.1 M sodium hydroxide* until a pink colour is obtained.

1 mL of *0.1 M sodium hydroxide* is equivalent to 39.26 mg of $C_{24}H_{40}O_4$.

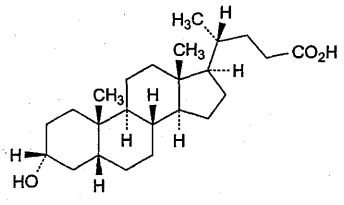
IMPURITIES



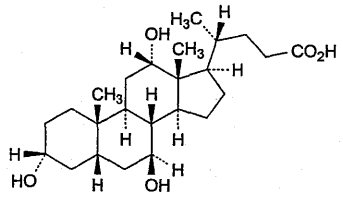
A. 3α-7β-dihydroxy-5β-cholan-24-oic acid (ursodeoxycholic acid),



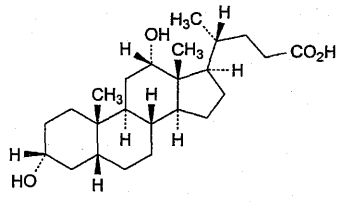
B. 3α,7α,12α-trihydroxy-5β-cholan-24-oic acid (cholic acid),



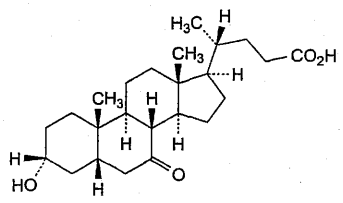
C. 3α-hydroxy-5β-cholan-24-oic acid (lithocholic acid),



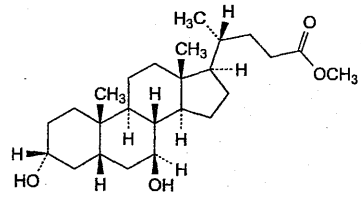
D. 3α,7β,12α-trihydroxy-5β-cholan-24-oic acid (ursocholic acid),



E. 3α,12α-dihydroxy-5β-cholan-24-oic acid (deoxycholic acid),



F. 3α-hydroxy-7-oxo-5β-cholan-24-oic acid,



G. methyl 3α,7β-dihydroxy-5β-cholan-24-oate.

Ph Eur

Chitosan Hydrochloride

(Ph. Eur. monograph 1774)

Ph Eur

DEFINITION

Chitosan hydrochloride is the chloride salt of an unbranched binary heteropolysaccharide consisting of the two units *N*-acetyl-D-glucosamine and D-glucosamine, obtained by partial deacetylation of chitin normally leading to a degree of deacetylation of 70.0 per cent to 95.0 per cent. Chitin is extracted from the shells of shrimp and crab.

PRODUCTION

The animals from which chitosan hydrochloride is derived must fulfil the requirements for the health of animals suitable for human consumption to the satisfaction of the competent authority. It must have been shown to what extent the method of production allows inactivation or removal of any contamination by viruses or other infectious agents.

CHARACTERS

Appearance

White or almost white, fine powder.

Solubility

Sparingly soluble in water, practically insoluble in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *chitosan hydrochloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

C. Dilute 50 mL of solution S (see Tests) to 250 mL with a 25 per cent *V/V* solution of *ammonia R*. A voluminous gelatinous mass is formed.

D. To 10 mL of solution S add 90 mL of *acetone R*. A voluminous gelatinous mass is formed.

TESTS

Solution S

Dissolve 1.0 g in 100 mL of *water R* and stir vigorously for 20 min with a mechanical stirrer.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Matter insoluble in water

Maximum 0.5 per cent.

Add 2.00 g to 400.0 mL of *water R* while stirring until no further dissolution takes place. Transfer the solution to a 2 L beaker, and add 200 mL of *water R*. Boil the solution gently for 2 h, covering the beaker during the operation. Filter through a sintered-glass filter (40) (*2.1.2*), wash the residue

with water and dry to constant weight in an oven at 100-105 °C. The residue weighs a maximum of 10 mg.

pH (*2.2.3*)
4.0 to 6.0 for solution S.

Viscosity (*2.2.10*)
80 per cent to 120 per cent of the value stated on the label, determined on solution S.

Determine the viscosity using a rotating viscometer at 20 °C with a spindle rotating at 20 r/min, using a suitable spindle for the range of the expected viscosity.

Degree of deacetylation
Test solution Dissolve 0.250 g in *water R* and dilute to 50.0 mL with the same solvent, stirring vigorously. Dilute 1.0 mL of this solution to 100.0 mL with *water R*. Measure the absorbance (*2.2.25*) from 200 nm to 205 nm as the first derivative of the absorbance curve. Determine the pH of the solution.

Reference solutions Prepare solutions of 1.0 µg/mL, 5.0 µg/mL, 15.0 µg/mL and 35.0 µg/mL of *N-acetylglucosamine R* in *water R*. Measure the absorbance (*2.2.25*) from 200 nm to 205 nm of each solution as the first derivative of the absorption curve. Make a standard curve by plotting the first derivative at 202 nm as a function of the concentration of *N*-acetylglucosamine, and calculate the slope of the curve by least squares linear regression. Use the standard curve to determine the equivalent amount of *N*-acetylglucosamine for the substance to be examined.

Calculate the degree of deacetylation (molar) using the following expression:

$$\frac{100 \times M_1 \times (C_1 - C_2)}{(M_1 \times C_1) - [(M_1 - M_3) \times C_2]}$$

C_1 = concentration of chitosan hydrochloride in the test solution in micrograms per millilitre;
C_2 = concentration of *N*-acetylglucosamine in the test solution, as determined from the standard curve prepared using the reference solution in micrograms per millilitre;
M_1 = 203 (relative molecular mass of *N*-acetylglucosamine unit ($C_8H_{13}NO_5$) in polymer);
M_3 = relative molecular mass of chitosan hydrochloride.

M_3 is calculated from the pH in solution, assuming a pKa value of 6.8, using the following equations:

$$M_3 = f \times M_2 + (1 - f) \times (M_2 + 36.5)$$

$$f = \frac{p}{1 + p}$$

$$p = 10^{(\mathrm{pH} - \mathrm{pKa})}$$

M_2 = 161 (relative molecular mass of deacetylated unit (glucosamine) ($C_6H_{11}NO_4$) in polymer).

Chlorides
10.0 per cent to 20.0 per cent.

Introduce 0.200 g into a 250 mL borosilicate flask fitted with a reflux condenser. Add 40 mL of a mixture of 1 volume of *nitric acid R* and 2 volumes of *water R*. Boil gently under a reflux condenser for 5 min. Cool and add 25 mL of *water R* through the condenser. Add 16.0 mL of *0.1 M silver nitrate*, shake vigorously and titrate with *0.1 M ammonium thiocyanate*, using 1 mL of *ferric ammonium sulfate solution R2* as indicator, and shaking vigorously towards the end-point. Carry out a blank titration.

1 mL of *0.1 M silver nitrate* is equivalent to 3.55 mg of Cl.

Loss on drying (*2.2.32*)
Maximum 10 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.0 g.

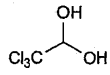
STORAGE
At a temperature of 2 °C to 8 °C, protected from moisture and light.

LABELLING
The label states the nominal viscosity in millipascal seconds for a 10 g/L solution in *water R*.

Ph Eur

Chloral Hydrate

(Ph. Eur. monograph 0265)



$C_2H_3Cl_3O_2$ 165.4 *302-17-0*

Action and use
Hypnotic.

Preparation
Chloral Hydrate Oral Solution

Ph Eur

DEFINITION
2,2,2-Trichloroethane-1,1-diol.

Content
98.5 per cent to 101.0 per cent.

CHARACTERS
Appearance
Colourless, transparent crystals.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION
A. To 10 mL of solution S (see Tests) add 2 mL of *dilute sodium hydroxide solution R*. The mixture becomes cloudy and, when heated, gives off an odour of chloroform.

B. To 1 mL of solution S add 2 mL of *sodium sulfide solution R*. A yellow colour develops which quickly becomes reddish-brown. On standing for a short time, a red precipitate may be formed.

TESTS
Solution S
Dissolve 3.0 g in *carbon dioxide-free water R* and dilute to 30 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
3.5 to 5.5 for solution S.

Chloral alcoholate
Warm 1.0 g with 10 mL of *dilute sodium hydroxide solution R*, filter the supernatant solution and add *0.05 M iodine* dropwise until a yellow colour is obtained. Allow to stand for 1 h. No precipitate is formed.

Chlorides (*2.4.4*)
Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Non-volatile residue
Maximum 0.1 per cent.

Evaporate 2.000 g on a water-bath. The residue weighs a maximum of 2 mg.

ASSAY
Dissolve 4.000 g in 10 mL of *water R* and add 40.0 mL of *1 M sodium hydroxide*. Allow to stand for exactly 2 min and titrate with *0.5 M sulfuric acid*, using 0.1 mL of *phenolphthalein solution R* as indicator. Titrate the neutralised solution with *0.1 M silver nitrate*, using 0.2 mL of *potassium chromate solution R* as indicator. Calculate the number of millilitres of *1 M sodium hydroxide* used by deducting from the volume of *1 M sodium hydroxide*, added at the beginning of the titration, the volume of *0.5 M sulfuric acid* used in the 1st titration and two-fifteenths of the volume of *0.1 M silver nitrate* used in the 2nd titration.

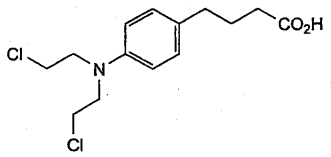
1 mL of *1 M sodium hydroxide* is equivalent to 0.1654 g of $C_2H_3Cl_3O_2$.

STORAGE
In an airtight container.

Ph Eur

Chlorambucil

(Ph. Eur. monograph 0137)



$C_{14}H_{19}Cl_2NO_2$ 304.2 *305-03-3*

Action and use
Cytotoxic alkylating agent.

Preparation
Chlorambucil Tablets

Ph Eur

DEFINITION
4-[4-[Bis(2-chloroethyl)amino]phenyl]butanoic acid.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *chlorambucil CRS*.

TESTS
Impurity G
Liquid chromatography (*2.2.29*). *The solutions are stable for 8 h at room temperature or for 24 h at 4-8 °C; protect them from light.*

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *chlorambucil with impurity G CRS* in *methanol R* and dilute to 10.0 mL with the same solvent.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R*, 1 per cent *V/V* solution of *trifluoroacetic acid R* (50:50 *V/V*).

Flow rate 1.8 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 µL.

Run time Twice the retention time of chlorambucil.

Relative retention With reference to chlorambucil (retention time = about 11 min): impurity G = about 1.2.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to chlorambucil and impurity G.

Limit:
— *impurity G*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Solvent mixture 10.3 g/L solution of *hydrochloric acid R*, *acetonitrile for chromatography R* (10:90 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *chlorambucil for system suitability CRS* (containing impurities B and E) in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 1.9 g/L solution of *ammonium acetate R* adjusted to pH 3.9 with *acetic acid R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	60	40
5 - 15	60 → 10	40 → 90
15 - 25	10	90

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *chlorambucil for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and E.

Relative retention With reference to chlorambucil (retention time = about 12 min): impurity B = about 0.5; impurity E = about 1.4.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity B and chlorambucil.

Limits:
— *impurity E*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *impurity B*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 10 mL of *acetone R* and add 10 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, using 0.1 mL of *phenolphthalein solution R* as indicator.

1 mL of *0.1 M sodium hydroxide* is equivalent to 30.42 mg of $C_{14}H_{19}Cl_2NO_2$.

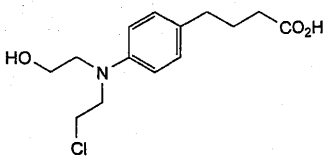
STORAGE

Protected from light.

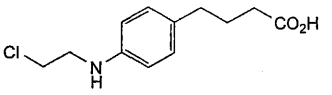
IMPURITIES

Specified impurities *B, E, G.*

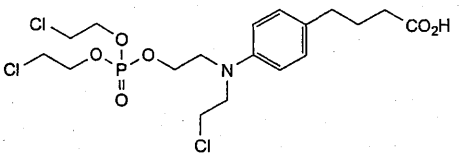
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, F.*



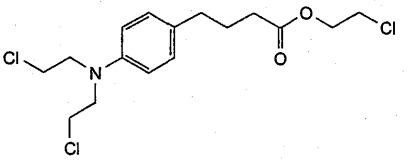
A. 4-[4-[(2-chloroethyl)(2-hydroxyethyl)amino]phenyl] butanoic acid,



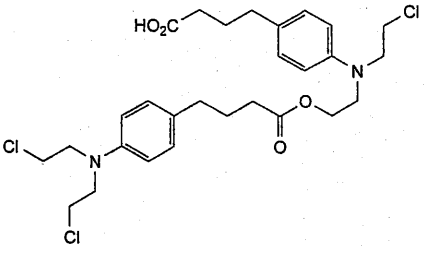
B. 4-[4-[(2-chloroethyl)amino]phenyl]butanoic acid,



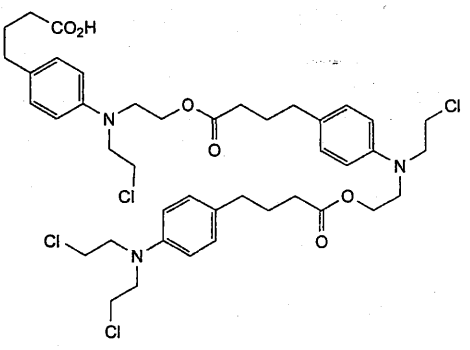
C. 4-[4-[[2-[[bis(2-chloroethoxy)phosphoryl]oxy]ethyl](2-chloroethyl)amino]phenyl]butanoic acid,



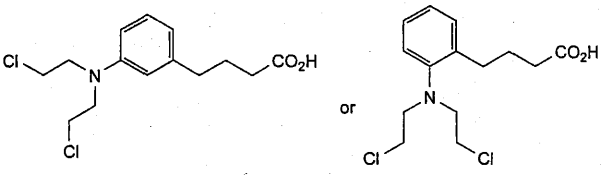
D. 2-chloroethyl 4-[4-[bis(2-chloroethyl)amino] phenyl]butanoate,



E. 4-[4-[[2-[[4-[4-[bis(2-chloroethyl)amino]phenyl]butanoyl] oxy]ethyl](2-chloroethyl)amino]phenyl]butanoic acid,



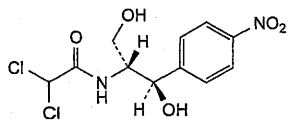
F. 4-[4-[[2-[[4-[4-[[2-[[4-[4-[bis(2-chloroethyl) amino]phenyl]butanoyl]oxy]ethyl](2-chloroethyl) amino]phenyl]butanoyl]oxy]ethyl](2-chloroethyl) amino]phenyl]butanoic acid,



G. 4-[2-[bis(2-chloroethyl)amino]phenyl]butanoic acid or 4-[3-[bis(2-chloroethyl)amino]phenyl]butanoic acid (*meta* or *ortho* chlorambucil).

Chloramphenicol

(Ph. Eur. monograph 0071)



$C_{11}H_{12}Cl_2N_2O_5$ 323.1 *56-75-7*

Action and use
Antibacterial.

Preparations
Chloramphenicol Capsules
Chloramphenicol Ear Drops
Chloramphenicol Eye Drops
Chloramphenicol Eye Ointment

Ph Eur

DEFINITION

2,2-Dichloro-*N*-[(1*R*,2*R*)-1,3-dihydroxy-1-(4-nitrophenyl) propan-2-yl]acetamide.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White, greyish-white or yellowish-white, fine, crystalline powder or fine crystals, needles or elongated plates.

Solubility
Slightly soluble in water, freely soluble in ethanol (96 per cent) and in propylene glycol.

A solution in anhydrous ethanol is dextrorotatory and a solution in ethyl acetate is laevorotatory.

IDENTIFICATION

First identification: A.

Second identification: B.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chloramphenicol CRS.*

B. Melting point (*2.2.14*).

Determination A Determine the melting point of the substance to be examined.

Results A 149 °C to 153 °C.

Determination B Mix equal parts of the substance to be examined and *chloramphenicol CRS* and determine the melting point of the mixture.

Results B The absolute difference between the melting point of the mixture and the value obtained in determination A is not greater than 2 °C.

TESTS

Acidity or alkalinity
To 0.1 g add 20 mL of *carbon dioxide-free water R*, shake and add 0.1 mL of *bromothymol blue solution R1*. Not more than 0.1 mL of *0.02 M hydrochloric acid* or *0.02 M sodium hydroxide* is required to change the colour of the indicator.

Specific optical rotation (*2.2.7*)
+ 18.5 to + 20.5.

Dissolve 1.50 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 2.0 g of *sodium heptanesulfonate R* in 900 mL of *water for chromatography R*. Add 6.8 g of *potassium dihydrogen phosphate R* and 5 mL of *triethylamine R*. Adjust to pH 2.5 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Test solution (a) Dissolve 20.0 mg of the substance to be examined in 10 mL of *methanol R* and dilute to 200.0 mL with mobile phase A.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in 5 mL of *methanol R* and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 20.0 mg of *chloramphenicol CRS* in 10 mL of *methanol R* and dilute to 200.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of test solution (b) to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 12.5 mg of *4-nitrobenzaldehyde R* (impurity B) in 2 mL of *methanol R* and dilute to 50 mL with mobile phase A.

Reference solution (d) Dissolve 5 mg of *chloramphenicol for peak identification CRS* (containing impurity A) in 1 mL of *methanol R*, add 1 mL of reference solution (c) and dilute to 10 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 25 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, solution A (32:68 *V/V*);
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 13	100	0
13 - 25	100 → 60	0 → 40
25 - 33	60	40

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 277 nm.

Injection 10 µL of test solution (b) and reference solutions (b) and (d).

Identification of impurities Use the chromatogram supplied with *chloramphenicol for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A and B.

Relative retention With reference to chloramphenicol (retention time = about 14 min): impurity A = about 0.7; impurity B = about 0.9.

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to impurity B and chloramphenicol.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity A by 0.7;
— for each impurity, use the concentration of chloramphenicol in reference solution (b).

Limits:
— *impurity A*: maximum 0.2 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Chlorides (*2.4.4*)
Maximum 100 ppm.

To 1.00 g add 10 mL of *nitric acid R* and 20 mL of *water R* and shake for 5 min. Filter through a filter paper previously washed by filtering 5 mL portions of *water R* until 5 mL of filtrate no longer becomes opalescent on addition of 0.1 mL of *nitric acid R* and 0.1 mL of *silver nitrate solution R1*. 15 mL of the filtrate complies with the test.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 2.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A.

Injection Test solution (a) and reference solution (a).

Run time 1.5 times the retention time of chloramphenicol.

Calculate the percentage content of $C_{11}H_{12}Cl_2N_2O_5$ taking into account the assigned content of *chloramphenicol CRS*.

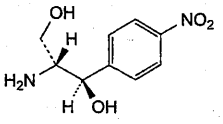
STORAGE

Protected from light. If the substance is sterile, the container is also sterile, airtight and tamper-proof.

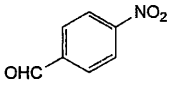
IMPURITIES

Specified impurities *A.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. (1*R*,2*R*)-2-amino-1-(4-nitrophenyl)propane-1,3-diol,

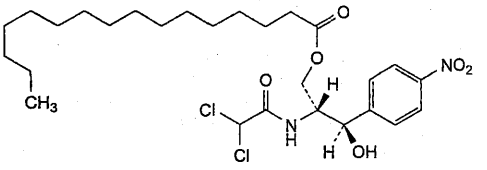


B. 4-nitrobenzaldehyde.

Ph Eur

Chloramphenicol Palmitate

(Ph. Eur. monograph 0473)



$C_{27}H_{42}Cl_2N_2O_6$ 561.6 *530-43-8*

Action and use
Antibacterial.

Ph Eur

DEFINITION

Chloramphenicol palmitate contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of (2*R*,3*R*)-2-[(dichloroacetyl)amino]-3-hydroxy-3-(4-nitrophenyl)propyl hexadecanoate, calculated with reference to the dried substance.

Semi-synthetic product derived from a fermentation product.

CHARACTERS

A white or almost white, fine, unctuous powder, practically insoluble in water, freely soluble in acetone, sparingly soluble in ethanol (96 per cent), very slightly soluble in hexane.

It melts at 87 °C to 95 °C.

It shows polymorphism (*5.9*). The thermodynamically stable form has low bioavailability following oral administration.

IDENTIFICATION

A. Examine by thin-layer chromatography (*2.2.27*), using *TLC silanised silica gel plate R*.

Test solution Dissolve 50 mg of the substance to be examined in a mixture of 1 mL of *1 M sodium hydroxide* and 5 mL of *acetone R* and allow to stand for 30 min. Add 1.1 mL of *1 M hydrochloric acid* and 3 mL of *acetone R*.

Reference solution (a) Dissolve 10 mg of *chloramphenicol CRS* in *acetone R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *palmitic acid R* in *acetone R* and dilute to 5 mL with the same solvent.

Reference solution (c) Dissolve 10 mg of the substance to be examined in *acetone R* and dilute to 5 mL with the same solvent.

Apply to the plate 4 µL of each solution. Develop over a path of 15 cm using a mixture of 30 volumes of a 100 g/L solution of *ammonium acetate R* and 70 volumes of *ethanol (96 per cent) R*. Allow the plate to dry in air and spray with a solution containing 0.2 g/L of *dichlorofluorescein R* and 0.1 g/L of *rhodamine B R* in *ethanol (96 per cent) R*. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. The chromatogram obtained with the test solution shows 3 spots corresponding in position to the principal spots in the chromatograms obtained with reference solutions (a), (b) and (c).

B. Dissolve 0.2 g in 2 mL of *pyridine R*, add 2 mL of a 100 g/L solution of *potassium hydroxide R* and heat on a water-bath. A red colour is produced.

C. Dissolve about 10 mg in 5 mL of *ethanol (96 per cent) R* and add 4.5 mL of *dilute sulfuric acid R* and 50 mg of *zinc powder R*. Allow to stand for 10 min and if necessary decant the supernatant or filter. Cool the solution in iced water and add 0.5 mL of *sodium nitrite solution R*. Allow to stand for

2 min and add 1 g of *urea R*, 2 mL of *strong sodium hydroxide solution R* and 1 mL of *β-naphthol solution R*. A red colour develops.

TESTS

Acidity

Dissolve 1.0 g in 5 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *ether R*, warming to 35 °C. Add 0.2 mL of *phenolphthalein solution R*. Not more than 0.4 mL of *0.1 M sodium hydroxide* is required to produce a pink colour persisting for 30 s.

Specific optical rotation (*2.2.7*)

Dissolve 1.25 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent. The specific optical rotation is + 22.5 to + 25.5.

Free chloramphenicol

Maximum 450 ppm. Dissolve 1.0 g, with gentle heating, in 80 mL of *xylene R*. Cool and shake with 3 quantities, each of 15 mL, of *water R*. Dilute the combined aqueous extracts to 50 mL with *water R* and shake with 10 mL of *toluene R*. Allow to separate and discard the toluene layer. Centrifuge a portion of the aqueous layer and measure the absorbance (*A*) (*2.2.25*) at the maximum at 278 nm using as the compensation liquid a blank solution having an absorbance not greater than 0.05.

Calculate the content of free chloramphenicol in parts per million from the expression:

$$\frac{A \times 10^4}{5.96}$$

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel GF_{254} R* as the coating substance.

Test solution Dissolve 0.1 g of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *chloramphenicol palmitate isomer CRS* in *acetone R* and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with *acetone R*.

Reference solution (b) Dissolve 20 mg of *chloramphenicol dipalmitate CRS* in *acetone R* and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with *acetone R*.

Reference solution (c) Dissolve 5 mg of *chloramphenicol CRS* in *acetone R* and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with *acetone R*.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of *methanol R*, 40 volumes of *chloroform R* and 50 volumes of *cyclohexane R*. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. In the chromatogram obtained with the test solution, any spots due to chloramphenicol palmitate isomer and chloramphenicol dipalmitate are not more intense than the corresponding spots in the chromatograms obtained with reference solutions (a) and (b) respectively (2.0 per cent) and any spot, apart from the principal spot and the spots due to chloramphenicol palmitate isomer and chloramphenicol dipalmitate, is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by heating at 80 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.1 kPa for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

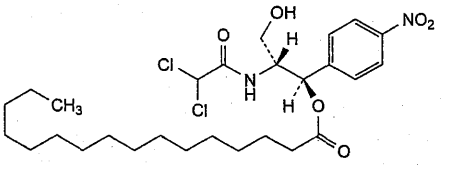
Dissolve 90.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 250.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) of the solution at the maximum at 271 nm.

Calculate the content of $C_{27}H_{42}Cl_2N_2O_6$ taking the specific absorbance to be 178.

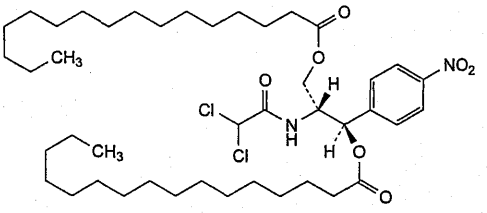
STORAGE

Protected from light.

IMPURITIES



A. (1*R*,2*R*)-2-[(dichloroacetyl)amino]-3-hydroxy-1-(4-nitrophenyl)propyl hexadecanoate (chloramphenicol palmitate isomer),

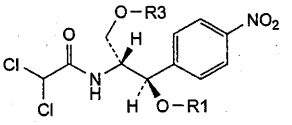


B. (1*R*,2*R*)-2-[(dichloroacetyl)amino]-1-(4-nitrophenyl) propane-1,3-diyl bishexadecanoate (chloramphenicol dipalmitate).

Ph Eur

Chloramphenicol Sodium Succinate

(Ph. Eur. monograph 0709)



1 isomer : R1 = CO-CH_2-CH_2-CO_2Na, R3 = H

3 isomer : R1 = H, R3 = CO-CH_2-CH_2-CO_2Na

$C_{15}H_{15}Cl_2N_2NaO_8$ 445.2

Action and use

Antibacterial.

Preparation

Chloramphenicol Sodium Succinate Injection

Ph Eur

DEFINITION

Mixture in variable proportions of sodium (2*R*,3*R*)-2-[(dichloroacetyl)amino]-3-hydroxy-3-(4-nitrophenyl)propyl butanedioate (3 isomer) and of sodium (1*R*,2*R*)-2-

[(dichloroacetyl)amino]-3-hydroxy-1-(4-nitrophenyl)propyl butanedioate (1 isomer).

Semi-synthetic product derived from a fermentation product.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or yellowish-white powder, hygroscopic.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 2 mL of *acetone R*.

Reference solution (a) Dissolve 20 mg of *chloramphenicol sodium succinate CRS* in 2 mL of *acetone R*.

Reference solution (b) Dissolve 20 mg of *chloramphenicol CRS* in 2 mL of *acetone R*.

Plate *TLC silica gel GF_{254} plate R*.

Mobile phase *dilute acetic acid R, methanol R, chloroform R* (1:14:85 *V/V/V*).

Application 2 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The 2 principal spots in the chromatogram obtained with the test solution are similar in position and size to the 2 principal spots in the chromatogram obtained with reference solution (a); their positions are different from that of the principal spot in the chromatogram obtained with reference solution (b).

B. Dissolve about 10 mg in 1 mL of *ethanol (50 per cent V/V) R*, add 3 mL of a 10 g/L solution of *calcium chloride R* and 50 mg of *zinc powder R* and heat on a water-bath for 10 min. Filter the hot solution and allow to cool. Add 0.1 mL of *benzoyl chloride R* and shake for 1 min. Add 0.5 mL of *ferric chloride solution R1* and 2 mL of *chloroform R* and shake. The upper layer is light violet-red or purple.

C. Dissolve 50 mg in 1 mL of *pyridine R*. Add 0.5 mL of *dilute sodium hydroxide solution R* and 1.5 mL of *water R*. Heat in a water-bath for 3 min. A red colour develops. Add 2 mL of *nitric acid R* and cool under running water. Add 1 mL of *0.1 M silver nitrate*. A white precipitate is formed slowly.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

pH (*2.2.3*)
6.4 to 7.0.

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 5.0 to + 8.0 (anhydrous substance).

Dissolve 0.50 g in *water R* and dilute to 10.0 mL with the same solvent.

Chloramphenicol and chloramphenicol disodium disuccinate
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *chloramphenicol CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase (solution A). Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10.0 mg of *chloramphenicol disodium disuccinate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase (solution B). Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 25 mg of the substance to be examined in the mobile phase, add 5 mL of solution A and 5 mL of solution B and dilute to 100 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 20 g/L solution of *phosphoric acid R*, *methanol R*, *water R* (5:40:55 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 µL.

System suitability Reference solution (c):
— the 2 peaks corresponding to those in the chromatograms obtained with reference solutions (a) and (b) are clearly separated from the peaks corresponding to the 2 principal peaks in the chromatogram obtained with the test solution; if necessary, adjust the methanol content of the mobile phase.

Limits:
— *chloramphenicol*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent);
— *chloramphenicol disodium disuccinate*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent).

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Pyrogens (*2.6.8*)
If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of pyrogens, it complies with the test for pyrogens. Inject per kilogram of the rabbit's mass 2.5 mL of a solution in *water for injections R* containing 2 mg of the substance to be examined per millilitre.

ASSAY

Dissolve 0.200 g in *water R* and dilute to 500.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *water R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 276 nm.

Calculate the content of $C_{15}H_{15}Cl_2N_2NaO_8$, taking the specific absorbance to be 220.

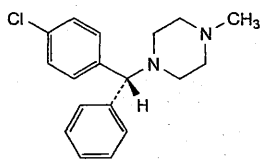
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container, protected from light.

Ph Eur

Chlorcyclizine Hydrochloride

(Ph. Eur. monograph 1086)



and enantiomer , HCl

$C_{18}H_{22}Cl_2N_2$ 337.3 *14362-31-3*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION
Chlorcyclizine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (*RS*)-1-[(4-chlorophenyl)phenylmethyl]-4-methylpiperazine hydrochloride, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, freely soluble in water and in methylene chloride, soluble in alcohol.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Dissolve 10.0 mg in a 5 g/L solution of *sulfuric acid R* and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with a 5 g/L solution of *sulfuric acid R*. Examined between 215 nm and 300 nm (*2.2.25*), the solution shows an absorption maximum at 231 nm. The specific absorbance at the maximum is 475 to 525, calculated with reference to the dried substance.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *chlorcyclizine hydrochloride CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for related substances (see Tests). The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
Dissolve 0.5 g in *water R* and dilute to 10 mL with the same solvent. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. The pH of the solution is 5.0 to 6.0.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using a plate coated with a suitable silica gel.

Test solution (a) Dissolve 0.20 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 5 mL of test solution (a) to 100 mL with *methanol R*.

Reference solution (a) Dissolve 10 mg of *chlorcyclizine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of methylpiperazine R in *methanol R* and dilute to 50 mL with the same solvent.

Reference solution (c) Dilute 1 mL of test solution (b) to 25 mL with *methanol R*.

Reference solution (d) Dissolve 10 mg of *hydroxyzine hydrochloride CRS* and 10 mg of *chlorcyclizine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Apply separately to the plate 10 µL of each solution and develop over a path of 15 cm using a mixture of 2 volumes of *concentrated ammonia R*, 13 volumes of *methanol R* and 85 volumes of *methylene chloride R*. Allow the plate to dry in air and expose it to iodine vapour for 10 min. In the chromatogram obtained with test solution (a): any spot corresponding to methylpiperazine is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent); any spot, apart from the principal spot and any spot corresponding to methylpiperazine, is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.2 per cent). The test is not valid unless the chromatogram obtained with reference solution (d) shows two clearly separated spots.

Loss on drying (*2.2.32*)
Not more than 1.0 per cent, determined on 1.000 g by drying in an oven at 130 °C.

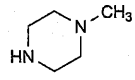
Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in a mixture of 1 mL of *0.1 M hydrochloric acid* and 50 mL of *methanol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the two points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 33.73 mg of $C_{18}H_{22}Cl_2N_2$.

STORAGE
Store protected from light.

IMPURITIES

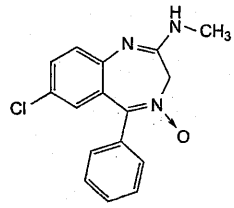


A. *N-methylpiperazine.*

Ph Eur

Chlordiazepoxide

(Ph. Eur. monograph 0656)



$C_{16}H_{14}ClN_3O$ 299.8 *58-25-3*

Action and use
Benzodiazepine.

Ph Eur

DEFINITION

7-Chloro-*N*-methyl-5-phenyl-3*H*-1,4-benzodiazepin-2-amine 4-oxide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
Almost white or light yellow, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chlordiazepoxide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from bright light and prepare the solutions immediately before use.*

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *chlordiazepoxide impurity A CRS* in the mobile phase, add 25.0 mL of the test solution and dilute to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 4.0 mg of *aminochlorobenzophenone R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R, water R* (50:50 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 6 times the retention time of chlordiazepoxide.

Relative retention With reference to chlordiazepoxide (retention time = about 3.6 min): impurity A = about 0.7; impurity B = about 2.3; impurity C = about 3.9.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity A and chlordiazepoxide.

Limits:
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent),
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent),
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g, with heating if necessary, in 80 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

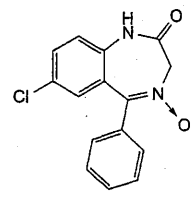
1 mL of *0.1 M perchloric acid* is equivalent to 29.98 mg of $C_{16}H_{14}ClN_3O$.

STORAGE

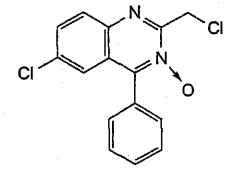
Protected from light.

IMPURITIES

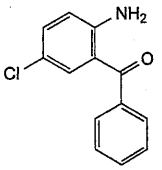
Specified impurities *A, B, C.*



A. 7-chloro-5-phenyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one 4-oxide,



B. 6-chloro-2-(chloromethyl)-4-phenylquinazoline 3-oxide,

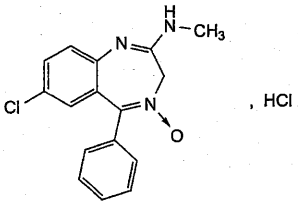


C. (2-amino-5-chlorophenyl)phenylmethanone (aminochlorobenzophenone).

Ph Eur

Chlordiazepoxide Hydrochloride

(Ph. Eur. monograph 0474)



$C_{16}H_{15}Cl_2N_3O$ 336.2 *438-41-5*

Action and use
Benzodiazepine.

Preparations
Chlordiazepoxide Capsules
Chlordiazepoxide Tablets

Ph Eur

DEFINITION
7-Chloro-*N*-methyl-5-phenyl-3*H*-1,4-benzodiazepin-2-amine 4-oxide hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder.

Solubility
Soluble in water, sparingly soluble in ethanol (96 per cent).
It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *chlordiazepoxide hydrochloride CRS.*
If the spectra obtained in the solid state show differences, dissolve 100 mg in 9 mL of *water R* and add 1 mL of *dilute sodium hydroxide solution R*. Extract with 10 mL of *methylene chloride R* in a separating funnel. Evaporate the organic layer and dry the residue obtained at 100-105 °C. Proceed in the same way with the reference substance. Record new spectra using the residues.

B. Dissolve 50 mg in 5 mL of *water R*, add 1 mL of *dilute ammonia R1*, mix, allow to stand for 5 min and filter. Acidify the filtrate with *dilute nitric acid R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).
Dissolve 2.5 g in *water R* and dilute to 25 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the following operations protected from bright light and prepare the solutions immediately before use.*
Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 5 mg of *chlordiazepoxide impurity A CRS* in the mobile phase, add 25.0 mL of the test solution and dilute to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 50.0 mL with the mobile phase.
Reference solution (c) Dissolve 4.0 mg of *aminochlorobenzophenone R* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).
Mobile phase *acetonitrile R, water R* (50:50 *V/V*).
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 10 µL.
Run time 6 times the retention time of chlordiazepoxide.
Relative retention With reference to chlordiazepoxide (retention time = about 3.6 min): impurity A = about 0.7; impurity B = about 2.3; impurity C = about 3.9.
System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity A and chlordiazepoxide.
Limits:
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent),
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent),
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 50 mL of *water R*. Titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

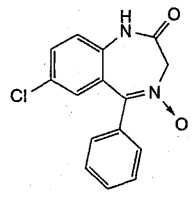
1 mL of *0.1 M silver nitrate* is equivalent to 33.62 mg of $C_{16}H_{15}Cl_2N_3O$.

STORAGE

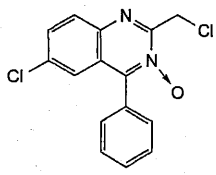
Protected from light.

IMPURITIES

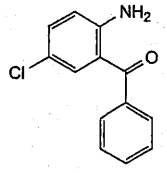
Specified impurities *A, B, C.*



A. 7-chloro-5-phenyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one 4-oxide,



B. 6-chloro-2-(chloromethyl)-4-phenylquinazoline 3-oxide,

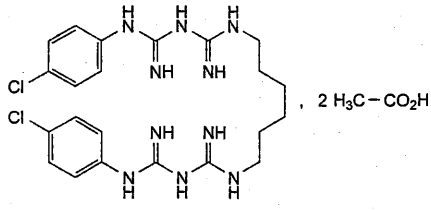


C. (2-amino-5-chlorophenyl)phenylmethanone (aminochlorobenzophenone).

Ph Eur

Chlorhexidine Acetate

(Chlorhexidine Diacetate, Ph. Eur. monograph 0657)



$C_{26}H_{38}Cl_2N_{10}O_4$ 626 *56-95-1*

Action and use

Antiseptic.

Preparation

Chlorhexidine Irrigation Solution

Ph Eur

DEFINITION

$N^1,N^{1\prime}$-(Hexane-1,6-diyl)bis[N^3-(4-chlorophenyl) imidodicarbonimidic diamide] diacetate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, microcrystalline powder.

Solubility

Sparingly soluble in water, soluble in ethanol (96 per cent), slightly soluble in glycerol and in propylene glycol.

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chlorhexidine diacetate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 5 mg of *chlorhexidine diacetate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous formic acid R, water R, ethanol (96 per cent) R, methylene chloride R* (7:10:40:50 *V/V/V/V*).

Application 5 µL; the volume may be adapted based on the type of plate used.

Development Over 2/3 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Treat with *copper sulfate solution R* and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of acetates (*2.3.1*).

TESTS

Impurity P (chloroaniline)

Maximum 500 ppm.

Test solution Dissolve 0.20 g in 25 mL of *water R* with shaking if necessary. Add 1 mL of *hydrochloric acid R* and dilute to 30 mL with *water R*. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol (96 per cent) R*; transfer quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Reference solutions Prepare reference solutions representing respectively 50 ppm, 100 ppm, 200 ppm, 500 ppm and 600 ppm of *chloroaniline R* (impurity P) in the test sample as follows: dilute 1.0 mL, 2.0 mL, 4.0 mL, 10.0 mL and 12.0 mL of a solution containing 0.010 g/L of *chloroaniline R* (impurity P) in *dilute hydrochloric acid R* to 20 mL with *water R*. Then, add 10 mL of *water R*. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol*

(96 per cent) R; transfer each solution quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Measure the absorbance (*2.2.25*) of each reference solution at 556 nm and plot a calibration curve.

Measure the absorbance (*2.2.25*) of the test solution at 556 nm. Determine the concentration of chloroaniline from the calibration curve.

Related substances

Liquid chromatography (*2.2.29*). *Store the solutions at a temperature not exceeding 12 °C.*

Test solution Dissolve 0.140 g of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *chlorhexidine for system suitability CRS* (containing impurities A, B, H, I, K, N and O) in 1.0 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: mix 20 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R* and 80 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water for chromatography R*;
- *mobile phase B*: mix 10 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water for chromatography R* and 90 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 32	100 → 80	0 → 20
32 - 37	80	20
37 - 47	80 → 70	20 → 30
47 - 54	70	30

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *chlorhexidine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, H, I, K, N and O.

Relative retention With reference to chlorhexidine (retention time = about 35 min): impurity N = about 0.35; impurity B = about 0.36; impurity A = about 0.6; impurity H = 0.85; impurity O = about 0.90; impurity I = about 0.91; impurity K = about 1.4.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity N.

Limits:
- *sum of impurities I and O*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *impurity K*: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurities A, H, N*: for each impurity, not more than 0.15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 3.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.15 per cent, determined on 1.0 g.

ASSAY

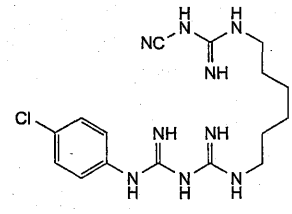
Dissolve 0.140 g in 100 mL of *anhydrous acetic acid R* and titrate with *0.1 M perchloric acid*. Determine the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 15.64 mg of $C_{26}H_{38}Cl_2N_{10}O_4$.

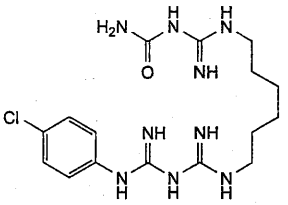
IMPURITIES

Specified impurities *A, H, I, K, N, O, P.*

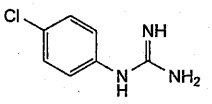
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, E, F, G, M.*



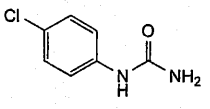
A. N^1-(4-chlorophenyl)-N^3-[6-[(*N*-cyanocarbamimidoyl) amino]hexyl]imidodicarbonimidic diamide,



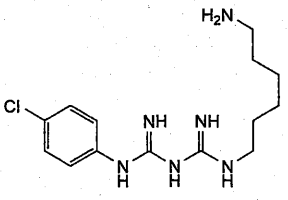
B. *N*-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl) carbamimidoyl]carbamimidoyl]amino]hexyl] carbamimidoyl]urea,



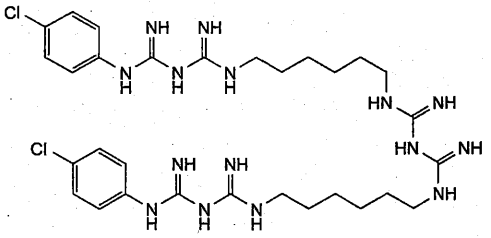
E. *N*-(4-chlorophenyl)guanidine,



F. *N*-(4-chlorophenyl)urea,

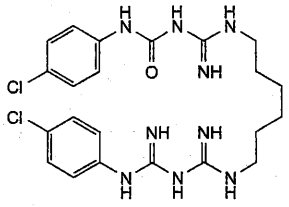


G. N^1-(6-aminohexyl)-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,

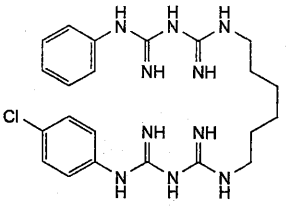


H. N^1,$N^{1\prime}$-[azanediylbis(carbonimidoylazanediylhexane-6,1-diyl)]bis[N^3-(4-chlorophenyl)imidodicarbonimidic diamide],

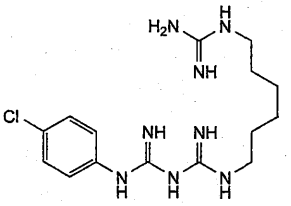
I. unknown structure,



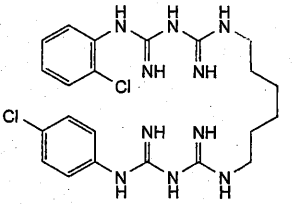
K. *N*-(4-chlorophenyl)-*N*′-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl]carbamimidoyl]amino] hexyl]carbamimidoyl]urea,



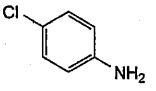
M. N^1-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl] carbamimidoyl]amino]hexyl]-N^3-phenylimidodicarbonimidic diamide,



N. N^1-[6-(carbamimidoylamino)hexyl]-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,



O. N^1-[6-[[*N*-[*N*-(2-chlorophenyl)carbamimidoyl] carbamimidoyl]amino]hexyl]-N^3-(4-chlorophenyl)imidodicarbonimidic diamide,

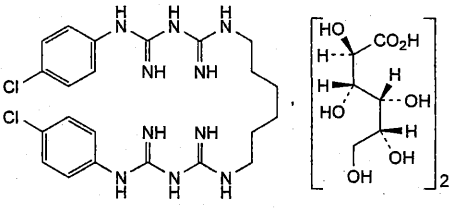


P. 4-chloroaniline.

Ph Eur

Chlorhexidine Gluconate Solution

(Chlorhexidine Digluconate Solution, Ph. Eur. monograph 0658)



$C_{34}H_{54}Cl_2N_{10}O_{14}$ 898 *18472-51-0*

Action and use
Antiseptic.

Preparations
Chlorhexidine Gluconate Eye Drops
Chlorhexidine Gluconate Gel
Chlorhexidine Irrigation Solution
Chlorhexidine Mouthwash
Lidocaine and Chlorhexidine Gel

Ph Eur

DEFINITION

N^1,$N^{1\prime}$-(Hexane-1,6-diyl)bis[N^3-(4-chlorophenyl) imidodicarbonimidic diamide] di-D-gluconate.

Content
190 g/L to 210 g/L.

CHARACTERS

Appearance
Almost colourless or pale-yellowish liquid.

Solubility
Miscible with water, with not more than 3 parts of acetone and with not more than 5 parts of ethanol (96 per cent).

IDENTIFICATION

First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation To 1 mL add 40 mL of *water R*, cool in iced water, make alkaline to *titan yellow paper R* by adding dropwise, and with stirring, *strong sodium hydroxide solution R* and add 1 mL in excess. Filter, wash the precipitate with *water R* until the washings are free from alkali and recrystallise from *ethanol (70 per cent V/V) R*. Dry at 100-105 °C. Examine the residue.

Comparison *chlorhexidine CRS*.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dilute 10.0 mL of the preparation to be examined to 50 mL with *water R*.

Reference solution Dissolve 25 mg of *calcium gluconate CRS* in 1 mL of *water R*.

Plate *TLC silica gel plate R*.

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).

Application 5 µL.

Development Over 1/2 of the plate.

Drying At 100 °C for 20 min and allow to cool.

Detection Spray with a solution containing 25 g/L of *ammonium molybdate R* and 10 g/L of *cerium sulfate R* in *dilute sulfuric acid R*, and heat at 110 °C for about 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 1 mL add 40 mL of *water R*, cool in iced water, make alkaline to *titan yellow paper R* by adding dropwise, and with stirring, *strong sodium hydroxide solution R* and add 1 mL in excess. Filter, wash the precipitate with *water R* until the washings are free from alkali and recrystallise from *ethanol (70 per cent V/V) R*. Dry at 100-105 °C. The residue melts (*2.2.14*) at 132 °C to 136 °C.

D. To 0.05 mL add 5 mL of a 10 g/L solution of *cetrimide R*, 1 mL of *strong sodium hydroxide solution R* and 1 mL of *bromine water R*; a deep red colour is produced.

TESTS

Relative density (*2.2.5*)
1.06 to 1.07.

pH (*2.2.3*)
5.5 to 7.0.

Dilute 5.0 mL to 100 mL with *carbon dioxide-free water R*.

Impurity P (chloroaniline)
Maximum 500 ppm, calculated with reference to chlorhexidine digluconate solution.

Test solution Dilute 0.20 g of the preparation to be examined to 30 mL with *water R*. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol (96 per cent) R*; transfer quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Reference solutions Prepare reference solutions representing respectively 50 ppm, 100 ppm, 200 ppm, 500 ppm and 600 ppm of *chloroaniline R* (impurity P) in the test sample as follows: dilute 1.0 mL, 2.0 mL, 4.0 mL, 10.0 mL and 12.0 mL of a solution containing 0.010 g/L of *chloroaniline R* (impurity P) in *dilute hydrochloric acid R* to 20 mL with *water R*. Then, add 10 mL of *water R*. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol (96 per cent) R*; transfer each solution quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Measure the absorbance (*2.2.25*) of each reference solution at 556 nm and plot a calibration curve.

Measure the absorbance (*2.2.25*) of the test solution at 556 nm. Determine the concentration of chloroaniline from the calibration curve.

Related substances
Liquid chromatography (*2.2.29*). *Store the solutions at a temperature not exceeding 12 °C.*

Test solution Dilute 1.0 mL of the preparation to be examined to 100.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *chlorhexidine for system suitability CRS* (containing impurities A, B, F, G, H, I, J, K, L, N and O) in 1.0 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: mix 20 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R* and 80 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water R*;
- *mobile phase B*: mix 10 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water R* and 90 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 32	100 → 80	0 → 20
32 - 37	80	20
37 - 47	80 → 70	20 → 30
47 - 54	70	30

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *chlorhexidine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, F, G, H, I, J, K, L, N and O.

Relative retention With reference to chlorhexidine (retention time = about 35 min): impurity L = about 0.23; impurity Q = about 0.24; impurity G = about 0.25; impurity N = about 0.35; impurity B = about 0.36; impurity F = about 0.5; impurity A = about 0.6; impurity H = about 0.85; impurity O = about 0.90; impurity I = about 0.91; impurity J = about 0.96; impurity K = about 1.4.

System suitability Reference solution (a):
- *resolution*: minimum 3.0 between the peaks due to impurities L and G;
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity N.

Limits:
- *impurity N*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *impurity H*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurities A, J, K*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *sum of impurities I and O*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *impurity G*: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurities B, F, L, Q*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

ASSAY

Determine the density (*2.2.5*) of the preparation to be examined. Transfer 1.00 g to a 250 mL beaker and add 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 22.44 mg of $C_{34}H_{54}Cl_2N_{10}O_{14}$.

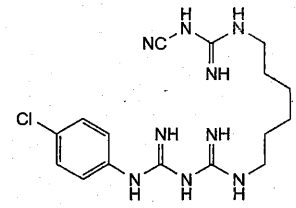
STORAGE

Protected from light.

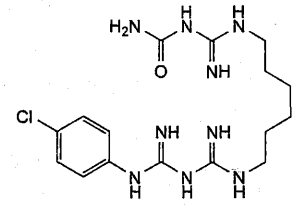
IMPURITIES

Specified impurities *A, B, F, G, H, I, J, K, L, N, O, P, Q.*

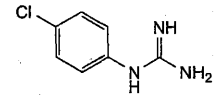
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, M.*



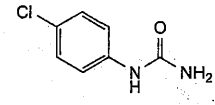
A. N^1-(4-chlorophenyl)-N^3-[6-[(*N*-cyanocarbamimidoyl)amino]hexyl]imidodicarbonimidic diamide,



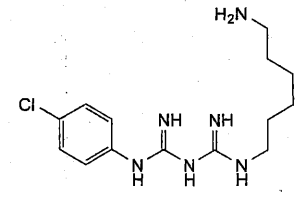
B. *N*-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl]carbamimidoyl]amino]hexyl]carbamimidoyl]urea,



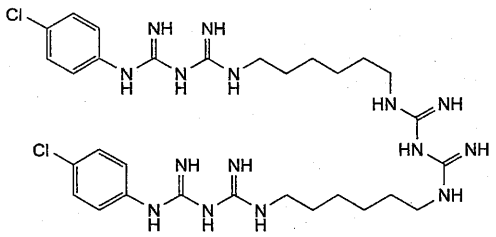
E. *N*-(4-chlorophenyl)guanidine,



F. *N*-(4-chlorophenyl)urea,

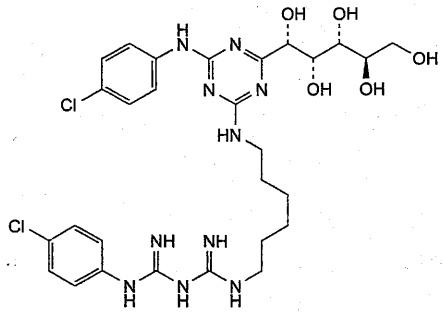


G. N^1-(6-aminohexyl)-N^3-(4-chlorophenyl)imidodicarbonimidic diamide,

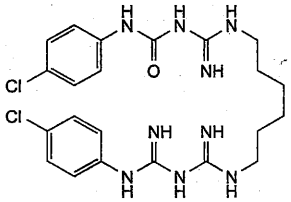


H. $N^1,N^{1\prime}$-[azanediylbis(carbonimidoylazanediylhexane-6,1-diyl)]bis[N^3-(4-chlorophenyl)imidodicarbonimidic diamide],

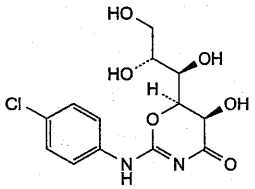
I. unknown structure,



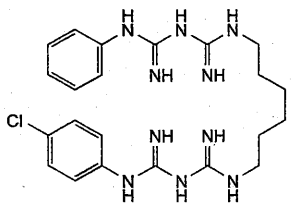
J. 1-(4-chlorophenyl)-5-[6-[[4-[(4-chlorophenyl)amino]-6-[(1*S*,2*R*,3*R*,4*R*)-1,2,3,4,5-pentahydroxypentyl]-1,3,5-triazin-2-yl]amino]hexyl]biguanide,



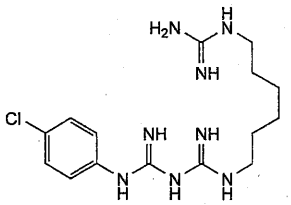
K. *N*-(4-chlorophenyl)-*N*′-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl) carbamimidoyl]carbamimidoyl]amino]hexyl] carbamimidoyl]urea,



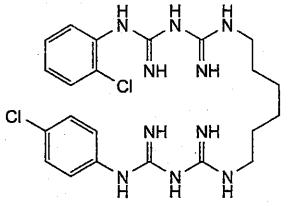
L. (5*R*,6*S*)-2-[(4-chlorophenyl)amino]-5-hydroxy-6-[(1*R*,2*R*)-1,2,3-trihydroxypropyl]-5,6-dihydro-4*H*-1,3-oxazin-4-one,



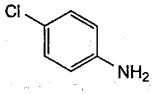
M. N^1-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl] carbamimidoyl]amino]hexyl]-N^3-phenylimidodicarbonimidic diamide,



N. N^1-[6-(carbamimidoylamino)hexyl]-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,



O. N^1-[6-[[*N*-[*N*-(2-chlorophenyl)carbamimidoyl] carbamimidoyl]amino]hexyl]-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,



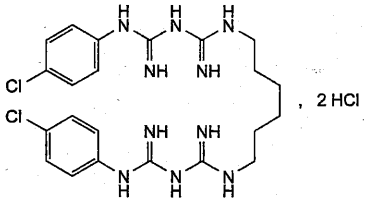
P. 4-chloroaniline,

Q. unknown structure.

Ph Eur

Chlorhexidine Hydrochloride

(Chlorhexidine Dihydrochloride, Ph. Eur. monograph 0659)



$C_{22}H_{32}Cl_4N_{10}$ 578.4 *3697-42-5*

Action and use
Antiseptic.

Ph Eur

DEFINITION
$N^1,N^{1\prime}$-(Hexane-1,6-diyl)bis[N^3-(4-chlorophenyl) imidodicarbonimidic diamide] dihydrochloride.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, slightly soluble in propylene glycol, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, C.
Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chlorhexidine dihydrochloride CRS*.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 5 mg of *chlorhexidine dihydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *anhydrous formic acid R, water R, ethanol (96 per cent) R, methylene chloride R* (7:10:40:50 *V/V/V/V*).

Application 5 µL; the volume may be adapted based on the type of plate used.

Development Over 2/3 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Treat with *copper sulfate solution R* and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Impurity P (chloroaniline)

Maximum 300 ppm.

Test solution To 0.20 g add 1 mL of *hydrochloric acid R*, shake for about 30 s, dilute to 30 mL with *water R* and shake until a clear solution is obtained. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol (96 per cent) R*; transfer quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Reference solutions Prepare reference solutions representing respectively 50 ppm, 100 ppm, 200 ppm, 300 ppm and 500 ppm of *chloroaniline R* (impurity P) in the test sample as follows: dilute 1.0 mL, 2.0 mL, 4.0 mL, 6.0 mL and 10.0 mL of a solution containing 0.010 g/L of *chloroaniline R* (impurity P) in *dilute hydrochloric acid R* to 20 mL with *water R*. Then, add 10 mL of *water R*. Add rapidly and with thorough mixing after each addition: 5 mL of a 103 g/L solution of *hydrochloric acid R*, 0.35 mL of *sodium nitrite solution R*, 2 mL of a 50 g/L solution of *ammonium sulfamate R*, 5 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and 1 mL of *ethanol (96 per cent) R*; transfer each solution quantitatively to a volumetric flask, dilute to 50.0 mL with *water R* and allow to stand for 30 min.

Measure the absorbance (*2.2.25*) of each reference solution at 556 nm and plot a calibration curve.

Measure the absorbance (*2.2.25*) of the test solution at 556 nm. Determine the concentration of chloroaniline from the calibration curve.

Related substances

Liquid chromatography (*2.2.29*). *Store the solutions at a temperature not exceeding 12 °C.*

Test solution Dissolve 0.130 g of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *chlorhexidine for system suitability CRS* (containing impurities A, B, H, I, K, N and O) in 1.0 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: mix 20 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R* and 80 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water for chromatography R*;
- *mobile phase B*: mix 10 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *water for chromatography R* and 90 volumes of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 32	100 → 80	0 → 20
32 - 37	80	20
37 - 47	80 → 70	20 → 30
47 - 54	70	30

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *chlorhexidine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, H, I, K, N and O.

Relative retention With reference to chlorhexidine (retention time = about 35 min): impurity N = about 0.35; impurity B = about 0.36; impurity A = about 0.6; impurity H = about 0.85; impurity O = about 0.90; impurity I = about 0.91; impurity K = about 1.4.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity N.

Limits:
- *impurity H*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurity K*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *sum of impurities I and O*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *impurities A, N*: for each impurity, not more than 0.15 times the area of the principal peak in the

chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

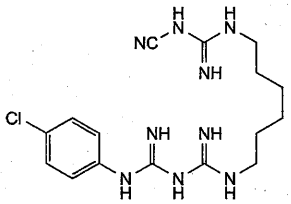
Dissolve 100.0 mg in 5 mL of *anhydrous formic acid R* and add 70 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 14.46 mg of $C_{22}H_{32}Cl_4N_{10}$.

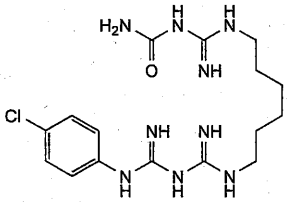
IMPURITIES

Specified impurities *A, H, I, K, N, O, P.*

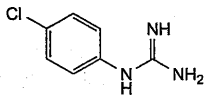
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, E, F, G, M.*



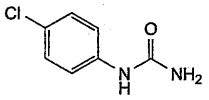
A. N^1-(4-chlorophenyl)-N^3-[6-[(*N*-cyanocarbamimidoyl)amino]hexyl]imidodicarbonimidic diamide,



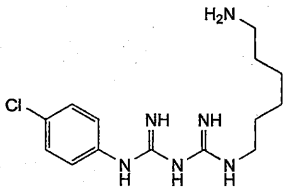
B. *N*-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl]carbamimidoyl]amino]hexyl]carbamimidoyl]urea,



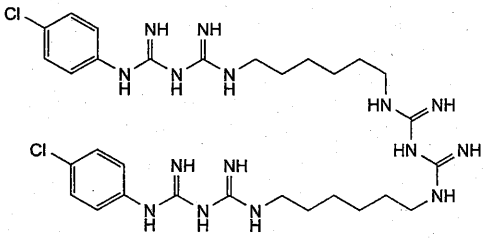
E. *N*-(4-chlorophenyl)guanidine,



F. *N*-(4-chlorophenyl)urea,

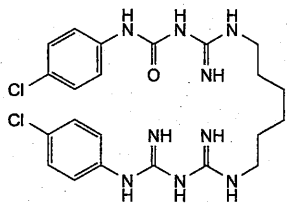


G. N^1-(6-aminohexyl)-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,

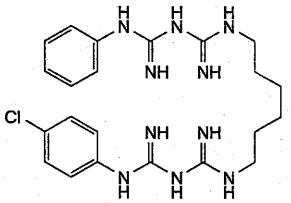


H. N^1,$N^{1\prime}$-[azanediylbis(carbonimidoylazanediylhexane-6,1-diyl)]bis[N^3-(4-chlorophenyl)imidodicarbonimidic diamide],

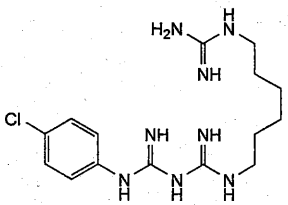
I. unknown structure,



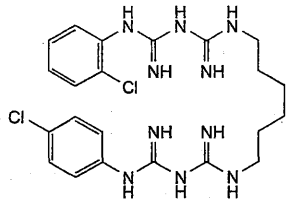
K. *N*-(4-chlorophenyl)-*N′*-[*N*-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl]carbamimidoyl]amino]hexyl]carbamimidoyl]urea,



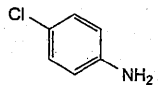
M. N^1-[6-[[*N*-[*N*-(4-chlorophenyl)carbamimidoyl]carbamimidoyl]amino]hexyl]-N^3-phenylimidodicarbonimidic diamide,



N. N^1-[6-(carbamimidoylamino)hexyl]-N^3-(4-chlorophenyl) imidodicarbonimidic diamide,



O. N^1-[6-[[*N*-[*N*-(2-chlorophenyl) carbamimidoyl]carbamimidoyl]amino]hexyl]-N^3-(4-chlorophenyl)imidodicarbonimidic diamide,



P. 4-chloroaniline.

Ph Eur

Chlorinated Lime

Action and use
Disinfectant.

DEFINITION
Chlorinated Lime contains not less than 30.0% w/w of available chlorine, Cl.

CHARACTERISTICS
A dull white powder.

Partly soluble in *water* and in *ethanol (96%)*.

IDENTIFICATION
A. Evolves chlorine copiously on the addition of 2M *hydrochloric acid.*

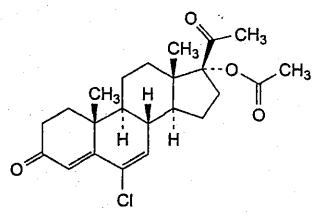
B. When shaken with *water* and filtered, the filtrate yields reaction C characteristic of *calcium salts* and reaction A characteristic of *chlorides*, Appendix VI.

ASSAY
Triturate 4 g with successive small quantities of *water*, dilute to 1000 mL with *water* and shake thoroughly. Mix 100 mL of the resulting suspension with a solution containing 3 g of *potassium iodide* in 100 mL of *water*, acidify with 5 mL of 6M *acetic acid* and titrate the liberated iodine with 0.1M *sodium thiosulfate VS*. Each mL of 0.1M *sodium thiosulfate VS* is equivalent to 3.545 mg of available chlorine, Cl.

STORAGE
On exposure to air Chlorinated Lime becomes moist and gradually decomposes, carbon dioxide being absorbed and chlorine evolved.

Chlormadinone Acetate

(Ph. Eur. monograph 2702)



$C_{23}H_{29}ClO_4$ 404.9 *302-22-7*

Action and use
Progestogen.

Ph Eur

DEFINITION
6-Chloro-3,20-dioxopregna-4,6-dien-17-yl acetate.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetonitrile, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison chlormadinone acetate CRS.

TESTS
Specific optical rotation (*2.2.7*)
−14.0 to −10.0 (dried substance).

Dissolve 0.200 g in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 20 mg of the substance to be examined in mobile phase B and dilute to 10.0 mL with mobile phase B.

Test solution (b) Dissolve 10.0 mg of the substance to be examined in mobile phase B and dilute to 20.0 mL with mobile phase B.

Reference solution (a) Dissolve 4 mg of *chlormadinone acetate for system suitability CRS* (containing impurities A, B, E and K) in mobile phase B and dilute to 2.0 mL with mobile phase B.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase B.

Reference solution (c) Dissolve 5.0 mg of *chlormadinone acetate impurity G CRS* in mobile phase B and dilute to 50.0 mL with mobile phase B. Dilute 1.0 mL of the solution to 50.0 mL with mobile phase B.

Reference solution (d) Dissolve 10.0 mg of *chlormadinone acetate CRS* in mobile phase B and dilute to 20.0 mL with mobile phase B.

Column:
— *size: l* = 0.15 m, Ø = 3.0 mm;
— *stationary phase: end-capped extra-dense bonded octadecylsilyl silica gel for chromatography R* (3.5 µm).

Mobile phase:
— *mobile phase A*: *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	60	40
8 - 30	60 → 5	40 → 95
30 - 33	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 236 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with *chlormadinone acetate for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, E and K; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.

Relative retention With reference to chlormadinone acetate (retention time = about 15 min): impurity K = about 0.75; impurity G = about 0.80; impurity A = about 0.95; impurity E = about 1.04; impurity B = about 1.2.

System suitability:
— *signal-to-noise ratio*: minimum 35 for the principal peak in the chromatogram obtained with reference solution (b);
— *peak-to-valley ratio*: minimum 1.6, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to chlormadinone acetate in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity K by 1.7;
— for impurities E, B and K, use the concentration of chlormadinone acetate in reference solution (b);
— for impurities other than E, B and K, use the concentration of impurity G in reference solution (c).

Limits:
— *impurity B*: maximum 0.2 per cent;
— *impurities A, E, G, K*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase B, mobile phase A (45:55 *V/V*).

Injection Test solution (b) and reference solution (d).

Run time Twice the retention time of chlormadinone acetate.

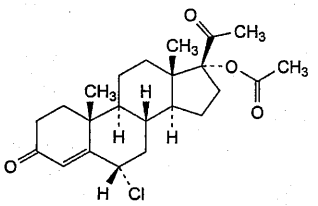
Retention time Chlormadinone acetate = about 12 min.

Calculate the percentage content of $C_{23}H_{29}ClO_4$ taking into account the assigned content of *chlormadinone acetate CRS*.

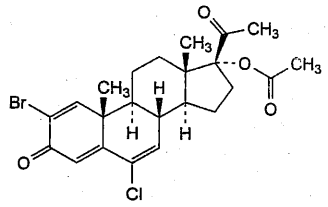
IMPURITIES

Specified impurities *A, B, E, G, K.*

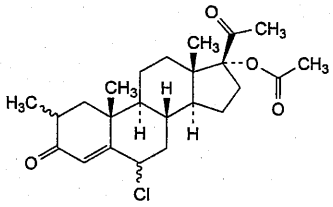
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, F, H, I, J, L.*



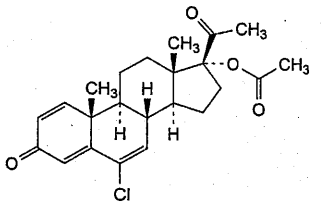
A. 6α-chloro-3,20-dioxopregn-4-en-17-yl acetate,



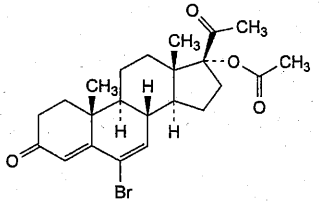
B. 2-bromo-6-chloro-3,20-dioxopregna-1,4,6-trien-17-yl acetate,



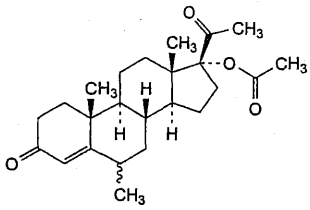
C. 6ξ-chloro-2ξ-methyl-3,20-dioxopregn-4-en-17-yl acetate,



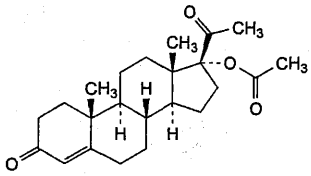
D. 6-chloro-3,20-dioxopregna-1,4,6-trien-17-yl acetate,



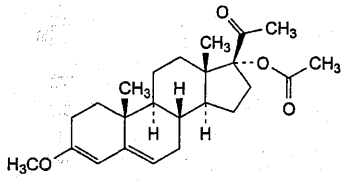
E. 6-bromo-3,20-dioxopregna-4,6-dien-17-yl acetate,



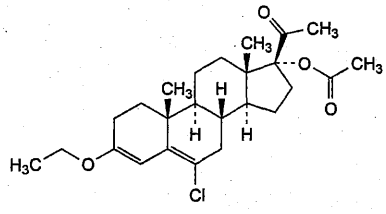
F. 6ξ-methyl-3,20-dioxopregn-4-en-17-yl acetate,



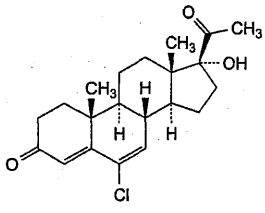
G. 3,20-dioxopregn-4-en-17-yl acetate,



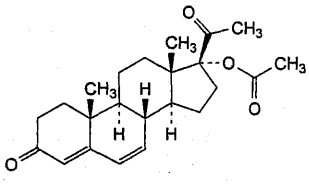
H. 3-methoxy-20-oxopregna-3,5-dien-17-yl acetate,



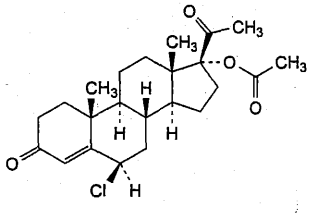
I. 6-chloro-3-ethoxy-20-oxopregna-3,5-dien-17-yl acetate,



J. 6-chloro-17-hydroxypregna-4,6-diene-3,20-dione,



K. 3,20-dioxopregna-4,6-dien-17-yl acetate,



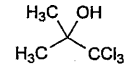
L. 6β-chloro-3,20-dioxopregn-4-en-17-yl acetate.

Ph Eur

Chlorobutanol

Anhydrous Chlorobutanol

(Ph. Eur. monograph 0382)



$C_4H_7Cl_3O$ 177.5 *57-15-8*

Action and use
Disinfectant preservative.

Ph Eur

DEFINITION
1,1,1-Trichloro-2-methylpropan-2-ol.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals, sublimes readily.

Solubility
Slightly soluble in water, very soluble in ethanol (96 per cent), soluble in glycerol (85 per cent).

mp
About 95 °C (without previous drying).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *chlorobutanol CRS.*

B. Water (see Tests).

TESTS
Solution S
Dissolve 5 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Acidity
To 4 mL of solution S add 15 mL of *ethanol (96 per cent) R* and 0.1 mL of *bromothymol blue solution R1*. Not more than 1.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Impurities A and B
Head-space gas chromatography (*2.2.28*).
Solvent mixture *water R, dimethylformamide R* (40:60 *V/V*).
Test solution Introduce 2.000 g of the substance to be examined into a vial, add 5.0 mL of the solvent mixture and close the vial immediately.
Reference solution (a) Dissolve 60.0 mg of *chloroform R* (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well. Dilute 4.0 mL of the solution to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.
Reference solution (b) Dissolve 0.500 g of *acetone R* (impurity B) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well.
Reference solution (c) Dilute 8.0 mL of reference solution (b) to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.

Reference solution (d) Dissolve 50.0 mg of *2-propanol R* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well. Mix 8.0 mL of the solution and 8.0 mL of reference solution (b), dilute to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 3 µm).

Carrier gas *nitrogen R.*

Flow rate 1.7 mL/min.

Split ratio 1:3.

Static head-space conditions that may be used:
— *equilibration temperature*: 90 °C;
— *equilibration time*: 20 min;
— *pressurisation time*: 30 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 25	40
	25 - 37	40 → 220
	37 - 42	220
Injection port		220
Detector		230

Detection Flame ionisation.

Injection 1.0 mL of the test solution and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (d) to identify the peak due to 2-propanol.

Relative retention With reference to impurity B (retention time = about 14 min): 2-propanol = about 1.1; impurity A = about 2.0.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to impurity B and 2-propanol.

Calculation of contents:
— for impurity A, use the concentration of impurity A in reference solution (a);
— for impurity B, use the concentration of impurity B in reference solution (c).

Limits:
— *impurity B*: maximum 0.10 per cent;
— *impurity A*: maximum 60 ppm.

Chlorides *(2.4.4)*
Maximum 300 ppm.

Dissolve 0.17 g in 5 mL of *ethanol (96 per cent) R* and dilute to 15 mL with *water R*. When preparing the standard, replace the 5 mL of *water R* by 5 mL of *ethanol (96 per cent) R*.

Water *(2.5.12)*
Maximum 1.0 per cent, determined on 2.00 g.

Sulfated ash *(2.4.14)*
Maximum 0.1 per cent, determined on 1.0 g.

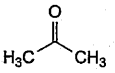
ASSAY
Dissolve 50.0 mg in 20 mL of *ethanol (96 per cent) R* in a 50 mL centrifuge tube. Add 10 mL of *dilute sodium hydroxide solution R*, cap tightly and heat in a water-bath for 10 min. Cool and transfer quantitatively to a titration vessel using 100 mL of *water R*. Add 20 mL of *dilute nitric acid R* and titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically *(2.2.20)*.

1 mL of *0.1 M silver nitrate* is equivalent to 5.92 mg of $C_4H_7Cl_3O$.

STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B.*

A. trichloromethane (chloroform),



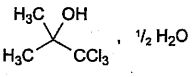
B. propan-2-one (acetone).

Ph Eur

Chlorobutanol Hemihydrate

(Ph. Eur. monograph 0383)

NOTE: The name Chlorobutanol was formerly used in the United Kingdom



$C_4H_7Cl_3O,½H_2O$ 186.5 *6001-64-5*

Action and use
Disinfectant preservative.

Ph Eur

DEFINITION
1,1,1-Trichloro-2-methylpropan-2-ol hemihydrate.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, sublimes readily.

Solubility
Slightly soluble in water, very soluble in ethanol (96 per cent), soluble in glycerol (85 per cent).

mp
About 78 °C (without previous drying).

IDENTIFICATION
A. Infrared absorption spectrophotometry *(2.2.24)*.
Comparison *chlorobutanol hemihydrate CRS.*
B. Water (see Tests).

TESTS

Solution S

Dissolve 5 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Acidity

To 4 mL of solution S add 15 mL of *ethanol (96 per cent) R* and 0.1 mL of *bromothymol blue solution R1*. Not more than 1.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Impurities A and B

Head-space gas chromatography (*2.2.28*).

Solvent mixture *water R, dimethylformamide R* (40:60 *V/V*).

Test solution Introduce 2.000 g of the substance to be examined into a vial, add 5.0 mL of the solvent mixture and close the vial immediately.

Reference solution (a) Dissolve 60.0 mg of *chloroform R* (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well. Dilute 4.0 mL of the solution to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.

Reference solution (b) Dissolve 0.500 g of *acetone R* (impurity B) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well.

Reference solution (c) Dilute 8.0 mL of reference solution (b) to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.

Reference solution (d) Dissolve 50.0 mg of *2-propanol R* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Shake well. Mix 8.0 mL of the solution and 8.0 mL of reference solution (b), dilute to 200.0 mL with the solvent mixture and transfer 5.0 mL to an injection vial.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.53 mm;
- *stationary phase*: *poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 3 µm).

Carrier gas *nitrogen R*.

Flow rate 1.7 mL/min.

Split ratio 1:3.

Static head-space conditions that may be used:
- *equilibration temperature*: 90 °C;
- *equilibration time*: 20 min;
- *pressurisation time*: 30 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 25	40
	25 - 37	40 → 220
	37 - 42	220
Injection port		220
Detector		230

Detection Flame ionisation.

Injection 1.0 mL of the test solution and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (d) to identify the peak due to 2-propanol.

Relative retention With reference to impurity B (retention time = about 14 min): 2-propanol = about 1.1; impurity A = about 2.0.

System suitability Reference solution (d):
- *resolution*: minimum 1.5 between the peaks due to impurity B and 2-propanol.

Calculation of contents:
- for impurity A, use the concentration of impurity A in reference solution (a);
- for impurity B, use the concentration of impurity B in reference solution (c).

Limits:
- *impurity B*: maximum 0.10 per cent;
- *impurity A*: maximum 60 ppm.

Chlorides (*2.4.4*)

Maximum 100 ppm.

To 1 mL of solution S add 4 mL of *ethanol (96 per cent) R* and dilute to 15 mL with *water R*. When preparing the standard, replace the 5 mL of *water R* by 5 mL of *ethanol (96 per cent) R*.

Water (*2.5.12*)

4.5 per cent to 5.5 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 50.0 mg in 20 mL of *ethanol (96 per cent) R* in a 50 mL centrifuge tube. Add 10 mL of *dilute sodium hydroxide solution R*, cap tightly and heat in a water-bath for 10 min. Cool and transfer quantitatively to a titration vessel using 100 mL of *water R*. Add 20 mL of *dilute nitric acid R* and titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 5.92 mg of $C_4H_7Cl_3O$.

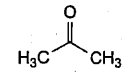
STORAGE

In an airtight container.

IMPURITIES

Specified impurities *A, B.*

A. trichloromethane (chloroform),

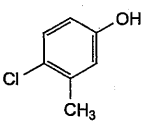


B. propan-2-one (acetone).

Ph Eur

Chlorocresol

(Ph. Eur. monograph 0384)



C_7H_7ClO 142.6 *59-50-7*

Action and use
Antiseptic; antimicrobial preservative.

Ph Eur

DEFINITION
4-Chloro-3-methylphenol.

Content
98.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
White or almost white, crystalline powder or compacted crystalline masses supplied as pellets or colourless or white crystals.

Solubility
Slightly soluble in water, very soluble in ethanol (96 per cent), freely soluble in fatty oils. It dissolves in solutions of alkali hydroxides.

IDENTIFICATION
A. Melting point (*2.2.14*): 64 °C to 67 °C.

B. To 0.1 g add 0.2 mL of *benzoyl chloride R* and 0.5 mL of *dilute sodium hydroxide solution R*. Shake vigorously until a white, crystalline precipitate is formed. Add 5 mL of *water R* and filter. The precipitate, recrystallised from 5 mL of *methanol R* and dried at 70 °C, melts (*2.2.14*) at 85 °C to 88 °C.

C. To 5 mL of solution S (see Tests) add 0.1 mL of *ferric chloride solution R1*. A bluish colour is produced.

TESTS

Solution S
To 3.0 g, finely powdered, add 60 mL of *carbon dioxide-free water R*, shake for 2 min and filter.

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 1.25 g in *ethanol (96 per cent) R* and dilute to 25 mL with the same solvent.

Acidity
To 10 mL of solution S add 0.1 mL of *methyl red solution R*. The solution is orange or red. Not more than 0.2 mL of *0.01 M sodium hydroxide* is required to produce a pure yellow colour.

Related substances
Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 1.0 g of the substance to be examined in *acetone R* and dilute to 100 mL with the same solvent.

Reference solution Dilute 1.0 mL of the test solution to 100.0 mL with *acetone R*. Dilute 5.0 mL of this solution to 100.0 mL with *acetone R*.

Column:
— *material*: glass;
— *size*: *l* = 1.80 m, Ø = 3-4 mm;
— *stationary phase*: *silanised diatomaceous earth for gas chromatography R* impregnated with 3-5 per cent *m/m* of *polymethylphenylsiloxane R*.

Carrier gas *nitrogen for chromatography R*.

Flow rate 30 mL/min.

Temperature:
— *column*: 125 °C;
— *injection port*: 210 °C;
— *detector*: 230 °C.

Detection Flame ionisation.

Run time 3 times the retention time of chlorocresol.

Retention time Chlorocresol = about 8 min.

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1 per cent;
— *disregard limit*: the area of the principal peak in the chromatogram obtained with the reference solution (0.05 per cent).

Non-volatile matter
Maximum 0.1 per cent.

Evaporate 2.0 g to dryness on a water-bath and dry the residue at 100-105 °C. The residue weighs not more than 2 mg.

ASSAY
In a ground-glass-stoppered flask, dissolve 70.0 mg in 30 mL of *glacial acetic acid R*. Add 25.0 mL of *0.0167 M potassium bromate*, 20 mL of a 150 g/L solution of *potassium bromide R* and 10 mL of *hydrochloric acid R*. Allow to stand protected from light for 15 min. Add 1 g of *potassium iodide R* and 100 mL of *water R*. Titrate with *0.1 M sodium thiosulfate*, shaking vigorously and using 1 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank titration.

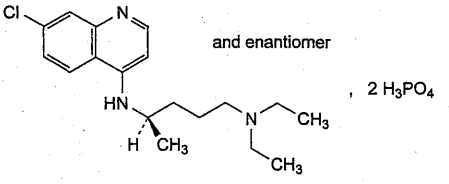
1 mL of *0.0167 M potassium bromate* is equivalent to 3.565 mg of C_7H_7ClO.

STORAGE
Protected from light.

Ph Eur

Chloroquine Phosphate

(Ph. Eur. monograph 0544)



$C_{18}H_{32}ClN_3O_8P_2$ 515.9 *50-63-5*

Action and use
Antiprotozoal (malaria).

Preparation
Chloroquine Phosphate Tablets

Ph Eur

DEFINITION

Chloroquine phosphate contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of N^4-(7-chloroquinolin-4-yl)-N^1,N^1-diethylpentane-1,4-diamine bis (dihydrogen phosphate), calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, hygroscopic, freely soluble in water, very slightly soluble in alcohol and in methanol.

It exists in 2 forms, one of which melts at about 195 °C and the other at about 218 °C.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Dissolve 0.100 g in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *water R*. Examined between 210 nm and 370 nm (*2.2.25*), the solution shows absorption maxima at 220 nm, 235 nm, 256 nm, 329 nm and 342 nm. The specific absorbances at the maxima are respectively 600 to 660, 350 to 390, 300 to 330, 325 to 355 and 360 to 390.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with the base isolated from *chloroquine sulfate CRS*. Record the spectra using solutions prepared as follows: dissolve separately 0.1 g of the substance to be examined and 80 mg of the reference substance in 10 mL of *water R*, add 2 mL of *dilute sodium hydroxide solution R* and shake with 2 quantities, each of 20 mL, of *methylene chloride R*; combine the organic layers, wash with *water R*, dry over *anhydrous sodium sulfate R*, evaporate to dryness and dissolve the residues separately, each in 2 mL of *methylene chloride R*.

C. Dissolve 25 mg in 20 mL of *water R* and add 8 mL of *picric acid solution R1*. The precipitate, washed with *water R*, with *alcohol R* and finally with *methylene chloride R*, melts (*2.2.14*) at 206-209 °C.

D. Dissolve 0.1 g in 10 mL of *water R*, add 2 mL of *dilute sodium hydroxide solution R* and shake with 2 quantities, each of 20 mL, of *methylene chloride R*. The aqueous layer, acidified by the addition of *nitric acid R*, gives reaction (b) of phosphates (*2.3.1*).

TESTS

Solution S

Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 or GY_5 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of solution S is 3.8 to 4.3.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution Dissolve 0.50 g of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 100 mL with *water R*.

Reference solution (b) Dilute 5 mL of reference solution (a) to 10 mL with *water R*.

Apply to the plate 2 µL of each solution. Develop over a path of 12 cm using a mixture of 10 volumes of *diethylamine R*, 40 volumes of *cyclohexane R* and 50 volumes of *chloroform R*. Allow the plate to dry in air. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (1.0 per cent) and not more than one such spot is more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Loss on drying (*2.2.32*)

Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.200 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 25.79 mg of $C_{18}H_{32}ClN_3O_8P_2$.

STORAGE

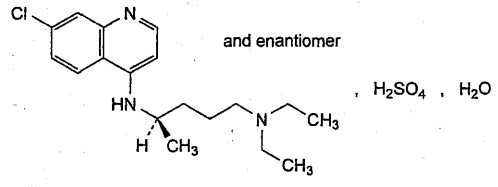
In an airtight container, protected from light.

Ph Eur

Chloroquine Sulfate

Chloroquine Sulphate

(Ph. Eur. monograph 0545)



$C_{18}H_{28}ClN_3O_4S,H_2O$ 436.0

Action and use

Antiprotozoal (malaria).

Preparation

Chloroquine Sulfate Tablets

Ph Eur

DEFINITION

Chloroquine sulfate contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of N^4-(7-chloroquinolin-4-yl)-N^1,N^1-diethylpentane-1,4-diamine sulfate, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder, freely soluble in water and in methanol, very slightly soluble in ethanol (96 per cent).

It melts at about 208 °C (instantaneous method).

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Dissolve 0.100 g in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL

with *water R*. Examined between 210 nm and 370 nm (*2.2.25*), the solution shows absorption maxima at 220 nm, 235 nm, 256 nm, 329 nm and 342 nm. The specific absorbances at the maxima are respectively 730 to 810, 430 to 470, 370 to 410, 400 to 440 and 430 to 470.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with the base isolated from *chloroquine sulfate CRS*. Record the spectra using solutions prepared as follows: dissolve separately 0.1 g of the substance to be examined and of the reference substance in 10 mL of *water R*, add 2 mL of *dilute sodium hydroxide solution R* and shake with 2 quantities, each of 20 mL, of *methylene chloride R*; combine the organic layers, wash with *water R*, dry over *anhydrous sodium sulfate R*, evaporate to dryness and dissolve the residues separately each in 2 mL of *methylene chloride R*.

C. Dissolve 25 mg in 20 mL of *water R* and add 8 mL of *picric acid solution R1*. The precipitate, washed with *water R*, with *ethanol (96 per cent) R* and finally with *ether R*, melts (*2.2.14*) at 206 °C to 209 °C.

D. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 or GY_5 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of solution S is 4.0 to 5.0.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel GF_{254} R* as the coating substance.

Test solution Dissolve 0.50 g of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 100 mL with *water R*.

Reference solution (b) Dilute 5 mL of reference solution (a) to 10 mL with *water R*.

Apply separately to the plate 2 µL of each solution. Develop over a path of 12 cm using a mixture of 10 volumes of *diethylamine R*, 40 volumes of *cyclohexane R* and 50 volumes of *methylene chloride R*. Allow the plate to dry in air. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (1.0 per cent) and not more than one such spot is more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Water (*2.5.12*)

3.0 per cent to 5.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

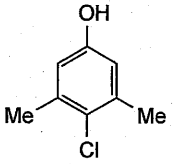
1 mL of *0.1 M perchloric acid* is equivalent to 41.8 mg of $C_{18}H_{28}ClN_3O_4S$.

STORAGE

Store in an airtight container, protected from light.

Ph Eur

Chloroxylenol



C_8H_9ClO 156.6 *88-04-0*

Action and use

Antiseptic.

Preparation

Chloroxylenol Solution

DEFINITION

Chloroxylenol is 4-chloro-3,5-xylenol. It contains not less than 98.0% and not more than 103.0% of C_8H_9ClO.

CHARACTERISTICS

White or cream crystals or crystalline powder. It is volatile in steam.

Very slightly soluble in *water*; freely soluble in *ethanol (96%)*; soluble in *ether*, in terpenes and in fixed oils. It dissolves in solutions of the alkali hydroxides.

IDENTIFICATION

The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of chloroxylenol *(RS 055)*.

TESTS

Melting point

114° to 116°, Appendix V A.

Tetrachloroethylene

Carry out the method for *gas chromatography*, Appendix III B, using the following solutions. Prepare a 0.2% v/v solution of *butanol* (internal standard) in *methanol* (solution A).

(1) To 4 g of the substance being examined, add 5 mL of solution A and dilute to 25 mL with *methanol*.

(2) To 5 mL of a 0.2% v/v solution of *tetrachloroethylene* in *methanol*, add 5 mL of solution A and dilute to 25 mL with *methanol* (equivalent to 0.06488% w/v of *tetrachloroethylene* in *methanol*).

CHROMATOGRAPHIC CONDITIONS

(a) Use a fused silica capillary column (30 m × 0.53 mm) bonded with a 1 µm film of *polyethylene glycol 20,000* (RH-Wax is suitable).

(b) Use *hydrogen* as the carrier gas at 2 mL per minute.

(c) Use the gradient conditions described below.

(d) Use an inlet temperature of 240°.

(e) Use a flame ionisation detector at a temperature of 280°.

(f) Inject 0.5 µL of each solution.

(g) Use a split ratio of 1:20.

Time (Minutes)	Temperature	Comment
0 - 4	70°	isothermal
4 - 5	70°→210°	linear increase
5 - 15	210	isothermal
15 - 18	210°→70°	linear gradient
18 - 20	70°	re-equilibration

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (2), the *resolution* between the peaks due to tetrachloroethylene and the internal standard is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1), the ratio of any peak due to tetrachloroethylene to that of the internal standard is not greater than the corresponding ratio obtained in the chromatogram obtained with solution (2) (0.4%).

Related substances

Carry out the method for *gas chromatography*, Appendix III B, using the following solutions in *chloroform*.

(1) 2.0% w/v of the substance being examined.

(2) 2.0% w/v of the substance being examined and 0.040% w/v of *4-chloro-o-cresol* (internal standard).

CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with *acid-washed diatomaceous support* (80 to 100 mesh) coated with 3% w/w of polyethylene glycol (Carbowax 20M is suitable).

(b) Use *nitrogen* as the carrier gas at 40 mL per minute.

(c) Use isothermal conditions maintained at 160°.

(d) Use an inlet temperature of 200°.

(e) Use a flame ionisation detector at a temperature of 300°.

(f) Inject 1 µL of each solution.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (2), the *resolution* between the peaks due to chloroxylenol and 4-chloro-o-cresol is at least 1.5

LIMITS

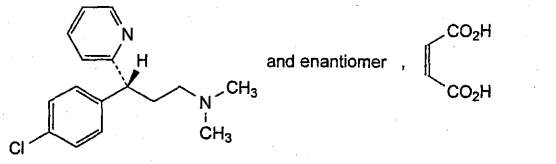
In the chromatogram obtained with solution (2) the sum of the areas of any *secondary peaks* is not greater than the area of the peak due to the internal standard.

ASSAY

Dissolve 70 mg in 30 mL of *glacial acetic acid*, add 25 mL of *0.0167M potassium bromate VS*, 20 mL of a 15% w/v solution of *potassium bromide* and 10 mL of *hydrochloric acid*, stopper the flask and allow to stand protected from light for 15 minutes. Add 1 g of *potassium iodide* and 100 mL of *water* and titrate with 0.1M *sodium thiosulfate VS*, shaking vigorously and using 1 mL of *starch solution*, added towards the end of the titration, as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of potassium bromate required. Each mL of *0.0167M potassium bromate VS* is equivalent to 3.915 mg of C_8H_9ClO.

Chlorphenamine Maleate

(Ph. Eur. monograph 0386)



$C_{20}H_{23}ClN_2O_4$ 390.9 *113-92-8*

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparations

Chlorphenamine Injection

Chlorphenamine Oral Solution

Chlorphenamine Tablets

Ph Eur

DEFINITION

(3*RS*)-3-(4-Chlorophenyl)-*N*,*N*-dimethyl-3-(pyridin-2-yl) propan-1-amine hydrogen (*Z*)-butenedioate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Melting point (*2.2.14*): 130 °C to 135 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chlorphenamine maleate CRS*.

C. Optical rotation (see Tests).

TESTS

Solution S

Dissolve 2.0 g in *water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)

−0.10° to + 0.10°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *chlorphenamine impurity C CRS* in 5 mL of the test solution and dilute to 50.0 mL with the mobile phase. Dilute 2 mL of this solution to 20 mL with the mobile phase.

Reference solution (d) Dissolve 5 mg of *2,2'-dipyridylamine R* (impurity B) in the mobile phase and dilute to 100 mL with the mobile phase.

Reference solution (e) Dissolve the contents of a vial of *chlorphenamine impurity A CRS* in 2 mL of the test solution. Sonicate for 5 min.

Column:
— *size*: *l* = 0.30 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm).

Mobile phase Mix 20 volumes of *acetonitrile R* and 80 volumes of a 8.57 g/L solution of *ammonium dihydrogen phosphate R* previously adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL.

Run time 3.5 times the retention time of chlorphenamine.

Relative retention With reference to chlorphenamine (retention time = about 11 min): maleic acid = about 0.2; impurity A = about 0.3; impurity B = about 0.4; impurity C = about 0.9; impurity D = about 3.0.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity C and chlorphenamine.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.5; impurity B = 1.4;
— *impurity A*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurities B, C, D*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to maleic acid.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

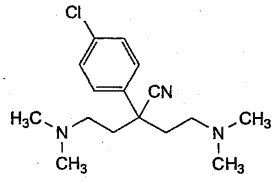
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 25 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

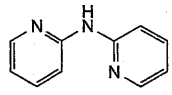
1 mL of *0.1 M perchloric acid* is equivalent to 19.54 mg of $C_{20}H_{23}ClN_2O_4$.

STORAGE
Protected from light.

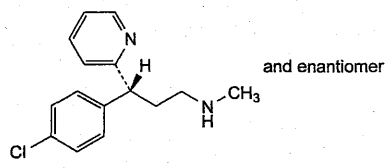
IMPURITIES
Specified impurities *A, B, C, D.*



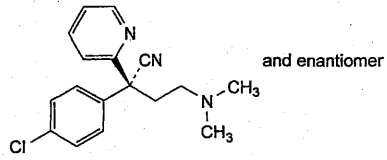
A. 2-(4-chlorophenyl)-4-(dimethylamino)-2-[2-(dimethylamino)ethyl]butanenitrile,



B. *N*-(pyridin-2-yl)pyridin-2-amine (2,2′-dipyridylamine),



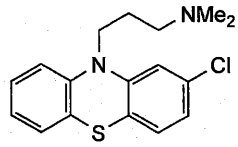
C. (3*RS*)-3-(4-chlorophenyl)-*N*-methyl-3-(pyridin-2-yl) propan-1-amine,



D. (2*RS*)-2-(4-chlorophenyl)-4-(dimethylamino)-2-(pyridin-2-yl)butanenitrile.

Ph Eur

Chlorpromazine



$C_{17}H_{19}ClN_2S$ 318.9 *50-53-3*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparation
Chlorpromazine Suppositories

DEFINITION
Chlorpromazine is [3-(2-chlorophenothiazin-10-yl)propyl]-dimethylamine. It contains not less than 99.0% and not more than 101.0% of $C_{17}H_{19}ClN_2S$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or creamy white powder or waxy solid.

Practically insoluble in *water*; freely soluble in *ethanol (96%)* and in *ether*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of chlorpromazine *(RS 056)*.

B. Complies with the test for *identification of phenothiazines*, Appendix III A, using *chlorpromazine hydrochloride BPCRS* to prepare reference solution.

TESTS

Melting point
56° to 58°, Appendix V A.

Related substances
Complies with the test for *related substances in phenothiazines*, Appendix III A, using *mobile phase A*.

Loss on drying
When dried to constant weight over *phosphorus pentoxide* at a pressure not exceeding 0.7 kPa, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY

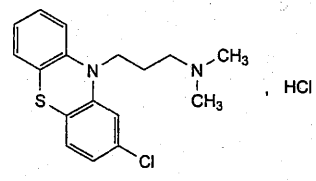
Dissolve 0.8 g in 300 mL of *acetone* and carry out Method I for *non-aqueous titration*, Appendix VIII A, using 3 mL of a saturated solution of *methyl orange* in *acetone* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 31.89 mg of $C_{17}H_{19}ClN_2S$.

STORAGE

Chlorpromazine should be protected from light.

Chlorpromazine Hydrochloride

(Ph. Eur. monograph 0475)



$C_{17}H_{20}Cl_2N_2S$ 355.3 *69-09-0*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparations
Chlorpromazine Injection
Chlorpromazine Oral Solution
Chlorpromazine Tablets

Ph Eur

DEFINITION

3-(2-Chloro-10*H*-phenothiazin-10-yl)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

It decomposes on exposure to air and light.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*). *Prepare the solutions protected from bright light and measure the absorbances immediately.*

Test solution Dissolve 50.0 mg in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 500.0 mL with the same solution. Dilute 5.0 mL of the solution to 100.0 mL with a 10.3 g/L solution of *hydrochloric acid R*.

Spectral range 230-340 nm.

Absorption maxima At 254 nm and 306 nm.

Specific absorbance at the absorption maximum at 254 nm 890 to 960.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation 60 g/L solutions in *methylene chloride R* using a 0.1 mm cell.

Comparison *chlorpromazine hydrochloride CRS.*

C. Identification of phenothiazines by thin-layer chromatography (*2.3.3*): use *chlorpromazine hydrochloride CRS* to prepare the reference solution.

D. It gives reaction (b) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)
3.5 to 4.5. *Carry out the test protected from light and use freshly prepared solutions.*

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Impurity F
Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use and protect from light.*

Solvent mixture *diethylamine R, methanol R* (5:95 *V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of *chlorpromazine impurity F CRS* in 2.0 mL of the solvent mixture.

Reference solution (b) Dilute 300 µL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 0.10 g of the substance to be examined in the solvent mixture, add 1.0 mL of reference solution (a) and dilute to 5.0 mL with the solvent mixture.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetone R, diethylamine R, cyclohexane R* (10:10:80 *V/V/V*).

Application 10 µL of the test solution and reference solutions (b) and (c).

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Retardation factors Impurity F = about 0.5; chlorpromazine = about 0.6.

System suitability Reference solution (c):
— the chromatogram shows 2 clearly separated spots due to impurity F and chlorpromazine.

Limit:
— *impurity F*: any spot due to impurity F is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.15 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 4 mg of *chlorpromazine impurity D CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. To 1 mL of the solution add 1 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 4.0 mg of *chlorpromazine impurity A CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve 4 mg of *promazine hydrochloride CRS* (impurity C) and 4.0 mg of *chlorpromazine impurity E CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 0.2 volumes of thiodiethylene glycol R with 50 volumes of *acetonitrile R* and 50 volumes of a 0.5 per cent *V/V* solution of *trifluoroacetic acid R* previously adjusted to pH 5.3 with *tetramethylethylenediamine R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 4 times the retention time of chlorpromazine.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities C and E; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to chlorpromazine (retention time = about 8 min): impurity A = about 0.4; impurity B = about 0.5; impurity C = about 0.7; impurity D = about 0.9; impurity E = about 3.4.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity D and chlorpromazine.

Limits:
— *impurities B, C, D*: for each impurity, not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurity A*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
— *impurity E*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: maximum 1.0 per cent;
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of *0.1 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

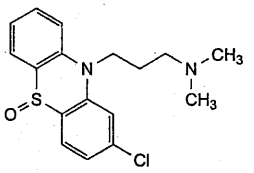
1 mL of *0.1 M sodium hydroxide* is equivalent to 35.53 mg of $C_{17}H_{20}Cl_2N_2S$.

STORAGE

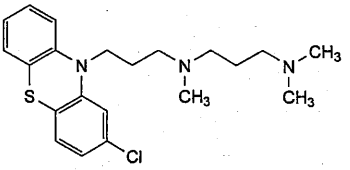
In an airtight container, protected from light.

IMPURITIES

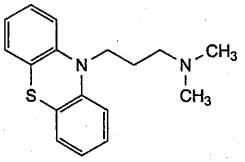
Specified impurities *A, B, C, D, E, F.*



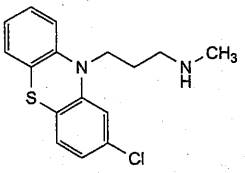
A. 3-(2-chloro-10*H*-phenothiazin-10-yl)-*N*,*N*-dimethylpropan-1-amine *S*-oxide (chlorpromazine sulfoxide),



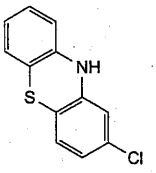
B. *N*-[3-(2-chloro-10*H*-phenothiazin-10-yl)propyl]-*N*,*N*′,*N*′-trimethylpropane-1,3-diamine,



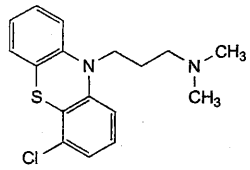
C. 3-(10*H*-phenothiazin-10-yl)-*N*,*N*-dimethylpropan-1-amine (promazine),



D. 3-(2-chloro-10*H*-phenothiazin-10-yl)-*N*-methylpropan-1-amine (desmethylchlorpromazine),



E. 2-chloro-10*H*-phenothiazine,

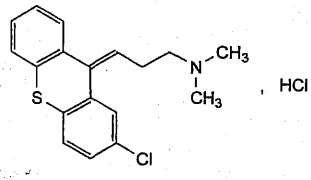


F. 3-(4-chloro-10*H*-phenothiazin-10-yl)-*N*,*N*-dimethylpropan-1-amine.

Ph Eur

Chlorprothixene Hydrochloride

(Ph. Eur. monograph 0815)



$C_{18}H_{19}Cl_2NS$ 352.3 *6469-93-8*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
(3*Z*)-3-(2-Chloro-9*H*-thioxanthen-9-ylidene)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water and in ethanol (96 per cent), slightly soluble in methylene chloride.

mp
About 220 °C.

IDENTIFICATION
First identification: A, E.

Second identification: B, C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 25 mg in 1 mL of *water R*, add 0.1 mL of *dilute sodium hydroxide solution R* and shake with 2 mL of *methylene chloride R*; separate the organic layer and wash with 0.5 mL of *water R*; evaporate the organic layer to dryness and dry the residue at 40-50 °C; examine the residue as a disc.

Comparison Repeat the operations using 25 mg of *chlorprothixene hydrochloride CRS*.

B. Dissolve 0.2 g in a mixture of 5 mL of *dioxan R* and 5 mL of a 1.5 g/L solution of *sodium nitrite R*. Add 0.8 mL of *nitric acid R*. After 10 min, add the solution to 20 mL of *water R*. Filter the precipate formed after 1 h. The filtrate is used immediately for identification test C. Dissolve the precipitate by warming in about 15 mL of *ethanol (96 per cent) R* and add the solution to 10 mL of *water R*. Filter and dry the precipitate at 100-105 °C for 2 h. The melting point (*2.2.14*) is 152 °C to 154 °C.

C. To 1 mL of the filtrate obtained in identification test B, add 0.2 mL of a suspension of 50 mg of *fast red B salt R* in 1 mL of *ethanol (96 per cent) R*. Add 1 mL of *0.5 M alcoholic potassium hydroxide*. A dark red colour is produced. Carry out a blank test.

D. Dissolve about 20 mg in 2 mL of *nitric acid R*. A red colour is produced. Add 5 mL of *water R* and examine in ultraviolet light at 365 nm. The solution shows green fluorescence.

E. Dissolve 20 mg in 2 mL of *water R*, acidify with *dilute nitric acid R* and allow to stand for 5 min. Centrifuge. The supernatant gives reaction (a) of chlorides (*2.3.1*) starting from 'add 0.4 mL of *silver nitrate solution R1*'.

TESTS
Solution S
Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.4 to 5.2 for solution S.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from bright light.*

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *chlorprothixene for system suitability CRS* (containing impurities C and F) in 1 mL of the mobile phase.

Column:
— *size: l* = 0.10 m, Ø = 4.0 mm;
— *stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Solution containing 6.0 g/L of *potassium dihydrogen phosphate R*, 2.9 g/L of *sodium laurilsulfate R* and 9 g/L of *tetrabutylammonium bromide R* in a mixture of 50 volumes of *methanol R*, 400 volumes of *acetonitrile R* and 550 volumes of *water for chromatography R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration For about 30 min with the mobile phase.

Injection 20 µL.

Run time Twice the retention time of chlorprothixene.

Identification of impurities Use the chromatogram supplied with *chlorprothixene for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and F.

Relative retention With reference to chlorprothixene (retention time = about 10 min): impurity C = about 1.25; impurity F = about 1.33.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to chlorprothixene and impurity C.

Limits:
— *impurity F*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

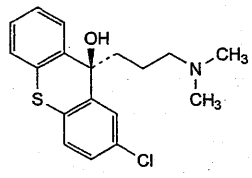
ASSAY
Dissolve 0.300 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 35.23 mg of $C_{18}H_{19}Cl_2NS$.

STORAGE
Protected from light.

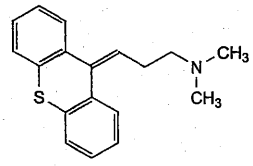
IMPURITIES
Specified impurities *F.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E.*

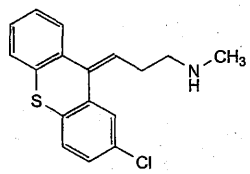


and enantiomer

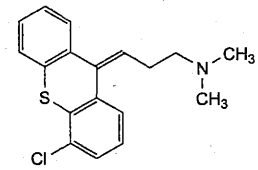
A. (3*RS*)-2-chloro-9-[3-(dimethylamino)propyl]-9*H*-thioxanthen-9-ol,



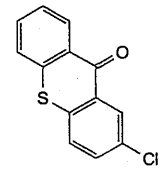
B. *N*,*N*-dimethyl-3-(9*H*-thioxanthen-9-ylidene)propan-1-amine,



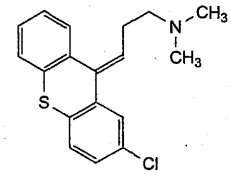
C. (3*Z*)-3-(2-chloro-9*H*-thioxanthen-9-ylidene)-*N*-methylpropan-1-amine,



D. (3*Z*)-3-(4-chloro-9*H*-thioxanthen-9-ylidene)-*N*,*N*-dimethylpropan-1-amine,



E. 2-chloro-9*H*-thioxanthen-9-one,

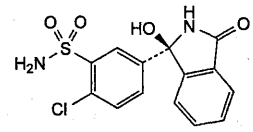


F. (3*E*)-3-(2-chloro-9*H*-thioxanthen-9-ylidene)-*N*,*N*-dimethylpropan-1-amine ((*E*)-chlorprothixene).

Ph Eur

Chlortalidone

(Ph. Eur. monograph 0546)



and enantiomer

$C_{14}H_{11}ClN_2O_4S$ 338.8 *77-36-1*

Action and use
Thiazide-like diuretic.

Preparations
Chlortalidone Tablets
Co-tenidone Tablets

Ph Eur

DEFINITION
2-Chloro-5-[(1*RS*)-1-hydroxy-3-oxo-2,3-dihydro-1*H*-isoindol-1-yl]benzenesulfonamide.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish-white powder.

Solubility
Very slightly soluble in water, soluble in acetone and in methanol, practically insoluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *chlortalidone CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Acidity

Dissolve 1.0 g with heating in a mixture of 25 mL of *acetone R* and 25 mL of *carbon dioxide-free water R*. Cool. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Not more than 0.75 mL of *0.1 M sodium hydroxide* is required.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mix 2 volumes of a 2 g/L solution of *sodium hydroxide R*, 48 volumes of mobile phase B and 50 volumes of mobile phase A.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *chlortalidone for peak identification CRS* (containing impurities B, G and J) in 1 mL of the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *chlortalidone CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 10.0 mL of this solution to 100.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: dissolve 1.32 g of *ammonium phosphate R* in about 900 mL of *water R* and adjust to pH 5.5 with *dilute phosphoric acid R*; dilute to 1000 mL with *water R*;
- *mobile phase B*: *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 16	65	35
16 - 21	65 → 50	35 → 50
21 - 35	50	50
35 - 45	50 → 65	50 → 35

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram obtained with reference solution (b) and the chromatogram supplied with *chlortalidone for peak identification CRS* to identify the peaks due to impurities B, G and J.

Relative retention With reference to chlortalidone (retention time = about 7 min): impurity B = about 0.7; impurity J = about 0.9; impurity G = about 6.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurity J and chlortalidone.

Limits:
- *impurity B*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
- *impurity J*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity G*: not more than 2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 12 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.2 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)

Maximum 350 ppm.

Triturate 0.3 g finely, add 30 mL of *water R*, shake for 5 min and filter. 15 mL of the filtrate complies with the test. Prepare the standard using 10 mL of *chloride standard solution (5 ppm Cl) R*.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

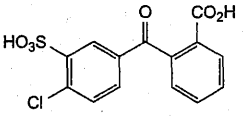
Injection 20 µL of test solution (b) and reference solution (c).

Calculate the percentage content of $C_{14}H_{11}ClN_2O_4S$ from the declared content of *chlortalidone CRS*.

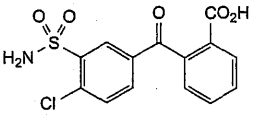
IMPURITIES

Specified impurities *B, G, J.*

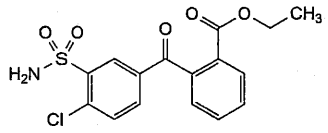
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, F, H, I.*



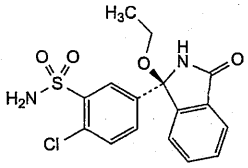
A. 2-(4-chloro-3-sulfobenzoyl)benzoic acid,



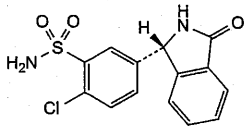
B. 2-(4-chloro-3-sulfamoylbenzoyl)benzoic acid,



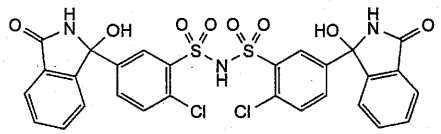
C. ethyl 2-(4-chloro-3-sulfamoylbenzoyl)benzoate,



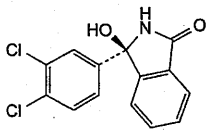
D. 2-chloro-5-[(1*RS*)-1-ethoxy-3-oxo-2,3-dihydro-1*H*-isoindol-1-yl]benzenesulfonamide,



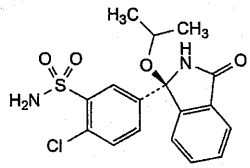
E. 2-chloro-5-[(1*RS*)-3-oxo-2,3-dihydro-1*H*-isoindol-1-yl] benzenesulfonamide,



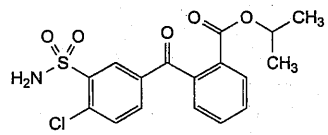
F. bis[2-chloro-5-(1-hydroxy-3-oxo-2,3-dihydro-1*H*-isoindol-1-yl)benzenesulfonyl]amine,



G. (3*RS*)-3-(3,4-dichlorophenyl)-3-hydroxy-2,3-dihydro-1*H*-isoindol-1-one,



H. 2-chloro-5-[(1*RS*)-1-(1-methylethoxy)-3-oxo-2,3-dihydro-1*H*-isoindol-1-yl]benzenesulfonamide,



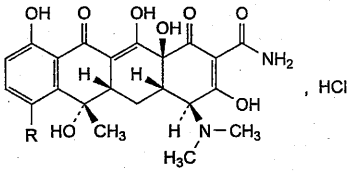
I. 1-methylethyl 2-(4-chloro-3-sulfamoylbenzoyl)benzoate,

J. impurity of unknown structure with a relative retention of about 0.9.

Ph Eur

Chlortetracycline Hydrochloride

(Ph. Eur. monograph 0173)



Compound	R	Molecular formula	M_r
Chlortetracycline hydrochloride	Cl	$C_{22}H_{24}Cl_2N_2O_8$	515.3
Tetracycline hydrochloride	H	$C_{22}H_{25}ClN_2O_8$	480.9

Chlortetracycline hydrochloride *64-72-2*

Tetracycline hydrochloride *64-75-5*

Action and use
Tetracycline antibacterial.

Ph Eur

DEFINITION

Mixture of antibiotics, the main component being the hydrochloride of (4*S*,4a*S*,5a*S*,6*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (chlortetracycline hydrochloride), a substance produced by the growth of certain strains of *Streptomyces aureofaciens* or obtained by any other means.

Content

— *chlortetracycline hydrochloride* ($C_{22}H_{24}Cl_2N_2O_8$): minimum 89.5 per cent (anhydrous substance);
— *tetracycline hydrochloride* ($C_{22}H_{25}ClN_2O_8$): maximum 6.0 per cent (anhydrous substance);
— *sum of the contents of chlortetracycline hydrochloride and tetracycline hydrochloride*: 94.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Yellow powder.

Solubility
Slightly soluble in water and in ethanol (96 per cent).
It dissolves in solutions of alkali hydroxides and carbonates.

IDENTIFICATION

First identification: C, D.

Second identification: A, B, C.

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *chlortetracycline hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *chlortetracycline hydrochloride CRS*, 5 mg of *demeclocycline hydrochloride R* and 5 mg of *doxycycline R* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase Mix 20 volumes of *acetonitrile R*, 20 volumes of *methanol R* and 60 volumes of a 63 g/L solution of *oxalic*

acid R previously adjusted to pH 2 with *concentrated ammonia R*.

Application 1 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of *sulfuric acid R*. A deep blue colour develops and becomes bluish-green. Add the solution to 2.5 mL of *water R*. The colour becomes brownish.

C. It gives reaction (a) of chlorides (*2.3.1*).

D. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

pH (*2.2.3*)

2.3 to 3.3.

Dissolve 0.1 g in 10 mL of *carbon dioxide-free water R*, heating slightly.

Specific optical rotation (*2.2.7*)

−250 to −235 (anhydrous substance).

Dissolve 0.125 g in *water R* and dilute to 50.0 mL with the same solvent.

Absorbance (*2.2.25*)

Maximum 0.40 at 460 nm.

Dissolve 0.125 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase B and dilute to 25.0 mL with mobile phase B.

Reference solution (a) Dissolve 25.0 mg of *chlortetracycline hydrochloride CRS* in mobile phase B and dilute to 25.0 mL with mobile phase B.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase B.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with mobile phase B.

Reference solution (d) Dissolve 5 mg of *chlortetracycline for system suitability CRS* (containing impurities A, B, D, E, G, H, J, K and L) in mobile phase B and dilute to 5 mL with mobile phase B.

Reference solution (e) Dissolve 25.0 mg of *tetracycline hydrochloride CRS* in mobile phase B and dilute to 25.0 mL with mobile phase B. Dilute 5.0 mL of this solution to 100.0 mL with mobile phase B.

Column:

— *size*: *l* = 0.075 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octylsilyl silica gel for chromatography with polar incorporated groups R* (3.5 µm);

— *temperature*: 45 °C.

Mobile phase:

— *mobile phase A*: to 725 mL of *water R* add 50 mL of *perchloric acid solution R*, shake and add 225 mL of *dimethyl sulfoxide R*;

— *mobile phase B*: to 250 mL of *water R* add 50 mL of *perchloric acid solution R*, shake and add 700 mL of *dimethyl sulfoxide R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 46	100 → 0	0 → 100

Flow rate 0.4 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *chlortetracycline for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, D, E, G, H, J, K and L.

Relative retention With reference to chlortetracycline (retention time = about 26 min): impurity D = about 0.5; tetracycline = about 0.6; impurity E = about 0.7; impurity B = about 0.8, impurity A = about 0.86; impurity G = about 0.9; impurity H = about 1.1; impurity J = about 1.4, impurity K = about 1.67; impurity L = about 1.71.

System suitability Reference solution (d):

— *resolution*: minimum 1.5 between the peaks due to tetracycline and impurity E; minimum 1.5 between the peaks due to impurities A and G; minimum 1.5 between the peaks due to impurities K and L; if necessary, adjust the concentration of dimethyl sulfoxide in mobile phase A.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity G = 1.4; impurity J = 0.3; impurity K = 0.4; impurity L = 0.4;

— *impurity A*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (4.0 per cent);

— *impurities B, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *impurity J*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— *impurities D, G, H, L*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *impurity K*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *sum of impurities other than A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 1 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solutions (a) and (e).

Calculate the percentage content of $C_{22}H_{24}Cl_2N_2O_8$ using the chromatogram obtained with reference solution (a) and taking into account the assigned content of *chlortetracycline hydrochloride CRS*. Calculate the percentage content of $C_{22}H_{25}ClN_2O_8$ using the chromatogram obtained with reference solution (e) and taking into account the assigned content of *tetracycline hydrochloride CRS*.

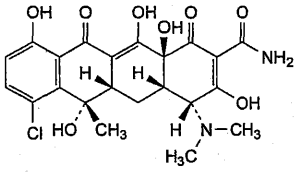
STORAGE

Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

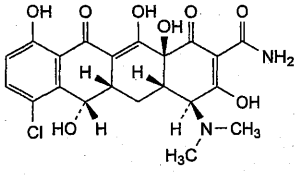
IMPURITIES

Specified impurities *A, B, D, E, G, H, J, K, L.*

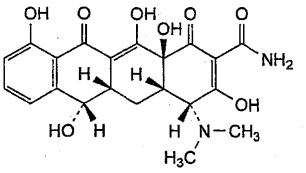
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, F, I.*



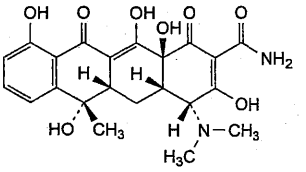
A. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epichlortetracycline),



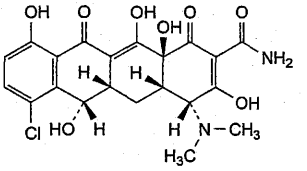
B. (4*S*,4a*S*,5a*S*,6*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (demeclocycline),



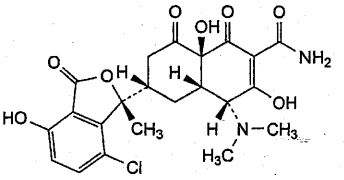
C. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidemethyltetracycline),



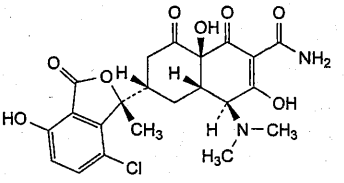
D. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epitetracycline),



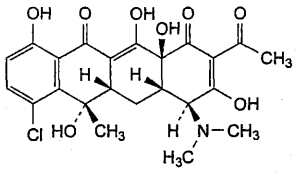
E. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidemethylchlortetracycline),



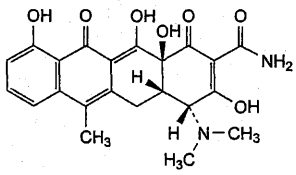
F. (4*R*,4a*S*,6*S*,8a*S*)-6-[(1*R*)-7-chloro-4-hydroxy-1-methyl-3-oxo-1,3-dihydro-2-benzofuran-1-yl]-4-(dimethylamino)-3,8a-dihydroxy-1,8-dioxo-1,4,4a,5,6,7,8,8a-octahydronaphthalene-2-carboxamide (4-epiisochlortetracycline),



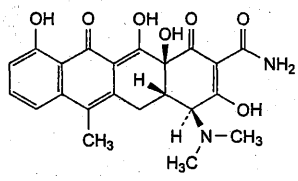
G. (4*S*,4a*S*,6*S*,8a*S*)-6-[(1*R*)-7-chloro-4-hydroxy-1-methyl-3-oxo-1,3-dihydro-2-benzofuran-1-yl]-4-(dimethylamino)-3,8a-dihydroxy-1,8-dioxo-1,4,4a,5,6,7,8,8a-octahydronaphthalene-2-carboxamide (isochlortetracycline),



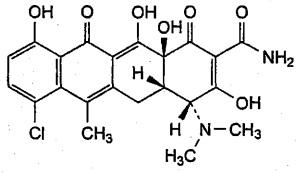
H. (4*S*,4a*S*,5a*S*,6*S*,12a*S*)-2-acetyl-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-4a,5a,6,12a-tetrahydrotetracene-1,11(4*H*,5*H*)-dione (2-acetyl-2-decarboxamidochlortetracycline),



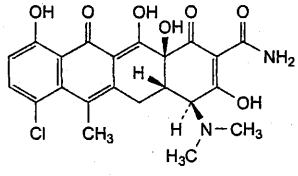
I. (4*R*,4a*S*,12a*S*)-4-(dimethylamino)-3,10,12,12a-tetrahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,11,12a-hexahydrotetracene-2-carboxamide (4-epianhydrotetracycline),



J. (4*S*,4a*S*,12a*S*)-4-(dimethylamino)-3,10,12,12a-tetrahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,11,12a-hexahydrotetracene-2-carboxamide (anhydrotetracycline),



K. (4*R*,4a*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,10,12,12a-tetrahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,11,12a-hexahydrotetracene-2-carboxamide (4-epianhydrochlortetracycline),

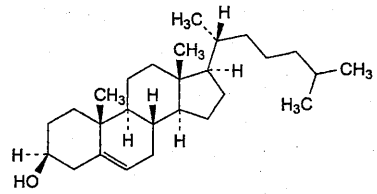


L. (4*S*,4a*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,10,12,12a-tetrahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,11,12a-hexahydrotetracene-2-carboxamide (anhydrochlortetracycline).

Ph Eur

Cholesterol

(Ph. Eur. monograph 0993)



$C_{27}H_{46}O$ 386.7 *57-88-5*

Action and use
Excipient.

Ph Eur

DEFINITION
Cholest-5-en-3β-ol.

Content
— *cholesterol*: minimum 95.0 per cent (dried substance);
— *total sterols*: 97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in acetone and in ethanol (96 per cent).

It is sensitive to light.

IDENTIFICATION
A. Melting point (*2.2.14*): 147 °C to 150 °C.

B. Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution Dissolve 10 mg of the substance to be examined in *ethylene chloride R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 10 mg of *cholesterol CRS* in *ethylene chloride R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *ethyl acetate R, toluene R* (33:66 *V/V*).

Application 20 µL.

Development Immediately, protected from light, over a path of 15 cm.

Drying In air.

Detection Spray 3 times with *antimony trichloride solution R*; examine within 3-4 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve about 5 mg in 2 mL of *methylene chloride R*. Add 1 mL of *acetic anhydride R*, 0.01 mL of *sulfuric acid R* and shake. A pink colour is produced which rapidly changes to red, then to blue and finally to brilliant green.

TESTS
Solubility in ethanol (96 per cent)
In a stoppered flask, dissolve 0.5 g in 50 mL of *ethanol (96 per cent) R* at 50 °C. Allow to stand for 2 h. No deposit or turbidity is formed.

Acidity
Dissolve 1.0 g in 10 mL of *ether R*, add 10.0 mL of *0.1 M sodium hydroxide* and shake for about 1 min. Heat gently to eliminate ether and then boil for 5 min. Cool, add 10 mL of *water R* and 0.1 mL of *phenolphthalein solution R* as indicator and titrate with *0.1 M hydrochloric acid* until the pink colour just disappears, stirring the solution vigorously throughout the titration. Carry out a blank titration. The difference between the volumes of *0.1 M hydrochloric acid* required to change the colour of the indicator in the blank and in the test is not more than 0.3 mL.

Loss on drying (*2.2.32*)
Maximum 0.3 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.100 g of *pregnenolone isobutyrate CRS* in *heptane R* and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 25.0 mg of the substance to be examined in the internal standard solution and dilute to 25.0 mL with the same solution.

Reference solution Dissolve 25.0 mg of *cholesterol CRS* in the internal standard solution and dilute to 25.0 mL with the same solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R.*

Flow rate 2 mL/min.

Split ratio 1:25.

Temperature:
— *column*: 275 °C;
— *injection port*: 285 °C;
— *detector*: 300 °C.

Detection Flame ionisation.

Injection 1.0 µL.

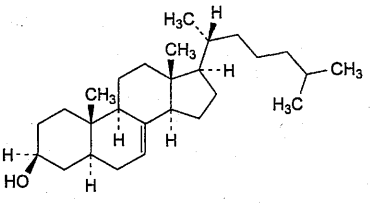
System suitability Reference solution:
— *resolution*: minimum 10.0 between the peaks due to pregnenolone isobutyrate and cholesterol;
— *symmetry factor*: minimum 0.6 for the peak due to cholesterol.

Calculate the percentage content of cholesterol taking into account the assigned content of *cholesterol CRS*. Calculate the percentage content of total sterols by adding together the contents of cholesterol and other substances with a retention time less than or equal to 1.5 times the retention time of cholesterol. Disregard the peaks due to the internal standard and the solvent.

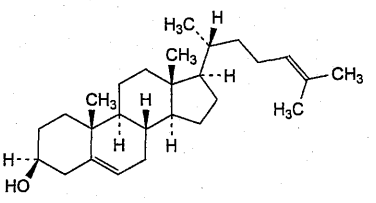
STORAGE
Protected from light.

LABELLING
The label states the source material for the production of cholesterol (for example bovine brain and spinal cord, wool fat or chicken eggs).

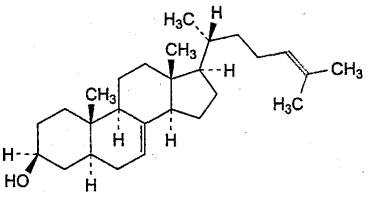
IMPURITIES



A. 5α-cholest-7-en-3β-ol (lathosterol),



B. cholesta-5,24-dien-3β-ol (desmosterol),

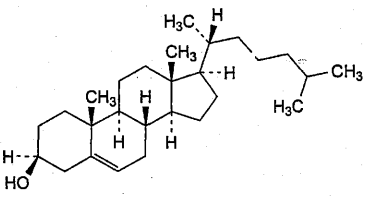


C. 5α-cholesta-7,24-dien-3β-ol.

Ph Eur

Cholesterol for Parenteral Use

(Ph. Eur. monograph 2397)



$C_{27}H_{46}O$ 386.7 *57-88-5*

Action and use
Excipient.

Ph Eur

DEFINITION
Cholest-5-en-3β-ol obtained from *Wool fat (0134)*.

Content
— *cholesterol*: 99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in acetone and in ethanol (96 per cent).

It is sensitive to light.

IDENTIFICATION
A. Melting point (*2.2.14*): 147 °C to 150 °C.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

C. Dissolve about 5 mg in 2 mL of *methylene chloride R*. Add 1 mL of *acetic anhydride R* and 0.01 mL of *sulfuric acid R* and shake. A pink colour is produced which rapidly changes to red, then to blue and finally to bright green.

TESTS

Solubility in ethanol (96 per cent)

In a stoppered flask, dissolve 0.5 g in 50 mL of *ethanol (96 per cent) R* at 50 °C. Allow to stand for 2 h. The solution is clear.

Acidity

Dissolve 1.0 g in 10 mL of *ether R*, add 10.0 mL of *0.1 M sodium hydroxide* and shake for about 1 min. Heat gently to eliminate the ether and then boil for 5 min. Cool, add 10 mL of *water R* and 0.1 mL of *phenolphthalein solution R* as indicator and titrate with *0.1 M hydrochloric acid* until the pink colour just disappears, stirring the solution vigorously throughout the titration. Carry out a blank titration. The difference between the volumes of *0.1 M hydrochloric acid* required to change the colour of the indicator in the blank titration and in the test is not more than 0.1 mL.

Peroxide value (*2.5.5, Method A*)

Maximum 10.

Other sterols

Gas chromatography (*2.2.28*): use the normalisation procedure.

Internal standard solution Dissolve 0.100 g of *pregnenolone isobutyrate CRS* in *heptane R* and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 25.0 mg of the substance to be examined in the internal standard solution and dilute to 25.0 mL with the same solution.

Reference solution Dissolve 25.0 mg of *cholesterol CRS* in the internal standard solution and dilute to 25.0 mL with the same solution.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.25 mm;
- *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Flow rate 2 mL/min.

Split ratio 1:25.

Temperature:
- *column*: 275 °C;
- *injection port*: 285 °C;
- *detector*: 300 °C.

Detection Flame ionisation.

Injection 1.0 µL.

Relative retention With reference to cholesterol (retention time = about 8.5 min): pregnenolone isobutyrate = about 0.8.

System suitability Reference solution:
- *resolution*: minimum 10.0 between the peaks due to pregnenolone isobutyrate and cholesterol.

Limits:
- *total of other substances with a retention time less than or equal to 1.5 times the retention time of cholesterol*: maximum 0.5 per cent;
- *disregard limit*: 0.05 per cent; disregard the peak due to the internal standard.

Benzoyl ureas

Liquid chromatography (*2.2.29*).

Test solution Dissolve 1.0 g of the substance to be examined in 200 mL of *heptane R* using a magnetic stirrer and add 10 mL of *acetonitrile R*. Shake and allow the layers to separate. Isolate the lower layer (acetonitrile) and add 10 mL of *acetonitrile R* to the heptane layer and extract again. Combine the lower layers and evaporate to dryness by suitable means. Add 0.5 mL of *acetonitrile R* then 0.5 mL of *water R* to the residue. Suspend with the aid of ultrasound for about 5 min. Centrifuge the suspension for 5 min and use the supernatant.

Reference solution (a) Dissolve 10.0 mg of *diflubenzuron R* (impurity A) and 10.0 mg of *triflumuron R* (impurity B) in *acetonitrile R* and dilute to 100.0 mL with the same solvent. Dilute 0.1 mL of the solution to 100.0 mL with *acetonitrile R*.

Reference solution (b) Mix 0.5 mL of reference solution (a) and 0.5 mL of *water R*.

Reference solution (c) Dissolve 1.0 g of the substance to be examined in 200 mL of *heptane R* using a magnetic stirrer. Add 0.5 mL of reference solution (a) and 9.5 mL of *acetonitrile R*. Shake and allow the layers to separate. Isolate the lower layer (acetonitrile) and add 10 mL of *acetonitrile R* to the heptane layer and extract again. Combine the lower layers and evaporate to dryness by suitable means. Add 0.5 mL of *acetonitrile R* then 0.5 mL of *water R* to the residue. Suspend with the aid of ultrasound for about 5 min. Centrifuge the suspension for 5 min and use the supernatant.

Column:
- *size*: *l* = 0.25 m, Ø = 3 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R*, *water R* (50:50 *V/V*);
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	100	0
20 - 20.5	100 → 0	0 → 100
20.5 - 30	0	100

After elution of the components, a gradient is applied to prevent a strong drifting baseline due to cholesterol during the following run.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 100 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Retention time Impurity A = about 10 min; impurity B = about 18 min.

Limits:
— *impurity A*: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.05 ppm);
— *impurity B*: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.05 ppm).

Loss on drying (*2.2.32*)
Maximum 0.1 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

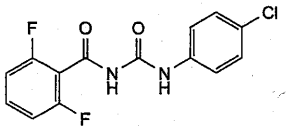
Bacterial endotoxins (*2.6.14*)
Less than 0.1 IU/mg.

ASSAY
Gas chromatography (*2.2.28*) as described in the test for other sterols.

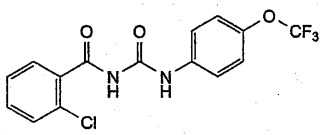
Calculate the percentage content of $C_{27}H_{46}O$ from the declared content of *cholesterol CRS*.

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B.*



A. 1-(4-chlorophenyl)-3-(2,6-difluorobenzoyl)urea (diflubenzuron),



B. 1-(2-chlorobenzoyl)-3-[(4-trifluoromethoxy)phenyl]urea (triflumuron).

Ph Eur

Choline Salicylate Solution

Action and use
Salicylate; non-selective cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Choline Salicylate Ear Drops
Choline Salicylate Oromucosal Gel

DEFINITION
Choline Salicylate Solution is an aqueous solution of choline salicylate. It contains not less than 47.5% w/v and not more than 52.5% w/v of choline salicylate, $C_{12}H_{19}NO_4$. It may contain a suitable antimicrobial preservative.

CHARACTERISTICS
A clear, colourless liquid.

IDENTIFICATION
A. Mix 0.5 mL with 10 mL of *methanol*, dry with *anhydrous sodium sulfate*, filter and evaporate the *methanol* to dryness. The *infrared absorption spectrum* of the residue, Appendix II A, is concordant with the *reference spectrum* of choline salicylate *(RS 059)*.

B. Dilute 5 mL to 25 mL with *water*. The resulting solution yields the reactions characteristic of *salicylates*, Appendix VI.

TESTS
Acidity
Dilute 4 mL to 20 mL with *water* and add 0.1 mL of *phenol red solution*. The solution is yellow and not more than 0.4 mL of *0.1M sodium hydroxide VS* is required to change the colour of the solution to reddish violet.

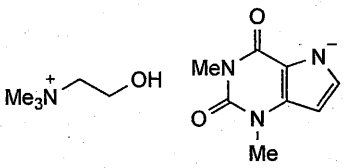
Clarity and colour of solution
Dilute 1 volume of the solution to 5 volumes with *water*. The resulting solution is *clear*, Appendix IV A, and *colourless*, Appendix IV B, Method II.

Weight per mL
1.070 to 1.110 g, Appendix V G.

Chloride
Mix 0.2 mL with 10 mL of *water* and add carefully, with mixing, 0.1 mL of a mixture of 10 volumes of *silver nitrate solution* and 1 volume of *nitric acid*. The resulting solution is not more opalescent than a standard prepared by treating 10 mL of a 0.00164% w/v solution of *sodium chloride* in the same manner beginning at the words 'add carefully ...' (0.1%).

ASSAY
To 1 g add 50 mL of *1,4-dioxan* and 5 mL of *acetic anhydride* and carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.25 mL of *methyl orange-xylene cyanol FF solution* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 24.13 mg of $C_{12}H_{19}NO_4$. Use the *weight per mL* to calculate the percentage of $C_{12}H_{19}NO_4$, weight in volume.

Choline Theophyllinate



$C_{12}H_{21}N_5O_3$ 283.3 *4499-40-5*

Action and use
Non-selective phosphodiesterase inhibitor (xanthine); treatment of reversible airways obstruction.

Preparation
Choline Theophyllinate Tablets

DEFINITION
Choline Theophyllinate is choline 1,2,3,6-tetrahydro-1,3-dimethyl-2,6-dioxo-7*H*-purin-7-ide. It contains not less than 41.9% and not more than 43.6% of choline, $C_5H_{15}NO_2$, and not less than 61.7% and not more than 65.5% of theophylline, $C_7H_9N_4O_2$, each calculated with reference to the dried substance.

CHARACTERISTICS
A white, crystalline powder. It melts between 187° and 192°, Appendix V A.

Very soluble in *water*; soluble in *ethanol (96%)*; very slightly soluble in *ether*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in 0.01M *sodium hydroxide* exhibits a maximum only at 275 nm. The *absorbance* at 275 nm is about 0.83.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of choline theophyllinate *(RS 060)*.

TESTS
Clarity and colour of solution
50 mL of a 10% w/v solution is *clear*, Appendix IV A, and not more intensely coloured than *reference solution* GY_4, Appendix IV B, Method I.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions of the substance being examined in *ethanol (96%)*.

(1) 1.0% w/v of the substance being examined.

(2) 0.010% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating *silica gel* HF_{254}.

(b) Use the mobile phase as described below.

(c) Apply 5 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, and examine under *ultraviolet light (254 nm)*.

MOBILE PHASE

5 volumes of *ethanol (96%)* and 95 volumes of *chloroform*.

LIMITS

Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (1%).

Loss on drying
When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
For choline
Dissolve 0.6 g in 50 mL of *water* and titrate with *0.05M sulfuric acid VS*, using *methyl red mixed solution* as indicator, until a violet end point is obtained. Each mL of *0.05M sulfuric acid VS* is equivalent to 12.12 mg of choline, $C_5H_{15}NO_2$.

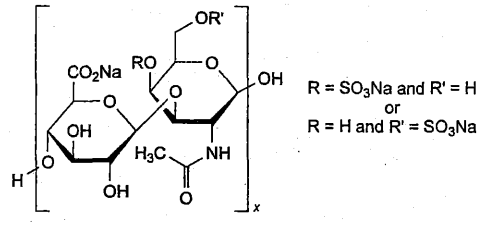
For theophylline
To the solution obtained in the Assay for choline, add 25 mL of *0.1M silver nitrate VS* and warm on a water bath for 15 minutes. Cool in ice for 30 minutes, filter and wash the residue with three 10 mL quantities of *water*. Titrate the combined filtrate and washings with *0.1M sodium hydroxide VS*. Each mL of *0.1M sodium hydroxide VS* is equivalent to 18.02 mg of theophylline, $C_7H_8N_4O_2$.

STORAGE
Choline Theophyllinate should be protected from light.

Chondroitin Sulfate Sodium

Chondroitin Sulphate Sodium

(Ph. Eur. monograph 2064)



$H_2O(C_{14}H_{19}NNa_2O_{14}S)_x$

Action and use
Acid mucopolysaccharide; treatment of osteoarthritis.

Ph Eur

DEFINITION
Natural copolymer based mainly on the 2 disaccharides: [4)-(β-D-glucopyranosyluronic acid)-(1→3)-[2-(acetylamino)-2-deoxy-β-D-galactopyranosyl 4-sulfate]-(1→] and [4)-(β-D-glucopyranosyluronic acid)-(1→3)-[2-(acetylamino)-2-deoxy-β-D-galactopyranosyl 6-sulfate]-(1→], sodium salt.
On complete hydrolysis it liberates D-galactosamine, D-glucuronic acid, acetic acid and sulfuric acid. It is obtained from cartilage of both terrestrial and marine origins.
Depending on the animal species of origin, it shows different proportions of 4-sulfate and 6-sulfate groups.

Content
95 per cent to 105 per cent (dried substance).

PRODUCTION
The animals from which chondroitin sulfate sodium is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS
Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry *(2.2.24)*.

Preparation Discs of *potassium bromide R*.

Comparison For chondroitin sulfate sodium of terrestrial origin use *chondroitin sulfate sodium CRS* and for chondroitin sulfate sodium of marine origine use *chondroitin sulfate sodium (marine) CRS*.

B. Solution S1 (see Tests) gives reaction (b) of sodium *(2.3.1)*.

C. Examine the electropherograms obtained in the test for related substances.

Results The principal band in the electropherogram obtained with the test solution is similar in position to the principal band in the electropherogram obtained with reference solution (a).

TESTS
Solution S1
Dissolve 2.500 g in 50.0 mL of *carbon dioxide-free water R*.

Solution S2
Dilute 1.0 mL of solution S1 to 10.0 mL with *water R*.

pH (*2.2.3*)
5.5 to 7.5 for solution S1.

Specific optical rotation (*2.2.7*)
−20 to −30 (terrestrial origin) or −12 to −19 (marine origin) (dried substance), determined on solution S1.

Intrinsic viscosity
0.01 m^3/kg to 0.15 m^3/kg.

Test solution (a) Weigh 5.000 g (m_{0p}) of the substance to be examined and add about 80 mL of an 11.7 g/L solution of *sodium chloride R* at room temperature. Dissolve by shaking at room temperature for 30 min. Dilute to 100.0 mL with an 11.7 g/L solution of *sodium chloride R*. Filter through a membrane filter (nominal pore size 0.45 µm) and discard the first 10 mL. The concentration of test solution (a) is only indicative and must be adjusted after an initial measurement of the viscosity of test solution (a).

Test solution (b) To 15.0 mL of test solution (a) add 5.0 mL of an 11.7 g/L solution of *sodium chloride R*.

Test solution (c) To 10.0 mL of test solution (a) add 10.0 mL of an 11.7 g/L solution of *sodium chloride R*.

Test solution (d) To 5.0 mL of test solution (a) add 15.0 mL of an 11.7 g/L solution of *sodium chloride R*.

Determine the flow-time (*2.2.9*) for an 11.7 g/L solution of *sodium chloride R* (t_0) and the flow times for the 4 test solutions (t_1, t_2, t_3 and t_4), at 25.00 ± 0.03 °C. Use an appropriate suspended level viscometer (specifications: viscometer constant = about 0.005 mm^2/s^2, kinematic viscosity range = 1-5 mm^2/s, internal diameter of tube *R* = 0.53 mm, volume of bulb *C* = 5.6 mL, internal diameter of tube *N* = 2.8-3.2 mm) with a funnel-shaped lower capillary end. Use the same viscometer for all measurements; measure all outflow times in triplicate. The test is not valid unless the results do not differ by more than 0.35 per cent from the mean and if the flow time t_1 is not less than 1.6 × t_0 and not more than 1.8 × t_0. If this is not the case, adjust the concentration of test solution (a) and repeat the procedure.

Calculation of the relative viscosities

Since the densities of the chondroitin sulfate solutions and of the solvent are almost equal, the relative viscosities η_{ri} (being η_{r1}, η_{r2}, η_{r3} and η_{r4}) can be calculated from the ratio of the flow times for the respective solutions t_i (being t_1, t_2, t_3 and t_4) to the flow time of the solvent t_0, but taking into account the kinetic energy correction factor for the capillary (B = 30 800 s^3), as shown below:

$$\frac{t_i - \frac{B}{t_i^2}}{t_0 - \frac{B}{t_0^2}}$$

Calculation of the concentrations

Calculate the concentration c_1 (expressed in kg/m^3) of chondroitin sulfate sodium in test solution (a) using the following expression:

$$m_{0p} \times \frac{x}{100} \times \frac{100 - h}{100} \times 10$$

x = percentage content of chondroitin sulfate sodium as determined in the assay;
h = loss on drying as a percentage.

Calculate the concentration c_2 (expressed in kg/m^3) of chondroitin sulfate sodium in test solution (b) using the following expression:

$$c_1 \times 0.75$$

Calculate the concentration c_3 (expressed in kg/m^3) of chondroitin sulfate sodium in test solution (c) using the following expression:

$$c_1 \times 0.50$$

Calculate the concentration c_4 (expressed in kg/m^3) of chondroitin sulfate sodium in test solution (d) using the following expression:

$$c_1 \times 0.25$$

Calculation of the intrinsic viscosity

The specific viscosity η_{si} of the test solution (being η_{s1}, η_{s2}, η_{s3} and η_{s4}) is calculated from the relative viscosities η_{ri} (being η_{r1}, η_{r2}, η_{r3} and η_{r4}) according to the following expression:

$$\eta_{ri} - 1$$

The intrinsic viscosity [η], defined as

$$[\eta] = \lim_{c \to 0} \left(\frac{\eta_s}{c}\right)$$

is calculated by linear least-squares regression analysis using the following equation:

$$\frac{\eta_{si}}{c_i} = c_i \times k_H + [\eta]$$

c_i = concentration of the substance to be examined expressed in kg/m^3;
k_H = Huggins' constant.

Related substances
Electrophoresis (*2.2.31*).

Buffer solution A (0.1 M barium acetate pH 5.0). Dissolve 25.54 g of *barium acetate R* in 900 mL of *water R*. Adjust to pH 5.0 with *glacial acetic acid R* and dilute to 1000.0 mL with *water R*.

Buffer solution B (1 M barium acetate pH 5.0). Dissolve 255.43 g of *barium acetate R* in 900 mL of *water R*. Adjust to pH 5.0 with *glacial acetic acid R* and dilute to 1000.0 mL with *water R*.

Staining solution Dissolve 1.0 g of *toluidine blue R* and 2.0 g of *sodium chloride R* in 1000 mL of *0.01 M hydrochloric acid*. Filter.

Test solution Prepare a 30 mg/mL solution of the substance to be examined in *water R*.

Reference solution (a) Prepare a 30 mg/mL solution of *chondroitin sulfate sodium CRS* in *water R*.

Reference solution (b) Dilute 2.0 mL of reference solution (a) to 100.0 mL with *water R*.

Reference solution (c) Mix equal volumes of reference solution (b) and *water R*.

Procedure Allow the electrophoresis support to cool the plate to 10 °C. Pre-equilibrate the agarose gel for 1 min in buffer solution A. Remove excess liquid by careful decanting. Dry the gel for approximately 5 min. Place 400 mL of buffer solution B into each of the containers of the electrophoresis equipment. Transfer 1 µL of each solution to the slots of the agarose gel. Pipette a few millilitres of a 50 per cent *V/V* solution of *glycerol R* onto the cooled plate of the

electrophoresis equipment and place the gel in the middle of the ceramic plate. Place a wick, saturated with buffer solution B, at the positive and negative sides of the agarose gel. Ensure that there is good contact between the electrophoresis buffer and the agarose gel. Perform the electrophoresis under the following conditions: 75 mA/gel, resulting in a voltage of 100-150 V (maximum 300-400 V) for a gel of about 12 cm × 10 cm. Carry out the electrophoresis for 12 min. Place the gel in a mixture consisting of 10 volumes of *anhydrous ethanol R* and 90 volumes of buffer solution A for 2 min. Carry out the electrophoresis for 20 min. Place the gel in a mixture consisting of 30 volumes of *anhydrous ethanol R* and 70 volumes of buffer solution A for 2 min. Carry out the electrophoresis for 20 min. Stain the gel in the staining solution for 10 min. Destain the gel for 15 min under running tap water followed by 10-15 min with *water R* until the band in the electropherogram obtained with reference solution (c) is visible. Allow the gel to dry.

System suitability:
- the electropherogram obtained with reference solution (c) shows a visible band;
- the band in the electropherogram obtained with reference solution (b) is clearly visible and similar in position to the band in the electropherogram obtained with reference solution (a).

Results Any secondary band in the electropherogram obtained with the test solution is not more intense than the band in the electropherogram obtained with reference solution (b) (2 per cent).

Protein (*2.5.33, Method 2*)
Maximum 3.0 per cent (dried substance).

Test solution Dilute 1.0 mL of solution S1 to 50.0 mL with *0.1 M sodium hydroxide*.

Reference solutions Dissolve about 0.100 g of *bovine albumin R*, accurately weighed, in *0.1 M sodium hydroxide* and dilute to 50.0 mL with the same solvent. Carry out all additional dilutions using *0.1 M sodium hydroxide*.

Chlorides (*2.4.4*)
Maximum 0.5 per cent.

Dilute 1 mL of solution S2 to 15 mL with *water R*. Do not add diluted nitric acid. Prepare the standard using 5 mL of *chloride standard solution (5 ppm Cl) R* and 10 mL of *water R*.

Loss on drying (*2.2.32*)
Maximum 12.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Staphylococcus aureus* (*2.6.13*).

Absence of *Pseudomonas aeruginosa* (*2.6.13*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

Absence of bile-tolerant gram-negative bacteria (*2.6.13*).

ASSAY

Test solution (a) Weigh 0.100 g (m_1) of the substance to be examined, dissolve in *water R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with *water R*.

Reference solution (a) Weigh 0.100 g (m_0) of *chondroitin sulfate sodium CRS*, previously dried as described in the test for loss on drying, dissolve in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with *water R*.

Titrant solution (a) Weigh 4.000 g of *cetylpyridinium chloride monohydrate R* and dilute to 1000 mL with *water R*.

Titrant solution (b) Weigh 1.000 g of *cetylpyridinium chloride monohydrate R* and dilute to 1000 mL with *water R*.

Perform either visual or photometric titration as follows:

Visual titration Titrate 40.0 mL of reference solution (a) and 40.0 mL of test solution (a) with titrant solution (a). The solution becomes turbid. At the end point, the liquid appears clear, with an almost-white precipitate in suspension. The precipitate is more apparent if 0.1 mL of a 1 per cent solution of *methylene blue R* is added before starting the titration. The precipitated particles are more apparent against the blue background.

Photometric titration Titrate 50.0 mL of reference solution (b) and 50.0 mL of test solution (b) with titrant solution (b). To determine the end point, use a suitable autotitrator equipped with a phototrode at a suitable wavelength (none is critical) in the visible range.

Calculate the percentage content of chondroitin sulfate sodium using the following expression:

$$\frac{v_1 \times m_0}{v_0 \times m_1} \times \frac{100}{100 - h} \times Z$$

v_0 = volume of appropriate titrant solution when titrating the appropriate reference solution, in millilitres;
v_1 = volume of appropriate titrant solution when titrating the appropriate test solution, in millilitres;
h = loss on drying of the substance to be examined, as a percentage;
Z = percentage content of $H_2O(C_{14}H_{19}NNa_2O_{14}S)_x$ in *chondroitin sulfate sodium CRS*.

STORAGE
In an airtight container, protected from light.

LABELLING
The label states the origin of the substance (marine or terrestrial).

Ph Eur

Chorionic Gonadotrophin

(Ph. Eur. monograph 0498)

Action and use
Gonadotrophic hormone.

Preparation
Chorionic Gonadotrophin Injection

Ph Eur

DEFINITION
Chorionic gonadotrophin is a dry preparation of placental glycoproteins which have luteinising activity. It is extracted from the urine of pregnant women. The potency is not less than 2500 IU/mg.

PRODUCTION
Chorionic gonadotrophin is extracted using a suitable fractionation procedure. It is either dried under reduced pressure or freeze-dried.

CHARACTERS

Appearance
White or yellowish-white, amorphous powder.

Solubility
Soluble in water.

IDENTIFICATION

When administered to immature rats as prescribed in the assay, it causes an increase in the mass of the seminal vesicles and of the prostate gland.

TESTS

Water (*2.5.32*)
Maximum 5.0 per cent.

Bacterial endotoxins (*2.6.14*)
Less than 0.02 IU per IU of chorionic gonadotrophin, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

The potency of chorionic gonadotrophin is estimated by comparing under given conditions its effect of increasing the mass of the seminal vesicles (or the prostate gland) of immature rats with the same effect of the International Standard of chorionic gonadotrophin or of a reference preparation calibrated in International Units.

The International Unit is the activity contained in a stated amount of the International Standard, which consists of a mixture of a freeze-dried extract of chorionic gonadotrophin from the urine of pregnant women with lactose.
The equivalence in International Units of the International Standard is stated by the World Health Organization.

Use immature male rats of the same strain, 19 to 28 days old, differing in age by not more than 3 days and having body masses such that the difference between the heaviest and the lightest rat is not more than 10 g. Assign the rats at random to 6 equal groups of at least 5 animals. If sets of 6 litter mates are available, assign one litter mate from each set to each group and mark according to litter.

Choose 3 doses of the reference preparation and 3 doses of the preparation to be examined such that the smallest dose is sufficient to produce a positive response in some of the rats and the largest dose does not produce a maximal response in all the rats. Use doses in geometric progression and as an initial approximation total doses of 4 IU, 8 IU and 16 IU may be tried although the dose will depend on the sensitivity of the animals used, which may vary widely.

Dissolve separately the total quantities of the preparation to be examined and of the reference preparation corresponding to the daily doses to be used in sufficient *phosphate-albumin buffered saline pH 7.2 R* such that the daily dose is administered in a volume of about 0.5 mL. Add a suitable antimicrobial preservative such as 4 g/L of phenol or 0.02 g/L of thiomersal. Store the solutions at 5 ± 3 °C.

Inject subcutaneously into each rat the daily dose allocated to its group, on 4 consecutive days at the same time each day. On the 5th day, about 24 h after the last injection, euthanise the rats and remove the seminal vesicles. Remove any extraneous fluid and tissue and weigh the vesicles immediately. Calculate the results by the usual statistical methods, using the mass of the vesicles as the response. (The precision of the assay may be improved by a suitable correction of the organ mass with reference to the body mass of the animal from which it was taken; an analysis of covariance may be used).

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits (P = 0.95) of the estimated potency are not less than 64 per cent and not more than 156 per cent of the stated potency.

STORAGE

In an airtight, tamper-proof container, protected from light at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

The label states:
— the number of International Units per container,
— the potency in International Units per milligram.

Ph Eur

Chymotrypsin

(Ph. Eur. monograph 0476)

9004-07-3

Action and use
Proteolytic enzyme.

Ph Eur

DEFINITION

Chymotrypsin is a proteolytic enzyme obtained by the activation of chrymotrypsinogen extracted from the pancreas of beef (*Bos taurus* L.). It has an activity of not less than 5.0 microkatals per milligram. In solution it has maximal enzymic activity at about pH 8; the activity is reversibly inhibited at pH 3, the pH at which it is most stable.

PRODUCTION

The animals from which chymotrypsin is derived must fulfil the requirements for the health of animals suitable for human consumption. Furthermore, the tissues used shall not include any specified risk material as defined by any relevant international or, where appropriate, national legislation.

The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Histamine (*2.6.10*)
Not more than 1 µg (calculated as histamine base) per 5 microkatals of chymotrypsin activity. Before carrying out the test, heat the solution of the substance to be examined on a water-bath for 30 min.

CHARACTERS

Appearance
White or almost white, crystalline or amorphous powder, hygroscopic if amorphous.

Solubility
Sparingly soluble in water.

IDENTIFICATION

A. Dilute 1 mL of solution S (see Tests) to 10 mL with *water R*. In a depression in a white spot-plate, mix 0.05 mL of this solution with 0.2 mL of the substrate solution.
A purple colour develops.

Substrate solution To 24.0 mg of *acetyltyrosine ethyl ester R* add 0.2 mL of *ethanol (96 per cent) R* and swirl to dissolve. Add 2.0 mL of *0.067 M phosphate buffer solution pH 7.0 R* and 1 mL of *methyl red mixed solution R* and dilute to 10.0 mL with *water R*.

B. Dilute 0.5 mL of solution S to 5 mL with *water R*. Add 0.10 mL of a 20 g/L solution of *tosylphenylalanylchloromethane R* in *ethanol (96 per cent) R*. Adjust to pH 7.0 and shake for 2 h. In a depression in a white spot-plate, mix 0.05 mL of this solution with 0.2 mL of the substrate solution (see Identification test A). No colour develops within 3 min of mixing.

TESTS

Solution S

Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*).

pH (*2.2.3*)

3.0 to 5.0 for solution S.

Specific absorbance (*2.2.25*)

18.5 to 22.5, determined at the absorption maximum at 281 nm; maximum 8, determined at the absorption minimum at 250 nm.

Dissolve 30.0 mg in *0.001 M hydrochloric acid* and dilute to 100.0 mL with the same acid.

Trypsin

Substrate solution To 98.5 mg of *tosylarginine methyl ester hydrochloride R*, suitable for assaying trypsin, add 5 mL of *tris (hydroxymethyl) aminomethane buffer solution pH 8.1 R* and swirl to dissolve. Add 2.5 mL of *methyl red mixed solution R* and dilute to 25.0 mL with *water R*.

Test solution Transfer to a depression in a white spot-plate 0.01 mL of *tris(hydroxymethyl)aminomethane buffer solution pH 8.1 R* and 0.1 mL of solution S. Add 0.2 mL of the substrate solution.

Reference solution At the same time and in the same manner as for the test solution, prepare a solution using the substance to be examined to which not more than 1 per cent *m/m* of *trypsin BRP* has been added.

Start a timer. No colour appears in the test solution within 3-5 min after the addition of the substrate solution. A purple colour is produced in the control solution.

Loss on drying (*2.2.32*)

Not more than 5.0 per cent, determined on 0.100 g by drying at 60 °C at a pressure not exceeding 0.7 kPa for 2 h.

ASSAY

The activity of chymotrypsin is determined by comparing the rate at which it hydrolyses *acetyltyrosine ethyl ester R* with the rate at which *chymotrypsin BRP* hydrolyses the same substrate under the same conditions.

Apparatus Use a reaction vessel of about 30 mL capacity provided with:

- a device that will maintain a temperature of 25.0 ± 0.1 °C;
- a stirring device, for example a magnetic stirrer;
- a lid with holes for the insertion of electrodes, the tip of a burette, a tube for the admission of nitrogen and the introduction of reagents.

An automatic or manual titration apparatus may be used. For the latter, the burette is graduated in 0.005 mL and the pH meter is provided with a wide-range scale and glass-silver-silver chloride or other suitable electrodes.

Test solution Dissolve 25.0 mg of the substance to be examined in *0.001 M hydrochloric acid* and dilute to 250.0 mL with the same acid.

Reference solution Dissolve 25.0 mg of *chymotrypsin BRP* in *0.001 M hydrochloric acid* and dilute to 250.0 mL with the same acid.

Store the solutions at 0-5 °C. Warm 1 mL of each solution to about 25 °C over 15 min and use 50 µL of each solution (corresponding to about 25 nanokatals) for each titration. Carry out the titration in an atmosphere of nitrogen. Transfer 10.0 mL of 0.01 M calcium chloride solution R to the reaction vessel and, while stirring, add 0.35 mL of *0.2 M acetyltyrosine ethyl ester R*. When the temperature is steady at 25.0 ± 0.1 °C (after about 5 min), adjust to pH 8.0 exactly with *0.02 M sodium hydroxide*. Add 50 µL of the test solution (equivalent to about 5 µg of the substance to be examined) and start a timer. Maintain at pH 8.0 by the addition of *0.02 M sodium hydroxide*, noting the volume added every 30 s. Calculate the volume of *0.02 M sodium hydroxide* used per second between 30 s and 210 s. Carry out a titration in the same manner using the reference solution and calculate the volume of *0.02 M sodium hydroxide* used per second.

Calculate the activity in microkatals per milligram using the following expression:

$$\frac{m' \times V}{m \times V'} \times A$$

m	=	mass of the substance to be examined, in milligrams;
m'	=	mass of *chymotrypsin BRP*, in milligrams;
V	=	volume of *0.02 M sodium hydroxide* used per second by the test solution;
V'	=	volume of *0.02 M sodium hydroxide* used per second by the reference solution;
A	=	activity of *chymotrypsin BRP*, in microkatals per milligram.

STORAGE

In an airtight container at 2 °C to 8 °C, protected from light.

LABELLING

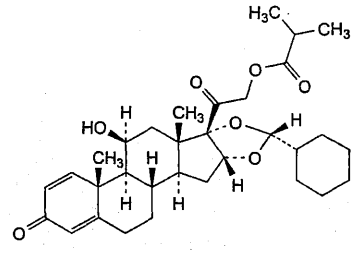
The label states:

- the quantity of chymotrypsin and the total activity in microkatals per container;
- for the amorphous substance, that it is hygroscopic.

Ph Eur

Ciclesonide

(Ph. Eur. monograph 2703)



$C_{32}H_{44}O_7$ 540.7 *126544-47-6*

Action and use

Glucocorticoid.

Ph Eur

DEFINITION

(2′*R*)-2′-Cyclohexyl-11β-hydroxy-3,20-dioxo-16β*H*-[1,3]dioxolo[4′,5′:16,17]pregna-1,4-dien-21-yl 2-methylpropanoate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or yellowish-white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble to soluble in acetone and in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciclesonide CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *anhydrous ethanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *ciclesonide CRS* in *anhydrous ethanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dissolve 3 mg of *ciclesonide impurity B CRS*, 3 mg of *ciclesonide impurity C CRS* and 5 mg of *ciclesonide containing impurity A CRS* in *anhydrous ethanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (c) Dissolve 50 mg of the substance to be examined in *anhydrous ethanol R*, add 1.0 mL of reference solution (b) and dilute to 50.0 mL with *anhydrous ethanol R*.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with *anhydrous ethanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 60 °C.

Mobile phase *water R, anhydrous ethanol R* (38:62 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 243 nm.

Injection 20 µL of the test solution and reference solutions (c) and (d).

Run time 2.2 times the retention time of ciclesonide.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to ciclesonide (retention time = about 16 min): impurity B = about 0.4; impurity C = about 0.9; impurity A = about 1.4.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity C and ciclesonide.

Calculation of percentage contents:
— for each impurity, use the concentration of ciclesonide in reference solution (d).

Limits:
— *impurity A*: maximum 1.0 per cent;
— *impurities B, C*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total of unspecified impurities*: maximum 0.2 per cent;
— *total*: maximum 1.2 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

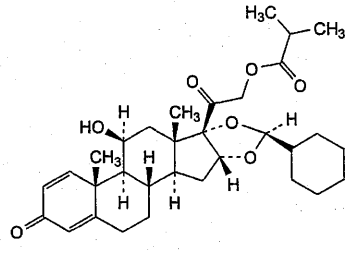
Run time 1.6 times the retention time of ciclesonide.

System suitability Reference solution (a):
— *symmetry factor*: maximum 2.2 for the peak due to ciclesonide.

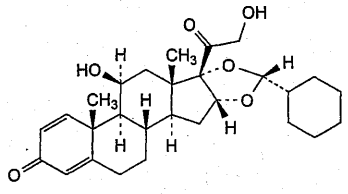
Calculate the percentage content of $C_{32}H_{44}O_7$ taking into account the assigned content of *ciclesonide CRS*.

IMPURITIES

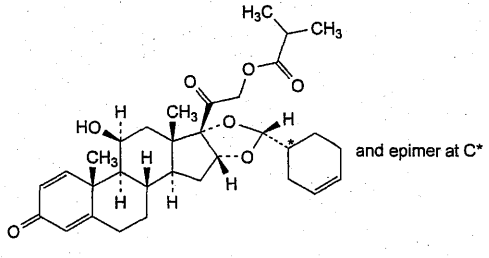
Specified impurities *A, B, C.*



A. (2′*S*)-2′-cyclohexyl-11β-hydroxy-3,20-dioxo-16β*H*-[1,3]dioxolo[4′,5′:16,17]pregna-1,4-dien-21-yl 2-methylpropanoate (*S*-epimer of ciclesonide),



B. (2′*R*)-2′-cyclohexyl-11β,21-dihydroxy-16β*H*-[1,3]dioxolo[4′,5′:16,17]pregna-1,4-diene-3,20-dione,

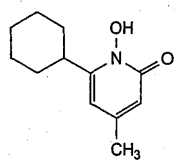


C. (2′*R*)-2′-[(1*RS*)-cyclohex-3-enyl]-11β-hydroxy-3,20-dioxo-16β*H*-[1,3]dioxolo[4′,5′:16,17]pregna-1,4-dien-21-yl 2-methylpropanoate.

Ph Eur

Ciclopirox

(Ph. Eur. monograph 1407)



$C_{12}H_{17}NO_2$ 207.3 *29342-05-0*

Action and use
Antifungal.

Ph Eur

DEFINITION
6-Cyclohexyl-1-hydroxy-4-methylpyridin-2(1*H*)-one.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish-white, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 140 °C to 145 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciclopirox CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *ciclopirox CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Pretreatment Before use, predevelop with the mobile phase until the solvent front has migrated to the top of the plate. Allow to dry in air for 5 min.

Mobile phase *concentrated ammonia R, water R, ethanol (96 per cent) R* (10:15:75 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air for 10 min.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Spray with a 20 g/L solution of *ferric chloride R* in *anhydrous ethanol R.*

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 2.0 g in *methanol R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test avoiding exposure to actinic light. All materials in direct contact with the substance to be examined like column materials, reagents, solvents, etc. should contain only very low amounts of extractable metal cations.*

Solvent mixture *acetonitrile R,* mobile phase (10:90 *V/V*).

Test solution Dissolve 30.0 mg of the substance to be examined in 15 mL of the solvent mixture, using an ultrasonic bath if necessary, and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 15.0 mg of *ciclopirox impurity A CRS* and 15.0 mg of *ciclopirox impurity B CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 200.0 mL with the solvent mixture.

Reference solution (c) Dilute 2.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Mix 5.0 mL of reference solution (a) and 5.0 mL of the test solution.

Column:
— *size: l* = 0.08 m, Ø = 4 mm;
— *stationary phase: nitrile silica gel for chromatography R2* (5 µm).

In order to ensure desorption of interfering metal ions, every new column is to be rinsed with the rinsing solution over a period of not less than 15 h and then with the mobile phase for not less than 5 h at a flow rate of 0.2 mL/min.

Rinsing solution *glacial acetic acid R, acetylacetone R, acetonitrile R, water R* (0.1:0.1:50:50 *V/V/V/V*).

Mobile phase *glacial acetic acid R, acetonitrile R,* 0.96 g/L solution of *sodium edetate R* (0.01:23:77 *V/V/V*).

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 220 nm and at 298 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d); inject the solvent mixture as a blank.

Run time 2.5 times the retention time of ciclopirox.

Retention time Ciclopirox = 8 min to 11 min; if necessary adjust the ratio of the 0.96 g/L solution of sodium edetate to acetonitrile in the mobile phase.

Relative retention With reference to ciclopirox: impurity A = about 0.5; impurity C = about 0.9; impurity B = about 1.3.

System suitability At 298 nm:
— *resolution*: minimum 2.0 between the peaks due to ciclopirox and impurity B in the chromatogram obtained with reference solution (d);
— *symmetry factor*: 0.8 to 2.0 for the principal peak in the chromatogram obtained with the test solution.

Limits:
— *impurity A at 220 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurities B, C at 298 nm*: for each impurity, not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities at 298 nm*: for each impurity, not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *sum of impurities other than B at 298 nm*: not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit at 298 nm*: 0.5 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C over *diphosphorus pentoxide R*.

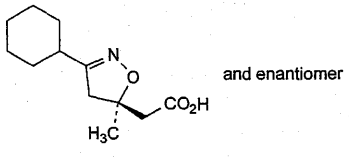
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 20 mL of *methanol R*. Add 20 mL of *water R* and titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

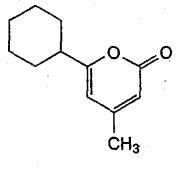
1 mL of *0.1 M sodium hydroxide* is equivalent to 20.73 mg of $C_{12}H_{17}NO_2$.

STORAGE
Protected from light.

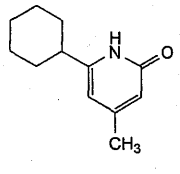
IMPURITIES
Specified impurities *A, B, C.*



A. [(5*RS*)-3-cyclohexyl-5-methyl-4,5-dihydro-1,2-oxazol-5-yl] acetic acid,



B. 6-cyclohexyl-4-methyl-2*H*-pyran-2-one,

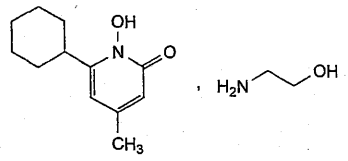


C. 6-cyclohexyl-4-methylpyridin-2(1*H*)-one.

Ph Eur

Ciclopirox Olamine

(Ph. Eur. monograph 1302)



$C_{14}H_{24}N_2O_3$ 268.4 *41621-49-2*

Action and use
Antifungal.

Ph Eur

DEFINITION
6-Cyclohexyl-1-hydroxy-4-methylpyridin-2(1*H*)-one and 2-aminoethanol.

Content
— *ciclopirox* ($C_{12}H_{17}NO_2$; M_r 207.3): 76.0 per cent to 78.5 per cent (dried substance);
— *2-aminoethanol* (C_2H_7NO; M_r 61.1): 22.2 per cent to 23.3 per cent (dried substance).

CHARACTERS
Appearance
White or pale yellow, crystalline powder.

Solubility
Sparingly soluble in water, very soluble in ethanol (96 per cent) and in methylene chloride, slightly soluble in ethyl acetate, practically insoluble in cyclohexane.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A.

Second identification: B.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciclopirox olamine CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethyl acetate R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 25 mg of *ciclopirox olamine CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Pretreatment Before use, predevelop 2 plates with the mobile phase until the solvent front has migrated to the top of the plates. Allow to dry in air for 5 min.

Mobile phase *concentrated ammonia R, water R, anhydrous ethanol R* (10:15:75 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air for 10 min.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with the reference solution.

Detection B Spray 1 plate with *ferric chloride solution R3*.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

Detection C Spray the 2nd second plate with *ninhydrin solution R*. Heat at 110 °C until the spots appear.

Results C The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 2.0 g in *methanol R* and dilute to 20 mL with the same solvent.

pH (*2.2.3*)

8.0 to 9.0.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test avoiding exposure to actinic light. All materials in direct contact with the substance to be examined, such as column materials, reagents, solvents, etc. should contain only small amounts of extractable metal cations.*

Solvent mixture *acetonitrile R*, mobile phase (10:90 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined (corresponding to about 30 mg of ciclopirox) in a mixture of 20 µL of *anhydrous acetic acid R*, 2 mL of *acetonitrile R*, and 15 mL of the mobile phase, using an ultrasonic bath if necessary. Dilute the solution to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 15.0 mg of *ciclopirox impurity A CRS* and 15.0 mg of *ciclopirox impurity B CRS* in a mixture of 1 mL of *acetonitrile R* and 7 mL of the mobile phase, and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 200.0 mL with the solvent mixture.

Reference solution (c) Dilute 2.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Mix 5.0 mL of reference solution (a) and 5.0 mL of the test solution.

Column:
- *size*: *l* = 80 mm, Ø = 4 mm;
- *stationary phase*: *nitrile silica gel for chromatography R* (5 µm).

In order to ensure desorption of interfering metal ions, every new column is to be rinsed with the rinsing solution over a period of not less than 15 h and then with the mobile phase for not less than 5 h at a flow rate of 0.2 mL/min.

Rinsing solution *acetylacetone R, anhydrous acetic acid R, acetonitrile R, water R* (0.1:0.1:50:50 *V/V/V/V*).

Mobile phase *anhydrous acetic acid R, acetonitrile R*, 0.96 g/L solution of *sodium edetate R* (0.01:23:77 *V/V/V*).

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 220 nm and at 298 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Run time 2.5 times the retention time of ciclopirox.

Retention time Ciclopirox = 8 min to 11 min; if necessary adjust the ratio of the 0.96 g/L solution of sodium edetate to acetonitrile in the mobile phase.

Relative retention With reference to ciclopirox: impurity A = about 0.5; impurity C = about 0.9; impurity B = about 1.3.

System suitability At 298 nm:
- *resolution*: minimum of 2.0 between the peaks due to impurity B and ciclopirox in the chromatogram obtained with reference solution (d);
- *symmetry factor*: 0.8 to 2.0 for the principal peak in the chromatogram obtained with the test solution.

Limits:
- *impurity A at 220 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurities B, C at 298 nm*: for each impurity, not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *unspecified impurities at 298 nm*: for each impurity, not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (c) (0.10 per cent);
- *sum of impurities other than B at 298 nm*: not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *disregard limit at 298 nm*: 0.5 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.5 per cent, determined on 1.000 g by drying under high vacuum.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

2-Aminoethanol

Dissolve 0.250 g in 25 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 6.108 mg of C_2H_7NO.

Ciclopirox

Dissolve 0.200 g in 2 mL of *methanol R*. Add 38 mL of *water R*, swirl and titrate immediately with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

Use *0.1 M sodium hydroxide*, the titre of which has been determined under the conditions prescribed above using 0.100 g of *benzoic acid RV*.

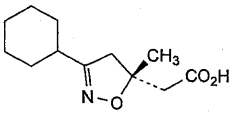
1 mL of *0.1 M sodium hydroxide* is equivalent to 20.73 mg of $C_{12}H_{17}NO_2$.

STORAGE

Protected from light.

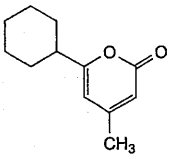
IMPURITIES

Specified impurities *A, B, C.*

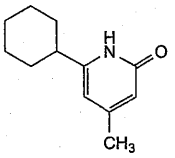


and enantiomer

A. [(5*RS*)-3-cyclohexyl-5-methyl-4,5-dihydro-1,2-oxazol-5-yl] acetic acid,



B. 6-cyclohexyl-4-methyl-2*H*-pyran-2-one,

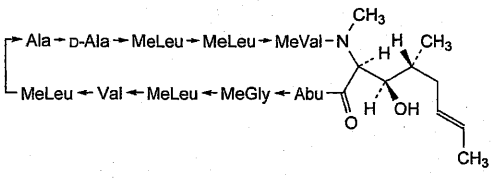


C. 6-cyclohexyl-4-methylpyridin-2(1*H*)-one.

Ph Eur

Ciclosporin

(Ph. Eur. monograph 0994)



$C_{62}H_{111}N_{11}O_{12}$ 1203 *59865-13-3*

Action and use
Calcineurin inhibitor; immunosuppressant.

Preparations
Ciclosporin Capsules
Ciclosporin Eye Drops
Sterile Ciclosporin Concentrate
Ciclosporin Oral Solution

Ph Eur

DEFINITION

Cyclo[[(2*S*,3*R*,4*R*,6*E*)-3-hydroxy-4-methyl-2-(methylamino)oct-6-enoyl]-L-2-aminobutanoyl-*N*-methylglycyl-*N*-methyl-L-leucyl-L-valyl-*N*-methyl-L-leucyl-L-alanyl-D-alanyl-*N*-methyl-L-leucyl-*N*-methyl-L-leucyl-*N*-methyl-L-valyl] (ciclosporin A).

Substance produced by *Beauveria nivea (Tolypocladium inflatum* Gams) or obtained by any other means.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciclosporin CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5, BY_5 or R_7 (*2.2.2, Method II*).

Dissolve 1.5 g in *anhydrous ethanol R* and dilute to 15 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−193 to −185 (dried substance).

Dissolve 0.125 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (50:50 *V/V*).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 30.0 mg of *ciclosporin CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dilute 2.0 mL of reference solution (a) to 200.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *ciclosporin for system suitability CRS* in 5.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3-5 µm);
— *temperature*: 80 °C.

The column is connected to the injection port by a steel capillary tube about 1 m long, having an internal diameter of 0.25 mm and maintained at 80 °C.

Mobile phase *phosphoric acid R, 1,1-dimethylethyl methyl ether R, acetonitrile R, water R* (0.1:5:43:52 *V/V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 1.7 times the retention time of ciclosporin.

System suitability Reference solution (c):
— *retention time*: ciclosporin = 25 min to 30 min; if necessary, adjust the ratio of acetonitrile to water in the mobile phase;
— *peak-to-valley ratio*: minimum 1.4, where H_p = height above the baseline of the peak due to ciclosporin U and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to ciclosporin; if necessary, adjust the ratio of 1,1-dimethylethyl methyl ether to acetonitrile in the mobile phase.

Limits:
— *any impurity*: for each impurity, not more than 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g at 60 °C at a pressure not exceeding 15 Pa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.84 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. Dissolve 50 mg of the substance to be examined in a mixture of 280 mg of *ethanol (96 per cent) R* and 650 mg of *polyoxyethylated castor oil R* and dilute to the required concentration using water for BET.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

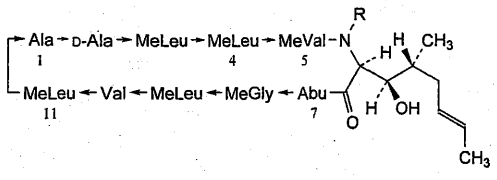
Injection Test solution and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

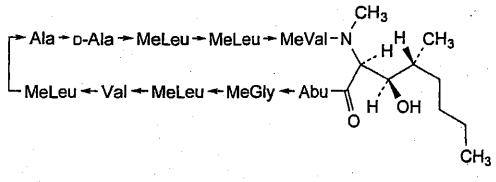
Calculate the percentage content of $C_{62}H_{111}N_{11}O_{12}$ taking into account the assigned content of *ciclosporin CRS*.

STORAGE
In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES



A. different ciclosporins [difference from ciclosporin (R = CH_3: ciclosporin A)]: ciclosporin B [7-L-Ala]; ciclosporin C [7-L-Thr]; ciclosporin D [7-L-Val]; ciclosporin E [5-L-Val]; ciclosporin G [7-(L-2-aminopentanoyl)]; ciclosporin H [5-D-MeVal]; ciclosporin L [R = H]; ciclosporin T [4-L-Leu]; ciclosporin U [11-L-Leu]; ciclosporin V [1-L-Abu],



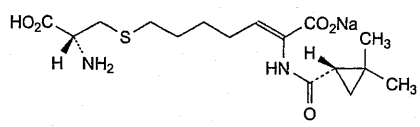
B. [6-[(2*S*,3*R*,4*R*)-3-hydroxy-4-methyl-2-(methylamino)octanoic acid]]ciclosporin A,

C. isociclosporin A.

Ph Eur

Cilastatin Sodium

(Ph. Eur. monograph 1408)



$C_{16}H_{25}N_2NaO_5S$ 380.4 *81129-83-1*

Action and use
Dehydropeptidase-I inhibitor; inhibition of the renal metabolism of imipenem.

Preparation
Cilastatin and Imipenem for Infusion

Ph Eur

DEFINITION
Sodium (*Z*)-7-[[(2*R*)-2-amino-2-carboxyethyl]sulfanyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-2-enoate.

Content
98.0 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS
Appearance
White or light yellow, amorphous, hygroscopic powder.

Solubility
Very soluble in water and in methanol, slightly soluble in anhydrous ethanol, very slightly soluble in dimethyl sulfoxide, practically insoluble in acetone and in methylene chloride.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cilastatin sodium CRS*.

C. It gives reaction (a) of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
6.5 to 7.5 for solution S.

Specific optical rotation (*2.2.7*)
+ 41.5 to + 44.5 (anhydrous substance).

Dissolve 0.250 g in a mixture of 1 volume of *hydrochloric acid R* and 120 volumes of *methanol R*, then dilute to 25.0 mL with the same mixture of solvents.

Related substances
Liquid chromatography (*2.2.29*). Prepare the solutions immediately before use.

Test solution Dissolve 32 mg of the substance to be examined in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 3 mg of *cilastatin for system suitability 1 CRS* (containing impurities A, B, E, F, G (epimer 2) and H) in *water R* and dilute to 2.0 mL with the same solvent.

Reference solution (c) Dissolve 3 mg of *cilastatin for system suitability 2 CRS* (containing impurities C and G (epimer 1)) in *water R* and dilute to 2.0 mL with the same solvent.

Reference solution (d) Dissolve 32 mg of *mesityl oxide R* (impurity D) in 100.0 mL of *water R*. Dilute 1.0 mL of the solution to 50.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: *phosphate buffer solution pH 3.25 R*;
- *mobile phase B*: *acetonitrile R1*, *phosphate buffer solution pH 3.25 R* (50:50 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 28	100 → 90	0 → 10
28 - 38	90	10
38 - 63	90 → 50	10 → 50
63 - 78	50 → 30	50 → 70
78 - 88	30	70

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *cilastatin for system suitability 1 CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, E, F, G (epimer 2) and H; use the chromatogram supplied with *cilastatin for system suitability 2 CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and G (epimer 1); use the chromatogram obtained with reference solution (d) to identify the peak due to impurity D.

Relative retention With reference to cilastatin (retention time = about 50 min): impurity E = about 0.2; impurity A (epimer 1) = about 0.60; impurity A (epimer 2) = about 0.62; impurity D = about 0.9; impurity F = about 0.98; impurity G (epimer 1) = about 1.02; impurity G (epimer 2) = about 1.05; impurity H = about 1.06; impurity B = about 1.17; impurity C = about 1.23.

System suitability:
- *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cilastatin in the chromatogram obtained with reference solution (b);
- *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity G (epimer 1) and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cilastatin in the chromatogram obtained with reference solution (c).

Calculation of percentage contents:
- for all impurities, use the concentration of cilastatin in reference solution (a);
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.3; impurity E = 3.3; impurity G (epimer 1) and impurity G (epimer 2) = 1.6.

Limits:
- *impurities A (sum of the epimers)*: maximum 0.5 per cent;
- *impurity C*: maximum 0.4 per cent;
- *impurities E*: maximum 0.3 per cent;
- *impurities B, F, H*: for each impurity, maximum 0.1 per cent;
- *impurity G*: for each epimer, maximum 0.1 per cent;
- *unspecified impurities*: for each impurity, maximum 0.05 per cent;
- *total*: maximum 1.0 per cent;
- *reporting threshold*: 0.03 per cent; disregard any peak due to impurity D in the chromatogram obtained with reference solution (d).

Impurity D, acetone and methanol

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.5 mL of *propanol R* in *water R* and dilute to 1000 mL with the same solvent.

Test solution Dissolve 0.200 g of the substance to be examined in *water R*, add 2.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*.

Reference solution Dissolve 2.0 mL of *acetone R*, 0.5 mL of *methanol R* and 0.5 mL of *mesityl oxide R* (impurity D) in *water R* and dilute to 1000 mL with the same solvent. To 2.0 mL of this solution add 2.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*. This solution contains 316 µg of acetone, 79 µg of methanol and 86 µg of impurity D per millilitre.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.53 mm;
- *stationary phase*: *macrogol 20 000 R* (film thickness 1.0 µm).

Carrier gas *helium for chromatography R*.

Flow rate 9 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2.5	50
	2.5 - 5	50 → 70
	5 - 5.5	70
Injection port		160
Detector		220

Detection Flame ionisation.

Injection 1 µL.

Calculate the percentage contents of acetone, methanol and impurity D using the following expression:

$$\left(\frac{C}{W}\right) \times \left(\frac{R_u}{R_s}\right)$$

C = concentration of the solvent in the reference solution, in µg/mL;
W = quantity of cilastatin sodium in the test solution, in milligrams;
R_u = ratio of the area of the solvent peak to the area of the propanol peak in the chromatogram obtained with the test solution;
R_s = ratio of the area of the solvent peak to the area of the propanol peak in the chromatogram obtained with the reference solution.

Limits:
- *acetone*: maximum 1.0 per cent *m/m*;
- *methanol*: maximum 0.5 per cent *m/m*;
- *impurity D*: maximum 0.4 per cent *m/m*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.17 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Dissolve 0.100 g in 30 mL of *methanol R* and add 5 mL of *water R*. Add *0.1 M hydrochloric acid* to a pH of about 3.0. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Three jumps of potential are observed. Read the volume added between the 1st and the 3rd point of inflexion.

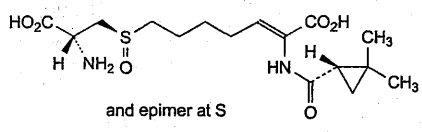
1 mL of *0.1 M sodium hydroxide* is equivalent to 19.02 mg of $C_{16}H_{25}N_2NaO_5S$.

STORAGE

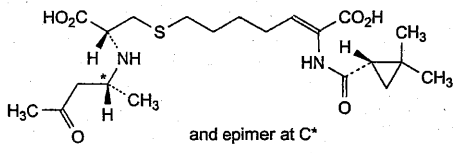
In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

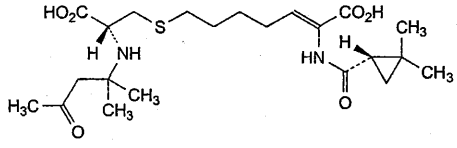
Specified impurities *A, B, C, D, E, F, G, H.*



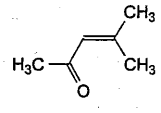
A. (*Z*)-7-[(*RS*)-[(2*R*)-2-amino-2-carboxyethyl]sulfinyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-2-enoic acid,



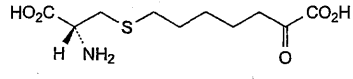
B. (*Z*)-7-[[(2*R*)-2-carboxy-2-[[(1*RS*)-1-methyl-3-oxobutyl]amino]ethyl]sulfanyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-2-enoic acid,



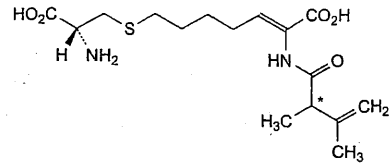
C. (*Z*)-7-[[(2*R*)-2-carboxy-2-[(1,1-dimethyl-3-oxobutyl)amino]ethyl]sulfanyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-2-enoic acid,



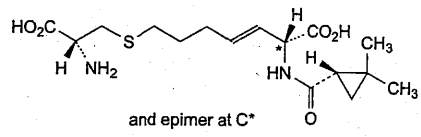
D. 4-methylpent-3-en-2-one (mesityl oxide),



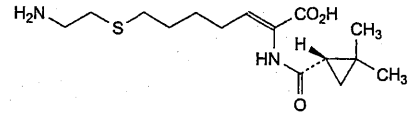
E. 7-[[(2*R*)-2-amino-2-carboxyethyl]sulfanyl]-2-oxoheptanoic acid,



F. (*Z*)-7-[[(2*R*)-2-amino-2-carboxyethyl]sulfanyl]-2-[(2,3-dimethylbut-3-enoyl)amino]hept-2-enoic acid,



G. (*E*)-(2*RS*)-7-[[(2*R*)-2-amino-2-carboxyethyl]sulfanyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-3-enoic acid,

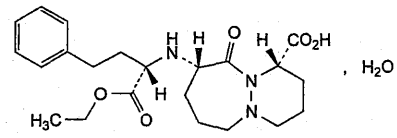


H. (*Z*)-7-[(2-aminoethyl)sulfanyl]-2-[[[(1*S*)-2,2-dimethylcyclopropyl]carbonyl]amino]hept-2-enoic acid.

Ph Eur

Cilazapril

(Ph. Eur. monograph 1499)



$C_{22}H_{31}N_3O_5,H_2O$ 435.5 *92077-78-6*

Action and use
Angiotensin converting enzyme inhibitor.

Ph Eur

DEFINITION

(1*S*,9*S*)-9-[[(1*S*)-1-(Ethoxycarbonyl)-3-phenylpropyl]amino]-10-oxooctahydro-6*H*-pyridazino[1,2-*a*][1,2]diazepine-1-carboxylic acid monohydrate.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in methanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cilazapril CRS*.

B. Specific optical rotation (see Tests).

TESTS

Specific optical rotation (*2.2.7*)
−383 to −399 (anhydrous substance).

Dissolve 0.200 g in *0.067 M phosphate buffer solution pH 7.0 R*, with the aid of ultrasound if necessary, and dilute to 50.0 mL with the same buffer solution. Carry out the determination at 365 nm.

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.20 g of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of *cilazapril impurity A CRS* in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *cilazapril impurity A CRS* and 5 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, hexane R, methanol R, ethyl acetate R* (5:5:15:15:60 *V/V/V/V/V*).

Application 5 µL.

Development Over a path of 10 cm.

Drying In a current of cold air for 10 min.

Detection Spray with a freshly prepared mixture of 1 volume of *potassium iodobismuthate solution R* and 10 volumes of *dilute acetic acid R* and then with *dilute hydrogen peroxide solution R.*

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Limit:
— *impurity A*: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram obtained with reference solution (a) (0.1 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *cilazapril impurity D CRS* in the test solution and dilute to 10.0 mL with the test solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 10 volumes of *triethylamine R* and 750 volumes of *water R*, adjust to pH 2.30 with *phosphoric acid R*, and add 200 volumes of *tetrahydrofuran R.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Run time Twice the retention time of cilazapril; when impurity A is present, it may be necessary to continue the chromatography until it is eluted.

Relative retention With reference to cilazapril: impurity B = about 0.6; impurity D = about 0.9; impurity C = about 1.6; impurity A = 4 to 5.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurity D and cilazapril;
— *symmetry factor*: maximum 3.0 for the peak due to cilazapril.

Limits:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity D*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity C*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to impurity A.

Water (*2.5.12*)

3.5 per cent to 5.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 10 mL of *anhydrous ethanol R* and add 50 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

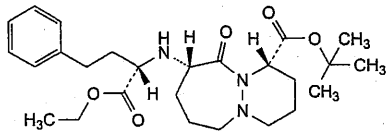
1 mL of *0.1 M sodium hydroxide* is equivalent to 41.75 mg of $C_{22}H_{31}N_3O_5$.

STORAGE

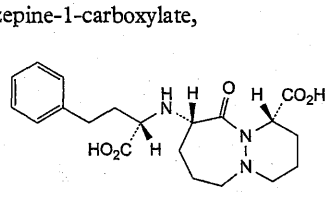
Protected from light.

IMPURITIES

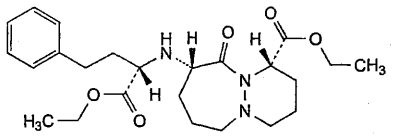
Specified impurities *A, B, C, D.*



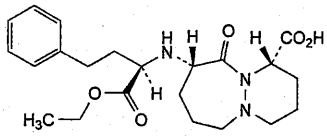
A. 1,1-dimethylethyl (1*S*,9*S*)-9-[[(*S*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]-10-oxooctahydro-6*H*-pyridazino[1,2-*a*][1,2]diazepine-1-carboxylate,



B. (1*S*,9*S*)-9-[[(*S*)-1-carboxy-3-phenylpropyl]amino]-10-oxooctahydro-6*H*-pyridazino[1,2-*a*][1,2]diazepine-1-carboxylic acid,



C. ethyl (1*S*,9*S*)-9-[[(*S*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]-10-oxooctahydro-6*H*-pyridazino[1,2-*a*][1,2]diazepine-1-carboxylate,

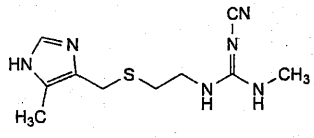


D. (1*S*,9*S*)-9-[[(*R*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]-10-oxooctahydro-6*H*-pyridazino[1,2-*a*][1,2]diazepine-1-carboxylic acid.

Ph Eur

Cimetidine

(Ph. Eur. monograph 0756)



$C_{10}H_{16}N_6S$ 252.3 *51481-61-9*

Action and use
Histamine H_2 receptor antagonist; treatment of peptic ulceration.

Preparations
Cimetidine Injection
Cimetidine Oral Solution
Cimetidine Oral Suspension
Cimetidine Tablets

Ph Eur

DEFINITION
2-Cyano-1-methyl-3-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]guanidine.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Slightly soluble in water, soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in dilute mineral acids.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 139 °C to 144 °C.

If necessary, dissolve the substance to be examined in *2-propanol R*, evaporate to dryness and determine the melting point again.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cimetidine CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *2-propanol R*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *cimetidine CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R* (15:20:65 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In a current of cold air.

Detection Expose to iodine vapour until maximum contrast has been obtained and examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 3.0 g in 12 mL of *1 M hydrochloric acid* and dilute to 20 mL with *water R.*

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 2.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *cimetidine for system suitability CRS* (containing impurities B, C, D, E, G and H) in 1.0 mL of mobile phase A.

Reference solution (c) Dissolve 4 mg of *cimetidine for peak identification CRS* (containing impurity F) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm).

Mobile phase A Mix 0.4 volumes of *diethylamine R* and 780 volumes of a 1.1 g/L solution of *sodium hexanesulfonate R*; adjust to pH 2.8 with *phosphoric acid R*; add 250 volumes of *methanol R2*;

Mobile phase B *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	100	0
60 - 65	100 → 90	0 → 10
65 - 120	90	10

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 µl.

Identification of impurities Use the chromatogram supplied with *cimetidine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D, E, G and H; use the chromatogram supplied with *cimetidine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to cimetidine (retention time = about 18 min): impurity G = about 0.2; impurity E = about 0.4; impurity D = about 1.5; impurity C = about 1.6; impurity B = about 2.0; impurity H = about 2.3; impurity F = about 4.6.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the peaks due to impurities D and C.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 2.5; impurity D = 3.3; impurity E = 0.7; impurity G = 0.6.

— *impurities B, C, D, E, F, G, H*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying *(2.2.32)*

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash *(2.4.14)*

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end point potentiometrically *(2.2.20)*.

1 mL of *0.1 M perchloric acid* is equivalent to 25.23 mg of $C_{10}H_{16}N_6S$

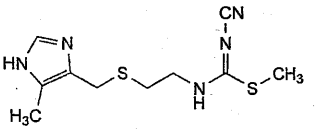
STORAGE

Protected from light.

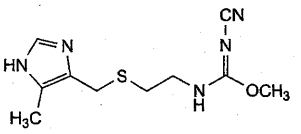
IMPURITIES

Specified impurities *B, C, D, E, F, G, H.*

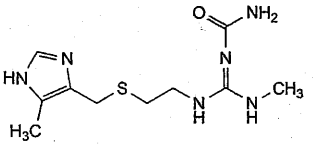
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, I, J.*



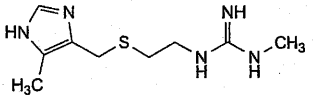
A. methyl 3-cyano-1-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]carbamimidothioate,



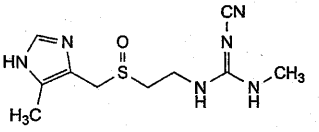
B. methyl 3-cyano-1-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]carbamimidate,



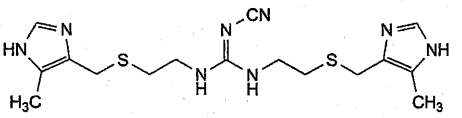
C. 1-[(methylamino)[[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]amino]methylidene]urea,



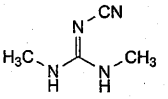
D. 1-methyl-3-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]guanidine,



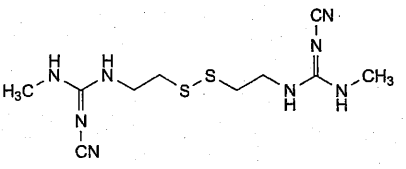
E. 2-cyano-1-methyl-3[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfinyl]ethyl]guanidine,



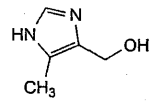
F. 2-cyano-1,3-bis[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]guanidine,



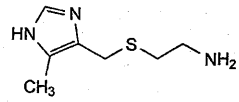
G. 2-cyano-1,3-dimethylguanidine,



H. 1,1′-(disulfanediyldiethylene)bis(2-cyano-3-methylguanidine),



I. (5-methyl-1*H*-imidazol-4-yl)methanol,

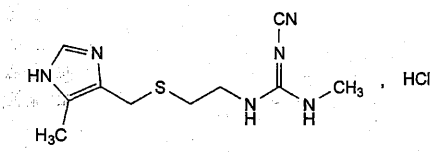


J. 2-[[(5-methyl-1*H*-imidazol-4-yl)methyl] sulfanyl]ethanamine.

Ph Eur

Cimetidine Hydrochloride

(Ph. Eur. monograph 1500)



$C_{10}H_{17}ClN_6S$ 288.8 *70059-30-2*

Action and use
Histamine H_2 receptor antagonist; treatment of peptic ulceration.

Preparation
Cimetidine Injection

Ph Eur

DEFINITION
2-Cyano-1-methyl-3-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl] sulfanyl]ethyl]guanidine hydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in anhydrous ethanol.

IDENTIFICATION
First identification: B, D.
Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 70 mg in *0.2 M sulfuric acid* and dilute to 100.0 mL with the same acid. Dilute 2.0 mL of this solution to 100.0 mL with *0.2 M sulfuric acid.*

Specific absorbance at the absorption maximum at 218 nm
650 to 705.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cimetidine hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *cimetidine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R* (15:20:65 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In a current of cold air

Detection Expose to iodine vapour until maximum contrast has been obtained and examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 3.0 g in 12 mL of *1 M hydrochloric acid* and dilute to 20 mL with *water R.*

pH (*2.2.3*)
4.0 to 5.0.

Dissolve 100 mg in *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 2.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *cimetidine for system suitability CRS* (containing impurities B, C, D, E, G and H) in 1.0 mL of mobile phase A.

Reference solution (c) Dissolve 4 mg of *cimetidine for peak identification CRS* (containing impurity F) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm).

Mobile phase A Mix 0.4 volumes of *diethylamine R* and 780 volumes of a 1.1 g/L solution of *sodium hexanesulfonate R.* Adjust to pH 2.8 with *phosphoric acid R* and add 250 volumes of *methanol R2*;

Mobile phase B *methanol R2*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	100	0
60 - 65	100 → 90	0 → 10
65 - 120	90	10

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram supplied with *cimetidine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to the impurities B, C, D, E, G and H; use the chromatogram supplied with *cimetidine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to cimetidine (retention time = about 18 min): impurity G = about 0.2; impurity E = about 0.4; impurity D = about 1.5; impurity C = about 1.6; impurity B = about 2.0; impurity H = about 2.3; impurity F = about 4.6.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurities D and C.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 2.5; impurity D = 3.3; impurity E = 0.7; impurity G = 0.6;
— *impurities B, C, D, E, F, G, H*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 5 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 28.88 mg of $C_{10}H_{17}ClN_6S$.

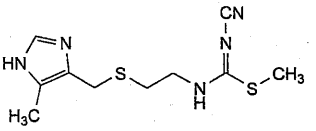
STORAGE

Protected from light.

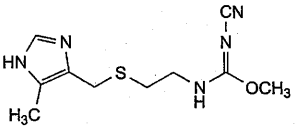
IMPURITIES

Specified impurities *B, C, D, E, F, G, H.*

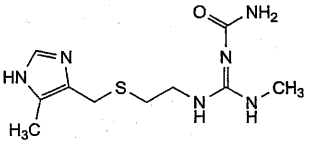
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, I, J.*



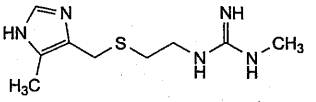
A. methyl 3-cyano-1-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]carbamimidothioate,



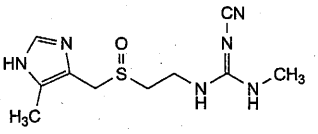
B. methyl 3-cyano-1-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]carbamimidate,



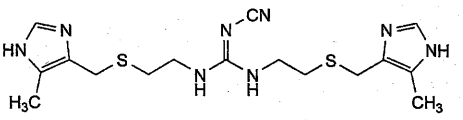
C. 1-[(methylamino)[[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]amino]methylidene]urea,



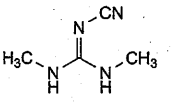
D. 1-methyl-3-[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfanyl]ethyl]guanidine,



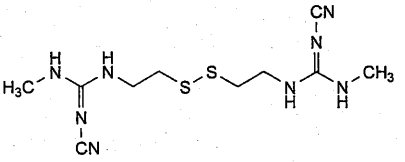
E. 2-cyano-1-methyl-3[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]sulfinyl]ethyl]guanidine,



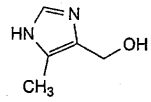
F. 2-cyano-1,3-bis[2-[[(5-methyl-1*H*-imidazol-4-yl)methyl]-sulfanyl]ethyl]guanidine.



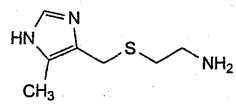
G. 2-cyano-1,3-dimethylguanidine,



H. 1,1′-(disulfanediyldiethylene)bis(2-cyano-3-methylguanidine),



I. (5-methyl-1*H*-imidazol-4-yl)methanol,

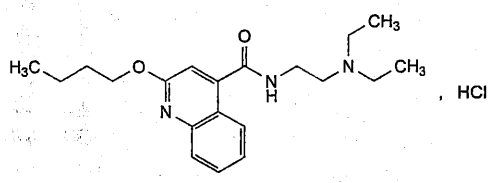


J. 2-[[(5-methyl-1*H*-imidazol-4-yl)methyl] sulfanyl]ethanamine.

Ph Eur

Cinchocaine Hydrochloride

(Ph. Eur. monograph 1088)



$C_{20}H_{30}ClN_3O_2$ 379.9 *61-12-1*

Action and use
Local anaesthetic.

Ph Eur

DEFINITION
2-Butoxy-*N*-[2-(diethylamino)ethyl]quinoline-4-carboxamide hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals, hygroscopic, very easily agglomerates.

Solubility
Very soluble in water, freely soluble in acetone, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 60.0 mg in a 103 g/L solution of *hydrochloric acid R* and dilute to 100 mL with the same acid solution. Dilute 2 mL of this solution to 100 mL with a 103 g/L solution of *hydrochloric acid R*.

Special range 220-350 nm.

Absorption maxima 246 nm and 319 nm.

Absorbance ratio A_{246}/A_{319} = 2.7 to 3.0.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cinchocaine hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 20 mg of *cinchocaine hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *ammonia R, methanol R, acetone R, toluene R* (1:5:30:50 *V/V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air for 15 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 0.5 g in 5 mL of *water R*. Add 1 mL of *dilute ammonia R2*. A white precipitate is formed. Filter, wash the precipitate with 5 quantities, each of 10 mL, of *water R* and dry in a desiccator. It melts at 64 °C to 66 °C (*2.2.14*).

E. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
5.0 to 6.0.

Dilute 10 mL of solution S to 50 mL with *carbon dioxide-free water R*.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase B, *water R* (10:90 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of *cinchocaine impurity A CRS* and 2 mg of *cinchocaine impurity B CRS* in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL of the solution to 10 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 3 mm;
— *stationary phase*: end-capped cross-linked *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: to 950 mL of *water for chromatography R* add 0.6 mL of *phosphoric acid R*, adjust to pH 2.0 with the same acid and dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	90	10
2 - 5.5	90 → 82	10 → 18
5.5 - 10.5	82 → 50	18 → 50
10.5 - 16.5	50 → 49	50 → 51
16.5 - 19.5	49 → 46	51 → 54
19.5 - 24.5	46 → 30	54 → 70

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 229 nm.

Injection 10 µL.

Relative retention With reference to cinchocaine (retention time = about 10.5 min): impurity B = about 0.5; impurity A = about 0.6.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to impurities B and A.

Calculation of percentage contents:
— for each impurity, use the concentration of cinchocaine hydrochloride in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.3 per cent;
— reporting threshold: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 0.500 g by drying *in vacuo* at 60 °C.

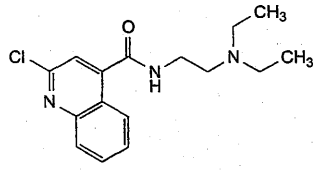
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in a mixture of 15.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

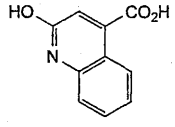
1 mL of *0.1 M sodium hydroxide* is equivalent to 37.99 mg of $C_{20}H_{30}ClN_3O_2$.

STORAGE
In an airtight container, protected from light.

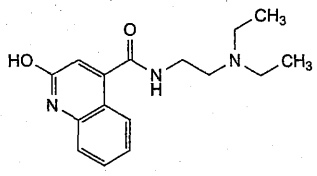
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D.*



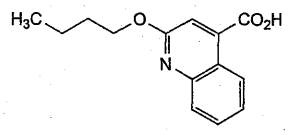
A. 2-chloro-*N*-[2-(diethylamino)ethyl]quinoline-4-carboxamide,



B. 2-hydroxyquinoline-4-carboxylic acid,



C. *N*-[2-(diethylamino)ethyl]-2-hydroxyquinoline-4-carboxamide,

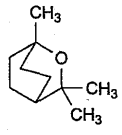


D. 2-butoxyquinoline-4-carboxylic acid.

Ph Eur

Cineole

(Ph. Eur. monograph 1973)



$C_{10}H_{18}O$ 154.3 *470-82-6*

Ph Eur

DEFINITION
1,3,3-Trimethyl-2-oxabicyclo[2.2.2]octane.

CHARACTERS
Appearance
Clear colourless liquid.

Solubility
Practically insoluble in water, miscible with alcohol and with methylene chloride.

It solidifies at about 0.5 °C.

IDENTIFICATION
A. Refractive index (see Tests).

B. Thin-layer chromatography (*2.2.27*).

Test solution Dilute 1 mL of solution S (see Tests) to 25 mL with *alcohol R*.

Reference solution Mix 80 mg of *cineole CRS* with *alcohol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R*.

Mobile phase *ethyl acetate R, toluene R* (10:90 *V/V*).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In a current of cold air.

Detection Spray with *anisaldehyde solution R*, heat at 100-105 °C for 5 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 0.1 mL add 4 mL of *sulfuric acid R*. An orange-red colour develops. Add 0.2 mL of *formaldehyde solution R*. The colour changes to deep brown.

TESTS

Solution S

Dilute 2.00 g to 10.0 mL with *alcohol R*.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method I*).

Chiral impurities

The optical rotation (*2.2.7*) of solution S is −0.10° to + 0.10°.

Refractive index (*2.2.6*)

1.456 to 1.460.

Related substances

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 1.0 g of *camphor R* in *heptane R* and dilute to 200 mL with the same solvent.

Test solution (a) Dissolve 2.5 g of the substance to be examined in *heptane R* and dilute to 25.0 mL with the same solvent.

Test solution (b) Dissolve 2.5 g of the substance to be examined in *heptane R*, add 5.0 mL of the internal standard solution and dilute to 25.0 mL with *heptane R*.

Reference solution (a) To 2.0 mL of test solution (a) add 20.0 mL of the internal standard solution and dilute to 100.0 mL with *heptane R*.

Reference solution (b) Dissolve 50 mg of *1,4-cineole R* and 50 mg of the substance to be examined in *heptane R* and dilute to 50.0 mL with the same solvent.

Column:

— *size*: *l* = 30 m, Ø = 0.25 mm,

— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Linear velocity 45 cm/s.

Split-ratio 1:70.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 10	50
	10 - 35	50 → 100
	35 - 45	100 → 200
	45 - 55	200
Injection port		220
Detector		250

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution (b):

— *resolution*: minimum 10 between the peaks due to impurity A and to cineole.

Limits:

— *total*: calculate the ratio (*R*) of the area of the peak due to cineole to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (a); from the chromatogram obtained with test solution (b), calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to internal standard: this ratio is not greater than *R* (2 per cent),

— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Residue on evaporation

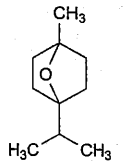
Maximum 0.1 per cent.

To 2.0 g add 5 mL of *water R*, evaporate to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs a maximum of 2 mg.

STORAGE

In an airtight container, protected from light.

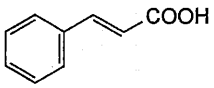
IMPURITIES



A. 1-methyl-4-(1-methylethyl)-7-oxabicyclo[2.2.1]heptane (1,4-cineole).

Ph Eur

Cinnamic Acid



$C_9H_8O_2$ 148.2 *621-82-9*

Action and use

Antimicrobial preservative; excipient.

DEFINITION

Cinnamic Acid is (*E*)-3-phenylprop-2-enoic acid. It contains not less than 99.0% and not more than 100.5% of $C_9H_8O_2$, calculated with reference to the dried substance.

CHARACTERISTICS

Colourless crystals.

Very slightly soluble in *water*; freely soluble in *ethanol (96%)*; soluble in *ether*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of cinnamic acid *(RS 062)*.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.0010% w/v solution in 0.1M *sodium hydroxide* exhibits a maximum only at 267 nm. The *absorbance* at 267 nm is about 1.4.

TESTS

Melting point

132° to 134°, Appendix V A.

Ethanol-insoluble matter

A 10% w/v solution in *ethanol (96%)* is *clear*, Appendix IV A.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in *methanol*.
(1) 5.0% w/v of the substance being examined.
(2) 0.025% w/v of the substance being examined.
CHROMATOGRAPHIC CONDITIONS
(a) Use as the coating *silica gel* GF_{254}.
(b) Use the mobile phase as described below.
(c) Apply 5 µL of each solution.
(d) Develop the plate to 15 cm.
(e) After removal of the plate, dry in air and examine under *ultraviolet light (254 nm)*.
MOBILE PHASE
10 volumes of *glacial acetic acid* and 90 volumes of *toluene*.
LIMITS
Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

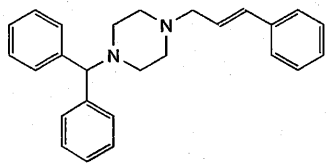
Loss on drying
When dried to constant weight at 60° at a pressure not exceeding 0.7 kPa, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.5 g in 15 mL of *ethanol (96%)* previously neutralised to *phenol red solution* and titrate with *0.1M sodium hydroxide VS* using *phenol red solution* as indicator. Each mL of *0.1M sodium hydroxide VS* is equivalent to 14.82 mg of $C_9H_8O_2$.

Cinnarizine

(Ph. Eur. monograph 0816)



$C_{26}H_{28}N_2$ 368.5 *298-57-7*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparation
Cinnarizine Tablets

Ph Eur

DEFINITION
(*E*)-1-(Diphenylmethyl)-4-(3-phenylprop-2-enyl)piperazine.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in acetone, slightly soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION
First identification: A, B.
Second identification: A, C, D.
A. Melting point (*2.2.14*): 118 °C to 122 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cinnarizine CRS.*
C. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 20 mL with the same solvent.
Reference solution (a) Dissolve 10 mg of *cinnarizine CRS* in *methanol R* and dilute to 20 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of *cinnarizine CRS* and 10 mg of *flunarizine dihydrochloride CRS* in *methanol R* and dilute to 20 mL with the same solvent.
Plate *TLC octadecylsilyl silica gel* F_{254} *plate R.*
Mobile phase 58.4 g/L solution of *sodium chloride R*, *methanol R*, *acetone R* (20:30:50 *V/V/V*).
Application 5 µL.
Development In an unsaturated tank, over 3/4 of the plate.
Drying In air.
Detection Examine in ultraviolet light at 254 nm.
System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).
D. Dissolve 0.2 g of *anhydrous citric acid R* in 10 mL of *acetic anhydride R* in a water-bath at 80 °C and maintain the temperature of the water-bath at 80 °C for 10 min. Add about 20 mg of the substance to be examined. A purple colour develops.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).
Dissolve 0.5 g in *methylene chloride R* and dilute to 20 mL with the same solvent.

Acidity or alkalinity
Suspend 0.5 g in 15 mL of *water R*. Boil for 2 min. Cool and filter. Dilute the filtrate to 20 mL with *carbon dioxide-free water R*. To 10 mL of this solution add 0.1 mL of *phenolphthalein solution R* and 0.25 mL of *0.01 M sodium hydroxide*. The solution is pink. To 10 mL of the solution add 0.1 mL of *methyl red solution R* and 0.25 mL of *0.01 M hydrochloric acid*. The solution is red.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 25.0 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.
Reference solution (a) Dissolve 12.5 mg of *cinnarizine CRS* and 15.0 mg of *flunarizine dihydrochloride CRS* in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 20.0 mL with *methanol R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 5.0 mL of this solution to 20.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.1 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 10 g/L solution of *ammonium acetate R*;
— *mobile phase B*: 0.2 per cent *V/V* solution of *glacial acetic acid R* in *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	75 → 10	25 → 90
20 - 25	10	90

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Relative retention With reference to cinnarizine (retention time = about 11 min): impurity A = about 0.4; flunarizine = about 1.05; impurity B = about 1.1; impurity C = about 1.2; impurity D = about 1.6; impurity E = about 1.8.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to cinnarizine and flunarizine.

Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven *in vacuo* at 60 °C for 4 h.

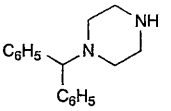
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

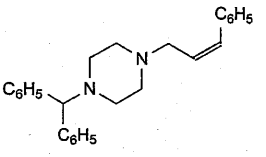
1 mL of *0.1 M perchloric acid* is equivalent to 18.43 mg of $C_{26}H_{28}N_2$.

STORAGE
Protected from light.

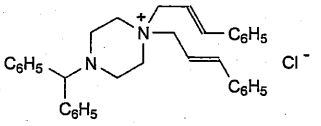
IMPURITIES
Specified impurities *A, B, C, D, E.*



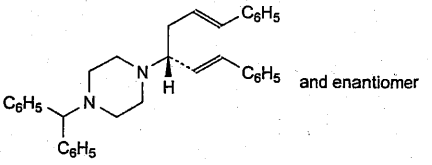
A. 1-(diphenylmethyl)piperazine,



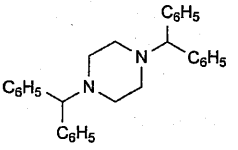
B. (*Z*)-1-(diphenylmethyl)-4-(3-phenylprop-2-enyl)piperazine,



C. 4-(diphenylmethyl)-1,1-bis[(*E*)-3-phenylprop-2-enyl] piperazinium chloride,



D. 1-(diphenylmethyl)-4-[(1*RS*,3*E*)-4-phenyl-1-[(*E*)-2-phenylethenyl]but-3-enyl]piperazine,

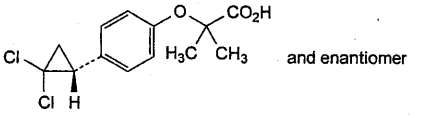


E. 1,4-bis(diphenylmethyl)piperazine.

Ph Eur

Ciprofibrate

(Ph. Eur. monograph 2013)



$C_{13}H_{14}Cl_2O_3$ 289.2 *52214-84-3*

Action and use
Fibrate; lipid-regulating drug.

Ph Eur

DEFINITION
2-[4-[(1*RS*)-2,2-Dichlorocyclopropyl]phenoxy]-2-methylpropanoic acid.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol, soluble in toluene.

mp
About 115 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciprofibrate CRS.*

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_4 (*2.2.2, Method II*).

Dissolve 1.0 g in *anhydrous ethanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.125 g of the substance to be examined in a mixture of equal volumes of *acetonitrile R* and *water R* and dilute to 50 mL with the same mixture of solvents.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of equal volumes of *acetonitrile R* and *water R*. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of equal volumes of *acetonitrile R* and *water R*.

Reference solution (b) Dissolve the contents of a vial of *ciprofibrate for system suitability CRS* in 2.0 mL of a mixture of equal volumes of *acetonitrile R* and *water R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 1.36 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.2 with *phosphoric acid R*,
— *mobile phase B*: *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	75 → 30	25 → 70
30 - 40	30	70
40 - 42	30 → 75	70 → 25

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *ciprofibrate for system suitability CRS* to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to ciprofibrate (retention time = about 18 min): impurity A = about 0.7; impurity B = about 0.8; impurity C = about 0.95; impurity D = about 1.3; impurity E = about 1.5.

System suitability Reference solution (b):
— *resolution*: baseline separation between the peaks due to impurity C and ciprofibrate.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 2.3,
— *impurities A, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *impurity E*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent),
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total of other impurities*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 350 ppm.

To 0.190 g add 20 mL of *water R* and treat in an ultrasonic bath for 8 min. Filter. 15 mL of the filtrate complies with the test.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

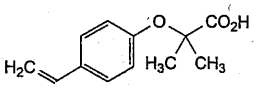
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in a mixture of 20 mL of *water R* and 40 mL of *anhydrous ethanol R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

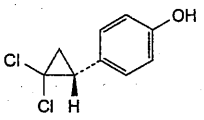
1 mL of *0.1 M sodium hydroxide* is equivalent to 28.92 mg of $C_{13}H_{14}Cl_2O_3$.

STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *A, B, C, D, E.*

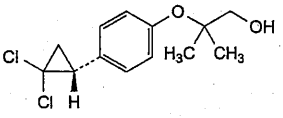


A. 2-(4-ethenylphenoxy)-2-methylpropanoic acid,



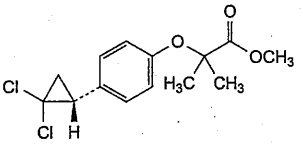
and enantiomer

B. 4-[(1*RS*)-2,2-dichlorocyclopropyl]phenol,



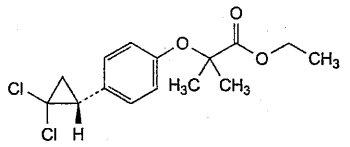
and enantiomer

C. 2-[4-[(1*RS*)-2,2-dichlorocyclopropyl]phenoxy]-2-methylpropan-1-ol,



and enantiomer

D. methyl 2-[4-[(1*RS*)-2,2-dichlorocyclopropyl]phenoxy]-2-methylpropanoate,



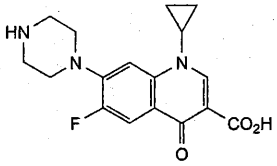
and enantiomer

E. ethyl 2-[4-[(1*RS*)-2,2-dichlorocyclopropyl]phenoxy]-2-methylpropanoate.

Ph Eur

Ciprofloxacin

(Ph. Eur. monograph 1089)



$C_{17}H_{18}FN_3O_3$ 331.4 *85721-33-1*

Action and use
Fluoroquinolone antibacterial.

Preparations
Ciprofloxacin Infusion
Ciprofloxacin Eye Drops

Ph Eur

DEFINITION
1-Cyclopropyl-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
Almost white or pale yellow, crystalline powder, slightly hygroscopic.

Solubility
Practically insoluble in water, very slightly soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *ciprofloxacin CRS*.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 (*2.2.2, Method II*).

Dissolve 0.25 g in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 20 mL with the same solution.

Impurity A
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *dilute ammonia R1* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 10 mg of *ciprofloxacin impurity A CRS* in a mixture of 0.1 mL of *dilute ammonia R1* and 90 mL of *water R* and dilute to 100 mL with *water R*. Dilute 2 mL of the solution to 10 mL with *water R*.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *acetonitrile R, concentrated ammonia R, methanol R, methylene chloride R* (10:20:40:40 *V/V/V/V*).

Application 5 µL.

Development At the bottom of a chromatographic tank, place an evaporating dish containing 50 mL of *concentrated ammonia R*. Expose the plate to the ammonia vapour for 15 min in the closed tank. Withdraw the plate, transfer to a 2nd chromatographic tank and develop over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Limit:
— *impurity A*: any spot due to impurity A is not more intense than the principal spot in the chromatogram obtained with the reference solution (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution To 25.0 mg of the substance to be examined add 0.2 mL of *dilute phosphoric acid R* and dilute to 50.0 mL with the mobile phase. Treat in an ultrasonic bath until a clear solution is obtained.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 5.0 mL with the mobile phase.

Reference solution (b) Dissolve 2.5 mg of *ciprofloxacin hydrochloride for peak identification CRS* (containing impurities B, C, D and E) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 13 volumes of *acetonitrile R* and 87 volumes of a 2.45 g/L solution of *phosphoric acid R*, previously adjusted to pH 3.0 with *triethylamine R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 278 nm.

Injection 50 µL.

Run time Twice the retention time of ciprofloxacin.

Identification of impurities Use the chromatogram supplied with *ciprofloxacin hydrochloride for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D and E.

Relative retention With reference to ciprofloxacin (retention time = about 9 min): impurity E = about 0.4; impurity B = about 0.6; impurity C = about 0.7; impurity D = about 1.2.

System suitability Reference solution (b):
— *resolution*: minimum 1.3 between the peaks due to impurities B and C.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.7; impurity C = 0.6; impurity D = 1.4; impurity E = 6.7;
— *impurities B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying under vacuum at 120 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

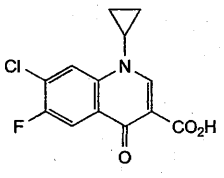
ASSAY
Dissolve 0.300 g in 80 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 33.14 mg of $C_{17}H_{18}FN_3O_3$.

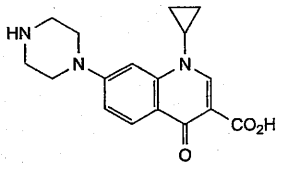
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *A, B, C, D, E.*

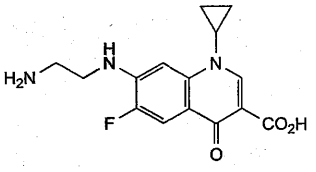
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F.*



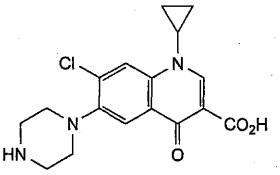
A. 7-chloro-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (fluoroquinolonic acid),



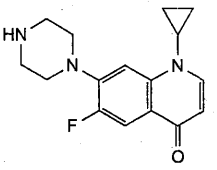
B. 1-cyclopropyl-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid (desfluoro compound),



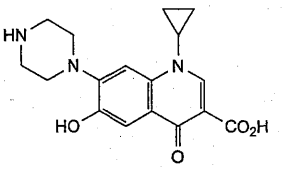
C. 7-[(2-aminoethyl)amino]-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (ethylenediamine compound),



D. 7-chloro-1-cyclopropyl-4-oxo-6-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid,



E. 1-cyclopropyl-6-fluoro-7-(piperazin-1-yl)quinolin-4(1*H*)-one (decarboxylated compound),

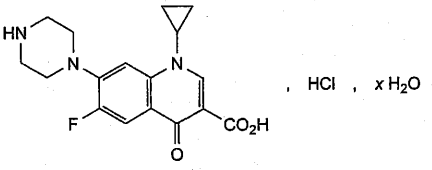


F. 1-cyclopropyl-6-hydroxy-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.

Ph Eur

Ciprofloxacin Hydrochloride

(Ph. Eur. monograph 0888)



$C_{17}H_{19}ClFN_3O_3,xH_2O$ 367.8
(anhydrous)

Action and use
Fluoroquinolone antibacterial.

Preparation
Ciprofloxacin Tablets

Ph Eur

DEFINITION
1-Cyclopropyl-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid hydrochloride. It contains a variable quantity of water.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
Pale yellow, crystalline, slightly hygroscopic powder.

Solubility
Soluble in water, slightly soluble in methanol, very slightly soluble in anhydrous ethanol, practically insoluble in acetone, in ethyl acetate and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ciprofloxacin hydrochloride CRS.*

B. 0.1 g gives reaction (b) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 (*2.2.2, Method II*).

Dilute 10 mL of solution S to 20 mL with *carbon dioxide-free water R.*

pH (*2.2.3*)

3.5 to 4.5 for solution S.

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 10 mg of *ciprofloxacin impurity A CRS* in a mixture of 0.1 mL of *dilute ammonia R1* and 90 mL of *water R* and dilute to 100 mL with *water R.* Dilute 2 mL of the solution to 10 mL with *water R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetonitrile R, concentrated ammonia R, methanol R, methylene chloride R* (10:20:40:40 *V/V/V/V*).

Application 5 µL.

Development At the bottom of a chromatographic tank, place an evaporating dish containing 50 mL of *concentrated ammonia R.* Expose the plate to the ammonia vapour for 15 min in the closed tank. Withdraw the plate, transfer to a 2nd chromatographic tank and develop over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Limit:

— *impurity A*: any spot due to impurity A is not more intense than the principal spot in the chromatogram obtained with the reference solution (0.2 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *ciprofloxacin hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 2.5 mg of *ciprofloxacin hydrochloride for peak identification CRS* (containing impurities B, C, D and E) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 40 °C.

Mobile phase Mix 13 volumes of *acetonitrile R* and 87 volumes of a 2.45 g/L solution of *phosphoric acid R* previously adjusted to pH 3.0 with *triethylamine R.*

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 278 nm.

Injection 50 µL of the test solution and reference solutions (b) and (c).

Run time 2.3 times the retention time of ciprofloxacin.

Identification of impurities Use the chromatogram supplied with *ciprofloxacin hydrochloride for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D and E.

Relative retention With reference to ciprofloxacin (retention time = about 9 min): impurity E = about 0.4; impurity B = about 0.6; impurity C = about 0.7; impurity D = about 1.2.

System suitability Reference solution (b):

— *resolution*: minimum 1.3 between the peaks due to impurities B and C.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.7; impurity C = 0.6; impurity D = 1.4; impurity E = 6.7;

— *impurity E*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— *impurities B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.12*)

Maximum 6.7 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $C_{17}H_{19}ClFN_3O_3$ taking into account the assigned content of *ciprofloxacin hydrochloride CRS.*

STORAGE

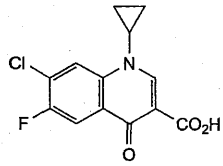
In an airtight container, protected from light.

IMPURITIES

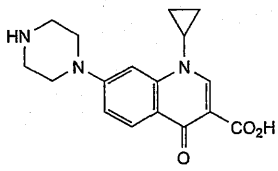
Specified impurities *A, B, C, D, E.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is

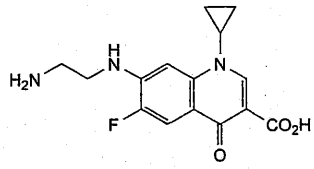
therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) F.



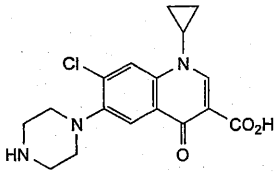
A. 7-chloro-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (fluoroquinolonic acid),



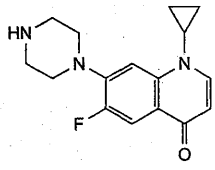
B. 1-cyclopropyl-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid (desfluoro compound),



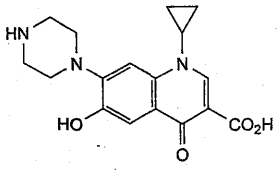
C. 7-[(2-aminoethyl)amino]-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (ethylenediamine compound),



D. 7-chloro-1-cyclopropyl-4-oxo-6-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid,



E. 1-cyclopropyl-6-fluoro-7-(piperazin-1-yl)quinolin-4(1*H*)-one (decarboxylated compound),

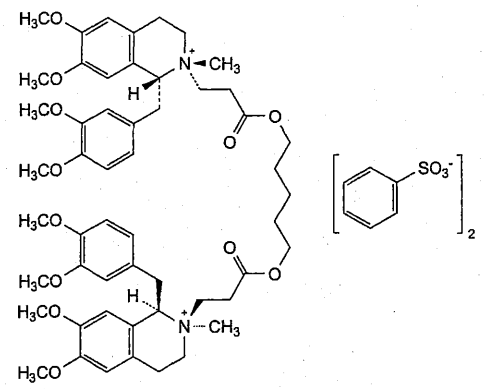


F. 1-cyclopropyl-6-hydroxy-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.

Ph Eur

Cisatracurium Besilate

(Ph. Eur. monograph 2763)



$C_{65}H_{82}N_2O_{18}S_2$ 1243 *96946-42-8*

Action and use
Non-depolarizing neuromuscular blocker.

Ph Eur

DEFINITION

2,2′-[Pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[(1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium] dibenzenesulfonate.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

It is considered that alkyl benzenesulfonate esters are genotoxic and are potential impurities in cisatracurium besilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation.
The general method *2.5.41. Methyl, ethyl and isopropyl benzenesulfonate in active substances* is available to assist manufacturers.

CHARACTERS

Appearance
White or yellowish, hygroscopic powder.

Solubility
Soluble in water, very soluble in anhydrous ethanol, freely soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cisatracurium besilate CRS.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 1.00 g in *water R* and dilute to 100 mL with the same solvent.

Diastereoisomers

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 0.100 g of the substance to be examined in a 0.1 per cent *V/V* solution of *diethylamine R* in *methanol R* and dilute to 25.0 mL with the same solution. Heat in a water-bath at 60 °C for 15 min then cool to room temperature.

Reference solution Dissolve 8 mg of *laudanosine CRS* in a 0.1 per cent *V/V* solution of *diethylamine R* in *methanol R* and dilute to 2.0 mL with the same solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *amylose derivative of silica gel for chiral separation R* (5 µm).

Mobile phase *propanol R1*, 0.1 per cent *V/V* solution of *diethylamine R* in *heptane R* (15:85 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 285 nm.

Injection 10 µL.

Run time 1.5 times the retention time of *R*-laudanosine.

Elution order *S*-laudanosine; *R*-laudanosine.

Retention time *R*-laudanosine = about 18 min.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to *S*-laudanosine and *R*-laudanosine.

Calculate the percentage content of the sum of impurities S, T, U and V by multiplying the percentage of *S*-laudanosine by 2.

Limit:
— *sum of impurities S, T, U, V*: maximum 0.5 per cent; disregard any peak other than *S*-laudanosine and *R*-laudanosine.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Solvent mixture Mix 20 volumes of *acetonitrile R* and 80 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.1 with *phosphoric acid R*.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of *cisatracurium besilate CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2.0 mg of *cisatracurium for system suitability CRS* (containing impurities D, F, N, O, P, Q and H and/or T and/or V) in 1.0 mL of the solvent mixture.

Reference solution (d) Dissolve 2.0 mg of *cisatracurium for peak identification CRS* (containing impurities A, C, I (epimer 1) and I (epimer 2) and/or K and/or L and/or M) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.10 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (2 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: mix 5 volumes of *methanol R*, 15 volumes of *acetonitrile R* and 80 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R*;
— *mobile phase B*: *methanol R*, *acetonitrile R* (30:70 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	92 → 82	8 → 18
1 - 8	82 → 80	18 → 20
8 - 10	80 → 70	20 → 30
10 - 12	70 → 45	30 → 55
12 - 20	45	55

Flow rate 0.5 mL/min.

Detection spectrophotometer at 280 nm.

Autosampler Set at 4 °C.

Injection 5 µL of test solution (a) and reference solutions (b), (c) and (d).

Identification of impurities use the chromatogram supplied with *cisatracurium for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D, F, H + T + V, N, O, P, Q; use the chromatogram supplied with *cisatracurium for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, C, I (epimer 1) and I (epimer 2) + K + L + M.

Relative retention With reference to cisatracurium (retention time = about 9 min): impurity A = about 0.3; impurity C = about 0.4; impurity D = about 0.45; impurity F = about 0.55; impurities H, T and V = about 0.97; impurity I (epimer 1) = about 1.25; impurities I (epimer 2), K, L and M = about 1.3; impurity N = about 1.4; impurity O = about 1.5; impurity P = about 1.6; impurity Q = about 1.65.

Impurity M can coelute or be separated from impurities I (epimer 2), K and L.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to impurity H and/or T and/or V and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cisatracurium.

Limits:
— *correction factor*: multiply the peak area of impurity C by 0.5;
— *sum of impurities H, T, V*: maximum 0.8 per cent;
— *impurity Q*: maximum 0.7 per cent;
— *impurities A, C, F, O, P*: for each impurity, maximum 0.5 per cent;
— *impurity D*: maximum 0.4 per cent;
— *impurity N*: maximum 0.3 per cent;
— *sum of impurities I, K, L, M*: maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 2.5 per cent;
— *reporting threshold*: 0.05 per cent (reference solution (b)).

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications:

Injection Test solution (b) and reference solution (a).

System suitability Reference solution (a):
— *symmetry factor*: maximum 2.5 for the principal peak.

Calculate the percentage content of $C_{65}H_{82}N_2O_{18}S_2$ taking into account the assigned content of *cisatracurium besilate CRS*.

STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

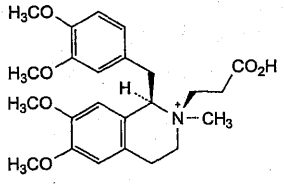
IMPURITIES

Test for diastereoisomers: S.

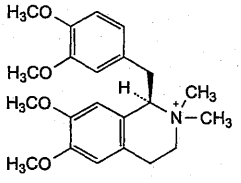
Test for related substances: *A, B, C, D, E, F, G, H, I, K, L, M, N, O, P, Q, R, T, U, V, W.*

Specified impurities *A, C, D, F, H, I, K, L, M, N, O, P, Q, S, T, V.*

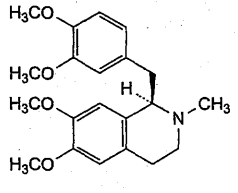
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, E, G, R, U, W.*



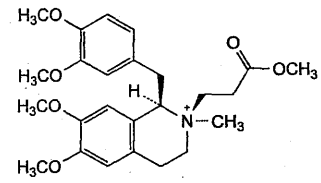
A. (1*R*,2*R*)-2-(2-carboxyethyl)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



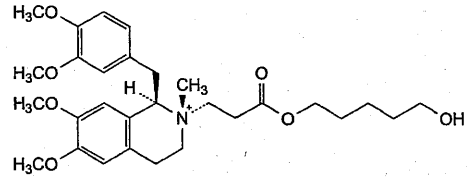
B. (1*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2,2-dimethyl-1,2,3,4-tetrahydroisoquinolinium,



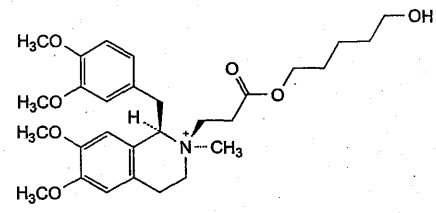
C. (1*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinoline (1*R*-laudanosine),



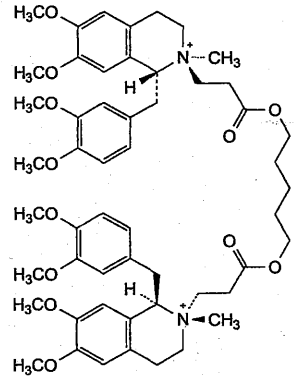
D. (1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-(3-methoxy-3-oxopropyl)-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



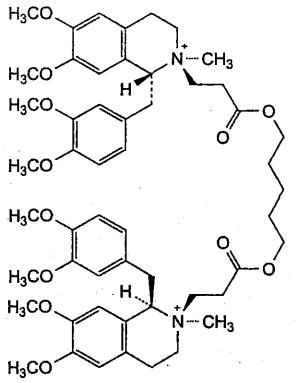
E. (1*R*,2*S*)-1-[(3,4-dimethoxyphenyl)methyl]-2-[3-[(5-hydroxypentyl)oxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



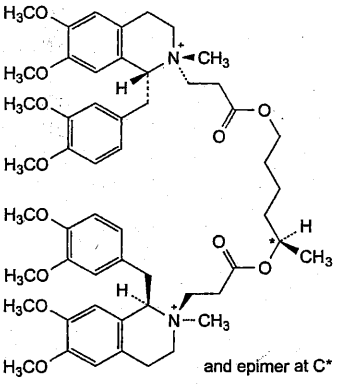
F. (1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-2-[3-[(5-hydroxypentyl)oxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



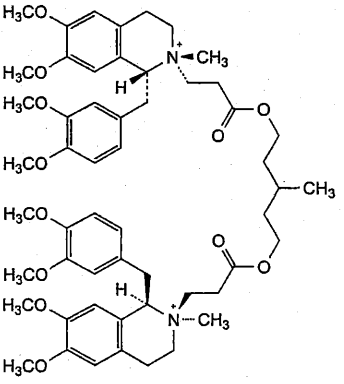
G. 2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[(1*R*,2*S*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



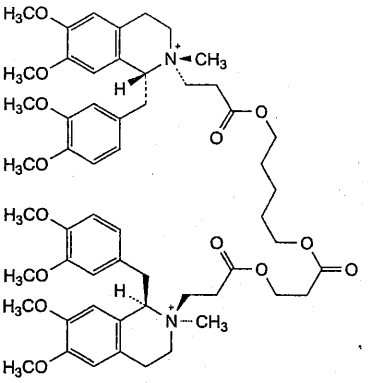
H. (1*R*,1′*R*,2*R*,2′*S*)-2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



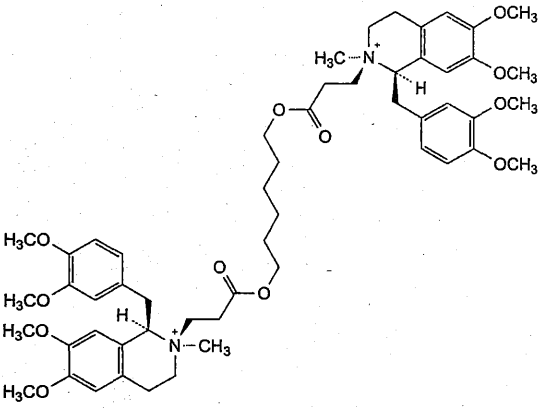
I. 2,2′-[[(1*RS*)-1-methylpentane-1,5-diyl]bis[oxy(3-oxopropane-3,1-diyl)]]bis[(1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



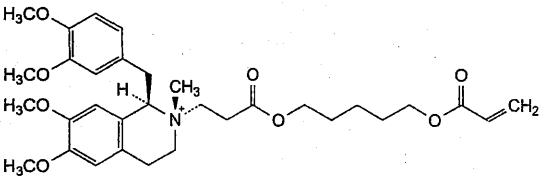
K. 2,2′-[(3-methylpentane-1,5-diyl)bis[oxy(3-oxopropane-3,1-diyl)]]bis[(1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



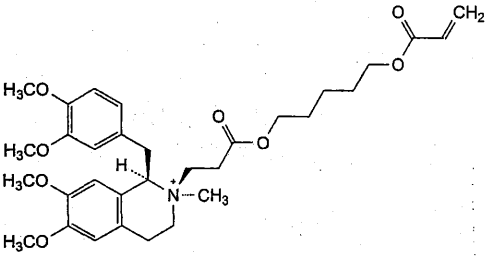
L. (1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-2-[3-[3-[[5-[[3-[(1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-3,4-dihydroisoquinolinium-2(1*H*)-yl]propanoyl]oxy]pentyl]oxy]-3-oxopropoxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



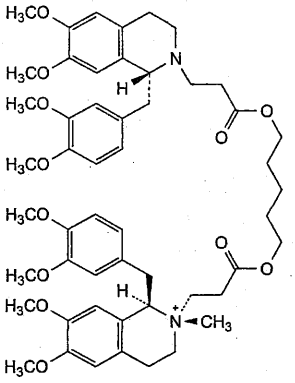
M. 2,2′-[hexane-1,6-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[(1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



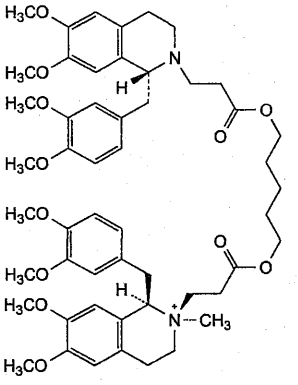
N. (1*R*,2*S*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-2-[3-[[5-(prop-2-enoyloxy)pentyl]oxy]-3-oxopropyl]-1,2,3,4-tetrahydroisoquinolinium,



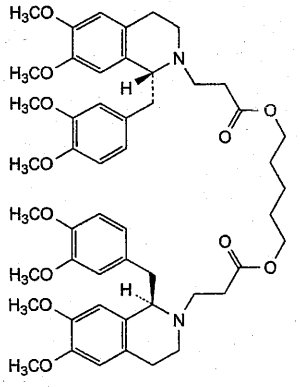
O. (1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-2-[3-[[5-(prop-2-enoyloxy)pentyl]oxy]-3-oxopropyl]-1,2,3,4-tetrahydroisoquinolinium,



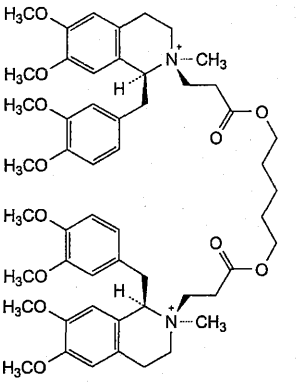
P. (1*R*,2*S*)-1-[(3,4-dimethoxyphenyl)methyl]-2-[3-[[5-[[3-[(1*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl]propanoyl]oxy]pentyl]oxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



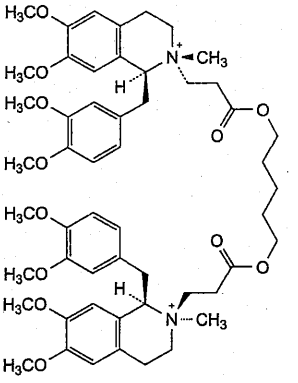
Q. (1*R*,2*R*)-1-[(3,4-dimethoxyphenyl)methyl]-2-[3-[[5-[[3-[(1*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl]propanoyl]oxy]pentyl]oxy]-3-oxopropyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium,



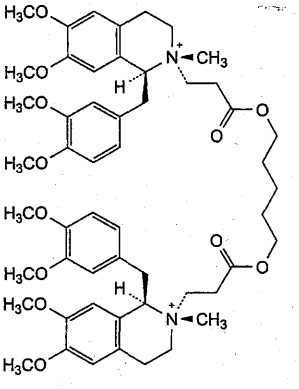
R. pentane-1,5-diyl bis[3-[(1*R*)-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl]propanoate],



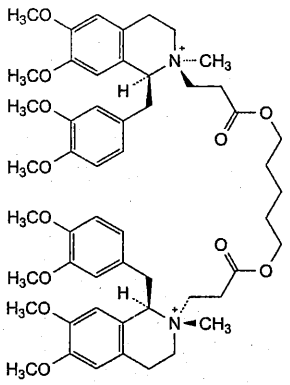
S. (1*R*,1′*S*,2*R*,2′*S*)-2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



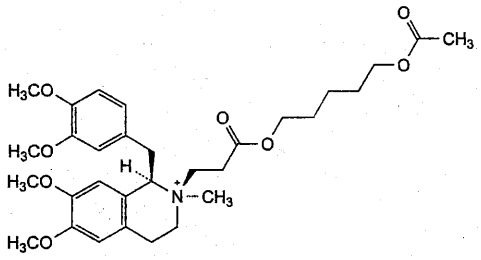
T. (1*R*,1′*S*,2*R*,2′*R*)-2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



U. (1*R*,1′*S*,2*S*,2′*R*)-2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],



V. (1*S*,1′*R*,2*S*,2′*S*)-2,2′-[pentane-1,5-diylbis[oxy(3-oxopropane-3,1-diyl)]]bis[1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinolinium],

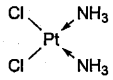


W. (1*R*,2*R*)-2-[3-[[5-(acetyloxy)pentyl]oxy]]-1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-2-methyl-3-oxopropyl-1,2,3,4-tetrahydroisoquinolinium.

Ph Eur

Cisplatin

(Ph. Eur. monograph 0599)



$PtCl_2(NH_3)_2$ 300.0 *15663-27-1*

Action and use
Platinum-containing cytotoxic.

Preparation
Cisplatin Injection

Ph Eur

DEFINITION
cis-Diamminedichloroplatinum(II).

Content
97.0 per cent to 102.0 per cent.

CHARACTERS
Appearance
Yellow powder, or yellow or orange-yellow crystals.

Solubility
Slightly soluble in water, sparingly soluble in dimethylformamide, practically insoluble in ethanol (96 per cent).

Carry out identification test B, the tests (except that for silver) and the assay protected from light.

IDENTIFICATION
First identification: A, B.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cisplatin CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dilute 1 mL of solution S2 (see Tests) to 10 mL with *dimethylformamide R.*

Reference solution Dissolve 10 mg of *cisplatin CRS* in 5 mL of *dimethylformamide R.*

Plate *cellulose for chromatography R1* as the coating substance.

Pretreatment Activate the plate by heating at 150 °C for 1 h.

Mobile phase *acetone R, dimethylformamide R* (10:90 *V/V*).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 50 g/L solution of *stannous chloride R* in a mixture of equal volumes of *dilute hydrochloric acid R* and *water R.* Examine after 1 h.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Add 50 mg to 2 mL of *dilute sodium hydroxide solution R* in a glass dish. Evaporate to dryness. Dissolve the residue in a mixture of 0.5 mL of *nitric acid R* and 1.5 mL of *hydrochloric acid R.* Evaporate to dryness. The residue is orange. Dissolve the residue in 0.5 mL of *water R* and add 0.5 mL of *ammonium chloride solution R.* A yellow, crystalline precipitate is formed.

TESTS
Solution S1
Dissolve 25 mg in a 9 g/L solution of *sodium chloride R* in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Solution S2
Dissolve 0.20 g in *dimethylformamide R* and dilute to 10 mL with the same solvent.

Appearance of solution S1
Solution S1 is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 (*2.2.2, Method II*).

Appearance of solution S2
Solution S2 is clear (*2.2.1*).

pH (*2.2.3*)
4.5 to 6.0 for solution S1, measured immediately after preparation.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light. Do not heat or sonicate any platinum-containing solution. All solutions are to be used within 4 h.*

Test solution Dissolve 25.0 mg of the substance to be examined in a 9.0 g/L solution of *sodium chloride R* and dilute to 25.0 mL with the same solution.

Reference solution (a) Dissolve 25.0 mg of *cisplatin CRS* in a 9.0 g/L solution of *sodium chloride R* and dilute to 25.0 mL with the same solution.

Reference solution (b) Dissolve 5.0 mg of *cisplatin impurity A CRS* in a 9.0 g/L solution of *sodium chloride R* and dilute to 50.0 mL with the same solution.

Reference solution (c) Dissolve 5.6 mg of *cisplatin impurity B CRS* in a 9.0 g/L solution of *sodium chloride R* and dilute to 100.0 mL with the same solution.

Reference solution (d) Mix 0.05 mL of the test solution with 5.0 mL of reference solution (b) and 5.0 mL of reference solution (c) and dilute to 25.0 mL with a 9.0 g/L solution of *sodium chloride R*.

Reference solution (e) Dilute 5.0 mL of reference solution (d) to 20.0 mL with a 9.0 g/L solution of *sodium chloride R*.

Blank solution 9.0 g/L solution of *sodium chloride R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (4 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 1.08 g of *sodium octanesulfonate R*, 1.70 g of *tetrabutylammonium hydrogen sulfate R* and 2.72 g of *potassium dihydrogen phosphate R* in *water for chromatography R* and dilute to 950 mL with the same solvent. Adjust to pH 5.9 with *1 M sodium hydroxide* and dilute to 1000 mL with *water for chromatography R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution, reference solutions (d) and (e), and the blank solution.

Run time 7 times the retention time of cisplatin.

The displacement peak is the latest eluting peak of the group of injection peaks in the chromatogram obtained with the blank solution.

Identification of cisplatin aquo complex Use the chromatogram supplied with *cisplatin CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to cisplatin aquo complex.

Relative retention With reference to cisplatin (retention time = about 3.8 min): displacement peak = about 0.5; impurity A = about 0.6; impurity B = about 0.7; cisplatin aquo complex = about 1.2.

System suitability Reference solution (d):
— *resolution*: minimum 2.5 between the peaks due to impurities A and B, the displacement peak and the peak due to impurity A are well separated.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (2.0 per cent);
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (1.0 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the peak due to cisplatin in the chromatogram obtained with reference solution (d) (0.10 per cent);
— *sum of impurities other than A and B*: not more than 2.5 times the area of the peak due to cisplatin in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *disregard limit*: the area of the peak due to cisplatin in the chromatogram obtained with reference solution (e) (0.05 per cent). Disregard any peak due to the cisplatin aquo complex.

Silver

Maximum 250 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 0.100 g in 15 mL of *nitric acid R*, heating to 80 °C. Cool and dilute to 25.0 mL with *water R*.

Reference solutions To suitable volumes (10 mL to 30 mL) of *silver standard solution (5 ppm Ag) R* add 50 mL of *nitric acid R* and dilute to 100.0 mL with *water R*.

Source Silver hollow-cathode lamp, preferably using a transmission band of 0.5 nm.

Wavelenth 328 nm.

Atomisation device Fuel-lean air-acetylene flame.

Carry out a blank determination.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $PtCl_2(NH_3)_2$ from the sum of the areas of the peaks due to cisplatin and cisplatin aquo complex and from the declared content of *cisplatin CRS*.

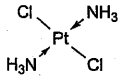
STORAGE

In an airtight container, protected from light.

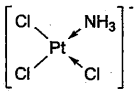
IMPURITIES

Specified impurities *A, B.*

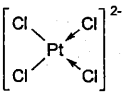
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



A. *trans*-diamminedichloroplatinum(II) (transplatin),



B. amminetrichloroplatinate(–),

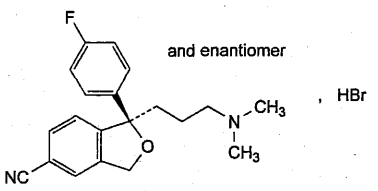


C. tetrachloroplatinate(2–).

Ph Eur

Citalopram Hydrobromide

(Ph. Eur. monograph 2288)



$C_{20}H_{22}BrFN_2O$ 405.3 *59729-32-7*

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Preparation
Citalopram Tablets

Ph Eur

DEFINITION
(1*RS*)-1-[3-(Dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-5-carbonitrile hydrobromide.

Content
99.0 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water and in anhydrous ethanol.

IDENTIFICATION
A. Optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *citalopram hydrobromide CRS.*

C. It gives reaction (a) of bromides (*2.3.1*).

TESTS
Optical rotation (*2.2.7*)
−0.10° to + 0.10°.

Dissolve 1.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A (solution A). Dilute 1.0 mL of solution A to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *citalopram for system suitability CRS* (containing impurities B, D and G) in 1.0 mL of solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (4 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.58 g of *ammonium formate R* in 500 mL of a mixture of 4 volumes of *acetonitrile for chromatography R*, 32 volumes of *methanol R1* and 64 volumes of *water for chromatography R*;
— *mobile phase B*: dissolve 1.58 g of *ammonium formate R* in 500 mL of a mixture of 32 volumes of *water for chromatography R* and 68 volumes of *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 25	100 → 40	0 → 60
25 - 30	40	60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm and, for impurity G, at 254 nm.

Injection 40 µL.

Identification of impurities Use the chromatogram supplied with *citalopram for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, D and G.

Relative retention With reference to citalopram (retention time = about 19 min): impurity G = about 0.5; impurity B = about 0.7; impurity D = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity D and citalopram at 230 nm.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity G by 0.6;
— *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity B*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *impurity G at 254 nm*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than G*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

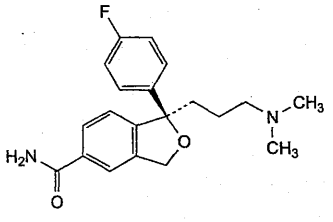
ASSAY
Dissolve 0.300 g in 50 mL of *ethanol (96 per cent) R* and add 0.5 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 40.53 mg of $C_{20}H_{22}BrFN_2O$.

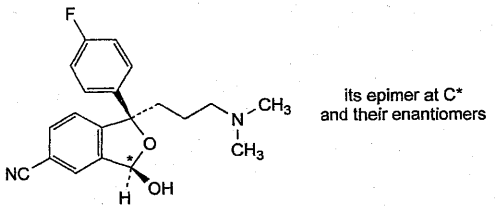
IMPURITIES
Specified impurities *B, D, G.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests

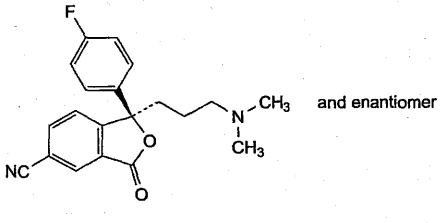
in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, E, F.*



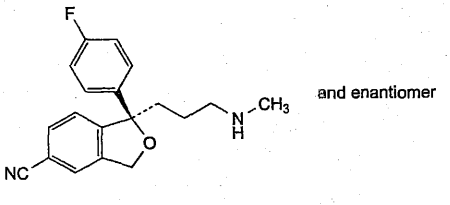
A. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-5-carboxamide,



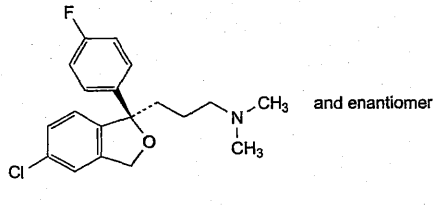
B. 1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3dihydroisobenzofuran-5-carbonitrile,



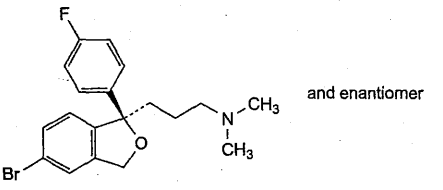
C. (3*RS*)-6-cyano-3-[3-(dimethylamino)propyl]-3-(4-fluorophenyl)isobenzofuran-1(3*H*)-one,



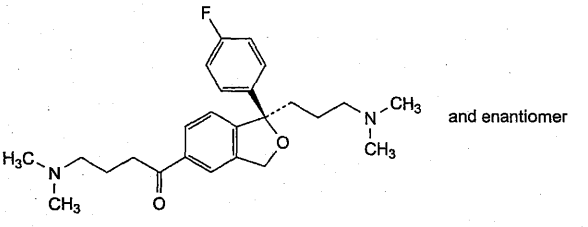
D. (1*RS*)-1-(4-fluorophenyl)-1-[3-(methylamino)propyl]-1,3-dihydroisobenzofuran-5-carbonitrile,



E. 3-[(1*RS*)-5-chloro-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine,



F. 3-[(1*RS*)-5-bromo-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine,

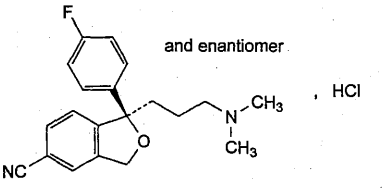


G. 4-(dimethylamino)-1-[(1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-5-yl]butan-1-one.

Ph Eur

Citalopram Hydrochloride

(Ph. Eur. monograph 2203)



$C_{20}H_{22}ClFN_2O$ 360.9 *85118-27-0*

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Preparation
Citalopram Oral Drops

Ph Eur

DEFINITION
(1*RS*)-1-[3-(Dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-5-carbonitrile hydrochloride.

Content
99.0 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in anhydrous ethanol.

IDENTIFICATION
A. Optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *citalopram hydrochloride CRS.*

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S, examined immediately after preparation, is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A (solution A). Dilute 1.0 mL of solution A to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *citalopram for system suitability CRS* (impurities B and D) in 1.0 mL of solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (4 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.58 g of *ammonium formate R* in 500 mL of a mixture of 4 volumes of *acetonitrile for chromatography R*, 32 volumes of *methanol R1* and 64 volumes of *water for chromatography R*;
— *mobile phase B*: dissolve 1.58 g of *ammonium formate R* in 500 mL of a mixture of 32 volumes of *water for chromatography R* and 68 volumes of *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 25	100 → 40	0 → 60
25 - 30	40	60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 40 µL.

Identification of impurities Use the chromatogram supplied with *citalopram for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and D.

Relative retention With reference to citalopram (retention time = about 19 min): impurity B = about 0.7; impurity D = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity D and citalopram.

Limits:
— *impurity B*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

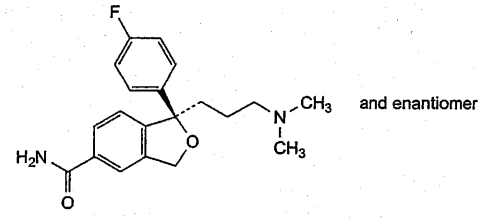
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 0.5 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

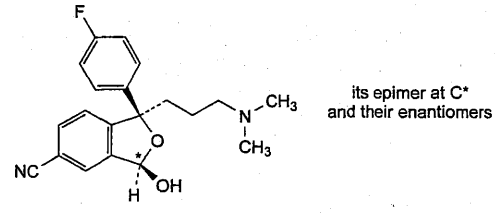
1 mL of *0.1 M sodium hydroxide* is equivalent to 36.09 mg of $C_{20}H_{22}ClFN_2O$.

IMPURITIES
Specified impurities *B.*

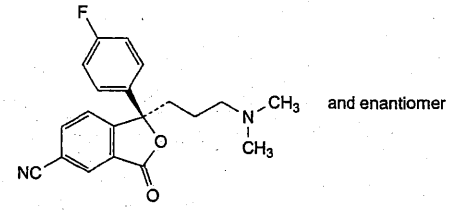
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, F.*



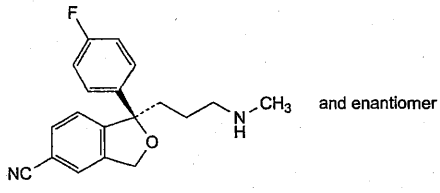
A. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-5-carboxamide,



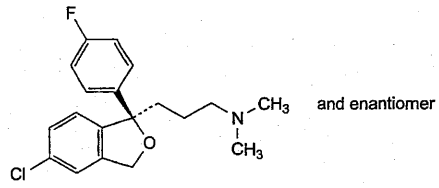
B. 1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3-dihydroisobenzofuran-5-carbonitrile,



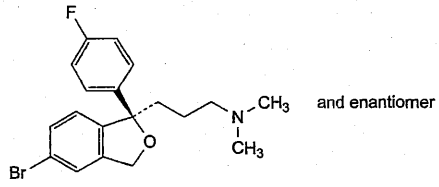
C. (3*RS*)-6-cyano-3-[3-(dimethylamino)propyl]-3-(4-fluorophenyl)isobenzofuran-1(3*H*)-one,



D. (1*RS*)-1-(4-fluorophenyl)-1-[3-(methylamino)propyl]-1,3-dihydroisobenzofuran-5-carbonitrile,



E. 3-[(1*RS*)-5-chloro-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine,



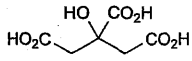
F. 3-[(1*RS*)-5-bromo-1-(4-fluorophenyl)-1,3-dihydroisobenzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine.

Ph Eur

Citric Acid[1]

Anhydrous Citric Acid

(Ph. Eur. monograph 0455)



$C_6H_8O_7$ 192.1 *77-92-9*

Ph Eur

DEFINITION

2-Hydroxypropane-1,2,3-tricarboxylic acid.

Content

99.5 per cent to 100.5 per cent (anhydrous substance).

♦CHARACTERS

Appearance

White or almost white, crystalline powder, colourless crystals or granules.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

mp

About 153 °C, with decomposition.♦

IDENTIFICATION

First identification: B,◊E.

Second identification: A, C, D, E.

A. Dissolve 1 g in 10 mL of *water R*. The solution is strongly acidic (*2.2.4*).◊

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dry the substance to be examined and the reference substance at 105 ± 2 °C for 2 h.

Comparison *anhydrous citric acid CRS*.

◊C. Add about 5 mg to a mixture of 1 mL of *acetic anhydride R* and 3 mL of *pyridine R*. A red colour develops.

D. Dissolve 0.5 g in 5 mL of *water R*, neutralise using *1 M sodium hydroxide* (about 7 mL), add 10 mL of *calcium chloride solution R* and heat to boiling. A white precipitate is formed.

E. Water (see Tests).◊

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless or not more intensely coloured than reference solution Y_7, BY_7 or GY_7 (*2.2.2, Method II*).

Dissolve 2.0 g in *water R* and dilute to 10 mL with the same solvent.

Readily carbonisable substances

To 1.0 g in a cleaned test tube add 10 mL of *sulfuric acid R* and immediately heat the mixture in a water-bath at 90 ± 1 °C for 60 min. Cool rapidly immediately afterwards. The solution is not more intensely coloured than a mixture of 1 mL of red primary solution and 9 mL of yellow primary solution (*2.2.2, Method I*).

Oxalic acid

Maximum 360 ppm, calculated as anhydrous oxalic acid.

Dissolve 0.80 g in 4 mL of *water R*. Add 3 mL of *hydrochloric acid R* and 1 g of *zinc R* in granules. Boil for 1 min. Allow to stand for 2 min. Transfer the supernatant to a test-tube containing 0.25 mL of a 10 g/L solution of *phenylhydrazine hydrochloride R* and heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an equal volume of *hydrochloric acid R* and 0.25 mL of a 50 g/L solution of *potassium ferricyanide R*. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using 4 mL of a 0.1 g/L solution of *oxalic acid R*.

Sulfates (*2.4.13*)

Maximum 150 ppm.

Dissolve 2.0 g in *distilled water R* and dilute to 30 mL with the same solvent.

Aluminium (*2.4.17*)

Maximum 0.2 ppm, if intended for use in the manufacture of dialysis solutions.

Prescribed solution Dissolve 20 g in 100 mL of *water R* and add 10 mL of *acetate buffer solution pH 6.0 R*.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al) R*, 10 mL of *acetate buffer solution pH 6.0 R* and 98 mL of *water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *water R*.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 2.000 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

♦Bacterial endotoxins (*2.6.14*)

Less than 0.5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.♦

[1] *This monograph has undergone* ***pharmacopoeial harmonisation.***
See chapter 5.8 ***Pharmacopoeial harmonisation.***

ASSAY

Dissolve 0.550 g in 50 mL of *water R*. Titrate with *1 M sodium hydroxide*, using 0.5 mL of *phenolphthalein solution R* as indicator.

1 mL of *1 M sodium hydroxide* is equivalent to 64.03 mg of $C_6H_8O_7$.

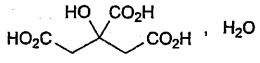
♦LABELLING

The label states, where applicable, that the substance is intended for use in the manufacture of dialysis solutions.♦

Ph Eur

Citric Acid Monohydrate[1]

(Ph. Eur. monograph 0456)



$C_6H_8O_7,H_2O$ 210.1 *5949-29-1*

Ph Eur

DEFINITION

2-Hydroxypropane-1,2,3-tricarboxylic acid monohydrate.

Content

99.5 per cent to 100.5 per cent (anhydrous substance).

♦CHARACTERS

Appearance

White or almost white, crystalline powder, colourless crystals or granules, efflorescent.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent). ♦

IDENTIFICATION

First identification: B,◊E.

Second identification: A, C, D, E.

A. Dissolve 1 g in 10 mL of *water R*. The solution is strongly acidic (*2.2.4*).◊

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dry the substance to be examined and the reference substance at 105 ± 2 °C for 2 h.

Comparison *citric acid monohydrate CRS*.

◊C. Add about 5 mg to a mixture of 1 mL of *acetic anhydride R* and 3 mL of *pyridine R*. A red colour develops.

D. Dissolve 0.5 g in 5 mL of *water R*, neutralise using *1 M sodium hydroxide* (about 7 mL), add 10 mL of *calcium chloride solution R* and heat to boiling. A white precipitate is formed.

E. Water (see Tests).◊

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless or not more intensely coloured than reference solution Y_7, BY_7 or GY_7 (*2.2.2, Method II*).

Dissolve 2.0 g in *water R* and dilute to 10 mL with the same solvent.

Readily carbonisable substances

To 1.0 g in a cleaned test tube add 10 mL of *sulfuric acid R* and immediately heat the mixture in a water-bath at 90 ± 1 °C for 60 min. Cool rapidly immediately afterwards. The solution is not more intensely coloured than a mixture of 1 mL of red primary solution and 9 mL of yellow primary solution (*2.2.2, Method I*).

Oxalic acid

Maximum 360 ppm, calculated as anhydrous oxalic acid.

Dissolve 0.80 g in 4 mL of *water R*. Add 3 mL of *hydrochloric acid R* and 1 g of *zinc R* in granules. Boil for 1 min. Allow to stand for 2 min. Transfer the supernatant to a test-tube containing 0.25 mL of a 10 g/L solution of *phenylhydrazine hydrochloride R* and heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an equal volume of *hydrochloric acid R* and 0.25 mL of a 50 g/L solution of *potassium ferricyanide R*. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using 4 mL of a 0.1 g/L solution of *oxalic acid R*.

Sulfates (*2.4.13*)

Maximum 150 ppm.

Dissolve 2.0 g in *distilled water R* and dilute to 30 mL with the same solvent.

Aluminium (*2.4.17*)

Maximum 0.2 ppm, if intended for use in the manufacture of dialysis solutions.

Prescribed solution Dissolve 20 g in 100 mL of *water R* and add 10 mL of *acetate buffer solution pH 6.0 R*.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al) R*, 10 mL of *acetate buffer solution pH 6.0 R* and 98 mL of *water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *water R*.

Water (*2.5.12*)

7.5 per cent to 9.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

♦Bacterial endotoxins (*2.6.14*)

Less than 0.5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.♦

ASSAY

Dissolve 0.550 g in 50 mL of *water R*. Titrate with *1 M sodium hydroxide*, using 0.5 mL of *phenolphthalein solution R* as indicator.

1 mL of *1 M sodium hydroxide* is equivalent to 64.03 mg of $C_6H_8O_7$.

♦STORAGE

In an airtight container.

LABELLING

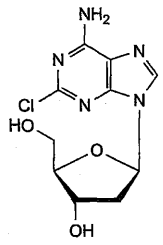
The label states, where applicable, that the substance is intended for use in the manufacture of dialysis solutions.♦

Ph Eur

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Cladribine

(Ph. Eur. monograph 2174)



$C_{10}H_{12}ClN_5O_3$ 285.7 *4291-63-8*

Action and use
Purine analogue; cytostatic.

Ph Eur

DEFINITION

2-Chloro-9-(2-deoxy-β-D-*erythro*-pentofuranosyl)-9*H*-purin-6-amine.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, soluble in dimethyl sulfoxide, slightly soluble in methanol, practically insoluble in acetonitrile.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cladribine CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined in the minimum volume of *methanol R* and evaporate to dryness. Dry the precipitate at 100 °C for 2 h and record a new spectrum using the residue.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Disperse 0.15 g in *water R*, dilute to 50 mL with the same solvent and sonicate until dissolution is complete.

Specific optical rotation (*2.2.7*)
−21.0 to −27.0 (anhydrous substance).

Dissolve 0.25 g in *dimethyl sulfoxide R* and dilute to 25.0 mL with the same solvent.

Impurity E
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 40.0 mg of the substance to be examined in *dimethylformamide R* and dilute to 2.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of 2-deoxy-D-ribose R (impurity E) in *dimethylformamide R* and dilute to 25.0 mL with the same solvent. Dilute 3.0 mL of this solution to 10.0 mL with *dimethylformamide R.*

Reference solution (b) Dissolve 10.0 mg of 2-deoxy-D-ribose R (impurity E) in *dimethylformamide R* and dilute to 5.0 mL with the same solvent. Mix 9 volumes of this solution with 1 volume of the test solution.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R, ethanol (96 per cent) R, ethyl acetate R* (20:40:40 *V/V/V*).

Application 5 µL as bands of 10 mm; thoroughly dry the points of application in a current of warm air.

Development Over 2/3 of the plate.

Drying In air, then heat at 45 °C for 10 min.

Detection Spray with a solution containing 0.5 g of *thymol R* in a mixture of 5 mL of *sulfuric acid R* and 95 mL of *ethanol (96 per cent) R*; heat at 110 °C for 20 min or until the spots appear.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Limit:
— *impurity E*: any spot due to impurity E is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.3 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (10:90 *V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of *cladribine CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 1.0 mg of *cladribine impurity C CRS* in reference solution (b) and dilute to 25.0 mL with the same solution.

Reference solution (e) Dilute 5.0 mL of reference solution (c) to 10.0 mL with the solvent mixture.

Reference solution (f) Dissolve 3 mg of *cladribine for peak identification CRS* (containing impurities A, B, C and D) in 2 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *water for chromatography R*;
— *mobile phase B*: *acetonitrile for chromatography R*;
— *mobile phase C*: 50 g/L solution of *phosphoric acid R* in *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 10	80 → 70	10 → 20	10
10 - 25	70 → 20	20 → 70	10
25 - 30	20	70	10

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 252 nm.

Injection 20 µL of test solution (a) and reference solutions (c), (d), (e) and (f).

Identification of impurities Use the chromatogram supplied with *cladribine for peak identification CRS* and the chromatogram obtained with reference solution (f) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to cladribine (retention time = about 10 min): impurity A = about 0.33; impurity B = about 0.44; impurity C = about 0.73; impurity D = about 0.92.

System suitability Reference solution (d):
— *resolution*: minimum 4.5 between the peaks due to impurity C and cladribine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.7; impurity C = 0.8;
— *impurities A, C*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurities B, D*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (e) (0.05 per cent).

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)
Less than 3 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{10}H_{12}ClN_5O_3$ from the declared content of *cladribine CRS*.

STORAGE

Protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

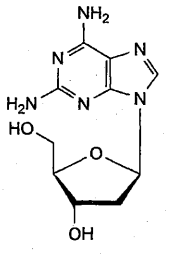
LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

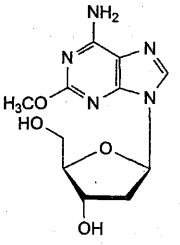
IMPURITIES

Specified impurities *A, B, C, D, E.*

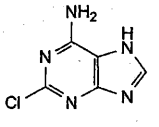
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G.*



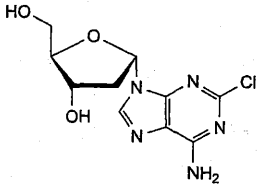
A. 9-(2-deoxy-β-D-*erythro*-pentofuranosyl)-9*H*-purin-2,6-diamine,



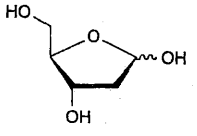
B. 9-(2-deoxy-β-D-*erythro*-pentofuranosyl)-2-methoxy-9*H*-purin-6-amine,



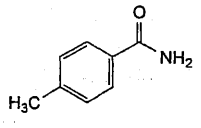
C. 2-chloro-7*H*-purin-6-amine (2-chloroadenine),



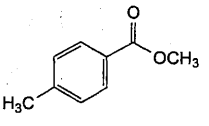
D. 2-chloro-9-(2-deoxy-α-D-*erythro*-pentofuranosyl)-9*H*-purin-6-amine,



E. 2-deoxy-D-*erythro*-pentofuranose (2-deoxy-D-ribose),



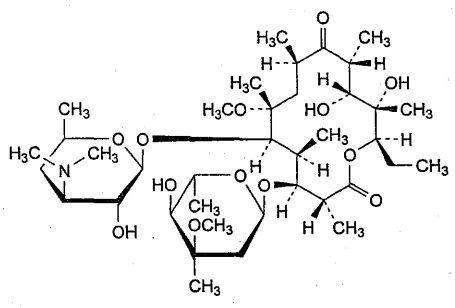
F. 4-methylbenzamide,



G. methyl 4-methylbenzoate.

Clarithromycin

(Ph. Eur. monograph 1651)



$C_{38}H_{69}NO_{13}$ 748 *81103-11-9*

Action and use
Macrolide antibacterial.

Preparations
Clarithromycin Granules for Oral Suspension
Clarithromycin for Infusion
Clarithromycin Tablets
Clarithromycin Prolonged-release Tablets

Ph Eur

DEFINITION

(3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-Dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-12,13-dihydroxy-7-methoxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (6-*O*-methylerythromycin A).

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetone and in methylene chloride, slightly soluble in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison: *clarithromycin CRS*.

TESTS

Solution S
Dissolve 0.500 g in *methylene chloride R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear or not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
−102 to −94 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 75.0 mg of the substance to be examined in 25 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R*.

Reference solution (a) Dissolve 75.0 mg of *clarithromycin CRS* in 25 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R*.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R*.

Reference solution (d) Dissolve 3.0 mg of *clarithromycin for peak identification CRS* in 1.0 mL of *acetonitrile R1* and dilute to 2.0 mL with *water R*.

Blank solution Dilute 25.0 mL of *acetonitrile R1* to 50.0 mL with *water R* and mix.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: a 4.76 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 4.4 with *dilute phosphoric acid R* or a 45 g/L solution of *potassium hydroxide R*, filtered through a C18 filtration kit;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 32	75 → 40	25 → 60
32 - 34	40	60

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 10 µL of the blank solution, the test solution and reference solutions (b), (c) and (d).

Relative retention r (not r_G) with reference to clarithromycin (retention time = about 11 min): impurity I = about 0.38; impurity A = about 0.42; impurity J = about 0.63; impurity L = about 0.74; impurity B = about 0.79; impurity M = about 0.81; impurity C = about 0.89; impurity D = about 0.96; impurity N = about 1.15; impurity E = about 1.27; impurity F = about 1.33; impurity P = about 1.35; impurity O = about 1.41; impurity K = about 1.59; impurity G = about 1.72; impurity H = about 1.82.

System suitability:
— *symmetry factor*: maximum 1.7 for the peak due to clarithromycin in the chromatogram obtained with reference solution (b);
— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clarithromycin in the chromatogram obtained with reference solution (d).

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity G = 0.27; impurity H = 0.15; use the chromatogram supplied with *clarithromycin for peak identification CRS* to identify the peaks;

— *any impurity*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent), and not more than 4 such peaks have an area greater than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.4 per cent);
— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent); disregard the peaks eluting before impurity I and after impurity H.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 0.5 g.

ASSAY

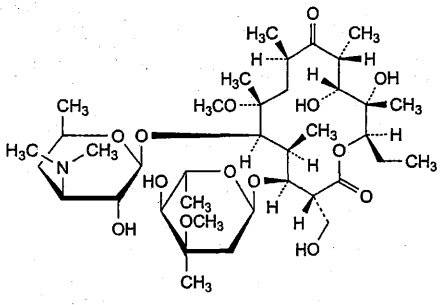
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

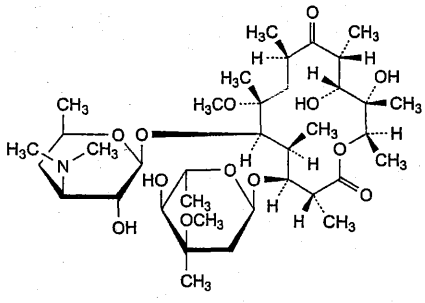
Calculate the percentage content of $C_{38}H_{69}NO_{13}$ taking into account the assigned content of *clarithromycin CRS*.

IMPURITIES

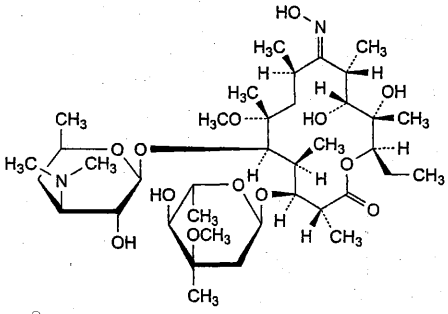
Specified impurities *A, B, C, D, E, F, G, H, I, J, K, L, M, N, O, P.*



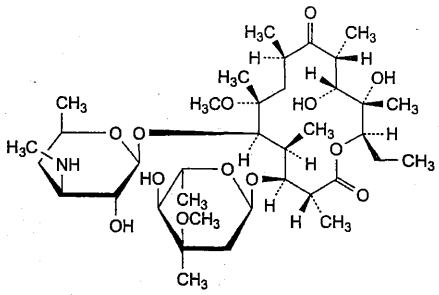
A. 2-demethyl-2-(hydroxymethyl)-6-*O*-methylerythromycin A (clarithromycin F),



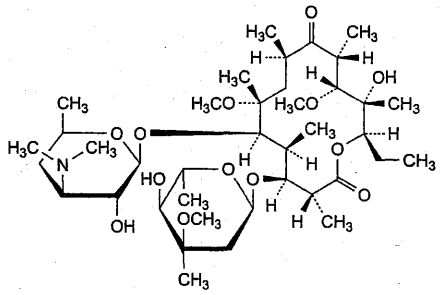
B. 6-*O*-methyl-15-norerythromycin A,



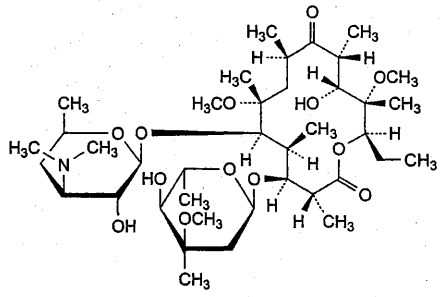
C. 6-*O*-methylerythromycin A (*E*)-9-oxime,



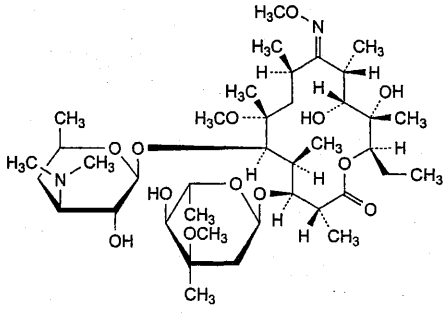
D. 3″-*N*-demethyl-6-*O*-methylerythromycin A,



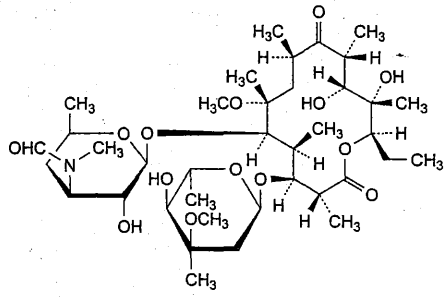
E. 6,11-di-*O*-methylerythromycin A,



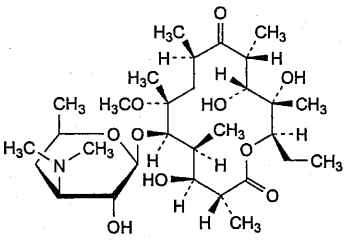
F. 6,12-di-*O*-methylerythromycin A,



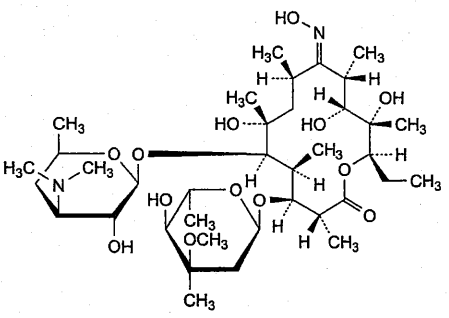
G. 6-*O*-methylerythromycin A (*E*)-9-(*O*-methyloxime),



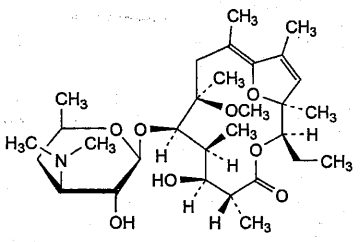
H. 3″-*N*-demethyl-3′-*N*-formyl-6-*O*-methylerythromycin A,



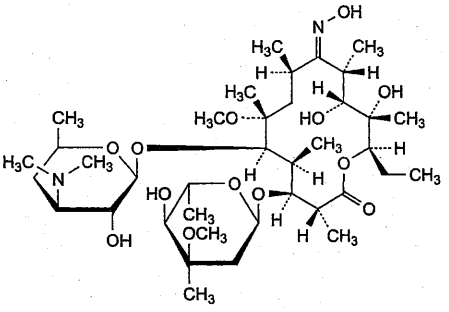
I. 3-*O*-decladinosyl-6-*O*-methylerythromycin A,



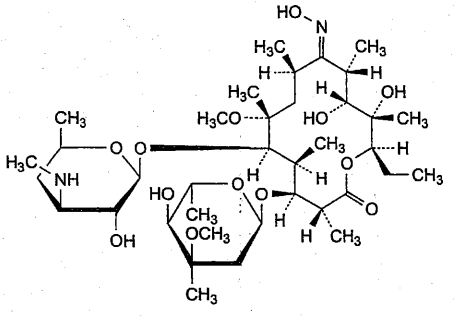
J. erythromycin A (*E*)-9-oxime,



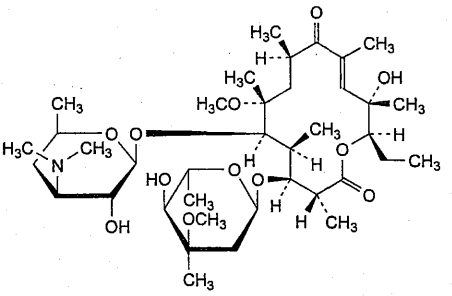
K. (1*S*,2*R*,5*R*,6*S*,7*S*,8*R*,9*R*,11*Z*)-2-ethyl-6-hydroxy-9-methoxy-1,5,7,9,11,13-hexamethyl-8-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-3,15-dioxabicyclo[10.2.1]pentadeca-11,13-dien-4-one (3-*O*-decladinosyl-8,9:10,11-dianhydro-6-*O*-methylerythromycin A-9,12-hemiketal),



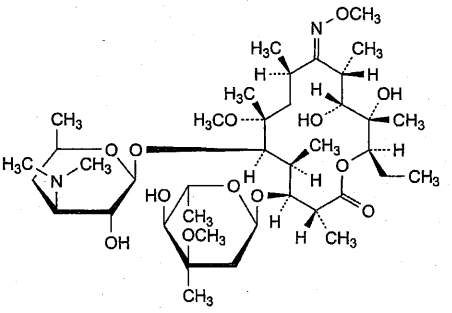
L. 6-*O*-methylerythromycin A (*Z*)-9-oxime,



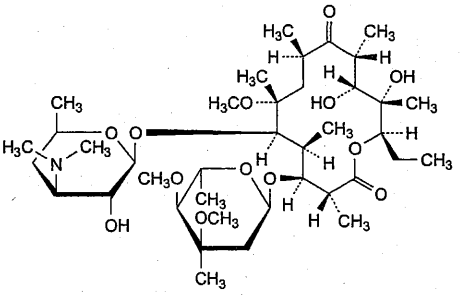
M. 3″-*N*-demethyl-6-*O*-methylerythromycin A (*E*)-9-oxime,



N. (10*E*)-10,11-didehydro-11-deoxy-6-*O*-methylerythromycin A,



O. 6-*O*-methylerythromycin A (*Z*)-9-(*O*-methyloxime),

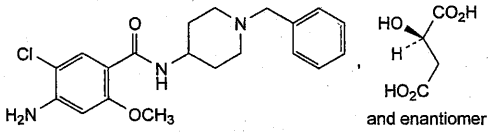


P. 4′,6-di-*O*-methylerythromycin A.

Ph Eur

Clebopride Malate

(Ph. Eur. monograph 1303)



$C_{24}H_{30}ClN_3O_7$ 508.0 *57645-91-7*

Action and use

Dopamine receptor antagonist; antiprotozoal (veterinary).

Ph Eur

DEFINITION

4-Amino-*N*-(1-benzylpiperidin-4-yl)-5-chloro-2-methoxybenzamide acid (*RS*)-2-hydroxybutanedioate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water and in methanol, slightly soluble in anhydrous ethanol, practically insoluble in methylene chloride.

mp

About 164 °C, with decomposition.

IDENTIFICATION

First identification: B, C.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 20.0 mg in *water R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *water R*.

Spectral range 230-350 nm.

Absorption maxima At 270 nm and 307 nm.

Specific absorbance at the absorption maxima:
— at 270 nm: 252 to 278;
— at 307 nm: 204 to 226.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clebopride malate CRS*.

C. Dissolve 20 mg in 1 mL of *sulfuric acid R*, add 1 mL of *β-naphthol solution R1* and mix. The solution examined in daylight is yellow with blue fluorescence.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *anhydrous ethanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *clebopride malate CRS* in *anhydrous ethanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *clebopride malate CRS* and 5 mg of *metoclopramide hydrochloride CRS* in *anhydrous ethanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *concentrated ammonia R, acetone R, methanol R, toluene R* (2:14:14:70 *V/V/V/V*).

Application 5 µL as bands of 10 mm by 3 mm.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated zones.

Results The principal zone in the chromatogram obtained with the test solution is similar in position and size to the principal zone in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S, examined immediately after preparation, is clear (*2.2.1*) and colourless (*2.2.2, Method I*).

pH (*2.2.3*)

3.8 to 4.2 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of *metoclopramide hydrochloride CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— *size*: l = 0.12 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 20 volumes of *acetonitrile R1* and 80 volumes of a 1 g/L solution of *sodium heptanesulfonate R* adjusted to pH 2.5 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time Twice the retention time of clebopride.

Relative retention With reference to clebopride (retention time = about 15 min): metoclopramide = about 0.45.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to metoclopramide and clebopride.

Limits:
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the 2 peaks eluting within the first 2 min.

Chlorides

Maximum 100 ppm.

Prepare the solutions at the same time.

Test solution Dissolve 0.530 g in 20.0 mL of *anhydrous acetic acid R*, add 6 mL of *dilute nitric acid R* and dilute to 50.0 mL with *water R*.

Reference solution To 1.5 mL of *0.001 M hydrochloric acid* add 20.0 mL of *anhydrous acetic acid R* and 6 mL of *dilute nitric acid R* and dilute to 50.0 mL with *water R*.

Transfer both recently prepared solutions to separate test-tubes. Add to each tube 1 mL of *silver nitrate solution R2*. Allow to stand for 5 min protected from light. Examine the tubes laterally against a black background. Any opalescence in the test solution is not more intense than that in the reference solution.

Sulfates

Maximum 100 ppm.

Prepare the solutions at the same time.

Test solution Dissolve 3.00 g in 20.0 mL of *glacial acetic acid R*, heating gently if necessary. Allow to cool and dilute to 50.0 mL with *water R*.

Reference solution To 9 mL of *sulfate standard solution (10 ppm SO_4) R1* add 6 mL of *glacial acetic acid R*.

Into 2 test-tubes introduce 1.5 mL of *sulfate standard solution (10 ppm SO_4) R1* and add 1 mL of a 250 g/L solution of *barium chloride R*. Shake and allow to stand for 1 min. To one of the tubes add 15 mL of the test solution and to the other add 15 mL of the reference solution. After 5 min, any opalescence in the tube containing the test solution is not more intense than that in the tube containing the reference solution.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

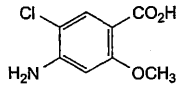
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.400 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

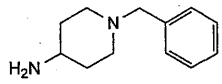
1 mL of *0.1 M perchloric acid* is equivalent to 50.80 mg of $C_{24}H_{30}ClN_3O_7$.

STORAGE
Protected from light.

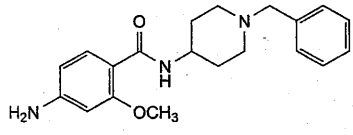
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



A. 4-amino-5-chloro-2-methoxybenzoic acid,



B. 1-benzylpiperidin-4-amine,

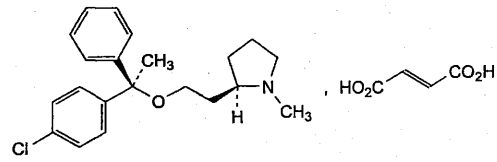


C. 4-amino-*N*-(1-benzylpiperidin-4-yl)-2-methoxybenzamide.

Ph Eur

Clemastine Fumarate

Clemastine Hydrogen Fumarate
(Ph. Eur. monograph 1190)



$C_{25}H_{30}ClNO_5$ 460.0 *14976-57-9*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparations
Clemastine Oral Solution
Clemastine Tablets

Ph Eur

DEFINITION
(2*R*)-2-[2-[(1*R*)-1-(4-Chlorophenyl)-1-phenylethoxy]ethyl]-1-methylpyrrolidine hydrogen (*E*)-butenedioate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, sparingly soluble in ethanol (70 per cent *V/V*), practically insoluble in heptane.

IDENTIFICATION
First identification: A, B.
Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *clemastine fumarate CRS.*

C. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 20.0 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.
Reference solution Dissolve 20.0 mg of *clemastine fumarate CRS* in *methanol R* and dilute to 10.0 mL with the same solvent.
Plate *TLC silica gel plate R.*
Mobile phase *concentrated ammonia R, methanol R, tetrahydrofuran R* (1:20:80 *V/V/V*).
Application 5 µL.
Development Over 2/3 of the plate.
Drying In a current of cold air for 5 min.
Detection Spray with a freshly prepared mixture of 1 volume of *potassium iodobismuthate solution R* and 10 volumes of *dilute acetic acid R* and then with *dilute hydrogen peroxide solution R*; cover the plate immediately with a glass plate of the same size and examine the chromatograms after 2 min.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 40 mg of the substance to be examined in *methanol R* and dilute to 2.0 mL with the same solvent.

Reference solution Dissolve 50 mg of *fumaric acid CRS* in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *water R, anhydrous formic acid R, di-isopropyl ether R* (5:25:70 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 100-105 °C for 30 min and allow to cool.

Detection Spray with a 16 g/L solution of *potassium permanganate R* and examine in daylight.

Results The principal spot with the highest R_F value in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Solution S

Dissolve 0.500 g in *methanol R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

pH (*2.2.3*)

3.2 to 4.2.

Suspend 1.0 g in 10 mL of *carbon dioxide-free water R.*

Specific optical rotation (*2.2.7*)

+ 15.0 to + 18.0 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Phosphate buffer solution pH 7.1 Mix 1.9 volumes of a 138 g/L solution of *sodium dihydrogen phosphate monohydrate R*, 6.8 volumes of an 89 g/L solution of *disodium hydrogen phosphate dihydrate R* and 91.3 volumes of *water for chromatography R.*

Solvent mixture *acetonitrile R1, water for chromatography R* (20:80 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in 30 mL of the solvent mixture with the aid of ultrasound and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *clemastine for system suitability CRS* (containing impurity B) in 1.0 mL of the solvent mixture with the aid of ultrasound for about 5 min.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped ethylene-bridged polar-embedded octadecylsilyl silica gel for chromatography (hybrid material) R* (3.5 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: phosphate buffer solution pH 7.1;
- *mobile phase B*: phosphate buffer solution pH 7.1, *acetonitrile R1* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	45	55
3 - 23	45 → 5	55 → 95
23 - 26	5	95

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 90 µL.

Identification of impurities Use the chromatogram supplied with *clemastine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to clemastine (retention time = about 17 min): fumaric acid = about 0.1; impurity B = about 0.9.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to impurity B and clemastine.

Calculation of percentage contents:
- for each impurity, use the concentration of clemastine in reference solution (a).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.2 per cent;
- *reporting threshold*: 0.05 per cent; disregard the peak due to fumaric acid.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 6 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

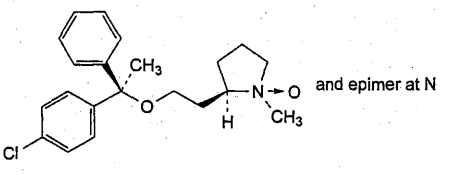
ASSAY

Dissolve 0.350 g in 60 mL of *anhydrous acetic acid R.* Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

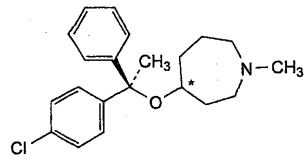
1 mL of *0.1 M perchloric acid* is equivalent to 46.00 mg of $C_{25}H_{30}ClNO_5$.

IMPURITIES

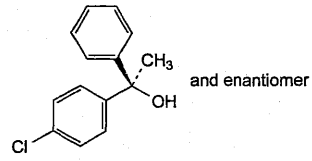
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



A. (1*RS*,2*R*)-2-[2-[(1*R*)-1-(4-chlorophenyl)-1-phenylethoxy]ethyl]-1-methylpyrrolidine 1-oxide,



B. 4-[(1*R*)-1-(4-chlorophenyl)-1-phenylethoxy]-1-methylazepane,

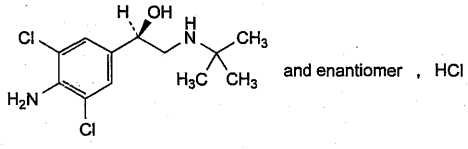


C. (1*RS*)-1-(4-chlorophenyl)-1-phenylethanol.

Ph Eur

Clenbuterol Hydrochloride

(Ph. Eur. monograph 1409)



$C_{12}H_{19}Cl_3N_2O$ 313.7 *21898-19-1*

Action and use
$Beta_2$-adrenoceptor agonist; bronchodilator.

Preparation
Clenbuterol Injection (VET)

Ph Eur

DEFINITION
(1*RS*)-1-(4-Amino-3,5-dichlorophenyl)-2-[(1,1-dimethylethyl) amino]ethanol hydrochloride.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water and in ethanol (96 per cent), slightly soluble in acetone.

mp
About 173 °C, with decomposition.

IDENTIFICATION
First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clenbuterol hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 10 mL of *methanol R.*

Reference solution Dissolve 10 mg of *clenbuterol hydrochloride CRS* in 10 mL of *methanol R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *ammonia R, anhydrous ethanol R, toluene R* (0.15:10:15 *V/V/V*).

Application 10 µL.

Development Over a path of 10 cm.

Drying In air.

Detection Spray with a 10 g/L solution of *sodium nitrite R* in *1 M hydrochloric acid* and dip after 10 min in a 4 g/L solution of *naphthylethylenediamine dihydrochloride R* in *methanol R.* Allow to dry in air.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 0.5 g in 10 mL of *carbon dioxide-free water R.*

Appearance of solution
Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
5.0 to 7.0 for solution S.

Optical rotation (*2.2.7*)
−0.10° to + 0.10°.

Dissolve 0.30 g in *water R* and dilute to 10.0 mL with the same solvent. Filter if necessary.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Disperse 100.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 0.1 mL of the test solution to 100.0 mL with *water R.*

Reference solution (b) Dissolve 5 mg of *clenbuterol impurity B CRS* in 10 mL of the mobile phase, add 2.5 mL of the test solution and dilute to 25.0 mL with the mobile phase.

Column:
— *size: l* = 0.125 m, Ø = 4 mm,
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm),
— *temperature:* 40 °C.

Mobile phase Mix 200 volumes of *acetonitrile R,* 200 volumes of *methanol R* and 600 volumes of a solution prepared as follows: dissolve 3.0 g of *sodium decanesulfonate R* and 5.0 g of *potassium dihydrogen phosphate R* in 900 mL of *water R,* adjust to pH 3.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R.*

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 5 µL.

Run time 1.5 times the retention time of clenbuterol.

Retention time Clenbuterol = about 29 min.

System suitability Reference solution (b):
— *resolution:* minimum 4.0 between the peaks due to impurity B and clenbuterol.

Limits:
— *impurities A, B, C, D, E, F:* for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),

— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

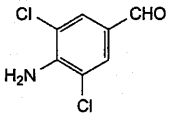
Water (*2.5.12*)
Maximum 1.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

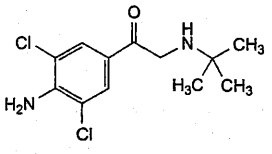
ASSAY
Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 31.37 mg of $C_{12}H_{19}Cl_3N_2O$.

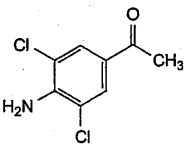
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



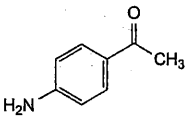
A. 4-amino-3,5-dichlorobenzaldehyde,



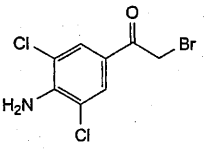
B. 1-(4-amino-3,5-dichlorophenyl)-2-[(1,1-dimethylethyl)amino]ethanone,



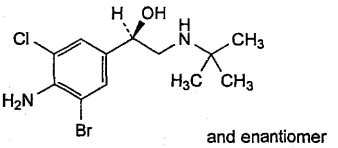
C. 1-(4-amino-3,5-dichlorophenyl)ethanone,



D. 1-(4-aminophenyl)ethanone,



E. 1-(4-amino-3,5-dichlorophenyl)-2-bromoethanone,

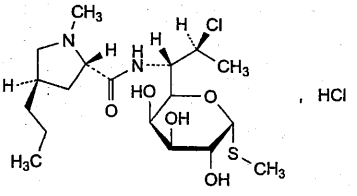


F. (1*RS*)-1-(4-amino-3-bromo-5-chlorophenyl)-2-[(1,1-dimethylethyl)amino]ethanol.

Ph Eur

Clindamycin Hydrochloride

(Ph. Eur. monograph 0582)



$C_{18}H_{34}Cl_2N_2O_5S$ 461.5 *21462-39-5*

Action and use
Lincosamide antibacterial.

Preparation
Clindamycin Capsules

Ph Eur

DEFINITION
Methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside hydrochloride. It contains a variable quantity of water.

Semi-synthetic product derived from a fermentation product.

Content
92.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder, slightly hygroscopic.

Solubility
Very soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clindamycin hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *clindamycin hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *clindamycin hydrochloride CRS* and 10 mg of *lincomycin hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase Mix 19 volumes of *2-propanol R*, 38 volumes of a 150 g/L solution of *ammonium acetate R* adjusted to pH 9.6 with *ammonia R*, and 43 volumes of *ethyl acetate R*. Use the upper layer.

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 1 g/L solution of *potassium permanganate R*.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 2 mL of *dilute hydrochloric acid R* and heat on a water-bath for 3 min. Add 3 mL of *sodium carbonate solution R* and 1 mL of a 20 g/L solution of *sodium nitroprusside R*. A violet-red colour develops.

D. Dissolve 0.1 g in *water R* and dilute to 10 mL with the same solvent. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)
3.0 to 5.0.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 135 to + 150 (anhydrous substance).

Dissolve 1.000 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *clindamycin hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 45 volumes of *acetonitrile R1* and 55 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.5 with a 250 g/L solution of *potassium hydroxide R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Run time Twice the retention time of clindamycin.

Relative retention With reference to clindamycin (retention time = about 10 min): impurity B = about 0.7; impurity C = about 0.8.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurities B and C; minimum 2.0 between the peaks due to impurity C and clindamycin.

Calculation of percentage contents:
— for each impurity, use the concentration of clindamycin hydrochloride in reference solution (b).

Limits:
— *impurity C*: maximum 4.0 per cent;
— *impurity B*: maximum 2.0 per cent;
— *any other impurity*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 6.0 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
3.0 per cent to 6.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{18}H_{34}Cl_2N_2O_5S$ taking into account the assigned content of *clindamycin hydrochloride CRS*.

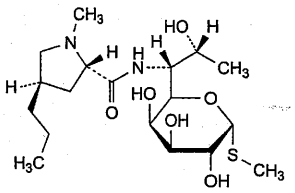
STORAGE

In an airtight container.

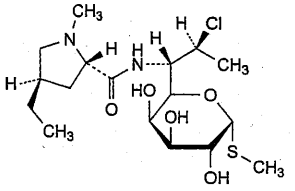
IMPURITIES

Specified impurities *B, C.*

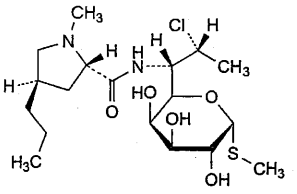
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, F.*



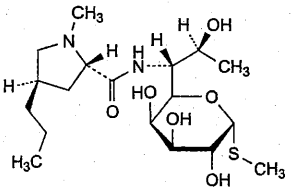
A. methyl 6,8-dideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (lincomycin),



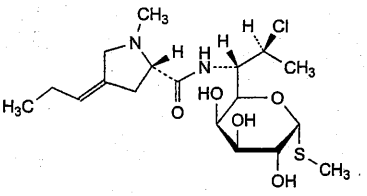
B. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-4-ethyl-1-methylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin B),



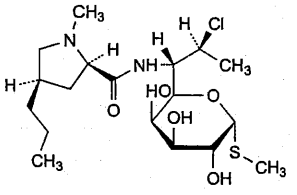
C. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (7-epiclindamycin),



D. methyl 6,8-dideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (7-epilincomycin),



E. methyl (5*R*)-5-[(1*S*,2*S*)-2-chloro-1-[[(4*Z*)-1-methyl-4-propylidene-L-prolyl]amino]propyl]-1-thio-β-L-arabinopyranoside,

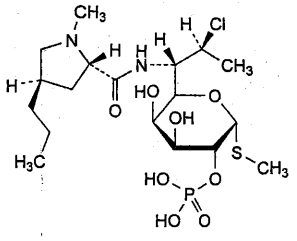


F. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*R*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside.

Ph Eur

Clindamycin Phosphate

(Ph. Eur. monograph 0996)



$C_{18}H_{34}ClN_2O_8PS$ 505.0 *24729-96-2*

Action and use
Lincosamide antibacterial.

Preparations
Benzoyl Peroxide and Clindamycin Gel
Clindamycin Injection

Ph Eur

DEFINITION
Methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-2-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside.

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, slightly hygroscopic powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clindamycin phosphate CRS.*

If the spectra obtained in the solid state show differences, dissolve 50 mg of the substance to be examined and the reference substance separately in 0.2 mL of *water R* and heat until completely dissolved. Evaporate to dryness under reduced pressure, dry the residues at 100-105 °C for 2 h and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *clindamycin phosphate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *lincomycin hydrochloride CRS* in 5 mL of reference solution (a).

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 100-105 °C for 30 min.

Detection Spray with a 1 g/L solution of *potassium permanganate R.*

System suitability Reference solution (b):
— the chromatogram shows 2 principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 2 mL of *dilute hydrochloric acid R* and heat in a water-bath for 3 min. Add 4 mL of *sodium carbonate solution R* and 1 mL of a 20 g/L solution of *sodium nitroprusside R.* Prepare a standard in the same manner using *clindamycin phosphate CRS.* The colour of the test solution corresponds to that of the standard.

D. Boil 0.1 g under a reflux condenser with a mixture of 5 mL of *strong sodium hydroxide solution R* and 5 mL of *water R* for 90 min. Cool and add 5 mL of *nitric acid R*. Extract with 3 quantities, each of 15 mL, of *methylene chloride R* and discard the extracts. Filter the upper layer through a paper filter. The filtrate gives reaction (b) of phosphates (*2.3.1*).

TESTS

Solution S

Dissolve 1.00 g in *carbon dioxide-free water R*. Heat gently if necessary. Cool and dilute to 25.0 mL with *carbon dioxide-free water R*.

Appearance of the solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 30.0 mg of *clindamycin phosphate CRS* in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 3.0 mg of *clindamycin phosphate for system suitability CRS* (containing impurities B, E, F, G, I, J, K and L) in mobile phase A and dilute to 1.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: mix 21 volumes of *acetonitrile R1* and 79 volumes of a 13.6 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 6.0 with a 450 g/L solution of *potassium hydroxide R*;
- *mobile phase B*: mix 40 volumes of a 13.6 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 6.0 with a 450 g/L solution of *potassium hydroxide R*, and 60 volumes of *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 13	100	0
13 - 18	100 → 50	0 → 50
18 - 39	50	50

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *clindamycin phosphate for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, E, F, G, I, J, K and L.

Relative retention With reference to clindamycin phosphate (retention time = about 12 min): impurity F = about 0.15; impurity G = about 0.19; impurity I = about 0.34; impurity B = about 0.45; impurity L = about 0.64; impurity J = about 1.20; impurity E = about 1.73; impurity K = about 1.90.

System suitability Reference solution (c):
- *resolution*: minimum 2.0 between the peaks due to impurities F and G.

Calculation of percentage contents:
- for each impurity, use the concentration of clindamycin phosphate in reference solution (b).

Limits:
- *impurities B, L*: for each impurity, maximum 1.0 per cent;
- *impurities E, F*: for each impurity, maximum 0.5 per cent;
- *impurities G, I, J, K*: for each impurity, maximum 0.2 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 2.0 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.200 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.6 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

System suitability Reference solution (a):
- *symmetry factor*: maximum 3.0 for the peak due to clindamycin phosphate.

Calculate the percentage content of $C_{18}H_{34}ClN_2O_8PS$ taking into account the assigned content of *clindamycin phosphate CRS*.

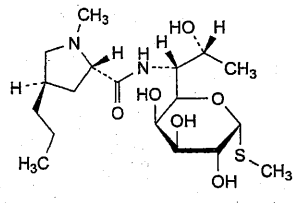
STORAGE

In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

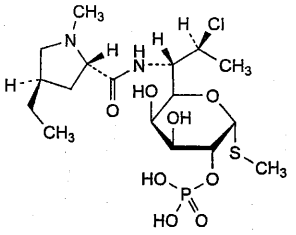
IMPURITIES

Specified impurities *B, E, F, G, I, J, K, L.*

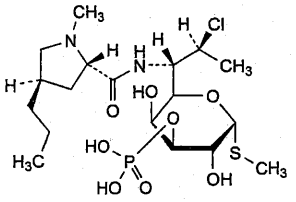
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, H.*



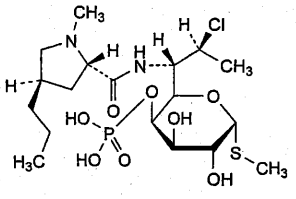
A. methyl 6,8-dideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (lincomycin),



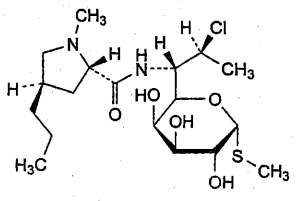
B. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-4-ethyl-1-methylpyrrolidin-2-yl]carbonyl]amino]-2-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin B 2-phosphate),



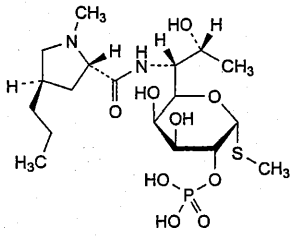
C. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-3-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin 3-phosphate),



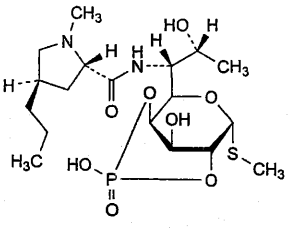
D. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-4-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin 4-phosphate),



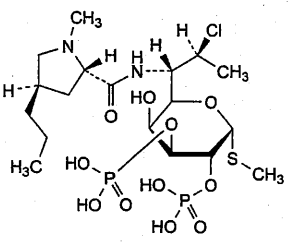
E. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin),



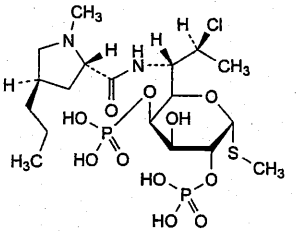
F. methyl 6,8-dideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-2-*O*-phosphono-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (lincomycin 2-phosphate),



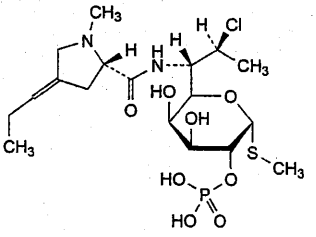
G. methyl 6,8-dideoxy-2,4-*O*-(hydroxyphosphoryl)-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (2,4-phosphatidyl lincomycin),



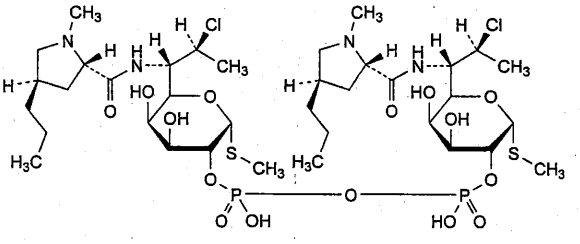
H. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-2,3-di-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin 2,3-bisphosphate),



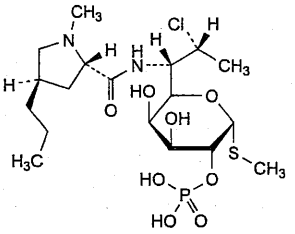
I. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-2,4-di-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (clindamycin 2,4-bisphosphate),



J. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*)-1-methyl-4-propylidenepyrrolidin-2-yl]carbonyl]amino]-2-*O*-phosphono-1-thio-L-*threo*-α-D-*galacto*-octopyranoside (propylidene analog of clindamycin 2-phosphate),



K. 2,2′-oxybis(hydroxyphosphoryl)bis[methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-*threo*-α-D-*galacto*-octopyranoside] (diclindamycin pyrophosphate),

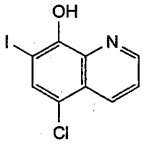


L. methyl 7-chloro-6,7,8-trideoxy-6-[[[(2*S*,4*R*)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-2-*O*-phosphono-1-thio-D-*erythro*-α-D-*galacto*-octopyranoside (7-epiclindamycin 2-phosphate).

Ph Eur

Clioquinol

(Ph. Eur. monograph 2111)



C_9H_5ClINO 305.5 *130-26-7*

Action and use
Antibacterial; antiprotozoal.

Preparations
Betamethasone and Clioquinol Cream
Betamethasone and Clioquinol Ointment
Flumetasone and Clioquinol Ear Drops
Hydrocortisone and Clioquinol Cream
Hydrocortisone and Clioquinol Ointment

Ph Eur

DEFINITION
5-Chloro-7-iodoquinolin-8-ol.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Almost white, light yellow, brownish-yellow or yellowish-grey powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, very slightly soluble or slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B.

Second identification: A, C, D.

A. Dissolve 40.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL to 100.0 mL with *methanol R* (solution A). Examined between 280 nm and 350 nm (*2.2.25*), solution A shows an absorption maximum at 321 nm. Dilute 10.0 mL of solution A to 100.0 mL with *methanol R* (solution B). Examined between 230 nm and 280 nm, solution B shows an absorption maximum at 255 nm. The specific absorbance at this absorption maximum is 1530 to 1660.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium bromide R.*

Comparison *clioquinol CRS.*

C. When heated, violet fumes are produced.

D. Dissolve about 1 mg in 5 mL of *ethanol (96 per cent) R.* Add 0.05 mL of *ferric chloride solution R1.* A dark green colour develops.

TESTS
Acidity or alkalinity
Shake 0.5 g with 10 mL of *carbon dioxide-free water R* and filter. To the filtrate add 0.2 mL of *phenolphthalein solution R.* The solution is colourless. Not more than 0.5 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *methanol R* and dilute to 50.0 mL with the same solvent, heating gently if necessary. Dilute 10.0 mL of the solution to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of 5-chloroquinolin-8-ol R, 10.0 mg of 5,7-dichloroquinolin-8-ol R, 5 mg of the substance to be examined and 10.0 mg of 5,7-diiodoquinolin-8-ol R in *methanol R*, heating gently if necessary and dilute to 20.0 mL with the same solvent. Dilute 4.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size: l* = 0.15 m, Ø = 3.9 mm,
— *stationary phase: octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 0.50 g of *sodium edetate R* in 350 mL of *water R*, add 4.0 mL of *hexylamine R* and mix. Adjust to pH 3.0 with *phosphoric acid R.* Add 600 mL of *methanol R* and dilute to 1000 mL with *water R.*

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 4 times the retention time of clioquinol.

Relative retention With reference to clioquinol (retention time = about 10 min): impurity A = about 0.4; impurity B = about 0.7; impurity C = about 1.3.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to clioquinol and impurity C.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (2.0 per cent),
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (1.0 per cent),
— *impurity C*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (1.0 per cent),

— *unspecified impurities*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent),
— *total of the nominal contents of impurities A, B, C and unspecified impurities*: maximum 3.0 per cent,
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Halides
Maximum 140 ppm, expressed as chlorides.

Shake 0.5 g with 25 mL of *water R* for 1 min and filter. To the filtrate add 0.5 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R2*. Allow to stand for 5 min. Any opalescence is not more intense than that in a standard prepared at the same time by adding 0.5 mL of *silver nitrate solution R2* to 25 mL of *water R* containing 0.2 mL of *0.01 M hydrochloric acid* and 0.5 mL of *dilute nitric acid R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 24 h.

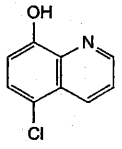
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 20 mL of *acetic anhydride R* and add 30 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

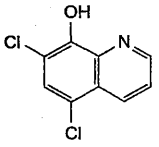
1 mL of *0.1 M perchloric acid* is equivalent to 30.55 mg of total quinolines, calculated as clioquinol.

STORAGE
Protected from light.

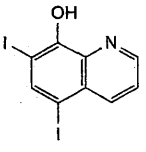
IMPURITIES
Specified impurities *A, B, C.*



A. 5-chloroquinolin-8-ol,



B. 5,7-dichloroquinolin-8-ol,

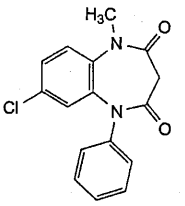


C. 5,7-diiodoquinolin-8-ol.

Ph Eur

Clobazam

(Ph. Eur. monograph 1974)



$C_{16}H_{13}ClN_2O_2$ 300.7 *22316-47-8*

Action and use
Benzodiazepine.

Preparations
Clobazam Capsules
Clobazam Oral Suspension

Ph Eur

DEFINITION
7-Chloro-1-methyl-5-phenyl-1,5-dihydro-3*H*-1,5-benzodiazepine-2,4-dione.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in methylene chloride, sparingly soluble in alcohol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of clobazam.*

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *clobazam impurity A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *chlordiazepoxide CRS* and 5 mg of *clonazepam CRS* in the mobile phase and dilute to 50 mL with the mobile phase. Dilute 1 mL of the solution to 100 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R, water R* (40:60 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 5 times the retention time of clobazam.

Retention time Clobazam = about 15 min.

System suitability Reference solution (b):
— *resolution*: minimum 1.3 between the peaks due to chlordiazepoxide and clonazepam.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *any other impurity*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent),
— *total of other impurities*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent),
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

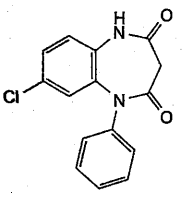
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

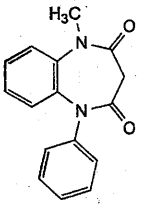
ASSAY
Dissolve 50.0 mg in *alcohol R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 250.0 mL with *alcohol R*. Measure the absorbance (*2.2.25*) at the maximum at 232 nm.

Calculate the content of $C_{16}H_{13}ClN_2O_2$ taking the specific absorbance to be 1380.

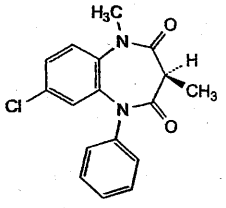
IMPURITIES



A. 7-chloro-5-phenyl-1,5-dihydro-3*H*-1,5-benzodiazepine-2,4-dione,

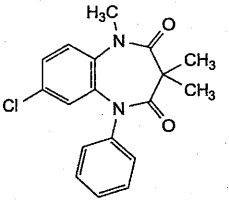


B. 1-methyl-5-phenyl-1,5-dihydro-3*H*-1,5-benzodiazepine-2,4-dione,

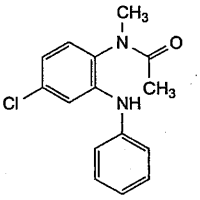


and enantiomer

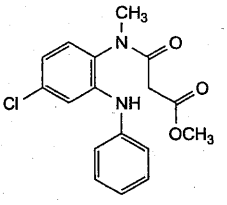
C. (3*RS*)-7-chloro-1,3-dimethyl-5-phenyl-1,5-dihydro-3*H*-1,5-benzodiazepine-2,4-dione,



D. 7-chloro-1,3,3-trimethyl-5-phenyl-1,5-dihydro-3*H*-1,5-benzodiazepine-2,4-dione,



E. *N*-[4-chloro-2-(phenylamino)phenyl]-*N*-methylacetamide,

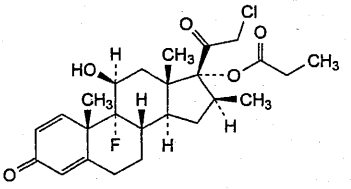


F. methyl 3-[[4-chloro-2-(phenylamino)phenyl] methylamino]-3-oxopropanoate.

Ph Eur

Clobetasol Propionate

(Ph. Eur. monograph 2127)



$C_{25}H_{32}ClFO_5$ 467.0 *25122-46-7*

Action and use
Glucocorticoid.

Preparations
Clobetasol Cutaneous Foam
Clobetasol Scalp Application
Clobetasol Shampoo

Ph Eur

DEFINITION
21-Chloro-9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clobetasol propionate CRS.*

TESTS
Specific optical rotation (*2.2.7*)
+ 112 to + 118 (dried substance).

Dissolve 0.500 g in *acetone R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of *clobetasol propionate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *clobetasol impurity J CRS* in 2.0 mL of the mobile phase. To 0.5 mL of this solution add 0.5 mL of test solution (b) and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *clobetasol for peak identification CRS* (containing impurities A, B, C, D, E, L and M) in 2 mL of the mobile phase.

Reference solution (d) Dilute 1.0 mL of test solution (a) to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Mix 10 volumes of *methanol R*, 42.5 volumes of a 7.85 g/L solution of *sodium dihydrogen phosphate monohydrate R* adjusted to pH 5.5 with a 100 g/L solution of *sodium hydroxide R* and 47.5 volumes of *acetonitrile R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL of test solution (a) and reference solutions (b), (c) and (d).

Run time 3 times the retention time of clobetasol propionate.

Identification of impurities Use the chromatogram supplied with *clobetasol for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D, E, L and M.

Relative retention With reference to clobetasol propionate (retention time = about 10 min): impurity A = about 0.4; impurity B = about 0.6; impurity C = about 0.9; impurity J = about 1.1; impurity D = about 1.2; impurity L = about 1.3; impurity M = about 1.6; impurity E = about 2.1.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to clobetasol propionate and impurity J in the chromatogram obtained with reference solution (b);
— the chromatogram obtained with reference solution (c) is similar to the chromatogram supplied with *clobetasol for peak identification CRS*.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.6; impurity C = 1.5;
— *impurity E*: not more than 1.4 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.7 per cent);
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *impurities B, C*: for each impurity, not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.3 per cent);
— *impurities A, L, M*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.10 per cent);
— *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (d) (2.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

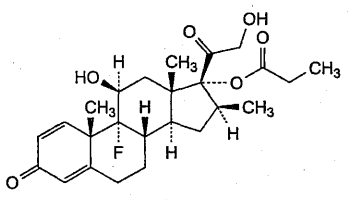
Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{25}H_{32}ClFO_5$ using the chromatogram obtained with reference solution (a) and the declared content of *clobetasol propionate CRS*.

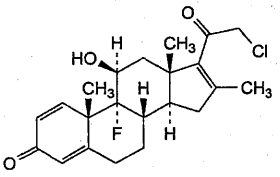
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C, D, E, L, M.*

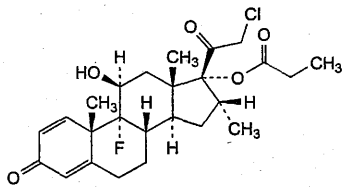
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G, H, I, J, K.*



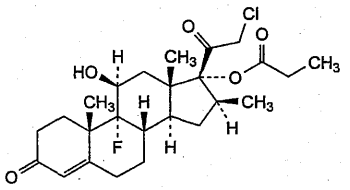
A. 9-fluoro-11β,21-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate (betamethasone 17-propionate),



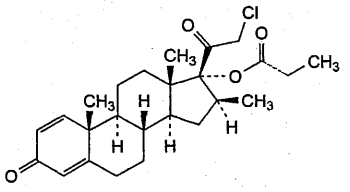
B. 21-chloro-9-fluoro-11β-hydroxy-16-methylpregna-1,4,16-triene-3,20-dione,



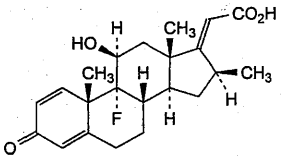
C. 21-chloro-9-fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate,



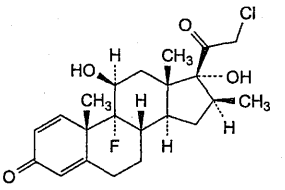
D. 21-chloro-9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregn-4-en-17-yl propanoate (1,2-dihydroclobetasol 17-propionate),



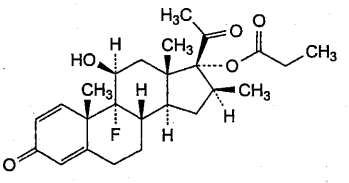
E. 21-chloro-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate,



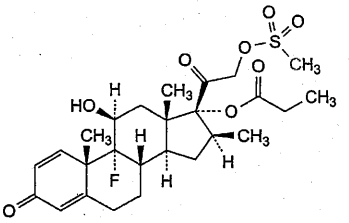
F. 9-fluoro-11β-hydroxy-16β-methyl-3-oxopregna-1,4,17(20)-trien-21-oic acid,



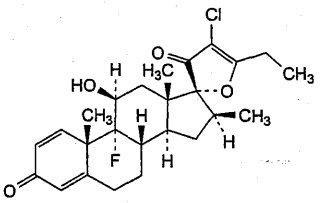
G. 21-chloro-9-fluoro-11β,17-dihydroxy-16β-methylpregna-1,4-diene-3,20-dione (clobetasol),



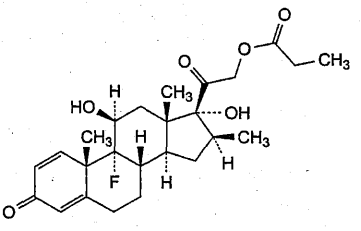
H. 9-fluoro-11β-hydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-17-yl propanoate,



I. 9-fluoro-11β-hydroxy-16β-methyl-21-[(methylsulfonyl)oxy]-3,20-dioxopregna-1,4-dien-17-yl propanoate,



J. (17*R*)-4′-chloro-5′-ethyl-9-fluoro-11β-hydroxy-16β-methylspiro[androsta-1,4-diene-17,2′(3′*H*)-furan]-3,3′-dione (17α-spiro compound),



K. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl propanoate (betamethasone 21-propionate),

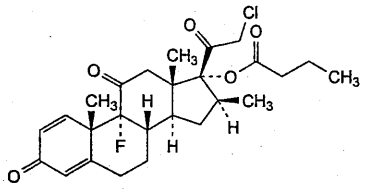
L. unknown structure,

M. unknown structure.

Ph Eur

Clobetasone Butyrate

(Ph. Eur. monograph 1090)



$C_{26}H_{32}ClFO_5$ 479.0 *25122-57-0*

Action and use
Glucocorticoid.

Preparations
Clobetasone Cream
Clobetasone Ointment

Ph Eur

DEFINITION

21-Chloro-9-fluoro-16β-methyl-3,11,20-trioxopregna-1,4-dien-17-yl butanoate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in methylene chloride, slightly soluble in ethanol (96 per cent).

mp
About 178 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *clobetasone butyrate CRS.*

TESTS

Specific optical rotation (*2.2.7*)
+ 131 to + 138 (dried substance).
Dissolve 0.250 g in *ethanol R1* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *anhydrous formic acid R, acetonitrile R, water R* (0.1:43:57 *V/V/V*).

Test solution Dissolve 65 mg of the substance to be examined in 5.0 mL of *acetonitrile R* and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 13 mg of *clobetasone butyrate for system suitability CRS* (containing impurity F) in 1 mL of *acetonitrile R* and dilute to 5.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: *anhydrous formic acid R, water R* (0.1:99.9 *V/V*);
— *mobile phase B*: *anhydrous formic acid R, acetonitrile R* (0.1:99.9 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	57	43
3 - 26	57 → 43	43 → 57

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 241 nm.
Injection 10 µL.
Identification of impurities Use the chromatogram supplied with *clobetasone butyrate for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity F.
Relative retention With reference to clobetasone butyrate (retention time = about 14 min): impurity F = about 0.9.
System suitability:
— *resolution*: minimum 3.5 between the peaks due to impurity F and clobetasone butyrate in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (b).
Limits:
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

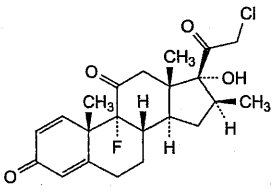
Dissolve 20.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 235 nm.
Calculate the content of $C_{26}H_{32}ClFO_5$, taking the specific absorbance to be 327.

STORAGE

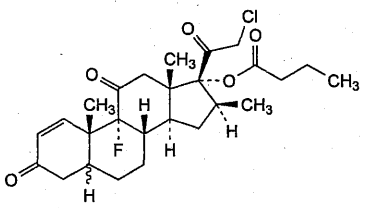
Protected from light.

IMPURITIES

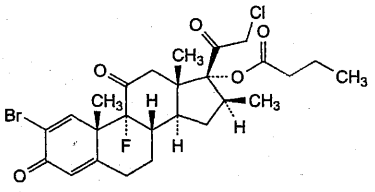
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, F, G, H, I.*



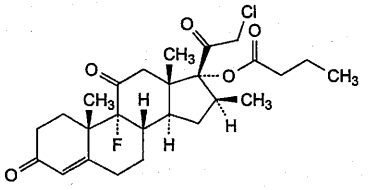
A. 21-chloro-9-fluoro-17-hydroxy-16β-methylpregna-1,4-diene-3,11,20-trione (clobetasone),



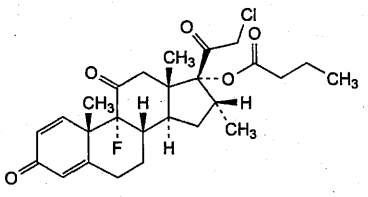
C. 21-chloro-9-fluoro-16β-methyl-3,11,20-trioxopregn-1-en-17-yl butanoate (4,5-dihydroclobetasone butyrate),



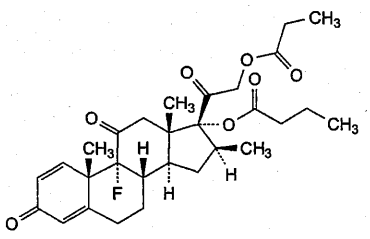
D. 2-bromo-21-chloro-9-fluoro-16β-methyl-3,11,20-trioxopregna-1,4-dien-17-yl butanoate (2-bromoclobetasone butyrate),



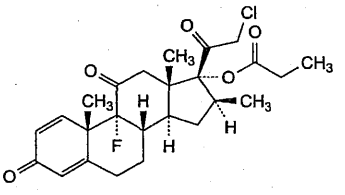
E. 21-chloro-9-fluoro-16β-methyl-3,11,20-trioxopregn-4-en-17-yl butanoate (1,2-dihydroclobetasone butyrate),



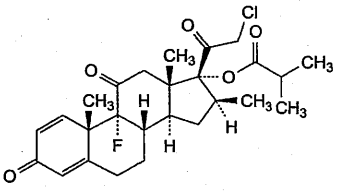
F. 21-chloro-9-fluoro-16α-methyl-3,11,20-trioxopregna-1,4-dien-17-yl butanoate (16α-methyl clobetasone butyrate),



G. 9-fluoro-16β-methyl-3,11,20-trioxo-21-(propanoyloxy)pregna-1,4-dien-17-yl butanoate,



H. 21-chloro-9-fluoro-16β-methyl-3,11,20-trioxopregna-1,4-dien-17-yl propanoate (17-*O*-propionyl clobetasone),

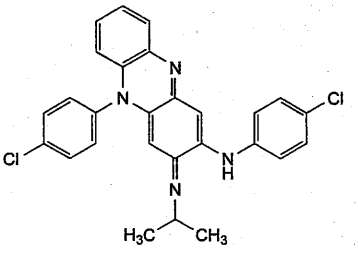


I. 21-chloro-9-fluoro-16β-methyl-3,11,20-trioxopregna-1,4-dien-17-yl 2-methylpropanoate (17-*O*-isobutyryl clobetasone).

Ph Eur

Clofazimine

(Ph. Eur. monograph 2054)



$C_{27}H_{22}Cl_2N_4$ 473.4 *2030-63-9*

Action and use
Antileprosy drug.

Preparation
Clofazimine Capsules

Ph Eur

DEFINITION

N,5-Bis(4-chlorophenyl)-3-[(1-methylethyl)imino]-3,5-dihydrophenazin-2-amine.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
Reddish-brown, fine powder.

Solubility
Practically insoluble in water, soluble in methylene chloride, very slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison clofazimine CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *clofazimine CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *propanol R, methylene chloride R* (6:85 *V/V*).

Application 5 µL.

First development Over 2/3 of the plate.

Drying Horizontally in air for 5 min.

Second development Over 2/3 of the plate.

Drying In air for 5 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 2 mg in 3 mL of *acetone R* and add 0.1 mL of *hydrochloric acid R*. An intense violet colour is produced. Add 0.5 mL of a 200 g/L solution of *sodium hydroxide R*; the colour changes to orange-red.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 100 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *clofazimine for system suitability CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm,
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 2.25 g of *sodium laurilsulfate R*, 0.85 g of *tetrabutylammonium hydrogen sulfate R* and 0.885 g of *disodium hydrogen phosphate dodecahydrate R* in *water R*. Adjust to pH 3.0 with *dilute phosphoric acid R* and dilute to 500 mL with *water R*. Mix 35 volumes of this solution and 65 volumes of *acetonitrile R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 3 times the retention time of clofazimine.

Identification of impurities Use the chromatogram supplied with *clofazimine for system suitability CRS* to identify the peak due to impurity B.

Relative retention With reference to clofazimine (retention time = about 15 min): impurity A = about 0.7; impurity B = about 0.8.

System suitability Reference solution (b):
- *resolution*: baseline separation between the peaks due to impurity B and clofazimine.

Limits:
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
- *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

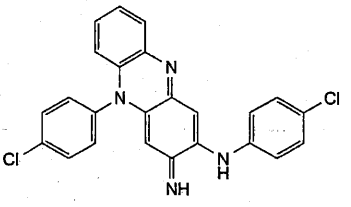
ASSAY

Dissolve 0.400 g in 5 mL of *methylene chloride R* and add 20 mL of *acetone R* and 5 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

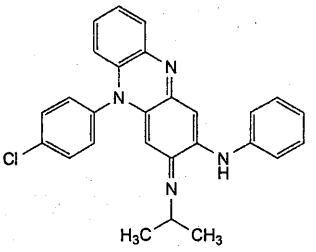
1 mL of *0.1 M perchloric acid* is equivalent to 47.34 mg of $C_{27}H_{22}Cl_2N_4$.

IMPURITIES

Specified impurities *A, B.*



A. *N*,5-bis(4-chlorophenyl)-3-imino-3,5-dihydrophenazin-2-amine,

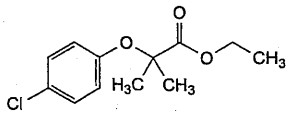


B. 5-(4-chlorophenyl)-3-[(1-methylethyl)imino]-*N*-phenyl-3,5-dihydrophenazin-2-amine.

Ph Eur

Clofibrate

(Ph. Eur. monograph 0318)



$C_{12}H_{15}ClO_3$ 242.7 *637-07-0*

Action and use
Fibrate; lipid-regulating drug.

Ph Eur

DEFINITION
Ethyl 2-(4-chlorophenoxy)-2-methylpropionate.

CHARACTERS
Appearance
Clear, almost colourless liquid.

Solubility
Very slightly soluble in water, miscible with ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *clofibrate CRS*.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).
Test solution (a) Dissolve 0.10 g in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with *methanol R*.
Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with *methanol R*.
Spectral range 250-350 nm for test solution (a); 220-250 nm for test solution (b).
Absorption maxima At 280 nm and 288 nm for test solution (a); at 226 nm for test solution (b).
Specific absorbances at the absorption maxima:
— at 226 nm: about 460 for test solution (b);
— at 280 nm: about 44 for test solution (a);
— at 288 nm: about 31 for test solution (a).

TESTS
Relative density (*2.2.5*)
1.138 to 1.147.

Refractive index (*2.2.6*)
1.500 to 1.505.

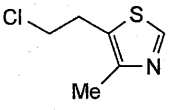
Acidity
To 1.0 g add 10 mL of *anhydrous ethanol R* and 0.1 mL of *phenol red solution R*. Not more than 1.0 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Volatile related substances
Gas chromatography (*2.2.28*).
Test solution To 10.0 g of the substance to be examined add a mixture of 10 mL of *dilute sodium hydroxide solution R* and 10 mL of *water R*. Shake, separate the lower (organic) layer, wash with 5 mL of *water R* and add the washings to the aqueous layer. Dry the organic layer with *anhydrous sodium sulfate R* and use as the test solution. Reserve the aqueous layer for the test for 4-chlorophenol.
Reference solution (a) Dissolve 0.12 g of the substance to be examined in *chloroform R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *chloroform R*.
Reference solution (b) Dissolve 0.12 g of *methyl 2-(4-chlorophenoxy)-2-methylpropionate CRS* in the substance to be examined and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with the substance to be examined. Dilute 1.0 mL of this solution to 10.0 mL with the substance to be examined.
Column:
— *size*: l = 1.5 m, Ø = 4 mm;
— *stationary phase*: *silanised diatomaceous earth for gas chromatography R* (250-420 µm) impregnated with 30 per cent *m/m* of *poly(dimethyl)siloxane R*; or *silanised diatomaceous earth for gas chromatography R* (150-180 µm) impregnated with 10 per cent *m/m* of *poly(dimethyl) siloxane R*;
— *temperature*: 185 °C.
Carrier gas *nitrogen for chromatography R*.
Detection Flame ionisation.
Injection 2 µL.
System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 4, where H_p = height above the baseline of the peak due to methyl 2-(4-chlorophenoxy)-2-methylpropionate and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clofibrate.
Limit:
— *total*: not more than 10 times the area of the peak due to clofibrate in the chromatogram obtained with reference solution (a) (0.1 per cent).

4-Chlorophenol
Gas chromatography (*2.2.28*) as described in the test for volatile related substances with the following modifications.
Test solution Shake the aqueous layer reserved in the test for volatile related substances with 2 quantities, each of 5 mL, of *chloroform R* and discard the organic layers. Acidify the aqueous layer by the dropwise addition of *hydrochloric acid R*. Shake with 3 quantities, each of 3 mL, of *chloroform R*. Combine the organic layers and dilute to 10.0 mL with *chloroform R*.
Reference solution Dissolve 0.25 g of *chlorophenol R* in *chloroform R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *chloroform R*.
Limit:
— *4-chlorophenol*: not more than the area of the peak due to 4-chlorophenol in the chromatogram obtained with the reference solution (25 ppm).

Ph Eur

Clomethiazole



C_6H_8ClNS 161.6 *533-45-9*

Action and use
Hypnotic.

Preparation
Clomethiazole Capsules

DEFINITION

Clomethiazole is 5-(2-chloroethyl)-4-methyl-thiazole.
It contains not less than 98.0% and not more than 101.0% of C_6H_8ClNS.

CHARACTERISTICS

A colourless to slightly yellowish brown liquid.

Slightly soluble in *water*, miscible with *ethanol (96%)* and with *ether*.

IDENTIFICATION

The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of clomethiazole *(RS 051)*.

TESTS

Acidity or alkalinity

pH of a 0.5% w/v solution, 5.5 to 7.0, Appendix V L.

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in the mobile phase.

(1) 0.20% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 100 volumes and dilute 1 volume of the resulting solution to 10 volumes.

(3) Dilute 1 volume of a 0.030% w/v solution of *clomethiazole impurity A BPCRS* (equivalent to 0.020% w/v of impurity A as base) in *methanol* (solution A) to 50 volumes.

(4) Dilute 1 volume of a 0.020% w/v solution of *clomethiazole impurity B BPCRS* (quaternary dimer) in *methanol* (solution B) to 50 volumes.

(5) Dilute 1 volume of a 0.020% w/v solution of *clomethiazole impurity C BPCRS* in *methanol* (solution C) to 50 volumes.

(6) Add 1 mL each of solutions A, B and C to 0.10 g of the substance being examined and dilute to 50 mL.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4 mm) packed with *octadecylsilyl silica gel for chromatography* (10 µm) (Lichrosorb RP18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 257 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

30 volumes of *methanol* and 70 volumes of a solution containing 0.13% w/v of *sodium hexanesulfonate* and 2.7% w/v of *tetramethylammonium hydrogen sulfate*, adjusted to pH 2.0 with 5m *sodium hydroxide*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (6), baseline separation is achieved between the peaks due to impurities A, B and C and also between the principal peak and the two adjacent specified impurity peaks.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to clomethiazole impurity A as base is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.2%);

the area of any peak corresponding to clomethiazole impurity B is not greater than the area of the principal peak in the chromatogram obtained with solution (4) (0.2%);

the area of any peak corresponding to clomethiazole impurity C is not greater than the area of the principal peak in the chromatogram obtained from solution (5) (0.2%);

the sum of the areas of impurities A, B and C is not greater than 0.5%;

the area of any other *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.1%);

the sum of the areas of any *secondary peaks*, other than impurities A, B and C, is not greater than 5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

Disregard any peak with an area less than half the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Sulfated ash

Not more than 0.1%, Appendix IX A. Use 1 g.

ASSAY

Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.3 g and determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 16.16 mg of C_6H_8ClNS.

STORAGE

Clomethiazole should be stored at a temperature of 2° to 8°.

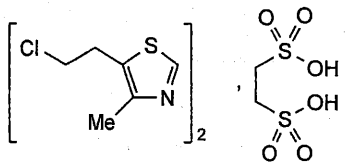
IMPURITIES

A. 4-methyl-5-vinylthiazole edisilate,

B. 5-(2-chloroethyl)-4-methyl-3-[2-(4-methylthiazol-5-yl) ethyl]-thiazolium chloride (quaternary dimer),

C. 4-methyl-5-(2-hydroxyethyl)thiazole.

Clomethiazole Edisilate



$(C_6H_8ClNS)_2,C_2H_6O_6S_2$ 513.5 *1867-58-9*

Action and use

Hypnotic.

Preparation

Clomethiazole Oral Solution

DEFINITION

Clomethiazole Edisilate is 5-(2-chloroethyl)-4-methylthiazole ethanedisulfonate. It contains not less than 99.0% and not more than 101.0% of $(C_6H_8ClNS)_2,C_2H_6O_6S_2$, calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder.

Freely soluble in *water*; soluble in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION

The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of clomethiazole edisilate *(RS 052)*.

TESTS

Calcium

10 mL of a 10.0% w/v solution diluted to 15 mL with *water* complies with the *limit test for calcium*, Appendix VII (100 ppm).

Chloride

10 mL of a 10% w/v solution diluted to 15 mL with *water* complies with the *limit test for chlorides*, Appendix VII (50 ppm).

Sulfate

10 mL of a 1.0% w/v solution diluted to 15 mL with *water* complies with the *limit test for sulfates*, Appendix VII (0.15%).

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in the mobile phase.

(1) The substance being examined containing 0.3% w/v of Clomethiazole Edisilate (equivalent to 0.2% w/v of clomethiazole base).

(2) Dilute 1 volume of solution (1) to 100 volumes and dilute 1 volume of the resulting solution to 10 volumes.

(3) Dilute 1 volume of a 0.030% w/v solution of *clomethiazole impurity A BPCRS* (equivalent to 0.02% w/v of impurity A as base) in methanol (solution A) to 50 volumes.

(4) Dilute 1 volume of a 0.020% w/v solution of *clomethiazole impurity B BPCRS* (quaternary dimer) in *methanol* (solution B) to 50 volumes.

(5) Dilute 1 volume of a 0.020% w/v solution of *clomethiazole impurity C BPCRS* in *methanol* (solution C) to 50 volumes.

(6) Add 1 mL each of solutions A, B and C to 0.10 g of the substance being examined and dilute to 50 mL.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4 mm) packed with *octadecylsilyl silica gel for chromatography* (10 µm) (Lichrosorb RP18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 257 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

30 volumes of *methanol* and 70 volumes of a solution containing 0.13% w/v of *sodium hexanesulfonate* and 2.7% w/v of *tetramethylammonium hydrogen sulfate*, adjusted to pH 2.0 with 5M *sodium hydroxide*.

SYSTEM SUITABILITY

The test is not valid unless in the chromatogram obtained with solution (6) closely resembles the chromatogram supplied with impurities A, B and C.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to impurity A is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.2%);

the area of any peak corresponding to impurity B is not greater than the area of the principal peak in the chromatogram obtained with solution (4) (0.2%);

the area of any peak corresponding to impurity C is not greater than the area of the principal peak in the chromatogram obtained from solution (5) (0.2%);

the area of any other *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained from solution (2) (0.1%);

the sum of the areas of impurities A, B and C is not greater than 0.5%;

the sum of the areas of any *secondary peaks*, other than impurities A, B and C, is not greater than 5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

Disregard any peak with an area less than half the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Loss on drying

When dried at 50° at a pressure not exceeding 0.7 kPa for 6 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.3%, Appendix IX A. Use 1 g.

ASSAY

Dissolve 0.4 g in 50 mL of *water* and titrate with *0.1M sodium hydroxide VS* using *phenolphthalein solution R1* as indicator. Each mL of *0.1M sodium hydroxide VS* is equivalent to 25.67 mg of $(C_6H_8ClNS)_2,C_2H_6O_6S_2$.

IMPURITIES

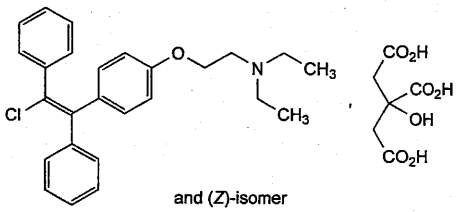
A. 4-methyl-5-vinylthiazole edisilate,

B. 5-(2-chloroethyl)-4-methyl-3-[2-(4-methylthiazol-5-yl) ethyl]-thiazolium chloride (quaternary dimer),

C. 4-methyl-5-(2-hydroxyethyl)thiazole.

Clomifene Citrate

(Ph. Eur. monograph 0997)



and (Z)-isomer

$C_{32}H_{36}ClNO_8$ 598.1 *50-41-9*

Action and use

Estrogen receptor modulator.

Preparation

Clomifene Tablets

Ph Eur

DEFINITION

Mixture of the (*E*)- and (*Z*)-isomers of 2-[4-(2-chloro-1,2diphenylethenyl)phenoxy]-*N*,*N*-diethylethanamine dihydrogen citrate.

Content

98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or pale yellow, crystalline powder.

Solubility
Slightly soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clomifene citrate CRS.*

B. Dissolve about 5 mg in 5 mL of a mixture of 1 volume of *acetic anhydride R* and 5 volumes of *pyridine R*, then heat in a water-bath. A deep red colour is produced.

TESTS

Prepare the solutions protected from light in brown-glass vessels. Ensure minimum exposure of the solutions to daylight until they are required for chromatography.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 12.5 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 12.5 mg of *clomifene citrate for performance test CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 400 mL of *acetonitrile for chromatography R* with 600 mL of *water for chromatography R* and add 8.0 mL of *diethylamine R*; adjust to pH 6.2 with about 1-2 mL of *phosphoric acid R*, taking care to reduce progressively the volume of each addition as the required pH is approached.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 233 nm.

Equilibration With the mobile phase for about 1 h.

Injection 10 µL.

Run time 4 times the retention time of clomifene.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 15, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clomifene; if necessary, adjust the concentration of acetonitrile in the mobile phase;
— the chromatogram obtained is similar to the chromatogram supplied with *clomifene citrate for performance test CRS.*

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
— *impurities B, C, D, E, F, G, H*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *total*: not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.5 per cent);
— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak with a retention time relative to the clomifene peak of 0.2 or less.

(*Z*)-isomer
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in 25 mL of *0.1 M hydrochloric acid*, add 5 mL of *1 M sodium hydroxide* and shake with 3 quantities, each of 25 mL, of *ethanol-free chloroform R*. Wash the combined extracts with 10 mL of *water R*, dry over *anhydrous sodium sulfate R* and dilute to 100 mL with *ethanol-free chloroform R*. To 20 mL of this solution add 0.1 mL of *triethylamine R* and dilute to 100 mL with *hexane R*.

Reference solution Dissolve 25 mg of *clomifene citrate CRS* in 25 mL of *0.1 M hydrochloric acid*, add 5 mL of *1 M sodium hydroxide* and shake with 3 quantities, each of 25 mL, of *ethanol-free chloroform R*. Wash the combined extracts with 10 mL of *water R*, dry over *anhydrous sodium sulfate R* and dilute to 100 mL with *ethanol-free chloroform R*. To 20 mL of this solution add 0.1 mL of *triethylamine R* and dilute to 100 mL with *hexane R*.

Column:
— *size*: *l* = 0.3 m, Ø = 4 mm;
— *stationary phase*: *silica gel for chromatography R* (10 µm).

Mobile phase *triethylamine R, ethanol-free chloroform R, hexane R* (1:200:800 *V/V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 302 nm.

Equilibration With the mobile phase for about 2 h.

Injection 50 µL.

Identification of peaks The chromatogram obtained with the reference solution shows a peak due to the (*E*)-isomer just before a peak due to the (*Z*)-isomer.

System suitability Reference solution:
— *resolution*: minimum 1.0 between the peaks due to the (*E*)- and (*Z*)-isomers; if necessary, adjust the relative proportions of ethanol-free chloroform and hexane in the mobile phase.

Measure the area of the peak due to the (*Z*)-isomer in the chromatograms obtained with the test solution and the reference solution. Calculate the content of the (*Z*)-isomer, as a percentage of the total clomifene citrate present, from the declared content of *clomifene citrate CRS.*

Limit:
— *(Z)-isomer*: 30.0 per cent to 50.0 per cent.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.000 g.

ASSAY

Dissolve 0.500 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

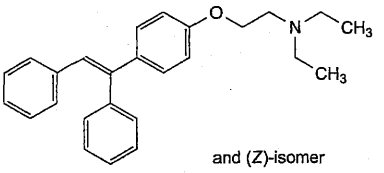
1 mL of *0.1 M perchloric acid* is equivalent to 59.81 mg of $C_{32}H_{36}ClNO_8$.

STORAGE

Protected from light.

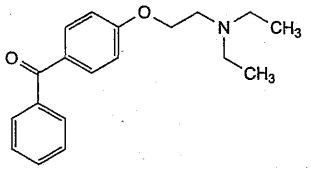
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H.*

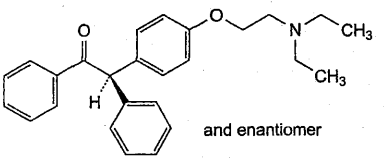


and (Z)-isomer

A. 2-[4-(1,2-diphenylethenyl)phenoxy]-*N*,*N*-diethylethanamine,

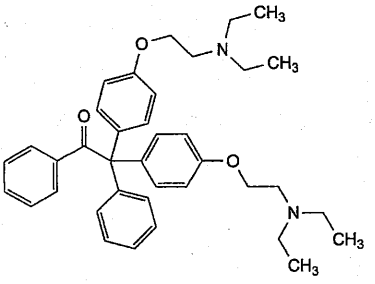


B. [4-[2-(diethylamino)ethoxy]phenyl]phenylmethanone,

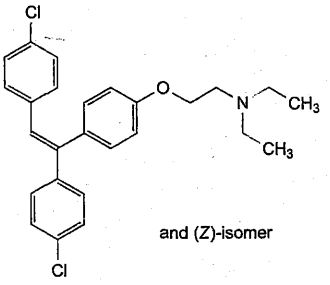


and enantiomer

C. (2*RS*)-2-[4-[2-(diethylamino)ethoxy]phenyl]-1,2-diphenylethanone,

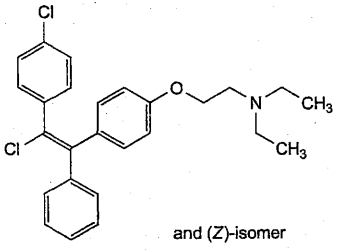


D. 2,2-bis[4-[2-(diethylamino)ethoxy]phenyl]-1,2-diphenylethanone,



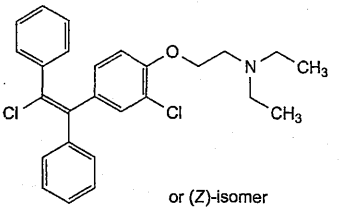
and (Z)-isomer

E. 2-[4-[1,2-bis(4-chlorophenyl)ethenyl]phenoxy]-*N*,*N*-diethylethanamine,



and (Z)-isomer

F. 2-[4-[2-chloro-2-(4-chlorophenyl)-1-phenylethenyl]phenoxy]-*N*,*N*-diethylethanamine,



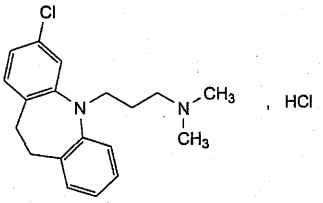
or (Z)-isomer

GH. 2-[2-chloro-4-(2-chloro-1,2-diphenylethenyl)phenoxy]-*N*,*N*-diethylethanamine (G. higher-melting-point isomer; H. lower-melting-point isomer).

Ph Eur

Clomipramine Hydrochloride

(Ph. Eur. monograph 0889)



$C_{19}H_{24}Cl_2N_2$ 351.3 *17321-77-6*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparations
Clomipramine Capsules
Clomipramine Prolonged-release Tablets

Ph Eur

DEFINITION
3-(3-Chloro-10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder, slightly hygroscopic.

Solubility
Freely soluble in water and in methylene chloride, soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clomipramine hydrochloride CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *2-propanol R*, evaporate to dryness and record new spectra using the residues.

B. Dissolve about 50 mg in 5 mL of *water R* and add 1 mL of *dilute ammonia R1*. Mix, allow to stand for 5 min and filter. Acidify the filtrate with *dilute nitric acid R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method I*).

pH (*2.2.3*)

3.5 to 5.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protected from light.*

Test solution Dissolve 20.0 mg of the substance to be examined in a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Reference solution (a) Dissolve 22.6 mg of *imipramine hydrochloride CRS* (impurity B), 4.0 mg of *clomipramine impurity C CRS*, 4.0 mg of *clomipramine impurity D CRS* and 2.0 mg of *clomipramine impurity F CRS* in a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A and dilute to 100.0 mL with the same mixture of mobile phases. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A.

Reference solution (c) Dissolve 10.0 mg of *clomipramine hydrochloride CRS* and 3.0 mg of *clomipramine impurity C CRS* in a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A and dilute to 20.0 mL with the same mixture of mobile phases. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 25 volumes of mobile phase B and 75 volumes of mobile phase A.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped cyanopropylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:

— *mobile phase A*: dissolve 1.2 g of *sodium dihydrogen phosphate R* in *water R*, add 1.1 mL of *nonylamine R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	75	25
10 - 20	75 → 65	25 → 35
20 - 32	65	35
32 - 34	65 → 75	35 → 25
34 - 44	75	25

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Relative retention With reference to clomipramine (retention time = about 8 min): impurity B = about 0.7; impurity C = about 0.9; impurity D = about 1.7; impurity F = about 3.4.

System suitability Reference solution (c):

— *resolution*: minimum 3.0 between the peaks due to clomipramine and impurity C.

Limits:

— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *impurities C, D*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity F*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *sum of unspecified impurities*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 35.13 mg of $C_{19}H_{24}Cl_2N_2$.

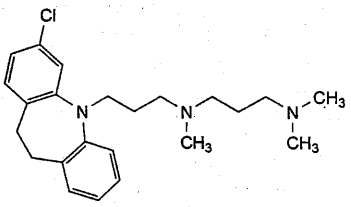
STORAGE

In an airtight container, protected from light.

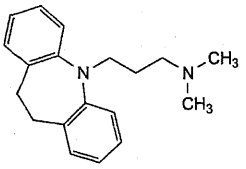
IMPURITIES

Specified impurities *B, C, D, F.*

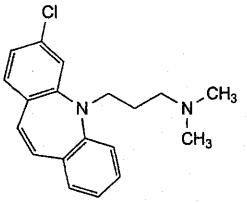
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, E, G.*



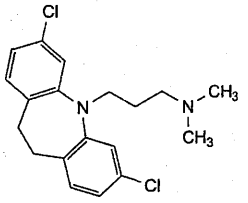
A. *N*-[3-(3-chloro-10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl) propyl]-*N*,*N*′,*N*′-trimethylpropane-1,3-diamine,



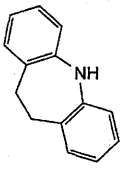
B. 3-(10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine (imipramine),



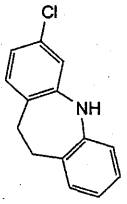
C. 3-(3-chloro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine,



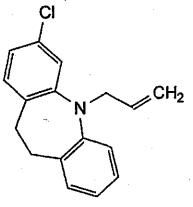
D. 3-(3,7-dichloro-10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine,



E. 10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepine (iminodibenzyl),



F. 3-chloro-10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepine,

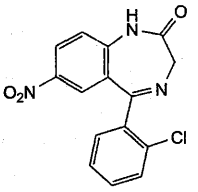


G. 3-chloro-5-(prop-2-en-1-yl)-10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepine.

Ph Eur

Clonazepam

(Ph. Eur. monograph 0890)



$C_{15}H_{10}ClN_3O_3$ 315.7 *1622-61-3*

Action and use
Benzodiazepine.

Preparations
Clonazepam Injection
Clonazepam Oral Suspension
Clonazepam Tablets

Ph Eur

DEFINITION

5-(2-Chlorophenyl)-7-nitro-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
Slightly yellowish, crystalline powder.

Solubility
Practically insoluble in water, slightly soluble in alcohol and in methanol.

mp
About 239 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of clonazepam.*

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Solvent mixture *tetrahydrofuran R, methanol R, water R* (10:42:48 *V/V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 20.0 mL with the same solvent. Dilute 1.0 mL to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *flunitrazepam R* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 1.0 mg of *clonazepam impurity B CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 10 volumes of *tetrahydrofuran R*, 42 volumes of *methanol R* and 48 volumes of a 6.6 g/L solution of *ammonium phosphate R* previously adjusted to pH 8.0 with a 40 g/L solution of *sodium hydroxide R* or *dilute phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 3 times the retention time of clonazepam.

Relative retention With reference to clonazepam (retention time = about 7 min): impurity B = about 2.1; impurity A = about 2.4.

System suitability Reference solution (b):
— *resolution*: minimum 1.8 between the peaks due to flunitrazepam and to clonazepam.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent)
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

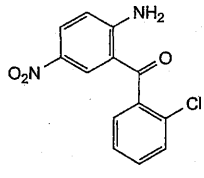
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.275 g in 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

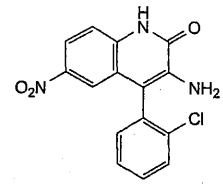
1 mL of *0.1 M perchloric acid* is equivalent to 31.57 mg of $C_{15}H_{10}ClN_3O_3$.

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B.*



A. (2-amino-5-nitrophenyl)(2-chlorophenyl)methanone,

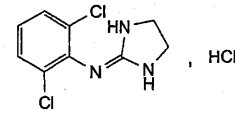


B. 3-amino-4-(2-chlorophenyl)-6-nitroquinolin-2(1*H*)-one.

Ph Eur

Clonidine Hydrochloride

(Ph. Eur. monograph 0477)



$C_9H_{10}Cl_3N_3$ 266.6 *4205-91-8*

Action and use
Alpha$_2$-adrenoceptor agonist; treatment of hypertension.

Preparations
Clonidine Injection
Clonidine Tablets

Ph Eur

DEFINITION
2,6-Dichloro-*N*-(imidazolidin-2-ylidene)aniline hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water and in anhydrous ethanol.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 30.0 mg in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid.

Spectral range 245-350 nm.

Absorption maxima At 272 nm and 279 nm.

Point of inflexion At 265 nm.

Specific absorbance at the absorption maxima:
— at 272 nm: about 18;
— at 279 nm: about 16.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clonidine hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 5 mg of *clonidine hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *glacial acetic acid R, butanol R, water R* (10:40:50 *V/V/V*); allow to separate, filter the upper layer and use the filtrate.

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *potassium iodobismuthate solution R2*. Allow to dry in air for 1 h. Spray again with *potassium iodobismuthate solution R2* and then immediately spray with a 50 g/L solution of *sodium nitrite R*.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)

4.0 to 5.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 50 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *clonidine impurity B CRS* in 2 mL of *acetonitrile R* and dilute to 5 mL with mobile phase A. To 1 mL of this solution, add 1 mL of the test solution and dilute to 10 mL with mobile phase A.

Column:
- *size*: *l* = 0.15 m, Ø = 3.0 mm;
- *stationary phase*: *propylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: dissolve 4 g of *potassium dihydrogen phosphate R* in 1000 mL of *water R*, and adjust to pH 4.0 with *phosphoric acid R*;
- *mobile phase B*: mobile phase A, *acetonitrile R1* (25:75 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0	90	10
0 - 15	90 → 30	10 → 70
15 - 15.1	30 → 90	70 → 10
15.1 - 20	90	10

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

System suitability Reference solution (b):
- *resolution*: minimum 5 between the peaks due to clonidine and impurity B.

Limits:
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

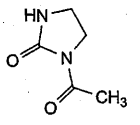
ASSAY

Dissolve 0.200 g in 70 mL of *ethanol (96 per cent) R*. Titrate with *0.1 M ethanolic sodium hydroxide* determining the end-point potentiometrically (*2.2.20*).

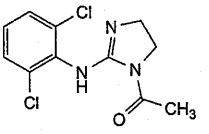
1 mL of *0.1 M sodium hydroxide* is equivalent to 26.66 mg of $C_9H_{10}Cl_3N_3$.

IMPURITIES

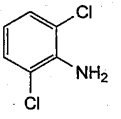
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



A. 1-acetylimidazolidin-2-one,



B. 1-acetyl-2-[(2,6-dichlorophenyl)amino]-4,5-dihydro-1*H*-imidazole,

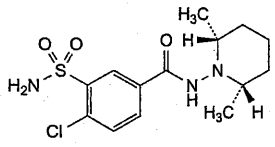


C. 2,6-dichloroaniline.

Ph Eur

Clopamide

(Ph. Eur. monograph 1747)



$C_{14}H_{20}ClN_3O_3S$ 345.8 *636-54-4*

Action and use
Thiazide-like diuretic.

Ph Eur

DEFINITION

4-Chloro-*N*-[(2*RS*,6*SR*)-2,6-dimethylpiperidin-1-yl]-3-sulfamoylbenzamide.

Content
99.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

The production method is evaluated to determine the potential for formation of an *N*-nitroso compound (*cis*-2,6-dimethyl-1-nitrosopiperidine). Where necessary, the production method is validated to demonstrate that the *N*-nitroso compound is absent in the final product.

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Slightly soluble in water and in anhydrous ethanol, sparingly soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clopamide CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *methanol R*, evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 100 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *clopamide for system suitability CRS* (containing impurities B, C and H) in 1.0 mL of *methanol R*.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 2.0 mL of this solution to 40.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 1.0 g of *ammonium acetate R* in 950 mL of *water R*, adjust to pH 2.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R*;
— *mobile phase C*: *water R*, *tetrahydrofuran for chromatography R* (20:80 *V/V*); this mobile phase allows adequate rinsing of the system;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 35	95 → 75	5 → 25	0
35 - 45	75 → 35	25 → 65	0
45 - 50	35 → 30	65 → 0	0 → 70
50 - 60	30	0	70

Flow rate 0.4 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *clopamide for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C and H.

Relative retention With reference to clopamide (retention time = about 33 min): impurity C = about 0.8; impurity H = about 1.2; impurity B = about 1.4.

System suitability Reference solution (a):
— *resolution*: minimum 3 between the peaks due to impurity C and clopamide.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.5; impurity H = 0.4;
— *impurities B, C, H*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 2.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.280 g in 70 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 34.58 mg of $C_{14}H_{20}ClN_3O_3S$.

STORAGE

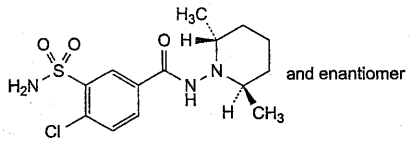
In an airtight container, protected from light.

IMPURITIES

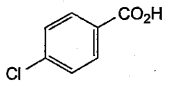
Specified impurities *B, C, H.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

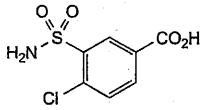
demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, G.*



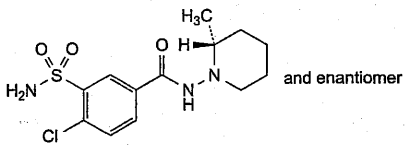
A. 4-chloro-*N*-[(2*RS*,6*RS*)-2,6-dimethylpiperidin-1-yl]-3-sulfamoylbenzamide (*trans*-clopamide),



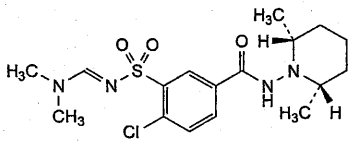
B. 4-chlorobenzoic acid,



C. 4-chloro-3-sulfamoylbenzoic acid,



G. 4-chloro-*N*-[(2*RS*)-2-methylpiperidin-1-yl]-3-sulfamoylbenzamide,

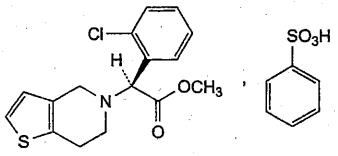


H. 4-chloro-3-[(*E*)-[(dimethylamino)methylene]sulfamoyl]-*N*-[(2*RS*,6*SR*)-2,6-dimethylpiperidin-1-yl]benzamide.

Ph Eur

Clopidogrel Besilate

(Ph. Eur. monograph 2790)



$C_{22}H_{22}ClNO_5S_2$ 480.0 *744256-69-7*

Action and use
Inhibitor of ADP-mediated platelet aggregation.

Ph Eur

DEFINITION
Methyl (2*S*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate benzenesulfonate.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION
It is considered that alkyl benzenesulfonate esters are genotoxic and are potential impurities in clopidogrel besilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general method *2.5.41. Methyl, ethyl and isopropyl benzenesulfonate in active substances* is not suitable for clopidogrel besilate since it was observed that methyl benzenesulfonate was obtained during the gas chromatography analysis as an artefact originating from degradation. Another suitable and validated method should be used. The content of each alkyl benzenesulfonate is not more than 3 ppm.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol, pratically insoluble in heptane.

IDENTIFICATION
Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (*2.2.7*): + 47.0 to + 51.0 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clopidogrel besilate CRS.*

C. Enantiomeric purity (see Tests).

D. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (d).

Results:
- the peak due to besilate in the chromatogram obtained with the test solution is similar in retention time to the corresponding peak in the chromatogram obtained with reference solution (d);
- the ratio of the area of the peak due to the besilate to the area of the peak due to clopidogrel in the chromatogram obtained with the test solution is minimum 0.1.

TESTS
Enantiomeric purity
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 46.0 mg of the substance to be examined in 10.0 mL of *anhydrous ethanol R* and dilute to 20.0 mL with *heptane R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with *heptane R*.

Reference solution (a) Dissolve 10 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in 2.5 mL of *anhydrous ethanol R* and dilute to 5.0 mL with *heptane R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *heptane R*. Dilute 1.5 mL of this solution to 10.0 mL with *heptane R*.

Reference solution (c) Dissolve 34.0 mg of *clopidogrel hydrochloride CRS* in 10.0 mL of *anhydrous ethanol R* and dilute to 20.0 mL with *heptane R*. Dilute 1.0 mL of the solution to 10.0 mL with *heptane R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *cellulose derivative of silica gel for chiral separation R* (10 µm).

Mobile phase *anhydrous ethanol R, heptane R* (15:85 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time 1.25 times the retention time of clopidogrel.

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and C.

Relative retention With reference to clopidogrel (retention time = about 18 min): impurity C = about 0.6; impurity B = about 0.7.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurities C and B.

Calculation of percentage content:
— for impurity C, use the concentration of clopidogrel in reference solution (b).

Limit:
— *impurity C*: maximum 0.15 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, *acetonitrile R1* (40:60 *V/V*).

Test solution Dissolve 74 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *clopidogrel impurity A CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dissolve 32 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in the solvent mixture, add 0.5 mL of reference solution (a) and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 25 mg of *sodium benzenesulfonate R* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 5 volumes of *methanol R2* and 95 volumes of a 0.96 g/L solution of *sodium pentanesulfonate monohydrate R* adjusted to pH 2.5 with *phosphoric acid R*;
— *mobile phase B*: *methanol R2, acetonitrile R1* (5:95 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	89.5	10.5
3 - 48	89.5 → 31.5	10.5 → 68.5
48 - 68	31.5	68.5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to clopidogrel (retention time = about 27 min): besilate = about 0.05; impurity A = about 0.4; impurity B = about 1.1.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clopidogrel.

Calculation of percentage contents:
— for each impurity, use the concentration of clopidogrel in reference solution (c).

Limits:
— *impurities A, B*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent; disregard the peak due to the besilate.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for enantiomeric purity with the following modification.

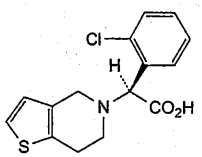
Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{22}H_{22}ClNO_5S_2$ taking into account the assigned content of *clopidogrel hydrochloride CRS*, and a conversion factor of 1.34.

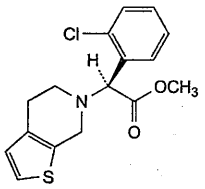
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C.*

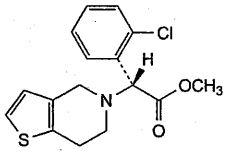
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E, F, G.*



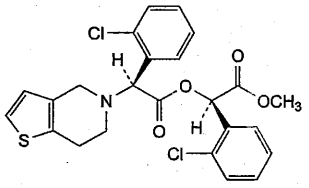
A. (2*S*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetic acid,



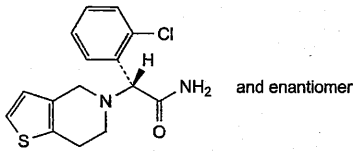
B. methyl (2*S*)-2-(2-chlorophenyl)-2-[4,7-dihydrothieno[2,3-*c*]pyridin-6(5*H*)-yl]acetate,



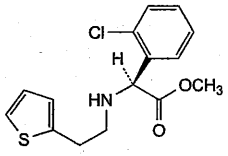
C. methyl (2*R*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate,



D. (1*R*)-1-(2-chlorophenyl)-2-methoxy-2-oxoethyl (2*S*)-2-(2-chlorophenyl)-2-(6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl)acetate,



E. (2*RS*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetamide,



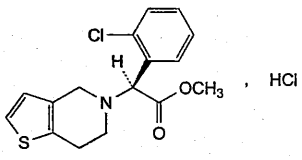
F. methyl (2*S*)-2-(2-chlorophenyl)-2-[[2-(thiophen-2-yl)ethyl]amino]acetate,

G. unknown structure.

Ph Eur

Clopidogrel Hydrochloride

(Ph. Eur. monograph 2791)



$C_{16}H_{17}Cl_2NO_2S$ 358.3 *120202-65-5*

Action and use
Inhibitor of ADP-mediated platelet aggregation.

Ph Eur

DEFINITION
Methyl (2*S*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate hydrochloride.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or yellowish powder.

Solubility
Practically insoluble in water, very soluble in anhydrous ethanol, pratically insoluble in heptane.

IDENTIFICATION
Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (*2.2.7*): + 65.0 to + 69.0 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clopidogrel hydrochloride CRS.*

C. Enantiomeric purity (see Tests).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Enantiomeric purity
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 34.0 mg of the substance to be examined in 10.0 mL of *anhydrous ethanol R* and dilute to 20.0 mL with *heptane R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with *heptane R*.

Reference solution (a) Dissolve 10 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in 2.5 mL of *anhydrous ethanol R* and dilute to 5.0 mL with *heptane R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *heptane R*. Dilute 1.5 mL of this solution to 10.0 mL with *heptane R*.

Reference solution (c) Dissolve 34.0 mg of *clopidogrel hydrochloride CRS* in 10.0 mL of *anhydrous ethanol R* and dilute to 20.0 mL with *heptane R*. Dilute 1.0 mL of the solution to 10.0 mL with *heptane R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *cellulose derivative of silica gel for chiral separation R* (10 µm).

Mobile phase *anhydrous ethanol R, heptane R* (15:85 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time 1.25 times the retention time of clopidogrel.

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and C.

Relative retention With reference to clopidogrel (retention time = about 18 min): impurity C = about 0.6; impurity B = about 0.7.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurities C and B.

Calculation of percentage content:
— for impurity C, use the concentration of clopidogrel in reference solution (b).

Limit:
— *impurity C*: maximum 0.15 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, *acetonitrile R1* (40:60 *V/V*).

Test solution Dissolve 55 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *clopidogrel impurity A CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dissolve 32 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in the solvent mixture, add 0.5 mL of reference solution (a) and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 5 volumes of *methanol R2* and 95 volumes of a 0.96 g/L solution of *sodium pentanesulfonate monohydrate R* adjusted to pH 2.5 with *phosphoric acid R*;
— *mobile phase B*: *methanol R2*, *acetonitrile R1* (5:95 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	89.5	10.5
3 - 48	89.5 → 31.5	10.5 → 68.5
48 - 68	31.5	68.5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to clopidogrel (retention time = about 27 min): impurity A = about 0.4; impurity B = about 1.1.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clopidogrel.

Calculation of percentage contents:
— for each impurity, use the concentration of clopidogrel in reference solution (c).

Limits:
— *impurities A, B*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.
Replace the solvent after each titration.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for enantiomeric purity with the following modification.

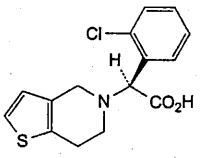
Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{16}H_{17}Cl_2NO_2S$ taking into account the assigned content of *clopidogrel hydrochloride CRS*.

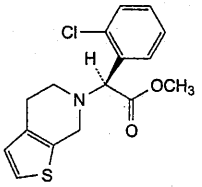
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C.*

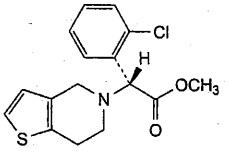
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E, F, G.*



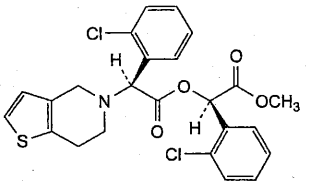
A. (2*S*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetic acid,



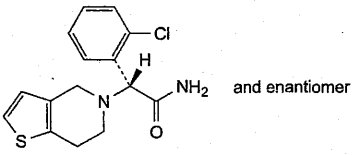
B. methyl (2*S*)-2-(2-chlorophenyl)-2-[4,7-dihydrothieno[2,3-*c*]pyridin-6(5*H*)-yl]acetate,



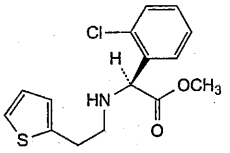
C. methyl (2*R*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate,



D. (1*R*)-1-(2-chlorophenyl)-2-methoxy-2-oxoethyl (2*S*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate,



E. (2*RS*)-2-(2-chlorophenyl)-2-[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetamide,



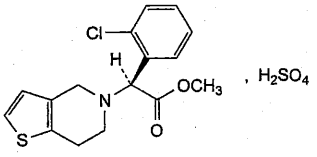
F. methyl (2*S*)-2-(2-chlorophenyl)-2-[[2-(thiophen-2-yl)ethyl]amino]acetate,

G. unknown structure.

Ph Eur

Clopidogrel Hydrogen Sulfate

(Ph. Eur. monograph 2531)



$C_{16}H_{18}ClNO_6S_2$ 419.9 *120202-66-6*

Action and use
Inhibitor of ADP-mediated platelet aggregation.

Ph Eur

DEFINITION
Methyl (2*S*)-(2-chlorophenyl)[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate sulfate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Freely soluble in water and in methanol, practically insoluble in cyclohexane.

It shows polymorphism (*5.9*).

IDENTIFICATION
Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (*2.2.7*): + 54.0 to + 58.0 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clopidogrel hydrogen sulfate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues (the substance may stick to the surface of the recipient used).

C. Enantiomeric purity (see Tests).

D. It gives reaction (a) of sulfates (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method I*).

Dissolve 1.0 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Enantiomeric purity
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 0.1 g of the substance to be examined in 25.0 mL of *anhydrous ethanol R* and dilute to 50.0 mL with *heptane R*.

Reference solution Dissolve 10 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in 2.5 mL of *anhydrous ethanol R* and dilute to 5.0 mL with *heptane R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel OJ for chiral separations R* (10 µm).

Mobile phase *anhydrous ethanol R, heptane R* (15:85 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 1.25 times the retention time of clopidogrel.

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with the reference solution to identify the peaks due to impurities B and C.

Relative retention With reference to clopidogrel (retention time = about 18 min): impurity C = about 0.6; impurity B = about 0.7.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to impurities C and B;
— *signal-to-noise ratio*: minimum 20 for the peak due to impurity C.

Limit:
— *impurity C*: maximum 0.5 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, *acetonitrile R1* (40:60 *V/V*).

Test solution Dissolve 65 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *clopidogrel impurity A CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dissolve 32 mg of *clopidogrel for system suitability CRS* (containing impurities B and C) in the solvent mixture, add 0.5 mL of reference solution (a) and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 3.9 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: mix 5 volumes of *methanol R2* and 95 volumes of a 0.96 g/L solution of *sodium pentanesulfonate monohydrate R* adjusted to pH 2.5 with *phosphoric acid R*;
- *mobile phase B*: *methanol R2, acetonitrile R1* (5:95 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	89.5	10.5
3 - 48	89.5 → 31.5	10.5 → 68.5
48 - 68	31.5	68.5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *clopidogrel for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to clopidogrel (retention time = about 25 min): impurity A = about 0.4; impurity B = about 1.1.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clopidogrel.

Limits:
- *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
- *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Replace the solvent after each titration.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.160 g in a mixture of 10 mL of *acetone R*, 10 mL of *methanol R* and 30 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). A precipitate may be formed during the titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 20.99 mg of $C_{16}H_{18}ClNO_6S_2$.

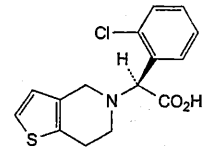
STORAGE

Protected from light.

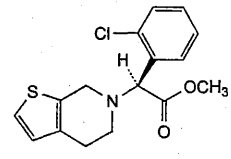
IMPURITIES

Specified impurities *A, B, C.*

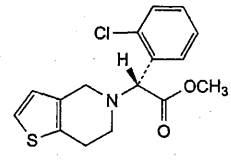
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*



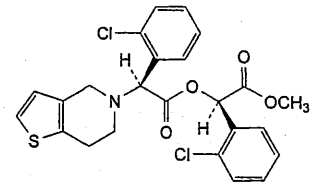
A. (2*S*)-(2-chlorophenyl)[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetic acid,



B. methyl (2*S*)-(2-chlorophenyl)[4,7-dihydrothieno[2,3-*c*]pyridin-6(5*H*)-yl]acetate,



C. methyl (2*R*)-(2-chlorophenyl)[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetate,

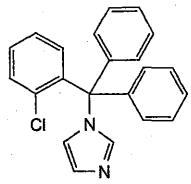


D. methyl (2*R*)-(2-chlorophenyl)[(2*S*)-(2-chlorophenyl)[6,7-dihydrothieno[3,2-*c*]pyridin-5(4*H*)-yl]acetyloxy]acetate.

Ph Eur

Clotrimazole

(Ph. Eur. monograph 0757)



$C_{22}H_{17}ClN_2$ 344.8 *23593-75-1*

Action and use
Antifungal.

Preparations
Clotrimazole and Hydrocortisone Acetate Cream
Clotrimazole Cream
Clotrimazole Eye Drops
Clotrimazole Pessaries
Clotrimazole Vaginal Tablets

Ph Eur

DEFINITION

1-[(2-Chlorophenyl)diphenylmethyl]-1*H*-imidazole.

Content
98.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance
White or pale yellow, crystalline powder.

Solubility
Practically insoluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 141 °C to 145 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clotrimazole CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *ethanol (96 per cent) R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 50 mg of *clotrimazole CRS* in *ethanol (96 per cent) R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R1, propanol R, toluene R* (0.5:10:90 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *acetonitrile R1* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R1*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R1*.

Reference solution (b) Dissolve the contents of a vial of *clotrimazole for peak identification CRS* (containing impurities A, B and F) in 1.0 mL of *acetonitrile R1*.

Reference solution (c) Dissolve 5.0 mg of *imidazole CRS* (impurity D) and 5.0 mg of *clotrimazole impurity E CRS* in *acetonitrile R1* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 25.0 mL with *acetonitrile R1*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.0 g of *potassium dihydrogen phosphate R* and 0.5 g of *tetrabutylammonium hydrogen sulfate R1* in *water R* and dilute to 1000 mL with the same solvent;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	75	25
3 - 25	75 → 20	25 → 80
25 - 30	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Relative retention With reference to clotrimazole (retention time = about 12 min): impurity D = about 0.1; impurity F = about 0.9; impurity B = about 1.1; impurity E = about 1.5; impurity A = about 1.8.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity F and clotrimazole;
— the chromatogram obtained is similar to the chromatogram supplied with *clotrimazole for peak identification CRS.*

Limits:
— *impurities A, B*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurities D, E*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *impurity F*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

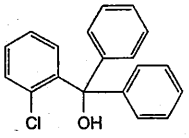
ASSAY
Dissolve 0.300 g in 80 mL of *anhydrous acetic acid R*. Using 0.3 mL of *naphtholbenzein solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from brownish-yellow to green.

1 mL of *0.1 M perchloric acid* is equivalent to 34.48 mg of $C_{22}H_{17}ClN_2$.

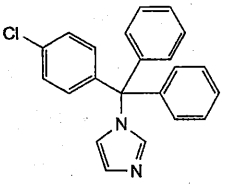
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, D, E, F.*

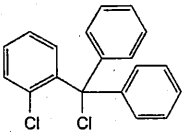
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



A. (2-chlorophenyl)diphenylmethanol,

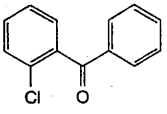


B. 1-[(4-chlorophenyl)diphenylmethyl]-1*H*-imidazole,

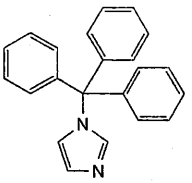


C. 1-chloro-2-(chlorodiphenylmethyl)benzene,

D. imidazole,



E. (2-chlorophenyl)phenylmethanone (2-chlorobenzophenone),

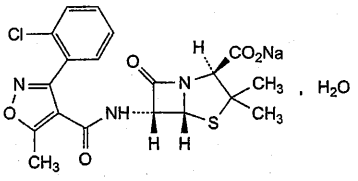


F. 1-(triphenylmethyl)-1*H*-imidazole (deschloroclotrimazole).

Ph Eur

Cloxacillin Sodium

(Ph. Eur. monograph 0661)



$C_{19}H_{17}ClN_3NaO_5S,H_2O$ 475.9 *7081-44-9*

Action and use
Penicillin antibacterial.

Ph Eur

DEFINITION
Sodium (2*S*,5*R*,6*R*)-6-[[[3-(2-chlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water and in methanol, soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *cloxacillin sodium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *cloxacillin sodium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *cloxacillin sodium CRS*, 25 mg of *dicloxacillin sodium CRS* and 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R*, then adjust to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear; examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R*. Mix the contents of the tube by swirling; the solution is slightly greenish-yellow. Place the test-tube in a water-bath for 1 min; the solution becomes yellow.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.04.

pH (*2.2.3*)

5.0 to 7.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 160 to + 169 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *cloxacillin sodium CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of test solution (b) to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *flucloxacillin sodium CRS* and 5 mg of *cloxacillin sodium CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 25 volumes of *acetonitrile R* and 75 volumes of a 2.7 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 5.0 with *dilute sodium hydroxide solution R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (a) and reference solutions (b) and (c).

Run time 5 times the retention time of cloxacillin.

System suitability Reference solution (c):
— *resolution*: minimum 2.5 between the peaks due to cloxacillin (1st peak) and flucloxacillin (2nd peak).

Limits:
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

N,*N*-Dimethylaniline (*2.4.26, Method B*)

Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)

Maximum 0.8 per cent *m/m*.

Water (*2.5.12*)

3.0 per cent to 4.5 per cent, determined on 0.300 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.20 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

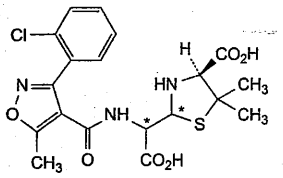
System suitability:
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections of reference solution (a).

Calculate the percentage content of $C_{19}H_{17}ClN_3NaO_5S$ from the declared content of *cloxacillin sodium CRS*.

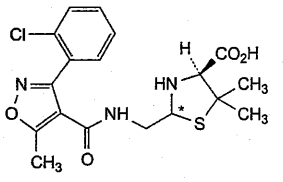
STORAGE

In an airtight container, at a temperature not exceeding 25 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

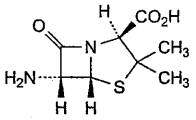
IMPURITIES



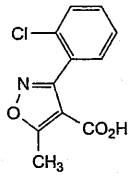
A. (4*S*)-2-[carboxy[[[3-(2-chlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acid of cloxacillin),



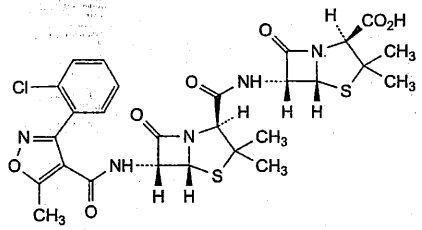
B. (2*RS*,4*S*)-2-[[[[3-(2-chlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penilloic acid of cloxacillin),



C. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



D. 3-(2-chlorophenyl)-5-methyl-1,2-oxazole-4-carboxylic acid,

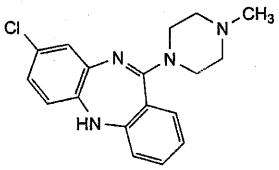


E. (2*S*,5*R*,6*R*)-6-[[[(2*S*,5*R*,6*R*)-6-[[[3-(2-chlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-APA cloxacillin amide).

Ph Eur

Clozapine

(Ph. Eur. monograph 1191)



$C_{18}H_{19}ClN_4$ 326.8 *5786-21-0*

Action and use
Dopamine D_4 receptor antagonist; neuroleptic.

Preparation
Clozapine Oral Suspension

Ph Eur

DEFINITION
8-Chloro-11-(4-methylpiperazin-1-yl)-5*H*-dibenzo[*b*,*e*][1,4]diazepine.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
Yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent). It dissolves in dilute acetic acid.

IDENTIFICATION
A. Melting point (*2.2.14*): 182 °C to 186 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *clozapine CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *water R, methanol R2* (20:80 *V/V*).

Solution A Dissolve 2.04 g of *potassium dihydrogen phosphate R* in 1000 mL of *water R* and adjust to pH 2.4 ± 0.05 with *dilute phosphoric acid R.*

Test solution Dissolve 75 mg of the substance to be examined in 80 mL of *methanol R2* and dilute to 100 mL with *water R.*

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *clozapine for peak identification CRS* (containing impurities A, B, C and D) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.125 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *acetonitrile for chromatography R, methanol R2*, solution A (1:1:8 *V/V/V*);
— *mobile phase B*: *acetonitrile for chromatography R, methanol R2*, solution A (4:4:2 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	100	0
4 - 24	100 → 0	0 → 100
24 - 29	0	100

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 257 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *clozapine for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to clozapine (retention time = about 11 min): impurity C = about 0.9; impurity D = about 1.1; impurity A = about 1.6; impurity B = about 1.7.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurity C and clozapine;
— the chromatogram obtained with reference solution (b) is similar to the chromatogram supplied with *clozapine for peak identification CRS.*

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 2.7;

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *impurities B, D*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

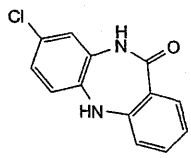
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

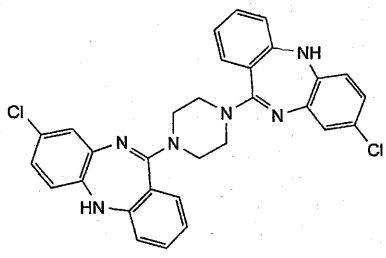
ASSAY
Dissolve 0.100 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 16.34 mg of $C_{18}H_{19}ClN_4$.

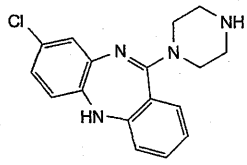
IMPURITIES
Specified impurities *A, B, C, D.*



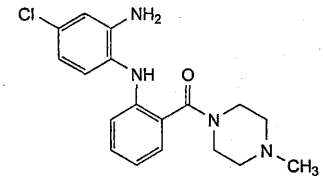
A. 8-chloro-5,10-dihydro-11*H*-dibenzo[*b*,*e*][1,4]diazepin-11-one,



B. 11,11′-(piperazine-1,4-diyl)bis(8-chloro-5*H*-dibenzo[*b*,*e*][1,4]diazepine),



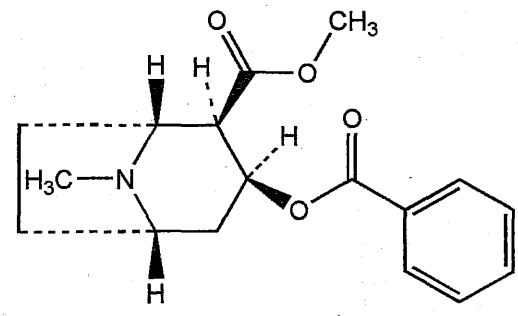
C. 8-chloro-11-(piperazin-1-yl)-5*H*-dibenzo[*b*,*e*][1,4]diazepine,



D. 1-[2-[(2-amino-4-chlorophenyl)amino]benzoyl]-4-methylpiperazine.

Ph Eur

Cocaine



$C_{17}H_{21}NO_4$ 303.4 *50-36-2*

Action and use
Local anaesthetic.

DEFINITION
Cocaine is methyl (1*R*,2*R*,3*S*,5*S*)-3-(benzoyloxy)-8-methyl-8-azabicyclo[3.2.1]octane-2-carboxylate and may be obtained from the leaves of *Erythroxylum coca* Lam. and other species of *Erythroxylum* or by synthesis. It contains not less than 98.0% and not more than 101.0% of $C_{17}H_{21}NO_4$, calculated with reference to the dried substance.

CHARACTERISTICS
Colourless crystals or a white, crystalline powder. Slightly volatile.

Practically insoluble in *water*; freely soluble in *ethanol (96%)* and in *ether*; soluble in arachis oil; slightly soluble in *liquid paraffin*.

IDENTIFICATION
The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of cocaine *(RS 071)*.

TESTS
Melting point
96° to 98°, Appendix V A.

Specific optical rotation
In a 2.4% w/v solution in 0.1M *hydrochloric acid*, −79 to −81, calculated with reference to the dried substance, Appendix V F.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) 0.05% w/v of the substance being examined in the mobile phase

(2) Dilute 1 volume of solution (1) to 50 volumes with the mobile phase, dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

(3) Dissolve 25 mg of the substance being examined in 0.01M *sodium hydroxide* and dilute to 100.0 mL with the same solvent. Allow the solution to stand for 15 minutes.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 4.6 mm) packed with *base-deactivated octadecylsilyl silica gel for chromatography* (5 µm) (Waters Symmetry is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use a column temperature of 35°.

(e) Use a detection wavelength of 216 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

1 volume of *triethylamine*, 200 volumes of *tetrahydrofuran*, 860 volumes of *acetonitrile* and 959 volumes of *water*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution factor* between the peaks due to cocaine (retention time, about 7 minutes) and the degradation product is at least 5.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak eluting after the principal peak is not greater than the area of the peak in the chromatogram obtained with solution (2) (0.1%);

the sum of the areas of any *secondary peaks* is not greater than 5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

Disregard any peak with an area less than 0.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Loss on drying

When dried to constant weight at 80°, loses not more than 0.5% of its weight, Appendix IX D. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

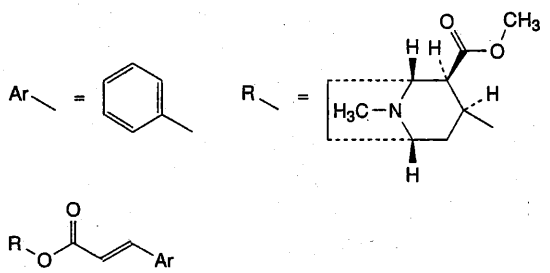
ASSAY

Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.7 g dissolved in 50 mL of *1,4-dioxan* and *crystal violet solution* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 30.34 mg of $C_{17}H_{21}NO_4$.

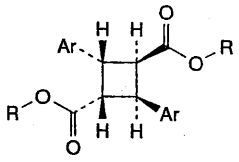
STORAGE

Cocaine should be stored protected from light.

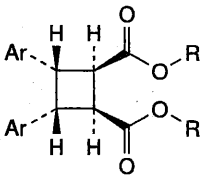
IMPURITIES



A. methyl (1*R*,2*R*,3*S*,5*S*)-8-methyl-3-[[(*E*)-3-phenylpropenoyl]oxy]-8-azabicyclo[3.2.1]octane-2-carboxylate (cinnamoylcocaine),



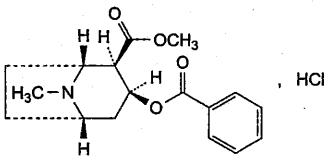
B. bis[(1*R*,2*R*,3*S*,5*S*)-2-(methoxycarbonyl)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl] (1*r*,2*c*,3*t*,4*t*)-2,4-diphenylcyclobutane-1,3-dicarboxylate (α-truxilline),



C. bis[(1*R*,2*R*,3*S*,5*S*)-2-(methoxycarbonyl)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl] (1*r*,2*c*,3*t*,4*t*)-3,4-diphenylcyclobutane-1,2-dicarboxylate (β-truxilline).

Cocaine Hydrochloride

(Ph. Eur. monograph 0073)



$C_{17}H_{22}ClNO_4$ 339.8 *53-21-4*

Action and use

Local anaesthetic.

Preparations

Adrenaline and Cocaine Intranasal Solution

Cocaine Eye Drops

Cocaine Paste

Ph Eur

DEFINITION

Methyl (1*R*,2*R*,3*S*,5*S*)-3-(benzoyloxy)-8-methyl-8-azabicyclo [3.2.1]octane-2-carboxylate hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very soluble in water, freely soluble in alcohol, slightly soluble in methylene chloride.

mp

About 197 °C, with decomposition.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D, E.

A. Dissolve 20.0 mg in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of the solution to 50.0 mL with *0.01 M hydrochloric acid*. Examined between

220 nm and 350 nm (*2.2.25*), the solution shows 2 absorption maxima, at 233 nm and 273 nm. The specific absorbance at 233 nm is 378 to 402.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of cocaine hydrochloride.*

C. Dissolve 0.1 g in 5 mL of *water R* and add 1 mL of *dilute ammonia R2*. A white precipitate is formed. Initiate crystallisation by scratching the wall of the tube with a glass rod. The crystals, washed with *water R* and dried *in vacuo*, melt (*2.2.14*) at 96 °C to 99 °C.

D. It gives reaction (a) of chlorides (*2.3.1*).

E. It gives the reaction of alkaloids (*2.3.1*).

TESTS

Solution S

Dissolve 0.5 g in *water R* and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity

To 10 mL of solution S add 0.05 mL of *methyl red solution R*. Not more than 0.2 mL of *0.02 M sodium hydroxide* is required to change the colour of the indicator.

Specific optical rotation (*2.2.7*)

−70 to −73 (dried substance).

Dissolve 0.50 g in *water R* and dilute to 20.0 mL with the same solvent.

Readily carbonisable substances

To 0.2 g add 2 mL of *sulfuric acid R*. After 15 min, the solution is not more intensely coloured than reference solution BY_5 (*2.2.2, Method I*).

Related substances

Examine by liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 25 mg of the substance to be examined in *0.01 M sodium hydroxide* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *0.01 M sodium hydroxide*. Allow the solution to stand for 15 min.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm) with a specific surface area of 335 m^2/g, a pore size of 10 nm and a carbon loading of 19.1 per cent,
— *temperature*: 35 °C.

Mobile phase *triethylamine R, tetrahydrofuran R, acetonitrile R, water R* (0.5:100:430:479.5 *V/V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 216 nm.

Injection 20 µL.

Relative retention With reference to cocaine (retention time = about 7.4 min): degradation product = about 0.7.

System suitability Reference solution (b):

— *resolution*: minimum of 5 between the peaks due to cocaine and to the degradation product.

Limits:

— *any impurity eluting after the principal peak*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on the residue from the test for loss on drying.

ASSAY

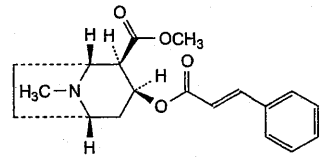
Dissolve 0.250 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 33.98 mg of $C_{17}H_{22}ClNO_4$.

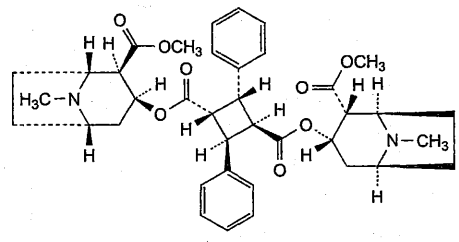
STORAGE

Protected from light.

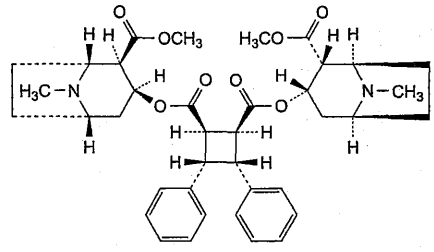
IMPURITIES



A. methyl (1*R*,2*R*,3*S*,5*S*)-8-methyl-3-[[(*E*)-3-phenylpropenoyl]oxy]-8-azabicyclo[3.2.1]octane-2-carboxylate (cinnamoylcocaine),



B. bis[(1*R*,2*R*,3*S*,5*S*)-2-(methoxycarbonyl)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl] (1*r*,2*c*,3*t*,4*t*)-2,4-diphenylcyclobutane-1,3-dicarboxylate (α-truxilline),



C. bis[(1*R*,2*R*,3*S*,5*S*)-2-(methoxycarbonyl)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl] (1*r*,2*c*,3*t*,4*t*)-3,4-diphenylcyclobutane-1,2-dicarboxylate (β-truxilline).

Cochineal

DEFINITION

Cochineal is the dried female insect, *Dactylopius coccus* Costa, containing eggs and larvae.

CHARACTERISTICS

Odour, characteristic.

Macroscopical Purplish black or purplish grey; about 3.5 to 5.5 mm long and 3 to 4.5 mm wide, plano-convex and somewhat oval in outline; the convex dorsal surface is transversely wrinkled and shows about 11 segments; the flat or slightly concave ventral surface carries upon the anterior part two seven-jointed straight antennae, three pairs of short legs, each terminating in a single claw, and a mouth from which projects the remains of a long filiform proboscis; these appendages are frequently more or less broken. Easily reduced to powder, which is dark red or puce.

Microscopical Scattered irregularly over the whole dermis are numerous solitary and grouped, short, tubular wax glands; within each insect are found numerous larvae, which are characterised by their proboscides appearing as two circular coils.

TESTS

Colour value

To 0.5 g in *moderately fine powder* add 60 mL of *phosphate buffer pH 8.0* and heat on a water bath for 30 minutes. Cool, add sufficient *phosphate buffer pH 8.0* to produce 100 mL and filter. Dilute 5 mL of the filtrate to 100 mL with *phosphate buffer pH 8.0*. The *absorbance* of the resulting solution at the maximum at 530 nm is not less than 0.25, Appendix II B.

Foreign matter

Complies with the test for *foreign matter*, Appendix XI D.

Water-insoluble matter

When the insects are placed in *water*, no insoluble powder separates.

Ash

Not more than 7.0%, Appendix XI J.

Microbial contamination

1 g is free from *Escherichia coli*; 10 g is free from Salmonella, Appendix XVI B1.

Coconut Oil

(Refined Coconut Oil, Ph. Eur. monograph 1410)

8001-31-8

Action and use

Excipient.

Ph Eur

DEFINITION

Fatty oil obtained from the dried, solid part of the endosperm of *Cocos nucifera* L., then refined.

CHARACTERS

Appearance

White or almost white, unctuous mass.

Solubility

Practically insoluble in water, freely soluble in methylene chloride and in light petroleum (bp: 65-70 °C), very slightly soluble in ethanol (96 per cent).

Refractive index

About 1.449, determined at 40 °C.

IDENTIFICATION

A. Melting point (see Tests).

B. Composition of fatty acids (see Tests).

TESTS

Melting point *(2.2.14)*

23 °C to 26 °C.

Acid value *(2.5.1)*

Maximum 0.5, determined on 20.0 g.

Peroxide value *(2.5.5, Method A)*

Maximum 5.0.

Unsaponifiable matter *(2.5.7)*

Maximum 1.0 per cent, determined on 5.0 g.

Alkaline impurities *(2.4.19)*

It complies with the test.

Composition of fatty acids *(2.4.22, Method B)*

Refined coconut oil is melted under gentle heating to a homogeneous liquid prior to sampling.

Reference solution Dissolve 15.0 mg of *tricaproin CRS*, 80.0 mg of *tristearin CRS*, 0.150 g of *tricaprin CRS*, 0.200 g of *tricaprylin CRS*, 0.450 g of *trimyristin CRS* and 1.25 g of *trilaurin CRS* in a mixture of 2 volumes of *methylene chloride R* and 8 volumes of *heptane R*, then dilute to 50 mL with the same mixture of solvents heating at 45-50 °C. Transfer 2 mL of this mixture to a 10 mL centrifuge tube with a screw cap and evaporate the solvent in a current of *nitrogen R*. Dissolve with 1 mL of *heptane R* and 1 mL of *dimethyl carbonate R* and mix vigorously under gentle heating (50-60 °C). Add, while still warm, 1 mL of a 12 g/L solution of *sodium R* in *anhydrous methanol R*, prepared with the necessary precautions, and mix vigorously for about 5 min. Add 3 mL of *distilled water R* and mix vigorously for about 30 s. Centrifuge for 15 min at 1500 *g*. Inject 1 µL of the organic phase.

Calculate the percentage content of each fatty acid using the following expression:

$$\frac{A_{x,s,c}}{\sum A_{x,s,c}} \times 100 \text{ per cent } m/m$$

$A_{x,s,c}$ is the corrected peak area of each fatty acid in the test solution:

$$A_{x,s,c} = A_{x,s} \times R_c$$

R_c is the relative correction factor:

$$R_c = \frac{m_{x,r} \times A_{1,r}}{A_{x,r} \times m_{1,r}}$$

for the peaks due to caproic, caprylic, capric, lauric and myristic acid methyl esters.

$m_{x,r}$	=	mass of tricaproin, tricaprylin, tricaprin, trilaurin or trimyristin in the reference solution, in milligrams;
$m_{1,r}$	=	mass of tristearin in the reference solution, in milligrams:
$A_{x,r}$	=	area of the peaks due to caproic, caprylic, capric, lauric and myristic acid methyl esters in the reference solution;
$A_{1,r}$	=	area of the peak due to stearic acid methyl ester in the reference solution;
$A_{x,s}$	=	area of the peaks due to any specified or unspecified fatty acid methyl esters;
R_c	=	1 for the peaks due to each of the remaining specified fatty acid methyl esters or any unspecified fatty acid methyl ester.

Composition of the fatty-acid fraction of the oil:
— *caproic acid (R_{Rt} 0.11)*: maximum 1.5 per cent,
— *caprylic acid (R_{Rt} 0.23)*: 5.0 per cent to 11.0 per cent,
— *capric acid (R_{Rt} 0.56)*: 4.0 per cent to 9.0 per cent,
— *lauric acid (R_{Rt} 0.75)*: 40.0 per cent to 50.0 per cent,
— *myristic acid (R_{Rt} 0.85)*: 15.0 per cent to 20.0 per cent,
— *palmitic acid (R_{Rt} 0.93)*: 7.0 per cent to 12.0 per cent,
— *stearic acid (R_{Rt} 1.00)*: 1.5 per cent to 5.0 per cent,
— *oleic acid (R_{Rt} 1.01)*: 4.0 per cent to 10.0 per cent,
— *linoleic acid (R_{Rt} 1.03)*: 1.0 per cent to 3.0 per cent,
— *linolenic acid (R_{Rt} 1.06)*: maximum 0.2 per cent,
— *arachidic acid (R_{Rt} 1.10)*: maximum 0.2 per cent,
— *eicosenoic acid (R_{Rt} 1.11)*: maximum 0.2 per cent.

Water (*2.5.32*)
Maximum 0.1 per cent, determined on 1.00 g.

STORAGE
In a well-filled container, protected from light.

Ph Eur

Cocoyl Caprylocaprate

(Ph. Eur. monograph 1411)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of esters of saturated C_{12} - C_{18} alcohols with caprylic (octanoic) and capric (decanoic) acids obtained by the reaction of these acids with vegetable saturated fatty alcohols.

CHARACTERS
Appearance
Slightly yellowish liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent) and with liquid paraffin.

Relative density
About 0.86.

Refractive index
About 1.445.

Viscosity
About 11 mPa·s.

IDENTIFICATION
A. Freezing point (*2.2.18*): maximum 15 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cocoyl caprylocaprate CRS.*
C. Composition of fatty acids and fatty alcohols (see Tests).

TESTS
Appearance
The substance to be examined is not more intensely coloured than reference solution Y_5 (*2.2.2, Method I*).

Acid value (*2.5.1*)
Maximum 0.5, determined on 5.00 g.

Hydroxyl value (*2.5.3, Method A*)
Maximum 5.0, determined on 4.0 g. Add 5.0 mL of acetylating reagent.

Iodine value (*2.5.4, Method A*)
Maximum 1.0.

Saponification value (*2.5.6*)
160 to 173.

Composition of fatty acids and fatty alcohols (*2.4.22, Method C*)
Use the chromatogram obtained with the following reference solution for identification of the peaks due to the fatty alcohols.

Reference solution Dissolve the amounts of the substances listed in the following table in 10 mL of *heptane R*.

Substance	Amount (mg)
Methyl caproate R	10
Methyl caprylate R	90
Methyl decanoate R	50
Methyl laurate R	20
Methyl myristate R	10
Methyl palmitate R	10
Methyl stearate R	10
Decanol R	10
Lauryl alcohol R	100
Myristyl alcohol R	40
Cetyl alcohol CRS	30
Stearyl alcohol CRS	20

Consider the sum of the areas of the peaks due to the fatty acids listed below to be equal to 100 and the sum of the areas of the peaks due to the fatty alcohols listed below to be equal to 100.

Composition of the fatty acid fraction of the substance:
— *caproic acid*: maximum 2.0 per cent,
— *caprylic acid*: 50.0 per cent to 80.0 per cent,
— *capric acid*: 20.0 per cent to 50.0 per cent,
— *lauric acid*: maximum 3.0 per cent,
— *myristic acid*: maximum 2.0 per cent.

Composition of the fatty alcohol fraction of the substance:
— *capric alcohol*: maximum 3.0 per cent,
— *lauryl alcohol*: 48.0 per cent to 63.0 per cent,
— *myristyl alcohol*: 18.0 per cent to 27.0 per cent,
— *cetyl alcohol*: 6.0 per cent to 13.0 per cent,
— *stearyl alcohol*: 9.0 per cent to 16.0 per cent.

Water (*2.5.12*)
Maximum 0.1 per cent, determined on 5.00 g.

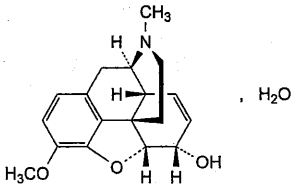
Total ash (*2.4.16*)
Maximum 0.1 per cent, determined on 1.0 g.

Ph Eur

Codeine Monohydrate

Codeine

(Ph. Eur. monograph 0076)



$C_{18}H_{21}NO_3,H_2O$ 317.4 *6059-47-8*

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION

4,5α-Epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol monohydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, C, F.

Second identification: A, B, D, E, F.

A. Melting point (*2.2.14*): 155 °C to 159 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution To 2.0 mL of solution S (see Tests) add 50 mL of *water R* then 10 mL of *1 M sodium hydroxide* and dilute to 100.0 mL with *water R*.

Spectral range 250-350 nm.

Absorption maximum At 284 nm.

Specific absorbance at the absorption maximum About 50 (dried substance).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *codeine CRS*.

D. To about 10 mg add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R2* and heat on a water-bath. A blue colour develops. Add 0.05 mL of *nitric acid R*. The colour changes to red.

E. It gives the reaction of alkaloids (*2.3.1*).

F. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 50 mg in *water R* and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

−142 to −146 (dried substance).

Dissolve 0.50 g in *ethanol (96 per cent) R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solution A 0.5 per cent *V/V* solution of *phosphoric acid R*.

Test solution Dissolve 0.160 g of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 3.0 mg of *codeine for system suitability CRS* (containing impurities A, B, C, D, E, F, G, H and I) in 1.0 mL of solution A.

Column:

— *size*: *l* = 0.075 m, Ø = 3.0 mm;

— *stationary phase*: *end-capped octadecylsilyl multi-layered organosilica polymer R* (1.9 µm).

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: mix 4 volumes of *acetonitrile R* and 96 volumes of a 20 g/L solution of *glacial acetic acid R* previously adjusted to pH 4.5 with a 500 g/L solution of *sodium hydroxide R*;

— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 7.33	100 → 93	0 → 7
7.33 - 10.33	93 → 67	7 → 33
10.33 - 12	67	33

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 3 µL.

Identification of impurities Use the chromatogram supplied with *codeine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G, H and I.

Relative retention With reference to codeine (retention time = about 6.0 min): impurity B = about 0.3; impurity E = about 0.4; impurity F = about 0.8; impurity H = about 0.9; impurity C = about 1.2; impurity I = about 1.4; impurity D = about 1.45; impurity A = about 1.5; impurity G = about 1.6.

System suitability Reference solution (b):

— *resolution*: minimum 2.5 between the peaks due to impurities F and H; minimum 1.5 between the peaks due to impurities D and A.

Calculation of percentage contents:

— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7; impurity G = 0.2; impurity I = 1.3;

— for each impurity, use the concentration of codeine in reference solution (a).

Limits:

— *impurity A*: maximum 1.0 per cent;

— *impurity H*: maximum 0.25 per cent;

— *impurities C, D, E*: for each impurity, maximum 0.2 per cent;

— *impurities B, F, G, I*: for each impurity, maximum 0.15 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 1.5 per cent;

— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

4.0 per cent to 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 29.94 mg of $C_{18}H_{21}NO_3$.

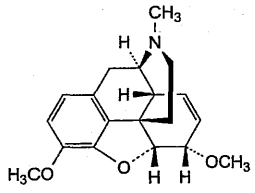
STORAGE

Protected from light.

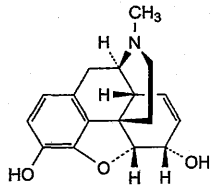
IMPURITIES

Specified impurities A, B, C, D, E, F, G, H, I.

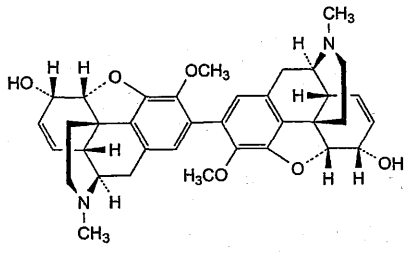
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) J, K, L, M.



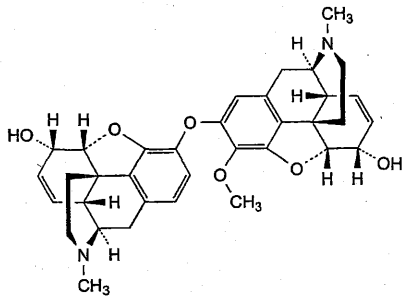
A. 4,5α-epoxy-3,6α-dimethoxy-17-methyl-7,8-didehydromorphinan (methylcodeine),



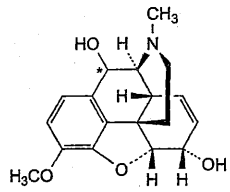
B. 4,5α-epoxy-17-methyl-7,8-didehydromorphinan-3,6α-diol (morphine),



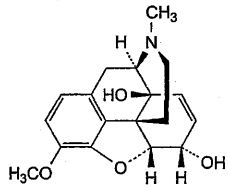
C. 4,5α:4′,5′α-diepoxy-3,3′-dimethoxy-17,17′-dimethyl-7,7′,8,8′-tetradehydro-2,2′-bimorphinan-6α,6′α-diol (codeine dimer),



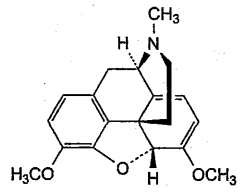
D. 2-[(4,5α-epoxy-6α-hydroxy-17-methyl-7,8-didehydromorphinan-3-yl)oxy]-4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol (3-*O*-(codein-2-yl) morphine),



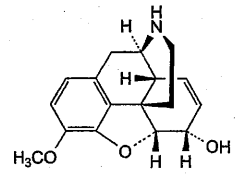
E. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,10ξ-diol,



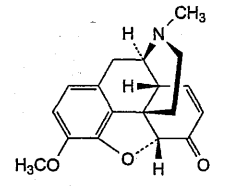
F. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,14-diol,



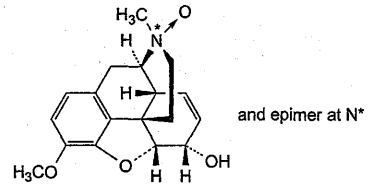
G. 4,5α-epoxy-3,6-dimethoxy-17-methyl-6,7,8,14-tetradehydromorphinan (thebaine),



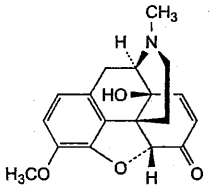
H. 4,5α-epoxy-3-methoxy-7,8-didehydromorphinan-6α-ol (norcodeine),



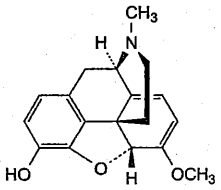
I. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (codeinone),



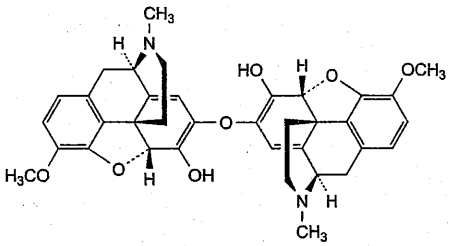
J. (17*RS*)-4,5α-epoxy-6α-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan 17-oxide (codeine *N*-oxide),



K. 4,5α-epoxy-14-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (14-hydroxycodeinone),



L. 4,5α-epoxy-6-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-3-ol (oripavine),

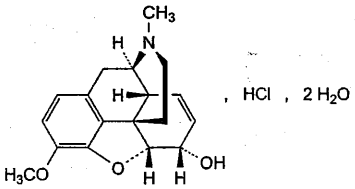


M. 7,7′-oxybis(4,5α-epoxy-3-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-6-ol) (7,7′-oxybis(6-*O*-demethylthebaine)).

Ph Eur

Codeine Hydrochloride

(Codeine Hydrochloride Dihydrate, Ph. Eur. monograph 1412)



$C_{18}H_{22}ClNO_3,2H_2O$ 371.9

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION
4,5α-Epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol hydrochloride dihydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or small, colourless crystals.

Solubility
Soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION
First identification: A, D, F.

Second identification: B, C, D, E, F.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *codeine hydrochloride dihydrate CRS.*

B. To 5 mL of solution S (see Tests) add 1 mL of a mixture of equal volumes of *strong sodium hydroxide solution R* and *water R* and initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod and cooling in iced water. Wash the precipitate with *water R* and dry at 100-105 °C. It melts (*2.2.15*) at 155 °C to 159 °C.

C. To about 10 mg add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R2* and heat on a water-bath. A blue colour develops. Add 0.05 mL of *nitric acid R*. The colour changes to red.

D. Solution S gives reaction (a) of chlorides (*2.3.1*).

E. It gives the reaction of alkaloids (*2.3.1*).

F. Water (see Tests).

TESTS
Solution S
Dissolve 2.00 g in *carbon dioxide-free water R*, prepared from *distilled water R*, and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 5 mL of solution S add 5 mL of *carbon dioxide-free water R*. Add 0.05 mL of *methyl red solution R* and 0.2 mL of *0.02 M hydrochloric acid*; the solution is red. Add 0.4 mL of *0.02 M sodium hydroxide*; the solution becomes yellow.

Specific optical rotation (*2.2.7*)
−117 to −121 (anhydrous substance).

Dilute 5.0 mL of solution S to 10.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Solution A 0.5 per cent *V/V* solution of *phosphoric acid R*.

Test solution Dissolve 0.190 g of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 3.0 mg of *codeine for system suitability CRS* (containing impurities A, B, C, D, E, F, G, H and I) in 1.0 mL of solution A.

Column:
— *size*: *l* = 0.075 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl multi-layered organosilica polymer R* (1.9 µm).
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: mix 4 volumes of *acetonitrile R* and 96 volumes of a 20 g/L solution of *glacial acetic acid R* previously adjusted to pH 4.5 with a 500 g/L solution of *sodium hydroxide R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 7.33	100 → 93	0 → 7
7.33 - 10.33	93 → 67	7 → 33
10.33 - 12	67	33

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 3 µL.

Identification of impurities Use the chromatogram supplied with *codeine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G, H and I.

Relative retention With reference to codeine (retention time = about 6.0 min): impurity B = about 0.3; impurity E = about 0.4; impurity F = about 0.8; impurity H = about 0.9; impurity C = about 1.2; impurity I = about 1.4; impurity D = about 1.45; impurity A = about 1.5; impurity G = about 1.6.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurities F and H; minimum 1.5 between the peaks due to impurities D and A.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7; impurity G = 0.2; impurity I = 1.3;
— for each impurity, use the concentration of codeine in reference solution (a).

Limits:
— *impurity A*: maximum 1.0 per cent;
— *impurity H*: maximum 0.25 per cent;
— *impurities C, D, E*: for each impurity, maximum 0.2 per cent;
— *impurities B, F, G, I*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

Dilute 5 mL of solution S to 20 mL with *distilled water R.*

Water (*2.5.12*)
8.0 per cent to 10.5 per cent, determined on 0.250 g.

ASSAY

Dissolve 0.300 g in a mixture of 5 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R.* Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 33.59 mg of $C_{18}H_{22}ClNO_3$.

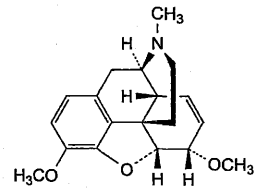
STORAGE

Protected from light.

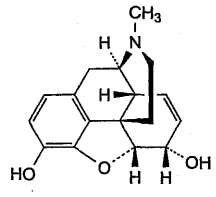
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H, I.*

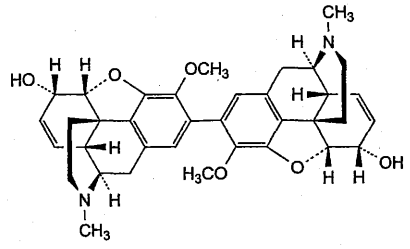
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *J, K, L, M.*



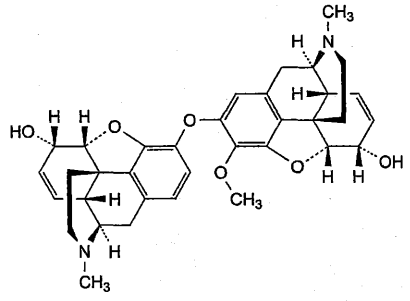
A. 4,5α-epoxy-3,6α-dimethoxy-17-methyl-7,8-didehydromorphinan (methylcodeine),



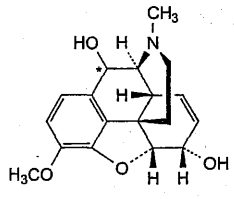
B. 4,5α-epoxy-17-methyl-7,8-didehydromorphinan-3,6α-diol (morphine),



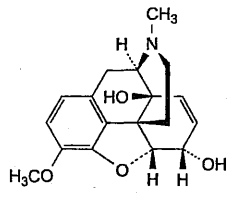
C. 4,5α:4′,5′α-diepoxy-3,3′-dimethoxy-17,17′-dimethyl-7,7′,8,8′-tetradehydro-2,2′-bimorphinan-6α,6′α-diol (codeine dimer),



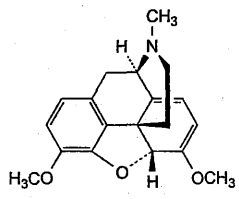
D. 2-[(4,5α-epoxy-6α-hydroxy-17-methyl-7,8-didehydromorphinan-3-yl)oxy]-4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol (3-*O*-(codein-2-yl) morphine),



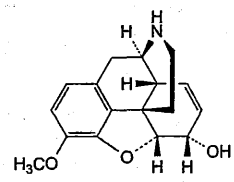
E. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,10ξ-diol,



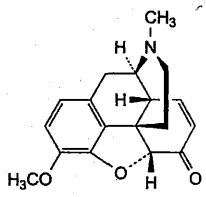
F. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,14-diol,



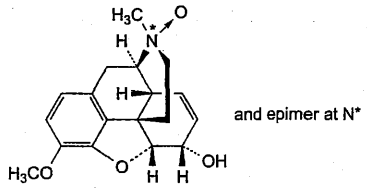
G. 4,5α-epoxy-3,6-dimethoxy-17-methyl-6,7,8,14-tetradehydromorphinan (thebaine),



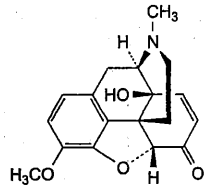
H. 4,5α-epoxy-3-methoxy-7,8-didehydromorphinan-6α-ol (norcodeine),



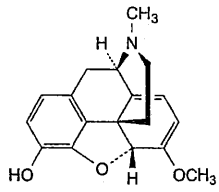
I. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (codeinone),



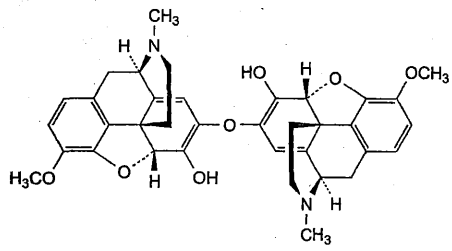
J. (17*RS*)-4,5α-epoxy-6α-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan 17-oxide (codeine *N*-oxide),



K. 4,5α-epoxy-14-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (14-hydroxycodeinone),



L. 4,5α-epoxy-6-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-3-ol (oripavine),

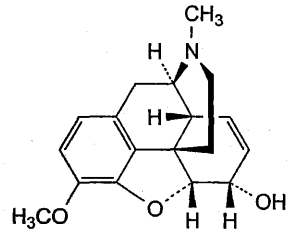


M. 7,7′-oxybis(4,5α-epoxy-3-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-6-ol) (7,7′-oxybis(6-*O*-demethylthebaine)).

Ph Eur

Codeine Phosphate

(Codeine Phosphate Hemihydrate, Ph. Eur. monograph 0074)



$C_{18}H_{24}NO_7P,½H_2O$ 406.4 *41444-62-6*

Action and use
Opioid receptor agonist; analgesic.

Preparations
Co-codamol Capsules
Co-codamol Tablets
Co-codamol Effervescent Tablets
Co-codaprin Tablets
Co-codaprin Dispersible Tablets
Codeine Linctus
Paediatric Codeine Linctus
Codeine Phosphate Injection
Codeine Phosphate Oral Solution
Codeine Phosphate Tablets
Paracetamol, Codeine Phosphate and Caffeine Capsules
Paracetamol, Codeine Phosphate and Caffeine Tablets

Ph Eur

DEFINITION
4,5α-Epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol phosphate hemihydrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or small, colourless crystals.

Solubility
Freely soluble in water, slightly soluble or very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: B, E, F.

Second identification: A, C, D, E, F, G.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dilute 1.0 mL of solution S (see Tests) to 100.0 mL with *water R*. To 25.0 mL of this solution add 25 mL of *water R* then 10 mL of *1 M sodium hydroxide* and dilute to 100.0 mL with *water R*.

Spectral range 250-350 nm.

Absorption maximum At 284 nm.

Specific absorbance at the absorption maximum About 38 (dried substance).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 0.20 g in 4 mL of *water R*. Add 1 mL of a mixture of equal volumes of *strong sodium hydroxide solution R* and *water R* and initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod and cooling in iced water. Wash the precipitate with *water R* and dry at 100-105 °C.

Comparison *codeine CRS*.

C. Dissolve 0.20 g in 4 mL of *water R*. Add 1 mL of a mixture of equal volumes of *strong sodium hydroxide solution R* and *water R* and initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod and cooling in iced water. The precipitate, washed with *water R* and dried at 100-105 °C, melts (*2.2.14*) at 155 °C to 159 °C.

D. To about 10 mg add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R2* and heat on a water-bath. A blue colour develops. Add 0.05 mL of *nitric acid R*. The colour changes to red.

E. Loss on drying (see Tests).

F. Solution S gives reaction (a) of phosphates (*2.3.1*).

G. It gives the reaction of alkaloids (*2.3.1*).

TESTS

Solution S
Dissolve 1.00 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 25.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)
−98 to −102 (dried substance).

Dilute 5.0 mL of solution S to 10.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Solution A 0.5 per cent *V/V* solution of *phosphoric acid R*.

Test solution Dissolve 0.190 g of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 3.0 mg of *codeine for system suitability CRS* (containing impurities A, B, C, D, E, F, G, H and I) in 1.0 mL of solution A.

Column:
— *size*: *l* = 0.075 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl multi-layered organosilica polymer R* (1.9 µm).
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: mix 4 volumes of *acetonitrile R* and 96 volumes of a 20 g/L solution of *glacial acetic acid R* previously adjusted to pH 4.5 with a 500 g/L solution of *sodium hydroxide R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 7.33	100 → 93	0 → 7
7.33 - 10.33	93 → 67	7 → 33
10.33 - 12	67	33

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 3 µL.

Identification of impurities Use the chromatogram supplied with *codeine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G, H and I.

Relative retention With reference to codeine (retention time = about 6.0 min): impurity B = about 0.3; impurity E = about 0.4; impurity F = about 0.8; impurity H = about 0.9; impurity C = about 1.2; impurity I = about 1.4; impurity D = about 1.45; impurity A = about 1.5; impurity G = about 1.6.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurities F and H; minimum 1.5 between the peaks due to impurities D and A.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7; impurity G = 0.2; impurity I = 1.3;
— for each impurity, use the concentration of codeine in reference solution (a).

Limits:
— *impurity A*: maximum 1.0 per cent;
— *impurity H*: maximum 0.25 per cent;
— *impurities C, D, E*: for each impurity, maximum 0.2 per cent;
— *impurities B, F, G, I*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

Dilute 5 mL of solution S to 20 mL with *distilled water R*.

Loss on drying (*2.2.32*)
1.5 per cent to 3.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.350 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 39.74 mg of $C_{18}H_{24}NO_7P$.

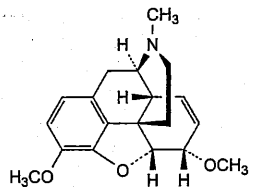
STORAGE

Protected from light.

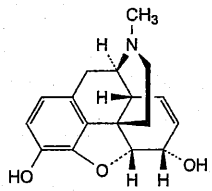
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H, I.*

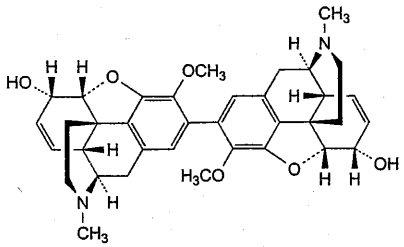
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *J, K, L, M.*



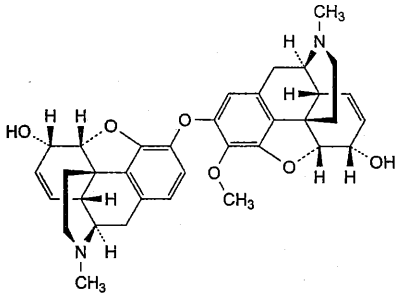
A. 4,5α-epoxy-3,6α-dimethoxy-17-methyl-7,8-didehydromorphinan (methylcodeine),



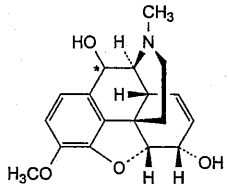
B. 4,5α-epoxy-17-methyl-7,8-didehydromorphinan-3,6α-diol (morphine),



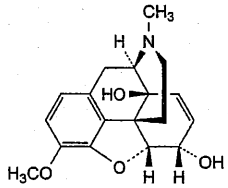
C. 4,5α:4′,5′α-diepoxy-3,3′-dimethoxy-17,17′-dimethyl-7,7′,8,8′-tetradehydro-2,2′-bimorphinan-6α,6′α-diol (codeine dimer),



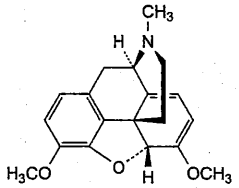
D. 2-[(4,5α-epoxy-6α-hydroxy-17-methyl-7,8-didehydromorphinan-3-yl)oxy]-4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α-ol (3-*O*-(codein-2-yl) morphine),



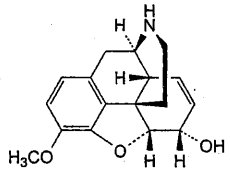
E. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,10ξ-diol,



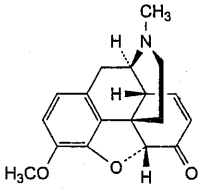
F. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6α,14-diol,



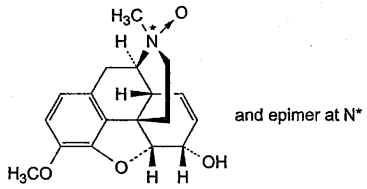
G. 4,5α-epoxy-3,6-dimethoxy-17-methyl-6,7,8,14-tetradehydromorphinan (thebaine),



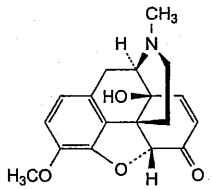
H. 4,5α-epoxy-3-methoxy-7,8-didehydromorphinan-6α-ol (norcodeine),



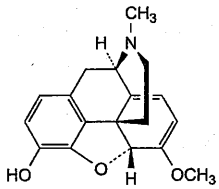
I. 4,5α-epoxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (codeinone),



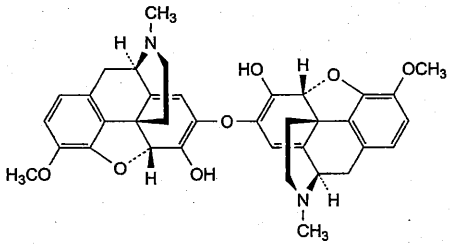
J. (17*RS*)-4,5α-epoxy-6α-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan 17-oxide (codeine *N*-oxide),



K. 4,5α-epoxy-14-hydroxy-3-methoxy-17-methyl-7,8-didehydromorphinan-6-one (14-hydroxycodeinone),



L. 4,5α-epoxy-6-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-3-ol (oripavine),

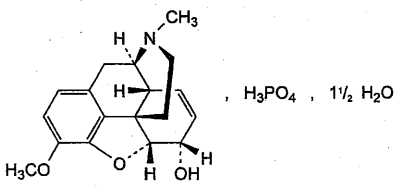


M. 7,7′-oxybis(4,5α-epoxy-3-methoxy-17-methyl-6,7,8,14-tetradehydromorphinan-6-ol) (7,7′-oxybis(6-*O*-demethylthebaine)).

Ph Eur

Codeine Phosphate Sesquihydrate

(Ph. Eur. monograph 0075)



$C_{18}H_{24}NO_7P,1½H_2O$ 424.4 *5913-76-8*

Action and use
Opioid receptor agonist; analgesic.

Preparations
Codeine Linctus
Paediatric Codeine Linctus
Codeine Phosphate Oral Solution
Codeine Phosphate Tablets

Ph Eur

DEFINITION
7,8-Didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α-ol phosphate sesquihydrate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or small, colourless crystals.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B, E, F.

Second identification: A, C, D, E, F, G.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dilute 1.0 mL of solution S (see Tests) to 100.0 mL with *water R*. To 25.0 mL of this solution add 25 mL of *water R* then 10 mL of *1 M sodium hydroxide* and dilute to 100.0 mL with *water R*.

Spectral range 250-350 nm.

Absorption maximum At 284 nm.

Specific absorbance at the absorption maximum About 38 (dried substance).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 0.20 g in 4 mL of *water R*. Add 1 mL of a mixture of equal volumes of *strong sodium hydroxide solution R* and *water R* and initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod and cooling in iced water. Wash the precipitate with *water R* and dry at 100-105 °C. Examine the dried precipitate prepared as discs using *potassium bromide R*.

Comparison *Ph. Eur. reference spectrum of codeine.*

C. Dissolve 0.20 g in 4 mL of *water R*. Add 1 mL of a mixture of equal volumes of *strong sodium hydroxide solution R* and *water R* and initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod and cooling in iced water. The precipitate, washed with *water R* and dried at 100-105 °C, melts (*2.2.14*) at 155 °C to 159 °C.

D. To about 10 mg add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R2* and heat on a water-bath. A blue colour develops. Add 0.05 mL of *nitric acid R*. The colour changes to red.

E. Loss on drying (see Tests).

F. Solution S gives reaction (a) of phosphates (*2.3.1*).

G. It gives the reaction of alkaloids (*2.3.1*).

TESTS
Solution S
Dissolve 1.00 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 25.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)
−98 to −102 (dried substance).

Dilute 5.0 mL of solution S to 10.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined and 0.100 g of *sodium octanesulfonate R* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *codeine impurity A CRS* in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (d) To 0.25 mL of the test solution add 2.5 mL of reference solution (a).

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 1.08 g of *sodium octanesulfonate R* in a mixture of 20 mL of *glacial acetic acid R* and 250 mL of *acetonitrile R* and dilute to 1000 mL with *water R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 10 µL.

Run time 10 times the retention time of codeine.

Relative retention With reference to codeine (retention time = about 6 min): impurity B = about 0.6; impurity E = about 0.7; impurity A = about 2.0; impurity C = about 2.3; impurity D = about 3.6.

System suitability Reference solution (d):
— *resolution*: minimum 3 between the peaks due to codeine and impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 0.25;
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *impurities B, C, D, E*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *sum of impurities other than A*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

Dilute 5 mL of solution S to 20 mL with *distilled water R*.

Loss on drying (*2.2.32*)
5.0 per cent to 7.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

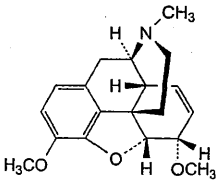
ASSAY
Dissolve 0.350 g in a mixture of 10 mL of *anhydrous acetic acid R* and 20 mL of *dioxan R*. Titrate with *0.1 M perchloric acid* using 0.05 mL of *crystal violet solution R* as indicator.

1 mL of *0.1 M perchloric acid* is equivalent to 39.74 mg of $C_{18}H_{24}NO_7P$.

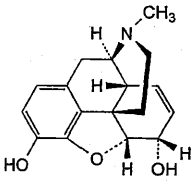
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C, D, E.*

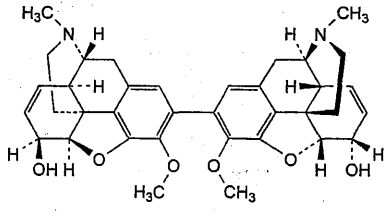
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G.*



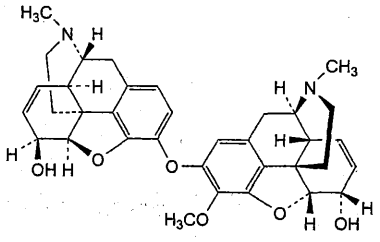
A. 7,8-didehydro-4,5α-epoxy-3,6α-dimethoxy-17-methylmorphinan (methylcodeine),



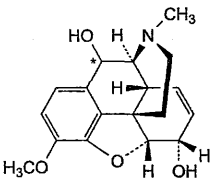
B. 7,8-didehydro-4,5α-epoxy-17-methylmorphinan-3,6α-diol (morphine),



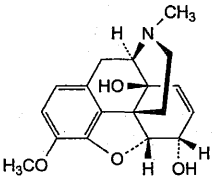
C. 7,7′,8,8′-tetradehydro-4,5α:4′,5′α-diepoxy-3,3′-dimethoxy-17,17′-dimethyl-2,2′-bimorphinanyl-6α,6′α-diol (codeine dimer),



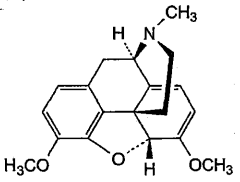
D. 7,8-didehydro-2-[(7,8-didehydro-4,5α-epoxy-6α-hydroxy-17-methylmorphinan-3-yl)oxy]-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α-ol (3-*O*-(codein-2-yl)morphine),



E. 7,8-didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α,10-diol,



F. 7,8-didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α,14-diol,

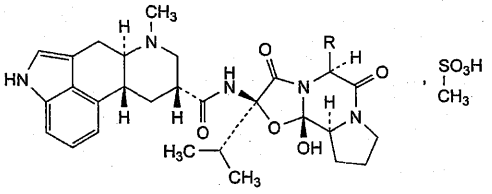


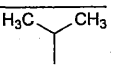
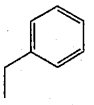
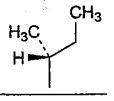
G. 6,7,8,14-tetradehydro-4,5α-epoxy-3,6-dimethoxy-17-methylmorphinan (thebaine).

Ph Eur

Codergocrine Mesilate

(Ph. Eur. monograph 2060)



Name	Mol. Formula	M_r	R
dihydroergocornine mesilate	$C_{32}H_{45}N_5O_8S$	660	
dihydroergocristine mesilate	$C_{36}H_{45}N_5O_8S$	708	
α-dihydroergocryptine mesilate	$C_{33}H_{47}N_5O_8S$	674	
β-dihydroergocryptine mesilate	$C_{33}H_{47}N_5O_8S$	674	

8067-24-1

Action and use
Vasodilator.

Ph Eur

DEFINITION

A mixture of:
- (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2,5-bis(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate (dihydroergocornine mesilate);
- (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate (dihydroergocristine mesilate);
- (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate (α-dihydroergocryptine mesilate);
- (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-[(1*RS*)-1-methylpropyl]-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate (β-dihydroergocryptine mesilate or epicriptine mesilate).

Content

98.0 per cent to 102.0 per cent (dried substance).

PRODUCTION

It is considered that alkylsulfonate esters are genotoxic and are potential impurities in codergocrine mesilate.
The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance

White or yellowish powder.

Solubility

Sparingly soluble in water, sparingly soluble to soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.20 g of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 0.20 g of *methanesulfonic acid R* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents.

Plate *TLC silica gel plate R.*

Mobile phase *water R, concentrated ammonia R, butanol R, acetone R* (5:10:20:65 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of cold air for not more than 1 min.

Detection Spray with a 1 g/L solution of *bromocresol purple R* in *methanol R*, adjusted to a violet-red colour with 0.05 mL of *dilute ammonia R1*.

Drying In a current of hot air at 100 °C.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the principal spot in the chromatogram obtained with the reference solution.

B. Examine the chromatograms obtained in the test for composition.

Results The 4 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 4 principal peaks in the chromatogram obtained with the reference solution.

TESTS

pH (*2.2.3*)

4.2 to 5.2.

Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Composition

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 20 mg of the substance to be examined in a mixture of 1 volume of *anhydrous ethanol R* and 2 volumes of a 10 g/L solution of *tartaric acid R* and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 20 mg of *codergocrine mesilate CRS* in a mixture of 1 volume of *anhydrous ethanol R* and 2 volumes of a 10 g/L solution of *tartaric acid R* and dilute to 10 mL with the same mixture of solvents.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *triethylamine R, acetonitrile R, water R* (2.5:25:75 *V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 20 min.

Elution order Dihydroergocornine, α-dihydroergocryptine, dihydroergocristine, β-dihydroergocryptine.

System suitability Test solution:
- *resolution*: minimum 3 between any 2 consecutive principal peaks.

Composition:
- *dihydroergocornine*: 30.0 per cent to 35.0 per cent;
- *α-dihydroergocryptine*: 20.0 per cent to 25.0 per cent;
- *dihydroergocristine*: 30.0 per cent to 35.0 per cent;
- *β-dihydroergocryptine*: 10.0 per cent to 13.0 per cent;
- *disregard limit*: 1.0 per cent.

Related substances

Thin-layer chromatography (*2.2.27*). *Perform the test as rapidly as possible and protected from direct light. Prepare the test solution last and immediately before application on the plate.*

Test solution Dissolve 0.40 g of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 40 mg of *dihydroergocristine mesilate CRS* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 10.0 mL with the same mixture of solvents. Dilute 3.0 mL of the solution to 50.0 mL with a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R*.

Reference solution (b) To 2.0 mL of reference solution (a), add 1.0 mL of a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R*.

Reference solution (c) To 1.0 mL of reference solution (a), add 2.0 mL of a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R*.

Reference solution (d) To 1.0 mL of reference solution (a), add 5.0 mL of a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R*.

Plate *TLC silica gel plate R*.

Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R, methylene chloride R* (1:3:50:50 *V/V/V/V*).

Application 10 µL.

Drying In the dark for 2 min after the application of the last solution.

First development In an unsaturated tank, over 2/3 of the plate.

Drying In a current of cold air for not more than 1 min.

Second development In an unsaturated tank, over 2/3 of the plate; use freshly prepared mobile phase.

Drying In a current of cold air for not more than 1 min.

Detection Spray thoroughly with *dimethylaminobenzaldehyde solution R7* and dry in a current of hot air until the spot in the chromatogram obtained with reference solution (d) is clearly visible.

System suitability Test solution:
- the chromatogram shows at least 3 separated secondary spots.

Limits:
- *any impurity*: any spots, apart from the principal spot, are not more intense than the spot in the chromatogram obtained with reference solution (a) (0.3 per cent); not more than 4 such spots are more intense than the spot in the chromatogram obtained with reference solution (c) (0.1 per cent) and 2 of these may be more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 0.500 g by drying at 120 °C under high vacuum.

ASSAY

Dissolve 0.500 g in 60 mL of *pyridine R*. Pass a stream of *nitrogen R* over the surface of the solution and titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 68.04 mg of codergocrine mesilate (average M_r = 680).

STORAGE

Protected from light.

Ph Eur

Farmed Cod-liver Oil

(Ph. Eur. monograph 2398)

Action and use
Source of vitamins A and D.

Ph Eur

DEFINITION

Purified fatty oil obtained from the fresh livers of farmed cod, *Gadus morhua* L., solid substances being removed by cooling and filtering.

Content

— *sum of the contents of EPA and DHA (expressed as triglycerides)*: 10.0 per cent to 28.0 per cent;
— *vitamin A*: 50 IU (15 µg) to 500 IU (150 µg) per gram;
— *vitamin D_3*: maximum 50 IU (1.3 µg) per gram.

A suitable antioxidant may be added.

PRODUCTION

The fish shall only be given feed with a composition that is in accordance with the relevant European Union or other applicable regulations.

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance
Clear, pale yellowish liquid.

Solubility
Practically insoluble in water, miscible with light petroleum, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Examine the ^{13}C NMR spectra obtained in the test for positional distribution (β(2)-acyl) of fatty acids (see Tests). The spectra contain peaks between 172 ppm and 173 ppm with shifts similar to those in the spectrum shown in Figure 2398.-1.

The positional distribution (β(2)-acyl) for cervonic (docosahexaenoic) acid (C22:6 n-3; DHA), timnodonic (eicosapentaenoic) acid (C20:5 n-3; EPA) and moroctic acid (C18:4 n-3) complies with the limits.

B. Linoleic acid (see Tests).

TESTS

Acid value *(2.5.1)*
Maximum 2.0.

Anisidine value *(2.5.36)*
Maximum 10.0.

Peroxide value *(2.5.5, Method B)*
Maximum 5.0.

Unsaponifiable matter *(2.5.7)*
Maximum 1.5 per cent, determined on 2.0 g, and extracting with 3 quantities, each of 50 mL, of *peroxide-free ether R.*

Stearin
Heat at least 10 mL to 60-90 °C then allow to cool for 3 h in a bath of iced water or a thermostatically controlled bath at 0 ± 0.5 °C. If necessary, to eliminate insoluble matter, filter the sample after heating. The sample remains clear.

Positional distribution (β(2)-acyl) of fatty acids
Nuclear magnetic resonance spectrometry *(2.2.33)*.

Test solution Dissolve 190-210 mg of the substance to be examined in 500 µL of *deuterated chloroform R.* Prepare at least 3 samples and examine within 3 days.

Apparatus High-resolution FT-NMR spectrometer operating at minimum 300 MHz.

Acquisition of ^{13}C NMR spectra The following parameters may be used:
— *sweep width*: 200 ppm (−5 ppm to 195 ppm);
— *irradiation frequency offset*: 95 ppm;
— *time domain*: 64 K;
— *pulse delay*: 2 s;
— *pulse program*: zgig 30 (inverse gated, 30° excitation pulse);
— *dummy scans*: 4;
— *number of scans*: 4096.

Processing and plotting The following parameters may be used:
— *size*: 64 K (zero-filling);
— *window multiplication*: exponential;
— *Lorentzian broadening factor*: 0.2 Hz.

Use the $CDCl_3$ signal for shift referencing. The shift of the central peak of the 1:1:1 triplet is set to 77.16 ppm.

Plot the spectral region δ 171.5-173.5 ppm. Compare the spectrum with the spectrum shown in Figure 2398.-1. The shift values lie within the ranges given in Table 2398.-1.

Table 2398.-1. – *Shift values*

Signal	Shift range (ppm)
β DHA	172.05 - 172.09
α DHA	172.43 - 172.47
β EPA	172.52 - 172.56
α EPA	172.90 - 172.94
β C18:4	172.56 - 172.60
α C18:4	172.95 - 172.99

System suitability:
— *signal-to-noise ratio*: minimum 5 for the smallest relevant peak corresponding to α C18:4 signal (in the range δ 172.95-172.99 ppm);
— *peak width at half-height*: maximum 0.02 ppm for the central $CDCl_3$ signal (at δ 77.16 ppm).

Calculation of positional distribution (β(2)-acyl) Use the following expression:

$$\frac{100 \times \beta}{\alpha + \beta}$$

α = peak area of the corresponding α-carbonyl peak;
β = peak area of β-carbonyl peak from C22:6 n-3, C20:5 n-3 or C18:4 n-3, respectively.

Limits:
— *positional distribution (β(2)-acyl)*:
  — *cervonic (docosahexaenoic) acid (C22:6 n-3; DHA)*: 71 per cent to 81 per cent;
  — *timnodonic (eicosapentaenoic) acid (C20:5 n-3 EPA)*: 32 per cent to 40 per cent;
  — *moroctic acid (C18:4 n-3)*: 28 per cent to 38 per cent.

Composition of fatty acids *(2.4.29)*
For identification of the peaks, see the chromatogram shown in Figure 2398.-2.

The 24 largest peaks of the methyl esters account for more than 90 per cent of the total area (these correspond to, in common elution order: 14:0, 15:0, 16:0, 16:1 n-7, 16:4 n-1, 18:0, 18:1 n-9, 18:1 n-7, 18:2 n-6, 18:3 n-3, 18:4 n-3, 20:1 n-11, 20:1 n-9, 20:1 n-7, 20:2 n-6, 20:4 n-6, 20:3 n-3, 20:4 n-3, 20:5 n-3, 22:1 n-11, 22:1 n-9, 21:5 n-3, 22:5 n-3, 22:6 n-3).

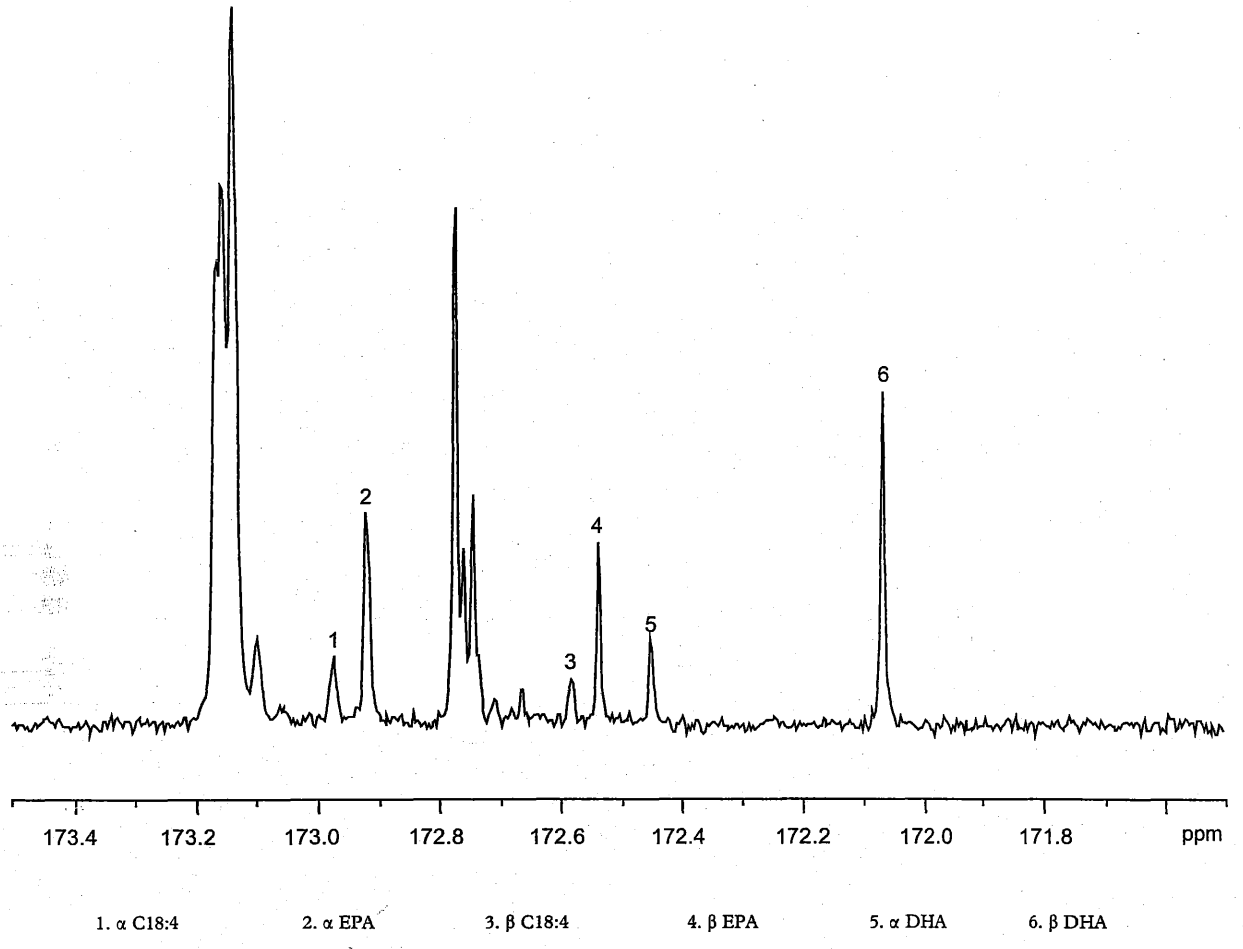


Figure 2398.-1. – *^{13}C NMR spectrum: carbonyl region of farmed cod-liver oil*

Linoleic acid (*2.4.29*)
3.0 per cent to 11.0 per cent.

ASSAY

EPA and DHA (*2.4.29*)
See the chromatogram shown in Figure 2398.-2.

Vitamin A

Carry out the test as rapidly as possible, avoiding exposure to actinic light and air, oxidising agents, oxidation catalysts (for example, copper and iron) and acids.

Use method A. If method A is found not to be valid, use method B.

METHOD A

Ultraviolet absorption spectrophotometry (*2.2.25*).

Test solution To 1.00 g in a round-bottomed flask, add 3 mL of a freshly prepared 50 per cent *m/m* solution of *potassium hydroxide R* and 30 mL of *anhydrous ethanol R*. Boil under reflux in a current of *nitrogen R* for 30 min. Cool rapidly and add 30 mL of *water R*. Extract with 50 mL of *ether R*. Repeat the extraction 3 times and discard the lower layer after complete separation. Wash the combined upper layers with 4 quantities, each of 50 mL, of *water R*, and evaporate to dryness under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C or in a rotary evaporator at a temperature not exceeding 30 °C under reduced pressure (water ejector). Dissolve the residue in sufficient *2-propanol R1* to give an expected concentration of vitamin A equivalent to 10-15 IU/mL.

Measure the absorbances of the solution at 300 nm, 310 nm, 325 nm and 334 nm and at the wavelength of maximum absorption with a suitable spectrophotometer in specially matched 1 cm cells, using *2-propanol R1* as the compensation liquid.

Calculate the content of vitamin A, as all-*trans*-retinol, in International Units per gram, using the following expression:

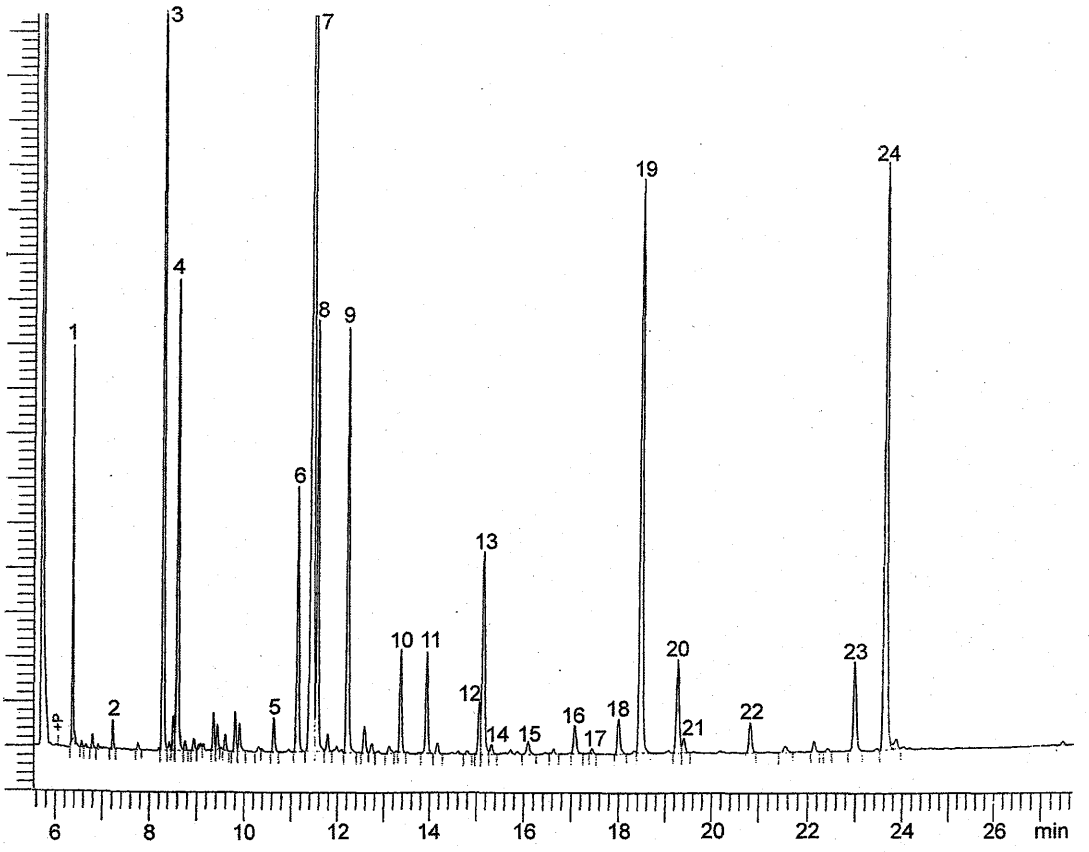
$$A_{325} \times \frac{1821}{100m} \times V$$

A_{325}	=	absorbance at 325 nm;
m	=	mass of the substance to be examined, in grams;
V	=	total volume of solution containing 10-15 IU of vitamin A per millilitre;
1821	=	conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

The above expression can be used only if A_{325} has a value not greater than $A_{325,corr}$ /0.970, where $A_{325,corr}$ is the corrected absorbance at 325 nm and is given by the following equation:

$$A_{325,\,corr} = 6.815A_{325} - 2.555A_{310} - 4.260A_{334}$$

A designates the absorbance at the wavelength indicated by the subscript.



1. C14:0	5. C16:4 n-1	9. C18:2 n-6	13. C20:1 n-9	17. C20:3 n-3	21. C22:1 n-9
2. C15:0	6. C18:0	10 C18:3 n-3	14. C20:1 n-7	18. C20:4 n-3	22. C21:5 n-3
3. C16:0	7. C18:1 n-9	11. C18:4 n-3	15. C20:2 n-6	19. C20:5 n-3	23. C22:5 n-3
4. C16:1 n-7	8. C18:1 n-7	12. C20:1 n-11	16. C20:4 n-6	20. C22:1 n-11	24. C22:6 n-3

Figure 2398.-2. – *Chromatogram for the test for composition of fatty acids of farmed cod-liver oil*

If A_{325} has a value greater than $A_{325,corr}$ /0.970, calculate the content of vitamin A using the following expression:

$$A_{325,\,corr} \times \frac{1821}{100m} \times V$$

The assay is not valid unless:
— the wavelength of maximum absorption lies between 323 nm and 327 nm;
— the absorbance at 300 nm relative to that at 325 nm is at most 0.73.

METHOD B

Liquid chromatography (*2.2.29*).

Test solution Prepare duplicates. To 2.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R*, 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 15 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R* and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively, evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in *2-propanol R*, transfer to a 25 mL volumetric flask and dilute to 25 mL with *2-propanol R*. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Reference solution (a) Prepare a solution of *retinol acetate CRS* in *2-propanol R1* so that 1 mL contains about 1000 IU of all-*trans*-retinol.

The exact concentration of reference solution (a) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (a) with *2-propanol R1* to a presumed concentration of 10-15 IU/mL and measure the absorbance at 326 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of vitamin A in International Units per millilitre of reference solution (a) using the following expression, taking into account the assigned content of *retinol acetate CRS*:

$$A_{326} \times \frac{1900 \times V_2}{100 \times V_1}$$

A_{326} = absorbance at 326 nm;
V_1 = volume of reference solution (a) used;
V_2 = volume of the diluted solution;
1900 = conversion factor for the specific absorbance of *retinol acetate CRS*, in International Units.

Reference solution (b) Proceed as described for the test solution but using 2.00 mL of reference solution (a) in place of the substance to be examined.

The exact concentration of reference solution (b) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (b) with *2-propanol R1* to a presumed all-*trans*-retinol concentration of 10-15 IU/mL and measure the absorbance at 325 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of all-*trans*-retinol in International Units per millilitre of reference solution (b), using the following expression:

$$A_{325} \times \frac{1821 \times V_3}{100 \times V_4}$$

A_{325} = absorbance at 325 nm;
V_3 = volume of the diluted solution;
V_4 = volume of reference solution (b) used;
1821 = conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase *water R, methanol R* (3:97 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL; inject in triplicate the test solution and reference solution (b).

Retention time All-*trans*-retinol = 5 ± 1 min.

System suitability:
— the chromatogram obtained with the test solution shows a peak corresponding to the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
— the results obtained with the duplicate test solutions do not differ by more than 5 per cent;
— the recovery of all-*trans*-retinol in reference solution (b) as assessed by direct absorption spectrophotometry is greater than 95 per cent.

Calculate the content of vitamin A using the following expression:

$$A_1 \times \frac{C \times V}{A_2} \times \frac{1}{m}$$

A_1 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with the test solution;
A_2 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
C = concentration of *retinol acetate CRS* in reference solution (a) as assessed prior to the saponification, in International Units per millilitre (= 1000 IU/mL);
V = volume of reference solution (a) treated (2.00 mL);
m = mass of the substance to be examined in the test solution (2.00 g).

Vitamin D_3

Liquid chromatography (*2.2.29*). *Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.*

Internal standard solution Dissolve 0.50 mg of *ergocalciferol CRS* in 100 mL of *anhydrous ethanol R*.

Test solution (a) To 4.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R*, 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 30 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool the solution to room temperature. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R*, and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively, evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in 1.5 mL of the mobile phase described under Purification. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Test solution (b) Prepare duplicates. To 4.00 g add 2.0 mL of the internal standard solution and proceed as described for test solution (a).

Reference solution (a) Dissolve 0.50 mg of *cholecalciferol CRS* in 100.0 mL of *anhydrous ethanol R*.

Reference solution (b) In a round-bottomed flask, add 2.0 mL of reference solution (a) and 2.0 mL of the internal standard solution and proceed as described for test solution (a).

PURIFICATION

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *nitrile silica gel for chromatography R* (10 µm).

Mobile phase *isoamyl alcohol R, hexane R* (1.6:98.4 *V/V*).

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 350 µL of reference solution (b) and test solutions (a) and (b). Collect each eluate from 2 min before until 2 min after the retention time of cholecalciferol, in a ground-glass-stoppered tube containing 1 mL of a 1 g/L solution of *butylhydroxytoluene R* in *hexane R*. Evaporate separately to dryness at a temperature not exceeding 30 °C under a gentle current of *nitrogen R*. Dissolve each residue in 1.5 mL of *acetonitrile R*.

DETERMINATION

Column:
— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *phosphoric acid R*, 96 per cent *V/V* solution of *acetonitrile R* (0.2:99.8 *V/V*).
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 265 nm.
Injection 2 quantities not exceeding 200 µL of each of the 3 solutions obtained under Purification.
System suitability:
— *resolution*: minimum 1.4 between the peaks due to ergocalciferol and cholecalciferol in the chromatogram obtained with reference solution (b);
— the results obtained with the test solution (b) duplicates do not differ by more than 5 per cent.

Calculate the content of vitamin D_3 in International Units per gram using the following expression, taking into account the assigned content of *cholecalciferol CRS*:

$$\frac{A_2}{A_6} \times \frac{A_3}{A_4 - \left[\frac{A_5}{A_1}\right] \times A_2} \times \frac{m_2}{m_1} \times \frac{V_2}{V_1} \times 40$$

m_1 = mass of the sample in test solution (b), in grams;
m_2 = total mass of *cholecalciferol CRS* used for the preparation of reference solution (a), in micrograms (500 µg);
A_1 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (a);
A_2 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (b);
A_3 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with reference solution (b);
A_4 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with test solution (b);
A_5 = area (or height) of a possible peak in the chromatogram obtained with test solution (a) with the same retention time as the peak co-eluting with ergocalciferol in test solution (b);
A_6 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (b);
V_1 = total volume of reference solution (a) (100 mL);
V_2 = volume of reference solution (a) used for preparing reference solution (b) (2.0 mL).

STORAGE

In an airtight and well-filled container, protected from light. If no antioxidant is added, store under an inert gas.
Once the container has been opened, its contents are used as soon as possible and any part of the contents not used at once is protected by an atmosphere of inert gas.

LABELLING

The label states:
— the concentration of EPA and DHA as a sum;
— the number of International Units of vitamin A per gram;
— the number of International Units of vitamin D_3 per gram.

Ph Eur

Cod-liver Oil (Type A)

(Ph. Eur. monograph 1192)

Action and use

Source of vitamins A and D.
Each IU of vitamin D_3 is equivalent to 0.025 µg of colecalciferol.

Ph Eur

DEFINITION

Purified fatty oil obtained from the fresh livers of wild cod, *Gadus morhua* L. and other species of *Gadidae*, solid substances being removed by cooling and filtering. A suitable antioxidant may be added.

Content

— *vitamin A*: 600 IU (180 µg) to 2500 IU (750 µg) per gram;
— *vitamin D_3*: 60 IU (1.5 µg) to 250 IU (6.25 µg) per gram.

PRODUCTION

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance

Clear, yellowish liquid.

Solubility

Practically insoluble in water, miscible with light petroleum, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, C.
Second identification: C, D.

A. In the assay for vitamin A using method A, the test solution shows an absorption maximum (*2.2.25*) at 325 ± 2 nm. In the assay for vitamin A using method B, the chromatogram obtained with the test solution shows a peak corresponding to the peak due to all-*trans*-retinol in the chromatogram obtained with the reference solution.

B. In the assay for vitamin D_3, the chromatogram obtained with test solution (a) shows a peak corresponding to the peak due to cholecalciferol in the chromatogram obtained with reference solution (b).

C. Composition of fatty acids (see Tests).

D. To 0.1 g add 0.5 mL of *methylene chloride R* and 1 mL of *antimony trichloride solution R*. Mix. A deep blue colour develops in about 10 s.

TESTS

Appearance

The substance to be examined is not more intensely coloured than a reference solution prepared as follows: to 3.0 mL of red primary solution add 25.0 mL of yellow primary solution and dilute to 50.0 mL with a 10 g/L solution of *hydrochloric acid R* (*2.2.2, Method II*).

Relative density (*2.2.5*)
0.917 to 0.930.

Refractive index (*2.2.6*)
1.477 to 1.484.

Acid value (*2.5.1*)
Maximum 2.0.

Anisidine value (*2.5.36*)
Maximum 30.0.

Iodine value (*2.5.4, Method B*)
150 to 180.
Use *starch solution R2*.

Peroxide value (*2.5.5, Method B*)
Maximum 10.0.

Unsaponifiable matter (*2.5.7*)
Maximum 1.5 per cent, determined on 2.0 g, and extracting with 3 quantities, each of 50 mL, of *peroxide-free ether R*.

Stearin

Heat at least 10 mL to 60-90 °C then allow to cool for 3 h in a bath of iced water or a thermostatically controlled bath at 0 ± 0.5 °C. If necessary, to eliminate insoluble matter, filter the sample after heating. The sample remains clear.

Composition of fatty acids
Gas chromatography (*2.2.28*).

Trivial name of fatty acid	Nomenclature	Lower limit area (per cent)	Upper limit area (per cent)
Saturated fatty acids:			
Myristic acid	14:0	2.0	6.0
Palmitic acid	16:0	7.0	14.0
Stearic acid	18:0	1.0	4.0
Mono-unsaturated fatty acids:			
Palmitoleic acid	16:1 n-7	4.5	11.5
cis-Vaccenic acid	18:1 n-7	2.0	7.0
Oleic acid	18:1 n-9	12.0	21.0
Gadoleic acid	20:1 n-11	1.0	5.5
Gondoic acid	20:1 n-9	5.0	17.0
Erucic acid	22:1 n-9	0	1.5
Cetoleic acid (22:1 n-11)	22:1 n-11+13	5.0	12.0
Poly-unsaturated fatty acids:			
Linoleic acid	18:2 n-6	0.5	3.0
α-Linolenic acid	18:3 n-3	0	2.0
Moroctic acid	18:4 n-3	0.5	4.5
Timnodonic (eicosapentaenoic) acid (EPA)	20:5 n-3	7.0	16.0
Cervonic (docosahexaenoic) acid (DHA)	22:6 n-3	6.0	18.0

Test solution Introduce about 0.45 g of the substance to be examined into a 10 mL volumetric flask, dissolve in *hexane R* containing 50 mg of *butylhydroxytoluene R* per litre and dilute to 10.0 mL with the same solvent. Transfer 2.0 mL of this solution into a quartz tube and evaporate the solvent with a gentle current of *nitrogen R*. Add 1.5 mL of a 20 g/L solution of *sodium hydroxide R* in *methanol R*, cover with *nitrogen R*, cap tightly with a polytetrafluoroethylene-lined cap, mix and heat on a water-bath for 7 min. Cool, add 2 mL of *boron trichloride-methanol solution R*, cover with *nitrogen R*, cap tightly, mix and heat on a water-bath for 30 min. Cool to 40-50 °C, add 1 mL of *trimethylpentane R*, cap and vortex or shake vigorously for at least 30 s. Immediately add 5 mL of *saturated sodium chloride solution R*, cover with *nitrogen R*, cap and vortex or shake vigorously for at least 15 s. Allow the upper layer to become clear and transfer it to a separate tube. Shake the methanol layer once more with 1 mL of *trimethylpentane R* and combine the trimethylpentane extracts. Wash the combined extracts with 2 quantities, each of 1 mL, of *water R* and dry over *anhydrous sodium sulfate R*. Prepare 2 solutions for each sample.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *hydrogen for chromatography R* or *helium for chromatography R*, where oxygen scrubber is applied.

Split ratio 1:200.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 55	170 → 225
	55 - 75	225
Injection port		250
Detector		280

Detection Flame ionisation.

Injection 1 µL, twice.

System suitability:
— the 15 fatty acids to be tested are satisfactorily identified from the chromatogram shown in Figure 1192.-1;
— injection of a mixture of equal amounts of *methyl palmitate R*, *methyl stearate R*, *methyl arachidate R* and *methyl behenate R* gives area percentages of 24.4, 24.8, 25.2 and 25.6 (± 0.5 per cent), respectively;
— *resolution*: minimum 1.3 between the peaks due to methyl oleate and methyl *cis*-vaccenate; the resolution between the pair due to methyl gadoleate and methyl gondoate is sufficient for purposes of identification and area measurement.

Calculate the area per cent for each fatty acid methyl ester using the following expression:

$$\frac{A_x}{A_t} \times 100$$

A_x = peak area of fatty acid x;
A_t = sum of the peak areas (up to C22:6 n-3).

The calculation is not valid unless:
— the total area is based only on peaks due solely to fatty acid methyl esters;
— the number of fatty acid methyl ester peaks exceeding 0.05 per cent of the total area is at least 24;
— the 24 largest peaks of the methyl esters account for more than 90 per cent of the total area (these correspond to, in common elution order: 14:0, 15:0, 16:0, 16:1 n-7, 16:4 n-1, 18:0, 18:1 n-9, 18:1 n-7, 18:2 n-6, 18:3 n-3, 18:4 n-3, 20:1 n-11, 20:1 n-9, 20:1 n-7, 20:2 n-6, 20:4 n-6, 20:3 n-3, 20:4 n-3, 20:5 n-3, 22:1 n-11, 22:1 n-9, 21:5 n-3, 22:5 n-3, 22:6 n-3).

ASSAY

Vitamin A

Carry out the test as rapidly as possible, avoiding exposure to actinic light and air, oxidising agents, oxidation catalysts (for example, copper and iron) and acids.

Use method A. If method A is found not to be valid, use method B.

METHOD A

Ultraviolet absorption spectrophotometry (*2.2.25*).

Test solution To 1.00 g in a round-bottomed flask, add 3 mL of a freshly prepared 50 per cent *m/m* solution of *potassium hydroxide R* and 30 mL of *anhydrous ethanol R*. Boil under reflux in a current of *nitrogen R* for 30 min. Cool rapidly and add 30 mL of *water R*. Extract with 50 mL of *ether R*. Repeat the extraction 3 times and discard the lower layer after complete separation. Wash the combined upper layers with 4 quantities, each of 50 mL, of *water R*, and evaporate to dryness under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C or in a rotary evaporator at a temperature not exceeding 30 °C under reduced pressure (water ejector). Dissolve the residue in sufficient *2-propanol R1* to give an expected concentration of vitamin A equivalent to 10-15 IU/mL.

Measure the absorbances of the solution at 300 nm, 310 nm, 325 nm and 334 nm and at the wavelength of maximum absorption with a suitable spectrophotometer in specially matched 1 cm cells, using *2-propanol R1* as the compensation liquid.

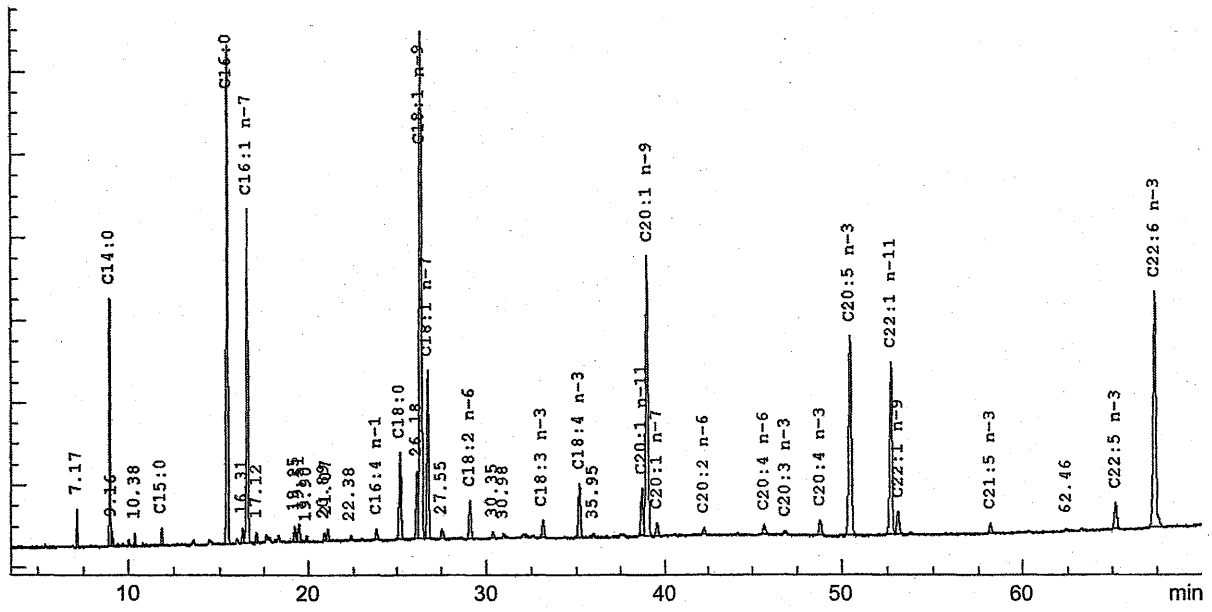


Figure 1192.-1. – *Chromatogram for the test for composition of fatty acids of cod-liver oil (type A)*

Calculate the content of vitamin A, as all-*trans*-retinol, in International Units per gram, using the following expression:

$$A_{325} \times \frac{1821}{100m} \times V$$

A_{325} = absorbance at 325 nm;
m = mass of the substance to be examined, in grams;
V = total volume of solution containing 10-15 IU of vitamin A per millilitre;
1821 = conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

The above expression can be used only if A_{325} has a value not greater than $A_{325,corr}/0.970$, where $A_{325,corr}$ is the corrected absorbance at 325 nm and is given by the following equation:

$$A_{325,corr} = 6.815A_{325} - 2.555A_{310} - 4.260A_{334}$$

A designates the absorbance at the wavelength indicated by the subscript.

If A_{325} has a value greater than $A_{325,corr}/0.970$, calculate the content of vitamin A using the following expression:

$$A_{325,corr} \times \frac{1821}{100m} \times V$$

The assay is not valid unless:
— the wavelength of the maximum absorption lies between 323 nm and 327 nm;
— the absorbance at 300 nm relative to that at 325 nm is at most 0.73.

METHOD B

Liquid chromatography (*2.2.29*).

Test solution Prepare duplicates. To 2.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R*, 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 15 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R* and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively, evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in *2-propanol R*, transfer to a 25 mL volumetric flask and dilute to 25 mL with *2-propanol R*. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Reference solution (a) Prepare a solution of *retinol acetate CRS* in *2-propanol R1* so that 1 mL contains about 1000 IU of all-*trans*-retinol.

The exact concentration of reference solution (a) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (a) with *2-propanol R1* to a presumed concentration of 10-15 IU/mL and measure the absorbance at 326 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of vitamin A in International Units per millilitre of reference solution (a) using the following expression, taking into account the assigned content of *retinol acetate CRS*:

$$A_{326} \times \frac{1900 \times V_2}{100 \times V_1}$$

A_{326} = absorbance at 326 nm;
V_1 = volume of reference solution (a) used;
V_2 = volume of the diluted solution;
1900 = conversion factor for the specific absorbance of *retinol acetate CRS*, in International Units.

Reference solution (b) Proceed as described for the test solution but using 2.00 mL of reference solution (a) in place of the substance to be examined.

The exact concentration of reference solution (b) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (b) with *2-propanol R1* to a presumed all-*trans*-retinol concentration of 10-15 IU/mL and measure the absorbance at 325 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of all-*trans*-retinol in International Units per millilitre of reference solution (b), using the following expression:

$$A_{325} \times \frac{1821 \times V_3}{100 \times V_4}$$

A_{325} = absorbance at 325 nm;
V_3 = volume of the diluted solution;
V_4 = volume of reference solution (b) used;
1821 = conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase *water R, methanol R* (3:97 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL; inject in triplicate the test solution and reference solution (b).

Retention time All-*trans*-retinol = 5 ± 1 min.

System suitability:
— the chromatogram obtained with the test solution shows a peak corresponding to the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
— the results obtained with the duplicate test solutions do not differ by more than 5 per cent;
— the recovery of all-*trans*-retinol in reference solution (b) as assessed by direct absorption spectrophotometry is greater than 95 per cent.

Calculate the content of vitamin A using the following expression:

$$A_1 \times \frac{C \times V}{A_2} \times \frac{1}{m}$$

A_1 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with the test solution;
A_2 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
C = concentration of *retinol acetate CRS* in reference solution (a) as assessed prior to the saponification, in International Units per millilitre (= 1000 IU/mL);
V = volume of reference solution (a) treated (2.00 mL);
m = mass of the substance to be examined in the test solution (2.00 g).

Vitamin D_3

Liquid chromatography (*2.2.29*). *Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.*

Internal standard solution Dissolve 0.50 mg of *ergocalciferol CRS* in 100 mL of *anhydrous ethanol R*.

Test solution (a) To 4.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R*, 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 30 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool the solution to room temperature. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R*, and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively, evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in 1.5 mL of the mobile phase described under Purification. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Test solution (b) Prepare duplicates. To 4.00 g add 2.0 mL of the internal standard solution and proceed as described for test solution (a).

Reference solution (a) Dissolve 0.50 mg of *cholecalciferol CRS* in 100.0 mL of *anhydrous ethanol R*.

Reference solution (b) Into a round-bottomed flask, add 2.0 mL of reference solution (a) and 2.0 mL of the internal standard solution and proceed as described for test solution (a).

PURIFICATION

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *nitrile silica gel for chromatography R* (10 µm).

Mobile phase *isoamyl alcohol R, hexane R* (1.6:98.4 *V/V*).

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 350 µL of reference solution (b) and test solutions (a) and (b). Collect each eluate from 2 min before until 2 min after the retention time of cholecalciferol, in a ground-glass-stoppered tube containing 1 mL of a 1 g/L solution of *butylhydroxytoluene R* in *hexane R*. Evaporate separately to dryness at a temperature not exceeding 30 °C under a gentle current of *nitrogen R*. Dissolve each residue in 1.5 mL of *acetonitrile R*.

DETERMINATION

Column:
— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *phosphoric acid R*, 96 per cent *V/V* solution of *acetonitrile R* (0.2:99.8 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 2 quantities not exceeding 200 µL of each of the 3 solutions obtained under Purification.

System suitability:
— *resolution*: minimum 1.4 between the peaks due to ergocalciferol and cholecalciferol in the chromatogram obtained with reference solution (b);

— the results obtained with test solution (b) duplicates do not differ by more than 5 per cent.

Calculate the content of vitamin D_3 in International Units per gram using the following expression, taking into account the assigned content of *cholecalciferol CRS*:

$$\frac{A_2}{A_6} \times \frac{A_3}{A_4 - \left[\frac{A_5}{A_1}\right] \times A_2} \times \frac{m_2}{m_1} \times \frac{V_2}{V_1} \times 40$$

m_1 = mass of the sample in test solution (b), in grams;
m_2 = total mass of *cholecalciferol CRS* used for the preparation of reference solution (a), in micrograms (500 µg);
A_1 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (a);
A_2 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (b);
A_3 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with reference solution (b);
A_4 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with test solution (b);
A_5 = area (or height) of a possible peak in the chromatogram obtained with test solution (a) with the same retention time as the peak co-eluting with ergocalciferol in test solution (b);
A_6 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (b);
V_1 = total volume of reference solution (a) (100 mL);
V_2 = volume of reference solution (a) used for preparing reference solution (b) (2.0 mL).

STORAGE

In an airtight and well-filled container, protected from light. If no antioxidant is added, store under an inert gas.

Once the container has been opened, its contents are used as soon as possible and any part of the contents not used at once is protected by an atmosphere of inert gas.

LABELLING

The label states:
— the number of International Units of vitamin A per gram;
— the number of International Units of vitamin D_3 per gram.

Ph Eur

Cod-liver Oil (Type B)

(Ph. Eur. monograph 1193)

Action and use

Source of vitamins A and D.

Each IU of vitamin D_3 is equivalent to 0.025 µg of colecalciferol.

Ph Eur

DEFINITION

Purified fatty oil obtained from the fresh livers of wild cod, *Gadus morhua* L. and other species of *Gadidae*, solid substances being removed by cooling and filtering. A suitable antioxidant may be added.

Content

— *vitamin A*: 600 IU (180 µg) to 2500 IU (750 µg) per gram;
— *vitamin D_3*: 60 IU (1.5 µg) to 250 IU (6.25 µg) per gram.

PRODUCTION

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance

Clear, yellowish liquid.

Solubility

Practically insoluble in water, miscible with light petroleum, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, C.

Second identification: C, D.

A. In the assay for vitamin A using method A, the test solution shows an absorption maximum (*2.2.25*) at 325 ± 2 nm. In the assay for vitamin A using method B, the chromatogram obtained with the test solution shows a peak corresponding to the peak due to all-*trans*-retinol in the chromatogram obtained with the reference solution.

B. In the assay for vitamin D_3, the chromatogram obtained with test solution (a) shows a peak corresponding to the peak due to cholecalciferol in the chromatogram obtained with reference solution (b).

C. Composition of fatty acids (see Tests).

D. To 0.1 g add 0.5 mL of *methylene chloride R* and 1 mL of *antimony trichloride solution R*. Mix. A deep blue colour develops in about 10 s.

TESTS

Appearance

The substance to be examined is not more intensely coloured than a reference solution prepared as follows: to 3.0 mL of red primary solution add 25.0 mL of yellow primary solution and dilute to 50.0 mL with a 10 g/L solution of *hydrochloric acid R* (*2.2.2, Method II*).

Relative density (*2.2.5*)
0.917 to 0.930.

Refractive index (*2.2.6*)
1.477 to 1.484.

Acid value (*2.5.1*)
Maximum 2.0.

Iodine value (*2.5.4, Method B*)
150 to 180.

Use *starch solution R2*.

Peroxide value (*2.5.5, Method B*)
Maximum 10.0.

Unsaponifiable matter (*2.5.7*)
Maximum 1.5 per cent, determined on 2.0 g and extracting with 3 quantities, each of 50 mL, of *peroxide-free ether R*.

Stearin

Heat at least 10 mL to 60-90 °C then allow to cool for 3 h in a bath of iced water or a thermostatically controlled bath at 0 ± 0.5 °C. If necessary, to eliminate insoluble matter, filter the sample after heating. The sample remains clear.

Composition of fatty acids

Gas chromatography (*2.2.28*).

Trivial name of fatty acid	Nomenclature	Lower limit area (per cent)	Upper limit area (per cent)
Saturated fatty acids:			
Myristic acid	14:0	2.0	6.0
Palmitic acid	16:0	7.0	14.0
Stearic acid	18:0	1.0	4.0
Mono-unsaturated fatty acids:			
Palmitoleic acid	16:1 n-7	4.5	11.5
cis-Vaccenic acid	18:1 n-7	2.0	7.0
Oleic acid	18:1 n-9	12.0	21.0
Gadoleic acid	20:1 n-11	1.0	5.5
Gondoic acid	20:1 n-9	5.0	17.0
Erucic acid	22:1 n-9	0	1.5
Cetoleic acid (22:1 n-11)	22:1 n-11+13	5.0	12.0
Poly-unsaturated fatty acids:			
Linoleic acid	18:2 n-6	0.5	3.0
α-Linolenic acid	18:3 n-3	0	2.0
Moroctic acid	18:4 n-3	0.5	4.5
Timnodonic (eicosapentaenoic) acid (EPA)	20:5 n-3	7.0	16.0
Cervonic (docosahexaenoic) acid (DHA)	22:6 n-3	6.0	18.0

Test solution Introduce about 0.45 g of the substance to be examined into a 10 mL volumetric flask, dissolve in *hexane R* containing 50 mg of *butylhydroxytoluene R* per litre and dilute to 10.0 mL with the same solvent. Transfer 2.0 mL of the solution into a quartz tube and evaporate the solvent with a gentle current of *nitrogen R*. Add 1.5 mL of a 20 g/L solution of *sodium hydroxide R* in *methanol R*, cover with *nitrogen R*, cap tightly with a polytetrafluoroethylene-lined cap, mix and heat on a water-bath for 7 min. Cool, add 2 mL of *boron trichloride-methanol solution R*, cover with *nitrogen R*, cap tightly, mix and heat on a water-bath for 30 min. Cool to 40-50 °C, add 1 mL of *trimethylpentane R*, cap and vortex or shake vigorously for at least 30 s. Immediately add 5 mL of *saturated sodium chloride solution R*, cover with *nitrogen R*, cap and vortex or shake thoroughly for at least 15 s. Allow the upper layer to become clear and transfer to a separate tube. Shake the methanol layer once more with 1 mL of *trimethylpentane R* and combine the trimethylpentane extracts. Wash the combined extracts with 2 quantities, each of 1 mL, of *water R* and dry over *anhydrous sodium sulfate R*. Prepare 2 solutions for each sample.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.25 mm;
- *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *hydrogen for chromatography R* or *helium for chromatography R*, where oxygen scrubber is applied.

Split ratio 1:200.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 55	170 → 225
	55 - 75	225
Injection port		250
Detector		280

Detection Flame ionisation.

Injection 1 µL, twice.

System suitability:
- the 15 fatty acids to be tested are satisfactorily identified from the chromatogram shown in Figure 1193.-1;
- injection of a mixture of equal amounts of *methyl palmitate R*, *methyl stearate R*, *methyl arachidate R*, and *methyl behenate R* give area percentages of 24.4, 24.8, 25.2 and 25.6 (± 0.5 per cent), respectively;
- *resolution*: minimum of 1.3 between the peaks due to methyl oleate and methyl *cis*-vaccenate; the resolution between the pair due to methyl gadoleate and methyl gondoate is sufficient for purposes of identification and area measurement.

Calculate the area per cent for each fatty acid methyl ester using the following expression:

$$\frac{A_x}{A_t} \times 100$$

A_x = peak area of fatty acid *x*;
A_t = sum of the peak areas (up to C22:6 n-3).

The calculation is not valid unless:
- the total area is based only on peaks due to solely fatty acids methyl esters;
- the number of fatty acid methyl ester peaks exceeding 0.05 per cent of the total area is at least 24;
- the 24 largest peaks of the methyl esters account for more than 90 per cent of the total area (these correspond to, in common elution order: 14:0, 15:0, 16:0, 16:1 n-7, 16:4 n-1, 18:0, 18:1 n-9, 18:1 n-7, 18:2 n-6, 18:3 n-3, 18:4 n-3, 20:1 n-11, 20:1 n-9, 20:1 n-7, 20:2 n-6, 20:4 n-6, 20:3 n-3, 20:4 n-3, 20:5 n-3, 22:1 n-11, 22:1 n-9, 21:5 n-3, 22:5 n-3, 22:6 n-3).

ASSAY

Vitamin A

Carry out the test as rapidly as possible, avoiding exposure to actinic light and air, oxidising agents, oxidation catalysts (for example, copper and iron) and acids.

Use method A. If method A is found not to be valid, use method B.

METHOD A

Ultraviolet absorption spectrophotometry (*2.2.25*).

Test solution To 1.00 g in a round-bottomed flask, add 3 mL of a freshly prepared 50 per cent *m/m* solution of *potassium hydroxide R* and 30 mL of *anhydrous ethanol R*. Boil under reflux in a current of *nitrogen R* for 30 min. Cool rapidly and add 30 mL of *water R*. Extract with 50 mL of *ether R*. Repeat the extraction 3 times and discard the lower layer after complete separation. Wash the combined upper layers with 4 quantities, each of 50 mL, of *water R* and evaporate to dryness under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C or in a rotary evaporator at a temperature not exceeding 30 °C under reduced pressure (water ejector). Dissolve the residue in sufficient *2-propanol R1* to give an expected concentration of vitamin A equivalent to 10-15 IU/mL.

Measure the absorbances of the solution at 300 nm, 310 nm, 325 nm and 334 nm and at the wavelength of maximum absorption with a suitable spectrophotometer in specially matched 1 cm cells, using *2-propanol R1* as the compensation liquid.

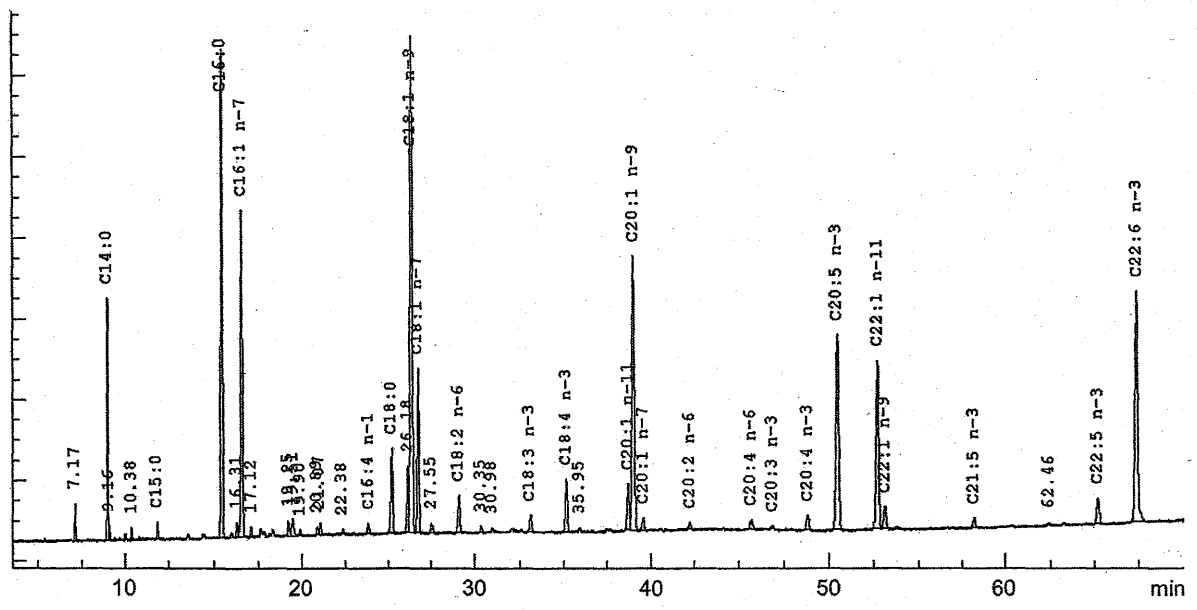


Figure 1193.-1. – *Chromatogram for the test for composition of fatty acids of cod-liver oil (type B)*

Calculate the content of vitamin A, as all-*trans*-retinol, in International Units per gram using the following expression:

$$A_{325} \times \frac{1821}{100m} \times V$$

A_{325} = absorbance at 325 nm;
m = mass of the substance to be examined, in grams;
V = total volume of solution containing 10-15 IU of vitamin A per millilitre;
1821 = conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

The above expression can be used only if A_{325} has a value not greater than $A_{325,corr}/0.970$ where $A_{325,corr}$ is the corrected absorbance at 325 nm and is given by the equation:

$$A_{325,corr} = 6.815A_{325} - 2.555A_{310} - 4.260A_{334}$$

A designates the absorbance at the wavelength indicated by the subscript.

If A_{325} has a value greater than $A_{325,corr}/0.970$, calculate the content of vitamin A using the following expression:

$$A_{325,corr} \times \frac{1821}{100m} \times V$$

The assay is not valid unless:
— the wavelength of maximum absorption lies between 323 nm and 327 nm;
— the absorbance at 300 nm relative to that at 325 nm is at most 0.73.

METHOD B

Liquid chromatography (*2.2.29*).

Test solution Prepare duplicates. To 2.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R* and 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 15 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R* and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in *2-propanol R*, transfer to a 25 mL volumetric flask and dilute to 25 mL with *2-propanol R*. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Reference solution (a) Prepare a solution of *retinol acetate CRS* in *2-propanol R1* so that 1 mL contains about 1000 IU of all-*trans*-retinol.

The exact concentration of reference solution (a) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (a) with *2-propanol R1* to a presumed concentration of 10-15 IU/mL and measure the absorbance at 326 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of vitamin A in International Units per millilitre of reference solution (a) using the following expression, taking into account the assigned content of *retinol acetate CRS*:

$$A_{326} \times \frac{1900 \times V_2}{100 \times V_1}$$

A_{326} = absorbance at 326 nm;
V_1 = volume of reference solution (a) used;
V_2 = volume of the diluted solution;
1900 = conversion factor for the specific absorbance of *retinol acetate CRS*, in International Units.

Reference solution (b) Proceed as described for the test solution but using 2.00 mL of reference solution (a) in place of the substance to be examined.

The exact concentration of reference solution (b) is assessed by ultraviolet absorption spectrophotometry (*2.2.25*). Dilute reference solution (b) with *2-propanol R1* to a presumed concentration of 10-15 IU/mL of all-*trans*-retinol and measure the absorbance at 325 nm in matched 1 cm cells using *2-propanol R1* as the compensation liquid.

Calculate the content of all-*trans*-retinol in International Units per millilitre of reference solution (b) from the expression:

$$A_{325} \times \frac{1821 \times V_3}{100 \times V_4}$$

A_{325} = absorbance at 325 nm;
V_3 = volume of the diluted solution;
V_4 = volume of reference solution (b) used;
1821 = conversion factor for the specific absorbance of all-*trans*-retinol, in International Units.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase *water R, methanol R* (3:97 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL; inject in triplicate the test solution and reference solution (b).

Retention time All-*trans*-retinol = 5 ± 1 min.

System suitability:
— the chromatogram obtained with the test solution shows a peak corresponding to the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
— the results obtained with the duplicate test solutions do not differ by more than 5 per cent;
— the recovery of all-*trans*-retinol in reference solution (b) as assessed by direct absorption spectrophotometry is greater than 95 per cent.

Calculate the content of vitamin A using the following expression:

$$A_1 \times \frac{C \times V}{A_2} \times \frac{1}{m}$$

A_1 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with the test solution;
A_2 = area of the peak due to all-*trans*-retinol in the chromatogram obtained with reference solution (b);
C = concentration of *retinol acetate CRS* in reference solution (a) as assessed prior to the saponification, in International Units per millilitre (= 1000 IU/mL);
V = volume of reference solution (a) treated (2.00 mL);
m = mass of the substance to be examined in the test solution (2.00 g).

Vitamin D_3

Liquid chromatography (*2.2.29*). *Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.*

Internal standard solution Dissolve 0.50 mg of *ergocalciferol CRS* in 100 mL of *anhydrous ethanol R*.

Test solution (a) To 4.00 g in a round-bottomed flask, add 5 mL of a freshly prepared 100 g/L solution of *ascorbic acid R*, 10 mL of a freshly prepared 800 g/L solution of *potassium hydroxide R* and 100 mL of *anhydrous ethanol R*. Boil under a reflux condenser on a water-bath for 30 min. Add 100 mL of a 10 g/L solution of *sodium chloride R* and cool the solution to room temperature. Transfer the solution to a 500 mL separating funnel, rinsing the round-bottomed flask with about 75 mL of a 10 g/L solution of *sodium chloride R* and then with 150 mL of a mixture of equal volumes of *ether R* and *light petroleum R1*. Shake for 1 min. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 30 g/L solution of *potassium hydroxide R* in a 10 per cent *V/V* solution of *anhydrous ethanol R*, and then with 3 quantities, each of 50 mL, of a 10 g/L solution of *sodium chloride R*. Filter the upper layer through 5 g of *anhydrous sodium sulfate R* on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distil them at a temperature not exceeding 30 °C under reduced pressure (water ejector) and fill with *nitrogen R* when evaporation is completed. Alternatively evaporate the solvent under a gentle current of *nitrogen R* at a temperature not exceeding 30 °C. Dissolve the residue in 1.5 mL of the mobile phase described under Purification. Gentle heating in an ultrasonic bath may be required. *A large fraction of the white residue is cholesterol, constituting approximately 50 per cent m/m of the unsaponifiable matter of cod-liver oil.*

Test solution (b) Prepare duplicates. To 4.00 g add 2.0 mL of the internal standard solution and proceed as described for test solution (a).

Reference solution (a) Dissolve 0.50 mg of *cholecalciferol CRS* in 100.0 mL of *anhydrous ethanol R*.

Reference solution (b) In a round-bottomed flask, add 2.0 mL of reference solution (a) and 2.0 mL of the internal standard solution and proceed as described for test solution (a).

PURIFICATION

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *nitrile silica gel for chromatography R* (10 µm).

Mobile phase *isoamyl alcohol R, hexane R* (1.6:98.4 *V/V*).

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 350 µL of reference solution (b) and test solutions (a) and (b). Collect each eluate from 2 min before until 2 min after the retention time of cholecalciferol, in a ground-glass-stoppered tube containing 1 mL of a 1 g/L solution of *butylhydroxytoluene R* in *hexane R*. Evaporate separately to dryness at a temperature not exceeding 30 °C under a gentle current of *nitrogen R*. Dissolve each residue in 1.5 mL of *acetonitrile R*.

DETERMINATION

Column:
— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *phosphoric acid R*, 96 per cent *V/V* solution of *acetonitrile R* (0.2:99.8 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 2 quantities not exceeding 200 µL of each of the 3 solutions obtained under Purification.

System suitability:
— *resolution*: minimum 1.4 between the peaks due to ergocalciferol and cholecalciferol in the chromatogram obtained with reference solution (b);

— the results obtained with the test solution (b) duplicates do not differ by more than 5 per cent.

Calculate the content of vitamin D_3 in International Units per gram using the following expression, taking into account the assigned content of *cholecalciferol CRS*:

$$\frac{A_2}{A_6} \times \frac{A_3}{A_4 - \left[\frac{A_5}{A_1}\right] \times A_2} \times \frac{m_2}{m_1} \times \frac{V_2}{V_1} \times 40$$

m_1 = mass of the sample in test solution (b), in grams;
m_2 = total mass of *cholecalciferol CRS* used for the preparation of reference solution (a), in micrograms (500 µg);
A_1 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (a);
A_2 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with test solution (b);
A_3 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with reference solution (b);
A_4 = area (or height) of the peak due to ergocalciferol in the chromatogram obtained with test solution (b);
A_5 = area (or height) of a possible peak in the chromatogram obtained with test solution (a) with the same retention time as the peak co-eluting with ergocalciferol in test solution (b);
A_6 = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (b);
V_1 = total volume of reference solution (a) (100 mL);
V_2 = volume of reference solution (a) used for preparing reference solution (b) (2.0 mL).

STORAGE

In an airtight and well-filled container, protected from light. If no antioxidant is added, store under an inert gas.

Once the container has been opened, its contents are used as soon as possible and any part of the contents not used at once is protected by an atmosphere of inert gas.

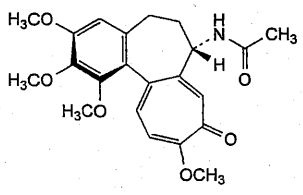
LABELLING

The label states:
— the number of International Units of vitamin A per gram;
— the number of International Units of vitamin D_3 per gram.

Ph Eur

Colchicine

(Ph. Eur. monograph 0758)



$C_{22}H_{25}NO_6$ 399.4 *64-86-8*

Action and use
Used in treatment of gout.

Preparation
Colchicine Tablets

Ph Eur

DEFINITION

N-[(7*S*,12a*M*)-1,2,3,10-tetramethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]acetamide.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Yellowish-white, amorphous or crystalline powder.

Solubility
Very soluble in water, rapidly recrystallising from concentrated solutions as the sesquihydrate, freely soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 5 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 25.0 mL with *ethanol (96 per cent) R*.

Spectral range 230-400 nm.

Absorption maxima 243 nm and 350 nm.

Absorbance ratio A_{243}/A_{350} = 1.7 to 1.9.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *colchicine CRS*.

C. To 0.5 mL of solution S (see Tests) add 0.5 mL of *dilute hydrochloric acid R* and 0.15 mL of *ferric chloride solution R1*. The solution is yellow and becomes dark green on boiling for 30 s. Cool, add 2 mL of *methylene chloride R* and shake. The organic layer is greenish-yellow.

D. Dissolve about 30 mg in 1 mL of *ethanol (96 per cent) R* and add 0.15 mL of *ferric chloride solution R1*. A brownish-red colour develops.

TESTS

Solution S
Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution GY_3 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *bromothymol blue solution R1*. Either the solution does not change colour or it becomes green. Not more than 0.1 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Specific optical rotation (*2.2.7*)
−250 to −235 (anhydrous substance).

Dissolve 50.0 mg in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, water R* (50:50 *V/V*).

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *colchicine for system suitability A CRS* (containing impurities A, E and G) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R1* (5 µm);
— *temperature*: 25 °C.

Mobile phase Mix 450 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* and 530 volumes of *methanol R*. After cooling to room temperature, adjust the volume to 1000 mL with *methanol R*. Adjust the apparent pH to 5.5 with *dilute phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 3 times the retention time of colchicine.

Identification of impurities Use the chromatogram supplied with *colchicine for system suitability A CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, E and G. (Impurity B is the conformational isomer of colchicine, which is formed *in situ* in solution).

Relative retention With reference to colchicine (retention time = about 7 min): impurity E = about 0.6; impurity B = about 0.9; impurity A = about 0.94; impurity G = about 1.4.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 2, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to colchicine; minimum 2, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity A.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity G by 1.6;
— for each impurity, use the concentration of colchicine in reference solution (b).

Limits:
— *impurity A*: maximum 3.0 per cent;
— *impurity G*: maximum 0.25 per cent;
— *impurity E*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 4.0 per cent;
— *reporting threshold*: 0.05 per cent; disregard the peak due to impurity B.

Impurity F
Maximum 0.2 per cent.

Dissolve 50 mg in *water R* and dilute to 5 mL with the same solvent. Add 0.1 mL of *ferric chloride solution R1*. The solution is not more intensely coloured than a mixture of 1 mL of red primary solution, 2 mL of yellow primary solution and 2 mL of blue primary solution (*2.2.2, Method II*).

Ethyl acetate (*2.4.24*)
Maximum 6.0 per cent *m/m*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 0.5 g.

ASSAY

Dissolve 0.250 g with gentle heating in a mixture of 20 mL of *acetic anhydride R* and 40 mL of *toluene R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 39.94 mg of $C_{22}H_{25}NO_6$.

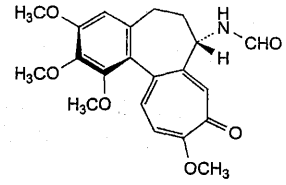
STORAGE

Protected from light.

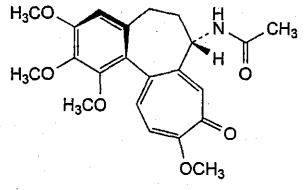
IMPURITIES

Specified impurities *A, E, F, G.*

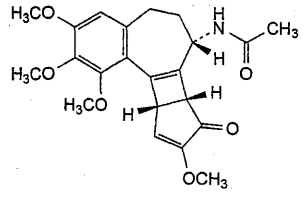
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D.*



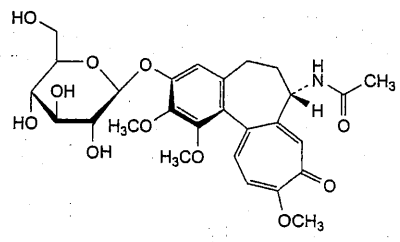
A. *N*-[(7*S*,12a*M*)-1,2,3,10-tetramethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]formamide (*N*-deacetyl-*N*-formylcolchicine),



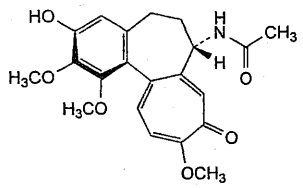
B. *N*-[(7*S*,12a*P*)-1,2,3,10-tetramethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]acetamide (conformational isomer),



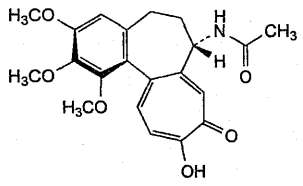
C. *N*-[(7*S*,7b*R*,10a*S*)-1,2,3,9-tetramethoxy-8-oxo-5,6,7,7b,8,10a-hexahydrobenzo[*a*]cyclopenta[3,4]cyclobuta[1,2-*c*]cyclohepten-7-yl]acetamide (β-lumicolchicine),



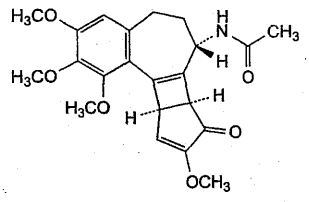
D. *N*-[(7*S*,12a*M*)-3-(β-D-glucopyranosyloxy)-1,2,10-trimethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]acetamide (colchicoside),



E. *N*-[(7*S*,12a*M*)-3-hydroxy-1,2,10-trimethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]acetamide (3-*O*-demethylcolchicine),



F. *N*-[(7*S*,12a*M*)-10-hydroxy-1,2,3-trimethoxy-9-oxo-5,6,7,9-tetrahydrobenzo[*a*]heptalen-7-yl]acetamide (colchiceine),

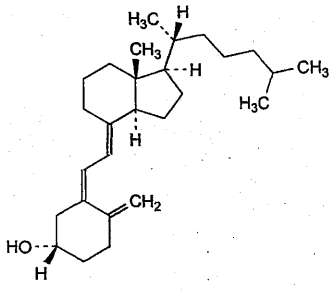


G. *N*-[(7*S*,7b*S*,10a*R*)-1,2,3,9-tetramethoxy-8-oxo-5,6,7,7b,8,10a-hexahydrobenzo[*a*]cyclopenta[3,4]cyclobuta[1,2-*c*]cyclohepten-7-yl]acetamide (γ-lumicolchicine).

Ph Eur

Colecalciferol

(Cholecalciferol, Ph. Eur. monograph 0072)



$C_{27}H_{44}O$ 384.6 *67-97-0*

Action and use
Vitamin D3 analogue.

Preparations
Calcium and Colecalciferol Tablets
Calcium and Colecalciferol Chewable Tablets
Colecalciferol Injection
Colecalciferol Tablets
Paediatric Vitamins A, C and D Oral Drops

When cholecalciferol or vitamin D3 is prescribed or demanded, Colecalciferol shall be dispensed or supplied. When calciferol or vitamin D is prescribed or demanded, Colecalciferol or Ergocalciferol shall be dispensed or supplied.

Ph Eur

DEFINITION
(5*Z*,7*E*)-9,10-Secocholesta-5,7,10(19)-trien-3β-ol.

Content
97.0 per cent to 102.0 per cent.

A reversible isomerisation to pre-cholecalciferol takes place in solution, depending on temperature and time. The activity is due to both compounds (see Assay).

1 mg of cholecalciferol is equivalent to 40 000 IU of antirachitic activity (vitamin D) in rats.

CHARACTERS
Appearance
White or almost white crystals.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in trimethylpentane and in fatty oils.

It is sensitive to air, heat and light. Solutions in solvents without an antioxidant are unstable and are to be used immediately.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cholecalciferol CRS.*

TESTS
Specific optical rotation (*2.2.7*)
+ 105 to + 112, determined within 30 min of preparing the solution.

Dissolve 0.200 g rapidly in *aldehyde-free alcohol R* without heating and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use, avoiding exposure to actinic light and air.*

Test solution Dissolve 10.0 mg of the substance to be examined in *trimethylpentane R* without heating and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *cholecalciferol CRS* in *trimethylpentane R* without heating and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of *cholecalciferol for system suitability CRS* (containing impurity A) to 5.0 mL with the mobile phase. Heat in a water-bath at 90 °C under a reflux condenser for 45 min and cool (formation of pre-cholecalciferol).

Reference solution (c) Dilute 10.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel for chromatography R* (5 μm).

Mobile phase pentanol R, *hexane R* (0.3:99.7 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 5 μL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of cholecalciferol.

Relative retention With reference to cholecalciferol (retention time = about 19 min): pre-cholecalciferol = about 0.5; impurity A = about 0.6.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to pre-cholecalciferol and impurity A.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard the peak due to pre-cholecalciferol.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{27}H_{44}O$ taking into account the assigned content of *cholecalciferol CRS* and, if necessary, the peak due to pre-cholecalciferol.

STORAGE

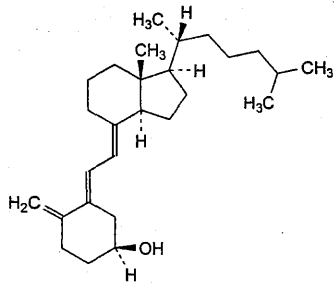
Under nitrogen, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

The contents of an opened container are to be used immediately.

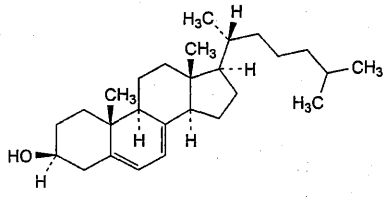
IMPURITIES

Specified impurities *A.*

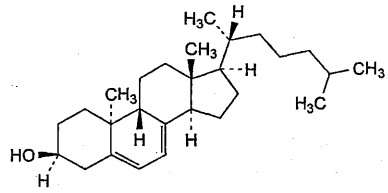
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E.*



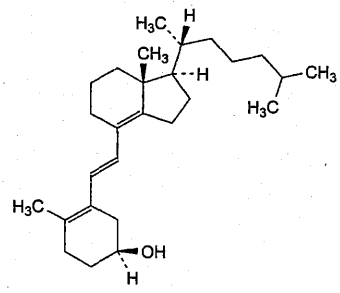
A. (5*E*,7*E*)-9,10-secocholesta-5,7,10(19)-trien-3β-ol (*trans*-cholecalciferol, *trans*-vitamin D_3),



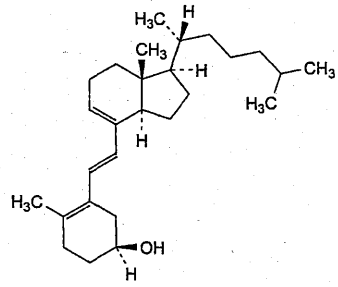
B. cholesta-5,7-dien-3β-ol (7,8-didehydrocholesterol, provitamin D_3),



C. 9β,10α-cholesta-5,7-dien-3β-ol (lumisterol$_3$),



D. (6*E*)-9,10-secocholesta-5(10),6,8(14)-trien-3β-ol (isotachysterol$_3$),



E. (6*E*)-9,10-secocholesta-5(10),6,8-trien-3β-ol (tachysterol$_3$).

Ph Eur

Colecalciferol Concentrate (Oily Form)

(Cholecalciferol Concentrate (Oily Form), Ph. Eur. monograph 0575)

Action and use

Vitamin D analogue (Vitamin D_3).

Ph Eur

DEFINITION

Solution of *Cholecalciferol (0072)* in a suitable vegetable fatty oil, authorised by the competent authority.

Content

90.0 per cent to 110.0 per cent of the cholecalciferol content stated on the label, which is not less than 500 000 IU/g.

It may contain suitable stabilisers such as antioxidants.

CHARACTERS

Appearance

Clear, yellow liquid.

Solubility

Practically insoluble in water, slightly soluble in anhydrous ethanol, miscible with solvents of fats.

Partial solidification may occur, depending on the temperature.

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Prepare a solution in *cyclohexane R* containing the equivalent of about 400 IU/mL.

Spectral range 250-300 nm.

Absorption maximum At 267 nm.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Acid value (*2.5.1*)

Maximum 2.0.

Dissolve 5.0 g in 25 mL of the prescribed mixture of solvents.

Peroxide value (*2.5.5, Method A*)

Maximum 20.

Related substances

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

ASSAY

Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.

Liquid chromatography (*2.2.29*).

Test solution Dissolve a quantity of the preparation to be examined, weighed with an accuracy of 0.1 per cent, equivalent to about 400 000 IU, in 10.0 mL of *toluene R* and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *cholecalciferol CRS* without heating in 10.0 mL of *toluene R* and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of *cholecalciferol for system suitability CRS* to 5.0 mL with the mobile phase. Heat in a water-bath at 90 °C under a reflux condenser for 45 min and cool.

Reference solution (c) Dissolve 0.10 g of *cholecalciferol CRS* without heating in *toluene R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dilute 5.0 mL of reference solution (c) to 50.0 mL with the mobile phase. Keep the solution in iced water.

Reference solution (e) Place 5.0 mL of reference solution (c) in a volumetric flask, add about 10 mg of *butylhydroxytoluene R* and displace air from the flask with *nitrogen R*. Heat in a water-bath at 90 °C under a reflux condenser protected from light and under *nitrogen R* for 45 min. Cool and dilute to 50.0 mL with the mobile phase.

Column:

— *size*: l = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *pentanol R, hexane R* (3:997 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection The chosen volume of each solution (the same volume for reference solution (a) and for the test solution); automatic injection device or sample loop recommended.

Relative retention With reference to cholecalciferol (retention time = about 19 min): pre-cholecalciferol = about 0.4; *trans*-cholecalciferol = about 0.5.

System suitability Reference solution (b):

— *resolution*: minimum 1.0 between the peaks due to pre-cholecalciferol and *trans*-cholecalciferol; if necessary adjust the proportions of the constituents and the flow rate of the mobile phase to obtain this resolution;

— *repeatability*: maximum relative standard deviation of 1.0 per cent for the peak due to cholecalciferol after 6 injections.

Calculate the conversion factor (f) using the following expression:

$$\frac{K - L}{M}$$

K = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (d);

L = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (e);

M = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with reference solution (e).

The value of f determined in duplicate on different days may be used during the entire procedure.

Calculate the content of cholecalciferol in International Units per gram using the following expression:

$$\frac{m'}{V'} \times \frac{V}{m} \times \frac{S_D + (f \times S_p)}{S'_D} \times 40\,000 \times 1000$$

m = mass of the preparation to be examined in the test solution, in milligrams;

m' = mass of *cholecalciferol CRS* in reference solution (a), in milligrams;

V = volume of the test solution (100 mL);

V' = volume of reference solution (a) (100 mL);

S_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with the test solution;

S'_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (a);

S_p = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with the test solution;

f = conversion factor.

STORAGE

In an airtight, well-filled container, protected from light. The contents of an opened container are to be used as soon as possible; any unused part is to be protected by an atmosphere of nitrogen.

LABELLING

The label states:

— the number of International Units per gram;

— the method of restoring the solution if partial solidification occurs.

Ph Eur

Colecalciferol Concentrate (Powder Form)

(Cholecalciferol Concentrate (Powder Form), Ph. Eur. monograph 0574)

Action and use

Vitamin D analogue (Vitamin D_3).

Ph Eur

DEFINITION

Powder concentrate obtained by dispersing an oily solution of *Cholecalciferol (0072)* in an appropriate matrix, which is

usually based on a combination of gelatin and carbohydrates of suitable quality, authorised by the competent authority.

Content

90.0 per cent to 110.0 per cent of the cholecalciferol content stated on the label, which is not less than 100 000 IU/g.

It may contain suitable stabilisers such as antioxidants.

CHARACTERS

Appearance

White or yellowish-white, small particles.

Solubility

Practically insoluble, swells, or forms a dispersion in water, depending on the formulation.

IDENTIFICATION

First identification: A, C.

Second identification: A, B.

A. Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution Place 10.0 mL of the test solution prepared for the assay in a suitable flask and evaporate to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 0.4 mL of *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R*.

Reference solution (a) Dissolve 10 mg of *cholecalciferol CRS* in *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R* and dilute to 4 mL with the same solution.

Reference solution (b) Dissolve 10 mg of *ergocalciferol CRS* in *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R* and dilute to 4 mL with the same solution.

Plate *TLC silica gel G plate R.*

Mobile phase A 0.1 g/L solution of *butylhydroxytoluene R* in a mixture of equal volumes of *cyclohexane R* and *peroxide-free ether R*.

Application 20 µL.

Development Immediately, protected from light, over a path of 15 cm.

Drying In air.

Detection Spray with *sulfuric acid R*.

Results The chromatogram obtained with the test solution shows immediately a bright yellow principal spot, which rapidly becomes orange-brown, then gradually greenish-grey, remaining so for 10 min. This spot is similar in position, colour and size to the spot in the chromatogram obtained with reference solution (a). The chromatogram obtained with reference solution (b) shows immediately at the same level an orange principal spot, which gradually becomes reddish-brown and remains so for 10 min.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Place 5.0 mL of the test solution prepared for the assay in a suitable flask and evaporate to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 50.0 mL of *cyclohexane R*.

Spectral range 250-300 nm.

Absorption maximum At 265 nm.

C. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

ASSAY

Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.

Liquid chromatography (*2.2.29*).

Test solution Introduce into a saponification flask a quantity of the preparation to be examined, weighed with an accuracy of 0.1 per cent, equivalent to about 100 000 IU. Add 5 mL of *water R*, 20 mL of *anhydrous ethanol R*, 1 mL of *sodium ascorbate solution R* and 3 mL of a freshly prepared 50 per cent *m/m* solution of *potassium hydroxide R*. Heat in a water-bath under a reflux condenser for 30 min. Cool rapidly under running water. Transfer the liquid to a separating funnel with the aid of 2 quantities, each of 15 mL, of *water R*, 1 quantity of 10 mL of *ethanol (96 per cent) R* and 2 quantities, each of 50 mL, of *pentane R*. Shake vigorously for 30 s. Allow to stand until the 2 layers are clear. Transfer the lower aqueous-alcoholic layer to a 2nd separating funnel and shake with a mixture of 10 mL of *ethanol (96 per cent) R* and 50 mL of *pentane R*. After separation, transfer the aqueous-alcoholic layer to a 3rd separating funnel and the pentane layer to the 1st separating funnel, washing the 2nd separating funnel with 2 quantities, each of 10 mL, of *pentane R* and adding the washings to the 1st separating funnel. Shake the aqueous-alcoholic layer with 50 mL of *pentane R* and add the pentane layer to the 1st funnel. Wash the pentane layer with 2 quantities, each of 50 mL, of a freshly prepared 30 g/L solution of *potassium hydroxide R* in *ethanol (10 per cent V/V) R*, shaking vigorously, then wash with successive quantities, each of 50 mL, of *water R* until the washings have an approximate pH of 7 to 8, using a *pH indicator strip R*. Transfer the washed pentane extract to a ground-glass-stoppered flask. Evaporate the contents of the flask to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 5.0 mL of *toluene R* and add 20.0 mL of the mobile phase to obtain a solution containing about 4000 IU/mL.

Reference solution (a) Dissolve 10.0 mg of *cholecalciferol CRS*, without heating, in 10.0 mL of *toluene R* and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of *cholecalciferol for system suitability CRS* to 5.0 mL with the mobile phase. Heat in a water-bath at 90 °C under a reflux condenser for 45 min and cool.

Reference solution (c) Dissolve 0.10 g of *cholecalciferol CRS*, without heating, in *toluene R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dilute 5.0 mL of reference solution (c) to 50.0 mL with the mobile phase. Keep the solution in iced water.

Reference solution (e) Place 5.0 mL of reference solution (c) in a volumetric flask, add about 10 mg of *butylhydroxytoluene R* and displace the air from the flask with *nitrogen R*. Heat in a water-bath at 90 °C under a reflux

condenser, protected from light and under *nitrogen R*, for 45 min. Cool and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase pentanol R, *hexane R* (3:997 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection The chosen volume of each solution (the same volume for reference solution (a) and for the test solution); automatic injection device or sample loop recommended.

Relative retention With reference to cholecalciferol: pre-cholecalciferol = about 0.4; *trans*-cholecalciferol = about 0.5.

System suitability Reference solution (b):
— *resolution*: minimum 1.0 between the peaks due to pre-cholecalciferol and *trans*-cholecalciferol; if necessary, adjust the proportions of the constituents and the flow rate of the mobile phase to obtain this resolution;
— *repeatability*: maximum relative standard deviation of 1.0 per cent for the peak due to cholecalciferol after 6 injections.

Calculate the conversion factor (f) using the following expression:

$$\frac{K - L}{M}$$

K = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (d);
L = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (e);
M = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with reference solution (e).

The value of f determined in duplicate on different days may be used during the entire procedure.

Calculate the content of cholecalciferol in International Units per gram using the following expression:

$$\frac{m'}{V'} \times \frac{V}{m} \times \frac{S_D + (f \times S_p)}{S'_D} \times 40\,000 \times 1000$$

m = mass of the preparation to be examined in the test solution, in milligrams;
m' = mass of *cholecalciferol CRS* in reference solution (a), in milligrams;
V = volume of the test solution (25 mL);
V' = volume of reference solution (a) (100 mL);
S_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with the test solution;
S'_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (a);
S_p = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with the test solution;
f = conversion factor.

STORAGE

In an airtight, well-filled container, protected from light. The contents of an opened container are to be used as soon as possible; any unused part is to be protected by an atmosphere of nitrogen.

LABELLING

The label states the number of International Units per gram.

Ph Eur

Colecalciferol Concentrate (Water-dispersible Form)

(Cholecalciferol Concentrate (Water-Dispersible Form), Ph. Eur. monograph 0598)

Action and use
Vitamin D analogue (Vitamin D_3).

Ph Eur

DEFINITION

Solution of *Cholecalciferol (0072)* in a suitable vegetable fatty oil, authorised by the competent authority, to which suitable solubilisers have been added.

Content
90.0 per cent to 115.0 per cent of the cholecalciferol content stated on the label, which is not less than 100 000 IU/g.

It may contain suitable stabilisers such as antioxidants.

CHARACTERS

Appearance
Slightly yellowish liquid of variable opalescence and viscosity.

Highly concentrated solutions may become cloudy at low temperatures or form a gel at room temperature.

IDENTIFICATION

First identification: A, C, D.

Second identification: A, B, D.

A. Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution Place 10.0 mL of the test solution prepared for the assay in a suitable flask and evaporate to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 0.4 mL of *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R*.

Reference solution (a) Dissolve 10 mg of *cholecalciferol CRS* in *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R* and dilute to 4 mL with the same solution.

Reference solution (b) Dissolve 10 mg of *ergocalciferol CRS* in *ethylene chloride R* containing 10 g/L of *squalane R* and 0.1 g/L of *butylhydroxytoluene R* and dilute to 4 mL with the same solution.

Plate *TLC silica gel G plate R.*

Mobile phase A 0.1 g/L solution of *butylhydroxytoluene R* in a mixture of equal volumes of *cyclohexane R* and *peroxide-free ether R*.

Application 20 µL.

Development Immediately, protected from light, over a path of 15 cm.

Drying In air.

Detection Spray with *sulfuric acid R*.

Results The chromatogram obtained with the test solution shows immediately a bright yellow principal spot, which rapidly becomes orange-brown, then gradually greenish-grey, remaining so for 10 min. This spot is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a). The chromatogram obtained with reference solution (b) shows immediately at the same level an orange principal spot, which gradually becomes reddish-brown and remains so for 10 min.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Place 5.0 mL of the test solution prepared for the assay in a suitable flask and evaporate to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 50.0 mL of *cyclohexane R*.

Spectral range 250-300 nm.

Absorption maximum At 265 nm.

C. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

D. Mix about 1 g with 10 mL of *water R* previously warmed to 50 °C, and cool to 20 °C. Immediately after cooling, a uniform, slightly opalescent and slightly yellow dispersion is obtained.

TESTS

Related substances

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

ASSAY

Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air.

Liquid chromatography (*2.2.29*).

Test solution Introduce into a saponification flask a quantity of the preparation to be examined, weighed with an accuracy of 0.1 per cent, equivalent to about 100 000 IU. Add 5 mL of *water R*, 20 mL of *anhydrous ethanol R*, 1 mL of *sodium ascorbate solution R* and 3 mL of a freshly prepared 50 per cent *m/m* solution of *potassium hydroxide R*. Heat in a water-bath under a reflux condenser for 30 min. Cool rapidly under running water. Transfer the liquid to a separating funnel with the aid of 2 quantities, each of 15 mL, of *water R*, 1 quantity of 10 mL of *ethanol (96 per cent) R* and 2 quantities, each of 50 mL, of *pentane R*. Shake vigorously for 30 s. Allow to stand until the 2 layers are clear. Transfer the aqueous-alcoholic layer to a 2nd separating funnel and shake with a mixture of 10 mL of *ethanol (96 per cent) R* and 50 mL of *pentane R*. After separation, transfer the aqueous-alcoholic layer to a 3rd separating funnel and the pentane layer to the 1st separating funnel, washing the 2nd separating funnel with 2 quantities, each of 10 mL, of *pentane R* and adding the washings to the 1st separating funnel. Shake the aqueous-alcoholic layer with 50 mL of *pentane R* and add the pentane layer to the 1st funnel. Wash the pentane layer with 2 quantities, each of 50 mL, of a freshly prepared 30 g/L solution of *potassium hydroxide R* in *ethanol (10 per cent V/V) R*, shaking vigorously, and then wash with successive quantities, each of 50 mL, of *water R* until the washings have an approximate pH of 7 to 8, using a *pH indicator strip R*. Transfer the washed pentane extract to a ground-glass-stoppered flask. Evaporate the contents of the flask to dryness under reduced pressure by swirling in a water-bath at 40 °C. Cool under running water and restore atmospheric pressure with *nitrogen R*. Dissolve the residue immediately in 5.0 mL of *toluene R* and add 20.0 mL of the mobile phase to obtain a solution containing about 4000 IU/mL.

Reference solution (a) Dissolve 10.0 mg of *cholecalciferol CRS*, without heating, in 10.0 mL of *toluene R* and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of *cholecalciferol for system suitability CRS* to 5.0 mL with the mobile phase. Heat in a water-bath at 90 °C under a reflux condenser for 45 min and cool.

Reference solution (c) Dissolve 0.10 g of *cholecalciferol CRS*, without heating, in *toluene R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dilute 5.0 mL of reference solution (c) to 50.0 mL with the mobile phase. Keep the solution in iced water.

Reference solution (e) Place 5.0 mL of reference solution (c) in a volumetric flask, add about 10 mg of *butylhydroxytoluene R* and displace the air from the flask with *nitrogen R*. Heat in a water-bath at 90 °C under a reflux condenser, protected from light and under *nitrogen R*, for 45 min. Cool and dilute to 50.0 mL with the mobile phase.

Column:
- *size*: l = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase pentanol R, *hexane R* (3:997 *V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection The chosen volume of each solution (the same volume for reference solution (a) and for the test solution); automatic injection device or sample loop recommended.

Relative retention With reference to cholecalciferol: pre-cholecalciferol = about 0.4; *trans*-cholecalciferol = about 0.5.

System suitability Reference solution (b):
- *resolution*: minimum 1.0 between the peaks due to pre-cholecalciferol and *trans*-cholecalciferol; if necessary, adjust the proportions of the constituents and the flow rate of the mobile phase to obtain this resolution;
- *repeatability*: maximum relative standard deviation of 1.0 per cent for the peak due to cholecalciferol after 6 injections.

Calculate the conversion factor (f) using the following expression:

$$\frac{K - L}{M}$$

K = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (d);
L = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (e);
M = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with reference solution (e).

The value of f determined in duplicate on different days may be used during the entire procedure.

Calculate the content of cholecalciferol in International Units per gram using the following expression:

$$\frac{m'}{V'} \times \frac{V}{m} \times \frac{S_D + (f \times S_p)}{S'_D} \times 40\,000 \times 1000$$

m = mass of the preparation to be examined in the test solution, in milligrams;
m' = mass of *cholecalciferol CRS* in reference solution (a), in milligrams;
V = volume of the test solution (25 mL);
V' = volume of reference solution (a) (100 mL);
S_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with the test solution;

S'_D = area (or height) of the peak due to cholecalciferol in the chromatogram obtained with reference solution (a);
S_p = area (or height) of the peak due to pre-cholecalciferol in the chromatogram obtained with the test solution;
f = conversion factor.

STORAGE
In an airtight, well-filled container, protected from light, at the temperature stated on the label.

The contents of an opened container are to be used as soon as possible; any unused part is to be protected by an atmosphere of inert gas.

LABELLING
The label states:
— the number of International Units per gram;
— the storage temperature.

Ph Eur

Colestipol Hydrochloride

37296-80-3

Action and use
Lipid-regulating drug.

Preparation
Colestipol Granules

DEFINITION
Colestipol Hydrochloride is a co-polymer of diethylenetriamine and 1-chloro-2,3-epoxypropane. Each g binds not less than 1.1 mEq and not more than 1.7 mEq of sodium cholate, determined in the test for Cholate binding capacity and calculated with reference to the dried material.

CHARACTERISTICS
Yellow to orange beads; hygroscopic.

Practically insoluble in *ethanol (96%)* and in *dichloromethane*; swells but does not dissolve in *water* and dilute aqueous solutions of acids and alkalis.

IDENTIFICATION
Carry out the method for *gas chromatography*, Appendix III B, using a suitable gas chromatograph fitted with a pyrolysis unit. Operate the unit in accordance with the manufacturer's instructions to obtain a pyrogram for *colestipol hydrochloride BPCRS* that is similar to that supplied with the reference material.

(1) To prepare the sample, mix 1 part of n-*eicosane* and 4 parts of the substance being examined and grind the mixture in a mortar with *chloroform* until the substance being examined is uniformly coated with the *n*-eicosane.

(2) Prepare the standard in the same manner but adding 4 parts of *colestipol hydrochloride BPCRS* in place of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use a a glass column (1.8 m × 3 mm) packed with *acid-washed, silanised diatomaceous support* (80 to 100 mesh) (Chromosorb W is suitable) coated with 0.25% w/w of *potassium hydroxide* and 5% w/w of *polyethylene glycol 20,000* (Carbowax 20M is suitable).

(b) Use *helium* as the carrier gas at 60 mL per minute.

(c) Use isothermal conditions maintained at 85°.

(d) Use a pyrolysis unit capable of attaining a temperature of about 1000° when fitted with a platinum ribbon probe.

(e) Use a detector at a temperature of 270°.

(f) Load the sample and the standard separately into the pyrolysis unit.

CONFIRMATION

The pyrogram obtained with the substance being examined is concordant with that obtained with *colestipol hydrochloride BPCRS*.

TESTS

Acidity or alkalinity
Shake a 10% w/w suspension in a stoppered vial at approximately 10-minute intervals for 1 hour and centrifuge. Transfer a portion of the clear supernatant liquid to a suitable container and record the pH as soon as the reading has stabilised. The pH is 6.0 to 7.5, Appendix V L.

Water-soluble substances
Place 5 g in a glass-stoppered, 125 mL conical flask, add 80 mL of *water*, close the flask and shake in a water bath at 36° to 38° for 72 hours. Filter the contents of the flask through a fine-porosity, sintered glass funnel or woven glass-fibre filter, collecting the filtrate in a tared 125 mL conical flask. Rinse any residual contents in the flask with two 5 mL quantities of *water*, filter the washings and combine the filtrates from the washings with the filtrate obtained previously. Evaporate the filtrate to dryness, using filtered air or nitrogen, if necessary, to aid in the evaporation. Dry the residue at 75° at a pressure of not more than 2 kPa for 1 hour, allow to cool in a desiccator and weigh. Repeat the procedure at the same time without the substance being examined beginning at the words 'add 80 mL of *water*...'. The difference in the weights of the residues is not more than 25 mg (0.5%).

Loss on drying
When dried at 75° at a pressure of not more than 2 kPa for 16 hours, loses not more than 1.0% of its weight.

Sulfated ash
Not more than 0.3%, Appendix IX A.

Chloride content
Not less than 6.5% and not more than 9.0% calculated with reference to the dried substance. Burn 20 mg by the method for *oxygen-flask combustion*, Appendix VIII C, using 10 mL of 0.05M *sodium hydroxide* as the absorbing liquid. When the process is complete shake the flask vigorously, allow to stand with frequent shaking for about 40 minutes or until no cloudiness is observed; add 20 mL of *ethanol (96%)* and 0.2 mL of *nitric acid*. Titrate the resulting solution with 0.05M *silver nitrate VS*, determining the end point potentiometrically using a silver-silver chloride electrode and a glass reference electrode (V mL). Repeat the procedure without the substance being examined adding 10 mL of 0.0075M *sodium chloride* to the solution in the flask (V_1 mL). Add 10 mL of 0.0075M *sodium chloride* to a flask containing a mixture of 10 mL of *water* and 20 mL of *ethanol (96%)*, add 0.2 mL of *nitric acid* and titrate with *0.05M silver nitrate VS*, determining the end point potentiometrically using a silver-silver chloride electrode and a glass reference electrode (V_2 mL). Determine the volume of *0.05M silver nitrate VS* required by the substance being examined using the following expression:

$V-(V_1-V_2)$

Each mL of *0.05M silver nitrate VS* is equivalent to 1.773 mg of Cl.

Water absorption capacity
Each g of Colestipol Hydrochloride absorbs not less than 3.3 g and not more than 5.3 g of *water* when determined in the following manner. Transfer 5 g to a dry, plastic container and add 80 g of *water*. Cover the container and allow the resulting suspension to equilibrate for 72 hours. Filter the resulting slurry through a medium-porosity fritted-glass funnel (KIMAX 60 mL-40M is suitable) at a pressure of 2kPa; collect the filtrate in a tared, plastic container, disconnecting the vacuum 2 minutes after collection of the last portion of the filtrate. Immediately weigh the container and the filtrate and determine the weight, in g, of the filtrate. Calculate the weight of water absorbed per g from the difference between the weight of the filtrate and the original weight of *water* used in the test.

Cholate binding capacity
Prepare a solution containing 1.0% w/v of *sodium cholate* and 0.9% w/v of *sodium chloride* and adjust to pH 6.4 by the drop wise addition of *hydrochloric acid* (solution A). Transfer 1 g (*m* g) of the substance being examined to a ground-glass-stoppered flask, add 100 mL of freshly prepared solution A and shake vigorously for 90 minutes with the flask positioned horizontally. Remove the flask from the shaker and allow the contents to settle for 5 minutes. Adjust the pH of a 20 mL aliquot of the supernatant liquid to 10.5 by the drop wise addition of 1M *sodium hydroxide* and titrate potentiometrically with 0.1M *hydrochloric acid* VS to the second inflection point of the pH curve. Determine the volume of titrant added between the inflection points. Carry out a blank titration on 20 mL of freshly prepared solution A. The difference between the titrations represents the amount of hydrochloric acid required (*V* mL). Calculate the cholate binding capacity of the substance being examined in milliequivalents from the expression $0.5V/m$.

Colestipol exchange capacity limit
Not less than 9.0 mEq per g and not more than 11.0 mEq per g determined in the following manner. Transfer not less than 2 g and 100 mL of 1M *sodium hydroxide* to a stoppered flask and shake for 4 hours. Filter the suspension through a coarse-porosity sintered-glass funnel and wash the resin with 500 mL of *water*. Transfer the resin to a 1000 mL beaker, add 200 mL of *water* and allow to stand for 10 minutes. Filter the suspension, check the pH of the filtrate and repeat the washing procedure with 200 mL quantities of *water* until the pH of the filtrate is less than 8 [5 litres may be required]. Dry the resin and funnel at 60° at a pressure of 2 kPa for at least 16 hours, breaking up any aggregates with a spatula and store in a dessicator.

Place 1 g (*w* g) of the free base resin prepared above and add 100 mL of *0.2M hydrochloric acid VS* in a stoppered flask and shake for not less than 2.5 hours. Filter a portion of the suspension through glass wool. Titrate 8 mL of the filtrate with *0.2M sodium hydroxide VS*, determining the end point potentiometrically (*a* mL). Carry out a blank titration on 5 mL of the *0.2M hydrochloric acid VS* used to equilibrate the free-base resin diluted with 5 mL of *water* (*b* mL). Calculate the exchange capacity in milliequivalents per g from the expression:

$(20/w)(b/5-a/8)$

STORAGE
Colestipol Hydrochloride should be kept in an airtight container.

Colestyramine

(Ph. Eur. monograph 1775)

11041-12-6

Action and use
Lipid-regulating drug.

Preparation
Colestyramine Oral Powder

Ph Eur

DEFINITION
Strongly basic anion-exchange resin in chloride form, consisting of styrene-divinylbenzene copolymer with quaternary ammonium groups.

Nominal exchange capacity 1.8 g to 2.2 g of sodium glycocholate per gram (dried substance).

CHARACTERS
Appearance
White or almost white, fine powder, hygroscopic.

Solubility
Insoluble in water, in methylene chloride and in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *colestyramine CRS*.
B. Chloride (see Tests).

TESTS
pH (*2.2.3*)
4.0 to 6.0.
Suspend 0.100 g in 10 mL of *water R* and allow to stand for 10 min.

Dialysable quaternary amines
Maximum 500 ppm, expressed as benzyltrimethylammonium chloride.

Test solution Place a 25 cm piece of cellulose dialysis tubing having a molecular weight cut-off of 12 000-14 000 and an inflated diameter of 3-6 cm (flat width of 5-9 cm) in *water R* to hydrate until pliable, appropriately sealing one end. Introduce 2.0 g of the substance to be examined into the tube and add 10 mL of *water R*. Seal the tube and completely immerse it in 100 mL of *water R* in a suitable vessel and stir the liquid for 16 h to effect dialysis. Use the dialysate as test solution.

Reference solution Prepare the reference solution in a similar manner but using 10 mL of a freshly prepared 0.1 g/L solution of *benzyltrimethylammonium chloride R* instead of the substance to be examined.

Transfer 5.0 mL of the test solution to a separating funnel and add 5 mL of a 3.8 g/L solution of *disodium tetraborate R*, 1 mL of a solution containing 1.5 g/L of *bromothymol blue R* and 4.05 g/L of *sodium carbonate R* and 10 mL of *chloroform R*. Shake the mixture vigourously for 1 min, allow the phases to separate and transfer the clear organic layer to a 25 mL volumetric flask. Repeat the extraction with a further 10 mL of *chloroform R*, combine the organic layers and dilute to 25 mL with *chloroform R*. Measure the absorbance (*2.2.25*) of the solution at the absorption maximum at 420 nm, using as compensation liquid a solution prepared in the same manner but using 5.0 mL of *water R* instead of the test solution.

Repeat the operation using 5.0 mL of the reference solution.

The absorbance obtained with the test solution is not greater than that obtained with the reference solution.

Impurity A

Liquid chromatography (*2.2.29*).

Test solution Shake 5.0 g with 10 mL of *acetone R* for 30 min. Centrifuge and use the supernatant.

Reference solution (a) Dissolve 5 mg of *styrene R* in *acetone R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *acetone R*.

Reference solution (b) Dissolve 0.35 mL of *styrene R* in *acetone R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *acetone R*.

Reference solution (c) Dissolve 0.35 mL of *toluene R* in *acetone R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Mix 1.0 mL of reference solution (b) and 1.0 mL of reference solution (c) with *acetone R* and dilute to 100.0 mL with the same solvent.

Column:
- *size*: *l* = 0.30 m, Ø = 3.9 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm) with a specific surface area of 330 m^2/g and a pore size of 12.5 nm.

Mobile phase *acetonitrile R, water R* (50:50 *V/V*).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of test solution and reference solutions (a) and (d).

System suitability Reference solution (d):
- *resolution*: minimum 1.5 between the peaks due to impurity A and toluene.

Limit:
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1 ppm).

Chloride

13.0 per cent to 17.0 per cent (dried substance).

To 0.2 g add 100 mL of *water R* and 50 mg of *potassium nitrate R*. Add, with stirring, 2 mL of *nitric acid R* and titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 3.55 mg of Cl.

Loss on drying (*2.2.32*)

Maximum 12 per cent, determined on 1.000 g by drying in an oven at 70 °C over *diphosphorus pentoxide R* at a pressure not exceeding 7 kPa for 16 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Exchange capacity

Liquid chromatography (*2.2.29*).

Solution A Dissolve 1.500 g of *sodium glycocholate R* in a solution containing 4 g/L of *potassium dihydrogen phosphate R* and 12 g/L of *dipotassium hydrogen phosphate R* and dilute to 100.0 mL with the same solution.

Test solution Add 20.0 mL of solution A to a quantity of the substance to be examined equivalent to about 0.100 g of the dried substance. Shake mechanically for 2 h and centrifuge for 15 min. Dilute 5.0 mL of the supernatant to 50.0 mL with *water R*.

Reference solution (a) Dilute 4.0 mL of solution A to 100.0 mL with *water R*.

Reference solution (b) Dissolve 60 mg of *sodium glycocholate R* and 30 mg of *sodium taurodeoxycholate R* in *water R* and dilute to 100 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 10.9 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 50 µL.

Run time Twice the retention time of glycocholate.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to glycocholate and taurodeoxycholate.

Calculate the nominal exchange capacity using the following expression:

$$\frac{(2.5\,A_1 - A_2) \times m_1 \times 1.2}{12.5 \times A_1 \times m_2}$$

A_1 = area of the peak due to glycocholate in the chromatogram obtained with reference solution (a),

A_2 = area of the peak due to glycocholate in the chromatogram obtained with the test solution,

m_1 = mass, in milligrams, of *sodium glycocholate R* used in the preparation of solution A,

m_2 = mass, in milligrams, of the dried substance to be examined used in the preparation of the test solution,

1.2 = correction factor to convert the true exchange capacity to the conventionally used nominal exchange capacity.

STORAGE

In an airtight container.

IMPURITIES

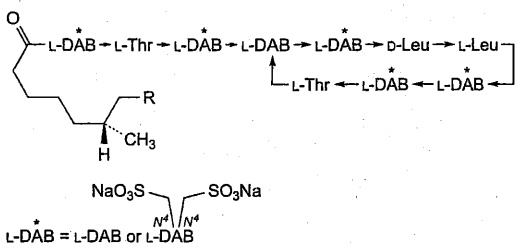
Specified impurities *A*.

A. styrene.

Ph Eur

Colistimethate Sodium

(Ph. Eur. monograph 0319)



L-DAB = L-DAB or L-DAB

DAB = 2,4-diaminobutanoic acid

Polymyxin E1 derivative: R = CH_3

Polymyxin E2 derivative: R = H

Between 2 and 5 of the L-DAB residues are disubstituted at N^4

CMS E1ASM8: principal polymyxin E1 with 4 disubstituted residues

CMS E1ASM6: principal polymyxin E1 with 3 disubstituted residues

CMS E1ASM4: principal polymyxin E1 with 2 disubstituted residues

CMS E2ASM8: principal polymyxin E2 with 4 disubstituted residues

CMS E2ASM6: principal polymyxin E2 with 3 disubstituted residues

CMS E2ASM4: principal polymyxin E2 with 2 disubstituted residues

8068-28-8

Action and use
Antibacterial.

Preparations
Colistimethate for Injection

Colistimethate Sodium Powder for Nebuliser Solution

Ph Eur

DEFINITION

Colistimethate sodium is prepared from colistin by the action of formaldehyde and sodium hydrogen sulfite to form a mixture of di to penta bis-sulfomethylated primary amine derivatives, mainly polymyxins E1 and E2.

Semi-synthetic product derived from a fermentation product.

Content
Minimum 11 500 IU/mg (dried substance).

PRODUCTION

The composition and purity of the colistin starting material used in the synthesis of colistimethate sodium are equivalent to that described in the monograph *Colistin sulfate (0320)*. In addition, the colistin starting material must have a polymyxin E1 content between 50 per cent and 75 per cent and a polymyxin E2 content between 5 per cent and 20 per cent, in order to comply with the composition limits for colistimethate sodium.

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Very soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for composition.

Results The peaks due to CMS E1ASM8, CMS E1ASM6, CMS E1ASM4, CMS E2ASM8, CMS E2ASM6 and CMS E2ASM4 in the chromatogram obtained with the test solution are similar in retention time to the corresponding peaks in the chromatogram obtained with reference solution (a).

B. It gives reaction (b) of sodium (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*).

Dissolve 0.16 g in 10 mL of *water R*.

pH (*2.2.3*)
6.5 to 8.5.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Measure after 30 min.

Free colistin
Dissolve 80 mg in 3 mL of *water R*. Add 0.1 mL of a 100 g/L solution of *silicotungstic acid R*; 10-20 s after addition of the reagent, the solution is not more opalescent than reference suspension II (*2.2.1*).

Composition
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Buffer solution 7.8 g/L solution of *sodium dihydrogen phosphate R* adjusted to pH 6.5 with *1 M sodium hydroxide*.

Test solution Dissolve 20 mg of the substance to be examined in 0.5 mL of *water R* and dilute to 10.0 mL with *methanol R*.

Reference solution (a) Dissolve 10 mg of *colistimethate sodium for peak identification CRS* in 0.25 mL of *water R* and dilute to 5.0 mL with *methanol R*.

Reference solution (b) Dissolve 2 mg of *E1 colistimethate sodium for peak identification CRS* in 0.25 mL of *water R* and dilute to 2.0 mL with *methanol R*.

Reference solution (c) Dissolve 1.5 mg of *E2 colistimethate sodium for peak identification CRS* in 0.25 mL of *water R* and dilute to 5.0 mL with *methanol R*.

Reference solution (d) Dilute 1.5 mL of reference solution (a) to 25.0 mL with *methanol R*.

Precolumn:
— *size*: *l* = 5 mm, Ø = 2.1 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (1.7 µm).

Column:
— *size*: *l* = 0.15 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (1.7 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, buffer solution (25:475 *V/V*);
— *mobile phase B*: *acetonitrile R1*, buffer solution (250:250 *V/V*);

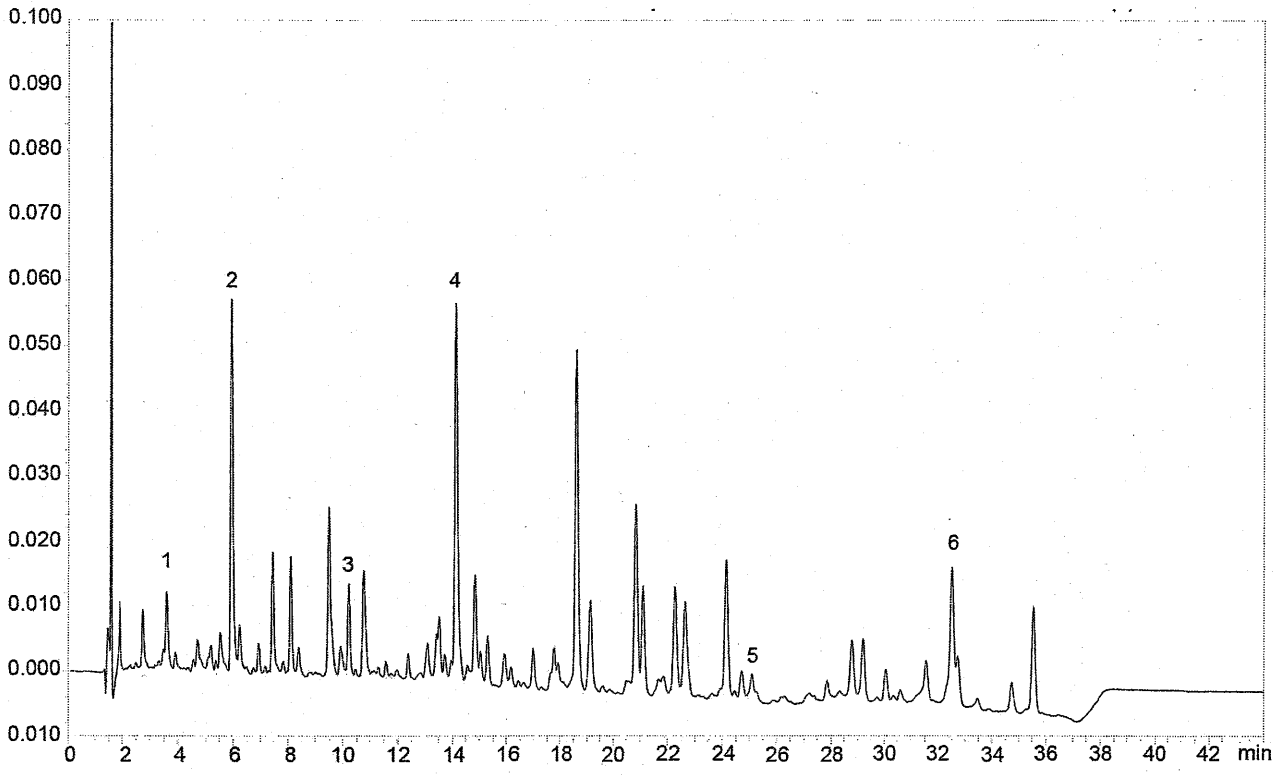
Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	80 → 68	20 → 32
10 - 35	68 → 53	32 → 47

Flow rate 0.30 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 5 °C.

Injection 2.0 µL.



1. CMS E2ASM8
2. CMS E1ASM8
3. CMS E2ASM6
4. CMS E1ASM6
5. CMS E2ASM4
6. CMS E1ASM4

Figure 0319.-1. – *Chromatogram for the test for composition of colistimethate sodium: reference solution (a)*

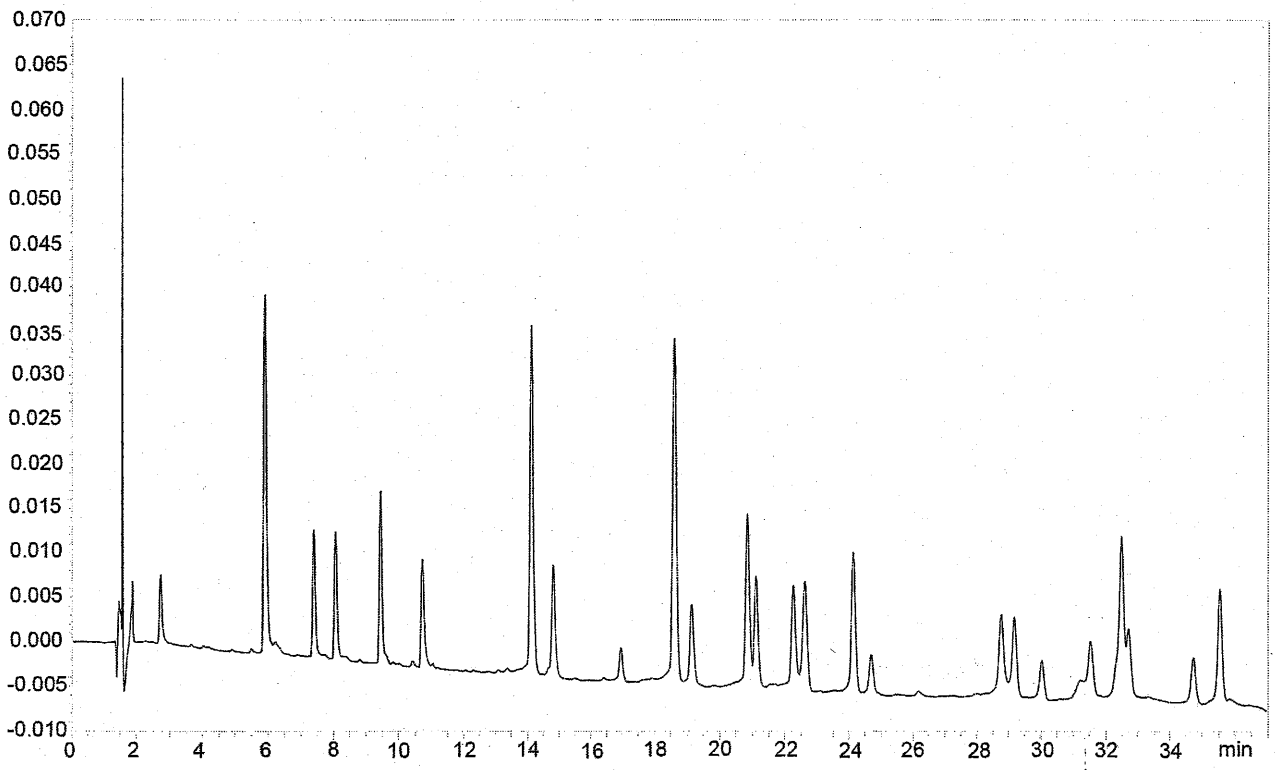


Figure 0319.-2. – *Chromatogram for the test for composition of colistimethate sodium: reference solution (b)*

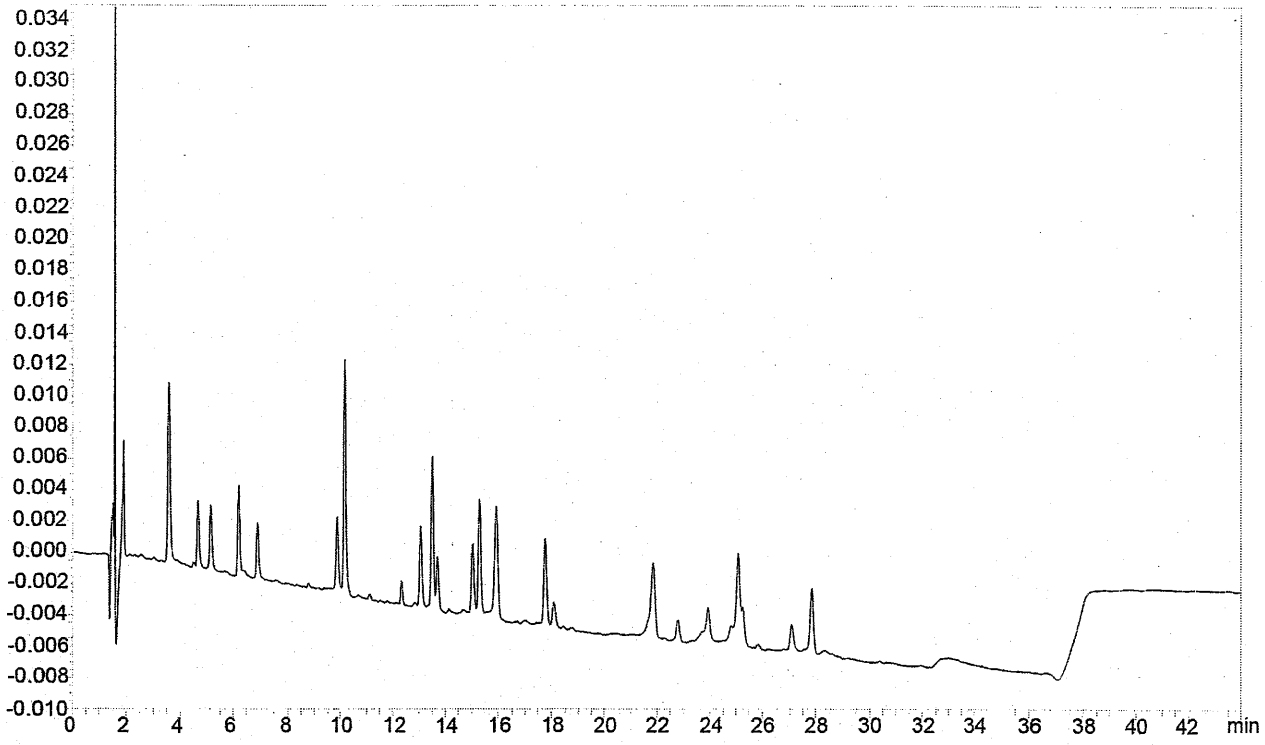


Figure 0319.-3. – *Chromatogram for the test for composition of colistimethate sodium: reference solution (c)*

Identification of peaks Use the chromatogram supplied with *colistimethate sodium for peak identification CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to CMS E1ASM8, CMS E1ASM6, CMS E1ASM4, CMS E2ASM8, CMS E2ASM6 and CMS E2ASM4 (see Figure 0319.-1).

The peak corresponding to the most abundant compound in the range of 11.0-14.5 min (CMS E1ASM6) in the chromatogram obtained with reference solution (a) is set as the identification reference peak (relative retention 1.00).

Identification of peaks related to CMS E1 and CMS E2 Use the chromatogram supplied with *E1 colistimethate sodium for peak identification CRS* and *E2 colistimethate sodium for peak identification CRS*, and the chromatograms obtained with reference solutions (b) and (c) to identify all peaks related to CMS E1 and CMS E2 in the chromatogram obtained with the test solution (see Figures 0319.-2 and 0319.-3).

Relative retention With reference to CMS E1ASM6 (retention time = about 13 min): CMS E2ASM8 = about 0.22; CMS E1ASM8 = about 0.39; CMS E2ASM6 = about 0.71; CMS E2ASM4 = about 1.77; CMS E1ASM4 = about 2.35.

Integrate all peaks above 0.05 per cent to establish the total area.

System suitability:
- the difference in the retention times of CMS E1ASM6 in 2 consecutive injections of reference solution (a) is less than 0.1 min; the drift in the retention time of CMS E1ASM6 from the beginning to the end of the sequence is less than 0.5 min;
- *peak-to-valley ratio*: minimum 1.2, where H_p = height above the baseline of the peak with a relative retention of about 2.37 and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to CMS E1ASM4 in the chromatogram obtained with reference solution (a);
- *number of theoretical plates*: minimum 50 000, calculated for the peak due to CMS E1ASM6 in the chromatogram obtained with reference solution (a);
- *signal-to-noise ratio*: minimum 50 for the peak due to CMS E1ASM6 in the chromatogram obtained with reference solution (d).

Limits:
- *CMS E1ASM8*: 5.0 per cent to 9.5 per cent;
- *CMS E1ASM6*: 6.5 per cent to 9.5 per cent;
- *CMS E1ASM4*: 2.0 per cent to 5.0 per cent;
- *CMS E2ASM8*: 0.5 per cent to 2.0 per cent;
- *CMS E2ASM6*: 0.5 per cent to 2.5 per cent;
- *CMS E2ASM4*: maximum 1.5 per cent;
- *sum of the peaks related to CMS E1 and CMS E2*: minimum 77.0 per cent;
- *disregard limit*: 0.50 per cent.

Related substances

Liquid chromatography (*2.2.29*) as described in the test for composition.

Limits:
- *any other impurity* (any peak not related to CMS E1 or CMS E2): for each impurity, maximum 1.5 per cent;
- *sum of impurities* (sum of all peaks not related to CMS E1 or CMS E2): maximum 5.5 per cent;
- *disregard limit*: 0.50 per cent.

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 670 Pa for 3 h.

Sulfated ash (*2.4.14*)

16 per cent to 21 per cent, determined on 0.50 g.

Pyrogens (*2.6.8*)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of pyrogens, it complies with the test. Inject, per

kilogram of the rabbit's mass, 1 mL of a solution in *water for injections R* containing 2.5 mg of the substance to be examined per millilitre.

ASSAY

Carry out the microbiological assay of antibiotics (*2.7.2*).

STORAGE

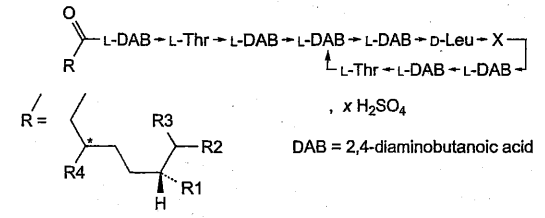
In an airtight container, protected from light. If the substance is sterile the container is also sterile and tamper-proof.

Ph Eur

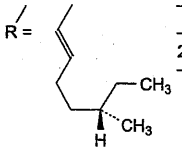
Colistin Sulfate

Colistin Sulphate

(Ph. Eur. monograph 0320)



polymyxin	X	R1	R2	R3	R4	Mol. Formula	M_r
E1	L-Leu	CH_3	CH_3	H	H	$C_{53}H_{100}N_{16}O_{13}$	1169
E2	L-Leu	CH_3	H	H	H	$C_{52}H_{98}N_{16}O_{13}$	1155
E3	L-Leu	H	CH_3	H	H	$C_{52}H_{98}N_{16}O_{13}$	1155
E4	L-Leu	H	H	H	H	$C_{51}H_{96}N_{16}O_{13}$	1141
E6	L-Leu	CH_3	CH_3	H	OH	$C_{53}H_{100}N_{16}O_{14}$	1185
E1-7MOA	L-Leu	H	CH_3	CH_3	H	$C_{53}H_{100}N_{16}O_{13}$	1169
E1-I	L-Ile	CH_3	CH_3	H	H	$C_{53}H_{100}N_{16}O_{13}$	1169
E1-Nva	L-Nva	CH_3	CH_3	H	H	$C_{52}H_{98}N_{16}O_{13}$	1155
E2-I	L-Ile	CH_3	H	H	H	$C_{52}H_{98}N_{16}O_{13}$	1155
E2-Val	L-Val	CH_3	H	H	H	$C_{51}H_{96}N_{16}O_{13}$	1141



polymyxin	X	Mol. Formula	M_r
2,3-dehydro E1	L-Leu	$C_{53}H_{98}N_{16}O_{13}$	1167

1264-72-8

Action and use

Antibacterial.

Preparation

Colistin Tablets

Ph Eur

DEFINITION

A mixture of the sulfates of polypeptides produced by certain strains of *Bacillus polymyxa* var. *colistinus*.

Content

Minimum 19 000 IU/mg (dried substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

First identification: B, E.

Second identification: A, C, D, E.

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in 1 mL of a mixture of equal volumes of *hydrochloric acid R* and *water R*. Heat at 135 °C in a sealed tube for 5 h. Evaporate to dryness on a water-bath and continue the heating until moistened *blue litmus paper R* does not turn red. Dissolve the residue in 0.5 mL of *water R*.

Reference solution (a) Dissolve 20 mg of *leucine R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *threonine R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dissolve 20 mg of *phenylalanine R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (d) Dissolve 20 mg of *serine R* in *water R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Carry out the following procedures protected from light.

Mobile phase *water R, phenol R* (25:75 *V/V*).

Application 5 µL as bands of 10 mm, then place the plate in the chromatographic tank so that it is not in contact with the mobile phase, and allow it to become impregnated with the vapour of the mobile phase for at least 12 h.

Development Over half of the plate.

Drying At 105 °C.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

Results The chromatogram obtained with the test solution shows zones corresponding to those in the chromatograms obtained with reference solutions (a) and (b), but shows no zones corresponding to those in the chromatograms obtained with reference solutions (c) and (d); the chromatogram obtained with the test solution also shows a zone with a very low R_F value (2,4-diaminobutyric acid).

B. Examine the chromatograms obtained in the test for composition.

Results The peaks due to polymyxin E1 and polymyxin E2 in the chromatogram obtained with the test solution are similar in retention time to the corresponding peaks in the chromatogram obtained with reference solution (a).

C. Dissolve about 5 mg in 3 mL of *water R*. Add 3 mL of *dilute sodium hydroxide solution R*. Shake and add 0.5 mL of a 10 g/L solution of *copper sulfate pentahydrate R*. A violet colour is produced.

D. Dissolve about 50 mg in 1 mL of *1 M hydrochloric acid* and add 0.5 mL of *0.01 M iodine*. The solution remains coloured.

E. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

pH (*2.2.3*)

4.0 to 6.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Composition

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 5.0 mg of the substance to be examined in 8 mL of *water R* and dilute to 10.0 mL with *acetonitrile R*.

Reference solution (a) Dissolve 5.0 mg of *colistin for system suitability CRS* in 8 mL of *water R* and dilute to 10.0 mL with *acetonitrile R*.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with a mixture of 20 volumes of *acetonitrile R* and 80 volumes of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.0 µm);
— *temperature*: 50 °C.

Mobile phase Mix 22 volumes of *acetonitrile R1* and 78 volumes of a solution prepared as follows: dissolve 4.46 g of *anhydrous sodium sulfate R* in 900 mL of *water for chromatography R*, adjust to pH 2.4 with *dilute phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 1.5 times the retention time of polymyxin E1.

Identification of peaks Use the chromatogram supplied with *colistin for system suitability CRS* to identify the peaks due to polymyxins E1, E2, E3, E4, E6, E1-7MOA, E1-I, E1-Nva, E2-I, E2-Val, 2,3-dehydro E1 and due to impurities A and B.

Relative retention With reference to polymyxin E1 (retention time = about 21 min): polymyxins E4 and E2-Val = about 0.28; polymyxin E6 = about 0.39; polymyxin E2-I = about 0.42; polymyxin E2 = about 0.50; impurity A = about 0.53; polymyxin E3 = about 0.56; polymyxin E1-Nva = about 0.59; polymyxin E1-I = about 0.82; polymyxin 2,3-dehydro E1 = about 0.90; polymyxin E1-7MOA = about 1.1; impurity B = about 1.3.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to polymyxin E6 and polymyxin E2-I, and minimum 3.0 between the peaks due to polymyxin 2,3-dehydro E1 and polymyxin E1 in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 10 for the peak due to polymyxin E1 in the chromatogram obtained with reference solution (b);
— *peak-to-valley ratio*: minimum 1.1, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to polymyxin E2 in the chromatogram obtained with reference solution (a).

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of polymyxin 2,3-dehydro E1 by 0.3;
— *polymyxin E1-I*: maximum 8.5 per cent;
— *polymyxin E3*: maximum 5.5 per cent;
— *polymyxin E1-7MOA*: maximum 5.0 per cent;
— *polymyxin E6*: maximum 4.5 per cent;
— *polymyxin E1-Nva*: maximum 4.5 per cent;
— *sum of polymyxins E4 and E2-Val*: maximum 3.0 per cent;
— *polymyxin E2-I*: maximum 2.5 per cent;
— *polymyxin 2,3-dehydro E1*: maximum 1.5 per cent;
— *sum of polymyxins E1, E2, E3, E4, E6, E1-7MOA, E1-I, E1-Nva, E2-I, E2-Val and 2,3-dehydro E1*: minimum 86.0 per cent;
— *disregard limit*: 0.35 per cent.

Related substances
Liquid chromatography (*2.2.29*) as described in the test for composition.

Limits:
— *impurity B*: maximum 4.0 per cent;
— *any other impurity*: for each impurity, maximum 2.5 per cent, and not more than 4 such impurities exceed 1.0 per cent;
— *total*: maximum 11.0 per cent;
— *disregard limit*: 0.35 per cent.

Sulfate
16.0 per cent to 18.0 per cent (dried substance).

Dissolve 0.250 g in 100 mL of *water R* and adjust to pH 11 with *concentrated ammonia R*. Add 10.0 mL of *0.1 M barium chloride* and about 0.5 mg of *phthalein purple R*. Titrate with *0.1 M sodium edetate*, adding 50 mL of *ethanol (96 per cent) R* when the colour of the solution begins to change and continuing the titration until the violet-blue colour disappears.

1 mL of *0.1 M barium chloride* is equivalent to 9.606 mg of SO_4.

Loss on drying (*2.2.32*)
Maximum 3.5 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.67 kPa for 3 h.

Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.0 g.

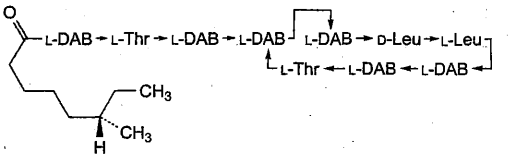
ASSAY
Carry out the microbiological assay of antibiotics (*2.7.2*).

STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *B*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A*.

A. unknown structure,

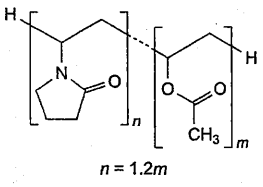


B. [N^4-Dab^5]polymyxin E1.

Ph Eur

Copovidone

(Ph. Eur. monograph 0891)



$n = 1.2m$

$(C_6H_9NO)_n$, $(C_4H_6O_2)_m$ M_r $(111.1)_n + (86.1)_m$ *25086-89-9*

Action and use
Excipient in pharmaceutical products.

Ph Eur

DEFINITION
Copovidone is a copolymer of 1-ethenylpyrrolidin-2-one and ethenyl acetate in the mass proportion 3:2.

Content
— nitrogen (N; A_r 14.01): 7.0 per cent to 8.0 per cent (dried substance),
— ethenyl acetate ($C_4H_6O_2$; M_r 86.10): 35.3 per cent to 42.0 per cent (dried substance).

K-value: 90.0 per cent to 110.0 per cent of the value stated on the label.

CHARACTERS
Aspect White or yellowish-white hygroscopic powder or flakes.

Solubility
Freely soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of copovidone.*

B. To 1 mL of solution S (see Tests) add 5 mL of *water R* and 0.2 mL of *0.05 M iodine*. A red colour appears.

C. Dissolve 0.7 g of *hydroxylamine hydrochloride R* in 10 mL of *methanol R*, add 20 mL of a 40 g/L solution of *sodium hydroxide R* and filter if necessary. To 5 mL of the solution add 0.1 g of the substance to be examined and boil for 2 min. Transfer 50 µL to a filter paper and add 0.1 mL of a mixture of equal volumes of *ferric chloride solution R1* and *hydrochloric acid R*. A violet colour appears.

TESTS

Solution S
Dissolve 10.0 g in *water R* and dilute to 100.0 mL with the same solvent. Add the substance to be examined to the *water R* in small portions with constant stirring.

Appearance of solution
Solution S is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution B_5, R_5 or BY_5 (*2.2.2, Method II*).

Viscosity, expressed as *K*-value
Dilute 5.0 mL of solution S to 50.0 mL with *water R*. Allow to stand for 1 h and determine the viscosity (*2.2.9*) of the solution at 25 ± 0.1 °C, using a size n° 1 viscometer with a minimum flow time of 100 s. Calculate the *K*-value using the following expression:

$$\frac{1.5\log_{10}\eta - 1}{0.15 + 0.003c} + \frac{\sqrt{300c\log_{10}\eta + (c + 1.5c\log_{10}\eta)^2}}{0.15c + 0.003c^2}$$

c = percentage concentration (g/100 mL) of the substance to be examined, calculated with reference to the dried substance;
η = viscosity of the solution relative to that of water.

Aldehydes
Maximum 500 ppm, expressed as acetaldehyde.

Test solution Dissolve 1.0 g of the substance to be examined in *phosphate buffer solution pH 9.0 R* and dilute to 100.0 mL with the same solvent. Stopper the flask and heat at 60 °C for 1 h. Allow to cool.

Reference solution Dissolve 0.140 g of *acetaldehyde ammonia trimer trihydrate R* in *water R* and dilute to 200.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *phosphate buffer solution pH 9.0 R*.

Into 3 identical spectrophotometric cells with a path length of 1 cm, introduce separately 0.5 mL of the test solution, 0.5 mL of the reference solution and 0.5 mL of *water R* (blank). To each cell add 2.5 mL of *phosphate buffer solution pH 9.0 R* and 0.2 mL of *nicotinamide-adenine dinucleotide solution R*. Mix and stopper tightly. Allow to stand at 22 ± 2 °C for 2-3 min and measure the absorbance (*2.2.25*) of each solution at 340 nm, using *water R* as the compensation liquid. To each cell, add 0.05 mL of *aldehyde dehydrogenase solution R*, mix and stopper tightly. Allow to stand at 22 ± 2 °C for 5 min. Measure the absorbance of each solution at 340 nm using *water R* as compensation liquid. Determine the content of aldehydes using the following expression:

$$\frac{(A_{t2} - A_{t1}) - (A_{b2} - A_{b1})}{(A_{s2} - A_{s1}) - (A_{b2} - A_{b1})} \times \frac{100\,000 \times C}{m}$$

A_{t1} = absorbance of the test solution before the addition of aldehyde dehydrogenase;
A_{t2} = absorbance of the test solution after the addition of aldehyde dehydrogenase;
A_{s1} = absorbance of the reference solution before the addition of aldehyde dehydrogenase;
A_{s2} = absorbance of the reference solution after the addition of aldehyde dehydrogenase;
A_{b1} = absorbance of the blank before the addition of aldehyde dehydrogenase;
A_{b2} = absorbance of the blank after the addition of aldehyde dehydrogenase;
m = mass of copovidone, in grams, calculated with reference to the dried substance;
C = concentration (mg/ml), of acetaldehyde in the reference solution, calculated from the weight of the acetaldehyde ammonia trimer trihydrate with the factor 0.72.

Peroxides
Maximum 400 ppm, expressed as H_2O_2.

Dilute 10 mL of solution S to 25 mL with *water R*. Add 2 mL of *titanium trichloride-sulfuric acid reagent R* and allow to stand for 30 min. The absorbance (*2.2.25*) of the solution, measured at 405 nm using a mixture of 25 mL of a 40 g/L solution of the substance to be examined and 2 mL of a 13 per cent *V/V* solution of *sulfuric acid R* as the compensation liquid, is not greater than 0.35.

Hydrazine
Thin-layer chromatography (*2.2.27*). *Use freshly prepared solutions.*

Test solution To 25 mL of solution S add 0.5 mL of a 50 g/L solution of *salicylaldehyde R* in *methanol R*, mix and heat in a water-bath at 60 °C for 15 min. Allow to cool, add 2.0 mL of *xylene R*, shake for 2 min and centrifuge. Use the clear supernatant layer.

Reference solution Dissolve 9 mg of *salicylaldehyde azine R* in *xylene R* and dilute to 100 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with *xylene R*.

Plate *TLC silanised silica gel plate R.*

Mobile phase *water R, methanol R* (20:80 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 365 nm.

Limit:

— *hydrazine*: any spot due to salicylaldehyde azine is not more intense than the spot in the chromatogram obtained with the reference solution (1 ppm).

Monomers

Maximum 0.1 per cent.

Dissolve 10.0 g in 30 mL of *methanol R* and add slowly 20.0 mL of *iodine bromide solution R*. Allow to stand for 30 min protected from light with repeated shaking. Add 10 mL of a 100 g/L solution of *potassium iodide R* and titrate with *0.1 M sodium thiosulfate* until a yellow colour is obtained. Continue titration dropwise until the solution becomes colourless. Carry out a blank titration. Not more than 1.8 mL of *0.1 M sodium thiosulfate* is used.

Impurity A

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 0.100 g of *2-pyrrolidone R* (impurity A) in *water R* and dilute to 100 mL with the same solvent. Dilute 1.0 mL to 100.0 mL with *water R*.

Precolumn:

— *size*: *l* = 0.025 m, Ø = 4 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Column:

— *size*: *l* = 0.25 m, Ø = 4 mm;

— *stationary phase*: *end-capped amidohexadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase *water R* adjusted to pH 2.4 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 205 nm. A detector is placed between the precolumn and the analytical column. A second detector is placed after the analytical column.

Injection 10 µL. When impurity A has left the precolumn (after about 1.2 min) switch the flow directly from the pump to the analytical column. Before the next chromatogram is run, wash the precolumn by reversed flow.

Limit:

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (0.5 per cent).

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Ethenyl acetate

Determine the saponification value (*2.5.6*) on 2.00 g of the substance to be examined. Multiply the result obtained by 0.1534 to obtain the percentage content of the ethenyl acetate component.

Nitrogen

Carry out the determination of nitrogen (*2.5.9*) using 30.0 mg of the substance to be examined and 1 g of a mixture of 3 parts of *copper sulfate pentahydrate R* and 997 parts of *dipotassium sulfate R*, heating until a clear, light green solution is obtained and then for a further 45 min.

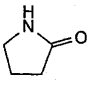
STORAGE

In an airtight container.

LABELLING

The label states the *K*-value.

IMPURITIES



A. pyrrolidin-2-one (2-pyrrolidone).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for copovidone used as binder in tablets and granules.

Viscosity (*2.2.9*)

Determine the dynamic viscosity using a capillary viscometer on a 10 per cent solution (dried substance) or on a 20 per cent solution (dried substance) at 25 °C. It is typically about 8 mPa·s or about 23 mPa·s, respectively.

Particle-size distribution (*2.9.31* or *2.9.38*)

Bulk and tapped density (*2.9.34*)

The following characteristic may be relevant for copovidone used as film former in coated dosage forms and in aerosols.

Viscosity (*2.2.9*)

See above.

Ph Eur

Copper Sulfate

Anhydrous Copper Sulfate

Anhydrous Copper Sulphate

(Ph. Eur. monograph 0893)

$CuSO_4$ 159.6 *7758-98-7*

Action and use
Used in treatment of copper deficiency.

Ph Eur

DEFINITION

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
Greenish-grey powder, very hygroscopic.

Solubility
Freely soluble in water, slightly soluble in methanol, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Add several drops of *dilute ammonia R2* to 1 mL of solution S (see Tests). A blue precipitate is formed. On further addition of *dilute ammonia R2* the precipitate dissolves and a dark blue colour is produced.

B. Loss on drying (see Tests).

C. Dilute 1 mL of solution S to 5 mL with *water R*. The solution gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S
Dissolve 1.6 g in *water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

Chlorides (*2.4.4*)
Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with *water R*.

Iron
Maximum 150 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 0.32 g in 10 mL of *water R*, add 2.5 mL of *lead-free nitric acid R* and dilute to 25.0 mL with *water R*.

Reference solutions Prepare the reference solutions using *iron standard solution (20 ppm Fe) R*, adding 2.5 mL of *lead-free nitric acid R* and diluting to 25.0 mL with *water R*.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Copper may form explosive acetylides with acetylene. Therefore, clean the burner thoroughly before any residues become dry.

Lead
Maximum 80 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 1.6 g in 10 mL of *water R*, add 2.5 mL of *lead-free nitric acid R* and dilute to 25.0 mL with *water R*.

Reference solutions Prepare the reference solutions using *lead standard solution (100 ppm Pb) R*, adding 2.5 mL of *lead-free nitric acid R* and diluting to 25.0 mL with *water R*.

Source Lead hollow-cathode lamp.

Wavelength 217.0 nm.

Atomisation device Air-acetylene flame.

Copper may form explosive acetylides with acetylene. Therefore, clean the burner thoroughly before any residues become dry.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 250 ± 10 °C.

ASSAY
Dissolve 0.125 g in 50 mL of *water R*. Add 2 mL of *sulfuric acid R* and 3 g of *potassium iodide R*. Titrate with *0.1 M sodium thiosulfate*, using 1 mL of *starch solution R*, added towards the end of the titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 15.96 mg of $CuSO_4$.

STORAGE
In an airtight container.

Ph Eur

Copper Sulfate Pentahydrate

Copper Sulphate Pentahydrate

(Ph. Eur. monograph 0894)

$CuSO_4,5H_2O$ 249.7 *7758-99-8*

Action and use
Used in treatment of copper deficiency.

Ph Eur

DEFINITION

Content
99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
Blue, crystalline powder or transparent, blue crystals.

Solubility
Freely soluble in water, soluble in methanol, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Add several drops of *dilute ammonia R2* to 1 mL of solution S (see Tests). A blue precipitate is formed. On further addition of *dilute ammonia R2* the precipitate dissolves and a dark blue colour is produced.

B. Loss on drying (see Tests).

C. Dilute 1 mL of solution S to 5 mL with *water R*. The solution gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S
Dissolve 5 g in *water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

Chlorides (*2.4.4*)
Maximum 100 ppm.

Dilute 10 mL of solution S to 15 mL with *water R*.

Iron
Maximum 100 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 0.5 g in 10 mL of *water R*, add 2.5 mL of *lead-free nitric acid R* and dilute to 25.0 mL with *water R*.

Reference solutions Prepare the reference solutions using *iron standard solution (20 ppm Fe) R*, adding 2.5 mL of *lead-free nitric acid R* and diluting to 25.0 mL with *water R*.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Copper may form explosive acetylides with acetylene. Therefore, clean the burner thoroughly before any residues become dry.

Lead

Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 2.5 g in 10 mL of *water R*, add 2.5 mL of *lead-free nitric acid R* and dilute to 25.0 mL with *water R*.

Reference solutions Prepare the reference solutions using *lead standard solution (100 ppm Pb) R*, adding 2.5 mL of *lead-free nitric acid R* and diluting to 25.0 mL with *water R*.

Source Lead hollow-cathode lamp.

Wavelength 217.0 nm.

Atomisation device Air-acetylene flame.

Copper may form explosive acetylides with acetylene. Therefore, clean the burner thoroughly before any residues become dry.

Loss on drying (*2.2.32*)

35.0 per cent to 36.5 per cent, determined on 0.500 g by drying in an oven at 250 ± 10 °C.

ASSAY

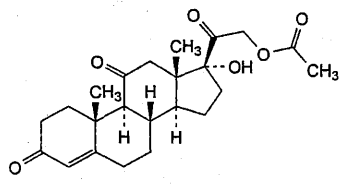
Dissolve 0.200 g in 50 mL of *water R*. Add 2 mL of *sulfuric acid R* and 3 g of *potassium iodide R*. Titrate with *0.1 M sodium thiosulfate*, adding 1 mL of *starch solution R* towards the end of the titration.

1 mL *0.1 M sodium thiosulfate* is equivalent to 24.97 mg of $CuSO_4,5H_2O$.

Ph Eur

Cortisone Acetate

(Ph. Eur. monograph 0321)



$C_{23}H_{30}O_6$ 402.5 *50-04-4*

Action and use

Corticosteroid.

Preparation

Cortisone Tablets

Ph Eur

DEFINITION

17-Hydroxy-3,11,20-trioxopregn-4-en-21-yl acetate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in dioxan, sparingly soluble in acetone, slightly soluble in ethanol (96 per cent) and in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, B.

Second identification: C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cortisone acetate CRS*.

If the spectra obtained in the solid state show differences, record new spectra using 50 g/L solutions in *methylene chloride R* in a 0.2 mm cell.

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *cortisone acetate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *hydrocortisone acetate R* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R*.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 25 mg of the substance to be examined in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of this solution to 10 mL with *methylene chloride R*.

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* briskly through the

solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C protected from light for 2.5 h. Allow to cool.

Reference solution (a) Dissolve 25 mg of *cortisone acetate CRS* in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of this solution to 10 mL with *methylene chloride R*.

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C protected from light for 2.5 h. Allow to cool.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R*.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R* and heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spots in the chromatograms obtained with test solution (b) and reference solution (b) have an R_F value distinctly lower than that of the principal spots in the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a faint yellow colour develops. Add this solution to 10 mL of *water R* and mix. The colour is discharged and a clear solution remains.

E. About 10 mg gives the reaction of acetyl (*2.3.1*).

TESTS

Specific optical rotation (*2.2.7*)

+ 211 to + 220 (dried substance).

Dissolve 0.250 g in *dioxan R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 25.0 mg of the substance to be examined in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of *cortisone acetate CRS* and 2 mg of *hydrocortisone acetate CRS* (impurity A) in *acetonitrile R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase In a 1000 mL volumetric flask mix 400 mL of *acetonitrile R* with 550 mL of *water R* and allow to equilibrate; dilute to 1000 mL with *water R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 30 min.

Injection 20 µL; inject *acetonitrile R* as a blank.

Run time Twice the retention time of cortisone acetate.

Retention time Impurity A = about 10 min; cortisone acetate = about 12 min.

System suitability Reference solution (a):

— *resolution*: minimum 4.2 between the peaks due to impurity A and cortisone acetate; if necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:

— *impurity A*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 237 nm.

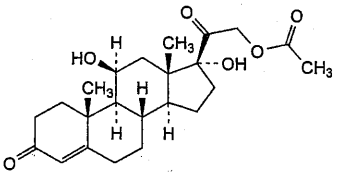
Calculate the content of $C_{23}H_{30}O_6$ taking the specific absorbance to be 395.

STORAGE

Protected from light.

IMPURITIES

Specified impurities *A*.



A. 11β,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate (hydrocortisone acetate).

Ph Eur

Hydrogenated Cottonseed Oil

(Ph. Eur. monograph 1305)

Ph Eur

DEFINITION

Product obtained by refining and hydrogenation of oil obtained from seeds of cultivated plants of various varieties of *Gossypium hirsutum* L. or of other species of *Gossypium*. The product consists mainly of triglycerides of palmitic and stearic acids.

CHARACTERS

Appearance
White or almost white mass or powder which melts to a clear, pale yellow liquid when heated.

Solubility
Practically insoluble in water, freely soluble in methylene chloride and in toluene, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Melting point (see Tests).

B. Composition of fatty acids (see Tests).

TESTS

Melting point (*2.2.14*)
57 °C to 70 °C.

Acid value (*2.5.1*)
Maximum 0.5.

Dissolve 10.0 g in 50 mL of a hot mixture of equal volumes of *ethanol (96 per cent) R* and *toluene R*, previously neutralised with *0.1 M potassium hydroxide* using 0.5 mL of *phenolphthalein solution R1* as indicator. Titrate the solution immediately while still hot.

Peroxide value (*2.5.5, Method A*)
Maximum 5.0.

Unsaponifiable matter (*2.5.7*)
Maximum 1.0 per cent, determined on 5.0 g.

Alkaline impurities
Dissolve by gentle heating 2.0 g in a mixture of 1.5 mL of *ethanol (96 per cent) R* and 3 mL of *toluene R*. Add 0.05 mL of a 0.4 g/L solution of *bromophenol blue R* in *ethanol (96 per cent) R*. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour to yellow.

Composition of fatty acids (*2.4.22, Method A*)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.25 mm;
— *stationary phase*: *poly(cyanopropyl)siloxane R* (film thickness 0.2 µm).

Carrier gas *helium for chromatography R.*

Flow rate 0.65 mL/min.

Split ratio 1:100.

Temperature:
— *column*: 180 °C for 35 min;
— *injection port and detector*: 250 °C.

Detection Flame ionisation.

Composition of the fatty-acid fraction of the oil:
— *saturated fatty acids of chain length less than* C_{14}: maximum 0.2 per cent;
— *myristic acid*: maximum 1.0 per cent;
— *palmitic acid*: 19.0 per cent to 26.0 per cent;
— *stearic acid*: 68.0 per cent to 80.0 per cent;
— *oleic acid and isomers*: maximum 4.0 per cent;
— *linoleic acid and isomers*: maximum 1.0 per cent;
— *arachidic acid*: maximum 1.0 per cent;
— *behenic acid*: maximum 1.0 per cent;
— *lignoceric acid*: maximum 0.5 per cent.

Nickel
Maximum 1 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Introduce 5.0 g into a platinum or silica crucible tared after ignition. Cautiously heat and introduce into the substance a wick formed from twisted ashless filter paper. Ignite the wick. When the substance ignites, stop heating. After combustion, ignite in a muffle furnace at about 600 ± 50 °C. Continue the incineration until white ash is obtained. After cooling, take up the residue with 2 quantities, each of 2 mL, of *dilute hydrochloric acid R* and transfer into a 25 mL graduated flask. Add 0.3 mL of *nitric acid R* and dilute to 25.0 mL with *distilled water R*.

Reference solutions Prepare 3 reference solutions by adding 1.0 mL, 2.0 mL and 4.0 mL of *nickel standard solution (0.2 ppm Ni) R* to 2.0 mL portions of the test solution, diluting to 10.0 mL with *distilled water R*.

Source Nickel hollow-cathode lamp.

Wavelength 232 nm.

Atomisation device Graphite furnace.

Carrier gas *argon R.*

STORAGE

Protected from light.

Ph Eur

Cresol

Action and use
Antiseptic; antimicrobial preservative.

DEFINITION

Cresol is a mixture of cresols and other phenols obtained from coal tar.

CHARACTERISTICS

An almost colourless to pale brownish yellow liquid.

Almost completely soluble in 50 volumes of *water*; freely soluble in *ethanol (96%)*, in *ether* and in fixed and volatile oils.

IDENTIFICATION

Shake 0.5 mL with 300 mL of *water* and filter. The filtrate complies with the following tests.

A. Add *iron(III) chloride solution R1*. A transient blue colour is produced.

B. Add *bromine water*. A pale yellow flocculent precipitate is produced.

TESTS

Acidity
A 2.0% w/v solution is neutral to *bromocresol purple solution*.

Distillation range
Not more than 2% v/v distils below 188° and not less than 80% v/v distils between 195° and 205°, Appendix V C.

Weight per mL
1.029 to 1.044 g, Appendix V G.

Hydrocarbons
Place 50 mL in a 500 mL round-bottomed flask, add 150 mL of 5M *sodium hydroxide* and 30 mL of *water* and mix thoroughly. Connect the flask to a splash-bulb and air-condenser about 60 cm long, with the end of the air-condenser fitting closely into the neck of a 250 mL pear-shaped separating funnel and passing well into the separating funnel, which has a cylindrical graduated portion above the stopcock. Fill the graduated portion of the separating funnel with *water*. Distil rapidly until 75 mL of distillate has been collected, cooling the separating funnel in running water if necessary. Allow the separating funnel to stand in a vertical

position until separation is complete and draw off the aqueous liquid into a titration flask for use in the test for Volatile bases.

Allow the separating funnel to stand for a few minutes, measure the volume of hydrocarbon oil in the graduated portion and warm, if necessary, to keep the oil in the liquid state. Subtract the volume of volatile bases in the hydrocarbon oil, as determined in the following test. Not more than 0.15% v/v of hydrocarbon oil is present.

Volatile bases

To the aqueous liquid reserved in the test for Hydrocarbons add any aqueous liquid still remaining in the separating funnel and neutralise, if necessary, with 0.1M *hydrochloric acid* using *phenolphthalein solution R1* as indicator. Titrate with *1M hydrochloric acid VS* using *methyl orange solution* as indicator. Wash the oil from the separating funnel into the titration flask with *water* and again titrate with *1M hydrochloric acid VS*. From the volume of additional *1M hydrochloric acid VS*, calculate the proportion of volatile bases in the hydrocarbon oil. From the total volume of *1M hydrochloric acid VS* used in both titrations calculate the volume of volatile bases in the substance being examined. Each mL of *1M hydrochloric acid VS* is equivalent to 0.080 mL of volatile bases. Not more than 0.15% v/v of volatile bases is present.

Hydrocarbons and volatile bases

The sum of the contents of hydrocarbon oil and volatile bases, as determined in the tests for Hydrocarbons and for Volatile bases, does not exceed 0.25% v/v.

Sulfur compounds

Place 20 mL in a small conical flask and over the mouth of the flask fix a piece of filter paper moistened with a 10% w/v solution of *lead(II) acetate*. Heat the flask on a water bath for 5 minutes. Not more than a light yellow colour is produced on the filter paper.

Non-volatile matter

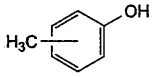
When evaporated on a water bath and dried at 105°, leaves not more than 0.1% w/v of residue.

STORAGE

Cresol should be protected from light. It darkens with age or on exposure to light.

Crude Cresol

(Ph. Eur. monograph 1628)



C_7H_8O 108.1

Action and use

Antiseptic.

Ph Eur

DEFINITION

Mixture of 2-, 3- and 4-methylphenol.

CHARACTERS

Appearance

Colourless or pale brown liquid.

Solubility

Sparingly soluble in water, miscible with alcohol and with methylene chloride.

IDENTIFICATION

A. To 0.5 mL add 300 mL of *water R*, mix and filter. To 10 mL of the filtrate add 1 mL of *ferric chloride solution R1*. A blue colour is produced.

B. To 10 mL of the filtrate obtained in identification test A, add 1 mL of *bromine water R*. A pale yellow flocculent precipitate is produced.

C. Relative density (see Tests).

TESTS

Solution S

To 2.5 g of the substance to be examined add 50 mL of *water R*, shake for 1 min and filter through a moistened filter.

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.3 mL of *0.01 M hydrochloric acid*. The solution is red.

Relative density *(2.2.5)*

1.029 to 1.044.

Distillation range *(2.2.11)*

A maximum of 2.0 per cent *V/V* distils below 188 °C and a minimum of 80 per cent *V/V* distils between 195 °C and 205 °C.

Sulfur compounds

Place 20 mL in a small conical flask. Over the mouth of the flask fix a piece of filter paper moistened with *lead acetate solution R*. Heat on a water-bath for 5 min. Not more than a light yellow colour is produced on the filter paper.

Residue on evaporation

Maximum 0.1 per cent.

Evaporate 2.0 g to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs not more than 2 mg.

STORAGE

Protected from light.

Ph Eur

Croscarmellose Sodium[1]

(Ph. Eur. monograph 0985)

Action and use

Excipient.

Ph Eur

DEFINITION

Cross-linked sodium carboxymethylcellulose.

Sodium salt of a cross-linked, partly *O*-carboxymethylated cellulose.

♦CHARACTERS

Appearance

White or greyish-white, hygroscopic powder.

Solubility

Practically insoluble in acetone, in anhydrous ethanol and in toluene.♦

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

IDENTIFICATION

A. Mix 1 g with 100 mL of a solution containing 4 ppm of *methylene blue R*, stir the mixture and allow it to settle. The substance to be examined absorbs the methylene blue and settles as a blue, fibrous mass.

B. Mix 1 g with 50 mL of *water R*. Transfer 1 mL of the mixture to a small test-tube and add 1 mL of *water R* and 0.05 mL of a freshly prepared 40 g/L solution of *α-naphthol R* in *methanol R*. Incline the test-tube and carefully add 2 mL of *sulfuric acid R* down the side so that it forms a lower layer. A reddish-violet colour develops at the interface.

♦C. To the residue obtained in the test for sulfated ash add 1 mL of *hydrochloric acid R* and evaporate on a water-bath. Take up the residue in 20 mL of *water R*. The solution gives reaction (a) of sodium (*2.3.1*).♦

TESTS

pH (*2.2.3*)

5.0 to 7.0 for the suspension.

Shake 1 g with 100 mL of *carbon dioxide-free water R* for 5 min.

♦Sodium chloride and sodium glycollate

Maximum 0.5 per cent (dried substance) for the sum of the percentage contents of sodium chloride and sodium glycollate.

Sodium chloride Place 5.00 g in a 250 mL conical flask, add 50 mL of *water R* and 5 mL of *strong hydrogen peroxide solution R* and heat on a water-bath for 20 min, stirring occasionally to ensure total hydration. Cool and add 100 mL of *water R* and 10 mL of *nitric acid R*. Titrate with *0.05 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*) using a silver indicator electrode and a double-junction reference electrode containing a 100 g/L solution of *potassium nitrate R* in the outer jacket and a standard filling solution in the inner jacket, and stirring constantly.

1 mL of *0.05 M silver nitrate* is equivalent to 2.922 mg of NaCl.

Sodium glycollate Place a quantity of the substance to be examined equivalent to 0.500 g of the dried substance in a 100 mL beaker. Add 5 mL of *glacial acetic acid R* and 5 mL of *water R* and stir to ensure total hydration (about 15 min). Add 50 mL of *acetone R* and 1 g of *sodium chloride R*. Stir for several minutes to ensure complete precipitation of the carboxymethylcellulose. Filter through a fast filter paper impregnated with *acetone R* into a volumetric flask, rinse the beaker and the filter with 30 mL of *acetone R* and dilute the filtrate to 100.0 mL with the same solvent. Allow to stand for 24 h without shaking. Use the clear supernatant to prepare the test solution.

Prepare the reference solutions as follows: in a 100 mL volumetric flask, dissolve 0.100 g of *glycollic acid R*, previously dried *in vacuo* over *diphosphorus pentoxide R* at room temperature overnight, in *water R* and dilute to 100.0 mL with the same solvent; use the solution within 30 days; transfer 1.0 mL, 2.0 mL, 3.0 mL and 4.0 mL of the solution to separate volumetric flasks, dilute the contents of each flask to 5.0 mL with *water R*, add 5 mL of *glacial acetic acid R*, dilute to 100.0 mL with *acetone R* and mix.

Transfer 2.0 mL of the test solution and 2.0 mL of each of the reference solutions to separate 25 mL volumetric flasks. Heat the uncovered flasks for 20 min on a water-bath to eliminate acetone. Allow to cool and add 5.0 mL of *2,7-dihydroxynaphthalene solution R* to each flask. Mix, add a further 15.0 mL of *2,7-dihydroxynaphthalene solution R* and mix again. Close the flasks with aluminium foil and heat on a water-bath for 20 min. Cool and dilute to 25.0 mL with *sulfuric acid R*.

Measure the absorbance (*2.2.25*) of each solution at 540 nm. Prepare a blank using 2.0 mL of a solution containing 5 per cent *V/V* each of *glacial acetic acid R* and *water R* in *acetone R*. Prepare a standard curve using the absorbances obtained with the reference solutions. From the standard curve and the absorbance of the test solution, determine the mass (*a*) of glycollic acid in the substance to be examined, in milligrams, and calculate the content of sodium glycollate using the following expression:

$$\frac{10 \times 1.29 \times a}{(100 - b)m}$$

1.29 = the factor converting glycollic acid to sodium glycollate;
b = loss on drying as a percentage;
m = mass of the substance to be examined, in grams.

Water-soluble substances

Maximum 10.0 per cent.

Disperse 10.00 g in 800.0 mL of *water R* and stir for 1 min every 10 min during the first 30 min. Allow to stand for 1 h and centrifuge if necessary. Decant 200.0 mL of the supernatant liquid onto a fast filter paper in a vacuum filtration funnel, apply vacuum and collect 150.0 mL of the filtrate. Evaporate to dryness and dry the residue at 100-105 °C for 4 h.♦

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 6 h.

Sulfated ash (*2.4.14*)

14.0 per cent to 28.0 per cent (dried substance), determined on 1.0 g.

♦Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).♦

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for croscarmellose sodium used as disintegrant.

Settling volume

Place 75 mL of *water R* in a 100 mL graduated cylinder and add 1.5 g of the substance to be examined in 0.5 g portions, shaking vigorously after each addition. Dilute to 100.0 mL with *water R* and shake again until the substance is homogeneously distributed. Allow to stand for 4 h.
The settling volume is between 10.0 mL and 30.0 mL.

Degree of substitution

0.60 to 0.85 (dried substance).

Place 1.000 g in a 500 mL conical flask, add 300 mL of a 100 g/L solution of *sodium chloride R* and 25.0 mL of *0.1 M sodium hydroxide*, stopper the flask and allow to stand for 5 min, shaking occasionally. Add 0.25 mL of *m-cresol purple solution R* and about 15 mL of *0.1 M hydrochloric acid* from a burette. Insert the stopper and shake. If the solution is violet, add *0.1 M hydrochloric acid* in 1 mL portions until the solution becomes yellow, shaking after each addition. Titrate with *0.1 M sodium hydroxide* until the colour turns to violet.

Calculate the number of milliequivalents (*M*) of base required to neutralise the equivalent of 1 g of dried substance.

Calculate the degree of acid carboxymethyl substitution (*A*) using the following expression:

$$\frac{1150M}{(7102 - 412M - 80C)}$$

C = sulfated ash as a percentage.

Calculate the degree of sodium carboxymethyl substitution (*S*) using the following expression:

$$\frac{(162 + 58A)C}{(7102 - 80C)}$$

The degree of substitution is the sum of *A* and *S*.

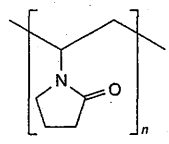
Particle size distribution (*2.9.31* or *2.9.38*)

Hausner ratio (*2.9.36*)

Ph Eur

Crospovidone[1]

(Ph. Eur. monograph 0892)



$(C_6H_9NO)_n$ M_r (111.1)$_n$ *9003-39-8*

Action and use
Excipient in pharmaceutical products.

Ph Eur

DEFINITION
Cross-linked homopolymer of 1-ethenylpyrrolidin-2-one.

Content
11.0 per cent to 12.8 per cent of N (A_r 14.01) (dried substance).

2 types of crospovidone are available, depending on the particle size: type A and type B.

♦CHARACTERS
Appearance
Hygroscopic, white or yellowish-white powder or flakes.

Solubility
Practically insoluble in water, in ethanol 96 per cent and in methylene chloride.♦

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

IDENTIFICATION
♦A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison crospovidone CRS.♦

B. Suspend 1 g in 10 mL of *water R*, add 0.1 mL of *0.05 M iodine* and shake for 30 s. Add 1 mL of *starch solution R* and shake. No blue colour develops within 30 s.

C. To 10 mL of *water R*, add 0.1 g and shake. A suspension is formed and no clear solution is obtained within 15 min.

D. *The analytical sieves must be clean and dry. For this purpose the sieves are washed in hot water and allowed to dry overnight in a drying cabinet at 105 °C.*

Place 20 g (dried substance) in a 1000 mL conical flask, add 500 mL of *water R* and shake the suspension for 30 min. Pour the suspension through a 63 µm analytical sieve, previously tared, and rinse the sieve with *water R* until the filtrate is clear. Dry the sieve and sample residue at 105 °C for 5 h in a drying cabinet without circulating air. Cool in a desiccator for 30 min and weigh.

Calculate the percentage sieving residue (fraction of sample particles having a diameter of more than 63 µm), using the following expression:

$$\frac{m_1 - m_3}{m_2} \times 100$$

m_1 = mass of the sieve and sample residue, after drying for 5 h, in grams;
m_2 = initial mass of the sample, in grams;
m_3 = mass of the sieve, in grams.

If the sieving residue fraction is more than 15 per cent, the substance is classified as type A; if the sieving residue fraction is less than or equal to 15 per cent, the substance is classified as type B.

TESTS
Peroxides
Type A: maximum 400 ppm expressed as H_2O_2; type B: maximum 1000 ppm expressed as H_2O_2.

Suspend 2.0 g in 50 mL of *water R*. To 25 mL of this suspension add 2 mL of *titanium trichloride-sulfuric acid reagent R*. Allow to stand for 30 min and filter.
The absorbance (*2.2.25*) of the filtrate, measured at 405 nm using a mixture of 25 mL of a filtered 40 g/L suspension of the substance to be examined and 2 mL of a 13 per cent *V/V* solution of *sulfuric acid R* as the compensation liquid, has a maximum of 0.35.

For type B use 10 mL of the suspension and dilute to 25 mL with *water R* for the test.

Water-soluble substances
Maximum 1.5 per cent.

Place 25.0 g in a 400 mL beaker, add 200 mL of *water R* and stir for 1 h using a magnetic stirrer. Transfer the suspension to a 250.0 mL volumetric flask, rinsing with *water R*, and dilute to volume with the same solvent. Allow the bulk of the solids to settle. Filter about 100 mL of the almost clear supernatant through a membrane filter (nominal pore size 0.45 µm), protected by superimposing a membrane filter (nominal pore size 3 µm). While filtering, stir the liquid above the membrane filter manually or by means of a mechanical stirrer, taking care not to damage the membrane filter. Transfer 50.0 mL of the clear filtrate to a tared 100 mL beaker, evaporate to dryness and dry at 105-110 °C for 3 h. The residue weighs a maximum of 75 mg.

Impurity A
Liquid chromatography (*2.2.29*).

Test solution Suspend 1.250 g in 50.0 mL of *methanol R* and shake for 60 min. Leave the bulk to settle and filter through a membrane filter (nominal pore size 0.2 µm).

Reference solution (a) Dissolve 50 mg of *1-vinylpyrrolidin-2-one R* (impurity A) in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *methanol R*. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *1-vinylpyrrolidin-2-one R* (impurity A) and 0.50 g of *vinyl acetate R* in *methanol R* and dilute to 100 mL with the same solvent. Dilute 1 mL of the solution to 100 mL with the mobile phase.

Precolumn:
— *size*: *l* = 0.025 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R* (5 µm);
— *temperature*: 40 °C.

Mobile phase *acetonitrile for chromatography R, water for chromatography R* (10:90 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 50 µL. After each injection of the test solution, wash the precolumn by passing the mobile phase backwards, at the same flow rate as applied in the test, for 30 min.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to impurity A and vinyl acetate in the chromatogram obtained with reference solution (b);
— *repeatability*: maximum relative standard deviation of 2.0 per cent determined on 6 injections of reference solution (a).

Calculation of percentage content:
— for impurity A, use the concentration of impurity A in reference solution (a).

Limit:
— *impurity A*: maximum 10 ppm.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Place 0.100 g of the substance to be examined (*m* mg) in a combustion flask and add 5 g of a mixture of 1 g of *copper sulfate pentahydrate R*, 1 g of *titanium dioxide R* and 33 g of *dipotassium sulfate R*, and 3 glass beads. Wash any adhering particles from the neck into the flask with a small quantity of *water R*. Add 7 mL of *sulfuric acid R*, allowing it to run down the inside wall of the flask. Gradually heat the flask until the solution has a clear, yellowish-green colour, and the inside wall of the flask is free from carbonised material, and then heat for a further 45 min. After cooling, cautiously add 20 mL of *water R*, and connect the flask to the distillation apparatus, which has been previously washed by passing steam through it. To the absorption flask add 30 mL of a 40 g/L solution of *boric acid R*, 0.15 mL of *bromocresol green-methyl red solution R* and sufficient *water R* to immerse the lower end of the condenser tube. Add 30 mL of *strong sodium hydroxide solution R* through a funnel, cautiously rinse the funnel with 10 mL of *water R*, immediately close the clamp attached to the rubber tube, then start the distillation with steam to obtain 80-100 mL of distillate. Remove the absorption flask from the lower end of the condenser tube, rinsing the end part with a small quantity of *water R*, and titrate the distillate with *0.025 M sulfuric acid* until the colour of the solution changes from green through pale greyish-blue to pale greyish red-purple. Carry out a blank determination and make any necessary correction.

1 mL of *0.025 M sulfuric acid* is equivalent to 0.700 mg of N.

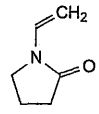
STORAGE

In an airtight container.

♦LABELLING

The label states the type of crospovidone (type A or type B).♦

IMPURITIES



A. 1-ethenylpyrrolidin-2-one (1-vinylpyrrolidin-2-one).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for crospovidone used as disintegrant.

Hydration capacity
Introduce 2.0 g into a 100 mL centrifuge tube and add 40 mL of *water R*. Shake vigorously until a suspension is obtained. Shake again 5 min and 10 min later, then centrifuge for 15 min at 750 *g*. Decant the supernatant and weigh the residue. The hydration capacity is the ratio of the mass of the residue to the initial mass of the sample. It is typically 3 to 9.

Particle-size distribution (*2.9.31*)

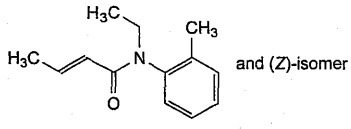
Powder flow (*2.9.36*)
The following characteristic may be relevant for crospovidone used as suspension stabiliser.

Settling volume
Introduce 10 g into a 100 mL graduated cylinder and add 90 mL of *water R*. Shake vigorously. Dilute to 100 mL with *water R*, washing the powder residues from the walls of the cylinder. Allow to stand for 24 h, then read the volume of the sediment. It is typically greater than 60 mL.

Ph Eur

Crotamiton

(Ph. Eur. monograph 1194)



$C_{13}H_{17}NO$ 203.3 *483-63-6*

Action and use
Acaricide.

Preparations
Crotamiton Cream
Crotamiton Lotion

Ph Eur

DEFINITION

N-Ethyl-*N*-(2-methylphenyl)but-2-enamide.

Content
— *sum of the (E)- and (Z)-isomers*: 96.0 per cent to 102.0 per cent;
— *(Z)-isomer*: maximum 15.0 per cent.

CHARACTERS

Appearance
Colourless or pale yellow, oily liquid.

Solubility
Slightly soluble in water, miscible with ethanol (96 per cent).

At low temperatures it may partly or completely solidify.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 25.0 mg in *cyclohexane R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *cyclohexane R*.

Spectral range 220-300 nm.

Absorption maximum At 242 nm.

Specific absorbance at the absorption maximum 300 to 330.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *crotamiton CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *anhydrous ethanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 25 mg of *crotamiton CRS* in *anhydrous ethanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase Shake 98 volumes of *methylene chloride R* with 2 volumes of *concentrated ammonia R*, dry over *anhydrous sodium sulfate R*, filter and mix 97 volumes of the filtrate with 3 volumes of *2-propanol R*.

Application 5 µL.

Development Over a 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 10 mL of a saturated solution add a few drops of a 3 g/L solution of *potassium permanganate R*. A brown colour is produced and a brown precipitate is formed on standing.

TESTS

Relative density (*2.2.5*)
1.006 to 1.011.

Refractive index (*2.2.6*)
1.540 to 1.542.

Free amines
Maximum 500 ppm, expressed as ethylaminotoluene.

Dissolve 5.00 g in 16 mL of *methylene chloride R* and add 4.0 mL of *glacial acetic acid R*. Add 0.1 mL of *metanil yellow solution R* and 1.0 mL of *0.02 M perchloric acid*. The solution is red-violet.

Chlorides
Maximum 100 ppm.

Boil 5.0 g under a reflux condenser for 1 h with 25 mL of *ethanol (96 per cent) R* and 5 mL of a 200 g/L solution of *sodium hydroxide R*. Cool, add 5 mL of *water R* and shake with 25 mL of *ether R*. Dilute the lower layer to 20 mL with *water R*; add 5 mL of *nitric acid R*, dilute to 50 mL with *water R* and add 1 mL of a freshly prepared 50 g/L solution of *silver nitrate R*. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of a freshly prepared 50 g/L solution of *silver nitrate R* and a solution prepared by diluting 5 mL of a 200 g/L solution of *sodium hydroxide R* to 20 mL with *water R* and adding 1.5 mL of *0.01 M hydrochloric acid*, 5 mL of *nitric acid R* and diluting to 50 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *crotamiton CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 15.0 mg of *crotamiton impurity A CRS* in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve 15 mg of *crotamiton impurity A CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with test solution (a).

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *tetrahydrofuran R, cyclohexane R* (8:92 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 242 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (c) and (d).

Run time 2.5 times the retention time of the (*E*)-isomer.

Relative retention With reference to the (*E*)-isomer: (*Z*)-isomer = about 0.5; impurity A = about 0.8.

System suitability Reference solution (d):
— *resolution*: minimum 4.5 between the peaks due to impurity A and the (*E*)-isomer.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the sum of the areas of the peaks due to the (*Z*)- and (*E*)- isomers in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *sum of impurities other than A*: not more than the sum of the areas of the peaks due to the (*Z*)- and (*E*)-isomers in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: 0.02 times the sum of the areas of the peaks due to the (*Z*)- and (*E*)-isomers in the chromatogram obtained with reference solution (c) (0.02 per cent).

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

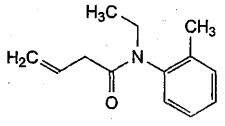
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{13}H_{17}NO$ from the sum of the areas of the peaks due to the (*Z*)- and (*E*)-isomers in the chromatograms obtained. Calculate the content of the (*Z*)-isomer, as a percentage of the total content of the (*E*)- and (*Z*)-isomers, from the chromatogram obtained with test solution (b).

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A*.

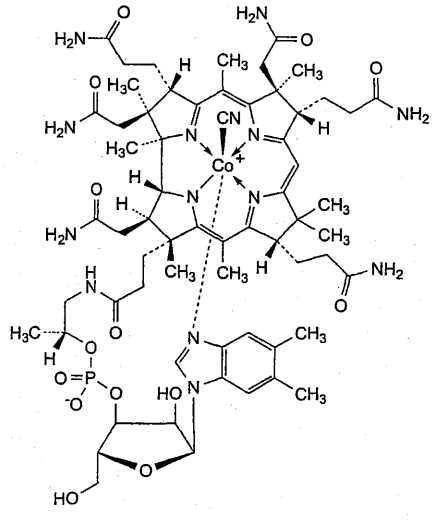


A. *N*-ethyl-*N*-(2-methylphenyl)but-3-enamide.

Ph Eur

Cyanocobalamin

(Ph. Eur. monograph 0547)



$C_{63}H_{88}CoN_{14}O_{14}P$ 1355 *68-19-9*

Action and use
Vitamin B12 analogue.

Preparations
Cyanocobalamin Oral Solution
Cyanocobalamin Tablets

Ph Eur

DEFINITION
α-(5,6-Dimethylbenzimidazol-1-yl)cobamide cyanide.

Content
96.0 per cent to 102.0 per cent (dried substance).

This monograph applies to cyanocobalamin produced by fermentation.

CHARACTERS

Appearance
Dark red, crystalline powder or dark red crystals.

Solubility
Sparingly soluble in water and in ethanol (96 per cent), practically insoluble in acetone.

The anhydrous substance is very hygroscopic.

IDENTIFICATION
A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 2.5 mg in *water R* and dilute to 100.0 mL with the same solvent.

Spectral range 260-610 nm.

Absorption maxima At 278 nm, 361 nm and from 547 nm to 559 nm.

Absorbance ratio:
— $A_{361} / A_{547\text{-}559}$ = 3.15 to 3.45;
— A_{361} / A_{278} = 1.70 to 1.90.

B. Thin-layer chromatography (*2.2.27*). *Carry out the test protected from light.*

Test solution Dissolve 2 mg of the substance to be examined in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Reference solution Dissolve 2 mg of *cyanocobalamin CRS* in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Plate *TLC silica gel G plate R.*

Mobile phase *dilute ammonia R1, methanol R, methylene chloride R* (9:30:45 *V/V/V*).

Application 10 µL.

Development In an unsaturated tank, over 2/3 of the plate.

Drying In air.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase. *Use within 1 h.*

Reference solution (a) Dilute 3.0 mL of the test solution to 100.0 mL with the mobile phase. *Use within 1 h.*

Reference solution (b) Dilute 5.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. *Use within 1 h.*

Reference solution (c) Dissolve 25 mg of the substance to be examined in 10 mL of *water R*, warming if necessary. Allow to cool and add 5 mL of a 1.0 g/L solution of *chloramine R* and 0.5 mL of *0.05 M hydrochloric acid*, then dilute to 25 mL with *water R*. Shake and allow to stand for 5 min. Dilute 1 mL of this solution to 10 mL with the mobile phase and inject immediately.

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 26.5 volumes of *methanol R* and 73.5 volumes of a 10 g/L solution of *disodium hydrogen phosphate dodecahydrate R* adjusted to pH 3.5 with *phosphoric acid R* and use within 2 days.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 361 nm.

Injection 20 µL.

Run time 3 times the retention time of cyanocobalamin.

System suitability:
- the chromatogram obtained with reference solution (c) shows 2 principal peaks;
- *resolution*: minimum 2.5 between the 2 principal peaks in the chromatogram obtained with reference solution (c);
- *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
- *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (3 per cent);
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying (*2.2.32*)

Maximum 12.0 per cent, determined on 40.00 mg by drying *in vacuo* at 105 °C for 2 h.

ASSAY

Dissolve 100.0 mg in *water R* and dilute to 500.0 mL with the same solvent. Dilute 25.0 mL of the solution to 200.0 mL with *water R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 361 nm.

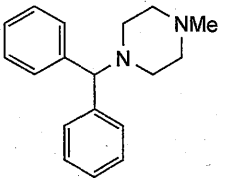
Calculate the content of $C_{63}H_{88}CoN_{14}O_{14}P$ taking the specific absorbance to be 207.

STORAGE

In an airtight container, protected from light.

Ph Eur

Cyclizine



$C_{18}H_{22}N_2$ 266.4 *82-92-8*

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Cyclizine Injection

DEFINITION

Cyclizine is 1-benzhydryl-4-methylpiperazine. It contains not less than 98.5% and not more than 101.0% of $C_{18}H_{22}N_2$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or creamy white, crystalline powder.

Practically insoluble in *water*. It dissolves in most organic solvents and in dilute acids.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of cyclizine *(RS 075)*.

B. *Melting point*, about 107°, Appendix V A.

TESTS

Alkalinity

Shake 1 g with 25 mL of *carbon dioxide-free water* for 5 minutes and filter. The pH of the filtrate is 7.6 to 8.6, Appendix V L.

Clarity of solution

A 1.0% w/v solution in *ether* and a 1.0% w/v solution in 2M *hydrochloric acid* are *clear*, Appendix IV A.

Chloride

Dissolve 0.20 g in 2 mL of *methanol* and dilute to 30 mL with 2M *nitric acid*. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (500 ppm).

Related substances

Carry out the method for *gas chromatography*, Appendix III B, using the following solutions in *methanol* prepared immediately before use.

(1) 0.5% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 100 volumes and further dilute 1 volume of the resulting solution to 10 volumes.

(3) 0.0025% w/v of *cyclizine hydrochloride BPCRS*, 0.0025% w/v of *1-methylpiperazine BPCRS* (impurity A) and 0.0025% w/v of *diphenylmethanol BPCRS* (impurity B).

CHROMATOGRAPHIC CONDITIONS

(a) Use a fused silica column (25 m × 0.33 mm) coated with a 0.5-µm film of *poly(dimethyl)(diphenyl)siloxane* (HP-5 is suitable).

(b) Use *helium* as the carrier gas at a flow rate of 1 mL per minute.

(c) Use the gradient conditions described below.

(d) Use a split injection ratio of 1:25.

(e) Use a flame ionisation detector at 290°.

(f) Inject 1 µL of each solution.

(g) The peaks elute in the order: methanol, 1-methylpiperazine, diphenylmethanol, cyclizine.

Time (minutes)	Temperature	Comment
0→14	100°→240°	linear gradient
14→16	240°→270°	linear gradient
16→30	270°	isocratic

SYSTEM SUITABILITY

Inject solution (3) 6 times. The relative standard deviation of each of the areas of the 3 principal peaks is not more than 5.0%.

The test is not valid unless, in the chromatogram obtained with solution (3);

the *peak-to-valley ratio* between methanol and 1-methylpiperazine (impurity A) is at least 50;

the *resolution factor* between diphenylmethanol (impurity B) and cyclizine is at least 18.

LIMITS

In the chromatogram obtained with solution (1):

the area of the peak corresponding to 1-methylpiperazine (impurity A) is not greater than the peak corresponding to 1-methylpiperazine in solution (3) (0.5%);

the area of the peak corresponding to diphenylmethanol (impurity B) is not greater than the peak corresponding to diphenylmethanol in solution (3) (0.5%);

the area of any other *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.1%);

the sum of the areas of all *secondary peaks* is not greater than 10 times the area of the principal peak in the chromatogram obtained with solution (2) (1.0%).

Disregard any peak with an area less than 0.5 times that of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Loss on drying

When dried to constant weight at 80°, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

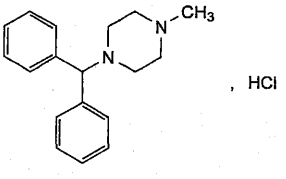
Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.1 g and determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 13.32 mg of $C_{18}H_{22}N_2$.

Cyclizine Hydrochloride

(Ph. Eur. monograph 1092)



$C_{18}H_{23}ClN_2$ 302.8 *305-25-3*

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Cyclizine Tablets

Ph Eur

DEFINITION

1-(Diphenylmethyl)-4-methylpiperazine hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution (a) Dissolve 20.0 mg in a 5 g/L solution of *sulfuric acid R* and dilute to 100.0 mL with the same acid solution.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with a 5 g/L solution of *sulfuric acid R*.

Spectral range 240-350 nm for test solution (a); 210-240 nm for test solution (b).

Resolution (*2.2.25*): minimum 1.7.

Absorption maxima At 258 nm and 262 nm for test solution (a); at 225 nm for test solution (b).

Absorbance ratio A_{262}/A_{258} = 1.0 to 1.1.

Specific absorbance at the absorption maximum at 225 nm 370 to 410 for test solution (b).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cyclizine hydrochloride CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *cyclizine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *concentrated ammonia R, methanol R, methylene chloride R* (2:13:85 *V/V/V*).

Application 20 µL.

Development Over 2/3 of the plate.

Drying In air for 30 min.

Detection Expose to iodine vapour for 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 0.5 g in 10 mL of *ethanol (60 per cent V/V) R*, heating if necessary. Cool in iced water. Add 1 mL of *dilute sodium hydroxide solution R* and 10 mL of *water R*. Filter, wash the precipitate with *water R* and dry at 60 °C at a pressure not exceeding 0.7 kPa for 2 h. The melting point (*2.2.14*) is 105 °C to 108 °C.

E. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)

4.5 to 5.5.

Dissolve 0.5 g in a mixture of 40 volumes of *ethanol (96 per cent) R* and 60 volumes of *carbon dioxide-free water R* and dilute to 25 mL with the same mixture of solvents.

Related substances

Gas chromatography (*2.2.28*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.250 g of the substance to be examined in 4.0 mL of *methanol R* and dilute to 5.0 mL with *1 M sodium hydroxide*.

Reference solution (a) Dissolve 25 mg of the substance to be examined in 10.0 mL of *methanol R*. Dilute 1.0 mL of this solution to 50.0 mL with *methanol R*.

Reference solution (b) Dissolve 5 mg of the substance to be examined, 5.0 mg of *cyclizine impurity A CRS* and 5.0 mg of *cyclizine impurity B CRS* in *methanol R* and dilute to 20.0 mL with the same solvent.

Column:

— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.33 mm;
— *stationary phase*: *poly(dimethyl) (diphenyl)siloxane R* (film thickness 0.50 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1.0 mL/min.

Split ratio 1:25.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 14	100 → 240
	14 - 16	240 → 270
	16 - 30	270
Injection port		250
Detector		290

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to cyclizine (retention time = about 15 min): impurity A = about 0.2; impurity B = about 0.7.

System suitability Reference solution (b):

— *peak-to-valley ratio*: minimum 50, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to methanol.

Limits:

— *impurities A, B*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 130 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.120 g in 15 mL of *anhydrous formic acid R* and add 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

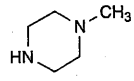
1 mL of *0.1 M perchloric acid* is equivalent to 15.14 mg of $C_{18}H_{23}ClN_2$.

STORAGE

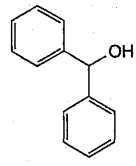
Protected from light.

IMPURITIES

Specified impurities *A, B.*



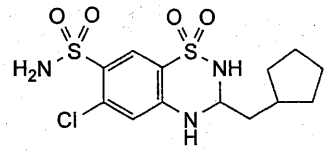
A. 1-methylpiperazine,



B. diphenylmethanol (benzhydrol).

Ph Eur

Cyclopenthiazide



$C_{13}H_{18}ClN_3O_4S_2$ 379.9 *742-20-1*

Action and use

Thiazide-diuretic.

DEFINITION

Cyclopenthiazide is 6-chloro-3-cyclopentylmethyl-3,4-dihydro-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide. It contains not less than 98.0% and not more than 102.0% of $C_{13}H_{18}ClN_3O_4S_2$, calculated with reference to the dried substance.

CHARACTERISTICS

A white powder.

Practically insoluble in *water*; soluble in *acetone* and in *ethanol (96%)*; very slightly soluble in *ether*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of cyclopenthiazide *(RS 077)*.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in 0.01M *sodium hydroxide* exhibits two maxima, at 273 nm and 320 nm. The *absorbance* at 273 nm is about 0.88 and at 320 nm is about 0.12.

C. Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel* GF_{254} as the coating substance and *ethyl acetate* as the mobile phase. Apply separately to the plate 5 µL of each of two solutions in *acetone* containing (1) 0.1% w/v of the substance being examined and (2) 0.1% w/v of *cyclopenthiazide BPCRS*. After removal of the plate, dry it in a current of air, examine under *ultraviolet light (254 nm)* and then reveal the spots by *Method I*. By each method of visualisation the principal spot in the chromatogram obtained with solution (1) corresponds in colour and intensity to that in the chromatogram obtained with solution (2).

TESTS

Related substances

Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and *ethyl acetate* as the mobile phase. Apply separately to the plate 5 µL of each of two solutions of the substance being examined in *acetone* containing (1) 0.50% w/v and (2) 0.0050% w/v. After removal of the plate, dry it in a current of air and reveal the spots by *Method I*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

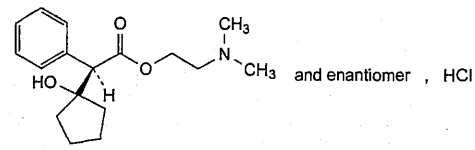
Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.5 g in 50 mL of *butylamine* and carry out Method II for *non-aqueous titration*, Appendix VIII A, using 0.1M *tetrabutylammonium hydroxide VS* as titrant and *magneson solution* as indicator; titrate to a pure blue end point. Each mL of 0.1M *tetrabutylammonium hydroxide VS* is equivalent to 18.99 mg of $C_{13}H_{18}ClN_3O_4S_2$.

Cyclopentolate Hydrochloride

(Ph. Eur. monograph 1093)



$C_{17}H_{26}ClNO_3$ 327.8 *5870-29-1*

Action and use

Anticholinergic.

Preparation

Cyclopentolate Eye Drops

Ph Eur

DEFINITION

2-(Dimethylamino)ethyl (2*RS*)-(1-hydroxycyclopentyl)(phenyl)acetate hydrochloride.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 135 °C to 141 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium chloride R.*

Comparison *cyclopentolate hydrochloride CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *ethanol (96 per cent) R*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 5 mL of *ethanol (96 per cent) R.*

Reference solution Dissolve 10 mg of *cyclopentolate hydrochloride CRS* in *ethanol (96 per cent) R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, water R, butyl acetate R, 2-propanol R* (5:15:30:50 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *alcoholic solution of sulfuric acid R* and heat at 120 °C for 30 min; examine in ultraviolet light at 365 nm.

Result The principal spot in the chromatogram obtained with the test solution is similar in position, fluorescence and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)
4.5 to 5.5.

Dissolve 0.2 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 20 mg of the substance to be examined in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 5.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 10 mg of *cyclopentolate for system suitability CRS* (containing impurity C) in *water R* and dilute to 10.0 mL with the same solvent.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: spherical *end-capped hexylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 0.66 g of *ammonium phosphate R* in *water R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*; mix and filter; mix 55 volumes of this solution and 45 volumes of *acetonitrile R1*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µl.

Run time 2.5 times the retention time of cyclopentolate.

Identification of impurities Use the chromatogram supplied with *cyclopentolate for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to cyclopentolate (retention time = about 4 min): impurity C = about 0.9.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 6, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to cyclopentolate.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 2.0;
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

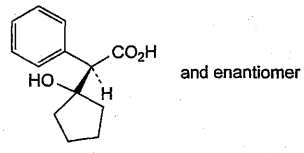
Dissolve 0.250 g in a mixture of 1.0 mL of *0.1 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 32.79 mg of $C_{17}H_{26}ClNO_3$.

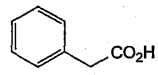
IMPURITIES

Specified impurities *C*.

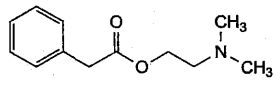
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*



A. (2*RS*)-(1-hydroxycyclopentyl)(phenyl)acetic acid,



B. phenylacetic acid,

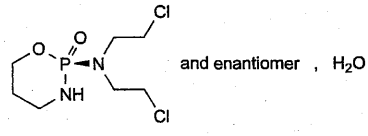


C. 2-(dimethylamino)ethyl phenylacetate.

Ph Eur

Cyclophosphamide

(Ph. Eur. monograph 0711)



$C_7H_{15}Cl_2N_2O_2P,H_2O$ 279.1 *6055-19-2*

Action and use
Cytotoxic alkylating agent.

Preparations
Cyclophosphamide Injection
Cyclophosphamide Oral Solution
Cyclophosphamide Tablets

Ph Eur

DEFINITION

Cyclophosphamide contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of (2*RS*)-*N*,*N*-bis(2-chloroethyl)tetrahydro-2*H*-1,3,2-oxazaphosphorin-2-

amine 2-oxide, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder, soluble in water, freely soluble in alcohol.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Determine the melting point (*2.2.14*) of the substance to be examined. Mix equal parts of the substance to be examined and *cyclophosphamide CRS* and determine the melting point of the mixture. The difference between the melting points (which are about 51 °C) is not greater than 2 °C.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *cyclophosphamide CRS*.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 10 mL of *water R* and add 5 mL of *silver nitrate solution R1*; the solution remains clear. Boil, a white precipitate is formed which dissolves in *concentrated ammonia R* and is reprecipitated on the addition of *dilute nitric acid R*.

TESTS

Solution S

Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of solution S is 4.0 to 6.0, determined immediately after preparation of the solution.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *alcohol R* and dilute to 5 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *alcohol R*.

Reference solution (a) Dissolve 10 mg of *cyclophosphamide CRS* in *alcohol R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dilute 0.1 mL of test solution (a) to 10 mL with *alcohol R*.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 2 volumes of *anhydrous formic acid R*, 4 volumes of *acetone R*, 12 volumes of *water R* and 80 volumes of *methyl ethyl ketone R*. Dry the plate in a current of warm air and heat at 110 °C for 10 min. At the bottom of a chromatographic tank, place an evaporating dish containing a 50 g/L solution of *potassium permanganate R* and add an equal volume of *hydrochloric acid R*. Place the plate whilst still hot in the tank and close the tank. Leave the plate in contact with the chlorine gas for 2 min. Withdraw the plate and place it in a current of cold air until the excess of chlorine is removed and an area of coating below the points of application gives at most a very faint blue colour with a drop of *potassium iodide and starch solution R*. Avoid prolonged exposure to cold air. Spray with *potassium iodide and starch solution R* and allow to stand for 5 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1.0 per cent). Disregard any spot remaining at the point of application.

Chlorides (*2.4.4*)

Dissolve 0.15 g in *water R* and dilute to 15 mL with the same solvent. The freshly prepared solution complies with the limit test for chlorides (330 ppm).

Phosphates (*2.4.11*)

Dissolve 0.10 g in *water R* and dilute to 100 mL with the same solvent. The solution complies with the limit test for phosphates (100 ppm).

Water (*2.5.12*)

6.0 per cent to 7.0 per cent, determined on 0.300 g by the semi-micro determination of water.

ASSAY

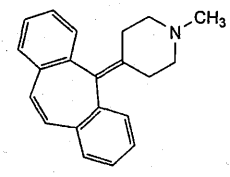
Dissolve 0.100 g in 50 mL of a 1 g/L solution of *sodium hydroxide R* in *ethylene glycol R* and boil under a reflux condenser for 30 min. Allow to cool and rinse the condenser with 25 mL of *water R*. Add 75 mL of *2-propanol R*, 15 mL of *dilute nitric acid R*, 10.0 mL of *0.1 M silver nitrate* and 2.0 mL of *ferric ammonium sulfate solution R2* and titrate with *0.1 M ammonium thiocyanate*.

1 mL of *0.1 M silver nitrate* is equivalent to 13.05 mg of $C_7H_{15}Cl_2N_2O_2P$.

Ph Eur

Cyproheptadine Hydrochloride

(Ph. Eur. monograph 0817)



, HCl , 1½ H_2O

$C_{21}H_{22}ClN,1½H_2O$ 350.9 *41354-29-4*

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Cyproheptadine Tablets

Ph Eur

DEFINITION

4-(5*H*-Dibenzo[*a*,*d*][7]annulen-5-ylidene)-1-methylpiperidine hydrochloride sesquihydrate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly yellow, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cyproheptadine hydrochloride CRS*.

B. A saturated solution gives reaction (b) of chlorides (*2.3.1*).

TESTS

Acidity

Dissolve 0.10 g in *water R* and dilute to 25 mL with the same solvent. Add 0.1 mL of *methyl red solution R*. Not more than 0.15 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 2.0 mg of *dibenzocycloheptene CRS* (impurity A), 2.0 mg of *dibenzosuberone CRS* (impurity B) and 2.0 mg of *cyproheptadine impurity C CRS* in mobile phase A, add 1.0 mL of the test solution and dilute to 100.0 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: dissolve 6.12 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 4.5 with *phosphoric acid R* and dilute to 1000 mL with *water R*; mix 60 volumes of this solution and 40 volumes of *acetonitrile for chromatography R*;
- *mobile phase B*: dissolve 6.12 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 4.5 with *phosphoric acid R* and dilute to 1000 mL with *water R*; mix 40 volumes of this solution and 60 volumes of *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10.0	100	0
10.0 - 10.1	100 → 0	0 → 100
10.1 - 35	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Relative retention With reference to cyproheptadine (retention time = about 8 min): impurity C = about 0.7; impurity B = about 2.6; impurity A = about 3.9.

System suitability Reference solution (b):
- *resolution*: minimum 7.0 between the peaks due to impurity C and cyproheptadine.

Limits:
- *impurities A, B, C*: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

7.0 per cent to 9.0 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

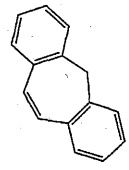
1 mL of *0.1 M sodium hydroxide* is equivalent to 32.39 mg of $C_{21}H_{22}ClN$.

STORAGE

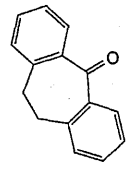
Protected from light.

IMPURITIES

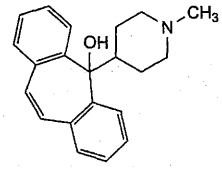
Specified impurities *A, B, C.*



A. 5*H*-dibenzo[*a,d*][7]annulene (dibenzocycloheptene),



B. 10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-one (dibenzosuberone),

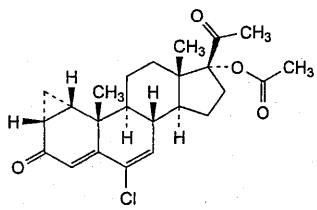


C. 5-(1-methylpiperidin-4-yl)-5*H*-dibenzo[*a,d*][7]annulen-5-ol.

Ph Eur

Cyproterone Acetate

(Ph. Eur. monograph 1094)



$C_{24}H_{29}ClO_4$ 416.9 *427-51-0*

Action and use
Antiandrogen.

Preparations
Co-cyprindiol Tablets
Cyproterone Tablets

Ph Eur

DEFINITION

6-Chloro-3,20-dioxo-1β,2β-dihydro-3′*H*-cyclopropa[1,2] pregna-1,4,6-trien-17-yl acetate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, very soluble in methylene chloride, freely soluble in acetone, soluble in methanol, sparingly soluble in anhydrous ethanol.

mp
About 210 °C.

IDENTIFICATION

First identification: A.

Second identification: B, C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cyproterone acetate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *cyproterone acetate CRS* in *methylene chloride R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *cyclohexane R, ethyl acetate R* (50:50 *V/V*).

Application 5 µL.

Development Twice over 3/4 of the plate; dry in air between the 2 developments.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. To about 1 mg add 2 mL of *sulfuric acid R* and heat on a water-bath for 2 min. A red colour develops. Cool. Add this solution cautiously to 4 mL of *water R* and shake. The solution becomes violet.

D. Incinerate about 30 mg with 0.3 g of *anhydrous sodium carbonate R* over a naked flame for about 10 min. Cool and dissolve the residue in 5 mL of *dilute nitric acid R*. Filter. To 1 mL of the filtrate add 1 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

E. It gives the reaction of acetyl (*2.3.1*).

TESTS

Specific optical rotation (*2.2.7*)
+ 152 to + 157 (dried substance).

Dissolve 0.25 g in *acetone R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10 mg of the substance to be examined in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R*.

Reference solution (b) Dissolve the contents of a vial of *cyproterone impurity mixture CRS* (impurities F and I) in 1.0 mL of the test solution.

Reference solution (c) Dissolve 2 mg of *cyproterone acetate for peak identification CRS* (containing impurities B, C, E and G) in 2.0 mL of *acetonitrile R*.

Column:
— *size*: *l* = 0.125 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase *acetonitrile R, water R* (40:60 *V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time Twice the retention time of cyproterone acetate.

Identification of impurities Use the chromatogram supplied with *cyproterone impurity mixture CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities F and I; use the chromatogram supplied with *cyproterone acetate for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C, E and G.

Relative retention With reference to cyproterone acetate (retention time = about 22 min): impurity E = about 0.27; impurity G = about 0.3; impurity F = about 0.5; impurity B = about 0.7; impurity I = about 0.9; impurity C = about 1.5.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity I and cyproterone acetate.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.8; impurity E = 0.7;
— *impurity F*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *impurity E*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurities B, C, G*: for each impurity, not more than 0.15 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying at 80 °C at a pressure not exceeding 0.7 kPa.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 50.0 mg in *methanol R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *methanol R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 282 nm.

Calculate the content of $C_{24}H_{29}ClO_4$ taking the specific absorbance to be 414.

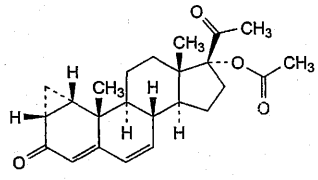
STORAGE

Protected from light.

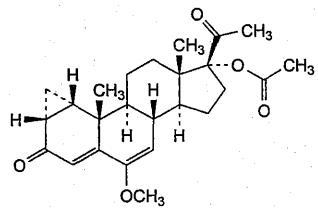
IMPURITIES

Specified impurities *B, C, E, F, G.*

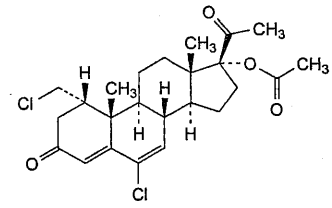
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, H, I, J.*



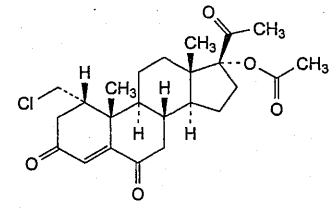
A. 3,20-dioxo-1β,2β-dihydro-3′*H*-cyclopropa[1,2]pregna-1,4,6-trien-17-yl acetate,



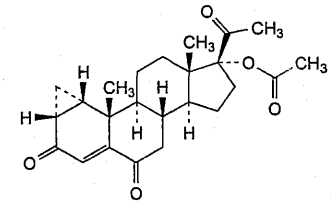
B. 6-methoxy-3,20-dioxo-1β,2β-dihydro-3′*H*-cyclopropa[1,2]pregna-1,4,6-trien-17-yl acetate,



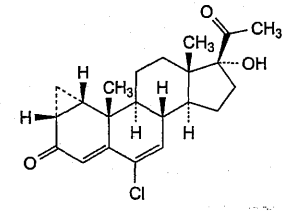
C. 6-chloro-1α-(chloromethyl)-3,20-dioxopregna-4,6-dien-17-yl acetate,



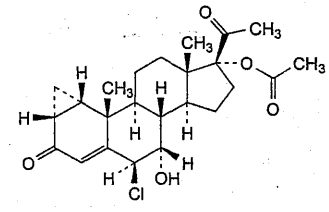
D. 1α-(chloromethyl)-3,6,20-trioxopregn-4-en-17-yl acetate,



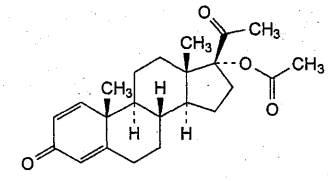
E. 3,6,20-trioxo-1β,2β-dihydro-3′*H*-cyclopropa[1,2]pregna-1,4-dien-17-yl acetate,



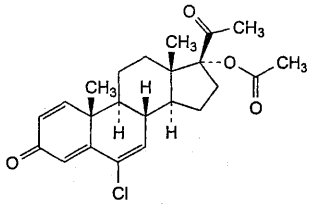
F. 6-chloro-17-hydroxy-1β,2β-dihydro-3′*H*-cyclopropa[1,2]pregna-1,4,6-triene-3,20-dione,



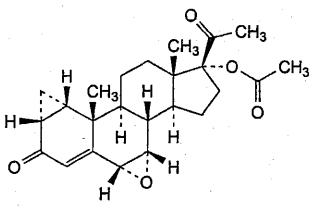
G. 6β-chloro-7α-hydroxy-3,20-dioxo-1β,2β-dihydro-3′*H*-cyclopropa[1,2]pregna-1,4-dien-17-yl acetate,



H. 3,20-dioxopregna-1,4-dien-17-yl acetate,



I. 6-chloro-3,20-dioxopregna-1,4,6-trien-17-yl acetate (delmadinone acetate),

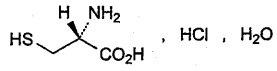


J. 6α,7α-epoxy-3,20-dioxo-1β,2β-dihydro-3′*H*-cyclopropa [1,2]pregna-1,4-dien-17-yl acetate.

Ph Eur

Cysteine Hydrochloride

(Cysteine Hydrochloride Monohydrate, Ph. Eur. monograph 0895)



$C_3H_8ClNO_2S,H_2O$ 175.6 *7048-04-6*

Action and use
Amino acid.

Ph Eur

DEFINITION

(2*R*)-2-Amino-3-sulfanylpropanoic acid hydrochloride monohydrate.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, E.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cysteine hydrochloride monohydrate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent. Add 10 mL of a 40 g/L solution of *N-ethylmaleimide R* in *ethanol (96 per cent) R*. Allow to stand for 5 min. Dilute 2 mL of the solution to 10 mL with *water R*.

Reference solution Dissolve 20 mg of *cysteine hydrochloride monohydrate CRS* in *water R* and dilute to 10 mL with the same solvent. Add 10 mL of a 40 g/L solution of *N-ethylmaleimide R* in *ethanol (96 per cent) R*. Allow to stand for 5 min. Dilute 2 mL of the solution to 10 mL with *water R*.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 80 °C for 30 min.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 5 mg in 1 mL of *dilute sodium hydroxide solution R*. Add 1 mL of a 30 g/L solution of *sodium nitroprusside R*. An intense violet colour develops which becomes brownish-red and then orange. Add 1 mL of *hydrochloric acid R*. The solution becomes green.

E. Dissolve about 50 mg in 5 mL of *water R*. Heat to about 60 °C on a water-bath and carefully add, dropwise, 5 mL of *strong hydrogen peroxide solution R*. Heat the water-bath to boiling and maintain the sample on the water-bath for 1 h. After cooling to room temperature reconstitute the sample to 10 mL with *water R*. 2 mL of the solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 2.5 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dilute 10 mL of solution S to 20 mL with *water R*.

Specific optical rotation (*2.2.7*)
+ 5.5 to + 7.0 (dried substance).

Dissolve 2.00 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances
Amino acid analysis (*2.2.56*). For analysis, use Method 1. *Prepare the solutions immediately before use.*

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *dilute hydrochloric acid R1* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *L-cystine R* (impurity A) in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (d) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (e) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (e):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for impurity A, use the concentration of impurity A in reference solution (b);
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of cysteine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (c); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *impurity A at 570 nm*: maximum 0.5 per cent;
— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (d) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 20 ppm.

In a separating funnel, dissolve 0.50 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
8.0 per cent to 12.0 per cent, determined on 1.000 g by drying at a pressure not exceeding 0.7 kPa for 24 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

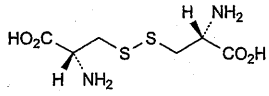
ASSAY
In a ground-glass stoppered flask dissolve 0.300 g of the substance to be examined and 4 g of *potassium iodide R* in 20 mL of *water R*. Cool the solution in iced water and add 3 mL of *hydrochloric acid R1* and 25.0 mL of *0.05 M iodine*. Stopper the flask and allow to stand in the dark for 20 min. Titrate with *0.1 M sodium thiosulfate* using 3 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank titration.

1 mL of *0.05 M iodine* is equivalent to 15.76 mg of $C_3H_8ClNO_2S$.

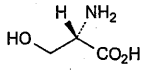
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B*.



A. (2*R*,2′*R*)-3,3′-disulfanediylbis(2-aminopropanoic acid) (cystine),

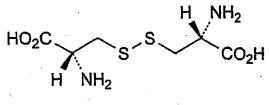


B. (2*S*)-2-amino-3-hydroxypropanoic acid (serine).

Ph Eur

Cystine

(Ph. Eur. monograph 0998)



$C_6H_{12}N_2O_4S_2$ 240.3 *56-89-3*

Action and use
Amino acid.

Ph Eur

DEFINITION
L-Cystine (3,3′-disulfanediylbis[(2*R*)-2-aminopropanoic acid]).

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *cystine CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in 1 mL of a 103 g/L solution of *hydrochloric acid R* and dilute to 50 mL with *water R.*

Reference solution Dissolve 10 mg of *cystine CRS* in 1 mL of a 103 g/L solution of *hydrochloric acid R* and dilute to 50 mL with *water R.*

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, 2-propanol R* (30:70 *V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 g carefully add 1 mL of *strong hydrogen peroxide solution R* and 0.1 mL of *ferric chloride solution R1.* Allow to cool. Add 1 mL of *dilute hydrochloric acid R* and 5 mL of *water R.* Add 1 mL of *barium chloride solution R1.* Turbidity or a white precipitate develops within 3 min.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 1.0 g in *dilute hydrochloric acid R* and dilute to 10 mL with the same acid.

Specific optical rotation (*2.2.7*)
−224 to −218 (dried substance).

Dissolve 0.50 g in a 103 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances
Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A A 10.3 g/L solution of *hydrochloric acid R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions (a), (b) and (d) into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of cystine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides
Maximum 200 ppm.

Dissolve 0.5 g in 5 mL of *dilute nitric acid R* and dilute to 10 mL with the same solvent. Add 10 mL of *strong hydrogen peroxide solution R* and heat on a water-bath for 30 min. Cool and dilute to 50 mL with *water R.* Add 1 mL of *silver nitrate solution R2* and mix. Allow to stand for 5 min protected from light. Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 2 mL of *chloride standard solution (50 ppm Cl) R.* Examine the tubes laterally against a black background.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dissolve 0.5 g in 5 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R.*

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In a flask with a ground-glass stopper, dissolve 0.100 g in a mixture of 2 mL of *dilute sodium hydroxide solution R* and 10 mL of *water R*. Add 10 mL of a 200 g/L solution of *potassium bromide R*, 50.0 mL of *0.0167 M potassium bromate* and 15 mL of *dilute hydrochloric acid R*. Stopper the flask and cool in iced water. Allow to stand in the dark for 10 min. Add 1.5 g of *potassium iodide R*. After 1 min, titrate with *0.1 M sodium thiosulfate*, using 2 mL of *starch solution R*, added towards the end-point, as indicator. Carry out a blank titration.

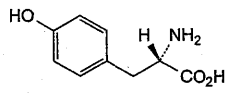
1 mL of *0.0167 M potassium bromate* is equivalent to 2.403 mg of $C_6H_{12}N_2O_4S_2$.

STORAGE

Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A.

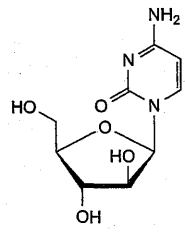


A. L-tyrosine ((2*S*)-2-amino-3-(4-hydroxyphenyl)propanoic acid).

Ph Eur

Cytarabine

(Ph. Eur. monograph 0760)



$C_9H_{13}N_3O_5$ 243.2 *147-94-4*

Action and use
Pyrimidine analogue; cytotoxic.

Preparation
Cytarabine Injection

Ph Eur

DEFINITION

4-Amino-1-β-D-arabinofuranosylpyrimidin-2(1*H*)-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

mp
About 215 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *cytarabine CRS*.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 1.0 g in *water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 154 to + 160 (dried substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 1.38 g of *sodium dihydrogen phosphate monohydrate R* and 1.42 g of *anhydrous disodium hydrogen phosphate R* in 950 mL of *water for chromatography R*, adjust to pH 7 with a 4 g/L solution of *sodium hydroxide R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *uracil arabinoside CRS* (impurity A) in *water R* and dilute to 100.0 mL with the same solvent. Dilute 0.5 mL of the solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 2 mg of *uridine R* (impurity B) and 2.5 mg of the substance to be examined in *water R* and dilute to 100 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, buffer solution (2:98 *V/V*);
— *mobile phase B*: *methanol R*, buffer solution (30:70 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	100	0
10 - 20	100 → 0	0 → 100
20 - 25	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to cytarabine (retention time = about 9 min): impurity B = about 1.2; impurity A = about 1.7.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to cytarabine and impurity B.

Calculation of percentage contents:
— for each impurity, use the concentration of impurity A in reference solution (a).

Limits:
— *impurity A*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.3 per cent;
— *reporting threshold*: 0.03 per cent.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 0.250 g by drying over *diphosphorus pentoxide R* at 60 °C at a pressure of 0.2 kPa to 0.7 kPa for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 60 mL of *anhydrous acetic acid R*, warming if necessary. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 24.32 mg of $C_9H_{13}N_3O_5$.

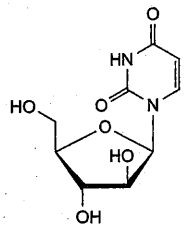
STORAGE

In an airtight container, protected from light.

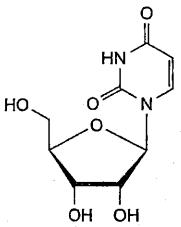
IMPURITIES

Specified impurities *A.*

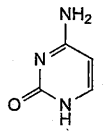
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H, I.*



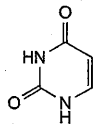
A. 1-β-D-arabinofuranosylpyrimidine-2,4(1*H*,3*H*)-dione (uracil arabinoside),



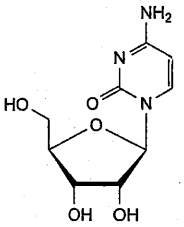
B. 1-β-D-ribofuranosylpyrimidine-2,4(1*H*,3*H*)-dione (uridine),



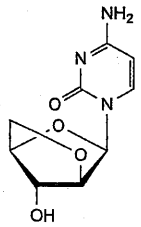
C. 4-aminopyrimidin-2(1*H*)-one (cytosine),



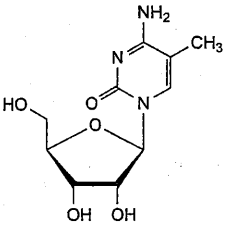
D. pyrimidin-2,4(1*H*,3*H*)-dione (uracil),



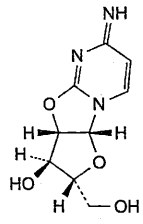
E. 4-amino-1-β-D-ribofuranosylpyrimidin-2(1*H*)-one (cytidine),



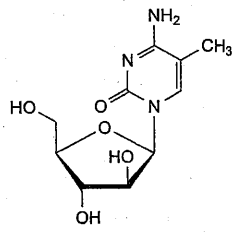
F. 4-amino-1-(2,5-anhydro-β-D-arabinofuranosyl)pyrimidin-2(1*H*)-one,



G. 4-amino-5-methyl-1-β-D-ribofuranosylpyrimidin-2(1*H*)-one (5-methylcytidine),



H. (2*R*,3*R*,3a*S*,9a*R*)-2-(hydroxymethyl)-6-imino-2,3,3a,9a-tetrahydro-6*H*-furo[2′,3′:4,5][1,3]oxazolo[3,2-*a*]pyrimidin-3-ol,

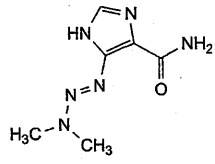


I. 4-amino-1-β-D-arabinofuranosyl-5-methylpyrimidin-2(1*H*)-one.

Ph Eur

Dacarbazine

(Ph. Eur. monograph 1691)



$C_6H_{10}N_6O$ 182.2 *4342-03-4*

Action and use
Cytotoxic alkylating agent.

Ph Eur

DEFINITION
5-[(1*E*)-3,3-Dimethyltriaz-1-enyl]-1*H*-imidazole-4-carboxamide.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or slightly yellowish, crystalline powder.

Solubility
Slightly soluble in water and in anhydrous ethanol, practically insoluble in methylene chloride.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 15.0 mg in 100.0 mL of *0.1 M hydrochloric acid.* Dilute 5.0 mL of this solution to 100.0 mL with *0.1 M hydrochloric acid.*

Spectral range 200-400 nm.

Absorption maximum At 323 nm.

Shoulder At 275 nm.

Specific absorbance at the absorption maximum 1024 to 1131.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dacarbazine CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 2.0 mg of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution Dissolve 2.0 mg of *dacarbazine CRS* in *methanol R* and dilute to 5.0 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (1:2:5 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.25 g in a 210 g/L solution of *citric acid monohydrate R* and dilute to 25.0 mL with the same solution.

Related substances
A. Liquid chromatography (*2.2.29*). *Use freshly prepared solutions and protect them from light.*

Test solution Dissolve 50.0 mg of the substance to be examined and 75 mg of *citric acid monohydrate R* in *distilled water R* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *dacarbazine impurity A CRS* in *distilled water R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of this solution to 25.0 mL with *distilled water R.*

Reference solution (b) Dissolve 5.0 mg of *dacarbazine impurity B CRS* in *distilled water R*, add 0.5 mL of the test solution and dilute to 10.0 mL with *distilled water R.* Dilute 1.0 mL of this solution to 50.0 mL with *distilled water R.*

Column:
- *size: l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 15.63 g/L solution of *glacial acetic acid R* containing 2.33 g/L of sodium dioctyl sulfosuccinate R. As the mobile phase contains sodium dioctyl sulfosuccinate, it must be freshly prepared every day, and the column must be flushed with a mixture of equal volumes of *methanol R* and *water R*, after all tests have been completed or at the end of the day, for at least 2 h.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 25 µL of the test solution and reference solution (a).

Run time 3 times the retention time of impurity A.

Retention time Impurity A = about 3 min.

Limits:
- — *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- — *unspecified impurities eluting after impurity A*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent).

B. Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modifications.

Mobile phase Mix 45 volumes of a 15.63 g/L solution of *glacial acetic acid R* containing 2.33 g/L of sodium dioctyl sulfosuccinate R with 55 volumes of *methanol R*.

Injection 10 µL of the test solution and reference solution (b).

Run time Twice the retention time of dacarbazine.

Relative retention With reference to dacarbazine (retention time = about 12 min): impurity B = about 0.7.

System suitability Reference solution (b):
- — *resolution*: minimum 1.5 between the peaks due to impurity B and dacarbazine.

Limits:
- — *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
- — *unspecified impurities*: for each impurity, not more than the area of the peak due to dacarbazine in the chromatogram obtained with reference solution (b) (0.10 per cent);
- — *total*: not more than 5 times the area of the peak due to dacarbazine in the chromatogram obtained with reference solution (b) (0.5 per cent);
- — *disregard limit*: 0.5 times the area of the peak due to dacarbazine in the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity D

Head-space gas chromatography (*2.2.28*).

Test solution Introduce 0.200 g of the substance to be examined into a 20 mL vial and firmly attach the septum and cap. Using a 10 µL syringe, inject 5 µL of *water R* into the vial.

Reference solution (a) Dilute 2.5 mL of *dimethylamine solution R* (impurity D) to 100.0 mL with *water R* (solution A). Firmly attach the septum and cap to a 20 mL vial. Using a 10 µL syringe, inject 10 µL of solution A into the vial.

Reference solution (b) Firmly attach the septum and cap to a 20 mL vial. Using a 10 µL syringe, inject 10 µL of solution A and 10 µL of a 10 g/L solution of *triethylamine R* into the vial.

Column:
- — *material*: fused silica;
- — *size*: *l* = 30.0 m, Ø = 0.53 mm;
- — *stationary phase*: base-deactivated polyethyleneglycol R (film thickness 1.0 µm).

Carrier gas *helium for chromatography R.*

Flow rate 13 mL/min.

Split ratio 1:1.

Static head-space conditions that may be used:
- — *equilibration temperature*: 60 °C;
- — *equilibration time*: 10 min;
- — *transfer-line temperature*: 90 °C;
- — *pressurisation time*: 30 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	35
	3 - 11	35 → 165
Injection port		180
Detector		220

Detection Flame ionisation.

Injection 1 mL.

System suitability Reference solution (b):
- — *resolution*: minimum 2.5 between the peaks due to impurity D and triethylamine.

Limit:
- — *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 18.22 mg of $C_6H_{10}N_6O$.

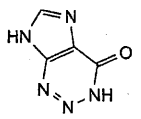
STORAGE

At a temperature of 2 °C to 8 °C, protected from light.

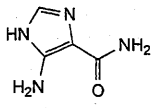
IMPURITIES

Specified impurities *A, B, D.*

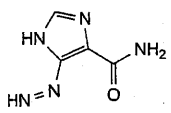
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



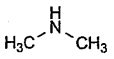
A. 3,7-dihydro-4*H*-imidazo[4,5-*d*]-1,2,3-triazin-4-one (2-azahypoxanthine),



B. 5-amino-1*H*-imidazole-4-carboxamide,



C. 5-diazenyl-1*H*-imidazole-4-carboxamide,

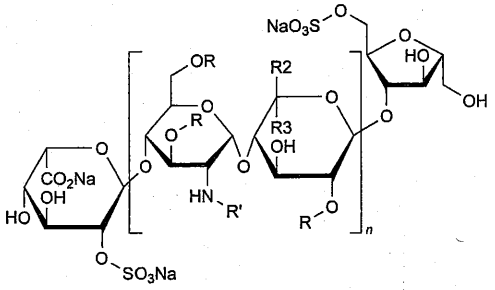


D. *N*-methylmethanamine.

Ph Eur

Dalteparin Sodium

(Ph. Eur. monograph 1195)



n = 3 to 20 , R = H or SO_3Na , R' = SO_3Na or CO-CH_3
R2 = H and R3 = CO_2Na or R2 = CO_2Na and R3 = H

Action and use
Low molecular weight heparin.

Preparation
Dalteparin Sodium Injection

Ph Eur

DEFINITION
Dalteparin sodium is the sodium salt of a low-molecular-mass heparin that is obtained by nitrous acid depolymerisation of heparin from porcine intestinal mucosa. The majority of the components have a 2-*O*-sulfo-α-L-idopyranosuronic acid structure at the non-reducing end and a 6-*O*-sulfo-2,5-anhydro-D-mannitol structure at the reducing end of their chain.

Dalteparin sodium complies with the monograph Low-molecular-mass heparins (0828) with the modifications and additional requirements below.

The mass-average relative molecular mass ranges between 5600 and 6400, with a characteristic value of about 6000.

The degree of sulfatation is 2.0 to 2.5 per disaccharide unit.

The potency is not less than 110 IU and not more than 210 IU of anti-factor Xa activity per milligram, calculated with reference to the dried substance. The anti-factor IIa activity is not less than 35 IU/mg and not more than 100 IU/mg, calculated with reference to the dried substance. The ratio of anti-factor Xa activity to anti-factor IIa activity is between 1.9 and 3.2.

PRODUCTION
Dalteparin sodium is produced by a validated manufacturing and purification procedure under conditions designed to minimise the presence of N-NO groups.

The manufacturing procedure must have been shown to reduce any contamination by N-NO groups to approved limits using an appropriate, validated quantification method.

IDENTIFICATION
Carry out identification test A as described in the monograph *Low-molecular-mass heparins (0828)* using *dalteparin sodium CRS*.

Carry out identification test C as described in the monograph *Low-molecular-mass heparins (0828)*. The following requirements apply.

The mass-average relative molecular mass ranges between 5600 and 6400. The mass percentage of chains lower than 3000 is not more than 13.0 per cent. The mass percentage of chains higher than 8000 ranges between 15.0 per cent and 25.0 per cent.

TESTS

Appearance of solution
Dissolve 1 g in 10 mL of *water R*. The solution is clear (*2.2.1*) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Nitrite
Not more than 5 ppm. Examine by liquid chromatography (*2.2.29*). *Rinse all volumetric flasks at least three times with water R before the preparation of the solutions.*

Test solution Dissolve 80.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent. Allow to stand for at least 30 min.

Reference solution (a) Dissolve 60.0 mg of *sodium nitrite R* in *water R* and dilute to 1000.0 mL with the same solvent.

For the preparation of reference solution (b), use a pipette previously rinsed with reference solution (a).

Reference solution (b) Dilute 1.00 mL of reference solution (a) to 50.0 mL with *water R*.

Before preparing reference solutions (c), (d) and (e), rinse all pipettes with reference solution (b).

Reference solution (c) Dilute 1.00 mL of reference solution (b) to 100.0 mL with *water R* (corresponding to 1 ppm of nitrite in the test sample).

Reference solution (d) Dilute 3.00 mL of reference solution (b) to 100.0 mL with *water R* (corresponding to 3 ppm of nitrite in the test sample).

Reference solution (e) Dilute 5.00 mL of reference solution (b) to 100.0 mL with *water R* (corresponding to 5 ppm of nitrite in the test sample).

The chromatographic procedure may be carried out using:
— a column 0.125 m long and 4.3 mm in internal diameter packed with a strong anion-exchange resin;
— as mobile phase at a flow rate of 1.0 mL/min a solution consisting of 13.61 g of *sodium acetate R* dissolved in *water R*, adjusted to pH 4.3 with *phosphoric acid R* and diluted to 1000 mL with *water R*;
— as detector an appropriate electrochemical device with the following characteristics and settings: a suitable working electrode, a detector potential of + 1.00 V versus Ag/AgCl reference electrode and a detector sensitivity of 0.1 μA full scale.

Inject 100 μL of reference solution (d). When the chromatograms are recorded in the prescribed conditions, the retention time for nitrite is 3.3 to 4.0 min. The test is not valid unless:
— the number of theoretical plates calculated for the nitrite peak is at least 7000 per metre per column (dalteparin sodium will block the binding sites of the stationary phase, which will cause shorter retention times and lower separation efficiency for the analyte; the initial performance of the column may be partially restored using a 58 g/L solution of *sodium chloride R* at a flow rate of 1.0 mL/min for 1 h; after regeneration the column is rinsed with 200 mL to 400 mL of *water R*);

— the symmetry factor for the nitrite peak is less than 3;
— the relative standard deviation of the peak area for nitrite obtained from 6 injections is less than 3.0 per cent.

Inject 100 µL each of reference solutions (c) and (e).
The test is not valid unless:
— the correlation factor for a linear relationship between concentration and response for reference solutions (c), (d) and (e) is at least 0.995;
— the signal-to-noise ratio for reference solution (c) is not less than 5 (if the noise level is too high, electrode recalibration is recommended);
— a blank injection of *water R* does not give rise to spurious peaks.

Inject 100 µL of the test solution. Calculate the content of nitrite from the peak areas in the chromatogram obtained with reference solutions (c), (d) and (e).

Boron

Not more than 1 ppm, determined by inductively coupled plasma atomic emission spectroscopy.

Boron is determined by measurement of the emission from an inductively coupled plasma (ICP) at a wavelength specific to boron. The emission line at 249.733 nm is used. Use an appropriate apparatus, whose settings have been optimised as directed by the manufacturer.

Test solution Dissolve 0.2500 g of the substance to be examined in about 2 mL of *water for chromatography R*, add 100 µL of *nitric acid R* and dilute to 10.00 mL with the same solvent.

Reference solution (a) Prepare a 1 per cent *V/V* solution of *nitric acid R* in *water for chromatography R* (blank).

Reference solution (b) Prepare a 11.4 µg/mL solution of *boric acid R* in a 1 per cent *V/V* solution of *nitric acid R* in *water for chromatography R* (STD_{cal}).

Reference solution (c) Dissolve 0.2500 g of a reference dalteparin sodium with no detectable boron in about 2 mL of *water for chromatography R*, add 100 µL of *nitric acid R* and dilute to 10.00 mL with the same solvent (STD_0).

Reference solution (d) Dissolve 0.2500 g of a reference dalteparin sodium with no boron detected in about 2 mL of a 1 per cent *V/V* solution of *nitric acid R* in *water for chromatography R*, add 10 µL of a 5.7 mg/mL solution of *boric acid R* and dilute to 10.00 mL with the same solvent (STD_1). This solution contains 1 µg/mL of boron.

Calculate the content of boron in the substance to be examined, using the following correction factor:

$$f = \frac{(STD_1 - STD_0) \times 2}{(STD_{cal} - \text{blank})}$$

Loss on drying (*2.2.32*)

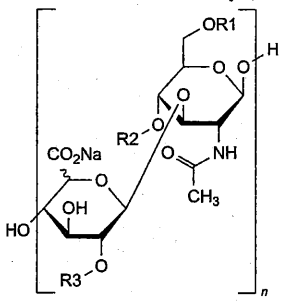
Not more than 5.0 per cent, determined on 1.000 g by drying in an oven at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 670 Pa for 3 h.

Ph Eur

Danaparoid Sodium

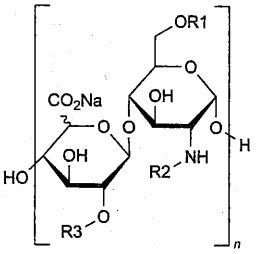
(Ph. Eur. monograph 2090)

Chondroitin sulfate and dermatan sulfate family



ΔDi	R1	R2	R3
-OS	H	H	H
-6S	SO_3Na	H	H
-4S	H	SO_3Na	H
-US	H	H	SO_3Na
-(U,6)S	SO_3Na	H	SO_3Na
-(U,4)S	H	SO_3Na	SO_3Na
-(4,6)S	SO_3Na	SO_3Na	H
-(U,4,6)S	SO_3Na	SO_3Na	SO_3Na

Heparan sulfate family



ΔDiHS	R1	R2	R3
-OS	H	Ac	H
-6S	SO_3Na	Ac	H
-NS	H	SO_3Na	H
-US	H	Ac	SO_3Na
-(U,N)S	H	SO_3Na	SO_3Na
-(6,N)S	SO_3Na	SO_3Na	H
-(U,N,6)S	SO_3Na	SO_3Na	SO_3Na

Action and use

Heparinoid; prevention of deep vein thrombosis.

Ph Eur

DEFINITION

Preparation containing the sodium salts of a mixture of sulfated glycosaminoglycans present in porcine tissues. Danaparoid sodium is prepared from the intestinal mucosa of pigs. Its major constituents are heparan sulfate and dermatan sulfate. On complete hydrolysis it liberates D-glucosamine, D-galactosamine, D-glucuronic acid, L-iduronic acid, acetic acid and sulfuric acid. It has the characteristic property of enhancing the inactivation of activated factor X (factor Xa) by antithrombin. It has a negligible effect on the inactivation rate of thrombin by antithrombin.

Potency

11.0 to 17.0 anti-factor Xa units per milligram (dried substance).

PRODUCTION

The animals from which danaparoid sodium is derived must fulfil the requirements for the health of animals suitable for human consumption. It is prepared using a process that ensures that the relative proportion of active sulfated glycosaminoglycans is consistent. It is produced by methods of manufacturing designed to minimise or eliminate endotoxins and hypotensive substances.

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water.

IDENTIFICATION

A. The ratio of anti-factor Xa activity to anti-factor IIa activity, determined as described under Assay and Tests respectively, is not less than 22.

B. Molecular mass distribution (see Tests): the mass-average relative molecular mass ranges between 4000 and 7000.

TESTS

pH (*2.2.3*)
5.5 to 7.0.

Dissolve 0.5 g of the dried substance to be examined in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Anti-factor IIa activity
Maximum 0.5 units per milligram (dried substance).

Test solutions Prepare 2 independent series of dilutions in geometric progression of the substance to be examined in *phosphate buffer solution pH 6.5 R* and in the concentration range of 0.0005 to 0.005 units of anti-factor IIa activity per millilitre.

Reference solutions Prepare 2 independent series of dilutions in geometric progression of *danaparoid sodium CRS* in *phosphate buffer solution pH 6.5 R* and in the concentration range of 0.0005 to 0.005 units of anti-factor IIa activity per millilitre.

Transfer 50 µL of each solution into the wells of a 96-well microtitre plate. To each well add 50 µL of *antithrombin III solution R3* and 50 µL of *human thrombin solution R1*. Shake the microtitre plate but do not allow bubbles to form. Incubate for 75 min. To each well add 50 µL of *chromogenic substrate R4*. Shake the microtitre plate. Measure the absorbances at 405 nm (*2.2.25*) using a suitable reading device, exactly 4 min after the addition of the chromogenic substrate. The reaction may be stopped using 75 µL of a 20 per cent *V/V* solution of *glacial acetic acid R*. Determine the blank amidolytic activity in a similar manner, using *phosphate buffer solution pH 6.5 R* as the blank solution (minimum 10 blanks per microtitre plate). Calculate the activity of the substance to be examined in units of anti-factor IIa activity per milligram using a suitable statistical method, for example the parallel-line assay.

Chondroitin sulfate and dermatan sulfate
Chondroitin sulfate: maximum 8.5 per cent (dried substance); dermatan sulfate: 8.0 per cent to 16.0 per cent (dried substance).

Determine by selective enzymatic degradation.

Test solutions Dry the substance to be examined at 60 °C over *diphosphorus pentoxide R* at a pressure of about 670 Pa for 3 h. Dissolve 0.200 g of the dried substance in 10.0 mL of *water R*. Dilute this solution as necessary to obtain 3 test solutions containing 20 mg/mL, 10 mg/mL and 5 mg/mL of the dried substance to be examined in *water R*.

Chondroitin sulfate reference solutions Dry *chondroitin sulfate sodium CRS* over *diphosphorus pentoxide R* at room temperature at a pressure of about 670 Pa for 16 h. Prepare solutions containing 1 mg/mL, 2 mg/mL and 3 mg/mL of dried *chondroitin sulfate sodium CRS* in *water R*.

Dermatan sulfate reference solutions Dry *dermatan sulfate CRS* over *diphosphorus pentoxide R* at room temperature at a pressure of about 670 Pa for 16 h. Prepare solutions containing 1 mg/mL, 2 mg/mL and 3 mg/mL of dried *dermatan sulfate CRS* in *water R*.

Chondroitinase ABC solution Dissolve *chondroitinase ABC R* in *tris-sodium acetate-sodium chloride buffer solution pH 8.0 R* to obtain an activity of 0.5-1.0 units per millilitre.

Chondroitinase AC solution Dissolve *chondroitinase AC R* in *tris-sodium acetate-sodium chloride buffer solution pH 7.4 R* to obtain an activity of 1.0-2.0 units per millilitre.

Procedure:

— *Degradation with chondroitinase ABC*: label 2 sets of 10 tubes in triplicate: T1, T2 and T3 for the test solutions; SD1, SD2 and SD3 for the dermatan sulfate reference solutions; SC1, SC2 and SC3 for the chondroitin sulfate reference solutions; and B for the blank (*water R*). To each tube add 1.25 mL of *tris-sodium acetate buffer solution pH 8.0 R* and 150 µL of the test solutions, dermatan sulfate reference solutions, chondroitin sulfate reference solutions or *water R*. To each tube in 1 set of tubes add 75 µL of chondroitinase ABC solution. To determine the blank response level, add 75 µL of *tris-sodium acetate-sodium chloride buffer solution pH 8.0 R* to each tube in the other set of tubes. Mix the contents of the tubes using a vortex mixer, cover with appropriate stoppers and incubate at 37 °C for at least 24 h.

— *Degradation with chondroitinase AC*: label 7 tubes in triplicate: T1, T2 and T3 for the test solutions; SC1, SC2 and SC3 for the chondroitin sulfate reference solutions; and B for the blank (*water R*). To each tube add 1.25 mL of *tris-sodium acetate buffer solution pH 7.4 R* and 150 µL of the test solutions, chondroitin sulfate reference solutions or *water R*. Add 75 µL of chondroitinase AC solution to each tube. Mix the contents of the tubes using a vortex mixer, cover with appropriate stoppers and incubate at 37 °C for at least 24 h. After the incubation period mix the contents of the tubes using a vortex mixer and dilute to 12 times with *water R*. Measure the absorbances (*2.2.25*) of the diluted solutions at 234 nm against *water R* using a suitable spectrophotometer.

Calculation Calculate the mean blank absorbance of each reference solution, i.e. the mean of the absorbances of the reference solutions to which no chondroitinase ABC has been added. Subtract the mean blank absorbance value from the individual absorbance of each reference solution. Calculate linear regression curves for the 2 chondroitin sulfate reference and the dermatan sulfate reference by plotting the blank-corrected absorbances against the concentrations.

Calculate the average percentage content of dermatan sulfate in the test solutions of all tested concentrations using the following expression:

$$\frac{A_2 - A_1 - \frac{(A_3 - A_1 - I_1) \times B_2}{B_1} - I_2 - I_3}{B_3 \times C} \times 100$$

A_1 = blank absorbance of the test solution;
A_2 = absorbance of the test solution with chondroitinase ABC;
A_3 = absorbance of the test solution with chondroitinase AC;
B_1 = gradient of the curve obtained with the chondroitin sulfate reference solutions with chondroitinase AC;
B_2 = gradient of the curve obtained with the chondroitin sulfate reference solutions with chondroitinase ABC;
B_3 = gradient of the curve obtained with the dermatan sulfate reference solutions with chondroitinase ABC;
C = concentration of the test solution, in milligrams per millilitre;
I_1 = y-intercept of the curve obtained with the chondroitin sulfate reference solutions with chondroitinase AC;
I_2 = y-intercept of the curve obtained with the chondroitin sulfate reference solutions with chondroitinase ABC;
I_3 = y-intercept of the curve obtained with the dermatan sulfate reference solutions with chondroitinase ABC.

Calculate the average percentage content of chondroitin sulfate in the test solutions for all tested concentrations using the following expression:

$$\frac{(A_3 - A_1 - I_1) \times 100}{B_1 \times C}$$

Molecular mass distribution
Size-exclusion chromatography (*2.2.30*).

Test solution Dissolve 10 mg of the substance to be examined in 2 mL of the mobile phase.

Reference solution Dissolve 10 mg of *danaparoid sodium CRS* in 2 mL of the mobile phase.

Column:
— *size*: *l* = 0.60 m, Ø = 7.5 mm;
— *stationary phase*: *hydrophilic silica gel for chromatography R* (10 µm) with a fractionation range for proteins with a relative molecular mass of approximately 5000-100 000;
— *temperature*: 30 °C.

Mobile phase 28.4 g/L solution of *anhydrous sodium sulfate R* adjusted to pH 5.0 with *dilute sulfuric acid R*.

Flow rate 0.9 mL/min ± 2 per cent.

Detection Spectrophotometer at 210 nm.

Injection 100 µL.

Run time For a period of time ensuring complete elution of sample and solvent peaks (about 40 min).

System suitability Inject the reference solution twice. The difference between the retention times corresponding to the maxima of the peaks is not more than 5 s.

Calibration Calibration is achieved by taking the relevant part of the chromatogram obtained with the reference solution, i.e. excluding the sharp peak at the end of the chromatogram, and matching the chromatogram obtained with the test solution with the calibration table obtained with the reference solution. From the calibration curve obtained, determine the molecular mass distribution of the sample. A calibration table is supplied with *danaparoid sodium CRS*.

Limits:
— *chains with a relative molecular mass less than 2000*: maximum 13 per cent;
— *chains with a relative molecular mass less than 4000*: maximum 39 per cent;
— *chains with a relative molecular mass between 4000 and 8000*: minimum 50 per cent;
— *chains with a relative molecular mass higher than 8000*: maximum 19 per cent;
— *chains with a relative molecular mass higher than 10 000*: maximum 11 per cent.

Nitrogen (*2.5.9*)
2.4 per cent to 3.0 per cent (dried substance).

Nucleic acids
Maximum 0.5 per cent (dried substance).

Test solution Weigh about 50 mg of the dried substance to be examined into a centrifuge tube and dissolve in 200 µL of *water R*.

Reference solution Dissolve about 50 mg of *ribonucleic acid CRS* in 5 mL of *0.1 M sodium hydroxide* and dilute to 20.0 mL with *water R*. Transfer 200 µL of the solution into a centrifuge tube.

Add 4.0 mL of a 50 g/L solution of *trichloroacetic acid R* to each tube and mix. Place all tubes in boiling water for 30 min. Allow to cool to room temperature. Add again 4.0 mL of a 50 g/L solution of *trichloroacetic acid R* to each tube and mix. If any of the test solutions is not clear, sonicate all the tubes in an ultrasonic bath for 10 min and centrifuge at 1500 *g* for 15 min. Dilute 1.0 mL of the clear supernatant to 4.0 mL with *water R*. Measure the absorbances of the diluted reference and test solutions at 265 nm (*2.2.25*) against a blank solution prepared in the same manner, and calculate the percentage nucleic acid content of the sample.

Total protein (*2.5.33, Method 2*)
Maximum 0.5 per cent.

Dissolve the substance to be examined in *water R*. Use *bovine albumin R* as the reference substance.

Sodium
9.0 per cent to 11.0 per cent (dried substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 0.125 g of the substance to be examined in 100.0 mL of a 1.27 mg/mL solution of *caesium chloride R* in *0.1 M hydrochloric acid*.

Reference solutions Prepare reference solutions containing 50 ppm, 100 ppm and 150 ppm of Na by diluting *sodium standard solution (1000 ppm Na) R* with a 1.27 mg/mL solution of *caesium chloride R* in *0.1 M hydrochloric acid*.

Source Sodium hollow-cathode lamp.

Wavelength 330.3 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 0.500 g by drying in an oven at 60 °C over *diphosphorus pentoxide R* at a pressure of 670 Pa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.02 IU per unit of anti-factor Xa activity, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

The anticoagulant activity of danaparoid sodium is determined *in vitro* by an assay which determines its ability to accelerate the inhibition of factor Xa by antithrombin III (anti-factor Xa assay).

Test solutions Prepare 2 independent series of dilutions in geometric progression of the substance to be examined in *tris (hydroxymethyl) aminomethane EDTA buffer solution pH 8.4 R* and in the concentration range of 0.1 to 0.32 units of anti-factor Xa activity per millilitre.

Reference solutions Prepare 2 independent series of dilutions in geometric progression of *danaparoid sodium CRS* in *tris (hydroxymethyl) aminomethane EDTA buffer solution pH 8.4 R* and in the concentration range of 0.08 to 0.35 units of anti-factor Xa activity per millilitre.

Transfer 40 µL of each solution into the wells of a 96-well microtitre plate. Add 40 µL of *antithrombin III solution R4* to each well and shake the microtitre plate but do not allow bubbles to form. Add 40 µL of *bovine factor Xa solution R1* to each well. Exactly 2 min after the addition of the factor Xa solution, add 80 µL of *chromogenic substrate R5*. Measure the absorbance at 405 nm (*2.2.25*) using a suitable reading device, exactly 4 min after the addition of the factor Xa solution. The reaction may be stopped using 75 µL of a 20 per cent *V/V* solution of *glacial acetic acid R*. Determine the blank amidolytic activity in the same manner, using *tris (hydroxymethyl) aminomethane EDTA buffer solution pH 8.4 R* as the blank (minimum 8 blanks per microtitre plate). Calculate the potency of the substance to be examined in units of anti-factor Xa activity per milligram using a suitable statistical method, for example the parallel-line assay.

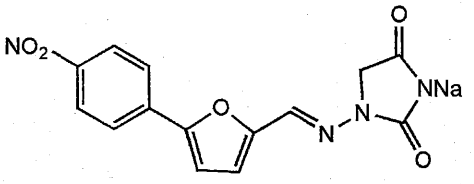
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING
The label states the number of units of anti-factor Xa activity per milligram.

Ph Eur

Dantrolene Sodium



, 3½ H_2O

$C_{14}H_9N_4NaO_5,3½H_2O$ 399.3 *24868-20-0*

Action and use
Skeletal muscle relaxant.

Preparation
Dantrolene Oral Suspension

DEFINITION
Dantrolene Sodium is 1-(5-*p*-nitrophenylfurfurylideneamino) hydantoin sodium. It contains not less than 98.0% and not more than 102.0% of $C_{14}H_9N_4NaO_5$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A yellowish-orange to orange crystalline powder.

Very slightly soluble in *water*; slightly soluble in *ethanol (96%)*; sparingly soluble in *methanol*; practically insoluble in *acetone*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of dantrolene sodium (*RS 422*).

B. In the Assay, the chromatogram obtained with solution (1) shows a peak with the same retention time as the principal peak in the chromatogram obtained with solution (2).

C. To 0.1 g of the substance being examined add 20 mL of *water* and 2 drops of *acetic acid*, shake well and filter. The filtrate yields the reactions characteristic of *sodium salts*, Appendix VI.

TESTS
Alkalinity
Shake 0.7 g in 10 mL of *water* for 5 minutes and centrifuge. To 5 mL of the supernatant add 45 mL of *water* and 3 drops of *phenolphthalein solution R1* and 0.1 mL of *0.1M hydrochloric acid VS*. A red colour is not produced.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) Dissolve 50 mg of the substance being examined in 20 mL of *tetrahydrofuran* and 2 mL of *glacial acetic acid* and dilute with sufficient *absolute ethanol* to produce 100 mL.

(2) Dilute 1 mL of solution (1) to 100 mL with *absolute ethanol*.

(3) Dissolve 5 mg of *dantrolene sodium BPCRS* and 0.1 g of *theophylline BPCRS* in 20 mL of *tetrahydrofuran* and 2 mL of *glacial acetic acid* and dilute with sufficient *absolute ethanol* to produce 100 mL. Further dilute 10 mL of this solution to 100 mL with *absolute ethanol*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 4.6 mm) packed with *silica gel for chromatography* (5 µm) (Zorbax Sil is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Adjust the flow rate of the mobile phase so that the retention time of the peak corresponding to Dantrolene Sodium is about 8 minutes.

(d) Use a column temperature of 30°.

(e) Use a detection wavelength of 300 nm.

(f) Inject 10 µL of each solution.

(g) For solution (1) allow the chromatography to proceed for at least twice the retention time of the principal peak.

MOBILE PHASE

9 volumes of *absolute ethanol*, 10 volumes of *glacial acetic acid* and 90 volumes of *hexane*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution* between the peaks corresponding to theophylline and dantrolene is at least 6.

LIMITS

In the chromatogram obtained with solution (1):

the total area of all the *secondary peaks* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%).

Water
14.5 to 17.0% w/w, Appendix IX C. Use 0.2 g

ASSAY
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) Dissolve 60 mg of the substance being examined in 50 mL of *dimethylformamide* and dilute 1 volume of the resulting solution to 100 volumes with the mobile phase.

(2) Dilute 1 volume of a 0.12% w/v solution of *dantrolene sodium BPCRS* in *dimethylformamide* to 100 volumes with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 4.6 mm) packed with spherical particles of silica, 5 µm in diameter, the surface of which has been modified with chemically-bonded nitrile groups (Spherisorb CN is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 262 nm.

(f) Inject 20 µL of each solution.

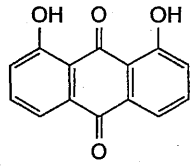
MOBILE PHASE

15 volumes of *acetonitrile* and 85 volumes of a phosphate buffer pH 6.8 prepared by dissolving 11.88 g of *disodium hydrogen orthophosphate* and 9.08 g of *potassium dihydrogen orthophosphate* in 1000 mL of *water*.

DETERMINATION OF CONTENT

Calculate the content of $C_{14}H_9N_4NaO_5$ in the substance being examined using the declared content of $C_{14}H_9N_4NaO_5$ in *dantrolene sodium BPCRS*.

Dantron



$C_{14}H_8O_4$ 240.2 *117-10-2*

Action and use
Anthraquinone stimulant laxative.

Preparation
Co-danthrusate Capsules

DEFINITION
Dantron is mainly 1,8-dihydroxyanthraquinone. It contains not less than 98.0% and not more than 102.0% of total phenols, calculated as $C_{14}H_8O_4$ and with reference to the dried substance.

CHARACTERISTICS
An orange, crystalline powder.

Practically insoluble in *water*, slightly soluble in *ether*, very slightly soluble in *ethanol (96%)*. It dissolves in solutions of alkali hydroxides.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of dantron *(RS 083)*.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.001% w/v solution in *dichloromethane* exhibits maxima at 255 nm and 285 nm and a less well-defined maximum at 275 nm. The *absorbance* at the maximum at 255 nm is about 0.82 and at the maximum at 285 nm is about 0.48, each calculated with reference to the dried substance.

C. Dissolve 5 mg in 5 mL of 1M *sodium hydroxide*. A clear red solution is produced immediately.

TESTS
Mercury
To 0.50 g in a Kjeldahl flask add 2.5 mL of *nitric acid* and allow to stand until the initial vigorous reaction has subsided. Add 2.5 mL of *sulfuric acid* and heat until dense white fumes are evolved. Cool, add 2.5 mL of *nitric acid* and heat until fumes are again evolved. Repeat the procedure with a further 2.5 mL of *nitric acid*, cool, add 50 mL of *water*, boil the solution until the volume has been reduced to about 25 mL and cool. Transfer to a separating funnel using *water*, dilute to about 50 mL with *water* and add 50 mL of 0.5M *sulfuric acid*. Add 100 mL of *water*, 2 g of *hydroxylamine hydrochloride*, 1 mL of 0.05M *disodium edetate*, 1 mL of *glacial acetic acid* and 5 mL of *dichloromethane*, shake, allow to separate and discard the dichloromethane layer. Titrate the aqueous layer with a 0.0008% w/v solution of *dithizone* in *dichloromethane*, shaking vigorously after each addition, allowing the layers to separate and discarding the dichloromethane layer, until the dichloromethane layer remains green. Repeat the operation using a solution prepared by diluting 1 mL of *mercury standard solution (5 ppm Hg)* to 100 mL with 0.5M *sulfuric acid* and beginning at the words 'Add 100 mL of *water*...'. The volume of the dithizone solution required by the substance being examined does not exceed that required by the mercury standard solution.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) Dissolve 50 mg of the substance being examined in 20 mL of *tetrahydrofuran* and dilute to 100 mL with the mobile phase.

(2) Dilute 1 volume of solution (1) to 50 volumes with the mobile phase.

(3) Dissolve 50 mg of *dantron impurity standard BPCRS* in 20 mL of *tetrahydrofuran* and dilute to 100 mL with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm x 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Nucleosil C18 is suitable).

(b) Use an isocratic system using the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for 1.5 times the retention time of the principal peak.

MOBILE PHASE

A mixture of 2.5 volumes of *glacial acetic acid*, 40 volumes of *tetrahydrofuran* and 60 volumes of *water*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3):
- — a peak due to 1-hydroxyanthraquinone appears immediately before the principal peak, as indicated in the reference chromatogram supplied with *dantron impurity standard BPCRS*;
- the height of the trough separating the two peaks is not greater than one third of the height of the peak due to 1-hydroxyanthraquinone.

LIMITS

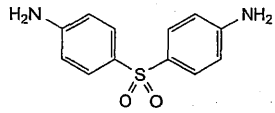
In the chromatogram obtained with solution (1):
- the area of any peak corresponding to 1-hydroxyanthraquinone is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with solution (2) (3.3% taking into account the correction factor of the impurity);
- — the sum of the areas of any other *secondary peaks* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (2%);
- disregard any peak with a retention time less than one third of that of the principal peak.

Loss on drying
When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

ASSAY
Dissolve 0.2 g in 50 mL of *anhydrous pyridine* and carry out Method II for *non-aqueous titration*, Appendix VIII A, using 0.1M *tetrabutylammonium hydroxide VS* as titrant and determining the end point potentiometrically. Each mL of 0.1M *tetrabutylammonium hydroxide VS* is equivalent to 24.02 mg of total phenols, calculated as $C_{14}H_8O_4$.

Dapsone

(Ph. Eur. monograph 0077)



$C_{12}H_{12}N_2O_2S$ 248.3 *80-08-0*

Action and use
Folic acid synthesis inhibitor; treatment of leprosy.

Preparation
Dapsone Tablets

Ph Eur

DEFINITION
Dapsone contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4,4′-sulfonyldianiline, calculated with reference to the dried substance.

CHARACTERS
A white or slightly yellowish-white, crystalline powder, very slightly soluble in water, freely soluble in acetone, sparingly soluble in alcohol. It dissolves freely in dilute mineral acids.

IDENTIFICATION
A. Melting point (*2.2.14*): 175 °C to 181 °C.

B. Dissolve 50.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *methanol R*. Examined between 230 nm and 350 nm (*2.2.25*), the solution shows 2 absorption maxima, at 260 nm and 295 nm. The specific absorbances at these maxima are 700 to 760 and 1150 to 1250, respectively.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dissolve 10 mg of *dapsone CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 10 mL with *methanol R*.

Reference solution (c) Dilute 2 mL of reference solution (b) to 10 mL with *methanol R*.

Apply separately to the plate 1 µL of test solution (b), 1 µL of reference solution (a), 10 µL of test solution (a), 10 µL of reference solution (b) and 10 µL of reference solution (c). Develop in an unsaturated tank over a path of 15 cm using a mixture of 1 volume of *concentrated ammonia R*, 6 volumes of *methanol R*, 20 volumes of *ethyl acetate R* and 20 volumes of *heptane R*. Allow the plate to dry in air. Spray the plate with a 1 g/L solution of *4-dimethylaminocinnamaldehyde R* in a mixture of 1 volume of *hydrochloric acid R* and 99 volumes of *alcohol R*. Examine in daylight. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1.0 per cent) and not more than 2 such spots are more intense than the spot in the chromatogram obtained with reference solution (c) (0.2 per cent).

Loss on drying (*2.2.32*)
Not more than 1.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.100 g in 50 mL of *dilute hydrochloric acid R*. Carry out the determination of primary aromatic amino-nitrogen (*2.5.8*).

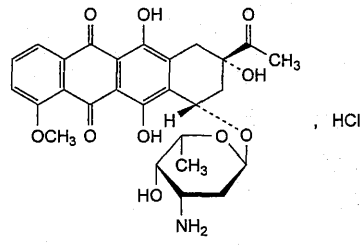
1 mL of *0.1 M sodium nitrite* is equivalent to 12.42 mg of $C_{12}H_{12}N_2O_2S$.

STORAGE
Store protected from light.

Ph Eur

Daunorubicin Hydrochloride

(Ph. Eur. monograph 0662)



$C_{27}H_{30}ClNO_{10}$ 564.0 *23541-50-6*

Action and use
Cytostatic; anthracycline antibacterial.

Ph Eur

DEFINITION
(8*S*,10*S*)-8-Acetyl-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione hydrochloride.

Substance produced by certain strains of *Streptomyces coeruleorubidus* or of *Streptomyces peucetius* or obtained by any other means.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION
It is produced by methods of manufacture designed to eliminate or minimise the presence of histamine.

CHARACTERS
Appearance
Crystalline, orange-red powder, hygroscopic.

Solubility
Freely soluble in water and in methanol, slightly soluble in alcohol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *daunorubicin hydrochloride CRS.*

B. Dissolve about 10 mg in 0.5 mL of *nitric acid R*, add 0.5 mL of *water R* and heat over a flame for 2 min. Allow to cool and add 0.5 mL of *silver nitrate solution R1*. A white precipitate is formed.

TESTS

pH (*2.2.3*)

4.5 to 6.5.

Dissolve 50 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *daunorubicin hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *doxorubicin hydrochloride CRS* and 10 mg of *epirubicin hydrochloride CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of *daunorubicinone CRS* and 5.0 mg of *doxorubicin hydrochloride CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 200.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm,
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mixture of equal volumes of *acetonitrile R* and a solution containing 2.88 g/L of *sodium laurilsulfate R* and 2.25 g/L of *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL; inject the test solution and reference solutions (b), (c) and (d).

Run time Twice the retention time of daunorubicin.

Relative retention With reference to daunorubicin (retention time = about 15 min): impurity A = about 0.4; impurity D = about 0.5; epirubicin = about 0.6; impurity B = about 0.7.

System suitability Reference solution (b):
- *resolution*: minimum of 2.0 between the peaks due to impurity D and epirubicin.

Limits:
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
- *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (d) (1.5 per cent),
- *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
- *any other impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent),
- *total of other impurities*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (2.5 per cent),
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Butanol (*2.4.24, System B*)

Maximum 1.0 per cent.

Water (*2.5.12*)

Maximum 3.0 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)

Less than 4.3 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances.

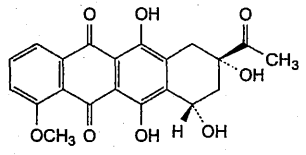
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{27}H_{30}ClNO_{10}$.

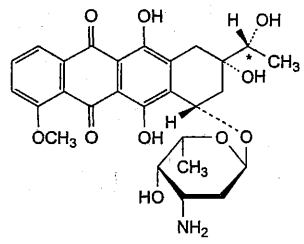
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

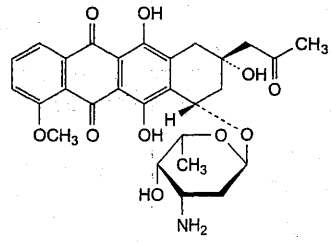


A. (8*S*,10*S*)-8-acetyl-6,8,10,11-tetrahydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (daunorubicin aglycone, daunorubicinone),

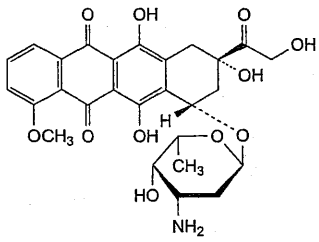


and epimer at C*

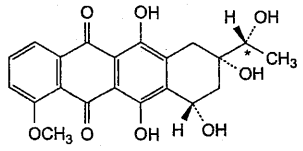
B. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-[(1*RS*)-1-hydroxyethyl]-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (daunorubicinol),



C. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-8-(2-oxopropyl)-7,8,9,10-tetrahydrotetracene-5,12-dione (feudomycin B),

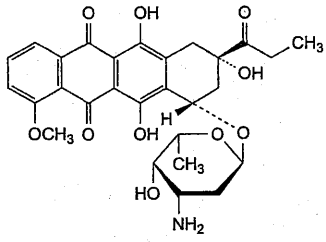


D. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (doxorubicin),



and epimer at C*

E. (8*S*,10*S*)-6,8,10,11-tetrahydroxy-8-[(1*RS*)-1-hydroxyethyl]-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (13-dihydrodaunorubicinone),



F. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-8-propanoyl-7,8,9,10-tetrahydrotetracene-5,12-dione (8-ethyldaunorubicin).

Ph Eur

Decyl Oleate

(Ph. Eur. monograph 1307)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture consisting of decyl esters of fatty acids, mainly oleic (*cis*-9-octadecenoic) acid.

A suitable antioxidant may be added.

CHARACTERS
Appearance
Clear, pale yellow or colourless liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent), with methylene chloride and with light petroleum (bp: 40-60 °C).

IDENTIFICATION
A. Relative density (see Tests).

B. Saponification value (see Tests).

C. Oleic acid (see Tests).

TESTS
Relative density (*2.2.5*)
0.860 to 0.870.

Acid value (*2.5.1*)
Maximum 1.0, determined on 10.0 g.

Iodine value (*2.5.4, Method A*)
55 to 70.

Peroxide value (*2.5.5, Method A*)
Maximum 10.0.

Saponification value (*2.5.6*)
130 to 140, determined on 2.0 g.

Oleic acid (*2.4.22, Method A*)
Minimum 60.0 per cent in the fatty acid fraction of the substance.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.00 g.

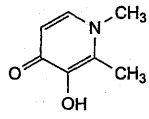
Total ash (*2.4.16*)
Maximum 0.1 per cent, determined on 2.0 g.

STORAGE
Protected from light.

Ph Eur

Deferiprone

(Ph. Eur. monograph 2236)



$C_7H_9NO_2$ 139.2 *30652-11-0*

Action and use
Chelating agent (iron).

Preparations
Deferiprone Oral Solution
Deferiprone Tablets

Ph Eur

DEFINITION
3-Hydroxy-1,2-dimethylpyridin-4-(1*H*)-one.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or pinkish-white powder.

Solubility
Sparingly soluble in water, slightly soluble in anhydrous ethanol, practically insoluble in heptane.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *deferiprone CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Use only colourless glassware. Protect the solutions from light.*

Solution A Dissolve 2.91 g of *sodium edetate R*, 4.01 g of *sodium octanesulfonate monohydrate R* and 6.20 g of *dipotassium*

hydrogen phosphate R in *water for chromatography R* and dilute to 2000 mL with the same solvent; adjust to pH 3.0 with *phosphoric acid R*.

Solution B Dissolve 0.73 g of *sodium edetate R*, 1.0 g of *sodium octanesulfonate monohydrate R* and 1.55 g of *dipotassium hydrogen phosphate R* in *water for chromatography R* and dilute to 2000 mL with the same solvent; adjust to pH 3.0 with *phosphoric acid R*.

Solvent mixture *acetonitrile R, water R* (10:90 *V/V*).

Test solution (a) Dissolve 0.100 g of the substance to be examined in a volume of the mobile phase corresponding to about 2/3 of the final volume and dilute to 100.0 mL with the mobile phase.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in a volume of the solvent mixture corresponding to about 2/3 of the final volume and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 200.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *maltol R* (impurity B) in the mobile phase and dilute to 100.0 mL with the mobile phase. To 2.5 mL of the solution add 10 mL of test solution (a) and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 50.0 mg of *deferiprone CRS* in a volume of the solvent mixture corresponding to about 2/3 of the final volume and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 200.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *acetonitrile R*, solution A (10:90 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Preconditioning of the column Rinse for 20 min with the mobile phase before each series of injections.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Run time 3.5 times the retention time of deferiprone.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity B.

Relative retention With reference to deferiprone (retention time = about 12 min): impurity B = about 0.5.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to impurity B and deferiprone.

Calculation of percentage contents:
— for each impurity, use the concentration of deferiprone in reference solution (b).

Limits:
— *impurity B*: maximum 0.10 per cent;
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.03 per cent.

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 0.100 g by direct sample introduction.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase *acetonitrile R*, solution B (10:90 *V/V*).

Injection 10 µL of test solution (b) and reference solution (c).

Run time Twice the retention time of deferiprone.

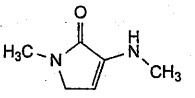
Retention time Deferiprone = about 7.7 min.

Calculate the percentage content of $C_7H_9NO_2$ taking into account the assigned content of *deferiprone CRS*.

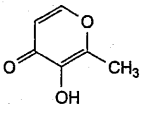
IMPURITIES

Specified impurities *B.*

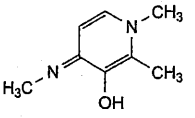
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C.*



A. 1-methyl-3-(methylamino)-1,5-dihydro-2*H*-pyrrol-2-one,



B. 3-hydroxy-2-methyl-4*H*-pyran-4-one (maltol),

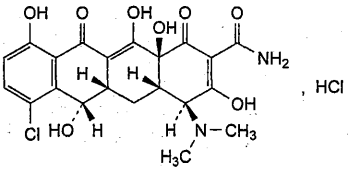


C. 1,2-dimethyl-4-[(*E*)-methylimino]-1,4-dihydropyridin-3-ol.

Ph Eur

Demeclocycline Hydrochloride

(Ph. Eur. monograph 0176)



$C_{21}H_{22}Cl_2N_2O_8$ 501.3 *64-73-3*

Action and use
Tetracycline antibacterial.

Preparation
Demeclocycline Capsules

Ph Eur

DEFINITION

(4*S*,4a*S*,5a*S*,6*S*,12a*S*)-7-Chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide hydrochloride.

Substance produced by certain strains of *Streptomyces aureofaciens*.

Content

89.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow powder.

Solubility

Soluble or sparingly soluble in water, slightly soluble in alcohol, very slightly soluble in acetone. It dissolves in solutions of alkali hydroxides and carbonates.

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *demeclocycline hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *demeclocycline hydrochloride CRS*, 5 mg of *chlortetracycline hydrochloride R* and 5 mg of *tetracycline hydrochloride R* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase Mix 20 volumes of *acetonitrile R*, 20 volumes of *methanol R* and 60 volumes of a 63 g/L solution of *oxalic acid R* previously adjusted to pH 2 with *concentrated ammonia R*.

Application 1 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of *sulfuric acid R*. A violet colour develops. Add the solution to 2.5 mL of *water R*. The colour becomes yellow.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)

2.0 to 3.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Buffer 1 22.2 g/L solution of *sodium edetate R* adjusted to pH 7.5 with a 40 g/L solution of *sodium hydroxide R*.

Buffer 2 17.0 g/L solution of *tetrapropylammonium hydrogen sulfate R* adjusted to pH 7.5 with a 40 g/L solution of *sodium hydroxide R*.

Test solution Dissolve 25.0 mg of the substance to be examined in a 1.0 g/L solution of *hydrochloric acid R* and dilute to 50.0 mL with the same solution.

Reference solution (a) Dissolve 25.0 mg of *demeclocycline hydrochloride CRS* in a 1.0 g/L solution of *hydrochloric acid R* and dilute to 50.0 mL with the same solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a 1.0 g/L solution of *hydrochloric acid R*.

Reference solution (c) Dissolve 5 mg of *demeclocycline for system suitability CRS* (containing impurities A, B, C, E and G) in a 1.0 g/L solution of *hydrochloric acid R* and dilute to 10 mL with the same solution.

Column:
- *size*: *l* = 0.075 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octylsilyl silica gel for chromatography with polar incorporated groups R* (3.5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R*, *water R*, buffer 1, buffer 2 (2:28:35:35 *V/V/V/V*);
- *mobile phase B*: *acetonitrile R*, buffer 1, buffer 2 (30:35:35 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	83	17
5 - 15	83 → 30	17 → 70
15 - 25	30	70

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *demeclocycline for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, E and G.

Relative retention With reference to demeclocycline (retention time = about 14 min): impurity C = about 0.3; impurity B = about 0.7; impurity A = about 0.8; impurity E = about 1.2; impurity G = about 1.6.

System suitability Reference solution (c):
- *resolution*: minimum 2.5 between the peaks due to impurities A and B.

Calculation of percentage contents:
- for each impurity, use the concentration of demeclocycline in reference solution (b).

Limits:
- *impurities A, B*: for each impurity, maximum 5.0 per cent;
- *impurities C, G*: for each impurity, maximum 0.3 per cent;
- *impurity E*: maximum 0.2 per cent;
- *any other impurity*: for each impurity, maximum 0.15 per cent;
- *total*: maximum 10.0 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

Maximum 3.0 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $C_{21}H_{22}Cl_2N_2O_8$ using the chromatogram obtained with reference solution (a) and taking into account the assigned content of *demeclocycline hydrochloride CRS*.

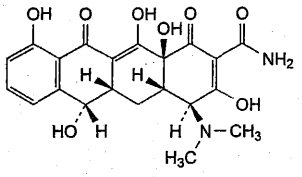
STORAGE

Protected from light.

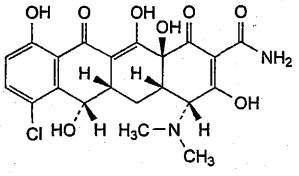
IMPURITIES

Specified impurities *A, B, C, E, G.*

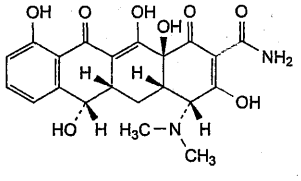
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, F.*



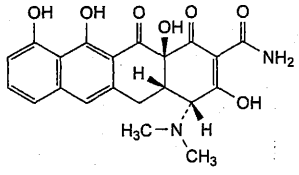
A. (4*S*,4a*S*,5a*S*,6*S*,12a*S*)-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (demethyltetracycline),



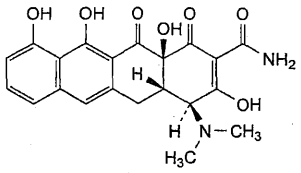
B. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidemeclocycline),



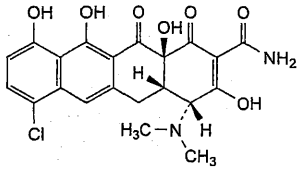
C. (4*R*,4a*S*,5a*S*,6*S*,12a*S*)-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidemethyltetracycline),



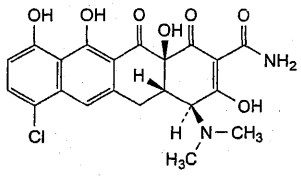
D. (4*R*,4a*S*,12a*S*)-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-1,12-dioxo-1,4,4a,5,12,12a-hexahydrotetracene-2-carboxamide (4-epianhydrodemethyltetracycline),



E. (4*S*,4a*S*,12a*S*)-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-1,12-dioxo-1,4,4a,5,12,12a-hexahydrotetracene-2-carboxamide (anhydrodemethyltetracycline),



F. (4*R*,4a*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-1,12-dioxo-1,4,4a,5,12,12a-hexahydrotetracene-2-carboxamide (4-epianhydrodemeclocycline),

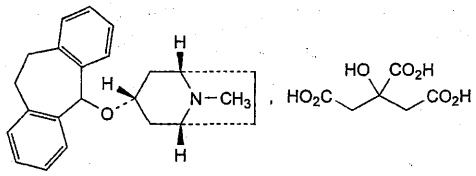


G. (4*S*,4a*S*,12a*S*)-7-chloro-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-1,12-dioxo-1,4,4a,5,12,12a-hexahydrotetracene-2-carboxamide (anhydrodemeclocycline).

Ph Eur

Deptropine Citrate

(Ph. Eur. monograph 1308)



$C_{29}H_{35}NO_8$ 525.6 *2169-75-7*

Action and use

Histamine H_1 receptor antagonist; anticholinergic.

Ph Eur

DEFINITION

Deptropine citrate contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of (1*R*,3*r*,5*S*)-3-(10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-yloxy)-8-methyl-8-azabicyclo[3.2.1]octane dihydrogen citrate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, microcrystalline powder, very slightly soluble in water and in ethanol, practically insoluble in methylene chloride.

It melts at about 170 °C, with decomposition.

IDENTIFICATION

First identification: A.

Second identification: B, C, D, E.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *deptropine citrate CRS*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (b).

C. To about 1 mg add 0.5 mL of *sulfuric acid R*. A stable red-orange colour develops.

D. Dissolve about 1 mg in 0.25 mL of *perchloric acid R* and warm gently until the solution becomes turbid. Add 5 mL of *glacial acetic acid R*; a pink colour with an intense green fluorescence appears.

E. To about 5 mg add 1 mL of *acetic anhydride R* and 5 mL of *pyridine R*. A purple colour develops.

TESTS

pH (*2.2.3*)

Suspend 0.25 g in *carbon dioxide-free water R*, dilute to 25 mL with the same solvent and filter. The pH of the solution is 3.7 to 4.5.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using as the coating substance a suitable silica gel with a fluorescent indicator having an optimal intensity at 254 nm.

Test solution (a) Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with *methanol R*.

Reference solution (b) Dissolve 20 mg of *deptropine citrate CRS* in *methanol R* and dilute to 2 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with *methanol R*.

Reference solution (c) Dissolve 5 mg of *tropine CRS* in *methanol R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dissolve 10 mg of *deptropine citrate CRS* and 10 mg of *tropine CRS* in *methanol R* and dilute to 25 mL with the same solvent.

Apply to the plate 40 µL of each solution. Develop over a path of 10 cm using a mixture of 8 volumes of *concentrated ammonia R* and 92 volumes of *butanol R*. Dry the plate at 100 °C to 105 °C until the ammonia has completely evaporated. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (1 per cent). Spray with *dilute potassium iodobismuthate solution R* and then with a 10 g/L solution of *sodium nitrite R*. Expose the plate to iodine vapours. Examine in daylight and in ultraviolet light at 254 nm. In the chromatogram obtained with test solution (a): any spot corresponding to tropine is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent); any spot, apart from the principal spot and any spot corresponding to tropine, is not more intense than the spot in the chromatogram obtained with reference solution (a) (1 per cent). The test is not valid unless the chromatogram obtained with reference solution (d) shows two clearly separated spots.

Loss on drying (*2.2.32*)

Not more than 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

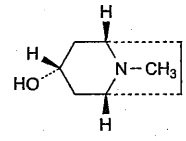
Dissolve 0.400 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 52.56 mg of $C_{29}H_{35}NO_8$.

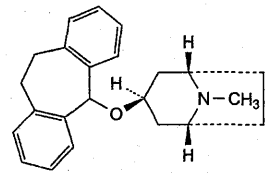
STORAGE

Store protected from light.

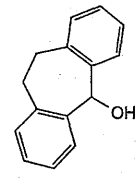
IMPURITIES



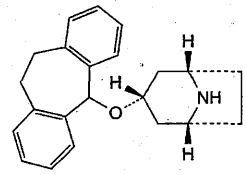
A. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]octan-3-ol (tropine),



B. (1*R*,3*s*,5*S*)-3-(10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-yloxy)-8-methyl-8-azabicyclo[3.2.1]octane (pseudodeptropine),



C. 10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-ol (dibenzocycloheptadienol),

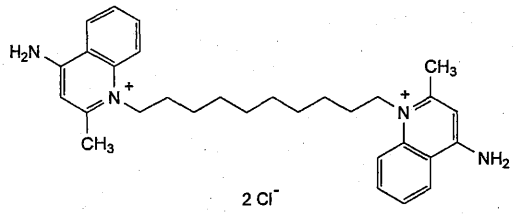


D. (1*R*,3*r*,5*S*)-3-(10,11-dihydro-5*H*-dibenzo[*a*,*d*][7]annulen-5-yloxy)-8-azabicyclo[3.2.1]octane (demethyldeptropine).

Ph Eur

Dequalinium Chloride

(Ph. Eur. monograph 1413)



$C_{30}H_{40}Cl_2N_4$ 527.6 *522-51-0*

Action and use
Antiseptic.

Ph Eur

DEFINITION
1,1′-(Decane-1,10-diyl)bis(4-amino-2-methylquinolinium) dichloride (dried substance).

Content
95.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
White or yellowish-white powder, hygroscopic.

Solubility
Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve about 10 mg in *water R* and dilute to 100 mL with the same solvent. Dilute 10 mL of the solution to 100 mL with *water R*.

Spectral range 230-350 nm.

Absorption maxima At 240 nm and 326 nm.

Shoulder At 336 nm.

Absorbance ratios:
— A_{240}/A_{326} = 1.56 to 1.80;
— A_{326}/A_{336} = 1.12 to 1.30.

B. Infrared absorption spectrophotometry (*2.2.24*).

Spectral range 600-2000 cm^{-1}.

Comparison *dequalinium chloride CRS*.

C. To 5 mL of solution S (see Tests) add 5 mL of *potassium ferricyanide solution R*. A yellow precipitate is formed.

D. To 10 mL of solution S add 1 mL of *dilute nitric acid R*. A white precipitate is formed. Filter and reserve the filtrate for identification test E.

E. The filtrate from identification test D gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.2 g in 90 mL of *carbon dioxide-free water R*, heating if necessary, and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 5 mL of solution S add 0.1 mL of *bromothymol blue solution R1*. Not more than 0.2 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *dequalinium chloride for performance test CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10.0 mg of *dequalinium chloride CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R*.

Mobile phase Dissolve 2 g of *sodium hexanesulfonate R* in 300 mL of *water R*; adjust to pH 4.0 with *acetic acid R* and add 700 mL of *methanol R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 5 times the retention time of dequalinium chloride.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dequalinium chloride. If necessary, adjust the concentration of methanol in the mobile phase.

Limits:
— *impurity A*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *total of impurities other than A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (10 per cent);
— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Readily carbonisable substances
Dissolve 20 mg in 2 mL of *sulfuric acid R*. After 5 min the solution is not more intensely coloured than reference solution BY_4 (*2.2.2, Method I*).

Loss on drying (*2.2.32*)
Maximum 7.0 per cent, determined on 1.000 g by drying at 105 °C at a pressure not exceeding 0.7 kPa.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.200 g in 5 mL of *anhydrous formic acid R* and add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 26.38 mg of $C_{30}H_{40}Cl_2N_4$.

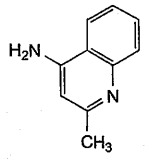
STORAGE

In an airtight container.

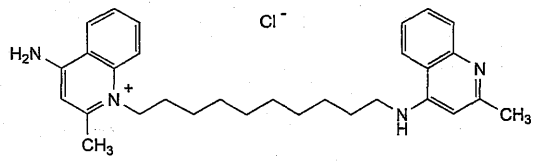
IMPURITIES

Specified impurities *A.*

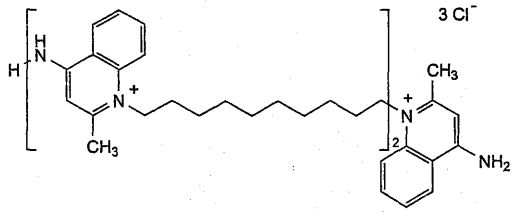
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C.*



A. 2-methylquinolin-4-amine,



B. 4-amino-1-[10-[(2-methylquinolin-4-yl)amino]decyl]-2-methylquinolinium chloride,



C. 1-[10-(4-amino-2-methylquinolinio)decyl]-4-[[10-(4-amino-2-methylquinolinio)decyl]amino]-2-methylquinolinium trichloride.

Ph Eur

3-*O*-Desacyl-4′-Monophosphoryl Lipid A

(Ph. Eur. monograph 2537)

Ph Eur

DEFINITION

3-*O*-Desacyl-4′-monophosphoryl lipid A is a detoxified derivative of the lipopolysaccharide (LPS) of *Salmonella minnesota*, strain R595, which retains the immunostimulatory activities of the parent LPS. It consists of a mixture of congeners, all containing a backbone of β1′→6-linked disaccharide of 2-deoxy-2-aminoglucose phosphorylated at the 4′-position, but differing in the fatty acid substitutions at the 2, 2′ and 3′ positions. The immunostimulatory activities of 3-*O*-desacyl-4′-monophosphoryl lipid A combined with the vaccine include up-regulation of co-stimulatory molecules on antigen-presenting cells and secretion of pro-inflammatory cytokines, resulting in an enhanced immune response of the Th1-type against the antigens. 3-*O*-desacyl-4′-monophosphoryl lipid A is a lyophilised powder or a sterile liquid.

Requirements given in the sections up to and including the section Triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A also apply to formulations that do not proceed to the 3-*O*-desacyl-4′-monophosphoryl lipid A liquid bulk.

PRODUCTION

GENERAL PROVISIONS

The production method shall have been shown to yield consistently a 3-*O*-desacyl-4′-monophosphoryl lipid A comparable in structure and function with a preparation of 3-*O*-desacyl-4′-monophosphoryl lipid A used as adjuvant in the particular vaccine of proven clinical efficacy and safety in man.

During development studies, and wherever revalidation is necessary, a test for residual endotoxin activity is carried out by injecting intravenously 12-day-old embryonated hens' eggs with 0.1 mL of dilutions of the test sample (8 eggs per dilution) of 3-*O*-desacyl-4′-monophosphoryl lipid A. Eggs are candled and read for mortality at 18-24 hours post-inoculation and the chick embryo 50 per cent lethal dose ($CELD_{50}$) is calculated. The residual endotoxin activity of the 3-*O*-desacyl-4′-monophosphoryl lipid A is acceptable if the $CELD_{50}$ is more than 100 μg.

An endotoxin standard of *Salmonella typhimurium* is prepared and selected dilutions are injected into each group of 8 eggs.

For a test to be valid, the $CELD_{50}$ of the endotoxin standard must not be more than 0.05 μg.

Reference preparation A batch of 3-*O*-desacyl-4′-monophosphoryl lipid A shown to be comparable in structure and function with a preparation of 3-*O*-desacyl-4′-monophosphoryl lipid A used as adjuvant in the particular vaccine of proven clinical efficacy and safety in man or a batch representative thereof.

BACTERIAL SEED LOTS

The bacterial strain used for master seed lots shall be identified by historical records that include information on its origin and the tests used to characterise the strain, in particular genotypic and phenotypic information. Only a working seed lot that complies with the following requirements may be used.

Identification

The working seed lot is identified by suitable methods such as Gram staining and fatty acid profiling (*5.1.6*).

Microbial Purity

Each seed lot complies with the requirements for absence of contaminating organisms. Purity of bacterial cultures is verified by methods of suitable sensitivity and specificity.

PROPAGATION AND HARVEST

The bacteria is grown using a suitable liquid medium. At the end of cultivation, the culture is tested for purity and yield. The culture medium is separated from the bacterial mass by a suitable method, for example filtration. Only a harvest that is consistent with respect to the profiles for growth rate, pH, and O_2-consumption may be used for the extraction of LPS.

TRIETHYLAMINE SALT OF 3-O-DESACYL-4′-MONOPHOSPHORYL LIPID A

LPS is extracted from the bacterial cells by successive alcohol and chloroform-methanol extractions and is then converted to 3-*O*-desacyl-4′-monophosphoryl lipid A by hydrolysis, then purified and salified by triethanolamine before freeze-

drying. The freeze-dried triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A must comply with the following requirements.

Appearance
A visual description of the particular preparation after freeze-drying is established and approved by the competent authority; each batch of freeze-dried triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A must comply with this description.

Protein
Less than 0.5 per cent *m/m*, determined using a suitable method, for example a reversed-phase HPLC method for amino acid analysis (*2.2.56*). The total amino acid content in micrograms is calculated by comparison to amino acid standards and is equal to the protein concentration.

Nucleic acid
Maximum 0.3 per cent *m/m*, determined using a suitable method. For example, a fluorimetric method may be used where nucleic acids are extracted from the freeze-dried triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A, using a solution containing NH_4OH and a suitable non-ionic detergent, and stained by a suitable fluorescent dye.
The nucleic acid content in the test sample is interpolated from a calibration curve.

Hexosamine (*2.5.20*)
1000 nmol/mg to 1450 nmol/mg.

Phosphorus (*2.5.18*)
0.5 µmol/mg to 0.8 µmol/mg.

Congener distribution
The relative amount of tetraacyl, pentaacyl, hexaacyl and heptaacyl congener groups are determined by a suitable method, for example reversed-phase HPLC analysis (*2.2.29*).

The relative amount of each congener group in the triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A is:
— tetraacyl: 15 per cent to 35 per cent;
— pentaacyl: 35 per cent to 60 per cent;
— hexaacyl: 20 per cent to 40 per cent;
— heptaacyl: less than 0.5 per cent.

Triethylamine
4.2 to 5.8 per cent *m/m*, determined by a suitable method, for example gas chromatography (*2.2.28*).

Water (*2.5.12*)
Maximum 6.7 per cent *m/m*.

Free fatty acids
Maximum 2.6 per cent *m/m*, determined by a suitable method, for example reversed-phase HPLC analysis (*2.2.29*).

2-Keto-3-deoxyoctonate
Less than 0.5 per cent *m/m*, determined by a suitable method. For example, a colorimetric method may be used where 2-keto-3-deoxyoctonate is released by hydrolysis (0.2 N H_2SO_4 at 100 °C for 30 min), oxidised by periodic acid, and reacted with sodium arsenite to yield β-formylpyruvic acid, which subsequently is coupled to thiobarbituric acid to give a red coloured chromophore with absorption maximum at 550 nm. The amount of 2-keto-3-deoxyoctonate is interpolated from a calibration curve.

Identity
The test for congener distribution also serves to identify the product.

Microbial contamination
TAMC: acceptance criterion 10^1 CFU/10 mg (*2.6.12*).

Pyrogens (*2.6.8*)
The triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A complies with the test for pyrogens. Inject into each rabbit per kilogram of body mass 3 mL of a solution containing 2.5 µg of 3-*O*-desacyl-4′-monophosphoryl lipid A.

3-O-DESACYL-4′-MONOPHOSPHORYL LIPID A LIQUID BULK

The triethylamine salt of 3-*O*-desacyl-4′-monophosphoryl lipid A is dispersed in a liquid suitable for the subsequent processing steps at a defined target concentration. If the salt is not soluble in water a microfluidisation step is necessary to prepare a stable aqueous suspension.

The liquid bulk is sterilised by filtration through a bacteria-retentive filter.

Only a 3-*O*-desacyl-4′-monophosphoryl lipid A liquid bulk that complies with the requirements given below under Identification, Tests and Assay and that is within the limits approved for the particular product may be used for the preparation of 3-*O*-desacyl-4′-monophosphoryl lipid A in the final lots.

CHARACTERS

When dispersed in an aqueous solution: slightly turbid suspension.

When dissolved in an organic solvent: a description of its appearance is established and approved by the competent authority; the 3-*O*-desacyl-4′-monophosphoryl lipid A liquid bulk complies with this description.

IDENTIFICATION

Congener distribution (see Tests).

TESTS

Particle size
Where applicable, the particle size in the microfluidised liquid bulk is determined by a suitable method, for example dynamic light scattering. The particle size for each batch of liquid bulk is within the limits approved for the particular product.

Sterility (*2.6.1*)
It complies with the test, carried out using 10 mL for each medium.

Congener distribution
The relative amount of tetraacyl, pentaacyl, hexaacyl and heptaacyl congener groups are determined by a suitable method, for example reversed-phase HPLC analysis (*2.2.29*).

The relative amount of each congener group in the 3-*O*-desacyl-4′-monophosphoryl lipid A liquid bulk is:
— tetraacyl: 15 per cent to 35 per cent;
— pentaacyl: 35 per cent to 60 per cent;
— hexaacyl: 20 per cent to 40 per cent;
— heptaacyl: less than 0.5 per cent.

ASSAY

The 3-*O*-desacyl-4′-monophosphoryl lipid A content is determined by a suitable method, for example gas chromatographic quantification (*2.2.28*) of trifluoroacetic anhydride derivatised fatty acid methyl esters of the 3-*O*-desacyl-4′-monophosphoryl lipid A fatty acids dodecanoic acid (C12:0), tetradecanoic acid (C14:0), 3-hydroxy tetradecanoic acid (3-OH-C14:0) and hexadecanoic acid (C16:0) obtained by hydrolysis of 3-*O*-desacyl-4′-monophosphoryl lipid A in an aqueous/methanol (50:50 *V/V*) solution, containing 5 per cent of sodium hydroxide. To the test sample, a reference sample and the dilutions of the calibration curve, pentadecanoic acid (C15:0) is added as an internal standard. The temperature gradient applied must

allow the separation of the fatty acid methyl esters in about 40 min.

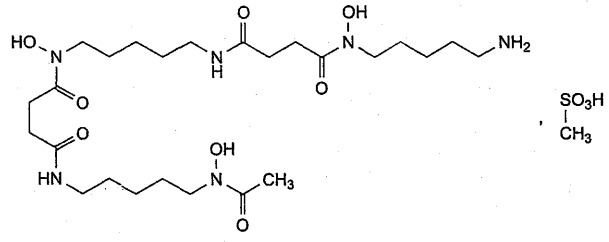
The sum of the ratios between the area for each individual fatty acid methyl ester (C12:0, C14:0, 3-OH-C14:0 and C16:0) and the area of the internal standard (ratio = area C_x / area C15:0) is calculated. The 3-*O*-desacyl-4′-monophosphoryl lipid A quantity corresponding to the sum ratio value on the calibration curve, established with the dilutions of the 3-*O*-desacyl-4′-monophosphoryl lipid A standard, is reported.

The content of 3-*O*-desacyl-4′-monophosphoryl lipid A is not less than 80 per cent and not greater than 120 per cent of the estimated content.

Ph Eur

Desferrioxamine Mesilate

(Deferoxamine Mesilate, Ph. Eur. monograph 0896)



$C_{26}H_{52}N_6O_{11}S$ 657 *138-14-7*

Action and use
Chelating agent (iron).

Preparation
Desferrioxamine Injection

Ph Eur

DEFINITION
N^1-[5-(4-[(5-aminopentyl)(hydroxy)amino]-4-oxobutanamido)pentyl]-N^1-hydroxy-N^4-[5-(*N*-hydroxyacetamido)pentyl]butanediamide methanesulfonate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION
It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in deferoxamine mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation.
The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Freely soluble in water, slightly soluble in methanol, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *deferoxamine mesilate CRS.*

B. Dissolve 0.1 g in 5 mL of *dilute hydrochloric acid R.* Add 1 mL of *barium chloride solution R2.* The solution is clear. In a porcelain crucible, mix 0.1 g with 1 g of *anhydrous sodium carbonate R*, heat and ignite over a naked flame. Allow to cool. Dissolve the residue in 10 mL of *water R*, heating if necessary, and filter. The filtrate gives reaction (a) of sulfates (*2.3.1*).

TESTS
Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 6.0.

Dilute 1 mL of freshly prepared solution S to 10 mL with *water R.*

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use, protected from light.*

A. Deferoxamine mesilate produced by fermentation.

Solvent mixture *acetonitrile for chromatography R, water for chromatography R* (6:94 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *deferoxamine for system suitability CRS* (containing impurity A) in mobile phase A and dilute to 5.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.075 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3.5 µm);
- *temperature*: 32 °C.

Mobile phase:
- *mobile phase A*: 1.32 g/L solution of *ammonium phosphate R* adjusted to pH 3.0 with *phosphoric acid R*;
- *mobile phase B*: mixture of equal volumes of *acetonitrile for chromatography R* and mobile phase A;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	88	12
4 - 20	88 → 80	12 → 20
20 - 35	80 → 57.5	20 → 42.5

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to deferoxamine (retention time = about 18 min): impurity A = about 0.9.

System suitability Reference solution (b):

— *resolution*: minimum 2.0 between the peaks due to impurity A and deferoxamine.

Calculation of percentage contents:
— for each impurity, use the concentration of deferoxamine in reference solution (a).

Limits:
— *impurity A*: maximum 2.0 per cent;
— *any other impurity*: for each impurity, maximum 0.8 per cent;
— *total*: maximum 5.0 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

B. Deferoxamine mesilate produced by a synthetic process.

Solvent mixture Mobile phase B, mobile phase A (20:80 *V/V*).

Test solution Dissolve 0.125 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 12.5 mg of *deferoxamine for peak identification CRS* (containing impurities F, G, H, I and J) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 25 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.605 g of *sodium edetate R* in 900 mL of *water for chromatography R*, add 1.0 mL of *phosphoric acid R* and adjust to pH 6.0 with *concentrated ammonia R*; dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: dissolve 0.780 g of *sodium edetate R* in 750 mL of *water for chromatography R*, add 1.0 mL of *phosphoric acid R*; add 250 mL of *acetonitrile for chromatography R*, mix thoroughly and adjust the apparent pH to 6.0 with *concentrated ammonia R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0	80	20
0 - 21.6	80 → 20	20 → 80
21.6 - 31.6	20	80

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *deferoxamine for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities F, G, H, I and J.

Relative retention With reference to deferoxamine (retention time = about 12.9 min): impurity I = about 0.5; impurity F = about 0.96; impurity G = about 0.98; impurity H = about 1.2; impurities J and K = about 1.37.

System suitability Reference solution (b):

— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to deferoxamine.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity I by 1.4;
— for each impurity, use the concentration of deferoxamine in reference solution (a).

Limits:
— *impurities F, H*: for each impurity, maximum 0.5 per cent;
— *impurities G, I*: for each impurity, maximum 0.3 per cent;
— *sum of impurities J and K*: maximum 0.5 per cent;
— *unspecified impurities*: for each impurity, maximum 0.20 per cent;
— *total*: maximum 2.0 per cent;
— *reporting threshold*: 0.03 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides (*2.4.4*)
Maximum 330 ppm.

Dilute 2 mL of solution S to 20 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 400 ppm.

Dilute 5 mL of solution S to 20 mL with *distilled water R*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.025 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. Since deferoxamine mesilate has an inhibitory effect on the bacterial endotoxins test, a suitable procedure is put in place to remove this inhibitory effect. The monocyte-activation test (*2.6.30*) has been found suitable to overcome this issue.

ASSAY

Dissolve 0.500 g in 50 mL of *water R*. Add 4 mL of *dilute sulfuric acid R1*. Titrate with 0.1 M ferric ammonium sulfate. To accelerate the titration, add 4.5 mL quickly, stop for 1.5 min and continue to titrate uniformly at a rate of about 0.2 mL/min. Determine the end-point potentiometrically (*2.2.20*) using a platinum indicator electrode and a suitable reference electrode.

1 mL of 0.1 M ferric ammonium sulfate is equivalent to 65.68 mg of $C_{26}H_{52}N_6O_{11}S$.

STORAGE

Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

The label states the origin of the substance:
— produced by fermentation;
— produced by a synthetic process.

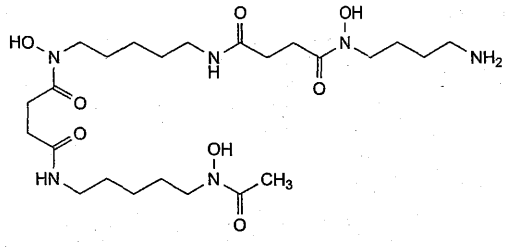
IMPURITIES

Test A for related substances: A, B, C, J.

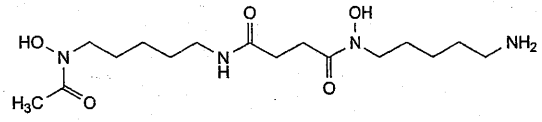
Test B for related substances: E, F, G, H, I, J, K.

Specified impurities *A, F, G, H, I, J, K.*

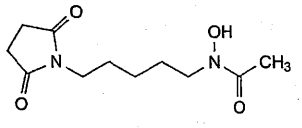
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, E.*



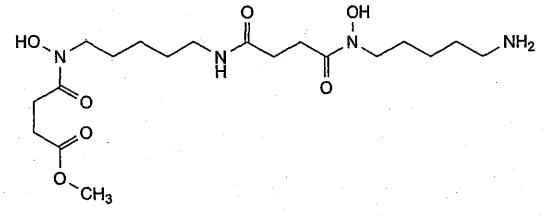
A. N^1-[5-(4-[(4-aminobutyl)(hydroxy)amino]-4-oxobutanamido)pentyl]-N^1-hydroxy-N^4-[5-(*N*-hydroxyacetamido)pentyl]butanediamide,



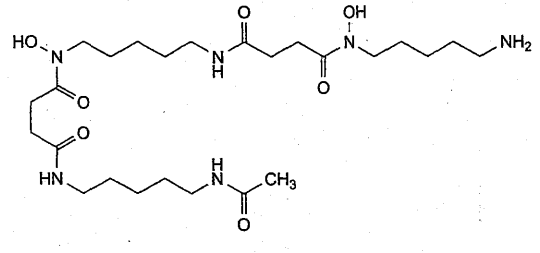
B. N^1-(5-aminopentyl)-N^1-hydroxy-N^4-[5-(*N*-hydroxyacetamido)pentyl]butanediamide,



C. *N*-[5-(2,5-dioxopyrrolidin-1-yl)pentyl]-*N*-hydroxyacetamide,

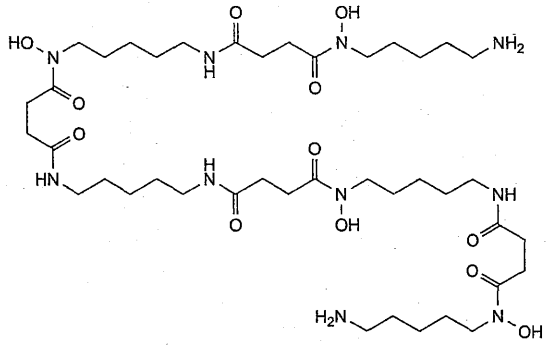


E. methyl 4-[[5-[[4-[(5-aminopentyl)(hydroxy)amino]-4-oxobutanamido]pentyl](hydroxy)amino]-4-oxobutanoate,

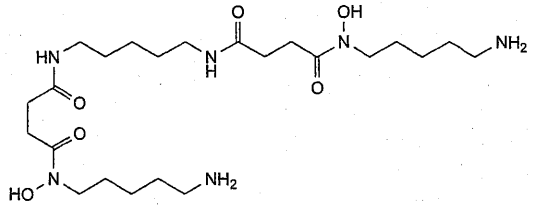


F. N^1-(5-acetamidopentyl)-N^4-[5-[4-[(5-aminopentyl)(hydroxyl)amino]-4-oxobutanamido]pentyl]-N^4-hydroxybutanediamide,

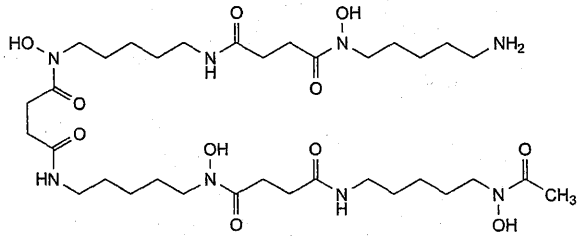
G. unknown structure,



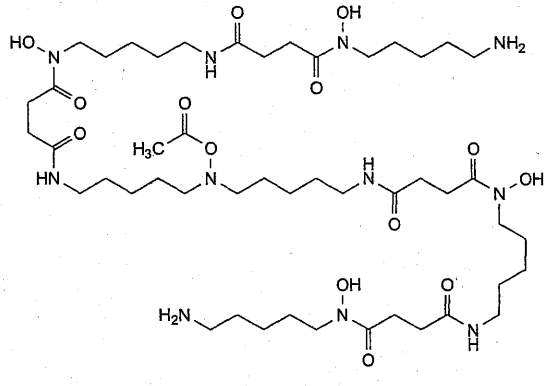
H. N^1,N^{37}-bis(5-aminopentyl)-N^1,N^{37},11,27-tetrahydroxy-4,12,15,23,26,34-hexaoxo-5,11,16,22,27,33-hexaazaheptatriacontane-1,37-diamide,



I. N^1-(5-aminopentyl)-N^4-[5-(4-[(5-aminopentyl)(hydroxy)amino]-4-oxobutanamido)pentyl]- N^1-hydroxybutanediamide,



J. N^1-(5-aminopentyl)-N^1,11,22-trihydroxy-N^{26}-[5-(*N*-hydroxyacetamido)pentyl]-4,12,15,23-tetraoxo-5,11,16,22-tetraazahexacosane-1,26-diamide,

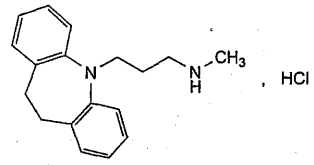


K. 22-(acetyloxy)-N^1,N^{43}-bis(5-aminopentyl)-N^1,N^{43},11,33-tetrahydroxy-4,12,15,29,32,40-hexaoxo-5,11,16,22,28,33,39-heptaazatritetracontane-1,43-diamide.

Ph Eur

Desipramine Hydrochloride

(Ph. Eur. monograph 0481)



$C_{18}H_{23}ClN_2$ 302.8 *58-28-6*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparation
Desipramine Tablets

Ph Eur

DEFINITION
Desipramine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 3-(10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*-methylpropan-1-amine hydrochloride, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, soluble in water and in alcohol.

It melts at about 214 °C.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Dissolve 40.0 mg in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of the solution to 100.0 mL with *0.01 M hydrochloric acid.* Examined between 230 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 251 nm and a shoulder at 270 nm. The specific absorbance at the maximum is 255 to 285.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *desipramine hydrochloride CRS.*

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 50 mg in 3 mL of *water R* and add 0.05 mL of a 25 g/L solution of *quinhydrone R* in *methanol R.* An intense pink colour develops within about 15 min.

E. To 0.5 mL of solution S (see Tests) add 1.5 mL of *water R.* The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 1.25 g in *carbon dioxide-free water R*, warming to not more than 30 °C if necessary, and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S, examined immediately after preparation, is not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *methyl red solution R* and 0.3 mL of *0.01 M sodium hydroxide.* The solution is yellow. Not more than 0.5 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Related substances
Carry out the test protected from bright light Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel plate R.*

Test solution (a) Dissolve 0.10 g of the substance to be examined in a mixture of equal volumes of *ethanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents. *Prepare immediately before use.*

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of equal volumes of *ethanol R* and *methylene chloride R.*

Reference solution (a) Dissolve 25 mg of *desipramine hydrochloride CRS* in a mixture of equal volumes of *ethanol R* and *methylene chloride R* and dilute to 25 mL with the same mixture of solvents. *Prepare immediately before use.*

Reference solution (b) Dilute 1 mL of reference solution (a) to 50 mL with a mixture of equal volumes of *ethanol R* and *methylene chloride R.*

Apply to the plate 5 µL of each solution. Develop over a path of 7 cm using a mixture of 1 volume of *water R*, 10 volumes of *anhydrous acetic acid R* and 10 volumes of *toluene R.* Dry the plate in a current of air for 10 min, spray with a 5 g/L solution of *potassium dichromate R* in a mixture of 1 volume of *sulfuric acid R* and 4 volumes of *water R* and examine immediately. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.2500 g in a mixture of 5 mL of *0.01 M hydrochloric acid* and 50 mL of *alcohol R.* Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide.* Read the volume added between the two points of inflexion.

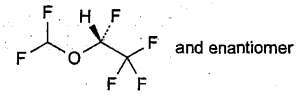
1 mL of *0.1 M sodium hydroxide* is equivalent to 30.28 mg of $C_{18}H_{23}ClN_2$.

STORAGE
Store protected from light.

Ph Eur

Desflurane

(Ph. Eur. monograph 1666)



$C_3H_2F_6O$ 168.0 *57041-67-5*

Ph Eur

DEFINITION
(2*RS*)-2-(Difluoromethoxy)-1,1,1,2-tetrafluoroethane.

CHARACTERS

Appearance
Clear, colourless, mobile, heavy liquid.

Solubility
Practically insoluble in water, miscible with anhydrous ethanol.

Relative density
1.47, determined at 15 °C.

bp
About 22 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Preparation Examine the substance in the gaseous state.

Comparison *Ph. Eur. reference spectrum of desflurane.*

TESTS

The substance to be examined must be cooled to a temperature below 10 °C and the tests must be carried out at a temperature below 20 °C.

Acidity or alkalinity
To 20 mL add 20 mL of *carbon dioxide-free water R*, shake for 3 min and allow to stand. Collect the upper layer and add 0.2 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.01 M sodium hydroxide* or 0.6 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Related substances
Gas chromatography (*2.2.28*).

Test solution The substance to be examined.

Reference solution (a) Introduce 25 mL of the substance to be examined into a 50 mL flask fitted with a septum, and add 0.50 mL of *desflurane impurity A CRS* and 1.0 mL of *isoflurane CRS* (impurity B). Add 50 µL of *acetone R* (impurity H), 10 µL of *chloroform R* (impurity F) and 50 µL of *methylene chloride R* (impurity E) to the solution, using an airtight syringe, and dilute to 50.0 mL with the substance to be examined. Dilute 5.0 mL of this solution to 50.0 mL with the substance to be examined. Store at a temperature below 10 °C.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the substance to be examined. Store at a temperature below 10 °C.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 25.0 mL with the substance to be examined. Store at a temperature below 10 °C.

Column:
— *material*: fused silica;
— *size*: *l* = 105 m, Ø = 0.32 mm;
— *stationary phase*: *poly[methyl(trifluoropropylmethyl)siloxane] R* (film thickness 1.5 µm).

Carrier gas *helium for chromatography R.*

Flow rate 2.0 mL/min.

Split ratio 1:25.

Temperature:
— *column*: 30 °C;
— *injection port*: 150 °C;
— *detector*: 200 °C.

Detection Flame ionisation.

Injection 2.0 µL.

Run time 35 min.

Relative retention With reference to desflurane (retention time = about 11.5 min): impurity C = about 1.06; impurity D = about 1.09; impurity A = about 1.14; impurity G = about 1.39; impurity E = about 1.5; impurity B = about 1.7; impurity F = about 2.2; impurity H = about 2.6.

System suitability Reference solution (a):
— *number of theoretical plates*: minimum 20 000, calculated for the peak due to impurity A;
— *symmetry factor*: maximum 2.0 for the peak due to impurity B.

Limits:
— *impurity B*: not more than the difference between the area of the corresponding peak in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution (0.2 per cent *V/V*);
— *impurity A*: not more than the difference between the area of the corresponding peak in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution (0.1 per cent *V/V*);
— *impurities C, D, G*: for each impurity, not more than the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (b) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.01 per cent *V/V*);
— *impurities E, H*: for each impurity, not more than the difference between the area of the corresponding peak in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution (0.01 per cent *V/V*);
— *impurity F*: not more than the difference between the area of the corresponding peak in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution (0.002 per cent *V/V*);
— *unspecified impurities*: for each impurity, not more than 0.5 times the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (b) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.005 per cent *V/V*);
— *sum of impurities other than A, B, C, D, E, F, G and H*: not more than the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (b) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.01 per cent *V/V*);
— *disregard limit*: the difference between the area of the peak due to impurity A in the chromatogram obtained with reference solution (c) and the area of the peak due to impurity A in the chromatogram obtained with the test solution (0.002 per cent *V/V*).

Fluorides
Maximum 10 ppm.

Potentiometry (*2.2.36, Method I*).

Test solution To 10.0 mL in a separating funnel, add 10 mL of a mixture of 30.0 mL of *dilute ammonia R2* and 70.0 mL of *distilled water R*. Shake for 1 min and collect the upper layer. Repeat this extraction procedure twice, collecting the upper layer each time. Adjust the combined upper layers to pH 5.2 with *dilute hydrochloric acid R*. Add 5.0 mL of *fluoride standard solution (1 ppm F) R* and dilute to 50.0 mL with *distilled water R*. To 20.0 mL of this solution add 20.0 mL of

total-ionic-strength-adjustment buffer R and dilute to 50.0 mL with *distilled water R*.

Reference solutions To each of 1.0 mL, 2.0 mL, 3.0 mL, 4.0 mL and 5.0 mL of *fluoride standard solution (10 ppm F) R* add 20.0 mL of *total-ionic-strength-adjustment buffer R* and dilute to 50.0 mL with *distilled water R*.

Indicator electrode Fluoride selective.

Reference electrode Silver-silver chloride.

Carry out the measurements on 20 mL of each solution. Calculate the concentration of fluorides using the calibration curve, taking into account the addition of fluoride to the test solution.

Non-volatile matter

Maximum 100 mg/L.

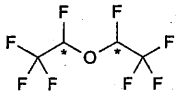
Evaporate 20.0 mL to dryness with the aid of a stream of *nitrogen R*. The residue weighs not more than 2.0 mg.

STORAGE

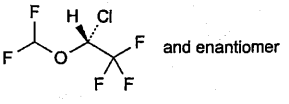
In a glass bottle fitted with a polyethylene-lined cap. Before opening the bottle, cool the contents to below 10 °C.

IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H.*



A. 1,1′-oxybis[(1Ξ)-1,2,2,2-tetrafluoroethane],



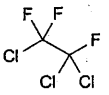
B. (2*RS*)-2-chloro-2-(difluoromethoxy)-1,1,1-trifluoroethane (isofluorane),

C. dichlorofluoromethane,

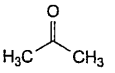
D. trichlorofluoromethane,

E. dichloromethane (methylene chloride),

F. trichloromethane (chloroform),



G. 1,1,2-trichloro-1,2,2-trifluoroethane,

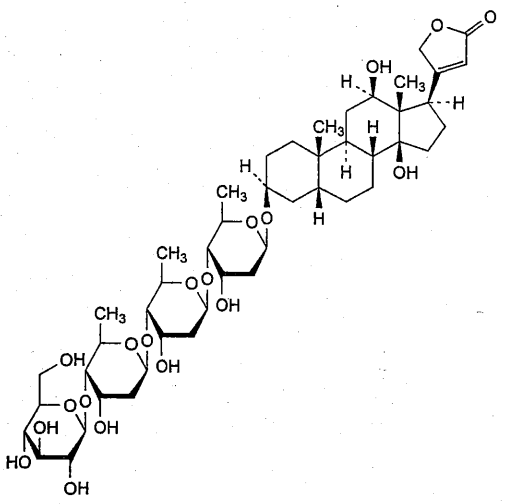


H. propan-2-one (acetone).

Ph Eur

Deslanoside

(Ph. Eur. monograph 0482)



$C_{47}H_{74}O_{19}$ 943 *17598-65-1*

Action and use

Na/K-ATPase inhibitor; cardiac glycoside.

Ph Eur

DEFINITION

Deslanoside contains not less than 95.0 per cent and not more than the equivalent of 105.0 per cent of 3β-[(*O*-β-D-glucopyranosyl-(1→4)-*O*-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-*O*-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β,14β-card-20(22)-enolide, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline or finely crystalline powder, hygroscopic, practically insoluble in water, very slightly soluble in alcohol. In an atmosphere of low relative humidity, it loses water.

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *deslanoside CRS*. When comparing the spectra, special attention is given to the absence of a distinct absorption maximum at about 1260 cm^{-1} and to the intensity of the absorption maximum at about 1740 cm^{-1}. Examine the substances in discs prepared by dissolving 1 mg of the substance to be examined or 1 mg of the reference substance in 0.3 mL of *methanol R* and triturating with about 0.4 g of dry, finely powdered *potassium bromide R* until the mixture is uniform and completely dry.

B. Examine the chromatograms obtained in the test for related substances. The principal zone in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal zone in the chromatogram obtained with reference solution (a).

C. Suspend about 0.5 mg in 0.2 mL of *alcohol (60 per cent V/V) R*. Add 0.1 mL of *dinitrobenzoic acid solution R* and 0.1 mL of *dilute sodium hydroxide solution R*. A violet colour develops.

D. Dissolve about 5 mg in 5 mL of *glacial acetic acid R* and add 0.05 mL of *ferric chloride solution R1*. Cautiously add 2 mL of *sulfuric acid R*, avoiding mixing the two liquids. Allow to stand; a brown but not reddish ring develops at the interface and a greenish-yellow, then bluish-green colour diffuses from it to the upper layer.

TESTS

Solution S

Dissolve 0.20 g in a mixture of equal volumes of *chloroform R* and *methanol R* and dilute to 10 mL with the same mixture of solvents.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

Dissolve 0.200 g in *anhydrous pyridine R* and dilute to 10.0 mL with the same solvent. The specific optical rotation is + 6.5 to + 8.5, calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Use solution S.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of equal volumes of *chloroform R* and *methanol R*.

Reference solution (a) Dissolve 20 mg of *deslanoside CRS* in a mixture of equal volumes of *chloroform R* and *methanol R* and dilute to 10 mL with the same mixture of solvents.

Reference solution (b) Dilute 2.5 mL of reference solution (a) to 10 mL with a mixture of equal volumes of *chloroform R* and *methanol R*.

Reference solution (c) Dilute 1 mL of reference solution (a) to 10 mL with a mixture of equal volumes of *chloroform R* and *methanol R*.

Apply separately to the plate as 10 mm bands 5 µL of each solution. Develop immediately over a path of 15 cm using a mixture of 3 volumes of *water R*, 36 volumes of *methanol R* and 130 volumes of *methylene chloride R*. Dry the plate in a current of warm air, spray with a mixture of 5 volumes of *sulfuric acid R* and 95 volumes of *alcohol R* and heat at 140 °C for 15 min. Examine in daylight. In the chromatogram obtained with test solution (a), any zone, apart from the principal zone, is not more intense than the zone in the chromatogram obtained with reference solution (b) (2.5 per cent) and at most two such zones are more intense than the zone in the chromatogram obtained with reference solution (c) (1.0 per cent).

Loss on drying (*2.2.32*)

Not more than 5.0 per cent, determined on 0.500 g by drying *in vacuo* at 105 °C.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

Dissolve 50.0 mg in *alcohol R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *alcohol R*. Prepare a reference solution in the same manner, using 50.0 mg of *deslanoside CRS* (undried). To 5.0 mL of each solution add 3.0 mL of *alkaline sodium picrate solution R* and allow to stand protected from bright light in a water-bath at 20 ± 1 °C for 40 min. Measure the absorbance (*2.2.25*) of each solution at the maximum at 484 nm, using as the compensation liquid a mixture of 3.0 mL of *alkaline sodium picrate solution R* and 5.0 mL of *alcohol R* prepared at the same time.

Calculate the content of $C_{47}H_{74}O_{19}$ from the absorbances measured and the concentrations of the solutions.

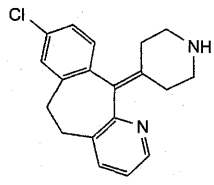
STORAGE

Store in an airtight, glass container, protected from light, at a temperature below 10 °C.

Ph Eur

Desloratadine

(Ph. Eur. monograph 2570)



$C_{19}H_{19}ClN_2$ 310.8 *100643-71-8*

Action and use

Histamine H1, receptor antagonist; antihistamine.

Ph Eur

DEFINITION

8-Chloro-11-(piperidin-4-ylidene)-6,11-dihydro-5*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridine.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble or pratically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble or very slightly soluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison desloratadine CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methyl isobutyl ketone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of *desloratadine CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 4 mg of *desloratadine for system suitability CRS* (containing impurities A and B) in the mobile phase and dilute to 5.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (4 µm);
— *temperature*: 35 °C.

Mobile phase Dissolve 0.865 g of *sodium dodecyl sulfate R* in *water R*, add 0.5 mL of *trifluoroacetic acid R* and dilute to 1000 mL with *water R*; mix 57 volumes of this solution and 43 volumes of *acetonitrile R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 100 µL of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of desloratadine.

Identification of impurities Use the chromatogram supplied with *desloratadine for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to desloratadine (retention time = about 21 min): impurity A = about 0.8; impurity B = about 0.9.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to impurity B and desloratadine.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.6; impurity B = 1.6;
— for each impurity, use the concentration of desloratadine in reference solution (b).

Limits:
— *impurity B*: maximum 0.3 per cent;
— *impurity A*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 0.250 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 0.5 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

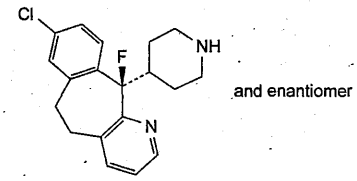
System suitability Reference solution (a):
— *symmetry factor*: 0.5 to 1.5 for the peak due to desloratadine.

Calculate the percentage content of $C_{19}H_{19}ClN_2$ taking into account the assigned content of *desloratadine CRS*.

IMPURITIES

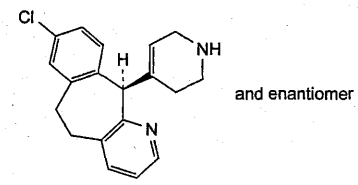
Specified impurities *A, B.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



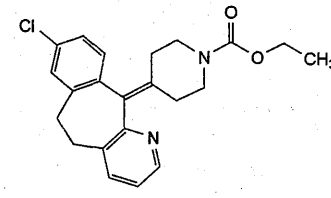
and enantiomer

A. (11*RS*)-8-chloro-11-fluoro-11-(piperidin-4-yl)-6,11-dihydro-5*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridine,



and enantiomer

B. (11*RS*)-8-chloro-11-(1,2,3,6-tetrahydropyridin-4-yl)-6,11-dihydro-5*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridine,

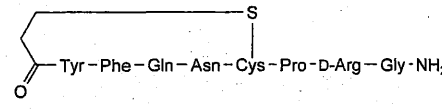


C. ethyl 4-(8-chloro-5,6-dihydro-11*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridin-11-ylidene)piperidine-1-carboxylate (loratadine).

Ph Eur

Desmopressin

(Ph. Eur. monograph 0712)



$C_{46}H_{64}N_{14}O_{12}S_2$ 1069 *16679-58-6*

Action and use
Vasopressin analogue; treatment of diabetes insipidus; nocturnal enuresis; haemophilia; von Willebrand's disease.

Preparations
Desmopressin Injection
Desmopressin Intranasal Solution

Desmopressin Nasal Spray
Desmopressin Oral Lyophilisate
Desmopressin Tablets

Ph Eur

DEFINITION

(3-Sulfanylpropanoyl)-L-tyrosyl-L-phenylalanyl-L-glutaminyl-L-asparaginyl-L-cysteinyl-L-prolyl-D-arginylglycinamide cyclic (1→6)-disulfide.

Synthetic cyclic nonapeptide, available as an acetate.

Content

95.0 per cent to 105.0 per cent (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance

White or almost white, fluffy powder.

Solubility

Soluble in water, in ethanol (96 per cent) and in glacial acetic acid.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The retention time and size of the principal peak in the chromatogram obtained with the test solution are approximately the same as those of the principal peak in the chromatogram obtained with the reference solution.

B. Amino acid analysis (*2.2.56*). For hydrolysis use Method 1 and for analysis use Method 1.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids, taking 1/6 of the sum of the number of moles of aspartic acid, glutamic acid, proline, glycine, arginine and phenylalanine as equal to 1. The values fall within the following limits: aspartic acid: 0.90 to 1.10; glutamic acid: 0.90 to 1.10; proline: 0.90 to 1.10; glycine: 0.90 to 1.10; arginine: 0.90 to 1.10; phenylalanine: 0.90 to 1.10; tyrosine: 0.70 to 1.05; half-cystine: 0.30 to 1.05. Lysine, isoleucine and leucine are absent; not more than traces of other amino acids are present.

TESTS

Specific optical rotation (*2.2.7*)

−72 to −82 (anhydrous and acetic acid-free substance).

Dissolve 10.0 mg in a 1 per cent *V/V* solution of *glacial acetic acid R* and dilute to 5.0 mL with the same acid.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 1.0 mg of the substance to be examined in 2.0 mL of *water R*.

Resolution solution Dissolve the contents of a vial of *oxytocin/desmopressin validation mixture CRS* in 500 µL of *water R*.

Column:
— *size*: *l* = 0.12 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *0.067 M phosphate buffer solution pH 7.0 R*; filter and degas;
— *mobile phase B*: *acetonitrile for chromatography R*, mobile phase A (50:50 *V/V*); filter and degas.

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	76	24
4 - 18	76 → 58	24 → 42
18 - 35	58 → 48	42 → 52
35 - 40	48 → 76	52 → 24
40 - 50	76	24

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 µL.

Retention time Desmopressin = about 16 min; oxytocin = about 17 min.

System suitability Resolution solution:
— *resolution*: minimum 1.5 between the peaks due to desmopressin and oxytoxin.

Limits:
— *unspecified impurities*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 1.5 per cent;
— *disregard limit*: 0.05 per cent.

Acetic acid (*2.5.34*)

3.0 per cent to 8.0 per cent.

Test solution Dissolve 20.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Water (*2.5.32*)

Maximum 6.0 per cent, determined on 20.0 mg.

Bacterial endotoxins (*2.6.14*)

Less than 500 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Reference solution Dissolve the contents of a vial of *desmopressin CRS* in *water R* to obtain a concentration of 0.5 mg/mL.

Mobile phase Mobile phase B, mobile phase A (40:60 *V/V*).

Flow rate 2.0 mL/min.

Retention time Desmopressin = about 5 min.

Calculate the content of desmopressin ($C_{46}H_{64}N_{14}O_{12}S_2$) from the declared content of $C_{46}H_{64}N_{14}O_{12}S_2$ in *desmopressin CRS*.

STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

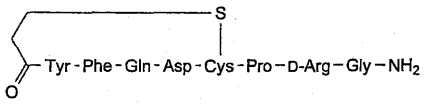
LABELLING

The label states:
— the mass of peptide per container;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

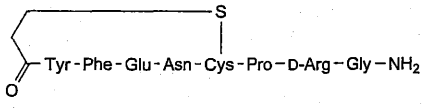
IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

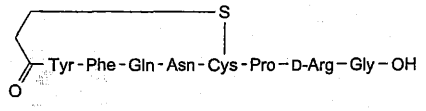
demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



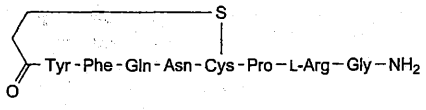
A. [5-L-aspartic acid]desmopressin,



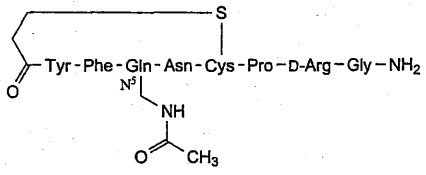
B. [4-L-glutamic acid]desmopressin,



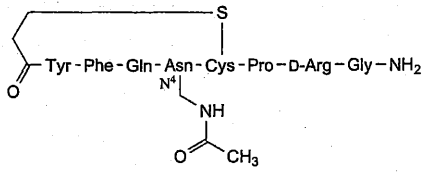
C. [9-glycine]desmopressin,



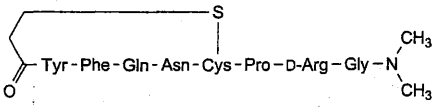
D. [8-L-arginine]desmopressin,



E. $N^{5.4}$-[(acetylamino)methyl]desmopressin,



F. $N^{4.5}$-[(acetylamino)methyl]desmopressin,

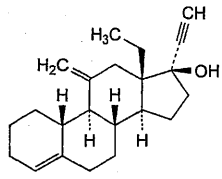


G. $N^{1.9}$,$N^{1.9}$-dimethyldesmopressin.

Ph Eur

Desogestrel

(Ph. Eur. monograph 1717)



$C_{22}H_{30}O$ 310.5 *54024-22-5*

Action and use
Progestogen.

Preparation
Desogestrel Tablets

Ph Eur

DEFINITION

13-Ethyl-11-methylidene-18,19-dinor-17α-pregn-4-en-20-yn-17-ol.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, very soluble in methanol, freely soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *desogestrel CRS.*

B. Specific optical rotation (see Tests).

TESTS

Specific optical rotation (*2.2.7*)
+ 53 to + 57 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in 25 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R.*

Reference solution (a) Dissolve 4 mg of *desogestrel for system suitability CRS* (containing impurities A, B, C and D) in 5 mL of *acetonitrile R1* and dilute to 10.0 mL with *water R.*

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R.*

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with a mixture of equal volumes of *acetonitrile R1* and *water R.*

Reference solution (d) Dissolve 20.0 mg of *desogestrel CRS* in 25 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R.*

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: sterically protected *octadecylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 50 °C.

Mobile phase *water R, acetonitrile R1* (27:73 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 15 µL of the test solution and reference solutions (a), (b) and (c).

Run time 2.5 times the retention time of desogestrel.

Identification of impurities Use the chromatogram supplied with *desogestrel for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to desogestrel (retention time = about 22 min): impurity E = about 0.2; impurity D = about 0.25; impurity B = about 0.7; impurity A = about 0.95; impurity C = about 1.05.

System suitability Reference solution (a):

— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to desogestrel.

Limits:

— *correction factors*: for the calculation of content, multiply the peak area of the following impurities by the corresponding correction factor: impurity A = 1.8, impurity D = 1.5;

— *impurities A, B, C*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at a pressure not exceeding 2 kPa.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

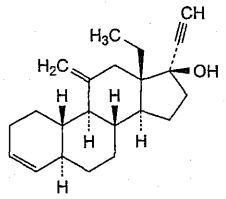
Injection Test solution and reference solution (d).

Calculate the percentage content of $C_{22}H_{30}O$ from the areas of the peaks and the declared content of *desogestrel CRS*.

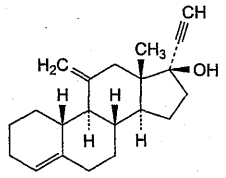
IMPURITIES

Specified impurities *A, B, C, D.*

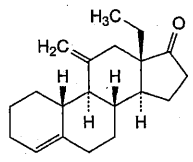
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E.*



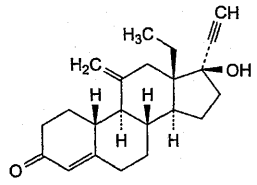
A. 13-ethyl-11-methylidene-18,19-dinor-5α,17α-pregn-3-en-20-yn-17-ol (desogestrel Δ^3-isomer),



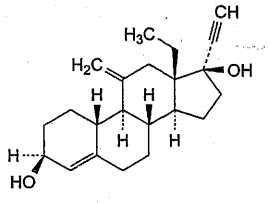
B. 11-methylidene-19-nor-17α-pregn-4-en-20-yn-17-ol,



C. 13-ethyl-11-methylidenegon-4-en-17-one,



D. 13-ethyl-17-hydroxy-11-methylidene-18,19-dinor-17α-pregn-4-en-20-yn-3-one,

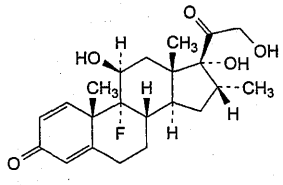


E. 13-ethyl-11-methylidene-18,19-dinor-17α-pregn-4-en-20-yne-3β,17-diol.

Ph Eur

Dexamethasone

(Ph. Eur. monograph 0388)



$C_{22}H_{29}FO_5$ 392.5 *50-02-2*

Action and use

Glucocorticoid.

Preparations
Dexamethasone Eye Drops, Suspension
Dexamethasone and Neomycin Ear Spray
Dexamethasone Tablets
Tobramycin and Dexamethasone Eye Drops, Solution

Ph Eur

DEFINITION

9-Fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene3,20-dione.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in anhydrous ethanol, slightly soluble in methylene chloride.

IDENTIFICATION

First identification: B, C.

Second identification: A, C, D, E.

A. Dissolve 10.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Place 2.0 mL of this solution in a stoppered test tube, add 10.0 mL of *phenylhydrazine-sulfuric acid solution R*, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (*2.2.25*) measured at the absorption maximum at 419 nm is not less than 0.4.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dexamethasone CRS.*

C. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *dexamethasone CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *betamethasone CRS* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *butanol R* saturated with *water R*, *toluene R*, *ether R* (5:10:85 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R.* Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 spots which may, however, not be completely separated.

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a faint reddish-brown colour develops. Add this solution to 10 mL of *water R* and mix; the colour is discharged.

E. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank solution is red.

TESTS

Specific optical rotation (*2.2.7*)
+ 86 to + 92 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution Dissolve 25 mg of the substance to be examined in 1.5 mL of *acetonitrile R* and add 5 mL of mobile phase A. Sonicate until dissolution is complete and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *dexamethasone for system suitability CRS* (containing impurities B, F and G) in 0.5 mL of *acetonitrile R* and add 1 mL of mobile phase A. Sonicate until dissolution is complete and dilute to 2.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *dexamethasone for peak identification CRS* (containing impurities J and K) in 0.5 mL of *acetonitrile R* and add 1 mL of mobile phase A. Sonicate until dissolution is complete and dilute to 2.0 mL with mobile phase A.

Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature:* 45 °C.

Mobile phase:
— *mobile phase A:* mix 250 mL of *acetonitrile R* with 700 mL of *water R* and allow to equilibrate; dilute to 1000.0 mL with *water R* and mix again;
— *mobile phase B: acetonitrile R;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100	0
15 - 40	100 → 0	0 → 100

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL; inject mobile phase A as a blank.

Identification of impurities Use the chromatogram supplied with *dexamethasone for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, F and G; use the chromatogram supplied with *dexamethasone for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities J and K.

Relative retention With reference to dexamethasone (retention time = about 15 min): impurity J = about 0.90; impurity B = about 0.94; impurity K = about 1.3; impurity F = about 1.5; impurity G = about 1.7.

System suitability Reference solution (a):

— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dexamethasone.

Limits:

— *impurity G*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurities B, F, J, K*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 238.5 nm.

Calculate the content of $C_{22}H_{29}FO_5$ taking the specific absorbance to be 394.

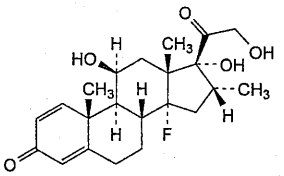
STORAGE

Protected from light.

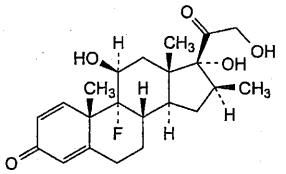
IMPURITIES

Specified impurities *B, F, G, J, K.*

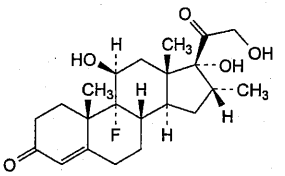
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, H, I.*



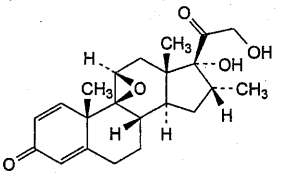
A. 14-fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione,



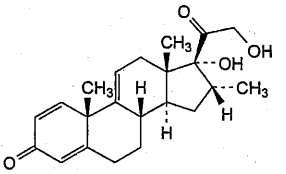
B. 9-fluoro-11β,17,21-trihydroxy-16β-methylpregna-1,4-diene-3,20-dione (betamethasone),



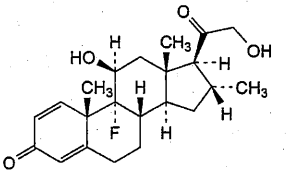
C. 9-fluoro-11β,17,21-trihydroxy-16α-methylpregn-4-ene-3,20-dione,



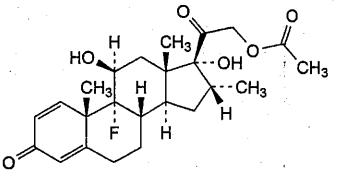
D. 9β,11β-epoxy-17,21-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione,



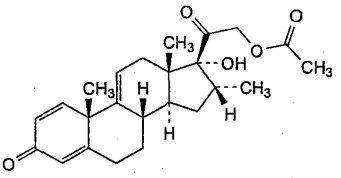
E. 17,21-dihydroxy-16α-methylpregna-1,4,9(11)-triene-3,20-dione,



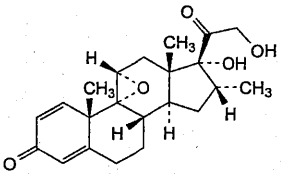
F. 9-fluoro-11β,21-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione,



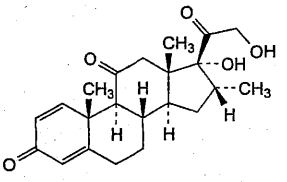
G. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (dexamethasone acetate),



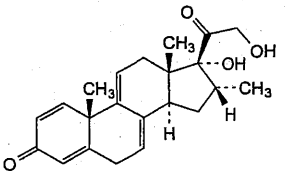
H. 17-hydroxy-16α-methyl-3,20-dioxopregna-1,4,9(11)-trien-21-yl acetate,



I. 9α,11α-epoxy-17,21-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione,



J. 17,21-dihydroxy-16α-methylpregna-1,4-diene-3,11,20-trione,

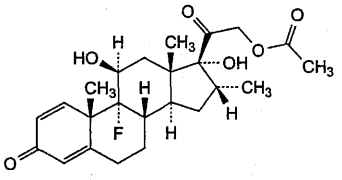


K. 17,21-dihydroxy-16α-methylpregna-1,4,7,9(11)-tetraene-3,20-dione.

Ph Eur

Dexamethasone Acetate

(Ph. Eur. monograph 0548)



$C_{24}H_{31}FO_6$ 434.5 *1177-87-3*

Action and use
Glucocorticoid.

Ph Eur

DEFINITION
9-Fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B, G.

Second identification: A, C, D, E, F.

A. Dissolve 10.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Place 2.0 mL of this solution in a ground-glass-stoppered tube, add 10.0 mL of *phenylhydrazine-sulfuric acid solution R*, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (*2.2.25*) measured at the absorption maximum at 419 nm is not less than 0.35.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dexamethasone acetate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *dexamethasone acetate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *cortisone acetate R* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*, heat at 120 °C for 10 min or until the spots appear, and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a faint reddish-brown colour develops. Add this solution to 10 mL of *water R* and mix. The colour is discharged and a clear solution remains.

E. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

F. About 10 mg gives the reaction of acetyl (*2.3.1*).

G. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (e).

TESTS

Specific optical rotation (*2.2.7*)

+ 94 to + 99 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution (a) Dissolve 25.0 mg of the substance to be examined in about 4 mL of *acetonitrile R* and dilute to 10.0 mL with *water for chromatography R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 5.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *dexamethasone CRS* (impurity A) and 2 mg of *betamethasone acetate CRS* (impurity D) in the mobile phase, using sonication for about 10 min, and dilute to 100.0 mL with the mobile phase. Mix 1.0 mL of this solution and 6.0 mL of test solution (a) and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *dexamethasone acetate impurity E CRS* in 1.0 mL of the mobile phase.

Reference solution (d) Dissolve 5 mg of *dexamethasone acetate for peak identification CRS* (containing impurity I) in about 0.8 mL of *acetonitrile R* and dilute to 2.0 mL with *water for chromatography R*.

Reference solution (e) Dissolve 25.0 mg of *dexamethasone acetate CRS* in about 4 mL of *acetonitrile R* and dilute to 10.0 mL with *water for chromatography R*. Dilute 1.0 mL of the solution to 5.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 380 mL of *acetonitrile R* and 550 mL of *water for chromatography R* and allow to equilibrate; dilute to 1000 mL with *water for chromatography R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b), (c) and (d).

Run time 2.5 times the retention time of dexamethasone acetate.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and D; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity E; use the chromatogram supplied with *dexamethasone acetate for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity I.

Relative retention With reference to dexamethasone acetate (retention time = about 22 min): impurity A = about 0.4; impurity D = about 0.9; impurity E = about 1.2; impurity I = about 1.4.

System suitability Reference solution (a):
- *resolution*: minimum 3.3 between the peaks due to impurity D and dexamethasone acetate.

Limits:
- *impurity I*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
- *impurity D*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurities A, E*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* in an oven at 105 °C.

ASSAY

Carry out the test protected from light.

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase *acetonitrile R, water for chromatography R* (45:55 *V/V*).

Injection Test solution (b) and reference solution (e).

Run time 1.5 times the retention time of dexamethasone acetate.

Retention time Dexamethasone acetate = about 13 min.

Calculate the percentage content of $C_{24}H_{31}FO_6$ taking into account the assigned content of *dexamethasone acetate CRS*.

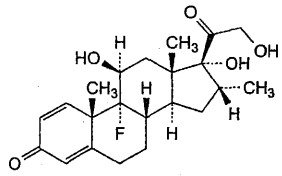
STORAGE

Protected from light.

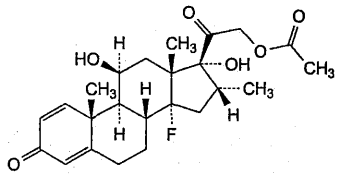
IMPURITIES

Specified impurities *A, D, E, I.*

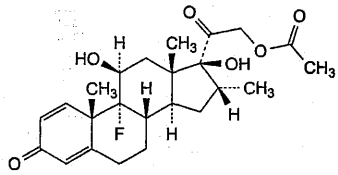
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, F, G, H.*



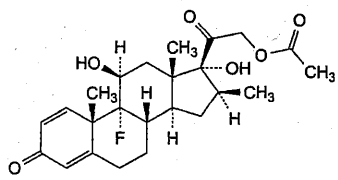
A. 9-fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione (dexamethasone),



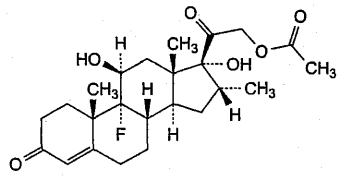
B. 14-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate,



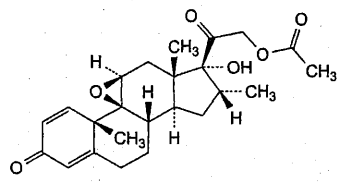
C. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxo-17α-pregna-1,4-dien-21-yl acetate,



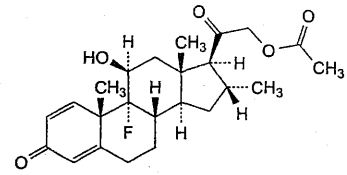
D. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (betamethasone acetate),



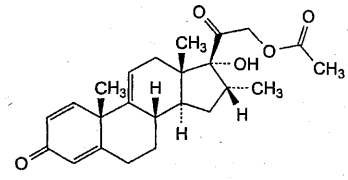
E. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregn-4-en-21-yl acetate,



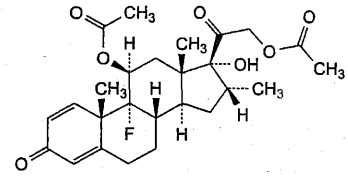
F. 9β,11β-epoxy-17-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate,



G. 9-fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate,



H. 17-hydroxy-16α-methyl-3,20-dioxopregna-1,4,9(11)-trien-21-yl acetate,

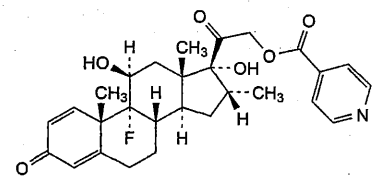


I. 9-fluoro-17-hydroxy-16α-methyl-3,20-dioxopregna-1,4-diene-11β,21-diyl diacetate (dexamethasone 11,21-diacetate).

Ph Eur

Dexamethasone Isonicotinate

(Ph. Eur. monograph 2237)



$C_{28}H_{32}FNO_6$ 497.6 *2265-64-7*

Action and use
Glucocorticoid.

Ph Eur

DEFINITION
9-Fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl pyridine-4-carboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white crystalline powder.

Solubility
Practically insoluble in water, slightly soluble in anhydrous ethanol and in acetone.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *dexamethasone isonicotinate CRS.*

TESTS

Specific optical rotation (*2.2.7*)

+ 142 to + 146 (dried substance).

Suspend 0.200 g in 4.0 mL of *ethyl acetate R* and dilute to 20.0 mL with *ethanol (96 per cent) R*. Treat in an ultrasonic bath until a clear solution is obtained.

Related substances

Liquid chromatography (*2.2.29*). *Prepare solutions immediately before use.*

Test solution Suspend 50.0 mg in 7 mL of *acetonitrile R* and dilute to 10.0 mL with *water R*. Treat in an ultrasonic bath until a clear solution is obtained.

Reference solution (a) Suspend 5.0 mg of *dexamethasone CRS* and 5.0 mg of *dexamethasone acetate CRS* in 70 mL of *acetonitrile R*, add 1.0 mL of the test solution and dilute to 100.0 mL with *water R*. Treat in an ultrasonic bath until a clear solution is obtained.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *water R*.

Reference solution (c) Suspend 5 mg of *dexamethasone isonicotinate for impurity C identification CRS* in 0.7 mL of *acetonitrile R* and dilute to 1 mL with *water R*. Treat in an ultrasonic bath until a clear solution is obtained.

Column:

— *size*: *l* = 0.125 m, Ø = 4.0 mm,

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: *water R*,

— *mobile phase B*: *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	68	32
2 - 20	68 → 50	32 → 50
20 - 25	50 → 68	50 → 32
25 - 35	68	32

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *dexamethasone isonicotinate for impurity C identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity C.

Relative retention With reference to dexamethasone isonicotinate (retention time = about 12 min): impurity A = about 0.4; impurity C = about 0.6; impurity B = about 0.8.

System suitability Reference solution (a):

— *resolution*: minimum 5.0 between the peaks due to impurity B and dexamethasone isonicotinate.

Limits:

— *impurity A*: not more than 5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent),

— *impurity B*: not more than 3 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.3 per cent),

— *impurity C*: not more than 3 times the area of the peak due to dexamethasone isonicotinate in the chromatogram obtained with reference solution (b) (0.3 per cent),

— *unspecified impurities*: for each impurity, not more than the area of the peak due to dexamethasone isonicotinate in the chromatogram obtained with reference solution (b) (0.1 per cent),

— *total*: not more than 8 times the area of the peak due to dexamethasone isonicotinate in the chromatogram obtained with reference solution (b) (0.8 per cent),

— *disregard limit*: 0.5 times the area of the peak due to dexamethasone isonicotinate in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 102 °C under high vacuum for 4 h.

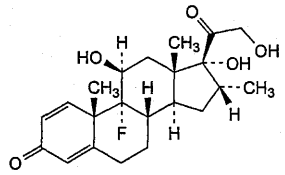
ASSAY

Dissolve 0.400 g in a mixture of 5 mL of *anhydrous formic acid R* and 50 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

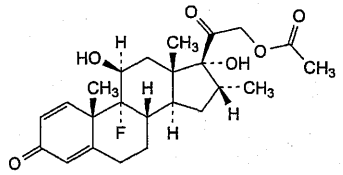
1 mL of *0.1 M perchloric acid* is equivalent to 49.76 mg of $C_{28}H_{32}FNO_6$.

IMPURITIES

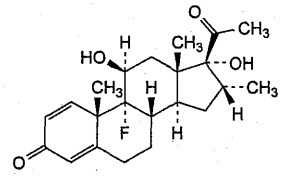
Specified impurities *A, B, C.*



A. 9-fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione (dexamethasone),



B. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (dexamethasone acetate),

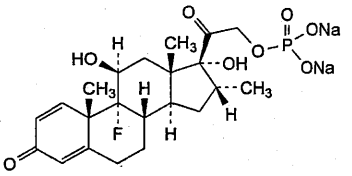


C. 9-fluoro-11β,17-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione (21-deoxydexamethasone).

Ph Eur

Dexamethasone Sodium Phosphate

(Ph. Eur. monograph 0549)



$C_{22}H_{28}FNa_2O_8P$ 516.4 *2392-39-4*

Action and use
Glucocorticoid.

Preparations
Dexamethasone Sodium Phosphate Eye Drops
Dexamethasone Sodium Phosphate Injection
Dexamethasone Sodium Phosphate Oral Solution

Ph Eur

DEFINITION
9-Fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl disodium phosphate.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, very hygroscopic powder.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B, G.

Second identification: A, C, D, E, F.

A. Dissolve 10.0 mg in 5 mL of *water R* and dilute to 100.0 mL with *anhydrous ethanol R*. Place 2.0 mL of this solution in a ground-glass-stoppered tube, add 10.0 mL of *phenylhydrazine-sulfuric acid solution R*, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (*2.2.25*) measured at the absorption maximum at 419 nm is at least 0.20.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dexamethasone sodium phosphate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethanol (96 per cent) R*, evaporate to dryness on a water-bath and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *dexamethasone sodium phosphate CRS* in *methanol R* and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *prednisolone sodium phosphate CRS* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*, heat at 120 °C for 10 min or until the spots appear, and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 spots which may, however, not be completely separated.

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a faint yellowish-brown colour develops. Add this solution to 10 mL of *water R* and mix. The colour fades and a clear solution remains.

E. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

F. To 40 mg add 2 mL of *sulfuric acid R* and heat gently until white fumes are evolved, add *nitric acid R* dropwise, continue the heating until the solution is almost colourless and cool. Add 2 mL of *water R*, heat until white fumes are again evolved, cool, add 10 mL of *water R* and neutralise to *red litmus paper R* with *dilute ammonia R1*. The solution gives reaction (a) of sodium (*2.3.1*) and reaction (b) of phosphates (*2.3.1*).

G. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (b).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).

pH (*2.2.3*)
7.5 to 9.5.

Dilute 1 mL of solution S to 5 mL with *carbon dioxide-free water R*.

Specific optical rotation (*2.2.7*)
+ 75 to + 83 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solution A Dissolve 7.0 g of *ammonium acetate R* in 1000 mL of *water R*.

Test solution Dissolve 10 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 2 mg of *betamethasone sodium phosphate CRS* (impurity B) and 2 mg of *dexamethasone sodium phosphate CRS* in mobile phase A, then dilute to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 2 mg of *dexamethasone sodium phosphate for peak identification CRS* (containing impurities A, C, D, E, F and G) in mobile phase A and dilute to 2.0 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.125 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 300 mL of solution A and 350 mL of *water R*, adjust to pH 3.8 with *acetic acid R*, then add 350 mL of *methanol R*;
— *mobile phase B*: adjust 300 mL of solution A to pH 4.0 with *acetic acid R*, then add 700 mL of *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3.5	90	10
3.5 - 23.5	90 → 60	10 → 40
23.5 - 34.5	60 → 5	40 → 95
34.5 - 50	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *dexamethasone sodium phosphate for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, C, D, E, F and G; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity B.

Relative retention With reference to dexamethasone sodium phosphate (retention time = about 22 min):
impurity C = about 0.5; impurity D = about 0.6;
impurity E = about 0.8; impurity F = about 0.92;
impurity B = about 0.95; impurity A = about 1.37;
impurity G = about 1.41.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity B and dexamethasone sodium phosphate.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.75;
— *impurity A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurity G*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurities B, C, D, E, F*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Inorganic phosphates
Maximum 1 per cent.

Dissolve 50 mg in *water R* and dilute to 100 mL with the same solvent. To 10 mL of this solution add 5 mL of *molybdovanadic reagent R*, mix and allow to stand for 5 min. Any yellow colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using 10 mL of *phosphate standard solution (5 ppm PO_4) R*.

Ethanol (*2.4.24, System A*)
Maximum 1.5 per cent.

Water (*2.5.12*)
Maximum 10.0 per cent, determined on 0.200 g.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *dexamethasone CRS* (impurity A) and 2 mg of *dexamethasone sodium phosphate CRS* in 2 mL of *tetrahydrofuran R*, then dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 30.0 mg of *dexamethasone sodium phosphate CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (7 µm).

Mobile phase Mix 520 mL of *water R* with 2 mL of *phosphoric acid R*. Adjust the temperature to 20 °C, then adjust to pH 2.6 with *sodium hydroxide R*. Mix this solution with 36 mL of *tetrahydrofuran R* and 364 mL of *methanol R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 3 times the retention time of dexamethasone sodium phosphate.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to dexamethasone sodium phosphate (retention time = about 8 min): impurity A = about 2.0.

System suitability Reference solution (a):
— *resolution*: minimum 6.0 between the peaks due to dexamethasone sodium phosphate and impurity A.

Calculate the percentage content of $C_{22}H_{28}FNa_2O_8P$ using the chromatogram obtained with reference solution (b) and taking into account the assigned content of *dexamethasone sodium phosphate CRS*.

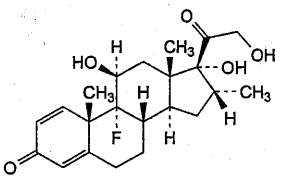
STORAGE

In an airtight container, protected from light.

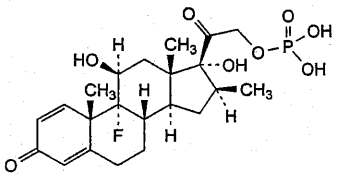
IMPURITIES

Specified impurities *A, B, C, D, E, F, G.*

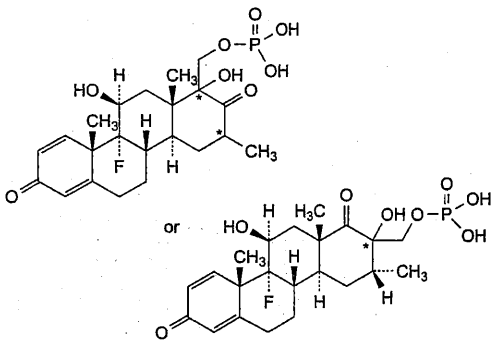
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H.*



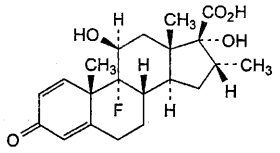
A. 9-fluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione (dexamethasone),



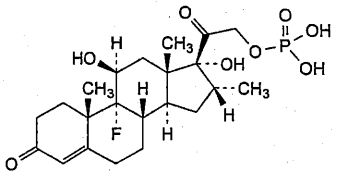
B. 9-fluoro-11β,17-dihydroxy-16β-methyl-3,20-dioxopregna-1,4-dien-21-yl dihydrogen phosphate (betamethasone phosphate),



C, D, E, F. for each impurity, one or more diastereoisomer(s) of (9-fluoro-11β,17a-dihydroxy-16-methyl-3,17-dioxo-*D*-homo-androsta-1,4-dien-17a-yl)methyl dihydrogen phosphate (undefined stereochemistry at C-16 and C-17a), or (9-fluoro-11β,17-dihydroxy-16α-methyl-3,17a-dioxo-*D*-homo-androsta-1,4-dien-17-yl)methyl dihydrogen phosphate (undefined stereochemistry at C-17),



G. 9-fluoro-11β,17-dihydroxy-16α-methyl-3-oxoandrosta-1,4-diene-17β-carboxylic acid,

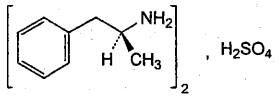


H. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregn-4-en-21-yl dihydrogen phosphate.

Ph Eur

Dexamfetamine Sulfate

(Ph. Eur. monograph 2752)



$C_{18}H_{28}N_2O_4S$	368.5	*51-63-8*

Action and use
Amfetamine.

Preparation
Dexamfetamine Tablets

Ph Eur

DEFINITION

Bis[(2*S*)-1-phenylpropan-2-amine] sulfate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, B, D.

Second identification: C, D.

A. Enantiomeric purity (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dexamfetamine sulfate CRS.*

C. To 50 mL of solution S add 5 mL of *strong sodium hydroxide solution R* and 0.5 mL of *benzoyl chloride R* and shake. Continue to add *benzoyl chloride R* in portions of 0.5 mL, shaking after each addition, until no further precipitate is formed. Filter, wash the precipitate with *water R*, recrystallise twice from a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*, then dry at 100-105 °C. The crystals melt (*2.2.14*) at 155 °C to 160 °C.

D. Solution S (see Tests) gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 25 mL of solution S add 0.1 mL of *methyl red solution R*. Not more than 0.1 mL of *0.01 M hydrochloric acid* or *0.01 M sodium hydroxide* is required to change the colour of the indicator.

Enantiomeric purity

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in *methanol R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 20.0 mL with *methanol R*.

Reference solution (b) Dissolve 5 mg of *amfetamine sulfate CRS* (containing impurity E and dexamfetamine) in 5 mL of *methanol R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *vancomycin-bonded silica gel for chiral separation R* (5 µm);
- *temperature*: 25 °C.

Mobile phase *concentrated ammonia R1, glacial acetic acid R, methanol R2* (0.1:1:998.9 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Run time 1.5 times the retention time of dexamfetamine.

Identification of peaks Use the chromatogram supplied with *amfetamine sulfate CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to dexamfetamine and impurity E.

Relative retention With reference to dexamfetamine (retention time = about 10 min): impurity E = about 1.1.

System suitability Reference solution (b):
- *resolution*: minimum 2.5 between the peaks due to dexamfetamine and impurity E.

Calculation of percentage content:
- for impurity E, use the concentration of dexamfetamine sulfate in reference solution (a).

Limit:
- *impurity E*: maximum 5.0 per cent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mix 5 mL of *trifluoroacetic acid R* and 900 mL of *water for chromatography R*, adjust to pH 2.2 with *concentrated ammonia R* and dilute to 1000 mL with *acetonitrile R*.

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *1-phenylpropan-2-ol R* (impurity A) and 5 mg of *benzaldehyde R* (impurity D) in the solvent mixture and dilute to 10 mL with the solvent mixture. Dilute 1 mL of the solution to 100 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: solvent mixture;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	100	0
1 - 16	100 → 65	0 → 35
16 - 21	65 → 0	35 → 100
21 - 23	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 257 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and D.

Relative retention With reference to dexamfetamine (retention time = about 8 min): impurity D = about 1.6; impurity A = about 1.7.

System suitability Reference solution (b):
- *resolution*: minimum 4.0 between the peaks due to impurities D and A.

Calculation of percentage contents:
- for each impurity, use the concentration of dexamfetamine sulfate in reference solution (a).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.5 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 36.85 mg of $C_{18}H_{28}N_2O_4S$.

STORAGE

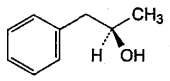
Protected from light.

IMPURITIES

Specified impurities *E.*

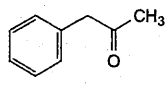
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D.*

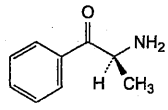


and enantiomer

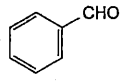
A. (2*RS*)-1-phenylpropan-2-ol,



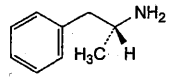
B. 1-phenylpropan-2-one,



C. (2*S*)-2-amino-1-phenylpropan-1-one (cathinone),



D. benzaldehyde,

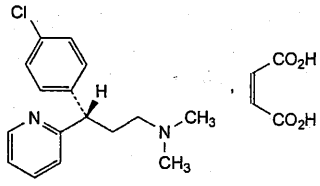


E. (2*R*)-1-phenylpropan-2-amine (levamfetamine).

Ph Eur

Dexchlorpheniramine Maleate

(Ph. Eur. monograph 1196)



$C_{20}H_{23}ClN_2O_4$ 390.9 *2438-32-6*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION
(3*S*)-3-(4-Chlorophenyl)-*N*,*N*-dimethyl-3-(pyridin-2-yl) propan-1-amine (*Z*)-butenedioate.

Content
98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent), in methanol and in methylene chloride.

IDENTIFICATION
First identification: A, C, E.

Second identification: A, B, D, E.

A. Specific optical rotation (see Tests).

B. Melting point (*2.2.14*): 110 °C to 115 °C.

C. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium bromide R.*

Comparison *dexchlorpheniramine maleate CRS.*

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution Dissolve 56 mg of *maleic acid R* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *water R, anhydrous formic acid R, methanol R, di-isopropyl ether R* (3:7:20:70 *V/V/V/V*).

Application 5 µL.

Development Over a path of 12 cm.

Drying In a current of air for a few minutes.

Detection Examine in ultraviolet light at 254 nm.

Results The chromatogram obtained with the test solution shows 2 clearly separated spots. The upper spot is similar in position and size to the spot in the chromatogram obtained with the reference solution.

E. To 0.15 g in a porcelain crucible add 0.5 g of *anhydrous sodium carbonate R.* Heat over an open flame for 10 min. Allow to cool. Take up the residue with 10 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R.* The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 2.0 g in *water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)
4.5 to 5.5.

Dissolve 0.20 g in 20 mL of *water R.*

Specific optical rotation (*2.2.7*)
+ 22 to + 23 (dried substance), determined on solution S.

Related substances
Gas chromatography (*2.2.28*).

Test solution Dissolve 10.0 mg of the substance to be examined in 1.0 mL of *methylene chloride R.*

Reference solution Dissolve 5.0 mg of *brompheniramine maleate CRS* in 0.5 mL of *methylene chloride R* and add 0.5 mL of the test solution. Dilute 0.5 mL of this solution to 50.0 mL with *methylene chloride R.*

Column:
— *material*: glass;
— *size*: *l* = 2.3 m, Ø = 2 mm;
— *stationary phase*: acid- and base-washed *silanised diatomaceous earth for gas chromatography R* (135-175 µm) impregnated with 3 per cent *m/m* of a mixture of 50 per cent of poly(dimethyl)siloxane and 50 per cent of poly(diphenyl)siloxane.

Carrier gas *nitrogen for chromatography R.*

Flow rate 20 mL/min.

Temperature:
— *column*: 205 °C;

— *injection port and detector*: 250 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 2.5 times the retention time of dexchlorpheniramine.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to dexchlorpheniramine and brompheniramine.

Limits:
— *impurity A*: not more than 0.8 times the area of the peak due to dexchlorpheniramine in the chromatogram obtained with the reference solution (0.4 per cent);
— *total*: not more than twice the area of the peak due to dexchlorpheniramine in the chromatogram obtained with the reference solution (1 per cent).

Enantiomeric purity

Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in 3 mL of *water R*. Add a few drops of *concentrated ammonia R* until an alkaline reaction is produced. Shake with 5 mL of *methylene chloride R*. Separate the layers. Evaporate the lower, methylene chloride layer to an oily residue on a water-bath. Dissolve the oily residue in *2-propanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *dexchlorpheniramine maleate CRS* in 3 mL of *water R*. Add a few drops of *concentrated ammonia R* until an alkaline reaction is produced. Shake with 5 mL of *methylene chloride R*. Separate the layers. Evaporate the lower, methylene chloride layer to an oily residue on a water-bath. Dissolve the oily residue in *2-propanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dissolve 10.0 mg of *chlorphenamine maleate CRS* in 3 mL of *water R*. Add a few drops of *concentrated ammonia R* until an alkaline reaction is produced. Shake with 5 mL of *methylene chloride R*. Separate the layers. Evaporate the lower, methylene chloride layer to an oily residue on a water-bath. Dissolve the oily residue in *2-propanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of the test solution to 50 mL with *2-propanol R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *amylose derivative of silica gel for chromatography R*.

Mobile phase *diethylamine R, 2-propanol R, hexane R* (3:20:980 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Under these conditions the peak due to the (*S*)-isomer appears first.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to the (*R*)-enantiomer (impurity B) and the (*S*)-enantiomer in the chromatogram obtained with reference solution (b);
— the retention times of the principal peaks in the chromatograms obtained with the test solution and reference solution (a) are identical ((*S*)-enantiomer).

Limits:
— *(R)-enantiomer* (impurity B): not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 65 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 25 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

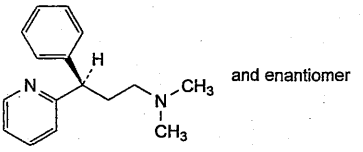
1 mL of *0.1 M perchloric acid* is equivalent to 19.54 mg of $C_{20}H_{23}ClN_2O_4$.

STORAGE

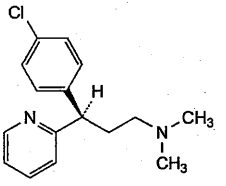
Protected from light.

IMPURITIES

Specified impurities *A, B.*



A. (3*RS*)-*N,N*-dimethyl-3-phenyl-3-(pyridin-2-yl)propan-1-amine,

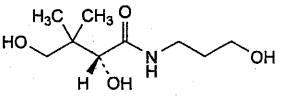


B. (3*R*)-3-(4-chlorophenyl)-*N,N*-dimethyl-3-(pyridin-2-yl) propan-1-amine ((*R*)-enantiomer).

Ph Eur

Dexpanthenol

(Ph. Eur. monograph 0761)



$C_9H_{19}NO_4$ 205.3 *81-13-0*

Action and use

Vitamin B_5 analogue.

Ph Eur

DEFINITION

Dexpanthenol contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of (2*R*)-2,4-dihydroxy-*N*-(3-hydroxypropyl)-3,3-dimethylbutanamide, calculated with reference to the anhydrous substance.

CHARACTERS
A colourless or slightly yellowish, viscous hygroscopic liquid, or a white or almost white, crystalline powder, very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *dexpanthenol CRS*. Examine the substances using discs prepared as follows: dissolve the substance to be examined and the reference substance separately in 1.0 mL of *anhydrous ethanol R* to obtain a concentration of 5 mg/mL. Place dropwise 0.5 mL of this solution on a disc of *potassium bromide R*. Dry the disc at 100-105 °C for 15 min.

C. Examine the chromatograms obtained in the test for 3-aminopropanol. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 1 mL of solution S (see Tests) add 1 mL of *dilute sodium hydroxide solution R* and 0.1 mL of *copper sulfate solution R*. A blue colour develops.

TESTS
Solution S
Dissolve 2.500 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_6 (*2.2.2, Method II*).

pH (*2.2.3*)
The pH of solution S is not greater than 10.5.

Specific optical rotation (*2.2.7*)
The specific optical rotation is + 29.0 to + 32.0, determined on solution S and calculated with reference to the anhydrous substance.

3-Aminopropanol
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.25 g of the substance to be examined in *anhydrous ethanol R* and dilute to 5 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *anhydrous ethanol R*.

Reference solution (a) Dissolve the contents of a vial of *dexpanthenol CRS* in 1.0 mL of *anhydrous ethanol R* to obtain a concentration of 5 mg/mL.

Reference solution (b) Dissolve 25 mg of 3-aminopropanol R in *anhydrous ethanol R* and dilute to 100 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 20 volumes of *concentrated ammonia R*, 25 volumes of *methanol R* and 55 volumes of *butanol R*. Allow the plate to dry in air, spray with a 100 g/L solution of *trichloroacetic acid R* in *methanol R* and heat at 150 °C for 10 min. Spray with a 1 g/L solution of *ninhydrin R* in *methanol R* and heat at 120 °C until a colour appears. Any spot due to 3-aminopropanol in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Water (*2.5.12*)
Not more than 1.0 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
To 0.400 g add 50.0 mL of *0.1 M perchloric acid*. Boil under a reflux condenser for 5 h protected from humidity. Allow to cool. Add 50 mL of *dioxan R* by rinsing the condenser, protected from humidity. Add 0.2 mL of *naphtholbenzein solution R* and titrate with 0.1 M potassium hydrogen phthalate until the colour changes from green to yellow. Carry out a blank titration.

1 mL of *0.1 M perchloric acid* is equivalent to 20.53 mg of $C_9H_{19}NO_4$.

STORAGE
In an airtight container.

Ph Eur

Dextran 1 for Injection

(Ph. Eur. monograph 1506)

Action and use
Plasma substitute.

Ph Eur

DEFINITION
Low-molecular-weight fraction of dextran, consisting of a mixture of isomaltooligosaccharides.

Average relative molecular mass About 1000.

PRODUCTION
It is obtained by hydrolysis and fractionation of dextrans produced by fermentation of sucrose using *Leuconostoc mesenteroides* strain NRRL B-512 = CIP 78.59 or substrains thereof (for example *L. mesenteroides* B-512 F = NCTC 10817).

It is prepared in conditions designed to minimise the risk of microbial contamination.

CHARACTERS
Appearance
White or almost white hygroscopic powder.

Solubility
Very soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Dissolve 3.000 g in *water R*, heat on a water-bath and dilute to 100.0 mL with the same solvent. The specific optical rotation (*2.2.7*) is + 148 to + 164, calculated with reference to the dried substance. Dry an aliquot of the solution first on a water-bath and then to constant weight *in vacuo* at 70 °C. Calculate the dextran content after correction for the content of sodium chloride.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation To 1-2 mg add 1 or a few drops of *water R*. Grind in an agate mortar for 1-2 min. Add about 300 mg of *potassium bromide R* and mix to a slurry but do not grind. Dry *in vacuo* at 40 °C for 15 min. Crush the residue. If it is not dry, dry for another 15 min. Prepare a disc with the residue.

Comparison Repeat the operations using *dextran 1 CRS*.

C. Molecular-mass distribution (see Tests).

TESTS

Solution S

Dissolve 7.5 g in *carbon dioxide-free water R*, heat on a water-bath and dilute to 50 mL with the same solvent.

Absorbance (2.2.25)

Maximum 0.12, determined at 375 nm on solution S.

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution is colourless. Add 0.2 mL of *0.01 M sodium hydroxide*. The solution is pink. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is colourless. Add 0.1 mL of *methyl red solution R*. The solution is red or orange.

Nitrogen-containing substances

Maximum 110 ppm of N.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.200 g and heating for 2 h. Collect the distillate in a mixture of 0.5 mL of *bromocresol green solution R*, 0.5 mL of *methyl red solution R* and 20 mL of *water R*. Titrate with *0.01 M hydrochloric acid*. Not more than 0.15 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Sodium chloride

Maximum 1.5 per cent.

Accurately weigh 3-5 g and dissolve in 100 mL of *water R*. Titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 5.844 mg of NaCl.

Molecular-mass distribution

Size-exclusion chromatography (*2.2.30*).

Test solution Dissolve 6.0-6.5 mg of the substance to be examined in 1.0 mL of the mobile phase.

Reference solution (a) Dissolve 6.0-6.5 mg of *dextran 1 CRS* in 1.0 mL of the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *isomaltooligosaccharide CRS* in 1 mL of the mobile phase, and mix. This corresponds to approximately 45 µg of isomaltotriose (3 glucose units), approximately 45 µg of isomaltononaose (9 glucose units), and approximately 60 µg of sodium chloride per 100 µL.

Column 2 columns coupled in series:
- *size*: l = 0.30 m, Ø = 10 mm;
- *stationary phase*: dextran covalently bound to highly cross-linked porous agarose beads, allowing resolution of oligosaccharides in the molecular mass range of 180 to 3000;
- *temperature*: 20-25 °C.

Mobile phase 2.92 g/L solution of *sodium chloride R*.

Flow rate 0.07-0.08 mL/min maintained constant to ± 1 per cent.

Detection Differential refractometer.

Injection 100 µL.

Identification of peaks Use the chromatogram obtained with reference solution (b) to identify the peaks due to isomaltotriose, isomaltononaose and sodium chloride.

Determine the peak areas. Disregard any peak due to sodium chloride. Calculate the average relative molecular mass M_w and the amount of the fraction with less than 3 and more than 9 glucose units, of *dextran 1 CRS* and of the substance to be examined, using the following expression:

$$M_w = \sum w_i \times m_i$$

M_w = average molecular mass of the dextran;
m_i = molecular mass of oligosaccharide i;
w_i = weight proportion of oligosaccharide i.

Use the following m_i values for the calculation:

Oligosaccharide i	m_i
glucose	180
isomaltose	342
isomaltotriose	504
isomaltotetraose	666
isomaltopentaose	828
isomaltohexaose	990
isomaltoheptaose	1152
isomaltooctaose	1314
isomaltononaose	1476
isomaltodecaose	1638
isomaltoundecaose	1800
isomaltododecaose	1962
isomaltotridecaose	2124
isomaltotetradecaose	2286
isomaltopentadecaose	2448
isomaltohexadecaose	2610
isomaltoheptadecaose	2772
isomaltooctadecaose	2934
isomaltononadecaose	3096

System suitability The values obtained for *dextran 1 CRS* are within the values stated on the label.

Limits:
- *average molecular mass (M_w)*: 850 to 1150;
- *fraction with less than 3 glucose units*: less than 15 per cent;
- *fraction with more than 9 glucose units*: less than 20 per cent.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 5.000 g by drying in an oven at 105 °C for 5 h.

Bacterial endotoxins (2.6.14)

Less than 25 IU/g.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Ph Eur

Dextran 40 for Injection

(Ph. Eur. monograph 0999)

Action and use

Plasma substitute.

Preparation

Dextran 40 Infusion

Ph Eur

DEFINITION

Mixture of polysaccharides, principally of the α-1,6-glucan type.

Average relative molecular mass About 40 000.

PRODUCTION

It is obtained by hydrolysis and fractionation of dextrans produced by fermentation of sucrose using *Leuconostoc mesenteroides* strain NRRL B-512 = CIP 78.59 or substrains

thereof (for example *L. mesenteroides* B-512F = NCTC 10817).

It is prepared in conditions designed to minimise the risk of microbial contamination.

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 195 to + 201 (dried substance).

Dissolve 1.0 g in *water R*, heating on a water-bath, and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dextran CRS*.

C. Molecular-mass distribution (see Tests).

TESTS

Solution S
Dissolve 5.0 g in *distilled water R*, heating on a water-bath, and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution remains colourless. Add 0.2 mL of *0.01 M sodium hydroxide*. The solution is red. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is colourless.
Add 0.1 mL of *methyl red solution R*. The solution is red or orange.

Nitrogen-containing substances
Maximum 110 ppm N.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.200 g and heating for 2 h. Collect the distillate in a mixture of 0.5 mL of *bromocresol green solution R*, 0.5 mL of *methyl red solution R* and 20 mL of *water R*. Titrate with *0.01 M hydrochloric acid*. Not more than 0.15 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Residual solvents
Gas chromatography (*2.2.28*).

Internal standard *propanol R*.

Test solution Dissolve 5 g of the substance to be examined in 100 mL of *water R* and distil. Collect the first 45 mL of the distillate, add 1 mL of a 25 g/L solution of *propanol R* and dilute to 50 mL with *water R*.

Reference solution Mix 0.5 mL of a 25 g/L solution of *anhydrous ethanol R*, 0.5 mL of a 25 g/L solution of *propanol R* and 0.5 mL of a 2.5 g/L solution of *methanol R* and dilute to 25.0 mL with *water R*.

Column:
— *material*: stainless steel;
— *size*: *l* = 1.8 m, Ø = 2 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (125-150 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 25 mL/min.

Temperature:
— *column*: 190 °C;
— *injection port*: 240 °C;
— *detector*: 210 °C.

Detection Flame ionisation.

Injection The chosen volume of each solution.

Limits:
— *ethanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.5 per cent);
— *methanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.05 per cent);
— *sum of solvents other than ethanol, methanol and propanol*: not more than the area of the peak due to the internal standard (0.5 per cent, calculated as propanol).

Molecular-mass distribution (*2.2.39*)
The average molecular mass (M_w) is 35 000 to 45 000. The average molecular mass of the 10 per cent high fraction is not greater than 110 000. The average molecular mass of the 10 per cent low fraction is not less than 7000.

Loss on drying (*2.2.32*)
Maximum 7.0 per cent, determined on 0.200 g by heating in an oven at 105 ± 2 °C for 5 h.

Sulfated ash (*2.4.14*)
Maximum 0.3 per cent, determined on 0.50 g.

Bacterial endotoxins (*2.6.14*)
Less than 10 IU/g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Ph Eur

Dextran 60 for Injection

(Ph. Eur. monograph 1000)

Action and use
Plasma substitute.

Ph Eur

DEFINITION

Mixture of polysaccharides, principally of the α-1,6-glucan type.

Average relative molecular mass About 60 000.

PRODUCTION

It is obtained by hydrolysis and fractionation of dextrans produced by fermentation of sucrose using *Leuconostoc mesenteroides* strain NRRL B-512 = CIP 78.59 or substrains thereof (for example *L. mesenteroides* B-512F = NCTC 10817).

It is prepared in conditions designed to minimise the risk of microbial contamination.

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 195 to + 201 (dried substance).

Dissolve 1.0 g in *water R*, heating on a water-bath, and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dextran CRS*.

C. Molecular-mass distribution (see Tests).

TESTS

Solution S

Dissolve 5.0 g in *distilled water R*, heating on a water-bath, and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution remains colourless. Add 0.2 mL of *0.01 M sodium hydroxide*. The solution is red. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is colourless.
Add 0.1 mL of *methyl red solution R*. The solution is red or orange.

Nitrogen-containing substances

Maximum 110 ppm of N.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.200 g and heating for 2 h. Collect the distillate in a mixture of 0.5 mL of *bromocresol green solution R*, 0.5 mL of *methyl red solution R* and 20 mL of *water R*. Titrate with *0.01 M hydrochloric acid*. Not more than 0.15 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Residual solvents

Gas chromatography (*2.2.28*).

Internal standard *propanol R*.

Test solution Dissolve 5 g of the substance to be examined in 100 mL of *water R* and distil. Collect the first 45 mL of the distillate, add 1 mL of a 25 g/L solution of *propanol R* and dilute to 50 mL with *water R*.

Reference solution Mix 0.5 mL of a 25 g/L solution of *anhydrous ethanol R*, 0.5 mL of a 25 g/L solution of *propanol R* and 0.5 mL of a 2.5 g/L solution of *methanol R* and dilute to 25.0 mL with *water R*.

Column:
— *material*: stainless steel;
— *size*: *l* = 1.8 m, Ø = 2 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (125-150 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 25 mL/min.

Temperature:
— *column*: 190 °C;
— *injection port*: 240 °C;
— *detector*: 210 °C.

Detection Flame ionisation.

Injection The chosen volume of each solution.

Limits:
— *ethanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.5 per cent);
— *methanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.05 per cent);
— *sum of solvents other than ethanol, methanol and propanol*: not more than the area of the peak due to the internal standard (0.5 per cent, calculated as propanol).

Molecular-mass distribution (*2.2.39*)

The average molecular mass (M_w) is 54 000 to 66 000. The average molecular mass of the 10 per cent high fraction is not greater than 180 000. The average molecular mass of the 10 per cent low fraction is not less than 14 000.

Loss on drying (*2.2.32*)

Maximum 7.0 per cent, determined on 0.200 g by heating in an oven at 105 ± 2 °C for 5 h.

Sulfated ash (*2.4.14*)

Maximum 0.3 per cent, determined on 0.50 g.

Bacterial endotoxins (*2.6.14*)

Less than 16 IU/g.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Ph Eur

Dextran 70 for Injection

(Ph. Eur. monograph 1001)

Action and use

Plasma substitute.

Preparation

Dextran 70 Infusion

Ph Eur

DEFINITION

Mixture of polysaccharides, principally of the α-1,6-glucan type.

Average relative molecular mass About 70 000.

PRODUCTION

It is obtained by hydrolysis and fractionation of dextrans produced by fermentation of sucrose using *Leuconostoc mesenteroides* strain NRRL B-512 = CIP 78.59 or substrains thereof (for example *L. mesenteroides* B-512F = NCTC 10817).

It is prepared in conditions designed to minimise the risk of microbial contamination.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 195 to + 201 (dried substance).

Dissolve 1.0 g in *water R*, heating on a water-bath, and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dextran CRS*.

C. Molecular-mass distribution (see Tests).

TESTS

Solution S

Dissolve 5.0 g in *distilled water R*, heating on a water-bath, and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of *phenolphthalein solution R*. The solution remains colourless. Add 0.2 mL of *0.01 M sodium hydroxide*. The solution is red. Add 0.4 mL of

0.01 M hydrochloric acid. The solution is colourless. Add 0.1 mL of *methyl red solution R*. The solution is red or orange.

Nitrogen-containing substances
Maximum 110 ppm of N.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.200 g and heating for 2 h. Collect the distillate in a mixture of 0.5 mL of *bromocresol green solution R*, 0.5 mL of *methyl red solution R* and 20 mL of *water R*. Titrate with *0.01 M hydrochloric acid*. Not more than 0.15 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Residual solvents
Gas chromatography (*2.2.28*).

Internal standard *propanol R*.

Test solution Dissolve 5 g of the substance to be examined in 100 mL of *water R* and distil. Collect the first 45 mL of the distillate, add 1 mL of a 25 g/L solution of *propanol R* and dilute to 50 mL with *water R*.

Reference solution Mix 0.5 mL of a 25 g/L solution of *anhydrous ethanol R*, 0.5 mL of a 25 g/L solution of *propanol R* and 0.5 mL of a 2.5 g/L solution of *methanol R* and dilute to 25.0 mL with *water R*.

Column:
— *material*: stainless steel;
— *size*: *l* = 1.8 m, Ø = 2 mm;
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (125-150 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 25 mL/min.

Temperature:
— *column*: 190 °C;
— *injection port*: 240 °C;
— *detector*: 210 °C.

Detection Flame ionisation.

Injection The chosen volume of each solution.

Limits:
— *ethanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.5 per cent);
— *methanol*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.05 per cent);
— *sum of solvents other than ethanol, methanol and propanol*: not more than the area of the peak due to the internal standard (0.5 per cent, calculated as propanol).

Molecular-mass distribution (*2.2.39*)
The average molecular mass (M_w) is 64 000 to 76 000. The average molecular mass of the 10 per cent high fraction is not greater than 185 000. The average molecular mass of the 10 per cent low fraction is not less than 15 000.

Loss on drying (*2.2.32*)
Maximum 7.0 per cent, determined on 0.200 g by heating in an oven at 105 ± 2 °C for 5 h.

Sulfated ash (*2.4.14*)
Maximum 0.3 per cent, determined on 0.50 g.

Bacterial endotoxins (*2.6.14*)
Less than 16 IU/g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Ph Eur

Dextranomer

(Ph. Eur. monograph 2238)

56087-11-7

Action and use
Fluid absorber; treatment of burns, wounds and skin ulcers; preparation for skin grafting.

Ph Eur

DEFINITION
Three-dimensional network made of dextran chains *O,O'*-cross-linked with 2-hydroxypropane-1,3-diyl bridges and *O*-substituted with 2,3-dihydroxypropyl and 2-hydroxy-1-(hydroxymethyl)ethyl groups.

CHARACTERS
Appearance
White or almost white, spherical beads.

Solubility
Practically insoluble in water. It swells in water and in electrolyte solutions.

PRODUCTION
The absorption capacity is determined using a 9.0 g/L solution of *sodium chloride R* containing 20 µL/L of *polysorbate 20 R* or another suitable solution, with a suitable, validated method.

The particle size is controlled to a minimum of 80 per cent of the number of dry beads within 100-300 µm and a maximum of 7 per cent of their number below 100 µm using a suitable, validated method.

IDENTIFICATION
A. The substance to be examined is practically insoluble in *water R*. It swells in *water R*.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Grind the substance to be examined in *acetone R*. Evaporate the solvent at room temperature and use the residue.

Comparison *dextranomer CRS*.

TESTS
pH (*2.2.3*)
5.3 to 7.5.

Introduce 0.50 g to 30 mL of a freshly prepared 74.6 g/L solution of *potassium chloride R*. Allow to stand for 2 min. Determine the pH on the mucilage obtained.

Boron
Maximum 30 ppm.

Inductively coupled plasma-atomic emission spectrometry (ICP-AES) (*2.2.57*).

Test solution Introduce 3.0 g into a platinum dish and moisten with 5 mL of a 32.1 g/L solution of *magnesium nitrate R* in a mixture of equal volumes of *ethanol (96 per cent) R* and *distilled water R*. Evaporate to dryness on a water-bath. Ignite at 550 °C for 5 h. Take up the residue with 5 mL of *6 M hydrochloric acid R* and transfer to a 50 mL volumetric flask. Add about 20 mL of *distilled water R* and allow to digest for 1 h on a water-bath. Allow to cool and dilute to 50.0 mL with *distilled water R*.

Reference solutions Prepare the reference solutions using a solution of *boric acid R* containing 10 ppm of boron. Proceed as described for the test solution.

Wavelength 249.773 nm.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 15 h.

Sulfated ash (*2.4.14*)
Maximum 0.4 per cent, determined on 1.0 g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*), determined using the pour-plate method.

Ph Eur

Dextrin

(Ph. Eur. monograph 1507)

9004-53-9

Action and use
Excipient.

Ph Eur

DEFINITION
Maize, potato or cassava starch partly hydrolysed and modified by heating with or without the presence of acids, alkalis or pH-control agents.

CHARACTERS
Appearance
White or almost white, free-flowing powder.

Solubility
Very soluble in boiling water forming a mucilaginous solution, slowly soluble in cold water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Suspend 1 g in 50 mL of *water R*, boil for 1 min and cool. To 1 mL of the solution add 0.05 mL of *iodine solution R1*. A dark blue or reddish-brown colour is produced, which disappears on heating.

B. Centrifuge 5 mL of the mucilage obtained in identification test A. To the upper layer add 2 mL of *dilute sodium hydroxide solution R* and, dropwise with shaking, 0.5 mL of *copper sulfate solution R* and boil. A red precipitate is produced.

C. It is very soluble in boiling *water R*, forming a mucilaginous solution.

TESTS
pH (*2.2.3*)
2.0 to 8.0.

Disperse 5.0 g in 100 mL of *carbon dioxide-free water R*.

Chlorides
Maximum 0.2 per cent.

Dissolve 2.5 g in 50 mL of boiling *water R*, dilute to 100 mL with *water R* and filter. Dilute 1 mL of the filtrate to 15 mL with *water R* and add 1 mL of *dilute nitric acid R*. Pour the mixture as a single addition into 1 mL of *silver nitrate solution R2* and allow to stand for 5 min protected from light. When viewed transversely against a black background any opalescence produced is not more intense than that obtained by treating a mixture of 10 mL of *chloride standard solution (5 ppm Cl) R* and 5 mL of *water R*, prepared in the same manner.

Reducing sugars
Maximum 10 per cent, calculated as glucose $C_6H_{12}O_6$.

To a quantity of dextrin equivalent to 2.0 g (dried substance) add 100 mL of *water R*, shake for 30 min, dilute to 200.0 mL with *water R* and filter. To 10.0 mL of alkaline *cupri-tartaric solution R* add 20.0 mL of the filtrate, mix, and heat on a hot plate adjusted to bring the solution to boil within 3 min. Boil for 2 min, and cool immediately. Add 5 mL of a 300 g/L solution of *potassium iodide R* and 10 mL of *dilute sulfuric acid R*, mix, and titrate immediately with *0.1 M sodium thiosulfate*, using *starch solution R*, added towards the end of the titration, as indicator. Repeat the procedure beginning with "To 10.0 mL of...", using, in place of the filtrate, 20.0 mL of a 1 g/L solution of *glucose R*, accurately prepared. Perform a blank titration. ($V_B - V_U$) is not greater than ($V_B - V_S$), in which V_B, V_U and V_S are the number of millilitres of *0.1 M sodium thiosulfate* consumed in the titrations of the blank, the dextrin and the glucose, respectively.

Loss on drying (*2.2.32*)
Maximum 13.0 per cent, determined on 1.000 g by drying at 130 ± 5 °C for 90 min.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for dextrin used as filler and binder, in tablets and capsules.

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)
The following characteristic may be relevant for dextrin used as viscosity-increasing agent.

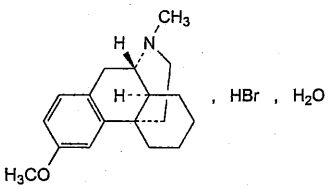
Apparent viscosity (*2.2.10*)
Typically 100 mPa·s to 350 mPa·s (dried substance), depending on the grade of dextrin.

In a beaker, prepare a 10-50 per cent slurry so that the viscosity value ranges from 100 mPa·s to 350 mPa·s. The total mass of the sample plus water must be 600 g. Mix with a plastic rod to obtain a homogeneous slurry. Place the beaker in a water-bath at 100 ± 1 °C. Introduce the paddle of a stirrer into the beaker and close the beaker with a lid. Start agitation at 250 r/min as rapidly as possible and carry on for exactly 30 min. Transfer the paste immediately to the beaker to be used for viscosity measurement, placed in a water-bath at 40 ± 1 °C. Stir until the temperature in the beaker is 40 ± 1 °C then measure the apparent viscosity using spindle no. 2 and a rotation speed of 100 r/min.

Ph Eur

Dextromethorphan Hydrobromide

(Ph. Eur. monograph 0020)



$C_{18}H_{26}BrNO,H_2O$ 370.3 *6700-34-1*

Action and use
Opioid receptor agonist; cough suppressant.

Ph Eur

DEFINITION

ent-3-Methoxy-17-methylmorphinan hydrobromide monohydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Almost white, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in ethanol (96 per cent).

mp
About 125 °C, with decomposition.

IDENTIFICATION

First identification: A, B, D.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dextromethorphan hydrobromide CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 25 mg of *dextromethorphan hydrobromide CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *concentrated ammonia R, methylene chloride R, methanol R, ethyl acetate R, toluene R* (2:10:13:20:55 *V/V/V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *potassium iodobismuthate solution R2.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 1.0 g in *ethanol (96 per cent) R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
Dissolve 0.4 g in *carbon dioxide-free water R* with gentle heating, cool and dilute to 20 mL with the same solvent. Add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Specific optical rotation (*2.2.7*)
+ 28 to + 30 (anhydrous substance).

Dissolve 0.200 g in *0.1 M hydrochloric acid* and dilute to 10.0 mL with the same acid.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *dextromethorphan impurity A CRS* in 2 mL of the test solution and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 3.11 g of *docusate sodium R* in a mixture of 400 mL of *water R* and 600 mL of *acetonitrile R*, add 0.56 g of *sodium nitrate R* and adjust to apparent pH 2.0 with *glacial acetic acid R.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time Twice the retention time of dextromethorphan.

Relative retention With reference to dextromethorphan (retention time = about 22 min): impurity B = about 0.4; impurity C = about 0.8; impurity D = about 0.9; impurity A = about 1.1.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to dextromethorphan and impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 0.2;
— *impurities A, B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent), and not more than 1 such peak has an area greater than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

N,N-Dimethylaniline
Maximum 10 ppm.

Dissolve 0.5 g with heating in 20 mL of *water R*. Allow to cool, add 2 mL of *dilute acetic acid R* and 1 mL of a 10 g/L solution of *sodium nitrite R* and dilute to 25 mL with *water R*. The solution is not more intensely coloured than a reference solution prepared at the same time and in the same manner using 20 mL of a 0.25 mg/L solution of *N,N-dimethylaniline R*.

Water (*2.5.12*)
4.0 per cent to 5.5 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 20 mL of *ethanol (96 per cent) R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

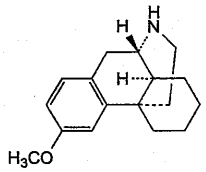
1 mL of *0.1 M sodium hydroxide* is equivalent to 35.23 mg of $C_{18}H_{26}BrNO$.

STORAGE

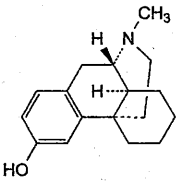
Protected from light.

IMPURITIES

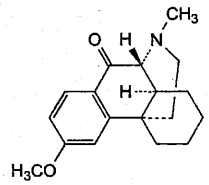
Specified impurities *A, B, C, D.*



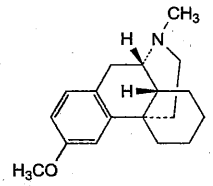
A. *ent*-3-methoxymorphinan,



B. *ent*-17-methylmorphinan-3-ol,



C. *ent*-3-methoxy-17-methylmorphinan-10-one,

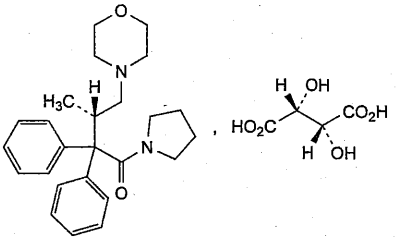


D. *ent*-(14*S*)-3-methoxy-17-methylmorphinan.

Ph Eur

Dextromoramide Tartrate

(Ph. Eur. monograph 0021)



$C_{29}H_{38}N_2O_8$ 542.6 *2922-44-3*

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION

Dextromoramide tartrate contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of 1-[(3*S*)-3-methyl-4-(morpholin-4-yl)-2,2-diphenylbutanoyl] pyrrolidine hydrogen (2*R*,3*R*)-2,3-dihydroxybutanedioate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, amorphous or crystalline powder, soluble in water, sparingly soluble in alcohol.

It melts at about 190 °C, with slight decomposition.

IDENTIFICATION

A. Dissolve 75 mg in *1 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Examined between 230 nm and 350 nm (*2.2.25*), the solution shows 3 absorption maxima, at 254 nm, 259 nm and 264 nm. The specific absorbances at the maxima are about 6.9, 7.7 and 6.5, respectively.

B. Dissolve about 50 mg in *water R* and dilute to 10 mL with the same solvent. To 2 mL of the solution add 3 mL of *ammoniacal silver nitrate solution R* and heat on a water-bath. A grey or black precipitate is formed.

C. It gives reaction (b) of tartrates (*2.3.1*).

TESTS

pH (*2.2.3*)
Dissolve 0.2 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent. The pH of the solution is 3.0 to 4.0.

Specific optical rotation (*2.2.7*)
Dissolve 0.50 g in *0.1 M hydrochloric acid* and dilute to 10.0 mL with the same acid. The specific optical rotation is + 21 to + 23.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution Dissolve 0.2 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dilute 1 mL of the test solution to 100 mL with *methanol R*.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using *methanol R*. Allow the plate to dry in air and spray with *dilute potassium iodobismuthate solution R*. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense

than the spot in the chromatogram obtained with the reference solution (1.0 per cent).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.00 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

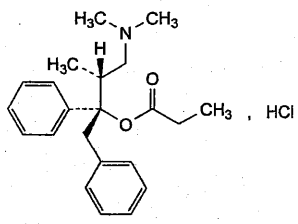
ASSAY
Dissolve 0.250 g in 30 mL of *anhydrous acetic acid R*. Titrate with *0.05 M perchloric acid* using 0.15 mL of *naphtholbenzein solution R* as indicator.

1 mL of *0.05 M perchloric acid* is equivalent to 27.13 mg of $C_{29}H_{38}N_2O_8$.

Ph Eur

Dextropropoxyphene Hydrochloride

(Ph. Eur. monograph 0713)



$C_{22}H_{30}ClNO_2$ 375.9 *1639-60-7*

Action and use
Opioid receptor agonist; analgesic.

Preparation
Co-proxamol Tablets

Ph Eur

DEFINITION
(1*S*,2*R*)-1-Benzyl-3-(dimethylamino)-2-methyl-1-phenylpropyl propanoate hydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

mp
About 165 °C.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *dextropropoxyphene hydrochloride CRS*.

C. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 1.5 g in *carbon dioxide-free water R* and dilute to 30 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
Dilute 10 mL of solution S to 25 mL with *carbon dioxide-free water R*. To 10 mL of this solution add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*.
The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is red.

Specific optical rotation (*2.2.7*)
+ 52 to + 57.

Dissolve 0.100 g in *water R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, methanol R* (50:50 *V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of *dextropropoxyphene for system suitability CRS* (containing impurities A, B, C and D) in 1.0 mL of the solvent mixture.

Reference solution (c) Dilute 1.0 mL of *toluene R* to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 2.5 g of *ammonium phosphate R* in *water R*, adjust to pH 5.6 with *dilute phosphoric acid R* and dilute to 1000 mL with the same solvent;
— *mobile phase B*: *acetonitrile R1*.

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	85	15
2 - 7	85 → 75	15 → 25
7 - 24	75 → 50	25 → 50
24 - 32	50 → 40	50 → 60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *dextropropoxyphene for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and D. Use the chromatogram obtained with reference solution (c) to identify the peak due to toluene.

Relative retention With reference to dextropropoxyphene (retention time = about 18 min): impurity A = about 0.8; impurity B = about 0.9; impurity D = about 1.1; impurity C = about 1.2.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dextropropoxyphene.

Limits:
- — *impurities A, B*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- — *impurities C, D*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- — *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- — *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
- — *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to toluene (relative retention = about 1.24).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

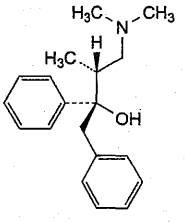
ASSAY
Dissolve 0.270 g in 60 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 37.59 mg of $C_{22}H_{30}ClNO_2$.

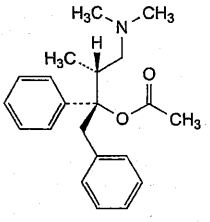
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C, D.*

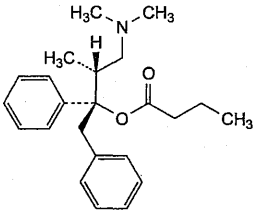
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F.*



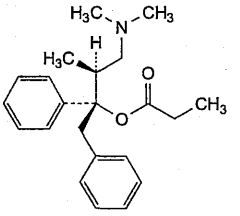
A. (2*S*,3*R*)-4-(dimethylamino)-1,2-diphenyl-3-methyl-butan-2-ol (oxyphene),



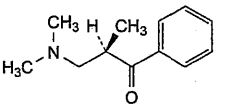
B. (1*S*,2*R*)-1-benzyl-3-(dimethylamino)-2-methyl-1-phenylpropyl acetate (acetoxyphene),



C. (1*S*,2*R*)-1-benzyl-3-(dimethylamino)-2-methyl-1-phenylpropyl butanoate (butyroxyphene),



D. (1*S*,2*S*)-1-benzyl-3-(dimethylamino)-2-methyl-1-phenylpropyl propanoate (isopropoxyphene),

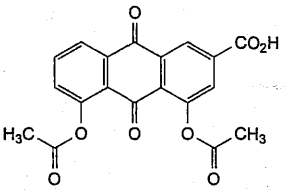

and enantiomer

F. (2*RS*)-3-(dimethylamino)-2-methyl-1-phenylpropan-1-one.

Ph Eur

Diacerein

(Ph. Eur. monograph 2409)



$C_{19}H_{12}O_8$ 368.3 *13739-02-1*

Action and use
Anti-inflammatory used in the treatment of arthritis and osteoarthritis.

Ph Eur

DEFINITION
4,5-Diacetoxy-9,10-dioxo-9,10-dihydroanthracene-2-carboxylic acid.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Yellow, crystalline powder.

Solubility
Practically insoluble in water, soluble in dimethylacetamide, slightly soluble in tetrahydrofuran, practically insoluble in anhydrous ethanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diacerein CRS.*

TESTS

Impurities B and H

Liquid chromatography (*2.2.29*).

Carry out the test protected from light.

Solution A Dissolve 10 g of *sodium hydroxide R* in 500 mL of *water R.*

Solution B Dissolve 14.7 g of *sodium chloride R* and 18.8 g of *glycine R* in 500 mL of *water R.*

Solution C Mix 25.3 volumes of solution A and 74.6 volumes of solution B. If necessary, adjust to pH 9.5 using *dilute sodium hydroxide solution R* or *dilute sulfuric acid R.*

Solution D Dilute 5 mL of *dilute sulfuric acid R* to 500 mL with *water R.*

Test solution Dissolve 0.100 g of the substance to be examined in 30 mL of solution A, mix for 10 min. Add 70 mL of solution B and adjust to pH 9.5 with *dilute sodium hydroxide solution R* or *dilute sulfuric acid R*, if necessary. Extract with 3 quantities, each of 25 mL, of *methylene chloride R.* Combine the methylene chloride extracts and wash with 2 quantities, each of 8 mL, of solution C and then once with 10 mL of solution D. Evaporate the organic layer to dryness at 33 °C, completing the drying procedure using compressed air. Dissolve the residue in 2.0 mL of the mobile phase.

Reference solution (a) Dissolve 7.5 mg of *diacerein impurity B CRS* in *tetrahydrofuran R* and dilute to 25.0 mL with the same solvent. Sonicate for not more than 30 s. Dilute 1.0 mL of the solution to 100.0 mL with solution A. Dilute 5.0 mL of this solution to 50.0 mL with solution A. Mix 5.0 mL of this solution with 25 mL of solution A for 10 min. Add 70 mL of solution B and adjust to pH 9.5 with *dilute sodium hydroxide solution R* or *dilute sulfuric acid R*, if necessary. Perform the extraction as described for the test solution. *Care should be taken that the time between dissolution of diacerein impurity B in tetrahydrofuran and extraction does not exceed 30 min.*

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 5.0 mL with the mobile phase.

Column:
- *size: l* = 0.125 m, Ø = 4.6 mm;
- *stationary phase:* irregular *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature:* 16 ± 1 °C.

Mobile phase *tetrahydrofuran R, acetonitrile R*, 4 g/L solution of *citric acid monohydrate R* (8:27.5:64.5 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 100 µL.

Run time 2.5 times the retention time of impurity B.

Retention time Impurity B = about 11 min.

System suitability Reference solution (b):
- *signal-to-noise ratio:* minimum 10 for the principal peak.

Limit:
- *sum of impurities B and H:* not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) (15 ppm).

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture Mobile phase A, mobile phase B (50:50 *V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in 50 mL of *tetrahydrofuran R* and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *tetrahydrofuran R.* Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) In order to prepare impurities D and E *in situ*, add 10.0 mL of *0.01 M sodium hydroxide* to 0.100 g of the substance to be examined. Add 40 mL of *tetrahydrofuran R* and dilute to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *diacerein impurity mixture CRS* (impurities C and F) in a mixture of 0.5 mL of *tetrahydrofuran R* and 0.5 mL of the solvent mixture.

Column:
- *size: l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm);
- *temperature:* 30 °C.

Mobile phase:
- *mobile phase A:* to 353 mL of *water R* add 147 mL of *phosphoric acid R* and mix; dilute 2 mL of the solution to 1000 mL with *water R;*
- *mobile phase B: acetonitrile R;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	80	20
3 - 13	80 → 60	20 → 40
13 - 20	60	40

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *diacerein impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and F; use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities D and E.

Relative retention With reference to diacerein (retention time = about 13.5 min): impurity D = about 1.1; impurity E = about 1.15; impurity C = about 1.2; impurity F = about 1.3.

System suitability:
- *resolution:* minimum 1.5 between the peaks due to impurities D and E in the chromatogram obtained with reference solution (b);
- *signal-to-noise ratio:* minimum 100 for the principal peak in the chromatogram obtained with reference solution (a).

Limits:
- *correction factors:* for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.4; impurity D = 1.3; impurity E = 1.3; impurity F = 9.5;
- *impurities D, E:* for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurity C:* not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurity F*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 20 times the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chromium

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution In a digestion bomb, dissolve 0.25 g of the substance to be examined in a mixture of 2 mL of *strong hydrogen peroxide solution R* and 6 mL of *nitric acid R*. Mineralise using a microwave oven with a power-incrementing system. Transfer quantitatively to a volumetric flask with *water R* and dilute to 50.0 mL with *water R*. Centrifuge. Dilute 5.0 mL of the clear supernatant to 50.0 mL with *water R*.

Blank solution Prepare as described for the test solution, omitting the substance to be examined.

Stock solution Dilute 5.0 mL of *chromium standard solution (100 ppm Cr) R* to 50.0 mL with *water R*. Dilute 5.0 mL of this solution to 100.0 mL with *water R*. Dilute 2.0 mL of this solution to 100.0 mL with a 0.12 per cent *V/V* solution of *dilute nitric acid R*.

Reference solutions Prepare the reference solutions using the stock solution, diluting with the blank solution.

Source Chromium hollow-cathode lamp using a transmission band preferably of 0.2 nm.

Wavelength 357.9 nm.

Atomisation device Graphite furnace.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Test solution Dissolve 60.0 mg of the substance to be examined in *tetrahydrofuran R* and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 25.0 mL with the solvent mixture.

Reference solution Dissolve 60.0 mg of *diacerein CRS* in *tetrahydrofuran R* and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 25.0 mL with the solvent mixture.

Calculate the percentage content of $C_{19}H_{12}O_8$ taking into account the assigned content of *diacerein CRS*.

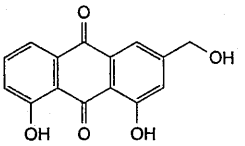
STORAGE

In an airtight container, protected from light.

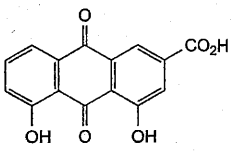
IMPURITIES

Specified impurities *B, C, D, E, F, H.*

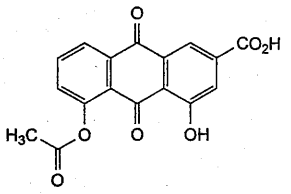
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G.*



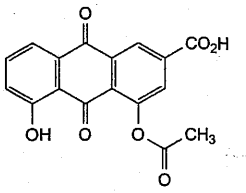
B. 1,8-dihydroxy-3-(hydroxymethyl)-anthracene-9,10-dione (aloe-emodin),



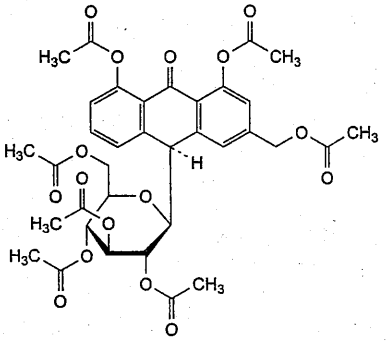
C. 4,5-dihydroxy-9,10-dioxo-9,10-dihydroanthracene-2-carboxylic acid (rhein),



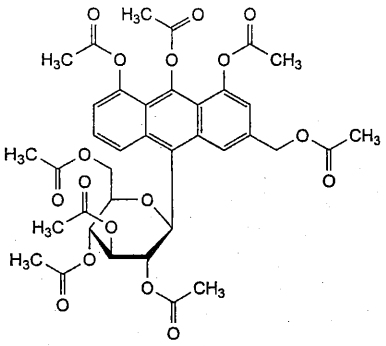
D. 5-acetoxy-4-hydroxy-9,10-dioxo-9,10-dihydroanthracene-2-carboxylic acid (monoacetyl rhein isomer A),



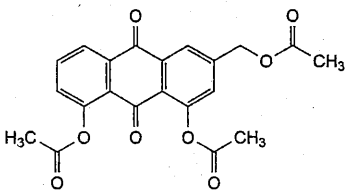
E. 4-acetoxy-5-hydroxy-9,10-dioxo-9,10-dihydroanthracene-2-carboxylic acid (monoacetyl rhein isomer B),



F. (10*S*)-3-(acetoxymethyl)-10-(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)-9-oxo-9,10-dihydroanthracene-1,8-diyl diacetate (heptaacetyl aloin, heptaacetyl barbaloin),



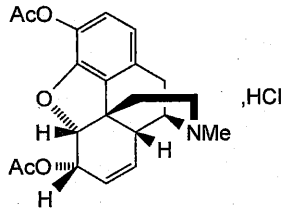
G. 3-(acetoxymethyl)-10-(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)anthracene-1,8,9-triyl triacetate,



H. 3-(acetoxymethyl)-9,10-dioxo-9,10-dihydroanthracene-1,8-diyl diacetate (triacetyl aloe-emodin).

Ph Eur

Diamorphine Hydrochloride



$C_{21}H_{23}NO_5,HCl,H_2O$ 423.9 *1502-95-0*

Action and use
Opioid receptor agonist; analgesic.

Preparations
Bupivacaine and Diamorphine Injection
Diamorphine Tablets
Diamorphine Injection

DEFINITION
Diamorphine Hydrochloride is 4,5-epoxy-17-methylmorphinan-3,6-diyl diacetate hydrochloride monohydrate. It contains not less than 98.0% and not more than 102.0% of $C_{21}H_{23}NO_5,HCl$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Freely soluble in *water*; soluble in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION
A. Dissolve a sufficient quantity in the minimum volume of *dichloromethane* and evaporate to dryness. The *infrared absorption spectrum* of the residue, Appendix II A, is concordant with the *reference spectrum* of diamorphine hydrochloride *(RS 093)*.

B. Yields reaction A characteristic of *chlorides*, Appendix VI.

TESTS
Acidity
Dissolve 0.2 g in 10 mL of *carbon dioxide-free water* and titrate with *0.02M sodium hydroxide VS* using *methyl red solution* as indicator. Not more than 0.2 mL of *0.02M sodium hydroxide VS* is required.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) 0.5% w/v of the substance being examined in *water*.

(2) Dilute 1 volume of solution (1) to 50 volumes with *water*.

(3) A freshly prepared solution containing 0.1% w/v of the substance being examined in 0.01M *sodium hydroxide*.

(4) Dilute 1 volume of solution (2) to 20 volumes with *water*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (12.5 cm × 4.6 mm) packed with *base-deactivated octylsilyl silica gel for chromatography*, (5 µm) (Lichrospher RP-select B is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 283 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for twice the retention time of the peak due to diamorphine hydrochloride.

MOBILE PHASE

0.11 % w/v of *sodium octanesulfonate* in a mixture of 10 volumes of *glacial acetic acid*, 10 volumes of *methanol*, 115 volumes of *acetonitrile* and 365 volumes of *water*.

SYSTEM SUITABILITY

The test is not valid unless:

the chromatogram obtained with solution (3) exhibits two *secondary peaks* with retention times relative to the principal peak of about 0.23 (morphine) and 0.43 (6-*O*-acetylmorphine);

in the chromatogram obtained with solution (3), the *resolution factor* between the peaks due to morphine and 6-*O*-acetylmorphine is at least 2.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to 6-*O*-acetylmorphine is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (2%);

the sum of the areas of any other *secondary peaks* is not greater than 0.25 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

Disregard any peak with an area less than the area of the principal peak in the chromatogram obtained with solution (4) (0.1%).

Loss on drying
When dried to constant weight at 105°, loses 3.0 to 4.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.40 g in 50 mL of *ethanol (96%)* and add 5.0 mL of *0.01M hydrochloric acid VS*. Titrate with *0.1M sodium*

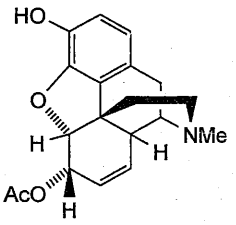
hydroxide VS , determining the end point potentiometrically. Measure the volume of titrant required between the two points of inflection. Each mL of *0.1M sodium hydroxide VS* is equivalent to 40.59 mg of $C_{21}H_{23}NO_5,HCl$.

STORAGE

Diamorphine Hydrochloride should be protected from light.

IMPURITIES

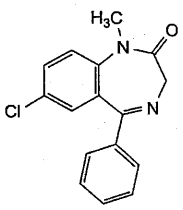
The impurity limited by this monograph is:



A. 6-*O*-acetylmorphine.

Diazepam

(Ph. Eur. monograph 0022)



$C_{16}H_{13}ClN_2O$ 284.7 *439-14-5*

Action and use

Benzodiazepine.

Preparations

Diazepam Injection

Diazepam Oral Solution

Diazepam Rectal Solution

Diazepam Tablets

Ph Eur

DEFINITION

7-Chloro-1-methyl-5-phenyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison: diazepam CRS.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions protected from bright light.*

Test solution Dissolve 25.0 mg of the substance to be examined in 0.5 mL of *acetonitrile R* and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *diazepam for system suitability CRS* (containing impurities A, B and E) in 1.0 mL of the mobile phase.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: spherical *end-capped octylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase Mix 22 volumes of *acetonitrile R*, 34 volumes of *methanol R* and 44 volumes of a 3.4 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 5.0 with *dilute sodium hydroxide solution R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time About 4 times the retention time of diazepam.

Identification of impurities Use the chromatogram supplied with *diazepam for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and E.

Relative retention With reference to diazepam (retention time = about 9 min): impurity E = about 0.7; impurity A = about 0.8; impurity B = about 1.3.

System suitability Reference solution (b):

— *resolution*: minimum 2.5 between the peaks due to impurities E and A and minimum 6.0 between the peaks due to impurity A and diazepam.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.3; impurity E = 1.3;

— *impurities A, B, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

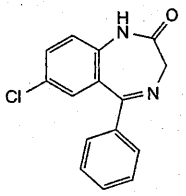
ASSAY

Dissolve 0.200 g in 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

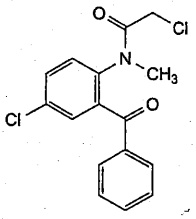
1 mL of *0.1 M perchloric acid* is equivalent to 28.47 mg of $C_{16}H_{13}ClN_2O$.

STORAGE
Protected from light.

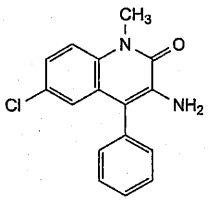
IMPURITIES
Specified impurities *A, B, E.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, F.*



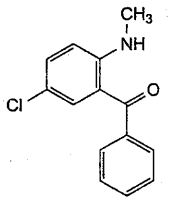
A. 7-chloro-5-phenyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one (nordazepam),



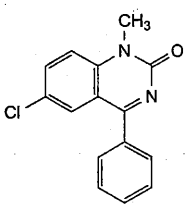
B. *N*-(2-benzoyl-4-chlorophenyl)-2-chloro-*N*-methylacetamide,



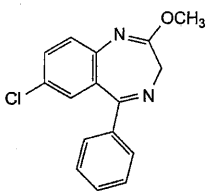
C. 3-amino-6-chloro-1-methyl-4-phenylquinolin-2(1*H*)-one,



D. [5-chloro-2-(methylamino)phenyl]phenylmethanone,



E. 6-chloro-1-methyl-4-phenylquinazolin-2(1*H*)-one,

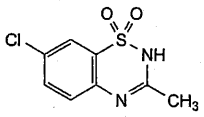


F. 7-chloro-2-methoxy-5-phenyl-3*H*-1,4-benzodiazepine.

Ph Eur

Diazoxide

(Ph. Eur. monograph 0550)



$C_8H_7ClN_2O_2S$ 230.7 *364-98-7*

Action and use
Vasodilator; Treatment of hypertension.

Preparations
Diazoxide Injection
Diazoxide Tablets

Ph Eur

DEFINITION
Diazoxide contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of 7-chloro-3-methyl-2*H*-1,2,4-benzothiadiazine 1,1-dioxide, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, fine or crystalline powder, practically insoluble in water, freely soluble in dimethylformamide, slightly soluble in alcohol. It is very soluble in dilute solutions of the alkali hydroxides.

IDENTIFICATION
First identification: B.
Second identification: A, C, D.

A. Dissolve 50.0 mg in 5 mL of *1 M sodium hydroxide* and dilute to 50.0 mL with *water R*. Dilute 1.0 mL of this solution to 100.0 mL with *0.1 M sodium hydroxide*. Examined between 230 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 280 nm and a shoulder at 304 nm. The specific absorbance at the maximum is 570 to 610.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *diazoxide CRS*. Examine the substances prepared as discs using *potassium bromide R*.

C. Examine the chromatograms obtained in the test for related substances in ultraviolet light at 254 nm.
The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (b).

D. Dissolve about 20 mg in a mixture of 5 mL of *hydrochloric acid R* and 10 mL of *water R*. Add 0.1 g of *zinc powder R*. Boil for 5 min, cool and filter. To the filtrate add 2 mL of a 1 g/L solution of *sodium nitrite R* and mix. Allow to stand for

1 min and add 1 mL of a 5 g/L solution of *naphthylethylenediamine dihydrochloride R*. A red or violet-red colour develops.

TESTS

Appearance of solution

Dissolve 0.4 g in 2 mL of *1 M sodium hydroxide* and dilute to 20 mL with *water R*. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Acidity or alkalinity

To 0.5 g of the powdered substance to be examined add 30 mL of *carbon dioxide-free water R*, shake for 2 min and filter. To 10 mL of the filtrate add 0.2 mL of *0.01 M sodium hydroxide* and 0.15 mL of *methyl red solution R*. The solution is yellow. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel GF_{254} R* as the coating substance.

Test solution (a) Dissolve 0.1 g of the substance to be examined in a mixture of 0.5 mL of *1 M sodium hydroxide* and 1 mL of *methanol R* and dilute to 5 mL with *methanol R*.

Test solution (b) Dilute 1 mL of test solution (a) to 5 mL with a mixture of 1 volume of *1 M sodium hydroxide* and 9 volumes of *methanol R*.

Reference solution (a) Dilute 0.5 mL of test solution (a) to 100 mL with a mixture of 1 volume of *1 M sodium hydroxide* and 9 volumes of *methanol R*.

Reference solution (b) Dissolve 20 mg of *diazoxide CRS* in a mixture of 0.5 mL of *1 M sodium hydroxide* and 1 mL of *methanol R* and dilute to 5 mL with *methanol R*.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 7 volumes of *concentrated ammonia R*, 25 volumes of *methanol R* and 68 volumes of *chloroform R*. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

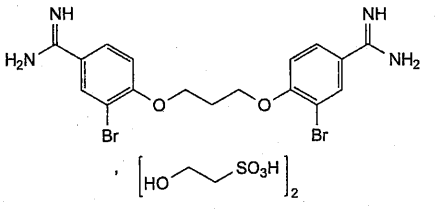
Dissolve 0.200 g with gentle heating in 50 mL of a mixture of 1 volume of *water R* and 2 volumes of *dimethylformamide R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 23.07 mg of $C_8H_7ClN_2O_2S$.

Ph Eur

Dibrompropamidine Isetionate

(Dibrompropamidine Diisetionate, Ph. Eur. monograph 2300)



$C_{21}H_{30}Br_2N_4O_{10}S_2$ 722 *614-87-9*

Action and use

Antiseptic.

Ph Eur

DEFINITION

3,3′-Dibromo-4,4′-[propane-1,3-diylbis(oxy)] dibenzimidamide bis(2-hydroxyethanesulfonate).

Content

99.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

It is considered that alkyl 2-hydroxyethanesulfonate esters are potential impurities in dibrompropamidine diisetionate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation including, where necessary, demonstration that alkyl 2-hydroxyethanesulfonate esters are not detectable in the final product.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble or soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison dibrompropamidine diisetionate CRS.

B. Mix 0.1 g with 0.5 g of *anhydrous sodium carbonate R*, ignite and take up the residue with 20 mL of *water R*. Filter and neutralise the filtrate to *blue litmus paper R* with *nitric acid R*. The filtrate gives reaction (a) of bromides (*2.3.1*).

TESTS

pH (*2.2.3*)

5.0 to 6.0.

Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture anhydrous formic acid R, methanol R, ethyl acetate R (0.01:8:12 *V/V/V*).

Test solution To 8 mL of *methanol R* add 20.0 mg of the substance to be examined and dissolve with the aid of an ultrasonic bath. Add 11 mL of *ethyl acetate R* then 10 µL of *anhydrous formic acid R* and mix. Dilute to 20.0 mL with *ethyl acetate R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *dibrompropamidine for system suitability CRS* (containing impurities A and B) in 4 mL of *methanol R* using an ultrasonic bath. Add 5 mL of *ethyl acetate R*, then 5 µL of *anhydrous formic acid R* and mix. Dilute to 10.0 mL with *ethyl acetate R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *strong cation-exchange silica gel for chromatography R* (5 µm).

Mobile phase Mix 4 volumes of a 25 g/L solution of *ammonium formate R* in *methanol R* and 6 volumes of *ethyl acetate R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 40 µL.

Run time 1.5 times the retention time of dibrompropamidine.

Identification of impurities Use the chromatogram supplied with *dibrompropamidine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to dibrompropamidine (retention time = about 20 min): impurity A = about 0.4; impurity B = about 1.1.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dibrompropamidine.

Limits:
— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity B*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

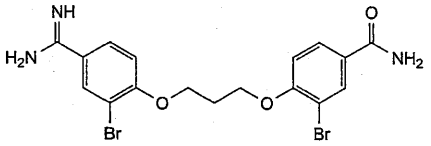
ASSAY

Dissolve 0.250 g in 50 mL of *dimethylformamide R*. Titrate with 0.1 M tetrabutylammonium hydroxide under a current of *nitrogen R*, determining the end-point potentiometrically (*2.2.20*).

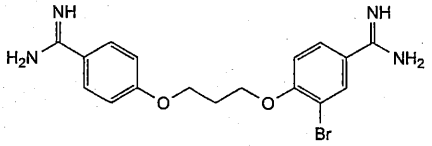
1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 36.12 mg of $C_{21}H_{30}Br_2N_4O_{10}S_2$.

IMPURITIES

Specified impurities *A, B.*



A. 3-bromo-4-[3-(2-bromo-4-carbamimidoylphenoxy)propoxy]benzamide,

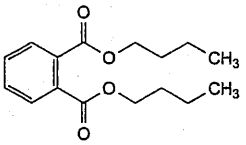


B. 3-bromo-4-[3-(4-carbamimidoylphenoxy) propoxy]benzimidamide.

Ph Eur

Dibutyl Phthalate

(Ph. Eur. monograph 0762)



$C_{16}H_{22}O_4$ 278.3 *84-74-2*

Action and use
Insect repellent.

Ph Eur

DEFINITION

Dibutyl benzene-1,2-dicarboxylate.

Content
99.0 per cent *m/m* to 101.0 per cent *m/m*.

CHARACTERS

Appearance
Clear, oily liquid, colourless or very slightly yellow.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent).

IDENTIFICATION

First identification: B, C.

Second identification: A, D, E.

A. Relative density (*2.2.5*): 1.043 to 1.048.

B. Refractive index (*2.2.6*): 1.490 to 1.495.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dibutyl phthalate CRS.*

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *ether R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of *dibutyl phthalate CRS* in *ether R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *heptane R, ether R* (30:70 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

E. To about 0.1 mL add 0.25 mL of *sulfuric acid R* and 50 mg of *resorcinol R*. Heat in a water-bath for 5 min. Allow to cool. Add 10 mL of *water R* and 1 mL of *strong sodium hydroxide solution R*. The solution becomes yellow or brownish-yellow and shows a green fluorescence.

TESTS

Appearance

The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity

Dissolve 20.0 g in 50 mL of *ethanol (96 per cent) R* previously neutralised to *phenolphthalein solution R1*. Add 0.2 mL of *phenolphthalein solution R1*. Not more than 0.50 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Related substances

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 60 mg of *bibenzyl R* in *methylene chloride R* and dilute to 20 mL with the same solvent.

Test solution (a) Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 20.0 mL with the same solvent.

Test solution (b) Dissolve 1.0 g of the substance to be examined in *methylene chloride R*, add 2.0 mL of the internal standard solution and dilute to 20.0 mL with *methylene chloride R*.

Reference solution To 1.0 mL of test solution (a) add 10.0 mL of the internal standard solution and dilute to 100.0 mL with *methylene chloride R*.

Column:
- *material*: glass;
- *size*: *l* = 1.5 m, Ø = 4 mm;
- *stationary phase*: *silanised diatomaceous earth for gas chromatography R* (150-180 µm) impregnated with 3 per cent *m/m* of *polymethylphenylsiloxane R*.

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Temperature:
- *column*: 190 °C;
- *injection port and detector*: 225 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 3 times the retention time of dibutyl phthalate.

Elution order Bibenzyl, dibutyl phthalate.

Retention time Dibutyl phthalate = about 12 min.

System suitability:
- *resolution*: minimum 12 between the peaks due to bibenzyl and dibutyl phthalate in the chromatogram obtained with the reference solution;
- in the chromatogram obtained with test solution (a), there is no peak with the same retention time as the internal standard.

Limit:
- *total*: calculate the ratio (*R*) of the area of the peak due to dibutyl phthalate to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with test solution (b), calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than *R* (1.0 per cent).

Water (*2.5.12*)

Maximum 0.2 per cent, determined on 10.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Introduce 0.750 g into a 250 mL borosilicate glass flask. Add 25.0 mL of 0.5 M alcoholic potassium hydroxide and a few glass beads. Heat in a water-bath under a reflux condenser for 1 h. Add 1 mL of *phenolphthalein solution R1* and titrate immediately with *0.5 M hydrochloric acid* until the colour changes from red to colourless. Carry out a blank titration. Calculate the volume of potassium hydroxide used in the saponification.

1 mL of 0.5 M alcoholic potassium hydroxide is equivalent to 69.59 mg of $C_{16}H_{22}O_4$.

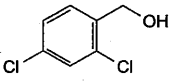
STORAGE

In an airtight container.

Ph Eur

2,4-Dichlorobenzyl Alcohol

(Ph. Eur. monograph 2410)



$C_7H_6Cl_2O$ 177.0 *1777-82-8*

Ph Eur

DEFINITION

(2,4-Dichlorophenyl)methanol.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, very soluble in ethanol (96 per cent).

mp

About 59 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *2,4-dichlorobenzyl alcohol CRS.*

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water R* (50:50 *V/V*).

Buffer solution Dissolve 0.68 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000.0 mL with *water R*.

Test solution (a) Dissolve 0.100 g of the substance to be examined in 10.0 mL of *acetonitrile R1* and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 20.0 mg of *2,4-dichlorobenzyl alcohol impurity A CRS* and 20.0 mg of *2,4-dichlorobenzyl alcohol impurity C CRS* in 100.0 mL of *acetonitrile R1*. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 0.100 g of *2,4-dichlorobenzyl alcohol CRS* in 10.0 mL of *acetonitrile R1* and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *methanol R2, acetonitrile R1*, buffer solution (20:30:50 *V/V/V*);
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	100	0
7 - 17	100 → 20	0 → 80
17 - 30	20	80

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Relative retention With reference to 2,4-dichlorobenzyl alcohol (retention time = about 7 min): impurity C = about 0.87; impurity A = about 0.91.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 4, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity A.

Calculation of percentage contents:
- for each impurity, use the concentration of 2,4-dichlorobenzyl alcohol in reference solution (a).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)

Maximum 0.2 per cent, determined on 0.500 g using the evaporation technique:
- *temperature*: 120 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

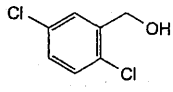
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

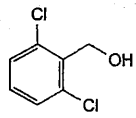
Calculate the percentage content of $C_7H_6Cl_2O$ taking into account the assigned content of *2,4-dichlorobenzyl alcohol CRS*.

IMPURITIES

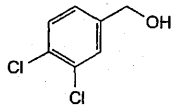
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G.*



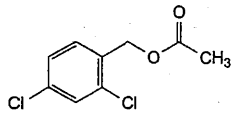
A. (2,5-dichlorophenyl)methanol,



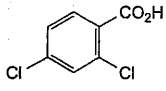
B. (2,6-dichlorophenyl)methanol,



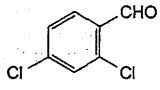
C. (3,4-dichlorophenyl)methanol,



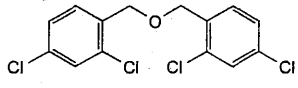
D. 2,4-dichlorobenzyl acetate,



E. 2,4-dichlorobenzoic acid,



F. 2,4-dichlorobenzaldehyde,



G. 1,1′-(oxydimethylene)bis(2,4-dichlorobenzene).

Ph Eur

Dichloromethane

(Methylene Chloride, Ph. Eur. monograph 0932)

CH_2Cl_2 84.9 *75-09-2*

Action and use
Excipient.

Ph Eur

DEFINITION
Dichloromethane.

It may contain maximum 2.0 per cent *V/V* of anhydrous ethanol and/or maximum 0.03 per cent *V/V* of 2-methylbut-2-ene as stabiliser.

CHARACTERS
Appearance
Clear, colourless, volatile liquid.

Solubility
Sparingly soluble in water, miscible with ethanol (96 per cent).

IDENTIFICATION
First identification: B, C.

Second identification: A, D, E.

A. Relative density (see Tests).

B. Refractive index (see Tests).

C. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Films.

Comparison *methylene chloride CRS.*

D. Heat 2 mL with 2 g of *potassium hydroxide R* and 20 mL of *ethanol (96 per cent) R* under a reflux condenser for 30 min. Allow to cool. Add 15 mL of *dilute sulfuric acid R* and filter. To 1 mL of the filtrate add 1 mL of a 15 g/L solution of *chromotropic acid, sodium salt R*, 2 mL of *water R* and 8 mL of *sulfuric acid R*. A violet colour is produced.

E. 2 mL of the filtrate obtained in identification test D gives reaction (a) of chlorides (*2.3.1*).

TESTS
Appearance
It is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity
To 50 mL of *methanol R* previously neutralised to 0.1 mL of *bromothymol blue solution R1*, add 50 g of the substance to be examined. Not more than 0.15 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to blue.

Relative density (*2.2.5*)
1.320 to 1.332.

Refractive index (*2.2.6*)
1.423 to 1.425.

Ethanol, 2-methylbut-2-ene and volatile impurities
Gas chromatography (*2.2.28*).

Test solution The substance to be examined.

Reference solution (a) Dilute 100 µL of *carbon tetrachloride R* (impurity A), 500 µL of *chloroform R* (impurity B), 3.0 mL of *2-methylbut-2-ene R* and 5.0 mL of *methanol R* (impurity D) to 100.0 mL with the test solution.

Reference solution (b) Dilute 2.0 mL of *anhydrous ethanol R* and 1.0 mL of reference solution (a) to 100.0 mL with the test solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 1.8 µm).

Carrier gas *nitrogen for chromatography R.*

Flow rate 1.0 mL/min, constant flow.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 5	40
	5 - 12.5	40 → 55
	12.5 - 18	55 → 100
	18 - 20	100
Injection port		260
Detector		300

Detection Flame ionisation; make-up gas flow rate: 25 mL/min.

Injection 2 µL of the test solution and reference solution (b).

Relative retention With reference to methylene chloride (retention time = about 7 min): impurity D = about 0.6; ethanol = about 0.8; 2-methylbut-2-ene = about 0.9; impurity B = about 1.7; impurity A = about 1.8.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to ethanol and 2-methylbut-2-ene;
— *signal-to-noise ratio*: minimum 5 for the peak due to impurity A.

Calculation of percentage contents:
— for ethanol, 2-methylbut-2-ene and impurities A and B, use the respective concentration of these substances in reference solution (b); correct the areas of the peaks in the chromatogram obtained with reference solution (b) by subtracting the area of the corresponding peak in the chromatogram obtained with the test solution;
— for any other impurity, use the concentration of impurity D in reference solution (b); correct the area of the peak due to impurity D in the chromatogram obtained with reference solution (b) by subtracting the area of the corresponding peak in the chromatogram obtained with the test solution.

Limits:
— *ethanol*: maximum 2.0 per cent *V/V*;
— *2-methylbut-2-ene*: maximum 300 ppm *V/V*;
— *impurity A*: maximum 10 ppm *V/V*;
— *impurity B*: maximum 50 ppm *V/V*;
— *sum of impurities other than ethanol and 2-methylbut-2-ene*: maximum 0.1 per cent *V/V*;
— *reporting threshold*: 50 ppm *V/V*; the reporting threshold does not apply to impurity A.

Free chlorine
Place 5 mL in a ground-glass-stoppered tube. Add 5 mL of a 100 g/L solution of *potassium iodide R* and 0.2 g of *soluble starch R*. Shake for 30 s and allow to stand for 5 min. No blue colour develops.

Residue on evaporation
Maximum 20 ppm.

Evaporate 50.0 g to dryness on a water-bath and dry at 100-105 °C for 30 min. The residue weighs a maximum of 1 mg.

Water (*2.5.32*)
Maximum 0.02 per cent *m/m*, determined on 10.00 g.

STORAGE
In an airtight container, protected from light.

LABELLING
The label states the name and concentration of any stabilisers.

IMPURITIES
Specified impurities *A, B.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*

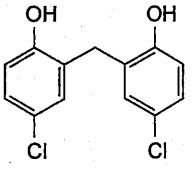
A. carbon tetrachloride,

B. trichloromethane (chloroform),

D. methanol.

Ph Eur

Dichlorophen



$C_{13}H_{10}Cl_2O_2$ 269.1 *97-23-4*

Action and use
Antihelminthic.

Preparation
Dichlorophen Tablets

DEFINITION
Dichlorophen is 4,4′-dichloro-2,2′-methylenediphenol.
It contains not less than 97.0% and not more than 101.0% of $C_{13}H_{10}Cl_2O_2$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or not more than slightly cream powder.
Practically insoluble in *water*; very soluble in *ether*; freely soluble in *ethanol (96%)*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 220 to 350 nm of a 0.002% w/v solution in 0.1M *sodium hydroxide* exhibits two maxima, at 245 nm and 304 nm.
The *absorbances* at the maxima are about 1.3 and about 0.54, respectively.

B. Dissolve 0.2 g in a mixture of 5 mL of *water* and 5 mL of 5M *sodium hydroxide*, cool in ice and add a solution prepared by mixing 1 mL of *sodium nitrite solution* with a cold solution containing 0.15 mL of *aniline* in a mixture of 4 mL of *water* and 1 mL of *hydrochloric acid*. A reddish brown precipitate is produced.

C. Fuse 0.5 g with 2 g of *anhydrous sodium carbonate*, cool, extract the residue with *water* and filter. The filtrate yields reaction A characteristic of *chlorides*, Appendix VI.

D. *Melting point*, about 175°, Appendix V A.

TESTS

Chloride
Dissolve 1.0 g in 2 mL of *ethanol (96%)*, dilute to 100 mL with *water*, allow to stand for 5 minutes and filter through a slow filter paper (Whatman No. 42 is suitable). 15 mL of the filtrate complies with the *limit test for chlorides*, Appendix VII (350 ppm).

Sulfate
Shake 0.8 g with 16 mL of *water* for 2 minutes, filter and dilute 5 mL of the filtrate to 15 mL with *water*. The solution complies with the *limit test for sulfates*, Appendix VII (600 ppm).

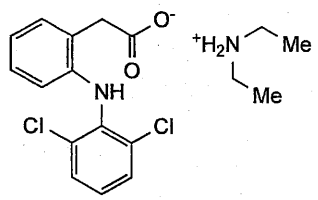
Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using three solutions in the mobile phase containing (1) 1.0% w/v of *dichlorophen impurity standard BPCRS*, (2) 1.0% w/v of the substance being examined and (3) 0.0010% w/v of *4-chlorophenol*.
The chromatographic procedure may be carried out using (a) a stainless steel column (20 cm × 5 mm) packed with *octadecylsilyl silica gel for chromatography* (10 µm) (Spherisorb ODS 1 is suitable), (b) as the mobile phase with a flow rate of 1.5 mL per minute a mixture of 25 volumes of *water* and 1 volume of *glacial acetic acid* and sufficient *methanol* to produce a chromatogram with solution (1) closely resembling the reference chromatogram supplied with the impurity standard (75 volumes of *methanol* is usually suitable) and (c) a detection wavelength of 280 nm. Record the chromatograms until all of the peaks named on the reference chromatogram have emerged.
In the chromatogram obtained with solution (2) the area of any peak corresponding to 4-chlorophenol is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.1%). The content of 4,4′-dichloro-2,2′-(2-hydroxy-4-chloro-*m*-xylene-α,α′-diyl)diphenol in the substance being examined does not exceed 8.0% w/w and the sum of the contents of any other impurities, excluding 4-chlorophenol, is not greater than 2.0% w/w calculated using the declared content of 4,4′-dichloro-2,2′-(2-hydroxy-4-chloro-*m*-xylene-α,α′-diyl)diphenol in *dichlorophen impurity standard BPCRS*.

Loss on drying
When dried to constant weight at 105°, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.5 g in 20 mL of *propan-2-ol* and carry out Method II for *non-aqueous titration*, Appendix VIII A, using 0.1M *tetrabutylammonium hydroxide VS* as titrant and determining the end point potentiometrically. Each mL of 0.1M *tetrabutylammonium hydroxide VS* is equivalent to 26.91 mg of $C_{13}H_{10}Cl_2O_2$.

Diclofenac Diethylamine



$C_{18}H_{22}Cl_2N_2O_2$ 369.29 *78213-16-8*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparation
Diclofenac Gel

DEFINITION
Diclofenac Diethylamine is diethylammonium 2-[(2,6-dichloroanilino)phenyl]acetate. It contains not less than 99.0% and not more than 101.0% of $C_{18}H_{22}Cl_2N_2O_2$, calculated with reference to the dried substance.

CHARACTERISTICS
A white to light beige, crystalline powder.

Sparingly soluble in *water* and in *acetone*; freely soluble in *ethanol (96%)* and in *methanol*; practically insoluble in 1M *sodium hydroxide*.

It melts at about 154°, with decomposition.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of diclofenac diethylamine *(RS 371)*.

B. Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in *methanol*.

(1) 5.0% w/v of the substance being examined.

(2) 5.0% w/v of *diclofenac diethylamine BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a silica gel precoated plate (Macherey Nagel SIL G-25 HR or silica gel $60F_{254}$ HPTLC plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 2 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry it in a stream of warm air for 10 minutes. Spray with *ninhydrin solution* and heat at 110° for 15 minutes.

MOBILE PHASE

1 volume of *hydrochloric acid*, 1 volume of *water*, 6 volumes of *glacial acetic acid* and 11 volumes of *ethyl acetate*.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (2) shows two clearly separated spots.

CONFIRMATION

The two principal spots in the chromatogram obtained with solution (1) are similar in position, colour and size to the corresponding spots in the chromatogram obtained with solution (2).

TESTS
Acidity or alkalinity
pH of a 1% w/v solution in *ethanol (10%)*, 6.4 to 8.4, Appendix V L.

Clarity and colour of solution
A 5% w/v solution in *methanol* is *clear*, Appendix IV A. The *absorbance* of the solution measured at 440 nm is not greater than 0.05, Appendix II B.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in the mobile phase.

(1) 0.10% w/v of the substance being examined.

(2) Dilute 2 volumes of solution (1) to 100 volumes and dilute 1 volume of this solution to 10 volumes.

(3) Dissolve 1 mg of *diclofenac impurity A BPCRS* in 1 mL of solution (1) and dilute to 200 mL.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with *end-capped octylsilyl silica gel for chromatography* (5 µm) (end-capped Zorbax C8 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for 1.5 times the retention time of diclofenac.

MOBILE PHASE

34 volumes of a mixture of equal volumes of a 0.1% w/v solution of *orthophosphoric acid* and a 0.16% w/v solution of *sodium dihydrogen orthophosphate* adjusted to pH 2.5 and 66 volumes of *methanol*.

When the chromatograms are recorded under the prescribed conditions, the retention times are about 25 minutes for diclofenac and about 12 minutes for diclofenac impurity A.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution* between the peaks corresponding to diclofenac and diclofenac impurity A is at least 6.5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.2%);

the sum of the areas of any *secondary peaks* is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

Disregard any peak with an area less than 0.25 times the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Loss on drying
When dried at a pressure not exceeding 1 kPa for 24 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A, Method II. Use 1 g.

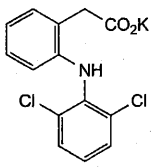
ASSAY
Dissolve 0.5 g in 30 mL of *anhydrous acetic acid* and carry out Method I for *non-aqueous titration*, Appendix VIII A, determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 36.93 mg of $C_{18}H_{22}Cl_2N_2O_2$.

STORAGE
Diclofenac Diethylamine should be kept in an airtight container and protected from light.

IMPURITIES
The impurities limited by the requirements of this monograph include impurities A, B, C, D and E listed under Diclofenac Sodium.

Diclofenac Potassium

(Ph. Eur. monograph 1508)



$C_{14}H_{10}Cl_2KNO_2$ 334.2 *15307-81-0*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
Potassium [2-[(2,6-dichlorophenyl)amino]phenyl]acetate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellowish, slightly hygroscopic, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent), slightly soluble in acetone.

IDENTIFICATION
First identification: A, D.
Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *diclofenac potassium CRS.*

B. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.
Reference solution (a) Dissolve 25 mg of *diclofenac potassium CRS* in *methanol R* and dilute to 5 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of *indometacin R* in reference solution (a) and dilute to 2 mL with the same solution.
Plate *TLC silica gel GF_{254} plate R.*
Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R* (10:10:80 *V/V/V*).
Application 5 µL.
Development Over 1/2 of the plate.
Drying In air.
Detection Examine in ultraviolet light at 254 nm.
System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 10 mL of *ethanol (96 per cent) R.* To 1 mL of this solution add 0.2 mL of a mixture, prepared immediately before use, of equal volumes of a 6 g/L solution of *potassium ferricyanide R* and a 9 g/L solution of *ferric chloride R.* Allow to stand protected from light for 5 min. Add 3 mL of a 10 g/L solution of *hydrochloric acid R.* Allow to stand protected from light for 15 min. A blue colour develops and a precipitate is formed.

D. Suspend 0.5 g in 10 mL of *water R.* Stir and add *water R* until the substance is dissolved. Add 2 mL of *hydrochloric acid R1,* stir for 1 h and filter with the aid of vacuum. Neutralise with *sodium hydroxide solution R.* The solution gives reaction (b) of potassium (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and its absorbance (*2.2.25*) at 440 nm is not greater than 0.05.
Dissolve 1.25 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve the contents of a vial of *diclofenac for system suitability CRS* (containing impurities A and F) in 1.0 mL of the mobile phase.
Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).
Mobile phase Mix 34 volumes of a solution containing 0.5 g/L of *phosphoric acid R* and 0.8 g/L of *sodium dihydrogen phosphate R,* previously adjusted to pH 2.5 with *phosphoric acid R,* and 66 volumes of *methanol R.*
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 20 µL.
Run time 1.6 times the retention time of diclofenac.
Identification of impurities Use the chromatogram supplied with *diclofenac for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and F.
Relative retention With reference to diclofenac (retention time = about 25 min): impurity A = about 0.4; impurity F = about 0.8.
System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to impurity F and diclofenac.
Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7; impurity F = 0.3;
— for each impurity, use the concentration of diclofenac in reference solution (a).

Limits:
— *impurities A, F*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

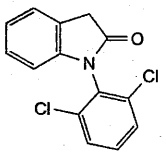
ASSAY
Dissolve 0.250 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 33.42 mg of $C_{14}H_{10}Cl_2KNO_2$.

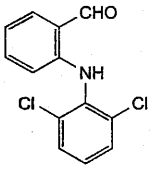
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *A, F.*

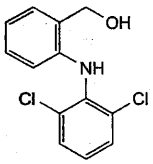
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E.*



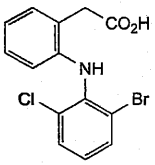
A. 1-(2,6-dichlorophenyl)-1,3-dihydro-2*H*-indol-2-one,



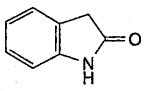
B. 2-[(2,6-dichlorophenyl)amino]benzaldehyde,



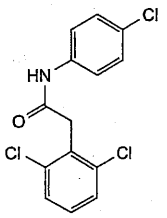
C. [2-[(2,6-dichlorophenyl)amino]phenyl]methanol,



D. [2-[(2-bromo-6-chlorophenyl)amino]phenyl]acetic acid,



E. 1,3-dihydro-2*H*-indol-2-one,

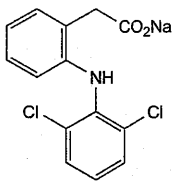


F. *N*-(4-chlorophenyl)-2-(2,6-dichlorophenyl)acetamide.

Ph Eur

Diclofenac Sodium

(Ph. Eur. monograph 1002)



$C_{14}H_{10}Cl_2NNaO_2$ 318.1 *15307-79-6*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Diclofenac Prolonged-release Capsules
Diclofenac Gastro-resistant Tablets
Diclofenac Prolonged-release Tablets

Ph Eur

DEFINITION
Sodium [2-[(2,6-dichlorophenyl)amino]phenyl]acetate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellowish, slightly hygroscopic, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent), slightly soluble in acetone.

mp
About 280 °C, with decomposition.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diclofenac sodium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of *diclofenac sodium CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *indometacin R* in reference solution (a) and dilute to 2 mL with reference solution (a).

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R* (10:10:80 *V/V/V*).

Application 5 µL.

Development Over 1/2 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 10 mL of *ethanol (96 per cent) R*. To 1 mL of this solution add 0.2 mL of a mixture, prepared immediately before use, of equal volumes of a 6 g/L solution of *potassium ferricyanide R* and a 9 g/L solution of *ferric chloride R*. Allow to stand protected from light for 5 min. Add 3 mL of a 10 g/L solution of *hydrochloric acid R*. Allow to stand, protected from light, for 15 min. A blue colour develops and a precipitate is formed.

D. Dissolve 60 mg in 0.5 mL of *methanol R* and add 0.5 mL of *water R*. The solution gives reaction (b) of sodium (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and its absorbance (*2.2.25*) at 440 nm is not greater than 0.05.

Dissolve 1.25 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *diclofenac for system suitability CRS* (containing impurities A and F) in 1.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 34 volumes of a solution containing 0.5 g/L of *phosphoric acid R* and 0.8 g/L of *sodium dihydrogen phosphate R*, previously adjusted to pH 2.5 with *phosphoric acid R*, and 66 volumes of *methanol R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 1.6 times the retention time of diclofenac.

Identification of impurities Use the chromatogram supplied with *diclofenac for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and F.

Relative retention With reference to diclofenac (retention time = about 25 min): impurity A = about 0.4; impurity F = about 0.8.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to impurity F and diclofenac.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7; impurity F = 0.3;
— for each impurity, use the concentration of diclofenac in reference solution (a).

Limits:
— *impurity A*: maximum 0.2 per cent;
— *impurity F*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Dissolve 0.250 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 31.81 mg of $C_{14}H_{10}Cl_2NNaO_2$.

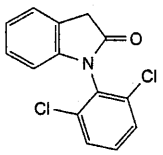
STORAGE

In an airtight container, protected from light.

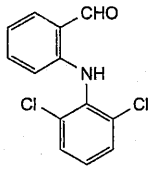
IMPURITIES

Specified impurities *A, F.*

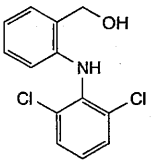
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E.*



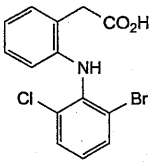
A. 1-(2,6-dichlorophenyl)-1,3-dihydro-2*H*-indol-2-one,



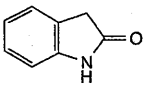
B. 2-[(2,6-dichlorophenyl)amino]benzaldehyde,



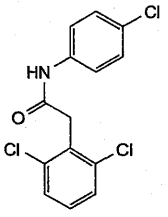
C. [2-[(2,6-dichlorophenyl)amino]phenyl]methanol,



D. [2-[(2-bromo-6-chlorophenyl)amino]phenyl]acetic acid,



E. 1,3-dihydro-2*H*-indol-2-one,

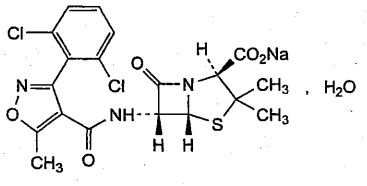


F. *N*-(4-chlorophenyl)-2-(2,6-dichlorophenyl)acetamide.

Ph Eur

Dicloxacillin Sodium

(Ph. Eur. monograph 0663)



$C_{19}H_{16}Cl_2N_3NaO_5S,H_2O$ 510.3 *13412-64-1*

Action and use
Penicillin antibacterial.

Ph Eur

DEFINITION

Sodium (2*S*,5*R*,6*R*)-6-[[[3-(2,6-dichlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *dicloxacillin sodium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *dicloxacillin sodium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *cloxacillin sodium CRS*, 25 mg of *dicloxacillin sodium CRS* and 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and about 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the solution is slightly greenish-yellow. Place the test-tube in a water-bath for 1 min; a yellow colour develops.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.04.

pH (*2.2.3*)
5.0 to 7.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 128 to + 143 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *dicloxacillin sodium CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of test solution (b) to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *flucloxacillin sodium CRS* and 5 mg of *dicloxacillin sodium CRS* in the mobile phase, then dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 25 volumes of *acetonitrile R* and 75 volumes of a 2.7 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 5.0 with *dilute sodium hydroxide solution R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (a) and reference solutions (b) and (c).

Run time 5 times the retention time of dicloxacillin.

Retention time Dicloxacillin = about 10 min.

System suitability Reference solution (c):
— *resolution*: minimum 2.5 between the peaks due to flucloxacillin (1st peak) and dicloxacillin (2nd peak).

Limits:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)
Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.8 per cent *m/m*.

Water (*2.5.12*)
3.0 per cent to 4.5 per cent, determined on 0.300 g.

Pyrogens (*2.6.8*)
If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of pyrogens, it complies with the test for pyrogens. Inject per kilogram of the rabbit's mass 1 mL of a solution in *water for injections R* containing 20 mg of the substance to be examined per millilitre.

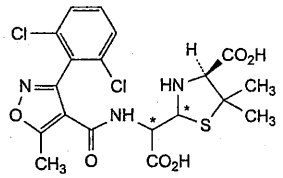
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

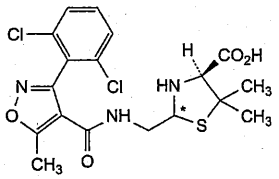
System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

STORAGE
In an airtight container, at a temperature not exceeding 25 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

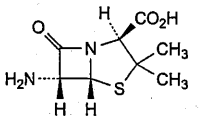
IMPURITIES



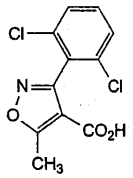
A. (4*S*)-2-[carboxy[[[3-(2,6-dichlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acids of dicloxacillin),



B. (2*RS*,4*S*)-2-[[[[3-(2,6-dichlorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penilloic acids of dicloxacillin),



C. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),

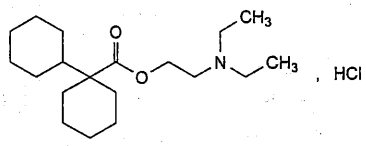


D. 3-(2,6-dichlorophenyl)-5-methyl-1,2-oxazole-4-carboxylic acid.

Ph Eur

Dicycloverine Hydrochloride

(Ph. Eur. monograph 1197)



$C_{19}H_{36}ClNO_2$ 346.0

Action and use
Anticholinergic.

Preparations
Dicycloverine Oral Solution
Dicycloverine Tablets

Ph Eur

DEFINITION

2-(Diethylamino)ethyl 1,1′-(bicyclohexane)-1-carboxylate hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dicycloverine hydrochloride CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *methanol R* and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of *dicycloverine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R*

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, propanol R* (5:10:10:75 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Spray with *dilute potassium iodobismuthate solution R.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 3 mL of a 1.0 g/L solution of *sodium laurilsulfate R* add 5 mL of *methylene chloride R* and 0.05 mL of a 2.5 g/L solution of *methylene blue R*, mix gently and allow to separate; the lower layer is blue. Add 2 mL of a 20 g/L solution of the substance to be examined, mix gently and allow to separate; the upper layer is blue and the lower layer is colourless.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

pH (*2.2.3*)

5.0 to 5.5.

Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *dicycloverine for system suitability CRS* (containing impurity B) in 1.0 mL of the mobile phase.

Reference solution (c) Dissolve 7.5 mg of *dicycloverine impurity A CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 25 °C.

Mobile phase Mix 15 volumes of a 4.62 g/L solution of *ammonium acetate R* previously adjusted to pH 8.0 with *2 M sodium hydroxide R* and 85 volumes of *methanol R2.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

Run time 1.5 times the retention time of dicycloverine.

Identification of impurities Use the chromatogram supplied with *dicycloverine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to dicycloverine (retention time = about 22 min): impurity A = about 0.5; impurity B = about 0.8.

System suitability Reference solution (b):

— *resolution*: minimum 2.0 between the peaks due to impurity B and dicycloverine.

Calculation of percentage contents:

— *correction factor*: multiply the peak area of impurity B by 0.4;

— for impurity A, use the concentration of impurity A in reference solution (c);

— for impurities other than A, use the concentration of dicycloverine in reference solution (a).

Limits:

— *impurity B*: maximum 0.5 per cent;

— *impurity A*: maximum 0.15 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 0.7 per cent;

— *reporting threshold:* 0.05 per cent.

Water (*2.5.32*): maximum 1.0 per cent, determined on 1.00 g by direct introduction of the sample.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

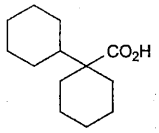
ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R.* Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

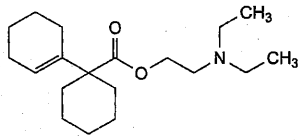
1 mL of *0.1 M sodium hydroxide* is equivalent to 34.60 mg of $C_{19}H_{36}ClNO_2$.

IMPURITIES

Specified impurities *A, B.*



A. 1,1′-bi(cyclohexane)-1-carboxylic acid,

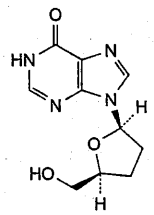


B. 2-(diethylamino)ethyl 1,1′-bi(cyclohexan)-1′-ene-1-carboxylate.

Ph Eur

Didanosine

(Ph. Eur. monograph 2200)



$C_{10}H_{12}N_4O_3$ 236.2 *69655-05-6*

Action and use
Nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Ph Eur

DEFINITION

9-(2,3-Dideoxy-β-D-*glycero*-pentofuranosyl)-1,9-dihydro-6*H*-purin-6-one (2′,3′-dideoxyinosine).

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in dimethyl sulfoxide, slightly soluble in methanol and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *didanosine CRS*.

B. Specific optical rotation (*2.2.7*): −28.2 to −24.2 (anhydrous substance).

Dissolve 0.100 g in *water R* and dilute to 10.0 mL with the same solvent.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture Mix 8 volumes of mobile phase B and 92 volumes of mobile phase A.

Test solution Dissolve 25.0 mg of the substance to be examined in 50.0 mL of the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of *didanosine impurity A CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *didanosine for system suitability CRS* (containing impurities A to F) in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (d) Dissolve 5 mg of *didanosine impurity G CRS* in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL to 20 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 8 volumes of *methanol R* and 92 volumes of a 3.86 g/L solution of *ammonium acetate R* adjusted to pH 8.0 with *concentrated ammonia R*;
— *mobile phase B*: mix 30 volumes of *methanol R* and 70 volumes of a 3.86 g/L solution of *ammonium acetate R* adjusted to pH 8.0 with *concentrated ammonia R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 18	100	0
18 - 25	100 → 0	0 → 100
25 - 45	0	100
45 - 50	0 → 100	100 → 0
50 - 60	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *didanosine for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A to F and use the chromatogram obtained with reference solution (d) to identify the peak due to impurity G.

Relative retention With reference to didanosine (retention time = about 13-15 min): impurity A = about 0.3; impurity B = about 0.4; impurity C = about 0.44; impurity D = about 0.48; impurity E = about 0.5; impurity F = about 0.8; impurity G = about 1.6.

System suitability Reference solution (c):
— *resolution*: minimum 2.5 between the peaks due to impurity C and impurity D.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurities B, C, D, E, F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

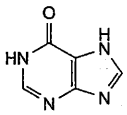
Dissolve 0.200 g in 50 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 23.62 mg of $C_{10}H_{12}N_4O_3$.

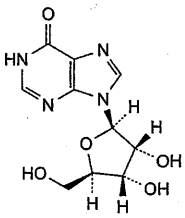
IMPURITIES

Specified impurities *A, B, C, D, E, F, G.*

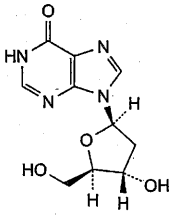
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H, I.*



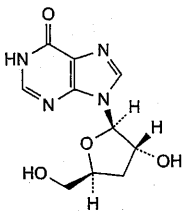
A. 1,7-dihydro-6*H*-purin-6-one (hypoxanthine),



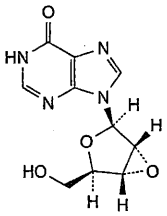
B. 9-β-D-ribofuranosyl-1,9-dihydro-6*H*-purin-6-one (inosine),



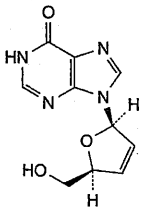
C. 9-(2-deoxy-β-D-*erythro*-pentofuranosyl)-1,9-dihydro-6*H*-purin-6-one (2′-deoxyinosine),



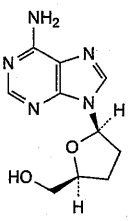
D. 9-(3-deoxy-β-D-*erythro*-pentofuranosyl)-1,9-dihydro-6*H*-purin-6-one (3′-deoxyinosine),



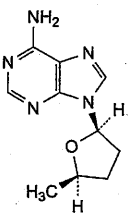
E. 9-(2,3-anhydro-β-D-ribofuranosyl)-1,9-dihydro-6*H*-purin-6-one (2′,3′-anhydroinosine),



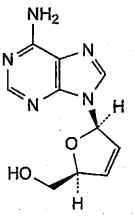
F. 9-(2,3-dideoxy-β-D-*glycero*-pent-2-enofuranosyl)-1,9-dihydro-6*H*-purin-6-one (2′,3′-dideoxy-2′,3′-didehydroinosine),



G. 9-(2,3-dideoxy-β-D-*glycero*-pentofuranosyl)-9*H*-purin-6-amine (2′,3′-dideoxyadenosine),



H. 9-(2,3,5-trideoxy-β-D-*glycero*-pentofuranosyl)-9*H*-purin-6-amine (2′,3′,5′-trideoxyadenosine),

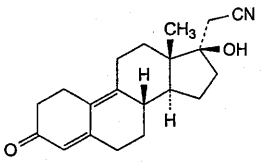


I. 9-(2,3-dideoxy-β-D-*glycero*-pent-2-enofuranosyl)-9*H*-purin-6-amine (2′,3′-dideoxy-2′,3′-didehydroadenosine).

Ph Eur

Dienogest

(Ph. Eur. monograph 2732)



$C_{20}H_{25}NO_2$ 311.4 *65928-58-7*

Action and use
Progestogen.

Ph Eur

DEFINITION

(17-Hydroxy-3-oxoestra-4,9-dien-17α-yl)acetonitrile.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White, almost white or slightly yellow, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in methanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dienogest CRS.*

TESTS

Specific optical rotation (*2.2.7*)
−352 to −344 (dried substance).

Dissolve 0.125 g in *methanol R2* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture *acetonitrile R1, water for chromatography R* (40:60 *V/V*).

Test solution (a) Dissolve 20 mg of the substance to be examined in 8 mL of *acetonitrile R1* and dilute to 20.0 mL with *water R.*

Test solution (b) Dissolve 30.0 mg of the substance to be examined in 40 mL of *acetonitrile R1* and dilute to 100.0 mL with *water R.* Dilute 5.0 mL of the solution to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2.5 mg of *dienogest for system suitability CRS* (containing impurity F) in 2 mL of *acetonitrile R1* and dilute to 5.0 mL with *water R.*

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 30.0 mg of *dienogest CRS* in 40 mL of *acetonitrile R1* and dilute to 100.0 mL with *water R.* Dilute 5.0 mL of the solution to 25.0 mL with the solvent mixture.

Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: end-capped extra-dense bonded octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A: acetonitrile R1, water for chromatography R* (10:90 *V/V*);
— *mobile phase B: water for chromatography R, acetonitrile R1* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	90	10
3 - 53	90 → 10	10 → 90

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 15 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *dienogest for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity F.

Relative retention With reference to dienogest (retention time = about 19 min): impurity F = about 1.1.

System suitability Reference solution (a):
— *resolution*: minimum 4.0 between the peaks due to dienogest and impurity F.

Calculation of percentage contents:
— for each impurity, use the concentration of dienogest in reference solution (b).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	75	25
3 - 14	75 → 25	25 → 75

Detection Spectrophotometer at 215 nm.

Injection Test solution (b) and reference solution (c).

Retention time Dienogest = about 9 min.

Calculate the percentage content of $C_{20}H_{25}NO_2$ taking into account the assigned content of *dienogest CRS.*

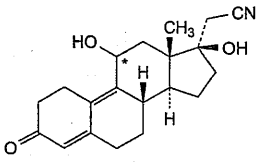
STORAGE

Protected from light.

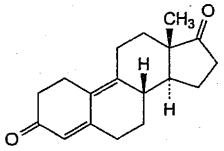
IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for

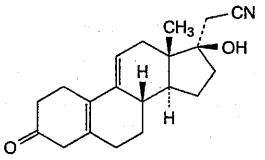
demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I, J, K, L, M.*



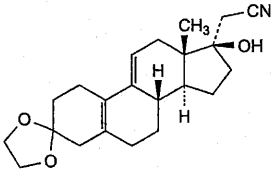
A. (11,17-dihydroxy-3-oxoestra-4,9-dien-17α-yl)acetonitrile,



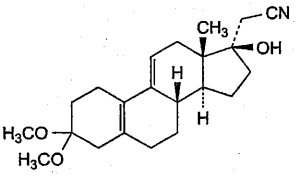
B. estra-4,9-diene-3,17-dione,



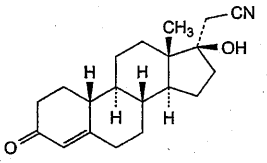
C. [17-hydroxy-3-oxoestra-5(10),9(11)-dien-17α-yl]acetonitrile,



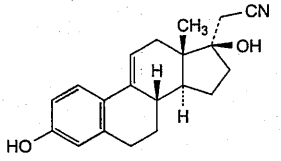
D. [17′-hydroxyspiro[1,3-dioxolan-2,3′-estra-5′(10′),9′(11′)-dien]-17′α-yl]acetonitrile,



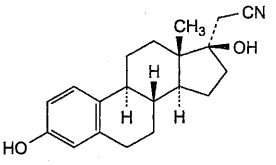
E. [17-hydroxy-3,3-dimethoxyestra-5(10),9(11)-dien-17α-yl]acetonitrile,



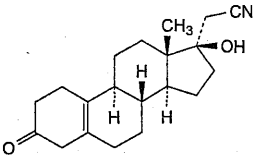
F. (17-hydroxy-3-oxoestr-4-en-17α-yl)acetonitrile,



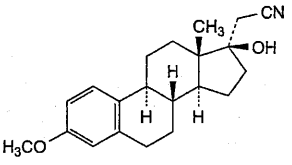
G. [3,17-dihydroxyestra-1,3,5(10),9(11)-tetraen-17α-yl]acetonitrile,



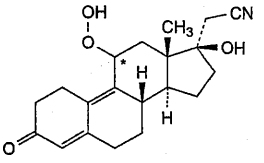
H. [3,17-dihydroxyestra-1,3,5(10)-trien-17α-yl]acetonitrile,



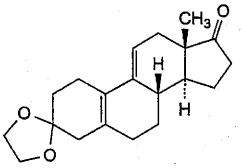
I. [17-hydroxy-3-oxoestr-5(10)-en-17α-yl]acetonitrile,



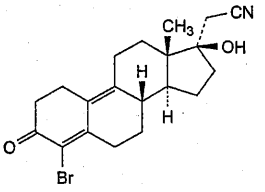
J. [17-hydroxy-3-methoxyestra-1,3,5(10)-trien-17α-yl]acetonitrile,



K. (11-hydroperoxy-17-hydroxy-3-oxoestra-4,9-dien-17α-yl)acetonitrile,



L. spiro[1,3-dioxolan-2,3′-estra-5′(10′),9′(11′)-dien]-17′-one,

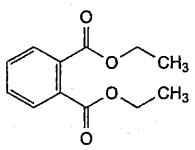


M. (4-bromo-17-hydroxy-3-oxoestra-4,9-dien-17α-yl)acetonitrile.

Ph Eur

Diethyl Phthalate

(Ph. Eur. monograph 0897)



$C_{12}H_{14}O_4$ 222.2 *84-66-2*

Action and use
Excipient.

Ph Eur

DEFINITION
Diethyl benzene-1,2-dicarboxylate.

Content
99.0 per cent *m/m* to 101.0 per cent *m/m*.

CHARACTERS
Appearance
Clear, colourless or very slightly yellow, oily liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent).

IDENTIFICATION
First identification: B, C.

Second identification: A, D, E.

A. Relative density (*2.2.5*): 1.117 to 1.121.

B. Refractive index (*2.2.6*): 1.500 to 1.505.

C. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Thin films.

Comparison *diethyl phthalate CRS.*

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *ether R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of *diethyl phthalate CRS* in *ether R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *heptane R, ether R* (30:70 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

E. To about 0.1 mL add 0.25 mL of *sulfuric acid R* and 50 mg of *resorcinol R*. Heat on a water-bath for 5 min. Allow to cool. Add 10 mL of *water R* and 1 mL of *strong sodium hydroxide solution R*. The solution becomes yellow or brownish-yellow and shows green fluorescence.

TESTS
Appearance
The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Acidity
Dissolve 20.0 g in 50 mL of *ethanol (96 per cent) R* previously neutralised to *phenolphthalein solution R1*. Add 0.2 mL of *phenolphthalein solution R1*. Not more than 0.1 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 60 mg of *naphthalene R* in *methylene chloride R* and dilute to 20 mL with the same solvent.

Test solution (a) Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 20.0 mL with the same solvent.

Test solution (b) Dissolve 1.0 g of the substance to be examined in *methylene chloride R*, add 2.0 mL of the internal standard solution and dilute to 20.0 mL with *methylene chloride R*.

Reference solution To 1.0 mL of test solution (a) add 10.0 mL of the internal standard solution and dilute to 100.0 mL with *methylene chloride R*.

Column:
— *material*: glass;
— *size*: *l* = 2 m, Ø = 2 mm;
— *stationary phase*: *silanised diatomaceous earth for gas chromatography R* (150-180 µm) impregnated with 3 per cent *m/m* of *polymethylphenylsiloxane R*.

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Temperature:
— *column*: 150 °C;
— *injection port and detector*: 225 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 3 times the retention time of diethyl phthalate.

Elution order Naphthalene, diethyl phtalate.

System suitability:
— *resolution*: minimum 10 between the peaks due to naphthalene and diethyl phthalate in the chromatogram obtained with the reference solution;
— in the chromatogram obtained with test solution (a), there is no peak with the same retention time as the internal standard.

Limit:
— *total*: calculate the ratio (*R*) of the area of the peak due to diethyl phthalate to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with test solution (b), calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than *R* (1.0 per cent).

Water (*2.5.12*)
Maximum 0.2 per cent, determined on 10.0 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Introduce 0.750 g into a 250 mL borosilicate glass flask. Add 25.0 mL of 0.5 M alcoholic potassium hydroxide and a few glass beads. Boil in a water-bath under a reflux condenser for 1 h. Add 1 mL of *phenolphthalein solution R1*

and titrate immediately with *0.5 M hydrochloric acid*. Carry out a blank titration. Calculate the volume of 0.5 M alcoholic potassium hydroxide used in the saponification.

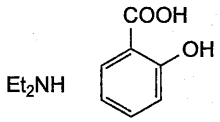
1 mL of 0.5 M alcoholic potassium hydroxide is equivalent to 55.56 mg of $C_{12}H_{14}O_4$.

STORAGE

In an airtight container.

Ph Eur

Diethylamine Salicylate



$C_{11}H_{17}NO_3$ 211.3 *4419-92-5*

Action and use

Counter-irritant.

Preparation

Diethylamine Salicylate Cream

DEFINITION

Diethylamine Salicylate contains not less than 99.0% and not more than 101.0% of $C_{11}H_{17}NO_3$.

CHARACTERISTICS

White or almost white crystals.

Very soluble in *water*; freely soluble in *ethanol (96%)*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of diethylamine salicylate *(RS 099)*.

B. To 0.2 g add 5 mL of 1M *sodium hydroxide*, heat to boiling point, cool and acidify with *2M hydrochloric acid*; a white precipitate is produced. The *melting point* of the precipitate, after recrystallisation from *water* and drying at 105°, is about 160°, Appendix V A.

TESTS

Acidity

Dissolve 2 g in 25 mL of *water* and titrate with *0.1M sodium hydroxide VS* using *phenol red solution* as indicator. Not more than 0.2 mL of *0.1M sodium hydroxide VS* is required to change the colour of the solution.

Clarity and colour of solution

A 50% w/v solution is *clear*, Appendix IV A, and not more intensely coloured than *reference solution* BY_5, Appendix IV B, Method II.

Melting point

100° to 102°, Appendix V A.

Sulfate

0.6 g complies with the *limit test for sulfates*, Appendix VII (250 ppm).

Loss on drying

When dried at 60° for 3 hours, loses not more than 0.1% of its weight. Use 1 g.

ASSAY

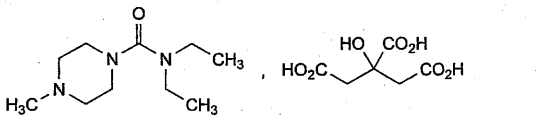
Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.4 g and *1-naphtholbenzein solution* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 21.13 mg of $C_{11}H_{17}NO_3$.

STORAGE

Diethylamine Salicylate should be protected from light. It should not be allowed to come into contact with iron or iron salts.

Diethylcarbamazine Citrate

(Ph. Eur. monograph 0271)



$C_{16}H_{29}N_3O_8$ 391.4 *1642-54-2*

Action and use

Antihelminthic.

Preparation

Diethylcarbamazine Tablets

Ph Eur

DEFINITION

N,N-Diethyl-4-methylpiperazine-1-carboxamide dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, slightly hygroscopic.

Solubility

Very soluble in water, soluble in ethanol (96 per cent), practically insoluble in acetone.

mp

About 138 °C, with decomposition.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diethylcarbamazine citrate CRS.*

B. Examine the chromatograms obtained in the test for impurities A and B.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in 5 mL of *water R*. The solution gives the reaction of citrates (*2.3.1*).

TESTS

Solution S

Shake 2.5 g with *water R* until dissolved and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Impurities A and B

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.5 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 0.1 g of *diethylcarbamazine citrate CRS* in *methanol R* and dilute to 2.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of methylpiperazine R (impurity A) in *methanol R* and dilute to 100 mL with the same solvent.

Reference solution (c) Dissolve 10 mg of *dimethylpiperazine R* (impurity B) in *methanol R* and dilute to 100 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, methyl ethyl ketone R, methanol R* (5:30:65 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying At 100-105 °C.

Detection Expose to iodine vapour for 30 min.

Retardation factors Impurity A = about 0.2; impurity B = about 0.5.

Limits:

— *impurity A*: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurity B*: any spot due to impurity B is not more intense than the corresponding spot in the chromatogram obtained with reference solution (c) (0.2 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Solution A Dissolve 31.2 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 1000 mL with the same solvent.

Test solution (a) Suspend 0.30 g of the substance to be examined in solution A and dilute to 100 mL with solution A. Filter or centrifuge and use the clear filtrate or supernatant.

Test solution (b) Dissolve 10.0 mg of the substance to be examined in solution A and dilute to 100.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 10 mg of *citric acid monohydrate R* in solution A and dilute to 10 mL with solution A.

Reference solution (c) To 3 mL of test solution (a) add 0.5 mL of *strong hydrogen peroxide solution R* and maintain at 80 °C for 3 h. Dilute to 100 mL with solution A.

Reference solution (d) Dissolve 5.0 mg of *diethylcarbamazine citrate CRS* in solution A and dilute to 50.0 mL with solution A.

Column:

— *size*: *l* = 0.15 m, Ø = 3.9 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 100 volumes of *methanol R2* and 900 volumes of a 10 g/L solution of *potassium dihydrogen phosphate R*.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Run time Twice the retention time of diethylcarbamazine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to the citrate.

Relative retention With reference to diethylcarbamazine (retention time = about 7 min): citrate = about 0.2; degradation product = about 1.6.

System suitability Reference solution (c):

— *resolution*: minimum 5 between the peaks due to diethylcarbamazine and the degradation product.

Limits:

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to the citrate.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 20 µL of test solution (b) and reference solution (d).

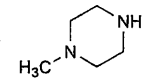
Calculate the percentage content of $C_{16}H_{29}N_3O_8$ from the declared content of *diethylcarbamazine citrate CRS*.

STORAGE

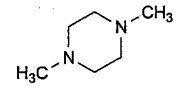
In an airtight container.

IMPURITIES

Specified impurities *A, B.*



A. 1-methylpiperazine,

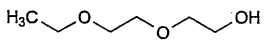


B. 1,4-dimethylpiperazine.

Ph Eur

Diethylene Glycol Monoethyl Ether

(Ph. Eur. monograph 1198)



$C_6H_{14}O_3$ 134.2 *111-90-0*

Action and use
Excipient.

Ph Eur

DEFINITION

2-(2-Ethoxyethoxy)ethanol, produced by condensation of ethylene oxide and alcohol, followed by distillation.

CHARACTERS

Appearance
Clear, colourless, hygroscopic liquid.

Solubility
Miscible with water, with acetone and with alcohol, miscible in certain proportions with vegetable oils, not miscible with mineral oils.

Relative density
About 0.991.

IDENTIFICATION

A. Refractive index (*2.2.6*): 1.426 to 1.428.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of diethylene glycol monoethyl ether.*

TESTS

Acid value (*2.5.1*)
Maximum 0.1.

Mix 30.0 mL with 30 mL of *alcohol R* previously neutralised with *0.1 M potassium hydroxide* using *phenolphthalein solution R* as indicator. Titrate with *0.01 M alcoholic potassium hydroxide.*

Peroxide value (*2.5.5*)
Maximum 8.0, determined on 2.00 g.

Related substances
Gas chromatography (*2.2.28*).

Internal standard solution Dilute 1.00 g of *decane R* to 100.0 mL with *methanol R.*

Test solution To 5.00 g of the substance to be examined, add 0.1 mL of the internal standard solution and dilute to 10.0 mL with *methanol R.*

Reference solution (a) Dilute 25.0 mg of *ethylene glycol monomethyl ether R*, 80.0 mg of *ethylene glycol monoethyl ether R*, 0.310 g of *ethylene glycol R* and 0.125 g of *diethylene glycol R* to 100.0 mL with *methanol R.* To 1.0 mL of this solution add 0.1 mL of the internal standard solution and dilute to 10.0 mL with *methanol R.*

Reference solution (b) Dilute 25.0 mg of *ethylene glycol monoethyl ether R* and 25.0 mg of *ethylene glycol R* to 100.0 mL with *methanol R.* Dilute 1.0 mL of this solution to 5.0 mL with *methanol R.*

Reference solution (c) Dilute 1.00 g of the substance to be examined to 100.0 mL with *methanol R.* To 1.0 mL of this solution add 0.1 mL of the internal standard solution and dilute to 10.0 mL with *methanol R.*

Column:
— *material:* fused silica,
— *size:* *l* = 30 m, Ø = 0.32 mm,
— *stationary phase: poly(cyanopropyl) (7) (phenyl) (7) (methyl) (86) siloxane R* (film thickness 1 µm).

Carrier gas *nitrogen for chromatography R* or *helium for chromatography R.*

Flow rate 2.0 mL/min.

Split ratio 1:80.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 1	120
	1 - 10	120 → 225
	10 - 12	225
Injection port		275
Detector		250

Detection Flame ionisation.

Injection 0.5 µL.

Relative retentions With reference to diethylene glycol monoethyl ether (retention time = about 4 min): ethylene glycol monomethyl ether = about 0.4; ethylene glycol monoethyl ether = about 0.5; ethylene glycol = about 0.55; diethylene glycol = about 1.1.

System suitability:
— *resolution:* minimum 3.0 between the peaks due to ethylene glycol monoethyl ether and to ethylene glycol in the chromatogram obtained with reference solution (b),
— *signal-to-noise ratio:* minimum 3.0 for the peak due to ethylene glycol monomethyl ether in the chromatogram obtained with reference solution (a),

Limits (take into account the impurity/internal standard peak area ratio):
— *ethylene glycol monomethyl ether:* not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (50 ppm),
— *ethylene glycol monoethyl ether:* not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (160 ppm),
— *ethylene glycol:* not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (620 ppm),
— *diethylene glycol:* not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (250 ppm),
— *total:* not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent).

Ethylene oxide
Head-space gas chromatography (*2.2.28*).

Test solution To 1.00 g of the substance to be examined in a vial, add 50 µL of *water R.*

Reference solution To 1.00 g of the substance to be examined in a vial, add 50 µL of *ethylene oxide solution R4* and close tightly.

Column:
— *material:* fused silica,
— *size:* *l* = 30 m, Ø = 0.32 mm,
— *stationary phase: poly(cyanopropyl) (7) (phenyl) (7) (methyl) (86) siloxane R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1.1 mL/min.

Static head-space conditions which may be used:
— *equilibration temperature:* 80 °C,

— *equilibration time*: 45 min,
— *transfer line temperature*: 110 °C,
— *pressurisation time*: 2 min,
— *injection time*: 12 s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 5	40
	5 - 18	40 → 200
Injection port		150
Detector		250

Detection Flame ionisation.

Injection 1.0 mL.

The peak due to ethylene oxide is identified by injecting solutions of ethylene oxide of increasing concentration.

Determine the content of ethylene oxide (ppm) in the substance to be examined using the following expression:

$$\frac{S_T \times C}{(S_S \times M_T) - (S_T \times M_S)}$$

S_T = area of the peak due to ethylene oxide in the chromatogram obtained with the test solution,
S_S = area of the peak due to ethylene oxide in the chromatogram obtained with the reference solution,
M_T = mass of the substance to be examined in the test solution, in grams,
M_S = mass of the substance to be examined in the reference solution, in grams,
C = mass of added ethylene oxide in the reference solution, in micrograms.

Limit:
— *ethylene oxide*: maximum 1 ppm.

Water (*2.5.12*)
Maximum 0.1 per cent, determined on 10.0 g.

STORAGE
Under an inert gas, in an airtight container.

LABELLING
The label states that the substance is stored under an inert gas.

Ph Eur

Diethylene Glycol Palmitostearate

(Ph. Eur. monograph 1415)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of diethylene glycol mono- and diesters of stearic (octadecanoic) and palmitic (hexadecanoic) acids.

It is produced by esterification of diethylene glycol and stearic acid 50 (see *Stearic acid (1474)*) of vegetable or animal origin.

Content
— *monoesters*: 45.0 per cent to 60.0 per cent;
— *diesters*: 35.0 per cent to 55.0 per cent.

CHARACTERS

Appearance
White or almost white, waxy solid.

Solubility
Practically insoluble in water, soluble in acetone and in hot ethanol (96 per cent).

IDENTIFICATION
A. Melting point (see Tests).
B. Composition of fatty acids (see Tests).
C. It complies with the limit of the assay (monoesters content).

TESTS

Melting point (*2.2.15*)
43 °C to 50 °C.

Acid value (*2.5.1*)
Maximum 4.0.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Saponification value (*2.5.6*)
155 to 180, determined on 2.0 g.

Composition of fatty acids (*2.4.22, Method A*)
Use the mixture of calibrating substances in Table 2.4.22.-1.

Composition of the fatty acid fraction of the substance:
— *stearic acid*: 40.0 per cent to 60.0 per cent;
— *sum of contents of palmitic acid and stearic acid*: minimum 90.0 per cent.

Free diethylene glycol
Maximum 2.5 per cent, determined as described in the assay.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY
Size-exclusion chromatography (*2.2.30*).

Test solution Into a 15 mL flask, weigh 0.200 g (*m*). Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Heat gently, if necessary. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into four 15 mL flasks, weigh, 2.5 mg, 5.0 mg, 10.0 mg and 20.0 mg respectively of *diethylene glycol R*. Add 5.0 mL of *tetrahydrofuran R*. Weigh the flasks again and calculate the concentration of diethylene glycol in milligrams per gram for each reference solution.

Column:
— *size*: l = 0.6 m, Ø = 7 mm,
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R.*

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to diethylene glycol: diesters = about 0.78; monoesters = about 0.84.

Calculations:
— *free diethylene glycol*: from the calibration curve obtained with the reference solutions, determine the concentration (C) of diethylene glycol in milligrams per gram in the test solution and calculate the percentage content of free diethylene glycol in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *monoesters*: calculate the percentage content of monoesters using the following expression:

$$\frac{A}{A+B} \times (100 - D)$$

A = area of the peak due to the monoesters,
B = area of the peak due to the diesters,
D = percentage content of free diethylene glycol + percentage content of free fatty acids.

Calculate the percentage content of free fatty acids using the following expression:

$$\frac{I_A \times 270}{561.1}$$

I_A = acid value.

— *diesters*: calculate the percentage content of diesters using the following expression:

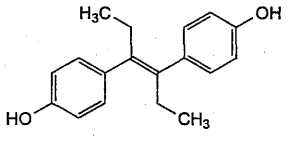
$$\frac{B}{A+B} \times (100 - D)$$

STORAGE
Protected from light.

Ph Eur

Diethylstilbestrol

(Ph. Eur. monograph 0484)



$C_{18}H_{20}O_2$ 268.4 *56-53-1*

Action and use
Estrogen.

Preparation
Diethylstilbestrol Tablets

Ph Eur

DEFINITION
Diethylstilbestrol contains not less than 97.0 per cent and not more than the equivalent of 101.0 per cent of (*E*)-4,4′-(1,2-diethylethene-1,2-diyl)diphenol, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, practically insoluble in water, freely soluble in alcohol. It dissolves in solutions of the alkali hydroxides.

It melts at about 172 °C.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Examined between 230 nm and 450 nm (*2.2.25*), the irradiated solution of the substance to be examined prepared as prescribed in the assay shows two absorption maxima, at 292 nm and 418 nm.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *diethylstilbestrol CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for mono-and dimethyl ethers. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 0.5 mg in 0.2 mL of *glacial acetic acid R*, add 1 mL of *phosphoric acid R* and heat on a water-bath for 3 min. A deep-yellow colour develops.

TESTS

4,4′-Dihydroxystilbene and related ethers
Dissolve 0.100 g in *ethanol R* and dilute to 10.0 mL with the same solvent. The absorbance (*2.2.25*) of the solution measured at 325 nm is not greater than 0.50.

Mono- and dimethyl ethers
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.2 g of the substance to be examined in 2 mL of *alcohol R*.

Test solution (b) Dilute 1 mL of test solution (a) to 20 mL with *alcohol R*.

Reference solution (a) Dissolve 10 mg of *diethylstilbestrol CRS* in 2 mL of *alcohol R*.

Reference solution (b) Dissolve 5 mg of *diethylstilbestrol monomethyl ether CRS* in *alcohol R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dissolve 5 mg of *diethylstilbestrol dimethyl ether CRS* in *alcohol R* and dilute to 10 mL with the same solvent.

Reference solution (d) Dissolve 10 mg of *dienestrol CRS* in 2 mL of *alcohol R*. To 1 mL of this solution add 1 mL of reference solution (a).

Apply to the plate 1 µL of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of *diethylamine R* and 90 volumes of *toluene R*. Allow the plate to dry in air, spray with *alcoholic solution of sulfuric acid R* and heat at 120 °C for 10 min. In the chromatogram obtained with test solution (a), any spots corresponding to diethylstilbestrol monomethyl ether and diethylstilbestrol dimethyl ether are not more intense than the spots in the chromatograms obtained with reference solutions (b) and (c) respectively (0.5 per cent). Diethylstilbestrol gives one or sometimes two spots. The test is not valid unless the chromatogram obtained with reference solution (d) shows at least two clearly separated spots having approximately the same intensity.

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 20.0 mg in *ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *ethanol R*. To 25.0 mL of the resulting solution add 25.0 mL of a solution of 1 g of *dipotassium hydrogen phosphate R* in 55 mL of *water R*. Prepare in the same manner a reference solution using 20.0 mg of *diethylstilbestrol CRS*. Transfer an equal volume of each solution to separate 1 cm quartz cells and close the cells; place the two cells about 5 cm from a low-pressure, short-wave 2 W to 20 W mercury lamp and irradiate for about

5 min. Measure the absorbance (*2.2.25*) of the irradiated solutions at the maximum at 418 nm, using *water R* as the compensation liquid. Continue the irradiation for successive periods of 3 min to 15 min, depending on the power of the lamp, and repeat the measurement of the absorbances at 418 nm until the maximum absorbance (about 0.7) is obtained. If necessary, adjust the geometry of the irradiation apparatus to obtain a maximum, reproducible absorbance at 418 nm.

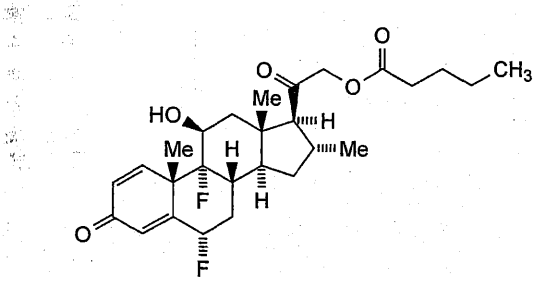
Calculate the content of $C_{18}H_{20}O_2$ from the measured absorbances and the concentrations of the solutions.

STORAGE

Store protected from light.

Ph Eur

Diflucortolone Valerate



$C_{27}H_{36}F_2O_5$ 478.6 *59198-70-8*

Action and use

Glucocorticoid.

Preparations

Diflucortolone Cream

Diflucortolone Oily Cream

Diflucortolone Ointment

DEFINITION

Diflucortolone Valerate is 6α,9α-difluoro-3,20-dioxo-11β-hydroxy-16α-methylpregna-1,4-dien-21-yl valerate.

It contains not less than 97.0% and not more than 102.0% of $C_{27}H_{36}F_2O_5$, calculated with reference to the dried substance.

CHARACTERISTICS

A white to creamy white, crystalline powder.

Practically insoluble in *water*; freely soluble in *dichloromethane* and in *1,4-dioxan*; slightly soluble in *methanol*; sparingly soluble in *ether*.

IDENTIFICATION

A. The *light absorption*, Appendix II B, in the range 210 to 350 nm of a 0.002% w/v solution in *methanol* exhibits a maximum only at 238 nm. The *absorbance* at the maximum at 238 nm is about 0.69.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of diflucortolone valerate *(RS 100)*.

TESTS

Specific optical rotation

In a 1% w/v solution in *1,4-dioxan*, +98 to +103, Appendix V F, calculated with reference to the dried substance.

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, injecting 20 µL of each of the following solutions in a mixture of 25 volumes of *water* and 75 volumes of *acetonitrile*. Solution (1) contains 0.060% w/v of the substance being examined. For solution (2) dilute 1 volume of solution (1) to 100 volumes. Solution (3) contains 0.060% w/v of *diflucortolone valerate impurity standard BPCRS*.

The chromatographic procedure may be carried out using (a) a stainless steel column (10 cm × 4.6 mm) packed with *end-capped octadecylsilyl silica gel for chromatography* (3 µm) (Spherisorb ODS 2 is suitable), (b) a mixture of equal volumes of *acetonitrile* and *water* as the mobile phase with a flow rate of 2 mL per minute and (c) a detection wavelength of 238 nm. If necessary the composition of the mobile phase may be altered so that the chromatogram obtained with solution (3) shows similar resolution to the reference chromatogram supplied with *diflucortolone valerate impurity standard BPCRS*.

Continue the chromatography for twice the retention time of the principal peak. In the chromatogram obtained with solution (1) the area of any *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%) and the sum of the areas of any *secondary peaks* is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (2%).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

ASSAY

Carry out the method for *liquid chromatography*, Appendix III D, using the following three solutions in a mixture of 25 volumes of *water* and 75 volumes of *acetonitrile* containing (1) 0.03% w/v of the substance being examined, (2) 0.03% w/v of *diflucortolone valerate BPCRS* and (3) 0.03% w/v of *diflucortolone valerate impurity standard BPCRS*.

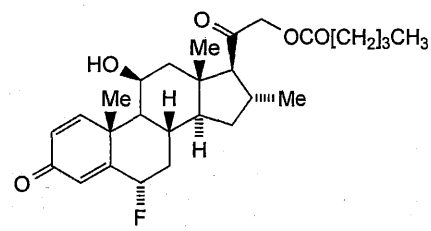
The chromatographic procedure described under Related substances may be used.

Calculate the content of $C_{27}H_{36}F_2O_5$ from the declared content of $C_{27}H_{36}F_2O_5$ in *diflucortolone valerate BPCRS*.

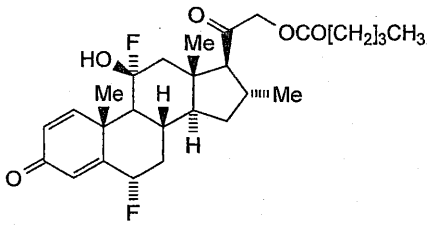
STORAGE

Diflucortolone Valerate should be protected from light.

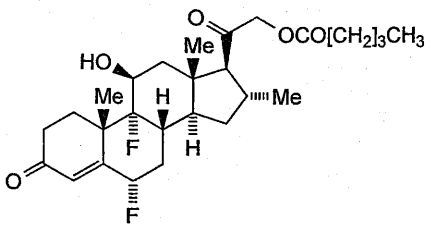
IMPURITIES



A. 6α-fluoro-3,20-dioxo-11β-hydroxy-16α-methylpregna-1,4-dien-21-yl valerate,



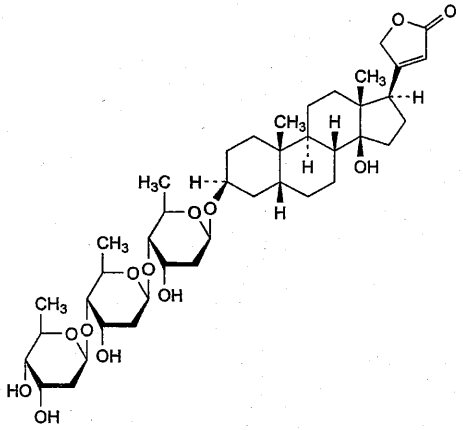
B. 6α,12α-difluoro-3,20-dioxo-11β-hydroxy-16α-methylpregna-1,4-dien-21-yl valerate,



C. 6α,9α-difluoro-3,20-dioxo-11β-hydroxy-16α-methylpregna-1,4-dien-21-yl valerate.

Digitoxin

(Ph. Eur. monograph 0078)



$C_{41}H_{64}O_{13}$ 765 *71-63-6*

Action and use
Na/K-ATPase inhibitor; cardiac glycoside.

Ph Eur

DEFINITION
Digitoxin contains not less than 95.0 per cent and not more than the equivalent of 103.0 per cent of 3β-[(*O*-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-*O*-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl) oxy]-14-hydroxy-5β,14β-card-20(22)-enolide, calculated with reference to the dried substance.

CHARACTERS
A white or almost white powder, practically insoluble in water, freely soluble in a mixture of equal volumes of methanol and methylene chloride, slightly soluble in alcohol and in methanol.

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *digitoxin CRS*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Suspend about 0.5 mg in 0.2 mL of *alcohol (60 per cent V/V) R*. Add 0.1 mL of *dinitrobenzoic acid solution R* and 0.1 mL of *dilute sodium hydroxide solution R*. A violet colour develops.

D. Dissolve about 0.5 mg in 1 mL of *glacial acetic acid R*, heating gently, allow to cool and add 0.05 mL of *ferric chloride solution R1*. Cautiously add 1 mL of *sulfuric acid R*, avoiding mixing the two liquids. A brown ring develops at the interface and on standing a green, then blue colour passes to the upper layer.

TESTS

Appearance of solution
Dissolve 50 mg in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method I*).

Specific optical rotation (*2.2.7*)
Dissolve 0.25 g in *chloroform R* and dilute to 10.0 mL with the same solvent. The specific optical rotation is + 16.0 to + 18.5.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel G plate R*.

Test solution Dissolve 20 mg of the substance to be examined in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 2 mL with the same mixture of solvents.

Reference solution (a) Dissolve 20 mg of *digitoxin CRS* in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 2 mL with the same mixture of solvents.

Reference solution (b) Dilute 0.5 mL of reference solution (a) to 50 mL with a mixture of equal volumes of *methanol R* and *methylene chloride R*.

Reference solution (c) Dissolve 10 mg of *gitoxin CRS* with stirring in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 50 mL with the same mixture of solvents.

Reference solution (d) Dilute 1 mL of reference solution (b) to 2 mL with a mixture of equal volumes of *methanol R* and *methylene chloride R*.

Reference solution (e) Mix 1 mL of reference solution (a) and 1 mL of reference solution (c).

Apply to the plate 5 μL of each solution. Develop immediately over a path of 15 cm using a mixture of 15 volumes of *methanol R*, 40 volumes of *cyclohexane R* and 90 volumes of *methylene chloride R*. Dry the plate in a stream of cold air for 5 min. Repeat the development and dry the plate in a stream of cold air for 5 min. Spray with a mixture of 1 volume of *sulfuric acid R* and 9 volumes of *alcohol R* and heat at 130 °C for 15 min. Examine in daylight.

Gitoxin Any spot corresponding to gitoxin in the chromatogram obtained with the test solution is not more

intense than the spot in the chromatogram obtained with reference solution (c) (2.0 per cent).

Other glycosides Any spot in the chromatogram obtained with the test solution, apart from the principal spot and the spot corresponding to gitoxin, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1.0 per cent).

The test is not valid unless the chromatogram obtained with reference solution (e) shows clearly separated spots corresponding to digitoxin, gitoxin and other glycosides and the spot in the chromatogram obtained with reference solution (d) is clearly visible.

Loss on drying (*2.2.32*)
Not more than 1.5 per cent, determined on 0.500 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on the residue from the test for loss on drying.

ASSAY

Dissolve 40.0 mg in *alcohol R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with *alcohol R*. Prepare a reference solution in the same manner, using 40.0 mg of *digitoxin CRS*. To 5.0 mL of each solution add 3.0 mL of *alkaline sodium picrate solution R*, allow to stand protected from bright light for 30 min and measure the absorbance (*2.2.25*) of each solution at the maximum at 495 nm, using as the compensation liquid a mixture of 5.0 mL of *alcohol R* and 3.0 mL of *alkaline sodium picrate solution R* prepared at the same time.

Calculate the content of $C_{41}H_{64}O_{13}$ from the absorbances measured and the concentrations of the solutions.

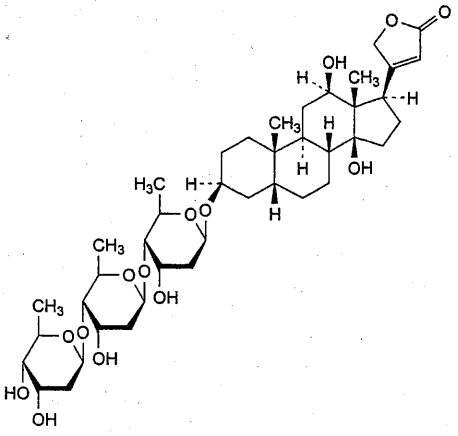
STORAGE

Store protected from light.

Ph Eur

Digoxin

(Ph. Eur. monograph 0079)



$C_{41}H_{64}O_{14}$ 781 *20830-75-5*

Action and use
Na/K-ATPase inhibitor; cardiac glycoside.

Preparations
Digoxin Injection
Paediatric Digoxin Injection
Paediatric Digoxin Oral Solution
Digoxin Tablets

Ph Eur

DEFINITION

3β-[(2,6-Dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide.

Content
96.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder, or colourless crystals.

Solubility
Practically insoluble in water, soluble in a mixture of equal volumes of methanol and methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *digoxin CRS*.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method I*).

Dissolve 50 mg in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Specific optical rotation (*2.2.7*)
+ 13.9 to + 15.9 (dried substance).

Dissolve 0.50 g in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 25.0 mL with the same mixture of solvents.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in 100.0 mL of *methanol R*.

Reference solution (a) Dissolve 10.0 mg of *digoxin CRS* in *methanol R* and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*.

Reference solution (c) Dissolve 2.5 mg of *digoxigenin CRS* (impurity C) in *methanol R* and dilute to 5.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (d) Dissolve 50.0 mg of *lanatoside C R* (impurity H) in *methanol R* and dilute to 100.0 mL with the same solvent. To 1.0 mL of this solution, add 1.0 mL of the test solution and dilute to 20.0 mL with *methanol R*.

Reference solution (e) Dissolve 5.0 mg of *digoxin for peak identification CRS* in *methanol R* and dilute to 10.0 mL with the same solvent.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *acetonitrile R, water R* (10:90 *V/V*);
— *mobile phase B*: *water R, acetonitrile R* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	78	22
5 - 15	78 → 30	22 → 70

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (b), (c), (d) and (e).

Identification of impurities Use the chromatogram supplied with *digoxin for peak identification CRS* and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities A, B, C, E, F, G and K.

Relative retention With reference to digoxin (retention time = about 4.3 min): impurity C = about 0.3; impurity E = about 0.5; impurity F = about 0.6; impurity G = about 0.8; impurity L = about 1.4; impurity K = about 1.6; impurity B = about 2.2; impurity A = about 2.6.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to impurity H and digoxin.

Limits:
— *impurity F*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.5 per cent);
— *impurity C*: not more than 5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *impurities E, K*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *impurity G*: not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);
— *impurities A, B*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurity L*: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *sum of impurities other than A, B, C, E, F, G, K, L*: not more than 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
— *total*: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

The thresholds indicated under Related Substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 0.500 g by drying *in vacuo* in an oven.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{41}H_{64}O_{14}$ from the declared content of *digoxin CRS*.

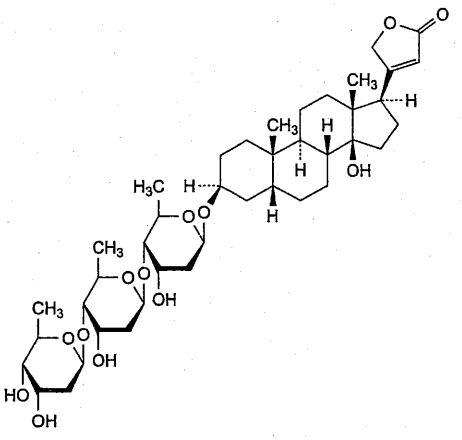
STORAGE

Protected from light.

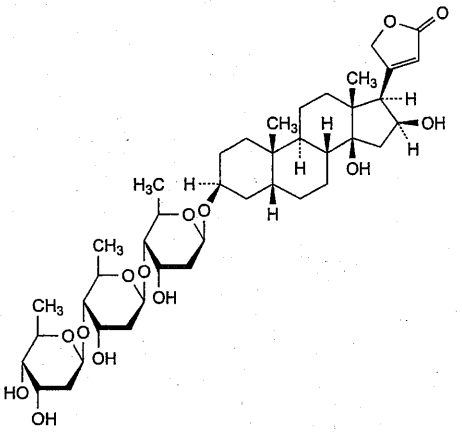
IMPURITIES

Specified impurities *A, B, C, E, F, G, K, L.*

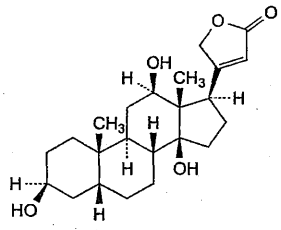
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, H, I, J.*



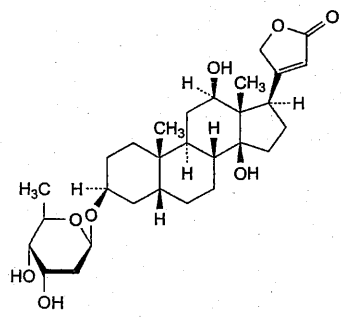
A. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14-hydroxy-5β-card-20(22)-enolide (digitoxin),



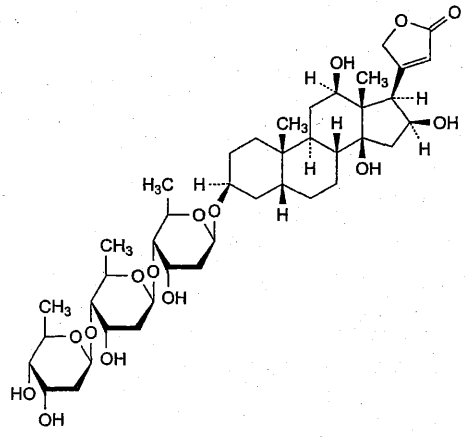
B. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-14,16β-dihydroxy-5β-card-20(22)-enolide (gitoxin),



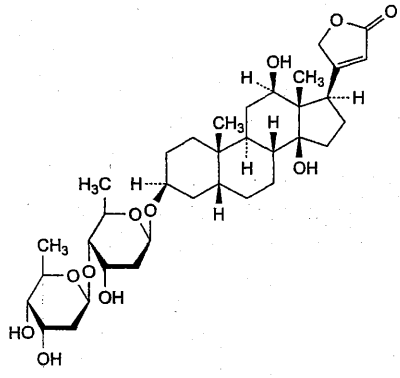
C. 3β,12β,14-trihydroxy-5β-card-20(22)-enolide (digoxigenin),



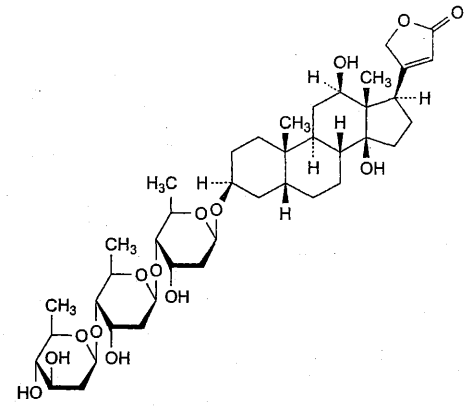
D. 3β-(2,6-dideoxy-β-D-*ribo*-hexopyranosyloxy)-12β,14-dihydroxy-5β-card-20(22)-enolide (digoxigenin monodigitoxoside),



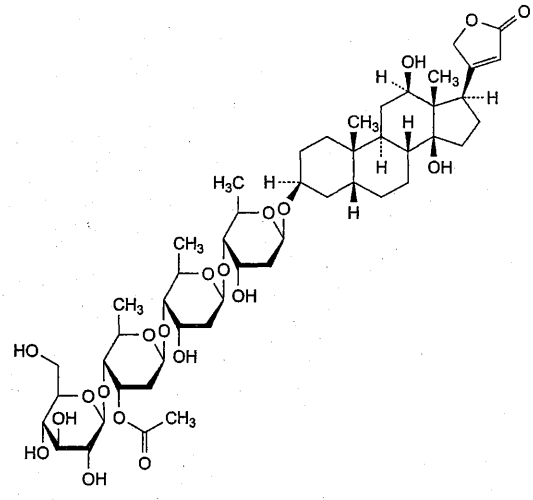
E. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]- 12β,14,16β-trihydroxy-5β-card-20(22)-enolide (diginatin),



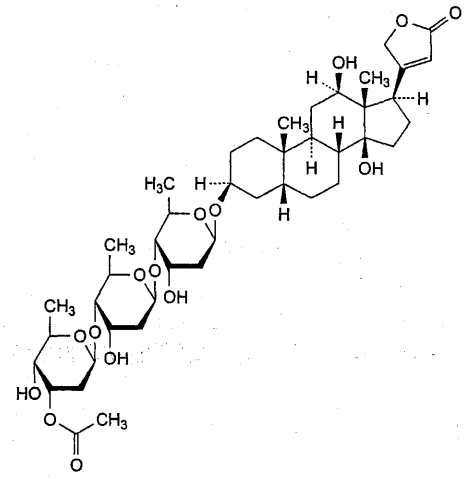
F. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (digoxigenin bisdigitoxoside),



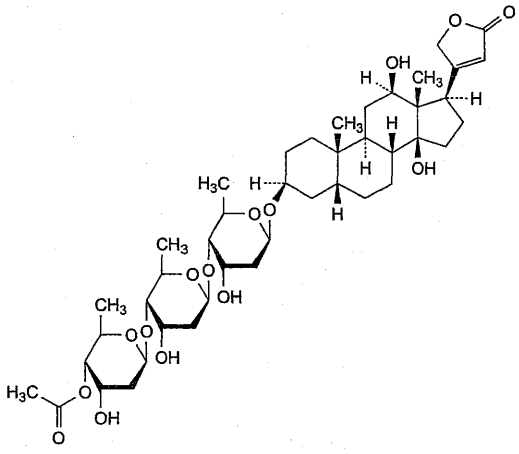
G. 3β-[(2,6-dideoxy-β-D-*arabino*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (neodigoxin),



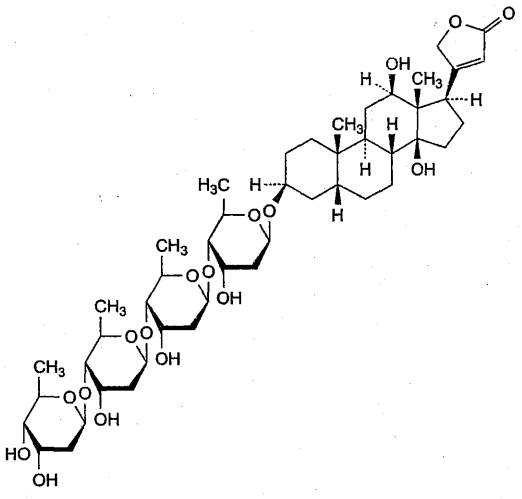
H. 3β-[(β-D-glucopyranosyl-(1→4)-3-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (lanatoside C),



I. 3β-[(3-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (α-acetyldigoxin),



J. 3β-[(4-*O*-acetyl-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (β-acetyldigoxin),



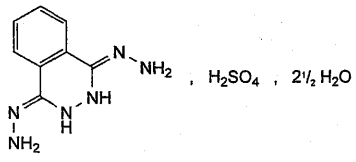
K. 3β-[(2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl-(1→4)-2,6-dideoxy-β-D-*ribo*-hexopyranosyl)oxy]-12β,14-dihydroxy-5β-card-20(22)-enolide (digoxigenin tetrakisdigitoxoside),

L. unknown structure.

Ph Eur

Hydrated Dihydralazine Sulfate

(Ph. Eur. monograph 1310)



$C_8H_{12}N_6O_4S,2½H_2O$ 333.3 *7327-87-9*

Dihydralazine sulfate, anhydrous

Action and use
Vasodilator.

Ph Eur

DEFINITION
(Phthalazine-1,4(2*H*,3*H*)-diylidene)dihydrazine sulfate 2.5-hydrate.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder.

Solubility
Slightly soluble in water, practically insoluble in anhydrous ethanol. It dissolves in dilute mineral acids.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of dihydralazine sulfate hydrated.*

B. Dissolve about 50 mg in 5 mL of *dilute hydrochloric acid R*. The solution gives reaction (a) of sulfates (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.20 g in *dilute nitric acid R* and dilute to 10 mL with the same acid.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in a 6 g/L solution of *glacial acetic acid R* and dilute to 50.0 mL with the same solution.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase containing 0.5 g/L of *sodium edetate R*. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase containing 0.5 g/L of *sodium edetate R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase containing 0.5 g/L of *sodium edetate R*.

Reference solution (c) Dissolve 5 mg of *dihydralazine for system suitability CRS* in a 6 g/L solution of *glacial acetic acid R* and dilute to 5.0 mL with the same solution.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *nitrile silica gel for chromatography R* (5 µm).

Mobile phase Mix 22 volumes of *acetonitrile R1* and 78 volumes of a solution containing 1.44 g/L of *sodium laurilsulfate R* and 0.75 g/L of *tetrabutylammonium bromide R*, then adjust to pH 3.0 with *dilute sulfuric acid R1*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time Twice the retention time of dihydralazine.

Relative retention With reference to dihydralazine: impurity A = about 0.8.

System suitability Reference solution (c):
— the peaks due to impurity A and dihydralazine are baseline separated as in the chromatogram supplied with *dihydralazine for system suitability CRS*.

Limits:
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent);
- *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *sum of impurities other than A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.01 per cent).

Impurity B

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 40.0 mg of *hydrazine sulfate R* (impurity B) in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 25.0 mL with *water R*. To 0.50 mL of this solution, add 0.200 g of the substance to be examined and dissolve in 6 mL of *dilute hydrochloric acid R*, then dilute to 10.0 mL with *water R*. In a centrifuge tube with a ground-glass stopper, place immediately 0.50 mL of this solution and 2.0 mL of a 60 g/L solution of *benzaldehyde R* in a mixture of equal volumes of *methanol R* and *water R*. Shake for 90 s. Add 1.0 mL of *water R* and 5.0 mL of *heptane R*. Shake for 1 min and centrifuge. Use the upper layer.

Reference solution Dissolve 40.0 mg of *hydrazine sulfate R* (impurity B) in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 25.0 mL with *water R*. To 0.50 mL of this solution, add 6 mL of *dilute hydrochloric acid R* and dilute to 10.0 mL with *water R*. In a centrifuge tube with a ground-glass stopper, place 0.50 mL of this solution and 2.0 mL of a 60 g/L solution of *benzaldehyde R* in a mixture of equal volumes of *methanol R* and *water R*. Shake for 90 s. Add 1.0 mL of *water R* and 5.0 mL of *heptane R*. Shake for 1 min and centrifuge. Use the upper layer.

Blank solution Prepare in the same manner as for the reference solution but replacing the 0.50 mL of hydrazine sulfate solution by 0.50 mL of *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 0.3 g/L solution of *sodium edetate R*, *acetonitrile R* (30:70 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 305 nm.

Injection 20 µL.

Relative retention With reference to benzaldehyde: benzaldehyde azine (benzalazine) corresponding to impurity B = about 1.8.

Limit:
- *impurity B*: the area of the peak due to benzaldehyde azine is not greater than twice the area of the corresponding peak in the chromatogram obtained with the reference solution (10 ppm).

Iron (*2.4.9*)

Maximum 20 ppm.

To the residue obtained in the test for sulfated ash add 0.2 mL of *sulfuric acid R* and heat carefully until the acid is almost completely eliminated. Allow to cool and dissolve the residue with heating in 5.5 mL of *hydrochloric acid R1*. Filter the hot solution through a filter previously washed 3 times with *dilute hydrochloric acid R*. Wash the crucible and the filter with 5 mL of *water R*. Combine the filtrate and the washings and neutralise with about 3.5 mL of *strong sodium hydroxide solution R*. Adjust to pH 3-4 with *acetic acid R* and dilute to 20 mL with *water R*. Prepare the standard with 5 mL of *iron standard solution (2 ppm Fe) R* and 5 mL of *water R*.

Loss on drying (*2.2.32*)

13.0 per cent to 15.0 per cent, determined on 1.000 g by drying in an oven at 50 °C at a pressure not exceeding 0.7 kPa for 5 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

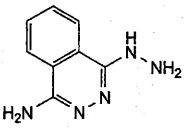
ASSAY

Dissolve 60.0 mg in 25 mL of *water R*. Add 35 mL of *hydrochloric acid R* and titrate slowly with *0.05 M potassium iodate*, determining the end-point potentiometrically (*2.2.20*), using a suitable reference electrode and a platinum indicator electrode.

1 mL of *0.05 M potassium iodate* is equivalent to 7.208 mg of $C_8H_{12}N_6O_4S$.

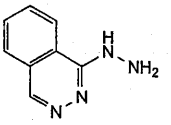
IMPURITIES

Specified impurities *A, B, C.*



A. 4-hydrazinophthalazin-1-amine,

B. $H_2N\text{-}NH_2$: hydrazine,

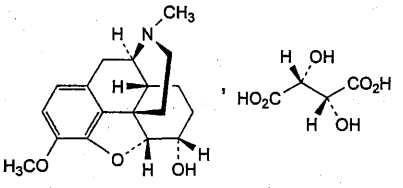


C. (phthalazin-1-yl)hydrazine (hydralazine).

Ph Eur

Dihydrocodeine Tartrate

(Dihydrocodeine Hydrogen Tartrate, Ph. Eur. monograph 1776)



$C_{22}H_{29}NO_9$ 451.5 *5965-13-9*

Action and use

Opioid receptor agonist; analgesic.

Preparations

Co-dydramol Tablets

Dihydrocodeine Injection
Dihydrocodeine Oral Solution
Dihydrocodeine Tablets
Dihydrocodeine Prolonged-release Tablets

Ph Eur

DEFINITION

4,5α-Epoxy-3-methoxy-17-methylmorphinan-6α-ol hydrogen (2*R*,3*R*)-2,3-dihydroxybutanedioate.

Content
98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in alcohol, practically insoluble in cyclohexane.

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of dihydrocodeine hydrogen tartrate.*

B. To about 0.1 g add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R1* and heat on a water-bath. A brownish-yellow colour develops. Add 0.05 mL of *dilute nitric acid R*. The colour does not become red.

C. To 1 mL of solution S (see Tests) add 5 mL of *picric acid solution R*. Heat on a water-bath until a clear solution is obtained. Allow to cool. A precipitate is formed. Filter, wash with 5 mL of *water R* and dry at 100-105 °C. The crystals melt (*2.2.14*) at 220 °C to 223 °C.

D. It gives reaction (b) of tartrates (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

pH (*2.2.3*)
3.2 to 4.2 for solution S.

Specific optical rotation (*2.2.7*)
−70.5 to −73.5 (anhydrous substance).

Dilute 10.0 mL of solution S to 20.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of *codeine phosphate R* in 2.0 mL of the test solution and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 200 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase To 1.0 g of *sodium heptanesulfonate R*, add 10.0 mL of *glacial acetic acid R* and 4.0 mL of a solution of 5.0 mL of *triethylamine R* diluted to 25.0 mL with a mixture of equal volumes of *water R* and *acetonitrile R*. Add 170 mL of *acetonitrile R* and dilute to 1000 mL with *water R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 284 nm.

Injection 20 µL.

Run time 5 times the retention time of dihydrocodeine.

Retention time Dihydrocodeine = about 14 min.

System suitability Reference solution (a):
— *resolution*: minimum of 2 between the peaks due to dihydrocodeine and to impurity A.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *any other peak*: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent); disregard any peak due to tartaric acid (relative retention with reference to dihydrocodeine = about 0.25),
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.7 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

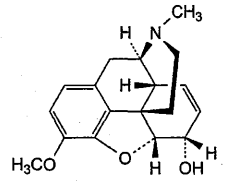
Dissolve 0.350 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 45.15 mg of $C_{22}H_{29}NO_9$.

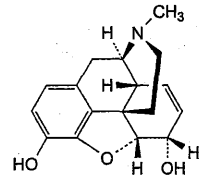
STORAGE

Protected from light.

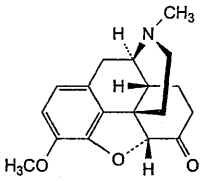
IMPURITIES



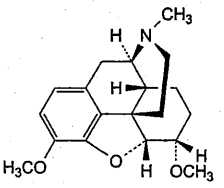
A. 7,8-didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α-ol (codeine),



B. 7,8-didehydro-4,5α-epoxy-17-methylmorphinan-3,6α-diol (morphine),



C. 4,5α-epoxy-3-methoxy-17-methylmorphinan-6-one (hydrocodone),

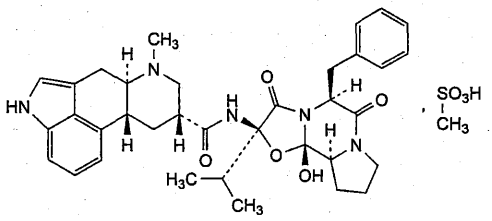


D. 4,5α-epoxy-3,6α-dimethoxy-17-methylmorphinan (tetrahydrothebaine).

Ph Eur

Dihydroergocristine Mesilate

(Ph. Eur. monograph 1416)



$C_{36}H_{45}N_5O_8S$ 708 *24730-10-7*

Action and use
Vasodilator.

Ph Eur

DEFINITION

(6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-Benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxo-octahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate.

Content
98.0 per cent to 102.0 per cent (dried substance).

PRODUCTION

It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in dihydroergocristine mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation.
The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance
White or almost white, fine crystalline powder.

Solubility
Slightly soluble in water, soluble in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *dihydroergocristine mesilate CRS*.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 0.10 g of *dihydroergocristine mesilate CRS* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *concentrated ammonia R, dimethylformamide R, ether R* (2:15:85 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate protected from light.

Drying In a current of cold air for 5 min.

Detection Spray with *dimethylaminobenzaldehyde solution R7* and dry in a current of hot air for 2 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.20 g of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 0.20 g of *methanesulfonic acid R* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5 mL with the same mixture of solvents. Dilute 1 mL of the solution to 10 mL with a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R*.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *water R, concentrated ammonia R, butanol R, acetone R* (5:10:20:65 *V/V/V/V*).

Application 10 µL.

Development Over a path of 10 cm protected from light.

Drying In a current of cold air for not more than 1 min.

Detection Spray with a 1 g/L solution of *bromocresol purple R* in *methanol R*, adjusting the colour to violet-red with one drop of *dilute ammonia R1* and dry the plate in a current of hot air at 100 °C.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).

Dissolve 0.50 g in *methanol R* and dilute to 25.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.0.

Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−43 to −37 (dried substance).

Dissolve 0.250 g in *anhydrous pyridine R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test and preparation of the solutions protected from bright light.*

Test solution Dissolve 75.0 mg of the substance to be examined in 10 mL of *acetonitrile R*. Add 10 mL of a 1.0 g/L solution of *phosphoric acid R* and dilute to 50.0 mL with *water R*.

Reference solution Dissolve 20.0 mg of *codergocrine mesilate CRS* in 10 mL of *acetonitrile R*. Add 10 mL of a 1.0 g/L solution of *phosphoric acid R* and dilute to 50.0 mL with *water R*. Dilute 6.0 mL of the solution to 50.0 mL with a mixture of 20 volumes of *acetonitrile R*, 20 volumes of a 1.0 g/L solution of *phosphoric acid R* and 60 volumes of *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 10 nm and a carbon loading of 19 per cent.

Mobile phase:
— *mobile phase A*: mix 100 volumes of *acetonitrile R* with 900 volumes of *water R* and add 10 volumes of *triethylamine R*;
— *mobile phase B*: mix 100 volumes of *water R* with 900 volumes of *acetonitrile R* and add 10 volumes of *triethylamine R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	75	25
5 - 20	75 → 25	25 → 75

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Relative retention With reference to dihydroergocristine (retention time = about 13.7 min): impurity F = about 0.8; impurity H = about 0.9; impurity I = about 1.02.

System suitability Reference solution:
— the chromatogram shows 4 peaks;
— *resolution*: minimum 1 between the peaks due to dihydroergocristine and impurity I.

Limits:
— *any impurity*: not more than the area of the peak due to dihydroergocristine in the chromatogram obtained with the reference solution (1 per cent);
— *total*: not more than twice the area of the peak due to dihydroergocristine in the chromatogram obtained with the reference solution (2 per cent);
— *disregard limit*: 0.1 times the area of the peak due to dihydroergocristine in the chromatogram obtained with the reference solution (0.1 per cent).

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 0.500 g by drying under high vacuum at 80 °C.

ASSAY

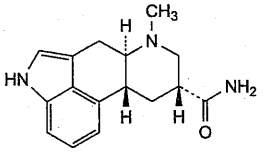
Dissolve 0.300 g in 60 mL of *pyridine R*. Pass a stream of *nitrogen R* over the surface of the solution and titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*). Note the volume used at the second point of inflexion.

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 35.39 mg of $C_{36}H_{45}N_5O_8S$.

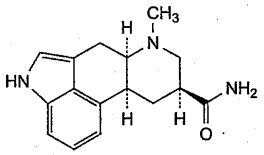
STORAGE

Store protected from light.

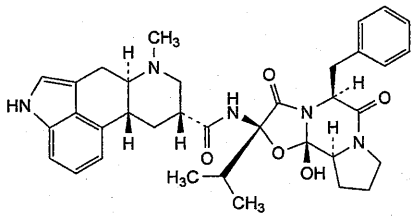
IMPURITIES



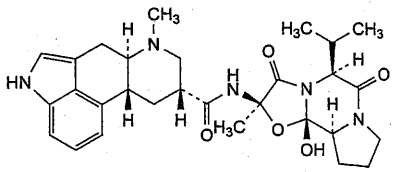
A. (6a*R*,9*R*,10a*R*)-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (6-methylergoline-8β-carboxamide),



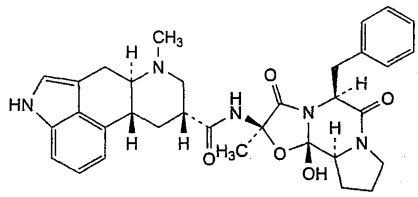
B. (6a*R*,9*S*,10a*S*)-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (6-methylisoergoline-8α-carboxamide),



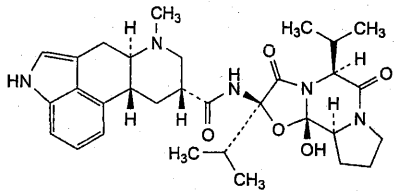
C. (6a*R*,9*R*,10a*R*)-*N*-[(2*S*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (2′-epidihydroergocristine),



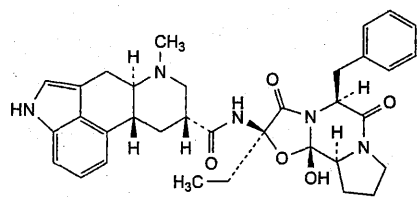
D. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-methyl-5-(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (dihydroergosine),



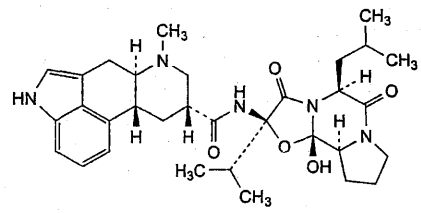
E. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline- 9-carboxamide (dihydroergotamine),



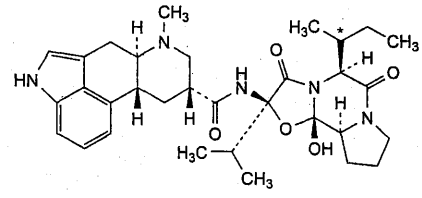
F. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)10b-hydroxy-2,5-bis(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (dihydroergocornine),



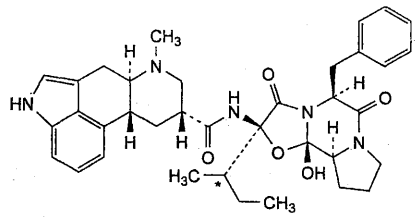
G. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-2-ethyl-10b-hydroxy-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (dihydroergostine),



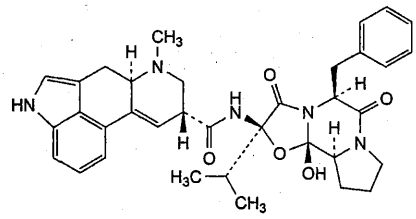
H. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-(2-methylpropyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (α-dihydroergocryptine),



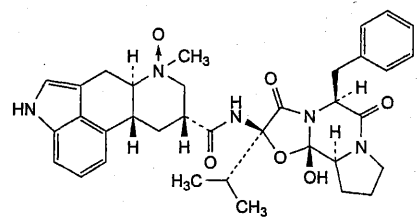
I. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-10b-hydroxy-2-(1-methylethyl)-5-[(1*RS*-1-methylpropyl]-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (β-dihydroergocryptine or epicriptine),



J. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-[(1*RS*)-1-methylpropyl]-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (dihydroergosedmine),



K. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergocristine),

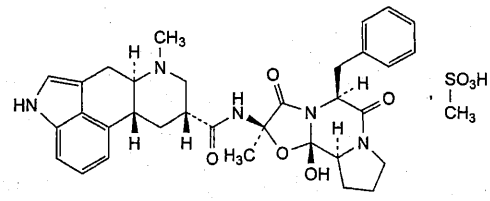


L. (6a*R*,7*RS*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide 7-oxide (dihydroergocristine 6-oxide).

Ph Eur

Dihydroergotamine Mesilate

(Ph. Eur. monograph 0551)



$C_{34}H_{41}N_5O_8S$ 680 *6190-39-2*

Action and use
Vasodilator.

Ph Eur

DEFINITION

(6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-Benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide methanesulfonate.

Content

98.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in dihydroergotamine mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation.

The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Slightly soluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: B, C.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 5.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent.

Spectral range 250-350 nm.

Absorption maxima At 281 nm and 291 nm.

Shoulder At 275 nm.

Absorbance Negligible above 320 nm.

Specific absorbance at the absorption maximum at 281 nm 95 to 105 (dried substance).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dihydroergotamine mesilate CRS.*

C. Thin-layer chromatography (*2.2.27*). *Prepare the reference solution and the test solution immediately before use.*

Solvent mixture *methanol R, methylene chloride R* (10:90 *V/V*).

Test solution Dissolve 5 mg of the substance to be examined in the solvent mixture and dilute to 2.5 mL with the solvent mixture.

Reference solution Dissolve 5 mg of *dihydroergotamine mesilate CRS* in the solvent mixture and dilute to 2.5 mL with the solvent mixture.

Plate *TLC silica gel G plate R.*

Mobile phase *concentrated ammonia R, methanol R, ethyl acetate R, methylene chloride R* (1:6:50:50 *V/V/V/V*).

Application 5 µL.

Development Protected from light, over a path of 15 cm; dry in a current of cold air for not longer than 1 min and repeat the development protected from light over a path of 15 cm using a freshly prepared amount of the mobile phase.

Drying In a current of cold air.

Detection Spray abundantly with *dimethylaminobenzaldehyde solution R7* and dry in a current of hot air for about 2 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 g of the substance to be examined, add 5 mL of *dilute hydrochloric acid R* and shake for about 5 min. Filter, then add 1 mL of *barium chloride solution R1*. The filtrate remains clear. Mix 0.1 g of the substance to be examined with 0.4 g of powdered *sodium hydroxide R*, heat to fusion and continue to heat for 1 min. Cool, add 5 mL of *water R*, boil and filter. Acidify the filtrate with *hydrochloric acid R1* and filter again. The filtrate gives reaction (a) of sulfates (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 or BY_7 (*2.2.2, Method II*).

Dissolve 0.10 g in a mixture of 0.1 mL of a 70 g/L solution of *methanesulfonic acid R* and 50 mL of *water R*.

pH (*2.2.3*)

4.4 to 5.4.

Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Specific optical rotation (*2.2.7*)

−47 to −42 (dried substance).

Dissolve 0.250 g in *anhydrous pyridine R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture *acetonitrile R, water R* (50:50 *V/V*).

Test solution Dissolve 70 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 7 mg of the substance to be examined and 6.8 mg of *ergotamine tartrate CRS* (impurity A) (equivalent to 7 mg of ergotamine mesilate) in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 5 mL of this solution to 10 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *dihydroergotamine for peak identification CRS* (containing impurities A, B, C, D and E) in the solvent mixture, add 100 µL of *dilute sulfuric acid R* and dilute to 5 mL with the solvent mixture.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);

— *temperature*: 25 °C.

Mobile phase:

— *mobile phase A*: 3 g/L solution of *sodium heptanesulfonate monohydrate R* adjusted to pH 2.0 with *phosphoric acid R*;

— *mobile phase B*: mobile phase A, *acetonitrile for chromatography R* (20:80 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	58 → 40	42 → 60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *dihydroergotamine for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to dihydroergotamine (retention time = about 6.5 min): impurity D = about 0.7; impurity C = about 0.86; impurity A = about 0.95; impurity B = about 1.2; impurity E = about 1.4.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity A and dihydroergotamine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.3; impurity C = 1.3;
— *impurities B, E*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities A, D*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 4.0 per cent, determined on 0.500 g by drying at 105 °C at a pressure not exceeding 0.1 kPa for 5 h.

ASSAY

Dissolve 0.500 g in a mixture of 10 mL of *anhydrous acetic acid R* and 70 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

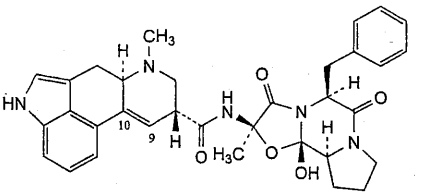
1 mL of *0.1 M perchloric acid* is equivalent to 68.00 mg of $C_{34}H_{41}N_5O_8S$.

STORAGE

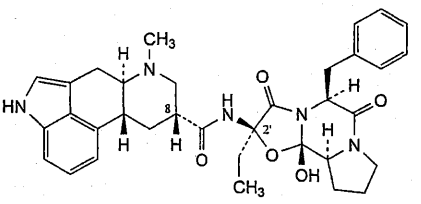
Protected from light.

IMPURITIES

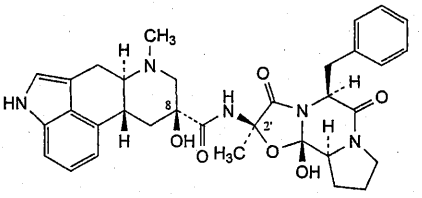
Specified impurities *A, B, C, D, E.*



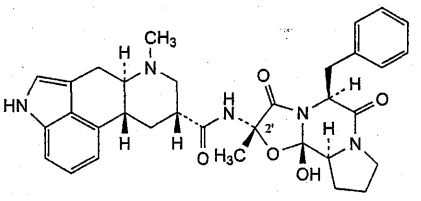
A. (6a*R*,9*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergotamine),



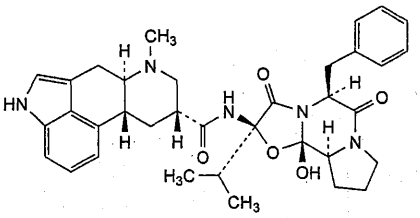
B. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-2-ethyl-10b-hydroxy-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (9,10-dihydroergostine),



C. (6a*R*,9*S*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-9-hydroxy-7-methyl-4,6,6a,7,8,9,10,10aoctahydroindolo[4,3-*fg*]quinoline-9-carboxamide (8-hydroxy-9,10-dihydroergotamine),



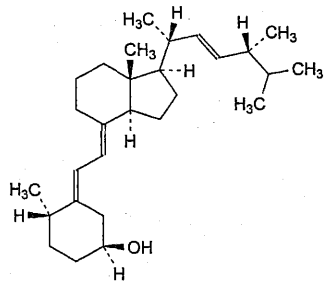
D. (6a*R*,9*R*,10a*R*)-*N*-[(2*S*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10aoctahydroindolo[4,3-*fg*]quinoline-9-carboxamide (2′-*epi*-9,10-dihydroergotamine),



E. (6a*R*,9*R*,10a*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-(1-methylethyl)-3,6-dioxo-octahydro-8*H*-oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-*fg*]quinoline-9-carboxamide (dihydroergocristine).

Dihydrotachysterol

(Ph. Eur. monograph 2014)



$C_{28}H_{46}O$ 398.7 *67-96-9*

Action and use
Vitamin D analogue.

Ph Eur

DEFINITION

(5*E*,7*E*,22*E*)-9,10-Seco-10α-ergosta-5,7,22-trien-3β-ol.

Content
97.0 per cent to 102.0 per cent.

CHARACTERS

Appearance
Colourless crystals or white or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in hexane, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dihydrotachysterol CRS.*

If the spectra obtained in the solid state show differences, record new spectra using the residues after recrystallisation from *methanol R.*

TESTS

Specific optical rotation (*2.2.7*)
+ 99 to + 103.

Dissolve 0.500 g in *ethanol (96 per cent) R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.00 mg of the substance to be examined in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 1.0 mg of *dihydrotachysterol for system suitability CRS* (containing impurities A, B and C) in *acetonitrile R* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dissolve 10.00 mg of *dihydrotachysterol CRS* in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (c) Dilute 5.0 mL of the test solution to 100.0 mL with *acetonitrile R.* Dilute 5.0 mL of this solution to 50.0 mL with *acetonitrile R.*

Column:
— *size: l* = 0.25 m, Ø = 3.0 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography for separation of polycyclic aromatic hydrocarbons R* (3 µm);
— *temperature:* 40 °C.

Mobile phase *decanol R, water for chromatography R, acetonitrile R1* (1:25:1000 *V/V/V*).

Flow rate 0.5 mL/min.

Detection Variable-wavelength spectrophotometer capable of operating at 251 nm and at 203 nm.

Injection 5 µL of the test solution and reference solutions (a) and (c).

Run time Twice the retention time of dihydrotachysterol.

Identification of impurities Reference solution (a):
— use the chromatogram obtained at 203 nm and the chromatogram obtained at 203 nm supplied with *dihydrotachysterol for system suitability CRS* to identify the peak due to impurity A;
— use the chromatogram obtained at 251 nm and the chromatogram obtained at 251 nm supplied with *dihydrotachysterol for system suitability CRS* to identify the peaks due to impurities B and C.

Relative retention With reference to dihydrotachysterol (retention time = about 15 min): impurity B = about 0.9; impurity C = about 1.2; impurity A (not visible at 251 nm, detected at 203 nm) = about 1.2.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 4, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dihydrotachysterol in the chromatogram obtained at 251 nm.

Examine the chromatogram obtained at 203 nm for impurity A and the chromatogram obtained at 251 nm for impurities other than A.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurities B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *total (including A)*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) at 251 nm (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.32*)
Maximum 0.10 per cent, determined on 40.0 mg.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 251 nm.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{28}H_{46}O$ taking into account the assigned content of *dihydrotachysterol CRS.*

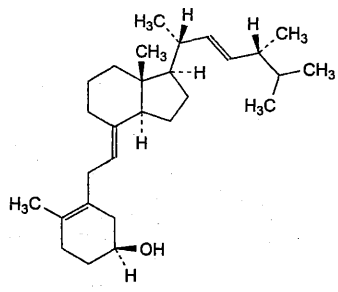
STORAGE

Under an inert gas, in an airtight container, at a temperature of 2 °C to 8 °C.

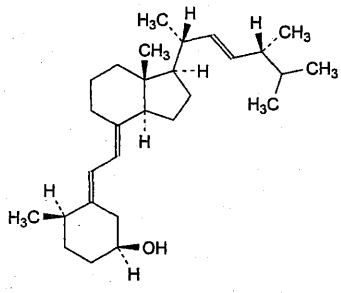
The contents of an opened container are to be used immediately.

IMPURITIES

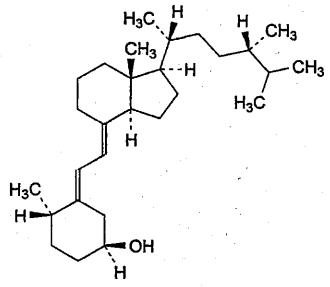
Specified impurities *A, B, C.*



A. (7*E*,22*E*)-9,10-secoergosta-5(10),7,22-trien-3β-ol (dihydrovitamin D_2-I),



B. (5*E*,7*E*,22*E*)-9,10-secoergosta-5,7,22-trien-3β-ol (dihydrovitamin D_2-IV),

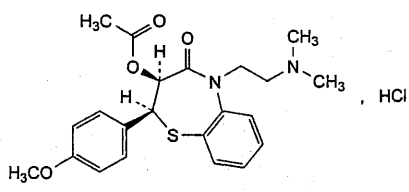


C. (5*E*,7*E*)-9,10-seco-10α-ergosta-5,7-dien-3β-ol (dihydrotachysterol$_4$).

Ph Eur

Diltiazem Hydrochloride

(Ph. Eur. monograph 1004)



$C_{22}H_{27}ClN_2O_4S$ 451.0 *33286-22-5*

Action and use

Calcium channel blocker.

Preparations

Diltiazem Oral Suspension

Diltiazem Prolonged-release Tablets

Ph Eur

DEFINITION

Hydrochloride of (2*S*,3*S*)-5-[2-(dimethylamino)ethyl]-2-(4-methoxyphenyl)-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepin-3-yl acetate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, in methanol and in methylene chloride, slightly soluble in anhydrous ethanol.

mp

About 213 °C, with decomposition.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diltiazem hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *methylene chloride R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve 50 mg of *diltiazem hydrochloride CRS* in *methylene chloride R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetic acid R, water R, methylene chloride R, anhydrous ethanol R* (1:3:10:12 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 50 mg in 5 mL of *water R*. Add 1 mL of *ammonium reineckate solution R*. A pink precipitate is produced.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.00 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

4.3 to 5.3.

Dilute 2.0 mL of solution S to 10.0 mL with *carbon dioxide-free water R*.

Specific optical rotation (*2.2.7*)

+ 115 to + 120 (dried substance).

Dilute 5.0 mL of solution S to 25.0 mL with *water R*.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 200.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *diltiazem for system suitability CRS* (containing impurity A) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *diltiazem impurity F CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Mix 5 volumes of *anhydrous ethanol R*, 25 volumes of *acetonitrile R* and 70 volumes of a solution containing 6.8 g/L of *potassium dihydrogen phosphate R* and 0.1 mL/L of *N,N-dimethyloctylamine R*, adjusted to pH 4.5 with *dilute phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL.

Run time 5 times the retention time of diltiazem.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to diltiazem (retention time = about 5 min): impurity F = about 0.5; impurity A = about 0.8.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurity A and diltiazem;
— *symmetry factor*: maximum 2.0 for the peak due to impurity A; if necessary, adjust the concentration of *N,N*-dimethyloctylamine in the mobile phase.

Limits:
— *impurity F*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in a mixture of 2 mL of *anhydrous formic acid R* and 60 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 45.1 mg of $C_{22}H_{27}ClN_2O_4S$.

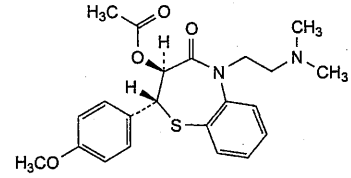
STORAGE

In an airtight container, protected from light.

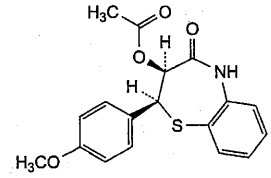
IMPURITIES

Specified impurities *F.*

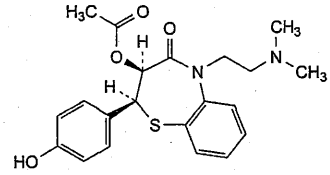
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E.*



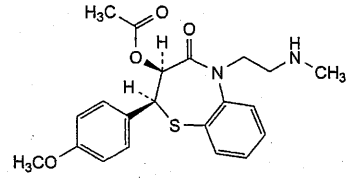
A. (2*R*,3*S*)-5-[2-(dimethylamino)ethyl]-2-(4-methoxyphenyl)-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepin-3-yl acetate,



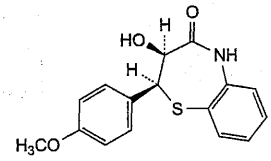
B. (2*S*,3*S*)-2-(4-methoxyphenyl)-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepin-3-yl acetate,



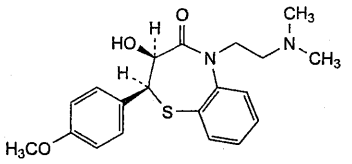
C. (2*S*,3*S*)-5-[2-(dimethylamino)ethyl]-2-(4-hydroxyphenyl)-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepin-3-yl acetate,



D. (2*S*,3*S*)-2-(4-methoxyphenyl)-5-[2-(methylamino)ethyl]-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepin-3-yl acetate,



E. (2*S*,3*S*)-3-hydroxy-2-(4-methoxyphenyl)-2,3-dihydro-1,5-benzothiazepin-4(5*H*)-one,

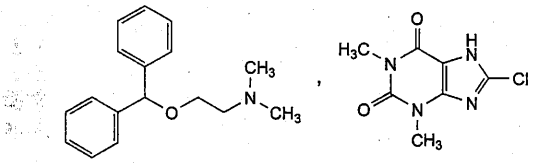


F. (2*S*,3*S*)-5-[2-(dimethylamino)ethyl]-3-hydroxy-2-(4-methoxyphenyl)-2,3-dihydro-1,5-benzothiazepin-4(5*H*)-one.

Ph Eur

Dimenhydrinate

(Ph. Eur. monograph 0601)



$C_{24}H_{28}ClN_5O_3$ 470.0 *523-87-5*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparation
Dimenhydrinate Tablets

Ph Eur

DEFINITION
Diphenhydramine [2-(diphenylmethoxy)-*N*,*N*-dimethylethanamine] 8-chlorotheophylline (8-chloro-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione).

Content
— *diphenhydramine* ($C_{17}H_{21}NO$; M_r 255.4): 53.0 per cent to 55.5 per cent (dried substance);
— *8-chlorotheophylline* ($C_7H_7ClN_4O_2$; M_r 214.6): 44.0 per cent to 46.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Slightly soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 102 °C to 106 °C.

B. Dissolve 0.1 g in a mixture of 3 mL of *water R* and 3 mL of *ethanol (96 per cent) R*, add 6 mL of *water R* and 1 mL of *dilute hydrochloric acid R* and cool in iced water for 30 min, scratching the wall of the tube with a glass rod if necessary to initiate crystallisation. Dissolve about 10 mg of the precipitate obtained in 1 mL of *hydrochloric acid R*, add 0.1 g of *potassium chlorate R* and evaporate to dryness in a porcelain dish. A reddish residue is obtained that becomes violet-red when exposed to ammonia vapour.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dimenhydrinate CRS.*

D. Dissolve 0.2 g in 10 mL of *ethanol (96 per cent) R.* Add 10 mL of *picric acid solution R* and initiate crystallisation by scratching the wall of the tube with a glass rod. The precipitate, washed with *water R* and dried at 100-105 °C, melts (*2.2.14*) at 130 °C to 134 °C.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in *ethanol (96 per cent) R* and dilute to 20 mL with the same solvent.

pH (*2.2.3*)
7.1 to 7.6 for the filtrate.

To 0.4 g add 20 mL of *carbon dioxide-free water R*, shake for 2 min and filter.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (18:82 *V/V*).

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 57 mg of *diphenhydramine hydrochloride CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of *diphenhydramine impurity A CRS* (impurity F) in 5.0 mL of reference solution (a) and dilute to 50.0 mL with the solvent mixture.

Reference solution (d) Dissolve the contents of a vial of *dimenhydrinate for peak identification CRS* (containing impurities A and E) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: dissolve 10.0 g of *triethylamine R2* in 950 mL of *water R*, adjust to pH 2.5 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Flow rate (mL/min)
0 - 2	82	18	1.2
2 - 15	82 → 50	18 → 50	1.2
15 - 20	50 → 20	50 → 80	1.2 → 2.0
20 - 30	20	80	2.0

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *dimenhydrinate for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A and E; use the chromatogram obtained with reference solution (c) to identify impurity F.

Relative retention With reference to diphenhydramine (retention time = about 13 min): impurity A = about 0.3; impurity E = about 0.7; impurity F = about 0.95.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity F and diphenhydramine.

Limits:
— *impurities A, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurity E*: not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo*.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Diphenhydramine

Dissolve 0.200 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 25.54 mg of $C_{17}H_{21}NO$.

8-Chlorotheophylline

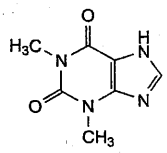
To 0.800 g add 50 mL of *water R*, 3 mL of *dilute ammonia R1* and 0.6 g of *ammonium nitrate R* and heat on a water-bath for 5 min. Add 25.0 mL of *0.1 M silver nitrate* and continue heating on a water-bath for 15 min with frequent swirling. Cool, add 25 mL of *dilute nitric acid R* and dilute to 250.0 mL with *water R*. Filter and discard the first 25 mL of the filtrate. Using 5 mL of *ferric ammonium sulfate solution R2* as indicator, titrate 100.0 mL of the filtrate with *0.1 M ammonium thiocyanate* until a yellowish-brown colour is obtained.

1 mL of *0.1 M silver nitrate* is equivalent to 21.46 mg of $C_7H_7ClN_4O_2$.

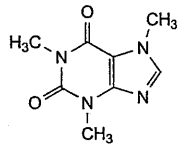
IMPURITIES

Specified impurities *A, E, F.*

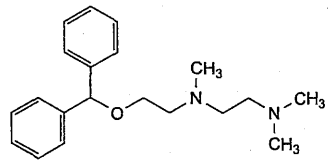
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, G, H, I, J, K.*



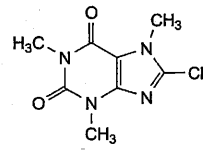
A. 1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theophylline),



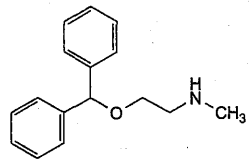
C. 1,3,7-trimethyl-3,7-dihydro-1*H*-purine-2,6-dione (caffeine),



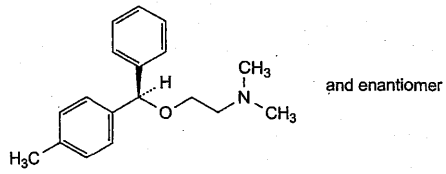
D. *N*-[2-(diphenylmethoxy)ethyl]-*N*,*N*′,*N*′-trimethylethane-1,2-diamine,



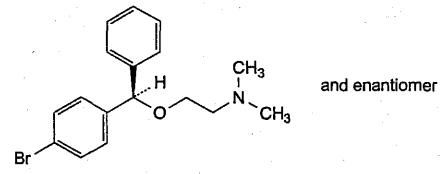
E. 8-chloro-1,3,7-trimethyl-3,7-dihydro-1*H*-purine-2,6-dione (8-chlorocaffeine),



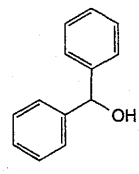
F. 2-(diphenylmethoxy)-*N*-methylethanamine (diphenhydramine impurity A),



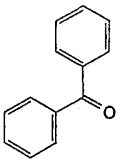
G. *N*,*N*-dimethyl-2-[(*RS*)-(4-methylphenyl)(phenyl)methoxy] ethanamine (4-methyldiphenhydramine),



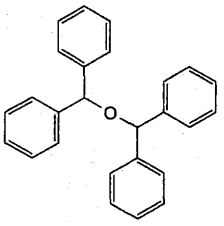
H. 2-[(*RS*)-(4-bromophenyl)-(phenyl)methoxy]-*N*,*N*-dimethylethanamine (4-bromodiphenhydramine),



I. diphenylmethanol (benzhydrol),



J. diphenylmethanone (benzophenone),

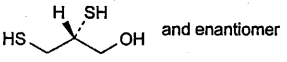


K. [oxybis(methanetriyl)]tetrabenzene.

Ph Eur

Dimercaprol

(Ph. Eur. monograph 0389)



$C_3H_8OS_2$ 124.2 *59-52-9*

Action and use
Chelating agent for use in heavy metal poisoning.

Preparation
Dimercaprol Injection

When B.A.L. is prescribed or demanded, Dimercaprol shall be dispensed or supplied.

Ph Eur

DEFINITION
(2*RS*)-2,3-Disulfanylpropan-1-ol.

Content
98.5 per cent to 101.5 per cent.

CHARACTERS

Appearance
Clear, colourless or slightly yellow liquid.

Solubility
Soluble in water and in arachis oil, miscible with ethanol (96 per cent) and with benzyl benzoate.

IDENTIFICATION
A. Dissolve 0.05 mL in 2 mL of *water R*. Add 1 mL of *0.05 M iodine*. The colour of the iodine is discharged immediately.

B. Dissolve 0.1 mL in 5 mL of *water R* and add 2 mL of *copper sulfate solution R*. A bluish-black precipitate is formed which quickly becomes dark grey.

C. In a ground-glass-stoppered tube, suspend 0.6 g of *sodium bismuthate R*, previously heated to 200 °C for 2 h, in a mixture of 2.8 mL of *dilute phosphoric acid R* and 6 mL of *water R*. Add 0.2 mL of the substance to be examined, mix and allow to stand for 10 min with frequent shaking. To 1 mL of the supernatant add 5 mL of a 4 g/L solution of *chromotropic acid, sodium salt R* in *sulfuric acid R* and mix. Heat in a water-bath for 15 min. A violet-red colour develops.

TESTS

Appearance
It is clear (*2.2.1*) and not more intensely coloured than reference solution B_6 or BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
Dissolve 0.2 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Add 0.25 mL of *bromocresol green solution R* and 0.3 mL of *0.01 M hydrochloric acid*. The solution is yellow. Not more than 0.5 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Refractive index (*2.2.6*)
1.568 to 1.574.

Halides
To 2.0 g add 25 mL of *alcoholic potassium hydroxide solution R* and boil under a reflux condenser for 2 h. Eliminate the ethanol by evaporation in a stream of hot air. Add 20 mL of *water R* and cool. Add 40 mL of *water R* and 10 mL of *strong hydrogen peroxide solution R*, boil gently for 10 min, cool and filter rapidly. Add 10 mL of *dilute nitric acid R* and 5.0 mL of *0.1 M silver nitrate*. Using 2 mL of *ferric ammonium sulfate solution R2* as indicator, titrate with *0.1 M ammonium thiocyanate* until a reddish-yellow colour is obtained. Carry out a blank titration. The difference between the titration volumes is not greater than 1.0 mL.

ASSAY
Dissolve 0.100 g in 40 mL of *methanol R*. Add 20 mL of *0.1 M hydrochloric acid* and 50.0 mL of *0.05 M iodine*. Allow to stand for 10 min and titrate with *0.1 M sodium thiosulfate*. Carry out a blank titration.

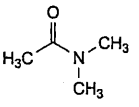
1 mL of *0.05 M iodine* is equivalent to 6.21 mg of $C_3H_8OS_2$.

STORAGE
In a well-filled, airtight container, protected from light, at a temperature of 2 °C to 8 °C.

Ph Eur

Dimethylacetamide

(Ph. Eur. monograph 1667)



C_4H_9NO 87.1 *127-19-5*

Action and use
Excipient.

Ph Eur

DEFINITION
N,*N*-Dimethylacetamide.

CHARACTERS

Appearance
Clear, colourless, slightly hygroscopic liquid.

Solubility
Miscible with water, with ethanol (96 per cent), and with most common organic solvents.

bp
About 165 °C.

IDENTIFICATION

First identification: C.

Second identification: A, B, D.

A. Relative density (*2.2.5*): 0.941 to 0.944.

B. Refractive index (*2.2.6*): 1.435 to 1.439.

C. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Films.

Comparison *Ph. Eur. reference spectrum of dimethylacetamide.*

D. Dilute 50 mg with 1 mL of *methanol R*. Add 1 mL of a 15 g/L solution of *hydroxylamine hydrochloride R* and mix. Add 1 mL of *dilute sodium hydroxide solution R*, mix and allow to stand for 30 min. Add 1 mL of *dilute hydrochloric acid R* and add 1 mL of a 100 g/L solution of *ferric chloride R* in *0.1 M hydrochloric acid*. A reddish-brown colour develops, reaching a maximum intensity after about 5 min.

TESTS

Appearance
The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Acidity
Dilute 50 mL with 50 mL of *water R* previously adjusted with *0.02 M potassium hydroxide* or *0.02 M hydrochloric acid* to a bluish-green colour, using 0.5 mL of *bromothymol blue solution R1* as indicator. Not more than 5.0 mL of *0.02 M potassium hydroxide* is required to restore the initial (bluish-green) colour.

Alkalinity
To 50 mL add 50 mL of *water R* previously adjusted with *0.02 M potassium hydroxide* or *0.02 M hydrochloric acid* to a yellow colour, using 0.5 mL of *bromothymol blue solution R1* as indicator. Not more than 0.5 mL of *0.02 M hydrochloric acid* is required to restore the initial (yellow) colour.

Related substances
Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution The substance to be examined.

Reference solution (a) Dilute a mixture of 1 mL of the substance to be examined and 1 mL of *dimethylformamide R* (impurity B) to 20 mL with *methylene chloride R*.

Reference solution (b) Dilute 1 mL of the substance to be examined to 20.0 mL with *methylene chloride R*. Dilute 0.1 mL of the solution to 10.0 mL with *methylene chloride R*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *nitrogen for chromatography R*.

Linear velocity 30 cm/s.

Split ratio 1:20.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 15	80 → 200
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 0.5 µL.

System suitability:
— *resolution*: minimum 5.0 between the peaks due to dimethylacetamide and impurity B in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
— *any impurity*: maximum 0.1 per cent;
— *total*: maximum 0.3 per cent;
— *disregard limit*: the area of the peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Non-volatile matter
Maximum 20 ppm.

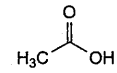
Evaporate 50 g to dryness by suitable means at a pressure not exceeding 1 kPa and on a water-bath. Dry the residue in an oven at 170-175 °C. The residue weighs not more than 1 mg.

Water (*2.5.32*)
Maximum 0.1 per cent, determined on 0.100 g.

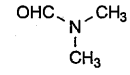
STORAGE

In an airtight container, protected from light.

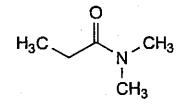
IMPURITIES



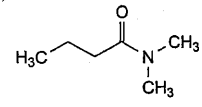
A. acetic acid,



B. *N*,*N*-dimethylformamide,



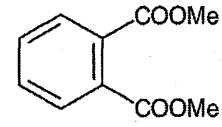
C. *N*,*N*-dimethylpropanamide,



D. *N*,*N*-dimethylbutanamide.

Ph Eur

Dimethyl Phthalate



$C_{10}H_{10}O_4$ 194.2 *131-11-3*

Action and use
Insect repellent.

DEFINITION

Dimethyl Phthalate contains not less than 99.0% and not more than 100.5% w/w of $C_{10}H_{10}O_4$.

CHARACTERISTICS

A colourless or faintly coloured liquid.

Slightly soluble in *water*; miscible with *ethanol (96%)*, with *ether* and with most organic solvents.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of dimethyl phthalate *(RS 105)*.

B. Gently boil 1 g with 5 mL of 2M *methanolic potassium hydroxide* for 10 minutes, add 5 mL of *water*, evaporate the mixture to half its volume and cool. Add 1 mL of *hydrochloric acid*, filter, melt the dried precipitate in a small tube, add 0.5 g of *resorcinol* and 0.05 mL of *chloroform* and heat to about 180° for 3 minutes. Cool, add 1 mL of 5M *sodium hydroxide* and pour into *water*. An intense yellowish green fluorescence is produced.

TESTS

Acidity

Mix 20 mL with 50 mL of *ethanol (96%)* previously neutralised to *phenolphthalein solution R1*. Not more than 0.1 mL of *0.1M sodium hydroxide VS* is required to neutralise the solution using *phenolphthalein solution R1* as indicator.

Refractive index

1.515 to 1.517, Appendix V E.

Weight per mL

1.186 to 1.192 g, Appendix V G.

Related substances

Prepare a 0.075% w/v solution of *phenyl benzoate* (internal standard) in *chloroform* (solution A). Carry out the method for *gas chromatography*, Appendix III B, using solutions of the substance being examined containing (1) 0.10% w/v in solution A, (2) 5.0% w/v in *chloroform* and (3) 5.0% w/v in solution A.

The chromatographic procedure may be carried out using a glass column (1.5 m × 4 mm) packed with *acid-washed, silanised diatomaceous support* (80 to 100 mesh) coated with 3% w/w of phenyl methyl silicone fluid (50% phenyl) (OV-17 is suitable) and maintained at 145°.

In the chromatogram obtained with solution (3) the ratio of the sum of the areas of any *secondary peaks* to the area of the peak due to the internal standard is not greater than the ratio of the area of the peak due to dimethyl phthalate to the area of the peak due to the internal standard in the chromatogram obtained with solution (1).

Sulfated ash

Not more than 0.1% w/w, Appendix IX A.

Water

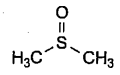
Not more than 0.1% w/w, Appendix IX C. Use 20 g.

ASSAY

In a borosilicate glass flask dissolve 1.5 g of the substance being examined in 5 mL of carbon dioxide-free ethanol prepared by boiling *ethanol (96%)* thoroughly and neutralising to *phenolphthalein solution R1*. Neutralise the free acid in the solution with *0.1M ethanolic potassium hydroxide VS* using 0.2 mL of *phenolphthalein solution R1* as indicator. Add 50 mL of *0.5M ethanolic potassium hydroxide VS* and boil under a reflux condenser on a water bath for 1 hour. Add 20 mL of *water* and titrate the excess of alkali with *0.5M hydrochloric acid VS* using a further 0.2 mL of *phenolphthalein solution R1* as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the alkali required to saponify the esters. Each mL of *0.5M ethanolic potassium hydroxide VS* is equivalent to 48.55 mg of $C_{10}H_{10}O_4$.

Dimethyl Sulfoxide

(Ph. Eur. monograph 0763)



C_2H_6OS 78.1 *67-68-5*

Action and use

Pharmaceutical solvent; excipient.

When dimethyl sulphoxide is demanded, Dimethyl Sulfoxide shall be supplied.

Ph Eur

DEFINITION

Sulfinylbismethane.

CHARACTERS

Appearance

Colourless liquid or colourless crystals, hygroscopic.

Solubility

Miscible with water and with ethanol (96 per cent).

IDENTIFICATION

First identification: C.

Second identification: A, B, D.

A. Relative density (see Tests).

B. Refractive index (see Tests).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dimethyl sulfoxide CRS.*

D. Dissolve 50 mg of nickel chloride R in 5 mL of the substance to be examined. The solution is greenish-yellow. Heat in a water-bath at 50 °C. The colour changes to green or bluish-green. Cool. The colour changes to greenish-yellow.

TESTS

Acidity

Dissolve 50.0 g in 100 mL of *carbon dioxide-free water R*. Add 0.1 mL of *phenolphthalein solution R1*. Not more than 5.0 mL of *0.01 M sodium hydroxide* is required to produce a pink colour.

Relative density (*2.2.5*)

1.100 to 1.104.

Refractive index (*2.2.6*)

1.478 to 1.480.

Freezing point (*2.2.18*)

Minimum 18.3 °C.

Absorbance (*2.2.25*)

Purge with *nitrogen R* for 15 min. The absorbance, measured using *water R* as the compensation liquid, is not more than 0.30 at 275 nm and not more than 0.20 at both 285 nm and 295 nm. Examined between 270 nm and 350 nm, the substance to be examined shows no absorption maximum.

Related substances

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.125 g of *bibenzyl R* in *acetone R* and dilute to 50 mL with the same solvent.

Test solution (a) Dissolve 5.0 g of the substance to be examined in *acetone R* and dilute to 10.0 mL with the same solvent.

Test solution (b) Dissolve 5.0 g of the substance to be examined in *acetone R*, add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *acetone R*.

Reference solution Dissolve 50.0 mg of the substance to be examined and 50 mg of *dimethyl sulfone R* in *acetone R*, add 10.0 mL of the internal standard solution and dilute to 100.0 mL with *acetone R*.

Column:
— *material*: glass;
— *size*: *l* = 1.5 m, Ø = 4 mm;
— *stationary phase*: diatomaceous earth for gas chromatography R (125-180 µm) impregnated with 10 per cent *m/m* of polyethyleneglycol adipate R.

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Temperature:
— *column*: 165 °C;
— *injection port and detector*: 190 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 4 times the retention time of dimethyl sulfoxide.

Elution order Dimethyl sulfoxide, dimethyl sulfone, bibenzyl.

Retention time Dimethyl sulfoxide = about 5 min.

System suitability:
— *resolution*: minimum 3 between the peaks due to dimethyl sulfoxide and dimethyl sulfone in the chromatogram obtained with the reference solution;
— in the chromatogram obtained with test solution (a) there is no peak with the same retention time as the internal standard.

Limit:
— *total*: calculate the ratio *R* of the area of the peak due to dimethyl sulfoxide to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with test solution (b), calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard to the area of the peak due to the internal standard: this ratio is not greater than *R* (0.1 per cent).

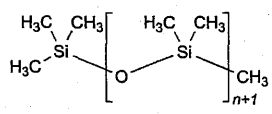
Water (*2.5.12*)
Maximum 0.2 per cent, determined on 10.0 g.

STORAGE
In an airtight, glass container, protected from light.

Ph Eur

Dimeticone

(Ph. Eur. monograph 0138)



9006-65-9

Action and use
Antifoaming agent; water repellent.

When dimethicone is demanded, Dimeticone shall be supplied.

Ph Eur

DEFINITION
α-Trimethylsilyl-ω-methylpoly[oxy(dimethylsilanediyl)].

This poly(dimethylsiloxane) is obtained by hydrolysis and polycondensation of dichlorodimethylsilane and chlorotrimethylsilane. Different grades of dimeticone exist which are distinguished by a number indicating the nominal kinematic viscosity placed after the name.

Their degree of polymerisation (n = 20 to 400) is such that their kinematic viscosities are nominally between 20 $mm^2 \cdot s^{-1}$ and 1300 $mm^2 \cdot s^{-1}$.

Dimeticones with a nominal viscosity of 50 $mm^2 \cdot s^{-1}$ or lower are intended for external use only.

CHARACTERS

Appearance
Clear, colourless liquid of various viscosities.

Solubility
Practically insoluble in water, very slightly soluble or practically insoluble in anhydrous ethanol, miscible with ethyl acetate, with methyl ethyl ketone and with toluene.

IDENTIFICATION
A. It is identified by its kinematic viscosity at 25 °C (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dimeticone CRS.*

The region of the spectrum from 850 cm^{-1} to 750 cm^{-1} is not taken into account.

C. Heat 0.5 g in a test-tube over a small flame until white fumes begin to appear. Invert the tube over a 2nd tube containing 1 mL of a 1 g/L solution of *chromotropic acid, sodium salt R* in *sulfuric acid R* so that the fumes reach the solution. Shake the 2nd tube for about 10 s and heat on a water-bath for 5 min. The solution is violet.

D. In a platinum crucible, prepare the sulfated ash (*2.4.14*) using 50 mg. The residue is a white powder that gives the reaction of silicates (*2.3.1*).

TESTS

Acidity
To 2.0 g add 25 mL of a mixture of equal volumes of *anhydrous ethanol R* and *ether R*, previously neutralised to 0.2 mL of *bromothymol blue solution R1*, and shake. Not more than 0.15 mL of *0.01 M sodium hydroxide* is required to change the colour of the solution to blue.

Viscosity (*2.2.9*)
90 per cent to 110 per cent of the nominal kinematic viscosity stated on the label, determined at 25 °C.

Mineral oils
Place 2 g in a test-tube and examine in ultraviolet light at 365 nm. The fluorescence is not more intense than that of a solution containing 0.1 ppm of *quinine sulfate R* in *0.005 M sulfuric acid* examined in the same conditions.

Phenylated compounds
Dissolve 5.0 g with shaking in 10 mL of *cyclohexane R*. At wavelengths from 250 nm to 270 nm, the absorbance (*2.2.25*) of the solution is not greater than 0.2.

Volatile matter
Maximum 0.3 per cent, for dimeticones with a nominal viscosity greater than 50 $mm^2 \cdot s^{-1}$, determined on 1.00 g by heating in an oven at 150 °C for 2 h. Carry out the test using a dish 60 mm in diameter and 10 mm deep.

LABELLING
The label states:
— the nominal kinematic viscosity by a number placed after the name of the product;

— where applicable, that the product is intended for external use.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

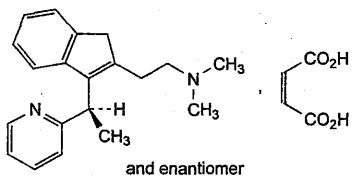
The following characteristic may be relevant for dimeticone used as emollient.

Viscosity
(see Tests).

Ph Eur

Dimetindene Maleate

(Ph. Eur. monograph 1417)



$C_{24}H_{28}N_2O_4$ 408.5 *3614-69-5*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION

N,*N*-Dimethyl-2-[3-[(*RS*)-1-(pyridin-2-yl)ethyl]-1*H*-inden-2-yl]ethanamine (*Z*)-butenedioate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, soluble in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *dimetindene maleate CRS.*

TESTS

Solution S
Dissolve 0.20 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than Y_6 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Gas chromatography (*2.2.28*).

Solvent mixture *acetone R, methylene chloride R* (50:50 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (a) Dilute 1 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of *2-ethylpyridine R* (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 10.0 mL of this solution to 100.0 mL with the solvent mixture.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.32 mm;
— *stationary phase*: *polymethylphenylsiloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R.*

Linear velocity About 30 cm/s.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 1	60
	1 - 34.3	60 → 260
	34.3 - 46.3	260
Injection port		240
Detector		260

Detection Flame ionisation.

Injection 2 µL; inject via a split injector with a split flow of 30 mL/min.

Run time 1.3 times the retention time of dimetindene.

Elution order Impurity A and maleic acid appear during the first 8 min.

System suitability Reference solution (a):
— *symmetry factor*: maximum 1.3 for the principal peak.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *impurities B, C, D, E, F, G, H, I*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *sum of impurities other than A*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to maleic acid.

Loss on drying (*2.2.32*)
Maximum 0.1 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 80 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

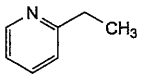
1 mL of *0.1 M perchloric acid* is equivalent to 20.43 mg of $C_{24}H_{28}N_2O_4$.

STORAGE

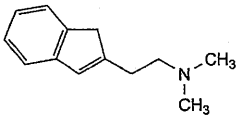
Protected from light.

IMPURITIES

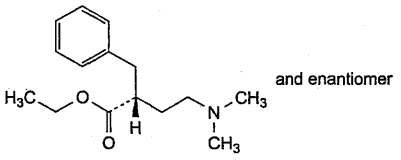
Specified impurities *A, B, C, D, E, F, G, H, I.*



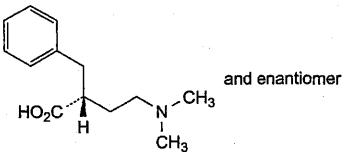
A. 2-ethylpyridine,



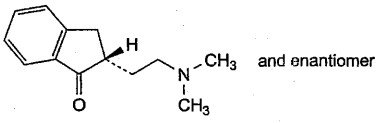
B. 2-(1*H*-inden-2-yl)-*N*,*N*-dimethylethanamine,



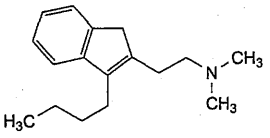
C. ethyl (2*RS*)-2-benzyl-4-(dimethylamino)butanoate,



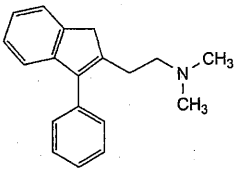
D. (2*RS*)-2-benzyl-4-(dimethylamino)butanoic acid,



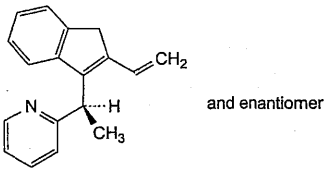
E. (2*RS*)-2-[2-(dimethylamino)ethyl]indan-1-one,



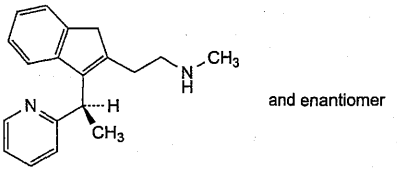
F. 2-(3-butyl-1*H*-inden-2-yl)-*N*,*N*-dimethylethanamine,



G. *N*,*N*-dimethyl-2-(3-phenyl-1*H*-inden-2-yl)ethanamine,



H. 2-[(1*RS*)-1-(2-ethenyl-1*H*-inden-3-yl)ethyl]pyridine,

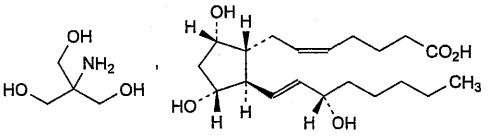


I. *N*-methyl-2-[3-[(1*RS*)-1-(pyridin-2-yl)ethyl]-1*H*-inden-2-yl]ethanamine.

Ph Eur

Dinoprost Trometamol

(Ph. Eur. monograph 1312)



$C_{24}H_{45}NO_8$ 475.6 *38562-01-5*

Action and use

Prostaglandin $F_{2\alpha}$($PGF_{2\alpha}$); inducer of uterine muscle contraction.

Ph Eur

DEFINITION

Trometamol (*Z*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]cyclopentyl]hept-5-enoate ($PGF_{2\alpha}$).

Content

96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in acetonitrile.

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 19 to + 26 (anhydrous substance).

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dinoprost trometamol CRS.*

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (23:77 *V/V*).

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a). Degradation of dinoprost trometamol to impurity B Dissolve 1 mg of the substance to be examined in 1 mL of the mobile phase and heat the solution on a water-bath at 85 °C for 5 min and cool.

Reference solution (b) Dilute 2.0 mL of the test solution to 20.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 20.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R1* (5 µm) with a pore size of 10 nm and a carbon loading of 19 per cent.

Mobile phase Dissolve 2.44 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1000 mL with *water R*; adjust to pH 2.5 with *phosphoric acid R* (about 0.6 mL); mix 770 mL of this solution with 230 mL of *acetonitrile R1*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

Run time 2.5 times the retention time of the principal peak (to elute degradation products formed during heating) for reference solution (a) and 10 min after the elution of dinoprost for the test solution and reference solution (b).

Retention time Impurity B = about 55 min; impurity A = about 60 min; dinoprost = about 66 min.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurities B and A and minimum 2.0 between the peaks due to impurity A and dinoprost; if necessary, adjust the composition of the mobile phase by increasing the concentration of acetonitrile to decrease the retention times;
— *symmetry factor*: maximum 1.2 for the peaks due to impurities A and B.

Limits:
— *impurity A*: not more than twice the area of the principal peak obtained with reference solution (b) (2 per cent);
— *impurities B, C, D*: for each impurity, not more than 1.5 times the area of the principal peak obtained with reference solution (b) (1.5 per cent) and not more than one such peak has an area greater than 0.5 times the area of the principal peak obtained with reference solution (b) (0.5 per cent);
— *sum of impurities other than A*: not more than twice the area of the principal peak obtained with reference solution (b) (2 per cent);
— *disregard limit*: 0.05 times the area of the principal peak obtained with reference solution (b) (0.05 per cent); disregard any peak due to trometamol (retention time = about 1.5 min).

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (23:77 *V/V*).

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution Dissolve 10.0 mg of *dinoprost trometamol CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R1* (5 µm) with a pore size of 10 nm and a carbon loading of 19 per cent.

Mobile phase Dissolve 2.44 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1000 mL with *water R*; adjust to pH 2.5 with *phosphoric acid R* (about 0.6 mL); mix 730 mL of this solution with 270 mL of *acetonitrile R1*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 µL.

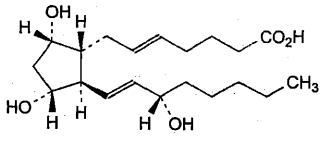
Retention time Dinoprost = about 23 min.

System suitability Reference solution:
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the peak due to dinoprost after 6 injections.

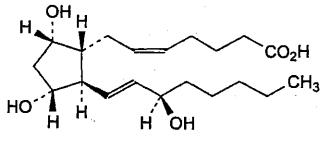
Calculate the percentage of dinoprost trometamol from the declared content of *dinoprost trometamol CRS*.

IMPURITIES

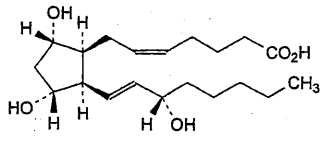
Specified impurities *A, B, C, D.*



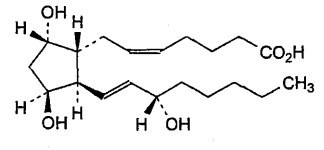
A. (*E*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]cyclopentyl]hept-5-enoic acid ((5*E*)-$PGF_{2\alpha}$; 5,6-*trans*-$PGF_{2\alpha}$),



B. (*Z*)-7-[(1*R*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(*E*)-(3*R*)-3-hydroxyoct-1-enyl]cyclopentyl]hept-5-enoic acid ((15*R*)-$PGF_{2\alpha}$;15-epi$PGF_{2\alpha}$),



C. (*Z*)-7-[(1*S*,2*R*,3*R*,5*S*)-3,5-dihydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]cyclopentyl]hept-5-enoic acid ((8*S*)-$PGF_{2\alpha}$; 8-epi$PGF_{2\alpha}$),

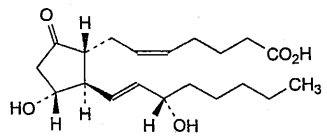


D. (*Z*)-7-[(1*R*,2*R*,3*S*,5*S*)-3,5-dihydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]cyclopentyl]hept-5-enoic acid (11β-$PGF_{2\alpha}$; 11-epi$PGF_{2\alpha}$).

Ph Eur

Dinoprostone

(Ph. Eur. monograph 1311)



$C_{20}H_{32}O_5$ 352.5 *363-24-6*

Action and use
Prostaglandin E_2(PGE_2); inducer of uterine muscle contraction.

Preparation
Dinoprostone Oral Solution

Ph Eur

DEFINITION
(*Z*)-7-[(1*R*,2*R*,3*R*)-3-Hydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (PGE_2).

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Practically insoluble in water, very soluble in methanol, freely soluble in alcohol.

The substance degrades at room temperature.

IDENTIFICATION
A. Specific optical rotation (*2.2.7*): −90 to −82 (anhydrous substance).

Immediately before use, dissolve 50.0 mg in *alcohol R* and dilute to 10.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dinoprostone CRS*.

TESTS
Prepare the solutions immediately before use.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 10.0 mg of the substance to be examined in a 58 per cent *V/V* solution of *methanol R2* and dilute to 2.0 mL with the same solvent.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in a 58 per cent *V/V* solution of *methanol R2* and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 1 mg of *dinoprostone CRS* and 1 mg of *dinoprostone impurity C CRS* in a 58 per cent *V/V* solution of *methanol R2* and dilute to 10.0 mL with the same solvent. Dilute 4.0 mL of the solution to 10.0 mL with a 58 per cent *V/V* solution of *methanol R2*.

Reference solution (b) Dilute 0.5 mL of test solution (a) to 10.0 mL with a 58 per cent *V/V* solution of *methanol R2*. Dilute 1.0 mL of the solution to 10.0 mL with a 58 per cent *V/V* solution of *methanol R2*.

Reference solution (c) In order to prepare *in situ* the degradation compounds (impurity D and impurity E), dissolve 1 mg of the substance to be examined in 100 µL of *1 M sodium hydroxide* (the solution becomes brownish-red), wait 4 min, add 150 µL of dilute acetic acid R1 (yellowish-white opalescent solution) and dilute to 5.0 mL with a 58 per cent *V/V* solution of *methanol R2*.

Reference solution (d) Dissolve 20 mg of *dinoprostone CRS* in a 58 per cent *V/V* solution of *methanol R2* and dilute to 20.0 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R*,
— *temperature*: 30 °C.

Mobile phase Mix 42 volumes of a 0.2 per cent *V/V* solution of *acetic acid R* and 58 volumes of *methanol R2*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL; inject test solution (a) and reference solutions (a), (b) and (c).

Relative retention With reference to dinoprostone (retention time = about 18 min): impurity C = about 1.2; impurity D = about 1.8; impurity E = about 2.0.

System suitability Reference solution (a):
— *resolution*: minimum of 3.8 between the peaks due to dinoprostone and to impurity C. If necessary adjust the concentration of the acetic acid solution and/or methanol (increase the concentration of the acetic acid solution to increase the retention time for dinoprostone and impurity C and increase the concentration of methanol to decrease the retention time for both compounds).

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.2; impurity E = 0.7,
— *impurity C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent),
— *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent),
— *impurity E*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *any other impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *total of other impurities*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent),
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

If any peak with a relative retention to dinoprostone of about 0.8 is greater than 0.5 per cent or if the total of other impurities is greater than 1.0 per cent, record the chromatogram of test solution (a) with a detector set at 230 nm. If the area of the peak at 230 nm is twice the area of the peak at 210 nm, multiply the area at 210 nm by 0.2 (correction factor for impurity F).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 0.50 g.

ASSAY

Prepare the solutions immediately before use.

Liquid chromatography (*2.2.29*) as described in the test for related substances.

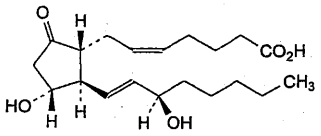
Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{20}H_{32}O_5$.

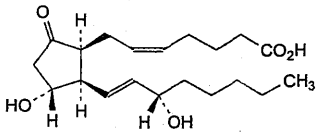
STORAGE

At a temperature not exceeding - 15 °C.

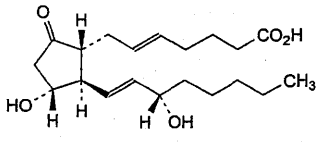
IMPURITIES



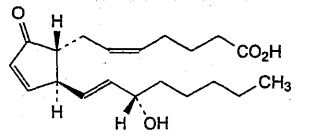
A. (*Z*)-7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(*E*)-(3*R*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (15-epiPGE$_2$; (15*R*)-PGE$_2$),



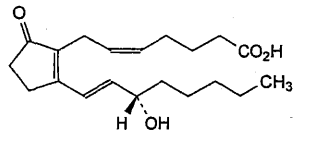
B. (*Z*)-7-[(1*S*,2*R*,3*R*)-3-hydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (8-epiPGE$_2$; (8S)-PGE$_2$),



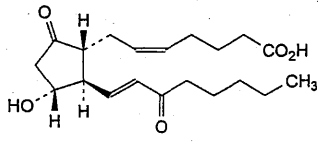
C. (*E*)-7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (5-*trans*-PGE$_2$; (5*E*)-PGE$_2$),



D. (*Z*)-7-[(1*R*,2*S*)-2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopent-3-enyl]hept-5-enoic acid (PGA$_2$),



E. (*Z*)-7-[2-[(*E*)-(3*S*)-3-hydroxyoct-1-enyl]-5-oxocyclopent-1-enyl]hept-5-enoic acid (PGB$_2$),

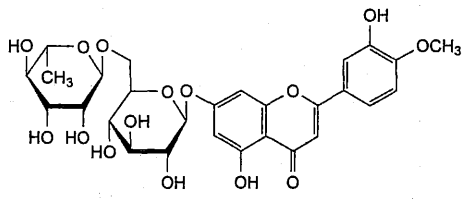


F. (*Z*)-7-[(1*R*,2*R*,3*R*)-3-hydroxy-2-[(*E*)-3-oxo-oct-1-enyl]-5-oxocyclopentyl]hept-5-enoic acid (15-oxo-PGE$_2$; 15-keto-PGE$_2$).

Ph Eur

Diosmin

(Ph. Eur. monograph 1611)



$C_{28}H_{32}O_{15}$ 609 *520-27-4*

Action and use

Chronic venous insufficiency (flavonoid).

Ph Eur

DEFINITION

7-[[6-*O*-(6-Deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-4*H*-1-benzopyran-4-one.

Substance obtained through iodine-assisted oxidation of (2*S*)-7-[[6-*O*-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-2,3-dihydro-4*H*-1-benzopyran-4-one (hesperidin) of natural origin.

Content 90.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Greyish-yellow or light yellow, hygroscopic powder.

Solubility

Practically insoluble in water, soluble in dimethyl sulfoxide, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diosmin CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Iodine

Maximum 0.1 per cent.

Determine the total content of iodine by potentiometry, using an iodide-selective electrode (*2.2.36*), after oxygen combustion (*2.5.10*).

Test solution Wrap 0.100 g of the substance to be examined in a piece of filter paper and place it in a sample carrier. Introduce into the flask 50 mL of a 0.2 g/L solution of *hydrazine R*. Flush the flask with oxygen for 10 min. Ignite the filter paper. Stir the contents of the flask immediately after the end of the combustion to dissolve completely the combustion products. Continue stirring for 1 h.

Reference solution Dilute 2.0 mL of a 16.6 g/L solution of *potassium iodide R* to 100.0 mL with *water R*. Dilute 10.0 mL of this solution to 100.0 mL with *water R*.

Introduce into a beaker 30 mL of a 200 g/L solution of *potassium nitrate R* in *0.1 M nitric acid*. Immerse the electrodes and stir for 10 min. The potential of the solution

(nT_1) must remain stable. Add 1 mL of the test solution and measure the potential (nT_2).

Introduce into a beaker 30 mL of a 200 g/L solution of *potassium nitrate R* in *0.1 M nitric acid*. Immerse the electrodes and stir for 10 min. The potential of the solution must remain stable (nR_1). Add 80 µL of the reference solution and measure the potential (nR_2).

The absolute value $|nT_2 - nT_1|$ is not higher than the absolute value $|nR_2 - nR_1|$.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in *dimethyl sulfoxide R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 25.0 mg of *diosmin CRS* in *dimethyl sulfoxide R* and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethyl sulfoxide R*.

Reference solution (c) Dissolve 5.0 mg of *diosmin for system suitability CRS* (containing impurities A, B, C, D, E and F) in *dimethyl sulfoxide R* and dilute to 5.0 mL with the same solvent.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 40 °C.

Mobile phase *acetonitrile R, glacial acetic acid R, methanol R, water R* (2:6:28:66 *V/V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 6 times the retention time of diosmin.

Identification of impurities Use the chromatogram supplied with *diosmin for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D, E and F.

Relative retention With reference to diosmin (retention time = about 4 min): impurity A = about 0.5; impurity B = about 0.6; impurity C = about 0.8; impurity D = about 2.2; impurity E = about 2.6; impurity F = about 4.5.

System suitability Reference solution (c):
- *resolution*: minimum 2.5 between the peaks due to impurities B and C.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.4; impurity F = 0.6;
- for each impurity, use the concentration of diosmin in reference solution (b).

Limits:
- *impurity B*: maximum 4.0 per cent;
- *impurities C, E*: for each impurity, maximum 3.0 per cent;
- *impurity F*: maximum 2.0 per cent;
- *impurity D*: maximum 0.6 per cent;
- *impurity A*: maximum 0.5 per cent;
- *unspecified impurities*: for each impurity, maximum 0.4 per cent;
- *total*: maximum 8.5 per cent;
- *reporting threshold*: 0.10 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Water (*2.5.12*)

Maximum 6.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{28}H_{32}O_{15}$ taking into account the assigned content of *diosmin CRS*.

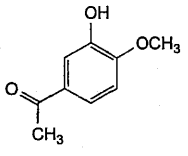
STORAGE

In an airtight container.

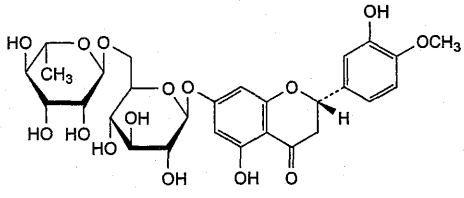
IMPURITIES

Specified impurities *A, B, C, D, E, F.*

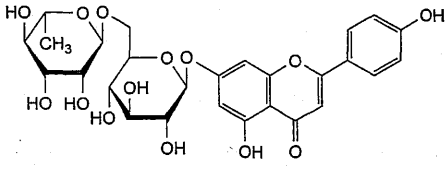
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G.*



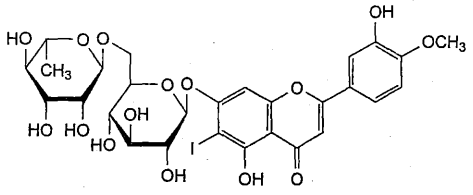
A. 1-(3-hydroxy-4-methoxyphenyl)ethan-1-one (acetoisovanillone),



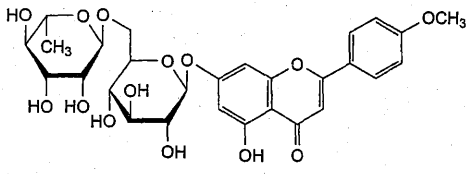
B. (2*S*)-7-[[6-*O*-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-2,3-dihydro-4*H*-1-benzopyran-4-one (hesperidin),



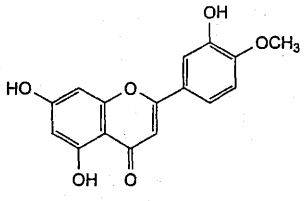
C. 7-[[6-*O*-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(4-hydroxyphenyl)-4*H*-1-benzopyran-4-one (isorhoifolin),



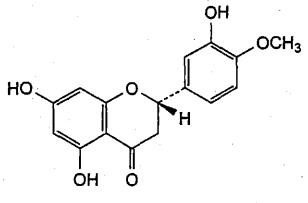
D. 7-[[6-*O*-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-6-iodo-4*H*-1-benzopyran-4-one (6-iododiosmin),



E. 7-[[6-*O*-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(4-methoxyphenyl)-4*H*-1-benzopyran-4-one (linarin),



F. 5,7-dihydroxy-2-(3-hydroxy-4-methoxyphenyl)-4*H*-1-benzopyran-4-one (diosmetin),

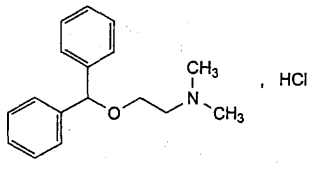


G. (2*S*)-5,7-dihydroxy-2-(3-hydroxy-4-methoxyphenyl)-2,3-dihydro-4*H*-1-benzopyran-4-one (hesperetin).

Ph Eur

Diphenhydramine Hydrochloride

(Ph. Eur. monograph 0023)



$C_{17}H_{22}ClNO$ 291.8 *147-24-0*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Preparations
Diphenhydramine Oral Solution
Diphenhydramine Tablets

Ph Eur

DEFINITION
2-(Diphenylmethoxy)-*N*,*N*-dimethylethanamine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: C, D.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 168 °C to 172 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent.

Spectral range 230-350 nm.

Absorption maxima At 253 nm, 258 nm and 264 nm.

Absorbance ratios:
— A_{258}/A_{253} = 1.1 to 1.3;
— A_{258}/A_{264} = 1.2 to 1.4.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diphenhydramine hydrochloride CRS.*

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S and a fivefold dilution of solution S are clear (*2.2.1*). Solution S is not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.15 mL of *methyl red solution R* and 0.25 mL of *0.01 M hydrochloric acid.* The solution is pink. Not more than 0.5 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to yellow.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 70 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 2.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *diphenhydramine impurity A CRS* and 5 mg of *diphenylmethanol R* in the mobile phase and dilute to 10.0 mL with the mobile phase. To 2.0 mL of this solution add 1.5 mL of the test solution and dilute to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 5.4 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.0 using *phosphoric acid R*.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 7 times the retention time of diphenhydramine.

Relative retention With reference to diphenhydramine (retention time = about 6 min): impurity A = about 0.9; impurity B = about 1.5; impurity C = about 1.8; impurity D = about 2.6; impurity E = about 5.1.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to diphenhydramine and to impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 0.7,
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *any other impurity*: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent),
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

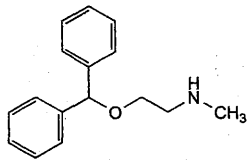
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

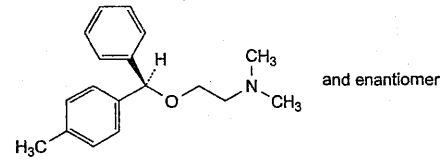
1 mL of *0.1 M sodium hydroxide* is equivalent to 29.18 mg of $C_{17}H_{22}ClNO$.

STORAGE
Protected from light.

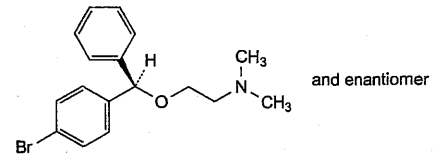
IMPURITIES
Specified impurities *A, B, C, D, E.*



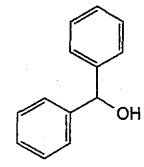
A. 2-(diphenylmethoxy)-*N*-methylethanamine,



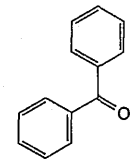
B. 2-[(*RS*)-(4-methylphenyl)phenylmethoxy]-*N*,*N*-dimethylethanamine,



C. 2-[(*RS*)-(4-bromophenyl)phenylmethoxy]-*N*,*N*-dimethylethanamine,



D. diphenylmethanol (benzhydrol),

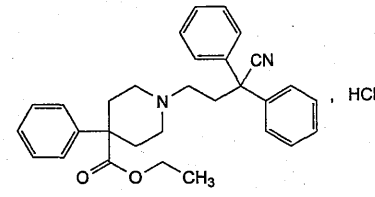


E. diphenylmethanone (benzophenone).

Ph Eur

Diphenoxylate Hydrochloride

(Ph. Eur. monograph 0819)



$C_{30}H_{33}ClN_2O_2$ 489.1 *3810-80-8*

Action and use
Opioid receptor agonist; treatment of diarrhoea.

Ph Eur

DEFINITION
Ethyl 1-(3-cyano-3,3-diphenylpropyl)-4-phenylpiperidine-4-carboxylate hydrochloride.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, freely soluble in methylene chloride, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *diphenoxylate hydrochloride CRS.*

B. Dissolve about 30 mg in 5 mL of *methanol R.* Add 0.25 mL of *nitric acid R* and 0.4 mL of *silver nitrate solution R1.* Shake and allow to stand. A curdled precipitate is formed. Centrifuge and rinse the precipitate with 3 quantities, each of 2 mL, of *methanol R.* Carry out this operation rapidly and protected from bright light. Suspend the precipitate in 2 mL of *water R* and add 1.5 mL of *ammonia R.* The precipitate dissolves easily.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 1.0 g in *methylene chloride R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solution A Adjust 900 mL of *water R* to pH 2.3 with *phosphoric acid R* and dilute to 1000.0 mL with *water R.*

Solvent mixture *acetonitrile R1*, solution A (50:50 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in 20 mL of the solvent mixture, sonicate for 2 min, cool and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of *diphenoxylate for system suitability CRS* (containing impurity A) in 2.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: solution A;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	75	25
5 - 40	75 → 15	25 → 85

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Relative retention With reference to diphenoxylate (retention time = about 16 min): impurity A = about 0.8.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity A and diphenoxylate.

Limits:
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 40 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid.* Carry out a potentiometric titration (*2.2.20*), using *0.1 M ethanolic sodium hydroxide.* Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M ethanolic sodium hydroxide* is equivalent to 48.91 mg of $C_{30}H_{33}ClN_2O_2$.

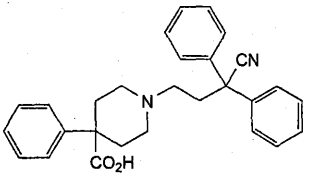
STORAGE

Protected from light.

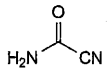
IMPURITIES

Specified impurities *A.*

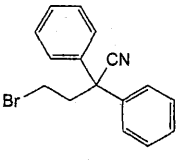
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C.*



A. 1-(3-cyano-3,3-diphenylpropyl)-4-phenylpiperidine-4-carboxylic acid (diphenoxylic acid),



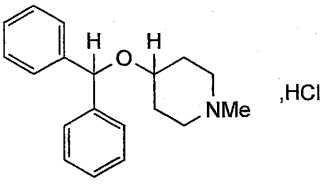
B. 1-cyanomethanamide,



C. 4-bromo-2,2-diphenylbutanenitrile.

Ph Eur

Diphenylpyraline Hydrochloride



$C_{19}H_{23}NO,HCl$ 317.9 *132-18-3*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

DEFINITION
Diphenylpyraline Hydrochloride is 4-benzhydryloxy-1-methylpiperidine hydrochloride. It contains not less than 98.0% and not more than 101.0% of $C_{19}H_{23}NO,HCl$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white powder; odourless or almost odourless.

Freely soluble in *water* and in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of diphenylpyraline hydrochloride *(RS 106)*.

B. Yields the reactions characteristic of *chlorides*, Appendix VI.

TESTS
Related substances
Carry out the method for *gas chromatography*, Appendix III B, using the following solutions.

(1) Dissolve 45 mg of *bibenzyl* (internal standard) in sufficient *dichloromethane* to produce 100 mL.

(2) Dissolve 0.20 g of the substance being examined in 20 mL of *water*, make the solution alkaline with 5M *ammonia* and extract with three 25-mL quantities of *dichloromethane*. Shake the combined extracts with 10 g of *anhydrous sodium sulfate*, filter, evaporate the filtrate to dryness at about 30° and dissolve the residue in 2 mL of *dichloromethane*.

(3) Prepare solution (3) in the same manner as solution (2) but dissolve the residue in 2 mL of solution (1).

CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with *silanised diatomaceous support* (80 to 100 mesh) coated with 3% w/w of phenyl methyl silicone fluid (50% phenyl) (OV-17 is suitable).

(b) Use *nitrogen* as the carrier gas.

(c) Use an oven temperature of 165°. Allow the chromatography to proceed for 3 times the retention time of bibenzyl.

(d) Increase the oven temperature to 240° to elute the diphenylpyraline from the column.

LIMITS

In the chromatogram obtained with solution (3):

the sum of the areas of any *secondary peaks* is not greater than the area of the peak due to the internal standard.

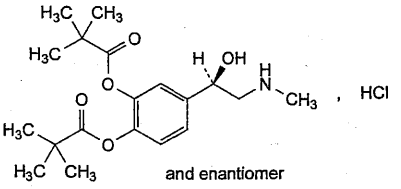
Loss on drying
When dried to constant weight at 105°, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.2 g, adding 5 mL of *mercury(II) acetate solution* and determining the end-point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 31.79 mg of $C_{19}H_{23}NO,HCl$.

Dipivefrine Hydrochloride

(Ph. Eur. monograph 1719)



$C_{19}H_{30}ClNO_5$ 387.9 *64019-93-8*

Action and use
Adrenaline prodrug; treatment of glaucoma.

Preparation
Dipivefrine Eye Drops

Ph Eur

DEFINITION
Hydrochloride of 4-[(1*RS*)-1-hydroxy-2-(methylamino)ethyl]-1,2-phenylene bis(2,2-dimethylpropanoate).

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, very soluble in methanol, freely soluble in ethanol (96 per cent) and in methylene chloride.

mp
About 160 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *dipivefrine hydrochloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Impurities A and B
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 10.0 mL with the same acid.

Reference solution Dissolve 10.0 mg of *adrenaline R* and 10.0 mg of *adrenalone hydrochloride R* in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 1.0 mL of this solution to 10.0 mL with *0.01 M hydrochloric acid*. *Protect this solution from light.*

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm).

Mobile phase:
— *mobile phase A*: 0.1 per cent *V/V* solution of *anhydrous formic acid R*;
— *mobile phase B*: *methanol R2, acetonitrile R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 5	100 → 40	0 → 60
5 - 10	40	60

Flow rate 1 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Retention times Impurity A = about 2.2 min; impurity B = about 3.2 min.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to impurities A and B.

Limits:
— *impurities A, B*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mix 40 volumes of *methanol R2* and 60 volumes of *acetonitrile R*. Mix 55 volumes of this mixture and 45 volumes of *0.01 M hydrochloric acid*.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *dipivefrine for system suitability CRS* (containing impurities C, D and E) in the solvent mixture and dilute to 2.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of *dipivefrine hydrochloride CRS* in the solvent mixture and dilute to 2.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 25.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm).

Mobile phase Mix 45 volumes of a 2.7 g/L solution of *concentrated ammonia R* adjusted to pH 10.0 with *dilute acetic acid R* and 55 volumes of a mixture of 40 volumes of *methanol R2* and 60 volumes of *acetonitrile R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Run time 2.5 times the retention time of dipivefrine.

Relative retention With reference to dipivefrine (retention time = about 7 min): impurities C and D = about 0.4; impurity E = about 1.3; impurity F = about 2.0.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to dipivefrine and impurity E.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurities C and D = 0.5; impurity E = 0.06;
— *sum of impurities C and D*: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities E, F*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak with a mass distribution ratio less than 0.5.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 6 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

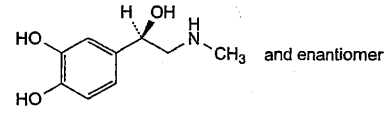
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 20 µL of reference solutions (a) and (c).

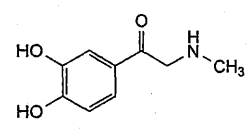
System suitability Reference solution (c):
— *symmetry factor*: maximum 2.0 for the peak due to dipivefrine.

Calculate the percentage content of $C_{19}H_{30}ClNO_5$ using the chromatograms obtained with reference solutions (a) and (c) and the declared content of *dipivefrine hydrochloride CRS*.

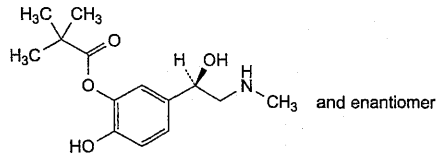
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



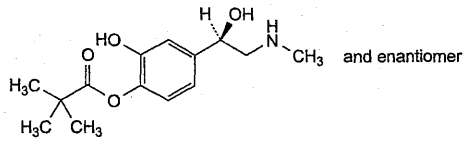
A. 4-[(1*RS*)-1-hydroxy-2-(methylamino)ethyl]benzene-1,2-diol ((±)-adrenaline),



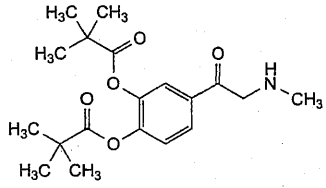
B. 1-(3,4-dihydroxyphenyl)-2-(methylamino)ethanone (adrenalone),



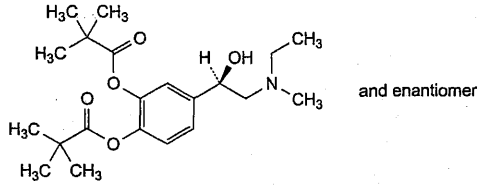
C. 2-hydroxy-5-[(1*RS*)-1-hydroxy-2-(methylamino)ethyl] phenyl 2,2-dimethylpropanoate,



D. 2-hydroxy-4-[(1*RS*)-1-hydroxy-2-(methylamino)ethyl] phenyl 2,2-dimethylpropanoate,



E. 4-[(methylamino)acetyl]-1,2-phenylene bis(2,2-dimethylpropanoate) (adrenalone dipivalate ester),



F. 4-[(1*RS*)-2-(ethylmethylamino)-1-hydroxyethyl]-1,2-phenylene bis(2,2-dimethylpropanoate).

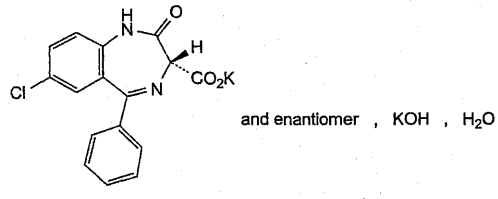
Ph Eur

Dipotassium Clorazepate Monohydrate

Dipotassium Clorazepate

Potassium Clorazepate

(Ph. Eur. monograph 0898)



$C_{16}H_{11}ClK_2N_2O_4,H_2O$ 426.9

Action and use

Hypnotic; anxiolytic.

Ph Eur

DEFINITION

Potassium (3*RS*)-7-chloro-2-oxo-5-phenyl-2,3-dihydro-1*H*-1,4-benzodiazepine-3-carboxylate compound with potassium hydroxide (1:1) monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or light yellow, crystalline powder, hygroscopic.

Solubility

Freely soluble to very soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

Solutions in water and in ethanol (96 per cent) are unstable and are to be used immediately.

IDENTIFICATION

First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution (a) Dissolve 10.0 mg in a 0.3 g/L solution of *potassium carbonate R* and dilute to 100.0 mL with the same solution.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with a 0.3 g/L solution of *potassium carbonate R.*

Spectral range 280-350 nm for test solution (a); 220-280 nm for test solution (b).

Absorption maxima About 315 nm (broad) for test solution (a); 230 nm for test solution (b).

Specific absorbance at the absorption maxima:

— 230 nm: 800 to 870;

— 315 nm: 49 to 56.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison Ph. Eur. reference spectrum of dipotassium clorazepate.

C. Dissolve about 20 mg in 2 mL of *sulfuric acid R.* Observed in ultraviolet light at 365 nm, the solution shows yellow fluorescence.

D. Dissolve 0.5 g in 5 mL of *water R.* Add 0.1 mL of *thymol blue solution R.* The solution is violet-blue.

E. Place 1.0 g in a crucible and add 2 mL of *dilute sulfuric acid R.* Heat at first on a water-bath, then ignite until all black particles have disappeared. Allow to cool. Take up the residue with *water R* and dilute to 20 mL with the same solvent. The solution gives reaction (b) of potassium (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_5 (*2.2.2, Method II*).

Rapidly dissolve 2.0 g with shaking in *water R* and dilute to 20.0 mL with the same solvent. Observe immediately.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 11.7 g of *dipotassium hydrogen phosphate R* in about 900 mL of *water R,* adjust to pH 7.0 with *phosphoric acid R* and dilute to 1000 mL with *water R.* Mix equal volumes of this solution and *methanol R.*

Solution B *methanol R,* 20 g/L solution of *potassium carbonate R* (30:70 *V/V*).

Test solution (a) Dissolve 10.0 mg of the substance to be examined in solution A and dilute to 20.0 mL with solution A.

Test solution (b) Dissolve 10.0 mg of the substance to be examined in solution B cooled to 0 °C and dilute to 20.0 mL with solution B.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 20.0 mL with solution A.

Reference solution (b) Dissolve 15 mg of *aminochlorobenzophenone R* (impurity A) and 15 mg of *ethyl clorazepate R* (impurity C) in *methanol R* and dilute to 20 mL with the same solvent. Dilute 1 mL of the solution to 20 mL with *methanol R*.

Reference solution (c) Dissolve 15 mg of *nordazepam R* (impurity B) in *methanol R* and dilute to 20 mL with the same solvent. Dilute 1 mL of the solution to 20 mL with solution B.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 1.17 g of *dipotassium hydrogen phosphate R* in about 580 mL of *water for chromatography R*, adjust to pH 8.0 with *phosphoric acid R*, dilute to 650 mL with *water for chromatography R*, and add 450 mL of *methanol R1*;
— *mobile phase B*: *methanol R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	80	20
5 - 20	80 → 50	20 → 50
20 - 25	50	50

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 230 nm.

Autosampler Set at 4 °C.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B.

Relative retention With reference to clorazepate (retention time = about 3 min): impurity B = about 4.6; impurity C = about 5.9; impurity A = about 6.5.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to impurities C and A.

Calculation of percentage contents:
— for each impurity, use the concentration of dipotassium clorazepate monohydrate in reference solution (a).

Limits:
— *impurity C*: maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent; disregard the peak due to impurity B.

Impurity B

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solutions (a) and (c).

Calculation of percentage content:
— *correction factor*: multiply the peak area of impurity B by 0.6;
— use the concentration of dipotassium clorazepate monohydrate in reference solution (a).

Limit:
— *impurity B*: maximum 0.2 per cent.

Water (*2.5.12*)

3.5 per cent to 5.5 per cent, determined on 0.250 g.

ASSAY

Dissolve 0.130 g in 10 mL of *anhydrous acetic acid R*. Add 30 mL of *methylene chloride R*. Titrate with *0.1 M perchloric acid*, determining the 2 points of inflexion by potentiometry (*2.2.20*).

At the 2nd point of inflexion, 1 mL of *0.1 M perchloric acid* is equivalent to 13.63 mg of $C_{16}H_{11}ClK_2N_2O_4$.

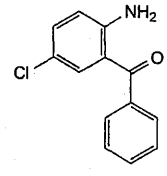
STORAGE

In an airtight container, protected from light.

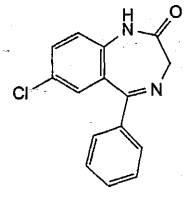
IMPURITIES

Specified impurities *B, C.*

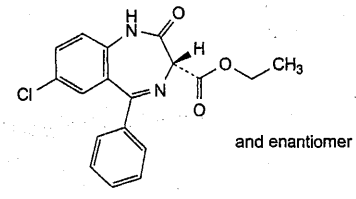
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A.*



A. (2-amino-5-chlorophenyl)(phenyl)methanone (aminochlorobenzophenone),



B. 7-chloro-5-phenyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one (nordazepam),



C. ethyl (3*RS*)-7-chloro-2-oxo-5-phenyl-2,3-dihydro-1*H*-1,4-benzodiazepine-3-carboxylate (ethyl clorazepate).

Ph Eur

Dipotassium Hydrogen Phosphate

(Dipotassium Phosphate, Ph. Eur. monograph 1003)

K_2HPO_4 174.2 *7758-11-4*

Action and use
Excipient.

Preparation
Dipotassium Hydrogen Phosphate Injection

Ph Eur

DEFINITION

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder or colourless crystals, very hygroscopic.

Solubility
Very soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is slightly alkaline (*2.2.4*).

B. Solution S gives reaction (b) of phosphates (*2.3.1*).

C. Solution S gives reaction (a) of potassium (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Reducing substances
To 5 mL of solution S add 5 mL of *dilute sulfuric acid R* and 0.25 mL of *0.02 M potassium permanganate* and heat on a water-bath for 5 min. The solution remains faintly pink.

Monopotassium phosphate
Maximum 2.5 per cent.

From the volume of *1 M hydrochloric acid* (10.0 mL) and of *1 M sodium hydroxide* (n_1 mL and n_2 mL) used in the assay, calculate the following ratio:

$$\frac{n_2 - 10}{10 - n_1}$$

This ratio is not greater than 0.025.

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 2.5 mL of solution S add 10 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

To 1.5 mL of solution S add 2 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 2 ppm, determined on 5 mL of solution S.

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S.

Sodium
Maximum 0.1 per cent, if intended for use in the manufacture of parenteral preparations.

Atomic emission spectrometry (*2.2.22, Method I*).

Test solution Dissolve 1.00 g in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using *sodium standard solution (200 ppm Na) R*, diluted as necessary with *water R*.

Wavelength 589 nm.

Loss on drying (*2.2.32*)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 125-130 °C.

Bacterial endotoxins (*2.6.14*)
Less than 1.1 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Dissolve 0.800 g (*m*) in 40 mL of *carbon dioxide-free water R* and add 10.0 mL of *1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*) using *1 M sodium hydroxide*. Read the volume added at the 1st inflexion point (n_1 mL). Continue the titration to the 2nd inflexion point (total volume of *1 M sodium hydroxide* required, n_2 mL).

Calculate the percentage content of K_2HPO_4 from the following expression:

$$\frac{1742(10 - n_1)}{m(100 - d)}$$

d = percentage loss on drying.

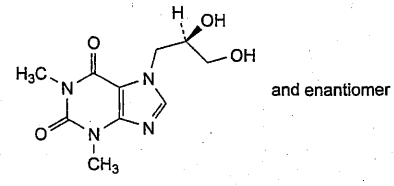
STORAGE
In an airtight container.

LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Diprophylline

(Ph. Eur. monograph 0486)



and enantiomer

$C_{10}H_{14}N_4O_4$ 254.2 *479-18-5*

Action and use
Non-selective phosphodiesterase inhibitor (xanthine); treatment of reversible airways obstruction.

Ph Eur

DEFINITION
7-[(2*RS*)-2,3-Dihydroxypropyl]-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison: *diprophylline CRS*.

TESTS
Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.25 mL of *bromothymol blue solution R1*. The solution is yellow or green. Not more than 0.4 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to blue.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 50 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.
Reference solution (b) Dissolve 5 mg of *etofylline CRS* (impurity C) in *water R* and dilute to 50.0 mL with the same solvent. Dilute 0.5 mL of the solution to 20.0 mL with the test solution.
Column:
— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with polar incorporated groups R* (3 µm);
— *temperature*: 30 °C.
Mobile phase *methanol R, water R* (10:90 *V/V*).
Flow rate 0.7 mL/min.
Detection Spectrophotometer at 272 nm.
Injection 10 µL.
Run time 3 times the retention time of diprophylline.
Relative retention With reference to diprophylline (retention time = about 18 min): impurity C = about 1.1.
System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to diprophylline.
Limits:
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 400 ppm.
Dilute 2.5 mL of solution S to 15 mL with *water R*.

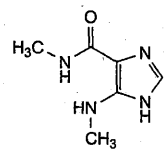
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

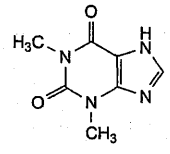
ASSAY
In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.
Dissolve 0.200 g in 3.0 mL of *anhydrous formic acid R* and add 50.0 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M perchloric acid* is equivalent to 25.42 mg of $C_{10}H_{14}N_4O_4$.

STORAGE
Protected from light.

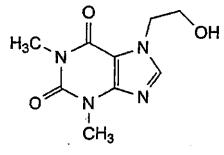
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D.*



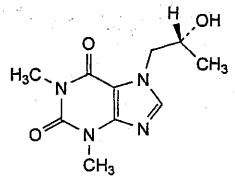
A. *N*-methyl-5-(methylamino)-1*H*-imidazole-4-carboxamide (theophyllidine),



B. 1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (theophylline),



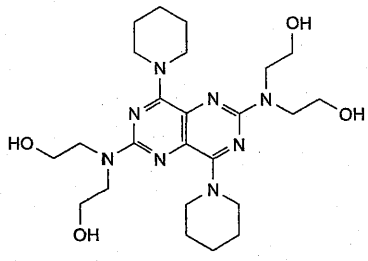
C. 7-(2-hydroxyethyl)-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (etofylline),



D. 7-[(2*RS*)-2-hydroxypropyl]-1,3-dimethyl-3,7-dihydro-1*H*-purine-2,6-dione (proxyphylline).

Dipyridamole

(Ph. Eur. monograph 1199)



$C_{24}H_{40}N_8O_4$ 504.6 *58-32-2*

Action and use
Adenosine reuptake inhibitor; inhibitor of platelet aggregation.

Preparations
Dipyridamole Prolonged-release Capsules
Dipyridamole Infusion
Dipyridamole Oral Suspension
Dipyridamole Tablets

Ph Eur

DEFINITION
2,2′,2″,2‴-[[4,8-Di(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidine-2,6-diyl]dinitrilo]tetraethanol.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
Bright yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in anhydrous ethanol. It dissolves in dilute mineral acids.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Preparation Discs of *potassium bromide R*.
Comparison *dipyridamole CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*
Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 50 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.
Reference solution (b) Dissolve the contents of a vial of *dipyridamole for peak identification CRS* (containing impurities A, B, C, D, E and F) in 1 mL of *methanol R*.
Column:
— *size*: *l* = 0.10 m, Ø = 4.0 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 45 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.0 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 7.0 with *0.5 M sodium hydroxide* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	40	60
5 - 19	40 → 5	60 → 95
19 - 24	5 → 40	95 → 60
24 - 29	40	60

Flow rate 1.2 mL/min.
Detection Spectrophotometer at 295 nm.
Injection 5 µL.
Identification of impurities Use the chromatogram supplied with *dipyridamole for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E and F.
Relative retention With reference to dipyridamole (retention time = about 8 min): impurity B = about 0.2; impurity F = about 0.3; impurity D = about 0.9; impurity E = about 1.3; impurity C = about 1.6; impurity A = about 2.2.
System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity D and dipyridamole;
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity B.
Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 1.7;
— *impurities A, B, C*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurities D, E*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 200 ppm.
To 0.250 g add 10 mL of *water R* and shake vigorously. Filter, rinse the filter with 5 mL of *water R* and dilute to 15 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 70 mL of *methanol R.* Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 50.46 mg of $C_{24}H_{40}N_8O_4$.

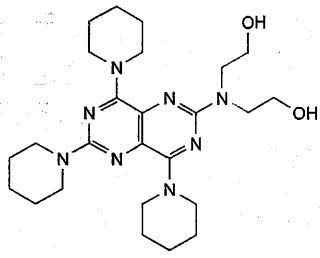
STORAGE

Protected from light.

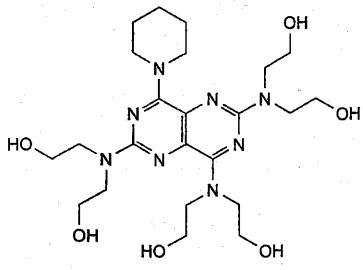
IMPURITIES

Specified impurities *A, B, C, D, E.*

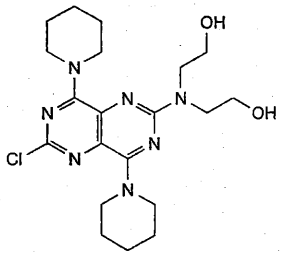
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G.*



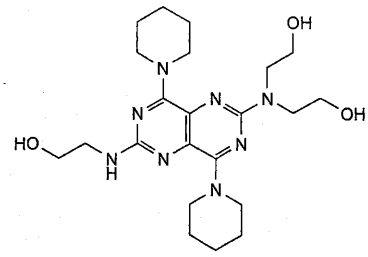
A. 2,2′-[[4,6,8-tri(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidin-2-yl]nitrilo]diethanol,



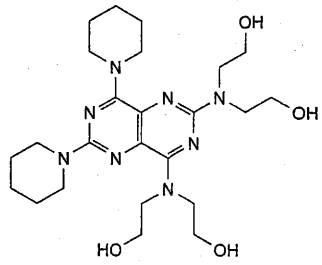
B. 2,2′,2″,2‴,2⁗,2⁗′[[8-(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidine-2,4,6-triyl]trinitrilo]hexaethanol,



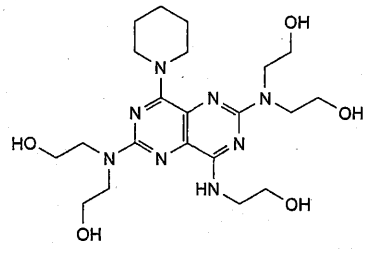
C. 2,2′-[[6-chloro-4,8-di(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidin-2-yl]nitrilo]diethanol,



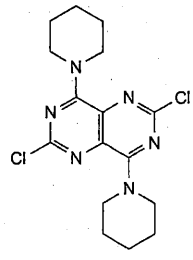
D. 2,2′-[[6-[(2-hydroxyethyl)amino]-4,8-di(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidin-2-yl]nitrilo]diethanol,



E. 2,2′,2″,2‴,2⁗-[[6,8-di(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidine-2,4-diyl]dinitrilo]tetraethanol,



F. 2,2′,2″,2‴,2⁗-[[4-[(2-hydroxyethyl)amino]-8-(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidine-2,6-diyl]dinitrilo]tetraethanol,

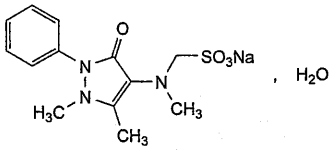


G. 2,6-dichloro-4,8-di(piperidin-1-yl)pyrimido[5,4-*d*]pyrimidine.

Ph Eur

Dipyrone

(Metamizole Sodium Monohydrate, Ph. Eur. monograph 1346)



$C_{13}H_{16}N_3NaO_4S,H_2O$ 351.4 *5907-38-0*

Action and use
Analgesic.

Ph Eur

DEFINITION

Sodium [(1,5-dimethyl-3-oxo-2-phenyl-2,3-dihydro-1*H*-pyrazol-4-yl)(methyl)amino]methanesulfonate monohydrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *metamizole sodium CRS.*

B. Dissolve 50 mg in 1 mL of *strong hydrogen peroxide solution R*. A blue colour is produced which fades rapidly and turns to intense red in a few minutes.

C. Place 0.10 g in a test tube, add some glass beads and dissolve the substance in 1.5 mL of *water R*. Add 1.5 mL of *dilute hydrochloric acid R* and place a filter paper wetted with a solution of 20 mg of *potassium iodate R* in 2 mL of *starch solution R* at the open end of the test tube. Heat gently, the evolving vapour of sulfur dioxide colours the filter paper blue. After heating gently for 1 min, take a glass rod with a drop of a 10 g/L solution of *chromotropic acid, sodium salt R* in *sulfuric acid R* and place in the opening of the tube. Within 10 min, a blue-violet colour develops in the drop of the reagent.

D. 0.5 mL of solution S (see Tests) gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 40 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and, immediately after preparation, not more intensely coloured than reference solution BY_6 (*2.2.2, Method I*).

Acidity or alkalinity
To 5 mL of solution S, add 0.1 mL of *phenolphthalein solution R1*. The solution is colourless. Not more than 0.1 mL of *0.02 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *metamizole impurity A CRS* in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase. Use 1.0 mL of this solution to dissolve the contents of a vial of *metamizole impurity E CRS*.

Reference solution (c) In order to prepare impurity C *in situ*, dissolve 40 mg of the substance to be examined in *methanol R*, dilute to 20 mL with the same solvent and boil under reflux for 10 min. Allow to cool to room temperature and dilute to 20 mL with *methanol R*.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *methanol R*.

Column:
— *size*: l = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (1.8 µm).

Mobile phase Mix 28 volumes of *methanol R* and 72 volumes of a buffer solution prepared as follows: mix 1000 volumes of a 6.0 g/L solution of *sodium dihydrogen phosphate R* and 1 volume of *triethylamine R*, then adjust to pH 7.0 with *strong sodium hydroxide solution R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL of the test solution and reference solutions (b), (c) and (d).

Run time 4.5 times the retention time of metamizole.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and E; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C.

Relative retention With reference to metamizole (retention time = about 2 min): impurity A = about 0.7; impurity E = about 0.8; impurity C = about 2.5.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity E.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity E by 1.5;
— *impurity C*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *impurity E*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);

— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.03 per cent).

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

Dissolve 0.150 g in *distilled water R* and dilute to 15 mL with the same solvent.

Loss on drying (*2.2.32*)
4.9 per cent to 5.3 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.200 g in 10 mL of *0.01 M hydrochloric acid* previously cooled in iced water and titrate immediately, dropwise, with *0.05 M iodine*. Before each addition of *0.05 M iodine* dissolve the precipitate by swirling. At the end of the titration, add 2 mL of *starch solution R* and titrate until the blue colour of the solution persists for at least 2 min.
The temperature of the solution during the titration must not exceed 10 °C.

1 mL of *0.05 M iodine* is equivalent to 16.67 mg of $C_{13}H_{16}N_3NaO_4S$.

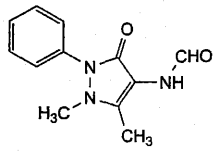
STORAGE

Protected from light.

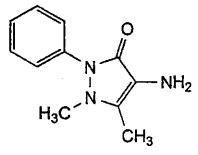
IMPURITIES

Specified impurities C, E.

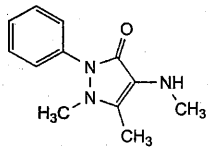
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B, D.



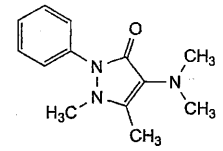
A. 4-(formylamino)-1,5-dimethyl-2-phenyl-2,3-dihydro-1*H*-pyrazol-3-one,



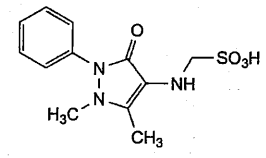
B. 4-amino-1,5-dimethyl-2-phenyl-2,3-dihydro-1*H*-pyrazol-3-one,



C. 1,5-dimethyl-4-(methylamino)-2-phenyl-2,3-dihydro-1*H*-pyrazol-3-one,



D. 1,5-dimethyl-4-(dimethylamino)-2-phenyl-2,3-dihydro-1*H*-pyrazol-3-one,

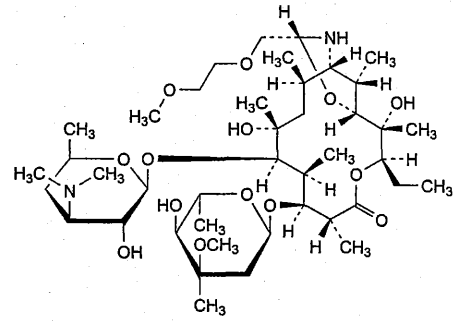


E. [(1,5-dimethyl-3-oxo-2-phenyl-2,3-dihydro-1*H*-pyrazol-4-yl)amino]methanesulfonic acid (4-*N*-desmethylmetamizole).

Ph Eur

Dirithromycin

(Ph. Eur. monograph 1313)



$C_{42}H_{78}N_2O_{14}$ 835 *62013-04-1*

Action and use
Macrolide antibacterial.

Ph Eur

DEFINITION

(1*R*,2*S*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*,12*R*,13*S*,15*R*,17*S*)-9-[[3-(Dimethylamino)-3,4,6-trideoxy-β-D-*xylo*-hexopyranosyl]oxy]-3-ethyl-2,10-dihydroxy-15-[(2-methoxyethoxy)methyl]-2,6,8,10,12,17-hexamethyl-7-[(3-*C*-methyl-3-*O*-methyl-2,6dideoxy-α-L-*ribo*-hexopyranosyl)oxy]-4,16-dioxa-14azabicyclo[11.3.1]heptadecan-5-one (or (9*S*)-9,11-[imino[(1*R*)-2-(2-methoxyethoxy)ethylidene]oxy]-9-deoxo-11-deoxyerythromycin).

Semi-synthetic product derived from a fermentation product.

Content
96.0 per cent to 102.0 per cent for the sum of the percentage contents of $C_{42}H_{78}N_2O_{14}$ and dirithromycin 15*S*-epimer (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, very soluble in methanol and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dirithromycin CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (a) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, acetonitrile R1* (30:70 *V/V*).

Test solution (a) Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 0.10 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of *dirithromycin CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 20 mg of *dirithromycin CRS* in the mobile phase and dilute to 10 mL with the mobile phase. Allow to stand for 24 h before use.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 9 volumes of *water R*, 19 volumes of *methanol R*, 28 volumes of a solution containing 1.9 g/L of *potassium dihydrogen phosphate R* and 9.1 g/L of *dipotassium hydrogen phosphate R* adjusted to pH 7.5 if necessary with a 100 g/L solution of *potassium hydroxide R*, and 44 volumes of *acetonitrile R1*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 10 µL of test solution (b) and reference solutions (b) and (c).

Run time 3 times the retention time of dirithromycin.

Relative retention With reference to dirithromycin: impurity A = about 0.7; 15*S*-epimer = about 1.1.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to dirithromycin and its 15*S*-epimer; if necessary, adjust the concentration of the organic modifiers in the mobile phase.

Limits:
— *impurity A*: not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *disregard limit*: disregard the peak due to the 15*S*-epimer.

Dirithromycin 15*S*-epimer

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (b).

System suitability Reference solution (b):
— *repeatability*: maximum relative standard deviation of 5.0 per cent after 6 injections.

Limit:
— *15S-epimer*: maximum 1.5 per cent.

Acetonitrile (*2.4.24, System A*)

Maximum 0.1 per cent.

Prepare the solutions using *dimethylformamide R* instead of *water R*.

Sample solution Dissolve 0.200 g of the substance to be examined in *dimethylformamide R* and dilute to 20.0 mL with the same solvent.

Static head-space injection conditions that may be used:
— *equilibration temperature*: 120 °C;
— *equilibration time*: 60 min;
— *transfer-line temperature*: 125 °C.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

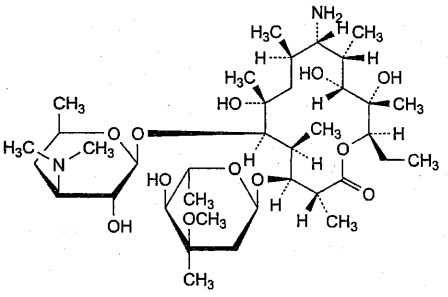
Injection Test solution (a) and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

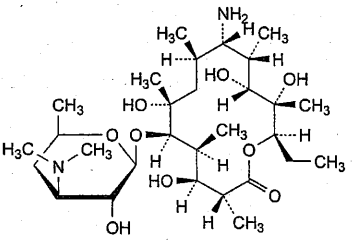
IMPURITIES

Specified impurities *A*.

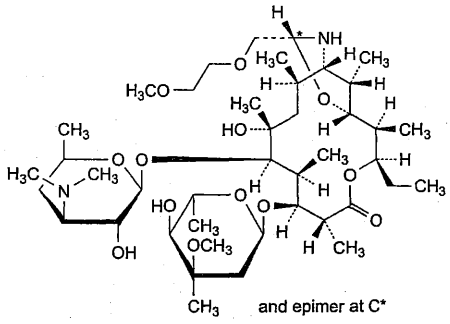
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E*.



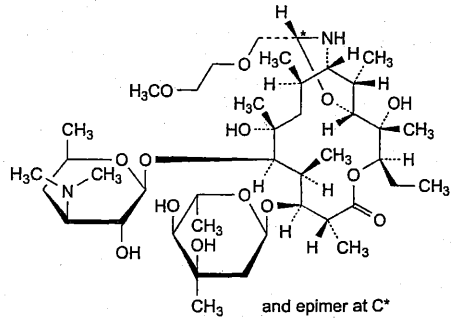
A. (9*S*)-9-amino-9-deoxoerythromycin,



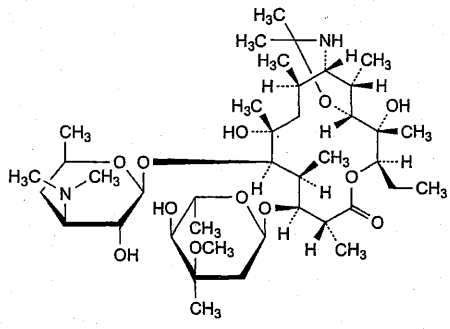
B. (9*S*)-9-amino-3-de(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)-9-deoxoerythromycin,



C. (9*S*)-9,11-[imino[(1*RS*)-2-(2-methoxyethoxy) ethylidene]oxy]-9-deoxo-11,12-dideoxyerythromycin (dirithromycin B),



D. (9*S*)-9,11-[imino[(1*RS*)-2-(2-methoxyethoxy) ethylidene]oxy]-3′-*O*-demethyl-9-deoxo-11-deoxyerythromycin (dirithromycin C),

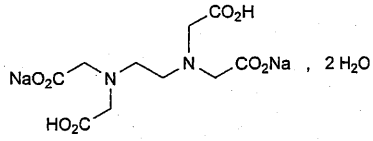


E. 9,11-[imino(1-methylethylidene)oxy]-9-deoxo-11-deoxyerythromycin.

Ph Eur

Disodium Edetate

(Ph. Eur. monograph 0232)



$C_{10}H_{14}N_2Na_2O_8,2H_2O$ 372.2

Action and use
Chelating agent.

Preparations
Disodium Edetate Eye Drops
Trisodium Edetate Infusion

Ph Eur

DEFINITION
Disodium dihydrogen (ethylenedinitrilo)tetraacetate dihydrate.

Content
98.5 per cent to 101.0 per cent.

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *disodium edetate CRS.*

B. Dissolve 2 g in 25 mL of *water R*, add 6 mL of *lead nitrate solution R*, shake and add 3 mL of *potassium iodide solution R*. No yellow precipitate is formed. Make alkaline to *red litmus paper R* by the addition of *dilute ammonia R2*. Add 3 mL of *ammonium oxalate solution R*. No precipitate is formed.

C. Dissolve 0.5 g in 10 mL of *water R* and add 0.5 mL of *calcium chloride solution R*. Make alkaline to *red litmus paper R* by the addition of *dilute ammonia R2* and add 3 mL of *ammonium oxalate solution R*. No precipitate is formed.

D. It gives the reactions of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 5.5 for solution S.

Impurity A
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture Dissolve 10.0 g of *ferric sulfate pentahydrate R* in 20 mL of *0.5 M sulfuric acid* and add 780 mL of *water R*. Adjust to pH 2.0 with *1 M sodium hydroxide* and dilute to 1000 mL with *water R*.

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution Dissolve 40.0 mg of *nitrilotriacetic acid R* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. To 1.0 mL of the solution add 0.1 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Column:
— *size:* l = 0.10 m, Ø = 4.6 mm,
— *stationary phase:* spherical *graphitised carbon for chromatography R1* (5 µm) with a specific surface area of 120 m^2/g and a pore size of 25 nm.

Mobile phase Dissolve 50.0 mg of *ferric sulfate pentahydrate R* in 50 mL of *0.5 M sulfuric acid* and add 750 mL of *water R*. Adjust to pH 1.5 with *0.5 M sulfuric acid* or *1 M sodium hydroxide*, add 20 mL of *ethylene glycol R* and dilute to 1000 mL with *water R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 µL; filter the solutions and inject immediately.

Run time 4 times the retention time of the iron complex of impurity A.

Retention times Iron complex of impurity A = about 5 min; iron complex of edetic acid = about 10 min.

System suitability Reference solution:
- *resolution*: minimum 7 between the peaks due to the iron complex of impurity A and the iron complex of edetic acid,
- *signal-to-noise ratio*: minimum 50 for the peak due to impurity A.

Limit:
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Iron (*2.4.9*)
Maximum 80 ppm.

Dilute 2.5 mL of solution S to 10 mL with *water R*. Add 0.25 g of *calcium chloride R* to the test solution and the standard before the addition of the *thioglycollic acid R*.

ASSAY

Dissolve 0.300 g in *water R* and dilute to 300 mL with the same solvent. Add 2 g of *hexamethylenetetramine R* and 2 mL of *dilute hydrochloric acid R*. Titrate with *0.1 M lead nitrate*, using about 50 mg of *xylenol orange triturate R* as indicator.

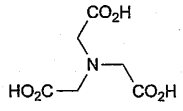
1 mL of *0.1 M lead nitrate* is equivalent to 37.22 mg of $C_{10}H_{14}N_2Na_2O_8,2H_2O$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities *A*.



A. nitrilotriacetic acid.

Ph Eur

Disodium Hydrogen Phosphate

Anhydrous Disodium Hydrogen Phosphate

(Disodium Phosphate, Ph. Eur. monograph 1509)

Na_2HPO_4 142.0 *7558-79-4*

Action and use
Excipient.

Preparation
Phosphates Enema

Ph Eur

DEFINITION

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder, hygroscopic.

Solubility
Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is slightly alkaline (*2.2.4*).

B. Loss on drying (see Tests).

C. Solution S gives reaction (b) of phosphates (*2.3.1*).

D. Solution S gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *distilled water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Reducing substances
To 10 mL of solution S add 5 mL of *dilute sulfuric acid R* and 0.25 mL of *0.02 M potassium permanganate* and heat on a water-bath for 5 min. The colour of the permanganate is not completely discharged.

Monosodium phosphate
Maximum 2.5 per cent.

From the volume of *1 M hydrochloric acid* (25 mL) and of *1 M sodium hydroxide* (n_1 mL and n_2 mL) used in the assay, calculate the following ratio:

$$\frac{n_2 - 25}{25 - n_1}$$

This ratio is not greater than 0.025.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *dilute nitric acid R*.

Sulfates (*2.4.13*)
Maximum 500 ppm.

To 6 mL of solution S add 2 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 2 ppm, determined on 10 mL of solution S.

Iron (*2.4.9*)
Maximum 20 ppm, determined on solution S.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

ASSAY

Dissolve 1.600 g (m) in 25.0 mL of *carbon dioxide-free water R* and add 25.0 mL of *1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*) using *1 M sodium hydroxide*. Read the volume added at the 1st inflexion point (n_1 mL). Continue the titration to the 2nd inflexion point (total volume of *1 M sodium hydroxide* required, n_2 mL).

Calculate the percentage content of Na_2HPO_4 from the following expression:

$$\frac{1420(25 - n_1)}{m(100 - d)}$$

d = percentage loss on drying.

STORAGE

In an airtight container.

Ph Eur

Disodium Hydrogen Phosphate Dihydrate

Sodium Phosphate Dihydrate

(Disodium Phosphate Dihydrate, Ph. Eur. monograph 0602)

$Na_2HPO_4,2H_2O$ 178.0 *10028-24-7*

Action and use
Excipient.

Preparations
Phosphates Enema

Phosphate Oral Solution

Ph Eur

DEFINITION

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder or colourless crystals.

Solubility
Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is slightly alkaline (*2.2.4*).

B. Loss on drying (see Tests).

C. Solution S gives reaction (b) of phosphates (*2.3.1*).

D. Solution S gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Reducing substances
To 5 mL of solution S add 5 mL of *dilute sulfuric acid R* and 0.25 mL of *0.02 M potassium permanganate* and heat on a water-bath for 5 min. The colour of the permanganate is not completely discharged.

Monosodium phosphate
Maximum 2.5 per cent.

From the volume of *1 M hydrochloric acid* (25 mL) and of *1 M sodium hydroxide* (n_1 mL and n_2 mL) used in the assay, calculate the following ratio:

$$\frac{n_2 - 25}{25 - n_1}$$

This ratio is not greater than 0.025.

Chlorides (*2.4.4*)
Maximum 400 ppm.

To 2.5 mL of solution S add 10 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 0.1 per cent.

To 3 mL of solution S add 2 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 4 ppm, determined on 5 mL of solution S.

Iron (*2.4.9*)
Maximum 40 ppm.

Dilute 5 mL of solution S to 10 mL with *water R*.

Loss on drying (*2.2.32*)
19.5 per cent to 21.0 per cent, determined on 1.000 g by drying in an oven at 130 °C.

ASSAY

Dissolve 2.000 g (*m*) in 50 mL of *water R* and add 25.0 mL of *1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*) using *1 M sodium hydroxide*. Read the volume added at the 1st inflexion point (n_1 mL). Continue the titration to the 2nd inflexion point (total volume of *1 M sodium hydroxide* required, n_2 mL).

Calculate the percentage content of Na_2HPO_4 from the following expression:

$$\frac{1420(25 - n_1)}{m(100 - d)}$$

d = percentage loss on drying.

Ph Eur

Disodium Hydrogen Phosphate Dodecahydrate

Disodium Hydrogen Phosphate

Sodium Phosphate

(Disodium Phosphate Dodecahydrate, Ph. Eur. monograph 0118)

$Na_2HPO_4,12H_2O$ 358.1 *10039-32-4*

Preparation
Phosphates Enema

Ph Eur

DEFINITION

Content
98.5 per cent to 102.5 per cent.

CHARACTERS

Appearance
Colourless, transparent crystals, very efflorescent.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is slightly alkaline (*2.2.4*).

B. Loss on drying (see Tests).

C. Solution S gives reaction (b) of phosphates (*2.3.1*).

D. Solution S gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Reducing substances
To 5 mL of solution S add 5 mL of *dilute sulfuric acid R* and 0.25 mL of *0.02 M potassium permanganate* and heat on a water-bath for 5 min. The colour of the permanganate is not completely discharged.

Monosodium phosphate
Maximum 2.5 per cent.

From the volume of *1 M hydrochloric acid* (n_3 mL) and of *1 M sodium hydroxide* (n_1 mL and n_2 mL) used in the assay, calculate the following ratio:

$$\frac{n_2 - n_3}{n_3 - n_1}$$

This ratio is not greater than 0.025.

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 2.5 mL of solution S add 10 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 500 ppm.

To 3 mL of solution S add 2 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)
Maximum 2 ppm, determined on 5 mL of solution S.

Iron (*2.4.9*)
Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with *water R*.

Loss on drying (*2.2.32*)
57.0 per cent to 61.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 4 h.

ASSAY
Dissolve 4.00 g (*m*) in 25 mL of *water R* and add 25.0 mL of *1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*) using *1 M sodium hydroxide*. Read the volume added at the 1st inflexion point (n_1 mL). Continue the titration to the 2nd inflexion point (total volume of *1 M sodium hydroxide* required, n_2 mL). Carry out a blank titration (n_3 mL).

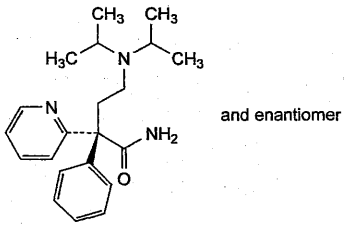
Calculate the percentage content of $Na_2HPO_4,12H_2O$ using the following expression:

$$\frac{3581(n_3 - n_1)}{m \times 100}$$

Ph Eur

Disopyramide

(Ph. Eur. monograph 1006)



and enantiomer

$C_{21}H_{29}N_3O$ 339.5 *3737-09-5*

Action and use
Class I antiarrhythmic.

Preparation
Disopyramide Capsules

Ph Eur

DEFINITION
Disopyramide contains not less than 98.5 per cent and not more than the equivalent of 101.5 per cent of (2*RS*)-4-[bis(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl) butanamide, calculated with reference to the dried substance.

CHARACTERS
A white or almost white powder, slightly soluble in water, freely soluble in methylene chloride, soluble in alcohol.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Dissolve 40.0 mg in a 5 g/L solution of *sulfuric acid R* in *methanol R* and dilute to 100.0 mL with the same solution. Dilute 5.0 mL of this solution to 50.0 mL with a 5 g/L solution of *sulfuric acid R* in *methanol R*. Examined between 240 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 269 nm and a shoulder at 263 nm. The specific absorbance at the maximum is 190 to 210.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *disopyramide CRS*. Examine the substances as discs prepared by placing 50 µL of a 50 g/L solution in *methylene chloride R* on a disc of *potassium bromide R*. Dry the discs at 60 °C for 1 h before use.

C. Examine the chromatograms obtained in the test for related substances in ultraviolet light at 254 nm.
The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a). Spray with *dilute potassium iodobismuthate solution R*. Examine in daylight. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel GF_{254} R* as the coating substance.

Test solution (a) Dissolve 0.20 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dissolve 20 mg of *disopyramide CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 0.5 mL of test solution (b) to 20 mL with *methanol R*.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of *concentrated ammonia R*, 30 volumes of *acetone R* and 30 volumes of *cyclohexane R*. Dry the plate in a current of warm air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying at 80 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 2 h.

Sulfated ash (*2.4.14*)
Not more than 0.2 per cent, determined on 1.0 g.

ASSAY

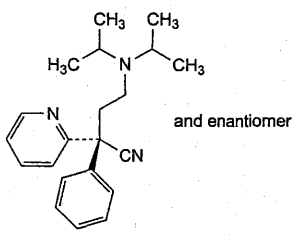
Dissolve 0.130 g in 30 mL of *anhydrous acetic acid R*.
Add 0.2 mL of *naphtholbenzein solution R*. Titrate with *0.1 M perchloric acid* until the colour changes from yellow to green.

1 mL of *0.1 M perchloric acid* is equivalent to 16.97 mg of $C_{21}H_{29}N_3O$.

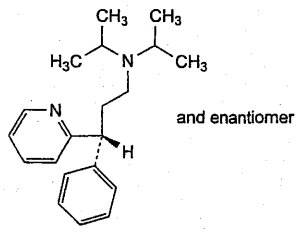
STORAGE

Store protected from light.

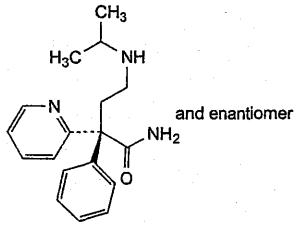
IMPURITIES



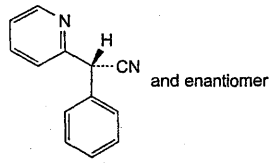
A. (2*RS*)-4-[bis(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl)butanenitrile (di-isopyronitrile),



B. (3*RS*)-*N*,*N*-bis(1-methylethyl)-3-phenyl-3-(pyridin-2-yl) propan-1-amine,



C. (2*RS*)-4-[(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl) butanamide,

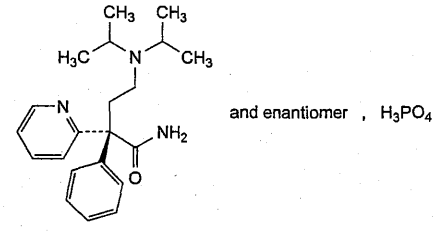


D. (*RS*)-phenyl(pyridin-2-yl)acetonitrile (pyronitrile).

Ph Eur

Disopyramide Phosphate

(Ph. Eur. monograph 1005)



$C_{21}H_{32}N_3O_5P$ 437.5 *22059-60-5*

Action and use

Antiarrhythmic.

Preparation

Disopyramide Phosphate Capsules

Ph Eur

DEFINITION

Disopyramide phosphate contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of (2*RS*)-4-[bis(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl) butanamide dihydrogen phosphate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white powder, soluble in water, sparingly soluble in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Dissolve 50.0 mg in a 5 g/L solution of *sulfuric acid R* in *methanol R* and dilute to 100.0 mL with the same solution. Dilute 5.0 mL of this solution to 50.0 mL with a 5 g/L solution of *sulfuric acid R* in *methanol R*. Examined between 240 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 269 nm and a shoulder at 263 nm. The specific absorbance at the maximum is 147 to 163.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *disopyramide phosphate CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for related substances in ultraviolet light at 254 nm.
The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a). Spray with *dilute potassium iodobismuthate solution R*. Examine in daylight. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Solution S (see Tests) gives reaction (a) of phosphates (*2.3.1*).

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
The pH of solution S is 4.0 to 5.0.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel GF_{254} R* as the coating substance.

Test solution (a) Dissolve 0.25 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dissolve 25 mg of *disopyramide phosphate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 20 mL with *methanol R*.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of *concentrated ammonia R*, 30 volumes of *acetone R* and 30 volumes of *cyclohexane R*. Dry the plate in a current of warm air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

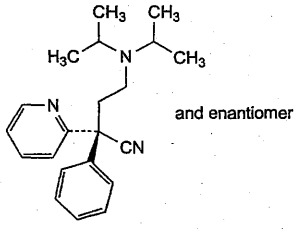
Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
Dissolve 0.180 g in 30 mL of *anhydrous acetic acid R*. Add 0.2 mL of *naphtholbenzein solution R*. Titrate with *0.1 M perchloric acid* until the colour changes from yellow to green.

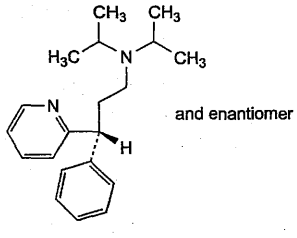
1 mL of *0.1 M perchloric acid* is equivalent to 21.88 mg of $C_{21}H_{32}N_3O_5P$.

STORAGE
Store protected from light.

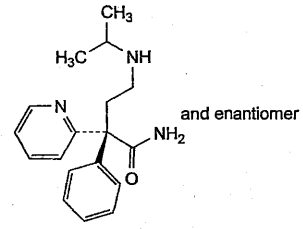
IMPURITIES



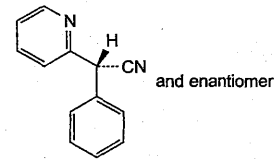
A. (2*RS*)-4-[bis(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl)butanenitrile (di-isopyronitrile),



B. (3*RS*)-*N*,*N*-bis(1-methylethyl)-3-phenyl-3-(pyridin-2-yl) propan-1-amine,



C. (2*RS*)-4-[(1-methylethyl)amino]-2-phenyl-2-(pyridin-2-yl) butanamide,

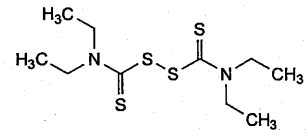


D. (*RS*)-phenyl(pyridin-2-yl)acetonitrile (pyronitrile).

Ph Eur

Disulfiram

(Ph. Eur. monograph 0603)



$C_{10}H_{20}N_2S_4$ 296.5 *97-77-8*

Action and use
Aldehyde dehyrogenase inhibitor; treatment of alcoholism.

Preparation
Disulfiram Tablets

Ph Eur

DEFINITION
Disulfiram contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of tetraethyldisulfanedicarbothioamide, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder, practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in alcohol.

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 70 °C to 73 °C.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *disulfiram CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 10 mL of *methanol R*. Add 2 mL of a 0.5 g/L solution of *cupric chloride R* in *methanol R*. A yellow colour develops which becomes greenish-yellow.

TESTS

Related substances

Examine by thin-layer chromatography (*2.2.27*), using as the coating substance a suitable silica gel with a fluorescent indicator having an optimal intensity at 254 nm.

Test solution (a) Dissolve 0.20 g of the substance to be examined in *ethyl acetate R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *ethyl acetate R*.

Reference solution (a) Dissolve 10 mg of *disulfiram CRS* in *ethyl acetate R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 20 mL with *ethyl acetate R*.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 30 volumes of *butyl acetate R* and 70 volumes of *hexane R*. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Diethyldithiocarbamate

Dissolve 0.20 g in 10 mL of *peroxide-free ether R*, add 5 mL of *buffer solution pH 8.0 R* and shake vigorously. Discard the upper layer and wash the lower layer with 10 mL of *peroxide-free ether R*. Add to the lower layer 0.2 mL of a 4 g/L solution of *copper sulfate pentahydrate R* and 5 mL of *cyclohexane R*. Shake. Any yellow colour in the upper layer is not more intense than that of a standard prepared at the same time using 0.2 mL of a freshly prepared 0.15 g/L solution of *sodium diethyldithiocarbamate R* (150 ppm).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 50 °C.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

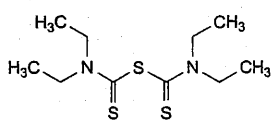
Dissolve 0.450 g in 80 mL of *acetone R* and add 20 mL of a 20 g/L solution of *potassium nitrate R*. Titrate with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*), using a silver electrode and a silver-silver chloride double-junction electrode saturated with potassium nitrate.

1 mL of *0.1 M silver nitrate* is equivalent to 59.30 mg of $C_{10}H_{20}N_2S_4$.

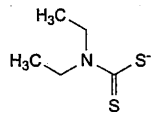
STORAGE

Store protected from light.

IMPURITIES



A. diethylthiocarbamic thioanhydride (sulfiram),

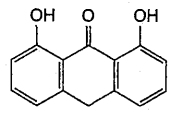


B. diethyldithiocarbamate.

Ph Eur

Dithranol

(Ph. Eur. monograph 1007)



$C_{14}H_{10}O_3$ 226.2 *1143-38-0*

Action and use

Coal tar extract; treatment of psoriasis.

Preparations

Dithranol Cream

Dithranol Ointment

Dithranol Paste

Dithranol and Salicylic Acid Ointment

Ph Eur

DEFINITION

1,8-Dihydroxyanthracen-9(10*H*)-one.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow or brownish-yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in methylene chloride, sparingly soluble in acetone, slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

Carry out all tests protected from bright light and use freshly prepared solutions.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 178 °C to 182 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison dithranol CRS.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *dithranol CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve about 5 mg of *dantron R* (impurity B) in 5 mL of reference solution (a).

Plate TLC silica gel plate R.

Mobile phase hexane R, methylene chloride R (50:50 *V/V*).

Application 10 µL.

Development Over a path of 12 cm.

Drying In air.

Detection Place the plate in a tank saturated with ammonia vapour until the spots appear. Examine in daylight.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to

the principal spot in the chromatogram obtained with reference solution (a).

D. To 5 mg add 0.1 g of *anhydrous sodium acetate R* and 1 mL of *acetic anhydride R*. Boil for 30 s. Add 20 mL of *ethanol (96 per cent) R*. Examined in ultraviolet light at 365 nm, the solution shows a blue fluorescence.

TESTS

Related substances

A. Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.200 g of the substance to be examined in 20 mL of *methylene chloride R*, add 1.0 mL of *glacial acetic acid R* and dilute to 100.0 mL with *hexane R*.

Reference solution Dissolve 5.0 mg of *anthrone R* (impurity A), 5.0 mg of *dantron R* (impurity B), 5.0 mg of *dithranol impurity C CRS* and 5.0 mg of *dithranol CRS* in *methylene chloride R* and dilute to 5.0 mL with the same solvent. To 1.0 mL of this solution, add 19.0 mL of *methylene chloride R* and 1.0 mL of *glacial acetic acid R*, and dilute to 50.0 mL with *hexane R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *glacial acetic acid R, methylene chloride R, hexane R* (1:5:82 *V/V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 µL.

Run time 1.5 times the retention time of impurity C.

Elution order Dithranol, impurity B, impurity A, impurity C.

System suitability Reference solution:
- *resolution*: minimum 2.0 between the peaks due to dithranol and impurity B.

Limits:
- *impurities A, B, C*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (1 per cent).

B. Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in 5 mL of *tetrahydrofuran R* and dilute to 25.0 mL with the mobile phase.

Reference solution Dissolve 5.0 mg of *dithranol impurity D CRS* and 5.0 mg of *dithranol CRS* in 5 mL of *tetrahydrofuran R* and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.20 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *glacial acetic acid R, tetrahydrofuran R, water R* (2.5:40:60 *V/V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 3 times the retention time of dithranol.

System suitability Reference solution:
- *resolution*: minimum 2.5 between the peaks due to impurity D and dithranol.

Limit:
- *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (2.5 per cent).

Total (tests A + B) Maximum 3.0 per cent for the sum of the contents of all impurities.

Chlorides (*2.4.4*)

Maximum 100 ppm.

Shake 1.0 g with 20 mL of *water R* for 1 min and filter. Dilute 10 mL of the filtrate to 15 mL with *water R*.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of *anhydrous pyridine R*. Titrate with 0.1 M tetrabutylammonium hydroxide under *nitrogen R*. Determine the end-point potentiometrically (*2.2.20*), using a glass indicator electrode and a suitable reference electrode.

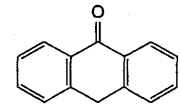
1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 22.62 mg of $C_{14}H_{10}O_3$.

STORAGE

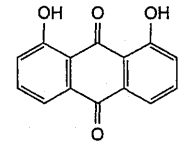
Protected from light.

IMPURITIES

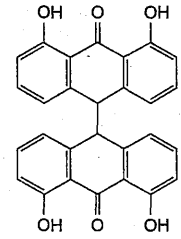
Specified impurities *A, B, C, D.*



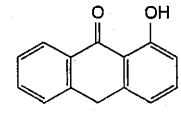
A. anthracen-9(10*H*)-one (anthrone),



B. 1,8-dihydroxyanthracene-9,10-dione (dantron),



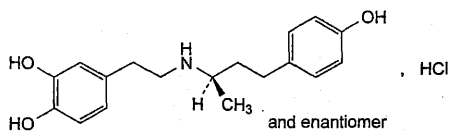
C. 4,4′,5,5′-tetrahydroxy-9,9′-bianthracenyl-10,10′(9*H*,9′*H*)-dione,



D. 1-hydroxyanthracen-9(10*H*)-one.

Dobutamine Hydrochloride

(Ph. Eur. monograph 1200)



$C_{18}H_{24}ClNO_3$ 337.8 *49745-95-1*

Action and use
$Beta_1$-adrenoceptor agonist.

Preparation
Dobutamine Infusion

Ph Eur

DEFINITION

4-[2-[[(2*RS*)-4-(4-Hydroxyphenyl)-butan-2-yl]amino] ethyl]benzene-1,2-diol hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dobutamine hydrochloride CRS.*

B. It gives reaction (a) of chlorides (*2.3.1*) using a mixture of equal volumes of *methanol R* and *water R*.

TESTS

Acidity or alkalinity
Dissolve 0.1 g in *water R* with gentle heating and dilute to 10 mL with the same solvent. Add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is red.

Absorbance (*2.2.25*)
Maximum 0.04 at 480 nm.

Dissolve 0.5 g in a mixture of equal volumes of *methanol R* and *water R* with heating, if necessary, at 30-35 °C and dilute to 25 mL with the same mixture of solvents. Cool quickly. Examine immediately.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase B, mobile phase A (35:65 *V/V*).

Test solution Dissolve 0.10 g of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 4.0 mL of the test solution to 100.0 mL with a 0.05 g/L solution of *anisaldehyde R* in the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *dobutamine impurity mixture CRS* (impurities A, B and C) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 2.60 g of *sodium octanesulfonate R* in 1000 mL of *water R*, add 3 mL of *triethylamine R* and adjust to pH 2.5 with *phosphoric acid R*;
— *mobile phase B*: *acetonitrile R*, *methanol R* (18:82 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	65	35
5 - 20	65 → 20	35 → 80
20 - 25	20	80

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *dobutamine impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to dobutamine (retention time = about 12 min): impurity A = about 0.3; impurity B = about 0.5; anisaldehyde = about 0.7; impurity C = about 1.4.

System suitability Reference solution (a):
— *resolution*: minimum 4.0 between the peaks due to dobutamine and anisaldehyde.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 1.4;
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.250 g in 10 mL of *anhydrous formic acid R*. Add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

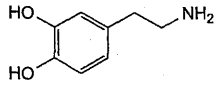
1 mL of *0.1 M perchloric acid* is equivalent to 33.79 mg of $C_{18}H_{24}ClNO_3$.

STORAGE

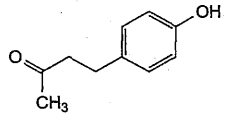
Protected from light.

IMPURITIES

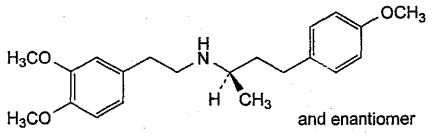
Specified impurities *A, B, C.*



A. 4-(2-aminoethyl)benzene-1,2-diol (dopamine),



B. 4-(4-hydroxyphenyl)butan-2-one,



and enantiomer

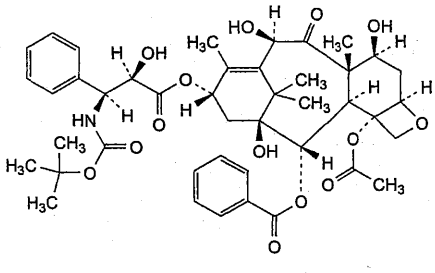
C. (2*RS*)-*N*-[2-(3,4-dimethoxyphenyl)ethyl]-4-(4-methoxyphenyl)butan-2-amine.

Ph Eur

Docetaxel

Anhydrous Docetaxel

(Ph. Eur. monograph 2593)



$C_{43}H_{53}NO_{14}$ 808 *114977-28-5*

Action and use

Taxane cytotoxic.

Ph Eur

DEFINITION

5β,20-Epoxy-1,7β,10β-trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate].

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline, hygroscopic powder.

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol, soluble in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *anhydrous docetaxel CRS.*

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution B_5 (*2.2.2, Method I*).

Dissolve 1.0 g in *anhydrous ethanol R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)

−41.5 to −38.5 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetic acid R, acetonitrile R1, water R* (0.05:50:50 *V/V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *anhydrous ethanol R* and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of *docetaxel trihydrate CRS* in 2.5 mL of *anhydrous ethanol R* and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *docetaxel for system suitability CRS* (containing impurities A, B and C) in 0.25 mL of *anhydrous ethanol R* and dilute to 5.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5 mg of *docetaxel impurity E CRS* in 2.5 mL of *anhydrous ethanol R* and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);

— *temperature*: 45 °C.

Mobile phase:

— *mobile phase A*: *water R*;

— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 9	72	28
9 - 39	72 → 28	28 → 72

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *docetaxel for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity E.

Relative retention With reference to docetaxel (retention time = about 27 min): impurity E = about 0.2;

impurity A = about 0.97; impurity B = about 1.08; impurity C = about 1.13.

System suitability Reference solution (c):

— *resolution*: minimum 3.0 between the peaks due to impurity A and docetaxel.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.6;

— *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *impurity E*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: maximum 0.8 per cent;

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.32*)

Maximum 1.5 per cent.

Inject 800 µL of a 25 mg/mL solution of the substance to be examined in *methanol R*.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.3 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $C_{43}H_{53}NO_{14}$ taking into account the assigned content of *docetaxel trihydrate CRS*.

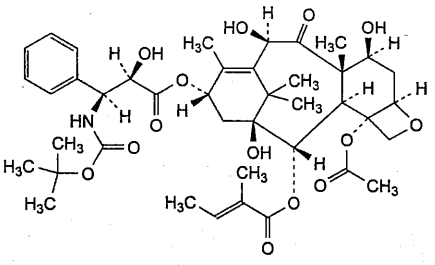
STORAGE

Protected from light, in an airtight container.

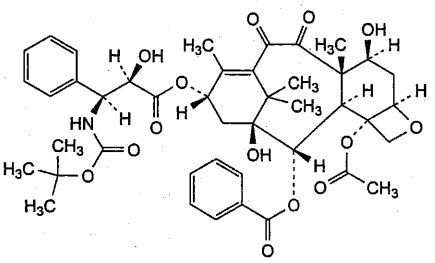
IMPURITIES

Specified impurities *A, B, C, E.*

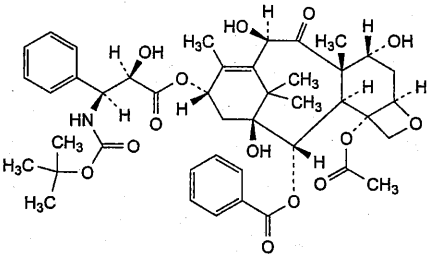
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, F, G.*



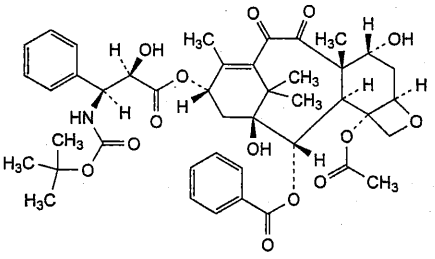
A. 5β,20-epoxy-1,7β,10β-trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] 2-[(2*E*)-2-methylbut-2-enoate] (2-*O*-desbenzoyl-2-*O*-tiglyldocetaxel),



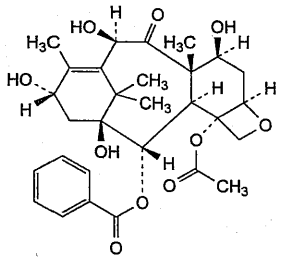
B. 5β,20-epoxy-1,7β-dihydroxy-9,10-dioxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (10-deoxy-10-oxodocetaxel),



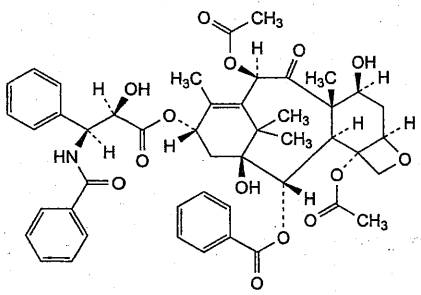
C. 5β,20-epoxy-1,7α,10β-trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (7-*epi*-docetaxel),



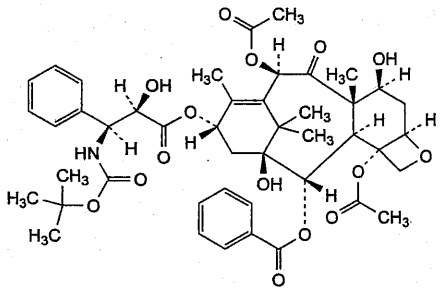
D. 5β,20-epoxy-1,7α-dihydroxy-9,10-dioxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (10-deoxy-10-oxo-7-*epi*-docetaxel),



E. 5β,20-epoxy-4-(acetyloxy)-1,7β,10β,13α-tetrahydroxy-9-oxotax-11-en-2α-yl benzoate (10-desacetyl-baccatin III),



F. 5β,20-epoxy-1,7β-dihydroxy-9-oxotax-11-ene-2α,4,10β,13α-tetrayl 4,10-diacetate 2-benzoate 13-[(2*R*,3*S*)-3-(benzoylamino)-2-hydroxy-3-phenylpropanoate] (paclitaxel),

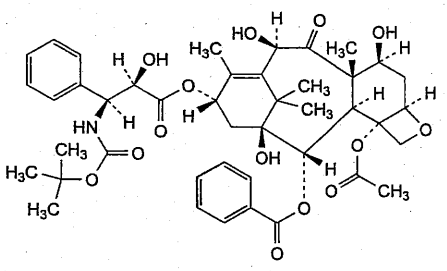


G. 5β,20-epoxy-1,7β-dihydroxy-9-oxotax-11-ene-2α,4,10β,13α-tetrayl 4,10-diacetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (10-acetyldocetaxel).

Ph Eur

Docetaxel Trihydrate

(Ph. Eur. monograph 2449)



$C_{43}H_{53}NO_{14},3H_2O$ 862 *148408-66-6*

Action and use
Taxane cytotoxic.

Ph Eur

DEFINITION

5β,20-epoxy-1,7β,10β-Trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] trihydrate.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in anhydrous ethanol, soluble in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *docetaxel trihydrate CRS.*

TESTS

Appearance of solution
The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution B_5 (*2.2.2, Method I*).

Dissolve 1.0 g in *anhydrous ethanol R* and dilute to 20 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−41.5 to −38.5 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetic acid R, acetonitrile R1, water R* (0.05:50:50 *V/V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *anhydrous ethanol R* and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of *docetaxel trihydrate CRS* in 2.5 mL of *anhydrous ethanol R* and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *docetaxel for system suitability CRS* (containing impurities A, B and C) in 0.25 mL of *anhydrous ethanol R* and dilute to 5.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 45 °C.

Mobile phase:
— *mobile phase A*: *water R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 9	72	28
9 - 39	72 → 28	28 → 72

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *docetaxel for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to docetaxel (retention time = about 27 min): impurity A = about 0.97; impurity B = about 1.08; impurity C = about 1.13.

System suitability Reference solution (c):
— *resolution*: minimum 3.0 between the peaks due to impurity A and docetaxel.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.6;
— *impurity A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurities B, C*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.32*)
5.0 per cent to 7.0 per cent.

Inject 200 µL of a 100 mg/mL solution of the substance to be examined in *dimethylformamide R*.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.3 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $C_{43}H_{53}NO_{14}$ taking into account the assigned content of *docetaxel trihydrate CRS*.

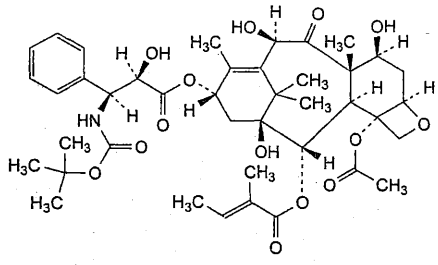
STORAGE

Protected from light.

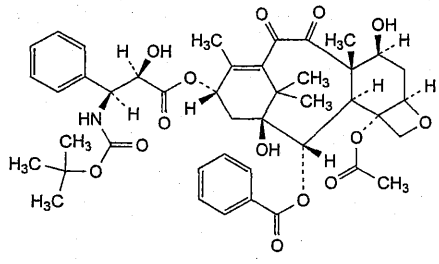
IMPURITIES

Specified impurities *A, B, C.*

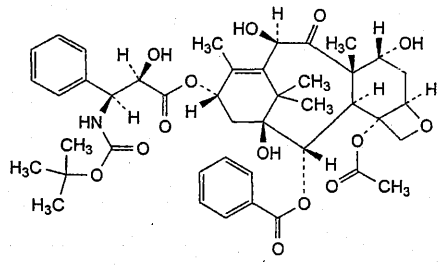
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*



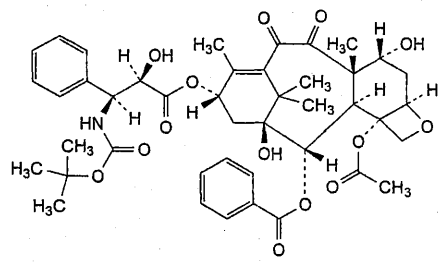
A. 5β,20-epoxy-1,7β,10β-trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 13-[(2*R*,3*S*)-3-[[(1,1dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] 2-[(2*E*)-2-methylbut-2-enoate] (2-*O*-desbenzoyl-2-*O*-tiglyldocetaxel),



B. 5β,20-epoxy-1,7β-dihydroxy-9,10-dioxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (10-deoxy-10-oxodocetaxel),



C. 5β,20-epoxy-1,7α,10β-trihydroxy-9-oxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (7-*epi*-docetaxel),



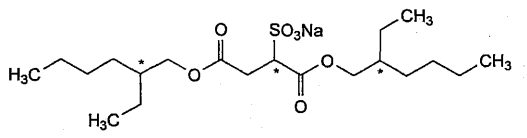
D. 5β,20-epoxy-1,7α-dihydroxy-9,10-dioxotax-11-ene-2α,4,13α-triyl 4-acetate 2-benzoate 13-[(2*R*,3*S*)-3-[[(1,1-dimethylethoxy)carbonyl]amino]-2-hydroxy-3-phenylpropanoate] (10-deoxy-10-oxo-7-*epi*-docetaxel).

Ph Eur

Docusate Sodium

Dioctyl Sodium Sulphosuccinate

(Ph. Eur. monograph 1418)



$C_{20}H_{37}NaO_7S$ 444.6 *577-11-7*

Action and use
Stimulant laxative; faecal softener.

Preparations
Co-danthrusate Capsules
Docusate Capsules
Compound Docusate Enema
Docusate Oral Solution
Paediatric Docusate Oral Solution

Ph Eur

DEFINITION
Sodium 1,4-bis[(2-ethylhexyl)oxy]-1,4-dioxobutane-2-sulfonate.

Content
98.0 to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, waxy masses or flakes, hygroscopic.

Solubility
Sparingly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Place about 3 mg of the substance to be examined on a sodium chloride plate, add 0.05 mL of *acetone R* and immediately cover with another sodium chloride plate. Rub the plates together to dissolve the substance to be examined, slide the plates apart and allow the acetone to evaporate.

Comparison *docusate sodium CRS.*

B. In a crucible, ignite 0.75 g in the presence of *dilute sulfuric acid R*, until an almost white residue is obtained. Allow to cool and take up the residue with 5 mL of *water R*. Filter. 2 mL of the filtrate gives reaction (a) of sodium (*2.3.1*).

TESTS
Alkalinity
Dissolve 1.0 g in 100 mL of a mixture of equal volumes of *methanol R* and *water R*, previously neutralised to *methyl red solution R*. Add 0.1 mL of *methyl red solution R*. Not more than 0.2 mL of *0.1 M hydrochloric acid* is required to change the colour of the indicator to red.

Related non-ionic substances
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 10 mg of *methyl behenate R* in *hexane R* and dilute to 50 mL with the same solvent.

Test solution (a) Dissolve 0.10 g of the substance to be examined in 2.0 mL of the internal standard solution and dilute to 5.0 mL with *hexane R*. Pass the solution, at a rate of about 1.5 mL/min, through a column 10 mm in internal diameter, packed with 5 g of *basic aluminium oxide R* and previously washed with 25 mL of *hexane R*. Elute with 5 mL of *hexane R* and discard the eluate. Elute with 20 mL of a mixture of equal volumes of *ether R* and *hexane R*. Evaporate the eluate to dryness and dissolve the residue in 2.0 mL of *hexane R*.

Test solution (b) Prepare as described for test solution (a) but dissolving 0.10 g of the substance to be examined in *hexane R*, diluting to 5.0 mL with the same solvent, and using a new column.

Reference solution Dilute 2.0 mL of the internal standard solution to 5.0 mL with *hexane R*.

Column:
— *material*: glass;
— *size*: *l* = 2 m, Ø = 2 mm;
— *stationary phase*: *silanised diatomaceous earth for gas chromatography R* impregnated with 3 per cent *m/m* of *polymethylphenylsiloxane R*.

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Temperature:
— *column*: 230 °C;
— *injection port and detector*: 280 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time 2.5 times the retention time of the internal standard.

System suitability There is no peak with the same retention time as the internal standard in the chromatogram obtained with test solution (b).

Limits Test solution (a):
— *any impurity*: for each impurity, not more than the area of the peak due to the internal standard (0.4 per cent).

Chlorides
Maximum 350 ppm.

Dissolve 5.0 g in 50 mL of *ethanol (50 per cent V/V) R*. Titrate with *0.01 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.01 M silver nitrate* is equivalent to 0.3545 mg of Cl.

Sodium sulfate
Maximum 2 per cent.

Dissolve 0.25 g in 40 mL of a mixture of 20 volumes of *water R* and 80 volumes of *2-propanol R*. Adjust the pH to between 2.5 and 4.0 using *perchloric acid solution R*. Add 0.4 mL of *naphtharson solution R* and 0.1 mL of a 0.125 g/L solution of *methylene blue R*. Not more than 1.5 mL of *0.025 M barium perchlorate* is required to change the colour of the indicator from yellowish-green to yellowish-pink.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 0.250 g.

ASSAY
To 1.000 g in a 250 mL conical flask fitted with a reflux condenser add 25.0 mL of 0.5 M alcoholic potassium hydroxide and heat on a water-bath under reflux for 45 min. Allow to cool. Add 0.25 mL of *phenolphthalein solution R1* and titrate with *0.5 M hydrochloric acid* until the red colour disappears. Carry out a blank titration.

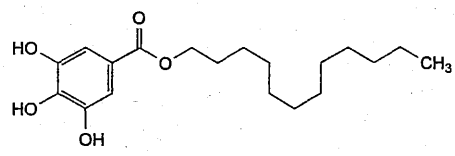
1 mL of 0.5 M alcoholic potassium hydroxide is equivalent to 0.1112 g of $C_{20}H_{37}NaO_7S$.

STORAGE
In an airtight container.

Ph Eur

Dodecyl Gallate

(Ph. Eur. monograph 2078)



$C_{19}H_{30}O_5$ 338.4 *1166-52-5*

Action and use
Antioxidant.

Ph Eur

DEFINITION
Dodecyl 3,4,5-trihydroxybenzoate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble or practically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION
A. Determine the melting point (*2.2.14*) of the substance to be examined. Mix equal parts of the substance to be examined and *dodecyl gallate CRS* and determine the melting point of the mixture. The difference between the melting points (which are about 96 °C) is not greater than 2 °C.

B. Examine the chromatograms obtained in the test for impurity A.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Impurity A
Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.20 g of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20 mL with *acetone R*.

Reference solution (a) Dissolve 10 mg of *dodecyl gallate CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *gallic acid R* in *acetone R* and dilute to 20 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10 mL with *acetone R*.

Reference solution (d) Dilute 1.0 mL of reference solution (b) to 5 mL with test solution (a).

Plate *TLC silica gel plate R*.

Mobile phase *anhydrous formic acid R, ethyl formate R, toluene R* (10:40:50 *V/V/V*).

Application 5 µL of test solutions (a) and (b) and reference solutions (a), (c) and (d).

Development Over 2/3 of the plate.

Drying In air for 10 min.

Detection Spray with a mixture of 1 volume of *ferric chloride solution R1* and 9 volumes of *ethanol (96 per cent) R*.

System suitability Reference solution (d):
— the chromatogram shows 2 clearly separated principal spots.

Limit Test solution (a):
— *impurity A*: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Chlorides (*2.4.4*)
Maximum 100 ppm.

To 1.65 g add 50 mL of *water R*. Shake for 5 min. Filter. 15 mL of the filtrate complies with the test.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 70 °C.

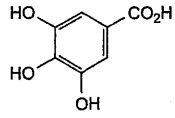
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.100 g in *methanol R* and dilute to 250.0 mL with the same solvent. Dilute 5.0 mL of the solution to 200.0 mL with *methanol R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 275 nm.

Calculate the content of $C_{19}H_{30}O_5$ taking the specific absorbance to be 321.

STORAGE
In a non-metallic container, protected from light.

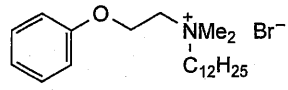
IMPURITIES
Specified impurities *A*.



A. 3,4,5-trihydroxybenzoic acid (gallic acid).

Ph Eur

Domiphen Bromide



$C_{22}H_{40}BrNO$ 414.5 *538-71-6*

Action and use
Antiseptic.

DEFINITION
Domiphen Bromide consists chiefly of dodecyldimethyl-2-phenoxyethylammonium bromide. It contains not less than 97.0% and not more than 100.5% of $C_{22}H_{40}BrNO$, calculated with reference to the dried substance.

CHARACTERISTICS

Colourless or faintly yellow, crystalline flakes.

Freely soluble in *water* and in *ethanol (96%)*; soluble in *acetone*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of domiphen bromide (*RS 383*).

B. Dissolve 10 mg in 10 mL of *water* and add 0.1 mL of a 0.5% w/v solution of *eosin* and 100 mL of *water*. An intense pink colour is produced.

C. Yields the reactions characteristic of *bromides*, Appendix VI.

TESTS

Acidity or alkalinity

Add 0.5 mL of *bromothymol blue solution R3* to each of 10 mL of *phosphate buffer pH 6.4* (solution A) and 10 mL of *phosphate buffer pH 7.6* (solution B). Dissolve 0.10 g in 10 mL of *carbon dioxide-free water* and add 0.5 mL of *bromothymol blue solution R3*. The resulting solution is not more yellow than solution A and not more blue than solution B.

Clarity and colour of solution

Dissolve 1.0 g in 10 mL of *carbon dioxide-free water*. The solution is not more opalescent than *reference suspension II*, Appendix IV A, and not more intensely coloured than *reference solution Y_7*, Appendix IV B, Method I.

Non-quaternary amines

Carry out the Assay described below using a further 25 mL of the original solution and 10 mL of 0.1M *hydrochloric acid* in place of the 0.1M *sodium hydroxide*. The difference between the volume of *0.05M potassium iodate VS* required in this titration and that required in the Assay is not more than 0.5 mL for each g of substance taken.

Loss on drying

When dried to constant weight at 70° at a pressure not exceeding 0.7 kPa, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

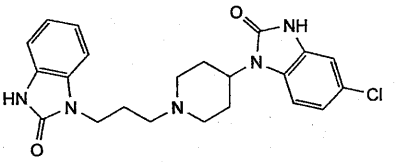
Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 2 g in sufficient *water* to produce 100 mL. Transfer 25 mL to a separating funnel and add 25 mL of *chloroform*, 10 mL of 0.1M *sodium hydroxide* and 10 mL of a freshly prepared 5% w/v solution of *potassium iodide*. Shake well, allow to separate and discard the chloroform layer. Wash the aqueous layer with three 10-mL quantities of *chloroform* and discard the chloroform solutions. Add 40 mL of *hydrochloric acid*, allow to cool and titrate with *0.05M potassium iodate VS* until the deep brown colour is discharged. Add 2 mL of *chloroform* and continue the titration, shaking vigorously, until the chloroform layer no longer changes colour. Carry out a blank titration on a mixture of 10 mL of the freshly prepared potassium iodide solution, 20 mL of *water* and 40 mL of *hydrochloric acid*. The difference between the titrations represents the amount of potassium iodate required. Each mL of *0.05M potassium iodate VS* is equivalent to 41.45 mg of $C_{22}H_{40}BrNO$.

Domperidone

(Ph. Eur. monograph 1009)



$C_{22}H_{24}ClN_5O_2$ 425.9 *57808-66-9*

Action and use

Peripheral dopamine receptor antagonist; antiemetic.

Ph Eur

DEFINITION

5-Chloro-1-[1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, soluble in dimethylformamide, slightly soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 244 °C to 248 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *domperidone CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *domperidone CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *domperidone CRS* and 20 mg of *droperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *ammonium acetate solution R, dioxan R, methanol R* (20:40:40 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear; examine in daylight.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives the reaction of non-nitrogen substituted barbiturates (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 0.20 g in *dimethylformamide R* and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.10 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *domperidone CRS* and 15.0 mg of *droperidol CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethylformamide R*. Dilute 5.0 mL of this solution to 20.0 mL with *dimethylformamide R*.

Column:
- *size*: *l* = 0.1 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
- *mobile phase A*: 5 g/L solution of *ammonium acetate R*;
- *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	70 → 0	30 → 100
10 - 12	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Relative retention With reference to domperidone (retention time = about 6.5 min): impurity A = about 0.4; impurity B = about 0.65; impurity C = about 0.7; droperidol = about 1.1; impurity D = about 1.15; impurity E = about 1.2; impurity F = about 1.3.

System suitability Reference solution (a):
- *resolution*: minimum 2.0 between the peaks due to domperidone and droperidol.

Limits:
- *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
- *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid* until the colour changes from orange-yellow to green using 0.2 mL of *naphtholbenzein solution R* as indicator.

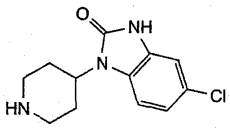
1 mL of *0.1 M perchloric acid* is equivalent to 42.59 mg of $C_{22}H_{24}ClN_5O_2$.

STORAGE

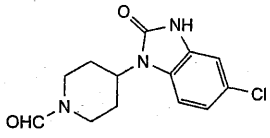
Protected from light.

IMPURITIES

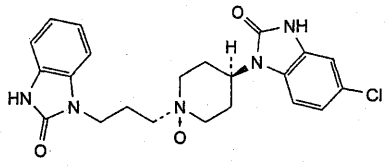
Specified impurities *A, B, C, D, E, F.*



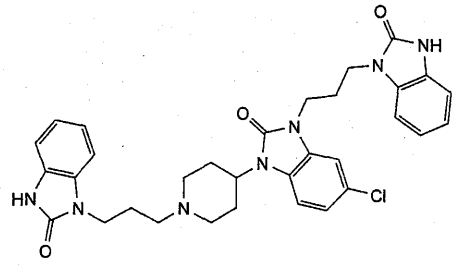
A. 5-chloro-1-(piperidin-4-yl)-1,3-dihydro-2*H*-benzimidazol-2-one,



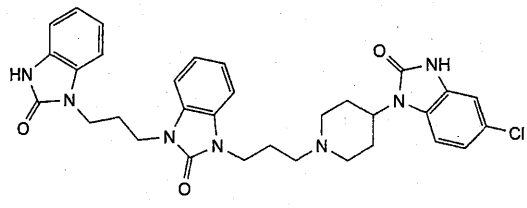
B. 4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-1-formylpiperidine,



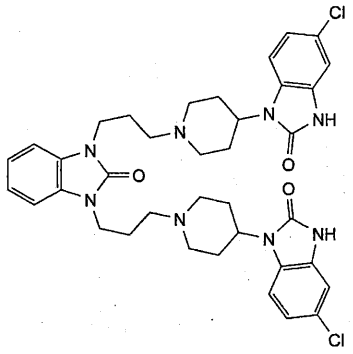
C. *cis*-4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)propyl] piperidine 1-oxide,



D. 5-chloro-3-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]-1-[1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one,



E. 1-[3-[4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)piperidin-1-yl]propyl]-3-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)propyl]-1,3-dihydro-2*H*-benzimidazol-2-one,

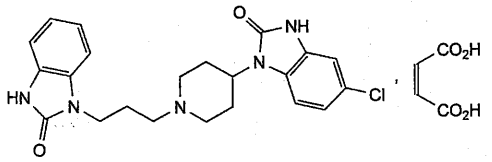


F. 1,3-bis[3-[4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)piperidin-1-yl]propyl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Ph Eur

Domperidone Maleate

(Ph. Eur. monograph 1008)



$C_{26}H_{28}ClN_5O_6$ 542.0 *83898-65-1*

Action and use
Peripheral dopamine receptor antagonist; antiemetic.

Preparation
Domperidone Tablets

Ph Eur

DEFINITION
5-Chloro-1-[1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one hydrogen (*Z*)-butenedioate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, sparingly soluble in dimethylformamide, slightly soluble in methanol, very slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *domperidone maleate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *2-propanol R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *domperidone maleate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *domperidone maleate CRS* and 20 mg of *droperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *ammonium acetate solution R, dioxan R, methanol R* (20:40:40 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear. Examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Triturate 0.1 g with a mixture of 1 mL of *strong sodium hydroxide solution R* and 3 mL of *water R*. Shake with 3 quantities, each of 5 mL, of *ether R*. To 0.1 mL of the aqueous layer add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min. No colour develops. To the remainder of the aqueous layer add 2 mL of *bromine solution R*. Heat on a water-bath for 15 min and then heat to boiling. Cool. To 0.1 mL of this solution add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min. A violet colour develops.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 0.20 g in *dimethylformamide R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.10 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *domperidone maleate CRS* and 15.0 mg of *droperidol CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethylformamide R*. Dilute 5.0 mL of this solution to 20.0 mL with *dimethylformamide R*.

Blank solution *dimethylformamide R.*

Column:
— *size: l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 5 g/L solution of *ammonium acetate R*;
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	70 → 0	30 → 100
10 - 12	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Equilibration With *methanol R* for at least 30 min and then with the mobile phase at the initial composition for at least 5 min.

Injection 10 µL.

Retention time Domperidone = about 6.5 min; droperidol = about 7 min.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to domperidone and droperidol; if necessary, adjust the concentration of methanol in the mobile phase or adjust the time programme for the linear gradient.

Limits:
— *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to maleic acid at the beginning of the chromatogram.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 50 mL of *anhydrous acetic acid R*. Using 0.2 mL of *naphtholbenzein solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from orange-yellow to green.

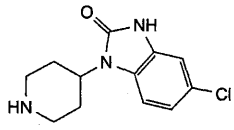
1 mL of *0.1 M perchloric acid* is equivalent to 54.20 mg of $C_{26}H_{28}ClN_5O_6$.

STORAGE

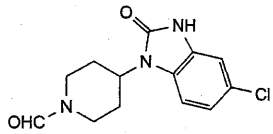
Protected from light.

IMPURITIES

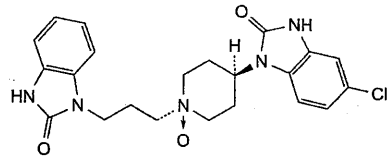
Specified impurities *A, B, C, D, E, F.*



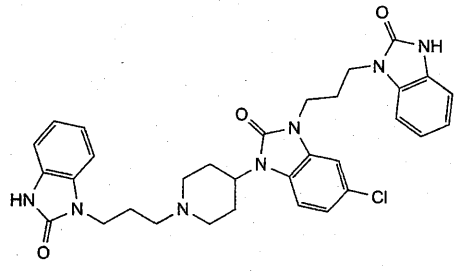
A. 5-chloro-1-(piperidin-4-yl)-1,3-dihydro-2*H*-benzimidazol-2-one,



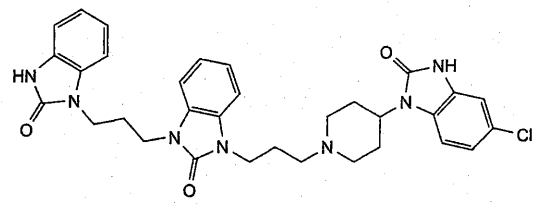
B. 4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-1-formylpiperidine,



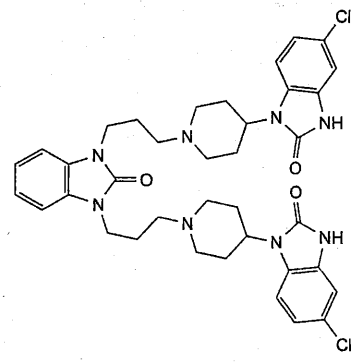
C. *cis*-4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)propyl] piperidine 1-oxide,



D. 5-chloro-3-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]-1-[1-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl) propyl]piperidin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one,



E. 1-[3-[4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)piperidin-1-yl]propyl]-3-[3-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)propyl]-1,3-dihydro-2*H*-benzimidazol-2-one,

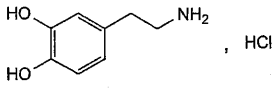


F. 1,3-bis[3-[4-(5-chloro-2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)piperidin-1-yl]propyl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Ph Eur

Dopamine Hydrochloride

(Ph. Eur. monograph 0664)



$C_8H_{12}ClNO_2$ 189.6 *62-31-7*

Action and use
Dopamine receptor antagonist; $beta_1$-adrenoceptor agonist; alpha-adrenoceptor agonist.

Preparation
Dopamine Infusion

Ph Eur

DEFINITION
4-(2-Aminoethyl)benzene-1,2-diol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent), sparingly soluble in acetone and in methylene chloride.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 40.0 mg in *0.1 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of this solution to 100.0 mL with *0.1 M hydrochloric acid.*

Spectral range 230-350 nm.

Absorption maximum At 280 nm.

Specific absorbance at the absorption maximum 136 to 150.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dopamine hydrochloride CRS.*

C. Dissolve about 5 mg in a mixture of 5 mL of *1 M hydrochloric acid* and 5 mL of *water R*. Add 0.1 mL of *sodium nitrite solution R* containing 100 g/L of *ammonium molybdate R*. A yellow colour develops which becomes red on the addition of *strong sodium hydroxide solution R.*

D. Dissolve about 2 mg in 2 mL of *water R* and add 0.2 mL of *ferric chloride solution R2*. A green colour develops which changes to bluish-violet on the addition of 0.1 g of *hexamethylenetetramine R.*

E. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_6 or Y_6 (*2.2.2, Method II*).

Dissolve 0.4 g in *water R* and dilute to 10 mL with the same solvent.

Acidity or alkalinity
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Add 0.1 mL of *methyl red solution R* and 0.75 mL of *0.01 M sodium hydroxide.* The solution is yellow. Add 1.5 mL of *0.01 M hydrochloric acid.* The solution is red.

Related substances
Liquid chromatography (*2.2.29*). *Protect the solutions from light.*

Buffer solution Dissolve 21 g of *citric acid monohydrate R* in 200 mL of *1 M sodium hydroxide* and dilute to 1000 mL with *water R*. To 600 mL of this solution add 400 mL of *0.1 M hydrochloric acid.*

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 25 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 10 mg of *3-O-methyldopamine hydrochloride R* (impurity B) and 10 mg of *4-O-methyldopamine hydrochloride R* (impurity A) in mobile phase A and dilute to 100 mL with mobile phase A. Dilute 6 mL of this solution to 25 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (4 µm).

Mobile phase:
— *mobile phase A*: dissolve 1.08 g of *sodium octanesulfonate R* in 880 mL of the buffer solution and add 50 mL of *methanol R* and 70 mL of *acetonitrile R*;
— *mobile phase B*: dissolve 1.08 g of *sodium octanesulfonate R* in 700 mL of the buffer solution and add 100 mL of *methanol R* and 200 mL of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90	10
5 - 20	90 → 40	10 → 60
20 - 25	40	60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Retention time Dopamine = about 5 min.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurities B and A.

Limits:
— *unspecificied impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout the titration and stop the titration immediately after the end-point has been reached.

Dissolve 0.150 g in 10 mL of *anhydrous formic acid R*. Add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

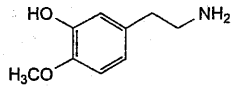
1 mL of *0.1 M perchloric acid* is equivalent to 18.96 mg of $C_8H_{12}ClNO_2$.

STORAGE

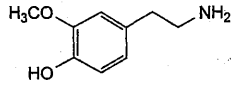
In an airtight container, under nitrogen, protected from light.

IMPURITIES

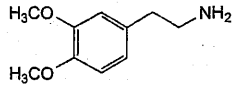
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



A. 5-(2-aminoethyl)-2-methoxyphenol (4-*O*-methyldopamine),



B. 4-(2-aminoethyl)-2-methoxyphenol (3-*O*-methyldopamine),

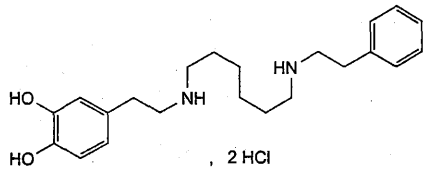


C. 2-(3,4-dimethoxyphenyl)ethanamine.

Ph Eur

Dopexamine Hydrochloride

(Dopexamine Dihydrochloride, Ph. Eur. monograph 1748)



$C_{22}H_{34}Cl_2N_2O_2$ 429.4 *86484-91-5*

Action and use

Beta$_2$-adrenoceptor agonist; bronchodilator.

Ph Eur

DEFINITION

4-[2-[[6-[(2-Phenylethyl)amino]hexyl]amino]ethyl] benzene1,2-diol dihydrochloride.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, sparingly soluble in ethanol (96 per cent) and in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dopexamine dihydrochloride CRS.*

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 0.10 g in *0.1 M hydrochloric acid* and dilute to 10 mL with the same acid.

pH (*2.2.3*)

3.7 to 5.7.

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *dopexamine impurity B CRS* in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *dopexamine impurity F CRS* in mobile phase A and dilute to 100 mL with mobile phase A.

Column:
- *size*: l = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 45 °C.

Mobile phase:
- *mobile phase A*: mix 5 volumes of *buffer solution pH 2.5 R* and 95 volumes of *water R*;
- *mobile phase B*: mix 5 volumes of *buffer solution pH 2.5 R* and 95 volumes of a 60 per cent *V/V* solution of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	81 → 77	19 → 23
10 - 25	77 → 50	23 → 50
25 - 30	50	50

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Preconditioning of the column Rinse for 5 min with a mixture of 19 volumes of mobile phase B and 81 volumes of mobile phase A.

Injection 20 µL.

Relative retention With reference to dopexamine (retention time = about 5 min): impurity A = about 0.5; impurity B = about 2.0; impurity C = about 2.3; impurity D = about 2.8; impurity E = about 2.9; impurity F = about 3.0; impurity I = about 3.6; impurity J = about 5.0; impurity K = about 5.9.

System suitability Reference solution (b):
— *resolution*: minimum 2 between the peaks due to dopexamine and impurity B.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.4; impurity F = 0.7;
— *impurities A, B, C, D, E, F, I, K*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity J

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Detection Spectrophotometer at 210 nm.

Limit:
— *impurity J*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent).

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)

Less than 10 IU/mg.

ASSAY

Carry out the titration immediately after preparation of the test solution [In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.]

Dissolve 0.150 g in 10 mL of *anhydrous formic acid R*. Add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 21.47 mg of $C_{22}H_{34}Cl_2N_2O_2$.

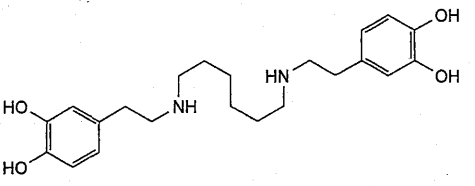
STORAGE

Protected from light.

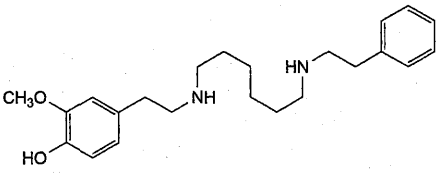
IMPURITIES

Specified impurities *A, B, C, D, E, F, I, J, K.*

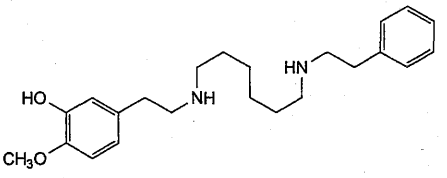
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, H.*



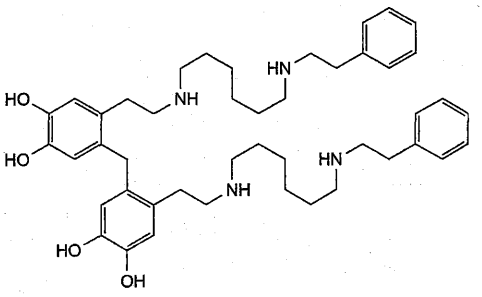
A. 4,4′-[hexane-1,6-diylbis(iminoethylene)]dibenzene-1,2-diol,



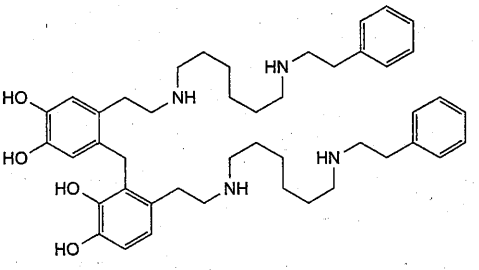
B. 2-methoxy-4-[2-[[6-[(2-phenylethyl)amino]hexyl]amino]ethyl]phenol,



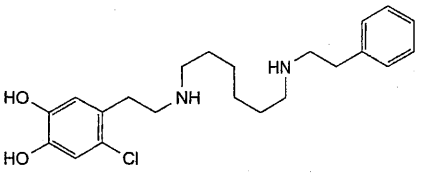
C. 2-methoxy-5-[2-[[6-[(2-phenylethyl)amino]hexyl]amino]ethyl]phenol,



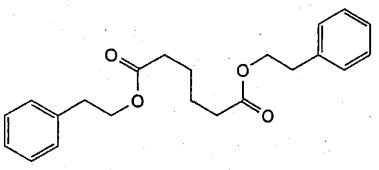
D. 4,4′-methylenebis[5-[2-[[6-[(2-phenylethyl)amino]hexyl]amino]ethyl]benzene-1,2-diol],



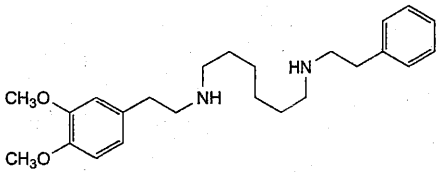
E. 3-[4,5-dihydroxy-2-[2-[[6-[(2-phenylethyl)amino]hexyl]amino]ethyl]benzyl]-4-[2-[[6-[(2-phenylethyl)amino]hexyl]amino]ethyl]benzene-1,2-diol,



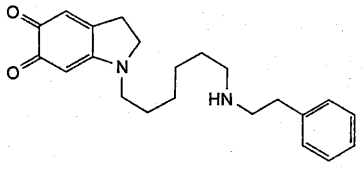
F. 4-chloro-5-[2-[[6-[(2-phenylethyl)amino]hexyl] amino]ethyl]benzene-1,2-diol,



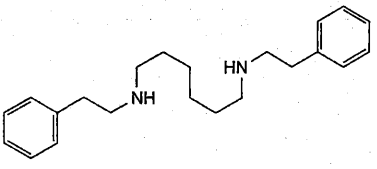
G. bis(2-phenylethyl) hexanedioate,



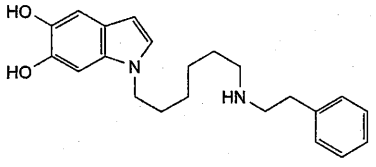
H. *N*-[2-(3,4-dimethoxyphenyl)ethyl]-*N'*-(2-phenylethyl) hexane-1,6-diamine,



I. 1-[6-[(2-phenylethyl)amino]hexyl]-2,3-dihydro-1*H*-indole5,6-dione (dopexamine aminochrome),



J. *N*,*N'*-bis(2-phenylethyl)hexane-1,6-diamine,

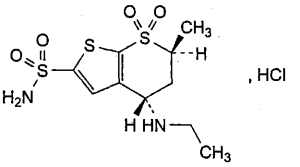


K. 1-[6-[(2-phenylethyl)amino]hexyl]-1*H*-indole-5,6-diol.

Ph Eur

Dorzolamide Hydrochloride

(Ph. Eur. monograph 2359)



$C_{10}H_{17}ClN_2O_4S_3$ 360.9 *130693-82-2*

Action and use
Carbonic anhydrase inhibitor; treatment of glaucoma and ocular hypertension.

Preparations
Dorzolamide Eye Drops
Dorzolamide and Timolol Eye Drops

Ph Eur

DEFINITION
(4*S*,6*S*)-4-(Ethylamino)-6-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-2-sulfonamide 7,7-dioxide hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, slightly soluble in methanol, very slightly soluble in anhydrous ethanol.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dorzolamide hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. It complies with the test for impurity A (see Tests).

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Impurity A
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R*, *glacial acetic acid R*, *1,1-dimethylethyl methyl ether R* (3:10:87 *V/V/V*).

Test solution In a centrifuge tube, dissolve 20.0 mg of the substance to be examined in 4 mL of *dilute ammonia R4*, add 4 mL of *ethyl acetate R*, and mix. Separate the organic layer and transfer it to a separate centrifuge tube. Add 4 mL of *ethyl acetate R* to the aqueous layer, mix, separate the organic layer, and combine it with the 1st extract. Evaporate the combined organic layers to dryness in a water-bath at 50 °C under a stream of *nitrogen R*. Dissolve the residue in 3 mL of *acetonitrile R*, add 0.06 mL of (*S*)-(−)-α-methylbenzyl isocyanate R, and heat in a water-bath at 50 °C for 5 min. Evaporate to dryness in a water-bath at 50 °C under a stream of *nitrogen R*. Dissolve the residue in 10 mL of the solvent mixture.

Reference solution In a centrifuge tube, dissolve 18.0 mg of *dorzolamide hydrochloride CRS* and 2.0 mg of *dorzolamide impurity A CRS* in 4 mL of *dilute ammonia R4*, and proceed

as indicated for the test solution beginning with "add 4 mL of *ethyl acetate R*, and mix".

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *water R, acetonitrile R, heptane R, 1,1-dimethylethyl methyl ether R* (0.2:2:35:63 *V/V/V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 3 times the retention time of dorzolamide.

Relative retention With reference to dorzolamide (retention time = about 10 min): impurity A = about 1.4.

System suitability Reference solution:
— *resolution*: minimum 4.0 between the peaks due to dorzolamide and impurity A.

Calculate the percentage content of impurity A using the following expression:

$$\frac{A}{A+B} \times 100$$

A = area of the peak due to impurity A in the chromatogram obtained with the test solution;
B = area of the peak due to dorzolamide in the chromatogram obtained with the test solution.

Limit:
— *impurity A*: maximum 0.5 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 30.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 2 mg of *dorzolamide for system suitability CRS* (containing impurity C) in 2 mL of mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: mix 65 mL of *acetonitrile R* and 935 mL of a 3.7 g/L solution of *potassium dihydrogen phosphate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100	0
15 - 30	100 → 50	0 → 50
30 - 37	50 → 100	50 → 0
37 - 44	100	0

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *dorzolamide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to dorzolamide (retention time = about 11 min): impurity C = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity C and dorzolamide.

Limits:
— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

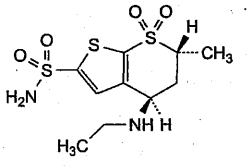
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*, using sonication if necessary. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 1st and the 3rd points of inflexion.

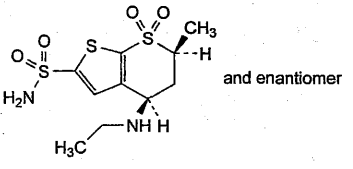
1 mL of *0.1 M sodium hydroxide* is equivalent to 18.05 mg of $C_{10}H_{17}N_2O_4S_3Cl$.

IMPURITIES
Specified impurities *A, C.*

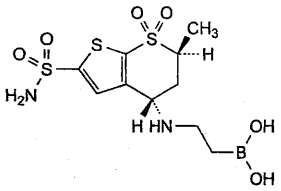
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D.*



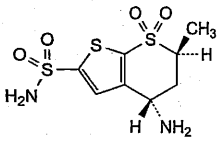
A. (4*R*,6*R*)-4-(ethylamino)-6-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-2-sulfonamide 7,7-dioxide,



B. (4*RS*,6*SR*)-4-(ethylamino)-6-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-2-sulfonamide 7,7-dioxide,



C. [2-[[(4*S*,6*S*)-6-methyl-7,7-dioxo-2-sulfamoyl-4,5,6,7-tetrahydro-7λ^6-thieno[2,3-*b*]thiopyran-4-yl]amino]ethyl] boronic acid,

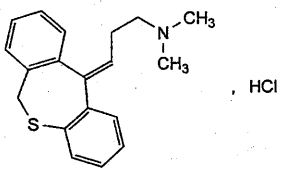


D. (4*S*,6*S*)-4-amino-6-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*] thiopyran-2-sulfonamide 7,7-dioxide.

Ph Eur

Dosulepin Hydrochloride

(Ph. Eur. monograph 1314)



$C_{19}H_{22}ClNS$ 331.9 *897-15-4*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparations
Dosulepin Capsules
Dosulepin Oral Solution
Dosulepin Tablets

Ph Eur

DEFINITION
(*E*)-3-(Dibenzo[*b*,*e*]thiepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or faintly yellow, crystalline powder.

Solubility
Freely soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 25.0 mg in a 1 g/L solution of *hydrochloric acid R* in *methanol R* and dilute to 100.0 mL with the same solution. Dilute 2.0 mL to 50.0 mL with a 1 g/L solution of *hydrochloric acid R* in *methanol R*.

Spectral range 220-350 nm.

Absorption maxima 231 nm and 306 nm.

Shoulder About 260 nm.

Specific absorbance at the absorption maximum at 231 nm 660 to 730.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dosulepin hydrochloride CRS.*

C. Dissolve about 1 mg in 5 mL of *sulfuric acid R*. A dark red colour is produced.

D. It gives reaction (b) of chlorides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 1 g in *water R* and dilute to 20 mL with the same solvent.

pH (*2.2.3*)
4.2 to 5.2.

Dissolve 1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 50.0 mg of the substance to be examined in 5 mL of *methanol R* and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 12.5 mg of *dosulepin impurity A CRS* in 5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 0.5 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *dosulepin for system suitability CRS* (containing impurity E) in 5 mL of *methanol R* and dilute to 20 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped cyanosilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase 0.83 per cent *V/V* solution of *perchloric acid R*, *propanol R*, *methanol R1*, *water for chromatography R* (1:10:30:60 *V/V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 229 nm.

Injection 5 µL.

Run time 2.5 times the retention time of dosulepin.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *dosulepin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity E.

Relative retention With reference to dosulepin (retention time = about 14 min): impurity A = about 0.3; impurity E = about 0.92.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 4, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to dosulepin.

Limits:
— *impurity E*: not more than 5 per cent of the sum of the areas of the peak due to impurity E and the principal peak in the chromatogram obtained with the test solution (5 per cent);
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than E*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

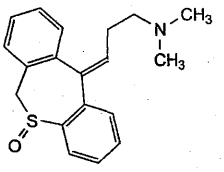
ASSAY
Dissolve 0.250 g in a mixture of 5 mL of *anhydrous acetic acid R* and 35 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 33.19 mg of $C_{19}H_{22}ClNS$.

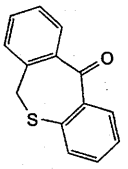
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, E.*

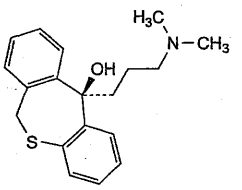
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D.*



A. (*E*)-3-(5-oxo-5λ⁴-dibenzo[*b*,*e*]thiepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine,

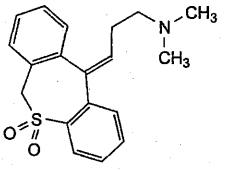


B. dibenzo[*b*,*e*]thiepin-11(6*H*)-one,

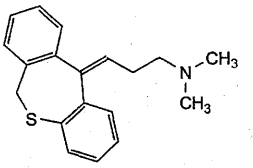


and enantiomer

C. (11*RS*)-11-[3-(dimethylamino)propyl]-6,11-dihydrodibenzo[*b*,*e*]thiepin-11-ol,



D. (*E*)-3-(5,5-dioxo-5λ⁶-dibenzo[*b*,*e*]thiepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine,

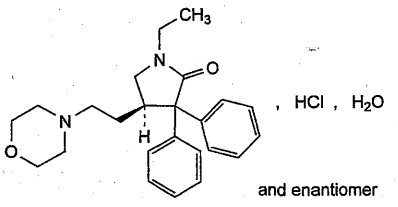


E. (*Z*)-3-(dibenzo[*b*,*e*]thiepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine.

Ph Eur

Doxapram Hydrochloride

(Ph. Eur. monograph 1201)



, HCl , H_2O

and enantiomer

$C_{24}H_{31}ClN_2O_2,H_2O$ 433.0 *7081-53-0*

Action and use
Respiratory stimulant.

Preparation
Doxapram Injection

Ph Eur

DEFINITION
(4*RS*)-1-Ethyl-4-[2-(morpholin-4-yl)ethyl]-3,3-diphenylpyrrolidin-2-one hydrochloride monohydrate.

Content
98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *doxapram hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *doxapram hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase Solution of *ammonia R* containing 17 g/L of NH_3, *2-propanol R*, *2-methylpropanol R* (10:10:80 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *dilute potassium iodobismuthate solution R* and examine immediately.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.000 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dilute 10 mL of solution S to 25 mL with *water R.*

pH (*2.2.3*)

3.5 to 5.0.

Dilute 5 mL of solution S to 25 mL with *carbon dioxide-free water R.*

Optical rotation (*2.2.7*)

−0.10° to + 0.10°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 5.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *doxapram impurity B CRS* in the mobile phase and dilute to 5.0 mL with the mobile phase. To 1.0 mL of the solution, add 1.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Column:

— *size: l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 50 volumes of *acetonitrile R* and 50 volumes of a 0.82 g/L solution of *sodium acetate R* adjusted to pH 4.5 with *glacial acetic acid R.*

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Run time 4 times the retention time of doxapram.

Retention time Doxapram = about 6 min.

System suitability Reference solution (c):

— *resolution*: minimum 3.0 between the peaks due to doxapram and impurity B.

Limits:

— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),

— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent),

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

3.0 per cent to 4.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

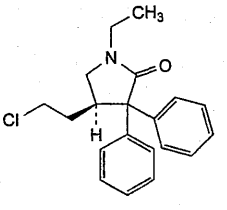
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 10 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R.* Carry out a potentiometric titration (*2.2.20*) using *0.1 M sodium hydroxide.* Read the volume added between the 2 points of inflexion.

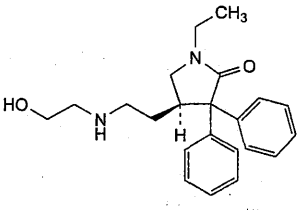
1 mL of *0.1 M sodium hydroxide* is equivalent to 41.50 mg of $C_{24}H_{31}ClN_2O_2$.

IMPURITIES



and enantiomer

A. (4*RS*)-4-(2-chloroethyl)-1-ethyl-3,3-diphenylpyrrolidin-2-one,



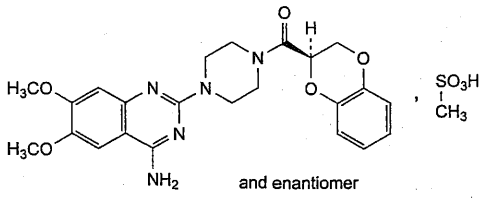
and enantiomer

B. (4*RS*)-1-ethyl-4-[2-[(2-hydroxyethyl)amino]ethyl]-3,3-diphenylpyrrolidin-2-one.

Ph Eur

Doxazosin Mesilate

(Ph. Eur. monograph 2125)



$C_{24}H_{29}N_5O_8S$ 547.6 *77883-43-3*

Action and use
Alpha$_1$-adrenoceptor antagonist.

Ph Eur

DEFINITION

[4-(4-Amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl][(2*RS*)-2,3-dihydro-1,4-benzodioxin-2-yl]methanone methanesulfonate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in doxazosin mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, soluble in a mixture of 15 volumes of water and 35 volumes of tetrahydrofuran, slightly soluble in methanol, practically insoluble in acetone.

It shows polymorphism (*5.9*); some forms may be hygroscopic.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *doxazosin mesilate CRS*.

If the spectra obtained in the solid state show differences, mix 1 part of the substance to be examined and 1 part of the reference substance separately with 10 parts of *anhydrous ethanol R* and heat to boiling. Continue heating the suspension under a reflux condenser for about 3 h. Cool and filter. Record new spectra using the previously dried residues on the filters.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 1.0 g in a mixture of 15 mL of *water R* and 35 mL of *tetrahydrofuran R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in 5 mL of mobile phase B, adding *water R*, and dilute to 50.0 mL with *water R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 25.0 mg of *doxazosin mesilate CRS* in 5 mL of mobile phase B, adding *water R*, and dilute to 50.0 mL with *water R*.

Reference solution (c) Dissolve 5 mg of *doxazosin impurity D CRS* and 5 mg of *doxazosin impurity F CRS* in 5 mL of mobile phase B, adding *water R*, and dilute to 50 mL with *water R*. Dilute 1 mL of this solution to 5 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated end-capped octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: 1.5 g/L solution of *phosphoric acid R*;
— *mobile phase B*: 1.5 g/L solution of *phosphoric acid R* in *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90	10
5 - 40	90 → 50	10 → 50
40 - 45	50	50

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D and F.

Relative retention With reference to doxazosin (retention time = about 30 min): impurity D = about 0.5; impurity F = about 0.6.

System suitability Reference solution (c):
— *resolution*: minimum 4.5 between the peaks due to impurities D and F.

Limits:
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 1.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

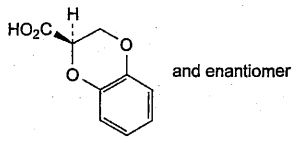
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

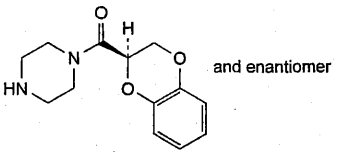
Calculate the percentage content of $C_{24}H_{29}N_5O_8S$ taking into account the assigned content of *doxazosin mesilate CRS*.

STORAGE
In an airtight container.

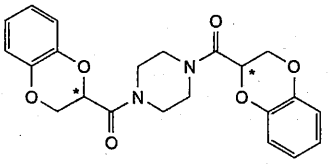
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H.*



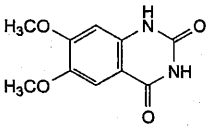
A. (2*RS*)-2,3-dihydro-1,4-benzodioxine-2-carboxylic acid,



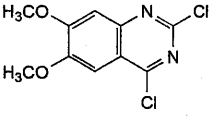
B. [(2*RS*)-2,3-dihydro-1,4-benzodioxin-2-yl](piperazin-1-yl) methanone,



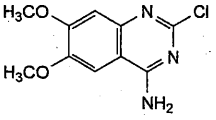
C. (piperazine-1,4-diyl)bis[[(2Ξ)-2,3-dihydro-1,4-benzodioxin-2-yl]methanone],



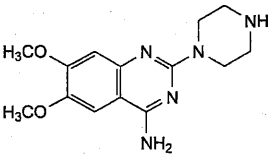
D. 6,7-dimethoxyquinazoline-2,4(1*H*,3*H*)-dione,



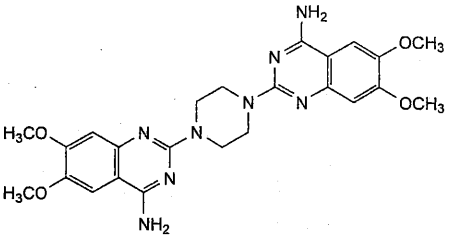
E. 2,4-dichloro-6,7-dimethoxyquinazoline,



F. 2-chloro-6,7-dimethoxyquinazolin-4-amine,



G. 6,7-dimethoxy-2-(piperazin-1-yl)quinazolin-4-amine,

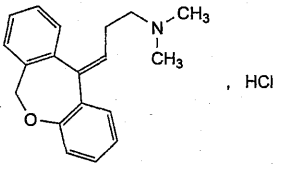


H. 2,2′-(piperazine-1,4-diyl)bis(6,7-dimethoxyquinazolin-4-amine).

Ph Eur

Doxepin Hydrochloride

(Ph. Eur. monograph 1096)



$C_{19}H_{22}ClNO$ 315.8 *1229-29-4*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparation
Doxepin Capsules

Ph Eur

DEFINITION
(*E*)-3-(Dibenzo[*b*,*e*]oxepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
98.0 per cent to 101.0 per cent of $C_{19}H_{22}ClNO$ (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: C, E.

Second identification: A, B, D, E.

A. Melting point (*2.2.14*): 185 °C to 191 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in a 1 g/L solution of *hydrochloric acid R* in *methanol R* and dilute to 100.0 mL with the same acid solution. Dilute 5.0 mL to 50.0 mL with a 1 g/L solution of *hydrochloric acid R* in *methanol R*.

Spectral range 230-350 nm.

Absorption maximum At 297 nm.

Specific absorbance at the absorption maximum 128 to 142.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *doxepin hydrochloride CRS.*

D. Dissolve about 5 mg in 2 mL of *sulfuric acid R*. A dark red colour is produced.

E. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.5 g in *carbon dioxide-free water R* and dilute to 30 mL with the same solvent.

Appearance of solution

Dilute 10 mL of solution S to 25 mL with *water R*. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity

To 10 mL of solution S add 0.1 mL of *methyl red solution R*. Not more than 0.1 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to yellow.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect them from light.*

Phosphate buffer solution Dissolve 1.42 g of *anhydrous disodium hydrogen phosphate R* in *water R*, adjust to pH 7.7 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture Mix 1 volume of *1 M sodium hydroxide* and 250 volumes of the mobile phase.

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *doxepin for system suitability CRS* (containing impurities A, B and C) in 1.0 mL of mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase *acetonitrile R1*, phosphate buffer solution, *methanol R1* (20:30:50 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 1.5 times the retention time of doxepin.

Identification of impurities Use the chromatogram supplied with *doxepin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to doxepin (retention time = about 18 min): impurity A = about 0.5; impurity C = about 0.6; impurity B = about 0.7; the peak due to doxepin might show a shoulder caused by the (*Z*)-isomer (impurity D).

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurities A and C, and minimum 1.5 between the peaks due to impurities C and B;

—the chromatogram obtained is similar to the chromatogram supplied with *doxepin for system suitability CRS*.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity B by 1.7;
- *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- *impurity C*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

(*Z*)-Isomer

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.12 m, Ø = 4 mm;
- *stationary phase*: spherical *octylsilyl silica gel for chromatography R* (5 µm) with a specific surface area of 220 m^2/g and a pore size of 80 nm;
- *temperature*: 50 °C.

Mobile phase Mix 30 volumes of *methanol R* and 70 volumes of a 30 g/L solution of *sodium dihydrogen phosphate R* previously adjusted to pH 2.5 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to the (*E*)-isomer (1st peak) and to the (*Z*)-isomer (2nd peak).

Results:
- calculate the ratio of the area of the peak due to the (*E*)-isomer to the area of the peak due to the (*Z*)-isomer: this ratio is 4.4 to 6.7 (13.0 per cent to 18.5 per cent of the (*Z*)-isomer).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5 mL of *anhydrous acetic acid R* and 35 mL of *acetic anhydride R*. Using 0.2 mL of *crystal violet solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from blue to green.

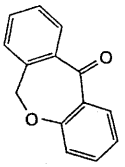
1 mL of *0.1 M perchloric acid* is equivalent to 31.58 mg of $C_{19}H_{22}ClNO$.

STORAGE

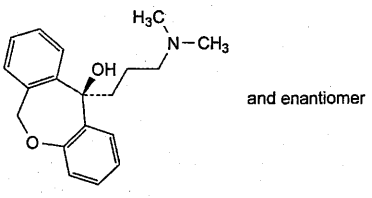
Protected from light.

IMPURITIES

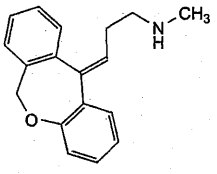
Specified impurities *A, B, C, D.*



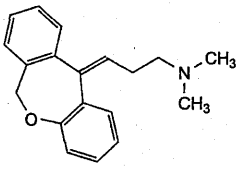
A. dibenzo[*b*,*e*]oxepin-11(6*H*)-one (doxepinone),



B. (11*RS*)-11-[3-(dimethylamino)propyl]-6,11-dihydrodibenzo[*b*,*e*]oxepin-11-ol (doxepinol),



C. (*E*)-3-(dibenzo[*b*,*e*]oxepin-11(6*H*)-ylidene)-*N*-methylpropan-1-amine (desmethyldoxepin),

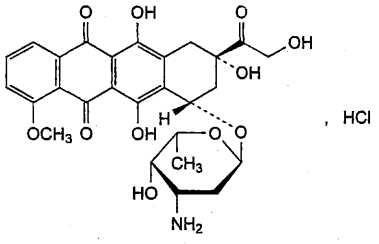


D. (*Z*)-3-(dibenzo[*b*,*e*]oxepin-11(6*H*)-ylidene)-*N*,*N*-dimethylpropan-1-amine.

Ph Eur

Doxorubicin Hydrochloride

(Ph. Eur. monograph 0714)



$C_{27}H_{30}ClNO_{11}$ 580.0 *25316-40-9*

Action and use
Anthracycline antibiotic; cytotoxic.

Preparations
Sterile Doxorubicin Concentrate
Doxorubicin for Infusion

Ph Eur

DEFINITION
(8*S*,10*S*)-10-[(3-Amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione hydrochloride.

Substance produced by certain strains of *Streptomyces coeruleorubidus* or *Streptomyces peucetius* or obtained by any other means.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Orange-red, crystalline powder, hygroscopic.

Solubility
Soluble in water, slightly soluble in methanol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *doxorubicin hydrochloride CRS.*

B. Dissolve about 10 mg in 0.5 mL of *nitric acid R*, add 0.5 mL of *water R* and heat over a flame for 2 min. Allow to cool and add 0.5 mL of *silver nitrate solution R1*. A white precipitate is formed.

TESTS

pH (*2.2.3*)
4.0 to 5.5.

Dissolve 50 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *doxorubicin hydrochloride CRS* and 10 mg of *epirubicin hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 50.0 mg of *doxorubicin hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm,
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix equal volumes of *acetonitrile R* and a solution containing 2.88 g/L of *sodium laurilsulfate R* and 2.25 g/L of *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 5 µL; inject test solution (a) and reference solutions (a) and (b).

Run time 3.5 times the retention time of doxorubicin.

Retention time Doxorubicin = about 8 min.

System suitability Reference solution (a):
— *resolution*: minimum of 2.0 between the peaks due to doxorubicin and to epirubicin.

Limits:
— *any impurity*: not more than the area of the peak due to doxorubicin in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *disregard limit*: 0.1 times the area of the peak due to doxorubicin in the chromatogram obtained with reference solution (b) (0.05 per cent).

Ethanol (*2.4.24, System B*)
Maximum 1.0 per cent.

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)
Less than 2.2 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

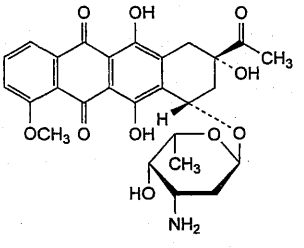
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances.

Injection Test solution (b) and reference solution (c).

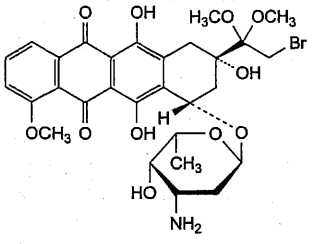
Calculate the percentage content of $C_{27}H_{30}ClNO_{11}$.

STORAGE
In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

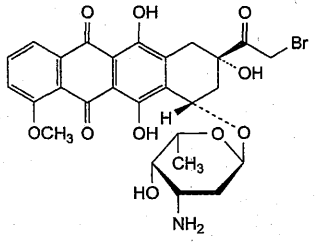
IMPURITIES



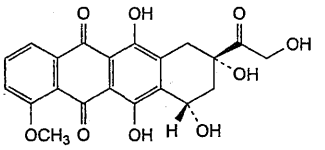
A. (8*S*,10*S*)-8-acetyl-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (daunorubicin),



B. (8*S*,10*S*)-10[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-8-(2-bromo-1,1-dimethoxyethyl)-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione,



C. (8*S*,10*S*)-10[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-8-(bromoacetyl)-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione,

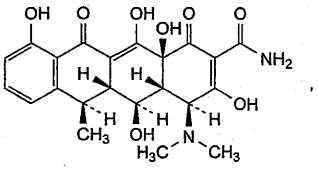


D. (8*S*,10*S*)-6,8,10,11-tetrahydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (doxorubicin aglycone, doxorubicinone).

Ph Eur

Doxycycline Hyclate

(Ph. Eur. monograph 0272)



, HCl , ½ C_2H_5-OH , ½ H_2O

$(C_{22}H_{25}ClN_2O_8)_2,½C_2H_6O,½H_2O$ 512.9 *24390-14-5*

Action and use
Tetracycline antibacterial.

Preparations
Doxycycline Capsules
Doxycycline Tablets

Ph Eur

DEFINITION
(4*S*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-4-(Dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide hydrochloride hemiethanol hemihydrate.

Substance obtained from oxytetracycline or metacycline or by any other means.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent of $C_{22}H_{25}ClN_2O_8$ (anhydrous and ethanol-free substance).

CHARACTERS
Appearance
Yellow, hygroscopic, crystalline powder.

Solubility
Freely soluble in water and in methanol, sparingly soluble in ethanol (96 per cent). It dissolves in solutions of alkali hydroxides and carbonates.

IDENTIFICATION
A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of *sulfuric acid R*. A yellow colour develops.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
pH (*2.2.3*)
2.0 to 3.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Absorbance (*2.2.25*)
Maximum 0.07 at 490 nm (anhydrous and ethanol-free substance).

Dissolve 0.10 g in a mixture of 1 volume of a 103 g/L solution of *hydrochloric acid R* and 99 volumes of *methanol R* and dilute to 10.0 mL with the same mixture of solvents. Carry out the measurement within 1 h of preparing the solution.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Suspend 111.6 g of *sodium edetate R* in 900 mL of *water R*. Adjust to pH 7.0 with *concentrated ammonia R* to achieve complete dissolution, then dilute to 1 L with *water R*.

Test solution Dissolve 20.0 mg of the substance to be examined in a 1 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same solution.

Reference solution (a) Dissolve 20.0 mg of *doxycycline hyclate CRS* in a 1 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same solution.

Reference solution (b) Dissolve 5 mg of *doxycycline for system suitability CRS* (containing impurities A, B, C and F) in a 1 g/L solution of *hydrochloric acid R* and dilute to 10 mL with the same solution.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with a 1 g/L solution of *hydrochloric acid R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm);
— *temperature*: 35 °C.

Mobile phase Mix 13 volumes of *acetonitrile R*, 17 volumes of *water R*, 35 volumes of a 67.9 g/L solution of *tetrabutylammonium hydrogen sulfate R* previously adjusted to pH 7.0 with *concentrated ammonia R* and 35 volumes of solution A.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of doxycycline.

Identification of impurities Use the chromatogram supplied with *doxycycline for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and F.

Relative retention With reference to doxycycline (retention time = about 21 min): impurity C = about 0.4; impurity A = about 0.7; impurity B = about 0.8; impurity F = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities A and B; minimum 2.0 between the peaks due to impurity B and doxycycline.

Calculation of percentage contents:
— for each impurity, use the concentration of doxycycline in reference solution (c).

Limits:
— *impurity A*: maximum 2.0 per cent;
— *impurity F*: maximum 1.2 per cent;
— *impurity B*: maximum 0.5 per cent;
— *impurity C*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 3.0 per cent;
— *reporting threshold*: 0.05 per cent.

Ethanol
Gas chromatography (*2.2.28*).

Internal standard solution Dilute 0.50 mL of *propanol R* to 1.0 L with *water R*.

Test solution Dissolve 0.10 g of the substance to be examined in the internal standard solution and dilute to 10.0 mL with the same solution.

Reference solution Dilute 0.5 mL of *anhydrous ethanol R* to 100.0 mL with the internal standard solution. Dilute 1.0 mL of this solution to 10.0 mL with the internal standard solution.

Column:
— *material*: fused silica;
— *size*: *l* = 50 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 5 µm).

Carrier gas *helium for chromatography R*.

Linear velocity 37 cm/s.

Split ratio 1:10.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 7	70
	7 - 13	70 → 160
	13 - 15	160
Injection port		200
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to propanol (retention time = about 7 min): ethanol = about 0.6.

Calculate the content of ethanol, taking the density (*2.2.5*) at 20 °C to be 0.790 g/mL.

Limit:
— *ethanol*: 4.3 per cent to 6.0 per cent.

Water (*2.5.12*)
1.4 per cent to 2.8 per cent, determined on 1.20 g.

Sulfated ash (*2.4.14*)
Maximum 0.4 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 1.14 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

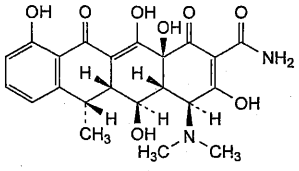
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{22}H_{25}ClN_2O_8$ taking into account the assigned content of *doxycycline hyclate CRS*.

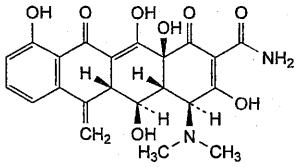
STORAGE
In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES
Specified impurities *A, B, C, F.*

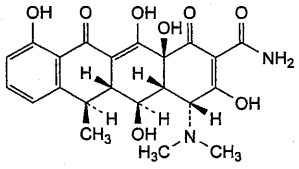
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E.*



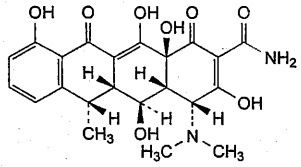
A. (4*S*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (6-epidoxycycline),



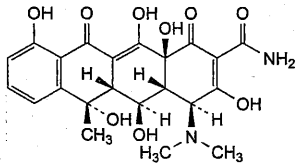
B. (4*S*,4a*R*,5*S*,5a*R*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methylene-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (metacycline),



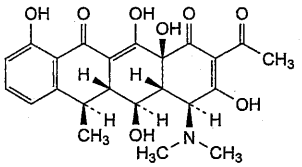
C. (4*R*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidoxycycline),



D. (4*R*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epi-6-epidoxycycline),



E. (4*S*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (oxytetracycline),

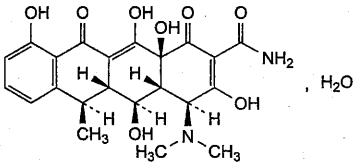


F. (4*S*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-2-acetyl-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-4a,5a,6,12a-tetrahydrotetracene-1,11(4*H*,5*H*)-dione (2-acetyl-2-decarbamoyldoxycycline).

Ph Eur

Doxycycline Monohydrate

(Ph. Eur. monograph 0820)



$C_{22}H_{24}N_2O_8,H_2O$ 462.5 *17086-28-1*

Action and use
Tetracycline antibacterial.

Preparations
Doxycycline Dispersible Tablets
Doxycycline Prolonged-release Capsules

Ph Eur

DEFINITION

(4*S*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-4-(Dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide monohydrate.

Substance obtained from oxytetracycline or metacycline or by any other means.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent of $C_{22}H_{24}N_2O_8$ (anhydrous substance).

CHARACTERS

Appearance
Yellow, crystalline powder.

Solubility
Very slightly soluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of mineral acids and in solutions of alkali hydroxides and carbonates.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of *sulfuric acid R*. A yellow colour develops.

C. Water (see Tests).

TESTS

pH *(2.2.3)*
5.0 to 6.5.

Suspend 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Absorbance (*2.2.25*)
Maximum 0.07 at 490 nm (anhydrous substance).

Dissolve 0.10 g in a mixture of 0.5 volumes of *hydrochloric acid R* and 99.5 volumes of *methanol R* and dilute to 10.0 mL with the same mixture of solvents. Carry out the measurement within 1 h of preparing the solution.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Suspend 111.6 g of *sodium edetate R* in 900 mL of *water R*. Adjust to pH 7.0 with *concentrated ammonia R* to achieve complete dissolution, then dilute to 1 L with *water R*.

Test solution Dissolve 20.0 mg of the substance to be examined in a 1 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same solution.

Reference solution (a) Dissolve 20.0 mg of *doxycycline hyclate CRS* in a 1 g/L solution of *hydrochloric acid R* and dilute to 25.0 mL with the same solution.

Reference solution (b) Dissolve 5 mg of *doxycycline for system suitability CRS* (containing impurities A, B, C and F) in a 1 g/L solution of *hydrochloric acid R* and dilute to 10 mL with the same solution.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with a 1 g/L solution of *hydrochloric acid R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm) ;
— *temperature*: 35 °C.

Mobile phase Mix 13 volumes of *acetonitrile R*, 17 volumes of *water R*, 35 volumes of a 67.9 g/L solution of *tetrabutylammonium hydrogen sulfate R* previously adjusted to pH 7.0 with *concentrated ammonia R* and 35 volumes of solution A.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of doxycycline.

Identification of impurities Use the chromatogram supplied with *doxycycline for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and F.

Relative retention With reference to doxycycline (retention time = about 21 min): impurity C = about 0.4; impurity A = about 0.7; impurity B = about 0.8; impurity F = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities A and B; minimum 2.0 between the peaks due to impurity B and doxycycline.

Calculation of percentage contents:
— for each impurity, use the concentration of doxycycline in reference solution (c).

Limits:
— *impurity A*: maximum 2.0 per cent;
— *impurity F*: maximum 1.2 per cent;
— *impurity B*: maximum 0.5 per cent;
— *impurity C*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 3.0 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
3.6 per cent to 4.6 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.4 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

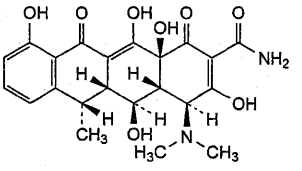
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{22}H_{24}N_2O_8$ taking into account the assigned content of *doxycycline hyclate CRS*.

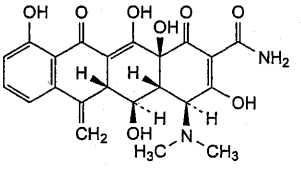
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C, F.*

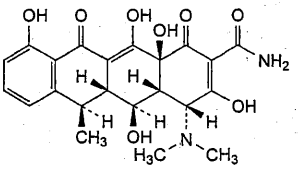
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E.*



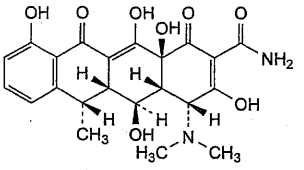
A. (4*S*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (6-epidoxycycline),



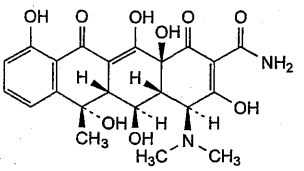
B. (4*S*,4a*R*,5*S*,5a*R*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methylene-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (metacycline),



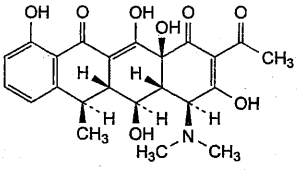
C. (4*R*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epidoxycycline),



D. (4*R*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epi-6-epidoxycycline),



E. (4*S*,4a*R*,5*S*,5a*R*,6*S*,12a*S*)-4-(dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (oxytetracycline),

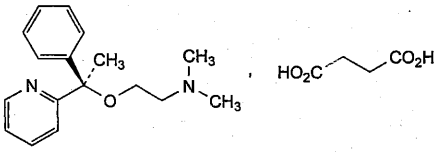


F. (4*S*,4a*R*,5*S*,5a*R*,6*R*,12a*S*)-2-acetyl-4-(dimethylamino)-3,5,10,12,12a-pentahydroxy-6-methyl-4a,5a,6,12a-tetrahydrotetracene-1,11(4*H*,5*H*)-dione (2-acetyl-2-decarbamoyldoxycycline).

Ph Eur

Doxylamine Succinate

(Doxylamine Hydrogen Succinate, Ph. Eur. monograph 1589)



and enantiomer

$C_{21}H_{28}N_2O_5$ 388.5 *562-10-7*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION
N,*N*-Dimethyl-2-[(1*RS*)-1-phenyl-1-(pyridin-2-yl)ethoxy] ethanamine hydrogen butanedioate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *doxylamine hydrogen succinate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.4 g in *water R* and dilute to 20 mL with the same solvent.

Related substances
Gas chromatography (*2.2.28*).

Test solution Dissolve 0.650 g of the substance to be examined in 20 mL of a 10.3 g/L solution of *hydrochloric acid R*. Add 3 mL of a 100 g/L solution of *sodium hydroxide R* and extract with 3 quantities, each of 25 mL, of *methylene chloride R*. Combine the methylene chloride extracts and filter using hydrophobic phase-separation filter paper. Rinse the filter with 10 mL of *methylene chloride R* and combine the rinsings with the methylene chloride extracts. Evaporate the solvent under reduced pressure at a temperature not exceeding 40 °C. Dissolve the residue in 20.0 mL of *anhydrous ethanol R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *anhydrous ethanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R*.

Reference solution (b) Dissolve 50 mg of *doxylamine for system suitability CRS* (containing impurity C) in 10 mL of a 10.3 g/L solution of *hydrochloric acid R*. Add 1.5 mL of a 100 g/L solution of *sodium hydroxide R* and extract with 3 quantities, each of 25 mL, of *methylene chloride R*. Combine the methylene chloride extracts and filter using hydrophobic phase-separation filter paper. Rinse the filter with 10 mL of *methylene chloride R* and combine the rinsings with the methylene chloride extracts. Evaporate the solvent under reduced pressure at a temperature not exceeding 40 °C. Dissolve the residue in 5.0 mL of *anhydrous ethanol R*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 1.5 µm).

Carrier gas *helium for chromatography R.*

Flow rate 7 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 12	160 → 220
	12 - 27	220
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL.

Identification of impurities Use the chromatogram supplied with *doxylamine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to doxylamine (retention time = about 12 min): impurity C = about 0.96.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurity C and doxylamine.

Limits:
— *impurity C*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 2.00 g.

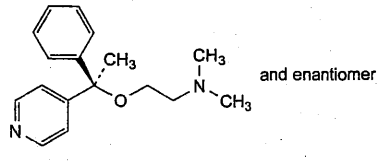
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

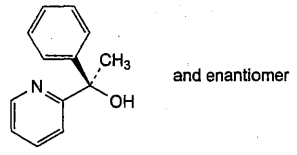
1 mL of *0.1 M perchloric acid* is equivalent to 19.43 mg of $C_{21}H_{28}N_2O_5$.

IMPURITIES
Specified impurities *C.*

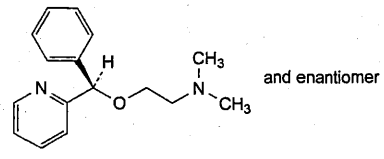
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, D.*



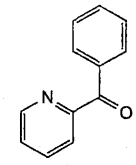
A. *N,N*-dimethyl-2-[(1*RS*)-1-phenyl-1-(pyridin-4-yl)ethoxy] ethanamine,



B. (1*RS*)-1-phenyl-1-(pyridin-2-yl)ethanol,



C. *N,N*-dimethyl-2-[(1*RS*)-1-phenyl-1-(pyridin-2-yl)methoxy] ethanamine,

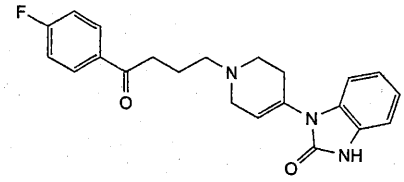


D. phenyl(pyridin-2-yl)methanone (2-benzoylpyridine).

Ph Eur

Droperidol

(Ph. Eur. monograph 1010)



$C_{22}H_{22}FN_3O_2$ 379.4 *548-73-2*

Action and use
Dopamine receptor antagonist; $beta_1$-adrenoceptor agonist; alpha-adrenoceptor agonist; neuroleptic.

Preparations
Droperidol Injection
Droperidol Tablets

Ph Eur

DEFINITION
1-[1-[4-(4-Fluorophenyl)-4-oxobutyl]-1,2,3,6-tetrahydropyridin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in dimethylformamide and in methylene chloride, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *droperidol CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *acetone R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (a) Dissolve 30 mg of *droperidol CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of *droperidol CRS* and 30 mg of *benperidol CRS* in the mobile phase and dilute to 10 mL with the mobile phase.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *acetone R, methanol R* (10:90 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 5 mL of *anhydrous ethanol R*. Add 0.5 mL of *dinitrobenzene solution R* and 0.5 mL of *2 M alcoholic potassium hydroxide R*. A violet colour is produced and becomes brownish-red after 20 min.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 0.20 g in *methylene chloride R* and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.10 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of *droperidol CRS* and 2.5 mg of *benperidol CRS* in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethylformamide R*. Dilute 5.0 mL of this solution to 20.0 mL with *dimethylformamide R*.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: *acetonitrile R*;
— *mobile phase B*: 10 g/L solution of *tetrabutylammonium hydrogen sulfate R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	0 → 40	100 → 60
15 - 20	40	60
20 - 25	40 → 0	60 → 100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 µL.

Relative retention With reference to droperidol (retention time = about 7 min): impurity A = about 0.2; impurity B = about 0.85; benperidol = about 0.9; impurity C = about 0.95; impurity D = about 1.2; impurity E = about 1.5.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to benperidol and droperidol.

Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *unspecified impurities*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Using 0.2 mL of *naphtholbenzein solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from orange-yellow to green.

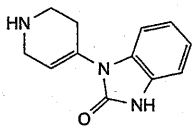
1 mL of *0.1 M perchloric acid* is equivalent to 37.94 mg of $C_{22}H_{22}FN_3O_2$.

STORAGE

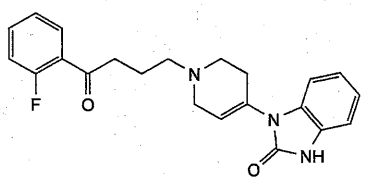
Protected from light.

IMPURITIES

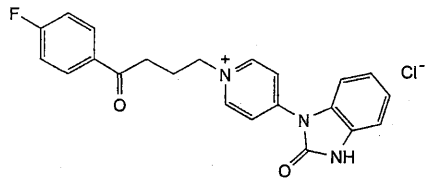
Specified impurities *A, B, C, D, E.*



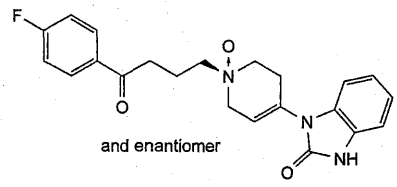
A. 1-(1,2,3,6-tetrahydropyridin-4-yl)-1,3-dihydro-2*H*-benzimidazol-2-one,



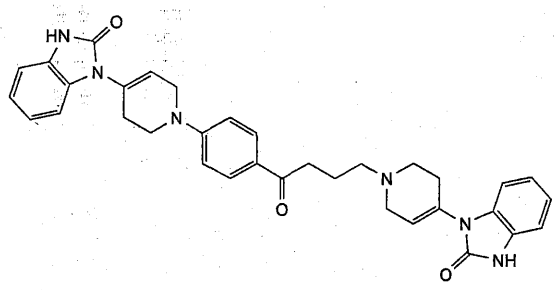
B. 1-[1-[4-(2-fluorophenyl)-4-oxobutyl]-1,2,3,6-tetrahydropyridin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one,



C. 1-[4-(4-fluorophenyl)-4-oxobutyl]-4-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)pyridinium chloride,



D. (1*RS*)-1-[4-(4-fluorophenyl)-4-oxobutyl]-4-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-1,2,3,6-tetrahydropyridine 1-oxide,

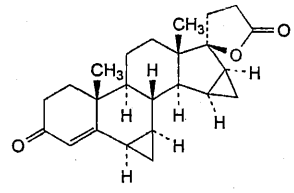


E. 1-[1-[4-[4-[4-(2-oxo-2,3-dihydro-1*H*-benzimidazol-1-yl)-3,6-dihydropyridin-1(2*H*)-yl]-1-oxobutyl]phenyl]-1,2,3,6-tetrahydropyridin-4-yl]-1,3-dihydro-2*H*-benzimidazol-2-one.

Ph Eur

Drospirenone

(Ph. Eur. monograph 2404)



$C_{24}H_{30}O_3$ 366.5 *67392-87-4*

Action and use
Aldosterone receptor antagonist.

Ph Eur

DEFINITION
3-Oxo-6α,7α,15α,16α-tetrahydro-3′*H*,3″*H*-dicyclopropa[6,7:15,16]-17α-pregn-4-en-21,17-carbolactone.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *drospirenone CRS*.

TESTS
Specific optical rotation (*2.2.7*)
−193 to −187 (dried substance).

Dissolve 0.100 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R*, *water R* (50:50 *V/V*).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Use 1.0 mL of this solution to dissolve the contents of a vial of *drospirenone impurity E CRS*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 30.0 mg of *drospirenone CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (d) Dissolve 3 mg of *drospirenone for peak identification CRS* (containing impurity A) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	63	37
2 - 16	63 → 52	37 → 48
16 - 23	52	48
23 - 31	52 → 20	48 → 80
31 - 39	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 10 µL of the test solution and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram supplied with *drospirenone for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E.

Relative retention With reference to drospirenone (retention time = about 22 min): impurity E = about 1.1; impurity A = about 1.2.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to drospirenone and impurity E.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.5;
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

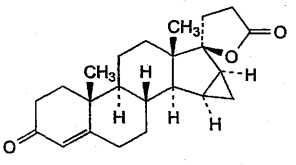
Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{24}H_{30}O_3$ taking into account the assigned content of *drospirenone CRS*.

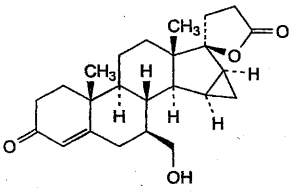
IMPURITIES

Specified impurities *A*.

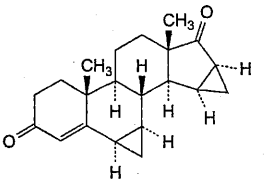
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H, I, K*.



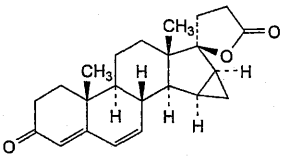
A. 3-oxo-15α,16α-dihydro-3′*H*-cyclopropa[15,16]-17α-pregn-4-ene-21,17-carbolactone (6,7-desmethylenedrospirenone),



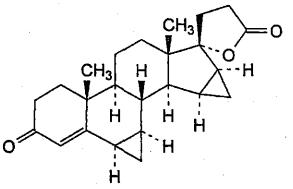
B. 7β-(hydroxymethyl)-3-oxo-15α,16α-dihydro-3′*H*-cyclopropa[15,16]-17α-pregn-4-ene-21,17-carbolactone (7β-hydroxymethyl derivative),



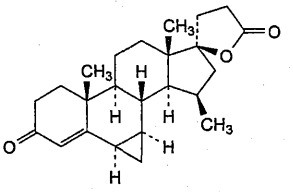
C. 6α,7α,15α,16α-tetrahydro-3′*H*,3″*H*-dicyclopropa[6,7:15,16]androst-4-ene-3,17-dione (17-keto derivative),



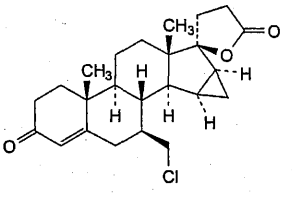
D. 3-oxo-15α,16α-dihydro-3′*H*-cyclopropa[15,16]-17α-pregna-4,6-diene-21,17-carbolactone (Δ6-drospirenone),



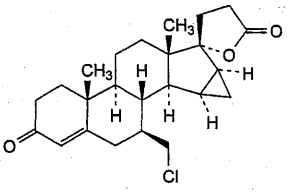
E. 3-oxo-6α,7α,15α,16α-tetrahydro-3′*H*,3″*H* dicyclopropa[6,7:15,16]pregn-4-ene-21,17-carbolactone (17-epidrospirenone),



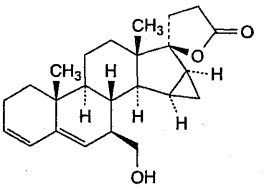
F. 15β-methyl-3-oxo-6α,7α-dihydro-3′*H*-cyclopropa[6,7]-17α-pregn-4-ene-21,17-carbolactone (3″-16-secodrospirenone),



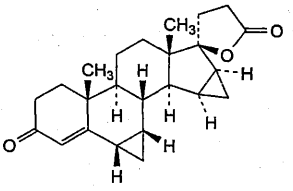
G. 7β-(chloromethyl)-3-oxo-15α,16α-dihydro-3′*H*-cyclopropa[15,16]-17α-pregn-4-ene-21,17-carbolactone (3′-chloro-3′,6-secodrospirenone),



H. 7β-(chloromethyl)-3-oxo-15α,16α-dihydro-3′*H*-cyclopropa[15,16]pregn-4-ene-21,17-carbolactone (3′-chloro-3′,6-seco-17-epidrospirenone),



I. 7β-(hydroxymethyl)-15α,16α-dihydro-3′*H*-cyclopropa[15,16]-17α-pregna-3,5-diene-21,17-carbolactone (7β-hydroxymethyldiene derivative),

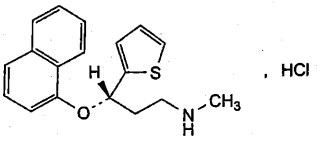


K. 3-oxo-6β,7β,15α,16α-tetrahydro-3′*H*,3″*H*-dicyclopropa[6,7:15,16]-17α-pregn-4-ene-21,17-carbolactone (6α,7α-drospirenone).

Ph Eur

Duloxetine Hydrochloride

(Ph. Eur. monograph 2594)



$C_{18}H_{20}ClNOS$ 333.9 *136434-34-9*

Action and use
Inhibition of 5HT and noradrenaline uptake; antidepressant.

Ph Eur

DEFINITION
(3*S*)-*N*-Methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl) propan-1-amine hydrochloride.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Sparingly soluble in water, freely soluble in methanol, practically insoluble in hexane.

IDENTIFICATION
Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (*2.2.7*): + 119 to + 127 (dried substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent. Examine within 30 min of preparing the solution.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *duloxetine hydrochloride CRS.*

C. Enantiomeric purity (see Tests).

D. Dissolve 25 mg in 5 mL of *methanol R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Enantiomeric purity
Liquid chromatography (*2.2.29*).

Test solution Dissolve 5.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *duloxetine impurity A CRS* and 5 mg of the substance to be examined in 100.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel OD for chiral separations R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Add 2.0 mL of *diethylamine R* to 1000 mL of a mixture of 17 volumes of *2-propanol R* and 83 volumes of *hexane R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Relative retention With reference to duloxetine (retention time = about 7 min): impurity A = about 1.3.

System suitability:
— *resolution*: minimum 3.5 between the peaks due to duloxetine and impurity A in the chromatogram obtained with reference solution (b);
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (a).

Limit:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light. Prepare the solutions immediately before use.*

Solvent mixture *acetonitrile R1, water R* (25:75 *V/V*).

Test solution (a) Dissolve 20 mg of the substance to be examined in 200.0 mL of the solvent mixture.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in 100.0 mL of the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 20 mg of *duloxetine for system suitability CRS* (containing impurity F) in the mobile phase and dilute to 200.0 mL with the mobile phase. In order to prepare impurities C and D *in situ*, heat the solution at 60 °C for 1 h (solution containing impurities C, D and F).

Reference solution (c) Dissolve 50.0 mg of *duloxetine hydrochloride CRS* in 100.0 mL of the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *octylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 40 °C.

Hexanesulfonate solution Dissolve 10.3 g of *sodium hexanesulfonate monohydrate for ion-pair chromatography R* in a solution prepared as follows and dilute to 1000.0 mL with the same solution: dissolve 2.9 g (1.7 mL) of *phosphoric acid R* in 900 mL of *water R*, adjust to pH 2.5 with *dilute sodium hydroxide solution R* and dilute to 1000 mL with *water R*.

Mobile phase *acetonitrile R1*, *propanol R*, hexanesulfonate solution (13:17:70 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Run time 2.5 times the retention time of duloxetine.

Identification of impurities Use the chromatogram supplied with *duloxetine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C, D and F.

Relative retention With reference to duloxetine (retention time = about 16 min): impurity C = about 0.4; impurity D = about 0.5; impurity F = about 1.1.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurities C and D;
- *peak-to-valley ratio*: minimum 4.0, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to duloxetine.

Limits:
- *impurity F*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying *(2.2.32)*
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash *(2.4.14)*
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography *(2.2.29)* as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{18}H_{20}ClNOS$ taking into account the assigned content of *duloxetine hydrochloride CRS*.

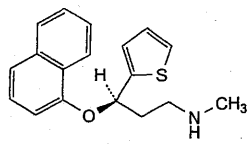
STORAGE

Protected from light.

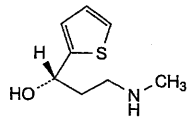
IMPURITIES

Specified impurities *A, F.*

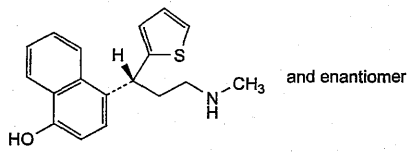
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, G.*



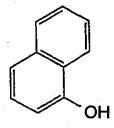
A. (3*R*)-*N*-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl) propan-1-amine,



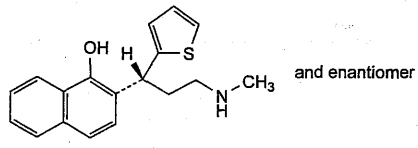
B. (1*S*)-3-(methylamino)-1-(thiophen-2-yl)propan-1-ol,



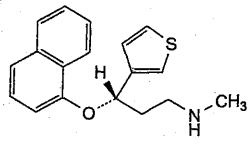
C. 4-[(1*RS*)-3-(methylamino)-1-(thiophen-2-yl)propyl] naphthalen-1-ol,



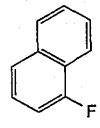
D. naphthalen-1-ol,



E. 2-[(1*RS*)-3-(methylamino)-1-(thiophen-2-yl)propyl] naphthalen-1-ol,



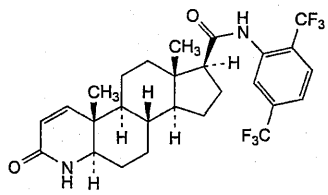
F. (3*S*)-*N*-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-3-yl) propan-1-amine,



G. 1-fluoronaphthalene.

Dutasteride

(Ph. Eur. monograph 2641)



$C_{27}H_{30}F_6N_2O_2$ 528.5 *164656-23-9*

Ph Eur

DEFINITION

N-[2,5-Bis(trifluoromethyl)phenyl]-3-oxo-4-aza-5α-androst-1-ene-17β-carboxamide.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or pale yellow powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble or sparingly soluble in anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *dutasteride CRS*.

TESTS

Specific optical rotation (*2.2.7*)

+ 33.0 to + 39.0 (anhydrous substance).

Dissolve 0.100 g in *anhydrous ethanol R* and dilute to 20.0 mL with the same solvent.

Related substances

A. Liquid chromatography (*2.2.29*).

Solvent mixture *water for chromatography R, acetonitrile R1* (40:60 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *dutasteride for system suitability CRS* (containing impurities A, B, C, E, F, G, H and I) in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *dutasteride CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 35 °C.

Mobile phase Mix 0.25 volumes of *trifluoroacetic acid R*, 480 volumes of *water for chromatography R* and 520 volumes of *acetonitrile R1*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (a) and (b).

Run time 1.6 times the retention time of dutasteride.

Identification of impurities Use the chromatogram supplied with *dutasteride for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, E, F and G.

Relative retention With reference to dutasteride (retention time = about 36 min): impurity A = about 0.10; impurity B = about 0.11; impurity C = about 0.4; impurity E = about 0.9; impurity F = about 1.1; impurity G = about 1.2.

System suitability:

— *resolution*: minimum 1.5 between the peaks due to impurity E and dutasteride and minimum 1.5 between the peaks due to impurities A and B in the chromatogram obtained with reference solution (b);

— *signal-to-noise ratio*: minimum 30 for the peak due to dutasteride in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:

— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.7; impurity F = 3.0;

— for each impurity, use the concentration of dutasteride in reference solution (a).

Limits:

— *impurity F*: maximum 0.4 per cent;

— *impurities E, G*: for each impurity, maximum 0.3 per cent;

— *impurities A, C*: for each impurity, maximum 0.2 per cent;

— *impurity B*: maximum 0.15 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *reporting threshold*: 0.05 per cent.

B. Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modifications.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water for chromatography R, acetonitrile R1* (20:80 *V/V*).

Injection 10 µL of the test solution and reference solutions (a) and (b).

Run time 5 times the retention time of dutasteride.

Identification of impurities Use the chromatogram supplied with *dutasteride for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities H and I.

Relative retention With reference to dutasteride (retention time = about 4 min): impurity H = about 3.4; impurity I = about 3.9.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the peaks due to impurities H and I.

Calculation of percentage contents:

— for each impurity, use the concentration of dutasteride in reference solution (a).

Limits:

— *impurity I*: maximum 0.5 per cent;

— *impurity H*: maximum 0.3 per cent;

— *unspecified impurities eluting after dutasteride*: for each impurity, maximum 0.10 per cent;
— *reporting threshold*: 0.05 per cent.

Limit:
— *total for tests A and B*: maximum 1.5 per cent.

Water (*2.5.32*)
Maximum 0.2 per cent, determined on 0.100 g using the evaporation technique at 180 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modification.

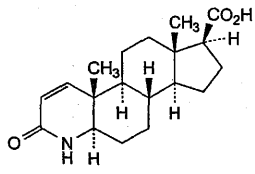
Injection 10 µL of the test solution and reference solution (c).

Calculate the percentage content of $C_{27}H_{30}F_6N_2O_2$ taking into account the assigned content of *dutasteride CRS*.

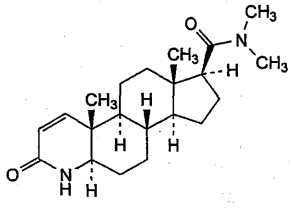
IMPURITIES

Specified impurities *A, B, C, E, F, G, H, I.*

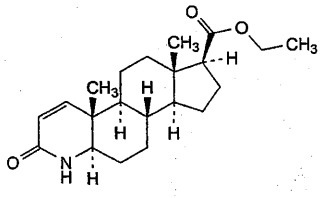
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*



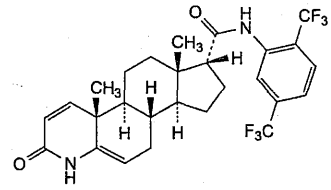
A. 3-oxo-4-aza-5α-androst-1-ene-17β-carboxylic acid,



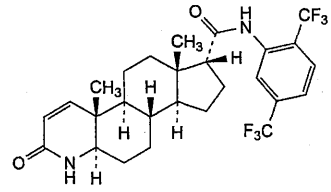
B. *N,N*-dimethyl-3-oxo-4-aza-5α-androst-1-ene-17β-carboxamide,



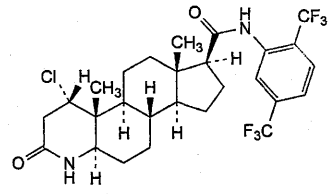
C. ethyl 3-oxo-4-aza-5α-androst-1-ene-17β-carboxylate,



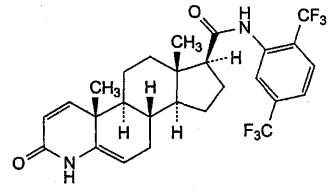
D. *N*-[2,5-bis(trifluoromethyl)phenyl]-3-oxo-4-azaandrost-1,5-diene-17α-carboxamide,



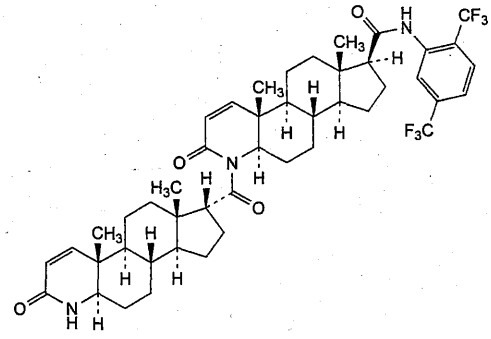
E. *N*-[2,5-bis(trifluoromethyl)phenyl]-3-oxo-4-aza-5α-androst-1-ene-17α-carboxamide,



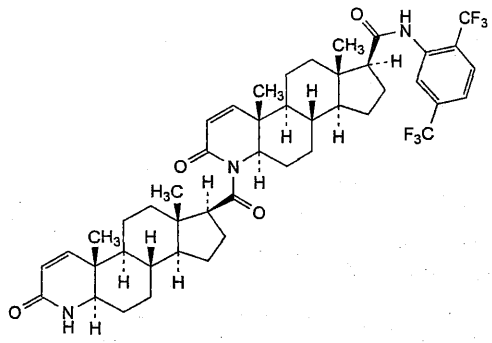
F. *N*-[2,5-bis(trifluoromethyl)phenyl]-1α-chloro-3-oxo-4-aza-5α-androstane-17β-carboxamide,



G. *N*-[2,5-bis(trifluoromethyl)phenyl]-3-oxo-4-azaandrost-1,5-diene-17β-carboxamide,



H. *N*-[2,5-bis(trifluoromethyl)phenyl]-3-oxo-4-[3-oxo-4-aza-5α-androst-1-ene-17α-carbonyl]-4-aza-5α-androst-1-ene-17β-carboxamide (dutasteride dimer 1),

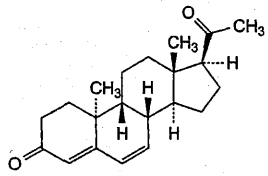


I. *N*-[2,5-bis(trifluoromethyl)phenyl]-3-oxo-4-[3-oxo-4-aza-5α-androst-1-ene-17β-carbonyl]-4-aza-5α-androst-1-ene-17β-carboxamide (dutasteride dimer 2).

Ph Eur

Dydrogesterone

(Ph. Eur. monograph 2357)



$C_{21}H_{28}O_2$ 312.5 *152-62-5*

Action and use
Progestogen.

Preparation
Dydrogesterone Tablets

Ph Eur

DEFINITION
9β,10α-Pregna-4,6-diene-3,20-dione.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetone, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *dydrogesterone CRS.*

TESTS
Specific optical rotation (*2.2.7*)
−469 to −485 (dried substance), measured at 25 °C.
Dissolve 0.100 g in *methylene chloride R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Test solution (b) Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 3.0 mg of *dydrogesterone impurity A CRS* in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 10 mg of the substance to be examined in 10 mL of reference solution (a).
Reference solution (d) Dissolve 10 mg of the substance to be examined in 30 mL of *ethanol (96 per cent) R*. Add 1 mL of a 8.4 g/L solution of *sodium hydroxide R* and heat at 85 °C for 10 min. Cool to room temperature, add 1 mL of a 20.6 g/L solution of *hydrochloric acid R*, add 20 mL of *acetonitrile R*, 2 mg of *dydrogesterone impurity B CRS*, dilute to 100 mL with *water R* and mix. This solution contains dydrogesterone and impurities B and C.
Reference solution (e) Dissolve 20.0 mg of *dydrogesterone CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 40 °C.
Mobile phase *acetonitrile R, ethanol (96 per cent) R, water R* (21:25:54 *V/V/V*).
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 280 nm and at 385 nm.
Injection 10 µL of test solution (a) and reference solutions (a), (b), (c) and (d).
Run time Twice the retention time of dydrogesterone.
Relative retention at 385 nm With reference to dydrogesterone (retention time = about 13 min): impurity A = about 0.9.
Relative retention at 280 nm With reference to dydrogesterone (retention time = about 13 min): impurity B = about 1.1; impurity C = about 1.2.
System suitability:
— *resolution at 385 nm*: minimum 1.1 between the peaks due to impurity A and dydrogesterone in the chromatogram obtained with reference solution (c);
— *resolution at 280 nm*: minimum 4.5 between the peaks due to dydrogesterone and impurity B and minimum 1.5 between the peaks due to impurity B and impurity C in the chromatogram obtained with reference solution (d).
Limits:
— *impurity A at 385 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity B at 280 nm*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *impurity C at 280 nm*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *unspecified impurities at 280 nm*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total at 280 nm*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit at 280 nm*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

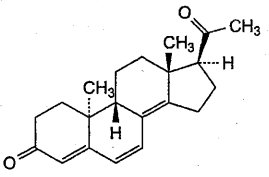
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 280 nm.

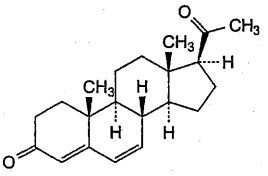
Injection Test solution (b) and reference solution (e).

Calculate the percentage content of $C_{21}H_{28}O_2$ from the declared content of *dydrogesterone CRS*.

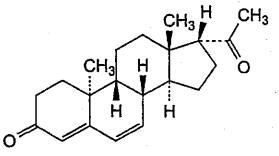
IMPURITIES
Specified impurities *A, B, C.*



A. 9β,10α-pregna-4,6,8(14)-triene-3,20-dione,



B. pregna-4,6-diene-3,20-dione,

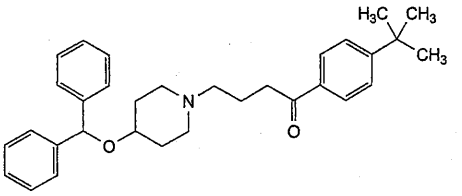


C. 9β,10α,17α-pregna-4,6-diene-3,20-dione.

Ph Eur

Ebastine

(Ph. Eur. monograph 2015)



$C_{32}H_{39}NO_2$ 469.7 *90729-43-4*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION
1-(4-*tert*-Butylphenyl)-4-[4-(diphenylmethoxy)piperidin-1-yl] butan-1-one.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, very soluble in methylene chloride, sparingly soluble in methanol.

mp
About 86 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ebastine CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solution A Dilute 12 mL of *phosphoric acid R* and 12 mL of *diethylamine R* in 800 mL of *water for chromatography R*, adjust to pH 6.0 with *diethylamine R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *bis(diphenylmethyl) ether R* (impurity H) in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 0.5 mL of the solution to 5.0 mL with the test solution. Dilute 3.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Solution A, *methanol R2*, *acetonitrile R1* (20:38:42 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 15 °C.

Injection 10 µL.

Run time 1.5 times the retention time of ebastine.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity H.

Relative retention With reference to ebastine (retention time = about 27 min): impurity H = about 0.9.

System suitability Reference solution (a):
— *resolution*: minimum 2 between the peaks due to impurity H and ebastine.

Calculation of percentage contents:
— for each impurity, use the concentration of ebastine in reference solution (b).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent.

Sulfates (*2.4.13*)
Maximum 100 ppm.

Suspend 2.5 g in 25 mL of *dilute nitric acid R*. Boil under a reflux condenser for 10 min. Cool and filter.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

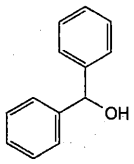
1 mL of *0.1 M perchloric acid* is equivalent to 46.97 mg of $C_{32}H_{39}NO_2$.

STORAGE

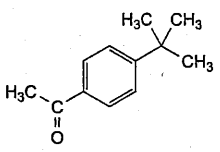
Protected from light.

IMPURITIES

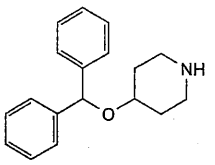
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H.*



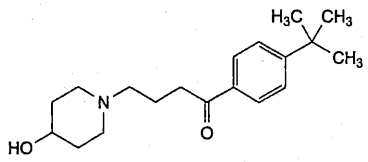
A. diphenylmethanol (benzhydrol),



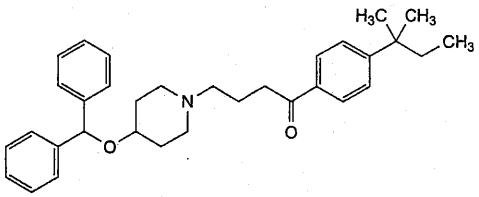
B. 1-(4-*tert*-butylphenyl)ethan-1-one,



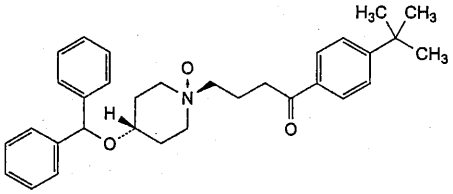
C. 4-(diphenylmethoxy)piperidine,



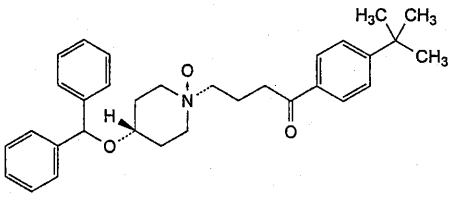
D. 1-(4-*tert*-butylphenyl)-4-(4-hydroxypiperidin-1-yl)butan-1-one,



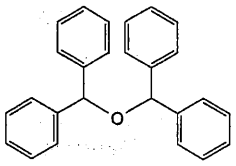
E. 4-[4-(diphenylmethoxy)piperidin-1-yl]-1-[4-(2-methylbutan-2-yl)phenyl]butan-1-one,



F. (1*s*,4*s*)-1-[4-(4-*tert*-butylphenyl)-4-oxobutyl]-4-(diphenylmethoxy)piperidine *N*-oxide,



G. (1*r*,4*r*)-1-[4-(4-*tert*-butylphenyl)-4-oxobutyl]-4-(diphenylmethoxy)piperidine *N*-oxide,

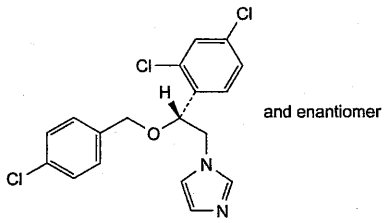


H. [oxybis(methanetriyl)]tetrakisbenzene (bis(diphenylmethyl) ether).

Ph Eur

Econazole

(Ph. Eur. monograph 2049)



$C_{18}H_{15}Cl_3N_2O$ 381.7 *27220-47-9*

Action and use
Antifungal.

Ph Eur

DEFINITION

1-[(2*RS*)-2-[(4-Chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, very soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Melting point (*2.2.14*): 88 °C to 92 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *econazole CRS*.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *econazole for system suitability CRS* (containing impurities A, B and C) in *methanol R* and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 25.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, 0.77 g/L solution of *ammonium acetate R* (20:80 *V/V*);
— *mobile phase B*: *methanol R*, *acetonitrile R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	60 → 10	40 → 90
25 - 27	10	90

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *econazole for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to econazole (retention time = about 15 min): impurity A = about 0.2; impurity B = about 0.6; impurity C = about 1.1.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to econazole.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 75 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

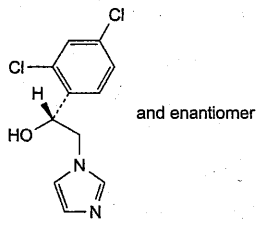
1 mL of *0.1 M perchloric acid* is equivalent to 38.17 mg of $C_{18}H_{15}Cl_3N_2O$.

STORAGE

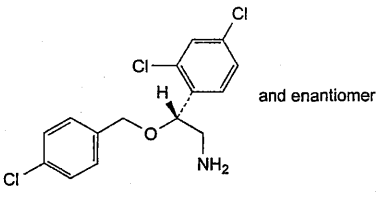
Protected from light.

IMPURITIES

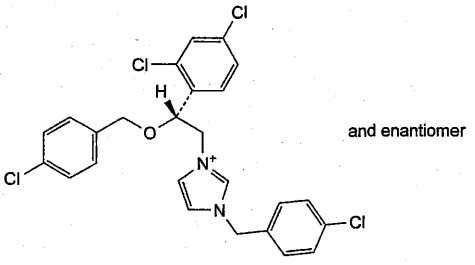
Specified impurities *A, B, C.*



A. (1*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl) ethanol,



B. (2*RS*)-2-[(4-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethanamine,

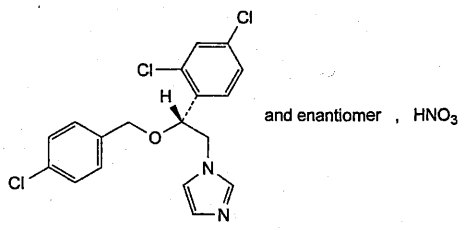


C. 1-(4-chlorobenzyl)-3-[(2*RS*)-2-[(4-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]imidazolium.

Ph Eur

Econazole Nitrate

(Ph. Eur. monograph 0665)



$C_{18}H_{16}Cl_3N_3O_4$ 444.7 *24169-02-6*

Action and use
Antifungal.

Preparations
Econazole Cream
Econazole Pessaries

Ph Eur

DEFINITION
1-[(2*RS*)-2-[(4-Chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole nitrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, soluble in methanol, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

mp
About 165 °C, with decomposition.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *econazole nitrate CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *econazole for system suitability CRS* (containing impurities A, B and C) in *methanol R* and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 25.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, 0.77 g/L solution of *ammonium acetate R* (20:80 *V/V*);
— *mobile phase B*: *methanol R*, *acetonitrile R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	60 → 10	40 → 90
25 - 27	10	90

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *econazole for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to econazole (retention time = about 15 min): impurity A = about 0.2; impurity B = about 0.6; impurity C = about 1.1.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to econazole.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to the nitrate ion at the beginning of the chromatogram.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

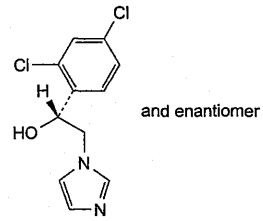
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.400 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

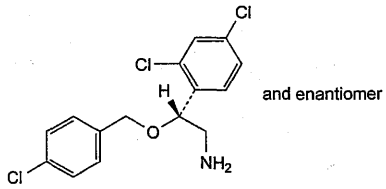
1 mL of *0.1 M perchloric acid* is equivalent to 44.47 mg of $C_{18}H_{16}Cl_3N_3O_4$.

STORAGE
Protected from light.

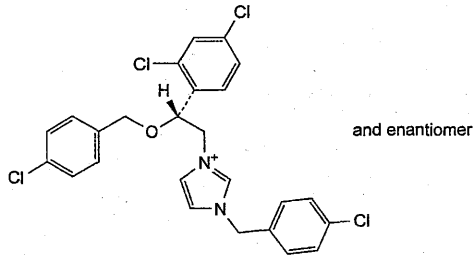
IMPURITIES
Specified impurities *A, B, C.*



A. (1*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl) ethanol,



B. (2*RS*)-2-[(4-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethanamine,

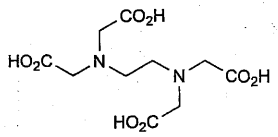


C. 1-(4-chlorobenzyl)-3-[(2*RS*)-2-[(4-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]imidazolium.

Ph Eur

Edetic Acid

(Ph. Eur. monograph 1612)



$C_{10}H_{16}N_2O_8$ 292.2 *60-00-4*

Action and use
Chelating agent.

Ph Eur

DEFINITION
(Ethylenedinitrilo)tetraacetic acid.

Content
98.0 per cent to 101.0 per cent.

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs, after drying the substance to be examined in an oven at 100-105 °C for 2 h.

Comparison *sodium edetate R*, treated as follows: dissolve 0.25 g of *sodium edetate R* in 5 mL of *water R*, add 1.0 mL of *dilute hydrochloric acid R*. Filter, wash the residue with 2 quantities, each of 5 mL, of *water R* and dry the residue in an oven at 100-105 °C for 2 h.

B. To 5 mL of *water R* add 0.1 mL of *ammonium thiocyanate solution R* and 0.1 mL of *ferric chloride solution R1* and mix. The solution is red. Add 0.5 mL of solution S (see Tests). The solution becomes yellowish.

C. To 10 mL of solution S add 0.5 mL of *calcium chloride solution R*. Make alkaline to *red litmus paper R* by the addition of *dilute ammonia R2* and add 3 mL of *ammonium oxalate solution R*. No precipitate is formed.

TESTS
Solution S
Dissolve 5.0 g in 20 mL of *dilute sodium hydroxide solution R* and dilute to 100 mL with *water R*.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Impurity A
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture Dissolve 10.0 g of *ferric sulfate pentahydrate R* in 20 mL of *0.5 M sulfuric acid* and add 780 mL of *water R*. Adjust to pH 2.0 with *1 M sodium hydroxide* and dilute to 1000 mL with *water R*.

Test solution Dissolve 0.100 g of the substance to be examined in 1.0 mL of *1 M sodium hydroxide* and dilute to 25.0 mL with the solvent mixture.

Reference solution Dissolve 40.0 mg of *nitrilotriacetic acid R* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. To 1.0 mL of the solution add 0.1 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: spherical *graphitised carbon for chromatography R1* (5 µm) with a specific surface area of 120 m^2/g and a pore size of 25 nm.

Mobile phase Dissolve 50.0 mg of *ferric sulfate pentahydrate R* in 50 mL of *0.5 M sulfuric acid* and add 750 mL of *water R*. Adjust to pH 1.5 with *0.5 M sulfuric acid* or *1 M sodium hydroxide*, add 20 mL of *ethylene glycol R* and dilute to 1000 mL with *water R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 µL; filter the solutions and inject immediately.

Run time 4 times the retention time of the iron complex of impurity A.

Retention time Iron complex of impurity A = about 5 min; iron complex of edetic acid = about 10 min.

System suitability Reference solution:
- *resolution*: minimum 7 between the peaks due to the iron complex of impurity A and the iron complex of edetic acid,
- *signal-to-noise ratio*: minimum 50 for the peak due to impurity A.

Limit:
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 10 mL of solution S add 8 mL of *nitric acid R* and stir for 10 min. A precipitate is formed. Filter and wash the filter with *water R*. Collect the filtrate and the washings and dilute to 20 mL with *water R*. Dilute 10 mL of this solution to 15 mL with *water R*.

Iron (*2.4.9*)
Maximum 80 ppm.

Dilute 2.5 mL of solution S to 10 mL with *water R* and add 0.25 g of *calcium chloride R* before adding the *thioglycollic acid R*. Allow to stand for 5 min. Also add 0.25 g of *calcium chloride R* to the standard.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 2.0 mL of *dilute sodium hydroxide solution R* and dilute to 300 mL with *water R*. Add 2 g of *hexamethylenetetramine R* and 2 mL of *dilute hydrochloric acid R*. Titrate with *0.1 M zinc sulfate* using about 50 mg of *xylenol orange triturate R* as indicator.

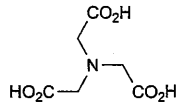
1 mL of *0.1 M zinc sulfate* corresponds to 29.22 mg of $C_{10}H_{16}N_2O_8$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities *A*.

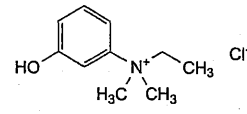


A. nitrilotriacetic acid.

Ph Eur

Edrophonium Chloride

(Ph. Eur. monograph 2106)



$C_{10}H_{16}ClNO$ 201.7 *116-38-1*

Action and use
Cholinesterase inhibitor.

Preparation
Edrophonium Injection

Ph Eur

DEFINITION

N-Ethyl-3-hydroxy-*N*,*N*-dimethylanilinium chloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *edrophonium chloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.5 g in *water R* and dilute to 25 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.0.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *3-dimethylaminophenol R* in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Mix 1.0 mL of the test solution and 1.0 mL of reference solution (a) and dilute to 100.0 mL with *water R*. Dilute 10.0 mL of this solution to 100.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm,
- *stationary phase*: *styrene-divinylbenzene copolymer R* (8-10 µm).

Mobile phase Mix 10 volumes of *acetonitrile R* and 90 volumes of a 7.7 g/L solution of *tetramethylammonium bromide R* previously adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 281 nm.

Injection 20 µL.

Run time Twice the retention time of edrophonium.

Relative retention With reference to edrophonium (retention time = about 3.8 min): impurity A = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to edrophonium and impurity A.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *any other impurity*: for each impurity, not more than the area of the peak due to edrophonium in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *total*: not more than 5 times the area of the peak due to edrophonium in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the peak due to edrophonium in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 24 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 8.3 IU/mg.

ASSAY

Dissolve 0.150 g in 60 mL of a mixture of equal volumes of *acetic anhydride R* and *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

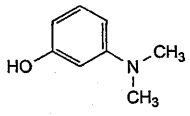
1 mL of *0.1 M perchloric acid* is equivalent to 20.17 mg of $C_{10}H_{16}ClNO$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities *A*.

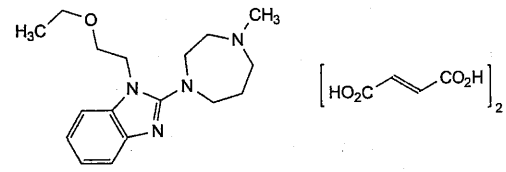


A. 3-(dimethylamino)phenol.

Ph Eur

Emedastine Fumarate

(Emedastine Difumarate, Ph. Eur. monograph 2242)



$C_{25}H_{34}N_4O_9$ 534.6 *87233-62-3*

Action and use
Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION

1-(2-Ethoxyethyl)-2-(4-methylhexahydro-1*H*-1,4-diazepin-1-yl)-1*H*-benzimidazole bis[hydrogen (2*E*)-butenedioate].

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or yellowish powder.

Solubility
Soluble in water, sparingly soluble in anhydrous ethanol, very slightly soluble in acetone.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *emedastine difumarate CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 2.50 g in *water R* and dilute to 50 mL with the same solvent.

pH (*2.2.3*)
3.0 to 4.5.

Dissolve 0.20 g in 100 mL of *carbon dioxide-free water R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *emedastine impurity E CRS* in the mobile phase and dilute to 25 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of the substance to be examined in the mobile phase. Add 0.5 mL of reference solution (a) and dilute to 10 mL with the mobile phase.

Reference solution (c) Dilute 5.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 3.9 g of *disodium hydrogen phosphate dodecahydrate R* and 2.5 g of *sodium dodecyl sulfate R* in *water R* and dilute to 1000.0 mL with the same solvent. Adjust to pH 2.4 with *phosphoric acid R*. Mix 550 volumes of this solution with 450 volumes of *acetonitrile R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of emedastine.

Relative retention With reference to emedastine (retention time = about 18 min): fumaric acid = about 0.1; impurity A = about 0.2; impurity B = about 0.3; impurity C = about 0.5; impurity D = about 0.7; impurity E = about 0.9; impurity F = about 1.4.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 4, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to emedastine.

Limits:
— *impurities A, B, C, D, E, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard the peak due to fumaric acid.

Loss on drying *(2.2.32)*
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

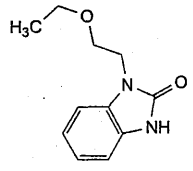
Sulfated ash *(2.4.14)*
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 50 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically *(2.2.20)*.

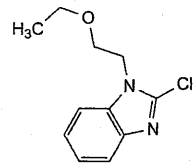
1 mL of *0.1 M perchloric acid* is equivalent to 26.73 mg of $C_{25}H_{34}N_4O_9$.

STORAGE
Protected from light.

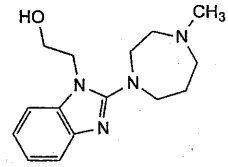
IMPURITIES
Specified impurities *A, B, C, D, E, F.*



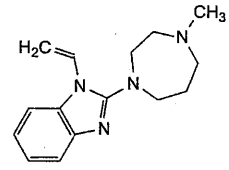
A. 1-(2-ethoxyethyl)-1,3-dihydro-2*H*-benzimidazol-2-one,



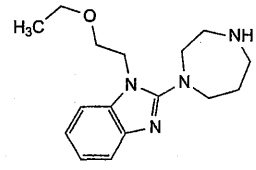
B. 2-chloro-1-(2-ethoxyethyl)-1*H*-benzimidazole,



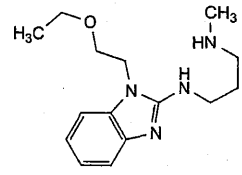
C. 2-[2-(4-methylhexahydro-1*H*-1,4-diazepin-1-yl)-1*H*-benzimidazol-1-yl]ethanol,



D. 1-ethenyl-2-(4-methylhexahydro-1*H*-1,4-diazepin-1-yl)-1*H*-benzimidazole,



E. 1-(2-ethoxyethyl)-2-(hexahydro-1*H*-1,4-diazepin-1-yl)-1*H*-benzimidazole,

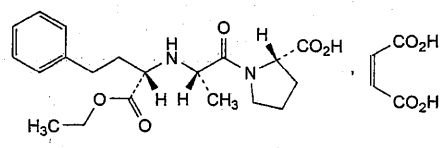


F. *N*-[1-(2-ethoxyethyl)-1*H*-benzimidazol-2-yl]-*N'*-methylpropane-1,3-diamine.

Ph Eur

Enalapril Maleate

(Ph. Eur. monograph 1420)



$C_{24}H_{32}N_2O_9$ 492.5 *76095-16-4*

Action and use
Angiotensin converting enzyme inhibitor.

Preparation
Enalapril Tablets

Ph Eur

DEFINITION

(2*S*)-1-[(2*S*)-2-[[(1*S*)-1-(Ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid hydrogen (*Z*)-butenedioate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, freely soluble in methanol, practically insoluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *enalapril maleate CRS*.

TESTS

Solution S

Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

2.4 to 2.9 for solution S.

Specific optical rotation (*2.2.7*)

−51 to −48 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Buffer solution A Dissolve 2.8 g of *sodium dihydrogen phosphate monohydrate R* in 950 mL of *water for chromatography R*, adjust to pH 2.5 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Buffer solution B Dissolve 2.8 g of *sodium dihydrogen phosphate monohydrate R* in 950 mL of *water for chromatography R*, adjust to pH 6.8 with *strong sodium hydroxide solution R* and dilute to 1000 mL with *water for chromatography R*.

Solvent mixture *acetonitrile R1*, buffer solution A (5:95 *V/V*).

Test solution (a) Dissolve 30 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Test solution (b) Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 3 mg of *enalapril for system suitability CRS* (containing impurity A) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *enalapril impurity mixture A CRS* (impurities C and H) in 1.0 mL of the solvent mixture.

Reference solution (d) Dissolve 2.0 mg of *enalapril impurity G CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 20.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 4.1 mm;
- *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm);
- *temperature*: 70 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R1*, buffer solution B (5:95 *V/V*);
- *mobile phase B*: buffer solution B, *acetonitrile R1* (34:66 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	95	5
3 - 23	95 → 40	5 → 60
23 - 30	40	60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 50 µL of test solution (a) and reference solutions (a), (b) and (c); for impurity G, test solution (b) and reference solution (d).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A; use the chromatogram supplied with *enalapril impurity mixture A CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and H; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity G.

Relative retention With reference to enalapril (retention time = about 13 min): maleic acid = about 0.1; impurity C = about 0.2; impurity A = about 1.1; impurity G = about 1.2; impurity H = about 1.3.

System suitability:
- *signal-to-noise ratio*: minimum 40 for the peak due to enalapril in the chromatogram obtained with reference solution (a); minimum 40 for the peak due to impurity G in the chromatogram obtained with reference solution (d);
- *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to enalapril in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity H by 2.0;
- for impurity G, use the concentration of impurity G in reference solution (d);
- for impurities other than G, use the concentration of enalapril in reference solution (a).

Limits:
- *impurity A*: maximum 0.5 per cent;
- *impurity H*: maximum 0.3 per cent;
- *impurities C, G*: for each impurity, maximum 0.2 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 1.0 per cent;
- *reporting threshold*: 0.05 per cent; disregard the peak due to maleic acid.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent. Titrate with *0.1 M sodium*

hydroxide, determining the end-point potentiometrically (*2.2.20*). Titrate to the 2nd point of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 16.42 mg of $C_{24}H_{32}N_2O_9$.

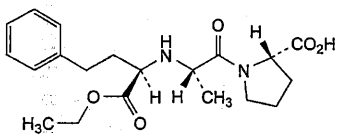
STORAGE

Protected from light.

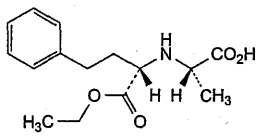
IMPURITIES

Specified impurities *A, C, G, H.*

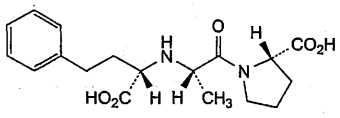
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, E, F, I.*



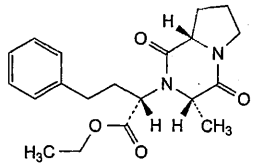
A. (2*S*)-1-[(2*S*)-2-[[(1*R*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,



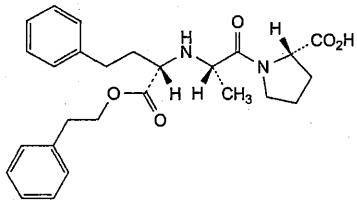
B. (2*S*)-2-[[(1*S*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid,



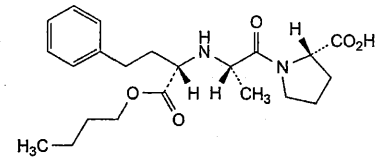
C. (2*S*)-1-[(2*S*)-2-[[(1*S*)-1-carboxy-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,



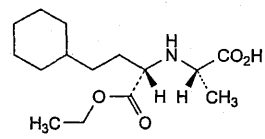
D. ethyl (2*S*)-2-[(3*S*,8a*S*)-3-methyl-1,4-dioxohexahydropyrrolo[1,2-*a*]pyrazin-2(1*H*)-yl]-4-phenylbutanoate,



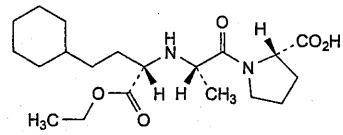
E. (2*S*)-1-[(2*S*)-2-[[(1*S*)-3-phenyl-1-[(2-phenylethoxy)carbonyl]propyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,



F. (2*S*)-1-[(2*S*)-2-[[(1*S*)-1-(butoxycarbonyl)-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,



G. (2*S*)-2-[[(1*S*)-3-cyclohexyl-1-(ethoxycarbonyl)propyl]amino]propanoic acid,



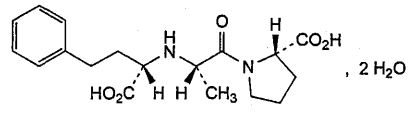
H. (2*S*)-1-[(2*S*)-2-[[(1*S*)-3-cyclohexyl-1-(ethoxycarbonyl)propyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,

I. 1*H*-imidazole.

Ph Eur

Enalaprilat Dihydrate

(Ph. Eur. monograph 1749)



$C_{18}H_{24}N_2O_5,2H_2O$ 384.4 *84680-54-6*

Action and use

Angiotensin converting enzyme inhibitor.

Ph Eur

DEFINITION

(2*S*)-1-[(2*S*)-2-[[(1*S*)-1-Carboxy-3-phenylpropyl]amino]propanoyl)pyrrolidine-2-carboxylic acid dihydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Very slightly soluble or slightly soluble in water, sparingly soluble in methanol, practically insoluble in acetonitrile.

It shows pseudopolymorphism (*5.9*).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Mulls in *liquid paraffin R*.

Comparison *enalaprilat dihydrate CRS*.

If the spectra obtained show differences, expose the substance to be examined and the reference substance to a 98 per cent relative humidity for 3 days using a chamber conditioned with a saturated solution of *calcium sulfate R*. Record new spectra.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.10 g in *water R* and dilute to 100.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)

−53.0 to −56.0 (anhydrous substance).

Dissolve 0.200 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions.*

Buffer solution Dissolve 1.36 g of *potassium dihydrogen phosphate R* in 950 mL of *water R*. Adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture Buffer solution, *acetonitrile R1*, *methanol R1* (1:2:2 *V/V/V*).

Dissolution mixture Solvent mixture, buffer solution (8:92 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in 2.5 mL of *methanol R1* and dilute to 25.0 mL with the dissolution mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the dissolution mixture. Dilute 5.0 mL of this solution to 10.0 mL with the dissolution mixture.

Reference solution (b) Dissolve 5 mg of *enalaprilat for system suitability CRS* (containing impurity C) in 0.5 mL of *methanol R1* and dilute to 5 mL with the dissolution mixture.

Reference solution (c) Dissolve the contents of a vial of *enalaprilat impurity G CRS* in 1 mL of the test solution.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 70 °C.

Mobile phase:
- *mobile phase A*: solvent mixture, buffer solution (10:90 *V/V*);
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	100	0
25 - 50	100 → 90	0 → 10
50 - 80	90	10

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *enalaprilat for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.

Relative retention With reference to enalaprilat (retention time = about 21 min): impurity C = about 1.2; impurity G = about 2.9.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to enalaprilat.

Limits:
- *impurities C, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)

7.0 per cent to 11.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.1 IU/mg.

ASSAY

Dissolve 0.300 g in *glacial acetic acid R* and dilute to 50 mL with the same solvent. Titrate with *0.1 M perchloric acid*, determining the end point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 34.84 mg of $C_{18}H_{24}N_2O_5$.

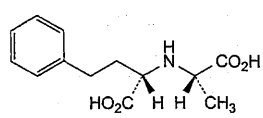
STORAGE

In an airtight container.

IMPURITIES

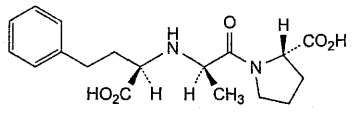
Specified impurities *C, G.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, D, E, F.*

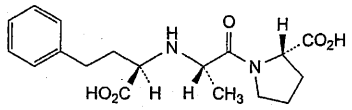


and enantiomer

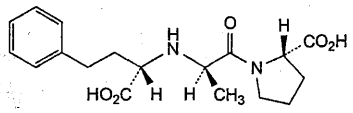
A. (2*SR*)-2-[[(1*SR*)-1-carboxyethyl]amino]-4-phenylbutanoic acid,


and enantiomer

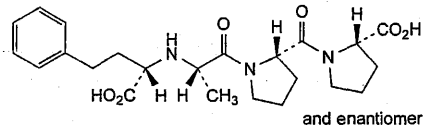
B. (2*SR*)-1-[(2*RS*)-2-[[(1*RS*)-1-carboxy-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,


and enantiomer

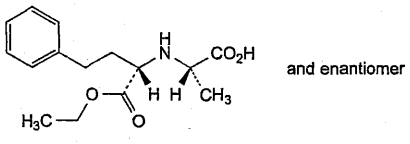
C. (2*SR*)-1-[(2*SR*)-2-[[(1*RS*)-1-carboxy-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,


and enantiomer

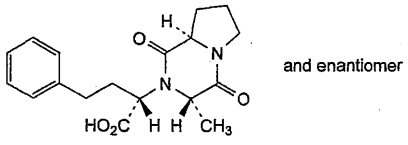
D. (2*SR*)-1-[(2*RS*)-2-[[(1*SR*)-1-carboxy-3-phenylpropyl]amino]propanoyl]pyrrolidine-2-carboxylic acid,


and enantiomer

E. (2*SR*)-1-[[(2*SR*)-1-[(2*SR*)-2-[[(1*SR*)-1-carboxy-3-phenylpropyl]amino]propanoyl]pyrrolidin-2-yl]carbonyl]pyrrolidine-2-carboxylic acid,


and enantiomer

F. (2*SR*)-2-[[(1*SR*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid,

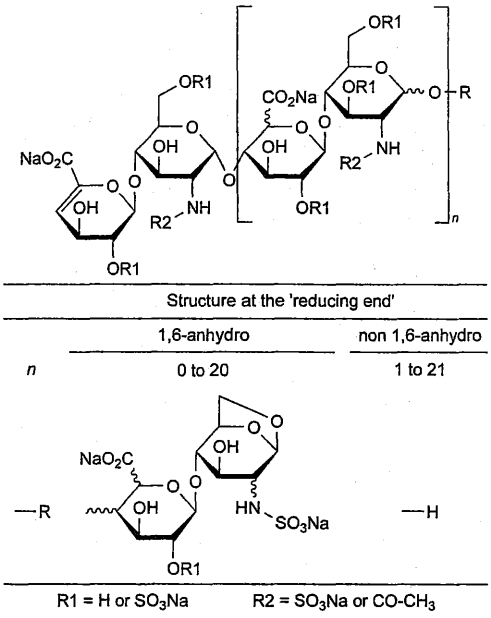

and enantiomer

G. (2*SR*)-2-[(3*SR*,8a*RS*)-3-methyl-1,4-dioxohexahydropyrrolo[1,2-*a*]pyrazin-2(1*H*)-yl]-4-phenylbutanoic acid.

Ph Eur

Enoxaparin Sodium

(Ph. Eur. monograph 1097)



Action and use
Low molecular weight heparin.

Preparation
Enoxaparin Sodium Injection

Ph Eur

DEFINITION

Enoxaparin sodium is the sodium salt of a low-molecular-mass heparin that is obtained by alkaline depolymerisation of the benzyl ester derivative of heparin from porcine intestinal mucosa. Enoxaparin consists of a complex set of oligosaccharides that have not yet been completely characterised. Based on current knowledge, the majority of the components have a 4-enopyranose uronate structure at the non-reducing end of their chain. 15 per cent to 25 per cent of the components have a 1,6-anhydro structure at the reducing end of their chain.

Enoxaparin sodium complies with the monograph Low-molecular-mass heparins (0828) with the modifications and additional requirements below.

The mass-average relative molecular mass ranges between 3800 and 5000, with a characteristic value of about 4500.

The degree of sulfatation is about 2 per disaccharide unit.

The potency is not less than 90 IU and not more than 125 IU of anti-factor Xa activity per milligram, calculated with reference to the dried substance. The anti-factor IIa activity is not less than 20.0 IU and not more than 35.0 IU per milligram, calculated with reference to the dried substance. The ratio of anti-factor Xa activity to anti-factor IIa activity is between 3.3 and 5.3.

PRODUCTION

Enoxaparin is produced by alkaline depolymerisation of benzyl ester derivatives of heparin from porcine intestinal mucosa under conditions that yield a product complying with the structural requirements stated under Definition.

IDENTIFICATION

A. Carry out identification test A as described in the monograph *Low-molecular-mass heparins (0828)* using *enoxaparin sodium CRS*.

B. Liquid chromatography (*2.2.29*): use the normalisation procedure.

Solution A Dissolve 12 mg of *sodium tetrahydroborate R* in 400 µL of *water R* and mix using a vortex mixer.

Heparinase solution (a) Dissolve *heparinase I R* in *potassium phosphate buffer solution pH 7.0 R* to obtain an activity of 0.4 IU/mL. Store the solution at – 20 °C until use.

Heparinase solution (b) Dissolve *heparinase II R* in *potassium phosphate buffer solution pH 7.0* to obtain an activity of 0.4 IU/mL. Store the solution at – 20 °C until use.

Heparinase solution (c) Dissolve *heparinase III R* in *potassium phosphate buffer solution pH 7.0 R* to obtain an activity of 0.4 IU/mL. Store the solution at – 20 °C until use.

Heparinase solution (d) Mix equal volumes of heparinase solution (a), heparinase solution (b) and heparinase solution (c).

Blank solution Gently mix by inversion 20 µL of *water R*, 70 µL of *sodium/calcium acetate buffer solution pH 7.0 R* and 100 µL of heparinase solution (d). Place in a water-bath at 25 °C for 48 h. Mix 60 µL of this solution and 10 µL of freshly prepared solution A. Mix and allow to stand at room temperature for 4 h.

Test solution (a) Dissolve 20 mg of the substance to be examined in 1 mL of *water R*.

Test solution (b) To 20 µL of test solution (a), add 70 µL of *sodium/calcium acetate buffer solution pH 7.0 R* and 100 µL of heparinase solution (d). Gently mix by inversion and place in a water-bath at 25 °C for 48 h.

Test solution (c) To 60 µL of test solution (b), add 10 µL of freshly prepared solution A. Mix and allow to stand at room temperature for 4 h.

Reference solution (a) Dissolve 20 mg of *enoxaparin sodium CRS* in 1 mL of *water R*.

Reference solution (b) To 20 µL of reference solution (a), add 70 µL of *sodium/calcium acetate buffer solution pH 7.0 R* and 100 µL of heparinase solution (d). Gently mix by inversion and place in a water-bath at 25 °C for 48 h.

Reference solution (c) To 60 µL of reference solution (b), add 10 µL of freshly prepared solution A. Mix and allow to stand at room temperature for 4 h.

NOTE: heparinase solutions (a), (b) and (c) can be stored for 3 months at – 20 °C. Test solutions (a) and (b) and reference solutions (a) and (b) must be prepared at the same time; depolymerised test solutions are stable for 1 month at – 20 °C. Test solution (c) and reference solution (c) must also be prepared at the same time.

Precolumn:
- *size*: *l* = 0.01 m, Ø = 4.6 mm;
- *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (5 µm).

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (5 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: dissolve 0.280 g of *sodium dihydrogen phosphate R* in 950 mL of *water R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
- *mobile phase B*: dissolve 140 g of *sodium perchlorate R* in 950 mL of mobile phase A, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with mobile phase A;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	97 → 65	3 → 35
20 - 50	65 → 0	35 → 100
50 - 60	0	100

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 234 nm.

Injection 18 µL of the blank solution, test solution (c) and reference solutions (b) and (c).

Identification of disaccharides Use the chromatogram supplied with *enoxaparin sodium CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to the disaccharides listed in Table 1097.-1; use the chromatogram obtained with reference solution (b) to confirm the identity of the peaks due to the 1,6-anhydro derivatives.

Table 1097.-1. – *Correlation between the relative retention of peaks observed in chromatograms obtained with solutions of depolymerised and reduced enoxaparin, with reference to reduced ΔIS (retention time = about 30 min), and molecular masses of the enoxaparin derivatives*

Derivatives	Relative retention	Molecular mass (Da)
Unidentified	< 0.20	741
Reduced ΔIVA	0.20	401
Unidentified	0.20 - 0.46	741
Reduced ΔIVS	0.46	461
Unidentified	0.46 - 0.48	483
Reduced ΔIIA	0.48	503
Unidentified	0.48 - 0.52	503
1,6-anhydro ΔIIS	0.52	443
Unidentified	0.52 - 0.57	503
Reduced ΔIIIA	0.57	503
Unidentified	0.57 - 0.66	533
Reduced ΔIIS	0.66	563
Unidentified	0.66 - 0.76	563
Reduced ΔIIIS	0.76	563
Unidentified	0.76 - 0.85	583
Reduced ΔIA	0.85	605
1,6-anhydro ΔIS	0.88	545
Unidentified	0.88 - 0.97	635
Reduced ΔIIA-IVSglu	0.97	1066
Reduced ΔIS	1.00	665
ΔIS	1.04	665
Unidentified	1.04 - 1.10	1228
Reduced ΔIIA-IISglu	1.10	1168

Derivatives	Relative retention	Molecular mass (Da)
Unidentified	1.10 - 1.28	1228
1,6-anhydro ΔIS-IS	1.28	1210
Unidentified	> 1.28	1228

NOTE: depending on the resolution of the column, 1,6-anhydro ΔIIS may be eluted in the form of 2 peaks (mannosamine and glucosamine forms), which are both taken into account as 1,6-anhydro ΔIIS.

Relative retention With reference to reduced ΔIS (retention time = about 30 min): see Table 1097.-1.

System suitability:

- *peak area ratio*: maximum 1.15 for the peaks due to 1,6-anhydro ΔIS-IS and 1,6-anhydro ΔIS in the chromatogram obtained with reference solution (b); maximum 0.02 for the peaks due to ΔIS and reduced ΔIS in the chromatogram obtained with reference solution (c);
- *resolution*: minimum 1.5 between the peaks due to reduced ΔIA and 1,6-anhydro ΔIS in the chromatogram obtained with reference solution (c);
- the content of 1,6-anhydro derivatives in *enoxaparin sodium CRS* is within 1.5 per cent of the assigned content.

Calculation:

Calculate the molar percentage of the 3 main 1,6-anhydro derivatives using the relative molecular masses given in Table 1097.-1 and the following expression:

$$100 \times \frac{Mw}{\sum Mw_x \cdot A_x} \times (A_1 + A_2 + A_3)$$

Mw = mass-average relative molecular mass of enoxaparin (as determined by identification test C);
Mw_x = relative molecular mass attributed to derivative x according to Table 1097.-1;
A_x = area of the peak due to derivative x;
A_1 = area of the peak due to 1,6-anhydro ΔIS;
A_2 = area of the peak due to 1,6-anhydro ΔIIS;
A_3 = area of the peak due to 1,6-anhydro ΔIS-IS.

Disregard any peak observed with the blank solution.

Correct the value to the nearest unit.

Limit 15 per cent to 25 per cent of components bearing the 1,6-anhydro structure at the reducing end of their chain.

C. Carry out identification test C as described in the monograph *Low-molecular-mass heparins (0828)*. The following requirements apply.

The mass-average relative molecular mass ranges between 3800 and 5000. The mass percentage of chains lower than 2000 ranges between 12.0 per cent and 20.0 per cent. The mass percentage of chains between 2000 and 8000 ranges between 68.0 per cent and 82.0 per cent.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Dissolve 1.0 g in 10 mL of *water R*.

pH (*2.2.3*)

6.2 to 7.7.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Specific absorbance (*2.2.25*)

14.0 to 20.0 (dried substance), determined at 231 nm.

Dissolve 50.0 mg in 100 mL of *0.01 M hydrochloric acid*.

Benzyl alcohol

Liquid chromatography (*2.2.29*).

Internal standard solution 1 g/L solution of *3,4-dimethylphenol R* in *methanol R*.

Test solution Dissolve about 0.500 g of the substance to be examined in 5.0 mL of *1 M sodium hydroxide*. Allow to stand for 1 h. Add 1.0 mL of *glacial acetic acid R* and 1.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*.

Reference solution Prepare a 0.25 g/L solution of *benzyl alcohol R* in *water R*. Mix 0.50 mL of this solution with 1.0 mL of the internal standard solution and dilute to 10.0 mL with *water R*.

Precolumn:

— *size*: l = 0.02 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Column:

— *size*: l = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, acetonitrile R, water R* (5:15:80 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 256 nm.

Injection 20 µL.

From the chromatogram obtained with the reference solution, calculate the ratio (R_1) of the height of the peak due to benzyl alcohol to the height of the peak due to the internal standard. From the chromatogram obtained with the test solution, calculate the ratio (R_2) of the height of the peak due to benzyl alcohol to the height of the peak due to the internal standard.

Calculate the percentage content *m/m* of benzyl alcohol using the following expression:

$$\frac{0.0125 \times R_2}{m \times R_1}$$

m = mass of the substance to be examined, in grams.

Limit:

— *benzyl alcohol*: maximum 0.1 per cent *m/m*.

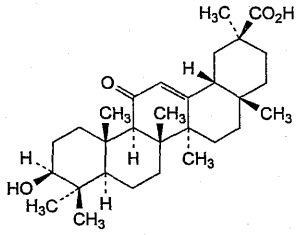
Sodium (*2.2.23, Method I*)

11.3 per cent to 13.5 per cent (dried substance).

Ph Eur

Enoxolone

(Ph. Eur. monograph 1511)



$C_{30}H_{46}O_4$ 470.7 *471-53-4*

Action and use
Treatment of benign peptic ulcer disease.

Ph Eur

DEFINITION
(20β)-3β-Hydroxy-11-oxo-olean-12-en-29-oic acid.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white crystalline powder.

Solubility
Practically insoluble in water, soluble in ethanol, sparingly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A.

Second identification: B, C.

A. Examine by infrared absorption spectrophotometry (*2.2.24*).

Comparison *enoxolone CRS.*

If the spectra obtained in the solid state show differences, dissolve 0.2 g of the substance to be examined and 0.2 g of the reference substance separately in 6 mL of *ethanol R.* Boil under a reflux condenser for 1 h and add 6 mL of *water R.* A precipitate is formed. Cool to about 10 °C and filter with the aid of vacuum. Wash the precipitate with 10 mL of *alcohol R,* dry in an oven at 80 °C and record new spectra.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *enoxolone CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, acetone R, methylene chloride R* (5:10:90 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air for 5 min.

Detection Spray with *anisaldehyde solution R* and heat at 100-105 °C for 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 50 mg in 10 mL of *methylene chloride R.* To 2 mL of this solution, add 1 mL of *acetic anhydride R* and 0.3 mL of *sulfuric acid R.* A pink colour is produced.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Dissolve 0.1 g in *ethanol R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 145 to + 154 (dried substance).

Dissolve 0.50 g in *dioxan R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.10 g of the substance to be examined in the mobile phaseand dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 0.1 g of *18α-glycyrrhetinic acid R* in *tetrahydrofuran R* and dilute to 100.0 mL with the same solvent. To 2.0 mL of the solution, add 2.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm),
— *temperature:* 30 °C.

Mobile phase Mix 430 volumes of *tetrahydrofuran R* and 570 volumes of a 1.36 g/L solution of *sodium acetate R* adjusted to pH 4.8 with *glacial acetic acid R.*

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 20 µL loop injector; inject the test solution and the reference solutions.

Run time 4 times the retention time of enoxolone.

System suitability:
— *resolution:* minimum of 2.0 between the peaks due to enoxolone and to 18α-glycyrrhetinic acid in the chromatogram obtained with reference solution (c).

Limits:
— *any impurity:* not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent),
— *total:* not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent),
— *disregard limit:* 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.330 g in 40 mL of *dimethylformamide R.* Titrate with 0.1 M tetrabutylammonium hydroxide, determining the

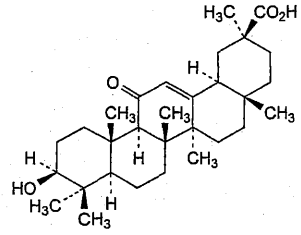
end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 47.07 mg of $C_{30}H_{46}O_4$.

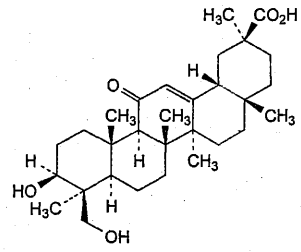
STORAGE

Protected from light.

IMPURITIES



A. (20β)-3β-hydroxy-11-oxo-18α-olean-12-en-29-oic acid,

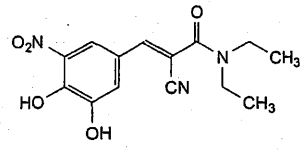


B. (4β,20β)-3β,23-dihydroxy-11-oxo-olean-12-en-29-oic acid.

Ph Eur

Entacapone

(Ph. Eur. monograph 2574)



$C_{14}H_{15}N_3O_5$ 305.3 *130929-57-6*

Action and use

Catechol-O-methyl transferase inhibitor; treatment of Parkinson's disease.

Ph Eur

DEFINITION

(2*E*)-2-Cyano-3-(3,4-dihydroxy-5-nitrophenyl)-*N*,*N*-diethylprop-2-enamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Greenish-yellow or yellow powder.

Solubility

Practically insoluble in water, soluble or sparingly soluble in acetone, slightly soluble in anhydrous ethanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *entacapone CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions.*

Solvent mixture *tetrahydrofuran R, methanol R* (30:70 *V/V*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *entacapone impurity A CRS* in the solvent mixture, add 5.0 mL of test solution (a) and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (b) to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *entacapone CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped propyl-2-phenylsilyl amorphous organosilica polymer R* (5 µm).

Mobile phase Mix 2 volumes of *tetrahydrofuran R*, 44 volumes of *methanol R* and 54 volumes of a 2.34 g/L solution of *sodium dihydrogen phosphate R* previously adjusted to pH 2.1 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 300 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time 2.5 times the retention time of entacapone.

Relative retention With reference to entacapone (retention time = about 17 min): impurity A = about 0.8.

System suitability Reference solution (a):

— *resolution*: minimum 3.0 between the peaks due to impurity A and entacapone.

Limits:

— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *sum of impurities other than A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{14}H_{15}N_3O_5$ from the declared content of *entacapone CRS*.

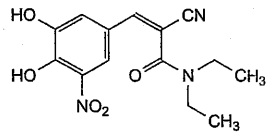
STORAGE

Protected from light.

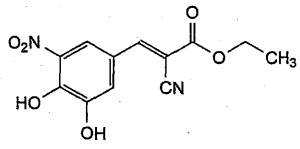
IMPURITIES

Specified impurities *A.*

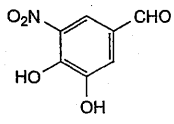
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H, I.*



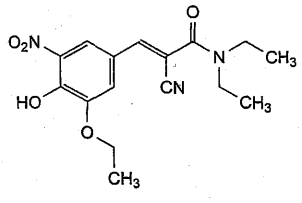
A. (2*Z*)-2-cyano-3-(3,4-dihydroxy-5-nitrophenyl)-*N*,*N*-diethylprop-2-enamide,



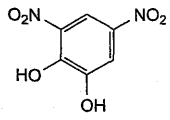
B. ethyl (2*E*)-2-cyano-3-(3,4-dihydroxy-5-nitrophenyl)prop-2-enoate,



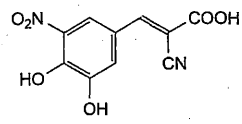
C. 3,4-dihydroxy-5-nitrobenzaldehyde,



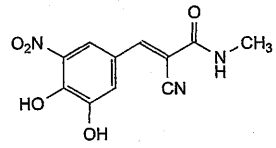
D. (2*E*)-2-cyano-3-(3-ethoxy-4-hydroxy-5-nitrophenyl)-*N*,*N*-diethylprop-2-enamide,



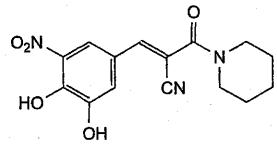
E. 3,5-dinitrobenzene-1,2-diol,



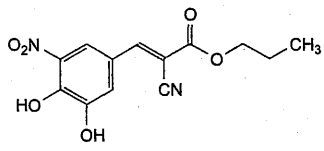
F. (2*E*)-2-cyano-3-(3,4-dihydroxy-5-nitrophenyl)prop-2-enoic acid,



G. (2*E*)-2-cyano-3-(3,4-dihydroxy-5-nitrophenyl)-*N*-methylprop-2-enamide,



H. (2*E*)-3-(3,4-dihydroxy-5-nitrophenyl)-2-(piperidin-1-ylcarbonyl)prop-2-ennitrile,

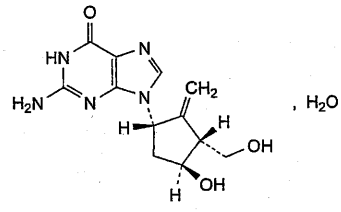


I. propyl (2*E*)-2-cyano-3-(3,4-dihydroxy-5-nitrophenyl)prop-2-enoate.

Ph Eur

Entecavir Monohydrate

(Ph. Eur. monograph 2815)



$C_{12}H_{15}N_5O_3,H_2O$ 295.3 *209216-23-9*

Ph Eur

DEFINITION

2-Amino-9-[(1*S*,3*R*,4*S*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-1,9-dihydro-6*H*-purin-6-one monohydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, in anhydrous ethanol and in heptane, slightly soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *entecavir monohydrate CRS.*

If the spectra obtained show differences, suspend 50 mg of the substance to be examined and 50 mg of the reference substance separately in 5 mL of hot *water R*. Swirl protected from light and heat until a clear solution is obtained. Allow to cool and filter the precipitates *in vacuo*. Dry the precipitates for 12 h in a desiccator and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)

+ 24 to + 30 (anhydrous substance), measured at 25 °C.

Dissolve 0.25 g in 10 mL of a mixture of equal volumes of *dimethylformamide R* and *methanol R* and dilute to 25.0 mL with the same mixture of solvents.

Impurity F

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in 10 mL of *methanol R* using sonication, and dilute to 25.0 mL with the same solvent.

Reference solution Dissolve 2.5 mg of *entecavir impurity F CRS* in 20 mL of *methanol R* using sonication, and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with *methanol R*.

Column:
- *size*: *l* = 0.05 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: 0.1 per cent *V/V* solution of *trifluoroacetic acid R*;
- *mobile phase B*: 0.1 per cent *V/V* solution of *trifluoroacetic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	65 → 53	35 → 47

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Retention time Impurity F = about 6 min.

Calculation of percentage content:
- for impurity F, use the concentration of impurity F in the reference solution.

Limit:
- *impurity F*: maximum 0.10 per cent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in 10 mL of *methanol R* using sonication, and dilute to 25.0 mL with the same solvent. Dilute 2.0 mL of the solution to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 25.0 mg of *entecavir monohydrate CRS* in 10 mL of *methanol R* using sonication, and dilute to 25.0 mL with the same solvent. Dilute 2.0 mL of the solution to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of *entecavir for system suitability CRS* (containing impurities A and C) in 1 mL of *methanol R* using sonication, and dilute to 5.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: *acetonitrile R, water for chromatography R* (3:97 *V/V*);
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	100	0
8 - 50	100 → 77	0 → 23
50 - 75	77 → 17	23 → 83

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *entecavir for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and C.

Relative retention With reference to entecavir (retention time = about 21 min): impurity A = about 0.9; impurity C = about 1.03.

System suitability Reference solution (c):
- *resolution*: minimum 3.5 between the peaks due to impurity A and entecavir; minimum 2.0 between the peaks due to entecavir and impurity C.

Calculation of percentage contents:
- for each impurity, use the concentration of entecavir monohydrate in reference solution (b).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *reporting threshold*: 0.05 per cent; disregard any peak with a relative retention with reference to entecavir of about 3.4 (impurity F).

Water (*2.5.32*)

5.5 per cent to 7.0 per cent, determined on 30.0 mg by direct sample introduction.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{12}H_{15}N_5O_3$ taking into account the assigned content of *entecavir monohydrate CRS*.

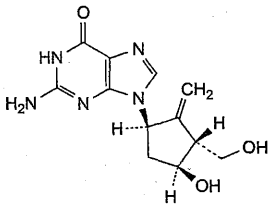
STORAGE

Protected from light.

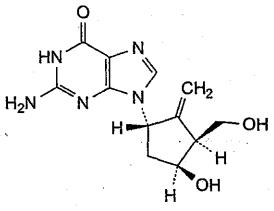
IMPURITIES

Specified impurities *F.*

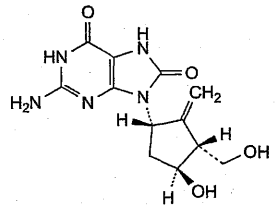
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E.*



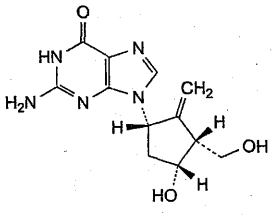
A. 2-amino-9-[(1*R*,3*R*,4*S*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-1,9-dihydro-6*H*-purin-6-one,



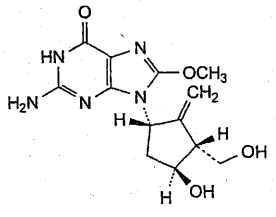
B. 2-amino-9-[(1*S*,3*S*,4*S*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-1,9-dihydro-6*H*-purin-6-one,



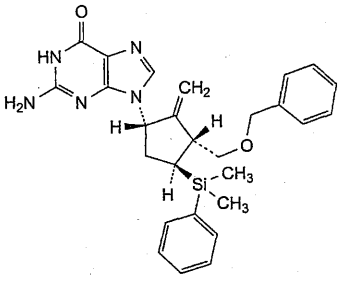
C. 2-amino-9-[(1*S*,3*R*,4*S*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-7,9-dihydro-1*H*-purine-6,8-dione,



D. 2-amino-9-[(1*S*,3*R*,4*R*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-1,9-dihydro-6*H*-purin-6-one,



E. 2-amino-9-[(1*S*,3*R*,4*S*)-4-hydroxy-3-(hydroxymethyl)-2-methylidenecyclopentyl]-8-methoxy-1,9-dihydro-6*H*-purin-6-one,



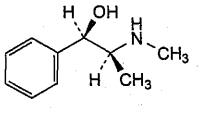
F. 2-amino-9-[(1*S*,3*R*,4*S*)-3-[(benzyloxy)methyl]-4-[dimethyl(phenyl)silyl]-2-methylidenecyclopentyl]-1,9-dihydro-6*H*-purin-6-one.

Ph Eur

Ephedrine

Anhydrous Ephedrine

(Ph. Eur. monograph 0488)



$C_{10}H_{15}NO$ 165.2 *299-42-3*

Action and use
Adrenoceptor agonist.

Ph Eur

DEFINITION
Ephedrine contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (1*R*,2*S*)-2-methylamino-1-phenylpropan-1-ol, calculated with reference to the anhydrous substance.

CHARACTERS
A white or almost white, crystalline powder or colourless crystals, soluble in water, very soluble in alcohol.

It melts at about 36 °C.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with the base isolated from *ephedrine hydrochloride CRS*. Examine the substances in discs prepared as follows: dissolve 40 mg of the substance to be examined in 1 mL of *water R*, add 1 mL of *dilute sodium hydroxide solution R* and 4 mL of *chloroform R* and shake; dry the organic layer over 0.2 g of *anhydrous sodium sulfate R*; prepare a blank disc using about 0.3 g of *potassium bromide R*; apply dropwise to the disc 0.1 mL of the organic layer, allowing the solvent to evaporate between applications; dry the disc at 50 °C for 2 min. Repeat the operations using 50 mg of *ephedrine hydrochloride CRS*.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 1 mL of *water R*. Add 0.2 mL of *strong sodium hydroxide solution R* and 0.2 mL of *copper sulfate solution R*. A violet colour is produced. Add 2 mL of *ether R* and shake. The ether layer is purple and the aqueous layer blue.

E. Water (see Tests).

TESTS

Appearance of solution

Dissolve 0.25 g in *water R* and dilute to 10 mL with the same solvent. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

Dissolve 2.25 g in 15 mL of *dilute hydrochloric acid R* and dilute to 50.0 mL with *water R*. The specific optical rotation is −41 to −43, calculated with reference to the anhydrous substance.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.2 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dissolve 25 mg of *ephedrine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 200 mL with *methanol R*.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 5 volumes of *chloroform R*, 15 volumes of *concentrated ammonia R* and 80 volumes of *2-propanol R*. Allow the plate to dry in air and spray with *ninhydrin solution R*. Heat at 110 °C for 5 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). Disregard any spot of lighter colour than the background.

Chlorides

Dissolve 0.17 g in 10 mL of *water R*. Add 5 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R1*. Allow to stand for 2 min, protected from bright light. Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of *chloride standard solution (5 ppm Cl) R*, 5 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R1* (290 ppm).

Water (*2.5.12*)

Not more than 0.5 per cent, determined on 2.000 g by the semi-micro determination of water.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 5 mL of *alcohol R* and add 20.0 mL of *0.1 M hydrochloric acid*. Using 0.05 mL of *methyl red solution R* as indicator, titrate with *0.1 M sodium hydroxide* until a yellow colour is obtained.

1 mL of *0.1 M hydrochloric acid* is equivalent to 16.52 mg of $C_{10}H_{15}NO$.

STORAGE

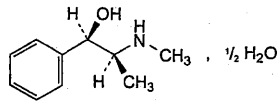
Store protected from light.

Ph Eur

Ephedrine Hemihydrate

(Ph. Eur. monograph 0489)

NOTE: The name Ephedrine was formerly used in the United Kingdom.



$C_{10}H_{15}NO,½H_2O$ 174.2 *50906-05-3*

Action and use

Adrenoceptor agonist.

Ph Eur

DEFINITION

Ephedrine hemihydrate contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (1*R*,2*S*)-2-(methylamino)-1-phenylpropan-1-ol, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, soluble in water, very soluble in alcohol.

It melts at about 42 °C, determined without previous drying.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with the base isolated from *ephedrine hydrochloride CRS*. Examine the substances in discs prepared as follows: dissolve 40 mg of the substance to be examined in 1 mL of *water R*, add 1 mL of *dilute sodium hydroxide solution R* and 4 mL of *chloroform R* and shake; dry the organic layer over 0.2 g of *anhydrous sodium sulfate R*; prepare a blank disc using about 0.3 g of *potassium bromide R*; apply dropwise to the disc 0.1 mL of the organic layer, allowing the solvent to evaporate between applications; dry the disc at 50 °C for 2 min. Repeat the operations using 50 mg of *ephedrine hydrochloride CRS*.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 1 mL of *water R*. Add 0.2 mL of *strong sodium hydroxide solution R* and 0.2 mL of *copper sulfate solution R*. A violet colour is produced. Add 2 mL of *ether R* and shake. The ether layer is purple and the aqueous layer blue.

E. Water (see Tests).

TESTS

Appearance of solution

Dissolve 0.25 g in *water R* and dilute to 10 mL with the same solvent. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

Dissolve 2.25 g in 15 mL of *dilute hydrochloric acid R* and dilute to 50.0 mL with *water R*. The specific optical rotation is −41 to −43, calculated with reference to the anhydrous substance.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.2 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dissolve 25 mg of *ephedrine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 200 mL with *methanol R*.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 5 volumes of *chloroform R*, 15 volumes of *concentrated ammonia R* and 80 volumes of *2-propanol R*. Allow the plate to dry in air and spray with *ninhydrin solution R*. Heat at 110 °C for 5 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). Disregard any spot of lighter colour than the background.

Chlorides
Dissolve 0.18 g in 10 mL of *water R*. Add 5 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R1*. Allow to stand for 2 min, protected from bright light. Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of *chloride standard solution (5 ppm Cl) R*, 5 mL of *dilute nitric acid R* and 0.5 mL of *silver nitrate solution R1* (280 ppm).

Water (*2.5.12*)
4.5 per cent to 5.5 per cent, determined on 0.300 g by the semi-micro determination of water.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 5 mL of *alcohol R* and add 20.0 mL of *0.1 M hydrochloric acid*. Using 0.05 mL of *methyl red solution R* as indicator, titrate with *0.1 M sodium hydroxide* until a yellow colour is obtained.

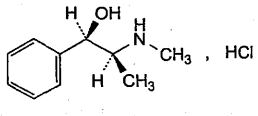
1 mL of *0.1 M hydrochloric acid* is equivalent to 16.52 mg of $C_{10}H_{15}NO$.

STORAGE
Store protected from light.

Ph Eur

Ephedrine Hydrochloride

(Ph. Eur. monograph 0487)



$C_{10}H_{16}ClNO$ 201.7 *50-98-6*

Action and use
Adrenoceptor agonist.

Preparations
Ephedrine Elixir
Ephedrine Nasal Drops
Ephedrine Hydrochloride Tablets
Ephedrine Injection

Ph Eur

DEFINITION
(1*R*,2*S*)-2-(Methylamino)-1-phenylpropan-1-ol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent).

mp
About 219 °C.

IDENTIFICATION
First identification: B, E.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ephedrine hydrochloride CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *ephedrine hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *methylene chloride R, concentrated ammonia R, 2-propanol R* (5:15:80 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R*; heat at 110 °C for 5 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 mL of solution S (see Tests) add 1 mL of *water R*, 0.2 mL of *copper sulfate solution R* and 1 mL of *strong sodium hydroxide solution R*. A violet colour is produced. Add 2 mL of *methylene chloride R* and shake. The lower (organic) layer is dark grey and the upper (aqueous) layer is blue.

E. To 5 mL of solution S (see Tests) add 5 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 5.00 g in *distilled water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.4 mL of *0.01 M hydrochloric acid*. The solution is red.

Specific optical rotation (*2.2.7*)
−33.5 to −35.5 (dried substance).

Dilute 12.5 mL of solution S to 25.0 mL with *water R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 75 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *pseudoephedrine hydrochloride CRS* in the mobile phase and dilute to 50 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *phenylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Mix 6 volumes of *methanol R* and 94 volumes of a 11.6 g/L solution of *ammonium acetate R* adjusted to pH 4.0 with *glacial acetic acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 257 nm.

Injection 20 µL.

Run time 2.5 times the retention time of ephedrine.

Relative retention With reference to ephedrine (retention time = about 8 min): impurity B = about 1.1; impurity A = about 1.4.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to ephedrine and impurity B.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.4;
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *sum of impurities other than A*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfates (*2.4.13*)
Maximum 100 ppm, determined on solution S.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

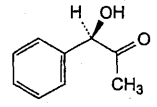
ASSAY
Dissolve 0.150 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 20.17 mg of $C_{10}H_{16}ClNO$.

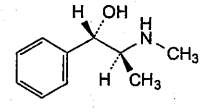
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B*.



A. (–)-(1*R*)-1-hydroxy-1-phenylpropan-2-one,

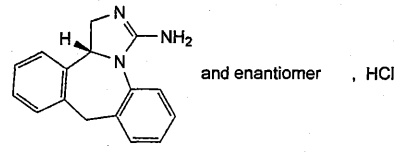


B. (1*S*,2*S*)-2-(methylamino)-1-phenylpropan-1-ol (pseudoephedrine).

Ph Eur

Epinastine Hydrochloride

(Ph. Eur. monograph 2411)



$C_{16}H_{16}ClN_3$ 285.8 *108929-04-0*

Action and use
Antihistamine.

Ph Eur

DEFINITION
(13b*RS*)-9,13b-Dihydro-1*H*-dibenzo[*c*,*f*]imidazo[1,5-*a*]azepin-3-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water and in methanol, sparingly soluble in methylene chloride, slightly soluble in acetonitrile.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison: *epinastine hydrochloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Acidity or alkalinity

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Add 0.1 mL of *methyl red mixed solution R* and 0.25 mL of *0.01 M sodium hydroxide*. The solution is green. Add 0.5 mL of *0.01 M hydrochloric acid*. The solution is reddish-violet.

Related substances

Liquid chromatography (*2.2.29*).

Buffer solution pH 4.4 Dissolve 3.8 g of *sodium pentanesulfonate monohydrate R* and 4.0 g of *potassium dihydrogen phosphate R* in *water R*, adjust to pH 4.4 with *phosphoric acid R* and dilute to 1000.0 mL with *water R*.

Solvent mixture Mobile phase B, mobile phase A (25:75 *V/V*).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 10.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *epinastine for system suitability CRS* (containing impurities A and B) in 10.0 mL of the solvent mixture.

Column:
- *size*: *l* = 0.10 m, Ø = 3.0 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 50 °C.

Mobile phase :
- *mobile phase A*: *methanol R2*, buffer solution pH 4.4 (15:85 *V/V*);
- *mobile phase B*: *methanol R2*, *acetonitrile R1* (15:85 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	80	20
4 - 13	80 → 30	20 → 70

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *epinastine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to epinastine (retention time = about 4 min): impurity A = about 1.2; impurity B = about 2.0.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to epinastine.

Limits:
- *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 100 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 2 volumes of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

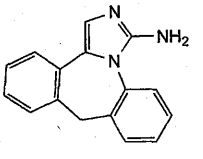
1 mL of *0.1 M perchloric acid* is equivalent to 28.58 mg of $C_{16}H_{16}ClN_3$.

STORAGE

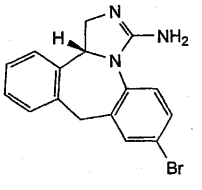
In an airtight container.

IMPURITIES

Specified impurities *A, B.*



A. 9*H*-dibenzo[*c*,*f*]imidazo[1,5-*a*]azepin-3-amine,



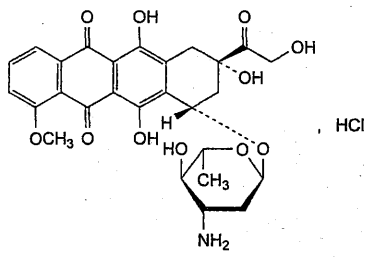
and enantiomer

B. (13b*RS*)-7-bromo-9,13b-dihydro-1*H*-dibenzo[*c*,*f*]imidazo[1,5-*a*]azepin-3-amine.

Ph Eur

Epirubicin Hydrochloride

(Ph. Eur. monograph 1590)



$C_{27}H_{30}ClNO_{11}$ 580.0 *56390-09-1*

Action and use
Cytotoxic.

Preparation
Epirubicin Injection

Ph Eur

DEFINITION

(8*S*,10*S*)-10-[(3-Amino-2,3,6-trideoxy-α-L-*arabino*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione hydrochloride.

Substance obtained by chemical transformation of a substance produced by certain strains of *Streptomyces peucetius*.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Orange-red powder.

Solubility
Soluble in water and in methanol, slightly soluble in anhydrous ethanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *epirubicin hydrochloride CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 0.5 mL of *nitric acid R*, add 0.5 mL of *water R* and heat over a flame for 2 min. Allow to cool and add 0.5 mL of *silver nitrate solution R1*. A white precipitate is formed.

TESTS

pH (*2.2.3*)
4.0 to 5.5.

Dissolve 50 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Allow the solutions to stand for 3 h before use.*

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *epirubicin hydrochloride CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of *epirubicin hydrochloride CRS* and 10 mg of *doxorubicin hydrochloride CRS* in the mobile phase and dilute to 100 mL with the mobile phase.

Reference solution (c) Dissolve 10 mg of *doxorubicin hydrochloride CRS* in a mixture of 5 mL of *water R* and 5 mL of *phosphoric acid R*. Allow to stand for 30 min. Adjust to pH 2.6 with an 80 g/L solution of *sodium hydroxide R*. Add 15 mL of *acetonitrile R* and 10 mL of *methanol R*. Mix.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *trimethylsilyl silica gel for chromatography R* (6 µm);
— *temperature*: 35 °C.

Mobile phase Mix 17 volumes of *methanol R*, 29 volumes of *acetonitrile R* and 54 volumes of a solution containing 3.7 g/L of *sodium laurilsulfate R* and 2.8 per cent *V/V* of *dilute phosphoric acid R*.

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Run time 3.5 times the retention time of epirubicin.

Identification of impurities Use the 2nd most abundant peak present in the chromatogram obtained with reference solution (c) to identify impurity A.

Relative retention With reference to epirubicin (retention time = about 9.5 min): impurity A = about 0.3; impurity B = about 0.4; impurity C = about 0.8; impurity E = about 1.1; impurity D = about 1.5; impurity F = about 1.7; impurity G = about 2.1.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity C and epirubicin.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.7;
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (1.0 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (1.0 per cent);
— *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (d) (2.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Acetone (*2.4.24*)
Maximum 1.5 per cent.

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 0.100 g.

Bacterial endotoxins (*2.6.14*)
Less than 1.1 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

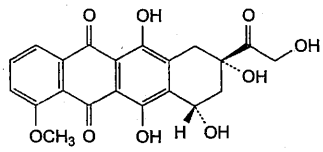
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

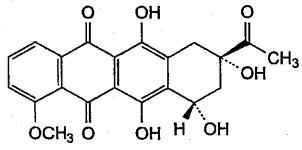
Calculate the percentage content of $C_{27}H_{30}ClNO_{11}$.

STORAGE
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

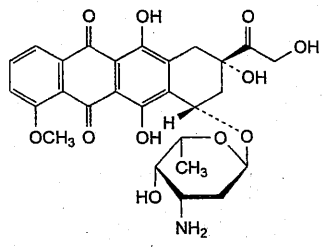
IMPURITIES



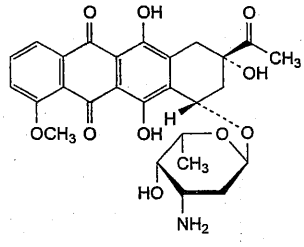
A. (8*S*,10*S*)-6,8,10,11-tetrahydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (doxorubicinone),



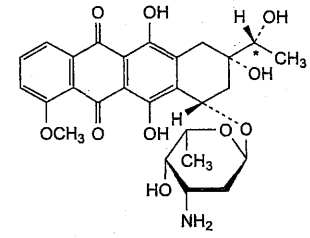
B. (8*S*,10*S*)-8-acetyl-6,8,10,11-tetrahydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (daunorubicinone),



C. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (doxorubicin),

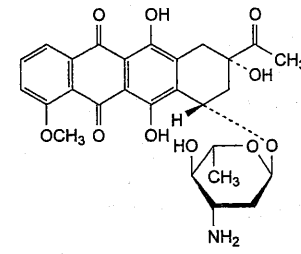


D. (8*S*,10*S*)-8-acetyl-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (daunorubicin),

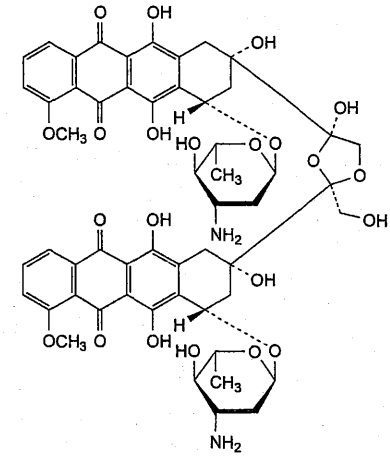


and epimer at C*

E. (8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*lyxo*-hexopyranosyl)oxy]-6,8,11-trihydroxy-8-[(1*RS*)-1-hydroxyethyl]-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (dihydrodaunorubicin),



F. (8*S*,10*S*)-8-acetyl-10-[(3-amino-2,3,6-trideoxy-α-L-*arabino*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione (*epi*-daunorubicin),

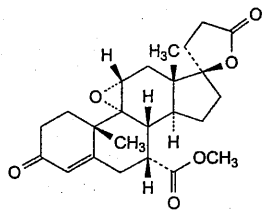


G. 8,8′-[(2*R*,4*R*)-4-hydroxy-2-(hydroxymethyl)-1,3-dioxolan-2,4-diyl]bis[(8*S*,10*S*)-10-[(3-amino-2,3,6-trideoxy-α-L-*arabino*-hexopyranosyl)oxy]-6,8,11-trihydroxy-1-methoxy-7,8,9,10-tetrahydrotetracene-5,12-dione] (epirubicin dimer).

Ph Eur

Eplerenone

(Ph. Eur. monograph 2765)



$C_{24}H_{30}O_6$ 414.5 *107724-20-9*

Action and use
Aldosterone receptor antagonist; antihypertensive.

Ph Eur

DEFINITION

9,11α-Epoxy-7α-(methoxycarbonyl)-3-oxo-17α-pregn-4-ene-21,17-carbolactone.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White, almost white or slightly yellow, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *eplerenone CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)
−16.0 to −14.0.

Dissolve 0.250 g in *acetonitrile R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, methanol R, water R* (25:25:50 *V/V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *eplerenone for system suitability CRS* (containing impurities A and D) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *eplerenone for peak identification CRS* (containing impurity B) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 25.0 mg of *eplerenone CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: 0.1 per cent *V/V* solution of *phosphoric acid R*;
— *mobile phase B*: *phosphoric acid R, acetonitrile R, methanol R* (0.1:40:60 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	54	46
25 - 32	54 → 40	46 → 60
32 - 45	40	60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with *eplerenone for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and D; use the chromatogram supplied with *eplerenone for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to eplerenone (retention time = about 9 min): impurity D = about 0.71; impurity A = about 0.74; impurity B = about 1.2.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity A.

Calculation of percentage contents:
— for each impurity, use the concentration of eplerenone in reference solution (c).

Limits:
— *impurities A, B*: for each impurity, maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.6 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

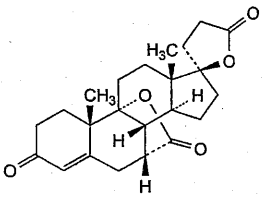
Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{24}H_{30}O_6$ taking into account the assigned content of *eplerenone CRS*.

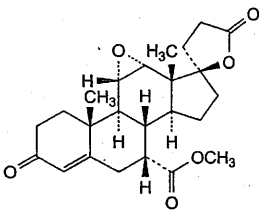
IMPURITIES

Specified impurities *A, B.*

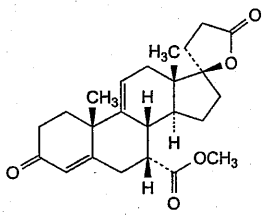
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) C, D, E, F, G.



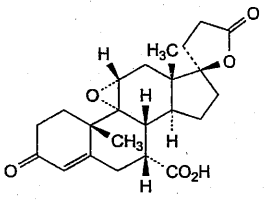
A. 3-oxo-17α-pregn-4-ene-7α,9:21,17-dicarbolactone,



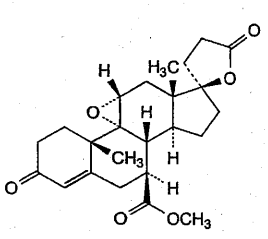
B. 11α,12α-epoxy-7α-(methoxycarbonyl)-3-oxo-17α-pregn-4-ene-21,17-carbolactone,



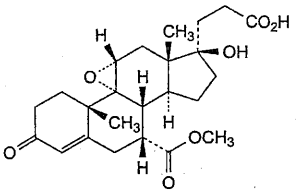
C. 7α-(methoxycarbonyl)-3-oxo-17α-pregna-4,9(11)-diene-21,17-carbolactone,



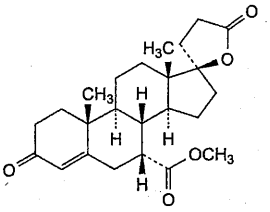
D. (2′*R*)-9,11α-epoxy-3,5′-dioxo-4′,5′-dihydro-3′*H*-spiro[androst-4-ene-17,2′-furan]-7α-carboxylic acid,



E. 9,11α-epoxy-7β-(methoxycarbonyl)-3-oxo-17α-pregn-4-ene-21,17-carbolactone,



F. 9,11α-epoxy-17-hydroxy-7α-(methoxycarbonyl)-3-oxo-17α-pregn-4-ene-21-carboxylic acid,

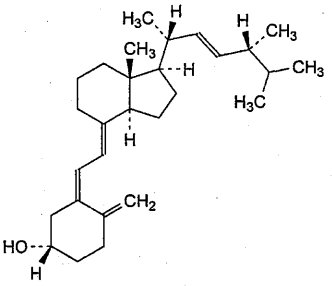


G. 7α-(methoxycarbonyl)-3-oxo-17α-pregn-4-ene-21,17-carbolactone.

Ph Eur

Ergocalciferol

(Ph. Eur. monograph 0082)



$C_{28}H_{44}O$ 396.7 *50-14-6*

Action and use

Vitamin D analogue (Vitamin D_2).

Preparations

Calcium and Ergocalciferol Tablets

Calcium and Ergocalciferol Chewable Tablets

Ergocalciferol Injection

Ergocalciferol Tablets

When vitamin D_2 is prescribed or demanded, Ergocalciferol shall be dispensed or supplied. When calciferol or vitamin D is prescribed or demanded, Ergocalciferol or Colecalciferol shall be dispensed or supplied.

Ph Eur

DEFINITION

(3*S*,5*Z*,7*E*,22*E*)-9,10-Secoergosta-5,7,10(19),22-tetraen-3-ol.

Content

97.0 per cent to 102.0 per cent.

A suitable antioxidant may be added.

A reversible isomerisation to pre-ergocalciferol takes place in solution, depending on temperature and time. The activity is due to both compounds (see Assay).

1 mg of ergocalciferol is equivalent to 40 000 IU of antirachitic activity (vitamin D) in rats.

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder or white or almost white crystals.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent) and in methanol, soluble in fatty oils.

It is sensitive to air, heat and light.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ergocalciferol CRS.*

TESTS

Specific optical rotation (*2.2.7*)

+ 103 to + 107.

Dissolve 0.200 g rapidly and without heating in *aldehyde-free alcohol R* and dilute to 25.0 mL with the same solvent. Examine within 30 min of preparing the solution.

Reducing substances

Dissolve 0.1 g in *aldehyde-free alcohol R* and dilute to 10.0 mL with the same solvent. Add 0.5 mL of a 5 g/L solution of *tetrazolium blue R* in *aldehyde-free alcohol R* and 0.5 mL of *dilute tetramethylammonium hydroxide solution R.* Allow to stand for exactly 5 min and add 1.0 mL of *glacial acetic acid R.* Prepare a reference solution at the same time and in the same manner using 10.0 mL of a solution containing 0.2 µg/mL of *hydroquinone R* in *aldehyde-free alcohol R.* Measure the absorbance (*2.2.25*) of the 2 solutions at 525 nm using as the compensation liquid 10.0 mL of *aldehyde-free alcohol R* treated in the same manner.
The absorbance of the test solution is not greater than that of the reference solution (20 ppm).

Impurity B

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use, avoiding exposure to actinic light and air.*

Test solution Dissolve 25.0 mg of the substance to be examined without heating in *methanol R* and dilute to 25.0 mL with the same solvent.

Reference solution Dissolve 5.0 mg of *ergosterol CRS* (impurity B) without heating in *methanol R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with *methanol R.*

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 25 °C.

Mobile phase *methanol R.*

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 20 µL.

Run time 2.5 times the retention time of ergocalciferol.

Relative retention With reference to ergocalciferol (retention time = about 7 min): impurity B = about 1.6.

Limit:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (0.2 per cent).

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use, avoiding exposure to actinic light and air.*

Test solution Dissolve 15.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *ergocalciferol for system suitability CRS* (containing impurities A, F and G) in 10 mL of the mobile phase. Heat in a water-bath at 90 °C under a reflux-condenser for 45 min and allow to cool (*in-situ* degradation to obtain pre-ergocalciferol). Dilute 3 mL of the solution to 25 mL with the mobile phase.

Reference solution (b) Dissolve 15.0 mg of *ergocalciferol CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 25 °C.

Mobile phase *methanol R, acetonitrile R* (10:90 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 20 µL of the test solution and reference solutions (a), (c) and (d).

Run time Twice the retention time of ergocalciferol.

Identification of impurities Use the chromatogram supplied with *ergocalciferol for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, F and G, and pre-ergocalciferol.

Relative retention With reference to ergocalciferol (retention time = about 13 min): impurity F = about 0.6; impurity A = about 0.8; pre-ergocalciferol = about 0.9; impurity G = about 1.2.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity A and pre-ergocalciferol; minimum 2.5 between the peaks due to pre-ergocalciferol and ergocalciferol.

Calculation of percentage contents:
— for impurities A, F and G, use the concentration of ergocalciferol in reference solution (c);
— for impurities other than A, F and G, use the concentration of ergocalciferol in reference solution (d).

Limits:
— *impurity G*: maximum 1.5 per cent;
— *impurities A, F*: for each impurity, maximum 0.5 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 2.0 per cent;
— *reporting threshold*: 0.05 per cent; disregard any peak due to pre-ergocalciferol or the antioxidant.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{28}H_{44}O$ taking into account the assigned content of *ergocalciferol CRS*, and if present, the peak due to pre-ergocalciferol in the test solution.

STORAGE

Under an inert gas, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

The contents of an opened container are to be used immediately.

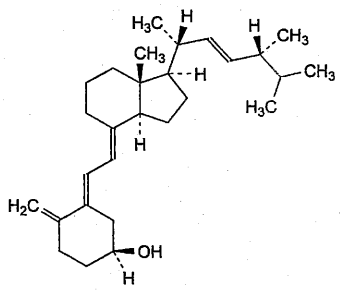
LABELLING

The label states the name and concentration of any added antioxidant.

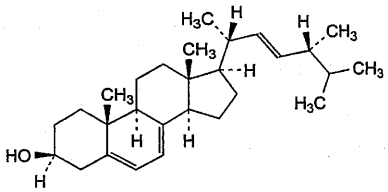
IMPURITIES

Specified impurities *A, B, F, G.*

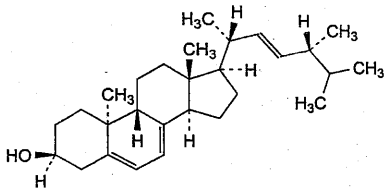
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E.*



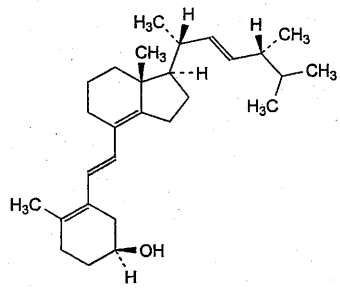
A. (3*S*,5*E*,7*E*,22*E*)-9,10-secoergosta-5,7,10(19),22-tetraen-3-ol (*trans*-vitamin D_2),



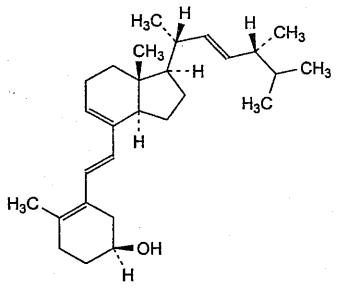
B. (22*E*)-ergosta-5,7,22-trien-3β-ol (ergosterol),



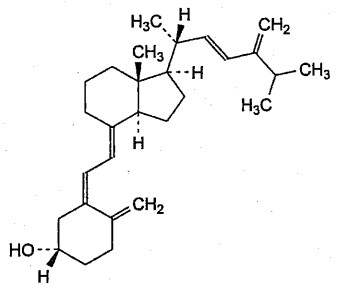
C. (22*E*)-9β,10α-ergosta-5,7,22-trien-3β-ol (lumisterol$_2$),



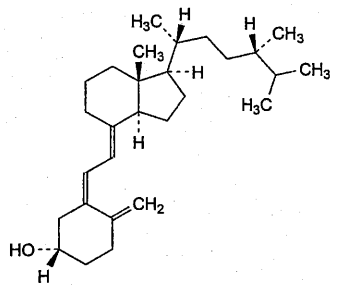
D. (3*S*,6*E*,22*E*)-9,10-secoergosta-5(10),6,8(14),22-tetraen-3-ol (iso-tachysterol$_2$),



E. (3*S*,6*E*,22*E*)-9,10-secoergosta-5(10),6,8,22-tetraen-3-ol (tachysterol$_2$),



F. (3*S*,5*Z*,7*E*,22*E*)-9,10-secoergosta-5,7,10(19),22,24(24^1)-pentaen-3-ol,

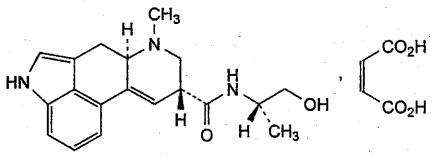


G. (3*S*,5*Z*,7*E*)-9,10-secoergosta-5,7,10(19)-trien-3-ol (vitamin D_4).

Ph Eur

Ergometrine Maleate

(Ph. Eur. monograph 0223)



$C_{23}H_{27}N_3O_6$ 441.5 *129-51-1*

Action and use

Oxytocic.

Preparations

Ergometrine Injection

Ergometrine and Oxytocin Injection

Ph Eur

DEFINITION

Ergometrine maleate contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of (6a*R*,9*R*)-

N-[(*S*)-2-hydroxy-1-methylethyl]-7-methyl-4,6,6a,7,8,9-hexahydro-indolo[4,3-*fg*]quinoline-9-carboxamide (*Z*)-butenedioate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white or slightly coloured, crystalline powder, sparingly soluble in water, slightly soluble in alcohol.

IDENTIFICATION

First identification: B, C.

Second identification: A, C, D, E.

A. Dissolve 30 mg in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with *0.01 M hydrochloric acid.* Examined between 250 nm and 360 nm (*2.2.25*), the solution shows an absorption maximum at 311 nm and a minimum at 265 nm to 272 nm. The specific absorbance at the maximum is 175 to 195.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *ergometrine maleate CRS*. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 0.1 mL of solution S (see Tests) add 1 mL of *glacial acetic acid R*, 0.05 mL of *ferric chloride solution R1* and 1 mL of *phosphoric acid R* and heat in a water-bath at 80 °C. After about 10 min, a blue or violet colour develops which becomes more intense on standing.

E. Dissolve 0.1 g in a mixture of 0.5 mL of *dilute sulfuric acid R* and 2.5 mL of *water R*. Add 5 mL of *ether R* and 1 mL of *strong sodium hydroxide solution R* and shake. Separate the aqueous layer and shake with two quantities, each of 5 mL, of *ether R*. To 0.1 mL of the aqueous layer add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min. No colour develops. To the rest of the aqueous layer add 1 mL of *bromine water R*. Heat on a water-bath for 10 min, then heat to boiling and cool. To 0.2 mL of this solution add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min. A pinkish-violet colour develops.

TESTS

Solution S

Dissolve 0.100 g, without heating and protected from light, in 9 mL of *carbon dioxide-free water R* and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 or BY_5 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of solution S is 3.6 to 4.4.

Specific optical rotation (*2.2.7*)

+ 50 to + 56, determined on solution S and calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using *silica gel G R* as the coating substance. *Carry out all operations as rapidly as possible, protected from light. Prepare the test and reference solutions immediately before use.*

Test solution (a) Dissolve 50 mg of the substance to be examined in a mixture of 1 volume of *concentrated ammonia R* and 9 volumes of *alcohol (80 per cent V/V) R* and dilute to 5.0 mL with the same mixture of solvents.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with a mixture of 1 volume of *concentrated ammonia R* and 9 volumes of *alcohol (80 per cent V/V) R.*

Reference solution (a) Dissolve 10 mg of *ergometrine maleate CRS* in a mixture of 1 volume of *concentrated ammonia R* and 9 volumes of *alcohol (80 per cent V/V) R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with a mixture of 1 volume of *concentrated ammonia R* and 9 volumes of *alcohol (80 per cent V/V) R.*

Reference solution (c) To 2.0 mL of reference solution (b) add 2.0 mL of a mixture of 1 volume of *concentrated ammonia R* and 9 volumes of *alcohol (80 per cent V/V) R.*

Apply separately to the plate 5 µL of each solution. Develop immediately over a path of 14 cm using a mixture of 3 volumes of *water R*, 25 volumes of *methanol R* and 75 volumes of *chloroform R*. Dry the plate in a current of cold air and spray with *dimethylaminobenzaldehyde solution R7.* Dry the plate in a current of warm air for about 2 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (1.0 per cent) and at most one such spot is more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Loss on drying (*2.2.32*)

Not more than 2.0 per cent, determined on 0.20 g by drying over *diphosphorus pentoxide R* at 80 °C at a pressure not exceeding 2.7 kPa for 2 h.

ASSAY

Dissolve 0.150 g in 40 mL of *anhydrous acetic acid R*. Titrate with *0.05 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.05 M perchloric acid* is equivalent to 22.07 mg of $C_{23}H_{27}N_3O_6$.

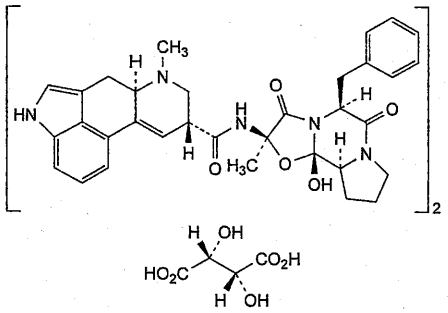
STORAGE

Store in an airtight, glass container, protected from light, at a temperature of 2 °C to 8 °C.

Ph Eur

Ergotamine Tartrate

(Ph. Eur. monograph 0224)



$C_{70}H_{76}N_{10}O_{16}$ 1313 *379-79-3*

Action and use
Oxytocic.

Preparation
Ergotamine Sublingual Tablets

Ph Eur

DEFINITION

Bis[(6a*R*,9*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-[1,3]oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide] (2*R*,3*R*)-2,3-dihydroxybutanedioate.

Content
98.0 per cent to 101.0 per cent (dried substance).

It may contain two molecules of methanol of crystallisation.

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, slightly hygroscopic.

Solubility
Slightly soluble in ethanol 96 per cent. Aqueous solutions slowly become cloudy owing to hydrolysis; this may be prevented by the addition of tartaric acid.

IDENTIFICATION

First identification: B.

Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50 mg in *0.01 M hydrochloric acid* and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with *0.01 M hydrochloric acid.*

Spectral range 250 nm and 360 nm.

Absorption maximum At 311 to 321 nm.

Absorption minimum At 265 to 275 nm.

Specific absorbance at the absorption maximum 118 to 128 (dried substance).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ergotamine tartrate CRS.*

Preparation As discs, triturate the substance to be examined and the reference substance separately with 0.2 mL of *methanol R* and then with *potassium bromide R* as prescribed in the general method.

C. Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution Dissolve 5 mg of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5.0 mL with the same mixture of solvents.

Reference solution Dissolve 5 mg of *ergotamine tartrate CRS* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 5.0 mL with the same mixture of solvents.

Plate *TLC silica gel plate R.*

Mobile phase *anhydrous ethanol R, methylene chloride R, dimethylformamide R, ether R* (5:10:15:70 *V/V/V/V*).

Application 5 µL; immediately expose the points of application to ammonia vapour for exactly 20 s by moving the line of application from side to side above a beaker 55 mm high and 45 mm in diameter containing about 20 mL of *concentrated ammonia R*; dry the line of application in a current of cold air for exactly 20 s.

Development Over 2/3 of the plate.

Drying In a current of cold air for about 2 min.

Detection A Examine for not more than 1 min in ultraviolet light at 365 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and fluorescence to the principal spot in the chromatogram obtained with the reference solution.

Detection B Spray the plate abundantly with *dimethylaminobenzaldehyde solution R7* and dry in a current of warm air for about 2 min; examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 mL of solution S (see Tests) add 1 mL of *glacial acetic acid R*, 0.05 mL of *ferric chloride solution R1* and 1 mL of *phosphoric acid R* and heat in a water-bath at 80 °C. After about 10 min, a blue or violet colour develops which becomes more intense on standing.

E. Dissolve about 10 mg in 1.0 mL of *0.1 M sodium hydroxide*. Transfer to a separating funnel and shake with 5 mL of *methylene chloride R*. Discard the organic layer. Neutralise the aqueous layer with a few drops of *dilute hydrochloric acid R*. 0.1 mL of this solution gives reaction (b) of tartrates (*2.3.1*). Pour the reaction mixture into 1 mL of *water R* to observe the colour change to red or brownish-red.

TESTS

Carry out all operations as rapidly as possible, protected from light.

Solution S
Triturate 50 mg finely with about 25 mg of *tartaric acid R* and dissolve with shaking in 10 mL of *water R.*

pH (*2.2.3*)
4.0 to 5.5 for the suspension.

Shake 25 mg, finely powdered, with 10 mL of *carbon dioxide-free water R.*

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Solvent mixture *water R, acetonitrile R* (10:90 *V/V*).

Solution A Dissolve 1.0 g of *potassium dihydrogen phosphate R* and 2.0 g of *sodium 1-propanesulfonate R* in 900 mL of *water R*, adjust to pH 4.3 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R.*

Test solution Dissolve 30 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution Dissolve 3 mg of *ergotamine for system suitability CRS* (containing impurities A and C) in the solvent mixture and dilute to 5.0 mL with the solvent mixture. In order to prepare impurity B *in situ*, maintain the solution at room temperature for 1 h.

Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (1.8 µm);
— *temperature*: 15 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R*, solution A (10:90 *V/V*);
— *mobile phase B*: solution A, *acetonitrile R* (20:80 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	80 → 50	20 → 50

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Autosampler Set at 4 °C.

Injection 1 µL.

Identification of impurities Use the chromatogram supplied with *ergotamine for system suitability CRS* and the chromatogram obtained with the reference solution to identify the peaks due to impurities A, B and C.

Relative retention With reference to ergotamine (retention time = about 3 min): impurity A = about 0.9; impurity B = about 1.1; impurity C = about 1.2.

System suitability Reference solution:
— *resolution*: minimum 1.5 between the peaks due to impurity A and ergotamine.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.3;
— *impurity C*: maximum 0.4 per cent;
— *impurities A, B*: for each impurity, maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.6 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 6.0 per cent, determined on 0.100 g by drying *in vacuo* at 95 °C for 6 h.

ASSAY

Dissolve 0.200 g in 40 mL of *anhydrous acetic acid R*. Titrate with *0.05 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.05 M perchloric acid* is equivalent to 32.84 mg of $C_{70}H_{76}N_{10}O_{16}$.

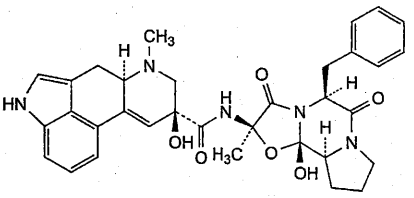
STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

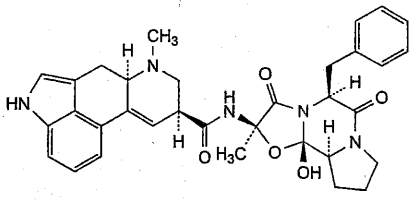
IMPURITIES

Specified impurities *A, B, C.*

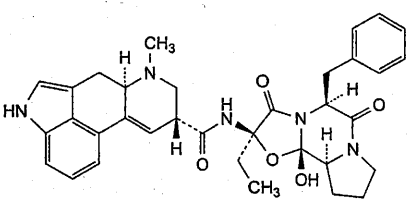
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*



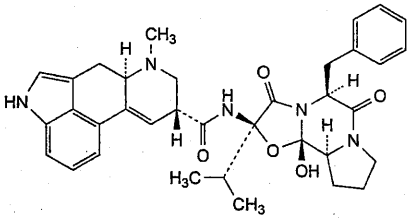
A. (6a*R*,9*S*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-[1,3]oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-9-hydroxy-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (8-hydroxyergotamine),



B. (6a*R*,9*S*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-2-methyl-3,6-dioxooctahydro-8*H*-[1,3]oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergotaminine),



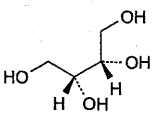
C. (6a*R*,9*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-2-ethyl-10b-hydroxy-3,6-dioxooctahydro-8*H*-[1,3]oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergostine),



D. (6a*R*,9*R*)-*N*-[(2*R*,5*S*,10a*S*,10b*S*)-5-benzyl-10b-hydroxy-3,6-dioxo-2-(propan-2-yl)octahydro-8*H*-[1,3]oxazolo[3,2-*a*]pyrrolo[2,1-*c*]pyrazin-2-yl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergocristine).

Erythritol

(Ph. Eur. monograph 1803)



$C_4H_{10}O_4$ 122.1 *149-32-6*

Action and use
Excipient.

Ph Eur

DEFINITION

(2*R*,3*S*)-Butane-1,2,3,4-tetrol (*meso*-erythritol).

Content
96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or free-flowing granules.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Melting point (*2.2.14*): 119 °C to 122 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *erythritol CRS.*

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 5.0 g in *water R* and dilute to 50 mL with the same solvent.

Conductivity (*2.2.38*)
Maximum 20 µS·cm^{-1}.

Dissolve 20.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution, while gently stirring with a magnetic stirrer.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.50 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.50 g of *erythritol CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with *water R*.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 100.0 mL with *water R*.

Reference solution (d) Dissolve 1.0 g of *erythritol R* and 1.0 g of *glycerol R* in *water R* and dilute to 20 mL with the same solvent.

Column:
— *size*: *l* = 0.3 m, Ø = 8.0 mm;
— *stationary phase*: *cation-exchange resin R* (9 µm);
— *temperature*: 70 °C.

Mobile phase 0.01 per cent *V/V* solution of *sulfuric acid R*.

Flow rate 0.8 mL/min.

Detection Differential refractometer maintained at a constant temperature (e.g. 35 °C).

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time 3 times the retention time of erythritol.

Relative retention With reference to erythritol (retention time = about 11 min): impurity A = about 0.77; impurity B = about 0.90; impurity C = about 0.94; impurity D = about 1.10.

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to erythritol and impurity D.

Limits:
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
— *disregard limit*: area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Microbial contamination
If intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

If not intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*);
— TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*);
— absence of *Escherichia coli* (*2.6.13*);
— absence of *Salmonella* (*2.6.13*).

Bacterial endotoxins (*2.6.14*)
If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins:
— less than 4 IU/g for parenteral preparations having a concentration of 100 g/L or less of erythritol;
— less than 2.5 IU/g for parenteral preparations having a concentration of more than 100 g/L of erythritol.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

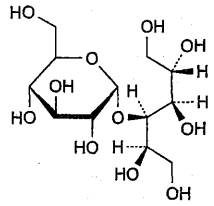
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_4H_{10}O_4$ taking into account the assigned content of *erythritol CRS*.

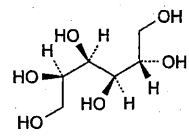
LABELLING

The label states where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

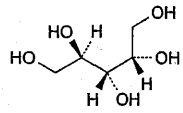
IMPURITIES



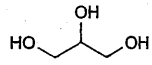
A. 4-*O*-α-D-glucopyranosyl-D-glucitol (D-maltitol),



B. D-glucitol (D-sorbitol),



C. (2*R*,3*s*,4*S*)-pentane-1,2,3,4,5-pentol (*meso*-ribitol),

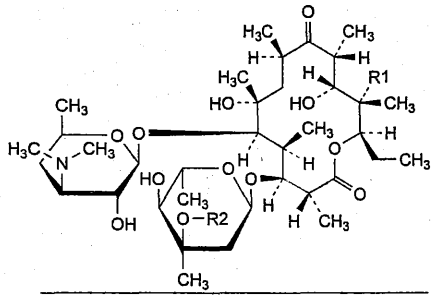


D. propane-1,2,3-triol (glycerol).

Ph Eur

Erythromycin

(Ph. Eur. monograph 0179)



Erythromycin	Mol. Formula	M_r	R1	R2
A	$C_{37}H_{67}NO_{13}$	734	OH	CH_3
B	$C_{37}H_{67}NO_{12}$	718	H	CH_3
C	$C_{36}H_{65}NO_{13}$	720	OH	H

Action and use
Macrolide antibacterial.

Preparations
Erythromycin Gastro-resistant Capsules
Erythromycin Gastro-resistant Tablets
Erythromycin and Zinc Acetate Lotion

Ph Eur

DEFINITION
Mixture of macrolide antibiotics produced by a strain of *Streptomyces erythreus*.

Main component (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin A).

Content
— *sum of erythromycins A, B and C*: 93.0 per cent to 102.0 per cent (anhydrous substance);
— *erythromycin B*: maximum 5.0 per cent (anhydrous substance);
— *erythromycin C*: maximum 5.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow powder or colourless or slightly yellow crystals, slightly hygroscopic.

Solubility
Slightly soluble in water (the solubility decreases as the temperature rises), freely soluble in ethanol (96 per cent), soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: A.

Second identification: B.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *erythromycin A CRS.*

Disregard any band in the region from 1980 cm^{-1} to 2050 cm^{-1}.

If the spectra obtained show differences, dissolve 50 mg of the substance to be examined and of the reference substance separately in 1.0 mL of *methylene chloride R*, dry at 60 °C for 3 h at a pressure not exceeding 0.67 kPa and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *erythromycin A CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *spiramycin CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase Mix 4 volumes of *2-propanol R*, 8 volumes of a 150 g/L solution of *ammonium acetate R* previously adjusted to pH 9.6 with *ammonia R*, and 9 volumes of *ethyl acetate R*; allow to settle and use the upper layer.

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *anisaldehyde solution R1* and heat at 110 °C for 5 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a), and its position and colour are different from those of the spots in the chromatogram obtained with reference solution (b).

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 11.5 g of *dipotassium hydrogen phosphate R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture *methanol R*, solution A (40:60 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 4 mg of *erythromycin for system suitability CRS* (containing impurities A, B, C, D, E, F, H and L) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (e) Dissolve 4 mg of *erythromycin for impurity M identification CRS* in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Column:
- *size*: l = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
- *temperature*: 65 °C; preheating the mobile phase may be required, for instance by extending the inlet tubing in the oven to 30 cm.

Mobile phase:
- *mobile phase A*: *phosphate buffer solution pH 7.0 R7, acetonitrile R1, water R* (5:35:60 *V/V/V*);
- *mobile phase B*: *phosphate buffer solution pH 7.0 R7, water R, acetonitrile R1* (5:45:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	100	0
t_R - (t_R + 2)	100 → 0	0 → 100
(t_R + 2) - (t_R + 15)	0	100

t_R = retention time of erythromycin B, determined by injecting 10 µL of reference solution (b) and eluting with mobile phase A

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 4 °C.

Injection 100 µL.

Identification of impurities Use the chromatogram supplied with *erythromycin for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E, F, H and L; use the chromatogram supplied with *erythromycin for impurity M identification CRS* and the chromatogram obtained with reference solution (e) to identify the peak due to impurity M; use the chromatogram obtained with reference solution (b) to identify the peaks due to erythromycins B and C.

Relative retention With reference to erythromycin A (retention time = about 23 min): impurity H = about 0.3; impurity A = about 0.4; impurity B = about 0.5; erythromycin C = about 0.55; impurity M = about 0.58; impurity L = about 0.63; impurity C = about 0.9; impurity D = about 1.61; erythromycin B = about 1.75; impurity F = about 1.81; impurity E = about 2.3.

System suitability Reference solution (d):
- *resolution*: minimum 1.2 between the peaks due to impurity B and erythromycin C;
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin B; minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin A. If necessary, adjust the concentration of *acetonitrile R1* in the mobile phases and/or the gradient to obtain the required separation.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 2; impurity E = 0.08; impurity F = 0.08; impurity L = 0.11;
- for each impurity, use the concentration of erythromycin A in reference solution (c).

Limits:
- *impurity C*: maximum 3.0 per cent;
- *impurities A, B*: for each impurity, maximum 2.0 per cent;
- *impurities D, E, F, H, M*: for each impurity, maximum 1.0 per cent;
- *impurity L*: maximum 0.4 per cent;
- *any other impurity*: for each impurity, maximum 0.4 per cent;
- *total*: maximum 7.0 per cent;
- *reporting threshold*: 0.2 per cent; disregard the peaks due to erythromycins B and C.

Thiocyanate

Maximum 0.3 per cent.

Prepare the solutions immediately before use and protect from actinic light.

Compensation liquid Dilute 1.0 mL of a 90 g/L solution of *ferric chloride R* to 50.0 mL with *methanol R*.

Test solution Dissolve 0.100 g of the substance to be examined in 20 mL of *methanol R*, add 1.0 mL of a 90 g/L solution of *ferric chloride R* and dilute to 50.0 mL with *methanol R*.

Prepare 2 independent reference solutions.

Reference solution Dissolve 0.100 g of *potassium thiocyanate R*, previously dried at 105 °C for 1 h, in *methanol R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *methanol R*. To 5.0 mL of this solution, add 1.0 mL of a 90 g/L solution of *ferric chloride R* and dilute to 50.0 mL with *methanol R*.

Measure the absorbance (*2.2.25*) of each reference solution (A_1, A_2) and of the test solution (A) at the absorption maximum (about 492 nm).

Suitability value:

$$S = \frac{m_2 \times A_1}{m_1 \times A_2}$$

m_1, m_2 = mass of potassium thiocyanate used to prepare reference solutions A_1 and A_2 respectively, in grams.

The test is not valid unless S is not less than 0.985 and not greater than 1.015.

Calculate the percentage content of thiocyanate using the following expression:

$$\frac{A \times 58.08 \times 0.5}{m \times 97.18} \times \left(\frac{m_1}{A_1} + \frac{m_2}{A_2}\right)$$

m	=	mass of the substance to be examined used to prepare the test solution, in grams;
58.08	=	relative molecular mass of the thiocyanate moiety;
97.18	=	relative molecular mass of potassium thiocyanate.

Water (*2.5.12*)
Maximum 6.5 per cent, determined on 0.200 g.
Use a 100 g/L solution of *imidazole R* in *anhydrous methanol R* as the solvent.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solutions (a) and (b).

System suitability Reference solution (a):
— *symmetry factor*: maximum 2.0 for the peak due to erythromycin A;
— *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

Calculate the percentage content of erythromycin A ($C_{37}H_{67}NO_{13}$) using the chromatogram obtained with reference solution (a). Calculate the percentage contents of erythromycin B ($C_{37}H_{67}NO_{12}$) and erythromycin C ($C_{36}H_{65}NO_{13}$) using the chromatogram obtained with reference solution (b).

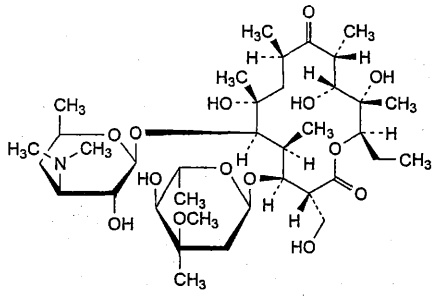
STORAGE

In an airtight container, protected from light.

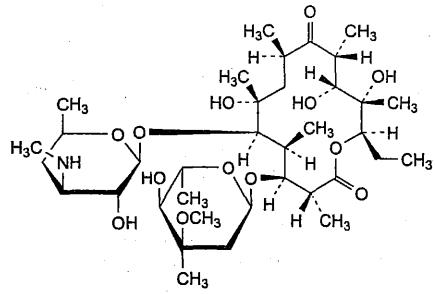
IMPURITIES

Specified impurities *A, B, C, D, E, F, H, L, M.*

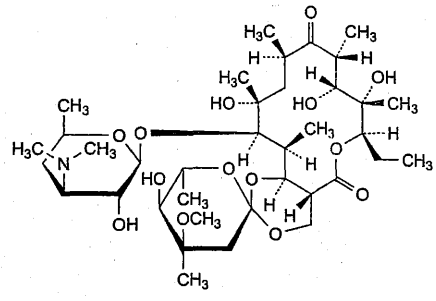
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *I, J, K, N.*



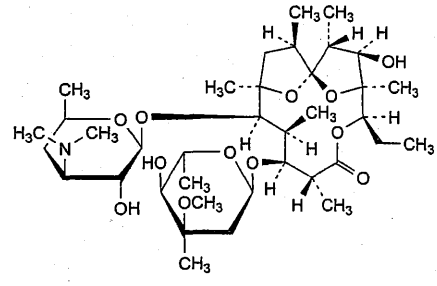
A. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin F),



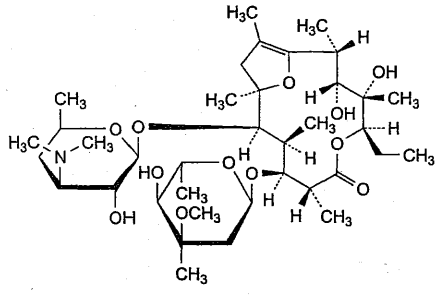
B. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethylerythromycin A),



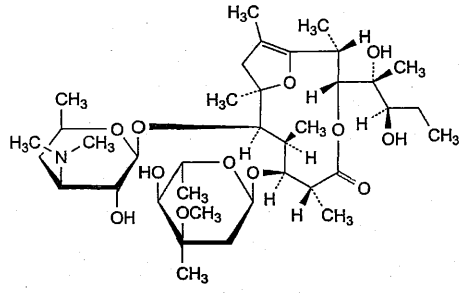
C. (2*S*,4a*R*,4′*R*,5′*S*,6′*S*,7*R*,8*S*,9*R*,10*R*,12*R*,14*R*,15*R*,16*S*,16a*S*)-7-ethyl-5′,8,9,14-tetrahydroxy-4′-methoxy-4′,6′,8,10,12,14,16-heptamethyl-15-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]hexadecahydrospiro[5*H*,11*H*-1,3-dioxino[5,4-*c*]oxacyclotetradecin-2,2′-pyrane]-5,11-dione (erythromycin E),



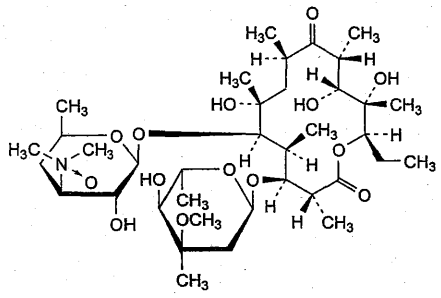
D. (1*S*,2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadecan-7-one (anhydroerythromycin A),



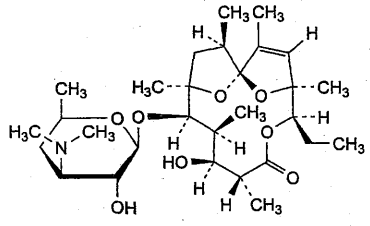
E. (2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3,4-dihydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15-dioxabicyclo[10.2.1]pentadec-1(14)-en-7-one (erythromycin A enol ether),



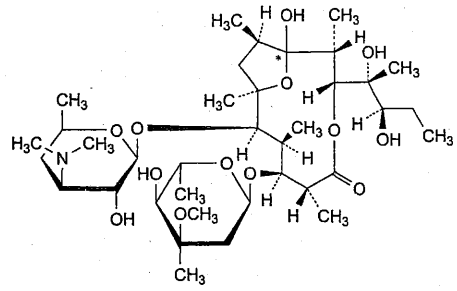
F. (2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridec-1(12)-en-5-one (pseudoerythromycin A enol ether),



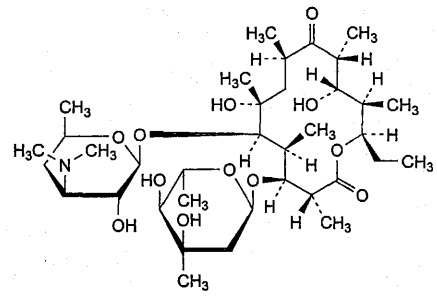
H. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione *N*-oxide (erythromycin A 3″-*N*-oxide),



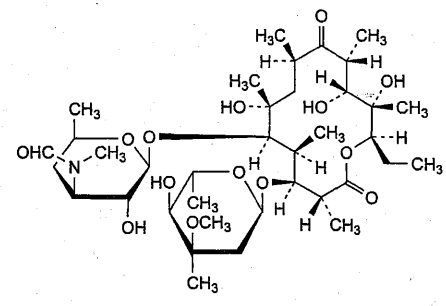
I. (1*S*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-5-ethyl-9-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadec-2-en-7-one (erythralosamine),



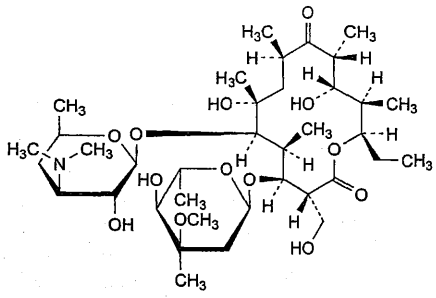
J. (1*RS*,2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*,12*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-1-hydroxy-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridecan-5-one (pseudoerythromycin A hemiketal),



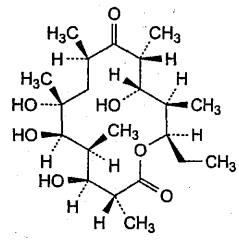
K. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy] oxacyclotetradecane-2,10-dione (erythromycin D),



L. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(formylmethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethyl-3″-*N*-formyl erythromycin A),



M. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin G),

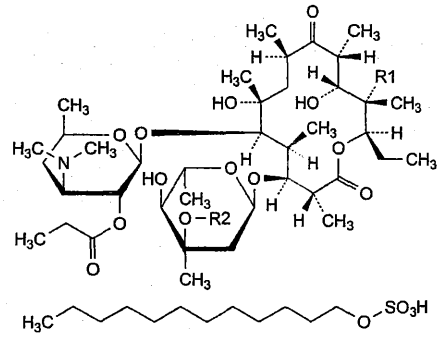


N. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-14-ethyl-4,6,7,12-tetrahydroxy-3,5,7,9,11,13-hexamethyloxacyclotetradecane-2,10-dione (erythronolide B).

Ph Eur

Erythromycin Estolate

(Ph. Eur. monograph 0552)



Erythromycin (estolate)	Mol. Formula	M_r	R1	R2
A	$C_{52}H_{97}NO_{18}S$	1056	OH	CH_3
B	$C_{52}H_{97}NO_{17}S$	1040	H	CH_3
C	$C_{51}H_{95}NO_{18}S$	1042	OH	H

Action and use
Macrolide antibacterial.

Preparation
Erythromycin Estolate Capsules

Ph Eur

DEFINITION
Mixture of the estolate esters of erythromycin.

Main component (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-2-*O*-propanyl-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione dodecyl sulfate (erythromycin A 2″-propionate dodecyl sulfate).

Semi-synthetic product derived from a fermentation product obtained using a strain of *Streptomycines erythreus.*

Content
— *sum of erythromycins A, B and C expressed as estolates*: 93.0 per cent to 102.0 per cent (anhydrous substance);
— *erythromycin B estolate*: maximum 5.0 per cent (anhydrous substance);
— *erythromycin C estolate*: maximum 5.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), soluble in acetone. It is practically insoluble in dilute hydrochloric acid.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *erythromycin estolate CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use (apart from the test solution).*

Solution A (hydrolysis solution). Dissolve 11.5 g of *dipotassium hydrogen phosphate R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R.*

Test solution Dissolve 0.150 g of the substance to be examined in 25 mL of *methanol R.* Add 20 mL of solution A, mix and allow to stand at room temperature for at least 12 h. Dilute to 50.0 mL with solution A.

Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in 12.0 mL of *methanol R* and dilute to 20.0 mL with solution A.

Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in 40.0 mL of *methanol R* and dilute to 100.0 mL with solution A.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with a mixture of 4 volumes of solution A and 6 volumes of *methanol R.*

Reference solution (d) Dissolve 4 mg of *erythromycin for system suitability CRS* (containing impurities A, B, C, D, E, F, H and L) in 0.4 mL of *methanol R* and add 0.6 mL of solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
— *temperature*: 65 °C; preheating the mobile phase may be required, for instance by extending the inlet tubing in the oven to 30 cm.

Mobile phase:
— *mobile phase A*: *phosphate buffer solution pH 7.0 R7, acetonitrile R1, water R* (5:35:60 *V/V/V*);

— *mobile phase B*: *phosphate buffer solution ph 7.0 R7, water R, acetonitrile R1* (5:45:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	100	0
t_R - (t_R + 2)	100 → 0	0 → 100
(t_R + 2) - (t_R + 15)	0	100

t_R = retention time of erythromycin B, determined by injecting 20 µL of reference solution (b) and eluting with mobile phase A

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 4 °C.

Injection 200 µL.

Identification of impurities Use the chromatogram supplied with *erythromycin for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E, F and L; use the chromatogram obtained with reference solution (b) to identify the peaks due to erythromycins B and C.

Relative retention With reference to erythromycin A (retention time = about 23 min): impurity A = about 0.4; impurity B = about 0.5; erythromycin C = about 0.55; impurity L = about 0.63; impurity C = about 0.9; impurity D = about 1.61; erythromycin B = about 1.75; impurity F = about 1.81; impurity E = about 2.3.

System suitability Reference solution (d):
— *resolution*: minimum 1.2 between the peaks due to impurity B and erythromycin C;
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin B; minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin A. If necessary, adjust the concentration of *acetonitrile R1* in the mobile phases and/or the gradient to obtain the required separation.

Calculation of percentage contents:
— *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 2; impurity E = 0.08; impurity F = 0.08; impurity L = 0.11;
— for each impurity, use the concentration of erythromycin A in reference solution (c).

Limits:
— *impurity C*: maximum 3.0 per cent;
— *impurities A, B*: for each impurity, maximum 2.0 per cent;
— *impurities D, F*: for each impurity, maximum 1.0 per cent;
— *impurities eluting with relative retention between 2.1 and 2.4*: not more than 4 impurities; for 1 impurity, maximum 3.5 per cent; for each other impurity, maximum 1.0 per cent;
— *impurity L*: maximum 0.4 per cent;
— *any other impurity*: for each impurity, maximum 0.4 per cent;
— *total*: maximum 7.0 per cent;
— *reporting threshold*: 0.2 per cent; disregard the peaks due to erythromycins B and C and any peak with a relative retention to erythromycin A of less than 0.3 (hydrolysis peaks).

Free erythromycin

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.250 g of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution Dissolve 75.0 mg of *erythromycin A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 25.0 mL with *acetonitrile R1*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Mix 35 volumes of *acetonitrile R1* and 65 volumes of a solution containing 3.4 g/L of *potassium dihydrogen phosphate R* and 2.0 g/L of *triethylamine R*, previously adjusted to pH 3.0 with *dilute phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 195 nm.

Injection 20 µL.

Run time Twice the retention time of erythromycin A for the reference solution and 4.5 times the retention time of the 1st peak of erythromycin propionate for the test solution.

Retention time Erythromycin A = about 5 min; 1st peak of erythromycin propionate = about 10 min.

Limit:
— *free erythromycin*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (6.0 per cent).

Dodecyl sulfate

23.0 per cent to 25.5 per cent of $C_{12}H_{26}O_4S$ (anhydrous substance).

Dissolve 0.500 g in 25 mL of *dimethylformamide R*. Titrate with 0.1 M sodium methoxide using 0.05 mL of a 3 g/L solution of *thymol blue R* in *methanol R* as indicator.

1 mL of 0.1 M sodium methoxide is equivalent to 26.64 mg of $C_{12}H_{26}O_4S$.

Water (*2.5.12*)

Maximum 4.0 per cent, determined on 0.300 g.

Use a 100 g/L solution of *imidazole R* in *anhydrous methanol R* as the solvent.

Sulfated ash (*2.4.14*)

Maximum 0.5 per cent, determined on 0.5 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solutions (a) and (b).

System suitability Reference solution (a):
— *symmetry factor*: maximum 2.0 for the peak due to erythromycin A;
— *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

Calculate the percentage content of erythromycin A ($C_{37}H_{67}NO_{13}$) using the chromatogram obtained with reference solution (a). Calculate the percentage contents of erythromycin B ($C_{37}H_{67}NO_{12}$) and erythromycin C ($C_{36}H_{65}NO_{13}$) using the chromatogram obtained with reference solution (b). Express the result as erythromycin A estolate, erythromycin B estolate and erythromycin C estolate by multiplying the percentage content of erythromycin A

by 1.4387, the percentage content of erythromycin B by 1.4485 and the percentage content of erythromycin C by 1.4472.

For the calculation of content of erythromycin estolate, use the sum of erythromycins A, B and C expressed as estolates as described above.

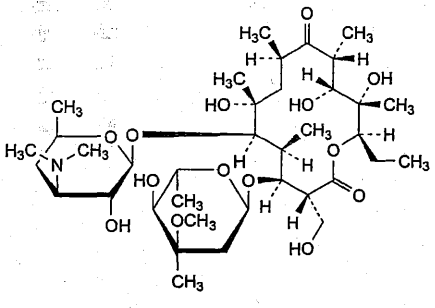
STORAGE

Protected from light.

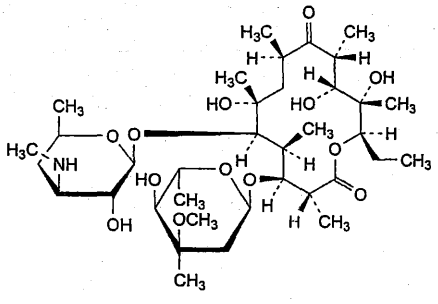
IMPURITIES

Specified impurities *A, B, C, D, E, F, L.*

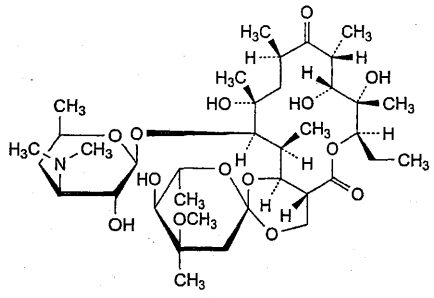
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, H, I, J, K, M, N.*



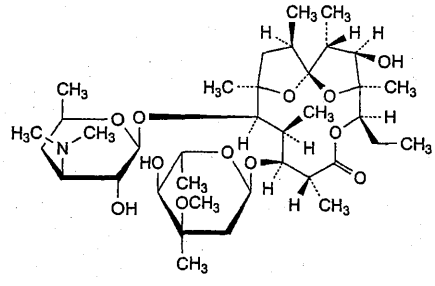
A. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin F),



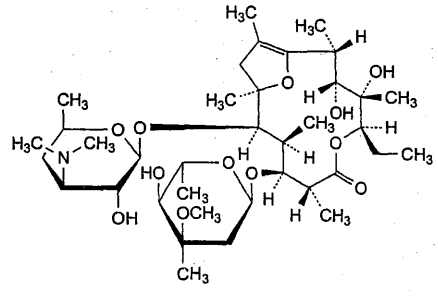
B. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3′′-*N*-demethylerythromycin A),



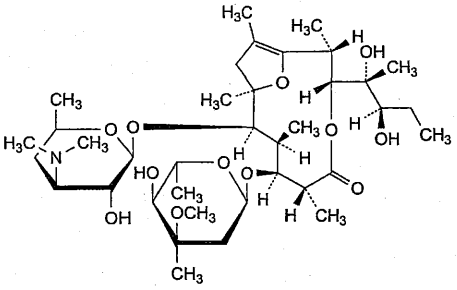
C. (2*S*,4a*R*,4′*R*,5′*S*,6′*S*,7*R*,8*S*,9*R*,10*R*,12*R*,14*R*,15*R*,16*S*,16a*S*)-7-ethyl-5′,8,9,14-tetrahydroxy-4′-methoxy-4′,6′,8,10,12,14,16-heptamethyl-15-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]hexadecahydrospiro[5*H*,11*H*-1,3-dioxino[5,4-*c*]oxacyclotetradecin-2,2′-pyrane]-5,11-dione (erythromycin E),



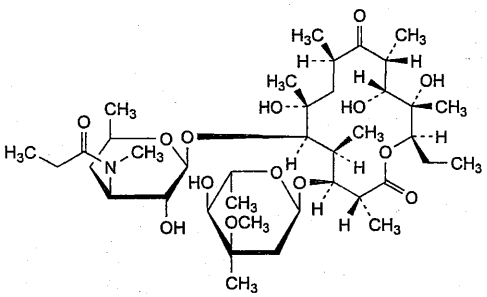
D. (1*S*,2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadecan-7-one (anhydroerythromycin A),



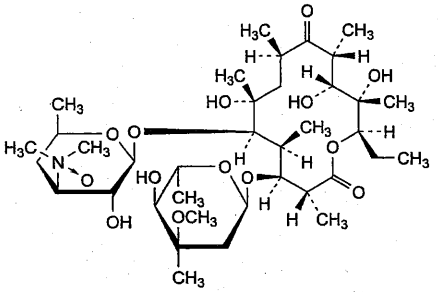
E. (2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3,4-dihydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15-dioxabicyclo[10.2.1]pentadec-1(14)-en-7-one (erythromycin A enol ether),



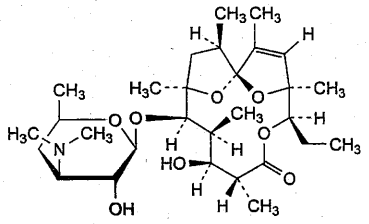
F. (2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridec-1(12)-en-5-one (pseudoerythromycin A enol ether),



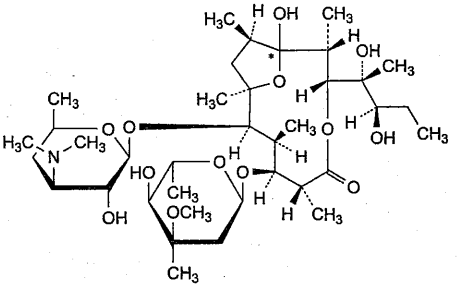
G. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylpropanoylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethyl-3″-*N*-propanoylerythromycin A),



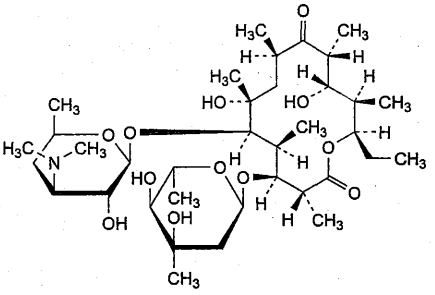
H. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione *N*-oxide (erythromycin A 3″-*N*-oxide),



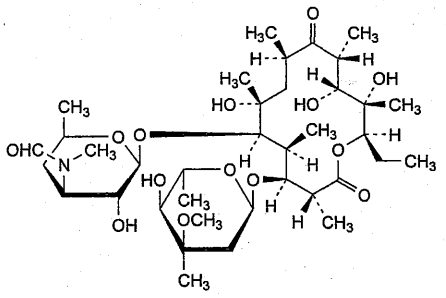
I. (1*S*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-5-ethyl-9-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadec-2-en-7-one (erythralosamine),



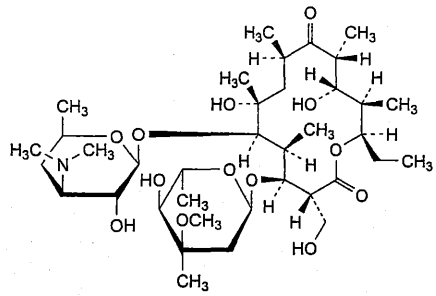
J. (1*RS*,2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*,12*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-1-hydroxy-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridecan-5-one (pseudoerythromycin A hemiketal),



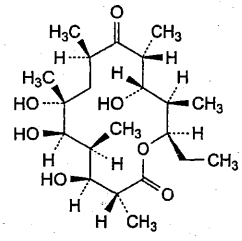
K. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin D),



L. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(formylmethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethyl-3″-*N*-formylerythromycin A),



M. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin G),

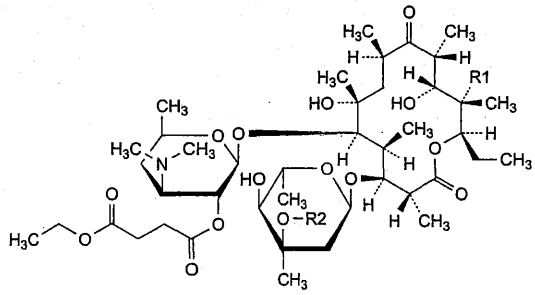


N. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-14-ethyl-4,6,7,12-tetrahydroxy-3,5,7,9,11,13-hexamethyloxacyclotetradecane-2,10-dione (erythronolide B).

Ph Eur

Erythromycin Ethyl Succinate

(Erythromycin Ethylsuccinate, Ph. Eur. monograph 0274)



Erythromycin (ethylsuccinate)	Mol. Formula	*M*r	R1	R2
A	$C_{43}H_{75}NO_{16}$	862	OH	CH_3
B	$C_{43}H_{75}NO_{15}$	846	H	CH_3
C	$C_{42}H_{73}NO_{16}$	848	OH	H

Action and use
Macrolide antibacterial.

Preparations
Erythromycin Ethyl Succinate Oral Suspension
Erythromycin Ethyl Succinate Tablets

Ph Eur

DEFINITION
Mixture of the ethylsuccinate esters of erythromycin.

Main component (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-2-*O*-(4-ethoxy-4-oxobutanoyl)-β-D-*xylo*-hexopyranosyl]oxy] oxacyclotetradecane-2,10-dione (erythromycin A 2″-(ethyl succinate)).

Semi-synthetic product derived from a fermentation product obtained using a strain of *Streptomyces erythreus*.

Content
— *sum of erythromycins A, B and C expressed as ethylsuccinates*: 93.0 per cent to 102.0 per cent (anhydrous substance);
— *erythromycin B ethylsuccinate*: maximum 5.0 per cent (anhydrous substance);
— *erythromycin C ethylsuccinate*: maximum 5.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder, hygroscopic.

Solubility
Practically insoluble in water, freely soluble in acetone, in anhydrous ethanol and in methanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *erythromycin ethylsuccinate CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*).
Hydrolysis solution A 20 g/L solution of *dipotassium hydrogen phosphate R* adjusted to pH 8.0 with *phosphoric acid R*.
Test solution Dissolve 0.115 g of the substance to be examined in 25 mL of *methanol R*. Add 20 mL of the hydrolysis solution, mix and allow to stand at room temperature for at least 12 h. Dilute to 50.0 mL with the hydrolysis solution.
Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in 10 mL of *methanol R* and dilute to 20.0 mL with the hydrolysis solution.
Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in 50 mL of *methanol R*. Add 5.0 mL of reference solution (a) and dilute to 100.0 mL with the hydrolysis solution.
Reference solution (c) Dissolve 2 mg of *N-demethylerythromycin A CRS* in 20 mL of reference solution (b).
Reference solution (d) Dilute 3.0 mL of reference solution (a) to 100.0 mL with a mixture of equal volumes of *methanol R* and the hydrolysis solution.
Reference solution (e) Dissolve 40 mg of *erythromycin A CRS*, previously heated at 130 °C for 3 h, in 10 mL of *methanol R* and dilute to 20 mL with the hydrolysis solution.
Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (8 µm) with a pore size of 100 nm;
— *temperature*: 70 °C using a water-bath for the column and at least one-third of the tubing preceding the column.
Mobile phase To 50 mL of a 35 g/L solution of *dipotassium hydrogen phosphate R* adjusted to pH 8.0 with *dilute phosphoric acid R*, add 400 mL of *water for chromatography R*, 165 mL of *2-methyl-2-propanol R* and 30 mL of *acetonitrile R1*, and dilute to 1000 mL with *water for chromatography R*.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 200 µL of the test solution and reference solutions (a), (c), (d) and (e).

Run time 5 times the retention time of erythromycin A; begin integration after the hydrolysis peak.

Relative retention With reference to erythromycin A (retention time = about 15 min): hydrolysis peak = less than 0.3; impurity B = about 0.45; erythromycin C = about 0.5; impurity C = about 0.9; impurity G = about 1.3; impurity D = about 1.4; impurity F = about 1.5; erythromycin B = about 1.8; impurity E = about 4.3.

System suitability Reference solution (c):
— *resolution*: minimum 0.8 between the peaks due to impurity B and erythromycin C and minimum 5.5 between the peaks due to impurity B and erythromycin A.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 0.09; impurity F = 0.15; impurity G = 0.14; use the chromatogram obtained with reference solution (e) to identify the peaks due to impurities E and F;
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (3.0 per cent);
— *total*: not more than 1.67 times the area of the principal peak in the chromatogram obtained with reference solution (d) (5.0 per cent);
— *disregard limit*: 0.02 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.06 per cent).

Free erythromycin

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.250 g of the substance to be examined in *acetonitrile R1* and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 75.0 mg of *erythromycin A CRS* in *acetonitrile R1* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 25.0 mL with *acetonitrile R1*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Mix 35 volumes of *acetonitrile R1* and 65 volumes of a solution containing 3.4 g/L of *potassium dihydrogen phosphate R* and 2.0 g/L of *triethylamine R*, previously adjusted to pH 3.0 with *dilute phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 195 nm.

Injection 20 µL.

Run time Twice the retention time of erythromycin A (retention time = about 8 min) for the reference solution; twice the retention time of erythromycin ethylsuccinate (retention time = about 24 min) for the test solution.

Limit:
— *free erythromycin*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (6.0 per cent).

Water (*2.5.12*)

Maximum 3.0 per cent, determined on 0.300 g.

Use a 100 g/L solution of *imidazole R* in *anhydrous methanol R* as the solvent.

Sulfated ash (*2.4.14*)

Maximum 0.3 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use (apart from the test solution).*

Solution A (hydrolysis solution). Dissolve 11.5 g of *dipotassium hydrogen phosphate R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture *methanol R*, solution A (40:60 *V/V*).

Test solution Dissolve 11.5 mg of the substance to be examined in 2.5 mL of *methanol R*. Add 2 mL of solution A, mix and allow to stand at room temperature for at least 12 h. Dilute to 5.0 mL with solution A.

Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in 10.0 mL of *methanol R* and dilute to 20.0 mL with solution A.

Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in 50.0 mL of *methanol R* and dilute to 100.0 mL with solution A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
— *temperature*: 65 °C; preheating the mobile phase may be required, for instance by extending the inlet tubing in the oven to 30 cm.

Mobile phase:
— *mobile phase A*: *phosphate buffer solution pH 7.0 R7, acetonitrile R1, water for chromatography R* (5:35:60 *V/V/V*);
— *mobile phase B*: *phosphate buffer solution pH 7.0 R7, water for chromatography R, acetonitrile R1* (5:45:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	100	0
t_R - (t_R + 2)	100 → 0	0 → 100
(t_R + 2) - (t_R + 15)	0	100

t_R = retention time of erythromycin B, determined by injecting 20 µL of reference solution (b) and eluting with mobile phase A

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 4 °C.

Injection 200 µL.

System suitability Reference solution (a):
— *symmetry factor*: maximum 2.0 for the peak due to erythromycin A;
— *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

Calculate the percentage content of erythromycin A ($C_{37}H_{67}NO_{13}$) using the chromatogram obtained with reference solution (a). Calculate the percentage contents of erythromycin B ($C_{37}H_{67}NO_{12}$) and erythromycin C ($C_{36}H_{65}NO_{13}$) using the chromatogram obtained with reference solution (b).

Express the results as erythromycin A ethylsuccinate, erythromycin B ethylsuccinate and erythromycin C ethylsuccinate by multiplying the percentage content of erythromycin A by 1.1744, the percentage content of

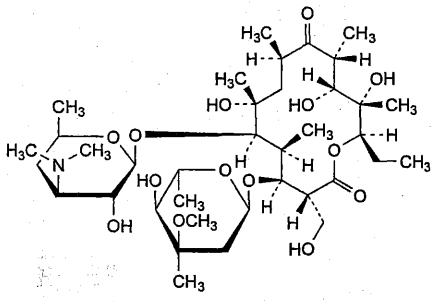
erythromycin B by 1.1783 and the percentage content of erythromycin C by 1.1777.

For the calculation of content of erythromycin ethylsuccinate, use the sum of erythromycins A, B and C expressed as ethylsuccinates as described above.

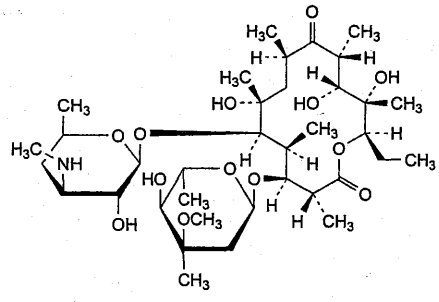
STORAGE

In an airtight container, protected from light.

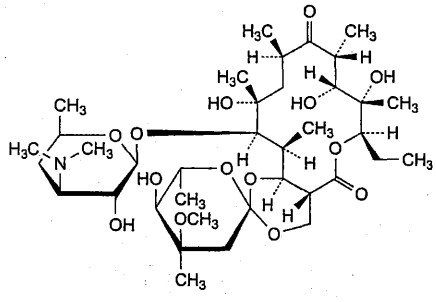
IMPURITIES



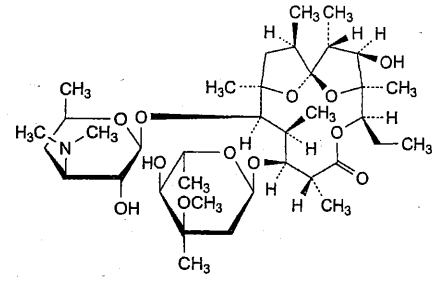
A. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin F),



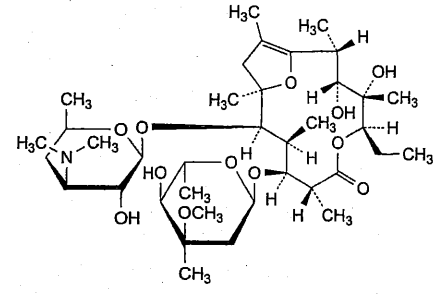
B. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethylerythromycin A),



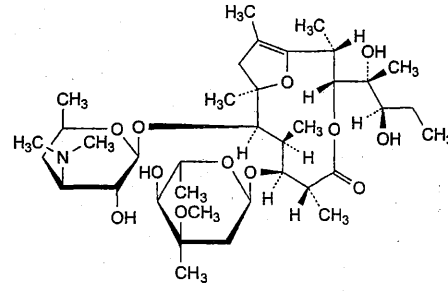
C. (2*S*,4a*R*,4′*R*,5′*S*,6′*S*,7*R*,8*S*,9*R*,10*R*,12*R*,14*R*,15*R*,16*S*,16a*S*)-7-ethyl-5′,8,9,14-tetrahydroxy-4′-methoxy-4′,6′,8,10,12,14,16-heptamethyl-15-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]hexadecahydrospiro[5*H*,11*H*-1,3-dioxino[5,4-*c*]oxacyclotetradecin-2,2′-pyrane]-5,11-dione (erythromycin E),



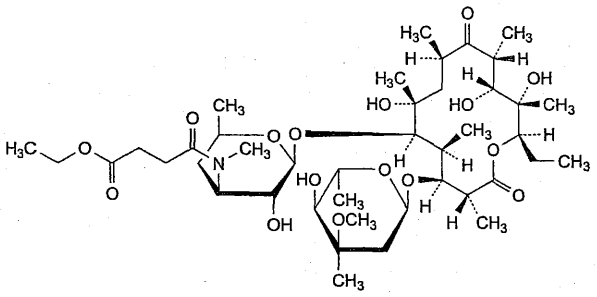
D. (1*S*,2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadecan-7-one (anhydroerythromycin A),



E. (2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3,4-dihydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15-dioxabicyclo[10.2.1]pentadec-1(14)-en-7-one (erythromycin A enol ether),



F. (2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridec-1(12)-en-5-one (pseudoerythromycin A enol ether),

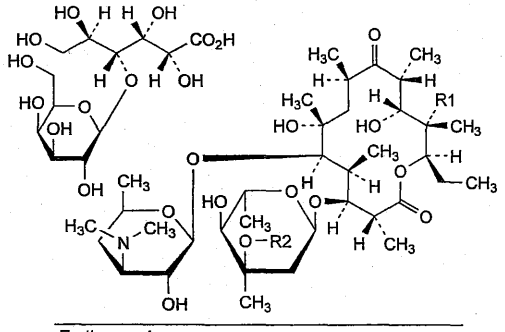


G. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-[(4-ethoxy-4-oxobutanoyl)methylamino]-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethyl-3″-*N*-(ethoxysuccinyl)erythromycin A).

Ph Eur

Erythromycin Lactobionate

(Ph. Eur. monograph 1098)



Erythromycin (lactobionate)	Mol. Formula	M_r	R1	R2
A	$C_{49}H_{89}NO_{25}$	1092	OH	CH_3
B	$C_{49}H_{89}NO_{24}$	1076	H	CH_3
C	$C_{48}H_{87}NO_{25}$	1078	OH	H

Action and use
Macrolide antibacterial.

Preparation
Erythromycin Lactobionate Infusion

Ph Eur

DEFINITION

Mixture of the lactobionate salts of erythromycin.

Main component (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione 4-*O*-β-D-galactopyranosyl-D-gluconate (erythromycin A lactobionate).

Salt of a product obtained by fermentation using a strain of *Streptomyces erythreus*.

Content
— *sum of erythromycins A, B and C expressed as lactobionates*: 93.0 per cent to 102.0 per cent (anhydrous substance);
— *erythromycin B lactobionate*: maximum 5.0 per cent (anhydrous substance);
— *erythromycin C lactobionate*: maximum 5.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellow, hygroscopic powder.

Solubility
Soluble in water, freely soluble in anhydrous ethanol and in methanol, very slightly soluble in acetone and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *erythromycin lactobionate CRS.*

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in 20 mL of *water R.*

pH (*2.2.3*)
6.5 to 7.5.

Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 11.5 g of *dipotassium hydrogen phosphate R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R.*

Solvent mixture *methanol R*, solution A (40:60 *V/V*).

Test solution Dissolve 60.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 4 mg of *erythromycin for system suitability CRS* (containing impurities A, B, C, D, E, F, H and L) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
— *temperature*: 65 °C; preheating the mobile phase may be required, for instance by extending the inlet tubing in the oven to 30 cm.

Mobile phase:
— *mobile phase A*: *phosphate buffer solution pH 7.0 R7, acetonitrile R1, water R* (5:35:60 *V/V/V*);
— *mobile phase B*: *phosphate buffer solution pH 7.0 R7, water R, acetonitrile R1* (5:45:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	100	0
t_R - (t_R + 2)	100 → 0	0 → 100
(t_R + 2) - (t_R + 15)	0	100

t_R = retention time of erythromycin B, determined by injecting 10 µL of reference solution (b) and eluting with mobile phase A

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 4 °C.

Injection 100 µL.

Identification of impurities Use the chromatogram supplied with *erythromycin for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E, F and L; use the chromatogram obtained with reference solution (b) to identify the peaks due to erythromycins B and C.

Relative retention With reference to erythromycin A (retention time = about 23 min): impurity A = about 0.4; impurity B = about 0.5; erythromycin C = about 0.55; impurity L = about 0.63; impurity C = about 0.9; impurity D = about 1.61; erythromycin B = about 1.75; impurity F = about 1.81; impurity E = about 2.3.

System suitability Reference solution (d):
- *resolution*: minimum 1.2 between the peaks due to impurity B and erythromycin C;
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin B; minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin A. If necessary, adjust the concentration of *acetonitrile R1* in the mobile phases and/or the gradient to obtain the required separation.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 2; impurity E = 0.08; impurity F = 0.08; impurity L = 0.11;
- for each impurity, use the concentration of erythromycin A in reference solution (c).

Limits:
- *impurity C*: maximum 3.0 per cent;
- *impurities A, B*: for each impurity, maximum 2.0 per cent;
- *impurities D, E, F*: for each impurity, maximum 1.0 per cent;
- *impurity L*: maximum 0.4 per cent,
- *any other impurity*: for each impurity, maximum 0.4 per cent;
- *total*: maximum 7.0 per cent;
- *reporting threshold*: 0.2 per cent; disregard the peaks due to erythromycins B and C.

Free lactobionic acid

Maximum 1.0 per cent of $C_{12}H_{22}O_{12}$ (anhydrous substance).

Dissolve 0.400 g in 50 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Calculate the volume of *0.1 M sodium hydroxide* required per gram of the substance to be examined (n_1 mL). Dissolve 0.500 g in 40 mL of *anhydrous acetic acid R* and titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Calculate the volume of *0.1 M perchloric acid* required per gram of the substance to be examined (n_2 mL).

Calculate the percentage content of $C_{12}H_{22}O_{12}$ using the following expression:

$$3.580(n_1 - n_2)$$

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.200 g.

Use a 100 g/L solution of *imidazole R* in *anhydrous methanol R* as the solvent.

Sulfated ash (*2.4.14*)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solutions (a) and (b).

System suitability Reference solution (a):
- *symmetry factor*: maximum 2.0 for the peak due to erythromycin A;
- *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

Calculate the percentage content of erythromycin A ($C_{37}H_{67}NO_{13}$) using the chromatogram obtained with reference solution (a). Calculate the percentage contents of erythromycin B ($C_{37}H_{67}NO_{12}$) and erythromycin C ($C_{36}H_{65}NO_{13}$) using the chromatogram obtained with reference solution (b).

Express the result as erythromycin A lactobionate, erythromycin B lactobionate and erythromycin C lactobionate by multiplying the percentage content of erythromycin A by 1.4877, the percentage content of erythromycin B by 1.4986 and the percentage content of erythromycin C by 1.4972.

For the calculation of content of erythromycin lactobionate, use the sum of erythromycins A, B and C expressed as lactobionates as described above.

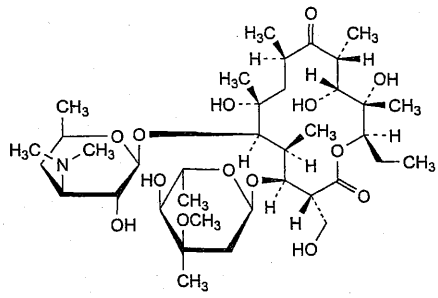
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

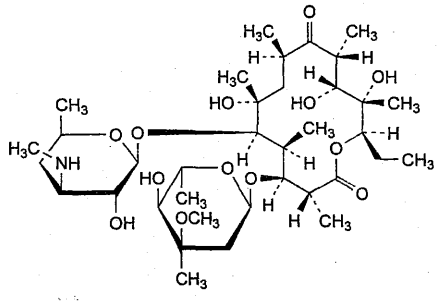
IMPURITIES

Specified impurities *A, B, C, D, E, F, L.*

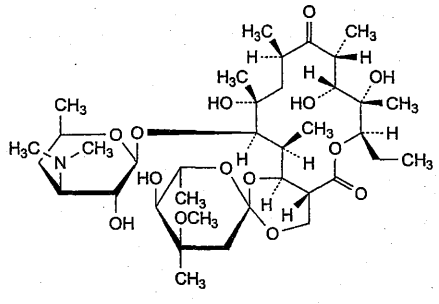
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H, I, J, K, M, N.*



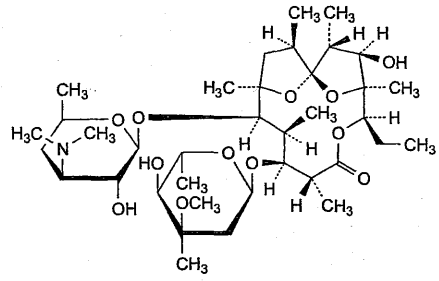
A. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin F),



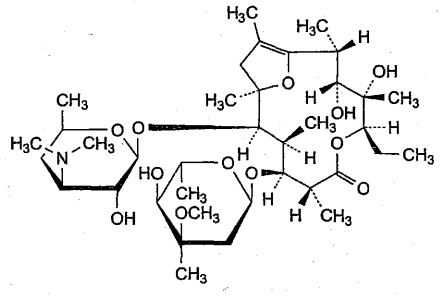
B. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethylerythromycin A),



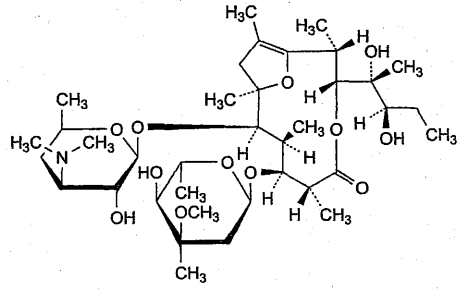
C. (2*S*,4a*R*,4′*R*,5′*S*,6′*S*,7*R*,8*S*,9*R*,10*R*,12*R*,14*R*,15*R*,16*S*,16a*S*)-7-ethyl-5′,8,9,14-tetrahydroxy-4′-methoxy-4′,6′,8,10,12,14,16-heptamethyl-15-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]hexadecahydrospiro[5*H*,11*H*-1,3-dioxino[5,4-*c*]oxacyclotetradecin-2,2′-pyrane]-5,11-dione (erythromycin E),



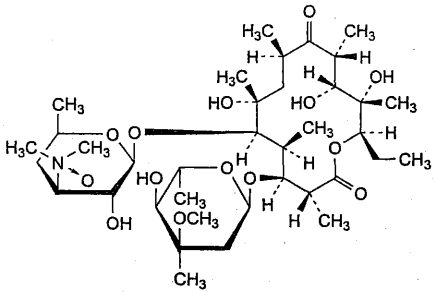
D. (1*S*,2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadecan-7-one (anhydroerythromycin A),



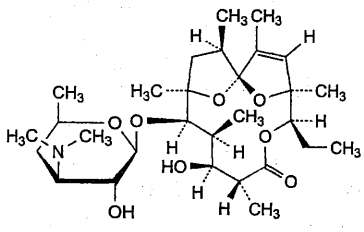
E. (2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3,4-dihydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15-dioxabicyclo[10.2.1]pentadec-1(14)-en-7-one (erythromycin A enol ether),



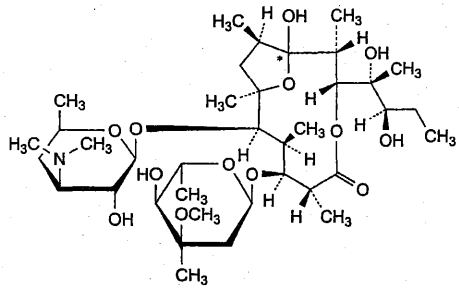
F. (2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridec-1(12)-en-5-one (pseudoerythromycin A enol ether),



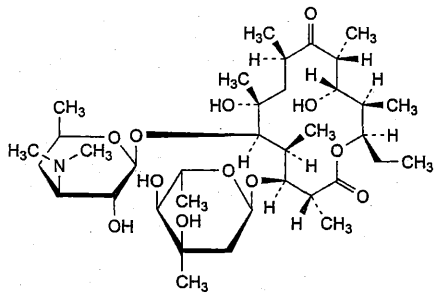
H. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione *N*-oxide (erythromycin A 3″-*N*-oxide),



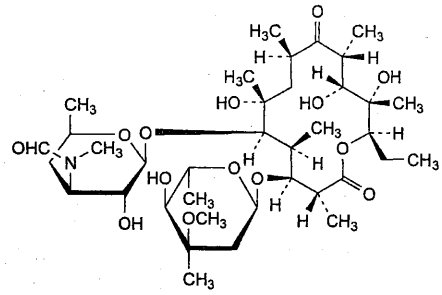
I. (1*S*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-5-ethyl-9-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadec-2-en-7-one (erythralosamine),



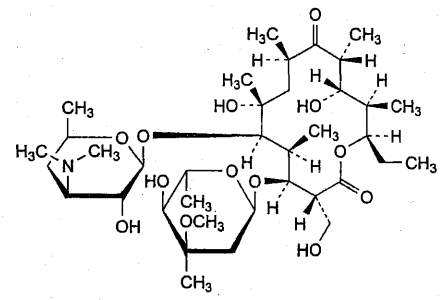
J. (1*RS*,2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*,12*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-1-hydroxy-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridecan-5-one (pseudoerythromycin A hemiketal),



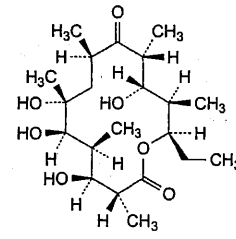
K. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin D),



L. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(formylmethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethyl-3″-*N*-formylerythromycin A),



M. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin G),

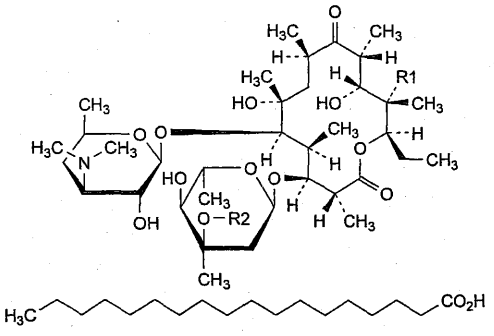


N. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-14-ethyl-4,6,7,12-tetrahydroxy-3,5,7,9,11,13-hexamethyloxacyclotetradecane-2,10-dione (erythronolide B).

Ph Eur

Erythromycin Stearate

(Ph. Eur. monograph 0490)



Erythromycin (stearate)	Mol. Formula	M_r	R1	R2
A	$C_{55}H_{103}NO_{15}$	1018	OH	CH_3
B	$C_{55}H_{103}NO_{14}$	1002	H	CH_3
C	$C_{54}H_{101}NO_{15}$	1004	OH	H

Action and use

Macrolide antibacterial.

Preparation

Erythromycin Stearate Tablets

Ph Eur

DEFINITION

Mixture of the stearate salts of erythromycin and stearic acid.

Main component (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione octadecanoate (erythromycin A stearate).

Salt of a product obtained by fermentation using a strain of *Streptomyces erythreus*.

Content

— *sum of erythromycins A, B and C expressed as stearates*: 79.0 per cent to 102.0 per cent (anhydrous substance);
— *erythromycin B stearate*: maximum 5.0 per cent (anhydrous substance);
— *erythromycin C stearate*: maximum 5.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone and in methanol.

Solutions may be opalescent.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *erythromycin stearate CRS.*

TESTS

Free stearic acid

Maximum 14.0 per cent (anhydrous substance) of $C_{18}H_{36}O_2$.

Dissolve 0.400 g in 50 mL of *methanol R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Calculate the volume of *0.1 M sodium hydroxide* required per gram of the substance to be examined (n_1 mL). Dissolve 0.500 g in 30 mL of *methylene chloride R*. If the solution is opalescent, filter and shake the residue with 3 quantities, each of 25 mL, of *methylene chloride R*. Filter, if necessary, and rinse the filter with *methylene chloride R*. Reduce the volume of the combined filtrate and rinsings to 30 mL by evaporation on a water-bath. Add 50 mL of *glacial acetic acid R* and titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Calculate the volume of *0.1 M perchloric acid* required per gram of the substance to be examined (n_2 mL).

Calculate the percentage content of $C_{18}H_{36}O_2$ using the following expression:

$$2.845(n_1 - n_2) \times \frac{100}{100 - h}$$

h = percentage water content.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 11.5 g of *dipotassium hydrogen phosphate R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture *methanol R*, solution A (40:60 *V/V*).

Test solution Dissolve 55.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Centrifuge and use the clear solution.

Reference solution (a) Dissolve 40.0 mg of *erythromycin A CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10.0 mg of *erythromycin B CRS* and 10.0 mg of *erythromycin C CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 4 mg of *erythromycin for system suitability CRS* (containing impurities A, B, C, D, E, F, H and L) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (e) Dissolve 4 mg of *erythromycin stearate for impurity S identification CRS* in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
— *temperature*: 65 °C; preheating the mobile phase may be required, for instance by extending the inlet tubing in the oven to 30 cm.

Mobile phase:
— *mobile phase A*: *phosphate buffer solution pH 7.0 R7, acetonitrile R1, water R* (5:35:60 *V/V/V*);
— *mobile phase B*: *phosphate buffer solution pH 7.0 R7, water R, acetonitrile R1* (5:45:50 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - t_R	100	0
t_R - (t_R + 2)	100 → 0	0 → 100
(t_R + 2) - (t_R + 15)	0	100

t_R = retention time of erythromycin B, determined by injecting 10 µL of reference solution (b) and eluting with mobile phase A

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 4 °C.

Injection 100 µL.

Identification of impurities Use the chromatogram supplied with *erythromycin for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E, F and L; use the chromatogram supplied with *erythromycin stearate for impurity S identification CRS* and the chromatogram obtained with reference solution (e) to identify the peak due to impurity S; use the chromatogram obtained with reference solution (b) to identify the peaks due to erythromycins B and C.

Relative retention With reference to erythromycin A (retention time = about 23 min): impurity A = about 0.4; impurity B = about 0.5; erythromycin C = about 0.55; impurity L = about 0.63; impurity C = about 0.9; impurity D = about 1.61; erythromycin B = about 1.75; impurity F = about 1.81; impurity S = about 2.1; impurity E = about 2.3.

System suitability Reference solution (d):
- *resolution*: minimum 1.2 between the peaks due to impurity B and erythromycin C;
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin B; minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to erythromycin A. If necessary, adjust the concentration of *acetonitrile R1* in the mobile phases and/or the gradient to obtain the required separation.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 2; impurity E = 0.08; impurity F = 0.08; impurity L = 0.11;
- for each impurity, use the concentration of erythromycin A in reference solution (c).

Limits:
- *impurity C*: maximum 3.0 per cent;
- *impurities A, B*: for each impurity, maximum 2.0 per cent;
- *impurities D, E, F, S*: for each impurity, maximum 1.0 per cent;
- *impurity L*: maximum 0.4 per cent;
- *any other impurity*: for each impurity, maximum 0.4 per cent;
- *total*: maximum 7.0 per cent;
- *reporting threshold*: 0.2 per cent; disregard the peaks due to erythromycins B and C.

Water (*2.5.12*)

Maximum 4.0 per cent, determined on 0.300 g.

Use a 100 g/L solution of *imidazole R* in *anhydrous methanol R* as the solvent.

Sulfated ash (*2.4.14*)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solutions (a) and (b).

System suitability Reference solution (a):
- *symmetry factor*: maximum 2.0 for the peak due to erythromycin A;
- *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

Calculate the percentage content of erythromycin A ($C_{37}H_{67}NO_{13}$) using the chromatogram obtained with reference solution (a). Calculate the percentage contents of erythromycin B ($C_{37}H_{67}NO_{12}$) and erythromycin C ($C_{36}H_{65}NO_{13}$) using the chromatogram obtained with reference solution (b).

Express the results as erythromycin A stearate, erythromycin B stearate and erythromycin C stearate by multiplying the percentage content of erythromycin A by 1.3869, the percentage content of erythromycin B by 1.3955 and the percentage content of erythromycin C by 1.3944.

For the calculation of content of erythromycin stearate, use the sum of erythromycins A, B and C expressed as stearates as described above.

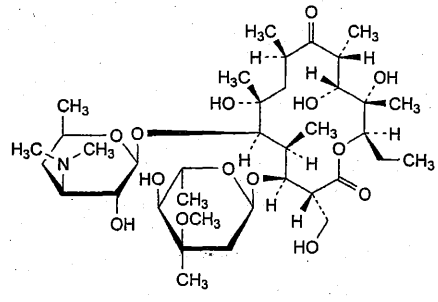
STORAGE

Protected from light.

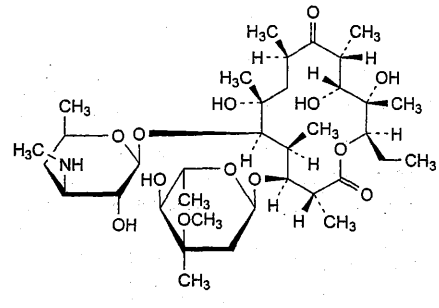
IMPURITIES

Specified impurities *A, B, C, D, E, F, L, S.*

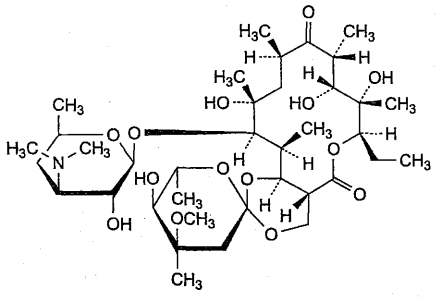
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H, I, J, K, M, N.*



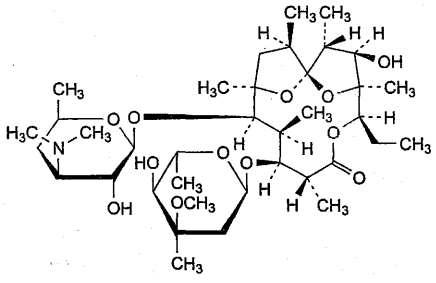
A. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin F),



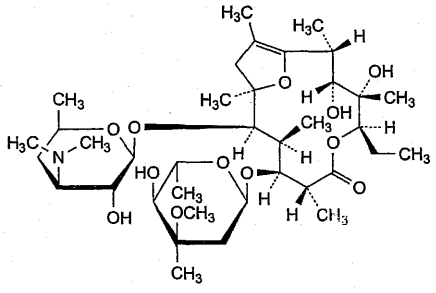
B. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(methylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3″-*N*-demethylerythromycin A),



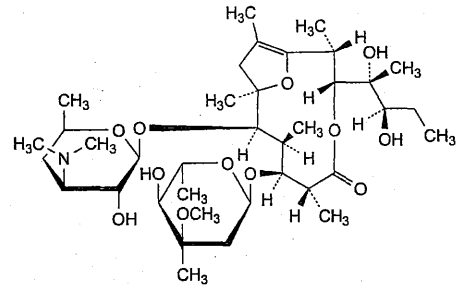
C. (2*S*,4a*R*,4′*R*,5′*S*,6′*S*,7*R*,8*S*,9*R*,10*R*,12*R*,14*R*,15*R*,16*S*,16a*S*)-7-ethyl-5′,8,9,14-tetrahydroxy-4′-methoxy-4′,6′,8,10,12,14,16-heptamethyl-15-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]hexadecahydrospiro[5*H*,11*H*-1,3-dioxino[5,4-*c*]oxacyclotetradecin-2,2′-pyrane]-5,11-dione (erythromycin E),



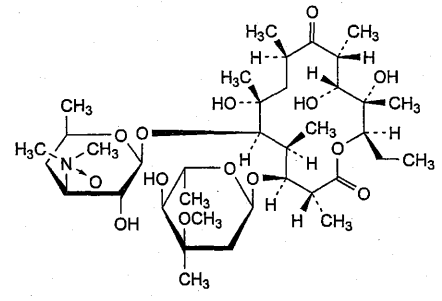
D. (1*S*,2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadecan-7-one (anhydroerythromycin A),



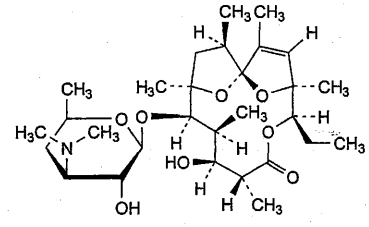
E. (2*R*,3*R*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*)-9-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-5-ethyl-3,4-dihydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15-dioxabicyclo[10.2.1]pentadec-1(14)-en-7-one (erythromycin A enol ether),



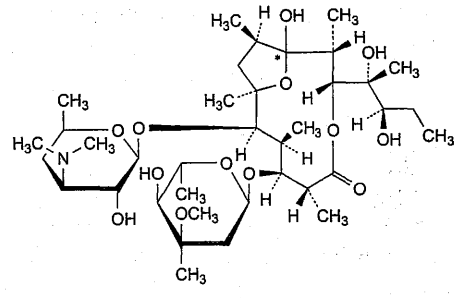
F. (2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridec-1(12)-en-5-one (pseudoerythromycin A enol ether),



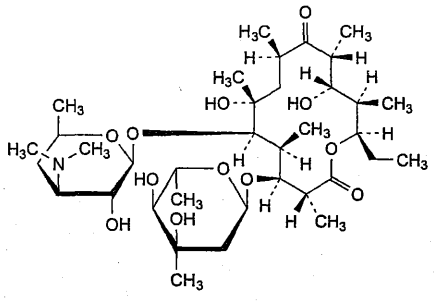
H. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione *N*-oxide (erythromycin A 3′′-*N*-oxide),



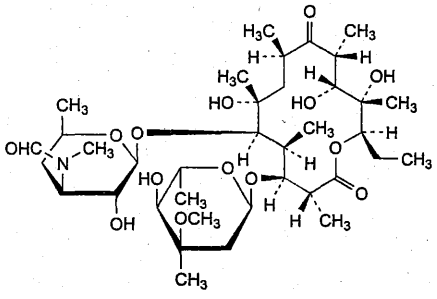
I. (1*S*,4*S*,5*R*,8*R*,9*S*,10*S*,11*R*,12*R*,14*R*)-5-ethyl-9-hydroxy-2,4,8,10,12,14-hexamethyl-11-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-6,15,16-trioxatricyclo[10.2.1.1^{1,4}]hexadec-2-en-7-one (erythralosamine),



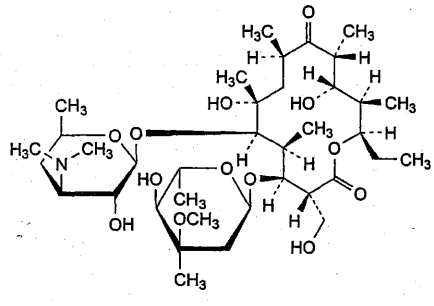
J. (1*RS*,2*R*,3*R*,6*R*,7*S*,8*S*,9*R*,10*R*,12*R*)-7-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-3-[(1*R*,2*R*)-1,2-dihydroxy-1-methylbutyl]-1-hydroxy-2,6,8,10,12-pentamethyl-9-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]-4,13-dioxabicyclo[8.2.1]tridecan-5-one (pseudoerythromycin A hemiketal),



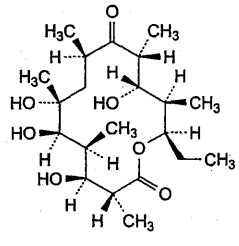
K. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin D),



L. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*R*,13*S*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(formylmethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (3′′-*N*-demethyl-3′′-*N*-formylerythromycin A),



M. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-4-[(2,6-dideoxy-3-*C*-methyl-3-*O*-methyl-α-L-*ribo*-hexopyranosyl)oxy]-14-ethyl-7,12-dihydroxy-3-(hydroxymethyl)-5,7,9,11,13-pentamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-*xylo*-hexopyranosyl]oxy]oxacyclotetradecane-2,10-dione (erythromycin G),



N. (3*R*,4*S*,5*S*,6*R*,7*R*,9*R*,11*R*,12*S*,13*R*,14*R*)-14-ethyl-4,6,7,12-tetrahydroxy-3,5,7,9,11,13-hexamethyloxacyclotetradecane-2,10-dione (erythronolide B),

S. unknown structure.

Ph Eur

Erythropoietin Concentrated Solution

(Ph. Eur. monograph 1316)

The label states (1) the type of erythropoeitin using the appropriate International Nonproprietary Name (Epoetin Alfa, Epoetin Beta, etc) and (2) the approved code in lower case letters indicative of the method of production.

APPRLICDSR	VLERYLLEAK	EAENITTGCA	EHCSLNENIT	40
VPDTKVNFYA	WKRMEVGQQA	VEVWQGLALL	SEAVLRGQAL	80
LVNSSQPWEP	LQLHVDKAVS	GLRSLTTLLR	ALGAQKEAIS	120
PPDAASAAPL	RTITADTFRK	LFRVYSNFLR	GKLKLYTGEA	160
CRTGD				165

disulfide bridges: 7-161, 29-33

M_r approx. 30 600
(glycosylated protein)

Action and use
Erythropoietin analogue.

Preparation
Erythropoietin Injection

Ph Eur

DEFINITION

Solution containing a family of closely-related glycoproteins which are indistinguishable from the naturally occurring human erythropoietin (urinary erythropoietin) in terms of amino acid sequence (165 amino acids) and average glycosylation pattern, at a concentration of 0.5-10 mg/mL. It may also contain buffer salts and other excipients. It has a potency of not less than 100 000 IU per milligram of active substance determined using the conditions described under Assay and in the test for protein.

PRODUCTION

Erythropoietin is produced in rodent cells *in vitro* by a method based on recombinant DNA technology.

Prior to batch release, the following tests are carried out on each batch of the final product, unless exemption has been granted by the competent authority.

Host cell-derived proteins
The limit is approved by the competent authority.

Host cell- and vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS

Appearance
Clear or slightly turbid, colourless solution.

IDENTIFICATION

A. It gives the appropriate response when examined using the conditions described under Assay.

B. Capillary zone electrophoresis (*2.2.47*).

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 1 mg/mL. Desalt 0.25 mL of the solution by passage through a micro-concentrator cartridge provided with a membrane with a molecular mass cut-off of not more than 10 000 Da. Add 0.2 mL of *water R* to the sample and desalt again. Repeat the desalting procedure once more. Dilute the sample with *water R*, determine its protein concentration as described under Tests and adjust to a concentration of approximately 1 mg/mL with *water R*.

Reference solution Dissolve the contents of a vial of *erythropoietin for physicochemical tests CRS* in 0.10 mL of *water R*. Proceed with desalting as described for the test solution.

Capillary:
— *material*: uncoated fused silica;
— *size*: effective length = about 100 cm, Ø = 50 µm.

Temperature 35 °C.

CZE buffer concentrate (0.1 M sodium chloride, 0.1 M tricine, 0.1 M sodium acetate) Dissolve 0.584 g of *sodium chloride R*, 1.792 g of *tricine R* and 0.820 g of *anhydrous sodium acetate R* in *water R* and dilute to 100.0 mL with the same solvent.

1 M putrescine solution Dissolve 0.882 g of *putrescine R* in 10 mL of *water R*. Distribute in 0.5 mL aliquots.

CZE buffer (0.01 M tricine, 0.01 M sodium chloride, 0.01 M sodium acetate, 7 M urea, 2.5 mM putrescine) Dissolve 21.0 g of *urea R* in 25 mL of *water R* by warming in a water-bath at 30 °C. Add 5.0 mL of CZE buffer concentrate and 125 µL of 1 M putrescine solution. Dilute to 50.0 mL with *water R*. Using *dilute acetic acid R*, adjust to pH 5.55 at 30 °C and filter through a membrane filter (nominal pore size 0.45 µm).

Detection Spectrophotometer at 214 nm.

Set the autosampler to store the samples at 4 °C during analysis.

Preconditioning of the capillary Rinse the capillary for 60 min with *0.1 M sodium hydroxide* filtered through a membrane filter (nominal pore size 0.45 µm) and for 60 min with CZE buffer. Apply voltage for 12 h (20 kV).

Between-run rinsing Rinse the capillary for 10 min with *water R*, for 5 min with *0.1 M sodium hydroxide* filtered through a membrane filter (nominal pore size 0.45 µm) and for 10 min with CZE buffer.

Injection Under pressure or vacuum.

Migration Apply a field strength of 143 V/cm (15.4 kV for capillaries of 107 cm total length) for 80 min, using CZE buffer as the electrolyte in both buffer reservoirs.

System suitability In the electropherogram obtained with the reference solution, a pattern of well separated peaks corresponding to the peaks in the electropherogram of erythropoietin supplied with *erythropoietin for physicochemical tests CRS* is seen, and the largest peak is at least 50 times greater than the baseline noise. If necessary, adjust the sample load to give peaks of sufficient height. Identify the peaks due to isoforms 1 to 8. Isoform 1 may not be visible. The peak due to isoform 8 is detected and the resolution between the peaks due to isoforms 5 and 6 is not less than 1. Repeat the separation at least 3 times. The baseline is stable, showing little drift, and the distribution of peaks is qualitatively and quantitatively similar to the distribution of peaks in the electropherogram of erythropoietin supplied with *erythropoietin for physicochemical tests CRS*. The relative standard deviation of the migration time of the peak due to isoform 2 is less than 2 per cent.

Limits Identify the peaks due to isoforms 1 to 8 in the electropherogram obtained with the test solution by comparison with the electropherogram obtained with the reference solution. Calculate the percentage content of each isoform from the corresponding peak area. The percentages are within the following ranges:

Isoform	Content (per cent)
1	0 - 15
2	0 - 15
3	1 - 20
4	10 - 35
5	15 - 40
6	10 - 35
7	5 - 25
8	0 - 15

C. Polyacrylamide gel electrophoresis and immunoblotting.

(a) Polyacrylamide gel electrophoresis (*2.2.31*).

Gel dimensions 0.75 mm thick, about 16 cm square.

Resolving gel 12 per cent acrylamide.

Sample buffer *concentrated SDS-PAGE sample buffer R.*

Test solution (a) Dilute the preparation to be examined in *water R* to obtain a concentration of 1.0 mg/mL.
To 1 volume of this solution add 1 volume of sample buffer.

Test solution (b) Dilute the preparation to be examined in *water R* to obtain a concentration of 0.1 mg/mL.
To 1 volume of this solution add 1 volume of sample buffer.

Reference solution (a) Dissolve the contents of a vial of *erythropoietin for physicochemical tests CRS* in 0.10 mL of *water R*. To 1 volume of this solution add 1 volume of sample buffer.

Reference solution (b) Dissolve the contents of a vial of *erythropoietin for physicochemical tests CRS* in *water R* and dilute with the same solvent to obtain a concentration of 0.1 mg/mL. To 1 volume of this solution add 1 volume of sample buffer.

Reference solution (c) A solution of molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 10-70 kDa.

Reference solution (d) A solution of pre-stained molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 10-70 kDa and suitable for the electrotransfer to an appropriate membrane.

Sample treatment Boil for 2 min.

Application 20 µL, in the following order: reference solution (c), reference solution (a), test solution (a), empty well, reference solution (b), test solution (b), reference solution (d).

At the end of the separation, remove the gel-cassette from the apparatus and cut the gel into 2 parts: the first part containing reference solution (c), reference solution (a) and test solution (a); the second part containing reference solution (b), test solution (b) and reference solution (d).

Detection By Coomassie staining on the first part of the gel.

System suitability Reference solution (c):
— the validation criteria are met.

Results The electropherogram obtained with test solution (a) shows a single diffuse band corresponding in position and intensity to the single band seen in the electropherogram obtained with reference solution (a).

(b) Immunoblotting.

Transfer the second part of the gel onto a membrane suitable for the immobilisation of proteins, using commercially available electrotransfer equipment and following the manufacturer's instructions. After electrotransfer, incubate the membrane in a neutral isotonic buffer containing a suitable blocking agent (for example, 50 g/L of dried milk or 10 per cent *V/V* foetal calf serum), for 1-2 h, followed by incubation for 1-14 h in the same blocking solution with a

suitable dilution of either a polyclonal or monoclonal anti-erythropoietin antibody. Detect erythropoietin-bound antibody using a suitable enzyme- or radiolabelled antibody (for example, an alkaline phosphatase-conjugated second antibody). The precise details of blocking agents, concentrations and incubation times should be optimised using the principles set out in *Immunochemical methods* (*2.7.1*).

System suitability In the electropherogram obtained with reference solution (d), the molecular mass markers are resolved on the membrane into discrete bands, with a linear relationship between distance migrated and logarithm$_{10}$ of the molecular mass.

Results The electropherogram obtained with test solution (b) shows a single broad band corresponding in position and intensity to the single band seen in the electropherogram obtained with reference solution (b).

D. Peptide mapping (*2.2.55*).

Selective cleavage of the peptide bonds

Test solution Dilute the preparation to be examined in *tris acetate buffer solution pH 8.5 R* to a concentration of 1.0 mg/mL. Equilibrate the solution in *tris acetate buffer solution pH 8.5 R* using a suitable procedure (dialysis against *tris acetate buffer solution pH 8.5 R*, or membrane filtration using the procedure described under Identification B, but reconstituting the desalted sample with *tris acetate buffer solution pH 8.5 R*, are suitable). Transfer the dialysed solution to a polypropylene centrifuge tube. Freshly prepare a solution of *trypsin for peptide mapping R* at a concentration of 1 mg/mL in *water R*, and add 2 µL to 0.10 mL of the dialysed solution. Cap the tube and place in a water-bath at 37 °C for 18 h. Remove the sample from the water-bath and stop the reaction immediately by freezing.

Reference solution Dissolve the contents of a vial of *erythropoietin for physicochemical tests CRS* in 0.10 mL of *water R*. Prepare as for the test solution, ensuring that all procedures are carried out simultaneously, and under identical conditions.

Chromatographic separation. Liquid chromatography (*2.2.29*).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase:
- *mobile phase A*: 0.06 per cent *V/V* solution of *trifluoroacetic acid R*;
- *mobile phase B*: to 100 mL of *water for chromatography R* add 0.6 mL of *trifluoroacetic acid R* and dilute to 1000 mL with *acetonitrile R1*;

Time (min)	Flow rate (mL/min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	0.75	100	0
10 - 125	0.75	100 → 39	0 → 61
125 - 135	1.25	39 → 17	61 → 83
135 - 145	1.25	17 → 0	83 → 100
145 - 150	1.25	100	0

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability The chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of erythropoietin digest supplied with *erythropoietin for physicochemical tests CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

E. *N*-terminal sequence analysis.

The first 15 amino acids are: Ala - Pro - Pro - Arg - Leu - Ile - (no recovered peak) - Asp - Ser - Arg - Val - Leu - Glu - Arg - Tyr.

Perform the Edman degradation using an automated solid-phase sequencer, operated in accordance with the manufacturer's instructions.

Desalt the equivalent of 50 µg of erythropoietin. For example, dilute a volume of the preparation to be examined equivalent to 50 µg of the active substance in 1 mL of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R*. Pre-wash a C18 reverse-phase sample preparation cartridge according to the instructions supplied and equilibrate the cartridge in a 0.1 per cent *V/V* solution of *trifluoroacetic acid R*. Apply the sample to the cartridge, and wash successively with a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* containing 0 per cent, 10 per cent and 50 per cent *V/V* of *acetonitrile R* according to the manufacturer's instructions. Lyophilise the 50 per cent *V/V* *acetonitrile R* eluate.

Redissolve the desalted sample in 50 µL of a 0.1 per cent *V/V* solution of *trifluoroacetic acid R* and couple to a sequencing cartridge using the protocol provided by the manufacturer. Run 15 sequencing cycles, using the reaction conditions for proline when running the 2nd and 3rd cycles.

Identify the phenylthiohydantoin (PTH)-amino acids released at each sequencing cycle by reverse-phase liquid chromatography. The procedure may be carried out using the column and reagents recommended by the manufacturer of the sequencing equipment for the separation of PTH-amino-acids.

The separation procedure is calibrated using:
- the mixture of PTH-amino acids provided by the manufacturer of the sequencer, with the gradient conditions adjusted as indicated to achieve optimum resolution of all amino acids;
- a sample obtained from a blank sequencing cycle obtained as recommended by the equipment manufacturer.

TESTS

Protein (*2.5.33, Method I*)

80 per cent to 120 per cent of the stated concentration.

Test solution Dilute the preparation to be examined in a 4 g/L solution of *ammonium hydrogen carbonate R*.

Record the absorbance spectrum between 250 nm and 400 nm. Measure the value at the absorbance maximum (276-280 nm), after correction for any light scattering, measured up to 400 nm. Calculate the concentration of erythropoietin taking the specific absorbance to be 7.43.

Dimers and related substances with molecular masses greater than that of erythropoietin

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Dilute the preparation to be examined in the mobile phase to obtain a concentration of 0.2 mg/mL.

Resolution solution Dissolve the contents of a vial of *erythropoietin for SEC system suitability CRS* in the mobile phase to obtain a concentration of 0.2 mg/mL.

Reference solution Dilute 20 µL of the resolution solution to 1.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.3 m, Ø = 7.5 mm;
— *stationary phase*: *hydrophilic silica gel for chromatography R* (5 µm), of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 20 000 to 200 000.

Mobile phase Dissolve 1.15 g of *anhydrous disodium hydrogen phosphate R*, 0.2 g of *potassium dihydrogen phosphate R* and 23.4 g of *sodium chloride R* in 900 mL of *water for chromatography R*, adjust to pH 7.4 if necessary, and dilute to 1000 mL with *water for chromatography R*.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 100 µL.

Run time Twice the retention time of erythropoietin monomer.

Relative retention With reference to erythropoietin monomer: erythropoietin dimer = about 0.9.

System suitability:
— the area of the principal peak in the chromatogram obtained with the reference solution is 1.5 per cent to 2.5 per cent of the area of the principal peak in the chromatogram obtained with the resolution solution;
— *resolution*: minimum 1.5 between the peaks due to erythropoietin dimer and erythropoietin monomer in the chromatogram obtained with the resolution solution.

Limit:
— *sum of the peaks eluted before the principal peak*: maximum 2.0 per cent; disregard any peak with a retention time greater than that of the principal peak.

Sialic acids

Minimum 10 mol of sialic acids (calculated as *N*-acetylneuraminic acid) per mole of erythropoietin.

Test solution (a) Dilute the preparation to be examined in the mobile phase used in the test for dimers and related substances with molecular masses greater than that of erythropoietin to obtain a concentration of 0.3 mg/mL.

Test solution (b) To 0.5 mL of test solution (a) add 0.5 mL of the mobile phase used in the test for dimers and related substances with molecular masses greater than that of erythropoietin.

Reference solution (a) Dissolve a suitable amount of *N-acetylneuraminic acid R* in *water R* to obtain a concentration of 0.1 mg/mL.

Reference solution (b) To 0.8 mL of reference solution (a) add 0.2 mL of *water R*.

Reference solution (c) To 0.6 mL of reference solution (a) add 0.4 mL of *water R*.

Reference solution (d) To 0.4 mL of reference solution (a) add 0.6 mL of *water R*.

Reference solution (e) To 0.2 mL of reference solution (a) add 0.8 mL of *water R*.

Reference solution (f) Use *water R*.

Carry out the test in triplicate. Transfer 100 µL of each of the test and reference solutions to 10 mL glass test tubes. To each tube add 1.0 mL of *resorcinol reagent R*. Stopper the tubes and incubate at 100 °C for 30 min. Cool on ice. To each tube, add 2.0 mL of a mixture of 12 volumes of *butanol R* and 48 volumes of *butyl acetate R*. Mix vigorously, and allow the 2 phases to separate. Ensuring that the upper phase is completely clear, remove the upper phase, taking care to exclude completely any of the lower phase. Measure the absorbance (*2.2.25*) of all samples at 580 nm. Using the calibration curve generated by the reference solutions, determine the content of sialic acids in test solutions (a) and (b) and calculate the mean. Calculate the number of moles of sialic acids per mole of erythropoietin assuming that the relative molecular mass of erythropoietin is 30 600 and that the relative molecular mass of *N*-acetylneuraminic acid is 309.

System suitability:
— the individual replicates agree to within ± 10 per cent of each other;
— the value obtained from reference solution (a) is between 1.5 and 3.3 times that obtained with test solution (a).

Bacterial endotoxins (*2.6.14*)

Less than 20 IU in the volume that contains 100 000 IU of erythropoietin.

ASSAY

The activity of the preparation is compared with that of *erythropoietin BRP* and expressed in International Units (IU).

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits of the estimated potency (P = 0.95) are not less than 64 per cent and not more than 156 per cent of the stated potency.

Carry out the determination of potency by Method A or B.

A. In polycythaemic mice

The activity of the preparation is estimated by examining, under given conditions, its effect in stimulating the incorporation of ^{59}Fe into circulating red blood cells of mice made polycythaemic by exposure to reduced atmospheric pressure.

The following schedule, using treatment in a hypobaric chamber, has been found to be suitable.

Induce polycythaemia in female mice of the same strain, weighing 16-18 g. Place the mice in a hypoxic chamber and reduce the pressure to 0.6 atmospheres. After 3 days at 0.6 atmospheres, further reduce the pressure to 0.4-0.5 atmospheres and maintain the animals at this pressure for a further 11 days (the partial vacuum is interrupted daily for a maximum of 1 h at about 11:00 a.m., in order to clean the cages and feed the animals). At the end of the specified period, return the mice to normal atmospheric conditions. Randomly distribute the mice into cages, each containing 6 animals, and mark them.

Test solution (a) Dilute the substance to be examined in *phosphate-albumin buffered saline pH 7.2 R1* to obtain a concentration of 0.2 IU/mL.

Test solution (b) Mix equal volumes of test solution (a) and *phosphate-albumin buffered saline pH 7.2 R1*.

Test solution (c) Mix equal volumes of test solution (b) and *phosphate-albumin buffered saline pH 7.2 R1*.

Reference solution (a) Dissolve *erythropoietin BRP* in *phosphate-albumin buffered saline pH 7.2 R1* to obtain a concentration of 0.2 IU/mL.

Reference solution (b) Mix equal volumes of reference solution (a) and *phosphate-albumin buffered saline pH 7.2 R1*.

Reference solution (c) Mix equal volumes of reference solution (b) and *phosphate-albumin buffered saline pH 7.2 R1*.

Radiolabelled ferric [^{59}Fe] chloride solution, concentrated Use a commercially available solution of [^{59}Fe]ferric chloride (approximate specific activity: 100-1000 MBq/mg of Fe).

Radiolabelled [^{59}Fe]ferric chloride solution Dilute the concentrated radiolabelled [^{59}Fe]ferric chloride solution in *Sodium citrate buffer solution pH 7.8 (0.034 M sodium citrate, 0.101 M sodium chloride) R* to obtain a solution with an activity of 3.7×10^4 Bq/mL.

The concentrations of the test solutions and reference solutions may need to be modified, based on the response range of the animals used.

3 days after returning the animals to atmospheric pressure, inject each animal subcutaneously with 0.2 mL of one of the solutions. The 6 animals in each cage must each receive one of the 6 different treatments (3 test solutions and 3 reference solutions), and the order of injection must be separately randomised for each cage. A minimum of 8 cages is recommended. 2 days after injection of the test or reference solution, inject each animal intraperitoneally with 0.2 mL of radiolabelled [^{59}Fe]ferric chloride solution. The order of the injections must be the same as that of the erythropoietin injections, and the time interval between administration of the erythropoietin and the radiolabelled ferric chloride solution must be the same for each animal. After a further 48 h, anaesthetise each animal by injection of a suitable anaesthetic, record body weights and withdraw blood samples (0.65 mL) into haematocrit capillaries from the bifurcation of the aorta. After determining the packed cell volume for each sample, measure the radioactivity.

Calculate the response (percentage of iron-59 in total circulating blood) for each mouse using the expression:

$$\frac{A_s \times M \times 7.5}{A_t \times V_s}$$

A_s = radioactivity in the sample;
A_t = total radioactivity injected;
7.5 = total blood volume as per cent body weight;
M = body weight, in grams;
V_s = sample volume.

Calculate the potency by the usual statistical methods for a parallel line assay. Eliminate from the calculation any animal where the packed cell volume is less than 54 per cent, or where the body weight is more than 24 g.

B. In normocythaemic mice

The assay is based on the measurement of stimulation of reticulocyte production in normocythaemic mice.

The assay may be carried out using the following procedure:

Test solution (a) Dilute the preparation to be examined in *phosphate-albumin buffered saline pH 7.2 R1* to obtain a concentration of 80 IU/mL.

Test solution (b) Mix equal volumes of test solution (a) and *phosphate-albumin buffered saline pH 7.2 R1.*

Test solution (c) Mix equal volumes of test solution (b) and *phosphate-albumin buffered saline pH 7.2 R1.*

Reference solution (a) Dissolve *erythropoietin BRP* in *phosphate-albumin buffered saline pH 7.2 R1* to obtain a concentration of 80 IU/mL.

Reference solution (b) Mix equal volumes of reference solution (a) and *phosphate-albumin buffered saline pH 7.2 R1.*

Reference solution (c) Mix equal volumes of reference solution (b) and *phosphate-albumin buffered saline pH 7.2 R1.*

The exact concentrations of the test solutions and reference solutions may need to be modified, based on the response range of the animals used.

At the beginning of the assay procedure, randomly distribute mice of a suitable age and strain (8-week old B6D2F1 mice are suitable) into 6 cages. A minimum of 8 mice per cage is recommended. Inject each animal subcutaneously with 0.5 mL of the appropriate treatment (one solution per cage) and put the animal in a new cage. Combine the mice in such a way that each cage housing the treated mice contains one mouse out of the 6 different treatments (3 test solutions and 3 reference solutions, 6 mice per cage). 4 days after the injections, collect blood samples from the animals and determine the number of reticulocytes using a suitable procedure.

The following method may be employed:

The volume of blood, dilution procedure and fluorescent reagent may need to be modified to ensure maximum development and stability of fluorescence.

Colorant solution, concentrated Use a solution of thiazole orange suitable for the determination of reticulocytes. Prepare at a concentration twice that necessary for the analysis.

Proceed with the following dilution steps. Dilute whole blood 500-fold in the buffer used to prepare the colorant solution. Dilute this solution 2-fold in the concentrated colorant solution. After staining for 3-10 min, determine the reticulocyte count microfluorometrically in a flow cytometer. The percentage of reticulocytes is determined using a biparametric histogram: number of cells/red fluorescence (620 nm).

Calculate the potency by the usual statistical methods for a parallel line assay.

STORAGE

In an airtight container at a temperature below −20 °C. Avoid repeated freezing and thawing.

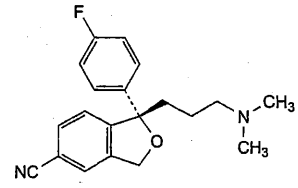
LABELLING

The label states:
— the erythropoietin content in milligrams per millilitre;
— the activity in International Units per millilitre;
— the name and the concentration of any other excipients.

Ph Eur

Escitalopram

(Ph. Eur. monograph 2758)



$C_{20}H_{21}FN_2O$ 324.4 *128196-01-0*

Action and use

Selective serotonin reuptake inhibitor; antidepressant.

Ph Eur

DEFINITION

(1*S*)-1-[3-(Dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carbonitrile.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol and in toluene.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *escitalopram CRS*.

B. Enantiomeric purity (see Tests).

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 3.4 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*; adjust to pH 3.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*.

Test solution Dissolve 25 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *escitalopram for system suitability CRS* (containing impurity D) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 45 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R*, buffer solution (10:90 *V/V*);
- *mobile phase B*: buffer solution, *acetonitrile R* (35:65 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Flow rate (mL/min)
0 - 2	95	5	1.0
2 - 37	95 → 65	5 → 35	1.0
37 - 47	65 → 0	35 → 100	1.0
47 - 62	0	100	2.0

Detection Spectrophotometer at 237 nm and at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *escitalopram for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to escitalopram (retention time = about 38 min): impurity D = about 0.98.

System suitability Reference solution (a) at 237 nm:
- *peak-to-valley ratio*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to escitalopram.

Calculation of percentage contents:
- for each impurity, use the concentration of escitalopram in reference solution (b) at 237 nm.

Limits:
- *unspecified impurities at 237 nm and at 254 nm*: for each impurity, maximum 0.10 per cent;
- *total at 237 nm*: maximum 0.5 per cent;
- *reporting threshold at 237 nm*: 0.05 per cent.

Enantiomeric purity

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *citalopram hydrobromide CRS* (containing equal amounts of impurity K and escitalopram) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *protein derivative of silica gel for chiral separation R* (5 µm);
- *temperature*: 30 °C.

Mobile phase *acetonitrile R, 0.05 M phosphate buffer solution pH 7.0 R* (15:85 *V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 15 µL.

Run time Twice the retention time of escitalopram.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity K.

Relative retention With reference to escitalopram (retention time = about 11 min): impurity K = about 1.2.

System suitability Reference solution (a):
- *resolution*: minimum 1.3 between the peaks due to escitalopram and impurity K;
- *symmetry factor*: maximum 4.0 for the peak due to escitalopram; maximum 4.0 for the peak due to impurity K.

Limits:
- *impurity K*: maximum 2.0 per cent;
- *reporting threshold*: 0.10 per cent (reference solution (b)).

Water (*2.5.12*)

Maximum 0.2 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of *acetic anhydride R* and 75 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

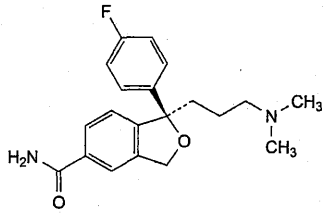
1 mL of *0.1 M perchloric acid* is equivalent to 32.44 mg of $C_{20}H_{21}FN_2O$.

IMPURITIES

Specified impurities *K*.

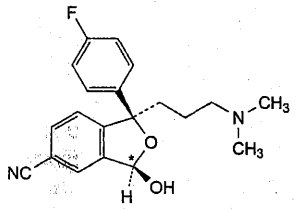
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities

in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I, J, L, M.*



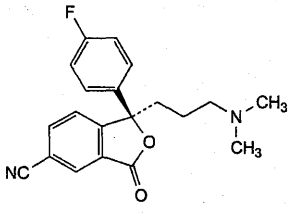
and enantiomer

A. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carboxamide,



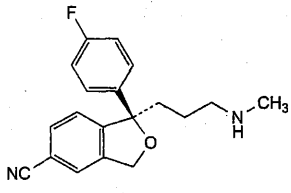
its epimer at C* and their enantiomers

B. mixture of (1*RS*,3*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile and (1*RS*,3*SR*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile,



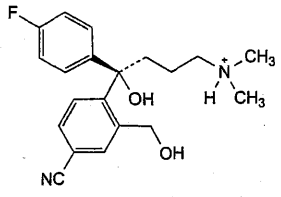
and enantiomer

C. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-oxo-1,3-dihydro-2-benzofuran-5-carbonitrile,



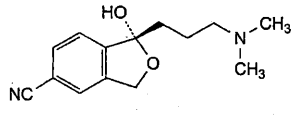
and enantiomer

D. (1*RS*)-1-(4-fluorophenyl)-1-[3-(methylamino)propyl]-1,3-dihydro-2-benzofuran-5-carbonitrile,



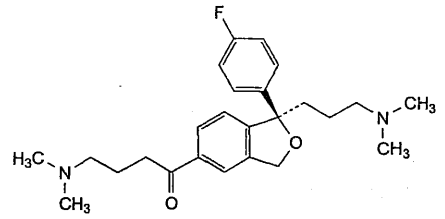
and enantiomer

E. (4*RS*)-4-[4-cyano-2-(hydroxymethyl)phenyl]-4-(4-fluorophenyl)-4-hydroxy-*N*,*N*-dimethylbutan-1-aminium,



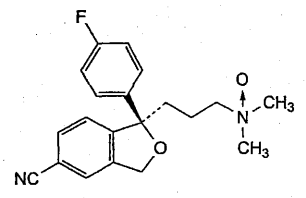
and enantiomer

F. (1*RS*)-1-[3-(dimethylamino)propyl]-1-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile,



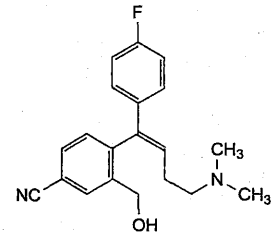
and enantiomer

G. 4-(dimethylamino)-1-[(1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-yl]butan-1-one,

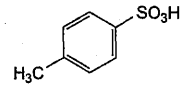


and enantiomer

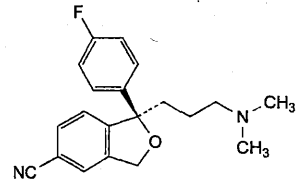
H. 3-[(1*RS*)-5-cyano-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine *N*-oxide,



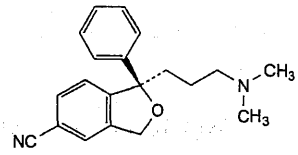
I. 4-[(1*Z*)-4-(dimethylamino)-1-(4-fluorophenyl)but-1-en-1-yl]-3-(hydroxymethyl)benzonitrile,



J. 4-methylbenzenesulfonic acid (*p*-toluenesulfonic acid),

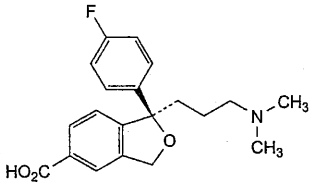


K. (1*R*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carbonitrile,



and enantiomer

L. (1*RS*)-1-[3-(dimethylamino)propyl]-1-phenyl-1,3-dihydro-2-benzofuran-5-carbonitrile,



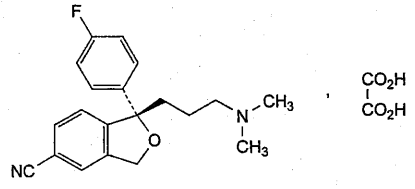
and enantiomer

M. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carboxylic acid.

Ph Eur

Escitalopram Oxalate

(Ph. Eur. monograph 2733)



$C_{22}H_{23}FN_2O_5$ 414.4 *219861-08-2*

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Ph Eur

DEFINITION
(1*S*)-1-[3-(Dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carbonitrile hydrogen oxalate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, freely soluble in methanol, slightly soluble in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *escitalopram oxalate CRS*.

B. Enantiomeric purity (see Tests).

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 3.4 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjust to pH 3.0 with *phosphoric acid R* and dilute to 1.0 L with *water R*.

Test solution Dissolve 25 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *escitalopram for system suitability CRS* (containing impurity D) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 45 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R*, buffer solution (10:90 *V/V*);
— *mobile phase B*: buffer solution, *acetonitrile R* (35:65 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Flow rate (mL/min)
0 - 2	95	5	1.0
2 - 37	95 → 65	5 → 35	1.0
37 - 47	65 → 0	35 → 100	1.0
47 - 62	0	100	2.0

Detection Spectrophotometer at 237 nm and at 254 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *escitalopram for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to escitalopram (retention time = about 38 min): impurity D = about 0.98.

System suitability Reference solution (a) at 237 nm:
— *peak-to-valley ratio*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to escitalopram.

Calculation of percentage contents:
— for each impurity, use the concentration of escitalopram in reference solution (b) at 237 nm.

Limits:
— *unspecified impurities at 237 nm and at 254 nm*: for each impurity, maximum 0.10 per cent;
— *total at 237 nm*: maximum 0.5 per cent;
— *reporting threshold at 237 nm*: 0.05 per cent.

Enantiomeric purity
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *citalopram hydrobromide CRS* (containing equal amounts of impurity K and escitalopram) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *protein derivative of silica gel for chiral separation R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *acetonitrile R, 0.05 M phosphate buffer solution pH 7.0 R* (15:85 *V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 15 µL.

Run time Twice the retention time of escitalopram.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity K.

Relative retention With reference to escitalopram (retention time = about 11 min): impurity K = about 1.2.

System suitability Reference solution (a):
— *resolution*: minimum 1.3 between the peaks due to escitalopram and impurity K;
— *symmetry factor*: maximum 4.0 for the peak due to escitalopram; maximum 4.0 for the peak due to impurity K.

Limits:
— *impurity K*: maximum 2.0 per cent;
— *reporting threshold*: 0.10 per cent (reference solution (b)).

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 0.250 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

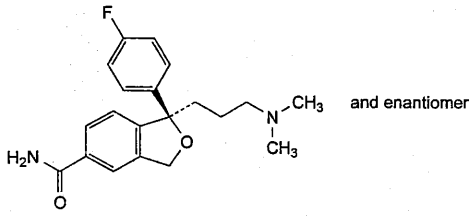
Dissolve 0.300 g in a mixture of 5.0 mL of *acetic anhydride R* and 75 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 41.44 mg of $C_{22}H_{23}FN_2O_5$.

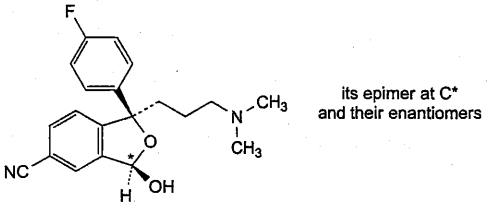
IMPURITIES

Specified impurities *K*.

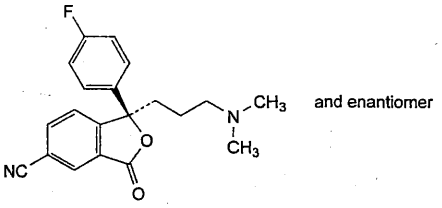
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I, J, L, M.*



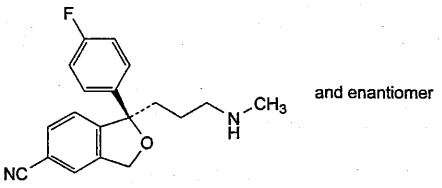
A. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carboxamide,



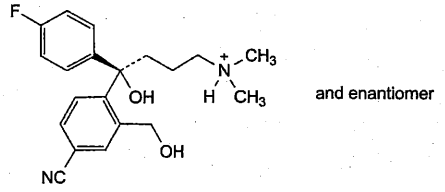
B. mixture of (1*RS*,3*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile and (1*RS*,3*SR*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile,



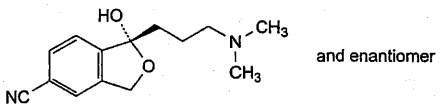
C. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-3-oxo-1,3-dihydro-2-benzofuran-5-carbonitrile,



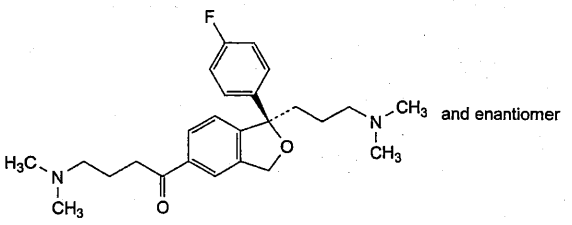
D. (1*RS*)-1-(4-fluorophenyl)-1-[3-(methylamino)propyl]-1,3-dihydro-2-benzofuran-5-carbonitrile,



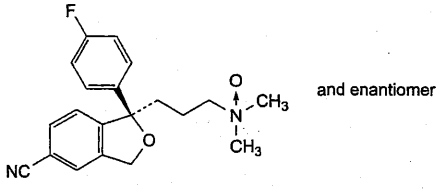
E. (4*RS*)-4-[4-cyano-2-(hydroxymethyl)phenyl]-4-(4-fluorophenyl)-4-hydroxy-*N*,*N*-dimethylbutan-1-aminium,



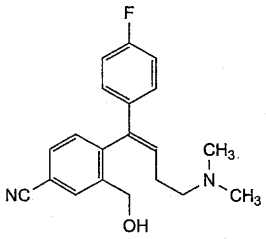
F. (1*RS*)-1-[3-(dimethylamino)propyl]-1-hydroxy-1,3-dihydro-2-benzofuran-5-carbonitrile,



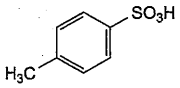
G. 4-(dimethylamino)-1-[(1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-yl]butan-1-one,



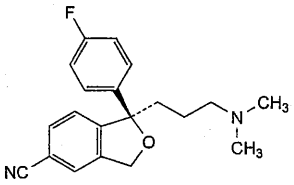
H. 3-[(1*RS*)-5-cyano-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-1-yl]-*N*,*N*-dimethylpropan-1-amine *N*-oxide,



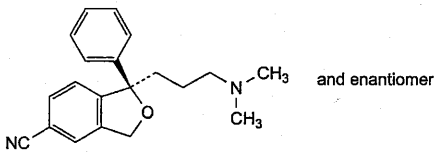
I. 4-[(1*Z*)-4-(dimethylamino)-1-(4-fluorophenyl)but-1-en-1-yl]-3-(hydroxymethyl)benzonitrile,



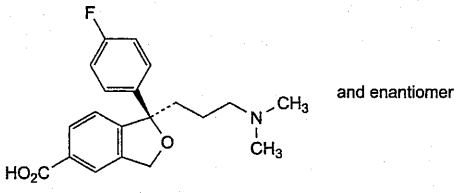
J. 4-methylbenzenesulfonic acid (*p*-toluenesulfonic acid),



K. (1*R*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carbonitrile,



L. (1*RS*)-1-[3-(dimethylamino)propyl]-1-phenyl-1,3-dihydro-2-benzofuran-5-carbonitrile,

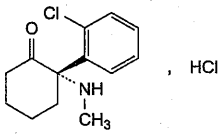


M. (1*RS*)-1-[3-(dimethylamino)propyl]-1-(4-fluorophenyl)-1,3-dihydro-2-benzofuran-5-carboxylic acid.

Ph Eur

Esketamine Hydrochloride

(Ph. Eur. monograph 1742)



$C_{13}H_{17}Cl_2NO$ 274.2 *33643-47-9*

Action and use
General anaesthetic.

Ph Eur

DEFINITION

(2*S*)-2-(2-Chlorophenyl)-2-(methylamino)cyclohexanone hydrochloride.

Content
99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in methanol, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 85.0 to + 95.0.

Dilute 12.5 mL of solution S (see Tests) to 40.0 mL with *water R*.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of esketamine hydrochloride.*

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 8.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
3.5 to 4.5.

Dilute 12.5 mL of solution S to 20 mL with *carbon dioxide-free water R*.

Impurity D
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *esketamine impurity D CRS* in *water R*, add 20 mL of the test solution and dilute to 50 mL with *water R*. Dilute 10 mL of this solution to 100 mL with *water R*.

Reference solution (b) Dilute 5.0 mL of the test solution to 25.0 mL with *water R*. Dilute 5.0 mL of this solution to 50.0 mL with *water R*.

Reference solution (c) Dilute 2.5 mL of reference solution (b) to 10.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Precolumn:
— *size*: *l* = 0.01 m, Ø = 3.0 mm;
— *stationary phase*: *silica gel AGP for chiral chromatography R* (5 µm);
— *temperature*: 30 °C.

Column:
— *size*: *l* = 0.125 m, Ø = 4.6 mm;
— *stationary phase*: *silica gel AGP for chiral chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Mix 16 volumes of *methanol R* and 84 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 7.0 with *potassium hydroxide R*.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 20 min.

Relative retention With reference to esketamine (retention time = about 10 min): impurity D = about 1.3.

System suitability:
— *resolution*: minimum 2.0 between the peaks due to esketamine and impurity D in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 3 for the principal peak in the chromatogram obtained with reference solution (c).

Limit:
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *ketamine impurity A CRS* in the mobile phase (using ultrasound, if necessary) and dilute to 10 mL with the mobile phase. To 1 mL of the solution add 0.5 mL of the test solution and dilute to 100 mL with the mobile phase. Prepare immediately before use.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 0.95 g of *sodium hexanesulfonate R* in 1000 mL of a mixture of 25 volumes of *acetonitrile R* and 75 volumes of *water R* and add 4 mL of *acetic acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 10 times the retention time of esketamine.

Relative retention With reference to esketamine: impurity A = about 1.6; impurity B = about 3.3; impurity C = about 4.6.

System suitability Reference solution (a):
— *retention time*: esketamine = 3.0 min to 4.5 min;
— *resolution*: minimum 1.5 between the peaks due to impurity A and esketamine.

Limits:
— *impurities A, B, C*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

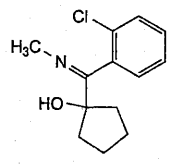
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 50 mL of *methanol R* and add 1.0 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

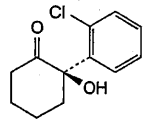
1 mL of *0.1 M sodium hydroxide* is equivalent to 27.42 mg of $C_{13}H_{17}Cl_2NO$.

STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C, D.*

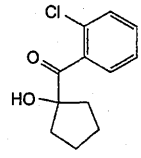


A. 1-[(2-chlorophenyl)(methylimino)methyl]cyclopentanol,

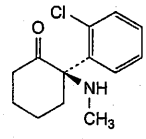


and enantiomer

B. (2*RS*)-2-(2-chlorophenyl)-2-hydroxycyclohexanone,



C. (2-chlorophenyl)(1-hydroxycyclopentyl)methanone,

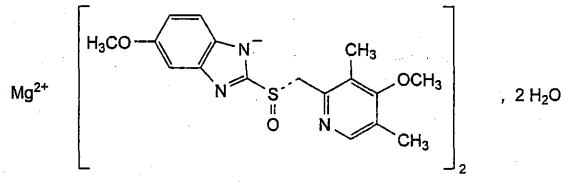


D. (2*R*)-2-(2-chlorophenyl)-2-(methylamino)cyclohexanone ((*R*)-ketamine).

Ph Eur

Esomeprazole Magnesium Dihydrate

(Ph. Eur. monograph 2787)



$C_{34}H_{36}MgN_6O_6S_2,2H_2O$ 749.2 *217087-10-0*

Action and use
Proton pump inhibitor; treatment of peptic ulcer disease.

Ph Eur

DEFINITION

Magnesium bis[5-methoxy-2-[(*S*)-[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1*H*-benzimidazol-1-ide] dihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly coloured powder, slightly hygroscopic.

Solubility

Slightly soluble in water, soluble in methanol, practically insoluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *esomeprazole magnesium dihydrate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Enantiomeric purity (see Tests).

C. Ignite about 0.5 g of the substance to be examined according to the procedure for the sulfated ash test (*2.4.14*). Dissolve the residue in 10 mL of *water R*. 2 mL of this solution gives the reaction of magnesium (*2.3.1*).

D. Water (see Tests).

TESTS

Absorbance (*2.2.25*)

Maximum 0.20 at 440 nm.

Dissolve 0.500 g in *methanol R* and dilute to 25.0 mL with the same solvent. Filter through a membrane filter (nominal pore size 0.45 µm).

Enantiomeric purity

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Buffer solution pH 6.0 Mix 20 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R* and 70 mL of a 156.0 g/L solution of *sodium dihydrogen phosphate R*, then dilute to 1000 mL with *water R*. Dilute 250 mL of this solution to 1000 mL with *water R*.

Buffer solution pH 11.0 Mix 11 mL of a 95.0 g/L solution of *trisodium phosphate dodecahydrate R* and 22 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R*, then dilute to 1000 mL with *water R*.

Test solution Dissolve 40 mg of the substance to be examined in 5 mL of *methanol R* and dilute to 50.0 mL with buffer solution pH 11.0. Dilute 1.0 mL of this solution to 25.0 mL with buffer solution pH 11.0.

Reference solution Dissolve 2 mg of *omeprazole CRS* in buffer solution pH 11.0 and dilute to 50.0 mL with the same buffer solution. Dilute 1.0 mL of the solution to 10.0 mL with buffer solution pH 11.0.

Column:
- *size*: *l* = 0.1 m, Ø = 4.0 mm;
- *stationary phase*: *α$_1$-acid-glycoprotein silica gel for chiral separation R* (5 µm).

Mobile phase *acetonitrile R*, buffer solution pH 6.0 (13:87 *V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 302 nm.

Injection 20 µL.

Relative retention With reference to esomeprazole (retention time = about 5 min): impurity F = about 0.7.

System suitability Reference solution:
- *resolution*: minimum 3.0 between the peaks due to impurity F and esomeprazole.

Limit:
- *impurity F*: maximum 0.6 per cent; disregard any peak other than impurity F and esomeprazole.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Test solution Dissolve 14 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 1 mg of *omeprazole CRS* and 1 mg of *omeprazole impurity D CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 3 mg of *omeprazole for peak identification CRS* (containing impurity E) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.125 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 27 volumes of *acetonitrile R* and 73 volumes of a 1.4 g/L solution of *disodium hydrogen phosphate dodecahydrate R* previously adjusted to pH 7.6 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 40 µL.

Run time 4 times the retention time of esomeprazole.

Identification of impurities Use the chromatogram supplied with *omeprazole for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to esomeprazole (retention time = about 9 min): impurity E = about 0.4; impurity D = about 0.7.

System suitability Reference solution (a):
- *resolution*: minimum 3.0 between the peaks due to impurity D and omeprazole.

Limits:
- *impurities D, E*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Magnesium

3.30 per cent to 3.55 per cent (anhydrous substance).

Dissolve 0.400 g in 25 mL of *methanol R*, sonicate until dissolution is complete. Add 25 mL of *water R*, 10 mL of *concentrated ammonia R*, 20.000 mL of 0.05 M sodium

edetate and about 50 mg of *mordant black 11 triturate R*. Titrate the excess of sodium edetate with *0.05 M zinc sulfate* until the colour changes from full blue to violet. Carry out a blank titration.

1 mL of 0.05 M sodium edetate corresponds to 1.21525 mg of Mg.

Water (*2.5.12*)

4.5 per cent to 6.1 per cent, determined on 0.200 g.

ASSAY

Liquid chromatography (*2.2.29*).

Buffer solution pH 11.0 Mix 11 mL of a 95.0 g/L solution of *trisodium phosphate dodecahydrate R* and 22 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R*, then dilute to 100.0 mL with *water R*.

Test solution Dissolve 10.0 mg of the substance to be examined in about 10 mL of *methanol R*, add 10 mL of buffer solution pH 11.0 and dilute to 200.0 mL with *water R*.

Reference solution Dissolve 10.0 mg of *omeprazole CRS* in about 10 mL of *methanol R*, add 10 mL of buffer solution pH 11.0 and dilute to 200.0 mL with *water R*.

Column:

— *size*: *l* = 0.125 m, Ø = 4 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 1.4 g/L solution of *disodium hydrogen phosphate dodecahydrate R* previously adjusted to pH 7.6 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 1.5 times the retention time of esomeprazole.

Retention time Esomeprazole = about 4 min.

Calculate the percentage content of $C_{34}H_{36}MgN_6O_6S_2$ taking into account the assigned content of *omeprazole CRS*.

1 g of omeprazole is equivalent to 1.032 g of esomeprazole magnesium.

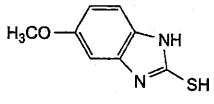
STORAGE

In an airtight container, protected from light.

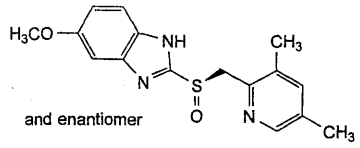
IMPURITIES

Specified impurities *D, E, F.*

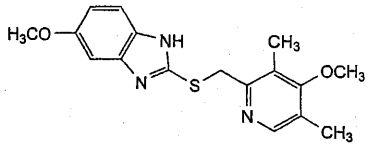
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



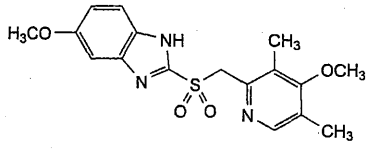
A. 5-methoxy-1*H*-benzimidazole-2-thiol,



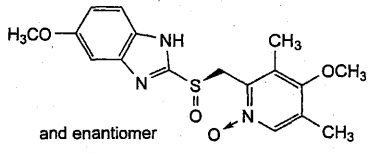
B. 2-[(*RS*)-[(3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-5-methoxy-1*H*-benzimidazole,



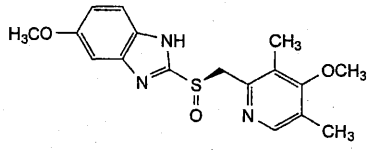
C. 5-methoxy-2-[[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfanyl]-1*H*-benzimidazole (ufiprazole),



D. 5-methoxy-2-[[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfonyl]-1*H*-benzimidazole (omeprazole sulfone),



E. 4-methoxy-2-[[(*RS*)-(5-methoxy-1*H*-benzimidazol-2-yl) sulfinyl]methyl]-3,5-dimethylpyridine 1-oxide,

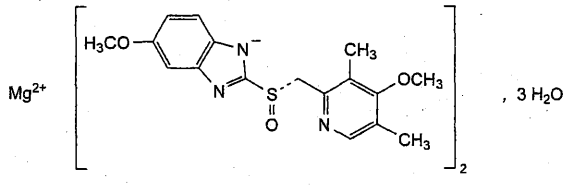


F. 5-methoxy-2-[(*R*)-[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfinyl]-1*H*-benzimidazole ((*R*)-omeprazole).

Ph Eur

Esomeprazole Magnesium Trihydrate

(Ph. Eur. monograph 2372)



$C_{34}H_{36}MgN_6O_6S_2,3H_2O$ 767.2 *217087-09-7*

Action and use

Proton pump inhibitor; treatment of peptic ulcer disease.

Ph Eur

DEFINITION

Magnesium bis[5-methoxy-2-[(*S*)-[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1*H*-benzimidazol-1-ide] trihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly coloured powder, slightly hygroscopic.

Solubility

Slightly soluble in water, soluble in methanol, practically insoluble in heptane.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *esomeprazole magnesium trihydrate CRS.*

B. Enantiomeric purity (see Tests).

C. Ignite about 0.5 g of the substance to be examined according to the procedure for the sulfated ash test (*2.4.14*). Dissolve the residue in 10 mL of *water R*. 2 mL of this solution gives the reaction of magnesium (*2.3.1*).

D. Water (see Tests).

TESTS

Absorbance (*2.2.25*)

Maximum 0.20 at 440 nm.

Dissolve 0.500 g in *methanol R* and dilute to 25.0 mL with the same solvent. Filter through a membrane filter (nominal pore size 0.45 µm).

Enantiomeric purity

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Buffer solution pH 6.0 Mix 20 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R* and 70 mL of a 156.0 g/L solution of *sodium dihydrogen phosphate R*, then dilute to 1000 mL with *water R*. Dilute 250 mL of this solution to 1000 mL with *water R*.

Buffer solution pH 11.0 Mix 11 mL of a 95.0 g/L solution of *trisodium phosphate dodecahydrate R* and 22 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R*, then dilute to 1000 mL with *water R*.

Test solution Dissolve 40 mg of the substance to be examined in 5 mL of *methanol R* and dilute to 50.0 mL with buffer solution pH 11.0. Dilute 1.0 mL of this solution to 25.0 mL with buffer solution pH 11.0.

Reference solution Dissolve 2 mg of *omeprazole CRS* in buffer solution pH 11.0 and dilute to 50.0 mL with the same buffer solution. Dilute 1.0 mL of the solution to 10.0 mL with buffer solution pH 11.0.

Column:
- *size*: *l* = 0.1 m, Ø = 4.0 mm;
- *stationary phase*: *α$_1$-acid-glycoprotein silica gel for chiral separation R* (5 µm).

Mobile phase *acetonitrile R*, buffer solution pH 6.0 (13:87 *V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 302 nm.

Injection 20 µL.

Relative retention With reference to esomeprazole (retention time = about 5 min): impurity F = about 0.7.

System suitability Reference solution:
- *resolution*: minimum 3.0 between the peaks due to impurity F and esomeprazole.

Limit:
- *impurity F*: maximum 0.2 per cent; disregard any peak other than impurity F and esomeprazole.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Test solution Dissolve 14 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 1 mg of *omeprazole CRS* and 1 mg of *omeprazole impurity D CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 3 mg of *omeprazole for peak identification CRS* (containing impurity E) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.125 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 27 volumes of *acetonitrile R* and 73 volumes of a 1.4 g/L solution of *disodium hydrogen phosphate dodecahydrate R* previously adjusted to pH 7.6 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 40 µL.

Run time 4 times the retention time of esomeprazole.

Identification of impurities Use the chromatogram supplied with *omeprazole for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to esomeprazole (retention time = about 9 min): impurity E = about 0.4; impurity D = about 0.7.

System suitability Reference solution (a):
- *resolution*: minimum 3.0 between the peaks due to impurity D and omeprazole.

Limits:
- *impurities D, E*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Magnesium

3.30 per cent to 3.55 per cent (anhydrous substance).

Dissolve 0.400 g in 25 mL of *methanol R*, sonicate until dissolution is complete. Add 25 mL of *water R*, 10 mL of *concentrated ammonia R*, 20.000 mL of 0.05 M sodium edetate and about 50 mg of *mordant black 11 triturate R*. Titrate the excess of sodium edetate with *0.05 M zinc sulfate* until the colour changes from full blue to violet. Carry out a blank titration.

1 mL of 0.05 M sodium edetate corresponds to 1.21525 mg of Mg.

Water (*2.5.12*)
6.2 per cent to 8.0 per cent, determined on 0.200 g.

ASSAY
Liquid chromatography (*2.2.29*).

Buffer solution pH 11.0 Mix 11 mL of a 95.0 g/L solution of *trisodium phosphate dodecahydrate R* and 22 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R*, then dilute to 100.0 mL with *water R*.

Test solution Dissolve 10.0 mg of the substance to be examined in about 10 mL of *methanol R*, add 10 mL of buffer solution pH 11.0 and dilute to 200.0 mL with *water R*.

Reference solution Dissolve 10.0 mg of *omeprazole CRS* in about 10 mL of *methanol R*, add 10 mL of buffer solution pH 11.0 and dilute to 200.0 mL with *water R*.

Column:
— *size*: *l* = 0.125 m, Ø = 4 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 1.4 g/L solution of *disodium hydrogen phosphate dodecahydrate R* previously adjusted to pH 7.6 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 1.5 times the retention time of esomeprazole.

Retention time Esomeprazole = about 4 min.

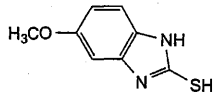
Calculate the percentage content of $C_{34}H_{36}MgN_6O_6S_2$ taking into account the assigned content of *omeprazole CRS*.

1 g of omeprazole is equivalent to 1.032 g of esomeprazole magnesium.

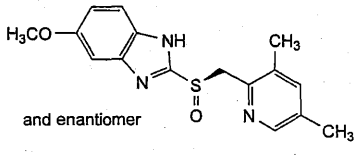
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *D, E, F.*

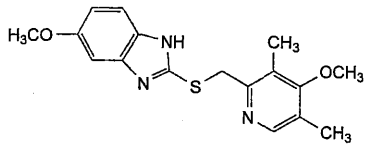
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



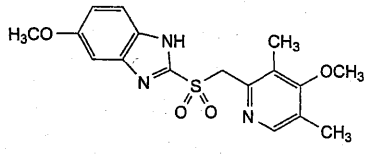
A. 5-methoxy-1*H*-benzimidazole-2-thiol,



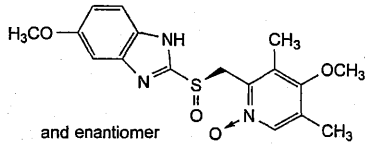
B. 2-[(*RS*)-[(3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-5-methoxy-1*H*-benzimidazole,



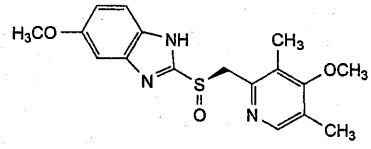
C. 5-methoxy-2-[[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfanyl]-1*H*-benzimidazole (ufiprazole),



D. 5-methoxy-2-[[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfonyl]-1*H*-benzimidazole (omeprazole sulfone),



E. 4-methoxy-2-[[(*RS*)-(5-methoxy-1*H*-benzimidazol-2-yl) sulfinyl]methyl]-3,5-dimethylpyridine 1-oxide,

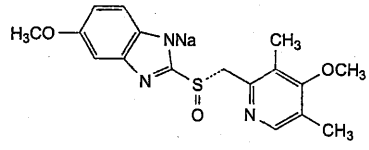


F. 5-methoxy-2-[(*R*)-[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfinyl]-1*H*-benzimidazole ((*R*)-omeprazole).

Ph Eur

Esomeprazole Sodium

(Ph. Eur. monograph 2923)



$C_{17}H_{18}N_3NaO_3S$ 367.4 *161796-78-7*

Action and use
Proton pump inhibitor; treatment of peptic ulcer disease.

Ph Eur

DEFINITION
Sodium 5-methoxy-2-[(*S*)-[(4-methoxy-3,5-dimethylpyridin-2-yl)methanesulfinyl]-1*H*-benzimidazol-1-ide.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, amorphous or crystalline powder, slightly hygroscopic.

Solubility
Freely soluble in water and in ethanol (96 per cent), soluble in propylene glycol, very slightly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *esomeprazole sodium CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *ethanol (96 per cent) R*, evaporate to dryness and record new spectra using the residues.

B. Enantiomeric purity (see Tests).

C. Ignite 1 g and cool. Add 1 mL of *water R* to the residue and neutralise with *hydrochloric acid R*. Filter and dilute the filtrate to 4 mL with *water R*. 0.1 mL of the solution gives reaction (b) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)
10.3 to 11.3 for solution S.

Enantiomeric purity
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*

Buffer solution pH 6.0 Mix 20 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R* and 70 mL of a 156.0 g/L solution of *sodium dihydrogen phosphate R*, then dilute to 1000.0 mL with *water for chromatography R*. Dilute 250 mL of this solution to 1000.0 mL with *water for chromatography R*.

Buffer solution pH 11.0 Mix 11 mL of a 95.0 g/L solution of *trisodium phosphate dodecahydrate R* and 22 mL of a 179.1 g/L solution of *disodium hydrogen phosphate dodecahydrate R*, then dilute to 1000.0 mL with *water R*.

Test solution Dissolve 40 mg of the substance to be examined in buffer solution pH 11.0 and dilute to 50.0 mL with the same buffer solution. Dilute 1.0 mL of this solution to 25.0 mL with buffer solution pH 11.0.

Reference solution (a) Dissolve 2 mg of *omeprazole CRS* in buffer solution pH 11.0 and dilute to 50 mL with the same buffer solution. Dilute 1 mL of the solution to 10 mL with buffer solution pH 11.0.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with buffer solution pH 11.0. Dilute 1.0 mL of this solution to 20.0 mL with buffer solution pH 11.0.

Column:
— *size*: *l* = 0.10 m, Ø = 4.0 mm;
— *stationary phase*: *α_1-acid-glycoprotein silica gel for chiral separation R* (5 µm).

Mobile phase *acetonitrile R*, buffer solution pH 6.0 (15:85 *V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 302 nm.

Injection 20 µL.

Run time 1.5 times the retention time of esomeprazole.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity B.

Relative retention With reference to esomeprazole (retention time = about 4 min): impurity B = about 0.7.

System suitability Reference solution (a):
— *resolution*: minimum 2.8 between the peaks due to impurity B and esomeprazole.

Limit:
— *impurity B*: maximum 0.2 per cent;
— *reporting threshold*: 0.05 per cent (reference solution (b));
disregard any peak other than impurity B and esomeprazole.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Buffer solution pH 7.6 Dissolve 5.7 g of *disodium hydrogen phosphate dihydrate R* and 0.7 g of *sodium dihydrogen phosphate monohydrate R* in *water for chromatography R* and dilute to 1000.0 mL with the same solvent.

Test solution Dissolve 10 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 1 mg of *omeprazole CRS* and 1 mg of *omeprazole impurity D CRS* in mobile phase A and dilute to 50 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (3.5 µm).

Mobile phase:
— *mobile phase A*: mix 100 mL of *acetonitrile R* and 100 mL of buffer solution pH 7.6 and dilute to 1000.0 mL with *water for chromatography R*;
— *mobile phase B*: mix 10 mL of buffer solution pH 7.6 and 800 mL of *acetonitrile R* and dilute to 1000.0 mL with *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 12	100 → 80	0 → 20
12 - 32	80 → 0	20 → 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 302 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to esomeprazole (retention time = about 16 min): impurity D = about 0.9.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity D and omeprazole.

Calculation of percentage contents:
— for each impurity, use the concentration of esomeprazole sodium in reference solution (b).

Limits:
— *impurity D*: maximum 0.2 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 0.300 g.

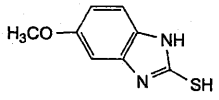
ASSAY
Dissolve 0.300 g in 50 mL of *carbon dioxide-free water R*. Titrate with *0.1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M hydrochloric acid* is equivalent to 36.74 mg of $C_{17}H_{18}N_3NaO_3S$.

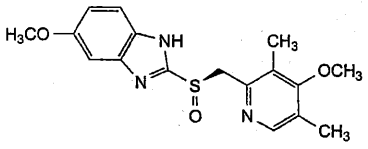
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *B, D.*

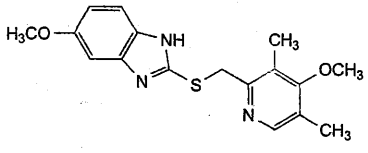
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, E, F, G.*



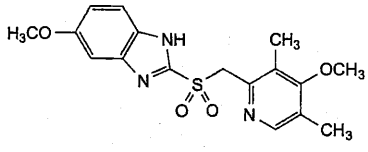
A. 5-methoxy-1*H*-benzimidazole-2-thiol,



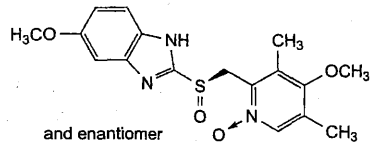
B. 5-methoxy-2-[(*R*)-(4-methoxy-3,5-dimethylpyridin-2-yl) methanesulfinyl]-1*H*-benzimidazole ((*R*)-omeprazole),



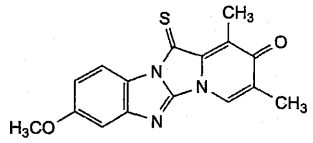
C. 5-methoxy-2-[[(4-methoxy-3,5-dimethylpyridin-2-yl) methyl]sulfanyl]-1*H*-benzimidazole (ufiprazole),



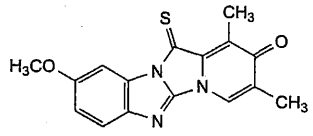
D. 5-methoxy-2-[(4-methoxy-3,5-dimethylpyridin-2-yl) methanesulfonyl]-1*H*-benzimidazole (omeprazole sulfone),



E. 4-methoxy-2-[[(*RS*)-5-methoxy-1*H*-benzimidazole-2-sulfinyl]methyl]-3,5-dimethylpyridine 1-oxide,



F. 8-methoxy-1,3-dimethyl-12-sulfanylidenepyrido[1′,2′:3,4] imidazo[1,2-a]benzimidazol-2(12*H*)-one,

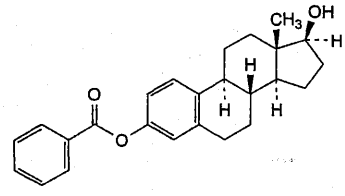


G. 9-methoxy-1,3-dimethyl-12-sulfanylidenepyrido[1′,2′:3,4] imidazo[1,2-*a*]benzimidazol-2(12*H*)-one.

Ph Eur

Estradiol Benzoate

(Ph. Eur. monograph 0139)



$C_{25}H_{28}O_3$ 376.5 *50-50-0*

Action and use
Estrogen.

Ph Eur

DEFINITION
17β-Hydroxyestra-1,3,5(10)-trien-3-yl benzoate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
Almost white, crystalline powder or colourless crystals.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in acetone, slightly soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *estradiol benzoate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference

substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)

+ 55.0 to + 59.0 (dried substance).

Dissolve 0.250 g in *acetone R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20 mg of the substance to be examined in *acetonitrile R1* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *estradiol benzoate for system suitability CRS* (containing impurities A, B, C, E and G) in *acetonitrile R1* and dilute to 2.5 mL with the same solvent.

Reference solution (b) Dilute 0.5 mL of the test solution to 100.0 mL with *acetonitrile R1*.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: *water R, acetonitrile R1* (40:60 *V/V*);

— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	100	0
20 - 21	100 → 10	0 → 90
21 - 31	10	90

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *estradiol benzoate for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, E and G.

Relative retention With reference to estradiol benzoate (retention time = about 19 min): impurity A = about 0.3; impurity E = about 1.1; impurity B = about 1.2; impurity G = about 1.3; impurity C = about 1.5.

System suitability Reference solution (a):

— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to estradiol benzoate.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 3.3; impurity C = 0.7;

— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurities B, E, G*: for each impurity, not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurity A*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

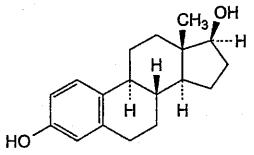
Dissolve 25.0 mg in *anhydrous ethanol R* and dilute to 250.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with *anhydrous ethanol R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 231 nm.

Calculate the content of $C_{25}H_{28}O_3$ taking the specific absorbance to be 500.

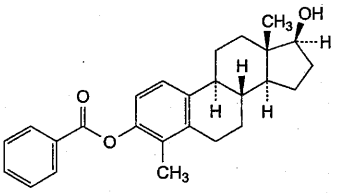
IMPURITIES

Specified impurities *A, B, C, E, G.*

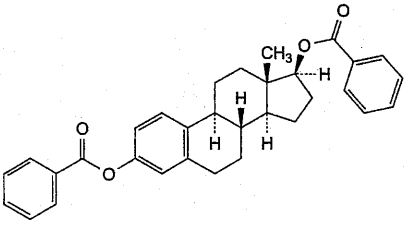
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, F, H.*



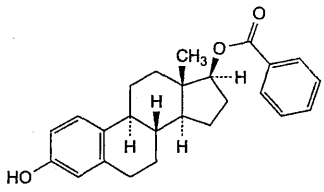
A. estra-1,3,5(10)-triene-3,17β-diol (estradiol),



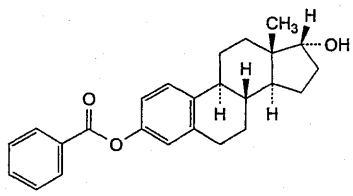
B. 17β-hydroxy-4-methylestra-1,3,5(10)-trien-3-yl benzoate,



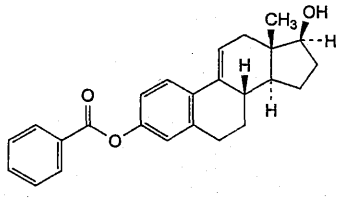
C. estra-1,3,5(10)-triene-3,17β-diyl dibenzoate,



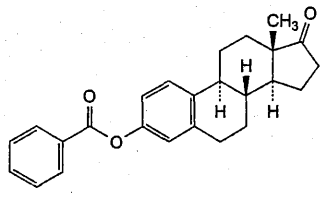
D. 3-hydroxyestra-1,3,5(10)-trien-17β-yl benzoate,



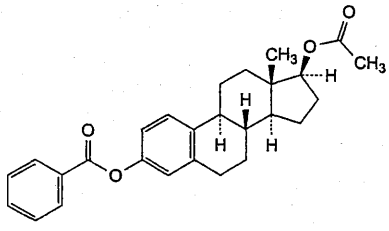
E. 17α-hydroxyestra-1,3,5(10)-trien-3-yl benzoate,



F. 17β-hydroxyestra-1,3,5(10),9(11)-tetraen-3-yl benzoate.



G. 17-oxoestra-1,3,5(10)-trien-3-yl benzoate (estrone benzoate),

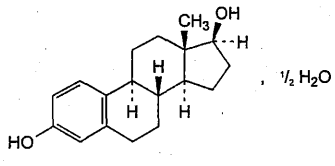


H. estra-1,3,5(10)-triene-3,17β-diyl 17-acetate 3-benzoate,

Ph Eur

Estradiol Hemihydrate

(Ph. Eur. monograph 0821)



$C_{18}H_{24}O_2,½H_2O$ 281.4

Action and use
Estrogen.

Preparations
Estradiol Transdermal Patches
Estradiol and Norethisterone Tablets
Estradiol and Norethisterone Acetate Tablets
Estradiol Vaginal Tablets

Ph Eur

DEFINITION
Estra-1,3,5(10)-triene-3,17β-diol hemihydrate.

Content
97.0 per cent to 103.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Practically insoluble in water, soluble in acetone, sparingly soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION
First identification: B.

Second identification: A, C, D, E.

A. Melting point (*2.2.14*): 175 °C to 180 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *estradiol hemihydrate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *methanol R* and dilute to 50 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *estradiol hemihydrate CRS* in *methanol R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of *ethinylestradiol CRS* in reference solution (a) and dilute to 25 mL with reference solution (a).

Plate *TLC silica gel plate R.*

Mobile phase *ethanol (96 per cent) R, toluene R* (20:80 *V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air until the solvent has evaporated.

Detection Heat at 110 °C for 10 min. Spray the hot plate with *alcoholic solution of sulfuric acid R.* Heat again at 110 °C for 10 min. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

System suitability The chromatogram obtained with reference solution (b) shows 2 spots which may however not be completely separated.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To about 1 mg add 0.5 mL of freshly prepared *sulfomolybdic reagent R2.* A blue colour develops which in ultraviolet light at 365 nm has an intense green fluorescence. Add 1 mL of *sulfuric acid R* and 9 mL of *water R.* The colour becomes pink with a yellowish fluorescence.

E. Water (see Tests).

TESTS

Specific optical rotation (*2.2.7*)
+ 76.0 to + 83.0 (anhydrous substance).

Dissolve 0.250 g in *ethanol (96 per cent) R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *acetonitrile R* and dilute to 25.0 mL with *methanol R2*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Mix equal volumes of a 1 mg/mL solution of the substance to be examined in *methanol R2* and of a 1 mg/mL solution of *2,3-dichloro-5,6-dicyanobenzoquinone R* in *methanol R2*. Allow to stand for 30 min before injection.

Reference solution (c) Dissolve 5 mg of *estradiol for peak identification CRS* (containing impurities A, B and C) in 2 mL of *acetonitrile R* and dilute to 5.0 mL with *methanol R2*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase To 400 mL of *acetonitrile R* add 50 mL of *methanol R2* and 400 mL of *water for chromatography R*; allow to stand for 10 min, dilute to 1000 mL with *water for chromatography R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time Twice the retention time of estradiol.

Identification of impurities Use the chromatogram supplied with *estradiol for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to estradiol (retention time = about 13 min): impurity D = about 0.9; impurity B = about 1.1; impurity A = about 1.4; impurity C = about 1.9.

System suitability Reference solution (c):
— *resolution*: minimum 2.5 between the peaks due to estradiol and impurity B.

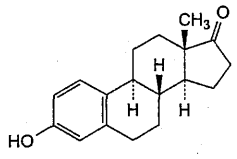
Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 0.4;
— *impurities A, B, C, D*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
2.9 per cent to 3.5 per cent, determined on 0.500 g.

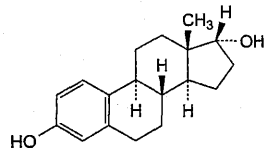
ASSAY
Dissolve 20.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *0.1 M sodium hydroxide*. Allow to cool to room temperature. Measure the absorbance (*2.2.25*) of the solution at the maximum at 238 nm.

Calculate the content of $C_{18}H_{24}O_2$ taking the specific absorbance to be 335.

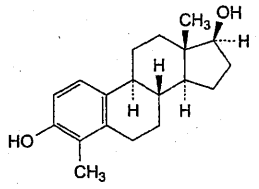
IMPURITIES
Specified impurities *A, B, C, D.*



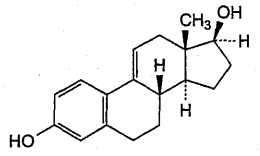
A. 3-hydroxyestra-1,3,5(10)-trien-17-one (estrone),



B. estra-1,3,5(10)-triene-3,17α-diol (17α-estradiol),



C. 4-methylestra-1,3,5(10)-triene-3,17β-diol,

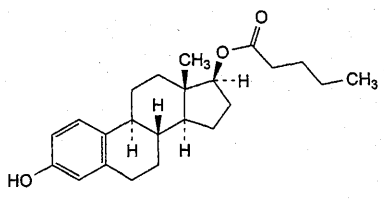


D. estra-1,3,5(10),9(11)-tetraene-3,17β-diol.

Ph Eur

Estradiol Valerate

(Ph. Eur. monograph 1614)



$C_{23}H_{32}O_3$ 356.5 *979-32-8*

Action and use
Estrogen.

Ph Eur

DEFINITION

3-Hydroxyestra-1,3,5(10)-trien-17β-yl pentanoate.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *estradiol valerate CRS.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

TESTS

Solution S

Dissolve 0.500 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 41 to + 47 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in *acetonitrile R1* and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 20.0 mL with *acetonitrile R1.*

Reference solution (a) Dissolve 3 mg of *estradiol valerate for system suitability CRS* (containing impurities A, C, D and E) in *acetonitrile R1* and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *acetonitrile R1.* Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R1.*

Reference solution (c) Dissolve 50.0 mg of *estradiol valerate CRS* in *acetonitrile R1* and dilute to 10.0 mL with the same solvent. Dilute 5.0 mL of the solution to 20.0 mL with *acetonitrile R1.*

Column:

— *size: l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase: end-capped solid core alkylsilyl silica gel for chromatography R* (2.6 µm).

Mobile phase:

— *mobile phase A: water R;*
— *mobile phase B: acetonitrile R1;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	48	52
1 - 10	48 → 35	52 → 65
10 - 17.5	35 → 0	65 → 100
17.5 - 26	0	100

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 5 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *estradiol valerate for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, C, D and E.

Relative retention With reference to estradiol valerate (retention time = about 9 min): impurity A = about 0.1; impurity C = about 0.9; impurity D = about 1.3; impurity E = about 1.7.

System suitability Reference solution (a):

— *resolution*: minimum 2.5 between the peaks due to impurity C and estradiol valerate.

Calculation of percentage contents:

— *correction factor*: multiply the peak area of impurity C by 0.5;
— for each impurity, use the concentration of estradiol valerate in reference solution (b).

Limits:

— *impurity D*: maximum 0.4 per cent;
— *impurities A, C*: for each impurity, maximum 0.2 per cent;
— *impurity E*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{23}H_{32}O_3$ taking into account the assigned content of *estradiol valerate CRS.*

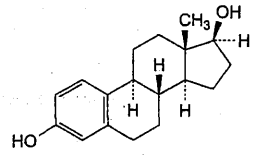
STORAGE

Protected from light.

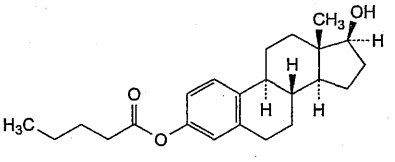
IMPURITIES

Specified impurities *A, C, D, E.*

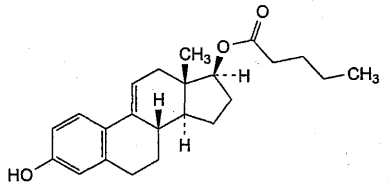
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, F, G, H, I, J.*



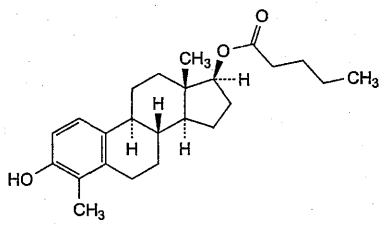
A. estra-1,3,5(10)-triene-3,17β-diol (estradiol),



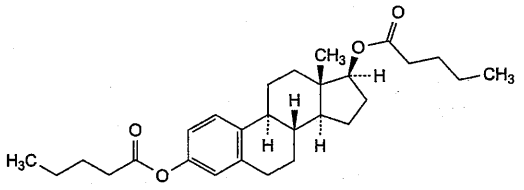
B. 17β-hydroxyestra-1,3,5(10)-trien-3-yl pentanoate,



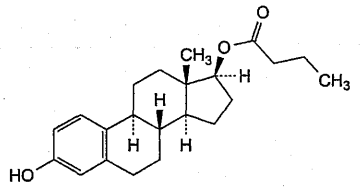
C. 3-hydroxyestra-1,3,5(10),9(11)-tetraen-17β-yl pentanoate,



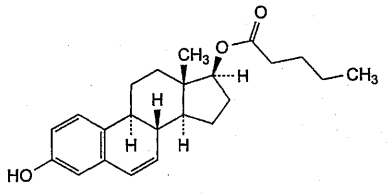
D. 3-hydroxy-4-methylestra-1,3,5(10)-trien-17β-yl pentanoate,



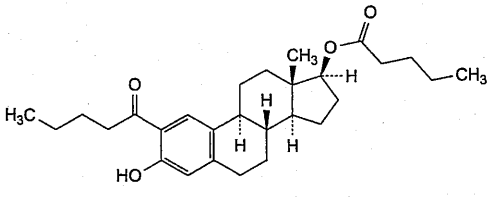
E. estra-1,3,5(10)-trien-3,17β-diyl dipentanoate,



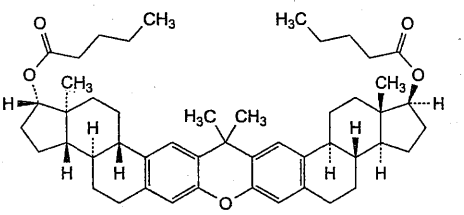
F. 3-hydroxyestra-1,3,5(10)-trien-17β-yl butanoate,



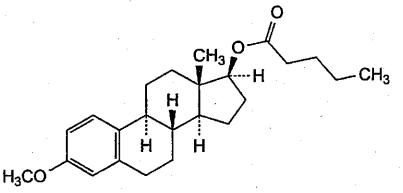
G. 3-hydroxyestra-1,3,5(10),6-tetraen-17β-yl pentanoate,



H. 3-hydroxy-2-pentanoylestra-1,3,5(10)-trien-17β-yl pentanoate,



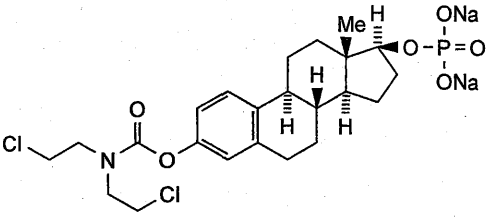
I. (1*S*,3a*S*,3b*R*,10a*R*,10b*S*,13*S*,13a*S*,15a*S*,18b*S*,20a*S*)-13a,17,17,20a-tetramethyl-2,3,3a,3b,4,5,9,10,10a,10b,11,12,13,13a,14,15,15a,17,18b,19,20,20a-docosahydro-1*H*-bis(cyclopenta[5,6]naphtho)[1,2-*b*:2′,1′-*i*]xanthene-1,13-diyl dipentanoate,



J. 3-methoxyestra-1,3,5(10)-trien-17β-yl pentanoate.

Ph Eur

Estramustine Sodium Phosphate



$C_{23}H_{30}Cl_2NNa_2O_6P$ 564.4 *52205-73-9*

Action and use
Cytotoxic alkylating agent.

Preparation
Estramustine Phosphate Capsules

DEFINITION
Estramustine Sodium Phosphate is disodium 3-[bis(2-chloroethyl)carbamoyloxy]estra-1,3,5(10)-trien-17β-yl orthophosphate. It contains not less than 97.0% and not more than 103.0% of $C_{23}H_{30}Cl_2NNa_2O_6P$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A white or almost white powder.

Freely soluble in *water* and in *methanol*; very slightly soluble in *absolute ethanol*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.05% w/v solution exhibits maxima at 267 nm and 275 nm. The *absorbance* at 267 nm is about 0.76 and at 275 nm is about 0.71.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of estramustine sodium phosphate *(RS 128)*. In preparing the potassium bromide disc precautions should be taken to exclude moisture and

avoid excessive grinding; if necessary heat the prepared disc at 90° for 2 minutes.

C. A 1% w/v solution yields the reactions characteristic of *sodium salts*, Appendix VI.

TESTS

Alkalinity

pH of a 0.5% w/v solution, 8.5 to 10.0, Appendix V L.

Clarity and colour of solution

A 5.0% w/v solution is not more opalescent than *reference suspension II*, Appendix IV A, and is *colourless*, Appendix IV B, Method I.

Specific optical rotation

In a 2% w/v solution, +11 to +13, Appendix V F, calculated with reference to the anhydrous substance.

Ionisable chlorine

Dissolve 0.10 g in 10 mL of *water*, add carefully, with mixing, 0.1 mL of a mixture of 10 volumes of *silver nitrate solution* and 1 volume of *nitric acid* and examine immediately. Any opalescence produced is not more intense than that obtained by treating a solution containing 13.4 µg of *sodium chloride* in 10 mL in the same manner (0.1%).

Estradiol 17β-phosphate

Dissolve 50 mg in 5 mL of 0.2M *sodium hydroxide*, add sufficient *ethanol (96%)* to produce 10 mL, mix and immediately measure the *absorbance* at the maxima at 300 nm and 350 nm, Appendix II B. The difference between the two absorbances is not more than 0.34 (1.0%).

Inorganic phosphate

Dissolve 25 mg in 10 mL of *water*, add 4 mL of 1M *sulfuric acid*, 1 mL of a 10% w/v solution of *ammonium molybdate* and 2 mL of *methylaminophenol-sulfite reagent* and allow to stand for 15 minutes. Add sufficient *water* to produce 25 mL, allow to stand for 15 minutes and filter. The *absorbance* of the filtrate at 730 nm, Appendix II B, is not greater than the *absorbance* at 730 nm of a solution obtained by repeating the operation using 10 mL of a 0.00180% w/v solution of *potassium dihydrogen orthophosphate* and beginning at the words 'add 4 mL of 1M *sulfuric acid*...'.

Volatile matter

Carry out the method for *gas chromatography*, Appendix III B using the following solutions in *water*.

(1) 4.0% w/v of the substance being examined and 0.020% v/v of the internal standard.

(2) 4.0% w/v of the substance being examined.

(3) 0.0040% v/v of *pyridine*, 0.020% v/v of *absolute ethanol* and 0.020% v/v of *butan-1-ol* (internal standard).

CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with *acid-washed, silanised diatomaceous support* (80 to 100 mesh) coated with 20% w/w of polyethylene glycol (Carbowax 20M is suitable).

(b) Use *nitrogen* as the carrier gas at 30 mL per minute.

(c) Use isothermal conditions maintained at 120°.

(d) Use an inlet temperature of 200°.

(e) Use a flame ionisation detector at a temperature of 300°.

(f) Inject 1 µL of each solution.

(g) The peaks elute in the order: ethanol, butan-1-ol, pyridine.

LIMITS

In the chromatogram obtained with solution (1):

the area of the peak due to pyridine is not greater than the area of any corresponding peak in the chromatogram obtained with solution (3) (0.1%);

the sum of the areas of any peaks with a retention time less than that of the peak due to the internal standard is not greater than the area of the peak due to ethanol in the chromatogram obtained with solution (3) (0.5%).

Related substances

Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in a mixture of 1 volume of *triethylamine* and 49 volumes of *methanol*.

(1) 4.0% w/v of the substance being examined.

(2) 0.020% w/v of the substance being examined.

(3) 0.080% w/v of *17β,17′β-bis{3-[bis(2-chloroethyl) carbamoyloxy]estra1,3,5(10)-trienyl} pyrophosphate BPCRS*.

(4) 0.040% of *estramustine BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating *silica gel*.

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, spray with *methanolic sulfuric acid (20%)* and heat at 110° for 10 minutes.

MOBILE PHASE

Equal volumes of *butan-2-one*, *propan-2-ol* and *triethylamine hydrogen carbonate solution*.

LIMITS

In the chromatogram obtained with solution (1):

any *secondary spot* due to 17β,17′β-bis{3-[bis(2-chloroethyl) carbamoyloxy]estra1,3,5(10)-trienyl} pyrophosphate is not more intense than the corresponding spot in the chromatogram obtained with solution (3) (2%);

any *secondary spot* due to estramustine is not more intense than the corresponding spot in the chromatogram obtained with solution (4) (1%);

Any other *secondary spot* is not more intense than the spot in the chromatogram obtained with solution (2) (0.5%).

Water

Not more than 5.0% w/w, Appendix IX C. Use 0.2 g.

ASSAY

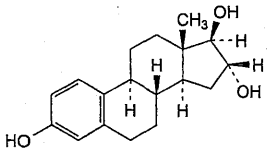
To 0.5 g add 40 mL of 1M *sodium hydroxide* and boil under a reflux condenser for 60 minutes. Cool and transfer the mixture to a 250 mL graduated flask with the aid of *water*. Add 100 mL of *0.1M silver nitrate VS* and 10 mL of *nitric acid*, dilute to 250 mL with *water* and mix. Filter and titrate the excess of silver nitrate in 50 mL of the filtrate with *0.1M ammonium thiocyanate VS* using 3 mL of *ammonium iron(III) sulfate solution R2* as indicator. Each mL of *0.1M silver nitrate VS* is equivalent to 28.22 mg of $C_{23}H_{30}Cl_2NNa_2O_6P$.

STORAGE

Estramustine Sodium Phosphate should be protected from light.

Estriol

(Ph. Eur. monograph 1203)



$C_{18}H_{24}O_3$ 288.4 *50-27-1*

Action and use
Estrogen.

Preparation
Estriol Cream

Ph Eur

DEFINITION

Estra-1,3,5(10)-triene-3,16α,17β-triol.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *estriol CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

TESTS

Specific optical rotation (*2.2.7*)
+ 60 to + 65 (dried substance).

Dissolve 80 mg in *anhydrous ethanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, water R* (50:50 *V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in 25 mL of *methanol R* and dilute to 50.0 mL with *water R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *estriol for system suitability CRS* (containing impurities A, D, E and F) in 5 mL of *methanol R* and dilute to 10.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of *estriol CRS* in 25 mL of *methanol R* and dilute to 50.0 mL with *water R*. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.10 m, Ø = 2.1 mm;
— *stationary phase*: end-capped, charged surface, ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R (1.7 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: *methanol R1, water for chromatography R* (28:72 *V/V*);
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 9	100	0
9 - 23	100 → 57	0 → 43
23 - 28	57	43

Flow rate 0.4 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *estriol for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, D, E and F.

Relative retention With reference to estriol (retention time = about 11 min): impurity A = about 0.95; impurity F = about 1.45; impurity E = about 1.5; impurity D = about 1.8.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 5.0, where H_P = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to estriol.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity A by 0.5;
— for each impurity, use the concentration of estriol in reference solution (b).

Limits:
— *impurity F*: maximum 0.5 per cent;
— *impurity E*: maximum 0.3 per cent;
— *impurities A, D*: for each impurity, maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90	10
5 - 5.5	90 → 30	10 → 70
5.5 - 7.5	30	70

Injection 5 µL of test solution (b) and reference solutions (a) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to estriol (retention time = about 4 min): impurity A = about 0.9.

System suitability Reference solution (a):

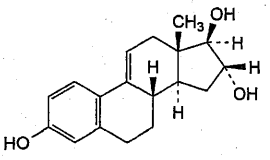
— *peak-to-valley ratio*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to estriol.

Calculate the percentage content of $C_{18}H_{24}O_3$ taking into account the assigned content of *estriol CRS*.

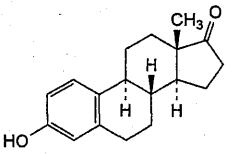
IMPURITIES

Specified impurities *A, D, E, F.*

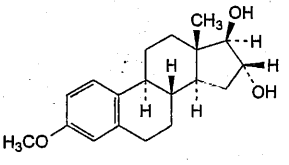
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, G, H, I, J, K.*



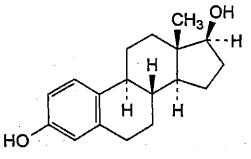
A. estra-1,3,5(10),9(11)-tetraene-3,16α,17β-triol (9,11-didehydroestriol),



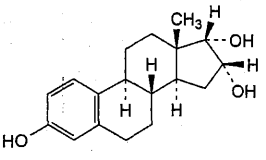
B. 3-hydroxyestra-1,3,5(10)-trien-17-one (estrone),



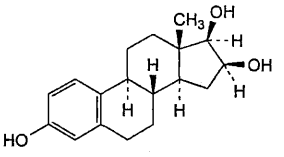
C. 3-methoxyestra-1,3,5(10)-triene-16α,17β-diol (estriol 3-methyl ether),



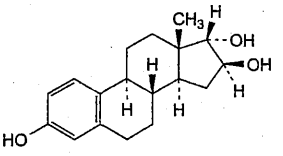
D. estra-1,3,5(10)-triene-3,17β-diol (estradiol),



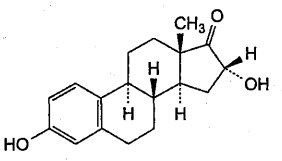
E. estra-1,3,5(10)-triene-3,16α,17α-triol (17-epi-estriol),



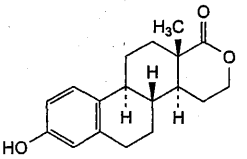
F. estra-1,3,5(10)-triene-3,16β,17β-triol (16-epi-estriol),



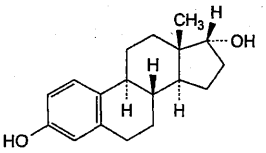
G. estra-1,3,5(10)-triene-3,16β,17α-triol (16,17-epi-estriol),



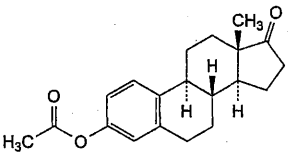
H. 3,16α-dihydroxyestra-1,3,5(10)-trien-17-one,



I. 3-hydroxy-17-oxa-17a-homoestra-1,3,5(10)-trien-17a-one,



J. estra-1,3,5(10)-triene-3,17α-diol (17-epi-estradiol),



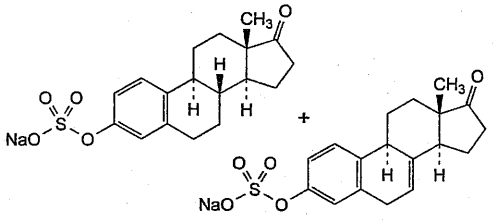
K. 17-oxoestra-1,3,5(10)-trien-3-yl acetate (estrone acetate).

Ph Eur

Conjugated Estrogens

Conjugated Oestrogens

(Ph. Eur. monograph 1512)



$C_{18}H_{21}O_5NaS + C_{18}H_{19}O_5NaS$ 372.4 + 370.4

Action and use
Estrogen.

Ph Eur

DEFINITION

Mixture of various conjugated forms of estrogens obtained from the urine of pregnant mares or by synthesis, dispersed in a suitable powdered diluent.

The 2 principal components are 17-oxoestra-1,3,5(10)-trien-3-yl sodium sulfate (sodium estrone sulfate) and 17-oxoestra-1,3,5(10),7-tetraen-3-yl sodium sulfate (sodium equilin sulfate). Concomitants are sodium 17α-estradiol sulfate, sodium 17α-dihydroequilin sulfate and sodium 17β-dihydroequilin sulfate.

Content (percentages related to the labelled content):
— *sodium estrone sulfate*: 52.5 per cent to 61.5 per cent;
— *sodium equilin sulfate*: 22.5 per cent to 30.5 per cent;
— *sodium 17α-estradiol sulfate*: 2.5 per cent to 9.5 per cent;
— *sodium 17α-dihydroequilin sulfate*: 13.5 per cent to 19.5 per cent;
— *sodium 17β-dihydroequilin sulfate*: 0.5 per cent to 4.0 per cent;
— *sum of sodium estrone sulfate and sodium equilin sulfate*: 79.5 per cent to 88.0 per cent.

CHARACTERS

Appearance
Almost white or brownish, amorphous powder.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks due to estrone and equilin in the chromatogram obtained with test solution (a) are similar in retention time and size to the 2 principal peaks in the chromatogram obtained with reference solution (a).

B. Examine the chromatogram obtained in the test for chromatographic profile.

Results The chromatogram obtained with test solution (b) exhibits additional peaks due to 17α-estradiol, 17α-dihydroequilin and 17β-dihydroequilin, at relative retentions with reference to 3-*O*-methylestrone (internal standard) of about 0.24, 0.30 and 0.35 respectively.

TESTS

Chromatographic profile
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 8 mg of *3-O-methylestrone R* in 10.0 mL of *anhydrous ethanol R*. Dilute 2.0 mL of this solution to 10.0 mL with *anhydrous ethanol R*.

Acetate buffer solution pH 5.2 Dissolve 10 g of *sodium acetate R* in 100 mL of *water R* and add 10 mL of *dilute acetic acid R*. Dilute to 500 mL with *water R* and adjust to pH 5.2 ± 0.1.

Test solution (a) Considering the labelled content, transfer an accurately weighed quantity corresponding to about 2 mg of conjugated estrogens to a 50 mL centrifuge tube containing 15 mL of the acetate buffer solution pH 5.2 and 1 g of *barium chloride R*. Cap the tube tightly and shake for 30 min. If necessary, adjust to pH 5.0 ± 0.5 with *acetic acid R* or a 120 g/L solution of *sodium acetate R*. Sonicate for 30 s, then shake for 30 min. Add a suitable sulfatase preparation equivalent to 2500 units and shake mechanically for 10 min in a water-bath at 50 ± 1 °C. Swirl the tube by hand, then shake mechanically for 10 min in the water-bath. Allow to cool. Add 15.0 mL of *ethylene chloride R* to the mixture, immediately cap the tube tightly and shake for 15 min. Centrifuge for 10 min or until the lower layer is clear. Draw out the organic layer to a screw-cap tube, add 5 g of *anhydrous sodium sulfate R* and shake. Allow the solution to stand until clear. Protect the solution from any loss due to evaporation. Transfer 3.0 mL of the clear solution to a suitable centrifuge tube fitted with a screw cap. Add 1.0 mL of the internal standard solution. Evaporate the mixture to dryness with the aid of a stream of *nitrogen R*, maintaining the temperature below 50 °C. To the dry residue add 15 µL of *anhydrous pyridine R* and 65 µL of *N,O-bis(trimethylsilyl)trifluoroacetamide R* containing 1 per cent of *chlorotrimethylsilane R*. Immediately cap the tube tightly, mix thoroughly and allow to stand for 15 min. Add 0.5 mL of *toluene R* and mix mechanically.

Test solution (b) Prepare as described in test solution (a), but do not add the sulfatase and use 6.0 mL of the upper layer instead of 3.0 mL. Prepare a blank in the same manner.

Reference solution (a) Dissolve separately 8 mg of *estrone CRS*, 7 mg of *equilin CRS* and 5 mg of *17α-dihydroequilin CRS* in 10.0 mL of *anhydrous ethanol R*. Dilute together 2.0 mL, 1.0 mL and 1.0 mL respectively of these solutions to 10.0 mL with *anhydrous ethanol R*. Transfer 1.0 mL of this solution and 1.0 mL of the internal standard solution to a centrifuge tube fitted with a screw cap. Evaporate the mixture to dryness with the aid of a stream of *nitrogen R*, maintaining the temperature below 50 °C. To the dry residue add 15 µL of *anhydrous pyridine R* and 65 µL of *N,O-bis(trimethylsilyl)trifluoroacetamide R* containing 1 per cent of *chlorotrimethylsilane R*. Immediately cap the tube tightly, mix and allow to stand for 15 min. Add 0.5 mL of *toluene R*.

Reference solution (b) Prepare as described in reference solution (a), but dilute tenfold with *anhydrous ethanol R* before adding the internal standard.

Column:
— *material*: fused silica;
— *size*: *l* = 15 m, Ø = 0.25 mm;
— *stationary phase*: *poly[(cyanopropyl)(methyl)][(phenyl)(methyl)]siloxane R* (film thickness 0.25 µm).

Carrier gas *hydrogen for chromatography R*.

Flow rate 2 mL/min.

Split ratio 1:20 to 1:30.

Temperature:
— *column*: 220 °C;
— *injection port and detector*: 260 °C.

Detection Flame ionisation.

Injection 1 µL.

Table 1512.-1

Relative retention (to 3-*O*-methylestrone)	Analyte	Quantified with reference to CRS	Present as
0.24	17α-estradiol	*17α-dihydroequilin CRS*	sodium sulfate
0.29	17β-estradiol	*estrone CRS*	sodium sulfate
0.30	17α-dihydroequilin	*17α-dihydroequilin CRS*	free steroid, sodium sulfate (assay)
0.35	17β-dihydroequilin	*17α-dihydroequilin CRS*	sodium sulfate
0.56	17α-dihydroequilenin	*estrone CRS*	sodium sulfate
0.64	17β-dihydroequilenin	*estrone CRS*	sodium sulfate
0.80	estrone	*estrone CRS*	free steroid, sodium sulfate (assay)
0.87	equilin	*equilin CRS*	free steroid, sodium sulfate (assay)
0.90	8,9-didehydroestrone	*estrone CRS*	sodium sulfate
1	3-*O*-methylestrone	(internal standard)	
1.3	equilenin	*estrone CRS*	sodium sulfate

Relative retention With reference to 3-*O*-methylestrone: 17α-dihydroequilin = about 0.30; estrone = about 0.80; equilin = about 0.87.

System suitability Reference solution (a):
— *resolution*: minimum 1.2 between the peaks due to estrone and equilin; if necessary, adjust the temperature and the flow rate of the carrier gas.

In the chromatogram obtained with reference solution (a), measure the areas of the peaks due to 17α-dihydroequilin, estrone and 3-*O*-methylestrone.

In the chromatogram obtained with test solution (a), locate the peaks with relative retentions with reference to 3-*O*-methylestrone of 1 and about 0.24, 0.29, 0.30, 0.35, 0.56, 0.64, 0.90 and 1.3 and measure their areas.

Calculate the percentage content of the components occurring as sodium sulfate salts using expression (1) below.

In the chromatogram obtained with reference solution (b), measure the areas of the peaks due to estrone and 3-*O*-methylestrone.

In the chromatogram obtained with test solution (b), locate the peaks with relative retentions with reference to 3-*O*-methylestrone of about 0.30, 0.80 and 0.87 and measure the sum of the areas.

Calculate the percentage content of 17α-dihydroequilin, estrone and equilin occurring as free steroids using expression (2) below.

$$\frac{S'_A \times S_I \times m_R \times 137.8 \times 1000}{S_R \times S'_I \times m \times LC} \qquad (1)$$

$$\frac{S'_{FS} \times S_I \times m_E \times 100 \times 1000}{S_E \times S'_I \times m \times LC} \qquad (2)$$

S_I = area of the peak due to the internal standard in the chromatogram obtained with the corresponding reference solution;
S'_I = area of the peak due to the internal standard in the chromatogram obtained with the corresponding test solution;
S_R = area of the peak due to the reference substance (Table 1512.-1) in the chromatogram obtained with the corresponding reference solution;
S'_A = area of the peak due to the analyte in the chromatogram obtained with the corresponding test solution;
m_R = mass of the reference substance (Table 1512.-1) in the corresponding reference solution, in milligrams;
m = mass of the substance to be examined in the corresponding test solution, in milligrams;
S'_{FS} = sum of the areas of the peaks due to 17α-dihydroequilin, estrone and equilin in the chromatogram obtained with the corresponding test solution;
S_E = area of the peak due to *estrone CRS* in the chromatogram obtained with the corresponding reference solution;
m_E = mass of *estrone CRS* in the corresponding reference solution, in milligrams;
LC = labelled content, in milligrams per gram.

The percentages are within the following ranges:
— *sodium 17α-estradiol sulfate*: 2.5 per cent to 9.5 per cent;
— *sodium 17α-dihydroequilin sulfate*: 13.5 per cent to 19.5 per cent;
— *sodium 17β-dihydroequilin sulfate*: 0.5 per cent to 4.0 per cent;
— *sodium 17β-estradiol sulfate*: maximum 2.25 per cent;
— *sodium 17α-dihydroequilenin sulfate*: maximum 3.25 per cent;
— *sodium 17β-dihydroequilenin sulfate*: maximum 2.75 per cent;
— *sodium 8,9-didehydroestrone sulfate*: maximum 6.25 per cent;
— *sodium equilenin sulfate*: maximum 5.5 per cent;
— *sum of estrone, equilin and 17α-dihydroequilin*: maximum 1.3 per cent.

ASSAY

Gas chromatography (*2.2.28*) as described in the test for chromatographic profile with the following modifications.

Injection Test solution (a) and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the ratio of the area of the peak due to estrone to that due to the internal standard after at least 6 injections.

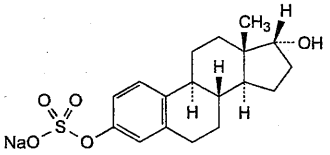
In the chromatogram obtained with reference solution (a), measure the areas of the peaks due to estrone or equilin and 3-*O*-methylestrone. In the chromatogram obtained with test solution (a), measure the areas of the peaks due to estrone, equilin and 3-*O*-methylestrone.

Calculate the percentage content of sodium estrone sulfate and sodium equilin sulfate using expression (1).

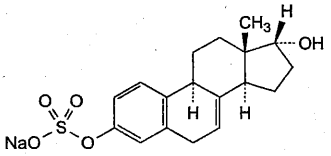
LABELLING

The label states:
— the name of the substance;
— the content of the substance;
— the nature of the diluent.

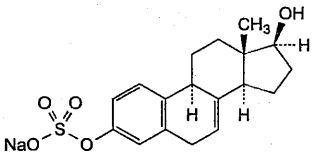
IMPURITIES AND CONCOMITANTS



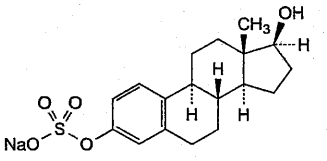
A. 17α-hydroxyestra-1,3,5(10)-trien-3-yl sodium sulfate (sodium 17α-estradiol sulfate),



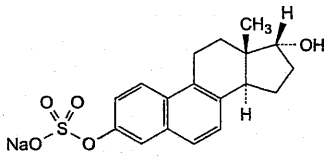
B. 17α-hydroxyestra-1,3,5(10),7-tetraen-3-yl sodium sulfate (sodium 17α-dihydroequilin sulfate),



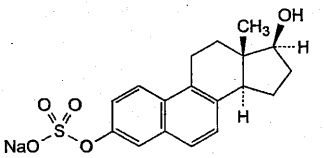
C. 17β-hydroxyestra-1,3,5(10),7-tetraen-3-yl sodium sulfate (sodium 17β-dihydroequilin sulfate),



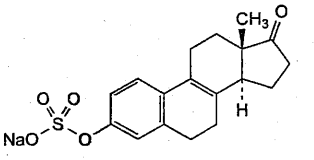
D. 17β-hydroxyestra-1,3,5(10)-trien-3-yl sodium sulfate (sodium 17β-estradiol sulfate),



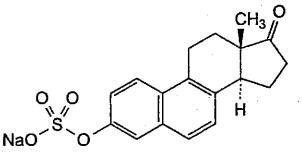
E. 17α-hydroxyestra-1,3,5(10),6,8-pentaen-3-yl sodium sulfate (sodium 17α-dihydroequilenin sulfate),



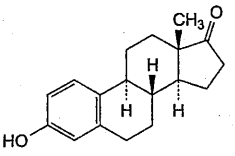
F. 17β-hydroxyestra-1,3,5(10),6,8-pentaen-3-yl sodium sulfate (sodium 17β-dihydroequilenin sulfate),



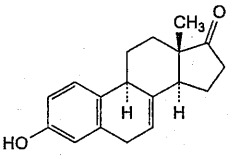
G. 17-oxoestra-1,3,5(10),8-tetraen-3-yl sodium sulfate (sodium 8,9-didehydroestrone sulfate),



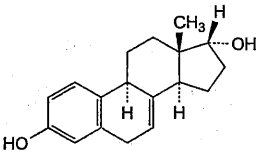
H. 17-oxoestra-1,3,5(10),6,8-pentaen-3-yl sodium sulfate (sodium equilenin sulfate),



I. 3-hydroxyestra-1,3,5(10)-trien-17-one (estrone),



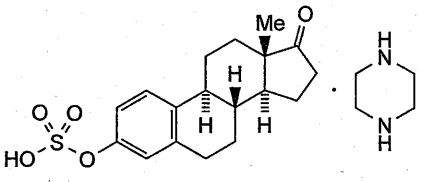
J. 3-hydroxyestra-1,3,5(10),7-tetraen-17-one (equilin),



K. estra-1,3,5(10),7-tetraene-3,17α-diol (17α-dihydroequilin).

Ph Eur

Estropipate



$C_{18}H_{22}O_5S,C_4H_{10}N_2$ 436.6 *7280-37-7*

Action and use
Estrogen.

DEFINITION
Estropipate is piperazine 17-oxoestra-1,3,5-(10)-trien-3-yl hydrogen sulfate (1:1). It contains not less than 97.0% and not more than 103.0% of $C_{18}H_{22}O_5S,C_4H_{10}N_2$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Very slightly soluble in *water*, in *ethanol (96%)* and in *ether*.

IDENTIFICATION
The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of estropipate *(RS 129)*.

TESTS

Free estrone

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol*.

(1) 0.10% w/v of the substance being examined.

(2) 0.0020% w/v of *estrone BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (30 cm × 3.9 mm) packed with *end-capped octadecylsilyl silica gel for chromatography*, (10 µm) (µBondapak C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.5 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 213 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

35 volumes of *acetonitrile* and 65 volumes of 0.025M *potassium dihydrogen orthophosphate*.

The peak due to estrone has a retention time, relative to the peak due to estropipate, of about 5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to estrone is not greater than the area of the peak in the chromatogram obtained with solution (2) (2%).

Loss on drying

When dried at 105° for 1 hour, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.5%, Appendix IX A. Use 1 g.

ASSAY

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol*.

(1) 0.01% w/v of the substance being examined.

(2) 0.01% w/v of *estropipate BPCRS*.

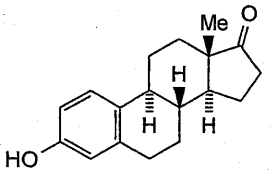
CHROMATOGRAPHIC CONDITIONS

The chromatographic procedure described under Free estrone may be used.

DETERMINATION OF CONTENT

Calculate the content of $C_{18}H_{22}O_5S,C_4H_{10}N_2$ in the substance being examined using the declared content of $C_{18}H_{22}O_5S,C_4H_{10}N_2$ in *estropipate BPCRS*.

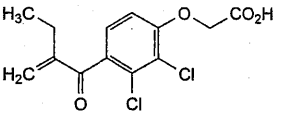
IMPURITIES



A. Estrone.

Etacrynic Acid

(Ph. Eur. monograph 0457)



$C_{13}H_{12}Cl_2O_4$ 303.1 *58-54-8*

Action and use

Loop diuretic.

Ph Eur

DEFINITION

[2,3-Dichloro-4-(2-methylenebutanoyl)phenoxy]acetic acid

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent). It dissolves in ammonia and in dilute solutions of alkali hydroxides and carbonates.

IDENTIFICATION

First identification: C.

Second identification: A, B, D, E.

A. Melting point (*2.2.14*): 121 °C to 124 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Solvent mixture 103 g/L solution of *hydrochloric acid R*, *methanol R* (1:99 *V/V*).

Test solution Dissolve 50.0 mg in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 10.0 mL of this solution to 100.0 mL with the solvent mixture.

Spectral range 230-350 nm.

Absorption maximum At 270 nm.

Shoulder At about 285 nm.

Specific absorbance at the absorption maximum 110 to 120.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *etacrynic acid CRS*.

D. Dissolve about 30 mg in 2 mL of *aldehyde-free alcohol R*. Dissolve 70 mg of *hydroxylamine hydrochloride R* in 0.1 mL of *water R*, add 7 mL of *alcoholic potassium hydroxide solution R* and dilute to 10 mL with *aldehyde-free alcohol R*. Allow to stand and add 1 mL of the supernatant to the solution of the substance to be examined. Heat the mixture on a water-bath for 3 min. After cooling, add 3 mL of *water R* and 0.15 mL of *hydrochloric acid R*. Examined in ultraviolet light at 254 nm, the mixture shows an intense blue fluorescence.

E. Dissolve about 25 mg in 2 mL of a 42 g/L solution of *sodium hydroxide R* and heat in a water-bath for 5 min. Cool and add 0.25 mL of a mixture of equal volumes of *sulfuric acid R* and *water R*. Add 0.5 mL of a 100 g/L solution of *chromotropic acid, sodium salt R* and, carefully, 2 mL of *sulfuric acid R*. An intense violet colour is produced.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R*, *water R* (40:60 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *etacrynic acid for system suitability CRS* (containing impurities A, B and C) in 5.0 mL of the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 25 °C.

Mobile phase:
- *mobile phase A*: 1 per cent *V/V* solution of *triethylamine R* adjusted to pH 6.8 with *phosphoric acid R*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0-2.5	70	30
2.5-3	70→65	30→35
3-6	65	35
6-7	65→45	35→55
7-22	45	55

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *etacrynic acid for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to etacrynic acid (retention time = about 9 min): impurity A = about 0.8; impurity B = about 1.3; impurity C = about 1.7.

System suitability Reference solution (b):
- *resolution*: minimum 4.0 between the peaks due to impurity A and etacrynic acid.

Limits:
- *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity B = 0.6; impurity C = 1.3;
- *impurity C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurities A, B*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

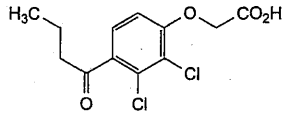
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 2.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure of 0.1-0.5 kPa.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

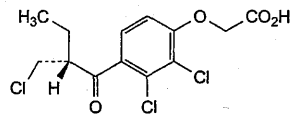
ASSAY
Dissolve 0.250 g in 100 mL of *methanol R* and add 5 mL of *water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 30.31 mg of $C_{13}H_{12}Cl_2O_4$.

IMPURITIES
Specified impurities *A, B, C.*

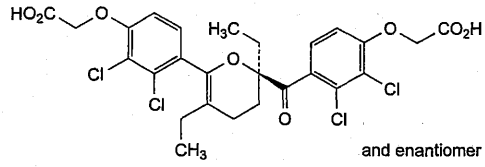


A. (4-butanoyl-2,3-dichlorophenoxy)acetic acid,



and enantiomer

B. [2,3-dichloro-4-[2-(chloromethyl)butanoyl]phenoxy]acetic acid,



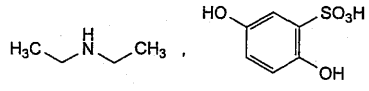
and enantiomer

C. [4-[2-[4-(carboxymethoxy)-2,3-dichlorobenzoyl]-2,5-diethyl-3,4-dihydro-2*H*-pyran-6-yl]-2,3-dichlorophenoxy] acetic acid.

Ph Eur

Etamsylate

(Ph. Eur. monograph 1204)



$C_{10}H_{17}NO_5S$ 263.3 *2624-44-4*

Action and use
Antifibrinolytic.

Ph Eur

DEFINITION
N-Ethylethanamine 2,5-dihydroxybenzenesulfonate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in methanol, soluble in anhydrous ethanol, practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 127 °C to 134 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *etamsylate CRS.*

C. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.100 g in *water R* and dilute to 200.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with *water R*. Examine immediately.

Spectral range 210-350 nm.

Absorption maxima At 221 nm and 301 nm.

Specific absorbance at the absorption maximum at 301 nm 145 to 151.

D. Into a test-tube, introduce 2 mL of freshly prepared solution S (see Tests) and 0.5 g of *sodium hydroxide R*. Warm the mixture and place a wet strip of *red litmus paper R* near the open end of the tube. The colour of the paper becomes blue.

TESTS

Solution S
Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S, when freshly prepared, is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.5 to 5.6 for solution S.

Related substances
Liquid chromatography (*2.2.29*). *Keep all solutions at 2-8 °C.*

Buffer solution Dissolve 1.2 g of *anhydrous sodium dihydrogen phosphate R* in 900 mL of *water for chromatography R*. Adjust to pH 6.5 with *disodium hydrogen phosphate solution R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of *hydroquinone R* (impurity A) in *water R* and dilute to 10 mL with the same solvent. Dilute 1 mL of the solution to 100 mL with *water R*.

Column:
— *size:* l = 0.25 m, Ø = 4.6 mm;
— *stationary phase:* spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R1*, buffer solution (10:90 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 11 times the retention time of etamsylate.

Relative retention With reference to etamsylate (retention time = about 6 min): impurity A = about 1.7.

System suitability Reference solution (b):
— *resolution*: minimum 8.0 between the peaks due to etamsylate and impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 0.5;
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* in an oven at 60 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 10 mL of *water R* and 40 mL of *dilute sulfuric acid R*. Titrate with *0.1 M cerium sulfate*, determining the end-point potentiometrically (*2.2.20*).

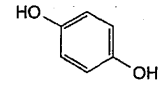
1 mL of *0.1 M cerium sulfate* is equivalent to 13.16 mg of $C_{10}H_{17}NO_5S$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities *A.*



A. benzene-1,4-diol (hydroquinone).

Ph Eur

Etanercept

(Ph. Eur. monograph 2895)

LPAQVAFTPY	APEPGSTCRL	REYYDQTAQM	CCSKCSPGQH	40
AKVFCTKTSD	TVCDSCEDST	YTQLWNWVPE	CLSCGSRCSS	80
DQVETQACTR	EQNRICTCRP	GWYCALSKQE	GCRLCAPLRK	120
CRPGFGVARP	GTETSDVVCK	PCAPGTFSNT	TSSTDICRPH	160
QICNVVAIPG	NASMDAVCTS	TSPTRSMAPG	AVHLPQPVST	200
RSQHTQPTPE	PSTAPSTSFL	LPMGPSPPAE	GSTGDEPKSC	240
DKTHTCPPCP	APELLGGPSV	FLFPPKPKDT	LMISRTPEVT	280
CVVVDVSHED	PEVKFNWYVD	GVEVHNAKTK	PREEQYNSTY	320
RVVSVLTVLH	QDWLNGKEYK	CKVSNKALPA	PIEKTISKAK	360
GQPREPQVYT	LPPSREEMTK	NQVSLTCLVK	GFYPSDIAVE	400
WESNGQPENN	YKTTPPVLDS	DGSFFLYSKL	TVDKSRWQQG	440
NVFSCSVMHE	ALHNHYTQKS	LSLSPGK		467

$_2$

disulfide bridges (the list is not exhaustive):
240-240′, 246-246′, 249-249′, 281-341, 387-445, 281′-341′, 387′-445′

N-glycosylation sites:
149, 171, 317, 149′, 171′, 317′

predominant *O*-glycosylation sites:
184, 199, 200, 205, 208, 212, 213, 216, 217, 226, 184′, 199′, 200′, 205′, 208′, 212′, 213′, 216′, 217′, 226′

$C_{2224}H_{3472}N_{618}O_{701}S_{36}$ (monomer) *185243-69-0*
M_r approx. 51 200 (monomer without glycosylation)

Action and use

Human tumour necrosis factor receptor fusion protein; treatment of rheumatoid arthritis and psoriasis.

Ph Eur

DEFINITION

Dimeric fusion protein consisting of the extracellular ligand-binding portion of the human tumour necrosis factor receptor (TNFR) linked to the Fc portion of human IgG1. The Fc component of etanercept contains the CH_2 domain, the CH_3 domain and the hinge region, but not the CH_1 domain of IgG1. Etanercept consists of 934 amino acids and has an apparent molecular mass of approximately 150 kDa.

Etanercept represents a glycosylated protein with multiple *N*- and *O*-linked glycosylation sites. It shows full occupancy of *N*-linked glycans at N149, N171 and N317.

It contains one or more suitable buffering and/or stabilising agents.

Content (milligrams of protein per millilitre) As approved by the competent authority.

Potency

1.0×10^6 to 2.9×10^6 IU per milligram of protein.

PRODUCTION

Etanercept is produced in a suitable mammalian cell expression system by a method based on recombinant DNA (rDNA) technology. During the course of product development, it must be demonstrated that the manufacturing process consistently produces a product with the expected *O*-glycan occupancy using a suitably qualified assay.

Prior to release, the following tests are carried out on each batch of etanercept, unless exemption has been granted by the competent authority.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell and vector-derived DNA

The limit is approved by the competent authority.

N-Glycan analysis

Use a suitable method developed according to general chapter *2.2.59. Glycan analysis of glycoproteins*, section 2-3:
— release the glycans using one of the agents described in Table *2.2.59.*-1, for example peptide *N*-glycosidase F (PNGase F);
— label the released glycans with one of the fluorescent labelling agents described in Table *2.2.59.*-2, for example 2-aminobenzamide;
— analyse the labelled glycans by liquid chromatography (*2.2.29*) using fluorescence detection.

The following procedure is given as an example.

Test solution To 4 µL of the preparation to be examined (about 25 mg/mL) add 21 µL of *water R*, 3 µL of *0.25 M sodium phosphate buffer solution pH 7.5 R* and 2 µL of a 500 000 U/mL solution of *peptide N-glycosidase F R*. Mix and incubate at 37 °C for 20-24 h. Label the released glycans with 2-aminobenzamide using a suitable procedure.
The procedure employs a combination of reagents optimised and validated for the efficient labelling of glycans, and for the subsequent extraction and recovery of the labelled glycans from the reaction. Resuspend or dilute the labelled glycans in 100 µL of *water R*.

Reference solution (a) Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of about 25 mg/mL. Carry out the release and labelling of glycans in the same manner as for the test solution. Resuspend or dilute the labelled glycans in 100 µL of *water R*.

Reference solution (b) Use a suitable etanercept in-house reference preparation shown to be representative of batches tested clinically and batches used to demonstrate consistency of production. Dilute, if necessary, with *water R* to obtain a concentration of about 25 mg/mL. Carry out the release and labelling of glycans in the same manner as for the test solution. Resuspend or dilute the labelled glycans in 100 µL of *water R*.

Blank solution Prepare at the same time and in the same manner as for the test solution but using *water R* instead of the preparation to be examined.

Analyse the labelled glycans by liquid chromatography (*2.2.29*).

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: an amide derivative of *silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

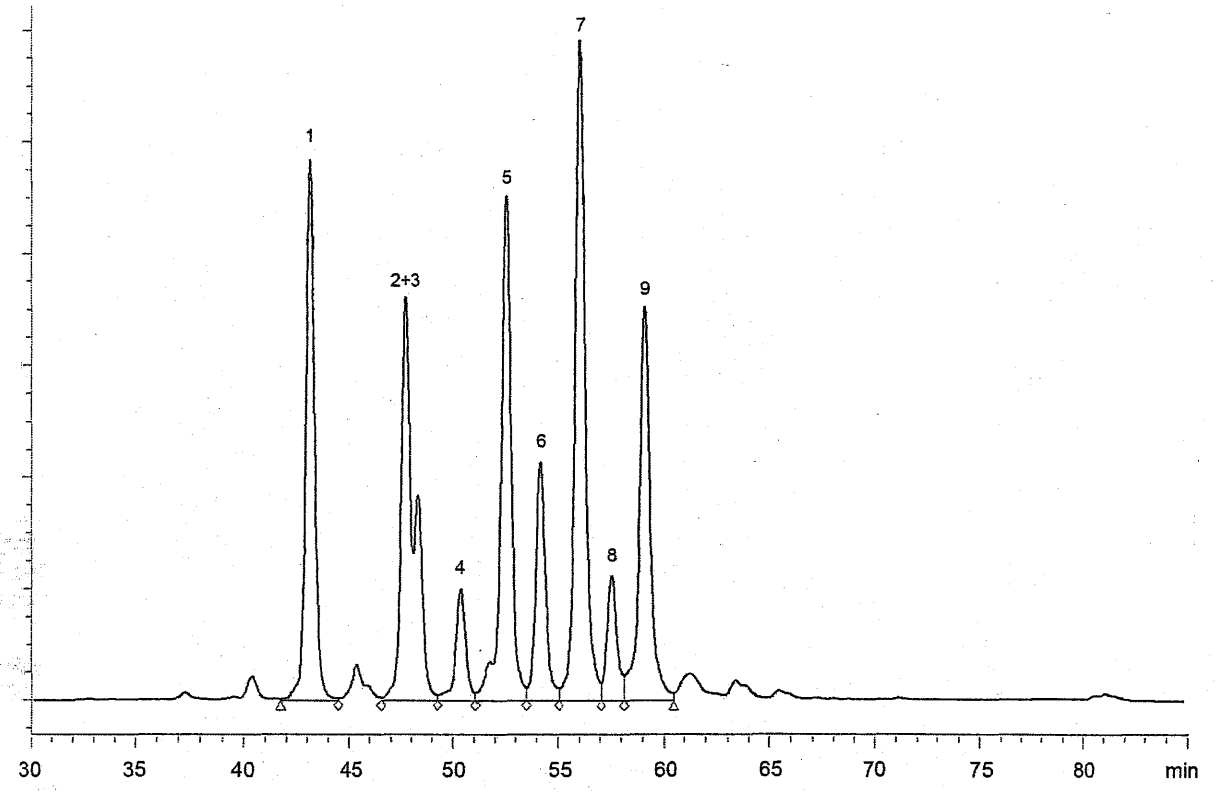
Mobile phase:
— *mobile phase A*: mix 9.8 mL of *anhydrous formic acid R* and 500 mL of *water for chromatography R*, adjust to pH 4.0 with *ammonia R* and dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	20 → 30	80 → 70
2 - 67.0	30 → 52	70 → 48
67.0 - 67.1	52 → 80	48 → 20
67.1 - 73.0	80	20

Flow rate 0.4 mL/min.

Detection Fluorimeter at 330 nm for excitation and 420 nm for emission.

Autosampler Set at 2-8 °C.



Peak	Charged	Structure	Peak	Charged	Structure	Peak	Charged	Structure	Peak	Charged	Structure
1.	No	Asialo-, agalacto-, biantennary, core-fucosylated	4.	No	Asialo-, galactosylated biantennary	6.	Yes	Monosialylated-, galactosylated biantennary	8.	Yes	Disialylated, galactosylated biantennary
2+3.	No	Asialo-, mono-galactosylated biantennary, core-fucosylated	5.	No	Asialo-, galactosylated biantennary, core-fucosylated	7.	Yes	Monosialylated-, galactosylated biantennary, core-fucosylated	9.	Yes	Disialylated-, galactosylated biantennary, core-fucosylated

Figure 2895.-1. – *Chromatogram for N-glycan analysis of etanercept*

Injection 10 µL.

Identification of peaks Use the chromatogram shown in Figure 2895.-1 to identify the 2 groups of oligosaccharides corresponding to neutral (peaks 1 to 5) and sialylated (peaks 6 to 9) *N*-glycans; record the retention time of each peak in both groups.

System suitability:
- the chromatogram obtained with reference solution (a) is qualitatively similar to the chromatogram supplied with *etanercept CRS* and peaks 1 to 9 are clearly visible;
- no significant peaks are observed in the chromatogram obtained with the blank solution.

Results:
- the profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with reference solution (b);
- the retention times of the peaks in the chromatogram obtained with the test solution correspond to those in the chromatogram obtained with reference solution (b);
- no additional peaks are observed in the chromatogram obtained with the test solution in comparison with the chromatogram obtained with reference solution (b).

Calculate the relative peak areas of the individual peaks corresponding to neutral and sialylated *N*-glycans with reference to the sum of the areas of all retained glycan peaks.

Calculate the percentage contents of the neutral and sialylated groups, using the following expressions:

$$\frac{A}{A+B} \times 100$$

$$\frac{B}{A+B} \times 100$$

A = sum of the areas of the peaks due to neutral *N*-glycans;
B = sum of the areas of the peaks due to sialylated *N*-glycans.

NOTE: peaks 2 and 3 are separate peaks, but they are integrated together.

Limits:
- *percentage of neutral N-glycans*: as approved by the competent authority;
- *percentage of sialylated N-glycans*: as approved by the competent authority.

CHARACTERS

Appearance

Clear, almost colourless, slightly yellow or slightly brown liquid.

IDENTIFICATION

A. It complies with the limits of the assay (potency).

B. Peptide mapping (*2.2.55*).

Selective cleavage of the peptide bonds

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of about 15 mg/mL.

Reference solution Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of about 15 mg/mL.

Reduction and alkylation To 200 µL of the test solution add 500 µL of *guanidine-tris(hydroxymethyl)aminomethane buffer solution pH 8.3 R* and 7 µL of a 154 g/L solution of *dithiothreitol R*. Mix and incubate at 65 °C for 15 min. Cool in an ice-bath for 5-10 min, then add 15.4 µL of a freshly prepared 185 g/L solution of *iodoacetamide R*. Mix and allow to stand protected from light for 10 min. Add 1.4 µL of a 154 g/L solution of *dithiothreitol R*. Mix and allow to stand protected from light for 10 min.

Digestion To 97 µL of the reduced test solution prepared previously, add 903 µL of *0.1 M tris-hydrochloride buffer solution pH 7.5 R*. Add 9.6 µL of a 500 000 U/mL solution of *peptide N-glycosidase F R* and incubate at 37 °C for 1 h. Add 40 µL of a 1 mg/mL solution of *trypsin for peptide mapping R* and incubate at 37 °C for 5 h. Heat at 95 °C for 5 min and cool in ice for 5 min. Adjust to pH 2 with about 30 µL of a 150 g/L solution of *trifluoroacetic acid R*.

Carry out the reduction/alkylation and digestion steps for the reference solution in the same manner as for the test solution.

Chromatographic separation. Liquid chromatography (*2.2.29*).

Column:

- *size*: *l* = 0.25 m, Ø = 3.2 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 10 nm;
- *temperature*: 30-35 °C.

Mobile phase:

- *mobile phase A*: mix 3 g of *trifluoroacetic acid R* and 2000 mL of *water for chromatography R*;
- *mobile phase B*: mix 2 g of *trifluoroacetic acid R*, 330 mL of *water for chromatography R* and 1320 mL of *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	98	2
5 - 125	98 → 50	2 → 50
125 - 140	50 → 5	50 → 95

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 220 nm.

Autosampler Set at 2-8 °C.

Injection 200 µL.

System suitability:

- the chromatogram obtained with the reference solution is qualitatively similar to the chromatogram supplied with *etanercept CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

TESTS

pH (*2.2.3*)

As approved by the competent authority.

Sialic acid

Use a suitable method developed according to general chapter *2.2.59. Glycan analysis of glycoproteins*, section 2-4:

- release the sialic acid by acid hydrolysis pre-treatment of the preparation to be examined;
- label the released sialic acid with a fluorescent labelling reagent, for example 1,2-diamino-4,5-methylenedioxybenzene, using a suitable procedure;
- analyse the labelled sialic acid by liquid chromatography (*2.2.29*) using fluorescence detection.

The following procedure is given as an example.

Solution A Dissolve 8.71 g of *arginine R* in 40 mL of *water R*. Add 0.5 mL of a 10 g/L solution of *polysorbate 80 R*, mix and adjust to pH 7.3 with *phosphoric acid R*. Dilute to 250 mL with *water R*.

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of about 5 mg/mL. Further dilute with solution A to obtain a concentration of about 1 mg/mL. To 50 µL of this solution add 50 µL of a 480 g/L solution of *glacial acetic acid R* and incubate at 90 °C for 65 min. Cool, briefly centrifuge and evaporate to dryness. Label the released sialic acid using a suitable procedure; for example, add 15 µL of a 1.6 g/L solution of *1,2-diamino-4,5-methylenedioxybenzene dihydrochloride R* containing 78.1 g/L of *2-mercaptoethanol R* and 3.1 g/L of *sodium dithionite R* and incubate at 50 °C for 3 h. Dilute to 1 mL with *water R*.

Reference solution (a) Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of about 5 mg/mL. Carry out the release and labelling of sialic acid in the same manner as for the test solution. Dilute to 1 mL with *water R*.

Reference solution (b) Dissolve *N-acetylneuraminic acid R* in *water R* to obtain a concentration of about 1 mg/mL. Mix 40 µL of the solution, 40 µL of a 1 mg/mL solution of *bovine albumin R1* and 120 µL of solution A. Use 50 µL of this solution to carry out the release and labelling of sialic acid in the same manner as for the test solution. Dilute with *water R* to obtain a concentration of 0.01 µg/µL.

Standard solutions Dilute reference solution (b) with *water R* to obtain a concentration of 2 ng/µL. Further dilute this solution to prepare a standard curve with concentrations in the range of 0.10-0.40 ng/µL (6 concentrations, typically 0.10 ng/µL, 0.15 ng/µL, 0.20 ng/µL, 0.25 ng/µL, 0.30 ng/µL, 0.40 ng/µL). Analyse the labelled sialic acid by liquid chromatography (*2.2.29*).

Column:

- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 8 nm;
- *temperature*: 35 °C.

Mobile phase Mix 8 mL of *acetonitrile R*, 500 mL of *methanol R* and 1820 mL of *water for chromatography R*; mix thoroughly.

Flow rate 1 mL/min.

Detection Fluorimeter at 374 nm for excitation and 448 nm for emission.

Autosampler Set at 2-8 °C.

Injection 20 µL.

Retention time Sialic acid = about 10.5 min.

Calculate the sialic acid content in the preparation to be examined using the standard curve and the content of sialic acid (*N*-acetylneuraminic acid) in the standard solutions. Report the molar ratio (number of moles of sialic acid per mole of etanercept), using the molar mass of the monomer.

System suitability:
- the peak due to sialic acid in the chromatogram obtained with reference solution (a) is visible and is similar to the corresponding peak in the chromatogram supplied with *etanercept CRS*;
- *repeatability*: maximum relative standard deviation of 15 per cent for the sialic acid content expressed as the molar ratio, determined on 3 consecutive injections of reference solution (a);
- the coefficient of determination (R^2) calculated for the standard curve is not less than 0.9995.

Results:
- the profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with reference solution (a).

Limit:
- 8 to 19 moles of sialic acid per mole of etanercept.

Related proteins

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of about 2 mg/mL.

Reference solution Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of about 2 mg/mL.

Column:
- *size*: *l* = 0.035 m, Ø = 4.6 mm;
- *stationary phase*: *resin for hydrophobic interaction chromatography R* (2.5 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: dissolve 28.4 g of *anhydrous disodium hydrogen phosphate R* and 475.9 g of *ammonium sulfate R* in *water for chromatography R* and dilute to 1950 mL with the same solvent; adjust to pH 7.0 with *phosphoric acid R* and dilute to 2000 mL with *water for chromatography R*;
- *mobile phase B*: dissolve 28.4 g of *anhydrous disodium hydrogen phosphate R* in *water for chromatography R* and dilute to 1950 mL with the same solvent; adjust to pH 7.0 with *phosphoric acid R* and dilute to 2000 mL with *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 50	100 → 0	0 → 100

Flow rate 1.0 mL/min.

Detection Fluorimeter at 278 nm for excitation and 350 nm for emission.

Autosampler Set at 10 °C.

Injection 5 µL; perform 3 injections.

Relative retention With reference to etanercept (retention time = about 28.5 min): peak 1 = 0.96; peak 3 = 1.12.

System suitability Reference solution:
- the chromatogram obtained is qualitatively similar to the chromatogram supplied with *etanercept CRS*;
- peaks 1 and 3 are clearly separated from the peak due to etanercept.

Results:
- no additional peaks are observed in the chromatogram obtained with the test solution in comparison with the chromatogram obtained with the reference solution.

Limits:
- *peak 1*: maximum 5 per cent;
- *peak 3*: maximum 28 per cent;
- *sum of all peaks other than the principal peak*: maximum 30 per cent.

Impurities with molecular masses greater than that of etanercept

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Solution A Dissolve 8.8 g of *sodium chloride R* and 15.6 g of *sodium dihydrogen phosphate R* in 900 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent.

Solution B Dissolve 8.75 g of *sodium chloride R* and 14.2 g of *anhydrous disodium hydrogen phosphate R* in 900 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent.

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 2.5 mg/mL.

Reference solution Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of 2.5 mg/mL.

Column:
- *size*: *l* = 0.30 m, Ø = 8.0 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5 µm) with a pore size of 30 nm and of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 1000 000.

Mobile phase Mix 220 mL of solution A and 780 mL of solution B, and adjust to pH 7.2 with solution A or solution B.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Autosampler Set at 2-8 °C.

Injection 14 µL; perform at least 3 injections.

Relative retention With reference to etanercept monomer (retention time = about 7.8 min): aggregates = 0.84; high molecular mass species = 0.89.

Any shoulder appearing on the descending part of the peak due to etanercept monomer is included in its area.

System suitability Reference solution:
- the chromatogram obtained is qualitatively similar to the chromatogram supplied with *etanercept CRS*;
- *resolution*: minimum 1.7 between the peaks due to the high molecular mass species and etanercept monomer;
- *number of theoretical plates*: minimum 3000 calculated for the peak due to etanercept monomer.

Limit:
- *sum of the peaks eluted before the principal peak*: maximum 8.0 per cent.

Impurities with molecular masses differing from that of etanercept

Polyacrylamide gel electrophoresis (*2.2.31*) under both reducing and non-reducing conditions.

Gel dimensions 1.0 mm thick.

Resolving gel 8-16 per cent acrylamide.

Sample buffer (non-reducing conditions) *concentrated SDS-PAGE sample buffer R.*

Sample buffer (reducing conditions) *concentrated SDS-PAGE sample buffer for reducing conditions R.*

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 0.2 mg/mL. Mix 1 volume of this solution and 1 volume of sample buffer.

Reference solution (a) Dissolve the contents of a vial of *etanercept CRS* in *water R* to obtain a concentration of 0.2 mg/mL. Mix 1 volume of the solution and 1 volume of sample buffer.

Reference solution (b) 0.01 mg/mL solution of *bovine albumin R.*

Reference solution (c) Mix 1 volume of reference solution (b) and 4 volumes of *concentrated SDS-PAGE sample buffer R.*

Reference solution (d) Mix 1 volume of reference solution (b) and 19 volumes of *concentrated SDS-PAGE sample buffer R.*

Reference solution (e) A solution of molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 5-200 kDa.

Sample treatment Heat at 90-105 °C for 5 min and load onto the gel within 15 min.

Application 10 µL.

Detection By silver staining.

Identification of bands:
- *non-reducing conditions*: the band corresponding to etanercept of an apparent molecular mass of approximately 150 kDa is present; related protein bands with apparent molecular masses of approximately 60 kDa, 100 kDa, 120 kDa, 225 kDa, 250 kDa may also be present;
- *reducing conditions*: the band corresponding to etanercept of an apparent molecular mass of approximately 76 kDa is present; related protein bands with apparent molecular masses of approximately 25 kDa, 35 kDa, 55 kDa, 200 kDa may also be present.

System suitability:
- the bands in the electropherogram obtained with reference solution (a) are clearly visible;
- the band in the electropherogram obtained with reference solutions (b), (c) and (d) is clearly visible;
- all expected bands in the electropherogram obtained with reference solution (e) are visible and clearly separated.

Results:
- the electropherogram obtained with the test solution is similar to the electropherogram obtained with reference solution (a);
- the electropherogram obtained with the test solution shows no additional band that is more intense than that of the band in the electropherogram obtained with reference solution (d).

Microbiological contamination (*2.6.12*)
Maximum 1 CFU/mL.

ASSAY

Protein (*2.5.33, Method 1*)

Test solution Dilute the preparation to be examined gravimetrically with a suitable buffer to obtain a concentration of 1.0 mg/mL. Prepare in triplicate.

Reference solution Dissolve the contents of a vial of *etanercept CRS* in a suitable buffer to obtain a concentration of 1.0 mg/mL.

Record the absorbance spectrum between 250 nm and 400 nm. Measure the value at the absorbance maximum of 280 nm after correction for any light scattering measured up to 320 nm. Calculate the protein concentration of etanercept taking into account the assigned content of *etanercept CRS.*

Potency

The potency of etanercept is determined by comparison of dilutions of the test preparation with the dilutions of *etanercept BRP* using a suitable cell-based assay based on the inhibitory action of etanercept on the biological activity of TNF-α and a suitable readout for assessing this inhibitory effect.

The following procedure is given as an example.

Carry out an apoptosis-based assay based on the ability of etanercept to inhibit TNF-α induced apoptosis in histiocytic lymphoma cell line U937 (ATCC No. CRL-1593.2) via caspase activation. The U937 cells are incubated with varying dilutions of test and reference preparations of etanercept in the presence of TNF-α. They are then incubated with Caspase-Glo 3/7 reagent, which results in caspase cleavage of a luminogenic substrate, subsequent release of a luciferase substrate and generation of a luminescent signal. The luminescence produced is proportional to the amount of caspase activity present.

Assay medium RPMI 1640 containing L-alanyl-L-glutamine, 6.0 g/L *HEPES R* (25 mM) and foetal bovine serum (7.5 per cent *V/V*).

Test solutions Dilute the preparation to be examined with assay medium to obtain a concentration of about 72 ng/mL. Use this solution to prepare 11 additional sample dilutions (dilution steps of 1.2 or 1.4 have been found suitable).

Reference solutions Reconstitute the contents of 1 ampoule of *etanercept BRP* with sterilised *water for injections R* to obtain a concentration of 10 000 IU/mL. Further dilute with assay medium to obtain a concentration of 144 IU/mL. Use this solution to prepare 11 additional dilutions to generate the standard curve (dilution steps of 1.2 or 1.4 have been found suitable).

TNF-α working solution Dissolve the contents of a vial of TNF-α according to the supplier's instructions. Further dilute with assay medium to obtain a suitable working concentration. As the biological activity of TNF-α is likely to vary between different suppliers and also between different batches from the same supplier, this should be controlled by use of an appropriate standard (e.g. WHO International Standard for TNF-α).

Method.

Plate preparation Add 600 µL of assay medium to the wells designated for cells only (column 1, rows A-D) on a cluster tube rack. Add 300 µL of assay medium and 300 µL of TNF-α working solution to the wells designated for the TNF-α controls (column 1, rows E-H). Add 300 µL of the test or reference solutions and 300 µL of TNF-α working solution to the sample wells (columns 2-12, rows A-H). Mix on a shaker for 5 min. Incubate at 36.0-38.0 °C for 30-60 min in a humidified incubator using 5 ± 2 per cent CO_2.

NOTE: when using deep-well or 96-well plates instead of cluster tubes, adapt the volumes of sample, TNF-α working solution and assay medium accordingly.

Cell preparation A cell density between 3.0×10^5 and 1.0×10^6 cells per millilitre is suitable, and cell viability is not less than 95 per cent.

Plating test solution, reference solution, controls and cells
Transfer 60 µL from each cluster tube and add to the corresponding wells. Mix the cell suspension thoroughly and add 60 µL to each well. Mix the contents of the plates on a

shaker for 5 min. Incubate the plates without lids at 36.0-38.0 °C for 2-2.5 h in a humidified incubator using 5 ± 2 per cent CO_2.

Addition of Caspase-Glo 3/7 assay system Reconstitute the Caspase-Glo 3/7 assay system according to the manufacturer's instructions and add 100 µL to each well of the assay plates. Shake the plates, covered with black lids, on a plate shaker for 10-15 min. Incubate at room temperature for 30-60 min. Place the uncovered plates in a luminometer and read the luminescence for a minimum of 1 second per well.

Calculate the potency of the preparation to be examined using the four-parameter logistic curve model (*5.3*).

System suitability:

— *maximum value (TNF-α control) to minimum value (cell only) ratio*: minimum 3.0.

Result The estimated potency is not less than 80 per cent and not more than 140 per cent relative to the reference solution. The confidence limits (P = 0.95) are not less than 80 per cent and not more than 125 per cent of the estimated potency.

STORAGE

In an airtight container at −20 °C or below.

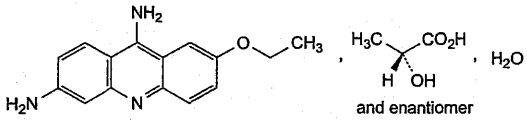
LABELLING

The label states the content, in milligrams of protein per millilitre.

Ph Eur

Ethacridine Lactate Monohydrate

(Ph. Eur. monograph 1591)



$C_{18}H_{21}N_3O_4,H_2O$ 361.4 *6402-23-9*

Action and use

Antiseptic.

Ph Eur

DEFINITION

7-Ethoxyacridine-3,9-diamine (2*RS*)-2-hydroxypropanoate monohydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow crystalline powder.

Solubility

Sparingly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ethacridine lactate monohydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in 2 mL of *water R* and dilute to 10 mL with *methanol R.*

Reference solution Dissolve 5 mg of *ethacridine lactate monohydrate CRS* in 2 mL of *water R* and dilute to 10 mL with *methanol R.*

Plate *TLC silica gel F_{254} plate R.*

(Merck

Mobile phase *glacial acetic acid R, water R, butanol R* (17:17:66 *V/V/V*).

Application 2 µL; the volume can be adapted based on the type of plate used.

Development Over 2/3 of the plate.

Drying At 100-105 °C.

Detection A Examine in daylight.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Examine in ultraviolet light at 254 nm and at 366 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 50 mL of solution S (see Tests) add 10 mL of *dilute sodium hydroxide solution R.* Filter. To 5 mL of the filtrate, add 1 mL of *dilute sulfuric acid R.* 5 mL of the solution obtained gives the reaction of lactates (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in *carbon dioxide-free water R* and dilute to 100.0 mL with the same solvent.

pH (*2.2.3*)

5.5 to 7.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 1.0 g of *sodium octanesulfonate R* in a mixture of 300 mL of *acetonitrile R* and 700 mL of *phosphate buffer solution pH 2.8 R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 268 nm.

Injection 10 µL.

Run time 3 times the retention time of ethacridine.

Retention time Ethacridine = about 15 min.

Limits:

— *any impurity*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
4.5 per cent to 5.5 per cent, determined on 1.000 g by drying *in vacuo* at 105 °C.

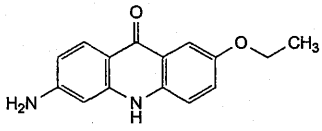
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.270 g in 5.0 mL of *anhydrous formic acid R*. Add 60.0 mL of *acetic anhydride R* and titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

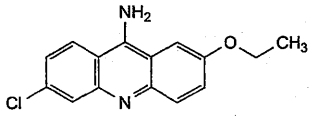
1 mL of *0.1 M perchloric acid* is equivalent to 34.34 mg of $C_{18}H_{21}N_3O_4$.

STORAGE
Protected from light.

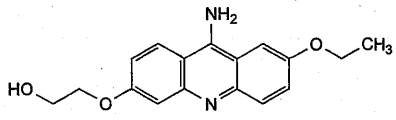
IMPURITIES



A. 6-amino-2-ethoxyacridin-9(10*H*)-one,



B. 6-chloro-2-ethoxyacridin-9-amine,

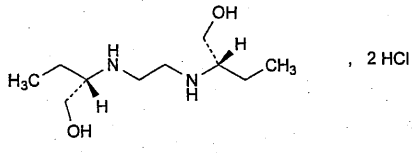


C. 2-[(9-amino-7-ethoxyacridin-3-yl)oxy]ethan-1-ol.

Ph Eur

Ethambutol Hydrochloride

(Ph. Eur. monograph 0553)



$C_{10}H_{26}Cl_2N_2O_2$ 277.2 *1070-11-7*

Action and use
Antituberculosis drug.

Preparations
Ethambutol Oral Solution
Ethambutol Tablets

Ph Eur

DEFINITION
(2*S*,2′*S*)-2,2′-(Ethylenediimino)dibutan-1-ol dihydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder, hygroscopic.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, D, E.
Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *ethambutol hydrochloride CRS.*

B. Examine the chromatograms obtained in the test for impurity A.
Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (b).

C. Dissolve 0.1 g in 10 mL of *water R*. Add 0.2 mL of *copper sulfate solution R* and 0.5 mL of *dilute sodium hydroxide solution R*; a blue colour is produced.

D. It gives reaction (a) of chlorides (*2.3.1*).

E. Related substances (see Tests).

TESTS

pH (*2.2.3*)
3.7 to 4.0.
Dissolve 0.2 g in 10 mL of *carbon dioxide-free water R*.

Impurity A
Thin-layer chromatography (*2.2.27*).
Test solution (a) Dissolve 0.50 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.
Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *methanol R*.
Reference solution (a) Dissolve 50.0 mg of *aminobutanol R* (impurity A) in *methanol R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.
Reference solution (b) Dissolve 50 mg of *ethambutol hydrochloride CRS* and 5 mg of *aminobutanol R* in *methanol R* and dilute to 10 mL with the same solvent.
Plate *TLC silica gel plate R.*
Mobile phase *concentrated ammonia R, water R, methanol R* (10:15:75 *V/V/V*).
Application 2 µL.
Development Over 2/3 of the plate.
Drying In air; heat at 110 °C for 10 min.
Detection Cool then spray with *ninhydrin solution R1*; heat at 110 °C for 5 min.
System suitability reference solution (b):
— the chromatogram shows 2 clearly separated spots.
Limit:
— *impurity A*: any spot due to impurity A in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (a) (1.0 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Suspend 4.0 mg of the substance to be examined in 4.0 mL of *acetonitrile R1* and add 100 µL of *triethylamine R*. Sonicate the mixture for 5 min. Add 15 µL of *(R)-(+)-α-methylbenzyl isocyanate R* and heat at 70 °C for 20 min.

Reference solution (a) Dilute 0.50 mL of the test solution to 100.0 mL with *acetonitrile R1*.

Reference solution (b) Treat 4.0 mg of *ethambutol for system suitability CRS* (containing impurity B) as described for the test solution.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: *methanol R, water R* (50:50 *V/V*);
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	71	29
30 - 35	71 → 0	29 → 100
35 - 37	0	100
37 - 38	0 → 71	100 → 29

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

Relative retention With reference to ethambutol (retention time = about 14 min): impurity B = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to ethambutol and impurity B.

Limits:
— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *unspecified impurities with a relative retention of 0.75 to 1.5 with reference to ethambutol*: for each impurity, not more than 0.2 times the area of the peak due to ethambutol in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total (impurity B and unspecified impurities with a relative retention of 0.75 to 1.5 with reference to ethambutol)*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the peak due to ethambutol in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity D (1,2-dichloroethane) (*2.4.24*)
Maximum 5 ppm.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

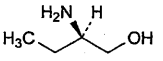
ASSAY
Dissolve 0.200 g in 50 mL of *water R* and add 1.0 mL of *0.1 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 27.72 mg of $C_{10}H_{26}Cl_2N_2O_2$.

STORAGE
In an airtight container.

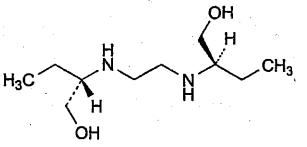
IMPURITIES
Specified impurities *A, B, D.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*

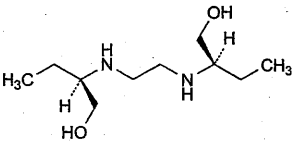


and enantiomer

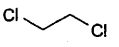
A. 2-aminobutan-1-ol,



B. (2*R*,2′*S*)-2,2′-(ethylenediimino)dibutan-1-ol (meso-ethambutol),



C. (2*R*,2′*R*)-2,2′-(ethylenediimino)dibutan-1-ol ((*R*,*R*)-ethambutol),

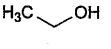


D. 1,2-dichloroethane (ethylene chloride).

Ph Eur

Ethanol[1]

Absolute Alcohol
Dehydrated Alcohol
(Anhydrous Ethanol, Ph. Eur. monograph 1318)



C_2H_6O 46.07 *64-17-5*

Ph Eur

DEFINITION
Content
Not less than 99.5 per cent *V/V* of C_2H_6O (99.2 per cent *m/m*), at 20 °C, calculated from the relative density using the alcoholimetric tables (*5.5*).

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

♦CHARACTERS

Appearance

Colourless, clear, volatile, flammable liquid, hygroscopic.

Solubility

Miscible with water and with methylene chloride.

It burns with a blue, smokeless flame.

bp

About 78 °C.♦

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Relative density (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of anhydrous ethanol.*

◊C. Mix 0.1 mL with 1 mL of a 10 g/L solution of *potassium permanganate R* and 0.2 mL of *dilute sulfuric acid R* in a test-tube. Cover immediately with a filter paper moistened with a freshly prepared solution containing 0.1 g of *sodium nitroprusside R* and 0.5 g of *piperazine hydrate R* in 5 mL of *water R*. After a few minutes, an intense blue colour appears on the paper and becomes paler after 10-15 min.

D. To 0.5 mL add 5 mL of *water R*, 2 mL of *dilute sodium hydroxide solution R*, then slowly add 2 mL of *0.05 M iodine*. A yellow precipitate is formed within 30 min.◊

TESTS

Appearance

It is clear (*2.2.1*) and colourless (*2.2.2, Method II*) when compared with *water R*. Dilute 1.0 mL to 20 mL with *water R*. After standing for 5 min, the dilution remains clear (*2.2.1*) when compared with *water R*.

Acidity or alkalinity

To 20 mL add 20 mL of *carbon dioxide-free water R* and 0.1 mL of *phenolphthalein solution R*. The solution is colourless. Add 1.0 mL of *0.01 M sodium hydroxide*. The solution is pink (30 ppm, expressed as acetic acid).

Relative density (*2.2.5*)

0.790 to 0.793.

Absorbance (*2.2.25*)

Maximum 0.40 at 240 nm, 0.30 between 250 nm and 260 nm, and 0.10 between 270 nm and 340 nm. The spectrum shows a steadily descending curve with no observable peaks or shoulders.

Examine between 235 nm and 340 nm in a 5 cm cell using *water R* as the compensation liquid.

Volatile impurities

Gas chromatography (*2.2.28*).

Test solution (a) The substance to be examined.

Test solution (b) Add 150 µL of *4-methylpentan-2-ol R* to 500.0 mL of the substance to be examined.

Reference solution (a) Dilute 100 µL of *anhydrous methanol R* to 50.0 mL with the substance to be examined. Dilute 5.0 mL of the solution to 50.0 mL with the substance to be examined.

Reference solution (b) Dilute 50 µL of *anhydrous methanol R* and 50 µL of *acetaldehyde R* to 50.0 mL with the substance to be examined. Dilute 100 µL of the solution to 10.0 mL with the substance to be examined.

Reference solution (c) Dilute 150 µL of *acetal R* to 50.0 mL with the substance to be examined. Dilute 100 µL of the solution to 10.0 mL with the substance to be examined.

Reference solution (d) Dilute 100 µL of *benzene R* to 100.0 mL with the substance to be examined. Dilute 100 µL of the solution to 50.0 mL with the substance to be examined.

Column:

— *material*: fused silica;
— *size*: l = 30 m, Ø = 0.32;
— *stationary phase*: *poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 1.8 µm).

Carrier gas *helium for chromatography R.*

Linear velocity 35 cm/s.

Split ratio 1:20.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 12	40
	12 - 32	40 → 240
	32 - 42	240
Injection port		200
Detector		280

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the 1st peak (acetaldehyde) and the 2nd peak (methanol).

Limits:

— *methanol* in the chromatogram obtained with test solution (a): not more than half the area of the corresponding peak in the chromatogram obtained with reference solution (a) (200 ppm *V/V*);
— *acetaldehyde + acetal*: maximum 10 ppm *V/V*, expressed as acetaldehyde.

Calculate the sum of the contents of acetaldehyde and acetal in parts per million *V/V* using the following expression:

$$\frac{10 \times A_E}{A_T - A_E} + \frac{30 \times C_E}{C_T - C_E} \times \frac{44.05}{118.2}$$

A_E = area of the peak due to acetaldehyde in the chromatogram obtained with test solution (a),
A_T = area of the peak due to acetaldehyde in the chromatogram obtained with reference solution (b),
C_E = area of the peak due to acetal in the chromatogram obtained with test solution (a),
C_T = area of the peak due to acetal in the chromatogram obtained with reference solution (c),
44.05 = molecular mass of acetaldehyde,
118.2 = molecular mass of acetal.

— *benzene*: maximum 2 ppm *V/V*.

Calculate the content of benzene in parts per million *V/V* using the following expression:

$$\frac{2B_E}{B_T - B_E}$$

B_E = area of the peak due to benzene in the chromatogram obtained with test solution (a),
B_T = area of the peak due to benzene in the chromatogram obtained with reference solution (d).

If necessary, the identity of benzene can be confirmed using another suitable chromatographic system (stationary phase with a different polarity).

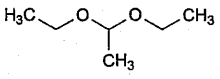
— *total of other impurities* in the chromatogram obtained with test solution (b): not more than the area of the peak due

to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (300 ppm);
— *disregard limit*: 0.03 times the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (9 ppm).

Residue on evaporation
Maximum 25 ppm *m/V*.
Evaporate 100 mL to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs a maximum of 2.5 mg.

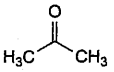
STORAGE
Protected from light.

◊IMPURITIES



A. 1,1-diethoxyethane (acetal),

B. acetaldehyde,

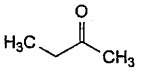


C. propan-2-one (acetone),

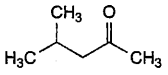
D. benzene,

E. cyclohexane,

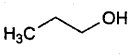
F. methanol,



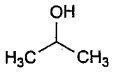
G. butan-2-one (methyl ethyl ketone),



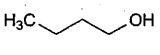
H. 4-methylpentan-2-one (methyl isobutyl ketone),



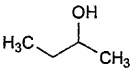
I. propan-1-ol (propanol),



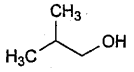
J. propan-2-ol (isopropyl alcohol),



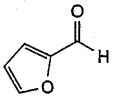
K. butan-1-ol (butanol),



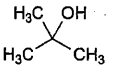
L. butan-2-ol,



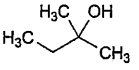
M. 2-methylpropan-1-ol (isobutanol),



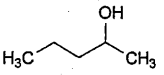
N. furane-2-carbaldehyde (furfural),



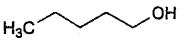
O. 2-methylpropan-2-ol (1,1-dimethylethyl alcohol),



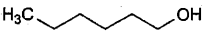
P. 2-methylbutan-2-ol,



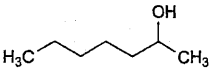
Q. pentan-2-ol,



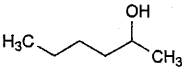
R. pentan-1-ol (pentanol),



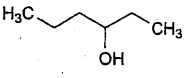
S. hexan-1-ol (hexanol),



T. heptan-2-ol,



U. hexan-2-ol,



V. hexan-3-ol.◊

Ethanol (96 per cent)[1]

Alcohol (96 per cent)

(Ph. Eur. monograph 1317)

Ph Eur

DEFINITION

Content

— *ethanol* (C_2H_6O; M_r 46.07): 95.1 per cent *V/V* (92.6 per cent *m/m*) to 96.9 per cent *V/V* (95.2 per cent *m/m*) at 20 °C, calculated from the relative density using the alcoholimetric tables (*5.5*);

— *water*.

♦CHARACTERS

Appearance

Colourless, clear, volatile, flammable liquid, hygroscopic.

Solubility

Miscible with water and with methylene chloride.

It burns with a blue, smokeless flame.

bp

About 78 °C.♦

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D.

A. Relative density (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of ethanol (96 per cent)*.

◊C. Mix 0.1 mL with 1 mL of a 10 g/L solution of *potassium permanganate R* and 0.2 mL of *dilute sulfuric acid R* in a test-tube. Cover immediately with a filter paper moistened with a freshly prepared solution containing 0.1 g of *sodium nitroprusside R* and 0.5 g of *piperazine hydrate R* in 5 mL of *water R*. After a few minutes, an intense blue colour appears on the paper and becomes paler after 10-15 min.

D. To 0.5 mL add 5 mL of *water R*, 2 mL of *dilute sodium hydroxide solution R*, then slowly add 2 mL of *0.05 M iodine*. A yellow precipitate is formed within 30 min.◊

TESTS

Appearance

It is clear (*2.2.1*) and colourless (*2.2.2, Method II*) when compared with *water R*. Dilute 1.0 mL to 20 mL with *water R*. After standing for 5 min, the dilution remains clear (*2.2.1*) when compared with *water R*.

Acidity or alkalinity

To 20 mL add 20 mL of *carbon dioxide-free water R* and 0.1 mL of *phenolphthalein solution R*. The solution is colourless. Add 1.0 mL of *0.01 M sodium hydroxide*. The solution is pink (30 ppm, expressed as acetic acid).

Relative density (*2.2.5*)

0.805 to 0.812.

Absorbance (*2.2.25*)

Maximum 0.40 at 240 nm, 0.30 between 250 nm and 260 nm, and 0.10 between 270 nm and 340 nm.

The spectrum shows a steadily descending curve with no observable peaks or shoulders.

Examine between 235 nm and 340 nm, in a 5 cm cell using *water R* as the compensation liquid.

Volatile impurities

Gas chromatography (*2.2.28*).

Test solution (a) The substance to be examined.

Test solution (b) Add 150 µL of *4-methylpentan-2-ol R* to 500.0 mL of the substance to be examined.

Reference solution (a) Dilute 100 µL of *anhydrous methanol R* to 50.0 mL with the substance to be examined. Dilute 5.0 mL of the solution to 50.0 mL with the substance to be examined.

Reference solution (b) Dilute 50 µL of *anhydrous methanol R* and 50 µL of *acetaldehyde R* to 50.0 mL with the substance to be examined. Dilute 100 µL of the solution to 10.0 mL with the substance to be examined.

Reference solution (c) Dilute 150 µL of *acetal R* to 50.0 mL with the substance to be examined. Dilute 100 µL of the solution to 10.0 mL with the substance to be examined.

Reference solution (d) Dilute 100 µL of *benzene R* to 100.0 mL with the substance to be examined. Dilute 100 µL of the solution to 50.0 mL with the substance to be examined.

Column:

— *material*: fused silica;

— *size*: *l* = 30 m, Ø = 0.32 mm;

— *stationary phase*: *poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 1.8 µm).

Carrier gas *helium for chromatography R*.

Linear velocity 35 cm/s.

Split ratio 1:20.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 12	40
	12 - 32	40 → 240
	32 - 42	240
Injection port		200
Detector		280

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the 1st peak (acetaldehyde) and the 2nd peak (methanol).

Limits:

— *methanol* in the chromatogram obtained with test solution (a): not more than half the area of the corresponding peak in the chromatogram obtained with reference solution (a) (200 ppm *V/V*);

— *acetaldehyde + acetal*: maximum 10 ppm *V/V*, expressed as acetaldehyde.

Calculate the sum of the contents of acetaldehyde and acetal in parts per million *V/V* using the following expression:

$$\frac{10 \times A_E}{A_T - A_E} + \frac{30 \times C_E}{C_T - C_E} \times \frac{44.05}{118.2}$$

A_E = area of the peak due to acetaldehyde in the chromatogram obtained with test solution (a),

A_T = area of the peak due to acetaldehyde in the chromatogram obtained with reference solution (b),

C_E = area of the peak due to acetal in the chromatogram obtained with test solution (a),

C_T = area of the peak due to acetal in the chromatogram obtained with reference solution (c),

44.05 = molecular mass of acetaldehyde,

118.2 = molecular mass of acetal.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

— *benzene*: maximum 2 ppm *V/V*.

Calculate the content of benzene in parts per million *V/V* using the following expression:

$$\frac{2B_E}{B_T - B_E}$$

B_E = area of the peak due to benzene in the chromatogram obtained with test solution (a),

B_T = area of the peak due to benzene in the chromatogram obtained with reference solution (d).

If necessary, the identity of benzene can be confirmed using another suitable chromatographic system (stationary phase with a different polarity).

— *total of other impurities* in the chromatogram obtained with test solution (b): not more than the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (300 ppm),

— *disregard limit*: 0.03 times the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (9 ppm).

Residue on evaporation

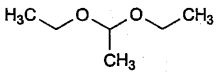
Maximum 25 ppm *m/V*.

Evaporate 100 mL to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs a maximum of 2.5 mg.

STORAGE

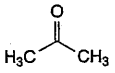
Protected from light.

◊IMPURITIES



A. 1,1-diethoxyethane (acetal),

B. acetaldehyde,

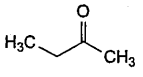


C. propan-2-one (acetone),

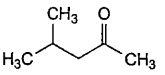
D. benzene,

E. cyclohexane,

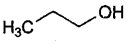
F. methanol,



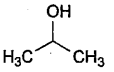
G. butan-2-one (methyl ethyl ketone),



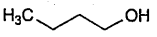
H. 4-methylpentan-2-one (methyl isobutyl ketone),



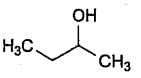
I. propan-1-ol (propanol),



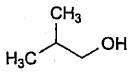
J. propan-2-ol (isopropyl alcohol),



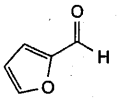
K. butan-1-ol (butanol),



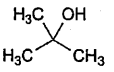
L. butan-2-ol,



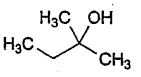
M. 2-methylpropan-1-ol (isobutanol),



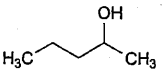
N. furane-2-carbaldehyde (furfural),



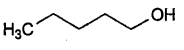
O. 2-methylpropan-2-ol (1,1-dimethylethyl alcohol),



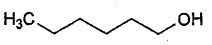
P. 2-methylbutan-2-ol,



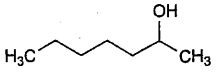
Q. pentan-2-ol,



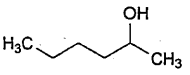
R. pentan-1-ol (pentanol),



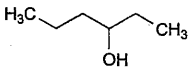
S. hexan-1-ol (hexanol),



T. heptan-2-ol,



U. hexan-2-ol,



V. hexan-3-ol.◊

Ph Eur

Dilute Ethanols

DEFINITION

The official Dilute Ethanols contain 90, 80, 70, 60, 50, 45, 25 and 20% v/v respectively of ethanol. They may be prepared as described below, the final adjustment of volume being made at the same temperature, 20°, as that at which the Ethanol (96 per cent) is measured.

NOTE *On mixing ethanol and water, contraction of volume and rise of temperature occur.*

TESTS

Acidity or alkalinity; Appearance; Volatile impurities; Residue on evaporation

Comply with the requirements stated under Ethanol (96 per cent).

ETHANOL (90 PER CENT)

Alcohol (90 per cent); Rectified Spirit

Dilute 934 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

89.6 to 90.5% v/v.

Apparent density

826.4 to 829.4 kg m^{-3}, Appendix V G.

ETHANOL (80 PER CENT)

Alcohol (80 per cent)

Dilute 831 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

79.5 to 80.3% v/v.

Apparent density

857.4 to 859.6 kg m^{-3}, Appendix V G.

ETHANOL (70 PER CENT)

Alcohol (70 per cent)

Dilute 727 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

69.5 to 70.4% v/v.

Apparent density

883.5 to 885.8 kg m^{-3}, Appendix V G.

ETHANOL (60 PER CENT)

Alcohol (60 per cent)

Dilute 623 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

59.7 to 60.2% v/v.

Apparent density

907.6 to 908.7 kg m^{-3}, Appendix V G.

ETHANOL (50 PER CENT)

Alcohol (50 per cent)

Dilute 519 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

49.6 to 50.2% v/v.

Apparent density

928.6 to 929.8 kg m^{-3}, Appendix V G.

ETHANOL (45 PER CENT)

Alcohol (45 per cent)

Dilute 468 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

44.7 to 45.3% v/v.

Apparent density

938.0 to 939.0 kg m^{-3}, Appendix V G.

ETHANOL (25 PER CENT)

Alcohol (25 per cent)

Dilute 259 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

24.6 to 25.4% v/v.

Apparent density

966.6 to 967.5 kg m^{-3}, Appendix V G.

ETHANOL (20 PER CENT)

Alcohol (20 per cent)

Dilute 207 ml of Ethanol (96 per cent) to 1000 ml with Purified Water.

Content of ethanol

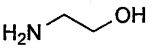
19.5 to 20.5% v/v.

Apparent density

972.0 to 973.1 kg m^{-3}, Appendix V G.

Ethanolamine

Monoethanolamine



C_2H_7NO 61.08 *141-43-5*

Action and use

Sclerosant.

Preparation

Ethanolamine Oleate Injection

DEFINITION

Ethanolamine is 2-aminoethanol. It contains not less than 98.0% and not more than 100.5% of C_2H_7NO.

CHARACTERISTICS

A clear, colourless or pale yellow liquid.

Miscible with *water* and with *ethanol (96%)*; slightly soluble in *ether*.

IDENTIFICATION

A. To 0.1 mL add 0.3 g of *picric acid* and 1 mL of *water* and evaporate to dryness on a water bath. The *melting point* of the residue, after recrystallisation from *ethanol (96%)* and drying at 105°, is about 160°, Appendix V A.

B. Distil, collect the second half of the distillate and allow to cool to room temperature. The second half of the distillate freezes at about 10°.

C. It is alkaline to *litmus solution*.

TESTS

Refractive index

1.453 to 1.459, Appendix V E.

Weight per mL
1.014 to 1.023 g, Appendix V G.

Related substances
Carry out the method for *gas chromatography*, Appendix III B. Prepare a 0.1% w/v solution of *3-aminopropan-1-ol* (internal standard) in *dichloromethane* (solution A).

(1) 0.05% w/v of *ethanolamine* and 0.1% w/v each of *diethanolamine* and *triethanolamine* in solution A. To 0.5 mL of this solution add 0.5 mL of *trifluoroacetic anhydride*, mix and allow to stand for 10 minutes.

(2) 10% w/v solution of the substance being examined in solution A. To 0.5 mL of this solution add 0.5 mL of *trifluoroacetic anhydride*, mix and allow to stand for 10 minutes.

CHROMATOGRAPHIC CONDITIONS

(a) Use a fused silica capillary column (25 m × 0.22 mm) bonded with a 0.25-µm layer of dimethylpolysiloxane.

(b) Use *helium* as the carrier gas at 1.0 mL per minute with a flow rate of the make up gas of 20 mL per minute.

(c) Maintain the temperature of the column at 80° for 2 minutes, then increase to 200° at a rate of 8° per minute and maintain this temperature for 10 minutes.

(d) Use an inlet temperature of 240°.

(e) Use a flame ionisation detector at a temperature of 250°.

(f) Inject 1 µL of each solution.

(g) Use a split ratio of 1:40.

In the chromatogram obtained with solution (1):

the peaks eluting after the solvent peak in order of emergence are due to (a) ethanolamine, (b) 3-aminopropan-1-ol, (c) diethanolamine and (d) triethanolamine.

LIMITS

In the chromatogram obtained with solution (2) calculate the content of diethanolamine and triethanolamine using the ratios of the peaks and by reference to the corresponding peaks in the chromatogram obtained with solution (1):

the content of diethanolamine is not more than 1.0% w/w,

the content of triethanolamine is not more than 1.0% w/w,

In the chromatogram obtained with solution (2), calculate the content of any other impurity using the ratios of the peaks and by reference to the peak due to ethanolamine:

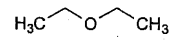
the content of any other impurity is not more than 0.5% w/w;

the sum of the contents of all the impurities is not more than 2.0% w/w.

ASSAY
Dissolve 2.5 g in 50 mL of *1M hydrochloric acid VS* and titrate the excess of acid with *1M sodium hydroxide VS* using *methyl red solution* as indicator. Each mL of *1M hydrochloric acid VS* is equivalent to 61.08 mg of C_2H_7NO.

Ether

(Ph. Eur. monograph 0650)



$C_4H_{10}O$ 74.1 *60-29-7*

Ph Eur

DEFINITION
Diethyl ether.

It may contain a suitable non-volatile antioxidant at a suitable concentration.

CHARACTERS

Appearance
Clear, colourless, volatile liquid.

Solubility
Soluble in water, miscible with ethanol (96 per cent), with methylene chloride and with fatty oils.

It is highly flammable.

IDENTIFICATION
A. Relative density (see Tests).

B. Distillation range (see Tests).

TESTS

Acidity
To 20 mL of *ethanol (96 per cent) R* add 0.25 mL of *bromothymol blue solution R1* and, dropwise, *0.02 M sodium hydroxide* until a blue colour persists for 30 s. Add 25 mL of the substance to be examined, shake and add, dropwise, *0.02 M sodium hydroxide* until the blue colour reappears and persists for 30 s. Not more than 0.4 mL of *0.02 M sodium hydroxide* is required.

Relative density *(2.2.5)*
0.714 to 0.716.

Distillation range *(2.2.11)*
Do not distil if the substance to be examined does not comply with the test for peroxides It distils completely between 34.0 °C and 35.0 °C. Carry out the test using a suitable heating device and taking care to avoid directly heating the flask above the level of the liquid.

Aldehydes
To 10.0 mL in a ground-glass-stoppered cylinder add 1 mL of *alkaline potassium tetraiodomercurate solution R* and shake for 10 s. Allow to stand for 5 min, protected from light. The lower layer may show a yellow or reddish-brown opalescence but not a grey or black opalescence.

Peroxides
Place 8 mL of *potassium iodide and starch solution R* in a 12 mL ground-glass-stoppered cylinder about 15 mm in diameter. Fill completely with the substance to be examined, mix and allow to stand protected from light for 5 min. No colour develops.

Non-volatile matter
Maximum 20 mg/L.

After ensuring that the substance to be examined complies with the test for peroxides Evaporate 50 mL to dryness on a water-bath and dry the residue in an oven at 100-105 °C. The residue weighs a maximum of 1 mg.

Substances with a foreign odour
Moisten a disc of filter paper 80 mm in diameter with 5 mL of the substance to be examined and allow to evaporate.

No foreign odour is perceptible immediately after the evaporation.

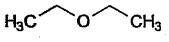
Water (*2.5.12*)
Maximum 2 g/L, determined on 20 mL.

STORAGE
In an airtight container, protected from light.

Ph Eur

Anaesthetic Ether

(Ph. Eur. monograph 0367)



$C_4H_{10}O$ 74.1 *60-29-7*

Ph Eur

DEFINITION
Diethyl ether.

It may contain a suitable non-volatile antioxidant at an appropriate concentration.

CHARACTERS
Appearance
Clear, colourless liquid, volatile, very mobile.

Solubility
Soluble in 15 parts of water, miscible with ethanol (96 per cent) and with fatty oils.

It is highly flammable.

IDENTIFICATION
A. Relative density (see Tests).

B. Distillation range (see Tests).

TESTS
Acidity
To 20 mL of *ethanol (96 per cent) R* add 0.25 mL of *bromothymol blue solution R1* and, dropwise, *0.02 M sodium hydroxide* until a blue colour persists for 30 s. Add 25 mL of the substance to be examined, shake and add, dropwise, *0.02 M sodium hydroxide* until the blue colour reappears and persists for 30 s. Not more than 0.4 mL of *0.02 M sodium hydroxide* is required.

Relative density (*2.2.5*)
0.714 to 0.716.

Distillation range (*2.2.11*)
Do not distil if the substance to be examined does not comply with the test for peroxides It distils completely between 34.0 °C and 35.0 °C. Carry out the test using a suitable heating device and taking care to avoid directly heating the flask above the level of the liquid.

Acetone and aldehydes
To 10.0 mL in a ground-glass-stoppered cylinder add 1 mL of *alkaline potassium tetraiodomercurate solution R* and shake for 10 s. Allow to stand for 5 min, protected from light. The lower layer shows only a slight opalescence.

If the substance to be examined does not comply with the test, distil 40 mL, *after ensuring that the substance to be examined complies with the test for peroxides*, until only 5 mL remains. Collect the distillate in a receiver cooled in a bath of iced water and repeat the test described above using 10.0 mL of the distillate.

Peroxides
Place 8 mL of *potassium iodide and starch solution R* in a 12 mL ground-glass-stoppered cylinder about 15 mm in diameter. Fill completely with the substance to be examined, shake vigorously and allow to stand protected from light for 30 min. No colour develops.

Non-volatile matter
Maximum 20 mg/L.

After ensuring that the substance to be examined complies with the test for peroxides Evaporate 50 mL to dryness on a water-bath and dry the residue in an oven at 100-105 °C. The residue weighs a maximum of 1 mg.

Substances with a foreign odour
Moisten a disc of filter paper 80 mm in diameter with 5 mL of the substance to be examined and allow to evaporate. No foreign odour is perceptible immediately after the evaporation.

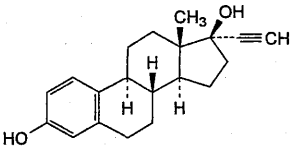
Water (*2.5.12*)
Maximum 2 g/L, determined on 20 mL.

STORAGE
In an airtight container, protected from light. The contents of a partly filled container may deteriorate rapidly.

Ph Eur

Ethinylestradiol

(Ph. Eur. monograph 0140)



$C_{20}H_{24}O_2$ 296.4 *57-63-6*

Action and use
Estrogen.

Preparations
Co-cyprindiol Tablets

Ethinylestradiol Tablets

Levonorgestrel and Ethinylestradiol Tablets

Ph Eur

DEFINITION
19-Nor-17α-pregna-1,3,5(10)-trien-20-yne-3,17-diol.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellowish-white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent). It dissolves in dilute alkaline solutions.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison ethinylestradiol CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference

substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (10:90 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 25 mL with the solvent mixture.

Reference solution Dissolve 25 mg of *ethinylestradiol CRS* in the solvent mixture and dilute to 25 mL with the solvent mixture.

Plate *TLC silica gel G plate R.*

Mobile phase *ethanol (96 per cent) R, toluene R* (10:90 *V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air until the solvent has evaporated.

Detection Heat at 110 °C for 10 min, spray the hot plate with *alcoholic solution of sulfuric acid R* and heat again at 110 °C for 10 min. Examine in daylight and in ultraviolet light at 365 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour, fluorescence and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *water R, acetonitrile R1* (40:60 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in 30 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of *estrone CRS* (impurity C) in 10.0 mL of the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture. Use 1.0 mL of this solution to dissolve the contents of a vial of *ethinylestradiol for system suitability CRS* (containing impurities B, F, H, I and K).

Reference solution (c) Dissolve 50.0 mg of *ethinylestradiol CRS* in 30 mL of *acetonitrile R1* and dilute to 50.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R1, water R* (30:70 *V/V*);
- *mobile phase B*: *water R, acetonitrile R1* (25:75 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 35	100	0
35 - 65	100 → 0	0 → 100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 30 µL of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *ethinylestradiol for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, F, H, I and K.

Relative retention With reference to ethinylestradiol (retention time = about 35 min): impurity F = about 0.2; impurity H = about 0.5; impurity I = about 0.8; impurity B = about 0.88; impurity C = about 0.92; impurity K = about 1.3.

System suitability Reference solution (b):
- *resolution*: minimum 1.2 between the peaks due to impurities I and B.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.7; impurity I = 0.4;
- *impurity B*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurities H, I, K*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurities C, F*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{20}H_{24}O_2$ from the declared content of *ethinylestradiol CRS*.

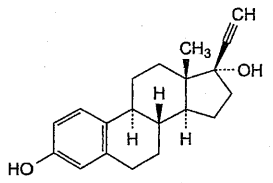
STORAGE

Protected from light.

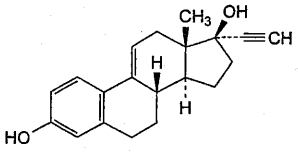
IMPURITIES

Specified impurities *B, C, F, H, I, K.*

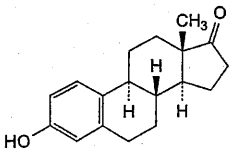
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, G, J, L, M.*



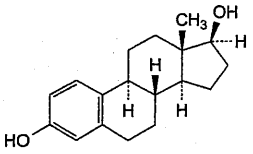
A. 19-norpregna-1,3,5(10)-trien-20-yne-3,17-diol (17β-ethinylestradiol),



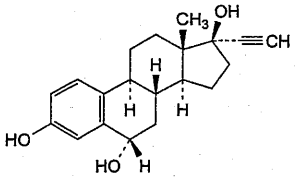
B. 19-nor-17α-pregna-1,3,5(10),9(11)-tetraen-20-yne-3,17-diol,



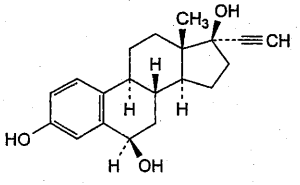
C. 3-hydroxyestra-1,3,5(10)-trien-17-one (estrone),



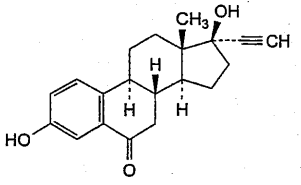
D. estra-1,3,5(10)-triene-3,17β-diol (estradiol),



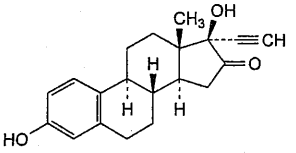
E. 19-nor-17α-pregna-1,3,5(10)-trien-20-yne-3,6α,17-triol (6α-hydroxy-ethinylestradiol),



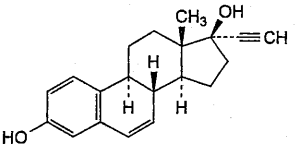
F. 19-nor-17α-pregna-1,3,5(10)-trien-20-yne-3,6β,17-triol (6β-hydroxy-ethinylestradiol),



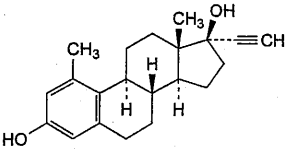
G. 3,17-dihydroxy-19-nor-17α-pregna-1,3,5(10)-trien-20-yn-6-one (6-oxo-ethinylestradiol),



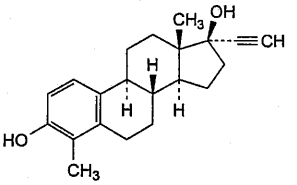
H. 3,17-dihydroxy-19-nor-17α-pregna-1,3,5(10)-trien-20-yn-16-one (16-oxo-ethinylestradiol),



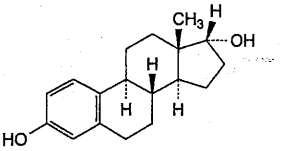
I. 19-nor-17α-pregna-1,3,5(10),6-tetraen-20-yne-3,17-diol,



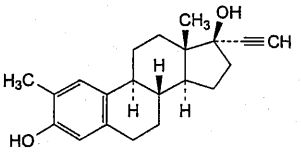
J. 1-methyl-19-nor-17α-pregna-1,3,5(10)-trien-20-yne-3,17-diol (1-methyl-ethinylestradiol),



K. 4-methyl-19-nor-17α-pregna-1,3,5(10)-trien-20-yne-3,17-diol (4-methyl-ethinylestradiol),



L. estra-1,3,5(10)-triene-3,17α-diol (17α-estradiol),

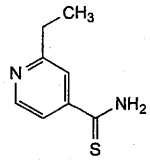


M. 2-methyl-19-nor-17α-pregna-1,3,5(10)-trien-20-yne-3,17-diol (2-methyl-ethinylestradiol).

Ph Eur

Ethionamide

(Ph. Eur. monograph 0141)



$C_8H_{10}N_2S$ 166.2 *536-33-4*

Action and use
Antituberculosis drug.

Ph Eur

DEFINITION
2-Ethylpyridine-4-carbothioamide.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance Yellow, crystalline powder or small, yellow crystals.

Solubility Practically insoluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 158 °C to 164 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 10.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with *methanol R*.

Spectral range 230-350 nm.

Absorption maximum At 290 nm.

Specific absorbance at the absorption maximum 380 to 440.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison: ethionamide CRS.

D. Dissolve about 10 mg in 5 mL of *methanol R*. Add 5 mL of *silver nitrate solution R2*. A dark brown precipitate is formed.

TESTS
Acidity
Dissolve 2.0 g in 20 mL of *methanol R*, heating to about 50 °C, and add 20 mL of *water R*. Cool slightly while shaking until crystallisation begins and then allow to cool to room temperature. Add 60 mL of *water R* and 0.2 mL of *cresol red solution R*. Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to red.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Dissolve 0.240 g of *ammonium dihydrogen phosphate R* in 700 mL of *water R* and add 300 mL of *acetonitrile R*.

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *pyridine-4-carbonitrile R* (impurity B) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *ethionamide for system suitability CRS* (containing impurity A) in reference solution (b) and dilute to 5.0 mL with reference solution (b).

Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.287 g of *ammonium dihydrogen phosphate R* in 850 mL of *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	85	15
30 - 32	85 → 70	15 → 30
32 - 55	70	30

Flow rate 1 mL/min.

Detection Spectrophotometer at 287 nm.

Injection 5 µL of the test solution and reference solutions (a) and (c).

Identification of impurities Use the chromatogram supplied with *ethionamide for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to ethionamide (retention time = about 9 min): impurity A = about 0.4; impurity B = about 0.5.

System suitability Reference solution (c):
— *resolution*: minimum 3.0 between the peaks due to impurities A and B.

Calculation of percentage contents:
— for each impurity, use the concentration of ethionamide in reference solution (a).

Limits:
— *impurity A*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

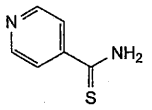
ASSAY
Dissolve 0.150 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 16.62 mg of $C_8H_{10}N_2S$.

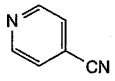
IMPURITIES

Specified impurities *A.*

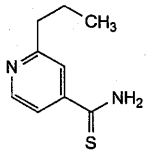
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F.*



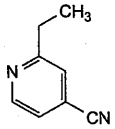
A. pyridine-4-carbothioamide,



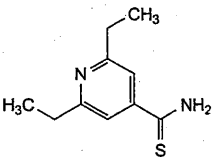
B. pyridine-4-carbonitrile,



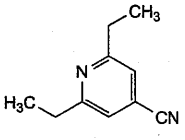
C. 2-propylpyridine-4-carbothioamide (prothionamide),



D. 2-ethylpyridine-4-carbonitrile,



E. 2,6-diethylpyridine-4-carbothioamide,

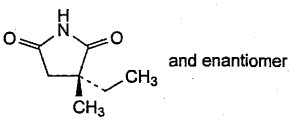


F. 2,6-diethylpyridine-4-carbonitrile.

Ph Eur

Ethosuximide

(Ph. Eur. monograph 0764)



$C_7H_{11}NO_2$ 141.2 *77-67-8*

Action and use

Antiepileptic.

Preparations

Ethosuximide Capsules

Ethosuximide Oral Solution

Ph Eur

DEFINITION

(3*RS*)-3-Ethyl-3-methylpyrrolidine-2,5-dione.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, powder or waxy solid.

Solubility

Freely soluble in water, very soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, B.

Second identification: A, C.

A. Melting point (*2.2.14*): 45 °C to 50 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ethosuximide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *methanol R.*

Reference solution Dissolve 20 mg of *ethosuximide CRS* in 1 mL of *methanol R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (17:17:66 *V/V/V*).

Application 5 µL.

Development Over 3/4 of the plate.

Drying At 100-105 °C for 10 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 2.5 g in *water R* and dilute to 25 mL with the same solvent.

Cyanide

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.50 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.125 g of *potassium cyanide R* in *water R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *water R*. Dilute 0.5 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 0.50 g of the substance to be examined in *water R*, add 0.5 mL of reference solution (a) and dilute to 10.0 mL with *water R*.

Column:
- *size*: l = 0.075 m, Ø = 7.5 mm,
- *stationary phase*: spherical *weak anion-exchange resin R* (10 µm).

Mobile phase Dissolve 2.1 g of *lithium hydroxide R* and 85 mg of *sodium edetate R* in *water for chromatography R*, and dilute to 1000 mL with the same solvent.

Flow rate 2.0 mL/min.

Detection Electrochemical detector (direct amperometry) with a silver working electrode, a silver-silver chloride reference electrode, held at + 0.05 V oxidation potential, and a detector sensitivity of 20 nA full scale.

Injection 20 µL of the test solution and reference solution (b).

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to cyanide and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to ethosuximide.

Limit:
- *cyanide*: not more than 0.5 times the height of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 ppm).

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.250 g of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A. Store the solution at room temperature for at least 30 min before injection (*in situ* transformation of impurity B to impurity A).

Reference solution (a) Dissolve 5.0 mg of *ethosuximide impurity A CRS* in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 20.0 mL with mobile phase A.

Reference solution (c) Mix 1 mL of reference solution (a) and 4 mL of the test solution.

Column:
- *size*: l = 0.10 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped solid core octadecylsilyl organosilica polymer compatible with 100 per cent aqueous mobile phases R* (2.6 µm);
- *temperature*: 25 °C.

Mobile phase:
- *mobile phase A*: 15.6 g/L solution of *sodium dihydrogen phosphate R* previously adjusted to pH 2.0 with *phosphoric acid R*;
- *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	90	10
10 - 11	90 → 30	10 → 70
11 - 15	30	70

Flow rate 0.25 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 3 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to ethosuximide (retention time = about 4 min): impurity A = about 1.7.

System suitability Reference solution (c):
- *resolution*: minimum 3.0 between the peaks due to ethosuximide and impurity A.

Calculation of percentage contents:
- for each impurity, use the concentration of impurity A in reference solution (b).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.2 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.120 g in 20 mL of *dimethylformamide R* and carry out a potentiometric titration (*2.2.20*) using *0.1 M tetrabutylammonium hydroxide*. Protect the solution from atmospheric carbon dioxide throughout the titration. Carry out a blank titration.

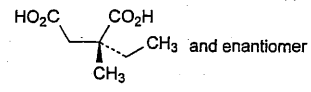
1 mL of *0.1 M tetrabutylammonium hydroxide* is equivalent to 14.12 mg of $C_7H_{11}NO_2$.

STORAGE

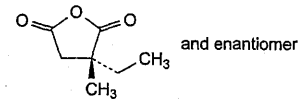
Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B.*



A. (2*RS*)-2-ethyl-2-methylbutanedioic acid,

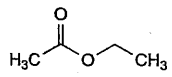


B. (3*RS*)-3-ethyl-3-methyldihydrofuran-2,5-dione.

Ph Eur

Ethyl Acetate

(Ph. Eur. monograph 0899)



$C_4H_8O_2$ 88.1 *141-78-6*

Action and use
Excipient.

Ph Eur

DEFINITION
Ethyl ethanoate.

CHARACTERS

Appearance
Clear, colourless, volatile liquid.

Solubility
Soluble in water, miscible with acetone, with ethanol (96 per cent) and with methylene chloride.

IDENTIFICATION
First identification: B.
Second identification: A, C, D.
A. Boiling point (*2.2.12*): 76 °C to 78 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of ethyl acetate.*
C. It gives the reaction of acetyl (*2.3.1*).
D. It gives the reaction of esters (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).
Mix 1 mL of the substance to be examined and 15 mL of *water R*.

Acidity
To 10 mL of *ethanol (96 per cent) R* add 0.1 mL of *phenolphthalein solution R* and *0.01 M sodium hydroxide* until the colour changes to pink. Add 5.5 mL of the substance to be examined and 0.25 mL of *0.02 M sodium hydroxide*. The solution remains pink for not less than 15 s.

Relative density (*2.2.5*)
0.898 to 0.902.

Refractive index (*2.2.6*)
1.370 to 1.373.

Reaction with sulfuric acid
Carefully add 2 mL to 10 mL of *sulfuric acid R*. After 15 min, the interface between the 2 liquids is not coloured.

Related substances
Gas chromatography (*2.2.28*).
Test solution The substance to be examined.
Column:
— *material:* glass;
— *size:* *l* = 2 m, Ø = 2 mm;
— *stationary phase: ethylvinylbenzene-divinylbenzene copolymer R* (136-173 µm).
Carrier gas *nitrogen for chromatography R.*
Flow rate 30 mL/min.
Temperature:

	Time (min)	Temperature (°C)
Column	0 - 18.8	90 → 240
	18.8 - 26.8	240
Injection port		240
Detector		240

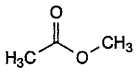
Detection Flame ionisation.
Injection 1 µL.
Limit:
— *total:* not more than 0.2 per cent of the area of the principal peak.

Residue on evaporation
Maximum 30 ppm.
Evaporate 100.0 g to dryness on a water-bath and dry in an oven at 100-105 °C. The residue weighs not more than 3 mg.

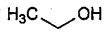
Water (*2.5.12*)
Maximum 0.1 per cent, determined on 10.0 mL.

STORAGE
Protected from light, at a temperature not exceeding 30 °C.

IMPURITIES



A. methyl ethanoate (methyl acetate),

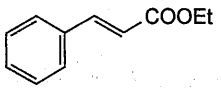


B. ethanol,

C. methanol.

Ph Eur

Ethyl Cinnamate



$C_{11}H_{12}O_2$ 176.2 *103-36-6*

DEFINITION
Ethyl Cinnamate is predominantly ethyl (*E*)-3-phenylprop-2-enoate. It contains not less than 99.0% and not more than 100.5% of $C_{11}H_{12}O_2$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A clear, colourless or almost colourless liquid.
Practically insoluble in *water*; miscible with most organic solvents.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of ethyl cinnamate *(RS 136)*.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.001% w/v solution in *ethanol (96%)* exhibits a maximum only at 276 nm. The *absorbance* at the maximum is about 1.23.

C. To 1 g add 25 mL of 1M *sodium hydroxide*, boil under a reflux condenser for 1 hour, cool and acidify with *hydrochloric acid*. The *melting point* of the resulting precipitate, after filtration, washing with *water* and drying at 60° at a pressure not exceeding 0.7 kPa, is about 133°, Appendix V A.

TESTS

Acidity
Mix 30 g with 150 mL of *ethanol (96%)* previously neutralised to *phenolphthalein solution R1*. Not more than 1.0 mL of *0.1M sodium hydroxide VS* is required for neutralisation using *phenolphthalein solution R1* as indicator.

Refractive index
1.558 to 1.560, Appendix V E.

Weight per mL
1.048 to 1.051 g, Appendix V G.

Related substances
Carry out the method for *gas chromatography*, Appendix III B, using the following solutions.

(1) Use the substance being examined.

(2) 1.0% w/v of the substance being examined in *chloroform*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with *acid-washed, silanised diatomaceous support* coated with 3% w/w of cyanopropylmethyl phenyl methyl silicone fluid (OV-225 is suitable).

(b) Use *helium* as the carrier gas at 1.7 mL per minute.

(c) Use isothermal conditions maintained at 150°.

(d) Use an inlet temperature of 150°.

(e) Use a flame ionisation detector at a temperature of 150°.

(f) Inject 1 µL of each solution.

LIMITS

In the chromatogram obtained with solution (1) the sum of the areas of any *secondary peaks* is not greater than 1% by *normalisation*.

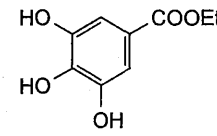
Sulfated ash
Not more than 0.1%, Appendix IX A.

Water
Not more than 0.1% w/w, Appendix IX C. Use 5 g.

ASSAY

In a borosilicate glass flask dissolve 2.5 g of the substance being examined in 5 mL of carbon dioxide-free ethanol prepared by boiling *ethanol (96%)* thoroughly and neutralising to *phenolphthalein solution R1*. Neutralise the free acid in the solution with *0.1M ethanolic potassium hydroxide VS* using 0.2 mL of *phenolphthalein solution R1* as indicator. Add 50 mL of *0.5M ethanolic potassium hydroxide VS* and boil under a reflux condenser on a water bath for 1 hour. Add 20 mL of *water* and titrate the excess of alkali with *0.5M hydrochloric acid VS* using a further 0.2 mL of *phenolphthalein solution R1* as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the alkali required to saponify the esters. Each mL of *0.5M ethanolic potassium hydroxide VS* is equivalent to 88.11 mg of $C_{11}H_{12}O_2$.

Ethyl Gallate



$C_9H_{10}O_5$ 198.2 *831-61-8*

Action and use
Antioxidant.

DEFINITION

Ethyl Gallate is ethyl 3,4,5-trihydroxybenzoate.

CHARACTERISTICS

A white to creamy white, crystalline powder.

Slightly soluble in *water*; freely soluble in *ethanol (96%)* and in *ether*; practically insoluble in arachis oil.

IDENTIFICATION

A. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in *methanol* exhibits a maximum only at 275 nm. The *absorbance* at the maximum is about 1.08.

B. Carry out the method for *gas chromatography*, Appendix III B using the following solutions.

(1) Boil 0.5 g with 50 mL of 5M *sodium hydroxide* under a reflux condenser for 10 minutes and distil 5 mL.

(2) *Absolute ethanol*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with *acid-washed, silanised diatomaceous support* (80 to 100 mesh) coated with 10% w/w of free fatty acid phase (Supelco FFAP is suitable).

(b) Use *nitrogen* as the carrier gas at 40 mL per minute.

(c) Use isothermal conditions maintained at 80°.

(d) Use a flame ionisation detector.

(e) Inject 1 µL of each solution.

CONFIRMATION

The chromatogram obtained with solution (1) shows a peak with the same retention time as the peak due to absolute ethanol in the chromatogram obtained with solution (2).

C. Dissolve 5 mg in a mixture of 25 mL of *acetone* and 25 mL of *water* and add 0.05 mL of *iron(III) chloride solution*. A purplish black colour is produced which rapidly becomes bluish black.

TESTS

Melting point
151° to 154°, Appendix V A.

Acidity
Dissolve 0.4 g in 100 mL of warm *carbon dioxide-free water*, cool and titrate with *0.1M sodium hydroxide VS* using *bromocresol green solution* as indicator. Not more than 0.1 mL of *0.1M sodium hydroxide VS* is required.

Chloride
Shake 0.50 g with 50 mL of *water* for 5 minutes and filter. 15 mL of the resulting solution complies with the *limit test for chlorides*, Appendix VII (330 ppm).

Loss on drying
When dried to constant weight at 105°, loses not more than 1.0% of its weight. Use 1 g.

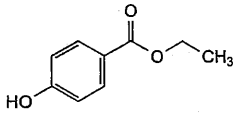
Sulfated ash
Not more than 0.1%, Appendix IX A.

STORAGE
Ethyl Gallate should be protected from light. Contact with metals should be avoided.

Ethyl Hydroxybenzoate

Ethylparaben

(Ethyl Parahydroxybenzoate, Ph. Eur. monograph 0900)



$C_9H_{10}O_3$ 166.2 *120-47-8*

Ph Eur

DEFINITION
Ethyl 4-hydroxybenzoate.

Content
98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION
First identification: A, B.
Second identification: A, C.
A. Melting point (*2.2.14*): 115 °C to 118 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *ethyl parahydroxybenzoate CRS.*
C. Thin-layer chromatography (*2.2.27*).
Test solution (a) Dissolve 0.10 g of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.
Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *acetone R.*
Reference solution (a) Dissolve 10 mg of *ethyl parahydroxybenzoate CRS* in *acetone R* and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of *methyl parahydroxybenzoate R* in 1 mL of test solution (a) and dilute to 10 mL with *acetone R.*
Plate *TLC octadecylsilyl silica gel F_{254} plate R.*
Mobile phase *glacial acetic acid R, water R, methanol R* (1:30:70 *V/V/V*).
Application 2 µL of test solution (b) and reference solutions (a) and (b).
Development Over 2/3 of the plate.
Drying In air.
Detection Examine in ultraviolet light at 254 nm.
System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated principal spots.
Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Solution S
Dissolve 1.0 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity
To 2 mL of solution S add 3 mL of *ethanol (96 per cent) R*, 5 mL of *carbon dioxide-free water R* and 0.1 mL of *bromocresol green solution R.* Not more than 0.1 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to blue.

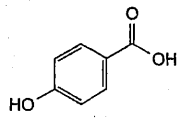
Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 5 mg of *4-hydroxybenzoic acid R* (impurity A), 5 mg of *methyl parahydroxybenzoate R* (impurity B) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 50.0 mg of *ethyl parahydroxybenzoate CRS* in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).
Mobile phase 6.8 g/L solution of *potassium dihydrogen phosphate R, methanol R* (35:65 *V/V*).
Flow rate 1.3 mL/min.
Detection Spectrophotometer at 272 nm.
Injection 10 µL of the test solution and reference solutions (a) and (c).
Run time 4 times the retention time of ethyl parahydroxybenzoate.
Relative retention With reference to ethyl parahydroxybenzoate (retention time = about 3.0 min): impurity A = about 0.5; impurity B = about 0.8.
System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity B and ethyl parahydroxybenzoate.
Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

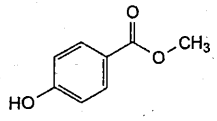
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.
Injection Test solution and reference solution (b).
Calculate the percentage content of $C_9H_{10}O_3$ from the declared content of *ethyl parahydroxybenzoate CRS*.

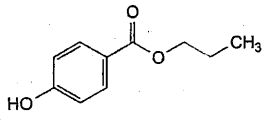
IMPURITIES
Specified impurities *A.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D.*



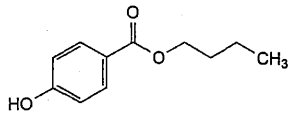
A. 4-hydroxybenzoic acid,



B. methyl 4-hydroxybenzoate (methyl parahydroxybenzoate),



C. propyl 4-hydroxybenzoate (propyl parahydroxybenzoate),



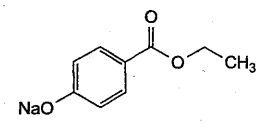
D. butyl 4-hydroxybenzoate (butyl parahydroxybenzoate).

Ph Eur

Ethyl Hydroxybenzoate Sodium

Ethylparaben Sodium

(Sodium Ethyl Parahydroxybenzoate, Ph. Eur. monograph 2134)



$C_9H_9NaO_3$ 188.2 *35285-68-8*

Ph Eur

DEFINITION
Sodium 4-(ethoxycarbonyl)phenolate.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water, soluble in anhydrous ethanol, practically insoluble in methylene chloride.

IDENTIFICATION
First identification: B, D.
Second identification: A, C, D.
A. Dissolve 0.5 g in 50 mL of *water R*. Immediately add 5 mL of *hydrochloric acid R1*. Filter and wash the precipitate with *water R*. Dry *in vacuo* at 80 °C for 2 h. It melts (*2.2.14*) at 115 °C to 118 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Preparation The precipitate obtained in identification A.
Comparison *ethyl parahydroxybenzoate CRS*.
C. Thin-layer chromatography (*2.2.27*).
Test solution (a) Dissolve 0.100 g of the substance to be examined in 10 mL of *water R*. Immediately add 2 mL of *hydrochloric acid R* and shake with 50 mL of *methylene chloride R*. Evaporate the lower layer to dryness and take up the residue with 10 mL of *acetone R*.
Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *acetone R*.
Reference solution (a) Dissolve 5 mg of *ethyl parahydroxybenzoate CRS* in *acetone R* and dilute to 5 mL with the same solvent.
Reference solution (b) Dissolve 5 mg of *methyl parahydroxybenzoate CRS* (impurity B) in 0.5 mL of test solution (a) and dilute to 5 mL with *acetone R*.
Plate *TLC octadecylsilyl silica gel F_{254} plate R.*
Mobile phase *glacial acetic acid R, water R, methanol R* (1:30:70 *V/V/V*).
Application 5 µL.
Development Over 2/3 of the plate.
Drying In air.
Detection Examine in ultraviolet light at 254 nm.
System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated principal spots.
Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 1 mL of solution S (see Tests) add 1 mL of *water R*. The solution gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S, examined immediately after preparation, is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)

9.5 to 10.5.

Dilute 1 mL of solution S to 100 mL with *carbon dioxide-free water R*.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *4-hydroxybenzoic acid R* (impurity A), 5 mg of *methyl parahydroxybenzoate R* (impurity B) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of *ethyl parahydroxybenzoate CRS* in 2.5 mL of *methanol R* and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 6.8 g/L solution of *potassium dihydrogen phosphate R*, *methanol R* (35:65 *V/V*).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Run time 4 times the retention time of ethyl parahydroxybenzoate.

Relative retention With reference to ethyl parahydroxybenzoate (retention time = about 3 min): impurity A = about 0.5; impurity B = about 0.8.

System suitability Reference solution (a):
- *resolution*: minimum 2.0 between the peaks due to impurity B and ethyl parahydroxybenzoate.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;
- *impurity A*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
- *sum of impurities other than A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
- *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Chlorides (*2.4.4*)

Maximum 350 ppm.

To 10 mL of solution S add 30 mL of *water R* and 1 mL of *nitric acid R* and dilute to 50 mL with *water R*. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with *water R*. Prepare the standard using a mixture of 1 mL of *water R* and 14 mL of *chloride standard solution (5 ppm Cl) R*.

Sulfates (*2.4.13*)

Maximum 300 ppm.

To 25 mL of solution S add 5 mL of *distilled water R* and 10 mL of *hydrochloric acid R* and dilute to 50 mL with *distilled water R*. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with *distilled water R*.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_9H_9NaO_3$ from the declared content of *ethyl parahydroxybenzoate CRS*, multiplied by a correction factor of 1.132.

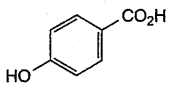
STORAGE

In an airtight container.

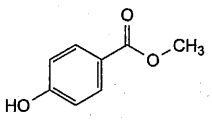
IMPURITIES

Specified impurities *A*.

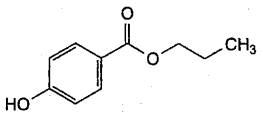
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D.*



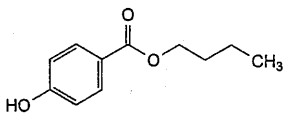
A. 4-hydroxybenzoic acid,



B. methyl 4-hydroxybenzoate (methyl parahydroxybenzoate),



C. propyl 4-hydroxybenzoate (propyl parahydroxybenzoate),



D. butyl 4-hydroxybenzoate (butyl parahydroxybenzoate).

Ethyl Oleate

(Ph. Eur. monograph 1319)

Ph Eur

DEFINITION

Mixture consisting of the ethyl esters of fatty acids, mainly oleic (*cis*-9-octadecenoic) acid.

A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, pale yellow or colourless liquid.

Solubility

Practically insoluble in water, miscible with ethanol (96 per cent), with methylene chloride and with light petroleum (bp: 40-60 °C).

IDENTIFICATION

A. Relative density (see Tests).

B. Saponification value (see Tests).

C. Oleic acid (see Tests).

TESTS

Relative density *(2.2.5)*

0.866 to 0.874.

Acid value *(2.5.1)*

Maximum 0.5, determined on 10.0 g.

Iodine value *(2.5.4, Method A)*

75 to 90.

Peroxide value *(2.5.5, Method A)*

Maximum 10.0.

Saponification value *(2.5.6)*

177 to 188, determined on 2.0 g.

Oleic acid *(2.4.22, Method A)*

Minimum 60.0 per cent in the fatty acid fraction of the substance to be examined.

Water *(2.5.12)*

Maximum 1.0 per cent, determined on 1.00 g.

Total ash *(2.4.16)*

Maximum 0.1 per cent, determined on 2.0 g.

STORAGE

Protected from light.

Ph Eur

Ethylcellulose[1]

(Ph. Eur. monograph 0822)

Action and use

Excipient.

Ph Eur

DEFINITION

Partly *O*-ethylated cellulose.

It may contain a suitable antioxidant.

Content

44.0 per cent to 51.0 per cent of ethoxy ($-OC_2H_5$) groups (dried substance).

♦CHARACTERS

Appearance

White or yellowish-white powder or granular powder.

Solubility

Practically insoluble in water, soluble in methylene chloride and in a mixture of 20 g of ethanol (96 per cent) and 80 g of toluene, slightly soluble in ethyl acetate and in methanol, practically insoluble in glycerol (85 per cent) and in propylene glycol. The solutions may show a slight opalescence.♦

IDENTIFICATION

A. Infrared absorption spectrophotometry *(2.2.24)*.

Preparation Dissolve 40 mg in 1 mL of *methylene chloride R*; spread 2 drops of this solution between 2 sodium chloride plates then remove one of the plates to evaporate the solvent.

Comparison *ethylcellulose CRS*.

◊B. It complies with the limits of the assay.◊

TESTS

Acidity or alkalinity

To 0.5 g add 25 mL of *carbon dioxide-free water R* and shake for 15 min. Filter through a sintered-glass filter (40) *(2.1.2)*. To 10 mL of the solution add 0.1 mL of *phenolphthalein solution R* and 0.5 mL of *0.01 M sodium hydroxide*. The solution is pink. To 10 mL of the solution add 0.1 mL of *methyl red solution R* and 0.5 mL of *0.01 M hydrochloric acid*. The solution is red.

Viscosity *(2.2.9)*

80.0 per cent to 120.0 per cent of that stated on the label for a nominal viscosity greater than 6 mPa·s; 75.0 per cent to 140.0 per cent of that stated on the label for a nominal viscosity not greater than 6 mPa·s.

Shake a quantity of the substance to be examined equivalent to 5.00 g of the dried substance with 95 g of a mixture of 20 g of *ethanol (96 per cent) R* and 80 g of *toluene R* until the substance is dissolved. Determine the viscosity in mPa·s at 25 °C using a capillary viscometer.

Acetaldehyde

Maximum 100 ppm.

Introduce 3.0 g into a 250 mL conical flask with a ground-glass stopper, add 10 mL of *water R* and stir mechanically for 1 h. Allow to stand for 24 h, filter and dilute the filtrate to 100.0 mL with *water R*. Transfer 5.0 mL of the filtrate to a 25 mL volumetric flask, add 5 mL of a 0.5 g/L solution of *methylbenzothiazolone hydrazone hydrochloride R* and heat in a water-bath at 60 °C for 5 min. Add 2 mL of *ferric chloride-sulfamic acid reagent R* and heat again in a water-bath at 60 °C for 5 min. Cool and dilute to 25.0 mL with *water R*. The solution is not more intensely coloured than a standard prepared at the same time and in the same manner using instead of the 5.0 mL of filtrate, 5.0 mL of a reference solution prepared by diluting 3.0 mL of *acetaldehyde standard solution (100 ppm C_2H_4O) R1* to 100.0 mL with *water R*.

Chlorides *(2.4.4)*

Maximum 0.1 per cent.

Disperse 0.250 g in 50 mL of *water R*, heat to boiling and allow to cool, shaking occasionally. Filter and discard the first 10 mL of the filtrate. Dilute 10 mL of the filtrate to 15 mL with *water R*.

Loss on drying *(2.2.32)*

Maximum 3.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*).

Internal standard solution To 10 mL of *o-xylene R* add 0.5 mL of *octane R* and dilute to 100.0 mL with *o-xylene R*.

Test solution To 30.0 mg (dried substance), add 60 mg of *adipic acid R* in a 5 mL pressure-tight reaction vial equipped with a pressure-tight membrane stopper coated with polytetrafluoroethylene and secured with an aluminium crimped cap or another sealing system providing a sufficient air-tightness. Add 2.00 mL of the internal standard solution and 1.0 mL of *hydriodic acid R* and close immediately. Accurately weigh the vial (total mass before heating). Do not mix the contents of the vial by hand before heating. Place the vial in an oven or heat in a suitable heater, with continuous mechanical agitation, maintaining the internal temperature of the vial at 115 ± 2 °C for 70 min. Allow to cool and accurately weigh the vial (total mass after heating). If the difference between the total mass before heating and the total mass after heating is more than 10 mg, prepare a new test solution. After phase separation, pierce through the septum of the vial with a cooled syringe and withdraw a sufficient volume of the upper layer as the test solution.

Reference solution Place 60 mg of *adipic acid R* and 2.00 mL of the internal standard solution in another 5 mL reaction vial, add 1.0 mL of *hydriodic acid R* and close immediately. Accurately weigh the vial then inject 25 µL of *iodoethane R* through the septum into the vial, weigh again accurately and mix. After phase separation, pierce through the septum of the vial with a cooled syringe and withdraw a sufficient volume of the upper layer as the reference solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 3 µm).

Carrier gas *helium for chromatography R.*

Flow rate 4.2 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	50
	3 - 8	50 → 100
	8 - 12	100 → 250
	12 - 20	250
Injection port		250
Detector		280

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to octane (retention time = about 10 min): iodoethane = about 0.6.

System suitability Reference solution:
— *resolution*: minimum 5.0 between the peaks due to iodoethane and octane;
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the response factor of the principal peak determined on 6 injections.

Calculate the response factor (*R*) of *iodoethane R* using the following expression:

$$\frac{A_1 \times W_1 \times C}{A_2 \times 100}$$

A_1 = area of the peak due to the internal standard in the chromatogram obtained with the reference solution;
A_2 = area of the peak due to iodoethane in the chromatogram obtained with the reference solution;
W_1 = mass of *iodoethane R* in the reference solution, in milligrams;
C = percentage content of *iodoethane R*.

Calculate the percentage content *m/m* of ethoxy groups using the following expression:

$$\frac{A_4 \times R \times M_1 \times 100}{A_3 \times W_2 \times M_2}$$

A_3 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_4 = area of the peak due to iodoethane in the chromatogram obtained with the test solution;
R = response factor;
M_1 = molar mass of the ethoxy group (45.1);
M_2 = molar mass of iodoethane (156.0);
W_2 = mass of the sample (dried substance) in the test solution, in milligrams.

LABELLING
The label states:
— the nominal viscosity in millipascal seconds for a 5 per cent *m/m* solution;
— the name and concentration of any added antioxidant.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for ethylcellulose used as binder and film former.

Viscosity
(see Tests).

Degree of substitution
(see Assay).

The following characteristics may be relevant for ethylcellulose used as matrix former in modified-release oral dosage forms.

Viscosity
(see Tests).

Degree of substitution
(see Assay).

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Ethylene Glycol Monopalmitostearate

Ethylene Glycol Monostearate

(Ph. Eur. monograph 1421)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of ethylene glycol mono- and diesters of stearic (octadecanoic) and palmitic (hexadecanoic) acids, produced from the condensation of ethylene glycol and stearic acid 50 of vegetable or animal origin (see *Stearic acid (1474)*).

Content
Minimum of 50.0 per cent of monoesters.

CHARACTERS
Appearance
White or almost white, waxy solid.

Solubility
Practically insoluble in water, soluble in acetone and in hot alcohol.

IDENTIFICATION
A. Melting point (see Tests).

B. Composition of fatty acids (see Tests).

C. It complies with the assay (monoesters content).

TESTS
Melting point *(2.2.15)*
54 °C to 60 °C.

Acid value *(2.5.1)*
Maximum 3.0, determined on 10.0 g.

Iodine value *(2.5.4)*
Maximum 3.0.

Saponification value *(2.5.6)*
170 to 195, determined on 2.0 g.

Composition of fatty acids *(2.4.22, Method A)*
The fatty acid fraction has the following composition:
— *stearic acid*: 40.0 per cent to 60.0 per cent,
— *sum of contents of palmitic acid and stearic acid*: minimum 90.0 per cent.

Free ethylene glycol
Maximum 5.0 per cent, determined as prescribed under Assay.

Total ash *(2.4.16)*
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Size-exclusion chromatography *(2.2.30)*.

Test solution Into a 15 mL flask, weigh about 0.2 g (*m*), to the nearest 0.1 mg. Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Heat gently, if necessary. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into four 15 mL flasks, weigh, to the nearest 0.1 mg, about 2.5 mg, 5.0 mg, 10.0 mg and 20.0 mg of *ethylene glycol R*. Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Weigh the flasks again and calculate the concentration of ethylene glycol in milligrams per gram for each reference solution.

Column:
— *size*: *l* = 0.6 m, Ø = 7 mm,
— *stationary phase*: *styrene-divinylbenzene copolymer R* (particle diameter 5 µm and pore size 10 nm).

Mobile phase *tetrahydrofuran R.*

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to ethylene glycol: diesters = about 0.76, monoesters = about 0.83.

Limits:
— *free ethylene glycol*: from the calibration curve obtained with the reference solutions, determine the concentration (*C*) in milligrams per gram in the test solution and calculate the percentage content in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *monoesters*: calculate the percentage content of monoesters using the following expression:

$$\frac{A}{A+B} \times (100 - D)$$

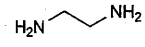
A = area of the peak due to the monoesters,
B = area of the peak due to the diesters,
D = percentage content of free ethylene glycol + percentage content of free fatty acids which may be determined using the following expression: $\frac{I_A \times 270}{561.1}$
I_A = acid value.

STORAGE
Protected from light.

Ph Eur

Ethylenediamine

(Ph. Eur. monograph 0716)



$C_2H_8N_2$ 60.1 *107-15-3*

Ph Eur

DEFINITION
Ethane-1,2-diamine.

Content
98.0 per cent to 101.0 per cent.

CHARACTERS
Appearance
Clear, colourless or slightly yellow liquid, hygroscopic.

Solubility
Miscible with water and with anhydrous ethanol.

On exposure to air, white fumes are evolved. On heating, it evaporates completely.

IDENTIFICATION
A. Relative density *(2.2.5)*: 0.895 to 0.905.

B. Boiling point *(2.2.12)*: 116 °C to 118 °C.

C. To 0.2 mL add 0.5 mL of *acetic anhydride R*. Boil. A crystalline mass forms after cooling, which dissolves in 5 mL of *2-propanol R* with heating. Cool the solution and add 5 mL of *ether R*. If necessary, initiate crystallisation by

scratching the walls of the test-tube with a glass rod. Filter through a sintered-glass filter (*2.1.2*), wash with several portions of *ether R* and dry at 100-105 °C. The residue melts (*2.2.14*) at 173 °C to 177 °C.

TESTS

Solution S
Mix 10 g with *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than the reference solution BY_6 (*2.2.2, Method II*).

Carbonate
A mixture of 4 mL of solution S and 6 mL of *calcium hydroxide solution R* is not more opalescent than reference suspension II (*2.2.1*).

Chlorides (*2.4.4*)
Maximum 100 ppm.

To 5 mL of solution S add 5 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*.

Ammonia and other bases
Dissolve 1.2 g in 20 mL of *ethanol (96 per cent) R* and add, dropwise with stirring, 4.5 mL of *hydrochloric acid R*. Evaporate to dryness on a water-bath, breaking up any resulting cake with a glass rod, and dry at 100-105 °C for 1 h. 1 g of the residue is equivalent to 0.4518 g of $C_2H_8N_2$. Calculate the percentage content of $C_2H_8N_2$: it does not vary by more than 0.5 from the percentage content determined in the assay.

Iron (*2.4.9*)
Maximum 10 ppm, determined on solution S.

Residue on evaporation
Maximum 0.3 per cent.

Evaporate 5.00 g to dryness on a water-bath and dry at 100-105 °C for 1 h. The residue weighs a maximum of 15 mg.

ASSAY
Place 25.0 mL of *1 M hydrochloric acid* and 0.2 mL of *methyl red mixed solution R* in a flask. Add 0.600 g of the substance to be examined. Titrate with *1 M sodium hydroxide* until the colour changes from violet-red to green.

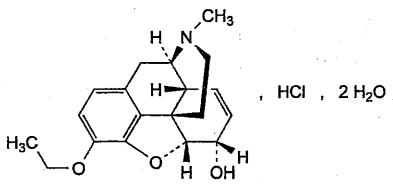
1 mL of *1 M hydrochloric acid* is equivalent to 30.05 mg of $C_2H_8N_2$.

STORAGE
In an airtight container, protected from light.

Ph Eur

Ethylmorphine Hydrochloride

(Ph. Eur. monograph 0491)



$C_{19}H_{24}ClNO_3,2H_2O$ 385.9

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION
7,8-Didehydro-4,5α-epoxy-3-ethoxy-17-methylmorphinan-6α-ol hydrochloride dihydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water and in alcohol, insoluble in cyclohexane.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison Ph. Eur. reference spectrum of ethylmorphine hydrochloride.

B. In a test-tube, dissolve 0.5 g in 6 mL of *water R* and add 15 mL of *0.1 M sodium hydroxide*. Scratch the wall of the tube with a glass rod. A white, crystalline precipitate is formed. Collect the precipitate, wash and dissolve in 20 mL of *water R* heated to 80 °C. Filter and cool in iced water. The crystals, after drying *in vacuo* for 12 h, melt (*2.2.14*) at 85 °C to 89 °C.

C. To about 10 mg add 1 mL of *sulfuric acid R* and 0.05 mL of *ferric chloride solution R2*. Heat on a water-bath. A blue colour develops. Add 0.05 mL of *nitric acid R*. The colour becomes red.

D. Solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.500 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.05 mL of *methyl red solution R* and 0.2 mL of *0.02 M hydrochloric acid*, the solution is red. Add 0.4 mL of *0.02 M sodium hydroxide*, the solution becomes yellow.

Specific optical rotation (*2.2.7*)
−102 to −105 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 25.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 12.5 mg of *codeine R* in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (c) Dilute 0.5 mL of reference solution (b) to 100.0 mL with the mobile phase.

Reference solution (d) To 1.0 mL of the test solution, add 1.0 mL of reference solution (b) and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 30 °C.

Mobile phase Add 1.25 g of *sodium heptanesulfonate R* to a mixture of 12.5 mL of *glacial acetic acid R* and 5 mL of a 20 per cent *V/V* solution of *triethylamine R* in a mixture of equal volumes of *methanol R* and *water R*. Dilute to 1000 mL with *water R*. To 550 mL of this solution add 450 mL of *methanol R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Run time 4 times the retention time of ethylmorphine.

Relative retention With reference to ethylmorphine (retention time = about 6.2 min): impurity B = about 0.7; impurity C = about 0.8; impurity D = about 1.3; impurity A = about 2.5.

System suitability Reference solution (d):
— *resolution*: minimum 5 between the peaks due to ethylmorphine and impurity C.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 0.4,
— *impurities A, B, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
— *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total of impurities other than C*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
8.0 per cent to 10.0 per cent, determined on 0.250 g.

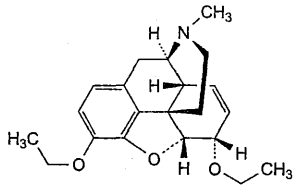
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in a mixture of 5 mL of *0.01 M hydrochloric acid* and 30 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

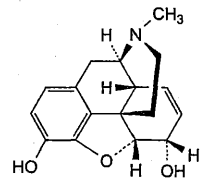
1 mL of *0.1 M sodium hydroxide* is equivalent to 34.99 mg of $C_{19}H_{24}ClNO_3$.

STORAGE
Protected from light.

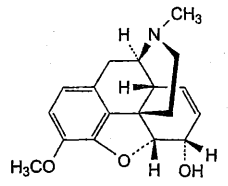
IMPURITIES
Specified impurities *A, B, C, D.*



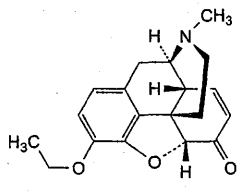
A. 7,8-didehydro-4,5α-epoxy-3,6α-diethoxy-17-methylmorphinan,



B. 7,8-didehydro-4,5α-epoxy-17-methylmorphinan-3,6α-diol (morphine),



C. 7,8-didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6α-ol (codeine),

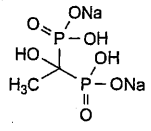


D. 7,8-didehydro-4,5α-epoxy-3-ethoxy-17-methylmorphinan-6-one (ethylmorphinone).

Ph Eur

Etidronate Disodium

(Ph. Eur. monograph 1778)



$C_2H_6Na_2O_7P_2$ 250.0 *7414-83-7*

Action and use
Bisphosphonate; treatment of osteoporosis; Paget's disease.

Ph Eur

DEFINITION
Disodium dihydrogen (1-hydroxyethylidene)bisphosphonate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or yellowish, hygroscopic powder.

Solubility
Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *etidronate disodium CRS*.
The transmittance at about 2000 cm^{-1} (5 µm) is not less than 40 per cent without compensation.
B. It gives reaction (a) of sodium (*2.3.1*).

TESTS
pH (*2.2.3*)
4.2 to 5.2.
Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent.

Impurities A and B
Liquid chromatography (*2.2.29*).
Test solution Dissolve 20.0 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.
Reference solution To 2.0 mL of a 0.3 g/L solution of *phosphoric acid R* add 2.0 mL of a 0.25 g/L solution of *phosphorous acid R* and dilute to 50.0 mL with *water R*.
Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *anion-exchange resin R* (5 µm);
— *temperature*: 35 °C.
Mobile phase Mix 0.2 mL of *anhydrous formic acid R* and 1000 mL of *water R*; adjust to pH 3.5 with an 80 g/L solution of *sodium hydroxide R*.
Flow rate 1.0 mL/min.
Detection Differential refractometer.
Injection 100 µL.
System suitability Reference solution:
— *resolution*: minimum 2.5 between the peaks due to impurity A and impurity B.
Limits:
— *impurities A, B*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.5 per cent).

Water (*2.5.32*)
Maximum 5.0 per cent.
Dissolve 50.0 mg in a mixture of equal volumes of *anhydrous acetic acid R* and *formamide R* and dilute to 5.0 mL with the same mixture of solvents. Use 1.0 mL of the solution.

ASSAY
Dissolve 0.100 g in 2 mL of *anhydrous formic acid R* and dilute to 50 mL with *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M perchloric acid* is equivalent to 12.50 mg of $C_2H_6Na_2O_7P_2$.

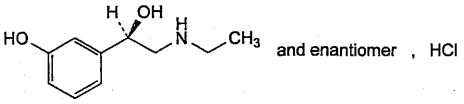
STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B*.
A. H_3PO_4: phosphoric acid,
B. H_3PO_3: phosphorous acid.

Ph Eur

Etilefrine Hydrochloride

(Ph. Eur. monograph 1205)



$C_{10}H_{16}ClNO_2$ 217.7 *943-17-9*

Action and use
Adrenoceptor agonist.

Ph Eur

DEFINITION
(1*RS*)-2-(Ethylamino)-1-(3-hydroxyphenyl)ethanol hydrochloride.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.
A. Melting point (*2.2.14*): 118 °C to 122 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Preparation Discs of *potassium chloride R*.
Comparison *etilefrine hydrochloride CRS*.
C. Thin-layer chromatography (*2.2.27*).
Prepare the solutions protected from bright light and develop the chromatograms protected from light.
Test solution Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.
Reference solution (a) Dissolve 25 mg of *etilefrine hydrochloride CRS* in *methanol R* and dilute to 5 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of *phenylephrine hydrochloride CRS* in 2 mL of reference solution (a) and dilute to 10 mL with *methanol R*.
Plate *TLC silica gel plate R*.
Mobile phase *concentrated ammonia R, methanol R, methylene chloride R* (5:25:70 *V/V/V*).
Application 5 µL.
Development Over a path of 15 cm.
Drying In a current of warm air.
Detection Spray with a 10 g/L solution of *potassium permanganate R*; examine in daylight after 15 min.
System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 0.2 mL of solution S (see Tests), add 1 mL of *water R*, 0.1 mL of *copper sulfate solution R* and 1 mL of *strong sodium hydroxide solution R*. A blue colour is produced. Add 2 mL of *ether R* and shake. The upper layer is colourless.

E. Dilute 1 mL of solution S to 10 mL with *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 2.50 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

Dilute 4 mL of solution S to 10 mL with *carbon dioxide-free water R*. Add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Optical rotation (*2.2.7*)

−0.10° to + 0.10°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with *water R*. Dilute 1.0 mL of this solution to 50.0 mL with *water R*.

Reference solution (b) Dissolve 10.0 mg of *etilefrine impurity A CRS* in *water R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with *water R*.

Reference solution (c) To 10.0 mL of reference solution (a) add 5.0 mL of reference solution (b) and dilute to 20.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm,
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 35 volumes of *acetonitrile R* and 65 volumes of a 1.1 g/L solution of *sodium laurilsulfate R* adjusted to pH 2.3 with *phosphoric acid R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time 5 times the retention time of etilefrine.

Relative retention With reference to etilefrine (retention time = about 9 min): impurity E = about 0.5; impurity C = about 0.8; impurity B = about 0.9; impurity A = about 1.2; impurity F = about 1.7; impurity D = about 4.5.

System suitability Reference solution (c):
- *resolution*: minimum 2.5 between the peaks due to etilefrine and impurity A.

Limits:
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent),
- *impurities B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
- *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *sum of impurities other than A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent),
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent); disregard any peak due to the solvent.

Sulfates (*2.4.13*)

Maximum 200 ppm, determined on 15 mL of solution S.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in a mixture of 20 mL of *anhydrous acetic acid R* and 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 21.77 mg of $C_{10}H_{16}ClNO_2$.

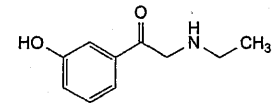
STORAGE

In an airtight container, protected from light.

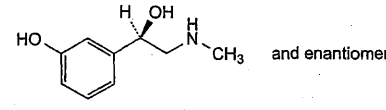
IMPURITIES

Specified impurities *A, B, C, D, E.*

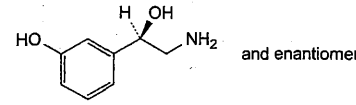
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F.*



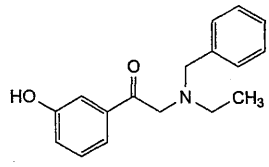
A. 2-(ethylamino)-1-(3-hydroxyphenyl)ethanone (etilefrone),



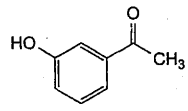
B. (1*RS*)-1-(3-hydroxyphenyl)-2-(methylamino)ethanol (phenylephrine),



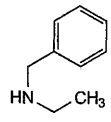
C. (1*RS*)-2-amino-1-(3-hydroxyphenyl)ethanol (norfenefrine),



D. 2-(benzylethylamino)-1-(3-hydroxyphenyl)ethanone (benzyletilefrone),



E. 1-(3-hydroxyphenyl)ethanone (3-hydroxyacetophenone),

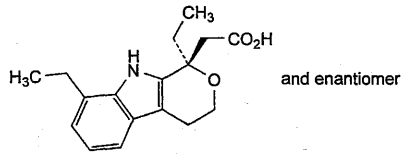


F. *N*-benzylethanamine (benzylethylamine).

Ph Eur

Etodolac

(Ph. Eur. monograph 1422)



$C_{17}H_{21}NO_3$ 287.4 *41340-25-4*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparation
Etodolac Capsules

Ph Eur

DEFINITION
[(1*RS*)-1,8-Diethyl-1,3,4,9-tetrahydropyrano[3,4-*b*]indol-1-yl] acetic acid.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 144 °C to 150 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *etodolac CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of *etodolac CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel GF_{254} plate R* previously activated by heating at 105 °C for 1 h.

Place the plate in an unsaturated tank containing a mixture of 20 volumes of a 25 g/L solution of *ascorbic acid R* and 80 volumes of *methanol R*. Allow the solution to ascend 1 cm above the line of application on the plate, remove the plate and allow it to dry for at least 30 min.

Mobile phase *glacial acetic acid R, anhydrous ethanol R, toluene R* (0.5:30:70 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in *acetonitrile R1* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with *acetonitrile R1*. Dilute 1.0 mL of this solution to 20.0 mL with *acetonitrile R1*.

Reference solution (b) Dissolve 4 mg of *etodolac impurity H CRS* in the test solution and dilute to 10 mL with the same solution. Dilute 0.5 mL of this solution to 50 mL with *acetonitrile R1*.

Reference solution (c) Dissolve the contents of a vial of *etodolac for peak identification CRS* (containing impurities A, B, C, D, E, F, G, H, I and K) in 1.0 mL of *acetonitrile R1*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: 0.77 g/L solution of *ammonium acetate R*;
— *mobile phase B*: mobile phase A, *acetonitrile R1* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	80 → 50	20 → 50
25 - 42	50	50
42 - 48	50 → 80	50 → 20

Flow rate 1 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *etodolac for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D, E, F, G, H, I and K.

Relative retention With reference to etodolac (retention time = about 16.7 min): impurity A = about 0.68; impurity B = about 0.83; impurity C = about 0.85; impurity H = about 1.09; impurity D = about 1.17; impurity G = about 1.19; impurity E = about 1.20; impurity F = about 1.22; impurity I = about 1.50; impurity K = about 2.37.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to etodolac and impurity H.

Limits:
— *impurity C*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurities A, B, D, E, F, G, H, I, K*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides
Maximum 300 ppm.

Dissolve 1.0 g of the substance to be examined in 60 mL of *methanol R*, add 10 mL of *water R* and 20 mL of *dilute nitric acid R*. Titrate with *0.01 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.01 M silver nitrate* is equivalent to 0.3545 mg of Cl.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

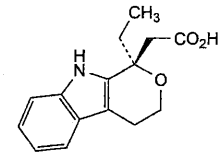
Dissolve 0.250 g in 60 mL of *methanol R*. Titrate with 0.1 M tetrabutylammonium hydroxide determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 28.74 mg of $C_{17}H_{21}NO_3$.

IMPURITIES

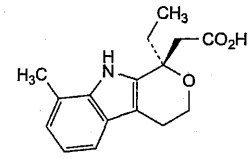
Specified impurities *A, B, C, D, E, F, G, H, I, K.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *J, L.*



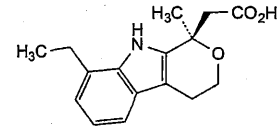
and enantiomer

A. [(1*RS*)-1-ethyl-1,3,4,9-tetrahydropyrano[3,4-*b*]indol-1-yl] acetic acid (8-desethyl etodolac),



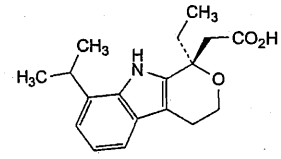
and enantiomer

B. [(1*RS*)-1-ethyl-8-methyl-1,3,4,9-tetrahydropyrano[3,4-*b*] indol-1-yl]acetic acid (8-methyl etodolac),



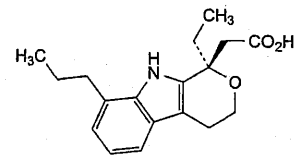
and enantiomer

C. [(1*RS*)-8-ethyl-1-methyl-1,3,4,9-tetrahydropyrano[3,4-*b*] indol-1-yl]acetic acid (1-methyl etodolac),



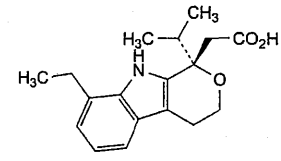
and enantiomer

D. [(1*RS*)-1-ethyl-8-(1-methylethyl)-1,3,4,9-tetrahydropyrano [3,4-*b*]indol-1-yl]acetic acid (8-isopropyl etodolac),



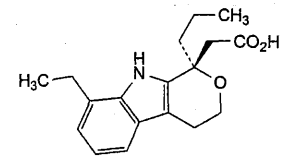
and enantiomer

E. [(1*RS*)-1-ethyl-8-propyl-1,3,4,9-tetrahydropyrano[3,4-*b*] indol-1-yl]acetic acid (8-propyl etodolac),



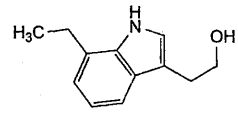
and enantiomer

F. [(1*RS*)-8-ethyl-1-(1-methylethyl)-1,3,4,9-tetrahydropyrano [3,4-*b*]indol-1-yl]acetic acid (1-isopropyl etodolac),

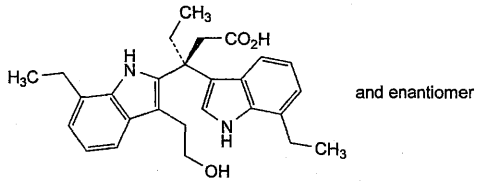


and enantiomer

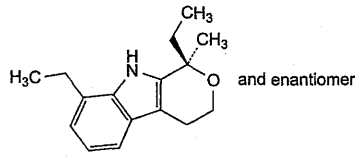
G. [(1*RS*)-8-ethyl-1-propyl-1,3,4,9-tetrahydropyrano[3,4-*b*] indol-1-yl]acetic acid (1-propyl etodolac),



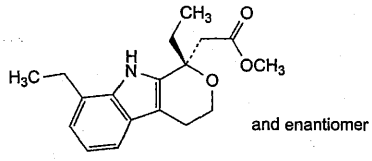
H. 2-(7-ethyl-1*H*-indol-3-yl)ethanol,



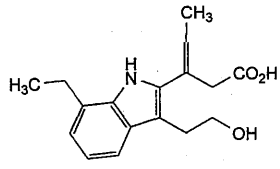
I. (3*RS*)-3-[7-ethyl-3-(2-hydroxyethyl)-1*H*-indol-2-yl]-3-(7-ethyl-1*H*-indol-3-yl)pentanoic acid (etodolac dimer),



J. (1*RS*)-1,8-diethyl-1-methyl-1,3,4,9-tetrahydropyrano[3,4-*b*]indole (decarboxy etodolac),



K. methyl [(1*RS*)-1,8-diethyl-1,3,4,9-tetrahydropyrano[3,4-*b*]indol-1-yl]acetate (etodolac methyl ester),

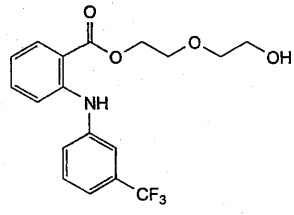


L. (*EZ*)-3-[7-ethyl-3-(2-hydroxyethyl)-1*H*-indol-2-yl]pent-3-enoic acid.

Ph Eur

Etofenamate

(Ph. Eur. monograph 1513)



$C_{18}H_{18}F_3NO_4$ 369.4 *30544-47-9*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
2-(2-Hydroxyethoxy)ethyl 2-[[3-(trifluoromethyl)phenyl]amino]benzoate.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS
Appearance
Yellowish, viscous liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent) and with ethyl acetate.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *etofenamate CRS.*

Preparation Films.

TESTS
Appearance
The substance to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution GY_1 (*2.2.2, Method II*).

Impurity F
Gas chromatography (*2.2.28*).

Internal standard *tetradecane R.*

Solution A Dissolve 6.0 mg of *tetradecane R* in *hexane R* and dilute to 10.0 mL with the same solvent.

Solution B To 6.0 mg of *diethylene glycol R* in a 10 mL volumetric flask add 3 mL of *N-methyltrimethylsilyl-trifluoroacetamide R* and heat for 30 min at 50 °C. After cooling dilute to 10.0 mL with *N-methyltrimethylsilyl-trifluoroacetamide R.*

Test solution To 0.200 g of the substance to be examined add 10 µL of solution A. Add 2 mL of *N-methyltrimethylsilyl-trifluoroacetamide R* and heat for 30 min at 50 °C.

Reference solution To 2.0 mL of *N-methyltrimethylsilyl-trifluoroacetamide R* add 10 µL of solution A and 10 µL of solution B.

Column:
— *size: l* = 25 m, Ø = 0.20 mm;
— *stationary phase: poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.33 µm).

Carrier gas *hydrogen for chromatography R.*

Flow rate 0.9 mL/min.

Temperature:

	Time (min)	Temperature (°C)	Rate (°C/min)
Column	0 - 13	60 → 150	7
	13 - 19	150 → 300	25
	19 - 34	300	
Injection port		150	
Detector		300	

Detection Flame ionisation.

Injection 0.2 µL.

Limit:
— *impurity F:* maximum 0.1 per cent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *water R, methanol R* (40:60 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in 30 mL of *methanol R* and dilute to 50.0 mL with *water R*.

Reference solution (a) Dissolve 10.0 mg of *etofenamate impurity G CRS* in *methanol R* and dilute to 20.0 mL with the same solvent. Dilute 0.2 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 0.2 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (c) To 5.0 mL of reference solution (a), add 5.0 mL of reference solution (b).

Reference solution (d) Dissolve 10.0 mg of *etofenamate for peak identification CRS* (containing impurities A, B, C, D and E) in 6.0 mL of *methanol R* and dilute to 10.0 mL with *water R*.

Column:
- *size*: *l* = 0.10 m, Ø = 4.0 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: dissolve 1.3 g of *ammonium phosphate R* and 4.0 g of *tetrabutylammonium hydroxide R* in 900 mL of *water R*, adjust to pH 8.0 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*;
- *mobile phase B*: *methanol R*;

Time	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 13	40	60
13 - 20	40 → 10	60 → 90
20 - 25	10	90

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 286 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *etofenamate for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D and E; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity G.

Relative retention With reference to etofenamate (retention time = about 13 min): impurity A = about 0.2; impurity C = about 0.7; impurity G = about 0.85; impurity E = about 1.5; impurity B = about 1.6; impurity D = about 1.7.

System suitability Reference solution (c):
- *resolution*: minimum 2.3 between the peaks due to impurity G and etofenamate.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.62; impurity C = 0.45; impurity D = 0.77;
- *impurity D*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurity A*: not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
- *impurities B, C, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *impurity G*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.2 per cent);
- *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

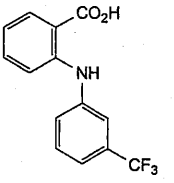
ASSAY

To 3.000 g add 20 mL of *2-propanol R* and 20.0 mL of *1 M sodium hydroxide* and heat under reflux for 2 h. Add 0.1 mL of *bromothymol blue solution R1*. Titrate after cooling with *1 M hydrochloric acid* until the colour disappears. Carry out a blank titration.

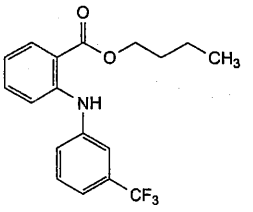
1 mL of *1 M sodium hydroxide* is equivalent to 0.3694 g of $C_{18}H_{18}F_3NO_4$.

IMPURITIES

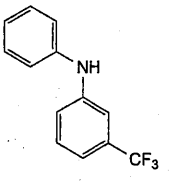
Specified impurities *A, B, C, D, E, F, G.*



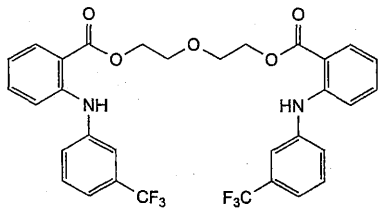
A. 2-[[3-(trifluoromethyl)phenyl]amino]benzoic acid (flufenamic acid),



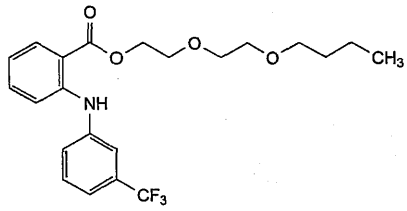
B. butyl 2-[[3-(trifluoromethyl)phenyl]amino]benzoate (butyl flufenamate),



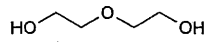
C. *N*-phenyl-3-(trifluoromethyl)aniline,



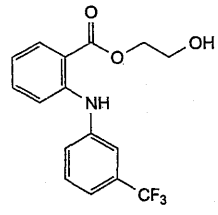
D. 2,2′-oxybis(ethylene) bis[2-[[3-(trifluoromethyl) phenyl]amino]benzoate],



E. 2-(2-butoxyethoxy)ethyl 2-[[3-(trifluoromethyl) phenyl]amino]benzoate,



F. 2,2′-oxydiethanol,

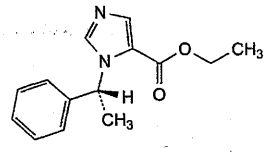


G. 2-hydroxyethyl 2-[[3-(trifluoromethyl)phenyl] amino]benzoate.

Ph Eur

Etomidate

(Ph. Eur. monograph 1514)



$C_{14}H_{16}N_2O_2$ 244.3 *33125-97-2*

Action and use
Intravenous general anaesthetic.

Ph Eur

DEFINITION
Ethyl 1-[(1*R*)-1-phenylethyl]-1*H*-imidazole-5-carboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

mp
About 68 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *etomidate CRS.*

B. Specific optical rotation (see Tests).

TESTS
Solution S
Dissolve 0.25 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
+ 67 to + 70 (dried substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *anhydrous ethanol R, water R* (50:50 *V/V*).
Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.
Reference solution (a) Dissolve 5.0 mg of *etomidate CRS* and 5.0 mg of *etomidate impurity B CRS* in the solvent mixture, then dilute to 250.0 mL with the solvent mixture.
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 25.0 mL with the solvent mixture.
Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).
Mobile phase:
— *mobile phase A*: 5 g/L solution of *ammonium carbonate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90 → 30	10 → 70
5 - 6	30 → 10	70 → 90
6 - 10	10	90

Flow rate 2.0 mL/min.
Detection Spectrophotometer at 235 nm.
Injection 10 µL.
Retention time Impurity B = about 4.5 min; etomidate = about 5.0 min.
System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to impurity B and etomidate; if necessary, adjust the concentration of ammonium carbonate in the mobile phase or the time programme of the linear gradient.
Limits:
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 40 °C for 4 h.

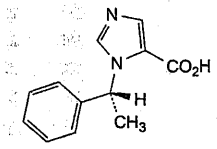
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid* using 0.2 mL of *naphtholbenzein solution R* as indicator.

1 mL of *0.1 M perchloric acid* is equivalent to 24.43 mg of $C_{14}H_{16}N_2O_2$.

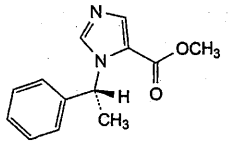
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, C.*



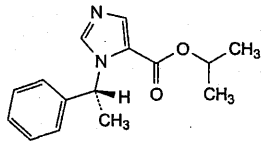
and enantiomer

A. 1-[(1*RS*)-1-phenylethyl]-1*H*-imidazole-5-carboxylic acid,



and enantiomer

B. methyl 1-[(1*RS*)-1-phenylethyl]-1*H*-imidazole-5-carboxylate (metomidate),



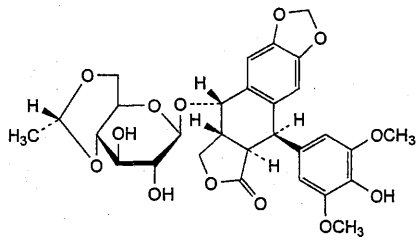
and enantiomer

C. 1-methylethyl 1-[(1*RS*)-1-phenylethyl]-1*H*-imidazole-5-carboxylate.

Ph Eur

Etoposide

(Ph. Eur. monograph 0823)



$C_{29}H_{32}O_{13}$ 588.6 *33419-42-0*

Action and use
Inhibitor of DNA topoisomerase type II; cytotoxic.

Preparations
Etoposide Capsules
Etoposide Infusion

Ph Eur

DEFINITION
(5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder, slightly hygroscopic.

Solubility
Practically insoluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: A, B.

Second identification: C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *etoposide CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 2 mL with the same mixture of solvents.

Reference solution Dissolve 10 mg of *etoposide CRS* in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 2 mL with the same mixture of solvents.

Plate *silica gel H R* as the coating substance.

Mobile phase *water R, glacial acetic acid R, acetone R, methylene chloride R* (1.5:8:20:100 *V/V/V/V*).

Application 5 µL as bands of 10 mm.

Development Immediately, over 6/7 of the plate.

Drying In a current of warm air for 5 min.

Detection Spray with a mixture of 1 volume of *sulfuric acid R* and 9 volumes of *ethanol (96 per cent) R* and heat at 140 °C for 15 min. Cover the plate immediately with a glass plate of the same size. Examine in daylight.

Results The principal zone in the chromatogram obtained with the test solution is similar in position, colour and size to the principal zone in the chromatogram obtained with the reference solution.

D. In a test-tube dissolve about 5 mg in 5 mL of *glacial acetic acid R* and add 0.05 mL of *ferric chloride solution R1*. Mix and cautiously add 2 mL of *sulfuric acid R*. Avoid mixing the 2 layers. Allow to stand for about 30 min; a pink to reddish-brown ring develops at the interface and the upper layer is yellow.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 or BY_6 (*2.2.2, Method II*).

Dissolve 0.6 g in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 20 mL with the same mixture of solvents.

Specific optical rotation (*2.2.7*)
−114 to −106 (anhydrous substance).

Dissolve 50.0 mg in a mixture of 1 volume of *methanol R* and 9 volumes of *methylene chloride R* and dilute to 10.0 mL with the same mixture of solvents.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mobile phase A, mobile phase B (50:50 *V/V*).

Test solution (a) Dissolve 40 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 4 mg of *etoposide for system suitability CRS* (containing impurities B, C, D, E, N and O) in 1.0 mL of the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *etoposide CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.125 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *anhydrous formic acid R, triethylamine R, water for chromatography R* (1:1:998 *V/V/V*);
- *mobile phase B*: *anhydrous formic acid R, triethylamine R, acetonitrile R* (1:1:998 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	75	25
7 - 23	75 → 27	25 → 73

Flow rate 1 mL/min.

Detection Spectrophotometer at 285 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *etoposide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D, E, N and O.

Relative retention With reference to etoposide (retention time = about 5 min): impurity D = about 0.4; impurity E = about 0.8; impurity C = about 1.1; impurity B = about 1.2; impurity N = about 3.1; impurity O = about 4.2.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to etoposide; and minimum 3.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity C.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity O by 1.7;
- *impurities B, C, D, E, N*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurity O*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to the solvent.

Water

(*2.5.32*)Maximum 6.0 per cent, determined on 0.150 g using the evaporation technique at 130 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (c).

System suitability:
- *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections of reference solution (c).

Calculate the percentage content of $C_{29}H_{32}O_{13}$ taking into account the assigned content of *etoposide CRS*.

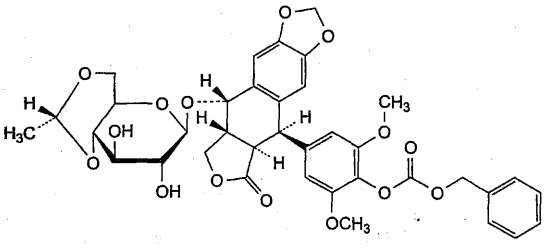
STORAGE

In an airtight container.

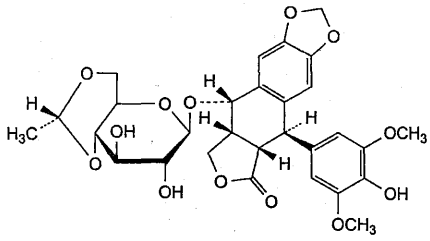
IMPURITIES

Specified impurities *B, C, D, E, N, O.*

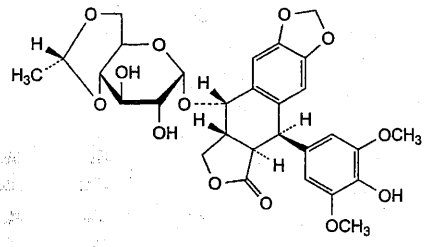
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, F, G, H, I, J, K, L, M, P, Q, R.*



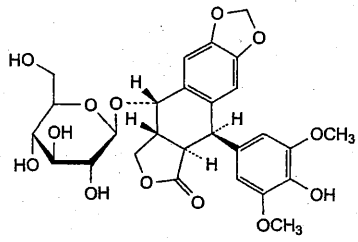
A. (5*R*,5a*R*,8a*R*,9*S*)-5-[4-[[(benzyloxy)carbonyl]oxy]-3,5-dimethoxyphenyl]-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4'-carbobenzoyloxyethylidene-lignan P),



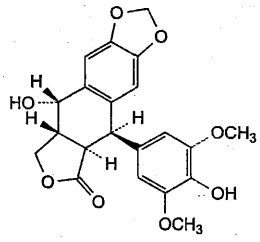
B. (5*R*,5a*S*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (picroethylidene-lignan P; *cis*-etoposide),



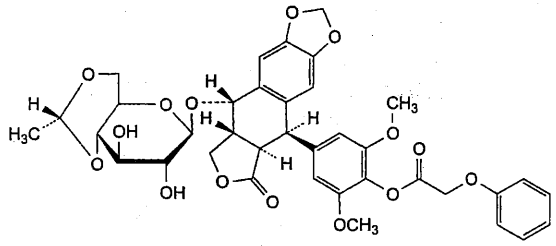
C. (5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-α-D-glucopyranosyl]oxy]-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (α-etoposide),



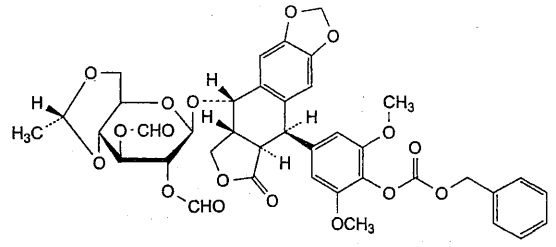
D. (5*R*,5a*R*,8a*R*,9*S*)-9-(β-D-glucopyranosyloxy)-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro-[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (lignan P),



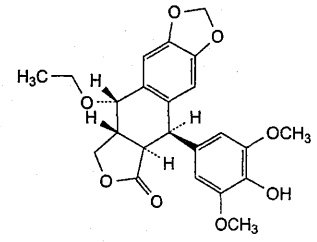
E. (5*R*,5a*R*,8a*R*,9*S*)-9-hydroxy-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4′-desmethylepipodophyllotoxin),



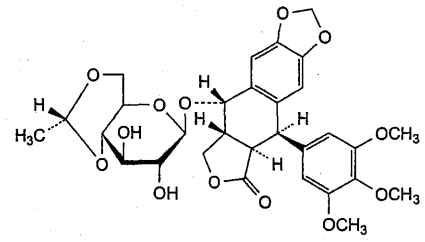
F. (5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-[4-[(phenoxyacetyl)oxy]-3,5-dimethoxyphenyl]-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4'-phenoxyacetyletoposide),



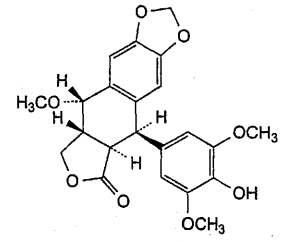
G. (5*R*,5a*R*,8a*R*,9*S*)-5-[4-[[(benzyloxy)carbonyl]oxy]-3,5-dimethoxyphenyl]-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-2,3-di-*O*-formyl-β-D-glucopyranosyl]oxy]-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4'-carbobenzoyloxydiformyl-ethylidene-lignan P),



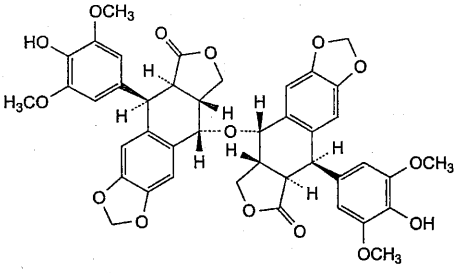
H. (5*R*,5a*R*,8a*R*,9*S*)-9-ethoxy-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4′-*O*-desmethyl-1-*O*-ethylepipodophyllotoxin),



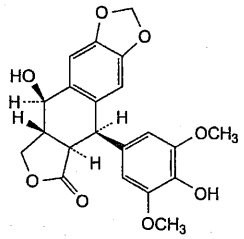
I. (5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-(3,4,5-trimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4-*O*-methylethylidene-lignan P),



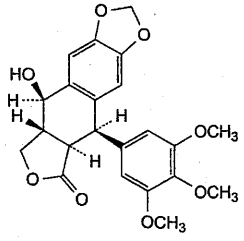
J. (5*R*,5a*R*,8a*R*,9*S*)-5-(4-hydroxy-3,5-dimethoxyphenyl)-9-methoxy-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4′-*O*-desmethyl-1-*O*-methylepipodophyllotoxin),



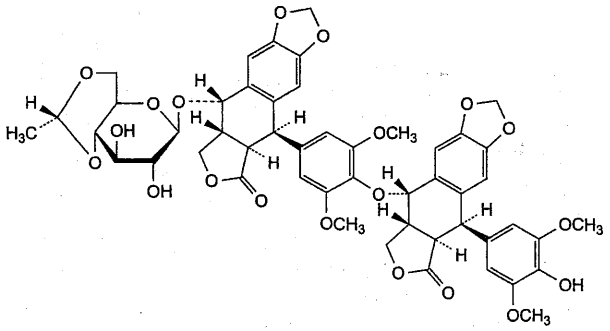
K. 9,9′-oxybis[(5*R*,5a*R*,8a*R*,9*S*)-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro-[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one] (di-4′-*O*-desmethylepipodophyllotoxin),



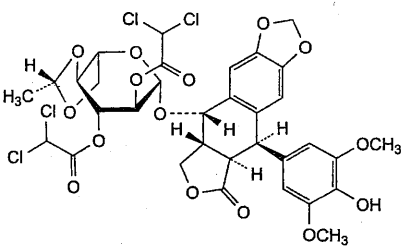
L. (5*R*,5a*R*,8a*R*,9*R*)-9-hydroxy-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (4′-*O*-desmethylpodophyllotoxin),



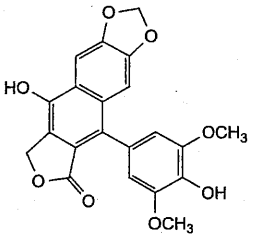
M. (5*R*,5a*R*,8a*R*,9*R*)-9-hydroxy-5-(3,4,5-trimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one (podophyllotoxin),



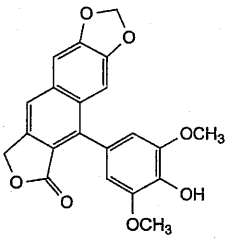
N. (5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-[4-[[(5*R*,5a*R*,8a*R*,9*S*)-5-(4-hydroxy-3,5-dimethoxyphenyl)-6-oxo-5,5a,6,8,8a,9-hexahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-9-yl]oxy]-3,5-dimethoxyphenyl]-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one,



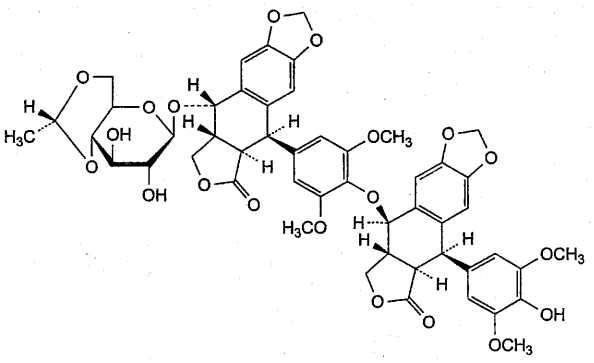
O. (5*R*,5a*R*,8a*R*,9*S*)-9-[[2,3-bis-*O*-(dichloroacetyl)-4,6-*O*-[(1*S*)-ethane-1,1-diyl]-β-L-glucopyranosyl]oxy]-5-(4-hydroxy-3,5-dimethoxyphenyl)-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one,



P. 9-hydroxy-5-(4-hydroxy-3,5-dimethoxyphenyl)[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(8*H*)-one,



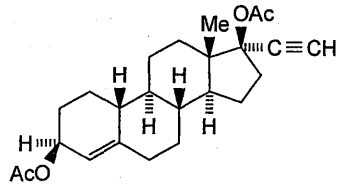
Q. 5-(4-hydroxy-3,5-dimethoxyphenyl)[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(8*H*)-one,



R. (5*R*,5a*R*,8a*R*,9*S*)-9-[[4,6-*O*-[(1*R*)-ethane-1,1-diyl]-β-D-glucopyranosyl]oxy]-5-[4-[[(5*R*,5a*R*,8a*R*,9*R*)-5-(4-hydroxy-3,5-dimethoxyphenyl)-6-oxo-5,5a,6,8,8a,9-hexahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-9-yl]oxy]-3,5-dimethoxyphenyl]-5,8,8a,9-tetrahydro[2]benzofuro[5,6-*f*][1,3]benzodioxol-6(5a*H*)-one.

Ph Eur

Etynodiol Diacetate



$C_{24}H_{32}O_4$ 384.5 *297-76-7*

Action and use
Progestogen.

Preparation
Etynodiol Tablets

DEFINITION
Etynodiol Diacetate is 19-nor-17α-pregn-4-en-20-yne-3β,17β-diyl diacetate. It contains not less than 97.0% and not more than 102.0% of $C_{24}H_{32}O_4$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Very slightly soluble in *water*; freely soluble in *ether*; soluble in *ethanol (96%)*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 220 to 350 nm of the solution obtained in the test for Light absorption exhibits a maximum only at 236 nm and shoulders at 229 and 243 nm.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of etynodiol diacetate *(RS 138)*.

C. Yields the reaction characteristic of *acetyl groups*, Appendix VI.

TESTS
Light absorption
Dissolve 50 mg in sufficient *methanol* to produce 50 mL (solution A). To 10 mL of solution A add 40 mL of *methanol* and a mixture of 3 mL of *hydrochloric acid* and 2 mL of *water*, mix and boil on a water bath for exactly 10 minutes. Cool, dilute to 100 mL with *methanol* and dilute 10 mL of the solution to 100 mL with *methanol*. The *absorbance* of the resulting solution at the maximum at 236 nm, Appendix II B, is 0.47 to 0.50, calculated with reference to the dried substance, using in the reference cell a solution prepared by diluting 1 mL of solution A to 100 mL with *methanol*.

Melting point
126° to 131°, Appendix V A.

Specific optical rotation
In a 1% w/v solution in *chloroform*, –70 to –76, Appendix V F, calculated with reference to the dried substance.

Conjugated compounds
Absorbance of a 0.050% w/v solution in *methanol* at 236 nm, not more than 0.47, calculated with reference to the dried substance, Appendix II B.

Loss on drying
When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

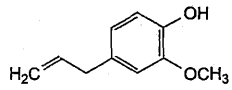
Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.2 g in 40 mL of *tetrahydrofuran*, add 10 mL of a 10% w/v solution of *silver nitrate* and titrate with *0.1M sodium hydroxide VS*, determining the end point potentiometrically. Each mL of *0.1M sodium hydroxide VS* is equivalent to 38.45 mg of $C_{24}H_{32}O_4$.

STORAGE
Etynodiol Diacetate should be protected from light.

Eugenol

(Ph. Eur. monograph 1100)



$C_{10}H_{12}O_2$ 164.2 *97-53-0*

Ph Eur

DEFINITION
2-Methoxy-4-(prop-2-enyl)phenol.

CHARACTERS
Appearance
Colourless or pale yellow, clear liquid, darkening on exposure to air.

It has a strong odour of clove.

Solubility
Practically insoluble in water, freely soluble in ethanol (70 per cent *V/V*), practically insoluble in glycerol, miscible with ethanol (96 per cent), with glacial acetic acid, with methylene chloride and with fatty oils.

IDENTIFICATION
First identification: B.

Second identification: A, C, D.

A. Refractive index (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *eugenol CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 µL of the substance to be examined in *ethanol (96 per cent) R* and dilute to 25 mL with the same solvent.

Reference solution Dissolve 50 µL of *eugenol CRS* in *ethanol (96 per cent) R* and dilute to 25 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *ethyl acetate R, toluene R* (10:90 *V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of cold air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Spray with *anisaldehyde solution R* and heat at 100-105 °C for 10 min.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 0.05 mL in 2 mL of *ethanol (96 per cent) R* and add 0.1 mL of *ferric chloride solution R1*. A dark green colour is produced which changes to yellowish-green within 10 min.

TESTS

Relative density (*2.2.5*)
1.066 to 1.070.

Refractive index (*2.2.6*)
1.540 to 1.542.

Dimeric and oligomeric compounds
Dissolve 0.150 g in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. The absorbance (*2.2.25*) of the solution at 330 nm is not greater than 0.25.

Related substances
Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 1.00 g of the substance to be examined in *anhydrous ethanol R* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *anhydrous ethanol R*.

Reference solution (b) Dissolve 50 mg of *vanillin R* (impurity H) in 1 mL of the test solution and dilute to 5 mL with *anhydrous ethanol R*.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.25 mm;
— *stationary phase*: *polymethylphenylsiloxane R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	80
	2 - 27	80 → 280
	27 - 47	280
Injection port		250
Detector		280

Detection Flame ionisation.

Injection 1 µL.

System suitability Reference solution (b):
— *relative retention* with reference to eugenol: impurity H = minimum 1.1.

Limits:
— *any impurity*: for each impurity, maximum 0.5 per cent;
— *sum of impurities with a relative retention greater than 2.0 with reference to eugenol*: maximum 1.0 per cent;
— *total*: maximum 3.0 per cent;
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

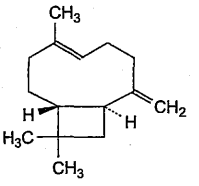
Hydrocarbons
Dissolve 1 mL in 5 mL of *dilute sodium hydroxide solution R* and add 30 mL of *water R* in a stoppered test-tube. Examined immediately, the solution is yellow and clear (*2.2.1*).

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

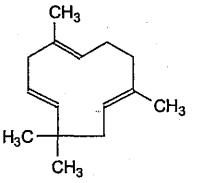
STORAGE

In a well-filled container, protected from light.

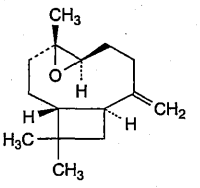
IMPURITIES



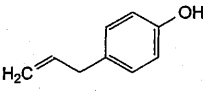
A. (1*R*,4*E*,9*S*)-4,11,11-trimethyl-8-methylenebicyclo[7.2.0]undec-4-ene (β-caryophyllene),



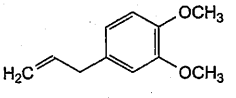
B. (1*E*,4*E*,8*E*)-2,6,6,9-tetramethylcycloundeca-1,4,8-triene (α-humulene, α-caryophyllene),



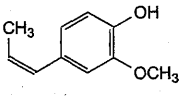
C. (1*R*,4*R*,6*R*,10*S*)-4,12,12-trimethyl-9-methylene-5-oxatricyclo[8.2.0.0^{4,6}]dodecane (β-caryophyllene oxide),



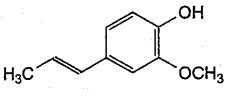
D. 4-(prop-2-enyl)phenol,



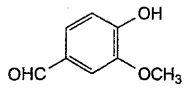
E. 1,2-dimethoxy-4-(prop-2-enyl)benzene (eugenol methyl ether),



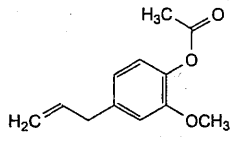
F. 2-methoxy-4-[(Z)-prop-1-enyl]phenol (*cis*-isoeugenol),



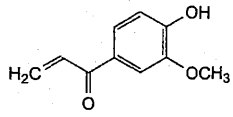
G. 2-methoxy-4-[(E)-prop-1-enyl]phenol (*trans*-isoeugenol),



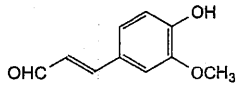
H. 4-hydroxy-3-methoxybenzaldehyde (vanillin),



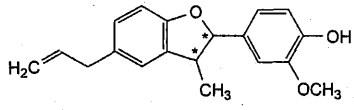
I. 2-methoxy-4-(prop-2-enyl)phenyl acetate (acetyleugenol),



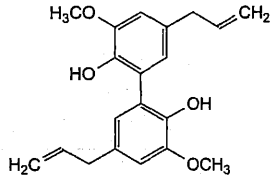
J. 1-(4-hydroxy-3-methoxyphenyl)prop-2-enone,



K. (*E*)-3-(4-hydroxy-3-methoxyphenyl)prop-2-enal (*trans*-coniferyl aldehyde),

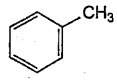


L. 2-methoxy-4-[3-methyl-5-(prop-2-enyl)-2,3-dihydrobenzofuran-2-yl]phenol (dehydrodi-isoeugenol),



M. 3,3′-dimethoxy-5,5′-bis(prop-2-enyl)biphenyl-2,2′-diol (dehydrodieugenol),

N. O. 2 further unknown dimeric compounds,



P. toluene.

Ph Eur

Refined Evening Primrose Oil

(Ph. Eur. monograph 2104)

Ph Eur

DEFINITION

Fatty oil obtained from seeds of *Oenothera biennis* L. or *Oenothera lamarckiana* L. by extraction and/or expression. It is then refined. A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, light yellow or yellow liquid.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum (bp: 40-60 °C).

Relative density

About 0.923.

Refractive index

About 1.478.

IDENTIFICATION

First identification: B.

Second identification: A.

A. Identification of fatty oils by thin-layer chromatography (*2.3.2*).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

B. Composition of fatty acids (see Tests).

TESTS

Acid value (*2.5.1*)

Maximum 0.5, or maximum 0.3 if intended for use in the manufacture of parenteral preparations.

Peroxide value (*2.5.5, Method A*)

Maximum 10.0, or maximum 5.0 if intended for use in the manufacture of parenteral preparations.

Unsaponifiable matter (*2.5.7*)

Maximum 2.5 per cent, determined on 5.0 g.

Alkaline impurities (*2.4.19*)

It complies with the test.

Composition of fatty acids (*2.4.22, Method A*)

Use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty-acid fraction of the oil:

— *saturated fatty acids of chain length less than C_{16}*: maximum 0.3 per cent;
— *palmitic acid*: 4.0 per cent to 10.0 per cent;
— *stearic acid*: 1.0 per cent to 4.0 per cent;
— *oleic acid*: 5.0 per cent to 12.0 per cent;
— *linoleic acid*: 65.0 per cent to 85.0 per cent;
— *gamma-linolenic acid*: 7.0 per cent to 14.0 per cent;
— *alpha-linolenic acid*: maximum 0.5 per cent.

Brassicasterol (*2.4.23*)

Maximum 0.3 per cent in the sterol fraction of the oil.

Water (*2.5.32*)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

Under an inert gas, in a well-filled, airtight container, protected from light.

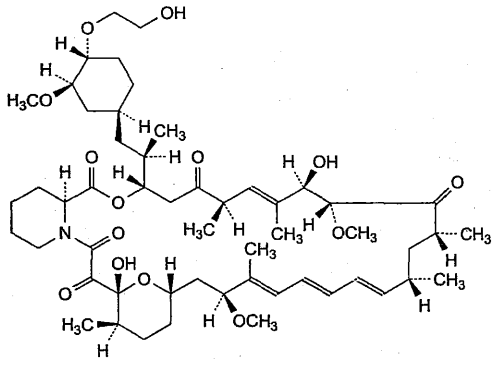
LABELLING

The label states, where applicable, that the oil is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Everolimus

(Ph. Eur. monograph 2918)



$C_{53}H_{83}NO_{14}$ 958 *159351-69-6*

Action and use
Calcineurin inhibitor; immunosuppressant.

Ph Eur

(1*R*,9*S*,12*S*,15*R*,16*E*,18*R*,19*R*,21*R*,23*S*,24*E*,26*E*,28*E*,30*S*,32*S*,35*R*)-1,18-Dihydroxy-12-[(2*R*)-1-[(1*S*,3*R*,4*R*)-4-(2-hydroxyethoxy)-3-methoxycyclohexyl]propan-2-yl]-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-11,36-dioxa-4-azatricyclo[30.3.1.$0^{4,9}$]hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentone.

Semi-synthetic product derived from a fermentation product. A suitable antioxidant may be added.

DEFINITION

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

A reversible isomerisation to everolimus tautomer takes place in solution. The activity is due to both compounds.

CHARACTERS

Appearance
White or faintly yellow, hygroscopic powder.

Solubility
Practically insoluble in water, very soluble in anhydrous ethanol, pratically insoluble in heptane.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *everolimus CRS*.

TESTS

Specific optical rotation (*2.2.7*)
− 153.0 to − 141.0 (anhydrous substance).

Dissolve 0.100 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Impurity A
Liquid chromatography (*2.2.29*). *Protect the solutions from light. Use plastic labware to minimise the formation of everolimus tautomer.*

Test solution Dissolve 50.0 mg of the substance to be examined in *acetonitrile R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 1 mg of *sirolimus R* (impurity A) in 5 mL of *acetonitrile R*. Dilute 0.8 mL of the solution to 10 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R*.

Column:
— *size*: *l* = 0.15 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped solid core octadecylsilyl silica gel for chromatography R* (2.7 µm);
— *temperature*: 60 °C.

Mobile phase:
— *mobile phase A*: *anhydrous formic acid R, methanol R*, 0.70 g/L solution of *ammonium formate R, acetonitrile R* (0.05:100:450:450 *V/V/V/V*);
— *mobile phase B*: *anhydrous formic acid R, methanol R*, 0.96 g/L solution of *ammonium formate R, acetonitrile R* (0.05:100:330:570 *V/V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 14	100	0
14 - 23	100 → 0	0 → 100
23 - 28	0	100

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 278 nm.

Autosampler Set at 6 °C.

Injection 3.5 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to everolimus (retention time = about 16 min): impurity A = about 0.9; everolimus tautomer = about 1.1.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to everolimus.

Calculation of percentage content:
— for impurity A, use the concentration of everolimus (and everolimus tautomer if present) in reference solution (b).

Limit:
— *impurity A*: maximum 0.8 per cent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light. Prepare the solutions immediately before use.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *everolimus for system suitability CRS* (containing impurities D, E, F, H, I and J) in 1 mL of *acetonitrile R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with *acetonitrile R*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R*.

Reference solution (c) Dissolve 25.0 mg of *everolimus CRS* in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (d) Dissolve 5 mg of *everolimus for impurity C identification CRS* in 1 mL of *acetonitrile R*.

Column:
— *size*: *l* = 0.25 m, Ø = 3.0 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, 0.27 g/L solution of *potassium dihydrogen phosphate R* (40:60 *V/V*);
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 7	100 → 72	0 → 28
7 - 25	72 → 60	28 → 40
25 - 33	60 → 0	40 → 100
33 - 43	0	100

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram supplied with *everolimus for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities D, E, F, H, I and J; use the chromatogram supplied with *everolimus for impurity C identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to everolimus (retention time = about 15 min): impurity H = about 0.78; impurity I = about 0.81; impurity C = about 0.87; impurity D = about 0.92; everolimus tautomer = about 1.1; impurity E = about 1.34; impurity J = about 1.38; impurity F = about 1.5.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurity D and everolimus; minimum 1.5 between the peaks due to everolimus and everolimus tautomer.

Calculation of percentage contents:
— for each impurity, use the sum of the concentrations of everolimus and everolimus tautomer in reference solution (b).

Limits:
— *impurities C, D, I*: for each impurity, maximum 0.8 per cent;
— *impurity H*: maximum 0.4 per cent;
— *sum of impurities E and J*: maximum 0.3 per cent;
— *impurity F*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.15 per cent;
— *total*: maximum 2.5 per cent;
— *reporting threshold*: 0.05 per cent; disregard any peak due to everolimus tautomer.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 0.400 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 275 nm.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{53}H_{83}NO_{14}$ taking into account the assigned content of *everolimus CRS* and, if present, the peak due to everolimus tautomer and by subtracting the content of impurity A obtained in the corresponding test.

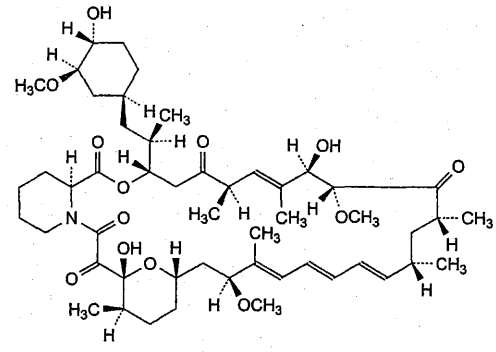
STORAGE

Under an inert gas, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

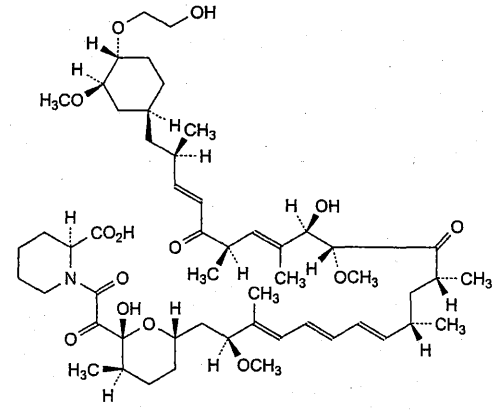
IMPURITIES

Specified impurities *A, C, D, E, F, H, I, J.*

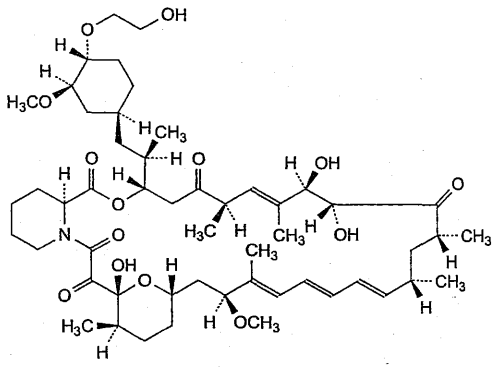
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



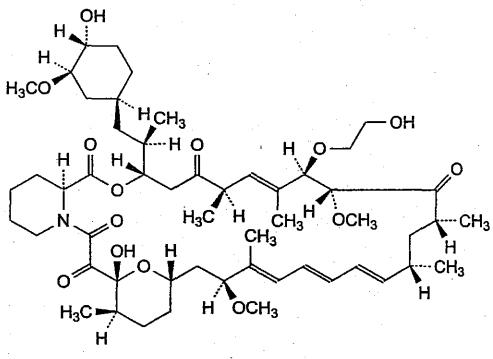
A. (1*R*,9*S*,12*S*,15*R*,16*E*,18*R*,19*R*,21*R*,23*S*,24*E*,26*E*,28*E*,30*S*,32*S*,35*R*)-1,18-dihydroxy-12-[(2*R*)-1-[(1*S*,3*R*,4*R*)-4-hydroxy-3-methoxycyclohexyl]propan-2-yl]-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-11,36-dioxa-4-azatricyclo[30.3.1.0^{4,9}]hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentone (sirolimus; rapamycin),



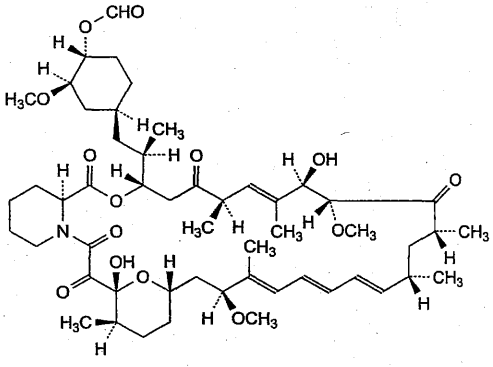
B. (2*S*)-1-[[(2*R*,3*R*,6*S*)-2-hydroxy-6-[(2*S*,3*E*,5*E*,7*E*,9*S*,11*R*,13*R*,14*R*,15*E*,17*R*,19*E*,21*R*)-14-hydroxy-22-[(1*S*,3*R*,4*R*)-4-(2-hydroxyethoxy)-3-methoxycyclohexyl]-2,13-dimethoxy-3,9,11,15,17,21-hexamethyl-12,18-dioxodocosa-3,5,7,15,19-pentaen-1-yl]-3-methyloxan-2-yl](oxo)acetyl]piperidine-2-carboxylic acid,



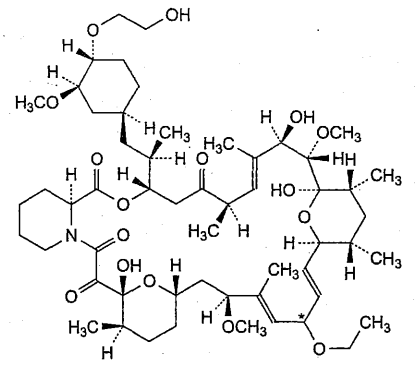
C. (1*R*,9*S*,12*S*,15*R*,16*E*,18*R*,19*R*,21*R*,23*S*,24*E*,26*E*,28*E*,30*S*,32*S*,35*R*)-1,18,19-trihydroxy-12-[(2*R*)-1-[(1*S*,3*R*,4*R*)-4-(2-hydroxyethoxy)-3-methoxycyclohexyl]propan-2-yl]-30-methoxy-15,17,21,23,29,35-hexamethyl-11,36-dioxa-4-azatricyclo[30.3.1.04,9]hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentone (19-*O*-demethyl everolimus),



D. (1*R*,9S,12S,15*R*,16*E*,18*R*,19*R*,21*R*,23*S*,24*E*,26*E*,28*E*,30*S*,32*S*,35*R*)-1-hydroxy-18-(2-hydroxyethoxy)-12-[(2*R*)-1-[(1*S*,3*R*,4*R*)-4-hydroxy-3-methoxycyclohexyl]propan-2-yl]-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-11,36-dioxa-4-azatricyclo[30.3.1.0^{4,9}]hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentone (18-*O*-hydroxyethyl sirolimus),



E. (1*R*,2*R*,4*S*)-4-[(2*R*)-2-[(1*R*,9*S*,12*S*,15*R*,16*E*,18*R*,19*R*,21*R*,23*S*,24*E*,26*E*,28*E*,30*S*,32*S*,35*R*)-1,18-dihydroxy-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-2,3,10,14,20-pentaoxo-11,36-dioxa-4-azatricyclo[30.3.1.0^{4,9}]hexatriaconta-16,24,26,28-tetraen-12-yl]propyl]-2-methoxycyclohexyl formate,



F. (1*R*,9S,12S,15*R*,16*E*,18*R*,19*S*,20*R*,21*R*,23*S*,24*S*,25*E*,27*E*,28*E*,30*S*,32*S*,35*R*)-27-ethoxy-1,18,20-trihydroxy-12-[(2*R*)-1-[(1S,3*R*,4*R*)-4-(2-hydroxyethoxy)-3-methoxycyclohexyl]propan-2-yl]-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-11,36,37-trioxa-4-azatetracyclo[30.3.1.1^{20,24}.0^{4,9}]heptatriaconta-16,25,28-triene-2,3,10,14-tetrone,

H. unknown structure,

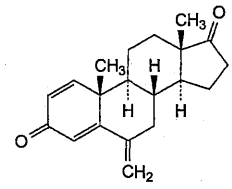
I. unknown structure,

J. unknown structure.

Ph Eur

Exemestane

(Ph. Eur. monograph 2766)



$C_{20}H_{24}O_2$ 296.4 *107868-30-4*

Action and use
Aromatase inhibitor; treatment of breast carcinoma.

Ph Eur

DEFINITION
6-Methylideneandrosta-1,4-diene-3,17-dione.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in methanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *exemestane CRS*.

TESTS
Specific optical rotation (*2.2.7*)
+ 290 to + 298 (dried substance).
Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Store the solutions protected from light.*

Solvent mixture *water R, acetonitrile R* (25:75 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *exemestane for system suitability CRS* (containing impurity G) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of *exemestane CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (3.5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *water R*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	75	25
5 - 35	75 → 55	25 → 45
35 - 45	55 → 5	45 → 95
45 - 50	5	95

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 247 nm.

Injection 10 µL of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *exemestane for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity G.

Relative retention With reference to exemestane (retention time = about 27 min): impurity G = about 1.03.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to exemestane.

Calculation of percentage contents:
- for each impurity, use the concentration of exemestane in reference solution (b).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

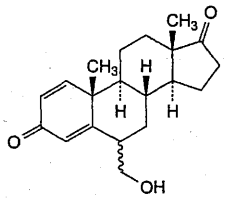
Calculate the percentage content of $C_{20}H_{24}O_2$ taking into account the assigned content of *exemestane CRS*.

STORAGE

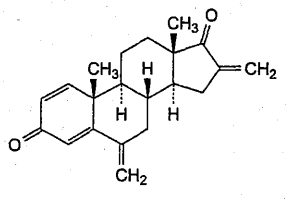
Protected from light.

IMPURITIES

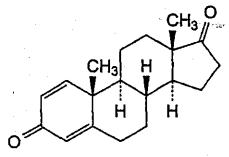
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G.*



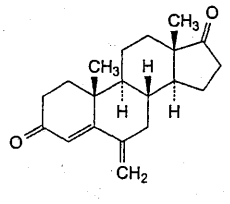
B. 6ξ-(hydroxymethyl)androsta-1,4-diene-3,17-dione,



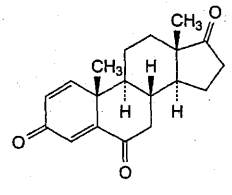
C. 6,16-dimethylideneandrosta-1,4-diene-3,17-dione,



D. androsta-1,4-diene-3,17-dione,



E. 6-methylideneandrost-4-ene-3,17-dione,

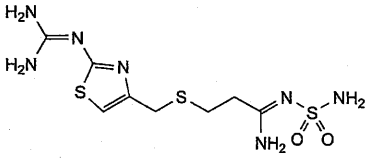


F. androsta-1,4-diene-3,6,17-trione,

G. unknown structure.

Famotidine

(Ph. Eur. monograph 1012)



$C_8H_{15}N_7O_2S_3$ 337.4 *76824-35-6*

Action and use
Histamine H_2 receptor antagonist; treatment of peptic ulceration.

Preparation
Famotidine Tablets

Ph Eur

DEFINITION

3-[[[2-[(Diaminomethylidene)amino]thiazol-4-yl]methyl]sulfanyl]-*N'*-sulfamoylpropanimidamide.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or yellowish-white, crystalline powder or crystals.

Solubility
Very slightly soluble in water, freely soluble in glacial acetic acid, very slightly soluble in anhydrous ethanol, practically insoluble in ethyl acetate. It dissolves in dilute mineral acids.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *famotidine CRS*.

If the spectra obtained show differences, suspend 0.10 g of the substance to be examined and 0.10 g of the reference substance separately in 5 mL of *water R*. Heat to boiling and allow to cool, scratching the wall of the tube with a glass rod to initiate crystallisation. Filter, wash the crystals with 2 mL of iced *water R* and dry in an oven at 80 °C at a pressure not exceeding 670 Pa for 1 h. Record new spectra using the residues.

TESTS

Appearance of solution
Dissolve 0.20 g in a 50 g/L solution of *hydrochloric acid R*, heating to 40 °C if necessary, and dilute to 20 mL with the same acid. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 12.5 mg of the substance to be examined in mobile phase A and dilute to 25.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 2.5 mg of *famotidine impurity D CRS* in *methanol R* and dilute to 10.0 mL with the same solvent. To 1.0 mL of the solution add 0.50 mL of the test solution and dilute to 100.0 mL with mobile phase A.

Reference solution (c) Dissolve 5.0 mg of *famotidine for system suitability CRS* (containing impurities A, B, C, D, F and G) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: mix 6 volumes of *methanol R*, 94 volumes of *acetonitrile R* and 900 volumes of a 1.882 g/L solution of *sodium hexanesulfonate R* previously adjusted to pH 3.5 with *acetic acid R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Flow rate (mL/min)
0 - 23	100 → 96	0 → 4	1
23 - 27	96	4	1 → 2
27 - 47	96 → 78	4 → 22	2

Detection Spectrophotometer at 265 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *famotidine for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, F and G; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to famotidine (retention time = about 21 min): impurity D = about 1.1; impurity C = about 1.2; impurity G = about 1.4; impurity F = about 1.5; impurity A = about 1.6; impurity B = about 2.0.

System suitability:
— *retention time*: famotidine = 19-23 min in all the chromatograms;
— *resolution*: minimum 3.5 between the peaks due to famotidine and impurity D in the chromatogram obtained with reference solution (b).

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.9; impurity B = 2.5; impurity C = 1.9; impurity F = 1.7; impurity G = 1.4;
— *impurities C, D*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities A, B, F, G*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 670 Pa for 5 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.120 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 16.87 mg of $C_8H_{15}N_7O_2S_3$.

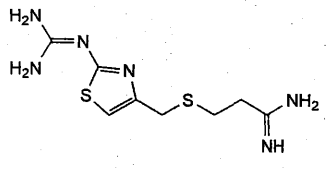
STORAGE

Protected from light.

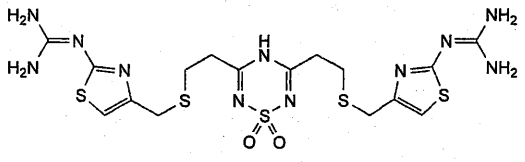
IMPURITIES

Specified impurities *A, B, C, D, F, G.*

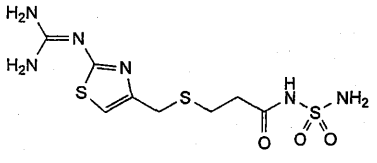
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, H, I, J.*



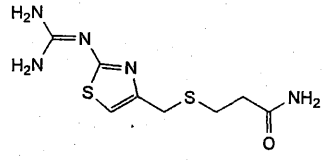
A. 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl]methyl] sulfanyl]propanimidamide,



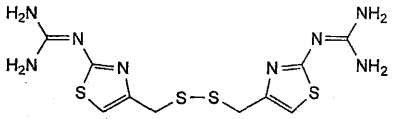
B. 3,5-bis[2-[[[2-[(diaminomethylidene)amino]thiazol-4-yl] methyl]sulfanyl]ethyl]-4*H*-1,2,4,6-thiatriazine 1,1-dioxide,



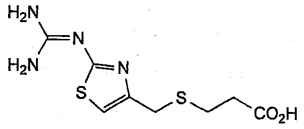
C. 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl]methyl] sulfanyl]-*N*-sulfamoylpropanamide,



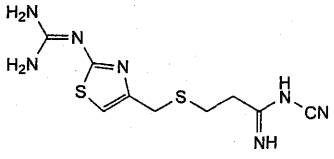
D. 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl]methyl] sulfanyl]propanamide,



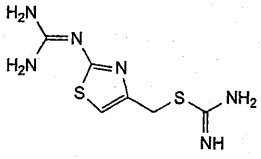
E. 2,2′-[disulfanediylbis(methylenethiazole-4,2-diyl)] diguanidine,



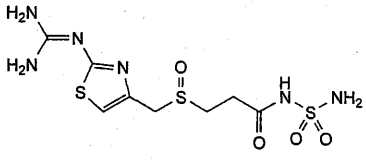
F. 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl]methyl] sulfanyl]propanoic acid,



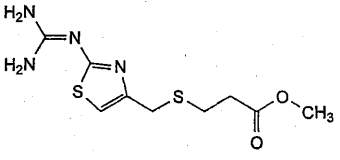
G. *N*-cyano-3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl] methyl]sulfanyl]propanimidamide,



H. [2-[(diaminomethylidene)amino]thiazol-4-yl]methyl carbamimidothioate,



I. 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl]methyl] sulfinyl]-*N*-sulfamoylpropanamide,



J. methyl 3-[[[2-[(diaminomethylidene)amino]thiazol-4-yl] methyl]sulfanyl]propanoate.

Ph Eur

Hard Fat

(Ph. Eur. monograph 0462)

Ph Eur

DEFINITION

Mixture of triglycerides, diglycerides and monoglycerides, which may be obtained either by esterification of hydrogenated fatty acids of vegetable origin with glycerol or by interesterification of hydrogenated vegetable oils.

Each type of hard fat is characterised by its melting point, its hydroxyl value and its saponification value.

It does not contain additives.

CHARACTERS

Appearance

White or almost white, waxy, brittle mass.

Solubility

Practically insoluble in water, slightly soluble in anhydrous ethanol and in methylene chloride.

When heated to 50 °C it melts, giving a colourless or slightly yellowish liquid.

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *ether R, methylene chloride R* (10:90 *V/V*).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

Results The chromatogram shows a spot due to triglycerides with an R_F value of about 0.7 (R_{st} 1) and may show spots due to 1,3-diglycerides (R_{st} 0.6), to 1,2-diglycerides (R_{st} 0.4) and to 1-monoglycerides (R_{st} 0.07). For substances with a low hydroxyl value, the spots due to monoglycerides and diglycerides may be faint or absent. In this case, the test for hydroxyl value (see Tests) is also carried out to confirm identification.

B. Soya bean lecithin, macrogol cetostearyl ether and polysorbate 65. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 0.100 g of *soya bean lecithin R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.100 g of *macrogol cetostearyl ether R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dissolve 0.100 g of *polysorbate 65 R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plates *TLC silica gel G plate R* (2 plates).

Mobile phase *water R, methanol R, methylene chloride R* (4:25:65 *V/V/V*).

Plate 1

Application 4 µL of the test solution and reference solution (a).

Development Over 3/4 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

Results:

- the chromatogram obtained with the test solution shows no spots corresponding to those of soya bean lecithin (R_F = about 0.3 and 0.5) in the chromatogram obtained with reference solution (a);
- the chromatogram obtained with the test solution shows a spot due to triglycerides with an R_F value of about 1.0.

Plate 2

Application 4 µL of the test solution and reference solutions (b) and (c).

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with *potassium iodobismuthate solution R4* and examine in daylight.

Results:

- the chromatogram obtained with the test solution shows no orange spot corresponding to that of macrogol cetostearyl ether (R_F = about 0.85) in the chromatogram obtained with reference solution (b);
- the chromatogram obtained with the test solution shows no orange spot corresponding to that of polysorbate 65 (R_F = about 1.0) in the chromatogram obtained with reference solution (c).

C. Beeswax. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 8 mL with the same solvent.

Reference solution Dissolve 0.100 g of *white beeswax R* in *methylene chloride R* and dilute to 8 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *ethyl acetate R, cyclohexane R* (10:90 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with a freshly prepared 100 g/L solution of *phosphomolybdic acid R* in *ethanol (96 per cent) R* and heat at 120 °C for 3 min; examine in daylight.

Results The chromatogram obtained with the test solution shows no black spot corresponding to that of white beeswax (R_F = about 0.9) in the chromatogram obtained with the reference solution.

TESTS

Alkaline impurities

Melt 10.00 g and while maintaining the temperature at about 50 °C, dissolve the melted mass in 40.0 mL of *ethanol (96 per cent) R*, mix and add 0.05 mL of *bromophenol blue solution R*. Not more than 0.75 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to yellow.

Melting point (*2.2.15*)

30 °C to 45 °C, and within 2 °C of the nominal value.

Introduce the melted substance into a capillary tube and allow to stand at a temperature below 10 °C for 24 h.

Acid value (*2.5.1*)

Maximum 0.5.

Melt 5.0 g and dissolve in 20 mL of the prescribed mixture of solvents.

Hydroxyl value (*2.5.3, Method A*)

Maximum 50, and within 5 units of the nominal value; maximum 5 if the nominal value is less than 5.

Iodine value (*2.5.4, Method A*)

Maximum 3.0.

Peroxide value (*2.5.5, Method A*)

Maximum 3.0.

Saponification value (*2.5.6*)

210 to 260, and within 5 per cent of the nominal value, determined on 2.0 g.

Nickel (*2.4.31*)
Maximum 1 ppm.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.0 g in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (*2.4.14*) on the residue obtained, starting from "Moisten the substance to be examined...".

STORAGE

Protected from light, at a temperature at least 5 °C below the nominal melting point.

LABELLING

The label states:
— the nominal melting point;
— the nominal hydroxyl value;
— the nominal saponification value.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hard fat used as basis for solid dosage forms.

Melting point
(see Tests).

Hydroxyl value
(see Tests).

Saponification value
(see Tests).

Ph Eur

Hard Fat with Additives

(Ph. Eur. monograph 2731)

Ph Eur

DEFINITION

Mixture of triglycerides, diglycerides and monoglycerides, which may be obtained either by esterification of hydrogenated fatty acids of vegetable origin with glycerol or by interesterification of hydrogenated vegetable oils.

Each type of hard fat with additives is characterised by its melting point, its hydroxyl value and its saponification value.

It contains additives such as lecithin, surfactants, beeswax or a mixture thereof.

CHARACTERS

Appearance
White or pale yellow, waxy, brittle mass.

Solubility
Practically insoluble in water, slightly soluble in anhydrous ethanol and in methylene chloride.

When heated to 50 °C it melts, giving a colourless or slightly yellowish liquid.

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *ether R, methylene chloride R* (10:90 *V/V*).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

Results The chromatogram shows a spot due to triglycerides with an R_F value of about 0.7 (R_{st} 1) and may show spots due to 1,3-diglycerides (R_{st} 0.6), to 1,2-diglycerides (R_{st} 0.4) and to 1-monoglycerides (R_{st} 0.07). For substances with a low hydroxyl value, the spots due to monoglycerides and diglycerides may be faint or absent. In this case, the test for hydroxyl value (see Tests) is also carried out to confirm identification.

B. Soya bean lecithin, macrogol cetostearyl ether and polysorbate 65. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 0.100 g of *soya bean lecithin R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.100 g of *macrogol cetostearyl ether R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dissolve 0.100 g of *polysorbate 65 R* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plates *TLC silica gel G plate R* (2 plates).

Mobile phase *water R, methanol R, methylene chloride R* (4:25:65 *V/V/V*).

Plate 1

Application 4 µL of the test solution and reference solution (a).

Development Over 3/4 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

Results:
— in the presence of soya bean lecithin, the chromatogram obtained with the test solution shows 2 spots (R_F = about 0.3 and 0.5) similar in position to the spots in the chromatogram obtained with reference solution (a);
— the chromatogram obtained with the test solution shows a spot due to triglycerides with an R_F value of about 1.0.

Plate 2

Application 4 µL of the test solution and reference solutions (b) and (c).

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with *potassium iodobismuthate solution R4* and examine in daylight.

Results:

— in the presence of macrogol cetostearyl ether, the chromatogram obtained with the test solution shows an orange spot (R_F = about 0.85) similar in position to the spot in the chromatogram obtained with reference solution (b);

— in the presence of polysorbate 65, the chromatogram obtained with the test solution shows an orange spot (R_F = about 1.0) similar in position to the spot in the chromatogram obtained with reference solution (c).

C. Beeswax. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 8 mL with the same solvent.

Reference solution Dissolve 0.100 g of *white beeswax R* in *methylene chloride R* and dilute to 8 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *ethyl acetate R, cyclohexane R* (10:90 *V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with a freshly prepared 100 g/L solution of *phosphomolybdic acid R* in *ethanol (96 per cent) R* and heat at 120 °C for 3 min; examine in daylight.

Results In the presence of beeswax, the chromatogram obtained with the test solution shows a black spot (R_F = about 0.9) similar in position to the spot in the chromatogram obtained with the reference solution.

TESTS

Alkaline impurities

Melt 10.00 g and while maintaining the temperature at about 50 °C, dissolve the melted mass in 40.0 mL of *ethanol (96 per cent) R*, mix and add 0.05 mL of *bromophenol blue solution R*. Not more than 0.75 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to yellow.

If the substance to be examined contains lecithins, not more than 3.75 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to yellow.

Melting point (*2.2.15*)
30 °C to 45 °C, and within 2 °C of the nominal value.

Introduce the melted substance into a capillary tube and allow to stand at a temperature below 10 °C for 24 h.

Acid value (*2.5.1*)
Maximum 1.0.

Melt 5.0 g and dissolve in 20 mL of the prescribed mixture of solvents.

Hydroxyl value (*2.5.3, Method A*)
Maximum 70, and within 5 units of the nominal value.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Peroxide value (*2.5.5, Method A*)
Maximum 3.0.

Saponification value (*2.5.6*)
210 to 260, and within 5 per cent of the nominal value, determined on 2.0 g.

Nickel (*2.4.31*)
Maximum 1 ppm.

Sulfated ash (*2.4.14*)
Maximum 0.4 per cent.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.0 g in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (*2.4.14*) on the residue obtained, starting from "Moisten the substance to be examined...".

STORAGE

Protected from light, at a temperature at least 5 °C below the nominal melting point.

LABELLING

The label states:

— the nominal melting point;

— the nominal hydroxyl value;

— the nominal saponification value.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hard fat with additives used as basis for solid dosage forms.

Melting point
(see Tests).

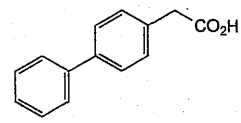
Hydroxyl value
(see Tests).

Saponification value
(see Tests).

Ph Eur

Felbinac

(Ph. Eur. monograph 2304)



$C_{14}H_{12}O_2$ 212.2 *5728-52-9*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION

(Biphenyl-4-yl)acetic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent).

mp

About 164 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *felbinac CRS.*

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Protect the solutions from light and inject within 20 min of preparation.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 5.0 mg of *felbinac impurity A CRS* and 5.0 mg of *biphenyl R* (impurity B) in *methanol R*, add 0.5 mL of the test solution and dilute to 50.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 45 volumes of a 0.1 per cent *V/V* solution of *glacial acetic acid R* and 55 volumes of *methanol R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 3.5 times the retention time of felbinac.

Relative retention With reference to felbinac (retention time = about 15 min): impurity A = about 1.3; impurity B = about 2.8.

System suitability Reference solution:

— *resolution*: minimum 3.0 between the peaks due to felbinac and impurity A.

Limits:

— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent);

— *impurity B*: not more than the area of the peak due to felbinac in the chromatogram obtained with the reference solution (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the peak due to felbinac in the chromatogram obtained with the reference solution (0.10 per cent);

— *total*: not more than twice the area of the peak due to felbinac in the chromatogram obtained with the reference solution (0.2 per cent);

— *disregard limit*: 0.5 times the area of the peak due to felbinac in the chromatogram obtained with the reference solution (0.05 per cent).

Chlorides

Maximum 110 ppm.

Dissolve 1.0 g in 40 mL of *acetone R*, add 6 mL of a 10 per cent *V/V* solution of *nitric acid R*, dilute to 50.0 mL with *water R* and mix. Pour 15.0 mL of this solution as a single addition into 1 mL of *0.1 M silver nitrate* and allow to stand for 5 min protected from light. When viewed horizontally against a black background, any opalescence produced is not more intense than that obtained by treating in the same manner 15.0 mL of a mixture of 1.5 mL of *0.002 M hydrochloric acid*, 40 mL of *acetone R*, 6 mL of 10 per cent *V/V* solution of *nitric acid R*, diluted to 50.0 mL with *water R*.

Sulfates

Maximum 130 ppm.

Dissolve 1.5 g in 40 mL of *dimethylformamide R*, add 1 mL of a 10 per cent *V/V* solution of *hydrochloric acid R*, dilute to 50.0 mL with *dimethylformamide R* and mix. To 15.0 mL of this solution add 2.0 mL of a 120 g/L solution of *barium chloride R* and allow to stand for 5 min. Any opalescence produced is not more intense than that of a standard prepared in the same manner but using 2.0 mL of *0.001 M sulfuric acid* instead of the substance to be examined.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

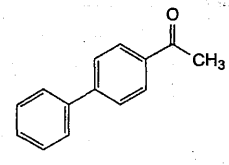
ASSAY

Dissolve 0.160 g in 50 mL of *methanol R*. Titrate with *0.1 M alcoholic potassium hydroxide* determining the end-point potentiometrically (*2.2.20*).

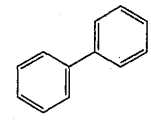
1 mL of *0.1 M alcoholic potassium hydroxide* is equivalent to 21.23 mg of $C_{14}H_{12}O_2$.

IMPURITIES

Specified impurities *A, B.*



A. 4-acetyl biphenyl,

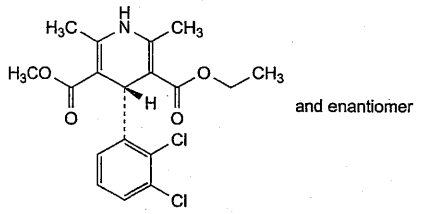


B. biphenyl.

Ph Eur

Felodipine

(Ph. Eur. monograph 1013)



and enantiomer

$C_{18}H_{19}Cl_2NO_4$ 384.3 *72509-76-3*

Action and use
Calcium channel blocker.

Preparation
Felodipine Prolonged-release Tablets

Ph Eur

DEFINITION
Ethyl methyl (4*RS*)-4-(2,3-dichlorophenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or light yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, in anhydrous ethanol, in methanol and in methylene chloride.

IDENTIFICATION
First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50 mg in *methanol R* and dilute to 100 mL with the same solvent. Dilute 3 mL of this solution to 100 mL with *methanol R*.

Spectral range 220-400 nm.

Absorption maxima At 238 nm and 361 nm.

Absorbance ratio A_{361} / A_{238} = 0.34 to 0.36.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *felodipine CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *felodipine CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *nifedipine CRS* in reference solution (a) and dilute to 5 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *ethyl acetate R, cyclohexane R* (40:60 *V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, fluorescence and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.150 g in a mixture of 25 mL of *2-methyl-2-propanol R* and 25 mL of *perchloric acid solution R*. Add 10 mL of *0.1 M cerium sulfate*, allow to stand for 15 min, add 3.5 mL of *strong sodium hydroxide solution R* and neutralise with *dilute sodium hydroxide solution R*. Shake with 25 mL of *methylene chloride R*. Evaporate the lower layer to dryness on a water-bath under nitrogen (the residue is also used in the test for related substances). Dissolve about 20 mg of the residue in *methanol R* and dilute to 50 mL with the same solvent. Dilute 2 mL of this solution to 50 mL with *methanol R*.

Spectral range 220-400 nm.

Absorption maximum At 273 nm.

TESTS
Solution S
Dissolve 1.00 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*).

Absorbance (*2.2.25*)
Maximum 0.10, determined at 440 nm on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 50.0 mg of the residue obtained in identification test D (impurity A) and 25.0 mg of *felodipine CRS* in the mobile phase, then dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.125-0.15 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 20 volumes of *methanol R*, 40 volumes of *acetonitrile R* and 40 volumes of a phosphate buffer solution pH 3.0 containing 0.8 g/L of *phosphoric acid R* and 8 g/L of *sodium dihydrogen phosphate R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time Twice the retention time of felodipine.

Elution order Impurity B, impurity A, felodipine, impurity C.

Retention time Felodipine = about 12 min.

System suitability Reference solution (c):

— *resolution*: minimum 2.5 between the peaks due to impurity A and felodipine.

Limits:

— *sum of impurities B and C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *sum of impurities other than B and C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

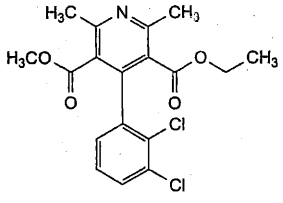
ASSAY
Dissolve 0.160 g in a mixture of 25 mL of *2-methyl-2-propanol R* and 25 mL of *perchloric acid solution R*.
Add 0.05 mL of *ferroin R*. Titrate with *0.1 M cerium sulfate* until the pink colour disappears. Titrate slowly towards the end of the titration.

1 mL of *0.1 M cerium sulfate* is equivalent to 19.21 mg of $C_{18}H_{19}Cl_2NO_4$.

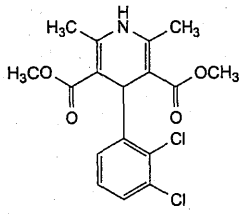
STORAGE
Protected from light.

IMPURITIES
Specified impurities *B, C.*

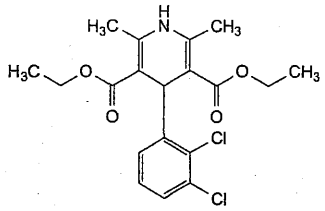
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A.*



A. ethyl methyl 4-(2,3-dichlorophenyl)-2,6-dimethylpyridine-3,5-dicarboxylate,



B. dimethyl 4-(2,3-dichlorophenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate,

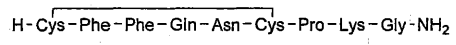


C. diethyl 4-(2,3-dichlorophenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate.

Ph Eur

Felypressin

(Ph. Eur. monograph 1634)



$C_{46}H_{65}N_{13}O_{11}S_2$ 1039 *56-59-7*

Action and use
Vasopressin analogue; vasoconstrictor in local anaesthesia.

Ph Eur

DEFINITION
L-Cysteinyl-L-phenylalanyl-L-phenylalanyl-L-glutaminyl-L-asparaginyl-L-cysteinyl-L-prolyl-L-lysylglycinamide cyclic (1,6)-disulfide.

Synthetic nonapeptide having a vasoconstricting activity. It is available as an acetate.

Content
95.0 per cent to 102.0 per cent (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance
White or almost white, powder or flakes.

Solubility
Freely soluble in water, practically insoluble in acetone and ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

B. Amino acid analysis (*2.2.56*). For hydrolysis use Method 1 and for analysis use Method 1.

Express the content of each amino acid in moles. Calculate the relative proportions of amino acids, taking one-seventh of the sum of the number of moles of glutamic acid, aspartic acid, proline, lysine, glycine and phenylalanine as equal to one. The values fall within the following limits: aspartic acid: 0.9 to 1.1; glutamic acid: 0.9 to 1.1; proline: 0.9 to 1.1; glycine: 0.9 to 1.1; phenylalanine: 1.8 to 2.2; half-cystine: 1.8 to 2.2; lysine: 0.9 to 1.1.

TESTS

Specific optical rotation (*2.2.7*)
−35 to −29, determined at 25 °C (anhydrous and acetic acid-free substance).

Dissolve 20.0 mg in a 1 per cent *V/V* solution of *glacial acetic acid R* and dilute to 10.0 mL with the same solution.

Related substances

Liquid chromatography (*2.2.29*); use the normalisation procedure. *The solutions are stable for 24 h at room temperature or for 1 week at 2-8 °C.*

Test solution (a) Dissolve 5.0 mg of the substance to be examined in 5.0 mL of mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 5.0 mL with mobile phase A.

Reference solution Dissolve the contents of a vial of *felypressin CRS* in mobile phase A to obtain a concentration of 0.2 mg/mL.

Column:
- *size*: *l* = 0.15 m, Ø = 3.9 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm),
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: dissolve 3.62 g of *tetramethylammonium hydroxide R* in 900 mL *water R*; adjust to pH 2.5 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
- *mobile phase B*: dissolve 1.81 g of *tetramethylammonium hydroxide R* in 450 mL of a 50 per cent *V/V* solution of *acetonitrile for chromatography R*; adjust to pH 2.5 with *phosphoric acid R* and dilute to 500 mL with a 50 per cent *V/V* solution of *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	80 → 50	20 → 50
20 - 25	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of test solution (a) and 50 µL of the reference solution.

Identification of impurities Use the chromatogram supplied with *felypressin CRS* to identify the peaks due to impurities A to F.

Relative retention With reference to felypressin: impurity A = about 0.9; impurity B = about 1.1; impurity F = about 1.2; impurity C = about 1.3; impurity D = about 1.4; impurity E = about 2.1.

System suitability Reference solution:
- *retention time*: felypressin = about 7.5 min;
- *resolution*: minimum 1.5 between the peaks due to impurity C and impurity D.

Limits:
- *impurities A, B, C, D, E, F*: for each impurity, maximum 0.5 per cent,
- *any other impurity*: for each impurity, maximum 0.1 per cent,
- *total*: maximum 3.0 per cent,
- *disregard limit*: 0.05 per cent.

Acetic acid (*2.5.34*)

9.0 per cent to 13.0 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Water (*2.5.32*)

Maximum 7.0 per cent.

Bacterial endotoxins (*2.6.14*)

Less than 100 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of test solution (b) and of the reference solution.

Calculate the content of felypressin ($C_{46}H_{65}N_{13}O_{11}S_2$) from the areas of the peaks and the declared content of $C_{46}H_{65}N_{13}O_{11}S_2$ in *felypressin CRS*.

STORAGE

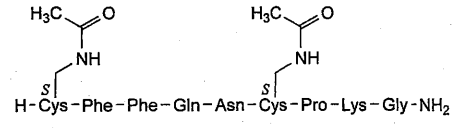
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

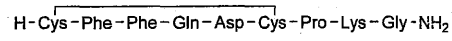
The label states the mass of peptide in the container.

IMPURITIES

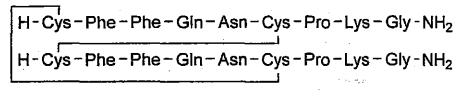
Specified impurities *A, B, C, D, E, F.*



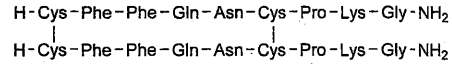
A. S^1,S^6-bis[(acetylamino)methyl]-(reduced felypressin),



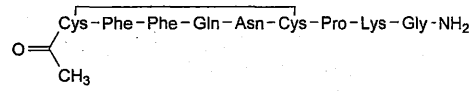
B. [5-aspartic acid]felypressin,



C. bis(reduced felypressin) (1,6′),(1′,6)-bis(disulfide),



D. bis(reduced felypressin) (1,1′),(6,6′)-bis(disulfide),



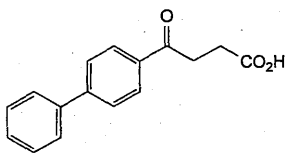
E. N^1-acetylfelypressin,

F. [4-glutamic acid]felypressin.

Ph Eur

Fenbufen

(Ph. Eur. monograph 1209)



$C_{16}H_{14}O_3$ 254.3 *36330-85-5*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION
4-(Biphenyl-4-yl)-4-oxobutanoic acid.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, fine, crystalline powder.

Solubility
Very slightly soluble in water, slightly soluble in acetone, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 186 °C to 189 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fenbufen CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *fenbufen CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *ketoprofen CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent. To 5 mL of this solution, add 5 mL of reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous acetic acid R, ethyl acetate R, hexane R* (5:25:75 *V/V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *dimethylformamide R*, mobile phase A (40:60 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 0.5 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 25 mg of *fenbufen CRS* and 6 mg of *ketoprofen CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture. Dilute 1 mL of this solution to 100 mL with the solvent mixture.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 32 volumes of *acetonitrile R* and 68 volumes of a mixture of 1 volume of *glacial acetic acid R* and 55 volumes of *water R*;
— *mobile phase B*: mix 45 volumes of *acetonitrile R* and 55 volumes of a mixture of 1 volume of *glacial acetic acid R* and 55 volumes of *water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 – 15	100	0
15 – 20	100 → 0	0 → 100
20 – 35	0	100

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to ketoprofen and fenbufen.

Limits:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

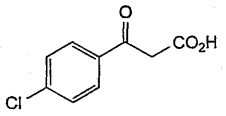
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

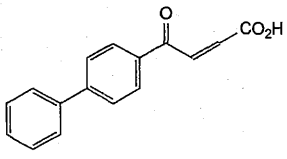
ASSAY
Dissolve 0.200 g in 75 mL of *acetone R* previously neutralised with *phenolphthalein solution R1* and add 50 mL of *water R*. Add 0.2 mL of *phenolphthalein solution R1* and titrate with *0.1 M sodium hydroxide*. Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 25.43 mg of $C_{16}H_{14}O_3$.

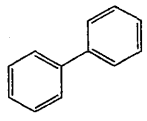
IMPURITIES



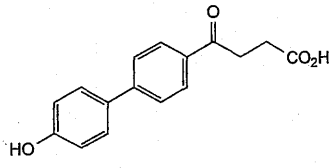
A. 3-(4-chlorophenyl)-3-oxopropanoic acid,



B. 4-(biphenyl-4-yl)-4-oxobut-2-enoic acid,



C. biphenyl,

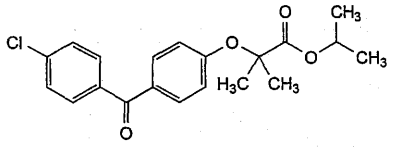


D. 4-(4′-hydroxybiphenyl-4-yl)-4-oxobutanoic acid.

Ph Eur

Fenofibrate

(Ph. Eur. monograph 1322)



$C_{20}H_{21}ClO_4$ 360.8 *49562-28-9*

Action and use
Fibrate; lipid-regulating drug.

Ph Eur

DEFINITION
1-Methylethyl 2-[4-(4-chlorobenzoyl)phenoxy]-2-methylpropanoate.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, very soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
A. Melting point (*2.2.14*): 79 °C to 82 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fenofibrate CRS.*

TESTS

Solution S
To 5.0 g, add 25 mL of *distilled water R* and heat at 50 °C for 10 min. Cool and dilute to 50.0 mL with *distilled water R.* Filter. Use the filtrate as solution S.

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.50 g in *acetone R* and dilute to 10.0 mL with the same solvent.

Acidity
Dissolve 1.0 g in 50 mL of *ethanol (96 per cent) R* previously neutralised using 0.2 mL of *phenolphthalein solution R1.* Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *fenofibrate CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *fenofibrate CRS,* 5.0 mg of *fenofibrate impurity A CRS,* 5.0 mg of *fenofibrate impurity B CRS* and 10.0 mg of *fenofibrate impurity G CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Column:
— *size: l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 30 volumes of *water R* acidified to pH 2.5 with *phosphoric acid R* and 70 volumes of *acetonitrile R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 286 nm.

Injection 20 µL of the test solution and reference solution (b).

Run time Twice the retention time of fenofibrate.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and G.

Relative retention With reference to fenofibrate (retention time = about 10 min): impurity A = about 0.34; impurity B = about 0.36; impurity G = about 1.35.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurities A and B.

Limits:
— *impurities A, B*: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *impurity G*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the peak due to fenofibrate in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 5 times the area of the peak due to fenofibrate in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the peak due to fenofibrate in the chromatogram obtained with reference solution (b) (0.05 per cent).

Halides expressed as chlorides (*2.4.4*)
Maximum 100 ppm.

To 5 mL of solution S add 10 mL of *distilled water R*.

Sulfates (*2.4.13*)
Maximum 100 ppm, determined on solution S.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

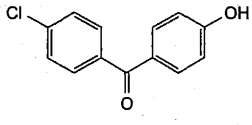
Injection 5 µL of the test solution and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 1.0 per cent determined on 6 injections.

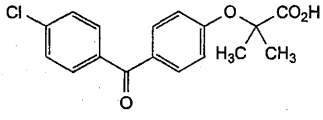
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B, G.*

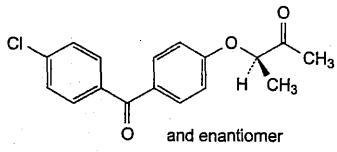
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, F.*



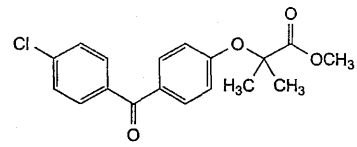
A. (4-chlorophenyl)(4-hydroxyphenyl)methanone,



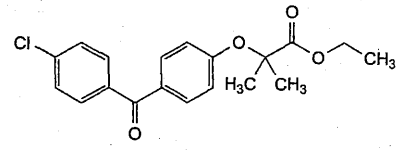
B. 2-[4-(4-chlorobenzoyl)phenoxy]-2-methylpropanoic acid (fenofibric acid),



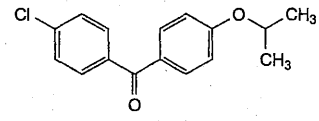
C. (3*RS*)-3-[4-(4-chlorobenzoyl)phenoxy]butan-2-one,



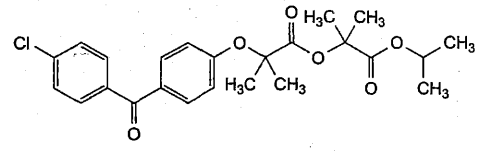
D. methyl 2-[4-(4-chlorobenzoyl)phenoxy]-2-methylpropanoate,



E. ethyl 2-[4-(4-chlorobenzoyl)phenoxy]-2-methylpropanoate,



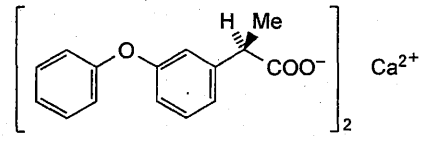
F. (4-chlorophenyl)[4-(1-methylethoxy)phenyl]methanone,



G. 1-methylethyl 2-[[2-[4-(4-chlorobenzoyl)phenoxy]-2-methylpropanoyl]oxy]-2-methylpropanoate.

Ph Eur

Fenoprofen Calcium



$(C_{15}H_{13}O_3)_2Ca,2H_2O$ 558.6 *34957-40-5*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparation
Fenoprofen Tablets

DEFINITION
Fenoprofen Calcium is calcium (*RS*)-2-(3-phenoxyphenyl) propionate dihydrate. It contains not less than 97.5% and not more than 101.0% of $(C_{15}H_{13}O_3)_2Ca$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Slightly soluble in *water*; soluble in *ethanol (96%)*.

IDENTIFICATION
A. Dissolve 0.1 g in 5 mL of *glacial acetic acid* and add sufficient *methanol* to produce 100 mL. Dilute 5 mL of this solution to 50 mL with *methanol*. The *light absorption* of the resulting solution, Appendix II B, in the range 230 to 350 nm exhibits two maxima, at 272 nm and 278 nm, and a

shoulder at 266 nm. The *absorbance* at the maximum at 272 nm is about 0.70 and at the maximum at 278 nm is about 0.65.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of fenoprofen calcium *(RS 142)*.

C. The residue on ignition yields the reactions characteristic of *calcium salts*, Appendix VI.

TESTS

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in mobile phase.

(1) 0.50% w/v of the substance being examined.

(2) 0.0025% w/v of the substance being examined.

(3) 0.04% w/v of *fenoprofen calcium* and 0.0015% w/v of *4,4′-dimethoxybenzophenone*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (7 to 8 µm) (Zorbax ODS is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 270 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for 3 times the retention time of the peak due to fenoprofen.

MOBILE PHASE

2 volumes of *glacial acetic acid*, 7 volumes of *tetrahydrofuran*, 30 volumes of *acetonitrile* and 61 volumes of *water*.

SYSTEM SUITABILITY

The test is valid if the *resolution factor* between the peaks corresponding to fenoprofen and 4,4′-dimethoxybenzophenone in the chromatogram obtained with solution (3) is at least 3.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any *secondary peak* is not greater than twice the area of the peak in the chromatogram obtained with solution (2) (1%);

not more than one *secondary peak* has an area greater than the area of the peak in the chromatogram obtained with solution (2) (0.5%);

the sum of the areas of all *secondary peaks* is not greater than four times the area of the peak in the chromatogram obtained with solution (2) (2%).

Water

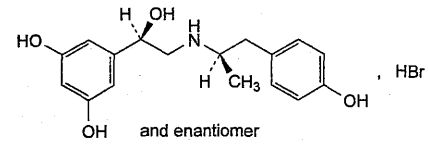
5.0 to 8.0% w/w, Appendix IX C. Use 0.2 g.

ASSAY

Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.5 g and determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 26.13 mg of $(C_{15}H_{13}O_3)_2Ca$.

Fenoterol Hydrobromide

(Ph. Eur. monograph 0901)



$C_{17}H_{22}BrNO_4$ 384.3 *1944-12-3*

Action and use

$Beta_2$-adrenoceptor agonist; bronchodilator.

Ph Eur

DEFINITION

5-[(1*RS*)-2-[(1*RS*)-2-(4-Hydroxyphenyl)-1-methylethyl]amino-1-hydroxyethyl]benzene-1,3-diol hydrobromide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fenoterol hydrobromide CRS*.

B. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S

Dissolve 2.00 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)

4.2 to 5.2 for solution S.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 6.0 mg of the substance to be examined in *water R* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 6 mg of *fenoterol hydrobromide CRS* (containing impurity A) in *water R* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dissolve the contents of a vial of *fenoterol for peak identification CRS* (containing impurities B and C) in 1.0 mL of *water R*.

Reference solution (c) Dilute 1.0 mL of the test solution to 25.0 mL with *water R*.

Reference solution (d) Dilute 0.5 mL of reference solution (c) to 10.0 mL with *water R*.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 24 g of *anhydrous disodium hydrogen phosphate R* in 1000 mL of *water R*. Mix 69 volumes of the

solution and 1 volume of a 9 g/L solution of *potassium dihydrogen phosphate R*, adjust to pH 8.5 with *phosphoric acid R* and add 35 volumes of *methanol R2*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 3 times the retention time of fenoterol.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *fenoterol for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

Relative retention With reference to fenoterol (retention time = about 7 min): impurity A = about 1.3; impurity B = about 2.0; impurity C = about 2.2.

System suitability:

— *resolution*: minimum 3 between the peaks due to fenoterol and impurity A in the chromatogram obtained with reference solution (a); minimum 1.5 between the peaks due to impurities B and C in the chromatogram obtained with reference solution (b).

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 0.6;

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (4.0 per cent);

— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.10 per cent);

— *sum of impurities other than A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Iron (*2.4.9*)
Maximum 10 ppm.

Dissolve the residue obtained in the test for sulfated ash in 2.5 mL of *dilute hydrochloric acid R* and dilute to 10 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.600 g in 50 mL of *water R* and add 5 mL of *dilute nitric acid R*, 25.0 mL of *0.1 M silver nitrate* and 2 mL of *ferric ammonium sulfate solution R2*. Shake and titrate with *0.1 M ammonium thiocyanate* until an orange colour is obtained. Carry out a blank titration.

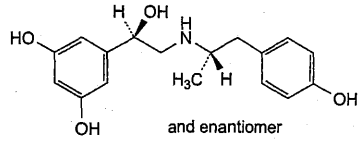
1 mL of *0.1 M silver nitrate* is equivalent to 38.43 mg of $C_{17}H_{22}BrNO_4$.

STORAGE

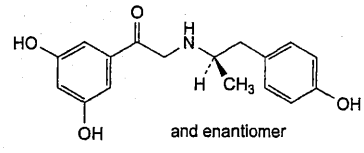
Protected from light.

IMPURITIES

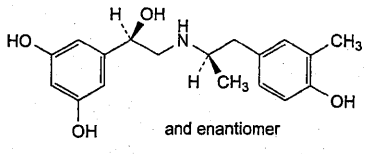
Specified impurities *A, B, C.*



A. 5-[(1*RS*)-2-[(1*SR*)-2-(4-hydroxyphenyl)-1-methylethyl]amino-1-hydroxyethyl]benzene-1,3-diol,



B. 1-(3,5-dihydroxyphenyl)-2-[[(1*RS*)-2-(4-hydroxyphenyl)-1-methylethyl]amino]ethan-1-one,

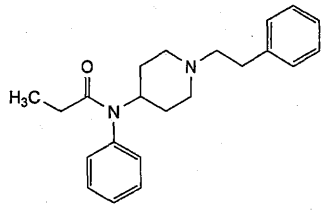


C. 5-[(1*RS*)-2-[(1*RS*)-2-(4-hydroxy-3-methylphenyl)-1-methylethyl]amino-1-hydroxyethyl]benzene-1,3-diol.

Ph Eur

Fentanyl

(Ph. Eur. monograph 1210)



$C_{22}H_{28}N_2O$ 336.5 *437-38-7*

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION

N-Phenyl-*N*-[1-(2-phenylethyl)piperidin-4-yl]propanamide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent), in methanol and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fentanyl CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *anhydrous ethanol R*, evaporate to dryness at room temperature in a current of air and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, water R* (50:50 *V/V*).

Test solution Dissolve 25 mg of the substance to be examined in 2.5 mL of *methanol R* and dilute to 5.0 mL with *water R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Dissolve the contents of a vial of *fentanyl impurity mixture CRS* (impurities C and D) in 1.0 mL of this solution, using sonication for 1 min if necessary.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.15 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped ethylene-bridged phenylsilyl silica gel for chromatography (hybrid material) R* (1.7 µm);
- *temperature*: 45 °C.

Mobile phase:
- *mobile phase A*: dissolve 0.8 g of *ammonium acetate R* in *water for chromatography R*, add 2 mL of *trifluoroacetic acid R* and dilute to 1000.0 mL with *water for chromatography R*;
- *mobile phase B*: add 2 mL of *trifluoroacetic acid R* to 950 mL of *acetonitrile R1* and shake to homogenise; dissolve 0.8 g of *ammonium acetate R* in this solution and shake until complete dissolution, then dilute to 1000.0 mL with *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	98	2
1 - 5	98 → 90	2 → 10
5 - 17	90 → 70	10 → 30
17 - 30	70 → 5	30 → 95
30 - 31	5	95

Flow rate 0.35 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *fentanyl impurity mixture CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities C and D.

Relative retention With reference to fentanyl (retention time = about 19 min): impurity D = about 0.8; impurity C = about 0.9.

System suitability Reference solution (a):
- *resolution*: minimum 5.0 between the peaks due to impurity C and fentanyl.

Calculation of percentage contents:
- for each impurity, use the concentration of fentanyl in reference solution (b).

Limits:
- *impurities C, D*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.4 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 50 °C.

ASSAY

Dissolve 0.200 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 33.65 mg of $C_{22}H_{28}N_2O$.

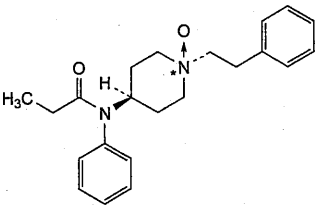
STORAGE

Protected from light.

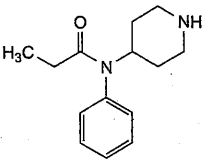
IMPURITIES

Specified impurities *C, D.*

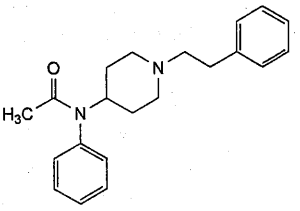
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E, F, G, H, I, J, K, L.*



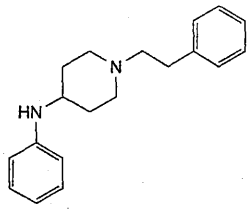
A. (1*rs*,4*rs*)-1-(2-phenylethyl)-4-(*N*-phenylpropanamido) piperidine *N*-oxide,



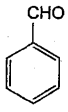
B. *N*-phenyl-*N*-(piperidin-4-yl)propanamide,



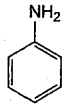
C. *N*-phenyl-*N*-[1-(2-phenylethyl)piperidin-4-yl]acetamide,



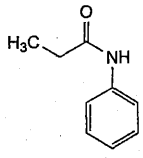
D. *N*-phenyl-1-(2-phenylethyl)piperidin-4-amine,



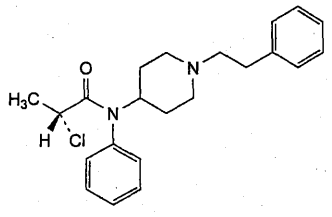
E. benzaldehyde,



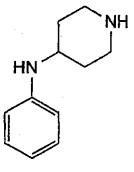
F. aniline (phenylamine),



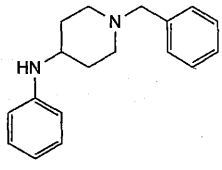
G. *N*-phenylpropanamide,



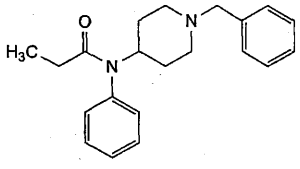
H. (2*RS*)-2-chloro-*N*-phenyl-*N*-[1-(2-phenylethyl)piperidin-4-yl]propanamide,



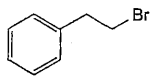
I. *N*-phenylpiperidin-4-amine,



J. 1-benzyl-*N*-phenylpiperidin-4-amine,



K. *N*-(1-benzylpiperidin-4-yl)-*N*-phenylpropanamide,

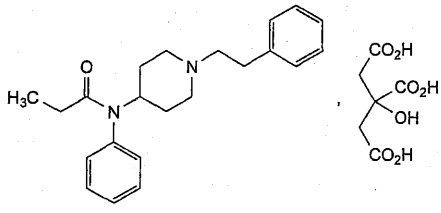


L. (2-bromoethyl)benzene.

Ph Eur

Fentanyl Citrate

(Ph. Eur. monograph 1103)



$C_{28}H_{36}N_2O_8$ 528.6 *990-73-8*

Action and use
Opioid receptor agonist; analgesic.

Preparations
Bupivacaine and Fentanyl Injection
Fentanyl Injection

Ph Eur

DEFINITION
N-Phenyl-*N*-[1-(2-phenylethyl)piperidin-4-yl]propanamide dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Soluble in water, freely soluble in methanol, sparingly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

mp
About 152 °C, with decomposition.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *fentanyl citrate CRS*.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.2 g of the substance to be examined in *water R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *methanol R, water R* (50:50 *V/V*).
Test solution Dissolve 39.3 mg of the substance to be examined in 2.5 mL of *methanol R* and dilute to 5.0 mL with *water R*.
Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the solvent mixture. Dissolve the contents of a vial of *fentanyl impurity mixture CRS* (impurities C and D) in

1.0 mL of this solution, using sonication for 1 min if necessary.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped ethylene-bridged phenylsilyl silica gel for chromatography (hybrid material) R* (1.7 µm);
— *temperature*: 45 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.8 g of *ammonium acetate R* in *water for chromatography R*, add 2 mL of *trifluoroacetic acid R* and dilute to 1000.0 mL with *water for chromatography R*;
— *mobile phase B*: add 2 mL of *trifluoroacetic acid R* to 950 mL of *acetonitrile R1* and shake to homogenise; dissolve 0.8 g of *ammonium acetate R* in this solution and shake until complete dissolution, then dilute to 1000.0 mL with *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	98	2
1 - 5	98 → 90	2 → 10
5 - 17	90 → 70	10 → 30
17 - 30	70 → 5	30 → 95
30 - 31	5	95

Flow rate 0.35 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied with *fentanyl impurity mixture CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities C and D.

Relative retention With reference to fentanyl (retention time = about 19 min): impurity D = about 0.8; impurity C = about 0.9.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to impurity C and fentanyl.

Calculation of percentage contents:
— for each impurity, use the concentration of fentanyl in reference solution (b).

Limits:
— *impurities C, D*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 52.86 mg of $C_{28}H_{36}N_2O_8$.

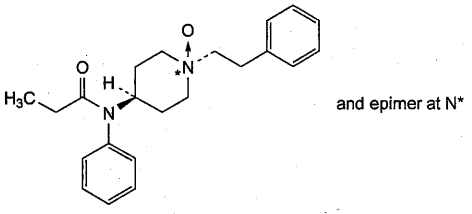
STORAGE

Protected from light.

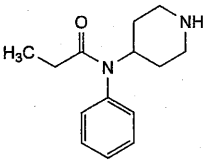
IMPURITIES

Specified impurities *C, D.*

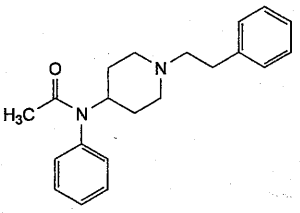
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E, F, G, H, I.*



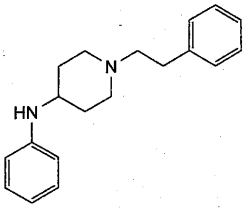
A. (1*rs*,4*rs*)-1-(2-phenylethyl)-4-(*N*-phenylpropanamido) piperidine *N*-oxide,



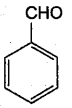
B. *N*-phenyl-*N*-(piperidin-4-yl)propanamide,



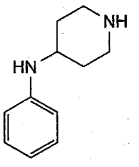
C. *N*-phenyl-*N*-[1-(2-phenylethyl)piperidin-4-yl]acetamide,



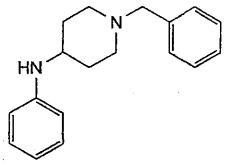
D. *N*-phenyl-1-(2-phenylethyl)piperidin-4-amine,



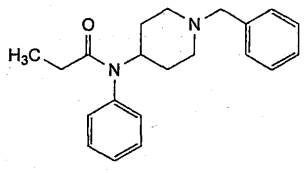
E. benzaldehyde,



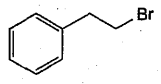
F. *N*-phenylpiperidin-4-amine,



G. 1-benzyl-*N*-phenylpiperidin-4-amine,



H. *N*-(1-benzylpiperidin-4-yl)-*N*-phenylpropanamide,

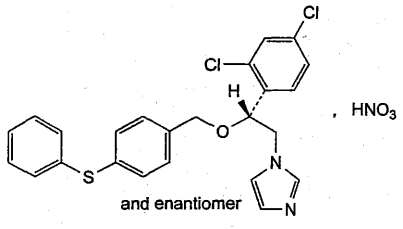


I. (2-bromoethyl)benzene.

Ph Eur

Fenticonazole Nitrate

(Ph. Eur. monograph 1211)



$C_{24}H_{21}Cl_2N_3O_4S$ 518.4 *73151-29-8*

Action and use
Antifungal.

Ph Eur

DEFINITION
1-[(2*RS*)-2-(2,4-Dichlorophenyl)-2-[[4-(phenylsulfanyl)benzyl]oxy]ethyl]-1*H*-imidazole nitrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in dimethylformamide and in methanol, sparingly soluble in anhydrous ethanol.

IDENTIFICATION
First identification: C, D.
Second identification: A, B, D.
A. Melting point (*2.2.14*): 134 °C to 137 °C.
B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).
Test solution Dissolve 20.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R*.
Spectral range 230-350 nm.
Absorption maximum At 252 nm.
Shoulder At about 270 nm.
Absorption minimum At 236 nm.
Specific absorbance at the absorption maximum 260 to 280.
C. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *fenticonazole nitrate CRS*.
D. It gives the reaction of nitrates (*2.3.1*).

TESTS

Optical rotation (*2.2.7*)
−0.10° to + 0.10°.
Dissolve 0.10 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.
Reference solution (b) Dilute 10.0 mL of reference solution (a) to 25.0 mL with the mobile phase.
Reference solution (c) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.
Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dissolve the contents of a vial of *fenticonazole impurity D CRS* in 1.0 mL of this solution.
Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).
Mobile phase Mix 70 volumes of *acetonitrile R1* and 30 volumes of a phosphate buffer solution prepared by dissolving 3.4 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*, adjusting to pH 3.0 with *phosphoric acid R* and diluting to 1000 mL with *water R*.
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 229 nm.
Injection 10 µL.
Run time 5.5 times the retention time of fenticonazole.
System suitability:
— *resolution*: minimum 2.0 between the peaks due to impurity D and fenticonazole in the chromatogram obtained with reference solution (d);
— *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (c).
Limits:
— *impurities A, B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard the peak due to the nitric ion (which corresponds to the dead volume of the column).

Toluene
Head-space gas chromatography (*2.2.28*): use the standard additions method.

Test solution Disperse 0.2 g of the substance to be examined in a 10 mL vial with 5 mL of *water R*.

Reference solution Mix 4 mg of *toluene R* with *water R* and dilute to 1000 mL with the same solvent. Place 5 mL of this solution in a 10 mL vial.

Column:
— *size*: *l* = 25 m, Ø = 0.32 mm;
— *stationary phase*: *poly(cyanopropyl) (7) (phenyl) (7) (methyl) (86)siloxane R* (film thickness 1.2 µm).

Carrier gas *helium for chromatography R.*

Split ratio 1:25.

Column head pressure 40 kPa.

Static head-space conditions which may be used:
— *equilibration temperature*: 90 °C;
— *equilibration time*: 1 h.

Temperature:
— *column*: 80 °C;
— *injection port*: 180 °C;
— *detector*: 220 °C.

Detection Flame ionisation.

Injection 1 mL of the gaseous phase.

Limit:
— *toluene*: maximum 100 ppm.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

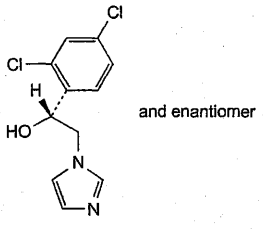
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.450 g in 50 mL of a mixture of equal volumes of *anhydrous acetic acid R* and *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

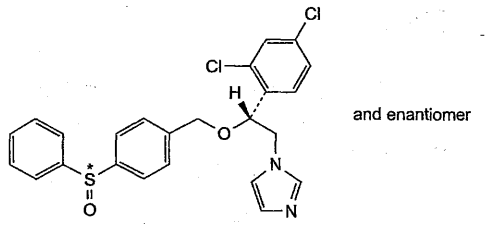
1 mL of *0.1 M perchloric acid* is equivalent to 51.84 mg of $C_{24}H_{21}Cl_2N_3O_4S$.

STORAGE
Protected from light.

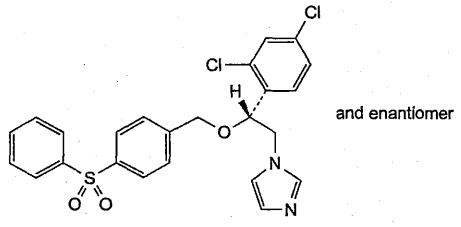
IMPURITIES
Specified impurities *A, B, C, D, E.*



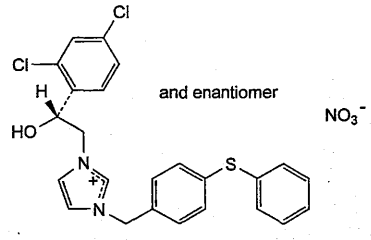
A. (*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl)ethanol,



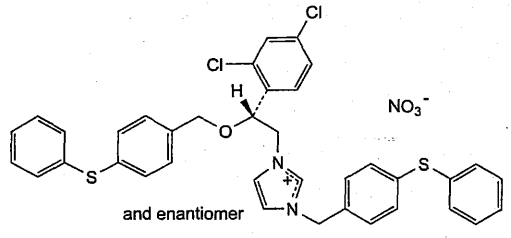
B. 1-[(2*RS*)-2-(2,4-dichlorophenyl)-2-[[4-(benzenesulfinyl)benzyl]oxy]ethyl]-1*H*-imidazole,



C. 1-[(2*RS*)-2-(2,4-dichlorophenyl)-2-[[4-(benzenesulfonyl)benzyl]oxy]ethyl]-1*H*-imidazole,



D. (*RS*)-1-[2-(2,4-dichlorophenyl)-2-hydroxyethyl]-3-[4-(phenylsulfanyl)benzyl]imidazolium nitrate,



E. (*RS*)-1-[2-(2,4-dichlorophenyl)-2-[4-(phenylsulfanyl)benzyloxy]ethyl]-3-[4-(phenylsulfanyl) benzyl]imidazolium nitrate.

Ph Eur

Products of Fermentation

(Ph. Eur. monograph 1468)

Ph Eur

This monograph applies to indirect gene products obtained by fermentation. It is not applicable to:
— *monographs in the Pharmacopoeia concerning vaccines for human or veterinary use;*
— *products derived from continuous cell lines of human or animal origin;*
— *direct gene products that result from the transcription and translation from nucleic acid to protein, whether or not subject to post-translational modification;*
— *products obtained by semi-synthesis from a product of fermentation and those obtained by biocatalytic transformation;*

— *whole broth concentrates or raw fermentation products.*

This monograph provides general requirements for the development and manufacture of products of fermentation. These requirements are not necessarily comprehensive in a given case and requirements complementary or additional to those prescribed in this monograph may be imposed in an individual monograph or by the competent authority.

DEFINITION

For the purposes of this monograph, products of fermentation are active or inactive pharmaceutical substances produced by controlled fermentation as indirect gene products. They are primary or secondary metabolites of micro-organisms such as bacteria, yeasts, fungi and micro-algae, whether or not modified by traditional procedures or recombinant DNA (rDNA) technology. Such metabolites include vitamins, amino acids, antibiotics, alkaloids and polysaccharides.

They may be obtained by batch or continuous fermentation processes followed by procedures such as extraction, concentration, purification and isolation.

PRODUCTION

Production is based on a process that has been validated and shown to be suitable. The extent of validation depends on the critical nature of the respective process step.

CHARACTERISATION OF THE PRODUCER MICRO-ORGANISM

The history of the micro-organism used for production is documented. The micro-organism is adequately characterised. This may include determination of the phenotype of the micro-organism, macroscopic and microscopic methods and biochemical tests and, if appropriate, determination of the genotype of the micro-organism and molecular genetic tests.

PROCESSES USING A SEED-LOT SYSTEM

The *master cell bank* is a homogeneous suspension or lyophilisate of the original cells distributed into individual containers for storage. The viability and productivity of the cells under the selected storage conditions and their suitability for initiating a satisfactory production process after storage must be demonstrated.

Propagation of the master cell bank may take place through a seed-lot system that uses a working cell bank.

The *working cell bank* is a homogeneous suspension or lyophilisate of the cell material derived from the master cell bank, distributed in equal volumes into individual containers for storage (for example, in liquid nitrogen).

Production may take place by batch or continuous culture and may be terminated under defined conditions.

All containers in a cell bank are stored under identical conditions. Once removed from storage, the individual ampoules, vials or culture straws are not returned to the cell bank.

PROCESSES USING STAGED GROWTH IN CULTURES

The contents of a container of the working cell bank are used, if necessary after resuspension, to prepare an inoculum in a suitable medium. After a suitable period of growth, the cultures are used to initiate the fermentation process, if necessary following preculture in a prefermentor.
The conditions to be used at each stage of the process are defined and must be met with each production run.

CHANGE CONTROL

If the production process is altered in a way that causes a significant change in the impurity profile of the product, the critical steps associated with this change in impurity profile are revalidated.

If a significant change has taken place in the micro-organism used for production that causes a significant change in the impurity profile of the product, the critical steps of the production process associated with this change, particularly the procedure for purification and isolation, are revalidated.

Revalidation includes demonstration that new impurities present in the product as a result of the change are adequately controlled by the test procedures. If necessary, additional or alternative tests must be introduced with appropriate limits. If the change in the process or in the micro-organism results in an increase in the level of an impurity already present, the acceptability of such an increase is addressed.

When a master cell bank is replaced, the critical steps of the production process must be revalidated to the extent necessary to demonstrate that no adverse change has occurred in the quality and safety of the product. Particular attention must be given to possible changes in the impurity profile of the product if a modified or new micro-organism is introduced into the process.

RAW MATERIALS

The raw materials employed in the fermentation and/or down-stream processing are of suitable quality for the intended purpose. They are tested to ensure that they comply with written specifications. Special attention must be paid to the levels of free histidine in fish peptones as the presence of free histidine may lead to histamine formation in certain conditions.

Levels of bioburden in media or in the inlet air for aeration are reduced to an adequately low level to ensure that if microbial contamination occurs, it does not adversely affect the quality, purity and safety of the product. Addition of components such as nutrients, precursors, and substrates during fermentation takes place aseptically.

IN-PROCESS CONTROLS

In-process controls are in place to ensure the consistency of the conditions during fermentation and down-stream processing and of the quality of the isolated product. Particular attention must be paid to ensure that any microbial contamination that adversely affects the quality, purity and safety of the product is detected by the controls applied.

Production conditions may be monitored, as appropriate, by suitable procedures for example to control and check:
— temperature,
— pH,
— rate of aeration,
— rate of agitation,
— pressure,
and to monitor the concentration of the required product.

DOWN-STREAM PROCESSING

At the end of fermentation, the producer micro-organism is inactivated or removed. Further processing is designed to reduce residues originating from the culture medium to an acceptable level and to ensure that the desired product is recovered with consistent quality.

Various purification processes may be used, for example, charcoal treatment, ultrafiltration and solvent extraction.

It must be demonstrated that the process or processes chosen reduce to a minimum or remove:
— residues from the producer micro-organism, culture media, substrates and precursors,
— unwanted transformation products of substrates and precursors.

If necessary, suitable tests are performed either as in-process controls or on the isolated product of fermentation.

IDENTIFICATION, TESTS AND ASSAY

The requirements with which the product must comply throughout its period of validity, as well as specific test methods, are stated in the individual monographs.

Ph Eur

Ferric Chloride Hexahydrate

(Ph. Eur. monograph 1515)

$FeCl_3,6H_2O$ 270.3 *10025-77-1*

Preparation

Ferric Chloride Injection

Ph Eur

DEFINITION

Content

98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

Crystalline mass or orange-yellow or brownish-yellow crystals, very hygroscopic.

Solubility

Very soluble in water and in ethanol (96 per cent), freely soluble in glycerol.

IDENTIFICATION

A. It gives reaction (a) of chlorides (*2.3.1*).

B. It gives reaction (c) of iron (*2.3.1*).

TESTS

Solution S

Dissolve 10 g in *distilled water R* and dilute to 100 mL with the same solvent.

Acidity

In a suitable polyethylene container, dissolve 3.0 g of *potassium fluoride R* in 15 mL of *water R*. Titrate with *0.1 M sodium hydroxide* using 0.1 mL of *phenolphthalein solution R* as indicator until a pink colour is obtained. Add 10 mL of solution S and allow to stand for 3 h. Filter and use 12.5 mL of the filtrate. Not more than 0.30 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Free chlorine

Heat 5 mL of solution S. The vapour does not turn *starch iodide paper R* blue.

Sulfates (*2.4.13*)

Maximum 100 ppm.

Heat 15 mL of solution S on a water-bath and add 5 mL of *strong sodium hydroxide solution R*. Allow to cool and filter. Neutralise the filtrate to *blue litmus paper R* using *hydrochloric acid R1* and evaporate to 15 mL.

Ferrous ions

Maximum 50 ppm.

To 10 mL of solution S, add 1 mL of *water R*, and 0.05 mL of *potassium ferricyanide solution R* followed by 4 mL of *phosphoric acid R*. After 10 min, any blue colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of *water R* and 1 mL of a freshly prepared 0.250 g/L solution of *ferrous sulfate R*.

ASSAY

In a conical flask with a ground-glass stopper, dissolve 0.200 g in 20 mL of *water R*. Add 10 mL of *dilute hydrochloric acid R* and 2 g of *potassium iodide R*. Allow the stoppered flask to stand for 1 h protected from light. Titrate with *0.1 M sodium thiosulfate*, adding 5 mL of *starch solution R* towards the end of the titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 27.03 mg of $FeCl_3,6H_2O$.

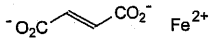
STORAGE

In an airtight container, protected from light.

Ph Eur

Ferrous Fumarate

(Ph. Eur. monograph 0902)



$C_4H_2FeO_4$ 169.9 *141-01-5*

Action and use

Used in prevention and treatment of anaemias.

Preparations

Ferrous Fumarate Capsules

Ferrous Fumarate Oral Suspension

Ferrous Fumarate Tablets

Ferrous Fumarate and Folic Acid Tablets

Ph Eur

DEFINITION

Iron(II) (*E*)-butenedioate.

Content

93.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Fine, reddish-orange or reddish-brown powder.

Solubility

Slightly soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Test solution To 1.0 g add 25 mL of a mixture of equal volumes of *hydrochloric acid R* and *water R* and heat on a water-bath for 15 min. Cool and filter. Use the filtrate for identification test C. Wash the residue with 50 mL of a mixture of 1 volume of *dilute hydrochloric acid R* and 9 volumes of *water R* and discard the washings. Dry the residue at 100-105 °C. Dissolve 20 mg of the residue in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *fumaric acid CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous formic acid R, methylene chloride R, butanol R, heptane R* (12:16:32:44 *V/V/V/V*).

Application 5 µL.

Development In an unsaturated tank, over a path of 10 cm.

Drying At 105 °C for 15 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

B. Mix 0.5 g with 1 g of *resorcinol R*. To 0.5 g of the mixture in a crucible add 0.15 mL of *sulfuric acid R* and heat gently. A dark red semi-solid mass is formed. Add the mass, with care, to 100 mL of *water R*. An orange-yellow colour develops and the solution shows no fluorescence.

C. The filtrate obtained during preparation of the test solution in identification test A gives reaction (a) of iron (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in a mixture of 10 mL of *lead-free hydrochloric acid R* and 80 mL of *water R*, heating slightly if necessary. Allow to cool, filter if necessary and dilute to 100 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 0.2 per cent.

Heat 0.15 g with 8 mL of *dilute hydrochloric acid R* and 20 mL of *distilled water R*. Cool in iced water, filter and dilute to 30 mL with *distilled water R*.

Arsenic (*2.4.2, Method A*)

Maximum 5 ppm.

Mix 1.0 g with 15 mL of *water R* and 15 mL of *sulfuric acid R*. Warm to precipitate the fumaric acid completely. Cool and add 30 mL of *water R*. Filter. Wash the precipitate with *water R*. Dilute the combined filtrate and washings to 125 mL with *water R*. 25 mL of the solution complies with the test.

Ferric ion

Maximum 2.0 per cent.

In a flask with a ground-glass stopper, dissolve 3.0 g in a mixture of 10 mL of *hydrochloric acid R* and 100 mL of *water R* by heating rapidly to boiling. Boil for 15 s. Cool rapidly, add 3 g of *potassium iodide R*, stopper the flask and allow to stand protected from light for 15 min. Add 2 mL of *starch solution R* as indicator. Titrate the liberated iodine with *0.1 M sodium thiosulfate*. Carry out a blank test.

The difference between the volumes used in the 2 titrations corresponds to the amount of iodine liberated by ferric ion.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 5.585 mg of ferric ion.

Cadmium

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *cadmium standard solution (0.1 per cent Cd) R* and diluting with a 10 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Air-acetylene flame.

Chromium

Maximum 200 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using chromium standard solution (0.1 per cent Cr) R and diluting with a 10 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Chromium hollow-cathode lamp.

Wavelength 357.9 nm.

Atomisation device Air-acetylene flame.

Lead

Maximum 20 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *lead standard solution (10 ppm Pb) R* and diluting with a 10 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm.

Atomisation device Air-acetylene flame.

Mercury

Maximum 1 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *mercury standard solution (10 ppm Hg) R* and diluting with a 25 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Mercury hollow-cathode lamp.

Wavelength 253.7 nm.

Following the recommendations of the manufacturer, introduce 5 mL of solution S or 5 mL of the reference solutions into the reaction vessel of the cold-vapour mercury assay accessory, add 10 mL of *water R* and 1 mL of *stannous chloride solution R1*.

Nickel

Maximum 200 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *nickel standard solution (10 ppm Ni) R* and diluting with a 10 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Nickel hollow-cathode lamp.

Wavelength 232 nm.

Atomisation device Air-acetylene flame.

Zinc

Maximum 500 ppm.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Solution S diluted to 10 volumes.

Reference solutions Prepare the reference solutions using *zinc standard solution (10 ppm Zn) R* and diluting with a 1 per cent *V/V* solution of *lead-free hydrochloric acid R*.

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
Dissolve with slight heating 0.150 g in 7.5 mL of *dilute sulfuric acid R*. Cool and add 25 mL of *water R*. Add 0.1 mL of *ferroin R*. Titrate immediately with *0.1 M cerium sulfate* until the colour changes from orange to light bluish-green.

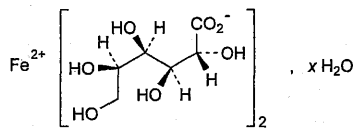
1 mL of *0.1 M cerium sulfate* is equivalent to 16.99 mg of $C_4H_2FeO_4$.

STORAGE
In an airtight container, protected from light.

Ph Eur

Ferrous Gluconate

(Ph. Eur. monograph 0493)



$C_{12}H_{22}FeO_{14},xH_2O$ 446.1
(anhydrous substance)

Action and use
Used in prevention and treatment of iron deficiency.

Preparation
Ferrous Gluconate Tablets

Ph Eur

DEFINITION
Iron(II) bis[(2*R*,3*S*,4*R*,5*R*)-2,3,4,5,6-pentahydroxyhexanoate] (iron(II) di(D-gluconate)).

Content
11.8 per cent to 12.5 per cent of iron(II) (dried substance).

It contains a variable quantity of water.

CHARACTERS
Appearance
Greenish-yellow or grey powder or granules.

Solubility
Freely but slowly soluble in water giving a greenish-brown solution, more readily soluble in hot water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 2 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Reference solution Dissolve 20 mg of *ferrous gluconate CRS* in 2 mL of *water R*, heating if necessary in a water-bath at 60 °C.

Plate *TLC silica gel plate R* (5-40 µm) [or *TLC silica gel plate R* (2-10 µm)].

Mobile phase *concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R* (10:10:30:50 *V/V/V/V*).

Application 1 µL.

Development Over 2/3 of the plate.

Drying At 105 °C for 20 min; allow to cool.

Detection Spray with a solution containing 10 g/L of *cerium sulfate R* and 25 g/L of *ammonium molybdate R* in *dilute sulfuric acid R* and heat at 105 °C for about 10 min.

Results After 5 min, the principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. 1 mL of solution S (see Tests) gives reaction (a) of iron (*2.3.1*).

TESTS
Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and heated to about 60 °C, allow to cool and dilute to 50 mL with *carbon dioxide-free water R* prepared from *distilled water R*.

Appearance of solution
The solution is clear (*2.2.1*).

Dilute 2 mL of solution S to 10 mL with *water R*. Examine the solution against the light.

pH (*2.2.3*)
4.0 to 5.5 for solution S, measured 3-4 h after preparation.

Sucrose and reducing sugars
Dissolve 0.5 g in 10 mL of warm *water R* and add 1 mL of *dilute ammonia R1*. Pass *hydrogen sulfide R* through the solution and allow to stand for 30 min. Filter and wash the precipitate with 2 quantities, each of 5 mL, of *water R*. Acidify the combined filtrate and washings to *blue litmus paper R* with *dilute hydrochloric acid R* and add 2 mL in excess. Boil until the vapour no longer darkens *lead acetate paper R* and continue boiling, if necessary, until the volume is reduced to about 10 mL. Cool, add 15 mL of *sodium carbonate solution R*, allow to stand for 5 min and filter. Dilute the filtrate to 100 mL with *water R*. To 5 mL of this solution add 2 mL of *cupri-tartaric solution R* and boil for 1 min. Allow to stand for 1 min. No red precipitate is formed.

Chlorides (*2.4.4*)
Maximum 0.06 per cent.

Dilute 0.8 mL of solution S to 15 mL with *water R*.

Oxalates
Dissolve 5.0 g in a mixture of 10 mL of *dilute sulfuric acid R* and 40 mL of *water R*. Shake the solution with 50 mL of *ether R* for 5 min. Separate the aqueous layer and shake it with 20 mL of *ether R* for 5 min. Combine the ether layers, evaporate to dryness and dissolve the residue in 15 mL of *water R*. Filter, boil the filtrate until the volume is reduced to 5 mL and add 1 mL of *dilute acetic acid R* and 1.5 mL of *calcium chloride solution R*. Allow to stand for 30 min. No precipitate is formed.

Sulfates (*2.4.13*)
Maximum 500 ppm.

To 3.0 mL of solution S add 3 mL of *acetic acid R* and dilute to 15 mL with *distilled water R*. Examine the solutions against the light.

Arsenic (*2.4.2, Method A*)
Maximum 2 ppm, determined on 0.5 g.

Barium
Dilute 10 mL of solution S to 50 mL with *distilled water R* and add 5 mL of *dilute sulfuric acid R*. Allow to stand for 5 min. Any opalescence in the solution is not more intense than that in a mixture of 10 mL of solution S and 45 mL of *distilled water R*.

Ferric ions
Maximum 1.0 per cent.

In a ground-glass-stoppered flask, dissolve 5.00 g in a mixture of 10 mL of *hydrochloric acid R* and 100 mL of *carbon dioxide-free water R*. Add 3 g of *potassium iodide R*, close the flask and allow to stand protected from light for 5 min. Titrate with *0.1 M sodium thiosulfate*, using 0.5 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank titration. Not more than 9.0 mL of *0.1 M sodium thiosulfate* is used.

Loss on drying (*2.2.32*)
5.0 per cent to 10.5 per cent, determined on 0.500 g by drying in an oven at 105 °C for 5 h.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

ASSAY
Dissolve 0.5 g of *sodium hydrogen carbonate R* in a mixture of 30 mL of *dilute sulfuric acid R* and 70 mL of *water R*. When the effervescence stops, dissolve 1.00 g of the substance to be examined with gentle shaking. Using 0.1 mL of *ferroin R* as indicator, titrate with *0.1 M ammonium and cerium nitrate* until the red colour disappears.

1 mL of *0.1 M ammonium and cerium nitrate* is equivalent to 5.585 mg of iron(II).

STORAGE
Protected from light.

Ph Eur

Dried Ferrous Sulfate

Dried Ferrous Sulphate

(Ph. Eur. monograph 2340)

$FeSO_4$ 151.9

Preparation
Ferrous Sulfate Prolonged-release Tablets

Ph Eur

DEFINITION
Hydrated ferrous sulfate from which part of the water of hydration has been removed by drying.

Content
86.0 per cent to 90.0 per cent.

CHARACTERS
Appearance
Greyish-white powder.

Solubility
Slowly but freely soluble in water, very soluble in boiling water, practically insoluble in ethanol (96 per cent).

It is oxidised in moist air, becoming brown.

IDENTIFICATION
A. It gives the reactions of sulfates (*2.3.1*).

B. It gives reaction (a) of iron (*2.3.1*).

C. It complies with the limits of the assay.

TESTS
Solution S
Dissolve 2.00 g in a 5 per cent *V/V* solution of *lead-free nitric acid R* and dilute to 100.0 mL with the same acid.

pH (*2.2.3*)
3.0 to 4.0.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Chlorides (*2.4.4*)
Maximum 300 ppm.

Dissolve 2.5 g in *water R*, add 0.5 mL of *dilute sulfuric acid R* and dilute to 50 mL with *water R*. Dilute 3.3 mL of this solution to 10 mL with *water R* and add 5 mL of *dilute nitric acid R*. Prepare the standard using a mixture of 10 mL of *chloride standard solution (5 ppm Cl) R* and 5 mL of *dilute nitric acid R*. Use 0.15 mL of *silver nitrate solution R2* in this test.

Chromium
Maximum 100 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *chromium standard solution (100 ppm Cr) R*, diluted as necessary with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Chromium hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 357.9 nm.

Atomisation device Air-acetylene flame.

Copper
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *copper standard solution (0.1 per cent Cu) R*, diluted as necessary with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Copper hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Ferric ions
Maximum 0.5 per cent.

In a ground-glass-stoppered flask, dissolve 5.00 g in a mixture of 10 mL of *hydrochloric acid R* and 100 mL of *carbon dioxide-free water R*. Add 3 g of *potassium iodide R*, close the flask and allow to stand in the dark for 5 min. Titrate the liberated iodine with *0.1 M sodium thiosulfate*, using 0.5 mL of *starch solution R*, added towards the end of titration, as indicator. Carry out a blank test in the same conditions. Not more than 4.5 mL of *0.1 M sodium thiosulfate* is used.

Manganese
Maximum 0.1 per cent.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Dilute 1.0 mL of solution S to 20.0 mL with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Reference solutions Prepare the reference solutions using *manganese standard solution (1000 ppm Mn) R*, diluted as necessary with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Manganese hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 279.5 nm.

Atomisation device Air-acetylene flame.

Nickel
Maximum 100 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *nickel standard solution (10 ppm Ni) R*, diluted as necessary with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Nickel hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 232.0 nm.

Atomisation device Air-acetylene flame.

Zinc
Maximum 100 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *zinc standard solution (100 ppm Zn) R*, diluted as necessary with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Zinc hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame.

ASSAY
Dissolve 2.5 g of *sodium hydrogen carbonate R* in a mixture of 150 mL of *water R* and 10 mL of *sulfuric acid R*. When the effervescence ceases, add to the solution 0.140 g of the substance to be examined and dissolve with gentle shaking. Add 0.1 mL of *ferroin R* and titrate with *0.1 M ammonium and cerium nitrate* until the red colour disappears.

1 mL of *0.1 M ammonium and cerium nitrate* is equivalent to 15.19 mg of $FeSO_4$.

STORAGE
In an airtight container.

Ph Eur

Ferrous Sulfate Heptahydrate

Ferrous Sulphate Heptahydrate

(Ph. Eur. monograph 0083)

$FeSO_4,7H_2O$ 278.0 *7782-63-0*

Action and use
Used in prevention and treatment of anaemias.

Preparation
Paediatric Ferrous Sulfate Oral Solution

Ph Eur

DEFINITION
Content
98.0 per cent to 105.0 per cent.

CHARACTERS
Appearance
Light green, crystalline powder or bluish-green crystals, efflorescent in air.

Solubility
Freely soluble in water, very soluble in boiling water, practically insoluble in ethanol (96 per cent).

Ferrous sulfate heptahydrate is oxidised in moist air, becoming brown.

IDENTIFICATION
A. It gives the reactions of sulfates (*2.3.1*).

B. It gives reaction (a) of iron (*2.3.1*).

C. It complies with the limits of the assay.

TESTS
Solution S
Dissolve 4.0 g in a 5 per cent *V/V* solution of *lead-free nitric acid R* and dilute to 100.0 mL with the same solution.

pH (*2.2.3*)
3.0 to 4.0.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 10 mL with *water R* and add 5 mL of *dilute nitric acid R*. Prepare the standard with a mixture of 2 mL of *water R*, 5 mL of *dilute nitric acid R* and 8 mL of *chloride standard solution (5 ppm Cl) R*. Use 0.15 mL of *silver nitrate solution R2* in this test.

Chromium
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *chromium standard solution (100 ppm Cr) R*, diluting with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Chromium hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 357.9 nm.

Atomisation device Air-acetylene flame.

Copper
Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *copper standard solution (0.1 per cent Cu) R*, diluting with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Copper hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 324.7 nm.

Atomisation device Air-acetylene flame.

Ferric ions
Maximum 0.3 per cent.

In a ground-glass-stoppered flask, dissolve 5.00 g in a mixture of 10 mL of *hydrochloric acid R* and 100 mL of *carbon dioxide-free water R*. Add 3 g of *potassium iodide R*, close the flask and allow to stand in the dark for 5 min. Titrate the liberated iodine with *0.1 M sodium thiosulfate*, using 0.5 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank test in the same conditions. Not more than 2.7 mL of *0.1 M sodium thiosulfate* is used, taking into account the blank titration.

Manganese
Maximum 0.1 per cent.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Dilute 1.0 mL of solution S to 20.0 mL with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Reference solutions Prepare the reference solutions using *manganese standard solution (1000 ppm Mn) R*, diluting with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Manganese hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 279.5 nm.

Atomisation device Air-acetylene flame.

Nickel

Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *nickel standard solution (10 ppm Ni) R*, diluting with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Nickel hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 232.0 nm.

Atomisation device Air-acetylene flame.

Zinc

Maximum 50 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Solution S.

Reference solutions Prepare the reference solutions using *zinc standard solution (100 ppm Zn) R*, diluting with a 5 per cent *V/V* solution of *lead-free nitric acid R*.

Source Zinc hollow-cathode lamp using a transmission band preferably of 1 nm.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame.

ASSAY

Dissolve 2.5 g of *sodium hydrogen carbonate R* in a mixture of 150 mL of *water R* and 10 mL of *sulfuric acid R*. When the effervescence ceases add to the solution 0.500 g of the substance to be examined and dissolve with gentle swirling. Add 0.1 mL of *ferroin R* and titrate with *0.1 M ammonium and cerium nitrate* until the red colour disappears.

1 mL of *0.1 M ammonium and cerium nitrate* is equivalent to 27.80 mg of $FeSO_4,7H_2O$.

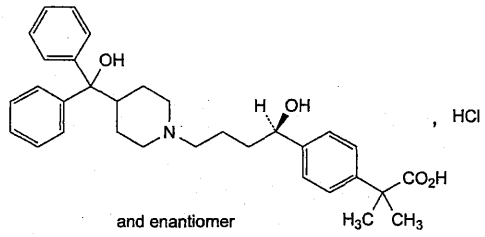
STORAGE

In an airtight container.

Ph Eur

Fexofenadine Hydrochloride

(Ph. Eur. monograph 2280)



$C_{32}H_{40}ClNO_4$ 538.1 *153439-40-8*

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Fexofenadine Tablets

Ph Eur

DEFINITION

2-[4-[(1*RS*)-1-Hydroxy-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butyl]phenyl]-2-methylpropanoic acid hydrochloride.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in methanol, very slightly soluble in acetone.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fexofenadine hydrochloride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Dissolve 30 mg of the substance to be examined in a mixture of equal volumes of *methanol R* and *water R*; sonicate if necessary and dilute to 2 mL with the same mixture of solvents. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Impurity B

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve the contents of a vial of *fexofenadine impurity B CRS* in the test solution and dilute to 2.0 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *silica gel BC for chiral chromatography R* (5 µm).

Mobile phase Mix 20 volumes of *acetonitrile for chromatography R* and 80 volumes of a buffer solution prepared as follows: to 1.15 mL of *glacial acetic acid R* add *water for chromatography R*, adjust to pH 4.0 ± 0.1 with *dilute ammonia R1* and dilute to 1000 mL with *water for chromatography R*.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time 1.2 times the retention time of fexofenadine.

Relative retention With reference to fexofenadine (retention time = about 20 min): impurity B = about 0.7.

System suitability Reference solution (a):

— *resolution*: minimum 3.0 between the peaks due to fexofenadine and impurity B.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 1.3;

— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 6.64 g of *sodium dihydrogen phosphate monohydrate R* and 0.84 g of *sodium perchlorate R* in *water for chromatography R*, adjust to pH 2.0 ± 0.1 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Solvent mixture Mix equal volumes of *acetonitrile for chromatography R* and the buffer solution.

Test solution (a) Dissolve 25.0 mg of the substance to be examined in 25.0 mL of the solvent mixture.

Test solution (b) Dilute 3.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *fexofenadine hydrochloride CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 3.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 1 mg each of *fexofenadine impurity A CRS* and *fexofenadine impurity C CRS* in 20 mL of reference solution (a) and dilute to 200.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 350 volumes of *acetonitrile for chromatography R* and 650 volumes of the buffer solution; add 3 volumes of *triethylamine R* and mix.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (b) and (c).

Relative retention With reference to fexofenadine (retention time = about 9 min): impurity A = about 1.7; impurity D = about 2.3; impurity C = about 3.2.

Run time 6 times the retention time of fexofenadine for test solution (a) and reference solution (c), twice the retention time of fexofenadine for reference solution (b).

System suitability Reference solution (c):
— *resolution*: minimum 10 between the peaks due to fexofenadine and impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.4;
— *impurities A, C, D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.32*)

Maximum 0.5 per cent.

Dissolve 1.000 g in *anhydrous methanol R* and dilute to 5.0 mL with the same solvent. Use 1.0 mL of this solution.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

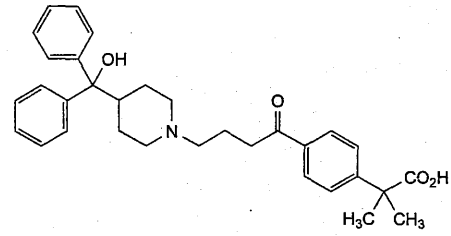
Run time Twice the retention time of fexofenadine.

Calculate the percentage content of fexofenadine hydrochloride from the declared content of *fexofenadine hydrochloride CRS*.

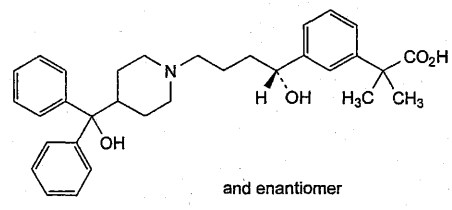
IMPURITIES

Specified impurities *A, B, C, D.*

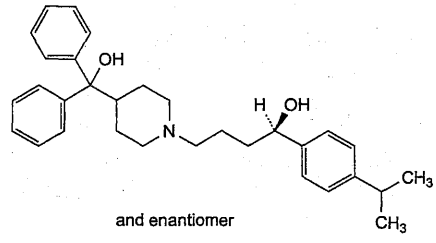
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F, G.*



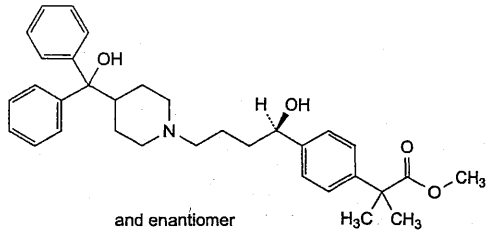
A. 2-[4-[4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butanoyl]phenyl]-2-methylpropanoic acid,



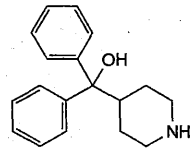
B. 2-[3-[(1*RS*)-1-hydroxy-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butyl]phenyl]-2-methylpropanoic acid,



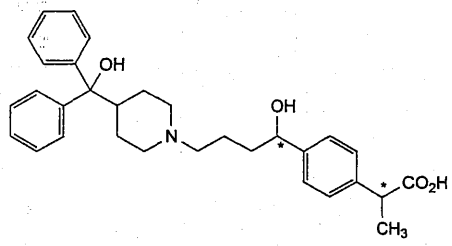
C. (1*RS*)-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]-1-[4-(1-methylethyl)phenyl]butan-1-ol,



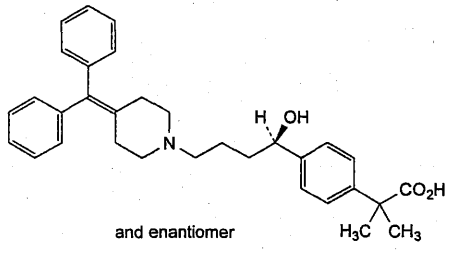
D. methyl 2-[4-[(1*RS*)-1-hydroxy-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butyl]phenyl]-2-methylpropanoate,



E. diphenyl(piperidin-4-yl)methanol,



F. 2-[4-[1-hydroxy-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butyl]phenyl]propanoic acid,



G. 2-[4-[(1*RS*)-4-[4-(diphenylmethylidene)piperidin-1-yl]-1-hydroxybutyl]phenyl]-2-methylpropanoic acid.

Ph Eur

Filgrastim Concentrated Solution

(Ph. Eur. monograph 2206)

```
MTPLGPASSL  PQSFLLKCLE  QVRKIQGDGA  ALQEKLCATY
KLCHPEELVL  LGHSLGIPWA  PLSSCPSQAL  QLAGCLSQLH
SGLFLYQGLL  QALEGISPEL  GPTLDTLQLD  VADFATTIWQ
QMEELGMAPA  LQPTQGAMPA  FASAFQRRAG  GVLVASHLQS
FLEVSYRVLR  HLAQP
```

$C_{845}H_{1339}N_{223}O_{243}S_9$ 18 799 *121181-53-1*

Action and use
Recombinant methionyl human granulocyte colony-stimulating factor.

Preparation
Filgrastim Injection

Ph Eur

DEFINITION
Solution of a protein having the primary structure of the 174-amino-acid isoform of human granulocyte colony-stimulating factor (HUG-CSF) plus 1 additional amino acid, an *N*-terminal methionine. In contrast to its natural counterpart, the protein is not glycosylated. Human G-CSF is produced and secreted by endothelial cells, monocytes and other immune cells. The protein stimulates the differentiation and proliferation of leucocyte stem cells into mature granulocytes.

Content
Minimum 0.9 mg of protein per millilitre.

Potency
Minimum 0.9×10^8 IU per milligram of protein.

PRODUCTION
Filgrastim concentrated solution is produced by a method based on recombinant DNA (rDNA) technology, using bacteria as host cells.

Prior to release, the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins
The limit is approved by the competent authority.

Host-cell- or vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS
Appearance
Clear, colourless or slightly yellowish liquid.

IDENTIFICATION
A. It shows the expected biological activity (see Assay).

B. Examine the electropherograms obtained in the test for impurities with charges differing from that of filgrastim.

Results The principal band in the electropherogram obtained with the test solution is similar in position to the principal band in the electropherogram obtained with reference solution (a).

C. Examine the chromatograms obtained in the test for impurities with molecular masses higher than that of filgrastim.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

D. Examine the electropherograms obtained under both reducing and non-reducing conditions in the test for impurities with molecular masses differing from that of filgrastim.

Results The principal band in the electropherogram obtained with test solution (a) is similar in position to the principal band in the electropherogram obtained with reference solution (b).

E. Examine the chromatograms obtained in the test for related proteins.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and shape to the principal peak in the chromatogram obtained with the reference solution.

F. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Introduce a volume of the preparation to be examined corresponding to 25 µg of protein into a polypropylene tube. Add 25 µL of a 0.1 mg/mL solution of *glutamyl endopeptidase for peptide mapping R*. Dilute to 100 µL with *0.02 M sodium phosphate buffer solution pH 8.0 R*, stopper the tube and incubate at about 37 °C for 17 h. Cool to 2-8 °C until analysis.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *filgrastim CRS* instead of the preparation to be examined.

CHROMATOGRAPHIC SEPARATION. Liquid chromatography (*2.2.29*).

Column:
- *size*: *l* = 0.10 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (5 µm) with a pore size of 20 nm;
- *temperature*: 60 °C.

Mobile phase:
- *mobile phase A*: dilute 0.5 mL of *trifluoroacetic acid R* in 950 mL of *water for chromatography R*, add 50 mL of *acetonitrile for chromatography R* and mix;
- *mobile phase B*: dilute 0.5 mL of *trifluoroacetic acid R* in 50 mL of *water for chromatography R*, add 950 mL of *acetonitrile for chromatography R* and mix;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	97 → 94	3 → 6
8 - 25	94 → 66	6 → 34
25 - 40	66 → 10	34 → 90
40 - 45	10	90

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

System suitability The chromatogram obtained with the reference solution is similar to the chromatogram of filgrastim digest supplied with *filgrastim CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

TESTS

Impurities with molecular masses higher than that of filgrastim

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Solution A Dissolve 4.1 g of *sodium acetate R* in 400 mL of *water R*, adjust to pH 4.0 with *acetic acid R* and dilute to 500 mL with *water R*.

Test solution Dilute the preparation to be examined with solution A to obtain a concentration of 0.4 mg/mL.

Reference solution Dilute *filgrastim CRS* with solution A to obtain a concentration of 0.4 mg/mL.

Resolution solution Mix a sample of the reference solution for about 30 s using a vortex mixer.

Column:
- *size*: *l* = 0.3 m, Ø = 7.8 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5 µm) of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000;
- *temperature*: 30 °C.

Mobile phase Dissolve 7.9 g of *ammonium hydrogen carbonate R* in 1000 mL of *water for chromatography R* and adjust to pH 7.0 with *phosphoric acid R*; dilute to 2000 mL with *water for chromatography R*.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Relative retention With reference to the filgrastim monomer (retention time = about 19 min): aggregates = about 0.60; filgrastim oligomer 1 = about 0.75; filgrastim oligomer 2 = about 0.80; filgrastim dimer = about 0.85.

System suitability Resolution solution:
- *retention time*: filgrastim monomer = 17 min to 20 min;
- *resolution*: minimum 3 between the peaks due to the filgrastim dimer and the filgrastim monomer.

Calculate the percentage content of the dimer, oligomers and aggregates.

Limits:
- *impurities with molecular masses higher than that of filgrastim, other than the dimer*: maximum 0.5 per cent;
- *total of impurities with molecular masses higher than that of filgrastim*: maximum 2 per cent.

Impurities with molecular masses differing from that of filgrastim

Polyacrylamide gel electrophoresis (*2.2.31*) under both reducing and non-reducing conditions.

Gel dimensions 1 mm thick.

Resolving gel 13 per cent acrylamide.

Sample buffer (non-reducing conditions) Mix equal volumes of *water R* and *concentrated SDS-PAGE sample buffer R*.

Sample buffer (reducing conditions) Mix equal volumes of *water R* and *concentrated SDS-PAGE sample buffer for reducing conditions R* containing 2-mercaptoethanol as the reducing agent.

Test solution (a) Dilute the preparation to be examined with sample buffer to obtain a concentration of 100 µg/mL.

Test solution (b) To 0.20 mL of test solution (a) add 0.20 mL of sample buffer.

Test solution (c) Dilute 0.20 mL of test solution (b) to 1 mL with sample buffer.

Test solution (d) Dilute 0.20 mL of test solution (c) to 1 mL with sample buffer.

Test solution (e) To 0.20 mL of test solution (d) add 0.20 mL of sample buffer.

Reference solution (a) Solution of molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 14.4-94 kDa.

Reference solution (b) Dilute *filgrastim CRS* with sample buffer to obtain a concentration of 100 µg/mL.

Sample treatment Boil for 5 min.

Application 20 µL.

Detection By silver staining.

System suitability:
- reference solution (a): the validation criteria are met;
- a band is seen in the electropherogram obtained with test solution (e);
- a gradation of intensity of staining is seen in the electropherograms obtained with test solutions (a) to (e).

Limit Test solution (a):

— *impurities with molecular masses lower or higher than that of filgrastim*: no band is more intense than the principal band in the electropherogram obtained with test solution (d) (2.0 per cent).

Impurities with charges differing from that of filgrastim

Isoelectric focusing (*2.2.54*).

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 0.3 mg/mL.

Reference solution (a) Dilute *filgrastim CRS* with *water R* to obtain a concentration of 0.3 mg/mL.

Reference solution (b) Dilute *filgrastim CRS* with *water R* to obtain a concentration of 0.03 mg/mL.

Reference solution (c) Use an isoelectric point (pI) calibration solution, in the pI range of 2.5-6.5, prepared according to the manufacturer's instructions.

Focusing:
— *pH gradient*: 4.5-8.0;
— *catholyte*: 1 M solution of *sodium hydroxide R*;
— *anolyte*: 0.04 M solution of *glutamic acid R* in a 0.0025 per cent *V/V* solution of *phosphoric acid R*;
— *application*: 20 µL.

Detection As described in *2.2.54*.

System suitability:
— in the electropherogram obtained with reference solution (c), the relevant isoelectric point markers are distributed along the entire length of the gel;
— in the electropherogram obtained with reference solution (a), the pI of the principal band is 5.7 to 6.3.

Limit:
— *any impurity*: no band is more intense than the principal band in the electropherogram obtained with reference solution (b) (10 per cent).

Related proteins

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dilute the preparation to be examined with *water for chromatography R* to obtain a concentration of 0.5 mg/mL.

Reference solution (a) Dilute the content of a vial of *filgrastim CRS* with *water for chromatography R* to obtain a concentration of 0.5 mg/mL.

Reference solution (b) To 250 µL of reference solution (a), add 2.5 µL of a 4.5 g/L solution of hydrogen peroxide. Mix and incubate at 25 ± 2 °C for 30 min, then add 1.9 mg of *L-methionine R*.

Reference solution (c) To 250 µL of reference solution (a), add 0.25 mg of *dithiothreitol R*. Mix and incubate at 35 ± 2 °C for 60 min.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm;
— *temperature*: 60 °C.

Mobile phase:
— *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* in 1000 mL of *water for chromatography R*;
— *mobile phase B*: dilute 1 mL of *trifluoroacetic acid R* in 100 mL of *water for chromatography R*, then add 900 mL of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	60 → 20	40 → 80
30 - 35	20	80
35 - 45	20 → 60	80 → 40

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 50 µL of the test solution and reference solutions (b) and (c).

Relative retention With reference to filgrastim (retention time = about 23 min): oxidised filgrastim (form 1) = about 0.84; oxidised filgrastim (form 2) = about 0.98; reduced filgrastim = about 1.04.

System suitability Reference solution (b):
— *symmetry factor*: maximum 1.8 for the peak due to filgrastim;
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to oxidised filgrastim (form 2) and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to filgrastim.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to filgrastim and reduced filgrastim;
— *symmetry factor*: maximum 1.8 for the peak due to filgrastim.

Limits:
— *any impurity*: for each impurity, maximum 1.0 per cent;
— *total*: maximum 2.0 per cent.

Bacterial endotoxins (*2.6.14*)

Less than 2 IU in the volume that contains 1.0 mg of protein.

ASSAY

Protein

Liquid chromatography (*2.2.29*) as described in the test for related proteins with the following modification.

Injection Test solution and reference solution (a).

Calculate the content of filgrastim ($C_{845}H_{1339}N_{223}O_{243}S_9$) taking into account the assigned content of $C_{845}H_{1339}N_{223}O_{243}S_9$ in *filgrastim CRS*.

Potency

The potency of the preparation to be examined is determined by comparison of the dilutions of the test preparation with the dilutions of the International Standard of filgrastim or with a reference preparation calibrated in International Units.

The International Unit is the activity contained in a stated amount of the appropriate International Standard. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Carry out the assay using a suitable method such as the following, which uses the conversion of a tetrazolium salt (MTS) as a staining method. Alternative methods of quantifying cell proliferation, such as measurement of intracellular ATP by luciferase bioluminescence, have also been found suitable, and may be used as the assay readout, subject to appropriate validation. The assay conditions (for example, cell concentration, incubation time and dilution steps) are then adapted accordingly.

Use an established cell line responsive to filgrastim. M-NFS-60 cells (ATCC No. CRL-1838) that have been made sensitive to G-CSF have been found suitable. Incubate with varying dilutions of test and reference preparations of

filgrastim. Then incubate with a solution of *tetrazolium salt R*. This cytochemical stain is converted by cellular dehydrogenases to a coloured formazan product. The formazan is then measured spectrophotometrically.

Add 50 µL of dilution medium to all wells of a 96-well microtitre plate. Add an additional 50 µL of this solution to the wells designed for the blanks. Add 50 µL of each solution to be tested in triplicate (test preparation and reference preparation at a concentration of about 800 IU/mL, plus a series of 10 twofold dilutions to obtain a standard curve). Prepare a suspension of M-NFS-60 cells containing 7×10^5 cells per millilitre. Immediately before use, add 2-mercaptoethanol to a final concentration of 0.1 mM, and add 50 µL of the prepared cell suspension to each well, maintaining the cells in a uniform suspension during addition.

Incubate the plate at 36.0-38.0 °C for 44-48 h in a humidified incubator using 6 ± 1 per cent CO_2. Add 20 µL of a 5.0 g/L sterile solution of *tetrazolium salt R* to each well and reincubate for 4 h. Estimate the quantity of formazan produced using a microtitre well plate reader at 490 nm.

Calculate the potency of the preparation to be examined using a suitable statistical method, for example the parallel line assay (*5.3*).

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency. The confidence limits (P = 0.95) are not less than 74 per cent and not more than 136 per cent of the estimated potency.

LABELLING

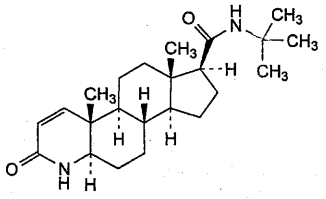
The label states:

— the content, in milligrams of protein per millilitre;
— the potency, in International Units per milligram of protein.

Ph Eur

Finasteride

(Ph. Eur. monograph 1615)



$C_{23}H_{36}N_2O_2$ 372.6 *98319-26-7*

Action and use

5-Alpha reductase inhibitor; treatment of benign prostatic hyperplasia.

Preparation

Finasteride Tablets

Ph Eur

DEFINITION

N-(1,1-Dimethylethyl)-3-oxo-4-aza-5α-androst-1-ene-17β-carboxamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethanol and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *finasteride CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Specific optical rotation (*2.2.7*)

+ 12.0 to + 14.0 (dried substance).

Dissolve 0.250 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1*, *water for chromatography R* (50:50 *V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dissolve 100.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of *finasteride CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *finasteride for peak identification CRS* (containing impurities A and C) in 1.0 mL of the solvent mixture.

Reference solution (c) Dilute 1.0 mL of test solution (b) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 60 °C.

Mobile phase *acetonitrile R1*, *tetrahydrofuran R*, *water for chromatography R* (10:10:80 *V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 15 µL of test solution (b) and reference solutions (b) and (c).

Run time Twice the retention time of finasteride.

Identification of impurities Use the chromatogram supplied with *finasteride for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C.

Relative retention With reference to finasteride (retention time = about 28 min): impurity A = about 0.9; impurity C = about 1.3.

System suitability:

— *signal-to-noise ratio*: minimum 40 for the principal peak in the chromatogram obtained with reference solution (c);

— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to finasteride in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity A by 2.4;
— for each impurity, use the concentration of finasteride in reference solution (c).

Limits:
— *impurities A, C*: for each impurity, maximum 0.3 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

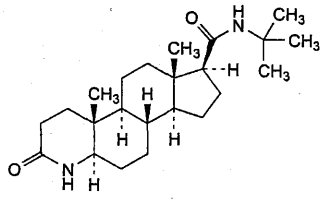
Injection Test solution (a) and reference solution (a).

Calculate the percentage content of $C_{23}H_{36}N_2O_2$ taking into account the assigned content of *finasteride CRS*.

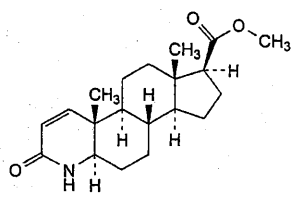
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, C.*

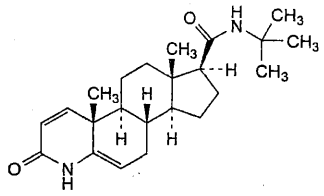
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. *N*-(1,1-dimethylethyl)-3-oxo-4-aza-5α-androstane-17β-carboxamide (dihydrofinasteride),



B. methyl 3-oxo-4-aza-5α-androst-1-ene-17β-carboxylate,

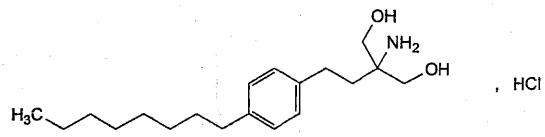


C. *N*-(1,1-dimethylethyl)-3-oxo-4-azaandrosta-1,5-diene-17β-carboxamide (Δ5-finasteride).

Ph Eur

Fingolimod Hydrochloride

(Ph. Eur. monograph 2988)



$C_{19}H_{34}ClNO_2$ 343.9 *162359-56-0*

Action and use
Sphingosine-1-phosphate receptor modulator; immunomodulator.

Ph Eur

DEFINITION
2-Amino-2-[2-(4-octylphenyl)ethyl]propane-1,3-diol hydrochloride.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Freely soluble in water and in ethanol (96 per cent), practically insoluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Mulls in *liquid paraffin R* if recording by transmission.

Comparison *fingolimod hydrochloride CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

B. It gives reaction (a) of chlorides (*2.3.1*).

If precipitation is observed after addition of *dilute nitric acid R*, centrifuge and use the supernatant in the remainder of the test.

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1*, 0.1 per cent *V/V* solution of *phosphoric acid R* (50:50 *V/V*).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 30.0 mg of *fingolimod hydrochloride CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 3 mg of *fingolimod for system suitability CRS* (containing impurities C and G) in the solvent mixture and dilute to 5 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 3.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R* (3 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: 0.1 per cent *V/V* solution of *phosphoric acid R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	80	20
2 - 22	80 → 5	20 → 95
22 - 25	5	95

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 5 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *fingolimod for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and G.

Relative retention With reference to fingolimod (retention time = about 6.5 min): impurity G = about 1.10; impurity C = about 1.13.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurities G and C.

Calculation of percentage contents:
— for each impurity, use the concentration of fingolimod hydrochloride in reference solution (b).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)
Maximum 0.3 per cent, determined on 50.0 mg by direct sample introduction.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

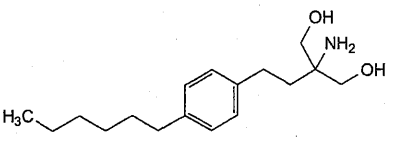
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

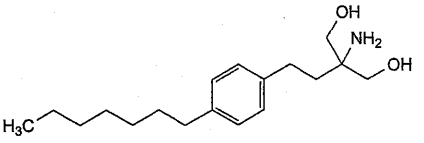
Calculate the percentage content of $C_{19}H_{34}ClNO_2$ taking into account the assigned content of *fingolimod hydrochloride CRS*.

IMPURITIES

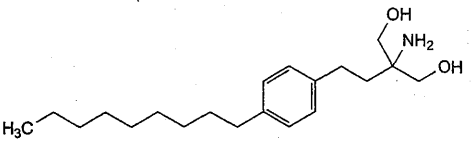
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I.*



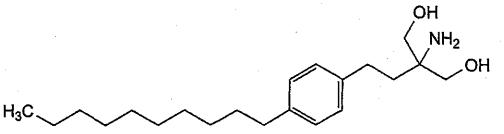
A. 2-amino-2-[2-(4-hexylphenyl)ethyl]propane-1,3-diol,



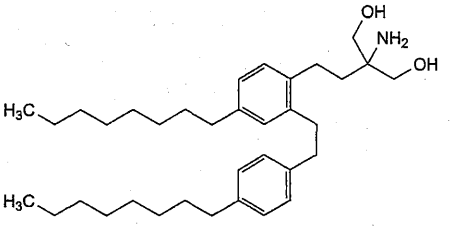
B. 2-amino-2-[2-(4-heptylphenyl)ethyl]propane-1,3-diol,



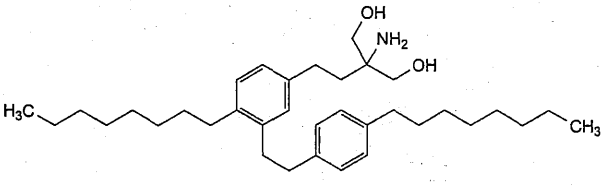
C. 2-amino-2-[2-(4-nonylphenyl)ethyl]propane-1,3-diol,



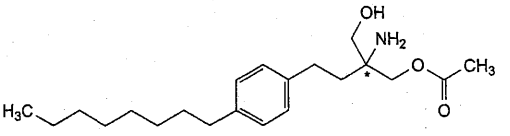
D. 2-amino-2-[2-(4-decylphenyl)ethyl]propane-1,3-diol,



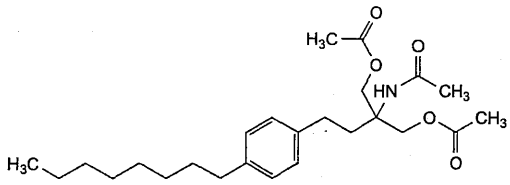
E. 2-amino-2-[2-[4-octyl-2-[2-(4-octylphenyl)ethyl]phenyl] ethyl]propane-1,3-diol,



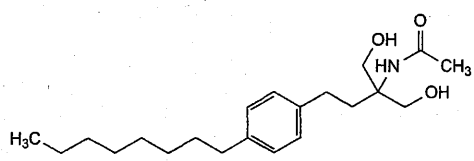
F. 2-amino-2-[2-[4-octyl-3-[2-(4-octylphenyl)ethyl]phenyl] ethyl]propane-1,3-diol,



G. (2*E*)-2-amino-2-(hydroxymethyl)-4-(4-octylphenyl)butyl acetate,



H. 2-acetamido-2-[(acetyloxy)methyl]-4-(4-octylphenyl)butyl acetate,



I. *N*-[1-hydroxy-2-(hydroxymethyl)-4-(4-octylphenyl)butan-2-yl]acetamide.

Ph Eur

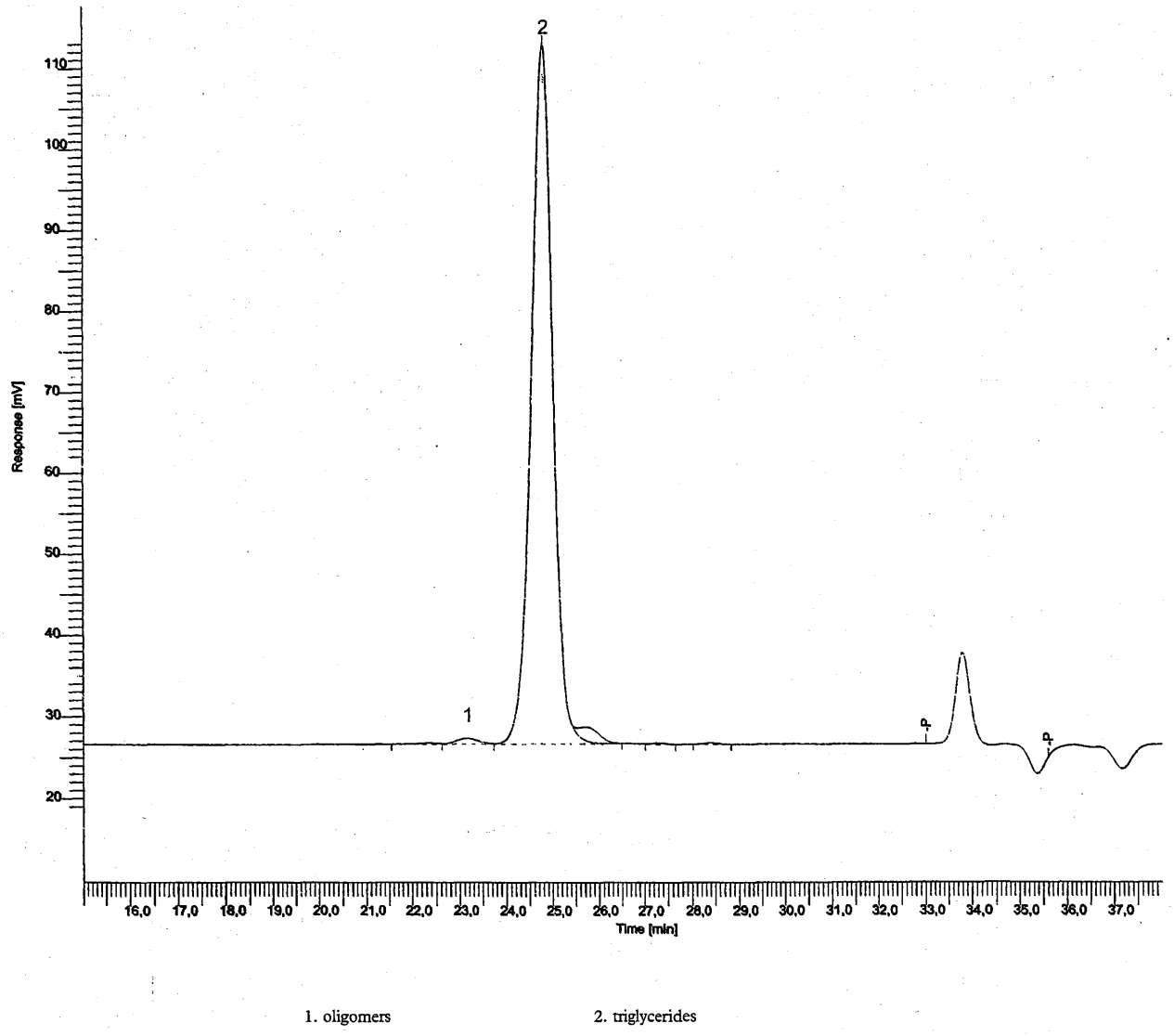
Fish Oil, Rich in Omega-3-Acids

(Ph. Eur. monograph 1912)

Ph Eur

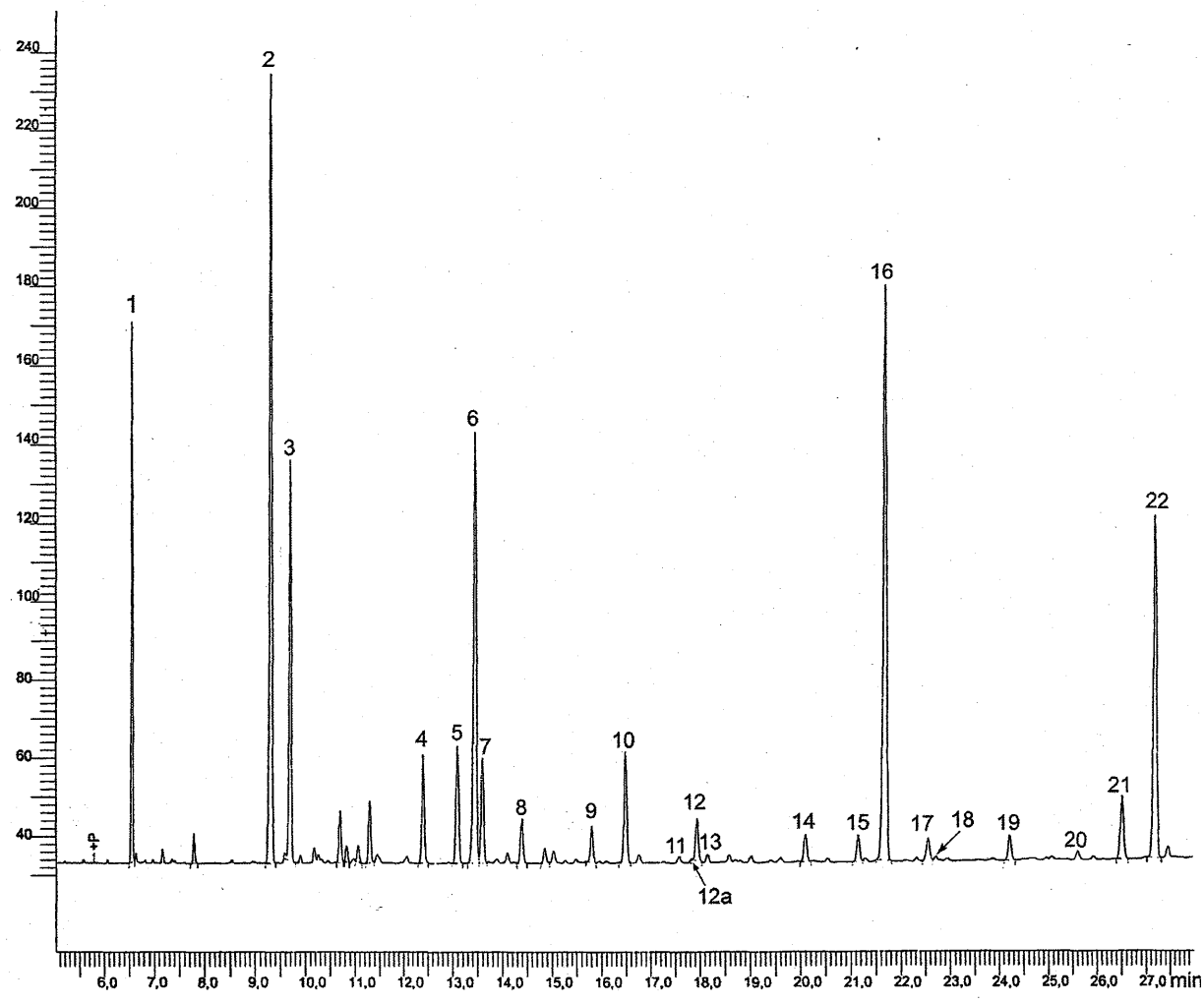
DEFINITION

Purified, winterised and deodorised fatty oil obtained from fish of families such as *Engraulidae*, *Carangidae*, *Clupeidae*, *Osmeridae*, *Scombridae* (except the genera *Thunnus* and *Sarda*) and *Ammodytidae* (type I), or from the genera *Thunnus* and *Sarda* within the family *Scombridae* (type II). The omega-3 acids are defined as the following acids: *alpha*-linolenic acid (C18:3 n-3), moroctic acid (C18:4 n-3), eicosatetraenoic acid (C20:4 n-3), timnodonic (eicosapentaenoic) acid (C20:5 n-3; EPA), heneicosapentaenoic acid (C21:5 n-3), clupanodonic acid (C22:5 n-3) and cervonic (docosahexaenoic) acid (C22:6 n-3; DHA).



1. oligomers 2. triglycerides

Figure 1912.-1. – *Chromatogram for the test for oligomers in fish oil rich in omega-3 acids*



1. C14:0	4. C16:4 n-1	7. C18:1 n-7	10. C18:4 n-3	12a. C20:1 n-11	15. C20:4 n-3	18. C22:1 n-9	21. C22:5 n-3
2. C16:0	5. C18:0	8. C18:2 n-6	11. C20:0	13. C20:1 n-7	16. C20:5 n-3	19. C21:5 n-3	22. C22:6 n-3
3. C16:1 n-7	6. C18:1 n-9	9. C18:3 n-3	12. C20:1 n-9	14. C20:4 n-6	17. C22:1 n-11	20. C22:5 n-6	

Figure 1912.-2. – *Chromatogram for the assay of total omega-3 acids in fish oil rich in omega-3 acids*

Content

	Type I	Type II
EPA, expressed as triglycerides	minimum 13 per cent	4 per cent to 12 per cent
DHA, expressed as triglycerides	minimum 9 per cent	minimum 20 per cent
Total omega-3 acids, expressed as triglycerides	minimum 28 per cent	minimum 28 per cent

A suitable antioxidant may be added.

PRODUCTION

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance

Pale yellow liquid.

Solubility

Practically insoluble in water, very soluble in acetone and in heptane, slightly soluble in anhydrous ethanol.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay for EPA and DHA.

Results The peaks due to eicosapentaenoic acid methyl ester and docosahexaenoic acid methyl ester in the chromatogram obtained with test solution (b) are similar in retention time to the corresponding peaks in the chromatogram obtained with reference solutions (a_1) and (a_2).

B. It complies with the limits of the assay for EPA (type I or II).

TESTS

Appearance

The substance to be examined is not more intensely coloured than a reference solution prepared as follows: to 3.0 mL of red primary solution add 25.0 mL of yellow primary solution and dilute to 50.0 mL with a 10 g/L solution of *hydrochloric acid R (2.2.2, Method II)*.

Absorbance (*2.2.25*)
Maximum 0.70 (type I) or maximum 0.50 (type II), at 233 nm.

Dilute 0.300 g of the substance to be examined to 50.0 mL with *trimethylpentane R*. Dilute 2.0 mL of the solution to 50.0 mL with *trimethylpentane R*.

Acid value (*2.5.1*)
Maximum 0.5, determined on 20.0 g.

Anisidine value (*2.5.36*)
Maximum 30.0 (type I) or maximum 15.0 (type II).

Peroxide value (*2.5.5, Method A*)
Maximum 10.0 (type I) or maximum 5.0 (type II).

Unsaponifiable matter (*2.5.7*)
Maximum 1.5 per cent, determined on 5.0 g.

Stearin
10 mL remains clear after cooling at 0 °C for 3 h.

Oligomers
Size-exclusion chromatography (*2.2.30*).

Test solution Dilute 50.0 mg of the substance to be examined to 10.0 mL with *tetrahydrofuran R*.

Reference solution In a 100 mL volumetric flask dissolve 50 mg of *monodocosahexaenoin R*, 30 mg of *didocosahexaenoin R* and 20 mg of *tridocosahexaenoin R* in *tetrahydrofuran R* and dilute to 100.0 mL with the same solvent.

Column 3 columns to be connected in series:
— *size*: l = 0.3 m, Ø = 7.8 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with the following pore sizes:
  — *column 1*: 50 nm;
  — *column 2*: 10 nm;
  — *column 3*: 5 nm;
— *connection sequence*: injector – column 1 – column 2 – column 3 – detector.

Mobile phase *tetrahydrofuran R*.

Flow rate 0.8 mL/min.

Detection Differential refractometer.

Injection 40 µL.

System suitability Reference solution:
— *elution order*: tridocosahexaenoin, didocosahexaenoin, monodocosahexaenoin;
— *resolution*: minimum 2.0 between the peaks due to didocosahexaenoin and monodocosahexaenoin and minimum 1.0 between the peaks due to tridocosahexaenoin and didocosahexaenoin.

Identify the peaks from the chromatogram (Figure 1912.-1). Calculate the percentage content of oligomers using the following expression:

$$\frac{B}{A} \times 100$$

A = sum of the areas of all the peaks in the chromatogram;
B = area of the peak with a retention time less than the retention time of the triglyceride peak.

Limit:
— *oligomers*: maximum 1.5 per cent.

ASSAY

EPA and DHA (*2.4.29*)
For identification of the peaks, see Figure 1912.-2.

Total omega-3 acids (*2.4.29*)
See Figure 1912.-2.

STORAGE

Under an inert gas, in a well-filled, airtight container, protected from light.

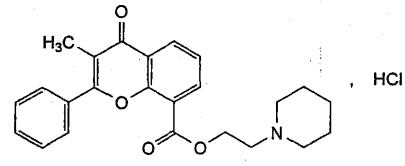
LABELLING

The label states:
— the concentration of EPA, DHA and total omega-3 acids, expressed as triglycerides;
— the type of fish oil rich in omega-3 acids (type I or II).

Ph Eur

Flavoxate Hydrochloride

(Ph. Eur. monograph 1692)



$C_{24}H_{26}ClNO_4$ 427.9 *3717-88-2*

Action and use
Anticholinergic.

Preparation
Flavoxate Tablets

Ph Eur

DEFINITION

2-(Piperidin-1-yl)ethyl 3-methyl-4-oxo-2-phenyl-4*H*-1-benzopyran-8-carboxylate hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *flavoxate hydrochloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Use freshly prepared solutions.*

Solvent mixture Mix 20 volumes of a 0.4 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 3.0 with *phosphoric acid R* and 80 volumes of *acetonitrile R*.

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 6.0 mg of *flavoxate impurity A CRS* and 3.0 mg of *flavoxate impurity B CRS* in the solvent mixture, add 2.0 mL of the test solution and

dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 0.435 g/L solution of *dipotassium hydrogen phosphate R* adjusted to pH 7.5 with *phosphoric acid R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	20	80
10 - 20	20 → 10	80 → 90
20 - 25	10	90

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Relative retention With reference to flavoxate (retention time = about 10 min): impurity A = about 0.2; impurity B = about 0.8.

System suitability Reference solution (c):
— *resolution*: minimum 4.0 between the peaks due to impurity B and flavoxate.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total of unspecified impurities*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.350 g in 10 mL of *anhydrous formic acid R* and add 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 42.79 mg of $C_{24}H_{26}ClNO_4$.

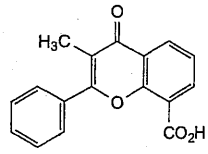
STORAGE

Protected from light.

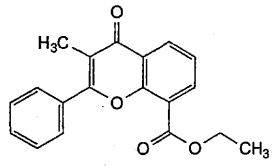
IMPURITIES

Specified impurities *A, B.*

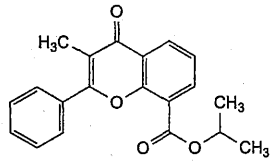
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*



A. 3-methyl-4-oxo-2-phenyl-4*H*-1-benzopyran-8-carboxylic acid,



B. ethyl 3-methyl-4-oxo-2-phenyl-4*H*-1-benzopyran-8-carboxylate,

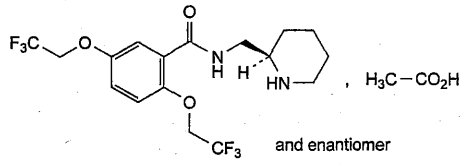


C. 1-methylethyl 3-methyl-4-oxo-2-phenyl-4*H*-1-benzopyran-8-carboxylate.

Ph Eur

Flecainide Acetate

(Ph. Eur. monograph 1324)



$C_{19}H_{24}F_6N_2O_5$ 474.4 *54143-56-5*

Action and use

Class I antiarrhythmic.

Preparations

Flecainide Injection

Flecainide Tablets

Ph Eur

DEFINITION

N-[(*RS*)-(Piperidin-2-ylmethyl)]-2,5-bis(2,2,2-trifluoroethoxy) benzamide acetate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, very hygroscopic, crystalline powder.

Solubility
Soluble in water and in anhydrous ethanol. It is freely soluble in dilute acetic acid and practically insoluble in dilute hydrochloric acid.

IDENTIFICATION

First identification: A, C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 146 °C to 152 °C, with a melting range not greater than 3 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50 mg in *ethanol (96 per cent) R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *ethanol (96 per cent) R*.

Spectral range 230-350 nm.

Absorption maximum At 298 nm.

Specific absorbance at the absorption maximum 61 to 65.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flecainide acetate CRS*.

D. It gives reaction (b) of acetates (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.50 g in *water R*, add 0.1 mL of *glacial acetic acid R* and dilute to 20 mL with *water R*.

pH (*2.2.3*)
6.7 to 7.1.

Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Impurity B
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in *methanol R* and dilute to 2.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of *flecainide impurity B CRS* in *methanol R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 0.10 g of the substance to be examined in reference solution (a) and dilute to 2.0 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R*.

Mobile phase Freshly prepared mixture of 5 volumes of *concentrated ammonia R* and 95 volumes of *acetone R*.

Application 5 µL.

Development Over 1/2 of the plate.

Drying At 100-105 °C until the ammonia has evaporated.

Detection Examine in ultraviolet light at 254 nm to establish the position of the flecainide spot, then spray with a freshly prepared 2 g/L solution of *ninhydrin R* in *methanol R* and heat at 100-110 °C for 2-5 min; examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Limit:
— *impurity B*: any spot due to impurity B is not more intense than the corresponding spot in the chromatogram obtained with reference solution (a) (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.25 g of the substance to be examined in *methanol R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (b) Dissolve 5 mg of *flecainide impurity A CRS* in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution (c) Dissolve 5 mg of *flecainide for system suitability CRS* (containing impurities C, D and E) in 1.0 mL of *methanol R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 2 mL of *concentrated ammonia R*, 4 mL of *triethylamine R* and 985 mL of *water R*; add 6 mL of *phosphoric acid R* and adjust to pH 2.8 with *concentrated ammonia R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	90	10
5 - 17	90 → 30	10 → 70
17 - 22	30	70

If a suitable baseline cannot be obtained, use another grade of triethylamine.

Flow rate 2 mL/min.

Detection Spectrophotometer at 300 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *flecainide for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C, D, and E; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to flecainide (retention time = about 11 min): impurity C = about 0.9; impurity A = about 1.1; impurity E = about 1.28; impurity D = about 1.32.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurities E and D.

Limits:
— *impurities A, C, D, E*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.6 kPa for 2 h.

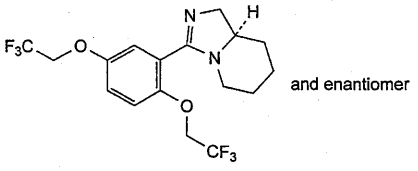
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.400 g in 25 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 47.44 mg of $C_{19}H_{24}F_6N_2O_5$.

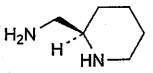
STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *A, B, C, D, E.*



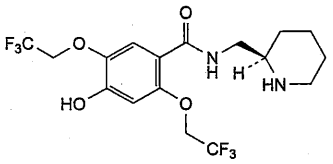
and enantiomer

A. (8a*RS*)-3-[2,5-bis(2,2,2-trifluoroethoxy)phenyl]-1,5,6,7,8,8a-hexahydroimidazo[1,5-*a*]pyridine,



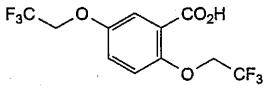
and enantiomer

B. (*RS*)-(piperidin-2-yl)methanamine,

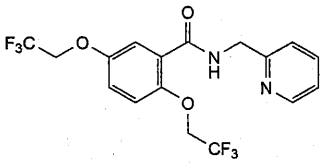


and enantiomer

C. (*RS*)-4-hydroxy-*N*-(piperidin-2-ylmethyl)-2,5-bis(2,2,2-trifluoroethoxy)benzamide,



D. 2,5-bis(2,2,2-trifluoroethoxy)benzoic acid,

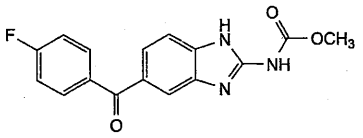


E. *N*-(pyridin-2-ylmethyl)-2,5-bis(2,2,2-trifluoroethoxy) benzamide.

Ph Eur

Flubendazole

(Ph. Eur. monograph 1721)



$C_{16}H_{12}FN_3O_3$ 313.3 *31430-15-6*

Action and use
Benzimadazole antihelminthic.

Ph Eur

DEFINITION
Methyl [5-(4-fluorobenzoyl)-1*H*-benzimidazol-2-yl] carbamate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, in alcohol and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*), without recrystallisation.

Comparison *flubendazole CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *flubendazole for system suitability CRS* in *dimethylformamide R* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *dimethylformamide R*. Dilute 5.0 mL of this solution to 20.0 mL with *dimethylformamide R*.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm),
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: 7.5 g/L solution of *ammonium acetate R*,
— *mobile phase B*: *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	90 → 75	10 → 25
15 - 30	75 → 45	25 → 55
30 - 32	45 → 10	55 → 90
32 - 37	10	90

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 µL.

System suitability Reference solution (a):
— the chromatogram obtained is similar to the chromatogram supplied with *flubendazole for system suitability CRS*.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.4; impurity C = 1.3; impurity D = 1.3; impurity G = 1.4,
— *impurities A, B, C, D, E, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent),
— *impurity F*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *any other impurity with a relative retention between 1.2 and 1.3*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent),
— *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent),
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying *(2.2.32)*
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C, for 4 h.

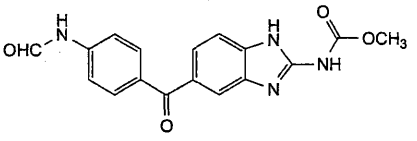
Sulfated ash *(2.4.14)*
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 3 mL of *anhydrous formic acid R* and add 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically *(2.2.20)*.

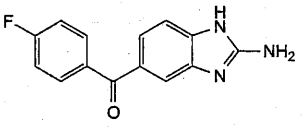
1 mL of *0.1 M perchloric acid* is equivalent to 31.33 mg of $C_{16}H_{12}FN_3O_3$.

STORAGE
Protected from light.

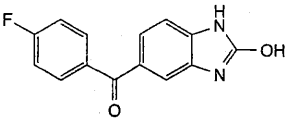
IMPURITIES
Specified impurities *A, B, C, D, E, F, G.*



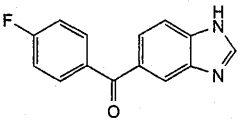
A. methyl [5-[4-(formylamino)benzoyl]-1*H*-benzimidazol-2-yl]carbamate,



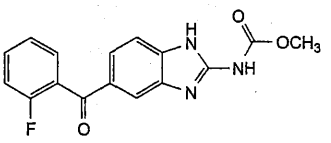
B. (2-amino-1*H*-benzimidazol-5-yl)(4-fluorophenyl)methanone,



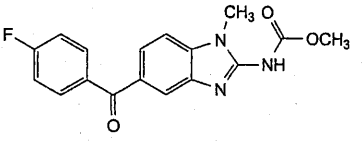
C. (4-fluorophenyl)(2-hydroxy-1*H*-benzimidazol-5-yl)methanone,



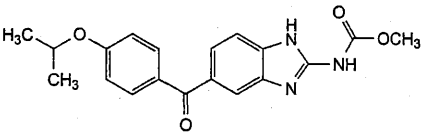
D. (1*H*-benzimidazol-5-yl)(4-fluorophenyl)methanone,



E. methyl [5-(2-fluorobenzoyl)-1*H*-benzimidazol-2-yl]carbamate,



F. methyl [5-(4-fluorobenzoyl)-1-methyl-1*H*-benzimidazol-2-yl]carbamate,



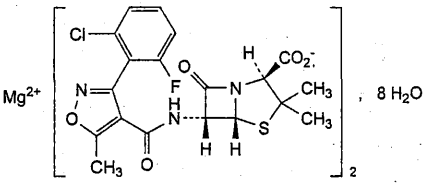
G. methyl [5-[4-(1-methylethoxy)benzoyl]-1*H*-benzimidazol-2-yl]carbamate.

Ph Eur

Flucloxacillin Magnesium Octahydrate

Flucloxacillin Magnesium

(Ph. Eur. monograph 2346)



$C_{38}H_{32}Cl_2F_2MgN_6O_{10}S_2,8H_2O$ 1074 *58486-36-5*

Action and use
Penicillin antibacterial.

Preparations
Flucloxacillin Oral Suspension
Co-fluampicil Oral Suspension

Ph Eur

DEFINITION

Magnesium bis[(2*S*,5*R*,6*R*)-6-[[[3-(2-chloro-6-fluorophenyl)-5-methylisoxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate] octahydrate.

Semi-synthetic product derived from a fermentation product.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methanol.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flucloxacillin magnesium octahydrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *cloxacillin sodium CRS*, 25 mg of *dicloxacillin sodium CRS* and 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* previously adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose the plate to iodine vapour until the spots appear.

System suitability Reference solution (b):

— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. It gives the reaction of magnesium (*2.3.1*).

TESTS

pH (*2.2.3*)

4.5 to 6.5.

Dissolve 0.25 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Specific optical rotation (*2.2.7*)

+ 163 to + 175 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *flucloxacillin sodium CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of test solution (b) to 50.0 mL with the mobile phase.

Reference solution (c) In order to prepare impurity A *in situ*, add 1 mL of *sodium carbonate solution R* to 10 mg of the substance to be examined, dilute to 25 mL with *water R* and place in an oven at 70 °C for 20 min.

Reference solution (d) Dilute 1 mL of reference solution (c) to 10 mL with a 27 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 3.5 with *dilute phosphoric acid R.*

Reference solution (e) In order to prepare impurity B *in situ*, add 5 mL of *dilute hydrochloric acid R* to 10 mL of reference solution (c), dilute to 25 mL with *water R* and place in an oven at 70 °C for 1 h. Dilute 1 mL of this solution to 5 mL with a 27 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 7.0 with *phosphoric acid R.*

Reference solution (f) Dilute 2 mL of reference solution (a) to 10 mL with reference solution (e).

Reference solution (g) Dissolve 1.5 mg of *flucloxacillin impurity C CRS* in 1 mL of the mobile phase and dilute to 50 mL with the mobile phase.

Reference solution (h) Dissolve 1 mg of *flucloxacillin impurity D CRS* in 100 mL of the mobile phase.

Reference solution (i) Dissolve 1 mg of *flucloxacillin impurity E CRS* in 100 mL of the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 40 °C.

Mobile phase Mix 25 volumes of *acetonitrile R1* and 75 volumes of a 2.7 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 5.0 with *dilute sodium hydroxide solution R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (d), (e), (f), (g), (h) and (i).

Run time 7 times the retention time of flucloxacillin.

Identification of impurities Use the chromatograms obtained with reference solutions (d), (e), (g), (h) and (i) to identify the peaks due to impurities A, B, C, D and E respectively.

Relative retention With reference to flucloxacillin (retention time = about 8 min): impurity C = about 0.2; impurity A (isomer 1) = about 0.3; impurity A (isomer 2) = about 0.5; impurity D = about 0.6; impurity B (isomer 1) = about 0.8; impurity B (isomer 2) = about 0.9; impurity E = about 6.

System suitability Reference solution (f):

— *resolution*: minimum 2.0 between the 2nd peak due to impurity B (isomer 2) and the peak due to flucloxacillin.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 3.3;

— *impurity A* (sum of the 2 isomers): the sum of the areas of the 2 peaks is not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— *impurity B* (sum of the 2 isomers): the sum of the areas of the 2 peaks is not more than the area of the principal

peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *impurities D, E*: for each impurity, not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *any other impurity*: for each impurity, not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.8 per cent *m/m*.

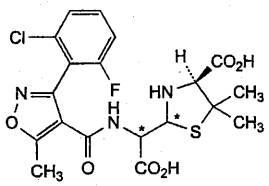
Water (*2.5.12*)
12.0 per cent to 15.0 per cent, determined on 0.100 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

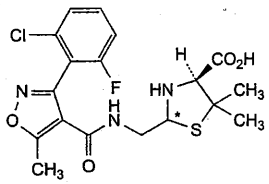
Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{38}H_{32}Cl_2F_2MgN_6O_{10}S_2$ from the declared content of *flucloxacillin sodium CRS*, multiplying by 0.9773.

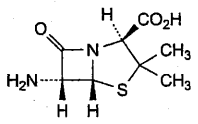
IMPURITIES
Specified impurities *A, B, C, D, E.*



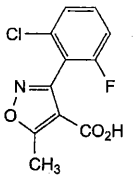
A. (4*S*)-2-[carboxy[[[3-(2-chloro-6-fluorophenyl)-5-methylisoxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of flucloxacillin),



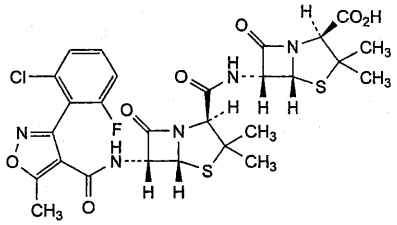
B. (2*RS*,4*S*)-2-[[[[3-(2-chloro-6-fluorophenyl)-5-methylisoxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of flucloxacillin),



C. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



D. 3-(2-chloro-6-fluorophenyl)-5-methylisoxazole-4-carboxylic acid,

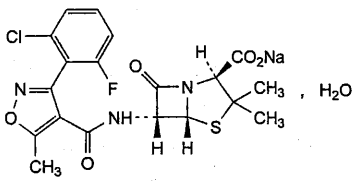


E. (2*S*,5*R*,6*R*)-6-[[[(2*S*,5*R*,6*R*)-6-[[[3-(2-chloro-6-fluorophenyl)-5-methylisoxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-APA flucloxacillin amide).

Ph Eur

Flucloxacillin Sodium

(Ph. Eur. monograph 0668)



$C_{19}H_{16}ClFN_3NaO_5S,H_2O$ 493.9

Action and use
Penicillin antibacterial.

Preparations
Flucloxacillin Capsules
Co-fluampicil Capsules
Flucloxacillin Injection
Flucloxacillin Oral Solution

Ph Eur

DEFINITION
Sodium (2*S*,5*R*,6*R*)-6-[[[3-(2-chloro-6-fluorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate monohydrate.

Semi-synthetic product derived from a fermentation product.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water and in methanol, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flucloxacillin sodium CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of *water R.*

Reference solution (a) Dissolve 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Reference solution (b) Dissolve 25 mg of *cloxacillin sodium CRS*, 25 mg of *dicloxacillin sodium CRS* and 25 mg of *flucloxacillin sodium CRS* in 5 mL of *water R.*

Plate *TLC silanised silica gel plate R.*

Mobile phase Mix 30 volumes of *acetone R* and 70 volumes of a 154 g/L solution of *ammonium acetate R* adjusted to pH 5.0 with *glacial acetic acid R.*

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of *water R* and add 2 mL of *sulfuric acid-formaldehyde reagent R.* Mix the contents of the tube by swirling; the colour of the solution is slightly greenish-yellow. Place the test-tube in a water-bath for 1 min; the solution becomes yellow.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and its absorbance (*2.2.25*) at 430 nm is not greater than 0.04.

pH (*2.2.3*)

5.0 to 7.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 158 to + 168 (anhydrous substance).

Dissolve 0.250 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *flucloxacillin sodium CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of *flucloxacillin sodium CRS* and 5 mg of *cloxacillin sodium CRS* in the mobile phase, then dilute to 50.0 mL with the mobile phase.

Column:
— *size:* l = 0.25 m, Ø = 4 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 25 volumes of *acetonitrile R1* and 75 volumes of a 2.7 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 5.0 with *dilute sodium hydroxide solution R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL of test solution (a) and reference solutions (b) and (c).

Run time 6 times the retention time of flucloxacillin.

System suitability Reference solution (c):
— *resolution:* minimum 2.5 between the peaks due to cloxacillin (1st peak) and flucloxacillin (2nd peak).

Limits:
— *impurities A, B, C, D, E:* for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *total:* not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5 per cent);
— *disregard limit:* 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

N,N-Dimethylaniline (*2.4.26, Method B*)

Maximum 20 ppm.

2-Ethylhexanoic acid (*2.4.28*)

Maximum 0.8 per cent *m/m.*

Water (*2.5.12*)

3.0 per cent to 4.5 per cent, determined on 0.300 g.

Pyrogens (*2.6.8*)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of pyrogens, it complies with the test. Inject per kilogram of the rabbit's mass 1 mL of a solution in *water for injections R* containing 20 mg of the substance to be examined per millilitre.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

System suitability Reference solution (a):
— *repeatability:* maximum relative standard deviation of 1.0 per cent after 6 injections.

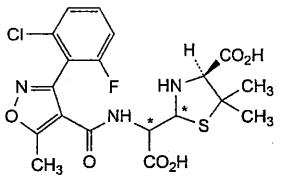
Calculate the percentage content of $C_{19}H_{16}ClFN_3NaO_5S$ from the declared content of *flucloxacillin sodium CRS.*

STORAGE

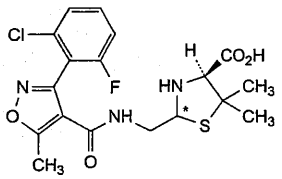
In an airtight container, at a temperature not exceeding 25 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

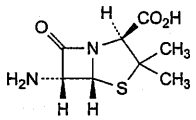
Specified impurities *A, B, C, D, E.*



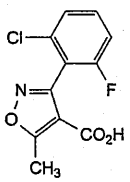
A. (4*S*)-2-[carboxy[[[3-(2-chloro-6-fluorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penicilloic acids of flucloxacillin),



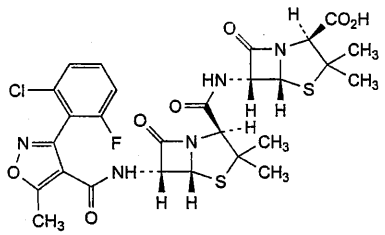
B. (2*RS*,4*S*)-2-[[[[3-(2-chloro-6-fluorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]methyl]-5,5-dimethyl-1,3-thiazolidine-4-carboxylic acid (penilloic acids of flucloxacillin),



C. (2*S*,5*R*,6*R*)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



D. 3-(2-chloro-6-fluorophenyl)-5-methyl-1,2-oxazole-4-carboxylic acid,

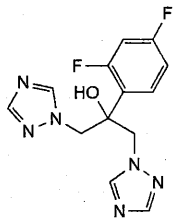


E. (2*S*,5*R*,6*R*)-6-[[[(2*S*,5*R*,6*R*)-6-[[[3-(2-chloro-6-fluorophenyl)-5-methyl-1,2-oxazol-4-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid.

Ph Eur

Fluconazole

(Ph. Eur. monograph 2287)



$C_{13}H_{12}F_2N_6O$ 306.3 *86386-73-4*

Action and use
Antifungal.

Preparations
Fluconazole Capsules
Fluconazole Oral Suspension
Fluconazole Infusion

Ph Eur

DEFINITION

2-(2,4-Difluorophenyl)-1,3-bis(1*H*-1,2,4-triazol-1-yl)propan-2-ol.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in methanol, soluble in acetone.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluconazole CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *methylene chloride R*, evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase, sonicate if necessary, and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *fluconazole for peak identification CRS* (containing impurity A) in the mobile phase, sonicate if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (c) Dissolve 3.0 mg of *fluconazole impurity B CRS* in the mobile phase, sonicate if necessary, and dilute to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve 3.0 mg of *fluconazole impurity C CRS* in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (e) To 1.0 mL of reference solution (d) add 1.0 mL of the test solution and dilute to 10.0 mL with the mobile phase.

Reference solution (f) Dilute 1.0 mL of reference solution (d) to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R1* (5 µm);
— *temperature*: 40 °C.

Mobile phase *acetonitrile R*, 0.63 g/L solution of *ammonium formate R* (14:86 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 µL of the test solution and reference solutions (a), (b), (c), (e) and (f).

Run time 3.5 times the retention time of fluconazole.

Identification of impurities Use the chromatogram supplied with *fluconazole for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B and the chromatogram obtained with reference solution (f) to identify the peak due to impurity C.

Relative retention With reference to fluconazole (retention time = about 11 min): impurity B = about 0.4; impurity A = about 0.5; impurity C = about 0.8.

System suitability Reference solution (e):
— *resolution*: minimum 3.0 between the peaks due to impurity C and fluconazole.

Limits:
— *impurity A*: not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (f) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: maximum 0.6 per cent;
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.125 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 15.32 mg of $C_{13}H_{12}F_2N_6O$.

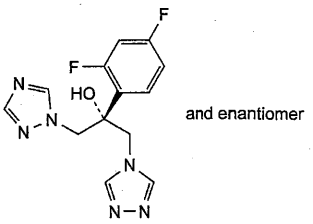
STORAGE

In an airtight container.

IMPURITIES

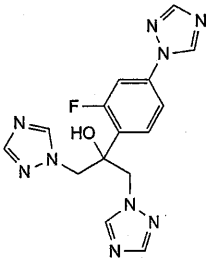
Specified impurities *A, B, C.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E, F, G, H, I.*

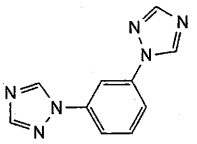


and enantiomer

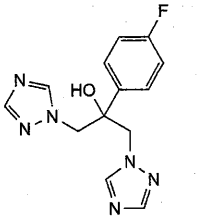
A. (2*RS*)-2-(2,4-difluorophenyl)-1-(1*H*-1,2,4-triazol-1-yl)-3-(4*H*-1,2,4-triazol-4-yl)propan-2-ol,



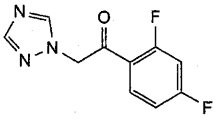
B. 2-[2-fluoro-4-(1*H*-1,2,4-triazol-1-yl)phenyl]-1,3-bis(1*H*-1,2,4-triazol-1-yl)propan-2-ol,



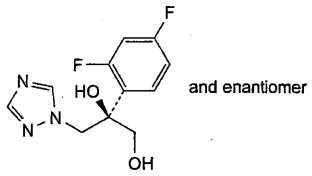
C. 1,1′-(1,3-phenylene)di-1*H*-1,2,4-triazole,



D. 2-(4-fluorophenyl)-1,3-bis(1*H*-1,2,4-triazol-1-yl)propan-2-ol,

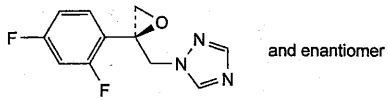


E. 1-(2,4-difluorophenyl)-2-(1*H*-1,2,4-triazol-1-yl)ethanone,



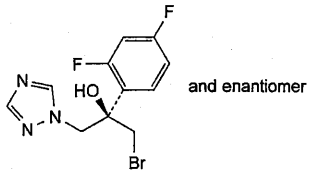
and enantiomer

F. (2*RS*)-2-(2,4-difluorophenyl)-3-(1*H*-1,2,4-triazol-1-yl) propane-1,2-diol,



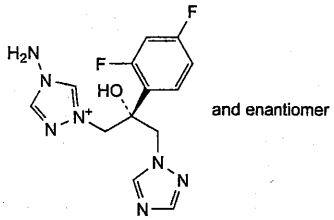
and enantiomer

G. 1-[[(2*RS*)-2-(2,4-difluorophenyl)oxiran-2-yl]methyl]-1*H*-1,2,4-triazole,



and enantiomer

H. (2*RS*)-1-bromo-2-(2,4-difluorophenyl)-3-(1*H*-1,2,4-triazol-1-yl)propan-2-ol,



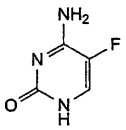
and enantiomer

I. 4-amino-1-[(2*RS*)-2-(2,4-difluorophenyl)-2-hydroxy-3(1*H*-1,2,4-triazol-1-yl)propyl]-4*H*-1,2,4-triazolium.

Ph Eur

Flucytosine

(Ph. Eur. monograph 0766)



$C_4H_4FN_3O$ 129.1 *2022-85-7*

Action and use
Antifungal.

Ph Eur

DEFINITION
4-Amino-5-fluoropyrimidin-2(1*H*)-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flucytosine CRS.*

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *water R, methanol R* (10:15 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution Dissolve 10 mg of *flucytosine CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous formic acid R, water R, methanol R, ethyl acetate R* (1:15:25:60 *V/V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate in an unsaturated tank with the mobile phase. Then allow the solvents to evaporate.

Detection At the bottom of a chromatography tank place an evaporating dish containing a mixture of 1 volume of *hydrochloric acid R1*, 1 volume of *water R* and 2 volumes of a 15 g/L solution of *potassium permanganate R*. Close the tank and allow to stand for 15 min. Place the dried plate in the tank and close the tank. Leave the plate in contact with the chlorine vapour for 5 min. Withdraw the plate and place it in a current of cold air until the excess of chlorine is removed and an area of the coating below the points of application does not give a blue colour with a drop of *potassium iodide and starch solution R*. Spray with *potassium iodide and starch solution R*. Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter and add to the filtrate a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The colour of the solution changes from red to yellow.

D. To 5 mL of solution S (see Tests) add 0.15 mL of *bromine water R* and shake. The colour of the solution is discharged.

TESTS
Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 or Y_7 (*2.2.2, Method II*).

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Dissolve 13.6 g of *potassium dihydrogen phosphate R* in 950 mL of *water R*. Add 50 mL of *methanol R*. Mix thoroughly.

Test solution Dissolve 15.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Mix well. Sonicate for 5 min. Mix thoroughly. Sonicate the solution for 5 min. Mix thoroughly.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 15.0 mg of *fluorouracil CRS* (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Mix well. Sonicate for 5 min. Mix thoroughly. Sonicate the solution for 5 min. Mix thoroughly. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *flucytosine for system suitability CRS* (containing impurity B) in 0.5 mL of the solvent mixture and add 0.5 mL of reference solution (b).

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 13.6 g of *potassium dihydrogen phosphate R* in 950 mL of *water R*. Filter through a membrane filter (nominal pore size 0.45 µm). Adjust to pH 2.0 by adding *phosphoric acid R* and add 50 mL of *methanol R*. Mix thoroughly.

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 µL of the test solution and reference solutions (a) and (c).

Run time 15 times the retention time of flucytosine.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to flucytosine (retention time = about 2 min): impurity A = about 1.7; impurity B = about 13.3.

System suitability:
- *resolution*: minimum 5.0 between the peaks due to flucytosine and impurity A in the chromatogram obtained with reference solution (c);
- *signal-to-noise ratio*: minimum 50 for the peak due to impurity B in the chromatogram obtained with reference solution (c);
- *symmetry factor*: maximum 2.0 for the peak due to flucytosine in the chromatogram obtained with reference solution (a).

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity B by 0.6;
- *impurity A*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
- *impurity B*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Fluorides

Maximum 200 ppm.

Potentiometry (*2.2.36, Method I*). *Prepare and store all solutions in plastic containers.*

Buffer solution Dissolve 58 g of *sodium chloride R* in 500 mL of *water R*. Add 57 mL of *glacial acetic acid R* and 200 mL of a 100 g/L solution of *cyclohexylenedinitrilotetra-acetic acid R* in *1 M sodium hydroxide*. Adjust the pH to 5.0-5.5 with a 200 g/L solution of *sodium hydroxide R* and dilute to 1000.0 mL with *water R*.

Test solution Dissolve 1.00 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solutions Dissolve 4.42 g of *sodium fluoride R*, previously dried at 120 °C for 2 h, in 300 mL of *water R* and dilute to 1000.0 mL with the same solvent (solution (a): 1.9 g/L of fluoride). Prepare 3 reference solutions by dilution of solution (a) 1 in 100, 1 in 1000 and 1 in 10 000 respectively.

Indicator electrode Fluoride selective.

Reference electrode Silver-silver chloride.

To 20.0 mL of the test solution and each reference solution, add 10.0 mL of the buffer solution and stir with a magnetic stirrer. Introduce the electrodes into the solution and allow to stand for 5 min with constant stirring.

Calculate the concentration of fluorides using the calibration curve.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.100 g in 40 mL of *anhydrous acetic acid R* and add 100 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).

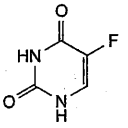
1 mL of *0.1 M perchloric acid* is equivalent to 12.91 mg of $C_4H_4FN_3O$.

STORAGE

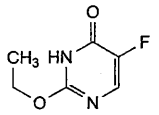
Protected from light.

IMPURITIES

Specified impurities *A, B.*



A. 5-fluoropyrimidine-2,4(1*H*,3*H*)-dione (fluorouracil),

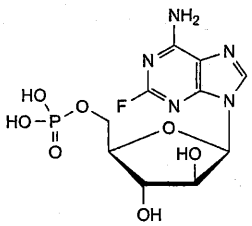


B. 2-ethoxy-5-fluoropyrimidin-4(3*H*)-one.

Ph Eur

Fludarabine Phosphate

(Ph. Eur. monograph 1781)



$C_{10}H_{13}FN_5O_7P$ 365.2 *75607-67-9*

Action and use
Purine analogue; cytotoxic.

Ph Eur

DEFINITION
2-Fluoro-9-(5-*O*-phosphono-β-D-arabinofuranosyl)-9*H*-purin-6-amine.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Slightly soluble in water, freely soluble in dimethylformamide, very slightly soluble in anhydrous ethanol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *fludarabine phosphate CRS.*

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).
Dissolve 50 mg in 5.0 mL of *dimethylformamide R* with the aid of ultrasound.

Specific optical rotation (*2.2.7*)
+ 10.0 to + 14.0 (anhydrous substance).
Dissolve 0.100 g in *water R* with the aid of ultrasound and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure. *Prepare the solutions immediately before use.*
Test solution Dissolve 20 mg of the substance to be examined in 50 mL of *water R* with the aid of ultrasound and dilute to 100.0 mL with the same solvent.
Reference solution (a) Dissolve 20 mg of *fludarabine phosphate CRS* in 50 mL of *water R* with the aid of ultrasound and dilute to 100.0 mL with the same solvent.
Reference solution (b) Dissolve 20 mg of the substance to be examined in 20 mL of *0.1 M hydrochloric acid* with the aid of ultrasound. Heat in a water-bath at 80 °C for 15 min, cool to room temperature, mix and dilute to 100.0 mL with *water R.*
Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *water R.* Dilute 1.0 mL of this solution to 20.0 mL with *water R.*
Reference solution (d) Dissolve 5 mg of *fludarabine for system suitability CRS* (containing impurities D, E and F) in 10 mL of *water R* with the aid of ultrasound and dilute to 25.0 mL with the same solvent.
Blank solution *0.02 M hydrochloric acid.*

A. Early eluting impurities.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 60 volumes of *methanol R* and 940 volumes of a 1.36 g/L solution of *potassium dihydrogen phosphate R.*
Flow rate 1 mL/min.
Detection Spectrophotometer at 260 nm and at 292 nm.
Injection 10 µL of the test solution and reference solutions (a), (b) and (c).
Run time 4.5 times the retention time of the principal peak in the chromatogram obtained with the test solution.
Identification of impurities Use the chromatogram obtained with reference solution (b) at 292 nm to identify the peaks due to impurities A and B, the response at 292 nm being much higher than at 260 nm; use the chromatogram supplied with *fludarabine phosphate CRS* and the chromatogram obtained with reference solution (a) at 260 nm to identify impurity C.
Relative retention With reference to fludarabine phosphate (retention time = about 9 min): impurity A = about 0.26; impurity B = about 0.34; impurity C = about 0.42.
System suitability Reference solution (b) at 292 nm:
- *resolution*: minimum 2.0 between the peaks due to impurities A and B.

Limits At 260 nm:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 4.0; impurity B = 2.5; impurity C = 1.9;
- *impurity A*: maximum 0.8 per cent;
- *impurity C*: maximum 0.4 per cent;
- *impurity B*: maximum 0.2 per cent;
- *unspecified impurities eluting before fludarabine phosphate*: for each impurity, maximum 0.10 per cent;
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak eluting after fludarabine phosphate.

B. Late eluting impurities.

Conditions as described under Test A with the following modifications.
Mobile phase Mix 200 volumes of *methanol R* and 800 volumes of a 1.36 g/L solution of *potassium dihydrogen phosphate R.*
Detection Spectrophotometer at 260 nm.
Injection 10 µL of the test solution and reference solutions (c) and (d).

Run time 8 times the retention time of the principal peak in the chromatogram obtained with the test solution.

Identification of impurities Use the chromatogram supplied with *fludarabine for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities D, E and F.

Relative retention With reference to fludarabine phosphate (retention time = about 2.5 min): impurity D = about 1.5; impurity E = about 1.9; impurity F = about 2.5.

System suitability Reference solution (d):
- *resolution*: minimum 5.0 between the peaks due to fludarabine phosphate and impurity D.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.5; impurity E = 0.6; impurity F = 1.8;
- *impurity E*: maximum 0.2 per cent;
- *impurity F*: maximum 0.2 per cent;
- *impurity D*: maximum 0.15 per cent;
- *unspecified impurities eluting after fludarabine phosphate*: for each impurity, maximum 0.10 per cent;
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak eluting before fludarabine phosphate.

Total of impurities eluting before fludarabine phosphate in test A, apart from impurities A, B and C, and after fludarabine phosphate in test B, apart from impurities D, E and F
Maximum 0.5 per cent.

Total of all impurities eluting before fludarabine phosphate in test A and after fludarabine phosphate in test B Maximum 2.0 per cent.

Ethanol (*2.4.24, System A*)
Maximum 1.0 per cent.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 0.200 g (ground to a very fine powder). Stir the substance in 15 mL of *anhydrous methanol R* for about 15 s before titrating.

ASSAY

Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modifications.

Test solution Dissolve 24.0 mg of the substance to be examined in 50 mL of *water R* with the aid of ultrasound and dilute to 100.0 mL with the same solvent. Dilute 25.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution Dissolve 24.0 mg of *fludarabine phosphate CRS* in 50 mL of *water R* with the aid of ultrasound and dilute to 100.0 mL with the same solvent. Dilute 25.0 mL of the solution to 100.0 mL with the mobile phase.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Calculate the percentage content of $C_{10}H_{13}FN_5O_7P$ taking into account the assigned content of *fludarabine phosphate CRS*.

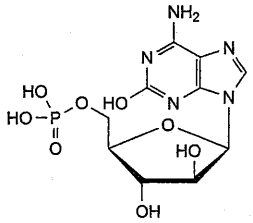
STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

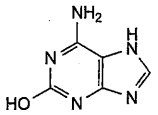
IMPURITIES

Specified impurities *A, B, C, D, E, F.*

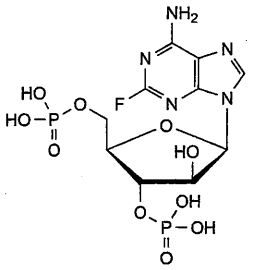
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, H, I, J.*



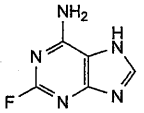
A. 6-amino-9-(5-*O*-phosphono-β-D-arabinofuranosyl)-9*H*-purin-2-ol,



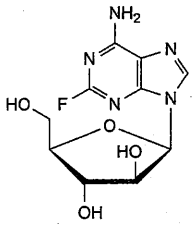
B. 6-amino-7*H*-purin-2-ol,



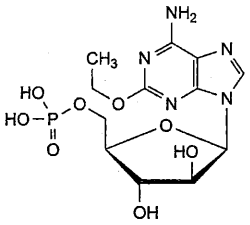
C. 9-(3,5-di-*O*-phosphono-β-D-arabinofuranosyl)-2-fluoro-9*H*-purin-6-amine,



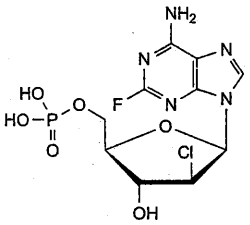
D. 2-fluoro-7*H*-purin-6-amine,



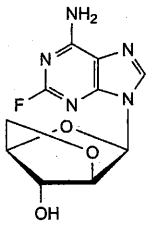
E. 9-β-D-arabinofuranosyl-2-fluoro-9*H*-purin-6-amine,



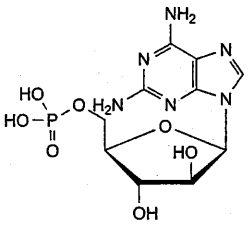
F. 2-ethoxy-9-(5-*O*-phosphono-β-D-arabinofuranosyl)-9*H*-purin-6-amine,



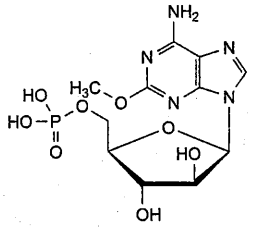
G. 9-(2-chloro-2-deoxy-5-*O*-phosphono-β-D-arabinofuranosyl)-2-fluoro-9*H*-purin-6-amine,



H. 9-(2,5-anhydro-β-D-arabinofuranosyl)-2-fluoro-9*H*-purin-6-amine,



I. 9-(5-*O*-phosphono-β-D-arabinofuranosyl)-9*H*-purine-2,6-diamine,

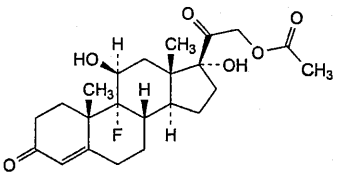


J. 2-methoxy-9-(5-*O*-phosphono-β-D-arabinofuranosyl)-9*H*-purin-6-amine.

Ph Eur

Fludrocortisone Acetate

(Ph. Eur. monograph 0767)



$C_{23}H_{31}FO_6$ 422.5 *514-36-3*

Action and use
Mineralocorticoid.

Preparation
Fludrocortisone Tablets

Ph Eur

DEFINITION
9-Fluoro-11β,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in anhydrous ethanol.

IDENTIFICATION
First identification: A, B.

Second identification: C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fludrocortisone acetate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *acetone R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of *fludrocortisone acetate CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *cortisone acetate CRS* in 5 mL of reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of the solution to 10 mL with *methylene chloride R*.

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* through the solution for 5 min. Stopper the tube. Heat on a water-bath at 45 °C protected from light for 2.5 h. Allow to cool.

Reference solution (a) Dissolve 25 mg of *fludrocortisone acetate CRS* in *methanol R* and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of the solution to 10 mL with *methylene chloride R*.

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* through the solution for 5 min. Stopper the tube. Heat on a water bath at 45 °C protected from light for 2.5 h. Allow to cool.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R*.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spots in the chromatograms obtained with test solution (b) and reference solution (b) have R_F values distinctly lower than those of the principal spots in the chromatograms obtained with test solution (a) and reference solution (a).

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter and add to the filtrate a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The colour of the solution to be examined changes from red to yellow.

E. About 10 mg gives the reaction of acetyl (*2.3.1*).

TESTS

Specific optical rotation (*2.2.7*)

+ 148 to + 156 (dried substance).

Dissolve 0.250 g in *dioxan R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of *fludrocortisone acetate CRS* and 2.0 mg of *hydrocortisone acetate CRS* in the mobile phase, then dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Column:
— *size*: l = 0.2 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *tetrahydrofuran R, water R* (35:65 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 30 min.

Injection 20 µL.

Run time Twice the retention time of fludrocortisone acetate.

Retention time Hydrocortisone acetate = about 8.5 min; fludrocortisone acetate = about 10 min.

System suitability Reference solution (a):
— *resolution*: minimum 1.0 between the peaks due to hydrocortisone acetate and fludrocortisone acetate; if necessary, adjust slightly the concentration of tetrahydrofuran in the mobile phase (an increase in the concentration of tetrahydrofuran reduces the retention times).

Limits:
— *any impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *total*: not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Dissolve 10.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the

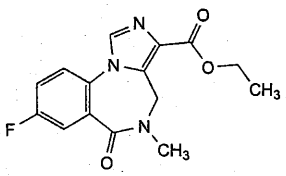
solution to 50.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 238 nm.

Calculate the content of $C_{23}H_{31}FO_6$ taking the specific absorbance to be 405.

Ph Eur

Flumazenil

(Ph. Eur. monograph 1326)



$C_{15}H_{14}FN_3O_3$ 303.3 *78755-81-4*

Action and use
Benzodiazepine receptor antagonist.

Ph Eur

DEFINITION

Ethyl 8-fluoro-5-methyl-6-oxo-5,6-dihydro-4*H*-imidazo[1,5-*a*][1,4]benzodiazepine-3-carboxylate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, freely soluble in methylene chloride, sparingly soluble in methanol.

mp
198 °C to 202 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of flumazenil.*

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and is not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 0.10 g in *methanol R* and dilute to 10 mL with the same solvent.

Impurity C
Maximum 1 per cent.

Dissolve 0.10 g in 0.5 mL of *methylene chloride R* and dilute to 10 mL with *butanol R*. To 5.0 mL of this solution add 2.0 mL of *ninhydrin solution R* and heat in a water-bath at 95 °C for 15 min. Any blue-purple colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 5.0 mL of a 0.1 g/L solution of *dimethylformamide diethylacetal R* in *butanol R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in 5 mL of *methanol R* and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of *flumazenil impurity B CRS* and 2.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 10.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase To 800 mL of *water R* adjusted to pH 2.0 with *phosphoric acid R*, add 130 mL of *methanol R* and 70 mL of *tetrahydrofuran R* and mix.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 3 times the retention time of flumazenil.

Relative retention With reference to flumazenil (retention time = about 14 min): impurity A = about 0.4; impurity D = about 0.5; impurity E = about 0.6; impurity B = about 0.7; impurity F = about 2.4.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurity B and flumazenil.

Limits:
— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

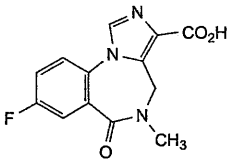
Dissolve 0.250 g in 50 mL of a mixture of 2 volumes of *acetic anhydride R* and 3 volumes of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 30.33 mg of $C_{15}H_{14}FN_3O_3$.

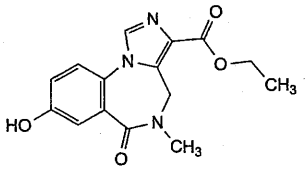
IMPURITIES

Specified impurities *B, C.*

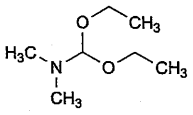
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, F.*



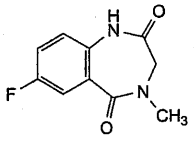
A. 8-fluoro-5-methyl-6-oxo-5,6-dihydro-4*H*-imidazo[1,5-*a*][1,4]benzodiazepine-3-carboxylic acid,



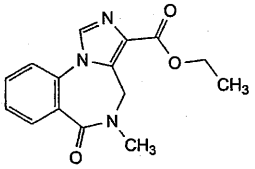
B. ethyl 8-hydroxy-5-methyl-6-oxo-5,6-dihydro-4*H*-imidazo[1,5-*a*][1,4]benzodiazepine-3-carboxylate,



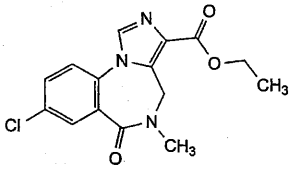
C. diethoxy-*N*,*N*-dimethylmethanamine,



D. 7-fluoro-4-methyl-3,4-dihydro-1*H*-1,4-benzodiazepine-2,5-dione,



E. ethyl 5-methyl-6-oxo-5,6-dihydro-4*H*-imidazo[1,5-*a*][1,4]benzodiazepine-3-carboxylate,

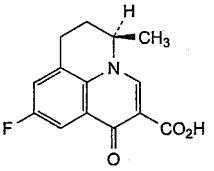


F. ethyl 8-chloro-5-methyl-6-oxo-5,6-dihydro-4*H*-imidazo[1,5-*a*][1,4]benzodiazepine-3-carboxylate.

Ph Eur

Flumequine

(Ph. Eur. monograph 1517)



and enantiomer

$C_{14}H_{12}FNO_3$ 261.3 *42835-25-6*

Action and use
Antibacterial.

Ph Eur

DEFINITION

(*RS*)-9-Fluoro-5-methyl-1-oxo-6,7-dihydro-1*H*,5*H*-benzo[*ij*]quinolizine-2-carboxylic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, microcrystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, very slightly soluble in methanol. It is freely soluble in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flumequine CRS.*

B. Optical rotation (see Tests).

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in 10 mL of *methylene chloride R.*

Reference solution Dissolve 5 mg of *flumequine CRS* in 10 mL of *methylene chloride R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *ammonia R, water R, ethanol (96 per cent) R* (10:10:90 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 2 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter and add to the filtrate a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution changes from red to yellow and the blank remains red.

TESTS

Solution S
Dissolve 5.00 g in *0.5 M sodium hydroxide* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 35.0 mg of the substance to be examined in *dimethylformamide R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of *flumequine impurity B CRS* in 2.0 mL of a 50 µg/mL solution of *flumequine CRS* in *dimethylformamide R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with *dimethylformamide R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R*, 1.36 g/L solution of *potassium dihydrogen phosphate R* (49:51 *V/V*).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 313 nm.

Injection 10 µL; inject *dimethylformamide R* as a blank.

Run time 3 times the retention time of flumequine.

Relative retention With reference to flumequine (retention time = about 13 min): impurity A = about 0.67; impurity B = about 0.85.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity B and flumequine.

Limits:
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

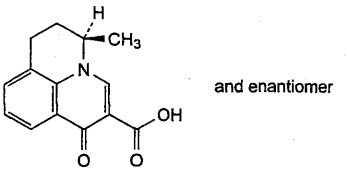
ASSAY

Dissolve 0.500 g in 50 mL of *dimethylformamide R*. Titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 26.13 mg of $C_{14}H_{12}FNO_3$.

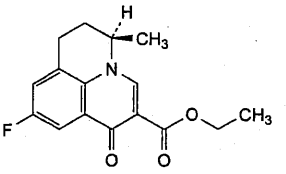
IMPURITIES

Specified impurities *A, B.*



and enantiomer

A. (*RS*)-5-methyl-1-oxo-6,7-dihydro-1*H*,5*H*-benzo[*i,j*]quinolizine-2-carboxylic acid (defluoroflumequine),



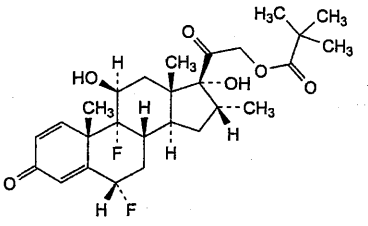
and enantiomer

B. ethyl (*RS*)-9-fluoro-5-methyl-1-oxo-6,7-dihydro-1*H*,5*H*-benzo[*i,j*]quinolizine-2-carboxylate (flumequine ethyl ester).

Ph Eur

Flumetasone Pivalate

(Ph. Eur. monograph 1327)



$C_{27}H_{36}F_2O_6$ 494.6 *2002-29-1*

Action and use
Glucocorticoid.

Preparation
Flumetasone and Clioquinol Ear Drops

Ph Eur

DEFINITION

6α,9-Difluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in acetone, slightly soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flumetasone pivalate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *flumetasone pivalate CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *desoxycortone acetate CRS* in *acetone R* and dilute to 10 mL with the same solvent. Dilute 5 mL of this solution to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R.* Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

C. Add about 2 mg to 2 mL of a mixture of 0.5 mL of *water R* and 1.5 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, a pink colour develops. Add this solution to 10 mL of *water R* and mix. The colour fades and a clear solution remains.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R* add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Solution S

Dissolve 0.50 g in *acetone R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)

+ 69 to + 77 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 10 mg of *dexamethasone pivalate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. To 5.0 mL of this solution, add 5.0 mL of the test solution, mix and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *tetrahydrofuran R, acetonitrile R, water R, methanol R* (5:30:30:35 *V/V/V/V*).

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 1.5 times the retention time of flumetasone pivalate.

Relative retention With reference to flumetasone pivalate: impurity C = about 1.1.

System suitability Reference solution (a):

— *resolution*: minimum 2.8 between the peaks due to flumetasone pivalate and impurity C; if necessary, adjust the concentration of tetrahydrofuran in the mobile phase.

Limits:

— *impurities A, B, C, D*: for each impurity, not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent);

— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C for 4 h.

ASSAY

Dissolve 50.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 239 nm.

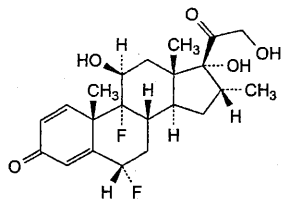
Calculate the content of $C_{27}H_{36}F_2O_6$ taking the specific absorbance to be 336.

STORAGE

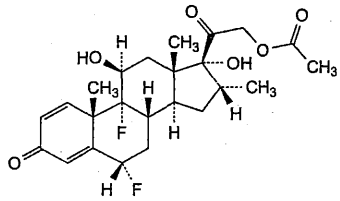
Protected from light.

IMPURITIES

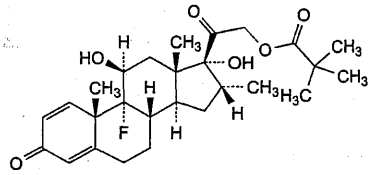
Specified impurities *A, B, C, D.*



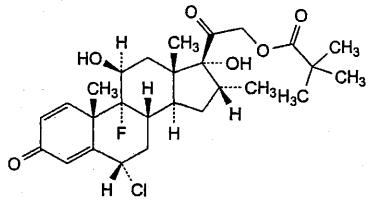
A. 6α,9-difluoro-11β,17,21-trihydroxy-16α-methylpregna-1,4-diene-3,20-dione (flumetasone),



B. 6α,9-difluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (flumetasone acetate),



C. 9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate (dexamethasone pivalate),

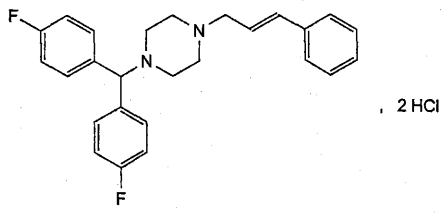


D. 6α-chloro-9-fluoro-11β,17-dihydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate (chlordexamethasone pivalate).

Ph Eur

Flunarizine Dihydrochloride

(Ph. Eur. monograph 1722)



$C_{26}H_{28}Cl_2F_2N_2$ 477.4 *30484-77-6*

Action and use
Calcium channel blocker.

Ph Eur

DEFINITION

1-[Bis(4-fluorophenyl)methyl]-4-[(2*E*)-3-phenylprop-2-enyl] piperazine dihydrochloride.

Content
99.0 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder, hygroscopic.

Solubility
Slightly soluble in water, sparingly soluble in methanol, slightly soluble in alcohol and in methylene chloride.

mp
About 208 °C, with decomposition.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of flunarizine dihydrochloride.*

B. Dissolve 25 mg in 2 mL of *methanol R* and add 0.5 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *flunarizine dihydrochloride for system suitability CRS* in *methanol R* and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 5.0 mL of this solution to 20.0 mL with *methanol R*.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: solution containing 23.8 g/L of *tetrabutylammonium hydrogen sulfate R* and 7 g/L of *ammonium acetate R*,
— *mobile phase B*: *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 12	80 → 40	20 → 60
12 - 15	40	60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to flunarizine,
— the chromatogram obtained is concordant with the chromatogram supplied with *flunarizine dihydrochloride for system suitability CRS*.

Limits:
- — *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.5,
- — *impurities A, D*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
- — *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
- — *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent),
- — *any other impurity*: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
- — *total*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent),
- — *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

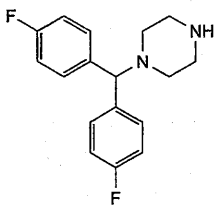
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.200 g in 70 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added at the second point of inflexion. Carry out a blank titration.

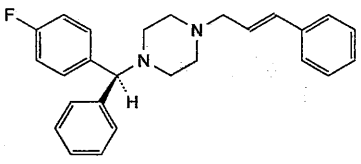
1 mL of *0.1 M sodium hydroxide* is equivalent to 23.87 mg of $C_{26}H_{28}Cl_2F_2N_2$.

STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities *A, B, C, D.*

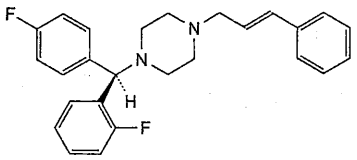


A. 1-[bis(4-fluorophenyl)methyl]piperazine,



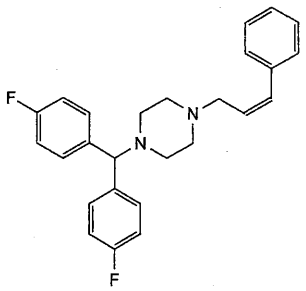
and enantiomer

B. 1-[(*RS*)-(4-fluorophenyl)phenylmethyl]-4-[(2*E*)-3-phenylprop-2-enyl]piperazine,



and enantiomer

C. 1-[(*RS*)-(2-fluorophenyl)(4-fluorophenyl)methyl]-4-[(2*E*)-3-phenylprop-2-enyl]piperazine,

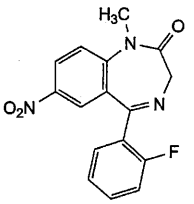


D. 1-[bis(4-fluorophenyl)methyl]-4-[(2*Z*)-3-phenylprop-2-enyl]piperazine.

Ph Eur

Flunitrazepam

(Ph. Eur. monograph 0717)



$C_{16}H_{12}FN_3O_3$ 313.3 *1622-62-4*

Action and use
Benzodiazepine.

Ph Eur

DEFINITION
5-(2-Fluorophenyl)-1-methyl-7-nitro-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish, crystalline powder.

Solubility
Practically insoluble in water, soluble in acetone, slightly soluble in alcohol.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *Ph. Eur. reference spectrum of flunitrazepam.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 100.0 mg of the substance to be examined in 10 mL of *acetonitrile R* and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 4 mg of the substance to be examined and 4 mg of *nitrazepam R* in 5 mL of *acetonitrile R* and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, acetonitrile R, water R* (50:305:645 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 6 times the retention time of flunitrazepam.

Relative retention With reference to flunitrazepam (retention time = about 11 min): impurity A = about 0.2; impurity B = about 0.6; impurity C = about 2.3; impurity D = about 4.0.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to nitrazepam and flunitrazepam.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 2.44,
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

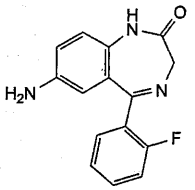
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 20 mL of *anhydrous acetic acid R* and add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

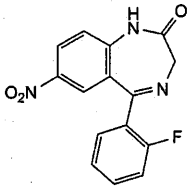
1 mL of *0.1 M perchloric acid* is equivalent to 31.33 mg of $C_{16}H_{12}FN_3O_3$.

STORAGE
Protected from light.

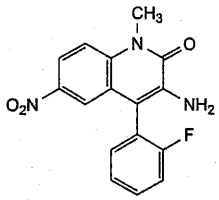
IMPURITIES



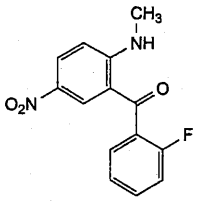
A. 7-amino-5-(2-fluorophenyl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one (7-aminodemethylflunitrazepam),



B. 5-(2-fluorophenyl)-7-nitro-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one (demethylflunitrazepam),



C. 3-amino-4-(2-fluorophenyl)-1-methyl-6-nitroquinolin-2(1*H*)-one,

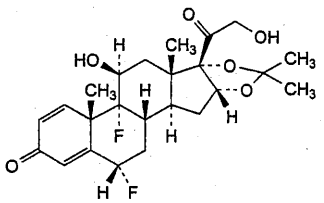


D. (2-fluorophenyl)[2-(methylamino)-5-nitrophenyl]methanone.

Ph Eur

Fluocinolone Acetonide

(Ph. Eur. monograph 0494)



$C_{24}H_{30}F_2O_6$ 452.5 *67-73-2*

Action and use
Glucocorticoid.

Preparations
Fluocinolone Cream
Fluocinolone Ointment

Ph Eur

DEFINITION

6α,9-Difluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone and in anhydrous ethanol, practically insoluble in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluocinolone acetonide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (d).

TESTS

Specific optical rotation (*2.2.7*)

+ 100 to + 104 (dried substance).

Dissolve 0.100 g in *anhydrous ethanol R* and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from lightand prepare the solutions immediately before use.*

Solvent mixture *acetonitrile R, water R* (20:80 *V/V*).

Buffer solution Dissolve 5.68 g of *anhydrous disodium hydrogen phosphate R* and 3.63 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 1000.0 mL with the same solvent.

Test solution (a) Dissolve 20.0 mg of the substance to be examined in 20 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of *fluocinolone acetonide for system suitability CRS* (containing impurities H, J and K) in 2 mL of *acetonitrile R* and dilute to 10.0 mL with *water R*.

Reference solution (b) Dissolve 2 mg of *fluocinolone acetonide for peak identification CRS* (containing impurities D and I) in 2 mL of *acetonitrile R* and dilute to 10.0 mL with *water R*.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 20.0 mg of *fluocinolone acetonide CRS* in 20 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: *acetonitrile R*, buffer solution (28:72 *V/V*);
- *mobile phase B*: *water R*, *acetonitrile R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 42	100 → 0	0 → 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 238 nm.

Injection 40 µL of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with *fluocinolone acetonide for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities H, J and K; use the chromatogram supplied with *fluocinolone acetonide for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities D and I.

Relative retention With reference to fluocinolone acetonide (retention time = about 18 min): impurity D = about 0.8; impurity H = about 0.90; impurity I = about 0.94; impurity J = about 1.05; impurity K = about 1.2.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 5.0, where H_p = height above the baseline of the peak due to impurity J and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to fluocinolone acetonide.

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity K by 1.3;
- for each impurity, use the concentration of fluocinolone acetonide in reference solution (c).

Limits:
- *impurity K*: maximum 0.3 per cent;
- *impurities D, J*: for each impurity, maximum 0.2 per cent;
- *impurities H, I*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.7 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 20 µL of test solution (b) and reference solution (d).

Calculate the percentage content of $C_{24}H_{30}F_2O_6$ taking into account the assigned content of *fluocinolone acetonide CRS*.

STORAGE

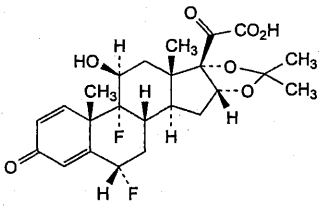
Protected from light.

IMPURITIES

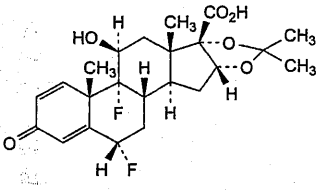
Specified impurities *D, H, I, J, K.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance

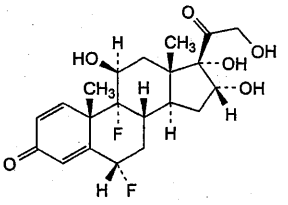
criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E, F, G, L, M.*



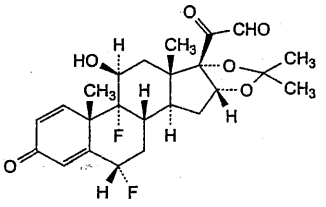
A. 6α,9-difluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregna-1,4-dien-21-oic acid,



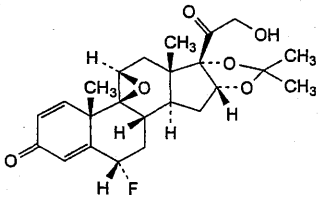
B. 6α,9-difluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3-oxoandrosta-1,4-diene-17β-carboxylic acid,



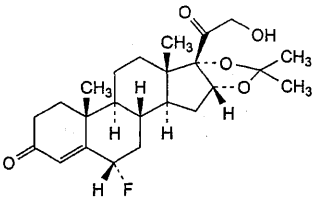
C. 6α,9-difluoro-11β,16α,17,21-tetrahydroxypregna-1,4-diene-3,20-dione (fluocinolone),



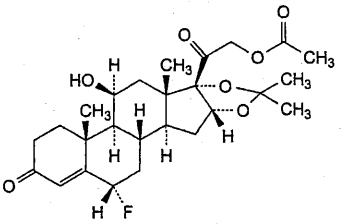
D. 6α,9-difluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregna-1,4-dien-21-al,



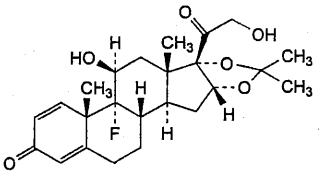
E. 9,11β-epoxy-6α-fluoro-21-hydroxy-16α,17-(1-methylethylidenedioxy)-9β-pregna-1,4-diene-3,20-dione,



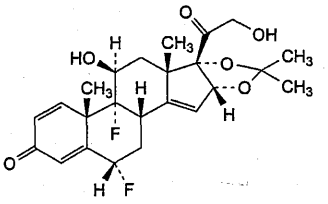
F. 6α-fluoro-21-hydroxy-16α,17-(1-methylethylidenedioxy) pregn-4-ene-3,20-dione,



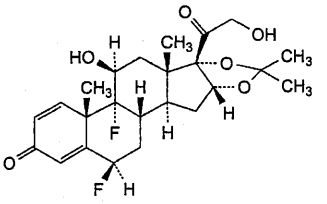
G. 6α-fluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregn-4-en-21-yl acetate,



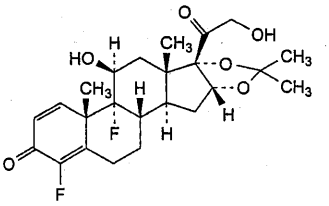
H. 9-fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione,



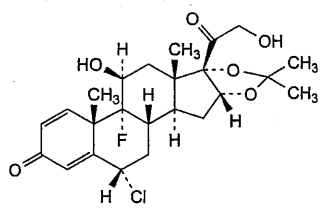
I. 6α,9-difluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4,14-triene-3,20-dione,



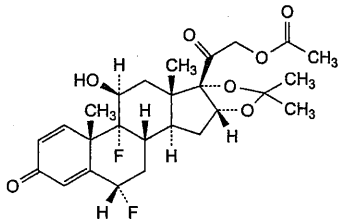
J. 6β,9-difluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione,



K. 4,9-difluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione,



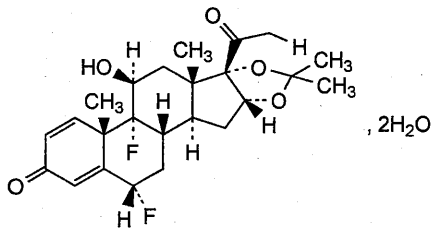
L. 6α-chloro-9-fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione,



M. 6α,9-difluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregna-1,4-dien-21-yl acetate.

Ph Eur

Fluocinolone Acetonide Dihydrate



$C_{24}H_{30}F_2O_6,2H_2O$ 488.5 (anhydrous) *67-73-2*

Action and use
Glucocorticoid.

Preparations
Fluocinolone Cream
Fluocinolone Ointment

DEFINITION
Fluocinolone Acetonide Dihydrate is 6α,9α-difluoro-11β,21-dihydroxy-16α-,17α-isopropylidenedioxypregna-1,4-diene-3,20-dione dihydrate. It contains not less than 96.0% and not more than 104.0% of $C_{24}H_{30}F_2O_6$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Practically insoluble in *water*; freely soluble in *acetone*; soluble in *absolute ethanol*; sparingly soluble in *dichloromethane* and in *methanol*; practically insoluble in *hexane*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of fluocinolone acetonide dihydrate *(RS 147)*.

B. Complies with the test for *identification of steroids*, Appendix III A, using *impregnating solvent I* and *mobile phase H*. Apply 5 µL of each of the three solutions.

C. Complies with the test for *identification of steroids*, Appendix III A, using the conditions specified in test B but using solutions prepared in the following manner.
For solution (1) dissolve 10 mg in 1.5 mL of *glacial acetic acid* in a separating funnel, add 0.5 mL of a 2% w/v solution of *chromium(VI) oxide* and allow to stand for 30 minutes. Add 5 mL of *water* and 2 mL of *dichloromethane* and shake vigorously for 2 minutes. Allow to separate and use the lower layer. Prepare solution (2) in the same manner but using 10 mg of *fluocinolone acetonide BPCRS*.

TESTS
Light absorption
Dissolve 15 mg in sufficient *absolute ethanol* to produce 100 mL. Dilute 10 mL of the solution to 100 mL with *absolute ethanol*. The A(1%, 1 cm) of the resulting solution at the maximum at 239 nm is 345 to 375, calculated with reference to the anhydrous substance, Appendix II B.

Specific optical rotation
In a 1% w/v solution in *1,4-dioxan*, +92 to +96, calculated with reference to the anhydrous substance, Appendix V F.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions.

(1) 0.25% w/v of the substance being examined in *acetonitrile*.

(2) 0.025% w/v each of *fluocinolone acetonide BPCRS* and *triamcinolone acetonide BPCRS* in 45% w/v of *acetonitrile*.

(3) Dilute 1 volume of solution (1) to 100 volumes with *acetonitrile*.

(4) Dilute 1 volume of solution (3) to 20 volumes with *acetonitrile*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with *base-deactivated end-capped octadecylsilyl silica gel for chromatography* (5 µm) (Hypersil BDS is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 238 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for 4 times the retention time of the principal peak.

MOBILE PHASE

45 volumes of *acetonitrile* and 55 volumes of *water*.

SYSTEM SUITABILITY

The test is not valid unless:

in the chromatogram obtained with solution (2), the *resolution factor* between the peaks due to triamcinolone acetonide and fluocinolone acetonide is at least 3.0;

in the chromatogram obtained with solution (4), the *signal-to-noise ratio* of the principal peak is at least 10.

LIMITS

In the chromatogram obtained with solution (1):

the area of any *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (1%);

the area of not more than one *secondary peak* is greater than 0.5 times the area of the principal peak in the chromatogram obtained with solution (3) (0.5%);

the sum of the areas of any *secondary peaks* is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with solution (3) (2.5%).

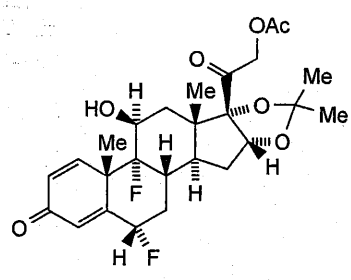
Disregard any peak with an area less than the area of the principal peak in the chromatogram obtained with solution (4) (0.05%).

Water
7.0 to 8.5% w/w, Appendix IX C. Use 0.5 g.

ASSAY
Carry out the *tetrazolium assay of steroids*, Appendix VIII J, and calculate the content of $C_{24}H_{30}F_2O_6$ from the *absorbance* obtained by repeating the operation using *fluocinolone acetonide BPCRS* in place of the substance being examined.

STORAGE
Fluocinolone Acetonide Dihydrate should be protected from light.

Fluocinonide



$C_{26}H_{32}F_2O_7$ 494.5 *356-12-7*

Action and use
Glucocorticoid.

Preparations
Fluocinonide Cream
Fluocinonide Ointment

DEFINITION
Fluocinonide is 6α,9α-difluoro-11β-hydroxy-16α,17α-isopropylidenedioxy-3,20-dioxopregna-1,4-dien-21-yl acetate. It contains not less than 97.0% and not more than 103.0% of $C_{26}H_{32}F_2O_7$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white, crystalline powder.

Practically insoluble in *water*; slightly soluble in *absolute ethanol*. It melts at about 220°, with decomposition.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of fluocinonide *(RS 148)*.

B. Carry out the method for *thin-layer chromatography*, Appendix III A, using a silica gel F_{254} precoated plate (Merck silica gel 60 F_{254} plates are suitable) and a mixture of 12 volumes of *water*, 80 volumes of *methanol*, 150 volumes of *ether* and 770 volumes of *dichloromethane* as the mobile phase; mix the water and the methanol before adding to the remaining components of the mobile phase. Apply separately to the plate 5 µL of each of the following solutions.
For solution (1) dissolve 25 mg of the substance being examined in 5 mL of *methanol* (solution A); dilute 2 mL of solution A to 10 mL with *chloroform*. For solution (2) transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper, add 10 mL of *saturated methanolic potassium hydrogen carbonate solution* and immediately pass a current of *nitrogen* briskly through the solution for 2 minutes. Stopper the tube, heat in a water bath at 45° protected from light for 2.5 hours and allow to cool. For solution (3) dissolve 25 mg of *fluocinonide BPCRS* in 5 mL of *methanol* (solution B); dilute 2 mL of solution B to 10 mL with *chloroform*. Prepare solution (4) in the same manner as solution (2) but use 2 mL of solution B in place of 2 mL of solution A. After removal of the plate, allow it to dry in air and examine under *ultraviolet light (254 nm)*. The principal spots in each of the chromatograms obtained with solutions (1) and (2) are similar in position and size to those in the chromatograms obtained with solutions (3) and (4), respectively.

TESTS
Specific optical rotation
In a 1.0% w/v solution in *chloroform*, +81 to +89, calculated with reference to the dried substance, Appendix V F.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using the precoated plate and the mobile phase specified in test B for Identification. Apply separately to the plate 5 µL of each of four solutions in *chloroform* containing (1) 0.50% w/v of the substance being examined, (2) 0.010% w/v of the substance being examined, (3) 0.0050% w/v of the substance being examined and (4) 0.010% w/v of *fluocinolone acetonide BPCRS*. After removal of the plate, allow it to dry in air, heat the plate at 105° for 10 minutes, cool and spray with *alkaline tetrazolium blue solution*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (2%) and not more than one such spot is more intense than the spot in the chromatogram obtained with solution (3) (1%). The test is not valid unless the principal spot in the chromatogram obtained with solution (2) has an R_f value relative to the spot in the chromatogram obtained with solution (4) of at least 1.5.

Loss on drying
When dried at 100° to 105° for 3 hours, loses not more than 1.0% of its weight. Use 1 g.

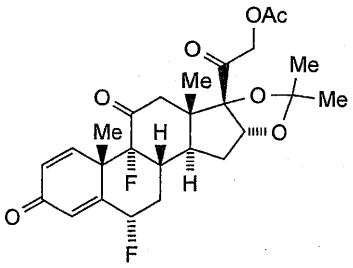
ASSAY
Carry out the method for *liquid chromatography*, Appendix III D, using two solutions in *methanol* containing (1) 0.012% w/v of the substance being examined and (2) 0.012% w/v of *fluocinonide BPCRS*.

The chromatographic procedure may be carried out using (a) a stainless steel column (20 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Spherisorb ODS 1 is suitable), (b) as the mobile phase with a flow rate of 1 mL per minute a mixture of 1 volume of *glacial acetic acid*, 450 volumes of *acetonitrile* and 550 volumes of *water* and (c) a detection wavelength of 238 nm.

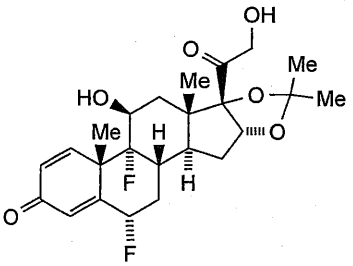
Calculate the content of $C_{26}H_{32}F_2O_7$ using the declared content of $C_{26}H_{32}F_2O_7$ in *fluocinonide BPCRS*.

STORAGE
Fluocinonide should be protected from light.

IMPURITIES

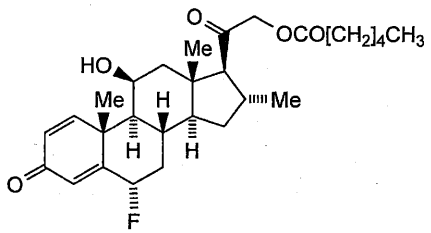


A. 6α,9α-difluoro-16α,17α-isopropylidene-3,11,20-trioxopregna-1,4-dien-21-yl acetate,



B. 6α,9α-difluoro-11β,21-dihydroxy-16α,17α-isopropylidenepregna-1,4-diene-3,20dione.

Fluocortolone Hexanoate



$C_{28}H_{39}FO_5$ 474.6 *303-40-2*

Action and use
Glucocorticoid.

Preparation
Fluocortolone Cream

DEFINITION
Fluocortolone Hexanoate is 6α-fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl hexanoate.
It contains not less than 97.0% and not more than 103.0% of $C_{28}H_{39}FO_5$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or creamy white, crystalline powder.

Practically insoluble in *water* and in *ether*; slightly soluble in *acetone* and in *1,4-dioxan*; very slightly soluble in *ethanol (96%)* and in *methanol*.

It exhibits polymorphism.

IDENTIFICATION
The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of fluocortolone hexanoate *(RS 149)*.

TESTS

Light absorption
Ratio of the *absorbance* of the solution prepared as directed in the Assay at the maximum at 242 nm to that at 263 nm, 2.15 to 2.35, Appendix II B.

Specific optical rotation
In a 1% w/v solution in *1,4-dioxan*, prepared with the aid of heat, +97 to +103, calculated with reference to the dried substance, Appendix V F.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions. Solution (1) contains 0.04% w/v of the substance being examined in a mixture of 1 volume of *water* and 9 volumes of *acetonitrile*. For solution (2) dilute 1 volume of solution (1) to 100 volumes with a mixture of 1 volume of *water* and 9 volumes of *acetonitrile*. Solution (3) contains 0.002% w/v each of *fluocortolone pivalate BPCRS* and *fluocortolone hexanoate BPCRS* in a mixture of 1 volume of *water* and 9 volumes of *acetonitrile*.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Spherisorb ODS 2 is suitable), (b) as the mobile phase with a flow rate of 1.5 mL per minute a mixture of 25 volumes of *methanol*, 32 volumes of *water* and 50 volumes of *acetonitrile* and (c) a detection wavelength of 242 nm.

Inject solution (2). Adjust the sensitivity of the system so that the height of the principal peak in the chromatogram obtained is at least 50% of the full scale of the recorder.

Inject solution (3). The test is not valid unless, in the chromatogram obtained, the *resolution factor* between the two principal peaks is at least 6.0.

Inject solution (1). Continue the chromatography for twice the retention time of fluocortolone hexanoate. In the chromatogram obtained with solution (1), the area of any *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%) and the sum of the areas of any *secondary peaks* is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (2%). Disregard any peak due to the solvent and any peak with an area less than 0.025 times that of the principal peak in the chromatogram obtained with solution (2) (0.025%).

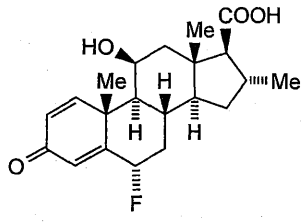
Loss on drying
When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

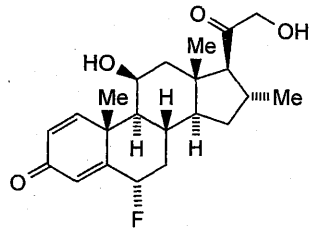
ASSAY
Dissolve 15 mg in sufficient *methanol* to produce 100 mL, dilute 20 mL to 100 mL with *methanol* and measure the *absorbance* of the resulting solution at the maximum at 242 nm, Appendix II B. Calculate the content of $C_{28}H_{39}FO_5$ taking 340 as the value of A(1%, 1 cm) at the maximum at 242 nm.

STORAGE
Fluocortolone Hexanoate should be protected from light.

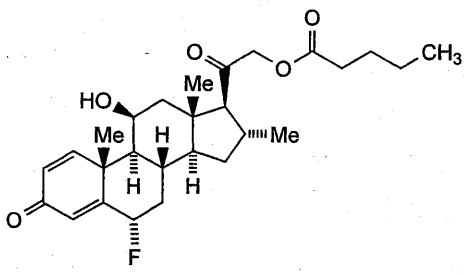
IMPURITIES



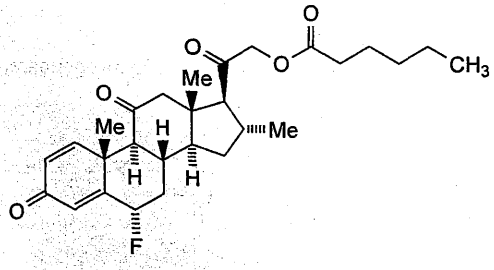
A. 6α-fluoro-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-diene-17β-carboxylic acid,



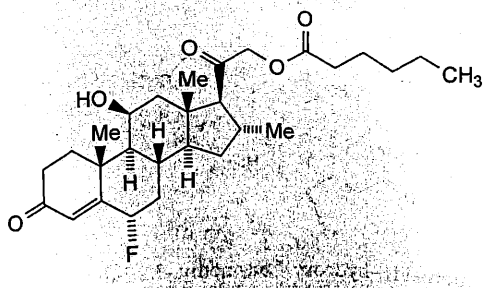
B. 6α-fluoro-11β,21-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione,



C. 6α-fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl pentanoate,



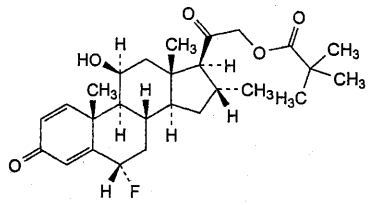
D. 6α-fluoro-16α-methyl-3,11,20-trioxopregna-1,4-dien-21-yl hexanoate,



E. 6α-fluoro-11β-hydroxy-16α-methyl-3,11-dioxopregnen-21-yl hexanoate.

Fluocortolone Pivalate

(Ph. Eur. monograph 1212)



$C_{27}H_{37}FO_5$ 460.6 *29205-06-9*

Action and use
Glucocorticoid.

Preparation
Fluocortolone Cream

Ph Eur

DEFINITION

6α-Fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride and in dioxan, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluocortolone pivalate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *fluocortolone pivalate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *norethisterone CRS* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

C. To about 1 mg add 2 mL of a mixture of 2 volumes of *glacial acetic acid R* and 3 volumes of *sulfuric acid R* and heat for 1 min on a water-bath. A red colour is produced. Add 5 mL of *water R*, the colour changes to violet-red.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter and add to the filtrate a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Specific optical rotation (*2.2.7*)
+ 100 to + 105 (dried substance).

Dissolve 0.25 g in *dioxan R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *acetonitrile R*.

Reference solution (b) Dissolve 2 mg of *fluocortolone pivalate CRS* and 2 mg of *prednisolone hexanoate CRS* in *acetonitrile R*, then dilute to 100 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, acetonitrile R, water R* (25:30:32 *V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 243 nm.

Injection 20 µL.

Run time Twice the retention time of fluocortolone pivalate.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to fluocortolone pivalate and prednisolone hexanoate.

Limits:
— *impurities A, B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (2 per cent);
— *disregard limit*: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.025 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 30.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *anhydrous ethanol R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 242 nm.

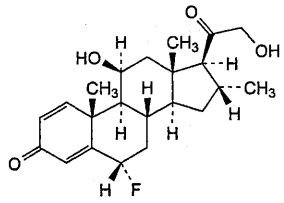
Calculate the content of $C_{27}H_{37}FO_5$ taking the specific absorbance to be 350.

STORAGE

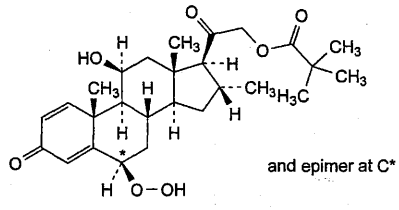
Protected from light.

IMPURITIES

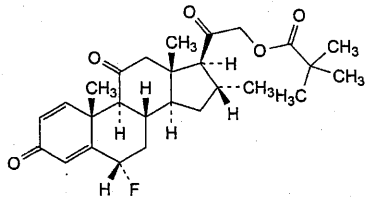
Specified impurities *A, B, C, D.*



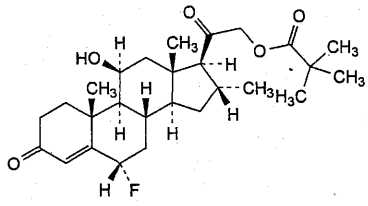
A. 6α-fluoro-11β,21-dihydroxy-16α-methylpregna-1,4-diene-3,20-dione (fluocortolone),



B. 6-hydroperoxy-11β-hydroxy-16α-methyl-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate,



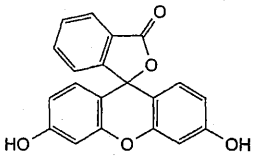
C. 6α-fluoro-16α-methyl-3,11,20-trioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate,



D. 6α-fluoro-11β-hydroxy-16α-methyl-3,20-dioxopregna-4-en-21-yl 2,2-dimethylpropanoate.

Fluorescein

(Ph. Eur. monograph 2348)



$C_{20}H_{12}O_5$ 332.3 *2321-07-5*

Action and use
Detection of corneal lesions, retinal angiography and pancreatic function testing.

Ph Eur

DEFINITION
3′,6′-Dihydroxy-3*H*-spiro[isobenzofuran-1,9′-xanthen]-3-one.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Orange-red, fine powder.

Solubility
Practically insoluble in water, soluble in hot ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluorescein CRS.*

Dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethanol (96 per cent) R*, evaporate to dryness and record the spectra using the residues.

B. Dilute 0.1 mL of solution S (see Tests) to 10 mL with *water R*. The solution shows a yellowish-green fluorescence. The fluorescence disappears on addition of 0.1 mL of *dilute hydrochloric acid R* and reappears on addition of 0.2 mL of *dilute sodium hydroxide solution R*.

C. The absorption by a piece of filter paper of 0.05 mL of the solution prepared for identification B (before the addition of *dilute hydrochloric acid R*) colours the paper yellow. On exposing the moist paper to bromine vapour for 1 min and then to ammonia vapour, the colour becomes deep pink.

D. Suspend 0.5 g in 50 mL of *water R* and shake for 10 min. The substance does not completely dissolve.

TESTS
Solution S
Suspend 1.0 g in 35.0 mL of *water R* and add dropwise with shaking 4.5 mL of *1 M sodium hydroxide*. Adjust to pH 8.5-9.0 with *1 M sodium hydroxide* and dilute to 50.0 mL with *water R* to obtain a clear solution.

Appearance of solution
Solution S is clear (*2.2.1*) and orange-yellow with yellowish-green fluorescence.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile for chromatography R*, mobile phase A (30:70 *V/V*).

Test solution (a) Disperse 50.0 mg of the substance to be examined in 15.0 mL of *ethanol (96 per cent) R*. Sonicate and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to 250.0 mL with the solvent mixture.

Reference solution (a) Disperse 50.0 mg of *fluorescein CRS* in 15.0 mL of *ethanol (96 per cent) R*. Sonicate and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 250.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10.0 mg of *phthalic acid CRS* (impurity B) and 10.0 mg of *resorcinol CRS* (impurity A) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 5.0 mL of test solution (b) to 20.0 mL with the solvent mixture.

Reference solution (d) Dilute 10.0 mL of reference solution (c) to 100.0 mL with the solvent mixture.

Reference solution (e) Dissolve the contents of a vial of *fluorescein impurity C CRS* in 1 mL of the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R3* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.610 g of *potassium dihydrogen phosphate R* in *water for chromatography R*, adjust to pH 2.0 with *phosphoric acid R* and dilute to 1000.0 mL with *water for chromatography R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	85 → 20	15 → 80
20 - 29	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (c), (d) and (e).

Identification of impurity C Use the chromatogram obtained with reference solution (e) to identify the peak due to impurity C.

Relative retention With reference to fluorescein (retention time = about 15 min): impurity A = about 0.42; impurity B = about 0.48; impurity C = about 0.86.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurities A and B.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.9;
— *impurity C*: not more than 1.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.6 per cent);
— *impurities A, B*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the

chromatogram obtained with reference solution (c) (0.10 per cent);
— *sum of impurities other than A, B and C*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 0.25 per cent.

To 10.0 mL of solution S add 90.0 mL of *water R* and 3.0 mL of *dilute nitric acid R*, wait for at least 30 min and filter. Dilute 10.0 mL of the filtrate to 15.0 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

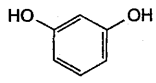
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

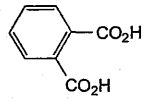
Calculate the percentage content of $C_{20}H_{12}O_5$ taking into account the assigned content of *fluorescein CRS*.

STORAGE
Protected from light.

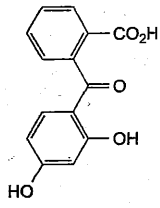
IMPURITIES
Specified impurities *A, B, C.*



A. benzene-1,3-diol (resorcinol),



B. benzene-1,2-dicarboxylic acid (phthalic acid),



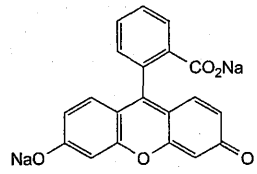
C. 2-(2,4-dihydroxybenzoyl)benzoic acid.

Ph Eur

Fluorescein Sodium

Soluble Fluorescein

(Ph. Eur. monograph 1213)



$C_{20}H_{10}Na_2O_5$ 376.3 *518-47-8*

Action and use
Detection of corneal lesions, retinal angiography and pancreatic function testing.

Preparations
Fluorescein Eye Drops
Fluorescein Injection

Ph Eur

DEFINITION
Disodium 2-(6-oxido-3-oxo-3*H*-xanthen-9-yl)benzoate.

Content
95.0 per cent to 103.0 per cent (dried substance).

CHARACTERS
Appearance
Orange-red, fine powder, hygroscopic.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent), practically insoluble in hexane and in methylene chloride.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Dilute 0.1 mL of solution S (see Tests) to 10 mL with *water R*. The solution shows yellowish-green fluorescence. The fluorescence disappears on addition of 0.1 mL of *dilute hydrochloric acid R* and reappears on addition of 0.2 mL of *dilute sodium hydroxide solution R*.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *Ph. Eur. reference spectrum of fluorescein sodium.*

C. The absorption by a piece of filter paper of 0.05 mL of the solution prepared for identification A (before the addition of *dilute hydrochloric acid R*) colours the paper yellow. On exposing the moist paper to bromine vapour for 1 min and then to ammonia vapour, the colour becomes deep pink.

D. Ignite 0.1 g in a porcelain crucible. Dissolve the residue in 5 mL of *water R* and filter. 2 mL of the filtrate gives reaction (a) of sodium (*2.3.1*).

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and orange-yellow with yellowish-green fluorescence.

pH (*2.2.3*)
7.0 to 9.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.100 g of the substance to be examined in a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A and dilute to 100.0 mL with the same mixture of solvents.

Test solution (b) Dilute 5.0 mL of test solution (a) to 250.0 mL with a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A.

Reference solution (a) Dissolve 55.0 mg of *diacetylfluorescein CRS* in a mixture of 1 mL of *2.5 M sodium hydroxide* and 5 mL of *ethanol (96 per cent) R*, heat on a water-bath for 20 min mixing frequently, cool and dilute to 50.0 mL with *water R*. Dilute 5.0 mL of the solution to 250.0 mL with a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A.

Reference solution (b) Dissolve 10.0 mg of *phthalic acid R* (impurity B) and 10.0 mg of *resorcinol R* (impurity A) in a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A and dilute to 100.0 mL with the same mixture of solvents. Dilute 5.0 mL of the solution to 100.0 mL with a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A.

Reference solution (c) Dilute 5.0 mL of test solution (b) to 20.0 mL with a mixture of 30 volumes of *acetonitrile R* and 70 volumes of mobile phase A.

Column:
- *size*: l = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: dissolve 0.610 g of *potassium dihydrogen phosphate R* in *water R* and dilute to 1000 mL with the same solvent; adjust to pH 2.0 with *phosphoric acid R*;
- *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	85 → 20	15 → 80
20 - 29	20	80
29 - 30	20 → 85	80 → 15
30 - 35	85	15

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (b) and (c).

Relative retention With reference to fluorescein (retention time = about 15 min): impurity A = about 0.4; impurity B = about 0.5; impurity C = about 0.9.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurity A and impurity B.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.6;
- *impurities A, B*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
- *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
- *sum of impurities other than A, B, C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
- *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Chlorides (*2.4.4*)

Maximum 0.25 per cent.

To 10 mL of solution S add 90 mL of *water R* and 1 mL of *dilute nitric acid R*, wait for at least 10 min and filter. Dilute 10 mL of the filtrate to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 1.0 per cent.

To 5 mL of solution S add 90 mL of *distilled water R*, 2.5 mL of *dilute hydrochloric acid R* and dilute to 100 mL with *distilled water R*. Filter.

Zinc

Dilute 5 mL of solution S to 10 mL with *water R*. Add 2 mL of *hydrochloric acid R1*, filter and add 0.1 mL of *potassium ferrocyanide solution R*. No turbidity or precipitate is formed immediately.

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{20}H_{10}Na_2O_5$ using the chromatogram obtained with reference solution (a) and the declared content of *diacetylfluorescein CRS*.

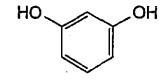
1 mg of *diacetylfluorescein CRS* is equivalent to 0.9037 mg of $C_{20}H_{10}Na_2O_5$.

STORAGE

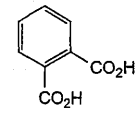
In an airtight container, protected from light.

IMPURITIES

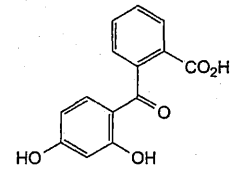
Specified impurities *A, B, C.*



A. benzene-1,3-diol (resorcinol),



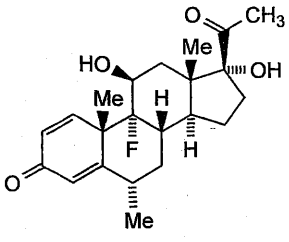
B. benzene-1,2-dicarboxylic acid (phthalic acid),



C. 2-(2,4-dihydroxybenzoyl)benzoic acid.

Ph Eur

Fluorometholone



$C_{22}H_{29}FO_4$ 376.5 *426-13-1*

Action and use
Glucocorticoid.

Preparation
Fluorometholone Eye Drops

DEFINITION
Fluorometholone is 9α-fluoro-11β,17α-dihydroxy-6α-methylpregna-1,4-diene-3,20-dione. It contains not less than 97.0% and not more than 103.0% of $C_{22}H_{29}FO_4$, calculated with reference to the dried substance.

CHARACTERISTICS
A white to yellowish white, crystalline powder. It melts at about 280°, with decomposition.

Practically insoluble in *water*; slightly soluble in *absolute ethanol* and in *ether*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of fluorometholone *(RS 152)*.

B. In the Assay, the principal peak in the chromatogram obtained with solution (1) has the same retention time as the principal peak in the chromatogram obtained with solution (2).

TESTS
Specific optical rotation
In a 1% w/v solution in *pyridine*, +52 to +60, Appendix V F, calculated with reference to the dried substance.

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol*.

(1) 0.010% w/v of the substance being examined.

(2) 0.00005% w/v of the substance being examined.

(3) 0.00005% w/v each of *deltamedrane BPCRS* and *fluorometholone BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (30 cm × 3.9 mm) packed with *octadecylsilyl silica gel for chromatography* (10 µm) (µBondapak C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

40 volumes of *water* and 60 volumes of *methanol*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution factor* between the peaks due to deltamedrane and fluorometholone is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.5%);

the sum of the areas of any *secondary peaks* is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (1%).

Loss on drying
When dried at 60° at a pressure not exceeding 0.7 kPa for 3 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol*.

(1) 0.005% w/v of the substance being examined.

(2) 0.005% w/v of *fluorometholone BPCRS*.

(3) 0.00005% w/v each of *deltamedrane BPCRS* and *fluorometholone BPCRS*.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related substances may be used.

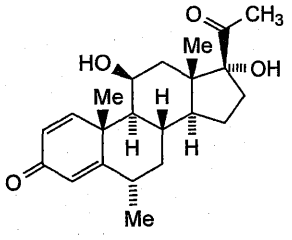
SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution factor* between the peaks due to deltamedrane and fluorometholone is at least 1.5.

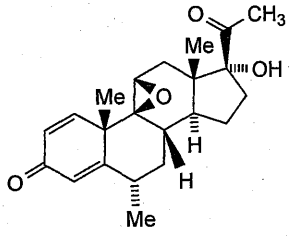
DETERMINATION OF CONTENT

Calculate the content of $C_{22}H_{29}FO_4$ from the chromatograms obtained and using the declared content of $C_{22}H_{29}FO_4$ in *fluorometholone BPCRS*.

IMPURITIES



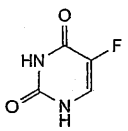
A. 11β,17α-dihydroxy-6α-methylpregna-1,4-diene-3,20-dione *(deltamedrane)*,



B. 9β,11β-epoxy-17α-hydroxy-6α-methylpregna-1,4-diene-3,20-dione *(epoxymedradiene)*.

Fluorouracil

(Ph. Eur. monograph 0611)



$C_4H_3FN_2O_2$ 130.1 *51-21-8*

Action and use
Pyrimidine analogue; cytotoxic.

Preparations
Fluorouracil Cream
Fluorouracil Injection

Ph Eur

DEFINITION

5-Fluoropyrimidine-2,4(1*H*,3*H*)-dione.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *fluorouracil CRS.*

TESTS

Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 or Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)
4.5 to 5.0 for solution S.

Impurities F and G
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.10 g of the substance to be examined in a mixture of equal volumes of *methanol R* and *water R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 5.0 mg of *fluorouracil impurity F CRS* in a mixture of equal volumes of *methanol R* and *water R* and dilute to 200.0 mL with the same mixture of solvents.

Reference solution (b) Dissolve 20.0 mg of *urea R* (impurity G) in *methanol R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *methanol R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *methanol R, water R, ethyl acetate R* (15:15:70 *V/V/V*).

Application 10 µL.

Development Over a path of 2/3 of the plate.

Drying In air.

Detection:
— *impurity F*: examine in ultraviolet light at 254 nm;
— *impurity G*: spray with a mixture of 200 mL of a 10 g/L solution of *dimethylaminobenzaldehyde R* in *anhydrous ethanol R* and 20 mL of *hydrochloric acid R*; dry in an oven at 80 °C for 3-4 min, then examine in daylight (impurity G produces a yellow spot and fluorouracil is not detected by the spray).

System suitability The chromatogram shows 2 clearly separated spots after both detections.

Limits:
— *impurity F*: any spot due to impurity F is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.25 per cent);
— *impurity G*: any spot due to impurity G is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *fluorouracil impurity C CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of *fluorouracil impurity A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 5.0 mg of *fluorouracil impurity B CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (e) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (f) To 1 mL of reference solution (a) add 1 mL of the test solution and dilute to 10 mL with the mobile phase.

Reference solution (g) Dissolve the contents of a vial of *fluorouracil impurity mixture CRS* (containing impurities D and E) in 1.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 6.805 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 5.7 ± 0.1 with a 5 M potassium hydroxide solution prepared from *potassium hydroxide R.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 20 µL.

Run time 3 times the retention time of fluorouracil.

Identification of impurities Use the chromatogram supplied with *fluorouracil impurity mixture CRS* and the chromatogram

obtained with reference solution (g) to identify the peaks due to impurities D and E.

Relative retention With reference to fluorouracil (retention time = about 6 min): impurity A = about 0.5; impurity B = about 0.7; impurity C = about 0.9; impurity D = about 1.6; impurity E = about 1.9.

System suitability Reference solution (f):
— *resolution*: minimum 2 between the peaks due to impurity C and fluorouracil.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 1.5; impurity E = 1.3;
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.1 per cent);
— *impurity C*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *impurities D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (e) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (e) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 4 h.

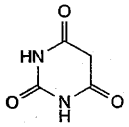
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.100 g in 80 mL of *dimethylformamide R*, warming gently. Cool and titrate with 0.1 M tetrabutylammonium hydroxide, using 0.25 mL of a 10 g/L solution of *thymol blue R* in *dimethylformamide R* as indicator. Carry out a blank titration.

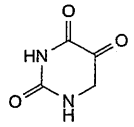
1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 13.01 mg of $C_4H_3FN_2O_2$.

STORAGE
Protected from light.

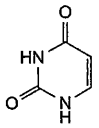
IMPURITIES
Specified impurities *A, B, C, D, E, F, G.*



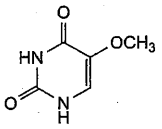
A. pyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione (barbituric acid),



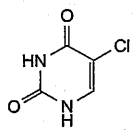
B. dihydropyrimidine-2,4,5(3*H*)-trione (isobarbituric acid or 5-hydroxyuracil),



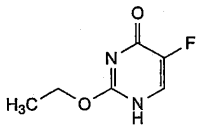
C. pyrimidine-2,4(1*H*,3*H*)-dione (uracil),



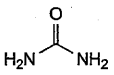
D. 5-methoxypyrimidine-2,4(1*H*,3*H*)-dione (5-methoxyuracil),



E. 5-chloropyrimidine-2,4(1*H*,3*H*)-dione (5-chlorouracil),



F. 2-ethoxy-5-fluoropyrimidin-4(1*H*)-one (2-ethoxy-5-fluorouracil),

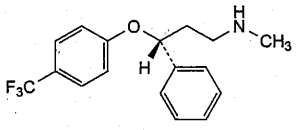


G. carbamide (urea).

Ph Eur

Fluoxetine Hydrochloride

(Ph. Eur. monograph 1104)



and enantiomer , HCl

$C_{17}H_{19}ClF_3NO$ 345.8 *56296-78-7*

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Preparations
Fluoxetine Capsules
Fluoxetine Oral Solution

Ph Eur

DEFINITION

(3*RS*)-*N*-Methyl-3-phenyl-3-[4-(trifluoromethyl) phenoxy]propan-1-amine hydrochloride.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, freely soluble in methanol, sparingly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluoxetine hydrochloride CRS.*

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 2.0 g in a mixture of 15 volumes of *water R* and 85 volumes of *methanol R*, then dilute to 100.0 mL with the same mixture of solvents.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

4.5 to 6.5.

Dissolve 0.20 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Optical rotation (*2.2.7*)

−0.05° to + 0.05°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 55 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Test solution (b) Dilute 2.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution Dissolve 22 mg of *fluoxetine hydrochloride CRS* in 10.0 mL of *0.5 M sulfuric acid.* Heat at about 85 °C for 3 h. Allow to cool. The resulting solution contains considerable quantities of impurity A and usually also contains 4-trifluoromethylphenol. To 0.4 mL of this solution add 28.0 mg of *fluoxetine hydrochloride CRS*, about 1 mg of *fluoxetine impurity B CRS* and about 1 mg of *fluoxetine impurity C CRS*, then dilute to 25.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 8 volumes of *methanol R*, 30 volumes of *tetrahydrofuran R* and 62 volumes of a solution of *triethylamine R* prepared as follows: to 10 mL of *triethylamine R*, add 980 mL of *water R*, mix and adjust to pH 6.0 with *phosphoric acid R* (about 4.5 mL) and dilute to 1000 mL with *water R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

Run time 3 times the retention time of fluoxetine.

Identification of impurities Use the chromatogram obtained with the reference solution to identify the peaks due to impurities A, B and C.

Relative retention With reference to fluoxetine: impurity A = about 0.24; impurity B = about 0.27; impurity C = about 0.9.

System suitability Reference solution:

— *retention time*: fluoxetine = 10 min to 18 min; 4-trifluoromethylphenol: maximum 35 min; if no peak due to 4-trifluoromethylphenol is observed, inject a 0.02 per cent solution of *4-trifluoromethylphenol R* in the mobile phase;

— *peak-to-valley ratio*: minimum 11, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to fluoxetine. If necessary, reduce the volume of methanol and increase the volume of the solution of triethylamine in the mobile phase.

Limit Test solution (b):

— *impurity C*: not more than 0.0015 times the area of the principal peak (0.15 per cent).

Limits Test solution (a):

— *impurities A, B*: for each impurity, not more than 0.0125 times the area of the principal peak in the chromatogram obtained with test solution (b) (0.25 per cent);

— *unspecified impurities*: for each impurity, not more than 0.005 times the area of the principal peak in the chromatogram obtained with test solution (b) (0.10 per cent);

— *total*: not more than 0.025 times the area of the principal peak in the chromatogram obtained with test solution (b) (0.5 per cent);

— *disregard limit*: 0.0025 times the area of the principal peak in the chromatogram obtained with test solution (b) (0.05 per cent).

Acetonitrile

Gas chromatography (*2.2.28*).

Test solution Dissolve 50 mg of the substance to be examined in *dimethylformamide R* and dilute to 5.0 mL with the same solvent.

Reference solution To 1.0 g of *acetonitrile R*, add *dimethylformamide R*, mix and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 1000.0 mL with *dimethylformamide R*.

Column:

— *material*: fused silica;

— *size*: *l* = 30 m, Ø = 0.53 mm;

— *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 10 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	35
	2 - 14.33	35 → 220
	14.33 - 24.33	220
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 1 µL; inject *dimethylformamide R* as a blank.

In the chromatogram obtained with *dimethylformamide R*, verify that there is no peak with the same retention time as acetonitrile.

Limit:

— *acetonitrile*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Test solution Dissolve 55.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution Dissolve 55.0 mg of *fluoxetine hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Detection Spectrophotometer at 227 nm.

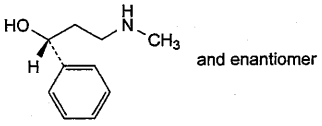
Retention time Fluoxetine = 10 min to 18 min; if necessary, adjust the volumes of methanol and of the solution of triethylamine in the mobile phase.

System suitability Reference solution:

— *symmetry factor*: maximum 2.0 calculated at 10 per cent of the height of the peak due to fluoxetine.

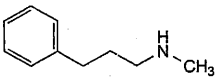
Calculate the content of $C_{17}H_{19}ClF_3NO$ from the declared content of *fluoxetine hydrochloride CRS*.

IMPURITIES
Specified impurities *A, B, C.*

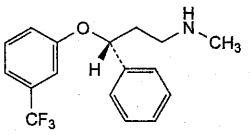


and enantiomer

A. (1*RS*)-3-(methylamino)-1-phenylpropan-1-ol,



B. *N*-methyl-3-phenylpropan-1-amine,

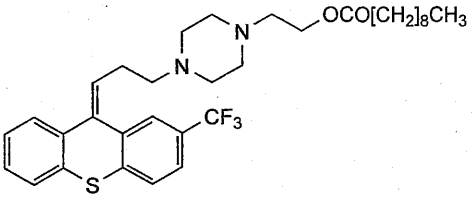


and enantiomer

C. (3*RS*)-*N*-methyl-3-phenyl-3-[3-(trifluoromethyl) phenoxy]propan-1-amine.

Ph Eur

Flupentixol Decanoate



$C_{33}H_{43}F_3N_2O_2S$ 588.82 *30909-51-4*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparation
Flupentixol Injection

DEFINITION
Flupentixol Decanoate is (*Z*)-2-{4-[3-(2-trifluoromethylthioxanthen-9-ylidene)propyl]piperazin-1-yl} ethyl decanoate. It contains not less than 98.0% and not more than 101.0% of $C_{33}H_{43}F_3N_2O_2S$, calculated with reference to the dried substance.

CHARACTERISTICS
A yellow, viscous oil.

Very slightly soluble in *water*; soluble in *ethanol (96%)*; freely soluble in *ether*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 210 to 350 nm of a 0.0015% w/v solution in *ethanol (96%)* exhibits two maxima at 230 nm and 264 nm. The *absorbances* at the maxima are about 0.85 and about 0.35, respectively.

B. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of flupentixol decanoate *(RS 154)*.

TESTS
Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *acetonitrile*, protected from light.

(1) 0.25% w/v of the substance being examined.

(2) 0.000625% w/v of cis-*flupentixol BPCRS*.

(3) 0.000625% w/v of *2-trifluoromethylthioxanthone BPCRS*.

(4) 0.0025% w/v of trans-*flupentixol decanoate dihydrochloride BPCRS*.

(5) 0.25% w/v of the substance being examined and 0.000625% w/v each of cis-*flupentixol BPCRS*, *2-trifluoromethylthioxanthone BPCRS* and 0.0025% w/v of trans-*flupentixol decanoate dihydrochloride BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Waters Symmetry C18 is suitable)

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use a column temperature of 40°.

(e) Use a detection wavelength of 270 nm.

(f) Inject 20 µL of each solution.

(g) Allow the chromatography to proceed for 1.5 times the retention time of the principal peak.

MOBILE PHASE

0.1 volume of *orthophosphoric acid,* 25 volumes of a 20 millimole solution of *dioctyl sodium sulfosuccinate* (prepared by dissolving 8.89 g of *dioctyl sodium sulfosuccinate* in 500 mL of *water,* stirring for 6 to 8 hours and diluting to 1000 mL with *water)* and *75* volumes of *ethanol (96%).*

The substances are eluted in the following order: 2-trifluoromethylthioxanthone, *cis*-flupentixol (free alcohol), flupentixol decanoate and *trans*-flupentixol decanoate.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (5), the peaks due to 2-trifluoromethylthioxanthone, *cis*-flupentixol, flupentixol decanoate and *trans*-flupentixol decanoate are clearly separated.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to *cis*-flupentixol is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.25%);

the area of any peak corresponding to 2-trifluoromethylthioxanthone is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.25%);

the area of any peak corresponding to *trans*-flupentixol decanoate dihydrochloride is not greater than the area of the principal peak in the chromatogram obtained with solution (4) (1%);

Loss on drying

When dried at 60° at a pressure of 0.7 kPa for 3 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

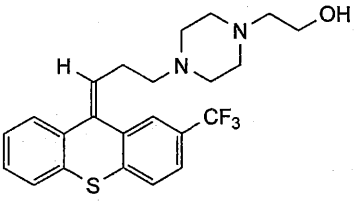
ASSAY

Carry out Method I for *non-aqueous titration,* Appendix VIII A, using 0.2 g in 50 mL of *anhydrous acetic acid* and determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 29.44 mg of $C_{33}H_{43}F_3N_2O_2S$.

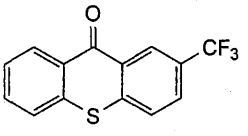
STORAGE

Flupentixol Decanoate should be protected from light and stored at a temperature below -15°.

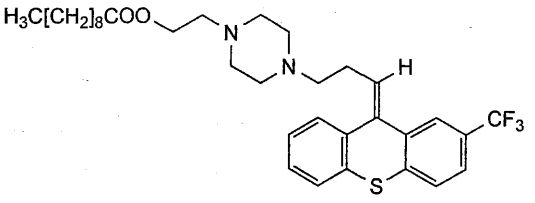
IMPURITIES



A. *cis*-flupentixol (free alcohol),



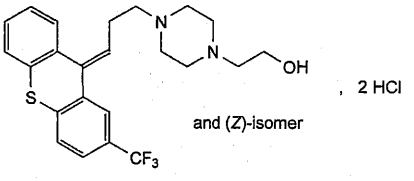
B. 2-trifluoromethylthioxanthone,



C. *trans*-flupentixol decanoate.

Flupentixol Hydrochloride

(Flupentixol Dihydrochloride, Ph. Eur. monograph 1693)



$C_{23}H_{27}Cl_2F_3N_2OS$ 507.4 *2413-38-9*

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Flupentixol Tablets

Ph Eur

DEFINITION

2-[4-[3-[(*EZ*)-2-(Trifluoromethyl)-9*H*-thioxanthen-9-ylidene] propyl]piperazin-1-yl]ethan-1-ol dihydrochloride.

Content

— *flupentixol dihydrochloride*: 99.0 per cent to 101.0 per cent (dried substance);
— *(Z)-isomer*: 42.0 per cent to 52.0 per cent.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very soluble in water, soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flupentixol dihydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *flupentixol dihydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *water R, diethylamine R, methyl ethyl ketone R* (1:4:95 *V/V/V*).

Application 2 µL.

Development Twice over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution. Doubling of the spot may be observed in both chromatograms.

Detection B Spray with *alcoholic solution of sulfuric acid R*; heat at 110 °C for 5 min and allow to cool; examine in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution. Doubling of the spot may be observed in both chromatograms.

C. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter and add to the filtrate a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow. The blank is red.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and its absorbance (*2.2.25*) at 410 nm is not greater than 0.125.

Dissolve 0.500 g in 5.0 mL of *water R*.

pH (*2.2.3*)

2.0 to 3.0.

Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Buffer solution Dissolve 6.3 g of *ammonium formate R* in about 900 mL of *water for chromatography R*, adjust to pH 8.2 with *concentrated ammonia R* and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 58.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *flupentixol for system suitability CRS* (containing impurities A, C, H and I) in 1 mL of mobile phase A.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: buffer solution, *acetonitrile for chromatography R*, *water for chromatography R* (58:420:522 *V/V/V*);
- *mobile phase B*: buffer solution, *acetonitrile for chromatography R* (100:900 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 40	90 → 5	10 → 95

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Identification of peaks Use the chromatogram supplied with *flupentixol for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to both isomers of flupentixol and impurities A, C + I, and H.

Relative retention With reference to flupentixol (*Z*)-isomer (retention time = about 11 min): impurity A = about 0.83; impurity H = about 0.90; impurities C and I = about 0.97; flupentixol (*E*)-isomer = about 1.04.

System suitability Reference solution (b):
- *resolution*: minimum 2.5 between the peaks due to impurities A and H;
- *peak-to-valley ratio*: minimum 10.0, where H_p = height above the baseline of the peak due to impurities C+I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to flupentixol (*Z*)-isomer.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurities C and I = 2.0; impurity H = 2.0;
- for each impurity, use the concentration of flupentixol dihydrochloride (both isomers) in reference solution (a).

Limits:
- *impurity H*: maximum 0.5 per cent;
- *sum of impurities C and I*: maximum 0.3 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 1.0 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Flupentixol dihydrochloride

Dissolve 0.200 g in 30 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 50.74 mg of $C_{23}H_{27}Cl_2F_3N_2OS$.

(*Z*)-Isomer

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution Dissolve 20.0 mg of *flupentixol dihydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: *silica gel for chromatography R* (5 µm).

Mobile phase *water for chromatography R, concentrated ammonia R, 2-propanol R, heptane R* (2:4:150:850 *V/V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 1.5 times the retention time of flupentixol (*Z*)-isomer.

Identification of peaks Use the chromatogram supplied with *flupentixol dihydrochloride CRS* and the chromatogram obtained with the reference solution to identify the peaks due to both isomers of flupentixol.

Relative retention With reference to flupentixol (*Z*)-isomer (retention time = about 6 min): flupentixol (*E*)-isomer = about 1.2.

System suitability Reference solution:

— *resolution*: minimum 3.0 between the peaks due to flupentixol (*Z*)-isomer and flupentixol (*E*)-isomer.

Calculate the percentage content of the (*Z*)-isomer of $C_{23}H_{27}Cl_2F_3N_2OS$ taking into account the assigned content of flupentixol (*Z*)-isomer in *flupentixol dihydrochloride CRS*.

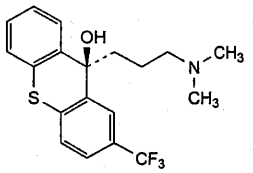
STORAGE

Protected from light.

IMPURITIES

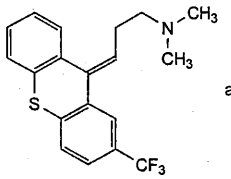
Specified impurities *C, H, I.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E, G.*



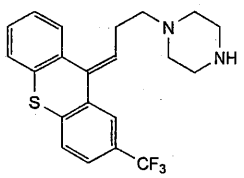
and enantiomer

A. (9*RS*)-9-[3-(dimethylamino)propyl]-2-(trifluoromethyl)-9*H*-thioxanthen-9-ol,



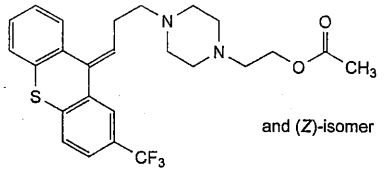
and (*Z*)-isomer

B. *N*,*N*-dimethyl-3-[(*EZ*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-ylidene]propan-1-amine,



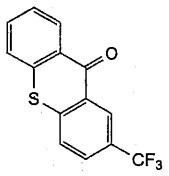
and (*Z*)-isomer

C. 1-[3-[(*EZ*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-ylidene]propyl]piperazine,

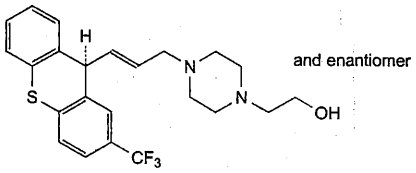


and (*Z*)-isomer

E. 2-[4-[3-[(*EZ*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-ylidene]propyl]piperazin-1-yl]ethyl acetate,

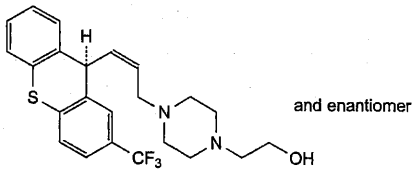


G. 2-(trifluoromethyl)-9*H*-thioxanthen-9-one,



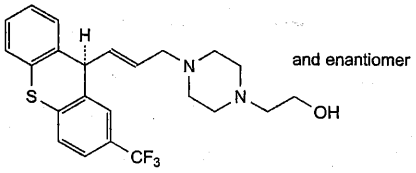
and enantiomer

or



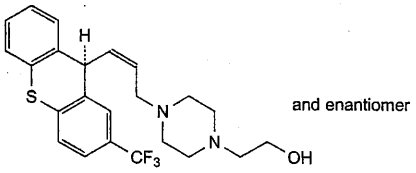
and enantiomer

H. either 2-[4-[(*E*)-3-[(9*RS*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-yl]prop-2-en-1-yl]piperazin-1-yl]ethan-1-ol or 2-[4-[(*Z*)-3-[(9*RS*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-yl]prop-2-en-1-yl]piperazin-1-yl]ethan-1-ol,



and enantiomer

or

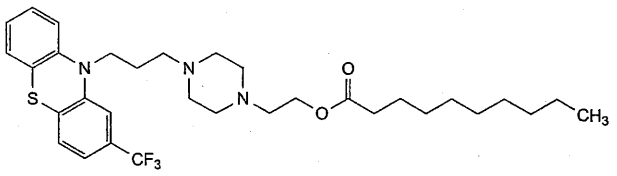


and enantiomer

I. either 2-[4-[(*E*)-3-[(9*RS*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-yl]prop-2-en-1-yl]piperazin-1-yl]ethan-1-ol or 2-[4-[(*Z*)-3-[(9*RS*)-2-(trifluoromethyl)-9*H*-thioxanthen-9-yl]prop-2-en-1-yl]piperazin-1-yl]ethan-1-ol.

Fluphenazine Decanoate

(Ph. Eur. monograph 1014)



$C_{32}H_{44}F_3N_3O_2S$ 591.8 *5002-47-1*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparation
Fluphenazine Decanoate Injection

Ph Eur

DEFINITION
2-[4-[3-[2-(Trifluoromethyl)-10*H*-phenothiazin-10-yl]propyl] piperazin-1-yl]ethyl decanoate.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
Pale yellow, viscous liquid or yellow solid.

Solubility
Practically insoluble in water, very soluble in ethanol and in methylene chloride, freely soluble in methanol.

IDENTIFICATION
First identification: B, C.

Second identification: A, C.

A. Dissolve 50.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL to 50.0 mL with *methanol R*. Examined between 230 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 260 nm and a broad absorption maximum at about 310 nm. The specific absorbance at the maximum at 260 nm is 570 to 630.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Apply 50 µL of a 25 g/L solution in *methylene chloride R* to a disc of *potassium bromide R*. Dry the discs at 60 °C for 1 h before use.

Comparison *fluphenazine decanoate CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *fluphenazine decanoate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *fluphenazine enantate CRS* in reference solution (a) and dilute to 5 mL with the same solution.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R1, water R, methanol R* (1:4:95 *V/V/V*).

Application 2 µL.

Development Over a path of 8 cm.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *fluphenazine octanoate CRS* and 5 mg of *fluphenazine enantate CRS* in *acetonitrile R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with a mixture of 5 volumes of mobile phase A and 95 volumes of mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 5 volumes of mobile phase A and 95 volumes of mobile phase B.

Reference solution (c) Dissolve 11.7 mg of *fluphenazine dihydrochloride CRS* and 5.0 mg of *fluphenazine sulfoxide CRS* in a mixture of 5 volumes of *water R* and 95 volumes of *acetonitrile R* and dilute to 100.0 mL with the same mixture of solvents. Dilute 1.0 mL to 50.0 mL with a mixture of 5 volumes of *water R* and 95 volumes of *acetonitrile R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: 10 g/L solution of *ammonium carbonate R* adjusted to pH 7.5 with *dilute hydrochloric acid R*,
— *mobile phase B*: mobile phase A, *acetonitrile R*, *methanol R* (7.5:45:45 *V/V/V*),

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	20	80
7 - 17	20 → 0	80 → 100
17 - 80	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Relative retention With reference to fluphenazine decanoate (retention time = about 34 min): impurity A = about 0.13; impurity B = about 0.33; impurity C = about 0.76; impurity D = about 0.82.

System suitability Reference solution (a):
— *resolution*: minimum 6 between the peaks due to impurity C and impurity D.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (1.0 per cent),
— *any other impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),

— *total*: not more than 2.0 per cent,
— *disregard limit for any other impurity*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.7 kPa for 3 h.

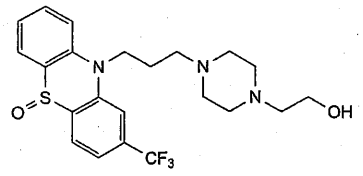
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.250 g in 30 mL of *glacial acetic acid R*. Using 0.05 mL of *crystal violet solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from violet to green.

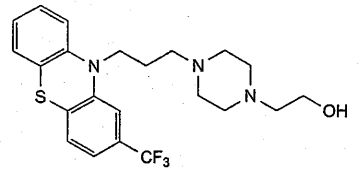
1 mL of *0.1 M perchloric acid* is equivalent to 29.59 mg of $C_{32}H_{44}F_3N_3O_2S$.

STORAGE
Protected from light.

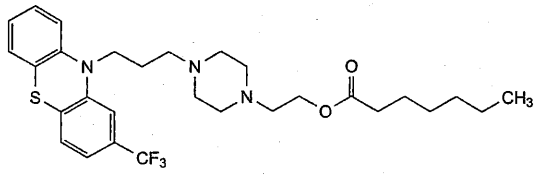
IMPURITIES



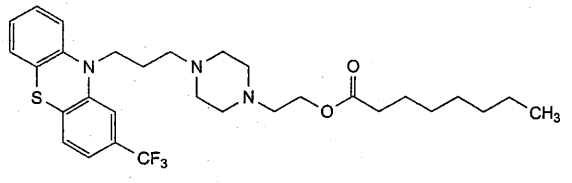
A. fluphenazine *S*-oxide,



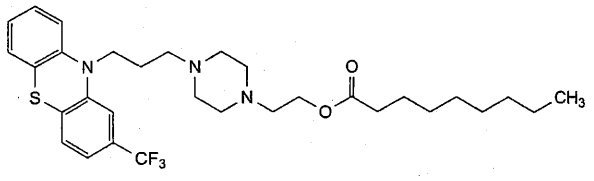
B. fluphenazine,



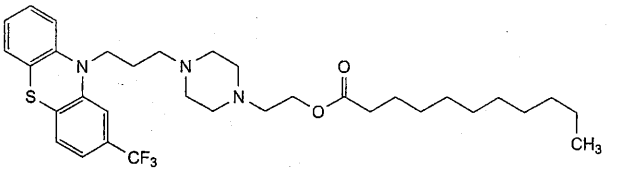
C. fluphenazine enantate,



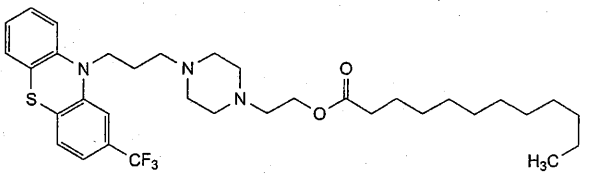
D. fluphenazine octanoate,



E. fluphenazine nonanoate,



F. fluphenazine undecanoate,

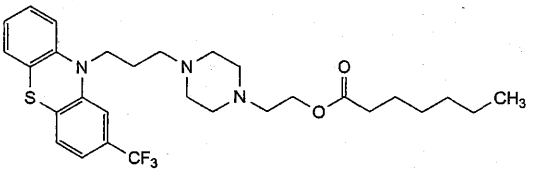


G. fluphenazine dodecanoate.

Ph Eur

Fluphenazine Enantate

(Ph. Eur. monograph 1015)



$C_{29}H_{38}F_3N_3O_2S$ 549.7 *2746-81-8*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
2-[4-[3-[2-(Trifluoromethyl)-10*H*-phenothiazin-10-yl]propyl] piperazin-1-yl]ethyl heptanoate.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
Pale yellow, viscous liquid or yellow solid.

Solubility
Practically insoluble in water, very soluble in ethanol and in methylene chloride, freely soluble in methanol.

IDENTIFICATION
First identification: B, C.
Second identification: A, C.

A. Dissolve 50.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL to 50.0 mL with *methanol R*. Examined between 230 nm and 350 nm (*2.2.25*), the solution shows an absorption maximum at 260 nm and a broad absorption maximum at about 310 nm. The specific absorbance at the maximum at 260 nm is 610 to 670.

B. Infrared absorption spectrophotometry (*2.2.24*).
Preparation Apply 50 µL of a 25 g/L solution in *methylene chloride R* to a disc of *potassium bromide R*. Dry the discs at 60 °C for 1 h before use.
Comparison *fluphenazine enantate CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *fluphenazine enantate CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *fluphenazine decanoate CRS* in reference solution (a) and dilute to 5 mL with the same solution.

Plate *TLC octadecylsilyl silica gel F_{254} plate R.*

Mobile phase *concentrated ammonia R1, water R, methanol R* (1:4:95 *V/V/V*).

Application 2 µL.

Development Over a path of 8 cm.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in *acetonitrile R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of *fluphenazine octanoate CRS* and 5 mg of *fluphenazine enantate CRS* in *acetonitrile R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with a mixture of 5 volumes of mobile phase A and 95 volumes of mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 5 volumes of mobile phase A and 95 volumes of mobile phase B.

Reference solution (c) Dissolve 5.0 mg of *fluphenazine sulfoxide CRS* in *acetonitrile R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL to 50.0 mL with *acetonitrile R.*

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm,
- *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: 10 g/L solution of *ammonium carbonate R* adjusted to pH 7.5 with *dilute hydrochloric acid R*,
- *mobile phase B*: mobile phase A, *acetonitrile R, methanol R* (7.5:45:45 *V/V/V*),

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	20	80
7 - 17	20 → 0	80 → 100
17 - 80	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Relative retention With reference to fluphenazine enantate (retention time = about 25 min): impurity A = about 0.2; impurity D = about 1.1.

System suitability Reference solution (a):
- *resolution*: minimum 6 between the peaks due to fluphenazine enantate and impurity D.

Limits:
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
- *any other impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
- *total*: not more than 1.6 per cent,
- *disregard limit for any other impurity*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.7 kPa for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

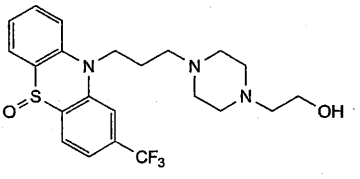
Dissolve 0.250 g in 30 mL of *glacial acetic acid R*. Using 0.05 mL of *crystal violet solution R* as indicator titrate with *0.1 M perchloric acid* until the colour changes from violet to green.

1 mL of *0.1 M perchloric acid* is equivalent to 27.49 mg of $C_{29}H_{38}F_3N_3O_2S$.

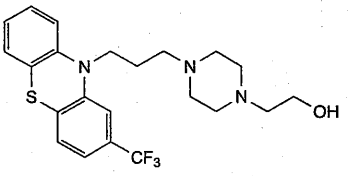
STORAGE

Protected from light.

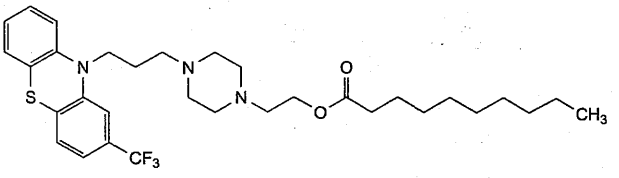
IMPURITIES



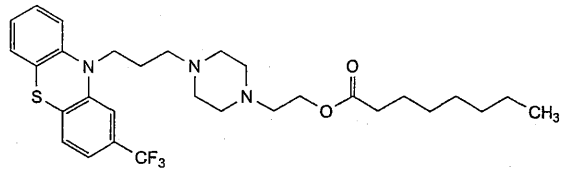
A. fluphenazine *S*-oxide,



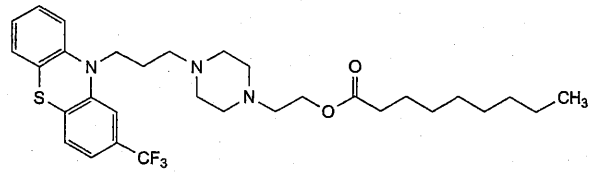
B. fluphenazine,



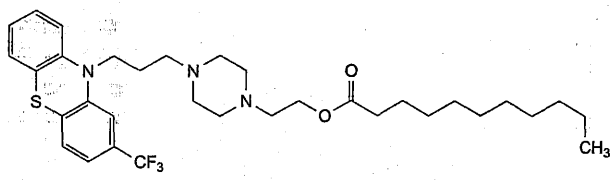
C. fluphenazine decanoate,



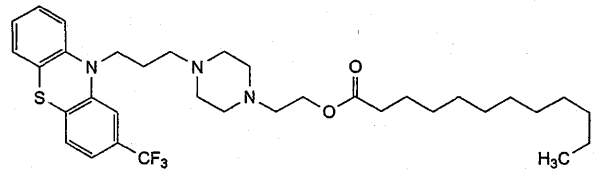
D. fluphenazine octanoate,



E. fluphenazine nonanoate,



F. fluphenazine undecanoate,

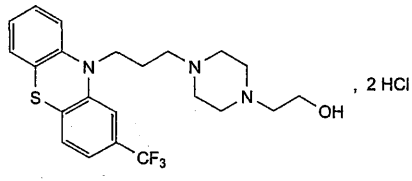


G. fluphenazine dodecanoate.

Ph Eur

Fluphenazine Hydrochloride

(Fluphenazine Dihydrochloride, Ph. Eur. monograph 0904)



$C_{22}H_{28}Cl_2F_3N_3OS$ 510.4 *146-56-5*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparation
Fluphenazine Tablets

Ph Eur

DEFINITION
2-[4-[3-[2-(Trifluoromethyl)-10*H*-phenothiazin-10-yl]propyl] piperazin-1-yl]ethanol dihydrochloride.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with *methanol R*.

Spectral range 230-350 nm.

Absorption maxima At 260 nm and at about 310 nm (broad band).

Specific absorbance at the absorption maximum at 260 nm 630 to 700.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluphenazine dihydrochloride CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *fluphenazine dihydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of *perphenazine CRS* in reference solution (a) and dilute to 5 mL with reference solution (a).

Plate *TLC octadecylsilyl silica gel F_{254} plate R*.

Mobile phase *concentrated ammonia R1, water R, methanol R* (1:4:95 *V/V/V*).

Application 2 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
pH (*2.2.3*)
1.9 to 2.4.

Dissolve 0.5 g in 10 mL of *water R*.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Solution A Mix 4 mL of *acetic acid R* and 996 mL of a 4.33 g/L solution of *sodium octanesulfonate R*.

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 25.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *fluphenazine impurity mixture CRS* (impurities A, B, C and D) in 5 mL of the test solution and sonicate for 1 min. Mix 1.0 mL of this solution and 1.0 mL of the test solution.

Reference solution (c) Dissolve 5.0 mg of *fluphenazine sulfoxide CRS* (impurity A) in mobile phase A and dilute to 50.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *acetic acid R, methanol R, acetonitrile R,* solution A (0.2:15:40:45 *V/V/V/V*);
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100	0
15 - 35	100 → 30	0 → 70
35 - 50	30	70

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 260 nm and at 274 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *fluphenazine impurity mixture CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and D.

Relative retention With reference to fluphenazine (retention time = about 19 min): impurity A = about 0.2; impurity B = about 0.3; impurity D = about 2.0; impurity C = about 2.1.

System suitability Reference solution (b):
— *resolution at 274 nm*: minimum 2.5 between the peaks due to impurities A and B.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.3; impurity C = 0.6;
— *impurity A at 274 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *impurity B at 274 nm*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurities C, D at 260 nm*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities at 260 nm*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of the impurities other than A and B at 260 nm and impurities A and B at 274 nm*: not more than 1.0 per cent;
— *disregard limit at 260 nm*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 65 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.220 g in a mixture of 10 mL of *anhydrous formic acid R* and 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 25.52 mg of $C_{22}H_{28}Cl_2F_3N_3OS$.

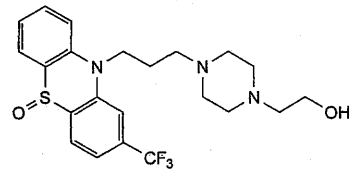
STORAGE

Protected from light.

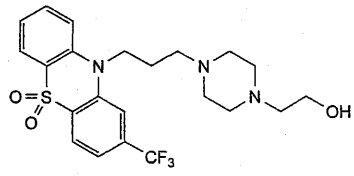
IMPURITIES

Specified impurities *A, B, C, D.*

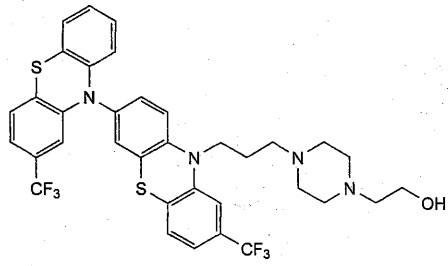
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F.*



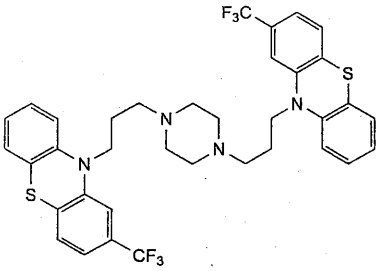
A. 2-[4-[3-[5-oxo-2-(trifluoromethyl)-10*H*-5λ⁴-phenothiazin-10-yl]propyl]piperazin-1-yl]ethanol (fluphenazine *S*-oxide),



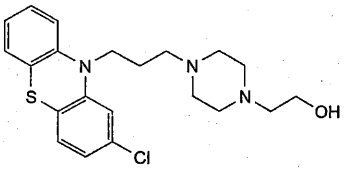
B. 2-[4-[3-[5,5-dioxo-2-(trifluoromethyl)-10*H*-5λ⁶-phenothiazin-10-yl]propyl]piperazin-1-yl]ethanol (fluphenazine *S,S*-dioxide),



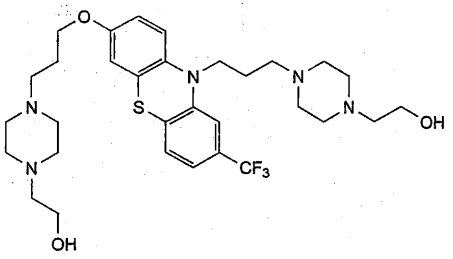
C. 2-[4-[3-[2′,8-bis(trifluoromethyl)-10*H*-3,10′-biphenothiazin-10-yl]propyl]piperazin-1-yl]ethanol,



D. 10,10′-[piperazine-1,4-diylbis(propane-3,1-diyl)]bis[2-(trifluoromethyl)-10*H*-phenothiazine],



E. 2-[4-[3-[2-chloro-10*H*-phenothiazin-10-yl] propyl]piperazin-1-yl]ethanol (perphenazine),

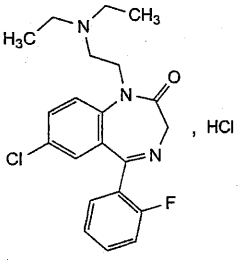


F. 2-[4-[3-[7-[3-[4-(2-hydroxyethyl)piperazin-1-yl]propoxy]-2-(trifluoromethyl)-10*H*-phenothiazin-10-yl] propyl]piperazin-1-yl)ethanol.

Ph Eur

Flurazepam Monohydrochloride

(Ph. Eur. monograph 0905)



$C_{21}H_{24}Cl_2FN_3O$ 424.3 *36105-20-1*

Action and use
Benzodiazepine.

Preparation
Flurazepam Capsules

Ph Eur

DEFINITION
7-Chloro-1-[2-(diethylamino)ethyl]-5-(2-fluorophenyl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one monohydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very soluble in water, freely soluble in alcohol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of flurazepam monohydrochloride.*

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
pH (*2.2.3*)
5.0 to 6.0.

Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *oxazepam R* in 10 mL of *acetonitrile R* and dilute to 50.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm,
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 350 volumes of *acetonitrile R* and 650 volumes of a 10.5 g/L solution of *potassium dihydrogen phosphate R* and ajust to pH 6.1 with a 40 g/L solution of *sodium hydroxide R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 239 nm.

Injection 20 µL.

Run time: 6 times the retention time of flurazepam.

Relative retention With reference to flurazepam (retention time = about 7 min): impurity C = about 1.5; impurity B = about 1.9; impurity A = about 2.4.

System suitability Reference solution (b):
— *resolution*: minimum of 4.5 between the peaks due to flurazepam and to oxazepam.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.61; impurity C = 0.65,
— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Fluorides (*2.4.5*)
Maximum 500 ppm.

0.10 g complies with the limit test for fluorides.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

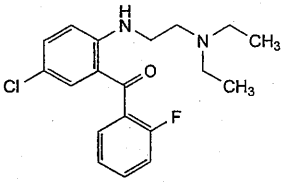
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.350 g in a mixture of 1.0 mL of *0.1 M hydrochloric acid* and 50 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

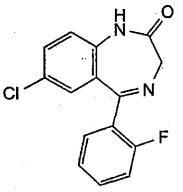
1 mL of *0.1 M sodium hydroxide* is equivalent to 42.43 mg of $C_{21}H_{24}Cl_2FN_3O$.

STORAGE
Protected from light.

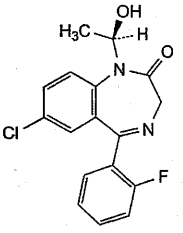
IMPURITIES



A. [5-chloro-2-[[2-(diethylamino)ethyl]amino]phenyl](2-fluorophenyl)methanone,



B. 7-chloro-5-(2-fluorophenyl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one,

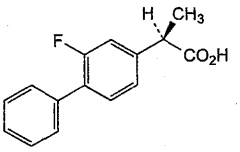


C. 7-chloro-5-(2-fluorophenyl)-1-[(1*RS*)-1-hydroxyethyl]-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Ph Eur

Flurbiprofen

(Ph. Eur. monograph 1519)



$C_{15}H_{13}FO_2$ 244.3 *5104-49-4*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Flurbiprofen Suppositories
Flurbiprofen Tablets

Ph Eur

DEFINITION
(2*RS*)-2-(2-Fluorobiphenyl-4-yl)propanoic acid.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride. It dissolves in aqueous solutions of alkali hydroxides and carbonates.

IDENTIFICATION
First identification: C, D.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 114 °C to 117 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 0.10 g in *0.1 M sodium hydroxide* and dilute to 100.0 mL with the same alkaline solution. Dilute 1.0 mL of this solution to 100.0 mL with *0.1 M sodium hydroxide*.

Spectral range 230-350 nm.

Absorption maximum At 247 nm.

Specific absorbance at the absorption maximum 780 to 820.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flurbiprofen CRS*.

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R* add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method I*).

Dissolve 1.0 g in *methanol R* and dilute to 10 mL with the same solvent.

Optical rotation (*2.2.7*)
−0.1° to + 0.1°.
Dissolve 0.50 g in *methanol R* and dilute to 20.0 mL with the same solvent.

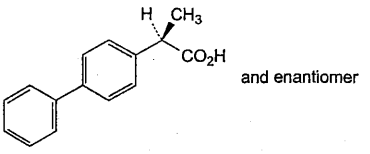
Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *acetonitrile R, water R* (45:55 *V/V*).
Test solution Dissolve 0.20 g of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.
Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.
Reference solution (b) Dissolve 10.0 mg of *flurbiprofen impurity A CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 10.0 mL of this solution to 100.0 mL with the solvent mixture.
Reference solution (c) Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with reference solution (b).
Column:
— *size*: *l* = 0.15 m, Ø = 3.9 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).
Mobile phase *glacial acetic acid R, acetonitrile R, water R* (5:35:60 *V/V/V*).
Flow rate 1 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 10 µL.
Run time Twice the retention time of flurbiprofen.
System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity A and flurbiprofen.
Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurities B, C, D, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *sum of impurities other than A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying at 60 °C at a pressure not exceeding 0.7 kPa for 3 h.

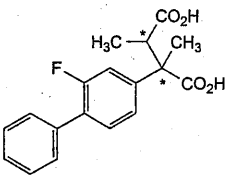
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Dissolve 0.200 g in 50 mL of *ethanol (96 per cent) R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M sodium hydroxide* is equivalent to 24.43 mg of $C_{15}H_{13}FO_2$.

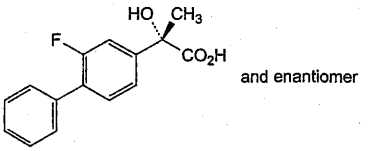
IMPURITIES
Specified impurities *A, B, C, D, E.*



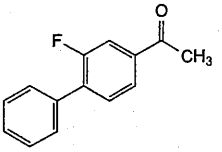
A. (2*RS*)-2-(biphenyl-4-yl)propanoic acid,



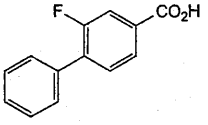
B. 2-(2-fluorobiphenyl-4-yl)-2,3-dimethylbutanedioic acid,



C. (2*RS*)-2-(2-fluorobiphenyl-4-yl)-2-hydroxypropanoic acid,



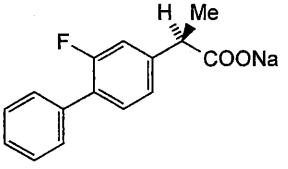
D. 1-(2-fluorobiphenyl-4-yl)ethanone,



E. 2-fluorobiphenyl-4-carboxylic acid.

Ph Eur

Flurbiprofen Sodium



$C_{15}H_{12}FNaO_2,2H_2O$ 302.3 *56767-76-1*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparation
Flurbiprofen Eye Drops

DEFINITION
Flurbiprofen Sodium is sodium (*RS*)-2-(2-fluorobiphenyl-4-yl)propionate dihydrate. It contains not less than 98.5% and not more than 101.5% of $C_{15}H_{12}FNaO_2$, calculated with reference to the dried substance.

CHARACTERISTICS

A white to creamy-white, crystalline powder.

Sparingly soluble in *water*; soluble in *ethanol (96%)*; practically insoluble in *dichloromethane*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of flurbiprofen sodium (*RS 157*).

B. Heat 0.2 g over a flame until charred and then heat at 600° for 2 hours. The residue yields the reactions characteristic of *sodium salts*, Appendix VI.

TESTS

Related substances

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in a mixture of 25 volumes of *water* and 50 volumes of *methanol*.

(1) 0.10% w/v of the substance being examined.

(2) 0.00020% w/v of the substance being examined.

(3) 0.00050% w/v of *2-(biphenyl-4-yl)propionic acid BPCRS*.

(4) 0.00050% w/v of the substance being examined and 0.00050% w/v of *2-(biphenyl-4-yl)propionic acid BPCRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 3.9 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Resolve 5µ is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

5 volumes of *glacial acetic acid*, 35 volumes of *acetonitrile* and 60 volumes of *water*.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (4), the *resolution factor* between the two principal peaks is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to 2-(biphenyl-4-yl) propionic acid is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.5%);

the area of any other *secondary peak* is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.2%);

the sum of the areas of any *secondary peaks* is not greater than five times the area of the peak in the chromatogram obtained with solution (2) (1%).

Loss on drying

11.3% to 12.5% when determined by drying over *phosphorus pentoxide* at 60° at a pressure of 2 kPa for 18 hours. Use 1 g.

ASSAY

Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in a mixture of 25 volumes of *water* and 50 volumes of *methanol*.

(1) 0.015% w/v of the substance being examined.

(2) 0.015% w/v of *flurbiprofen sodium BPCRS*.

(3) 0.00075% w/v of the substance being examined and 0.00075% w/v of *2-(biphenyl-4-yl)propionic acid BPCRS*.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related substances may be used.

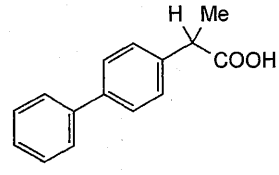
SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the *resolution factor* between the two principal peaks is at least 1.5.

DETERMINATION OF CONTENT

Calculate the content of $C_{15}H_{12}FNaO_2$ in the substance being examined using the declared content of $C_{15}H_{12}FNaO_2$ in *flurbiprofen sodium BPCRS*.

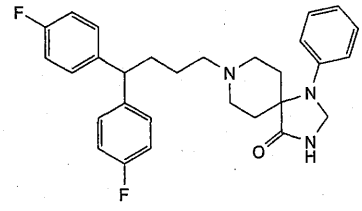
IMPURITIES



A. 2-(biphenyl-4-yl)propionic acid.

Fluspirilene

(Ph. Eur. monograph 1723)



$C_{29}H_{31}F_2N_3O$ 475.6 *1841-19-6*

Action and use

Antipsychotic.

Ph Eur

DEFINITION

8-[4,4-bis(4-Fluorophenyl)butyl]-1-phenyl-1,3,8-triazaspiro [4.5]decan-4-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluspirilene CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *methylene chloride R*, gently evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 0.25 g in 25 mL of *methylene chloride R*.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *dimethylformamide R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *fluspirilene impurity C CRS* in *dimethylformamide R*, add 0.5 mL of the test solution and dilute to 100.0 mL with *dimethylformamide R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with *dimethylformamide R*. Dilute 1.0 mL of this solution to 25.0 mL with *dimethylformamide R*.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
- *mobile phase A*: 13.6 g/L solution of *tetrabutylammonium hydrogen sulfate R*,
- *mobile phase B*: *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	75 → 70	25 → 30
15 - 20	70	30
20 - 22	70 → 0	30 → 100
22 - 30	0	100

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to fluspirilene (retention time = about 15 min): impurity A = about 0.8; impurity B = about 0.93; impurity C = about 0.97.

System suitability Reference solution (a):
- *resolution*: minimum 2.2 between the peaks due to impurity C and fluspirilene.

Limits:
- *impurities A, B, C*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent),
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent),
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.6 per cent),
- *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.350 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

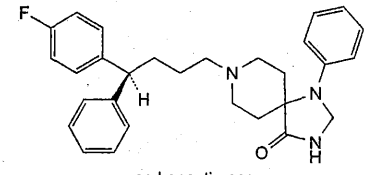
1 mL of *0.1 M perchloric acid* is equivalent to 47.56 mg of $C_{29}H_{31}F_2N_3O$.

STORAGE

Protected from light.

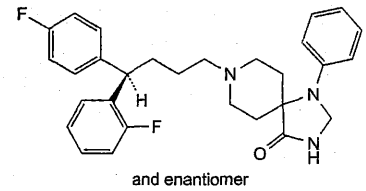
IMPURITIES

Specified impurities *A, B, C.*



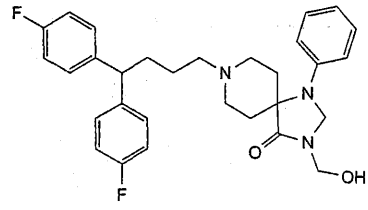
and enantiomer

A. 8-[(4*RS*)-4-(4-fluorophenyl)-4-phenylbutyl]-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one,



and enantiomer

B. 8-[(4*RS*)-4-(2-fluorophenyl)-4-(4-fluorophenyl)butyl]-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one,

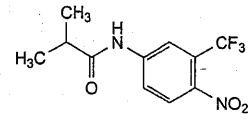


C. 8-[4,4-bis(4-fluorophenyl)butyl]-3-(hydroxymethyl)-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one.

Ph Eur

Flutamide

(Ph. Eur. monograph 1423)



$C_{11}H_{11}F_3N_2O_3$ 276.2 *13311-84-7*

Action and use

Antiandrogen.

Ph Eur

DEFINITION

2-Methyl-*N*-[4-nitro-3-(trifluoromethyl)phenyl]propanamide.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Pale yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent), practically insoluble in heptane.

mp

About 112 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *flutamide CRS.*

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *flutamide for system suitability CRS* (containing impurities A, B and C) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 20.0 mg of *flutamide CRS* in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.0 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R, water R* (50:50 *V/V*).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Run time 1.5 times the retention time of flutamide.

Identification of impurities Use the chromatogram supplied with *flutamide for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to flutamide (retention time = about 19 min): impurity B = about 0.5; impurity A = about 0.6; impurity C = about 0.7.

System suitability Reference solution (a):

— *resolution*: minimum 2.0 between the peaks due to impurities B and A.

Calculation of percentage contents:

— for each impurity, use the concentration of flutamide in reference solution (b).

Limits:

— *impurities A, B, C*: for each impurity, maximum 0.2 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 0.5 per cent;

— *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{11}H_{11}F_3N_2O_3$ taking into account the assigned content of *flutamide CRS*.

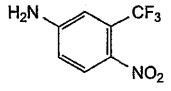
STORAGE

Protected from light.

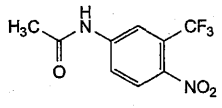
IMPURITIES

Specified impurities *A, B, C.*

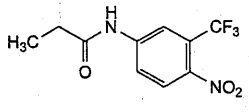
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E, F.*



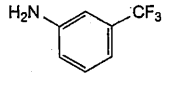
A. 4-nitro-3-(trifluoromethyl)aniline,



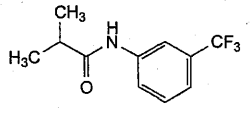
B. *N*-[4-nitro-3-(trifluoromethyl)phenyl]acetamide,



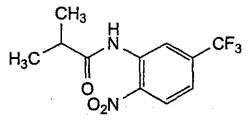
C. *N*-[4-nitro-3-(trifluoromethyl)phenyl]propanamide,



D. 3-(trifluoromethyl)aniline,



E. 2-methyl-*N*-[3-(trifluoromethyl)phenyl]propanamide,

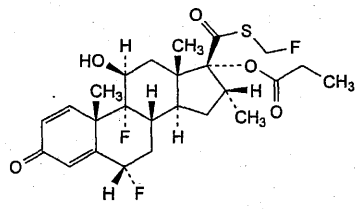


F. 2-methyl-*N*-[2-nitro-5-(trifluoromethyl)phenyl] propanamide.

Ph Eur

Fluticasone Propionate

(Ph. Eur. monograph 1750)



$C_{25}H_{31}F_3O_5S$ 500.6 *80474-14-2*

Action and use
Glucocorticoid.

Preparations
Fluticasone Cream
Fluticasone Inhalation Powder
Fluticasone Inhalation Powder, pre-metered
Fluticasone Nasal Drops
Fluticasone Nasal Spray
Fluticasone Ointment
Fluticasone Pressurised Inhalation
Fluticasone and Salmeterol Inhalation Powder, pre-metered
Fluticasone & Salmeterol Pressurised Inhalation, suspension

Ph Eur

DEFINITION

6α,9-Difluoro-17-[[(fluoromethyl)sulfanyl]carbonyl]-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-dien-17α-yl propanoate.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).
Comparison *fluticasone propionate CRS*.

TESTS

Specific optical rotation (*2.2.7*)
+ 32 to + 36 (anhydrous substance).
Dissolve 0.25 g in *methylene chloride R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Solvent mixture Mobile phase A, mobile phase B (50:50 *V/V*).

Test solution Dissolve 20 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 1 mg of *fluticasone impurity D CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 25.0 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of *fluticasone propionate for impurity G identification CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 2 mg of *fluticasone propionate for impurity C identification CRS* in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: solution containing 0.05 per cent *V/V* of *phosphoric acid R* and 3.0 per cent *V/V* of *methanol R* in *water R*;
— *mobile phase B*: solution containing 0.05 per cent *V/V* of *phosphoric acid R* and 3.0 per cent *V/V* of *methanol R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	57	43
2 - 42	57 → 45	43 → 55
42 - 62	45 → 10	55 → 90
62 - 72	10	90

Flow rate 1 mL/min.

Detection Spectrophotometer at 239 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D; use the chromatogram supplied with *fluticasone propionate for impurity G identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity G; use the chromatogram supplied with *fluticasone propionate for impurity C identification CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to fluticasone propionate (retention time = about 32 min): impurity C = about 0.8; impurity D = about 0.95; impurity G = about 1.3.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurity D and fluticasone propionate.

Limits:
— *impurities D, G*: for each impurity, maximum 0.3 per cent;
— *impurity C*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.8 per cent;
— *reporting threshold*: 0.05 per cent (reference solution (b)).

Acetone

Gas chromatography (*2.2.28*).

Internal standard solution Dilute 0.5 mL of *tetrahydrofuran R* to 1000 mL with *dimethylformamide R*.

Test solution Dissolve 0.50 g of the substance to be examined in the internal standard solution and dilute to 10.0 mL with the internal standard solution.

Reference solution Dilute 0.40 g of *acetone R* to 100.0 mL with the internal standard solution. Dilute 1.0 mL of this solution to 10.0 mL with the internal standard solution.

Column:
- *material*: fused silica;
- *size*: *l* = 25 m, Ø = 0.53 mm;
- *stationary phase*: cross-linked *macrogol 20 000 R* (film thickness 2 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 5.5 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3.5	60
	3.5 - 7.5	60 → 180
	7.5 - 10.5	180
Injection port		150
Detector		250

Detection Flame ionisation.

Injection 0.1 µL.

Limit:
- *acetone*: maximum 1.0 per cent *m/m*.

Water (*2.5.32*)

Maximum 0.5 per cent, determined on 0.100 g using the evaporation technique:
- *temperature*: 160 °C;
- *heating time*: 3 min.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve, using sonication, 20.0 mg of the substance to be examined in the mobile phase and dilute to 200.0 mL with the mobile phase. Dilute 4.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve, using sonication, 20.0 mg of *fluticasone propionate CRS* in the mobile phase and dilute to 200.0 mL with the mobile phase.

Reference solution (b) Dilute 4.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 1 mg of *fluticasone impurity D CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. To 1.0 mL of the solution add 2.0 mL of reference solution (a) and dilute to 5.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 40 °C.

Mobile phase Mix 15 volumes of *acetonitrile R*, 35 volumes of a 1.15 g/L solution of *ammonium dihydrogen phosphate R* adjusted to pH 3.5 and 50 volumes of *methanol R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 239 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of fluticasone propionate.

Relative retention With reference to fluticasone propionate (retention time = about 6 min): impurity D = about 1.1.

System suitability Reference solution (c):
- *resolution*: minimum 1.5 between the peaks due to fluticasone propionate and impurity D; if necessary, adjust the ratio of acetonitrile to methanol in the mobile phase.

Calculate the percentage content of $C_{25}H_{31}F_3O_5S$ using the chromatogram obtained with reference solution (b) and taking into account the assigned content of *fluticasone propionate CRS*.

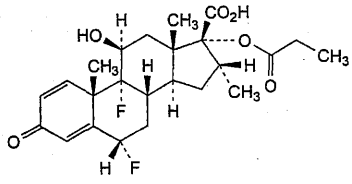
STORAGE

Protected from light.

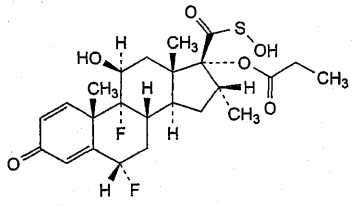
IMPURITIES

Specified impurities *C, D, G.*

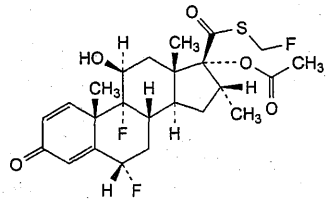
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E, F, H, I.*



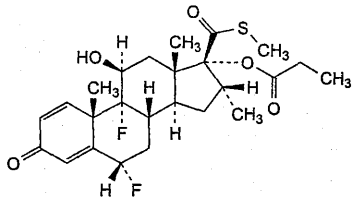
A. 6α,9-difluoro-11β-hydroxy-16α-methyl-3-oxo-17-(propanoyloxy)androsta-1,4-diene-17β-carboxylic acid,



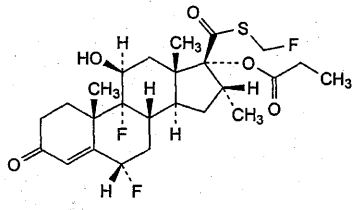
B. 6α,9-difluoro-11β-hydroxy-16α-methyl-3-oxo-17-(propanoyloxy)androsta-1,4-dien-17β-carbo(thioperoxoic) *SO*-acid,



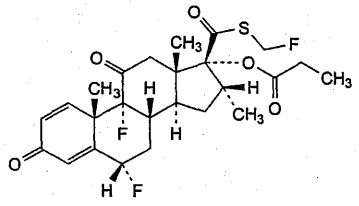
C. 6α,9-difluoro-17-[[(fluoromethyl)sulfanyl]carbonyl]-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-dien-17α-yl acetate,



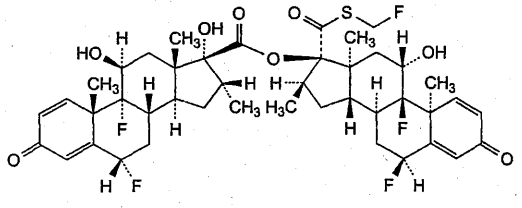
D. 6α,9-difluoro-11β-hydroxy-16α-methyl-17-[(methylsulfanyl)carbonyl]-3-oxoandrosta-1,4-dien-17α-yl propanoate,



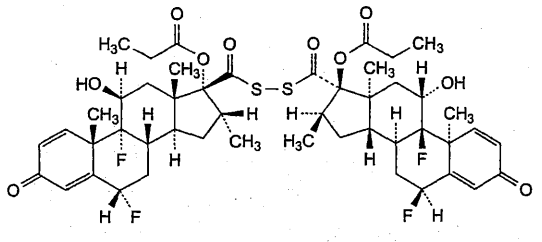
E. 6α,9-difluoro-17-[[(fluoromethyl)sulfanyl]carbonyl]-11β-hydroxy-16α-methyl-3-oxoandrost-4-en-17α-yl propanoate,



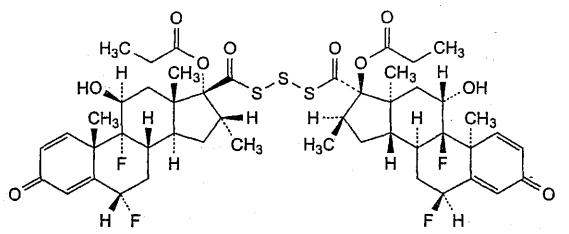
F. 6α,9-difluoro-17-[[(fluoromethyl)sulfanyl]carbonyl]-16α-methyl-3,11-dioxoandrosta-1,4-dien-17α-yl propanoate,



G. 6α,9-difluoro-17-[[(fluoromethyl)sulfanyl]carbonyl]-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-dien-17α-yl 6α,9-difluoro-11β,17-dihydroxy-16α-methyl-3-oxoandrosta-1,4-diene-17β-carboxylate,



H. 17,17′-(disulfanediyldicarbonyl)bis(6α,9-difluoro-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-dien-17α-yl) dipropanoate,

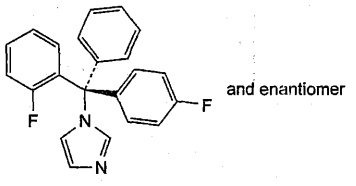


I. 17,17′-(trisulfanediyldicarbonyl)bis(6α,9-difluoro-11β-hydroxy-16α-methyl-3-oxoandrosta-1,4-dien-17α-yl) dipropanoate.

Ph Eur

Flutrimazole

(Ph. Eur. monograph 1424)



$C_{22}H_{16}F_2N_2$ 346.4 *119006-77-8*

Action and use
Antifungal.

Ph Eur

DEFINITION

(*RS*)-1-[(2-Fluorophenyl)(4-fluorophenyl)phenylmethyl]-1*H*-imidazole.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in tetrahydrofuran, soluble in methanol.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 161 °C to 166 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *flutrimazole CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *flutrimazole CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *flutrimazole CRS* and 10 mg of *metronidazole benzoate CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Pretreatment Heat the plate at 110 °C for 1 h.

Mobile phase *2-propanol R, ethyl acetate R* (10:90 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Mix about 5 mg with 45 mg of *heavy magnesium oxide R* and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of *water R*, 0.05 mL of *phenolphthalein solution R1* and about 1 mL of *dilute hydrochloric acid R* to render the solution colourless. Filter. Add 1.0 mL of the filtrate to a freshly prepared mixture of 0.1 mL of *alizarin S solution R* and 0.1 mL of *zirconyl nitrate solution R*. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Solution S

Dissolve 1.00 g in *methanol R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)

−0.05° to + 0.05°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *imidazole CRS* (impurity A) in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 30.0 mg of *flutrimazole impurity B CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Mix 2.0 mL of reference solution (a) and 2.0 mL of reference solution (b) and dilute to 50.0 mL with the mobile phase.

Reference solution (d) Dilute 10.0 mL of reference solution (c) to 50.0 mL with the mobile phase.

Reference solution (e) Mix 2.0 mL of the test solution and 10.0 mL of reference solution (c) and dilute to 50.0 mL with the mobile phase.

Reference solution (f) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.2 m, Ø = 4.6 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 0.03 M phosphate buffer solution pH 7.0 R, *acetonitrile R* (40:60 *V/V*).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time 2.5 times the retention time of flutrimazole.

System suitability Reference solution (e):

— *resolution*: minimum 2.0 between the peaks due to impurity A (1st peak) and impurity B (2nd peak); minimum 1.5 between the peaks due to impurity B and flutrimazole (3rd peak);

— *symmetry factors*: maximum 2.0 for the peaks due to impurities A and B.

Limits:

— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.1 per cent);

— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (f) (0.10 per cent);

— *sum of impurities other than B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (f) (0.3 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (f) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.300 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 34.64 mg of $C_{22}H_{16}F_2N_2$.

STORAGE

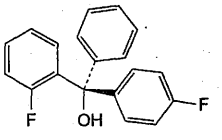
Protected from light.

IMPURITIES

Specified impurities *A, B.*

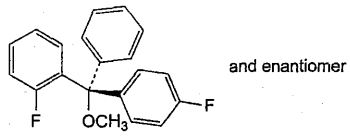
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C.*

A. imidazole,



and enantiomer

B. (*RS*)-(2-fluorophenyl)(4-fluorophenyl)phenylmethanol,

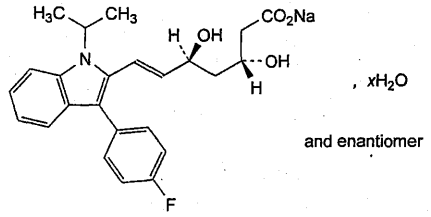


C. (*RS*)-(2-fluorophenyl)(4-fluorophenyl) methoxyphenylmethane.

Ph Eur

Fluvastatin Sodium

(Ph. Eur. monograph 2333)



$C_{24}H_{25}FNNaO_4,xH_2O$ 433.5 *93957-55-2*

(anhydrous substance)Anhydrous fluvastatin sodium

Action and use

HMG Co-A reductase inhibitor; lipid-regulating drug.

Preparations

Fluvastatin Capsules

Fluvastatin Prolonged-release Tablets

Ph Eur

DEFINITION

Sodium (3*RS*,5*SR*,6*E*)-7-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]-3,5-dihydroxyhept-6-enoate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

It may be anhydrous or contain a variable quantity of water.

CHARACTERS

Appearance

White or almost white, or pale yellow or pale reddish-yellow, very hygroscopic, amorphous or crystalline powder.

Solubility

Soluble in water, freely soluble in methanol, practically insoluble in acetonitrile.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluvastatin sodium CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness on a steam bath, protecting the solutions from light, and dry at 105 °C for 30 min. Cool and keep in a desiccator. Record new spectra using the residues.

B. 0.5 mL of solution S (see Tests) gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

pH (*2.2.3*)

8.0 to 10.0 for solution S.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution Dissolve 25 mg of the substance to be examined in 20 mL of mobile phase B and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of *fluvastatin for system suitability CRS* (containing impurities A, B and D) in 1.0 mL of a mixture of equal volumes of mobile phase A and mobile phase B.

Column:

— *size*: *l* = 0.10 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: to 880 mL of *water R* add 20 mL of a 250 g/L solution of *tetramethylammonium hydroxide R* and adjust quickly to pH 7.2 with *phosphoric acid R*; mix with 100 mL of a mixture of 40 volumes of *acetonitrile R* and 60 volumes of *methanol R*;

— *mobile phase B*: to 80 mL of *water R* add 20 mL of a 250 g/L solution of *tetramethylammonium hydroxide R* and adjust quickly to pH 7.2 with *phosphoric acid R*; mix with 900 mL of a mixture of 40 volumes of *acetonitrile R* and 60 volumes of *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	70	30
3 - 23	70 → 10	30 → 90
23 - 28	10	90

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 305 nm and at 365 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *fluvastatin for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and D.

Relative retention With reference to fluvastatin (retention time = about 14 min): impurity A = about 1.05; impurity D = about 1.1; impurity B = about 1.6.

System suitability Reference solution (b) at 305 nm:

— *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to fluvastatin.

Limits:

— *impurity A at 305 nm*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);

— *impurity B at 305 nm*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *impurity D at 365 nm*: not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 305 nm (0.15 per cent);

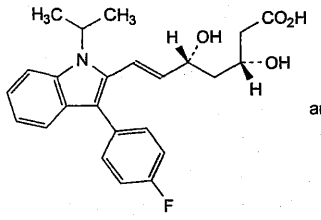
— *unspecified impurities at 305 nm*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities at 305 nm*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit at 305 nm*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
Maximum 12.0 per cent, determined on 0.200 g.

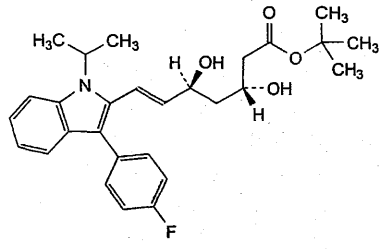
ASSAY
Dissolve 0.325 g in 50 mL of *glacial acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M perchloric acid* is equivalent to 43.35 mg of $C_{24}H_{25}FNNaO_4$.

STORAGE
In an airtight container, protected from light.

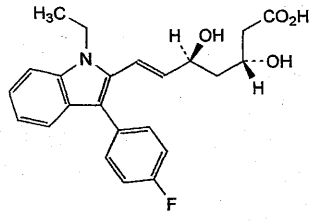
IMPURITIES
Specified impurities *A, B, D.*
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, F, G.*



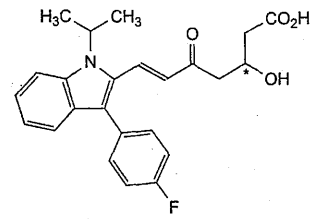
A. (3*RS*,5*RS*,6*E*)-7-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]-3,5-dihydroxyhept-6-enoic acid,



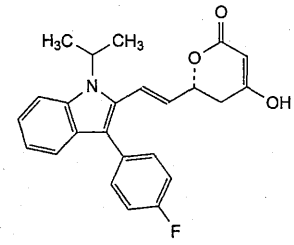
B. 1,1-dimethylethyl (3*R*,5*S*,6*E*)-7-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]-3,5-dihydroxyhept-6-enoate,



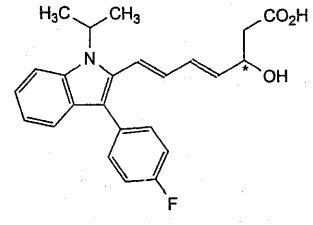
C. (3*R*,5*S*,6*E*)-7-[1-ethyl-3-(4-fluorophenyl)-1*H*-indol-2-yl]-3,5-dihydroxyhept-6-enoic acid,



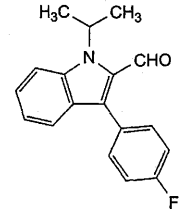
D. (6*E*)-7-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]-3-hydroxy-5-oxohept-6-enoic acid,



E. (6*R*)-6-[(*E*)-2-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]ethenyl]-4-hydroxy-5,6-dihydro-2*H*-pyran-2-one,



F. (4*E*,6*E*)-7-[3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indol-2-yl]-3-hydroxyhepta-4,6-dienoic acid,

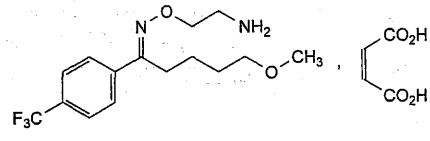


G. 3-(4-fluorophenyl)-1-(1-methylethyl)-1*H*-indole-2-carbaldehyde.

Ph Eur

Fluvoxamine Maleate

(Ph. Eur. monograph 1977)



$C_{19}H_{25}F_3N_2O_6$ 434.4 *61718-82-9*

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Preparation
Fluvoxamine Tablets

Ph Eur

DEFINITION

2-[[[(1*E*)-5-Methoxy-1-[4-(trifluoromethyl)phenyl]pentylidene]amino]oxy]ethanamine (*Z*)-butenedioate.

Content

99.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

The production method must be evaluated to determine the potential for formation of aziridine. Where necessary, a validated test for the substance is carried out or the production method is validated to demonstrate acceptable clearance.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fluvoxamine maleate CRS.*

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the test solution immediately before use.*

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 25 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *fluvoxamine for system suitability CRS* (containing impurities A, B, C and F) in 1.0 mL of the mobile phase.

Reference solution (c) Dissolve 3.0 mg of *fluvoxamine impurity D CRS* in 5 mL of the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 370 volumes of *acetonitrile R1* and 630 volumes of a buffer solution containing 1.1 g/L of *potassium dihydrogen phosphate R* and 1.9 g/L of *sodium pentanesulfonate R* in *water R*, previously adjusted to pH 3.0 with *phosphoric acid R*.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 234 nm.

Injection 20 µL.

Run time 6 times the retention time of fluvoxamine.

Identification of impurities Use the chromatogram supplied with *fluvoxamine for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and F.

Relative retention With reference to fluvoxamine (retention time = about 15 min): maleic acid = about 0.15; impurities F and G = about 0.5; impurity C = about 0.6; impurity B = about 0.8; impurity A = about 2.5; impurity D = about 5.4.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to impurities F and C.

Limits:
- *impurity B*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *impurity C*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
- *sum of impurities F and G*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to maleic acid.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

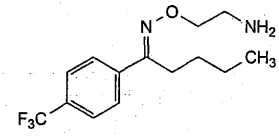
Dissolve 0.350 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 43.44 mg of $C_{19}H_{25}F_3N_2O_6$.

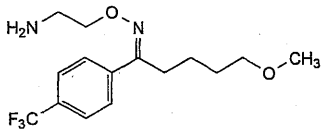
IMPURITIES

Specified impurities *A, B, C, D, F, G.*

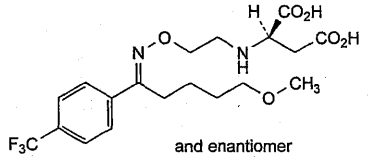
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, I, J.*



A. 2-[[[(1*E*)-1-[4-(trifluoromethyl)phenyl]pentylidene]amino]oxy]ethanamine,

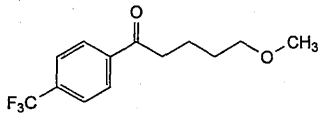


B. 2-[[[(1*Z*)-5-methoxy-1-[4-(trifluoromethyl)phenyl]pentylidene]amino]oxy]ethanamine,

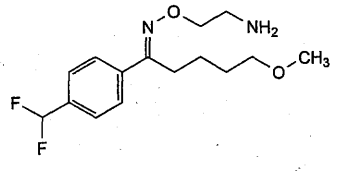


and enantiomer

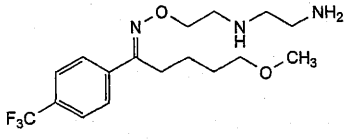
C. (2*RS*)-2-[[2-[[[(1*E*)-5-methoxy-1-[4-(trifluoromethyl)phenyl]pentylidene]amino]oxy]ethyl]amino]butanedioic acid,



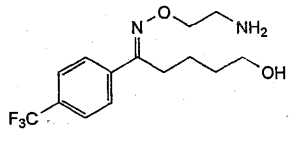
D. 5-methoxy-1-[4-(trifluoromethyl)phenyl]pentan-1-one,



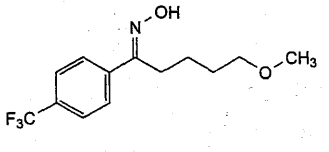
E. 2-[[[(1*E*)-1-[4-(difluoromethyl)phenyl]-5-methoxypentylidene]amino]oxy]ethanamine,



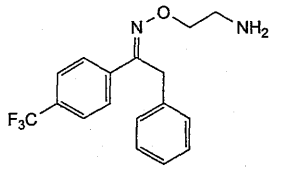
F. *N*-[2-[[[(1*E*)-5-methoxy-1-[4-(trifluoromethyl)phenyl]pentylidene]amino]oxy]ethyl]ethane-1,2-diamine,



G. (5*E*)-5-[(2-aminoethoxy)imino]-5-[4-(trifluoromethyl)phenyl]pentan-1-ol,



I. (*E*)-*N*-[5-methoxy-1-[4-(trifluoromethyl)phenyl]pentylidene]hydroxylamine,



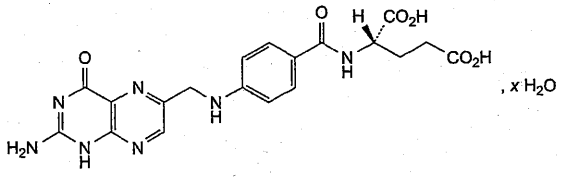
J. 2-[[[(1*E*)-2-phenyl-1-[4-(trifluoromethyl)phenyl]ethylidene]amino]oxy]ethanamine.

Ph Eur

Folic Acid Hydrate

Folic Acid

(Ph. Eur. monograph 0067)



$C_{19}H_{19}N_7O_6,xH_2O$ 441.4 *59-30-3*

(anhydrous substance)Anhydrous folic acid

Action and use

Vitamin B component.

Preparations

Folic Acid Injection

Folic Acid Tablets

Ferrous Fumarate and Folic Acid Tablets

Ph Eur

DEFINITION

(2*S*)-2-[4-[[(2-Amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid hydrate.

Content

96.0 per cent to 102.0 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS

Appearance

Yellowish or orange, crystalline powder.

Solubility

Practically insoluble in water and in most organic solvents. It dissolves in dilute acids and in alkaline solutions.

IDENTIFICATION

First identification: A, B, D.

Second identification: A, C.

A. Specific optical rotation (*2.2.7*): + 18 to + 22 (anhydrous substance).

Dissolve 0.25 g in a 4.2 g/L solution of *sodium hydroxide R* and dilute to 25.0 mL with the same solution.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *folic acid CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in a mixture of 2 volumes of *concentrated ammonia R* and 9 volumes of *methanol R*, and dilute to 100 mL with the same mixture of solvents.

Reference solution Dissolve 50 mg of *folic acid CRS* in a mixture of 2 volumes of *concentrated ammonia R* and 9 volumes of *methanol R*, and dilute to 100 mL with the same mixture of solvents.

Plate *TLC silica gel plate R.*

Mobile phase *concentrated ammonia R, propanol R, ethanol (96 per cent) R* (20:20:60 *V/V/V*).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 365 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, fluorescence and size to the principal spot in the chromatogram obtained with the reference solution.

D. Water (see Tests).

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solution A 28.6 g/L solution of *sodium carbonate R.*

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of solution A and dilute to 50.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of *folic acid CRS* in 2.5 mL of solution A and dilute to 50.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *folic acid for system suitability CRS* (containing impurities C, E, G and H) in 1 mL of solution A and dilute to 25.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 10.0 mg of *folic acid impurity A CRS* in 1 mL of solution A and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (e) Dissolve 4.0 mg of *folic acid impurity D CRS* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (f) Dissolve 5 mg of *folic acid for impurity I identification CRS* in 1 mL of solution A and dilute to 25.0 mL with the mobile phase.

Column:
- *size*: l = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: spherical *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 12 volumes of *methanol R* and 88 volumes of a solution containing 11.16 g/L of *potassium dihydrogen phosphate R* and 5.50 g/L of *dipotassium hydrogen phosphate R.*

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 5 µL of the test solution and reference solutions (b), (c), (d), (e) and (f).

Run time 3.3 times the retention time of folic acid.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peak due to impurity A; use the chromatogram supplied with *folic acid for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C, E, G and H; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity D; use the chromatogram supplied with *folic acid for impurity I identification CRS* and the chromatogram obtained with reference solution (f) to identify the peak due to impurity I.

Relative retention With reference to folic acid (retention time = about 8.5 min): impurity A = about 0.5; impurity C = about 0.9; impurity E = about 1.3; impurity D = about 1.5; impurity I = about 2.15; impurity G = about 2.4; impurity H = about 2.5.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to folic acid and impurity E;
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to folic acid; minimum 1.5, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity H.

Calculation of percentage contents:
- for impurity A, use the concentration of impurity A in reference solution (d);
- for impurity D, use the concentration of impurity D in reference solution (e);
- for impurities other than A and D, use the concentration of folic acid in reference solution (c).

Limits:
- *impurity A*: maximum 0.5 per cent;
- *impurity D*: maximum 0.4 per cent;
- *impurities C, E, G*: for each impurity, maximum 0.3 per cent;
- *impurities H, I*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 1.2 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

5.0 per cent to 8.5 per cent, determined on 0.150 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

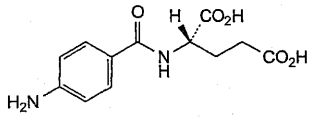
STORAGE

Protected from light, under inert gas.

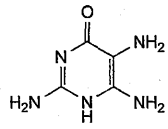
IMPURITIES

Specified impurities *A, C, D, E, G, H, I.*

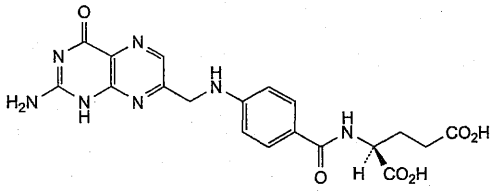
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, F.*



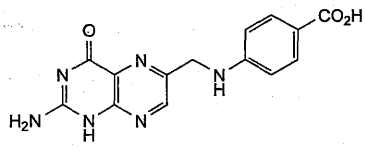
A. (2*S*)-2-(4-aminobenzamido)pentanedioic acid (*N*-(4-aminobenzoyl)-L-glutamic acid),



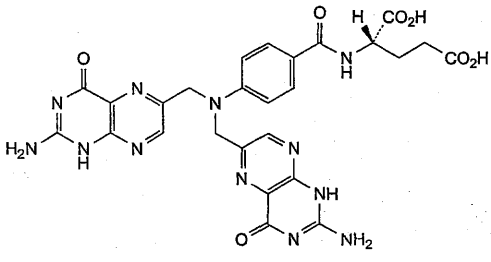
B. 2,5,6-triaminopyrimidin-4(1*H*)-one,



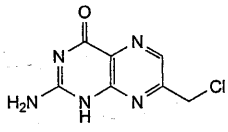
C. (2*S*)-2-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-7-yl)methyl]amino]benzamido]pentanedioic acid (isofolic acid),



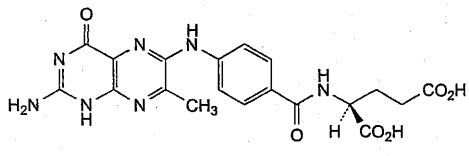
D. 4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzoic acid (pteroic acid),



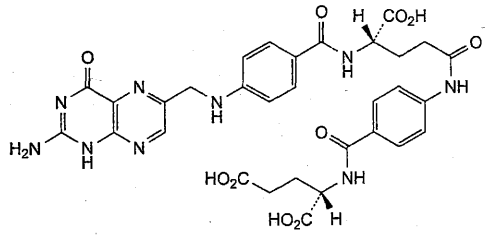
E. (2*S*)-2-[4-[bis[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzamido]pentanedioic acid (6-pterinylfolic acid),



F. 2-amino-7-(chloromethyl)pteridin-4(1*H*)-one,



G. (2*S*)-2-[4-[(2-amino-7-methyl-4-oxo-1,4-dihydropteridin-6-yl)amino]benzamido]pentanedioic acid,



H. (2*S*)-2-[4-[(4*S*)-4-[4-[[(2-amino-4-oxo-1,4-dihydropteridin-6-yl)methyl]amino]benzamido]-4-carboxybutanamido]benzamido]pentanedioic acid,

I. unknown structure.

Ph Eur

Follitropin

(Ph. Eur. monograph 2285)

α-subunit

```
APDVQDCPEC TLQENPFFSQ PGAPILQCMG CCFSRAYPTP   40
LRSKKTMLVQ KNVTSESTCC VAKSYNRVTV MGGFKVENHT   80
ACHCSTCYYH KS                                 92
```

β-subunit

```
NSCELTNITI AIEKEECRFC ISINTTWCAG YCYTRDLVYK  40*
DPARPKIQKT CTFKELVYET VRVPGCAHHA DSLYTYPVAT  80*
QCHCGKCDSD STDCTVRGLG PSYCSFGEMK E          111*
```

glycosylation sites:
Asn-52, Asn-78, Asn-7*, Asn-24*

disulfide bridges:
7-31, 10-60, 28-82, 32-84, 59-87, 3*-51*, 17*-66*, 20*-104*, 28*-82*, 32*-84*, 87*-94*

M_r approx. 30 000 - 40 000

Action and use
Recombinant human follicle stimulating hormone; treatment of female infertility.

Ph Eur

DEFINITION
Freeze-dried preparation of a heterodimeric glycoprotein having the structure of human follicle-stimulating hormone (FSH). It consists of 2 subunits: a 92-amino-acid α-chain common to other glycoprotein hormones and a specific 111-amino-acid β-chain.

Potency
9000 IU to 17 000 IU per milligram of protein.

PRODUCTION
Follitropin is produced in mammalian cells by a method based on recombinant DNA (rDNA) technology.

Follitropin complies with the following requirements.

Host-cell-derived proteins
The limit is approved by the competent authority.

Host-cell- and vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS

Appearance
White or almost white powder.

IDENTIFICATION
A. It complies with the requirements described under Assay.
B. Isoelectric focusing (*2.2.54*).

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of about 2 mg/mL, then desalt and concentrate using a suitably validated procedure. Reconstitute the recovered material in *water R* to obtain a concentration of 5 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in *water R* to obtain a concentration of 5 mg/mL.

Focusing:
- *pH gradient*: a combination of ampholytes and electrode buffers giving a functional separation in the isoelectric point (pI) range of 3.5-5.5 is selected, as defined by the system suitability criteria; where pre-cast gels are employed, proprietary electrode solutions may be used in conjunction; otherwise, suitable dilute mineral or organic acids and bases are employed at pH levels respectively lower and higher than the functional range of the ampholytes;
- *catholyte*: 20.0 g/L solution of *glycine R*;
- *anolyte*: solution containing 3.4 g/L of *aspartic acid R* and 3.6 g/L of *glutamic acid R*, adjusted to pH 2.8-3.8;
- *application*: 10 µL.

Detection: as described in *2.2.54.*

System suitability:
- in the electropherogram obtained with the reference solution, the number of bands seen in the pI region 3.5-5.5 corresponds to that shown in the electropherogram supplied with *follitropin CRS*; the distribution of bands in the pI region 3.5-5.5 is qualitatively similar to that shown in the electropherogram supplied with *follitropin CRS*.

Results Examine the electropherogram obtained with the test solution; identify the bands observed by comparison with the electropherogram obtained with the reference solution; the pattern of bands is qualitatively similar to that seen with the reference solution.

C. Examine the chromatograms obtained in the test for follitropin oligomers.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

D. Peptide mapping (*2.2.55*).

SEPARATION OF THE α- AND β-SUBUNITS Liquid chromatography (*2.2.29*).

Test solution Dissolve the substance to be examined in mobile phase A to obtain a concentration of about 0.4 mg/mL.

Reference solution Dilute *follitropin for peptide mapping and glycan analysis CRS* with mobile phase A to obtain a concentration of about 0.4 mg/mL.

Precolumn:
- *size*: *l* = 0.012 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *trifluoroacetic acid R, water for chromatography R, acetonitrile for chromatography R* (0.9:50:950 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 8	100 → 76	0 → 24
8 - 17	76	24
17 - 36	76 → 70	24 → 30
36 - 41	70 → 25	30 → 75
41 - 46	25	75
46 - 47	25 → 100	75 → 0
47 - 57	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 226 nm.

Injection 800 µL.

Retention time β-subunit = about 14 min; α-subunit = about 30 min.

Collect the fractions containing the α- and β-subunits and freeze-dry them.

REDUCTION, MODIFICATION AND DESALTING OF THE PURIFIED SUBUNITS

Reduction and modification

Solution A Dilute 10 µL of *tributylphosphine R* to 2 mL with *propanol R*. Saturate with nitrogen.

Solution B Dilute 20 µL of *4-vinylpyridine R* to 200 µL with *propanol R*. Saturate with nitrogen.

Test solutions Dissolve each of the α- and β-subunit fractions obtained from the test solution in the previous step in 300 µL of *guanidine-tris(hydroxymethyl)aminomethane-EDTA buffer solution pH 8.5 R* and incubate at 37 °C for 60 min in a thermostatically controlled water-bath. Add 100 µL of solution A, mix and saturate with nitrogen. Incubate at 37 °C for 90 min. Add 10 µL of solution B, mix and saturate with nitrogen. Incubate at 37 °C for 45 min. Add 100 µL of a 10 per cent *V/V* solution of *trifluoroacetic acid R* and mix.

Reference solutions Prepare at the same time and in the same manner as for the test solutions but using the α- and β-subunit fractions obtained from the reference solution in the previous step.

Desalting

Dilute the α- and β-subunit test and reference solutions to 840 µL with mobile phase A.

Column:
- *size*: *l* = 0.02 m, Ø = 4.6 mm;
- *stationary phase*: *butylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *trifluoroacetic acid R, water for chromatography R, acetonitrile for chromatography R* (1:300:700 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	100	0
7 - 27	100 → 0	0 → 100
27 - 27.01	0 → 100	100 → 0
27.01 - 32	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 226 nm.

Injection 800 µL.

For each solution the chromatogram shows a principal peak due to the monovinylpyridine-modified subunit and several minor peaks due to the di- and oligovinylpyridine-modified subunits. Only the fraction containing the monovinylpyridine-modified subunit is used for digestion in the following step.

Retention time *α-subunit solution*: monovinylpyridine-modified α-subunit = about 15 min; *β-subunit solution*: monovinylpyridine-modified β-subunit = about 16 min.

Collect the fractions containing the monovinylpyridine-modified subunits and freeze-dry them.

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Solution C (8 M urea solution). Dissolve 480 g of *urea R* in 600 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent. Add about 3-5 g of mixed-bed resin and stir for about 1 h. Filter through a glass filter before use.

Solution D Dissolve 15.8 g of *ammonium hydrogen carbonate R* and 8.3 g of *sodium edetate R* in 800 mL of *water for chromatography R*. Adjust to pH 7.8 (*2.2.3*) with an 80 g/L solution of *sodium hydroxide R* and dilute to 1000 mL with *water for chromatography R*.

Test solutions Dissolve each of the modified α- and β-subunits obtained from the test solutions in the previous step in 42.5 µL of solution C and incubate at room temperature for 30 min. Add 42.5 µL of solution D and mix. To 42.5 µL of these solutions add 35 µL of a solution containing about 23 mU/µL of endoproteinase Lys-C and mix. Incubate at 37 °C for 4 h, then add 35 µL of the same endoproteinase Lys-C solution and mix. Incubate at 37 °C overnight, then dilute to 420 µL with mobile phase A.

Reference solutions Prepare at the same time and in the same manner as for the test solutions but using the fractions obtained from the reference solutions in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Precolumn:
- *size*: *l* = 0.02 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *trifluoroacetic acid R*, *water for chromatography R*, *acetonitrile R1* (1:300:700 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	100	0
7 - 77	100 → 30	0 → 70
77 - 82	30 → 0	70 → 100
82 - 87	0	100
87 - 92	0 → 100	100 → 0
92 - 107	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 400 µL.

System suitability:

α-subunit:
- the chromatogram obtained with the reference solution is qualitatively similar to the chromatogram of follitropin α-subunit digest supplied with *follitropin for peptide mapping and glycan analysis CRS*; both chromatograms show peaks due to the L4, L6, L3, L5 and L1-2/L1 fragments;
- retention times obtained with the test and reference solutions differ by not more than 5 per cent for fragments L4, L6 and L3, not more than 3 per cent for fragment L5 and not more than 2 per cent for fragments L1-2/L1;

β-subunit:
- the chromatogram obtained with the reference solution is qualitatively similar to the chromatogram of follitropin β-subunit digest supplied with *follitropin for peptide mapping and glycan analysis CRS*; both chromatograms show peaks due to the L5, L7, L6, and L1-4 fragments;
- retention times obtained with the test and reference solutions differ by not more than 5 per cent for fragment L5, not more than 2 per cent for fragments L7 and L6 and not more than 1 per cent for fragments L1-4.

Results For each subunit, the profile of the chromatogram obtained with the test solution is similar to that of the chromatogram obtained with the corresponding reference solution.

E. Glycan analysis (*2.2.59*). Carry out either method A or method B.

METHOD A

PROTEIN DENATURATION

Test solution Dissolve 500 µg of the substance to be examined in 60 µL of *0.05 M phosphate buffer solution pH 7.5 R*. Add 6 µL of a 10 mg/mL solution of *sodium dodecyl sulfate R* and 35 µL of a 1 per cent *V/V* solution of *2-mercaptoethanol R*. Mix using a vortex mixer, centrifuge and incubate at 37 °C for 15 min.

Reference solution Freeze dry a sample of *follitropin for peptide mapping and glycan analysis CRS* that contains 500 µg of follitropin. Dissolve in 60 µL of *0.05 M phosphate buffer solution pH 7.5 R* and continue as for the test solution.

SELECTIVE RELEASE OF THE GLYCANS

Test solution To the test solution obtained in the previous step add 0.75 µL of octylphenyl-polyethylene glycol and mix using a vortex mixer. Add 25 mU of *peptide N-glycosidase F R*, mix using a vortex mixer and centrifuge. Incubate at 37 °C for 24 h. Remove the protein fraction using a suitable, validated procedure. The following method has been found to be appropriate. Add 600 µL of *anhydrous ethanol R*, previously cooled at – 20 °C for 45 min. Mix using a vortex mixer and centrifuge. Precipitate the proteins at – 20 °C for 15 min, then centrifuge at 10 600 *g* at 4 °C for 5 min. Transfer the supernatant to a separate tube and evaporate the ethanol for 15 min. Add 1 mL of *particle-free water R* and resume evaporating until the remaining volume is about 500-800 µL, then freeze-dry.

Label the liberated glycans contained in the sample with 2-aminobenzamide. The procedure employs a combination of reagents optimised and validated for the efficient labelling of glycans, and for the subsequent extraction and recovery of the labelled glycans from the reaction. Recover the sample in 1.5 mL of *particle-free water R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using the reference solution obtained in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Column:
- — *size*: *l* = 0.075 m, Ø = 7.5 mm;
- — *stationary phase*: *weak anion-exchange resin R* (10 µm);
- — *temperature*: 30 °C.

Mobile phase:
- — *mobile phase A*: *acetonitrile R*;
- — *mobile phase B*: *0.5 M ammonium acetate buffer solution pH 4.5 R*; filter through a membrane filter (nominal pore size 0.22 µm);
- — *mobile phase C*: *particle-free water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 5	20	0	80
5 - 21	20	0 → 4	80 → 76
21 - 61	20	4 → 25	76 → 55
61 - 62	20	25 → 50	55 → 30
62 - 71	20	50	30
71 - 72	20	50 → 0	30 → 80
72 - 117	20	0	80

Flow rate 0.4 mL/min.

Detection Fluorimeter at 330 nm for excitation and at 420 nm for emission.

Injection 50 µL.

System suitability Reference solution:
- — the chromatogram obtained is qualitatively similar to the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*;
- — by comparison with the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*, identify the peaks due to neutral, mono-, di-, tri- and tetra-sialylated forms; determine the area of each peak and express it as a percentage of the total; calculate the *Z* number using the following expression:

$$(A_0 \times 0) + (A_1 \times 1) + (A_2 \times 2) + (A_3 \times 3) + (A_4 \times 4)$$

A_0 = peak area percentage due to the neutral form;
A_1 = peak area percentage due to the mono-sialylated form;
A_2 = peak area percentage due to the di-sialylated form;
A_3 = peak area percentage due to the tri-sialylated form;
A_4 = peak area percentage due to the tetra-sialylated form.

The *Z* number obtained for the reference solution is in the range 177-233.

Examine the chromatogram obtained with the test solution and calculate the *Z* number as described above.

Result *Z* = 177-233.

METHOD B

PROTEIN DENATURATION

Solution A To 1.952 g of *2-[N-morpholino]ethanesulfonic acid R* and 57.32 g of *guanidine hydrochloride R*, add 1 mL of a 15.4 g/L solution of *dithiothreitol R*, 10 mL of an 18.61 g/L solution of *sodium edetate R* and 20 mL of *water for chromatography R*. Maintain in a water-bath at about 37 °C for 1 min to dissolve the components. Adjust to pH 8.1 (*2.2.3*) with an 80 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with *water for chromatography R*. Mix.

Solution B Dissolve 37 mg of *iodoacetamide R* in 1 mL of *water for chromatography R* and mix. Protect from light.

Solution C Dissolve 26.7 g of *disodium hydrogen phosphate dihydrate R* and 11.2 g of *sodium edetate R* in 3000 mL of *water for chromatography R* and mix. Adjust to pH 7.5 (*2.2.3*) with a 40 g/L solution of *sodium hydroxide R*.

Test solution Dissolve 1 mg of the substance to be examined in 0.2 mL of solution A and incubate in a water-bath at 37 ± 1 °C for 2 h. Add 20 µL of freshly prepared solution B, mix and incubate at 37 ± 1 °C for a further 2 h, protected from light. Add 10 µL of *2-mercaptoethanol R* and mix. Dialyse against 1000 mL of solution C. Add 200 µL of solution C and mix. Determine the protein content of the solution.

Reference solution (a) To a volume of *follitropin for peptide mapping and glycan analysis CRS* that contains 1 mg of follitropin, add 0.2 mL of solution A. Incubate in a water-bath at 37 ± 1 °C for 2 h. Continue as for the test solution. Determine the protein content of the solution.

Reference solution (b) Prepare at the same time and in the same manner as for the test solution but using fetuin instead of the substance to be examined. Determine the protein content of the solution.

SELECTIVE RELEASE OF THE GLYCANS

Test solution Dilute the test solution obtained in the previous step with solution C to obtain a concentration of 1.1 g/L. Add 1 U of *peptide N-glycosidase F R* to 500 µg of the solution, mix and incubate at 37 ± 1 °C for 24 h. Place the solution in ice. Precipitate the protein and salts with 3 volumes of ice-cold *anhydrous ethanol R* and allow to stand in ice for 10 min. Centrifuge at 16 000 *g* for about 5 min and transfer the supernatant to a separate tube. Add 3 µL of a 1 µg/µL solution of *maltotriose R*, then freeze-dry. Dissolve in 100 µL of *water for chromatography R*.

Reference solution (a) Prepare in the same manner as for the test solution but using the reference solution obtained with *follitropin for peptide mapping and glycan analysis CRS* in the previous step.

Reference solution (b) Prepare in the same manner as for the test solution but using the reference solution obtained with fetuin in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Precolumn:
- — *size*: *l* = 0.05 m, Ø = 4.0 mm;
- — *stationary phase*: *strongly basic anion-exchange resin for chromatography R*;

Column:
- — *size*: *l* = 0.25 m, Ø = 4.0 mm;
- — *stationary phase*: *strongly basic anion-exchange resin for chromatography R*.

Mobile phase:
- — *mobile phase A*: 20 g/L solution of *sodium hydroxide R*; maintain under helium;
- — *mobile phase B*: *water for chromatography R*; maintain under helium;
- — *mobile phase C*: dissolve 41 g of *anhydrous sodium acetate R* in 800 mL of *water for chromatography R*, dilute to 1000 mL with the same solvent, then mix; filter through a membrane filter (nominal pore size 0.45 µm); maintain under helium.

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 0.2	20	80	0
0.2 - 94.0	20	80 → 34	0 → 46
94.0 - 97.0	20	34	46
97.0 - 97.1	20	34 → 80	46 → 0
97.1 - 115.0	20	80	0

Flow rate 1.0 mL/min.

Detection Pulsed amperometric detector or equivalent with a gold indicator electrode, a silver-silver chloride reference electrode, and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V oxidation and −0.80 V reduction potentials, with pulse durations according to the instrument used.

Injection 45 µL.

System suitability:

— the chromatogram obtained with reference solution (b) is qualitatively similar to the chromatogram for fetuin supplied with *follitropin for peptide mapping and glycan analysis CRS*;

— the chromatograms obtained with the test solution and reference solution (a) are qualitatively similar to the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*;

— by comparison with the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*, identify the peaks due to neutral, mono-, di-, tri- and tetra-sialylated forms in the chromatogram obtained with reference solution (b); determine the area of each peak and express it as a percentage of the total; calculate the *Z* number using the following expression:

$$(A_0 \times 0) + (A_1 \times 1) + (A_2 \times 2) + (A_3 \times 3) + (A_4 \times 4)$$

A_0 = peak area percentage due to the neutral form;
A_1 = peak area percentage due to the mono-sialylated form;
A_2 = peak area percentage due to the di-sialylated form;
A_3 = peak area percentage due to the tri-sialylated form;
A_4 = peak area percentage due to the tetra-sialylated form.

The *Z* number obtained for reference solution (b) is in the range 290-325.

Examine the chromatogram obtained with the test solution and calculate the *Z* number as described above.

Result *Z* = 178-274.

TESTS

Follitropin oligomers

Size-exclusion chromatography (*2.2.30*). Use the normalisation procedure.

Solution A Dissolve 118 mg of *sodium dihydrogen phosphate R*, 1.65 g of *disodium hydrogen phosphate dihydrate R* and 30.0 g of *sucrose R* in 40 mL of *water for chromatography R* and dilute to 100.0 mL with the same solvent.

Solution B Dissolve 2.0 mg of *bovine albumin R* in 30 mL of solution A.

Test solution Dissolve the substance to be examined in solution A to obtain a concentration of 0.25 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in solution A to obtain a concentration of 0.5 mg/mL and mix equal volumes of this solution and solution B to obtain a concentration of 0.25 mg/mL.

Column:

— *size*: *l* = 0.3 m, Ø = 7.8 mm;

— *stationary phase*: *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000 (5 µm).

Mobile phase Dissolve 28.4 g of *anhydrous sodium sulfate R* in 2000 mL of *0.1 M phosphate buffer solution pH 6.7 R* and filter through a membrane filter (nominal pore size 0.45 µm).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 100 µL.

Retention time Follitropin = 14-16 min.

System suitability Reference solution:

— *resolution*: minimum 1.2 between the peaks due to bovine albumin and follitropin;

— no peak is detected between 5 min and 16 min in blank injections.

Limit:

— *sum of the peaks with a retention time less than that of the principal peak*: maximum 0.5 per cent.

Free subunits

Polyacrylamide gel electrophoresis (*2.2.31*) under non-reducing conditions.

Gel dimensions 1.5 mm thick.

Resolving gel 12 per cent acrylamide.

Sample buffer *Concentrated SDS-PAGE sample buffer R.*

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of 2 µg/µL. To 55 µL of the solution add 55 µL of the sample buffer. Allow to stand for 4 h at room temperature.

Reference solution (a) Dissolve the contents of a vial of *follitropin CRS* in *water R* to obtain a concentration of 2 µg/µL. To 25 µL of the solution add 25 µL of the sample buffer. To 40 µL of this solution add 180 µL of the sample buffer and 180 µL of *water R*. Allow to stand for 4 h at room temperature, then boil for 5 min.

Reference solution (b) A solution of molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 14.4-94 kDa.

Application:

Well	Solution(s)	Volume (µL)
1	Reference solution (a)	40
2	Reference solution (a)	30
3	Reference solution (a)	20
4	Reference solution (a)	15
5	Reference solution (a)	10
6	Reference solution (a)	5
7	Test solution	50
8	Test solution + reference solution (a)	50 + 25
9	Reference solution (b)	10

Detection By Coomassie staining.

System suitability:

— reference solution (b): the validation criteria are met (*2.2.31*);

— test solution + reference solution (a): the bands corresponding to the follitropin heterodimer and subunits are clearly separated;

— reference solution (a): no bands corresponding to the follitropin heterodimer are seen.

Limit:

— *free subunits*: maximum 3 per cent.

Oxidised follitropin

Liquid chromatography (*2.2.29*).

Solution A Dissolve about 3.3 mg of *2,4-dichlorobenzoic acid R* in 10.0 mL of *ethanol (96 per cent) R*.

Test solution Dissolve the substance to be examined in *water for chromatography R* to obtain a concentration of 300 µg/mL.

Reference solution (a) Dissolve the contents of a vial of *follitropin CRS* in *water for chromatography R* to obtain a concentration of 300 µg/mL.

Reference solution (b) Dilute 0.1 mL of *strong hydrogen peroxide solution R* to 30 mL with *water for chromatography R*. Dissolve the contents of a vial of *follitropin CRS* in this solution to obtain a concentration of 300 µg/mL. Incubate for 30-45 min. Add solution A to obtain a concentration in 2,4-dichlorobenzoic acid of about 17 µg/mL in the total volume and inject immediately.

Column:
- *size*: l = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *0.2 M phosphate buffer solution pH 2.5 R*;
- *mobile phase B*: *water for chromatography R, acetonitrile R1* (40:60 *V/V*);
- *mobile phase C*: *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 8.4	50	25 → 39	25 → 11
8.4 - 8.5	50	39 → 45	11 → 5
8.5 - 15	50	45	5
15 - 15.1	50	45 → 25	5 → 25
15.1 - 25	50	25	25

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 25 µL.

System suitability Reference solution (b):
- the peaks due to the oxidised follitropin α- and β-subunits are separated from the peaks due to the non-oxidised follitropin subunits and from the peak due to 2,4-dichlorobenzoic acid;
- the chromatogram obtained is similar to the chromatogram supplied with *follitropin CRS*.

Calculate the percentage of oxidation of the follitropin subunits using the following expression:

$$\frac{(A_2 + A_4) \times 100}{A_1 + A_2 + A_3 + A_4}$$

A_1 = area of the peak due to the follitropin α-subunit;
A_2 = area of the peaks due to the oxidised follitropin α-subunit;
A_3 = area of the peak due to the follitropin β-subunit;
A_4 = area of the peak due to the oxidised follitropin β-subunit.

Limit:
- *total oxidised forms*: maximum 6 per cent.

Bacterial endotoxins (*2.6.14*)

Less than 0.1 IU per International Unit of follitropin activity, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Protein

Size-exclusion chromatography (*2.2.30*).

Solution A Dissolve 100 mg of *poloxamer 188 R* in 900 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent.

Test solution Dissolve the substance to be examined in solution A to obtain a concentration of about 0.03 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in solution A to obtain a concentration of about 0.03 mg/mL.

Column:
- *size*: l = 0.3 m, Ø = 7.8 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000 (5 µm).

Mobile phase Mix 6.74 mL of *phosphoric acid R*, 14.2 g of *anhydrous sodium sulfate R* and 900 mL of *water for chromatography R*, adjust to pH 6.7 (*2.2.3*) with a 0.5 g/mL solution of *sodium hydroxide R* and dilute to 1000 mL with *water for chromatography R*; filter through a membrane filter (nominal pore size 0.45 µm).

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 100 µL.

System suitability Reference solution:
- *number of theoretical plates*: minimum 1300, calculated for the peak due to follitropin.

Calculate the content of follitropin taking into account the assigned content of *follitropin CRS*.

Potency

The follicle-stimulating activity of follitropin is estimated by comparing under given conditions its effect in enlarging the ovaries of immature rats treated with chorionic gonadotrophin with the same effect of the International Standard preparation of human recombinant follicle-stimulating hormone or of a reference preparation calibrated in International Units. The International Unit of FSH is the activity contained in stated amounts of the International Standard of human recombinant follicle-stimulating hormone. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Use immature female rats of the same strain, 19-28 days old, differing in age by not more than 3 days and having masses such that the difference between the heaviest and the lightest rat is not more than 10 g. Assign the rats at random to 6 equal groups of at least 5 rats. If sets of 6 litter mates are available, assign 1 litter mate from each set to each group and mark according to litter.

Choose 3 doses of the reference preparation and 3 doses of the preparation to be examined such that the smallest dose produces a positive response in some of the rats and the largest dose does not produce a maximal response in all of the rats. Use doses in geometric progression and as an initial approximation total doses of 1.5 IU, 3.0 IU and 6.0 IU may be tried, although the dose will depend on the sensitivity of the rats used, which may vary widely.

Dissolve separately the total quantities of the preparation to be examined and of the reference preparation corresponding to the daily doses to be used in sufficient *phosphate-albumin buffered saline pH 7.2 R* such that the daily dose is

administered in a volume of about 0.5 mL. The buffer solution shall contain in the daily dose not less than 14 IU of chorionic gonadotrophin to ensure complete luteinisation. Add a suitable antimicrobial preservative such as 4 g/L of phenol or 0.02 g/L of thiomersal. Store the solutions at 5 ± 3 °C.

Inject subcutaneously into each rat the daily dose allocated to its group. Repeat the injection of each dose 24 h and 48 h after the 1st injection. About 24 h after the last injection, euthanise the rats and remove the ovaries from each rat. Remove any extraneous fluid and tissue from the ovaries and weigh the 2 combined ovaries of each rat immediately. Calculate the results by the usual statistical methods (for example, *5.3*), using the mass of the 2 combined ovaries as the response. (The precision of the assay may be improved by a suitable correction of the organ mass with reference to the mass of the rat from which it was taken; an analysis of covariance may be used.)

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits (P = 0.95) of the estimated potency are not less than 64 per cent and not more than 156 per cent of the stated potency.

STORAGE

In an airtight container, at a temperature not exceeding – 20 °C.

LABELLING

The label states:
— the potency in International Units per milligram of protein;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Follitropin Concentrated Solution

(Ph. Eur. monograph 2286)

α-subunit
APDVQDCPEC TLQENPFFSQ PGAPILQCMG CCFSRAYPTP 40
LRSKKTMLVQ KNVTSESTCC VAKSYNRVTV MGGFKVENHT 80
ACHCSTCYYH KS 92

β-subunit
NSCELTNITI AIEKEECRFC ISINTTWCAG YCYTRDLVYK 40*
DPARPKIQKT CTFKELVYET VRVPGCAHHA DSLYTYPVAT 80*
QCHCGKCDSD STDCTVRGLG PSYCSFGEMK E 111*

glycosylation sites:
Asn-52, Asn-78, Asn-7*, Asn-24*

disulfide bridges:
7-31, 10-60, 28-82, 32-84, 59-87, 3*-51*, 17*-66*, 20*-104*, 28*-82*, 32*-84*, 87*-94*

M_r approx. 30 000 - 40 000

Action and use

Recombinant human follicle stimulating hormone; treatment of female infertility.

Ph Eur

DEFINITION

Solution of a heterodimeric glycoprotein having the structure of human follicle-stimulating hormone (FSH). It consists of 2 subunits: a 92-amino-acid α-chain common to other glycoprotein hormones and a specific 111-amino-acid β-chain.

Content

0.4 mg to 0.8 mg of protein per millilitre.

Potency

9000 IU to 17 000 IU per milligram of protein.

PRODUCTION

Follitropin is produced in mammalian cells by a method based on recombinant DNA (rDNA) technology.

Follitropin complies with the following requirements.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell- and vector-derived DNA

The limit is approved by the competent authority.

CHARACTERS

Appearance

Clear or slightly turbid, colourless liquid.

IDENTIFICATION

A. It complies with the requirements described under Assay.

B. Isoelectric focusing (*2.2.54*).

Test solution Desalt and concentrate the preparation to be examined using a suitably validated procedure. Reconstitute the recovered material in *water R* to obtain a concentration of 5 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in *water R* to obtain a concentration of 5 mg/mL.

Focusing:
— *pH gradient*: a combination of ampholytes and electrode buffers giving a functional separation in the isoelectric point (pI) range of 3.5-5.5 is selected, as defined by the system suitability criteria; where pre-cast gels are employed, proprietary electrode solutions may be used in conjunction; otherwise, suitable dilute mineral or organic acids and bases are employed at pH levels respectively lower and higher than the functional range of the ampholytes;
— *catholyte*: 20.0 g/L solution of *glycine R*;
— *anolyte*: solution containing 3.4 g/L of *aspartic acid R* and 3.6 g/L of *glutamic acid R*, adjusted to pH 2.8-3.8;
— *application*: 10 µL.

Detection: as described in *2.2.54*.

System suitability:
— in the electropherogram obtained with the reference solution, the number of bands seen in the pI region 3.5-5.5 corresponds to that shown in the electropherogram supplied with *follitropin CRS*; the distribution of bands in the pI region 3.5-5.5 is qualitatively similar to that shown in the electropherogram supplied with *follitropin CRS*.

Results Examine the electropherogram obtained with the test solution; identify the bands observed by comparison with the electropherogram obtained with the reference solution; the pattern of bands is qualitatively similar to that seen with the reference solution.

C. Examine the chromatograms obtained in the test for follitropin oligomers.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

D. Peptide mapping (*2.2.55*).

SEPARATION OF THE α- AND β-SUBUNITS Liquid chromatography (*2.2.29*).

Test solution Dilute the preparation to be examined with mobile phase A to obtain a concentration of about 0.4 mg/mL.

Reference solution Dilute *follitropin for peptide mapping and glycan analysis CRS* with mobile phase A to obtain a concentration of about 0.4 mg/mL

Precolumn:
- — *size*: *l* = 0.012 m, Ø = 4.6 mm;
- — *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm).

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- — *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- — *mobile phase B*: *trifluoroacetic acid R, water for chromatography R, acetonitrile for chromatography R* (0.9:50:950 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	100	0
2 - 8	100 → 76	0 → 24
8 - 17	76	24
17 - 36	76 → 70	24 → 30
36 - 41	70 → 25	30 → 75
41 - 46	25	75
46 - 47	25 → 100	75 → 0
47 - 57	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 226 nm.

Injection 800 µL.

Retention time β-subunit = about 14 min; α-subunit = about 30 min.

Collect the fractions containing the α- and β-subunits and freeze-dry them.

REDUCTION, MODIFICATION AND DESALTING OF THE PURIFIED SUBUNITS

Reduction and modification

Solution A Dilute 10 µL of *tributylphosphine R* to 2 mL with *propanol R*. Saturate with nitrogen.

Solution B Dilute 20 µL of *4-vinylpyridine R* to 200 µL with *propanol R*. Saturate with nitrogen.

Test solutions Dissolve each of the α- and β-subunit fractions obtained from the test solution in the previous step in 300 µL of *guanidine-tris(hydroxymethyl)aminomethane-EDTA buffer solution pH 8.5 R* and incubate at 37 °C for 60 min in a thermostatically controlled water-bath. Add 100 µL of solution A, mix and saturate with nitrogen. Incubate at 37 °C for 90 min. Add 10 µL of solution B, mix and saturate with nitrogen. Incubate at 37 °C for 45 min. Add 100 µL of a 10 per cent *V/V* solution of *trifluoroacetic acid R* and mix.

Reference solutions Prepare at the same time and in the same manner as for the test solutions but using the α- and β-subunit fractions obtained from the reference solution in the previous step.

Desalting

Dilute the α- and β-subunit test and reference solutions to 840 µL with mobile phase A.

Column:
- — *size*: *l* = 0.02 m, Ø = 4.6 mm;
- — *stationary phase*: *butylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- — *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- — *mobile phase B*: *trifluoroacetic acid R, water for chromatography R, acetonitrile for chromatography R* (1:300:700 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	100	0
7 - 27	100 → 0	0 → 100
27 - 27.01	0 → 100	100 → 0
27.01 - 32	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 226 nm.

Injection 800 µL.

For each solution the chromatogram shows a principal peak due to the monovinylpyridine-modified subunit and several minor peaks due to the di- and oligovinylpyridine-modified subunits. Only the fraction containing the monovinylpyridine-modified subunit is used for digestion in the following step.

Retention time *α-subunit solution*: monovinylpyridine-modified α-subunit = about 15 min; *β-subunit solution*: monovinylpyridine-modified β-subunit = about 16 min.

Collect the fractions containing the monovinylpyridine-modified subunits and freeze-dry them.

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Solution C (8 M urea solution). Dissolve 480 g of *urea R* in 600 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent. Add about 3-5 g of mixed-bed resin and stir for about 1 h. Filter through a glass filter before use.

Solution D Dissolve 15.8 g of *ammonium hydrogen carbonate R* and 8.3 g of *sodium edetate R* in 800 mL of *water for chromatography R*. Adjust to pH 7.8 (*2.2.3*) with an 80 g/L solution of *sodium hydroxide R* and dilute to 1000 mL with *water for chromatography R*.

Test solutions Dissolve each of the modified α- and β-subunits obtained from the test solutions in the previous step in 42.5 µL of solution C and incubate at room temperature for 30 min. Add 42.5 µL of solution D and mix. To 42.5 µL of these solutions add 35 µL of a solution containing about 23 mU/µL of endoproteinase Lys-C and mix. Incubate at 37 °C for 4 h, then add 35 µL of the same endoproteinase Lys-C solution and mix. Incubate at 37 °C overnight, then dilute to 420 µL with mobile phase A.

Reference solutions Prepare at the same time and in the same manner as for the test solutions but using the fractions obtained from the reference solutions in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Precolumn:
- — *size*: *l* = 0.02 m, Ø = 4.6 mm;
- — *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- — *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- — *mobile phase B*: *trifluoroacetic acid R*, *water for chromatography R*, *acetonitrile R1* (1:300:700 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 7	100	0
7 - 77	100 → 30	0 → 70
77 - 82	30 → 0	70 → 100
82 - 87	0	100
87 - 92	0 → 100	100 → 0
92 - 107	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 400 µL.

System suitability:

α-subunit:
- — the chromatogram obtained with the reference solution is qualitatively similar to the chromatogram of follitropin α-subunit digest supplied with *follitropin for peptide mapping and glycan analysis CRS*; both chromatograms show peaks due to the L4, L6, L3, L5 and L1-2/L1 fragments;
- — retention times obtained with the test and reference solutions differ by not more than 5 per cent for fragments L4, L6 and L3, not more than 3 per cent for fragment L5 and not more than 2 per cent for fragments L1-2/L1;

β-subunit:
- — the chromatogram obtained with the reference solutions is qualitatively similar to the chromatogram of follitropin β-subunit digest supplied with *follitropin for peptide mapping and glycan analysis CRS*; both chromatograms show peaks due to the L5, L7, L6, and L1-4 fragments;
- — retention times obtained with the test and reference solutions differ by not more than 5 per cent for fragment L5, not more than 2 per cent for fragments L7 and L6 and not more than 1 per cent for fragments L1-4.

Results For each subunit, the profile of the chromatogram obtained with the test solution is similar to that of the chromatogram obtained with the corresponding reference solution.

E. Glycan analysis (*2.2.59*). Carry out either method A or method B.

METHOD A

PROTEIN DENATURATION

Test solution Freeze-dry a sample of the preparation to be examined that contains 500 µg of follitropin. Dissolve in 60 µL of *0.05 M phosphate buffer solution pH 7.5 R*. Add 6 µL of a 10 mg/mL solution of *sodium dodecyl sulfate R* and 35 µL of a 1 per cent *V/V* solution of *2-mercaptoethanol R*. Mix using a vortex mixer, centrifuge and incubate at 37 °C for 15 min.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *follitropin for peptide mapping and glycan analysis CRS* instead of the freeze-dried preparation to be examined.

SELECTIVE RELEASE OF THE GLYCANS

Test solution To the test solution obtained in the previous step add 0.75 µL of octylphenyl-polyethylene glycol and mix using a vortex mixer. Add 25 mU of *peptide N-glycosidase F R*, mix using a vortex mixer and centrifuge. Incubate at 37 °C for 24 h. Remove the protein fraction using a suitable, validated procedure. The following method has been found to be appropriate. Add 600 µL of *anhydrous ethanol R*, previously cooled at −20 °C for 45 min. Mix using a vortex mixer and centrifuge. Precipitate the proteins at −20 °C for 15 min, then centrifuge at 10 600 *g* at 4 °C for 5 min. Transfer the supernatant to a separate tube and evaporate the ethanol for 15 min. Add 1 mL of *particle-free water R* and resume evaporating until the remaining volume is about 500-800 µL, then freeze-dry.

Label the liberated glycans contained in the sample with 2-aminobenzamide. The procedure employs a combination of reagents optimised and validated for the efficient labelling of glycans, and for the subsequent extraction and recovery of the labelled glycans from the reaction. Recover the sample in 1.5 mL of *particle-free water R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using the reference solution obtained in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Column:
- — *size*: *l* = 0.075 m, Ø = 7.5 mm;
- — *stationary phase*: *weak anion-exchange resin R* (10 µm);
- — *temperature*: 30 °C.

Mobile phase:
- — *mobile phase A*: *acetonitrile R*;
- — *mobile phase B*: *0.5 M ammonium acetate buffer solution pH 4.5 R*; filter through a membrane filter (nominal pore size 0.22 µm);
- — *mobile phase C*: *particle-free water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 5	20	0	80
5 - 21	20	0 → 4	80 → 76
21 - 61	20	4 → 25	76 → 55
61 - 62	20	25 → 50	55 → 30
62 - 71	20	50	30
71 - 72	20	50 → 0	30 → 80
72 - 117	20	0	80

Flow rate 0.4 mL/min.

Detection Fluorimeter at 330 nm for excitation and at 420 nm for emission.

Injection 50 µL.

System suitability Reference solution:
- — the chromatogram obtained is qualitatively similar to the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*;
- — by comparison with the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*, identify the peaks due to neutral, mono-, di-, tri- and tetra-sialylated forms; determine the area of each peak and express it as a percentage of the total; calculate the *Z* number using the following expression:

$$(A_0 \times 0) + (A_1 \times 1) + (A_2 \times 2) + (A_3 \times 3) + (A_4 \times 4)$$

A_0 = peak area percentage due to the neutral form;
A_1 = peak area percentage due to the mono-sialylated form;
A_2 = peak area percentage due to the di-sialylated form;
A_3 = peak area percentage due to the tri-sialylated form;
A_4 = peak area percentage due to the tetra-sialylated form.

The Z number obtained for the reference solution is in the range 177-233.

Examine the chromatogram obtained with the test solution and calculate the Z number as described above.

Result Z = 177-233.

METHOD B

PROTEIN DENATURATION

Solution A To 1.952 g of *2-[N-morpholino]ethanesulfonic acid R* and 57.32 g of *guanidine hydrochloride R*, add 1 mL of a 15.4 g/L solution of *dithiothreitol R*, 10 mL of an 18.61 g/L solution of *sodium edetate R* and 20 mL of *water for chromatography R*. Maintain in a water-bath at about 37 °C for 1 min to dissolve the components. Adjust to pH 8.1 (*2.2.3*) with an 80 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with *water for chromatography R*. Mix.

Solution B Dissolve 37 mg of *iodoacetamide R* in 1 mL of *water for chromatography R* and mix. Protect from light.

Solution C Dissolve 26.7 g of *disodium hydrogen phosphate dihydrate R* and 11.2 g of *sodium edetate R* in 3000 mL of *water for chromatography R* and mix. Adjust to pH 7.5 (*2.2.3*) with a 40 g/L solution of *sodium hydroxide R*.

Test solution To a volume of the preparation to be examined that contains 1 mg of follitropin add 0.2 mL of solution A and incubate in a water-bath at 37 ± 1 °C for 2 h. Add 20 µL of freshly prepared solution B, mix and incubate at 37 ± 1 °C for a further 2 h, protected from light. Add 10 µL of *2-mercaptoethanol R* and mix. Dialyse against 1000 mL of solution C. Add 200 µL of solution C and mix. Determine the protein content of the solution.

Reference solution (a) Prepare at the same time and in the same manner as for the test solution but using *follitropin for peptide mapping and glycan analysis CRS* instead of the preparation to be examined. Determine the protein content of the solution.

Reference solution (b) Prepare at the same time and in the same manner as for the test solution but using fetuin instead of the preparation to be examined. Determine the protein content of the solution.

SELECTIVE RELEASE OF THE GLYCANS

Test solution Dilute the test solution obtained in the previous step with solution C to obtain a concentration of 1.1 g/L. Add 1 U of *peptide N-glycosidase F R* to 500 µg of the solution, mix and incubate at 37 ± 1 °C for 24 h. Place the solution in ice. Precipitate the protein and salts with 3 volumes of ice-cold *anhydrous ethanol R* and allow to stand in ice for 10 min. Centrifuge at 16 000 *g* for about 5 min and transfer the supernatant to a separate tube. Add 3 µL of a 1 µg/µL solution of *maltotriose R* then freeze-dry. Dissolve in 100 µL of *water for chromatography R*.

Reference solution (a) Prepare in the same manner as for the test solution but using the reference solution obtained with *follitropin for peptide mapping and glycan analysis CRS* in the previous step.

Reference solution (b) Prepare in the same manner as for the test solution but using the reference solution obtained with fetuin in the previous step.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Precolumn:
- *size*: l = 0.05 m, Ø = 4.0 mm;
- *stationary phase*: *strongly basic anion-exchange resin for chromatography R*.

Column:
- *size*: l = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: *strongly basic anion-exchange resin for chromatography R*.

Mobile phase:
- *mobile phase A*: 20 g/L solution of *sodium hydroxide R*; maintain under helium;
- *mobile phase B*: *water for chromatography R*; maintain under helium;
- *mobile phase C*: dissolve 41 g of *anhydrous sodium acetate R* in 800 mL of *water for chromatography R*, dilute to 1000 mL with the same solvent, then mix; filter through a membrane filter (nominal pore size 0.45 µm); maintain under helium;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 0.2	20	80	0
0.2 - 94.0	20	80 → 34	0 → 46
94.0 - 97.0	20	34	46
97.0 - 97.1	20	34 → 80	46 → 0
97.1 - 115.0	20	80	0

Flow rate 1.0 mL/min.

Detection Pulsed amperometric detector or equivalent with a gold indicator electrode, a silver-silver chloride reference electrode, and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V oxidation and −0.80 V reduction potentials, with pulse durations according to the instrument used.

Injection 45 µL.

System suitability:
- the chromatogram obtained with reference solution (b) is qualitatively similar to the chromatogram for fetuin supplied with *follitropin for peptide mapping and glycan analysis CRS*;
- the chromatograms obtained with the test solution and reference solution (a) are qualitatively similar to the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*;
- by comparison with the chromatogram supplied with *follitropin for peptide mapping and glycan analysis CRS*, identify the peaks due to neutral, mono-, di-, tri- and tetra-sialylated forms in the chromatogram obtained with reference solution (b); determine the area of each peak and express it as a percentage of the total; calculate the Z number using the following expression:

$$(A_0 \times 0) + (A_1 \times 1) + (A_2 \times 2) + (A_3 \times 3) + (A_4 \times 4)$$

A_0 = peak area percentage due to the neutral form;
A_1 = peak area percentage due to the mono-sialylated form;
A_2 = peak area percentage due to the di-sialylated form;
A_3 = peak area percentage due to the tri-sialylated form;
A_4 = peak area percentage due to the tetra-sialylated form.

The Z number obtained for reference solution (b) is in the range 290-325.

Examine the chromatogram obtained with the test solution and calculate the Z number as described above.

Result Z = 178-274.

TESTS

Follitropin oligomers

Size-exclusion chromatography (*2.2.30*). Use the normalisation procedure.

Solution A Dissolve 118 mg of *sodium dihydrogen phosphate R*, 1.65 g of *disodium hydrogen phosphate dihydrate R*, and 30.0 g of *sucrose R* in 40 mL of *water for chromatography R* and dilute to 100.0 mL with the same solvent.

Solution B Dissolve 2.0 mg of *bovine albumin R* in 30 mL of solution A.

Test solution Dilute the preparation to be examined with solution A to obtain a concentration of 0.25 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in solution A to obtain a concentration of 0.5 mg/mL and mix equal volumes of this solution and solution B to obtain a concentration of 0.25 mg/mL.

Column:
- *size*: *l* = 0.3 m, Ø = 7.8 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000 (5 µm).

Mobile phase Dissolve 28.4 g of *anhydrous sodium sulfate R* in 2000 mL of *0.1 M phosphate buffer solution pH 6.7 R* and filter through a membrane filter (nominal pore size 0.45 µm).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 100 µL.

Retention time Follitropin = 14-16 min.

System suitability Reference solution:
- *resolution*: minimum 1.2 between the peaks due to bovine albumin and follitropin;
- no peak is detected between 5 min and 16 min in blank injections.

Limit:
- *sum of the peaks with a retention time less than that of the principal peak*: maximum 0.5 per cent.

Free subunits

Polyacrylamide gel electrophoresis (*2.2.31*) under non-reducing conditions.

Gel dimensions 1.5 mm thick.

Resolving gel 12 per cent acrylamide.

Sample buffer *Concentrated SDS-PAGE sample buffer R.*

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 2 µg/µL. To 55 µL of the solution add 55 µL of the sample buffer. Allow to stand for 4 h at room temperature.

Reference solution (a) Dissolve the contents of a vial of *follitropin CRS* in *water R* to obtain a concentration of 2 µg/µL. To 25 µL of the solution add 25 µL of the sample buffer. To 40 µL of this solution add 180 µL of the sample buffer and 180 µL of *water R*. Allow to stand for 4 h at room temperature, then boil for 5 min.

Reference solution (b) A solution of molecular mass markers suitable for calibrating SDS-polyacrylamide gels in the range of 14.4-94 kDa.

Application:

Well	Solution(s)	Volume (µL)
1	Reference solution (a)	40
2	Reference solution (a)	30
3	Reference solution (a)	20
4	Reference solution (a)	15
5	Reference solution (a)	10
6	Reference solution (a)	5
7	Test solution	50
8	Test solution + reference solution (a)	50 + 25
9	Reference solution (b)	10

Detection By Coomassie staining.

System suitability:
- reference solution (b): the validation criteria are met (*2.2.31*);
- test solution + reference solution (a): the bands corresponding to the follitropin heterodimer and subunits are clearly separated;
- reference solution (a): no bands corresponding to the follitropin heterodimer are seen.

Limit:
- *free subunits*: maximum 3 per cent.

Oxidised follitropin

Liquid chromatography (*2.2.29*).

Solution A Dissolve about 3.3 mg of *2,4-dichlorobenzoic acid R* in 10.0 mL of *ethanol (96 per cent) R.*

Test solution Dilute the preparation to be examined in *water for chromatography R* to obtain a concentration of 300 µg/mL.

Reference solution (a) Dissolve the contents of a vial of *follitropin CRS* in *water for chromatography R* to obtain a concentration of 300 µg/mL.

Reference solution (b) Dilute 0.1 mL of *strong hydrogen peroxide solution R* to 30 mL with *water for chromatography R.* Dissolve the contents of a vial of *follitropin CRS* in this solution to obtain a concentration of 300 µg/mL. Incubate for 30-45 min. Add solution A to obtain a concentration in 2,4-dichlorobenzoic acid of about 17 µg/mL in the total volume and inject immediately.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *0.2 M phosphate buffer solution pH 2.5 R*;
- *mobile phase B*: *water for chromatography R, acetonitrile R1* (40:60 *V/V*);
- *mobile phase C*: *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)
0 - 8.4	50	25 → 39	25 → 11
8.4 - 8.5	50	39 → 45	11 → 5
8.5 - 15	50	45	5
15 - 15.1	50	45 → 25	5 → 25
15.1 - 25	50	25	25

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 25 µL.

System suitability Reference solution (b):
- the peaks due to the oxidised follitropin α- and β-subunits are separated from the peaks due to the non-oxidised

follitropin subunits and from the peak due to 2,4-dichlorobenzoic acid;
— the chromatogram obtained is similar to the chromatogram supplied with *follitropin CRS*.

Calculate the percentage of oxidation of the follitropin subunits using the following expression:

$$\frac{(A_2 + A_4) \times 100}{A_1 + A_2 + A_3 + A_4}$$

A_1 = area of the peak due to the follitropin α-subunit;
A_2 = area of the peaks due to the oxidised follitropin α-subunit;
A_3 = area of the peak due to the follitropin β-subunit;
A_4 = area of the peak due to the oxidised follitropin β-subunit.

Limit:
— *total oxidised forms*: maximum 6 per cent.

Bacterial endotoxins (*2.6.14*)
Less than 0.1 IU per International Unit of follitropin activity, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Protein
Size-exclusion chromatography (*2.2.30*).

Solution A Dissolve 100 mg of *poloxamer 188 R* in 900 mL of *water for chromatography R* and dilute to 1000 mL with the same solvent.

Test solution Dilute the preparation to be examined with solution A to obtain a concentration of about 0.03 mg/mL.

Reference solution Dissolve the contents of a vial of *follitropin CRS* in solution A to obtain a concentration of about 0.03 mg/mL.

Column:
— *size*: l = 0.3 m, Ø = 7.8 mm;
— *stationary phase*: *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000 (5 µm).

Mobile phase Mix 6.74 mL of *phosphoric acid R*, 14.2 g of *anhydrous sodium sulfate R* and 900 mL of *water for chromatography R*, adjust to pH 6.7 (*2.2.3*) with a 0.5 g/mL solution of *sodium hydroxide R* and dilute to 1000 mL with *water for chromatography R*; filter through a membrane filter (nominal pore size 0.45 µm).

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 100 µL.

System suitability Reference solution:
— *number of theoretical plates*: minimum 1300, calculated for the peak due to follitropin.

Calculate the content of follitropin taking into account the assigned content of *follitropin CRS*.

Potency
The follicle-stimulating activity of follitropin is estimated by comparing under given conditions its effect in enlarging the ovaries of immature rats treated with chorionic gonadotrophin with the same effect of the International Standard preparation of human recombinant follicle-stimulating hormone or of a reference preparation calibrated in International Units. The International Unit of FSH is the activity contained in stated amounts of the International Standard of human recombinant follicle-stimulating hormone. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Use immature female rats of the same strain, 19-28 days old, differing in age by not more than 3 days and having masses such that the difference between the heaviest and the lightest rat is not more than 10 g. Assign the rats at random to 6 equal groups of at least 5 rats. If sets of 6 litter mates are available, assign 1 litter mate from each set to each group and mark according to litter.

Choose 3 doses of the reference preparation and 3 doses of the preparation to be examined such that the smallest dose produces a positive response in some of the rats and the largest dose does not produce a maximal response in all of the rats. Use doses in geometric progression and as an initial approximation total doses of 1.5 IU, 3.0 IU and 6.0 IU may be tried, although the dose will depend on the sensitivity of the rats used, which may vary widely.

Dilute and dissolve respectively the total quantities of the preparation to be examined and of the reference preparation corresponding to the daily doses to be used in sufficient *phosphate-albumin buffered saline pH 7.2 R* such that the daily dose is administered in a volume of about 0.5 mL.
The buffer solution shall contain in the daily dose not less than 14 IU of chorionic gonadotrophin to ensure complete luteinisation. Add a suitable antimicrobial preservative such as 4 g/L of phenol or 0.02 g/L of thiomersal. Store the solutions at 5 ± 3 °C.

Inject subcutaneously into each rat the daily dose allocated to its group. Repeat the injection of each dose 24 h and 48 h after the 1st injection. About 24 h after the last injection, euthanise the rats and remove the ovaries from each rat. Remove any extraneous fluid and tissue from the ovaries and weigh the 2 combined ovaries of each rat immediately. Calculate the results by the usual statistical methods (for example, *5.3*), using the mass of the 2 combined ovaries as the response. (The precision of the assay may be improved by a suitable correction of the organ mass with reference to the mass of the rat from which it was taken; an analysis of covariance may be used.)

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits (P = 0.95) of the estimated potency are not less than 64 per cent and not more than 156 per cent of the stated potency.

STORAGE

In an airtight container, at a temperature not exceeding −20 °C.

LABELLING

The label states:
— the content of protein in milligrams per millilitre;
— the potency in International Units per milligram of protein;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Formaldehyde Solution

Formalin

(Formaldehyde Solution (35 per cent), Ph. Eur. monograph 0826)

NOTE: The name Formalin as a synonym for Formaldehyde Solution may be used freely in many countries, including the United Kingdom, but in other countries exclusive proprietary rights in this name are claimed.

50-00-0

Action and use
When suitably diluted, used in the treatment of warts.

Ph Eur

DEFINITION

Content
34.5 per cent *m/m* to 38.0 per cent *m/m* of formaldehyde (CH_2O; M_r 30.03).

It contains methanol as stabiliser.

CHARACTERS

Appearance
Clear, colourless liquid.

Solubility
Miscible with water and with ethanol (96 per cent).

It may be cloudy after storage.

IDENTIFICATION

A. Dilute 1 mL of solution S (see Tests) to 10 mL with *water R*. To 0.05 mL of the solution add 1 mL of a 15 g/L solution of *chromotropic acid, sodium salt R*, 2 mL of *water R* and 8 mL of *sulfuric acid R*. A violet-blue or violet-red colour develops within 5 min.

B. To 0.1 mL of solution S add 10 mL of *water R*. Add 2 mL of a 10 g/L solution of *phenylhydrazine hydrochloride R*, prepared immediately before use, 1 mL of *potassium ferricyanide solution R* and 5 mL of *hydrochloric acid R*. An intense red colour is formed.

C. Mix 0.5 mL with 2 mL of *water R* and 2 mL of *silver nitrate solution R2* in a test-tube. Add *dilute ammonia R2* until slightly alkaline. Heat on a water-bath. A grey precipitate or a silver mirror is formed.

D. It complies with the limits of the assay.

TESTS

Solution S
Dilute 10 mL, filtered if necessary, to 50 mL with *carbon dioxide-free water R*.

Appearance of solution
Solution S is colourless (*2.2.2, Method II*).

Acidity
To 10 mL of solution S add 1 mL of *phenolphthalein solution R*. Not more than 0.4 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to red.

Methanol
Gas chromatography (*2.2.28*).

Internal standard solution Dilute 10 mL of *ethanol R1* to 100 mL with *water R*.

Test solution To 10.0 mL of the solution to be examined add 10.0 mL of the internal standard solution and dilute to 100.0 mL with *water R*.

Reference solution To 1.0 mL of *methanol R* add 10.0 mL of the internal standard solution and dilute to 100.0 mL with *water R*.

Column:
— *material*: glass,
— *size*: *l* = 1.5-2.0 m, Ø = 2-4 mm,
— *stationary phase*: *ethylvinylbenzene-divinylbenzene copolymer R* (150-180 µm).

Carrier gas *nitrogen for chromatography R*.

Flow rate 30-40 mL/min.

Temperature:
— *column*: 120 °C,
— *injection port and detector*: 150 °C.

Detection Flame ionisation.

Injection 1 µL of the test solution and the reference solution.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to methanol and ethanol.

Limit:
— *methanol*: 9.0 per cent *V/V* to 15.0 per cent *V/V*.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Into a 100 mL volumetric flask containing 2.5 mL of *water R* and 1 mL of *dilute sodium hydroxide solution R*, introduce 1.000 g of the solution to be examined, shake and dilute to 100.0 mL with *water R*. To 10.0 mL of the solution add 30.0 mL of *0.05 M iodine*. Mix and add 10 mL of *dilute sodium hydroxide solution R*. After 15 min, add 25 mL of *dilute sulfuric acid R* and 2 mL of *starch solution R*. Titrate with *0.1 M sodium thiosulfate*.

1 mL of *0.05 M iodine* is equivalent to 1.501 mg of CH_2O.

STORAGE

Protected from light, at a temperature of 15 °C to 25 °C.

Ph Eur

Formic Acid

(Ph. Eur. monograph 2809)

CH_2O_2 46.03 *64-18-6*

Ph Eur

DEFINITION

Content
98.0 per cent *m/m* to 100.5 per cent *m/m*.

CHARACTERS

Appearance
Clear, colourless, volatile liquid.

Solubility
Miscible with water, with ethanol (96 per cent) and with methylene chloride.

IDENTIFICATION

First identification: A, B.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Films.

Comparison *formic acid CRS*.

B. A 100 g/L solution is strongly acid (*2.2.4*).

C. To 2.0 mL of *dilute sodium hydroxide solution R* add 0.05 mL of *phenolphthalein solution R*. Mix, and add dropwise the substance to be examined until the solution becomes colourless. To 1 mL of this solution add 0.5 mL of *silver nitrate solution R2* and mix. Initially no change occurs. Slowly (quicker upon heating), the solution turns greyish and turbid due to the precipitation of metallic silver.

D. To the remainder of the neutralised solution obtained in identification test C, add 0.5 mL of *ferric chloride solution R1*. The solution turns orange-red or brownish-red.

TESTS

Solution S

Extract the residue obtained in the test for residue on evaporation by heating with 2 quantities, each of 15 mL, of *distilled water R*. After cooling, dilute the combined extracts to 50.0 mL with *distilled water R*.

Appearance

The substance to be examined is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Relative density (*2.2.5*)

1.217 to 1.223.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dilute 0.500 g of the substance to be examined to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 0.500 g of *anhydrous acetic acid R* (impurity A) to 100.0 mL with the mobile phase. Mix 2.0 mL of the solution and 1.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography compatible with 100 per cent aqueous mobile phases R* (4 µm).

Mobile phase 2.72 g/L solution of *potassium dihydrogen phosphate R* adjusted to pH 2.9 with *dilute phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time 3 times the retention time of formic acid.

Relative retention With reference to formic acid (retention time = about 3 min): impurity A = about 1.5.

System suitability Reference solution (a):
- *resolution*: minimum 5.0 between the peaks due to formic acid and impurity A.

Limits:
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (*2.4.4*)

Maximum 10 ppm.

Dilute 12.5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)

Maximum 50 ppm.

Dilute 7.5 mL of solution S to 15 mL with *distilled water R*.

Sulfites

Maximum 300 ppm.

To 0.5 mL add 12.0 mL of a 42 g/L solution of *sodium hydroxide R* in *carbon dioxide-free water R* and mix. Upon addition of 0.5 mL of *iodine solution R1* and 0.2 mL of *starch solution R* the solution remains blue.

Residue on evaporation

Maximum 0.01 per cent.

Condensing the vapour where possible, evaporate 20.0 g to dryness on a water-bath and dry at 105 °C. The residue weighs a maximum of 2.0 mg. The residue is used for the preparation of solution S.

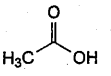
ASSAY

Weigh accurately a conical flask with a ground-glass stopper containing 20 mL of *water R*. Add 1.0 mL of the substance to be examined and weigh again accurately. Titrate with *1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *1 M sodium hydroxide* is equivalent to 46.03 mg of CH_2O_2.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A.

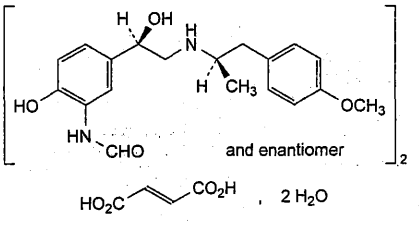


A. acetic acid.

Ph Eur

Formoterol Fumarate Dihydrate

(Ph. Eur. monograph 1724)



$C_{42}H_{52}N_4O_{12},2H_2O$ 841

Action and use

$Beta_2$-adrenoceptor agonist; bronchodilator.

Ph Eur

DEFINITION

N-[2-Hydroxy-5-[(1*RS*)-1-hydroxy-2-[[(1*RS*)-2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl] formamide (*E*)-butenedioate dihydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white or slightly yellow powder.

Solubility

Slightly soluble in water, soluble in methanol, slightly soluble in 2-propanol, practically insoluble in acetonitrile.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *formoterol fumarate dihydrate CRS.*

TESTS

pH (*2.2.3*)

5.5 to 6.5.

Dissolve 20 mg in *carbon dioxide-free water R* while heating to about 40 °C, allow to cool and dilute to 20 mL with the same solvent.

Optical rotation (*2.2.7*)

−0.10° to + 0.10°.

Dissolve 0.25 g in *methanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solution A Dissolve 6.10 g of *sodium dihydrogen phosphate monohydrate R* and 1.03 g of *disodium hydrogen phosphate dihydrate R* in *water R* and dilute to 1000 mL with the same solvent. The pH is 6.0 ± 0.1.

Solvent mixture *acetonitrile R*, solution A (16:84 *V/V*).

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture. *Inject within 4 h of preparation, or within 24 h if stored protected from light at 4 °C.*

Reference solution (a) Dissolve 5 mg of *formoterol fumarate for system suitability CRS* (containing impurities A, B, C, D, E, F and G) in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 25.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: spherical *octylsilyl silica gel for chromatography R3* (5 µm) with a pore size of 8 nm.

Mobile phase:

— *mobile phase A*: *acetonitrile R1*;

— *mobile phase B*: dissolve 3.73 g of *sodium dihydrogen phosphate monohydrate R* and 0.35 g of *phosphoric acid R* in *water R* and dilute to 1000 mL with the same solvent; the pH is 3.1 ± 0.1;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	16	84
10 - 37	16 → 70	84 → 30

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL; inject the solvent mixture until a repeatable profile is obtained.

Identification of impurities Use the chromatogram obtained with reference solution (a) and the chromatogram supplied with *formoterol fumarate for system suitability CRS* to identify the peaks.

Relative retention With reference to formoterol (retention time = about 12 min): impurity G = about 0.4; impurity A = about 0.5; impurity B = about 0.7; impurity C = about 1.2; impurity D = about 1.3; impurity E = about 1.8; impurity F = about 2.0; impurity H = about 2.2.

System suitability Reference solution (a):

— *resolution*: minimum 1.5 between the peaks due to impurity G and impurity A.

— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to formoterol.

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity A by 1.75;

— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurities B, C, D, F*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurity E*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity I

Liquid chromatography (*2.2.29*).

Test solution Dissolve 5.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent. Sonicate if necessary.

Reference solution (a) Dissolve 5.0 mg of *formoterol for impurity I identification CRS* in *water R* and dilute to 50.0 mL with the same solvent. Sonicate if necessary.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with *water R*. Dilute 1.0 mL of this solution to 25.0 mL with *water R*.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: octadecyl vinyl polymer for chromatography R.

Mobile phase Mix 12 volumes of *acetonitrile R1* with 88 volumes of a 5.3 g/L solution of *tripotassium phosphate trihydrate R* previously adjusted to pH 12.0 ± 0.1 with a 280 g/L solution of *potassium hydroxide R* or *phosphoric acid R*.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 µL.

Elution order Formoterol, impurity I.

System suitability Reference solution (a):

— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to formoterol.

Limit:

— *impurity I*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent).

Water (*2.5.12*)

4.0 per cent to 5.0 per cent, determined on 0.100 g.

ASSAY

Dissolve 0.350 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 40.24 mg of $C_{42}H_{52}N_4O_{12}$.

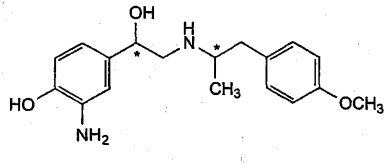
STORAGE

Protected from light.

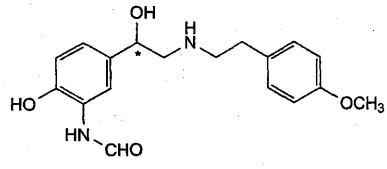
IMPURITIES

Specified impurities *A, B, C, D, E, F, I.*

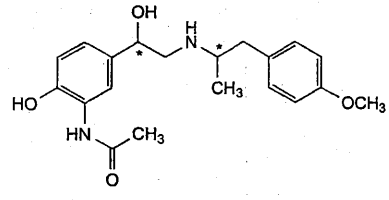
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, H.*



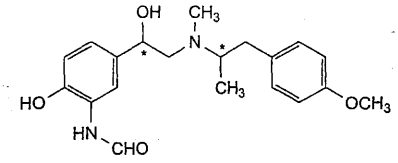
A. 1-(3-amino-4-hydroxyphenyl)-2-[[2-(4-methoxyphenyl)-1-methylethyl]amino]ethanol,



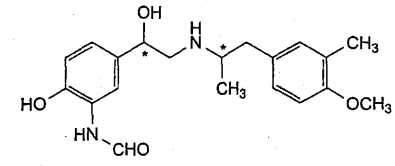
B. *N*-[2-hydroxy-5-[(1*RS*)-1-hydroxy-2-[[2-(4-methoxyphenyl)ethyl]amino]ethyl]phenyl]formamide,



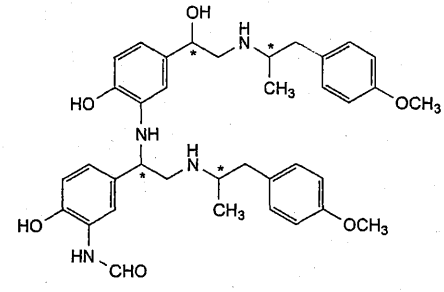
C. *N*-[2-hydroxy-5-[1-hydroxy-2-[[2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl]acetamide,



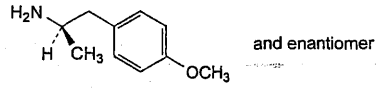
D. *N*-[2-hydroxy-5-[1-hydroxy-2-[methyl[2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl]formamide,



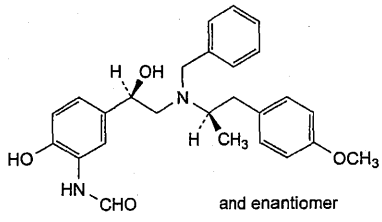
E. *N*-[2-hydroxy-5-[1-hydroxy-2-[[2-(4-methoxy-3-methylphenyl)-1-methylethyl]amino]ethyl]phenyl]formamide,



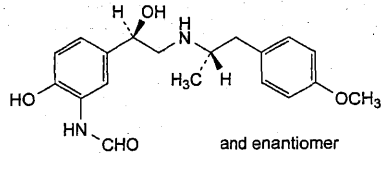
F. *N*-[2-hydroxy-5-[1-[[2-hydroxy-5-[1-hydroxy-2-[[2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl]amino]-2-[[2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl]formamide,



G. (2*RS*)-1-(4-methoxyphenyl)propan-2-amine,



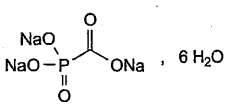
H. *N*-[5-[(1*RS*)-2-[benzyl[(1*RS*)-2-(4-methoxyphenyl)-1-methylethyl]amino]-1-hydroxyethyl]-2-hydroxyphenyl]formamide (monobenzyl analogue),



I. *N*-[2-hydroxy-5-[(1*RS*)-1-hydroxy-2-[[(1*SR*)-2-(4-methoxyphenyl)-1-methylethyl]amino]ethyl]phenyl]formamide (diastereoisomer).

Foscarnet Sodium

(Foscarnet Sodium Hexahydrate, Ph. Eur. monograph 1520)



$CNa_3O_5P,6H_2O$ 300.0 *34156-56-4*

Action and use
Antiviral (cytomegalovirus).

Preparation
Foscarnet Infusion

Ph Eur

DEFINITION
Trisodium phosphonatoformate hexahydrate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *foscarnet sodium hexahydrate CRS.*
B. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is not more opalescent than reference suspension I (*2.2.1*) and is colourless (*2.2.2, Method II*).

pH (*2.2.3*)
9.0 to 11.0 for solution S.

Impurity D
Gas chromatography (*2.2.28*).
Test solution Dissolve 0.250 g of the substance to be examined in 9.0 mL of a 6 g/L solution of *glacial acetic acid R* using a magnetic stirrer. Add 1.0 mL of *anhydrous ethanol R* and mix.
Reference solution Dissolve 25.0 mg of *foscarnet impurity D CRS* in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with *anhydrous ethanol R.*
Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.31 mm;
— *stationary phase*: *poly(dimethyl) (diphenyl) (divinyl)siloxane R* (film thickness 0.5 µm).
Carrier gas *helium for chromatography R.*
Split ratio 1:20.
Temperature:

	Time (min)	Temperature (°C)
Column	0 - 8	100 → 180
Injection port		200
Detector		250

Detection Flame ionisation.
Injection 3 µL
Limit:
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (0.1 per cent).

Related substances
Liquid chromatography (*2.2.29*).
Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 5 mg of *foscarnet impurity B CRS* in the mobile phase, add 2.0 mL of the test solution and dilute to 50.0 mL with the mobile phase.
Reference solution (c) Dissolve the contents of a vial of *foscarnet impurity mixture CRS* (impurities A and C) in 1.0 mL of mobile phase.
Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).
Mobile phase Dissolve 3.22 g of *sodium sulfate decahydrate R* in *water R*, add 3 mL of *glacial acetic acid R* and 6 mL of a 44.61 g/L solution of *sodium pyrophosphate R* and dilute to 1000 mL with *water R* (solution A); dissolve 3.22 g of *sodium sulfate decahydrate R* in *water R*, add 6.8 g of *sodium acetate R* and 6 mL of a 44.61 g/L solution of *sodium pyrophosphate R* and dilute to 1000 mL with *water R* (solution B). Mix about 700 mL of solution A and about 300 mL of solution B to obtain a solution of pH 4.4. To 1000 mL of this solution, add 0.25 g of *tetrahexylammonium hydrogen sulfate R* and 100 mL of *methanol R.*
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 230 nm.
Injection 40 µL.
Run time 2.5 times the retention time of foscarnet.
Identification of impurities Use the chromatogram supplied with *foscarnet impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and C; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.
Relative retention With reference to foscarnet (retention time = about 5 min): impurity A = about 0.7; impurity B = about 1.5; impurity C = about 2.0.
System suitability Reference solution (b):
— *resolution*: minimum 7.0 between the peaks due to foscarnet and impurity B.
Limits:
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.04 per cent); disregard any peak with a relative retention time less than 0.6.

Phosphate and phosphite
Liquid chromatography (*2.2.29*).

Test solution Dissolve 60.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 28 mg of *sodium dihydrogen phosphate monohydrate R* in *water R* and dilute to 100 mL with the same solvent.

Reference solution (b) Dissolve 43 mg of *sodium phosphite pentahydrate R* in *water R* and dilute to 100 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b) to 25 mL with *water R*.

Reference solution (d) Dilute 3 mL of reference solution (a) and 3 mL of reference solution (b) to 25 mL with *water R*.

Column:
— *size*: *l* = 0.05 m, Ø = 4.6 mm;
— *stationary phase*: *anion-exchange resin R*.

Mobile phase Dissolve 0.102 g of *potassium hydrogen phthalate R* in *water R*, add 2.5 mL of 1 M nitric acid and dilute to 1000 mL with *water R*.

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 290 nm (indirect detection).

Injection 20 µL of the test solution and reference solutions (c) and (d).

System suitability Reference solution (d):
— *resolution*: minimum 2.0 between the peaks due to phosphate (1st peak) and phosphite;
— *signal-to-noise ratio*: minimum 10 for the principal peak.

Limits:
— *phosphate*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *phosphite*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent).

Loss on drying (*2.2.32*)
35.0 per cent to 37.0 per cent, determined on 1.000 g by drying in an oven at 150 °C.

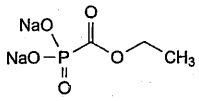
Bacterial endotoxins (*2.6.14*)
Less than 83.3 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Dissolve 0.200 g in 50 mL of *water R*. Titrate with *0.05 M sulfuric acid*, determining the end-point potentiometrically (*2.2.20*) at the 1st point of inflexion.

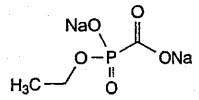
1 mL of *0.05 M sulfuric acid* is equivalent to 19.20 mg of CNa_3O_5P.

STORAGE
Protected from light.

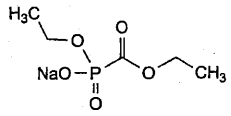
IMPURITIES
Specified impurities *A, B, C, D.*



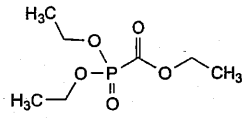
A. disodium (ethoxycarbonyl)phosphonate,



B. disodium (ethoxyoxydophosphanyl)formate,



C. ethyl sodium (ethoxycarbonyl)phosphonate,

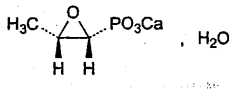


D. ethyl (diethoxyphosphoryl)formate.

Ph Eur

Fosfomycin Calcium

(Ph. Eur. monograph 1328)



$C_3H_5CaO_4P,H_2O$ 194.1 *26469-67-0*

Action and use
Phosphonic acid derivative; antibacterial.

Ph Eur

DEFINITION
Calcium (2*R*,3*S*)-(3-methyloxiran-2-yl)phosphonate monohydrate.

Substance produced by certain strains of *Streptomyces fradiae* or obtained by any other means.

Content
95.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Slightly soluble in water, practically insoluble in acetone, in methanol and in methylene chloride.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fosfomycin calcium CRS.*

B. Dissolve about 0.1 g in 3 mL of a 25 per cent *V/V* solution of *perchloric acid R*. Add 1 mL of *0.1 M sodium periodate* and heat on a water-bath for 30 min. Allow to cool and add 50 mL of *water R*. Neutralise with a saturated solution of *sodium hydrogen carbonate R* and add 1 mL of a freshly prepared 400 g/L solution of *potassium iodide R*. Prepare a blank at the same time and in the same manner. The test solution remains colourless and the blank is orange.

C. To about 8 mg add 2 mL of *water R*, 1 mL of *perchloric acid R* and 2 mL of *0.1 M sodium periodate*. Heat on a water-bath for 10 min and add, without cooling, 1 mL of *ammonium molybdate solution R5* and 1 mL of *aminohydroxynaphthalenesulfonic acid solution R*. Allow to stand for 30 min. A blue colour develops.

D. It gives reaction (a) of calcium (*2.3.1*).

TESTS

pH (*2.2.3*)
8.1 to 9.6.

Dissolve 20 mg in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−11.0 to −13.0 (anhydrous substance), determined at 405 nm using a mercury lamp.

Dissolve 2.5 g in a 125 g/L solution of *sodium edetate R* previously adjusted to pH 8.5 with *strong sodium hydroxide solution R*, and dilute to 50.0 mL with the same solution.

Impurity A
Maximum 1.5 per cent.

In a glass-stoppered flask, dissolve 0.200 g in 100.0 mL of *water R*. Add 50 mL of 0.5 M phthalate buffer solution pH 6.4 R and 5.0 mL of *0.005 M sodium periodate*, close and shake. Allow to stand protected from light for 90 min. Add 10 mL of a freshly prepared 400 g/L solution of *potassium iodide R*, close and shake for 2 min. Titrate with *0.0025 M sodium arsenite* until the yellow colour almost disappears. Add 2 mL of *starch solution R* and slowly continue the titration until the colour is completely discharged. Carry out a blank test under the same conditions.

Calculate the percentage content of $C_3H_7CaO_5P$ using the following expression:

$$\frac{(n_1 - n_2) \times c \times 97 \times 100}{m(100 - H)} \times 100$$

m	=	mass of the substance to be examined, in milligrams;
n_1	=	volume of *0.0025 M sodium arsenite* used in the blank titration, in millilitres;
n_2	=	volume of *0.0025 M sodium arsenite* used in the titration of the test solution, in millilitres;
c	=	molarity of the sodium arsenite solution;
H	=	percentage content of water.

Chlorides (*2.4.4*)
Maximum 0.2 per cent.

Dissolve 0.500 g in *water R*, add 2 mL of *nitric acid R* and dilute to 50 mL with the same acid. To 2.5 mL of this solution add 12.5 mL of *water R*.

Water (*2.5.12*)
8.5 per cent to 11.5 per cent, determined on 0.250 g. Use as the solvent a mixture of 1 volume of *pyridine R* and 3 volumes of *ethylene glycol R*.

ASSAY

In a glass-stoppered flask, dissolve 0.120 g in 20.0 mL of *0.1 M sodium periodate*. Add 5 mL of a 50 per cent *V/V* solution of *perchloric acid R* and shake. Heat in a water-bath at 37 °C for 105 min. Add 50 mL of *water R* and immediately adjust to pH 6.4 with a saturated solution of *sodium hydrogen carbonate R*. Add 10 mL of a freshly prepared 400 g/L solution of *potassium iodide R*, close and allow to stand for 2 min. Titrate with *0.1 M sodium arsenite* until the yellow colour almost disappears. Add 2 mL of *starch solution R* and slowly continue the titration until the colour is completely discharged. Carry out a blank test under the same conditions.

Calculate the percentage content of $C_3H_5CaO_4P$ using the following expression:

$$\frac{(n_1 - n_2) \times c \times 88 \times 100}{m(100 - H)} \times (100 - G)$$

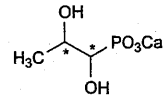
m	=	mass of the substance to be examined, in milligrams;
n_1	=	volume of *0.1 M sodium arsenite* used in the blank titration, in millilitres;
n_2	=	volume of *0.1 M sodium arsenite* used in the titration of the test solution, in millilitres;
c	=	molarity of the sodium arsenite solution;
G	=	percentage content of impurity A;
H	=	percentage content of water.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities *A*.

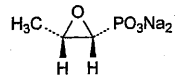


A. calcium (1,2-dihydroxypropyl)phosphonate.

Ph Eur

Fosfomycin Sodium

(Ph. Eur. monograph 1329)



$C_3H_5Na_2O_4P$ 182.0 *26016-99-9*

Action and use
Phosphonic acid derivative; antibacterial.

Ph Eur

DEFINITION

Disodium (2*R*,3*S*)-(3-methyloxiran-2-yl)phosphonate.

Substance produced by certain strains of *Streptomyces fradiae* or obtained by any other means.

Content
95.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, very hygroscopic powder.

Solubility
Very soluble in water, sparingly soluble in methanol, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs of *potassium bromide R.*

Comparison *Ph. Eur. reference spectrum of fosfomycin sodium.*

B. Dissolve about 0.1 g in 3 mL of a 25 per cent *V/V* solution of *perchloric acid R.* Add 1 mL of *0.1 M sodium periodate* and heat on a water-bath for 30 min. Allow to cool and add 50 mL of *water R.* Neutralise with a saturated solution of *sodium hydrogen carbonate R* and add 1 mL of a freshly prepared 400 g/L solution of *potassium iodide R.* Prepare a blank at the same time and in the same manner. The test solution remains colourless and the blank is orange.

C. To about 8 mg add 2 mL of *water R,* 1 mL of *perchloric acid R* and 2 mL of *0.1 M sodium periodate.* Heat on a water-bath for 10 min and add, without cooling, 1 mL of *ammonium molybdate solution R5* and 1 mL of *aminohydroxynaphthalenesulfonic acid solution R.* Allow to stand for 30 min. A blue colour develops.

D. It gives reaction (a) of sodium (*2.3.1*).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_9 (*2.2.2, Method II*).

pH (*2.2.3*)

9.0 to 10.5.

Dilute 10 mL of solution S to 20 mL with *carbon dioxide-free water R.*

Specific optical rotation (*2.2.7*)

−13.0 to −15.0 (anhydrous substance), determined at 405 nm using a mercury lamp.

Dissolve 2.5 g in *water R* and dilute to 50.0 mL with the same solvent.

Impurity A

Maximum 1.0 per cent.

In a glass-stoppered flask, dissolve 0.200 g in 100.0 mL of *water R.* Add 50 mL of 0.5 M phthalate buffer solution pH 6.4 R and 5.0 mL of *0.005 M sodium periodate,* close and shake. Allow to stand protected from light for 90 min. Add 10 mL of a freshly prepared 400 g/L solution of *potassium iodide R,* close and shake for 2 min. Titrate with *0.0025 M sodium arsenite* until the yellow colour almost disappears. Add 2 mL of *starch solution R* and slowly continue the titration until the colour is completely discharged. Carry out a blank test under the same conditions.

Calculate the percentage content of $C_3H_7Na_2O_5P$ using the following expression:

$$\frac{(n_1 - n_2) \times c \times 100 \times 100}{m(100 - H)} \times 100$$

m	=	mass of the substance to be examined, in milligrams;
n_1	=	volume of *0.0025 M sodium arsenite* used in the blank titration, in millilitres;
n_2	=	volume of *0.0025 M sodium arsenite* used in the titration of the test solution, in millilitres;
c	=	molarity of the sodium arsenite solution;
H	=	percentage content of water.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 0.50 g. Use as the solvent a mixture of 1 volume of *pyridine R* and 3 volumes of *ethylene glycol R.*

Bacterial endotoxins (*2.6.14*)

Less than 0.083 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

In a glass-stoppered flask, dissolve 0.120 g in 20.0 mL of *0.1 M sodium periodate.* Add 5 mL of a 50 per cent *V/V* solution of *perchloric acid R* and shake. Heat in a water-bath at 37 °C for 105 min. Add 50 mL of *water R* and immediately adjust to pH 6.4 with a saturated solution of *sodium hydrogen carbonate R.* Add 10 mL of a freshly prepared 400 g/L solution of *potassium iodide R,* close and allow to stand for 2 min. Titrate with *0.1 M sodium arsenite* until the yellow colour almost disappears. Add 2 mL of *starch solution R* and slowly continue the titration until the colour is completely discharged. Carry out a blank test under the same conditions.

Calculate the percentage content of $C_3H_5Na_2O_4P$ using the following expression:

$$\frac{(n_1 - n_2) \times c \times 91 \times 100}{m(100 - H)} \times (100 - G)$$

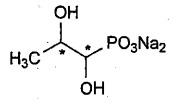
m	=	mass of the substance to be examined, in milligrams;
n_1	=	volume of *0.1 M sodium arsenite* used in the blank titration, in millilitres;
n_2	=	volume of *0.1 M sodium arsenite* used in the titration of the test solution, in millilitres;
c	=	molarity of the sodium arsenite solution;
G	=	percentage content of impurity A;
H	=	percentage content of water.

STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

Specified impurities *A.*

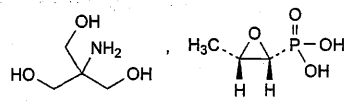


A. disodium (1,2-dihydroxypropyl)phosphonate.

Ph Eur

Fosfomycin Trometamol

(Ph. Eur. monograph 1425)



$C_7H_{18}NO_7P$ 259.2 *78964-85-9*

Action and use

Phosphonic acid derivative; antibacterial.

Ph Eur

DEFINITION

2-Amino-2-(hydroxymethyl)propane-1,3-diol hydrogen (2*R*,3*S*)-(3-methyloxiran-2-yl)phosphonate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Very soluble in water, slightly soluble in ethanol (96 per cent) and in methanol, practically insoluble in acetone.

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fosfomycin trometamol CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of *fosfomycin trometamol CRS* in *water R* and dilute to 10 mL with the same solvent.

Plate *cellulose for chromatography R* as the coating substance.

Mobile phase *concentrated ammonia R, water R, 2-propanol R* (10:20:70 *V/V/V*).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In a current of warm air.

Detection Expose to iodine vapour until the spots appear.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. To about 15 mg add 2 mL of *water R*, 1 mL of *perchloric acid R* and 2 mL of *0.1 M sodium periodate*. Heat on a water-bath for 10 min and add, without cooling, 1 mL of *ammonium molybdate solution R5* and 1 mL of *aminohydroxynaphthalenesulfonic acid solution R*. Allow to stand for 30 min. A blue colour develops.

TESTS

Solution S

Dissolve 1.00 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

pH (*2.2.3*)

3.5 to 5.5 for solution S.

Specific optical rotation (*2.2.7*)

−13.5 to −12.5 (anhydrous substance), determined on solution S at 365 nm using a mercury lamp.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.600 g of the substance to be examined in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (a) Dissolve 0.600 g of *fosfomycin trometamol CRS* in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 3.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Wet 0.3 g of the substance to be examined with 60 µL of *water R* and heat in an oven at 60 °C for 24 h. Dissolve the residue in the mobile phase and dilute to 20.0 mL with the mobile phase (solution A). Dissolve 0.6 g of the substance to be examined in solution A and dilute to 5.0 mL with the same solution (*in situ* degradation to obtain impurities A, B, C and D).

Column:

— *size*: l = 0.25 m, Ø = 4.6 mm,

— *stationary phase*: *aminopropylsilyl silica gel for chromatography R* (5 µm).

Mobile phase 10.89 g/L solution of *potassium dihydrogen phosphate R* in *water for chromatography R*.

Flow rate 1.0 mL/min.

Detection Differential refractometer at 35 °C.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of fosfomycin.

Relative retention With reference to fosfomycin (retention time = about 9 min): trometamol (2 peaks) = about 0.3; impurity B = about 0.48; impurity C = about 0.54; impurity A = about 0.88; impurity D = about 1.27.

System suitability Reference solution (c):

— *resolution*: minimum 1.5 between the peaks due to impurity A and fosfomycin,

— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity B.

Limits:

— *impurities A, B*: for each impurity, not more than the area of the peak due to fosfomycin in the chromatogram obtained with reference solution (b) (0.3 per cent),

— *impurities C, D*: for each impurity, not more than 0.33 times the area of the peak due to fosfomycin in the chromatogram obtained with reference solution (b) (0.1 per cent),

— *unspecified impurities*: for each impurity, not more than 0.33 times the area of the peak due to fosfomycin in the chromatogram obtained with reference solution (b) (0.1 per cent),

— *total*: not more than 1.67 times the area of the peak due to fosfomycin in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.17 times the area of the peak due to fosfomycin in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the 2 peaks due to trometamol.

Phosphates

Maximum 500 ppm.

Dissolve 0.1 g in 3 mL of *dilute nitric acid R* and dilute to 10 mL with *water R*. To 5 mL of this solution add 5 mL of *water R* and 5 mL of *molybdovanadic reagent R*. Shake vigorously. After 5 min, any colour in the test solution is not more intense than that in a standard prepared at the same time in the same manner, using 5 mL of *phosphate standard solution (5 ppm PO_4) R*.

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 0.500 g.

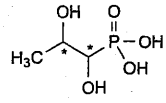
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 5 µL of the test solution and reference solution (a).

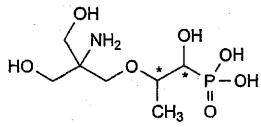
Calculate the percentage content of $C_7H_{18}NO_7P$ from the areas of the peaks due to fosfomycin and the declared content of *fosfomycin trometamol CRS*.

STORAGE
In an airtight container.

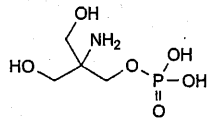
IMPURITIES
Specified impurities *A, B, C, D.*



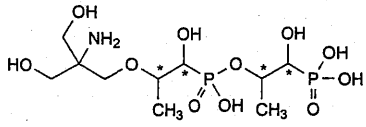
A. (1,2-dihydroxypropyl)phosphonic acid,



B. [2-[2-amino-3-hydroxy-2-(hydroxymethyl)propoxy]-1-hydroxypropyl]phosphonic acid,



C. 2-amino-3-hydroxy-2-(hydroxymethyl)propyl dihydrogen phosphate (trometamol phosphoric acid monoester),

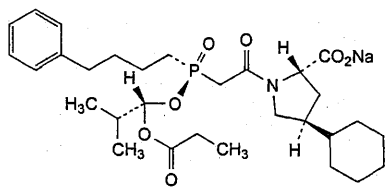


D. [2-[[[2-[2-amino-3-hydroxy-2-(hydroxymethyl)propoxy]-1-hydroxypropyl]hydroxyphosphoryl]oxy]-1-hydroxypropyl] phosphonic acid (trometamoyloxy fosfomycin dimer).

Ph Eur

Fosinopril Sodium

(Ph. Eur. monograph 1751)



$C_{30}H_{45}NNaO_7P$ 585.7 *88889-14-9*

Action and use
Angiotensin converting enzyme inhibitor.

Preparation
Fosinopril Tablets

Ph Eur

DEFINITION
Sodium (2*S*,4*S*)-4-cyclohexyl-1-[[(*R*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl] acetyl]pyrrolidine-2-carboxylate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in anhydrous ethanol, practically insoluble in hexane.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Specific optical rotation (*2.2.7*): −6.7 to −4.7 (anhydrous substance).

Dissolve 0.500 g in *methanol R* and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fosinopril sodium CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in a 2 per cent *V/V* solution of *water R* in *methanol R*, evaporate to dryness and record new spectra using the residues.

C. It gives reaction (a) of sodium (*2.3.1*).

TESTS
Related substances
A. Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Sonicate until dissolution is complete.

Reference solution (a) Dissolve 2 mg of the substance to be examined, 2 mg of *fosinopril impurity A CRS*, 2 mg of *fosinopril impurity B CRS*, 2 mg of *fosinopril impurity I CRS* and 2 mg of *fosinopril impurity K CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 10.0 mL with the mobile phase.

Column:
- *size*: *l* = 0.15 m, Ø = 3.9 mm;
- *stationary phase*: *silica gel for chromatography R* (5 µm);
- *temperature*: 33 °C.

Mobile phase *phosphoric acid R, water R, acetonitrile R1* (0.05:0.35:100 *V/V/V*).

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Run time 4 times the retention time of fosinopril.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, I and K.

Relative retention With reference to fosinopril (retention time = about 5 min): impurity K = about 0.3;

impurity I = about 0.5; impurities B, E and H = about 0.7; impurity A = about 2.0.

System suitability:
- — *resolution*: minimum 2.0 between the peaks due to impurity B and fosinopril in the chromatogram obtained with reference solution (a);
- — *signal-to-noise ratio*: minimum 40 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:
- — *correction factor*: for the calculation of content, multiply the peak area of impurity I by 1.3;
- — *sum of impurities B, E and H*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- — *impurity A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- — *impurities I, K*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- — *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent).

B. Impurities C and D. Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Sonicate until dissolution is complete.

Reference solution (a) Dissolve 5 mg of the substance to be examined and 5.0 mg of *fosinopril impurity C CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of *fosinopril impurity D CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (5 µm);
- — *temperature*: 45 °C.

Mobile phase *phosphoric acid R, water R, acetonitrile R1* (0.2:1.5:400 *V/V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Run time Twice the retention time of fosinopril.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to fosinopril (retention time = about 10 min): impurity C = about 1.2; impurity D = about 1.3.

System suitability Reference solution (a):
- — *resolution*: minimum 1.5 between the peaks due to fosinopril and impurity C.

Limits:
- — *impurity C*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- — *impurity D*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent).

C. Impurities E and F. Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *fosinopril impurity mixture CRS* (containing impurities E and F) in 1.0 mL of reference solution (a).

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm);
- — *temperature*: 45 °C.

Mobile phase 0.2 per cent *V/V* solution of *phosphoric acid R, acetonitrile R1* (44:56 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 3 times the retention time of fosinopril.

Identification of impurities Use the chromatogram supplied with *fosinopril impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities E and F.

Relative retention With reference to fosinopril (retention time = about 8 min): impurity E = about 0.8; impurity F = about 0.9.

System suitability Reference solution (c):
- — *resolution*: minimum 1.5 between the peaks due to impurity F and fosinopril.

Limits:
- — *correction factor*: for the calculation of content, multiply the peak area of impurity E by 0.7;
- — *impurity F*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- — *impurity E*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent).

2-Ethylhexanoic acid (*2.4.28*)
Maximum 0.2 per cent *m/m*.

Water (*2.5.12*)
Maximum 0.2 per cent, determined on 1.00 g.

ASSAY

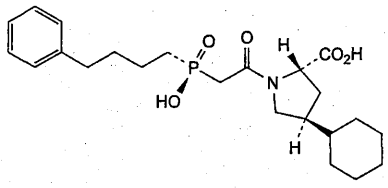
Dissolve 0.450 g in 50 mL of *water R*. Titrate with *0.1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M hydrochloric acid* is equivalent to 58.57 mg of $C_{30}H_{45}NNaO_7P$.

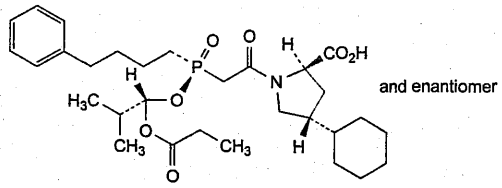
IMPURITIES

Specified impurities *A, B, C, D, E, F, H, I, K.*

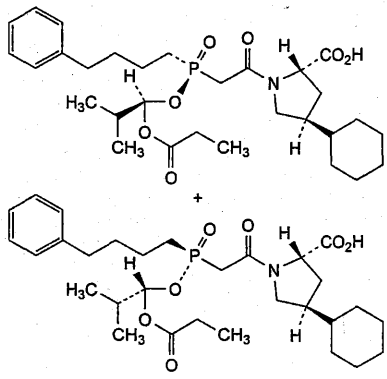
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) N.



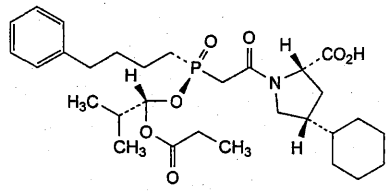
A. (2*S*,4*S*)-4-cyclohexyl-1-[[(*R*)-hydroxy(4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



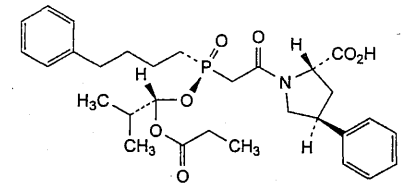
B. (2*RS*,4*RS*)-4-cyclohexyl-1-[[(*RS*)-[(1*SR*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



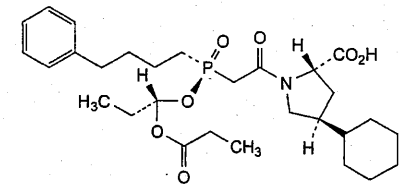
C. mixture of (2*S*,4*S*)-4-cyclohexyl-1-[[(*S*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid and (2*S*,4*S*)-4-cyclohexyl-1-[[(*R*)-[(1*R*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



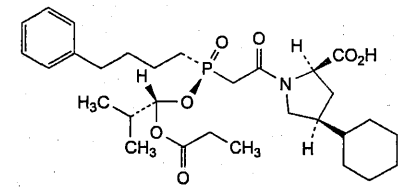
D. (2*S*,4*R*)-4-cyclohexyl-1-[[(*R*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



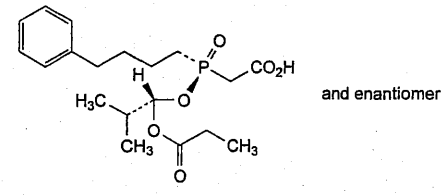
E. (2*S*,4*S*)-1-[[(*R*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]-4-phenylpyrrolidine-2-carboxylic acid (phenylfosinopril),



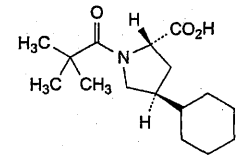
F. (2*S*,4*S*)-4-cyclohexyl-1-[[(*R*)-(4-phenylbutyl)[(1*S*)-1-(1-oxopropoxy)propoxy]phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



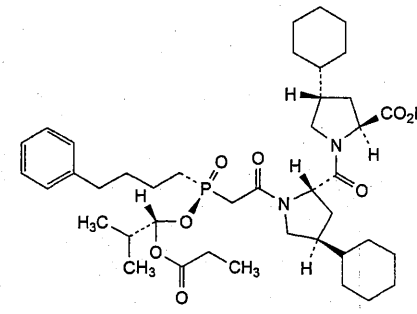
H. (2*R*,4*S*)-4-cyclohexyl-1-[[(*R*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidine-2-carboxylic acid,



I. [(*RS*)-[(1*SR*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetic acid,



K. (2*S*,4*S*)-4-cyclohexyl-1-(2,2-dimethyl-1-oxopropyl)pyrrolidine-2-carboxylic acid,



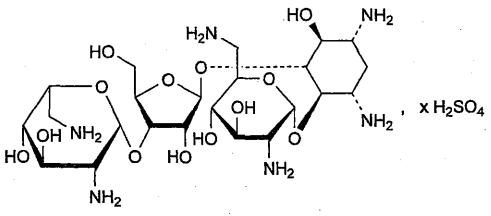
N. (2*S*,4*S*)-4-cyclohexyl-1-[[(2*S*,4*S*)-4-cyclohexyl-1-[[(*R*)-[(1*S*)-2-methyl-1-(1-oxopropoxy)propoxy](4-phenylbutyl)phosphoryl]acetyl]pyrrolidin-2-yl]carbonyl]pyrrolidine-2-carboxylic acid.

Ph Eur

Framycetin Sulfate

Framycetin Sulphate

(Ph. Eur. monograph 0180)



$C_{23}H_{46}N_6O_{13},xH_2SO_4$ 615 (base) *4146-30-9*

Action and use
Antibacterial.

Ph Eur

DEFINITION

Sulfate of 2-deoxy-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-5-*O*-[3-*O*-(2,6-diamino-2,6-dideoxy-β-L-idopyranosyl)-β-D-ribofuranosyl]-D-streptamine (neomycin B).

Substance produced by the growth of selected strains of *Streptomyces fradiae* or *Streptomyces decaris* or obtained by any other means.

Content
Minimum of 630 IU/mg (dried substance).

CHARACTERS

Appearance
White or yellowish-white powder, hygroscopic.

Solubility
Freely soluble in water, very slightly soluble in alcohol, practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for related substances.

Results:
- — the retention time of the principal peak in the chromatogram obtained with the test solution is approximately the same as that of the principal peak in the chromatogram obtained with reference solution (a),
- — it complies with the limit given for impurity C.

B. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

pH (*2.2.3*)
6.0 to 7.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 52.5 to + 55.5 (dried substance).

Dissolve 1.00 g in *water R* and dilute to 10.0 mL with the same solvent

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve the contents of a vial of *framycetin sulfate CRS* in the mobile phase and dilute with the mobile phase to obtain a solution containing 0.5 mg/mL.

Reference solution (b) Dilute 3.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve the contents of a vial of *neamine CRS* (corresponding to 0.5 mg) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (e) Dissolve 10 mg of *neomycin sulfate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm,
- — *stationary phase*: *base-deactivated octadecylsilyl silica gel for chromatography R* (5 µm),
- — *temperature*: 25 °C.

Mobile phase Mix 20.0 mL of *trifluoroacetic acid R*, 6.0 mL of *carbonate-free sodium hydroxide solution R* and 500 mL of *water R*, allow to equilibrate, dilute to 1000 mL with *water R* and degas.

Flow rate 0.7 mL/min.

Post-column solution *carbonate-free sodium hydroxide solution R* diluted 1 in 25 previously degassed, which is added pulse-less to the column effluent using a 375 µL polymeric mixing coil.

Flow rate 0.5 mL/min.

Detection Pulsed amperometric detector with a gold working electrode, a silver-silver chloride reference electrode and a stainless steel auxiliary electrode which is the cell body, held at respectively 0.00 V detection, + 0.80 V oxidation and −0.60 V reduction potentials, with pulse durations according to the instrument used.

Injection 10 µL.

Run time 1.5 times the retention time of neomycin B.

Relative retention With reference to neomycin B (retention time = about 10 min): impurity A = about 0.65; impurity C = about 0.9; impurity G = about 1.1.

System suitability:
- — *resolution*: minimum 2.0 between the peaks due to impurity C and to neomycin B in the chromatogram obtained with reference solution (e); if necessary, adjust the volume of the carbonate-free sodium hydroxide solution in the mobile phase,
- — *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:
- — *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) and taking into account the declared content of *neamine CRS* (1.0 per cent),
- — *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent),
- — *total of other impurities*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent),
- — *disregard limit*: area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent).

Sulfate
27.0 per cent to 31.0 per cent (dried substance).

Dissolve 0.250 g in 100 mL of *water R* and adjust the solution to pH 11 using *concentrated ammonia R*.
Add 10.0 mL of *0.1 M barium chloride* and about 0.5 mg of *phthalein purple R*. Titrate with *0.1 M sodium edetate* adding 50 mL of *alcohol R* when the colour of the solution begins to change and continuing the titration until the violet-blue colour disappears.

1 mL of *0.1 M barium chloride* is equivalent to 9.606 mg of SO_4.

Loss on drying (*2.2.32*)
Maximum 8.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.7 kPa for 3 h.

Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.0 g.

Sterility (*2.6.1*)
If intended for introduction into body cavities without a further appropriate sterilisation procedure, it complies with the test for sterility.

Bacterial endotoxins (*2.6.14, Method D*)
Less than 1.3 IU/mg if intended for introduction into body cavities without a further appropriate procedure for the removal of bacterial endotoxins.

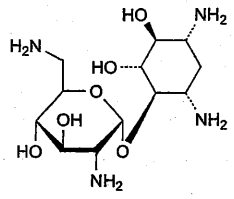
ASSAY

Carry out the microbiological assay of antibiotics (*2.7.2*).
Use *framycetin sulfate CRS* as the reference substance.

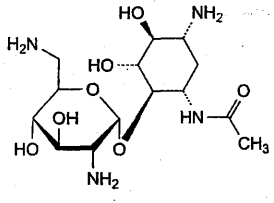
STORAGE

In an airtight container, protected from light. If the substance is intended for introduction into body cavities, store in a sterile, tamper-proof container.

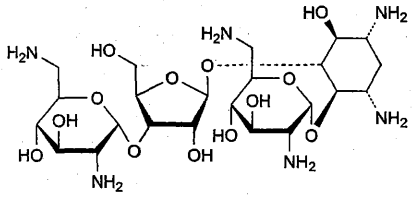
IMPURITIES



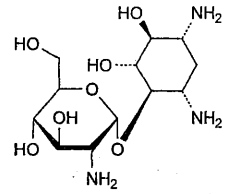
A. 2-deoxy-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-D-streptamine (neamine or neomycin A-LP),



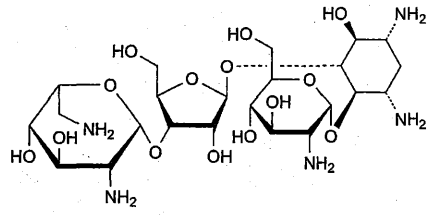
B. 3-*N*-acetyl-2-deoxy-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-D-streptamine (3-acetylneamine),



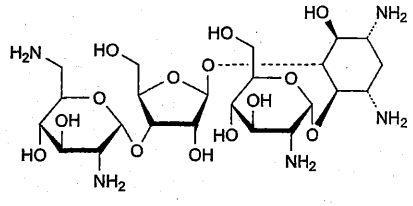
C. 2-deoxy-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-5-*O*-[3-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-β-D-ribofuranosyl]-D-streptamine (neomycin C),



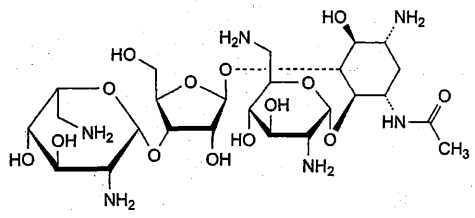
D. 4-*O*-(2-amino-2-deoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine (paromamine or neomycin D),



E. 4-*O*-(2-amino-2-deoxy-α-D-glucopyranosyl)-2-deoxy-5-*O*-[3-*O*-(2,6-diamino-2,6-dideoxy-β-L-idopyranosyl)-β-D-ribofuranosyl]-D-streptamine (paromomycin I or neomycin E),



F. 4-*O*-(2-amino-2-deoxy-α-D-glucopyranosyl)-2-deoxy-5-*O*-[3-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-β-D-ribofuranosyl]-D-streptamine (paromomycin II or neomycin F),

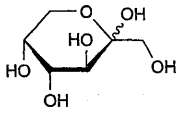


G. 3-*N*-acetyl-2-deoxy-4-*O*-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-5-*O*-[3-*O*-(2,6-diamino-2,6-dideoxy-β-L-idopyranosyl)-β-D-ribofuranosyl]-D-streptamine (neomycin B-LP).

Ph Eur

Fructose

(Ph. Eur. monograph 0188)



$C_6H_{12}O_6$ 180.2 *57-48-7*

Ph Eur

DEFINITION

D-*arabino*-Hex-2-ulopyranose (levulose).

The substance described in this monograph is not necessarily suitable for parenteral administration.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (*2.2.27*).

Solvent mixture *water R, methanol R* (2:3 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of *fructose CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *fructose R*, 10 mg of *glucose R*, 10 mg of *lactose monohydrate R* and 10 mg of *sucrose R* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Plate *TLC silica gel G plate R.*

Mobile phase *water R, methanol R, anhydrous acetic acid R, ethylene chloride R* (10:15:25:50 *V/V/V/V*). Measure the volumes accurately since a slight excess of water produces cloudiness.

Application 2 µL; thoroughly dry the points of application.

Development A Over 3/4 of the plate.

Drying A In a current of warm air.

Development B Immediately, over 3/4 of the plate, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with a solution of 0.5 g of *thymol R* in a mixture of 5 mL of *sulfuric acid R* and 95 mL of *ethanol (96 per cent) R*. Heat at 130 °C for 10 min.

System suitability Reference solution (b):

— the chromatogram shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

B. Dissolve 0.1 g in 10 mL of *water R*. Add 3 mL of *cupri-tartaric solution R* and heat. A red precipitate is formed.

C. To 1 mL of solution S (see Tests) add 9 mL of *water R*. To 1 mL of the solution add 5 mL of *hydrochloric acid R* and heat to 70 °C. A brown colour develops.

D. Dissolve 5 g in *water R* and dilute to 10 mL with the same solvent. To 0.5 mL of the solution add 0.2 g of *resorcinol R* and 9 mL of *dilute hydrochloric acid R* and heat on a water-bath for 2 min. A red colour develops.

TESTS

Solution S

Dissolve 10.0 g in *distilled water R* and dilute to 100 mL with the same solvent.

Appearance of solution

Dissolve 5.0 g in *water R* and dilute to 10 mL with the same solvent. The solution is clear (*2.2.1*). Add 10 mL of *water R*. The solution is colourless (*2.2.2, Method II*).

Acidity or alkalinity

Dissolve 6.0 g in 25 mL of *carbon dioxide-free water R* and add 0.3 mL of *phenolphthalein solution R*. The solution is colourless. Not more than 0.15 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Specific optical rotation (*2.2.7*)

−93.5 to −91.0 (anhydrous substance).

Dissolve 10.0 g in 80 mL of *water R*, add 0.2 mL of *dilute ammonia R1*, allow to stand for 30 min and dilute to 100.0 mL with *water R*.

Foreign sugars

Dissolve 5.0 g in *water R* and dilute to 10 mL with the same solvent. To 1 mL of the solution add 9 mL of *ethanol (96 per cent) R*. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of the initial solution and 9 mL of *water R*.

5-Hydroxymethylfurfural and related compounds

To 5 mL of solution S add 5 mL of *water R*. The absorbance (*2.2.25*) measured at 284 nm is not greater than 0.32.

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash

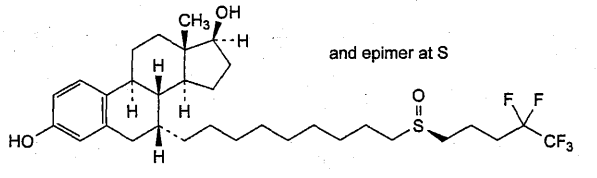
Maximum 0.1 per cent.

Dissolve 5.0 g in 10 mL of *water R*, add 2 mL of *sulfuric acid R*, evaporate to dryness on a water-bath and ignite to constant mass.

Ph Eur

Fulvestrant

(Ph. Eur. monograph 2443)



$C_{32}H_{47}F_5O_3S$ 607 *129453-61-8*

Action and use

Oestrogen receptor antagonist.

Ph Eur

DEFINITION

7α-[9-[(*RS*)-(4,4,5,5,5-Pentafluoropentyl)sulfinyl]nonyl]estra-1,3,5(10)-triene-3,17β-diol.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

Carry out either tests A, B or tests B, C.

A. Specific optical rotation (*2.2.7*): + 108 to + 115 (anhydrous substance), measured at 365 nm at a temperature of 25 °C.

Dissolve 0.50 g in *methanol R* and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fulvestrant CRS*.

C. Stereochemical purity (see Tests).

TESTS

Appearance of solution

The solution is clear (*2.2.1*).

Dissolve 0.1 g in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in *methanol R1* and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of *fulvestrant CRS* in *methanol R1* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *fulvestrant for system suitability CRS* (containing impurities A, B, C, D and F) in 1.0 mL of *methanol R1*.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R1*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R1*.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (3.5 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *methanol R2*, *acetonitrile R1*, *water for chromatography R* (27:32:41 *V/V/V*);
- *mobile phase B*: *water for chromatography R*, *methanol R2*, *acetonitrile R1* (10:41:49 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	100	0
25 - 55	100 → 0	0 → 100
55 - 65	0	100

Flow rate 2 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with *fulvestrant for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and F.

Relative retention With reference to fulvestrant (retention time = about 23 min): impurity F = about 0.4; impurity A = about 1.1; impurity B = about 1.2; impurity C = about 1.7; impurity D = about 1.9.

System suitability Reference solution (b):
- *resolution*: minimum 1.5 between the peaks due to fulvestrant and impurity A.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.7; impurity F = 0.3;
- for each impurity, use the concentration of fulvestrant in reference solution (c).

Limits:
- *impurity D*: maximum 0.6 per cent;
- *impurity C*: maximum 0.3 per cent;
- *impurity B*: maximum 0.2 per cent;
- *impurity F*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 1.0 per cent;
- *reporting threshold*: 0.05 per cent.

Stereochemical purity

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution Dissolve 5 mg of *fulvestrant CRS* in the mobile phase and dilute to 5 mL with the mobile phase.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *amylose derivative of silica gel for chiral separation R* (5 µm);
- *temperature*: 25 °C.

Mobile phase *propanol R1*, *heptane R* (15:85 *V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Run time Twice the retention time of fulvestrant epimer B.

Identification of peaks Use the chromatogram supplied with *fulvestrant CRS* and the chromatogram obtained with the reference solution to identify the peaks due to fulvestrant epimers A and B.

Relative retention With reference to fulvestrant epimer B (retention time = about 18 min): fulvestrant epimer A = about 1.2.

System suitability Reference solution:
- *resolution*: minimum 2.0 between the peaks due to fulvestrant epimer B and fulvestrant epimer A.

Limit:
- *fulvestrant epimer A*: 42 per cent to 48 per cent;
- *fulvestrant epimer B*: 52 per cent to 58 per cent.

Water (*2.5.32*)

Maximum 0.5 per cent, determined on 50 mg.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

Bacterial endotoxins (*2.6.14*)

Less than 1.25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

Test solution Dissolve 0.1 g of the substance to be examined in 1 mL of *ethanol (96 per cent) R* and dilute to 80 mL with water for BET.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{32}H_{47}F_5O_3S$ taking into account the assigned content of *fulvestrant CRS*.

STORAGE

Protected from light at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

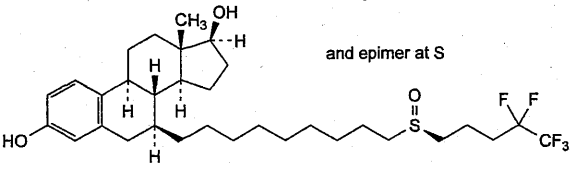
LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

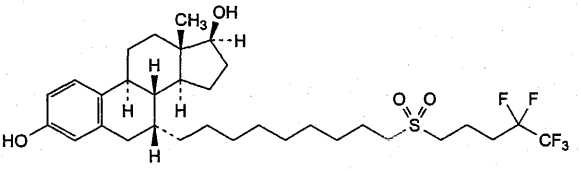
IMPURITIES

Specified impurities *B, C, D, F.*

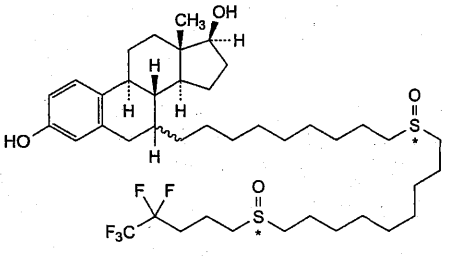
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, E.*



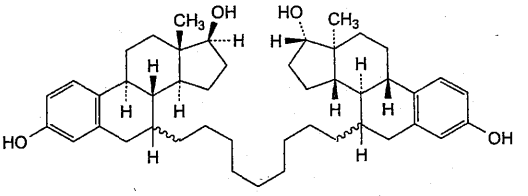
A. 7β-[9-[(*RS*)-(4,4,5,5,5-pentafluoropentyl)sulfinyl] nonyl]estra-1,3,5(10)-triene-3,17β-diol (7β-fulvestrant),



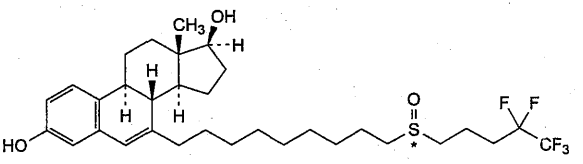
B. 7α-[9-[(4,4,5,5,5-pentafluoropentyl)sulfonyl]nonyl]estra-1,3,5(10)-triene-3,17β-diol,



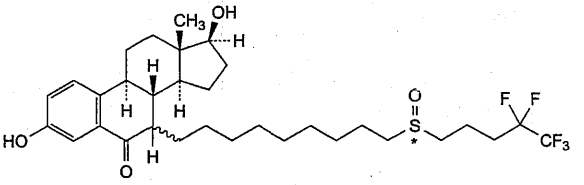
C. 7ξ-[9-[[9-[(4,4,5,5,5-pentafluoropentyl)sulfinyl] nonyl]sulfinyl]nonyl]estra-1,3,5(10)-triene-3,17β-diol,



D. 7ξ,7′ξ-nonane-1,9-diylbis[estra-1,3,5(10)-triene-3,17β-diol],



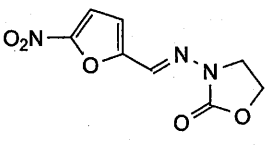
E. 7-[9-[(4,4,5,5,5-pentafluoropentyl)sulfinyl]nonyl]estra-1,3,5(10),6-tetraene-3,17β-diol (Δ6-fulvestrant),



F. 3,17β-dihydroxy-7ξ-[9-[(4,4,5,5,5-pentafluoropentyl) sulfinyl]nonyl]estra-1,3,5(10)-trien-6-one (6-keto-fulvestrant).

Ph Eur

Furazolidone



$C_8H_7N_3O_5$ 225.2 *67-45-8*

Action and use

Antiprotozoal; antibacterial.

DEFINITION

Furazolidone is 3-(5-nitrofurfurylideneamino)oxazolidin-2-one. It contains not less than 97.0% and not more than 103.0% of $C_8H_7N_3O_5$, calculated with reference to the dried substance.

CHARACTERISTICS

A yellow, crystalline powder.

Very slightly soluble in *water* and in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION

A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of furazolidone *(RS 164)*.

B. Dissolve 1 mg in 1 mL of *dimethylformamide* and add 0.05 mL of 1M *ethanolic potassium hydroxide*. A deep blue colour is produced.

TESTS

Acidity or alkalinity

Shake 1 g for 15 minutes with 100 mL of *carbon dioxide-free water* and filter. The pH of the filtrate is 4.5 to 7.0, Appendix V L.

Nitrofurfural diacetate

Carry out in subdued light the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and a mixture of 5 volumes of *1,4-dioxan* and 95 volumes of *toluene* as the mobile phase. Apply separately to the plate 20 µL of solution (1) and 10 µL of solution (2). For solution (1) dissolve 50 mg of the substance being examined in 5 mL of *dimethylformamide* by heating on a water bath for a few minutes, allow to cool and dilute to 10 mL with *acetone*. Solution (2) contains 0.010% w/v of *nitrofurfural diacetate BPCRS* in a mixture of equal volumes of *dimethylformamide* and *acetone*. After removal of the plate, heat it at 105° for 5 minutes and spray with a solution prepared by dissolving 0.75 g of *phenylhydrazine hydrochloride* in 10 mL of *ethanol (96%)*, diluting to 50 mL with *water*, adding *activated charcoal*, filtering and then adding 25 mL of *hydrochloric acid* and sufficient *water* to produce 200 mL. Any spot corresponding to nitrofurfural diacetate in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (1%).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

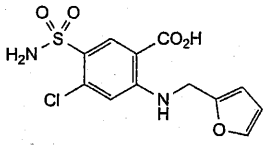
Carry out the following procedure protected from light. To 80 mg add 150 mL of *dimethylformamide*, swirl to dissolve and add sufficient *water* to produce 500 mL. Dilute 5 mL to 100 mL with *water* and mix. Measure the *absorbance* of the resulting solution at the maximum at 367 nm, Appendix II B. Calculate the content of $C_8H_7N_3O_5$ taking 750 as the value of A(1%, 1 cm) at the maximum at 367 nm.

STORAGE

Furazolidone should be protected from light.

Furosemide

(Ph. Eur. monograph 0391)



$C_{12}H_{11}ClN_2O_5S$ 330.7 *54-31-9*

Action and use

Loop diuretic.

Preparations

Co-amilofruse Tablets

Furosemide Injection

Furosemide Tablets

Ph Eur

DEFINITION

4-Chloro-2-[[(furan-2-yl)methyl]amino]-5-sulfamoylbenzoic acid.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: B.

Second identification: A.

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of *methanol R.*

Reference solution Dissolve 20 mg of *furosemide CRS* in 1 mL of *methanol R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *glacial acetic acid R, ethyl acetate R, toluene R* (5:45:50 *V/V/V*).

Application 2 µL; the volume may be adapted according to the type of plate used.

Development Over 2/3 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

Detection B Treat with *ninhydrin solution R*, dry at 100-105 °C until the spots appear and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *furosemide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Dissolve 0.5 g in *0.5 M sodium hydroxide* and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of *furosemide impurity A CRS* in the mobile phase, add 2.0 mL of the test solution and dilute to 20.0 mL with the mobile phase. Dilute 0.5 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 2 mg of *furosemide for peak identification CRS* (containing impurities C and D) in 2.0 mL of the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 2.0 g of *potassium dihydrogen phosphate R* and 2.5 g of *cetrimide R* in 700 mL of *water for chromatography R*, adjust to pH 7.0 with *ammonia R* and add 300 mL of *propanol R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 238 nm.

Injection 20 µL.

Run time 3 times the retention time of furosemide.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *furosemide for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and D.

Relative retention With reference to furosemide (retention time = about 9 min): impurity C = about 0.5; impurity A = about 0.8; impurity D = about 1.5.

System suitability:
— *resolution*: minimum 4.0 between the peaks due to impurity A and furosemide in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 40 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.4; impurity D = 2.0;
— *impurity C*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurity D*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 200 ppm.

To 0.5 g add a mixture of 0.2 mL of *nitric acid R* and 30 mL of *water R* and shake for 5 min. Allow to stand for 15 min and filter.

Sulfates (*2.4.13*)
Maximum 300 ppm.

To 1.0 g add a mixture of 0.2 mL of *acetic acid R* and 30 mL of *distilled water R* and shake for 5 min. Allow to stand for 15 min and filter.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 40 mL of *dimethylformamide R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M sodium hydroxide* is equivalent to 33.07 mg of $C_{12}H_{11}ClN_2O_5S$.

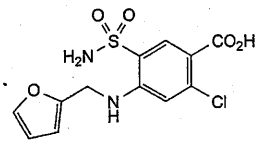
STORAGE

Protected from light.

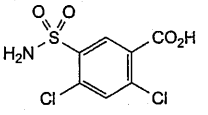
IMPURITIES

Specified impurities *C, D.*

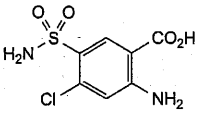
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E, F.*



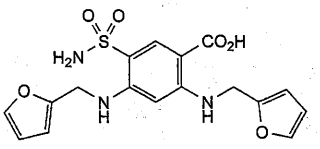
A. 2-chloro-4-[[(furan-2-yl)methyl]amino]-5-sulfamoylbenzoic acid,



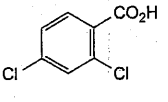
B. 2,4-dichloro-5-sulfamoylbenzoic acid,



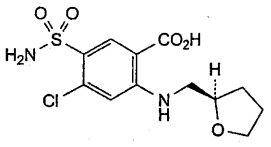
C. 2-amino-4-chloro-5-sulfamoylbenzoic acid,



D. 2,4-bis[[(furan-2-yl)methyl]amino]-5-sulfamoylbenzoic acid,



E. 2,4-dichlorobenzoic acid,

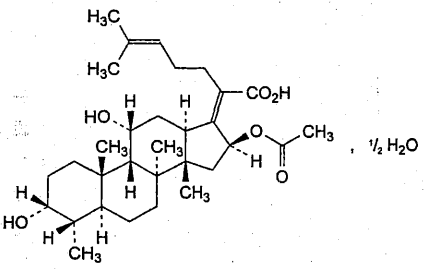


F. 4-chloro-5-sulfamoyl-2-[[[(2*RS*)-oxolan-2-yl]methyl] amino]benzoic acid.

Ph Eur

Fusidic Acid

(Ph. Eur. monograph 0798)



$C_{31}H_{48}O_6,½H_2O$ 525.7 *6990-06-3*

Action and use
Antibacterial.

Preparations
Fusidic Acid Cream
Fusidic Acid Eye Drops
Fusidic Acid Oral Suspension

Ph Eur

DEFINITION

ent-(17*Z*)-16α-(Acetyloxy)-3β,11β-dihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid hemihydrate.

Antimicrobial substance produced by fermentation of certain strains of *Fusidium coccineum* or by any other means.

Content
97.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *fusidic acid CRS.*

B. Ignite 1 g. The residue does not give reaction (a) of sodium (*2.3.1*).

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *methanol R*, 5 g/L solution of *phosphoric acid R*, *acetonitrile R* (10:40:50 *V/V/V*).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of *fusidic acid for peak identification CRS* (containing impurities A, B, C, D, F, G, H and N) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve the contents of a vial of *fusidic acid impurity mixture CRS* (containing impurities I, K, L and M) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3.5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: *methanol R*, *acetonitrile R*, 5 g/L solution of *phosphoric acid R* (20:40:40 *V/V/V*);
— *mobile phase B*: 5 g/L solution of *phosphoric acid R*, *methanol R*, *acetonitrile R* (10:20:70 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 28	100 → 0	0 → 100
28 - 33	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *fusidic acid for peak identification CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, F, G, H and N; use the chromatogram supplied with *fusidic acid impurity mixture CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities I, K, L and M.

Relative retention With reference to fusidic acid (retention time = about 18 min): impurity A = about 0.4; impurity B = about 0.5; impurity C = about 0.6; impurity D = about 0.63; impurity N = about 0.65; impurity F = about 0.7; impurity G = about 0.82; impurity H = about 0.85; impurity I = about 0.96; impurity K = about 1.18; impurity L = about 1.23; impurity M = about 1.4.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to impurities G and H.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7; impurity D = 0.7; impurity F = 0.3; impurity I = 0.6; impurity K = 0.6;
— *impurity M*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *impurity G*: not more than 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
— *impurity L*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *impurity B*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.4 per cent);
— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
— *impurities C, D, F, I, K, N*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (*2.5.12*)
1.4 per cent to 2.0 per cent, determined on 0.50 g.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 10 mL of *ethanol (96 per cent) R*.
Add 0.5 mL of *phenolphthalein solution R*. Titrate with *0.1 M sodium hydroxide* until a pink colour is obtained.

1 mL of *0.1 M sodium hydroxide* is equivalent to 51.67 mg of $C_{31}H_{48}O_6$.

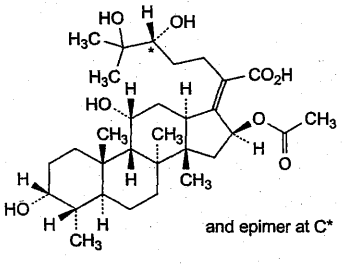
STORAGE

Protected from light, at a temperature of 2 °C to 8 °C.

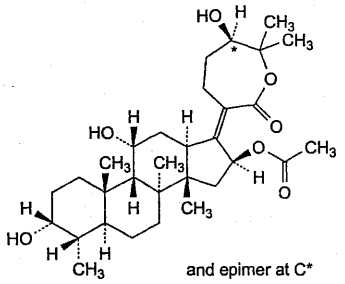
IMPURITIES

Specified impurities *A, B, C, D, F, G, I, K, L, M, N.*

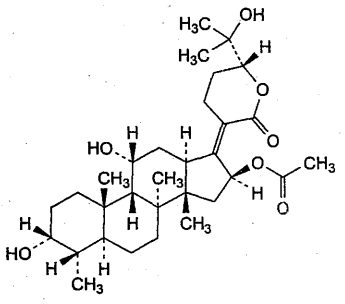
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, H, J, O.*



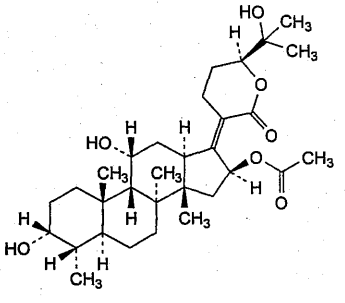
A. *ent*-(24*SR*,17*Z*)-16α-(acetyloxy)-3β,11β,24,25-tetrahydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholest-17(20)-en-21-oic acid (24,25-dihydro-24,25-dihydroxyfusidic acid),



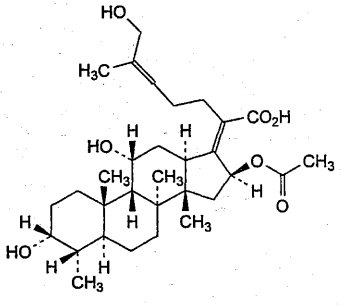
B. *ent*-(17*Z*)-3β,11β-dihydroxy-17-[(6*SR*)-6-hydroxy-7,7-dimethyl-2-oxooxepan-3-ylidene]-4β,8,14-trimethyl-18-nor-5β,10α-androstan-16α-yl acetate (24,25-dihydro-24,25-dihydroxyfusidic acid 21,25-lactone),



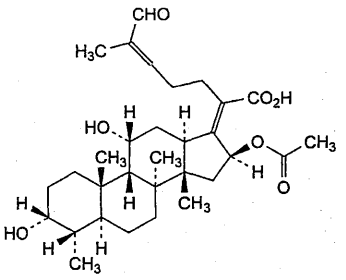
C. *ent*-(17*Z*)-3β,11β-dihydroxy-17-[(6*S*)-6-(1-hydroxy-1-methylethyl)-2-oxodihydro-2*H*-pyran-3(4*H*)-ylidene]-4β,8,14-trimethyl-18-nor-5β,10α-androstan-16α-yl acetate ((24*R*)-24,25-dihydro-24,25-dihydroxyfusidic acid 21,24-lactone),



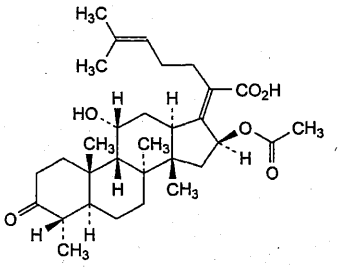
D. *ent*-(17*Z*)-3β,11β-dihydroxy-17-[(6*R*)-6-(1-hydroxy-1-methylethyl)-2-oxodihydro-2*H*-pyran-3(4*H*)-ylidene]-4β,8,14-trimethyl-18-nor-5β,10α-androstan-16α-yl acetate ((24*S*)-24,25-dihydro-24,25-dihydroxyfusidic acid 21,24-lactone),



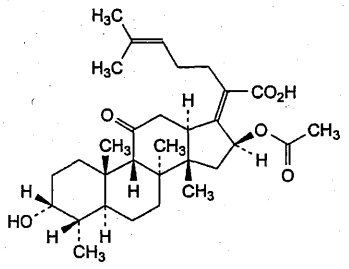
E. *ent*-(17*Z*,24*EZ*)-16α-(acetyloxy)-3β,11β,26-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (26-hydroxyfusidic acid),



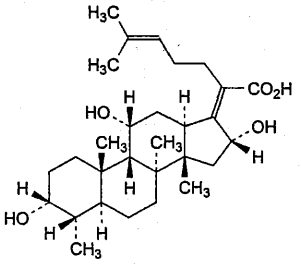
F. *ent*-(17*Z*,24*EZ*)-16α-(acetyloxy)-3β,11β-dihydroxy-4β,8,14-trimethyl-26-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (26-oxofusidic acid),



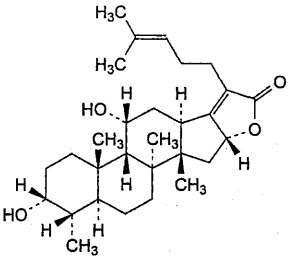
G. *ent*-(17*Z*)-16α-(acetyloxy)-11β-hydroxy-4β,8,14-trimethyl-3-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (3-didehydrofusidic acid),



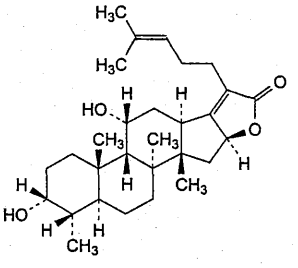
H. *ent*-(17*Z*)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-11-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (11-didehydrofusidic acid),



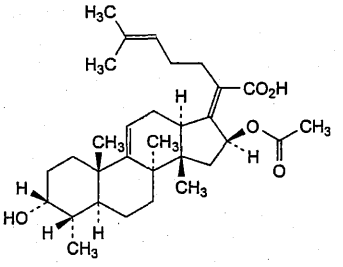
I. *ent*-(17*Z*)-3β,11β,16β-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (16-*epi*-deacetylfusidic acid),



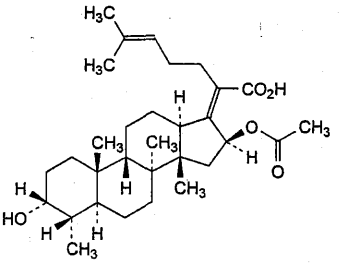
J. *ent*-(17*Z*)-3β,11β-dihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dieno-21(16β)-lactone (16-*epi*-deacetylfusidic acid 21,16-lactone),



K. *ent*-(17*Z*)-3β,11β-dihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dieno-21(16α)-lactone (deacetylfusidic acid 21,16-lactone),

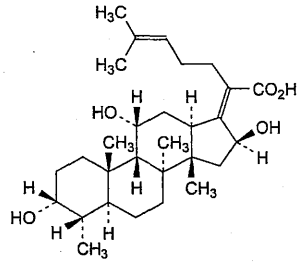


L. *ent*-(17*Z*)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-9(11),17(20),24-trien-21-oic acid (9,11-anhydrofusidic acid),



M. *ent*-(17*Z*)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (11-deoxyfusidic acid),

N. unknown structure,

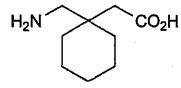


O. *ent*-(17*Z*)-3β,11β,16α-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (deacetylfusidic acid).

Ph Eur

Gabapentin

(Ph. Eur. monograph 2173)



$C_9H_{17}NO_2$ 171.2 *60142-96-3*

Action and use
Antiepileptic.

Preparations
Gabapentin Capsules
Gabapentin Oral Solution
Gabapentin Tablets

Ph Eur

DEFINITION
[1-(Aminomethyl)cyclohexyl]acetic acid.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in dilute acids and dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gabapentin CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1.50 g in a mixture of 0.5 mL of *acetic acid R*, 19.5 mL of *methanol R* and 30 mL of *water R*.

pH (*2.2.3*)
6.5 to 7.5.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Related substances
A. Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solution A Dissolve 2.32 g of *ammonium dihydrogen phosphate R* in 950 mL of *water R*, adjust to pH 2.0 with *phosphoric acid R*, and dilute to 1000 mL with *water R*.

Buffer solution Dissolve 0.58 g of *ammonium dihydrogen phosphate R* and 1.83 g of *sodium perchlorate R* in 950 mL of *water for chromatography R*, adjust to pH 1.8 with *perchloric acid R*, and dilute to 1000 mL with *water for chromatography R*.

Test solution Dissolve 0.140 g of the substance to be examined in solution A and dilute to 10.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 7.0 mg of *gabapentin impurity A CRS* and 10 mg of *gabapentin impurity B CRS* in *methanol R1* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with solution A.

Reference solution (c) Dissolve 0.140 g of *gabapentin CRS* in solution A and dilute to 10.0 mL with solution A.

Reference solution (d) Dissolve 7.0 mg of *gabapentin impurity D CRS* in 25 mL of *methanol R1* and dilute to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R* (5 µm);
- *temperature*: 40 °C.

Mobile phase *acetonitrile R1*, buffer solution (24:76 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL of the test solution and reference solutions (a) and (b).

Run time 4 times the retention time of gabapentin.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to gabapentin (retention time = about 4 min): impurity A = about 2.4; impurity B = about 2.8.

System suitability Reference solution (b):
- *resolution*: minimum 2.3 between the peaks due to impurities A and B.

To avoid memory effects between 2 chromatograms, the column may be washed using *acetonitrile R1*.

Limits:
- *impurity A*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply for this test.

B. Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modifications.

Mobile phase *methanol R2, acetonitrile R1*, buffer solution (30:35:35 *V/V/V*).

Injection 20 µL of the test solution and reference solution (d).

Run time 1.2 times the retention time of impurity D.

Retention time Impurity D = about 10 min.

Limits:

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent);

— *disregard limit*: 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.03 per cent); disregard any peak with a relative retention of not more than 0.4 with reference to impurity D.

Limit:

— *total for tests A and B*: maximum 0.5 per cent.

Chlorides

Maximum 100 ppm.

Dissolve 1.5 g in a mixture of 0.5 mL of *acetic acid R*, 19.5 mL of *methanol R* and 30 mL of *water R*. Titrate with *0.001 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.001 M silver nitrate* is equivalent to 0.03545 mg of chlorides.

Water (*2.5.32*)

Maximum 0.3 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in test A for related substances with the following modification.

Injection 20 µL of the test solution and reference solution (c).

System suitability Reference solution (c):

— *symmetry factor*: maximum 5.0 for the peak due to gabapentin.

Calculate the percentage content of $C_9H_{17}NO_2$ taking into account the assigned content of *gabapentin CRS*.

IMPURITIES

Test A for related substances

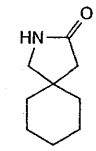
A, B, E, G.

Test B for related substances

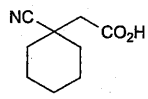
D.

Specified impurities *A.*

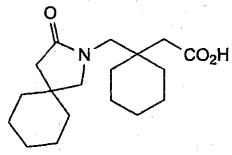
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, E, G.*



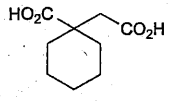
A. 2-azaspiro[4.5]decan-3-one,



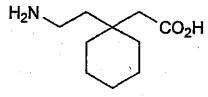
B. (1-cyanocyclohexyl)acetic acid,



D. [1-[(3-oxo-2-azaspiro[4.5]dec-2-yl)methyl] cyclohexyl]acetic acid,



E. 1-(carboxymethyl)cyclohexanecarboxylic acid,

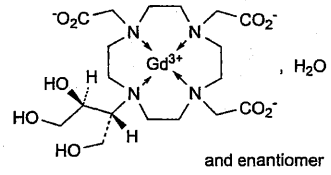


G. [1-(2-aminoethyl)cyclohexyl]acetic acid.

Ph Eur

Gadobutrol Monohydrate

(Ph. Eur. monograph 2735)



$C_{18}H_{31}GdN_4O_9,H_2O$ 623 *198637-52-4*

Ph Eur

DEFINITION

Gadolinium 2,2′,2′′-[10-[(1*RS*,2*SR*)-2,3-dihydroxy-1-(hydroxymethyl)propyl]-1,4,7,10-tetraazacyclododecane-1,4,7-triyl]triacetate monohydrate.

Content

— *gadobutrol*: 97.5 per cent to 102.5 per cent (anhydrous substance);

— *gadolinium*: 98.0 per cent to 102.0 per cent, calculated as gadobutrol (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility
Freely soluble in water, practically insoluble in anhydrous ethanol and in heptane.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gadobutrol monohydrate CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in 0.5 mL of *water R*, then add 5 mL of *anhydrous ethanol R*; evaporate to dryness and record new spectra using the residues.

B. Inductively coupled plasma-atomic emission spectrometry (*2.2.57*) as described in the assay for gadolinium.

TESTS

Appearance of solution
The solution is clear (*2.2.1*).

Dissolve 6.0 g in *water R* and dilute to 10.0 mL with the same solvent. Sonicate if necessary.

Specific optical rotation (*2.2.7*)
−0.05 to + 0.05 (anhydrous substance), using an instrument with a minimum sensitivity of ± 0.002°.

Dissolve 6.00 g in *water R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Use suitable plastic labware for the preparation of the solutions and plastic vials. Store the solutions at 10 °C and use them within 42 h.*

Test solution (a) Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 5.0 mL with mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 10 mg of *gadobutrol for peak identification CRS* (containing impurity C) in 1.0 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 20.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (c) Dissolve 50.0 mg of *gadobutrol monohydrate CRS* in mobile phase A and dilute to 50.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped phenylhexylsilyl silica gel for chromatography R* (3 µm) with a pore size of 10 nm.

Temperature:
— *autosampler*: 10 °C;
— *column*: 50 °C.

Mobile phase:
— *mobile phase A*: mix 5 volumes of *acetonitrile R1* and 995 volumes of *water for chromatography R* previously adjusted to pH 3.6 with a 50 per cent *V/V* solution of *anhydrous formic acid R* in *water for chromatography R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100	0
15 - 30	100 → 75	0 → 25

Flow rate 1.0 mL/min.

Detection Charged aerosol detector at 100 pA.

Injection 40 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *gadobutrol for peak identification CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to gadobutrol (retention time = about 11 min): impurity C = about 1.2.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to gadobutrol.

Calculation of percentage contents:
— for each impurity, use the concentration of gadobutrol in reference solution (b).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.3 per cent;
— *reporting threshold*: 0.03 per cent.

Free gadolinium
Maximum 0.01 per cent (anhydrous substance).

Solution A To 30.0 mL of *sodium acetate buffer solution pH 5.0 R* add 3.0 mL of *xylenol orange solution R* and dilute to 200.0 mL with *water R*.

Gadolinium sulfate solution Dissolve 93.35 mg of *gadolinium sulfate octahydrate R* in *water R* and dilute to 1000.0 mL with the same solvent.

Test solution Dissolve 0.250 g of the substance to be examined in 5.0 mL of the gadolinium sulfate solution and 30 mL of *water R* with the aid of ultrasound. Add 10.0 mL of solution A and adjust to pH 5.0 with a 10.3 g/L solution of *hydrochloric acid R*. Titrate with 0.00025 M sodium edetate, determining the end-point photometrically using a suitable autotitrator equipped with a photometric sensor at a wavelength of 570-574 nm (V_1 mL).

Standard solution Prepare a solution in the same manner as for the test solution but omitting the substance to be examined (V_2 mL).

Determine the free gadolinium content from the difference in the volumes of titrant consumed (V_1 - V_2).

1 mL of 0.00025 M sodium edetate is equivalent to 0.03931 mg of gadolinium.

Water (*2.5.32*)
2.0 per cent to 7.0 per cent, determined on 0.100 g using the evaporation technique at 220 °C.

ASSAY

Gadobutrol
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 195 nm.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{18}H_{31}GdN_4O_9$ taking into account the assigned content of *gadobutrol monohydrate CRS*.

Gadolinium
Inductively coupled plasma-atomic emission spectrometry (*2.2.57*).

Yttrium standard solution To 100.0 mL of a certified reference solution containing 1000 mg/L of Y add 50 mL of *nitric acid R* and dilute to 1000.0 mL with *water R*.

Zero solution To 1.5 mL of the yttrium standard solution add 0.5 mL of *nitric acid R* and dilute to 50.0 mL with *water R*.

Test solution Dissolve 50.0 mg in 150.0 mL of *water R*, add 7.5 mL of yttrium standard solution and 2.0 mL of *nitric acid R* and dilute to 250.0 mL with *water R*.

Reference solutions Into 3 volumetric flasks introduce respectively 2.0 mL, 2.5 mL and 3.0 mL of a certified reference solution containing 1000 mg/L of Gd. To each flask add 1.5 mL of the yttrium standard solution and 0.5 mL of *nitric acid R* and dilute to 50.0 mL with *water R*.

Wavelengths 217.069 nm, 217.774 nm, 219.103 nm, 226.109 nm (gadolinium), 224.306 nm (yttrium).

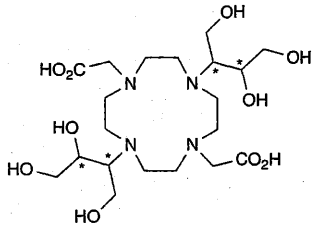
From the calibration curve obtained with the reference solutions, calculate the percentage content of gadolinium in the substance to be examined, using the mean of the results obtained with the different wavelengths and applying a conversion factor of 3.846.

STORAGE

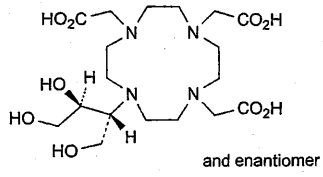
In an airtight container, protected from light.

IMPURITIES

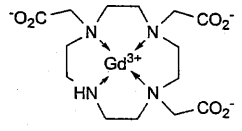
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C.*



A. 2,2′-[4,10-bis[2,3-dihydroxy-1-(hydroxymethyl)propyl]-1,4,7,10-tetraazacyclododecane-1,7-diyl]diacetic acid,



B. 2,2′,2″-[10-[(1*RS*,2*SR*)-2,3-dihydroxy-1-(hydroxymethyl)propyl]-1,4,7,10-tetraazacyclododecane-1,4,7-triyl]triacetic acid,

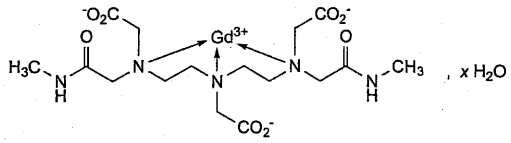


C. gadolinium 2,2′,2″-(1,4,7,10-tetraazacyclododecane-1,4,7-triyl)triacetate.

Ph Eur

Gadodiamide Hydrate

(Ph. Eur. monograph 2225)



$C_{16}H_{26}GdN_5O_8,xH_2O$ with x = 2 to 5 573.7 (anhydrous substance)

Anhydrous gadodiamide *131410-48-5*

Action and use

Paramagnetic contrast enhancing medium for magnetic resonance imaging.

Ph Eur

DEFINITION

6-(Carboxylatomethyl)-3,9-bis[(methylcarbamoyl)methyl]-3,6,9-triazaundecanedioato-κ$^3N^3$,N^6,N^9-gadolinium(III) hydrate (Gd-DTPA-BMA).

Content

— *gadolinium* (Gd; M_r 157.3): 26.0 per cent to 29.0 per cent (anhydrous substance);
— *gadodiamide* ($C_{16}H_{26}GdN_5O_8$; M_r 573.7): 96.5 per cent to 102.0 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS

Appearance

White or almost white powder, slightly hygroscopic.

Solubility

Freely soluble in water and in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gadodiamide CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Gadolinium (see Assay).

Results The test solution exhibits the characteristic emission of gadolinium at 342.247 nm.

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (*2.2.1*). If this is not the case, heat the solution at 60-70 °C for 2-3 min, cool to room temperature and compare the solutions again.

Dissolve 7.5 g (anhydrous substance) in *water R* and dilute to 25 mL with the same solvent.

pH (*2.2.3*)

4.0 to 6.5.

Dissolve 0.50 g (anhydrous substance) in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.200 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dissolve 60.0 mg of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *gadodiamide impurity A CRS* and 5.0 mg of *gadodiamide impurity B CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dissolve 20.0 mg of *gadodiamide CRS* in *water R*, add 0.5 mL of reference solution (a) and dilute to 10.0 mL with *water R*.

Reference solution (c) Dissolve 60.0 mg of *gadodiamide CRS* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dilute 1.0 mL of test solution (a) to 50.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (e) Dilute 1.0 mL of reference solution (a) to 20.0 mL with *water R*.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase To 980 mL of *water R*, add 287 µL of *glacial acetic acid R* and 700 µL of *triethylamine R*. Adjust to pH 6.8 with a 0.6 g/L solution of *acetic acid R* or with a 42 g/L solution of *sodium hydroxide R*, then dilute to 1000 mL with *water R*. Use within 2 days.

Flow rate 1.5 mL/min.

Post-column solution Dissolve 0.120 g of *arsenazo III R* in about 400 mL of *water R*. Add 0.650 g of *urea R* and mix to dissolve. Add 6.3 mL of *nitric acid R* and sonicate for 15 min. Filter through a membrane filter (nominal pore size 0.45 µm) and wash the filter with about 550 mL of *water R*. Dilute the filtrate and washings to 1000 mL with *water R*. Use within 1 week.

Flow rate of post-column solution 1.5 mL/min.

Detection Spectrophotometer at 658 nm.

Injection 10 µL of test solution (a) and reference solutions (b), (d) and (e).

Run time 5 times the retention time of gadodiamide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to gadodiamide (retention time = about 5 min): impurity A = about 2.6; impurity B = about 4.0. The peak due to gadodiamide shows a shoulder due to an isomer that is to be integrated with the main peak.

System suitability Reference solution (b):
— *resolution*: minimum 2.5 between the peaks due to gadodiamide and impurity A; minimum 2.5 between the peaks due to impurities A and B.

Calculation of percentage contents:
— for impurity A, use the concentration of impurity A in reference solution (e);
— for impurity B, use the concentration of impurity B in reference solution (e);
— for impurities other than A and B, use the concentration of gadodiamide in reference solution (d).

Limits:
— *impurity A*: maximum 1.0 per cent;
— *impurity B*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.2 per cent;
— *sum of unspecified impurities*: maximum 0.3 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.10 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Excess of gadolinium or DTPA-BMA

Maximum 20 ppm (anhydrous substance) for the excess of gadolinium and maximum 0.1 per cent (anhydrous substance) for the excess of diethylenetriaminepentaacetic acid bis-(methylamide) (DTPA-BMA).

Test solution Dissolve 2.0 g of the substance to be examined in 25.0 mL of *1 M morpholinoethanesulfonate buffer solution pH 6.0 R*. Add 50 µL of a 1.5 g/L solution of *arsenazo III R*.

Blank solution To 25.0 mL of *1 M morpholinoethanesulfonate buffer solution pH 6.0 R*, add 50 µL of a 1.5 g/L solution of *arsenazo III R*. The solution is violet-pink.

Standard solution Dissolve 18.6 g of *gadolinium chloride hexahydrate R* in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 1000.0 mL with the same acid solution. Dilute 20.0 mL of this solution to 1000.0 mL with *water R* (0.001 M solution). Standardise 10.0 mL of *0.001 M sodium edetate*, using a mixture of 25.0 mL of *1 M morpholinoethanesulfonate buffer solution pH 6.0 R* and 50 µL of a 1.5 g/L solution of *arsenazo III R*. Titrate with the standard solution until a greenish-blue colour is obtained.

Excess of DTPA-BMA If the test solution is violet-pink, there is no excess of gadolinium. Determine the excess of DTPA-BMA by titrating the test solution and the blank solution with the standard solution until a greenish-blue colour is obtained. Calculate the excess of DTPA-BMA, in per cent, using the following expression:

$$\frac{(V_1 - V_0) \times M_1 \times 419.43 \times 100}{m}$$

V_0 = volume of standard solution used in the blank titration, in millilitres;
V_1 = volume of standard solution used in the titration of the test solution, in millilitres;
M_1 = molarity of the gadolinium solution, in moles per litre;
m = mass of the substance to be examined (anhydrous substance) used to prepare the test solution, in milligrams.

Excess of gadolinium If the test solution is greenish-blue, there is no excess of DTPA-BMA. Determine the excess of gadolinium by titrating the test solution with *0.001 M sodium edetate* until a violet-pink colour is obtained. Calculate the excess of gadolinium, in parts per million, using the following expression:

$$\frac{V_2 \times M_2 \times 157.25 \times 10^6}{m}$$

V_2 = volume of *0.001 M sodium edetate* used in the titration of the test solution, in millilitres;
M_2 = molarity of the sodium edetate solution, in moles per litre;
m = mass of the substance to be examined (anhydrous substance) used to prepare the test solution, in milligrams.

Impurity C

Maximum 0.02 per cent (anhydrous substance).

Phthalaldehyde solution Introduce 0.100 g of *phthalaldehyde R* into a brown-glass vessel and dissolve in 3 mL of *methanol R*. Add 220 mL of *borate buffer solution*

pH 10.0 R and 100 µL of *2-mercaptoethanol R* and mix. Keep refrigerated and use within 24 h.

Test solution Dissolve 1.5 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solutions Prepare a series of reference solutions for calibration in the range of 1 µg/mL to 35 µg/mL of methylamine (impurity C) from a 0.220 g/L solution of *methylamine hydrochloride R*. Calculate the concentrations of methylamine by multiplying the concentrations of methylamine hydrochloride by 0.46.

Blank solution A recently prepared mixture of 3.0 mL of phthalaldehyde solution and 3.0 mL of *water R*.

Introduce 3.0 mL of the test solution and of each reference solution into separate test tubes. Add 3.0 mL of phthalaldehyde solution to each tube and mix. Immediately transfer each solution to a cell and immediately measure the absorbance (*2.2.25*) at 335 nm using the blank solution as compensation liquid. Calculate the linear regression curve from the absorbances obtained with the reference solutions: the coefficient of determination r^2 is not less than 0.99. If the absorbance obtained with the test solution is outside the calibration range, dilute the test solution with *water R* and repeat the procedure.

Water (*2.5.12*)
6.0 per cent to 13.0 per cent, determined on 0.150 g.

Microbial contamination
TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Bacterial endotoxins (*2.6.14*)
Less than 3.50 IU/g.

ASSAY

Gadolinium
Inductively coupled plasma-atomic emission spectrometry (*2.2.57*). An external calibration curve is used for quantification.

Test solution Dissolve 0.600 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with a 20 g/L solution of *nitric acid R*.

Reference solutions Into 3 volumetric flasks, introduce 10.0 mL, 15.0 mL and 20.0 mL of a certified reference solution containing 1000 µg/mL of gadolinium. Dilute each flask to 100.0 mL with a 20 g/L solution of *nitric acid R*. Mix to obtain reference solutions containing respectively 100 µg/mL, 150 µg/mL and 200 µg/mL of gadolinium.

The standard operating conditions prescribed by the manufacturer of the apparatus are to be followed.
The following operating conditions are cited as an example of conditions found suitable for a given apparatus:
— *flush time*: 15 s,
— *pure gas (N_2) flow*: 1 mL/min,
— *plasma flow*: 15 L/min,
— *auxiliary flow*: 1.0 mL/min,
— *nebuliser flow*: 1.0 mL/min,
— *pump rate*: 1.0 mL/min,
— *power*: 1160 W,
— *read delay*: 25 s,
— *wavelength*: 342.247 nm,
— *viewing height*: 14.0 mm,
— *background correction*: none,
— *nebuliser*: cross-flow nebuliser.

System suitability:
— the correlation coefficient of the calibration curve is not less than 0.999.

Gadodiamide
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of test solution (b) and reference solution (c).

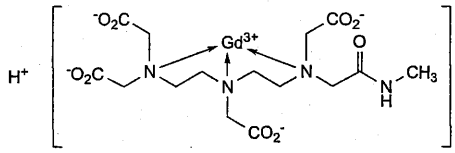
Calculate the percentage content of $C_{16}H_{26}GdN_5O_8$ taking into account the assigned content of *gadodiamide CRS*.

STORAGE

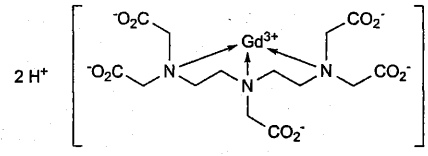
In an airtight container.

IMPURITIES

Specified impurities *A, B, C.*



A. hydrogen[3,6-bis(carboxylatomethyl)-9-[(methylcarbamoyl)methyl]-3,6,9-triazaundecanedioato-κ^3N^3,N^6,N^9-gadolinate](1-) (Gd-DTPA-MMA),



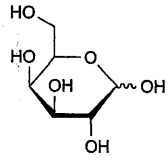
B. dihydrogen[3,6,9-tris(carboxylatomethyl)-3,6,9-triazaundecanedioato-κ^3N^3,N^6,N^9-gadolinate](2-) (Gd-DTPA),

C. $H_3C\text{-}NH_2$: methanamine (methylamine).

Ph Eur

Galactose

(Ph. Eur. monograph 1215)



$C_6H_{12}O_6$ 180.2 *59-23-4*

Ph Eur

DEFINITION

D-Galactopyranose.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline or finely granulated powder.

Solubility
Freely soluble or soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *galactose CRS*.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 2 volumes of *water R* and 3 volumes of *methanol R* and dilute to 20 mL with the same mixture of solvents.

Reference solution (a) Dissolve 10 mg of *galactose CRS* in a mixture of 2 volumes of *water R* and 3 volumes of *methanol R* and dilute to 20 mL with the same mixture of solvents.

Reference solution (b) Dissolve 10 mg each of *galactose R*, *glucose R* and *lactose monohydrate R* (impurity A) in a mixture of 2 volumes of *water R* and 3 volumes of *methanol R* and dilute to 20 mL with the same mixture of solvents.

Plate *TLC silica gel plate R*.

Mobile phase *water R*, *propanol R* (15:85 *V/V*).

Application 2 µL; thoroughly dry the points of application.

Development In an unsaturated tank over 3/4 of the plate.

Drying In a current of warm air.

Detection Spray with a solution of 0.5 g of *thymol R* in a mixture of 5 mL of *sulfuric acid R* and 95 mL of *ethanol (96 per cent) R*. Heat in an oven at 130 °C for 10 min.

System suitability Reference solution (b):
— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in 10 mL of *water R*. Add 3 mL of *cupri-tartaric solution R* and heat. An orange or red precipitate is formed.

TESTS

Solution S

Dissolve, with heating in a water-bath at 50 °C, 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_8 (*2.2.2, Method II*).

Acidity or alkalinity

To 30 mL of solution S add 0.3 mL of *phenolphthalein solution R*. The solution is colourless. Not more than 1.5 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Proteins

Maximum 0.1 mg/mL.

Dissolve 1.000 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent. Measure the absorbance (*2.2.25*) of the solution at 260 nm and at 280 nm and calculate the protein content, in milligrams per millilitre, using the following expression:

$$(A_{280} \times 1.45) - (A_{260} \times 0.74)$$

A_{280} = absorbance at 280 nm;
A_{260} = absorbance at 260 nm.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.250 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with *water R*.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with *water R*.

Reference solution (b) Dissolve 3 mg each of *1,6-galactosylgalactose R* (impurity B), *galacturonic acid R* (impurity C) and *lactose monohydrate R* (impurity A) in *water R* and dilute to 10.0 mL with the same solvent. To 100 µL of the solution add 900 µL of test solution (a).

Reference solution (c) Dissolve 25.0 mg of *galactose CRS* in *water R* and dilute to 50.0 mL with the same solvent.

Column 2 columns to be connected in series:
— *size*: *l* = 0.30 m, Ø = 6.5 mm;
— *stationary phase*: *strong cation-exchange resin (calcium form) R* (10 µm);
— *temperature*: 75 °C.

Mobile phase Dissolve 50 mg of *sodium calcium edetate R* in 900 mL of *water R*, add 1.0 mL of *sulfuric acid R* and dilute to 1000 mL with *water R*.

Flow rate 0.4 mL/min.

Detection Differential refractometer maintained at a constant temperature (about 40 °C).

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time Twice the retention time of galactose.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A + B and C.

Relative retention With reference to galactose (retention time = about 24 min): impurities A and B = about 0.8; impurity C = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to impurity C and galactose.

Calculation of percentage contents:
— for each impurity, use the concentration of galactose in reference solution (a).

Limits:
— *sum of impurities A and B*: maximum 1.0 per cent;
— *unspecified impurities*: for each impurity, maximum 0.3 per cent;
— *total*: maximum 2.0 per cent;
— *reporting threshold*: 0.2 per cent.

Barium

Dilute 5 mL of solution S to 10 mL with *distilled water R*. Add 1 mL of *dilute sulfuric acid R*. When examined immediately and after 1 h, any opalescence in the solution is not more intense than that in a mixture of 5 mL of solution S and 6 mL of *distilled water R*.

Lead (*2.4.10*)

Maximum 0.5 ppm.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 1.00 g.

Sulfated ash

Maximum 0.1 per cent.

To 5 mL of solution S add 2 mL of *sulfuric acid R*, evaporate to dryness on a water-bath and ignite to constant mass. The residue weighs a maximum of 1 mg.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

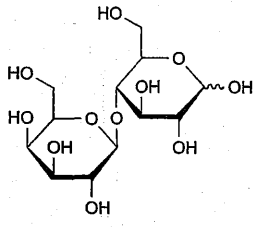
Injection 10 µL of test solution (b) and reference solution (c).

Calculate the percentage content of $C_6H_{12}O_6$ taking into account the assigned content of *galactose CRS*.

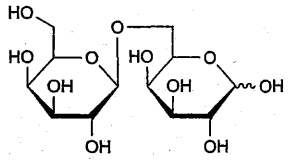
IMPURITIES

Specified impurities *A, B.*

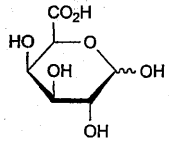
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E.*



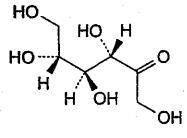
A. 4-*O*-β-D-galactopyranosyl-D-glucopyranose (lactose),



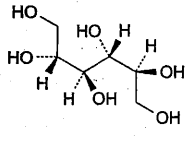
B. 6-*O*-β-D-galactopyranosyl-D-galactopyranose (1,6-galactosylgalactose),



C. D-galactopyranuronic acid (galacturonic acid),



D. D-tagatose,

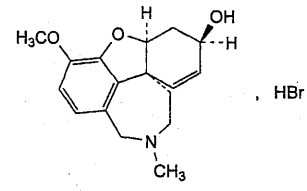


E. galactitol (dulcitol).

Ph Eur

Galantamine Hydrobromide

(Ph. Eur. monograph 2366)



$C_{17}H_{22}BrNO_3$ 368.3 *1953-04-4*

Action and use
Cholinesterase inhibitor; treatment of Alzheimer's disease.

Preparations
Galantamine Oral Solution
Galantamine Prolonged-release Capsules
Galantamine Tablets

Ph Eur

DEFINITION

(4a*S*,6*R*,8a*S*)-3-Methoxy-11-methyl-5,6,9,10,11,12-hexahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-ol hydrobromide.

It is isolated from natural sources or produced by a synthetic process.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline or amorphous powder.

Solubility
Sparingly soluble in water, very slightly soluble in anhydrous ethanol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *galantamine hydrobromide CRS.*

B. Specific optical rotation or enantiomeric purity (see Tests).

C. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 0.60 g in *carbon dioxide-free water R* and dilute to 30.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.5 for solution S.

Specific optical rotation (*2.2.7*)
For galantamine from natural sources: −90 to −100 (dried substance), determined on Solution S.

Enantiomeric purity
For galantamine produced by a synthetic process. Capillary electrophoresis (*2.2.47*). *Prepare the solutions immediately before use.*

Buffer electrolyte 8.9 g/L solution of *disodium hydrogen phosphate dihydrate R.*

Test solution Dissolve 25.0 mg of the substance to be examined in 50.0 mL of *water R* and filter through a membrane filter (nominal pore size 0.22 µm).

Reference solution (a) Dissolve 5 mg of *galantamine racemic mixture CRS* in 10.0 mL of *water R*. Dilute 1.0 mL of this

solution to 100.0 mL with *water R*. Filter through a membrane filter (nominal pore size 0.22 µm).

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 with *water R*. Filter through a membrane filter (nominal pore size 0.22 µm).

Blank solution Filter *water R* through a membrane filter (nominal pore size 0.22 µm).

Capillary:
— *material*: uncoated fused silica;
— *size*: effective length = about 0.50 m, Ø = 75 µm.

Temperature 20 °C.

CZE buffer Dissolve 0.196 g of *α-cyclodextrin R* in 10.0 mL of buffer electrolyte and filter through a membrane filter (nominal pore size 0.22 µm).

Detection Spectrophotometer at 214 nm.

Preconditioning of the capillary At 137.9 kPa, rinse the capillary for 5 min with *water R* and for 5 min with CZE buffer.

Injection Under pressure (3.45 kPa) for 4 s.

Migration A voltage of 15kV.

Run time 35 min.

Relative migration times with reference to galantamine (retention time = about 18 min): impurity F = about 1.05.

System suitability Reference solution (a):
— *resolution*: minimum 2.5 between the peaks due to galantamine and to impurity F.

Limit:
— *impurity F*: not more than 1.5 times the area of the principal peak in the electropherogram obtained with reference solution (b) (0.15 per cent).

Related substances

Liquid chromatography (*2.2.29*).

A. Galantamine from natural sources

Solvent mixture Mobile phase B, mobile phase A (10:90 *V/V*).

Test solution Dissolve 12 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the same solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *galantamine natural for system suitability CRS* (containing impurities A and E) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.15 g of *ammonium formate R* in 900 mL of *water for chromatography R*, adjust to pH 3.8 with *anhydrous formic acid R* and dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	95	5
5 - 20	95 → 80	5 → 20
20 - 23	80 → 50	20 → 50
23 - 31	50 → 20	50 → 80
31 - 35	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 287 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *galantamine natural for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and E.

Relative retention With reference to galantamine (retention time = about 12 min): impurity E = about 0.8; impurity A = about 1.5.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurity E and galantamine.

Limits:
— *impurity E*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

B. Galantamine produced by a synthetic process

Solvent mixture Dilute 50 mL of *methanol R* to 1000 mL with *water R*.

Test solution Dissolve 0.10 g of the substance to be examined in 50.0 mL of the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.5 mg of *galantamine synthetic for system suitability CRS* (containing impurities C and D) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (3.5 µm);
— *temperature*: 55 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.79 g of *disodium hydrogen phosphate dihydrate R* and 2.46 g of *sodium dihydrogen phosphate R* in *water for chromatography R* and dilute to 1000 mL with *water for chromatography R*; to 950 mL of this solution, add 50 mL of *methanol R1*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 6	100	0
6 - 20	100 → 95	0 → 5
20 - 35	95 → 85	5 → 15
35 - 50	85 → 80	15 → 20

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *galantamine synthetic for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and D.

Relative retention With reference to galantamine (retention time = about 16 min): impurity C = about 0.8; impurity D = about 2.1.

System suitability Reference solution (b):
— *resolution*: minimum 4.5 between the peak due to impurity C and galantamine.

Limits:
— *impurities C, D*: for each impurity, not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Palladium

Maximum 10 ppm for galantamine produced by a synthetic process.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Transfer 1.000 g into an appropriate digestion system and digest using *nitric acid R*. After digestion, heat to dryness. Add 0.125 mL of *nitric acid R*, 0.375 mL of *hydrochloric acid R* and 2 mL of *water R*. Warm gently to dissolve any residue and allow to cool. Transfer quantitatively, by rinsing with several millilitres of *water R*, and dilute to 10.0 mL with *water R*.

Reference solution Use solutions containing 0.2 µg, 1.0 µg and 2.0 µg of palladium per millilitre, freshly prepared by dilution of *palladium standard solution (20 ppm Pd) R* with a mixture of 0.25 volumes of *nitric acid R*, 0.75 volumes of *hydrochloric acid R* and 25.0 volumes of *water R*.

Source Palladium hollow cathode lamp.

Wavelength 247.6 nm.

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 2.0 g.

ASSAY

Dissolve 0.275 g in 40 mL of *water R*. Add 40 mL of *ethanol (96 per cent) R*. Add 5 mL of *0.01 M hydrochloric acid*. Titrate with *0.1 M sodium hydroxide* determining the end-point potentiometrically. Read the volume between the 2 points of inflection.

1 mL of *0.1 M sodium hydroxide* is equivalent to 36.83 mg of $C_{17}H_{22}BrNO_3$.

LABELLING

The label states the origin of the substance:
— isolated from natural sources;
— produced by a synthetic process.

IMPURITIES

Test A for related substances

A, B, E.

Specified impurities *A, E.*

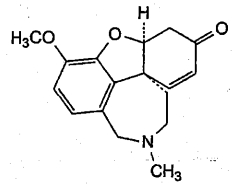
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*

Test B for related substances

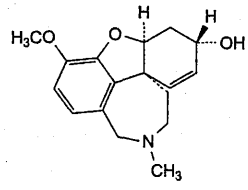
A, B, C, D, E, F.

Specified impurities *C, D, F.*

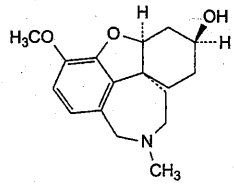
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, E.*



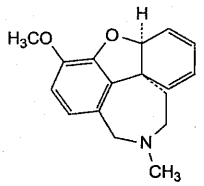
A. (4a*S*,8a*S*)-3-methoxy-11-methyl-4a,5,9,10,11,12-hexahydro-6*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-one (narwedine),



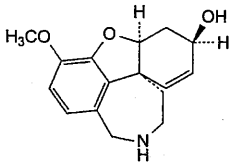
B. (4a*S*,6*S*,8a*S*)-3-methoxy-11-methyl-5,6,9,10,11,12-hexahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-ol (*epi*-galantamine),



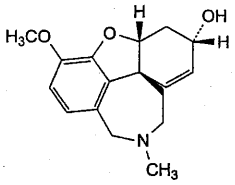
C. (4a*S*,6*S*,8a*R*)-3-methoxy-11-methyl-5,6,7,8,9,10,11,12-octahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-ol (dihydrogalantamine),



D. (4a*S*,8a*S*)-3-methoxy-11-methyl-9,10,11,12-tetrahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepine (anhydrogalantamine),



E. (4aS,6*R*,8a*S*)-3-methoxy-5,6,9,10,11,12-hexahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-ol (*N*-demethylgalantamine).

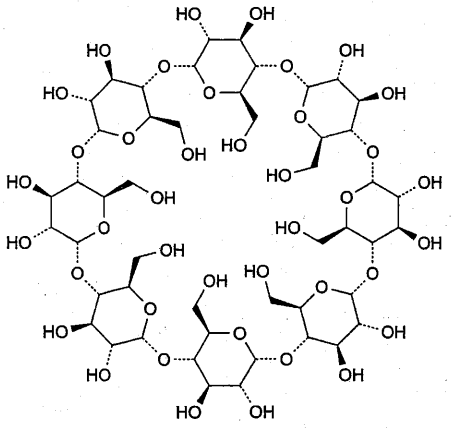


F. (4a*R*,6*S*,8a*R*)-3-methoxy-11-methyl-5,6,9,10,11,12-hexahydro-4a*H*-[1]benzofuro[3a,3,2-*ef*][2]benzazepin-6-ol (*ent*-galantamine).

Ph Eur

Gammadex

(Ph. Eur. monograph 2769)



$[C_6H_{10}O_5]_8$ 1297 *17465-86-0*

Ph Eur

DEFINITION

Cyclooctakis-(1→4)-(α-D-glucopyranosyl) (cyclomaltooctaose or γ-cyclodextrin).

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, amorphous or crystalline, hygroscopic powder.

Solubility

Freely soluble in water, very slightly soluble in propylene glycol, practically insoluble in anhydrous ethanol and in methylene chloride. When dissolved in water, it forms a colloidal dispersion over time.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gammacyclodextrin CRS.*

TESTS

Solution S

Dissolve 1.000 g in *carbon dioxide-free water R* and dilute to 100.0 mL with the same solvent.

pH (*2.2.3*)

5.0 to 8.0.

Mix 1 mL of a 223.6 g/L solution of *potassium chloride R* and 30 mL of solution S.

Specific optical rotation (*2.2.7*)

+ 174 to + 180 (dried substance), determined on solution S after filtration through a membrane filter (nominal pore size 0.45 µm).

Reducing sugars

Maximum 0.2 per cent.

Test solution To 1 mL of solution S add 1 mL of *cupri-tartaric solution R4*. Heat on a water-bath for 10 min and allow to cool to room temperature. Add 10 mL of *ammonium molybdate reagent R1* and allow to stand for 15 min.

Reference solution Prepare at the same time and in the same manner as for the test solution, using 1 mL of a 0.02 g/L solution of *glucose R*.

Measure the absorbances (*2.2.25*) of the 2 solutions at the absorption maximum at 740 nm using *water R* as the compensation liquid. The absorbance of the test solution is not greater than that of the reference solution.

Light-absorbing impurities

Examine solution S after filtration through a membrane filter (nominal pore size 0.45 µm). The absorbance (*2.2.25*) is not greater than 0.10 between 230 nm and 350 nm and not greater than 0.05 between 350 nm and 750 nm.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.250 g of the substance to be examined in *water R* with heating, cool and dilute to 25.0 mL with the same solvent.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 25.0 mg of *alfadex CRS* (impurity A), 25.0 mg of *betadex CRS* (impurity B) and 25.0 mg of *gammacyclodextrin CRS* (gammadex) in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with *water R*.

Reference solution (c) Dissolve 25.0 mg of *gammacyclodextrin CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *methanol R, water for chromatography R* (5:95 *V/V*).

Flow rate 1.5 mL/min.

Detection Differential refractometer maintained at a constant temperature (e.g. 35 °C).

Equilibration With the mobile phase for about 3 h.

Injection 50 µL of test solution (a) and reference solutions (a) and (b).

Run time 4 times the retention time of gammadex.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B.

Relative retention With reference to gammadex (retention time = about 7 min): impurity A = about 1.5; impurity B = about 3.2.

System suitability Reference solution (a):
— *resolution*: minimum 2.5 between the peaks due to gammadex and impurity A;
— *symmetry factor*: 0.8 to 1.8 for the peak due to gammadex.

Calculation of percentage contents:
— for impurities A and B, use the concentration of the corresponding impurity in reference solution (b);
— for impurities other than A and B, use the concentration of gammadex in reference solution (b).

Limits:
— *impurities A, B*: for each impurity, maximum 0.5 per cent;
— *sum of impurities other than A and B*: maximum 0.5 per cent;
— *reporting threshold*: 0.15 per cent.

Loss on drying (*2.2.32*)
Maximum 11.0 per cent, determined on 1.000 g by drying in an oven at 120 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

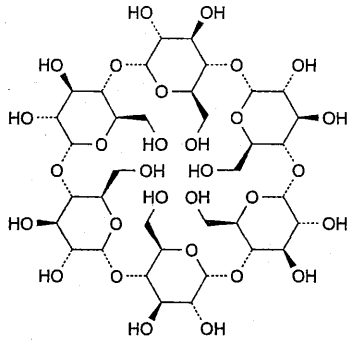
Injection Test solution (b) and reference solution (c).

System suitability Reference solution (c):
— *symmetry factor*: 0.8 to 1.8 for the peak due to gammadex;
— *repeatability*: maximum relative standard deviation of 2.0 per cent determined on 5 injections.

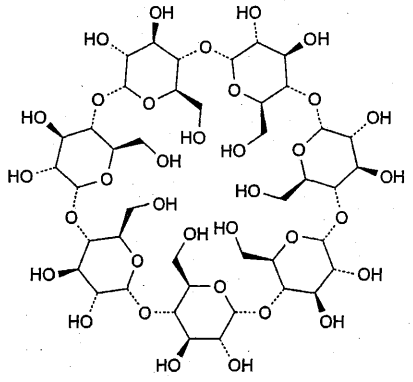
Calculate the percentage content of $[C_6H_{10}O_5]_8$ taking into account the assigned content of *gammacyclodextrin CRS*.

STORAGE
In an airtight container.

IMPURITIES
Specified impurities *A, B.*



A. cyclohexakis-(1→4)-(α-D-glucopyranosyl) (alfadex or cyclomaltohexaose or α-cyclodextrin),

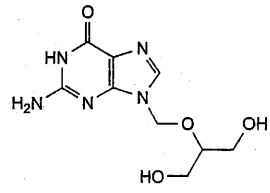


B. cycloheptakis-(1→4)-(α-D-glucopyranosyl) (betadex or cyclomaltoheptaose or β-cyclodextrin).

Ph Eur

Ganciclovir

(Ph. Eur. monograph 1752)



$C_9H_{13}N_5O_4$ 255.2 *82410-32-0*

Action and use
Antiviral (cytomegalovirus).

Ph Eur

DEFINITION
2-Amino-9-[[2-hydroxy-1-(hydroxymethyl)ethoxy]methyl]-1,9-dihydro-6*H*-purin-6-one.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Slightly soluble in water, very slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of mineral acids and alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ganciclovir CRS.*

If the spectra obtained in the solid state show differences, dissolve 0.10 g of the substance to be examined and the reference substance separately in about 3.6 mL of *water R* at 80 °C. Allow to cool in an ice-bath and filter the precipitate. Dry in an oven at 105 °C for 3 h and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 1.25 g in a 40 g/L solution of *sodium hydroxide R* and dilute to 25 mL with the same solution.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase with the aid of ultrasound and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 3 mg of *ganciclovir CRS* in the mobile phase with the aid of ultrasound and dilute to 5.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of *ganciclovir impurity mixture CRS* (impurities A, B, C, D, E and F) in 1.0 mL of reference solution (b).

Column:
- *size: l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase: strong cation-exchange silica gel for chromatography R* (10 µm);
- *temperature*: 40 °C.

Mobile phase Mix equal volumes of *acetonitrile R* and a 0.05 per cent *V/V* solution of *trifluoroacetic acid R.*

Flow rate 1.5 mL/min.

Detection Spectophotometer at 254 nm.

Injection 20 µL.

Run time 2.5 times the retention time of ganciclovir.

Identification of impurities Use the chromatogram supplied with *ganciclovir impurity mixture CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D, E and F.

Relative retention With reference to ganciclovir (retention time = about 14 min): impurity A = about 0.6; impurity B = about 0.67; impurity C = about 0.71; impurity D = about 0.8; impurity E = about 0.9; impurity F = about 2.0.

System suitability Reference solution (c):
- *peak-to-valley ratio*: minimum 5, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to ganciclovir.

Limits:
- *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.3; impurity F = 0.7;
- *impurity F*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
- *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *impurities A, C, D, E*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
- *total*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
- *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Water (*2.5.12*)

Maximum 4.0 per cent, determined on 0.300 g.

Use *methanol R* as solvent. The substance to be examined has limited solubility in methanol. The sample will appear as a slurry. Replace the solvent after each titration.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)

Less than 0.84 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Dissolve 0.200 g in 10 mL of *anhydrous formic acid R* and dilute to 60 mL with *glacial acetic acid R.* Titrate with *0.1 M perchloric acid*, determining the end point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 25.52 mg of $C_9H_{13}N_5O_4$.

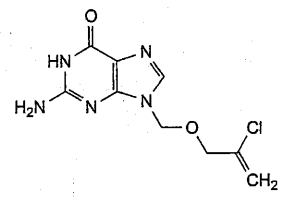
STORAGE

In an airtight container.

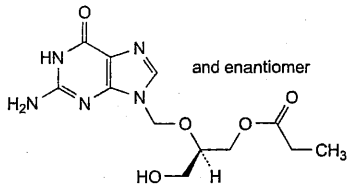
IMPURITIES

Specified impurities *A, B, C, D, E, F.*

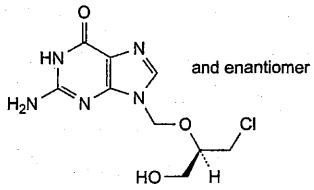
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *H, I, J.*



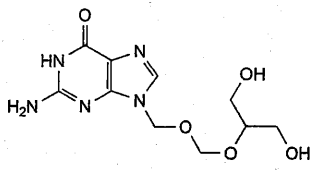
A. 2-amino-9-[[(2-chloroprop-2-en-1-yl)oxy]methyl]-1,9-dihydro-6*H*-purin-6-one,



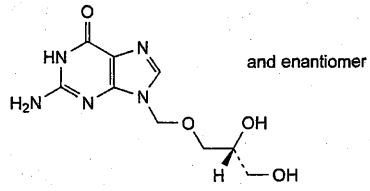
B. (2*RS*)-2-[(2-amino-6-oxo-1,6-dihydro-9*H*-purin-9-yl) methoxy]-3-hydroxypropyl propionate,



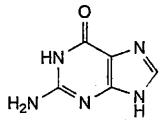
C. 2-amino-9-[[(1*RS*)-2-chloro-1-(hydroxymethyl) ethoxy]methyl]-1,9-dihydro-6*H*-purin-6-one,



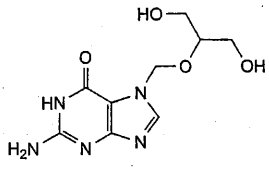
D. 2-amino-9-[[[2-hydroxy-1-(hydroxymethyl) ethoxy]methoxy]methyl]-1,9-dihydro-6*H*-purin-6-one,



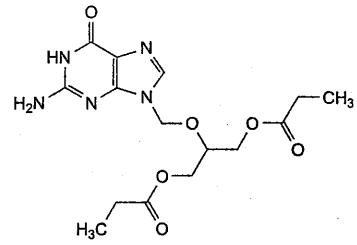
E. 2-amino-9-[[(2*RS*)-2,3-dihydroxypropoxy]methyl]-1,9-dihydro-6*H*-purin-6-one,



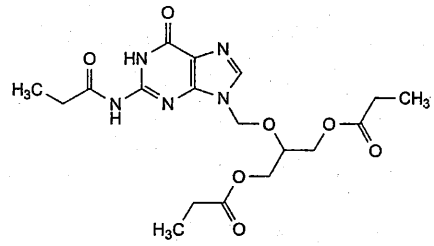
F. 2-amino-1,9-dihydro-6*H*-purin-6-one (guanine),



H. 2-amino-7-[[2-hydroxy-1-(hydroxymethyl)ethoxy]methyl]-1,7-dihydro-6*H*-purin-6-one,



I. 2-[(2-amino-6-oxo-1,6-dihydro-9*H*-purin-9-yl)methoxy] propane-1,3-diyl dipropanoate,

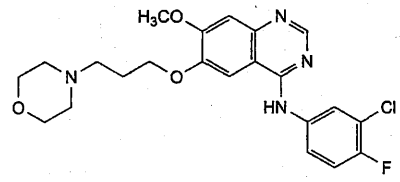


J. 2-[2-(propanoylamino)-6-oxo-1,6-dihydro-9*H*-purin-9-yl] methoxy]propane-1,3-diyl dipropanoate.

Ph Eur

Gefitinib

(Ph. Eur. monograph 2866)



$C_{22}H_{24}ClFN_4O_3$ 446.9 *184475-35-2*

Action and use
Cytotoxic.

Ph Eur

DEFINITION
N-(3-Chloro-4-fluorophenyl)-7-methoxy-6-[3-(morpholin-4-yl)propoxy]quinazolin-4-amine.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, slightly soluble in anhydrous ethanol, practically insoluble in heptane.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *gefitinib CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*).
Solvent mixture *acetonitrile R*, 0.2 per cent *V/V* solution of *trifluoroacetic acid R* (40:60 *V/V*).

Test solution Dissolve 35.0 mg of the substance to be examined in 85 mL of the solvent mixture, with the aid of ultrasound, and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 35.0 mg of *gefitinib CRS* in 85 mL of the solvent mixture, with the aid of ultrasound, and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *gefitinib for system suitability CRS* (containing impurity B) in 1.0 mL of the solvent mixture with the aid of ultrasound.

Column:
— *size*: *l* = 0.10 m, Ø = 3 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 60 °C.

Mobile phase *acetonitrile R*, 9.68 g/L solution of *ammonium acetate R* (38:62 *V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 247 nm.

Injection 5 µL of the test solution and reference solutions (b) and (c).

Run time 5 times the retention time of gefitinib.

Identification of impurities Use the chromatogram supplied with *gefitinib for system suitability CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity B.

Relative retention With reference to gefitinib (retention time = about 5.5 min): impurity B = about 1.3.

System suitability Reference solution (c):
— *resolution*: minimum 3.0 between the peaks due to gefitinib and impurity B.

Calculation of percentage contents:
— for each impurity, use the concentration of gefitinib in reference solution (b).

Limits:
— *impurity B*: maximum 0.2 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.4 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.32*)
Maximum 0.5 per cent, determined on 0.100 g by direct sample introduction.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

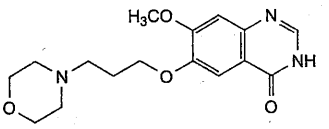
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{22}H_{24}ClFN_4O_3$ taking into account the assigned content of *gefitinib CRS*.

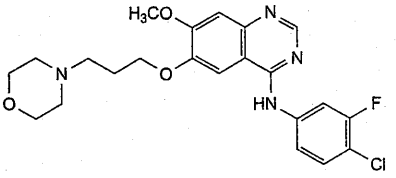
IMPURITIES

Specified impurities *B*.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A*.



A. 7-methoxy-6-[3-(morpholin-4-yl)propoxy]quinazolin-4(3*H*)-one,



B. *N*-(4-chloro-3-fluorophenyl)-7-methoxy-6-[3-(morpholin-4-yl)propoxy]quinazolin-4-amine.

Ph Eur

Gelatin[1]

(Ph. Eur. monograph 0330)

Action and use
Excipient.

Ph Eur

DEFINITION

Purified protein obtained from collagen of animals by partial alkaline and/or acid hydrolysis and/or enzymatic hydrolysis, or by thermal hydrolysis.

The hydrolysis leads to gelling or non-gelling product grades. This monograph covers both grades.

♦CHARACTERS

Appearance
— *gelling grade*: faintly yellow or light yellowish-brown solid, usually occurring as translucent sheets, shreds, granules or powder;
— *non-gelling grade*: faintly yellow or white granules or powder.

Solubility
— *gelling grade*: practically insoluble in common organic solvents; gelling grades swell in cold water and give on heating a colloidal solution which on cooling forms a more or less firm gel;
— *non-gelling grade*: soluble in cold or warm water, practically insoluble in common organic solvents.

Different gelatins form aqueous solutions that vary in clarity and colour. For a particular application, a suitable specification for clarity and colour may be required.♦

IDENTIFICATION

Gelling grade: A, B.

Non-gelling grade: A, B, C.

A. To 2 mL of solution S (see Tests) add 0.05 mL of *copper sulfate solution R*. Mix and add 0.5 mL of *dilute sodium hydroxide solution R*. A violet colour is produced.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

B. In a test-tube about 15 mm in internal diameter, place 0.5 g of the substance to be examined and add 10 mL of *water R*. Allow to stand for 10 min, heat at 60 °C for 15 min and keep the tube upright at 0 °C for 6 h. Invert the tube; the contents flow out immediately for non-gelling grades and do not flow out immediately for gelling grades.

C. To 0.5 g in a 250 mL bottle, add 10 mL of *water R* and 5 mL of *sulfuric acid R*. Place the bottle, partly but not completely closed (for example, using a watch glass), in an oven at 105 °C for 4 h. Allow to cool and add 200 mL of *water R*. Adjust to pH 6.0-8.0 using a 200 g/L solution of *sodium hydroxide R*. Place 2 mL of the solution in a test-tube and add 2 mL of a solution prepared immediately before use containing 14 g/L of *chloramine R* in phosphate buffer solution pH 6.8 R. Mix and allow to stand for 20 min. Add 2 mL of *dimethylaminobenzaldehyde solution R9*. Mix and place in a water-bath at 60 °C for 15 min. A red to violet colour develops.

TESTS

Solution S

Dissolve 1.00 g in *carbon dioxide-free water R* at about 55 °C, dilute to 100 mL with the same solvent and keep the solution at this temperature to carry out the tests.

pH (*2.2.3*)

3.8 to 7.6 for solution S, measured at 55 °C.

Conductivity (*2.2.38*)

Maximum 1 $mS \cdot cm^{-1}$, determined on a 1.0 per cent solution at 30 ± 1.0 °C (without the use of the temperature compensation device).

Sulfur dioxide (*2.5.29*)

Maximum 50 ppm.

Peroxides

Maximum 10 ppm, determined using *peroxide test strips R*.

Peroxidase transfers oxygen from peroxides to an organic redox indicator which is converted to a blue oxidation product. The intensity of the colour obtained is proportional to the quantity of peroxide and can be compared with a colour scale provided with the test strips, to determine the peroxide concentration.

Suitability test Dip a test strip for 1 s into *hydrogen peroxide standard solution (2 ppm H_2O_2) R*, such that the reaction zone is properly wetted. Remove the test strip, shake off excess liquid and after 15 s compare the reaction zone with the colour scale provided. The test strips are suitable if the colour matches that of the 2 ppm concentration.

Test Weigh 20.0 ± 0.1 g of the substance to be examined in a beaker and add 80.0 ± 0.2 mL of *water R*. Stir to moisten all the gelatin and allow the sample to stand at room temperature for 1-3 h. Cover the beaker with a watch-glass. If dissolution is not complete, place the beaker for 20 ± 5 min in a water-bath at 65 ± 2 °C to dissolve the sample. Stir the contents of the beaker with a glass rod to achieve a homogeneous solution. Dip a test strip for 1 s into the test solution, such that the reaction zone is properly wetted. Remove the test strip, shake off excess liquid and after 15 s compare the reaction zone with the colour scale provided. Multiply the concentration read from the colour scale by a factor of 5 to calculate the concentration in parts per million of peroxide in the substance to be examined.

Gel strength (Bloom value)

80 per cent to 120 per cent of the nominal value stated on the label for the gelling grade.

The gel strength is expressed as the mass in grams necessary to produce the force which, applied to a plunger 12.7 mm in diameter, makes a depression 4 mm deep in a gel having a concentration of 6.67 per cent *m/m* and matured at 10 °C.

Apparatus Texture analyser or gelometer with:
- a cylindrical piston 12.7 ± 0.1 mm in diameter with a plane pressure surface with a sharp bottom edge;
- a bottle 59 ± 1 mm in internal diameter and 85 mm high.

Adjust the apparatus according to the manufacturer's manual. Settings are: distance 4 mm, test speed 0.5 mm/s.

Method Place 7.5 g of the substance to be examined in a bottle. Add 105 mL of *water R*, close the bottle and allow to stand for 1-4 h. Heat in a water-bath at 65 ± 2 °C for 15 min. While heating, stir gently with a glass rod. Ensure that the solution is uniform and that any condensed water on the inner walls of the bottle is incorporated. Allow to cool at room temperature for 15 min and transfer the bottle to a thermostatically controlled bath at 10.0 ± 0.1 °C, and fitted with a device to ensure that the platform on which the bottle stands is perfectly horizontal. Close the bottle with a rubber stopper and allow to stand for 17 ± 1 h. Remove the bottle from the bath and quickly wipe the water from the exterior of the bottle. Centre the bottle on the platform of the apparatus so that the plunger contacts the sample as near to its midpoint as possible and start the measurement.

Iron

Maximum 30 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution To 5.00 g of the substance to be examined, in a conical flask, add 10 mL of *hydrochloric acid R*. Close the flask and place in a water-bath at 75-80 °C for 2 h (if necessary for proper solubilisation, the gelatin may be allowed to swell after addition of the acid and before heating, the heating time may be prolonged, and a higher temperature may be used). Allow to cool and adjust the contents of the flask to 100.0 g with *water R*.

Reference solutions Prepare the reference solutions using *iron standard solution (8 ppm Fe) R*, diluting with *water R*.

Wavelength 248.3 nm.

Chromium

Maximum 10 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Test solution described in the test for iron.

Reference solutions Prepare the reference solutions using *chromium standard solution (100 ppm Cr) R*, diluting with *water R*.

Wavelength 357.9 nm.

Zinc

Maximum 30 ppm.

Atomic absorption spectrometry (*2.2.23, Method II*).

Test solution Test solution described in the test for iron.

Reference solutions Prepare the reference solutions using *zinc standard solution (10 ppm Zn) R*, diluting with *water R*.

Wavelength 213.9 nm.

Loss on drying (*2.2.32*)

Maximum 15.0 per cent, determined on 5.000 g by drying in an oven at 105 °C for 16 h.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

STORAGE

Protect from heat and moisture.

LABELLING

The label states the gel strength (Bloom value) or that it is a non-gelling grade.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for gelling grade gelatin used as viscosity-increasing agent, binder or used for microencapsulation.

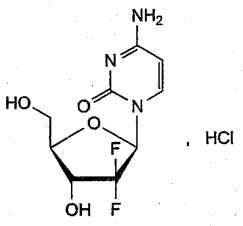
Gel strength (Bloom value)

(see Tests).

Ph Eur

Gemcitabine Hydrochloride

(Ph. Eur. monograph 2306)



$C_9H_{12}ClF_2N_3O_4$ 299.7 *122111-03-9*

Action and use

Pyrimidine analogue; cytotoxic.

Ph Eur

DEFINITION

4-Amino-1-(2-deoxy-2,2-difluoro-β-D-*erythro*-pentofuranosyl) pyrimidin-2(1*H*)-one hydrochloride.

Content

98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Soluble in water, slightly soluble in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gemcitabine hydrochloride CRS.*

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 1.00 g in *carbon dioxide-free water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

pH (*2.2.3*)

2.0 to 3.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 43.0 to + 50.0, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in *water R* and dilute to 200.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of the substance to be examined and 10.0 mg of *gemcitabine impurity A CRS* in *water R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with *water R*.

Reference solution (b) Dissolve 20.0 mg of *gemcitabine hydrochloride CRS* in *water R* and dilute to 200.0 mL with the same solvent.

Reference solution (c) Place 10 mg of the substance to be examined in a small vial. Add 4 mL of a 168 g/L solution of *potassium hydroxide R* in *methanol R*, sonicate for 5 min then seal with a cap. The mixture may be cloudy. Heat at 55 °C for a minimum of 6 h to produce impurity B. Allow to cool, then transfer the entire contents of the vial to a 100 mL volumetric flask by successively washing with a 1 per cent *V/V* solution of *phosphoric acid R*. Dilute to 100 mL with a 1 per cent *V/V* solution of *phosphoric acid R* and mix.

Reference solution (d) Dissolve 2 mg of *gemcitabine impurity C CRS* in *water R* and dilute to 5.0 mL with the same solvent.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: 13.8 g/L solution of *sodium dihydrogen phosphate monohydrate R* adjusted to pH 2.5 ± 0.1 with *phosphoric acid R*;

— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 8	97	3
8 - 13	97 → 50	3 → 50
13 - 20	50	50

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity C.

Relative retention With reference to gemcitabine (retention time = about 7 min): impurity A = about 0.4; impurity B = about 0.7; impurity C = about 1.7.

System suitability Reference solution (c):
— *resolution*: minimum 8.0 between the peaks due to impurity B and gemcitabine.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 2.4;
— *impurity C*: not more than twice the area of the peak due to gemcitabine in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity A*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the peak due to gemcitabine in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 4 times the area of the peak due to gemcitabine in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *disregard limit*: 0.5 times the area of the peak due to gemcitabine in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

Bacterial endotoxins (*2.6.14*)
Less than 0.05 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase *methanol R*, mobile phase A (3:97 *V/V*).

Injection Test solution (b) and reference solutions (b) and (c).

Relative retention With reference to gemcitabine (retention time = about 8 min): impurity B = about 0.5.

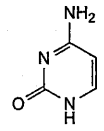
System suitability Reference solution (c):
— *resolution*: minimum 8.0 between the peaks due to impurity B and gemcitabine.

Calculate the percentage content of $C_9H_{12}ClF_2N_3O_4$ taking into account the assigned content of *gemcitabine hydrochloride CRS*.

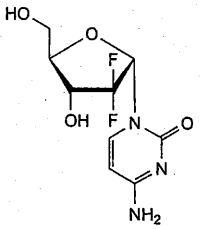
STORAGE
If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES
Specified impurities *A, C.*

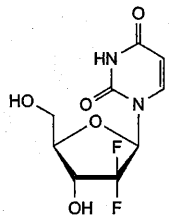
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B.*



A. 4-aminopyrimidin-2(1*H*)-one (cytosine),



B. 4-amino-1-(2-deoxy-2,2-difluoro-α-D-*erythro*-pentofuranosyl)pyrimidin-2(1*H*)-one (gemcitabine α-anomer),

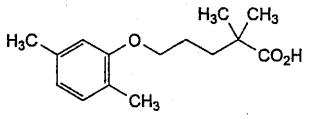


C. 1-(2-deoxy-2,2-difluoro-β-D-*erythro*-pentofuranosyl) pyrimidin-2,4(1*H*,3*H*)-dione (2′-deoxy-2′,2′-difluorouridine) (β-uridine).

Ph Eur

Gemfibrozil

(Ph. Eur. monograph 1694)



$C_{15}H_{22}O_3$ 250.3 *25812-30-0*

Action and use
Fibrate; lipid-regulating drug.

Preparations
Gemfibrozil Capsules
Gemfibrozil Tablets

Ph Eur

DEFINITION
5-(2,5-Dimethylphenoxy)-2,2-dimethylpentanoic acid.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, waxy, crystalline powder.

Solubility
Practically insoluble in water, very soluble in methylene chloride, freely soluble in anhydrous ethanol and in methanol.

IDENTIFICATION

A. Melting point (*2.2.14*): 58 °C to 61 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gemfibrozil CRS.*

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve the contents of a vial of *gemfibrozil for system suitability CRS* (containing impurities C, D and E) in 2.0 mL of *acetonitrile R.*

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of *2,5-dimethylphenol R* (impurity A) in mobile phase A and dilute to 10.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: dissolve 0.49 g of *potassium acetate R* in 400 mL of *water for chromatography R*, adjust to pH 4.0 with *phosphoric acid R* and add 600 mL of *acetonitrile R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	100	0
5 - 20	100 → 0	0 → 100
20 - 25	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *gemfibrozil for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities C, D and E; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to gemfibrozil (retention time = about 7 min): impurity A = about 0.4; impurity C = about 1.3; impurity D = about 1.5; impurity E = about 1.7; impurity I = about 2.0; impurity H = about 2.9.

System suitability Reference solution (a):
— *resolution*: minimum 6.0 between the peaks due to gemfibrozil and impurity C, and minimum 2.0 between the peaks due to impurities D and E.

Limits:
— *correction factors*: for the calculation of content multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.5; impurity D = 1.8; impurity E = 0.2; impurity H = 0.6;
— *impurities E, I*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurities A, D, H*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
Maximum 0.25 per cent, determined on 2.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 2.0 g. Allow to stand for 1 h after the first moistening before heating.

ASSAY

Dissolve 0.200 g in 40 mL of *methanol R.* Add 10 mL of *water R* and 1 mL of *0.1 M hydrochloric acid.* Carry out a potentiometric titration (*2.2.20*) using *0.1 M sodium hydroxide.* Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 25.03 mg of $C_{15}H_{22}O_3$.

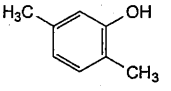
STORAGE

Protected from light.

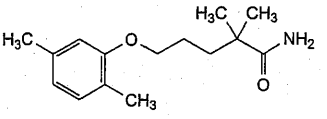
IMPURITIES

Specified impurities *A, D, E, H, I.*

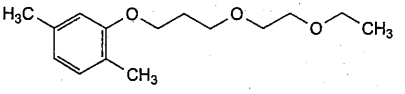
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, F, G.*



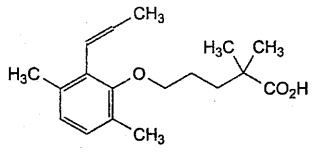
A. 2,5-dimethylphenol (*p*-xylenol),



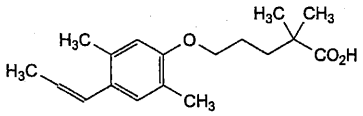
B. 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanamide,



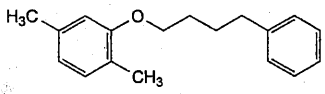
C. 2-[3-(2-ethoxyethoxy)propoxy]-1,4-dimethylbenzene,



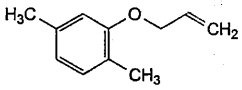
D. 5-[3,6-dimethyl-2-(prop-1-en-1-yl)phenoxy]-2,2-dimethylpentanoic acid,



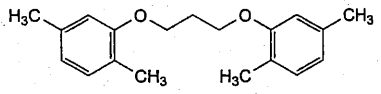
E. 5-[2,5-dimethyl-4-(prop-1-en-1-yl)phenoxy]-2,2-dimethylpentanoic acid,



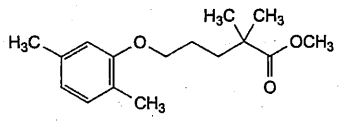
F. 1,4-dimethyl-2-(4-phenylbutoxy)benzene,



G. 1,4-dimethyl-2-(prop-2-en-1-yloxy)benzene,



H. 1,1′-[propane-1,3-diylbis(oxy)]bis(2,5-dimethylbenzene),

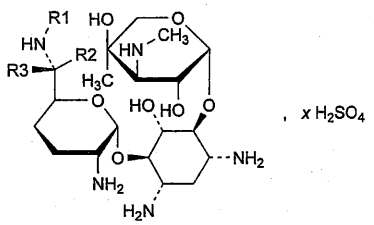


I. methyl 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoate.

Ph Eur

Gentamicin Sulfate

(Ph. Eur. monograph 0331)



Gentamicin	Mol. Formula	R1	R2	R3
C1	$C_{21}H_{43}N_5O_7$	CH_3	CH_3	H
C1a	$C_{19}H_{39}N_5O_7$	H	H	H
C2	$C_{20}H_{41}N_5O_7$	H	CH_3	H
C2a	$C_{20}H_{41}N_5O_7$	H	H	CH_3
C2b	$C_{20}H_{41}N_5O_7$	CH_3	H	H

1405-41-0

Action and use
Aminoglycoside antibacterial.

Preparations
Gentamicin Cream
Gentamicin Ear Drops
Gentamicin and Hydrocortisone Acetate Ear Drops
Gentamicin Eye Drops
Gentamicin Injection
Gentamicin Ointment

Ph Eur

DEFINITION
Mixture of the sulfates of antimicrobial substances produced by *Micromonospora purpurea*, the main components being gentamicins C1, C1a, C2, C2a and C2b.

Content
Minimum 590 IU/mg (anhydrous substance).

CHARACTERS
Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B, C.

Second identification: A, C.

A. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 25 mg of the substance to be examined in *water R* and dilute to 5 mL with the same solvent.

Reference solution Dissolve the contents of a vial of *gentamicin sulfate CRS* in *water R* and dilute to 5 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase The lower layer of a mixture of equal volumes of *concentrated ammonia R*, *methanol R* and *methylene chloride R.*

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R1* and heat at 110 °C for 5 min.

Results The 3 principal spots in the chromatogram obtained with the test solution are similar in position, colour and size to the 3 principal spots in the chromatogram obtained with the reference solution.

B. Examine the chromatograms obtained in the test for composition.

Results The chromatogram obtained with test solution (b) shows 5 principal peaks having the same retention times as the 5 principal peaks in the chromatogram obtained with reference solution (a).

C. It gives reaction (a) of sulfates (*2.3.1*).

TESTS
Solution S
Dissolve 0.8 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

pH (*2.2.3*)
3.5 to 5.5 for solution S.

Specific optical rotation (*2.2.7*)
+ 107 to + 121 (anhydrous substance).

Dissolve 2.5 g in *water R* and dilute to 25.0 mL with the same solvent.

Composition
Liquid chromatography (*2.2.29*): use the normalisation procedure taking into account only the peaks due to gentamicins C1, C1a, C2, C2a and C2b.

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *gentamicin for peak identification CRS* (containing impurity B) in the mobile phase and dilute to 25 mL with the mobile phase.

Reference solution (b) Dissolve 20.0 mg of *sisomicin sulfate CRS* (impurity A) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 100.0 mL with the mobile phase.

Reference solution (d) To 1 mL of reference solution (b), add 5 mL of test solution (a) and dilute to 50 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase To 900 mL of *carbon dioxide-free water R*, add 7.0 mL of *trifluoroacetic acid R*, 250.0 µL of *pentafluoropropanoic acid R* and 4.0 mL of *carbonate-free sodium hydroxide solution R*, allow to equilibrate and adjust to pH 2.6 using *carbonate-free sodium hydroxide solution R* diluted 1 to 25. Add 15 mL of *acetonitrile R* and dilute to 1000.0 mL with *carbon dioxide-free water R*.

Flow rate 1.0 mL/min.

Post-column solution *carbonate-free sodium hydroxide solution R* diluted 1 to 25, previously degassed, which is added pulse-less to the column effluent using a 375 µL polymeric mixing coil.

Flow rate of post-column solution 0.3 mL/min.

Detection Pulsed amperometric detector or equivalent with a gold indicator electrode, a silver-silver chloride reference electrode, and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V oxidation and −0.15 V reduction potentials, with pulse durations according to the instrument used.

Injection 20 µL of test solution (b) and reference solutions (a), (c) and (d).

Run time 1.2 times the retention time of gentamicin C1.

Identification of peaks Use the chromatogram supplied with *gentamicin for peak identification CRS* to identify the peaks due to gentamicins C1, C1a, C2, C2a and C2b.

Relative retention With reference to impurity A (retention time = about 23 min): gentamicin C1a = about 1.1; gentamicin C2 = about 1.8; gentamicin C2b = about 2.0; gentamicin C2a = about 2.3; gentamicin C1 = about 3.0.

System suitability:
— *resolution*: minimum 1.2 between the peaks due to impurity A and gentamicin C1a and minimum 1.5 between the peaks due to gentamicin C2 and gentamicin C2b in the chromatogram obtained with reference solution (d); if necessary, adjust the volume of *acetonitrile R* in the mobile phase, a total volume of up to 50 mL may be added per litre of mobile phase;
— *signal-to-noise ratio*: minimum 20 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:
— *gentamicin C1*: 25.0 per cent to 45.0 per cent;
— *gentamicin C1a*: 10.0 per cent to 30.0 per cent;
— *sum of gentamicins C2, C2a and C2b*: 35.0 per cent to 55.0 per cent.

Related substances
Liquid chromatography (*2.2.29*) as described in the test for composition with the following modifications; use reference solution (c) to calculate the percentage content of each impurity.

Injection 20 µL of test solution (a) and reference solutions (a) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity A; use the chromatogram supplied with *gentamicin for peak identification CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity B.

Limits:
— *impurities A, B*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
— *any other impurity*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (10 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent).

Methanol (*2.4.24, System B*)
Maximum 1.0 per cent.

Sulfate
32.0 per cent to 35.0 per cent (anhydrous substance).

Dissolve 0.250 g in 100 mL of *distilled water R* and adjust the solution to pH 11 using *concentrated ammonia R*.
Add 10.0 mL of *0.1 M barium chloride* and about 0.5 mg of *phthalein purple R*. Titrate with *0.1 M sodium edetate*, adding 50 mL of *ethanol (96 per cent) R* when the colour of the solution begins to change and continue the titration until the violet-blue colour disappears.

1 mL of *0.1 M barium chloride* is equivalent to 9.606 mg of SO_4.

Water (*2.5.12*)
Maximum 15.0 per cent, determined on 0.300 g.

Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 0.50 g.

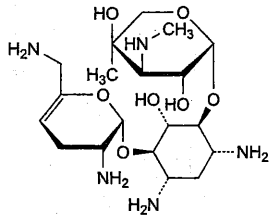
Bacterial endotoxins (*2.6.14*)
Less than 0.71 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Carry out the microbiological assay of antibiotics (*2.7.2*).

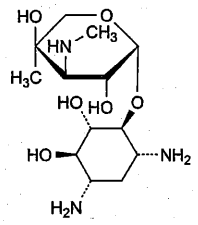
STORAGE
In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES
Specified impurities *A, B.*

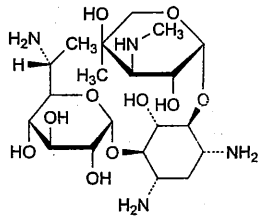
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E.*



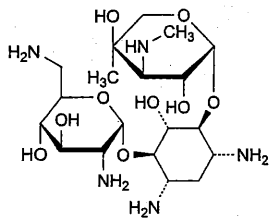
A. 2-deoxy-4-*O*-[3-deoxy-4-*C*-methyl-3-(methylamino)-β-L-arabinopyranosyl]-6-*O*-(2,6-diamino-2,3,4,6-tetradeoxy-α-D-*glycero*-hex-4-enopyranosyl)-L-streptamine (sisomicin),



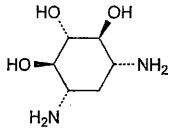
B. 2-deoxy-4-*O*-[3-deoxy-4-*C*-methyl-3-(methylamino)-β-L-arabinopyranosyl]-L-streptamine (garamine),



C. 4-*O*-(6-amino-6,7-dideoxy-D-*glycero*-α-D-*gluco*-heptopyranosyl)-2-deoxy-6-*O*-[3-deoxy-4-*C*-methyl-3-(methylamino)-β-L-arabinopyranosyl]-D-streptamine (gentamicin B_1),



D. 2-deoxy-4-*O*-[3-deoxy-4-*C*-methyl-3-(methylamino)-β-L-arabinopyranosyl]-6-*O*-(2,6-diamino-2,6-dideoxy-α-D-*gluco*-hexopyranosyl)-L-streptamine,

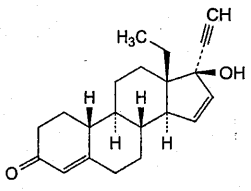


E. 2-deoxystreptamine.

Ph Eur

Gestodene

(Ph. Eur. monograph 1726)



$C_{21}H_{26}O_2$ 310.4 *60282-87-3*

Action and use
Progestogen.

Ph Eur

DEFINITION
13-Ethyl-17-hydroxy-18,19-dinor-17α-pregna-4,15-dien-20-yn-3-one.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or yellowish, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in methanol, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *gestodene CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS
Specific optical rotation (*2.2.7*)
−188 to −198 (dried substance).

Dissolve 0.100 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water R* (50:50 *V/V*).

Test solution (a) Dissolve 30.0 mg of the substance to be examined in 5 mL of *acetonitrile R1* and dilute to 10.0 mL with *water R*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 3 mg of *gestodene for system suitability CRS* (containing impurities A, B, C and L) in 0.5 mL of *acetonitrile R1* and dilute to 1.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 30.0 mg of *gestodene CRS* in 5 mL of *acetonitrile R1* and dilute to 10.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve the contents of a vial of *gestodene impurity I CRS* in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: spherical *end-capped octylsilyl silica gel for chromatography R* (3.5 µm).

Mobile phase:
— *mobile phase A*: *water R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	62	38
2 - 20	62 → 58	38 → 42
20 - 24	58 → 30	42 → 70
24 - 32	30	70

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm and at 254 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram supplied with *gestodene for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C and L; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity I.

Relative retention With reference to gestodene (retention time = about 12.5 min): impurity A = about 0.9; impurity C = about 1.1; impurity I = about 1.2; impurity L = about 1.46; impurity B = about 1.53.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity A and gestodene.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 2.2; impurity I = 1.3;
— *impurity A at 254 nm*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurity B at 205 nm*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurity C at 254 nm*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurities I, L at 205 nm*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— *unspecified impurities at 254 nm*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total at 254 nm*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit at 254 nm*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (c).

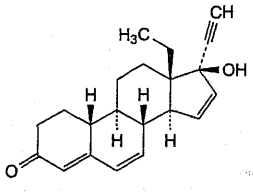
Detection Spectrophotometer at 254 nm.

Calculate the percentage content of $C_{21}H_{26}O_2$ from the declared content of *gestodene CRS*.

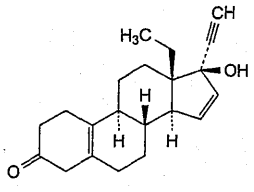
IMPURITIES

Specified impurities *A, B, C, I, L.*

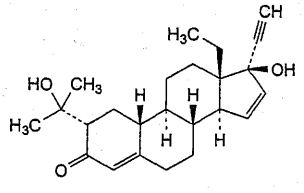
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use)
— *at 205 nm*: *G, J, K*;
— *at 254 nm*: *D, E, F, H.*



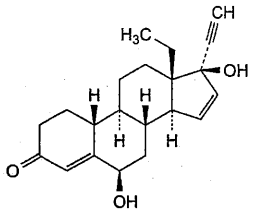
A. 13-ethyl-17-hydroxy-18,19-dinor-17α-pregna-4,6,15-trien-20-yn-3-one (Δ6-gestodene),



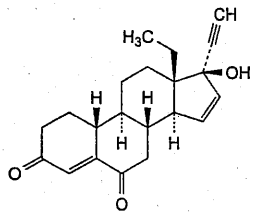
B. 13-ethyl-17-hydroxy-18,19-dinor-17α-pregna-5(10),15-dien-20-yn-3-one (Δ5(10)-gestodene),



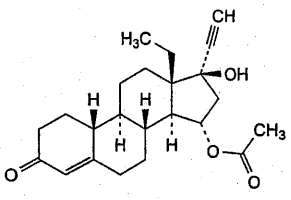
C. 13-ethyl-17-hydroxy-2α-(1-hydroxy-1-methylethyl)-18,19-dinor-17α-pregna-4,15-dien-20-yn-3-one (2-isopropanol-gestodene),



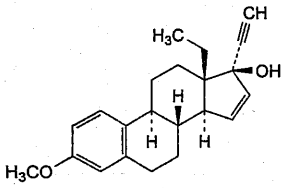
D. 13-ethyl-6β,17-dihydroxy-18,19-dinor-17α-pregna-4,15-dien-20-yn-3-one (6β-hydroxy-gestodene),



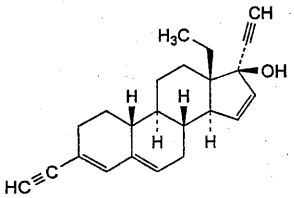
E. 13-ethyl-17-hydroxy-18,19-dinor-17α-pregna-4,15-dien-20-yne-3,6-dione (6-keto-gestodene),



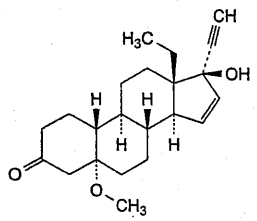
F. 13-ethyl-17-hydroxy-3-oxo-18,19-dinor-17α-pregn-4-en-20-yn-15α-yl acetate (15α-acetoxy-gestodene),



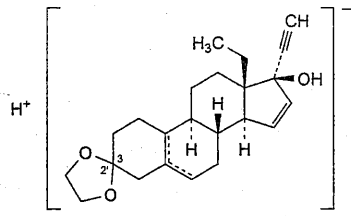
G. 13-ethyl-3-methoxy-18,19-dinor-17α-pregna-1,3,5(10),15-tetraen-20-yn-17-ol (*A*-aromatic-gestodene),



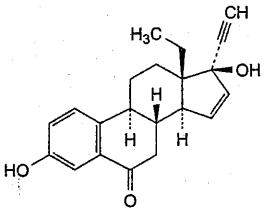
H. 13-ethyl-3-ethynyl-18,19-dinor-17α-pregna-3,5,15-trien-20-yn-17-ol (diethynyl-gestodene),



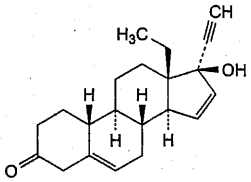
I. 13-ethyl-17-hydroxy-5-methoxy-18,19-dinor-5α,17α-pregn-15-en-20-yn-3-one (5-methoxy-gestodene),



J. 13-ethylspiro(18,19-dinor-17α-pregna-5,15-dien-20-yne-3,2′-[1,3]dioxolan)-17-ol and 13-ethylspiro(18,19-dinor-17α-pregna-5(10),15-dien-20-yne-3,2′-[1,3]dioxolan)-17-ol (gestodene ketal),



K. 13-ethyl-3,17-dihydroxy-18,19-dinor-17α-pregna-1,3,5(10),15-tetraen-20-yn-6-one (aromatic 6-keto-gestodene),

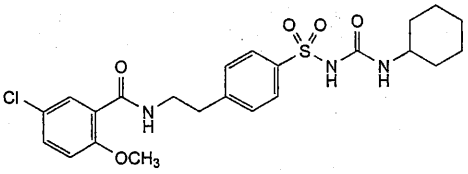


L. 13-ethyl-17-hydroxy-18,19-dinor-17α-pregna-5,15-dien-20-yn-3-one (Δ5(6)-gestodene).

Ph Eur

Glibenclamide

(Ph. Eur. monograph 0718)



$C_{23}H_{28}ClN_3O_5S$ 494.0 *10238-21-8*

Action and use
Inhibition of ATP-dependent potassium channels (sulfonylurea); treatment of diabetes mellitus.

Preparation
Glibenclamide Tablets

Ph Eur

DEFINITION
1-[[4-[2-[(5-Chloro-2-methoxybenzoyl)amino]ethyl]phenyl]sulfonyl]-3-cyclohexylurea.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent) and in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: C.

Second identification: A, B, D, E.

A. Melting point (*2.2.14*): 169 °C to 174 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *methanol R*, with the aid of ultrasound if necessary, and dilute to 50.0 mL with the same solvent. To 10.0 mL of the solution add 1.0 mL of a 103 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with *methanol R*.

Spectral range 230-350 nm.

Absorption maxima At 300 nm and a less intense maximum at 275 nm.

Specific absorbance at the absorption maxima:
- at 300 nm: 61 to 65;
- at 275 nm: 27 to 32.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glibenclamide CRS.*

If the spectra obtained show differences, moisten separately the substance to be examined and the reference substance with *methanol R*, triturate, dry at 100-105 °C and record the spectra again.

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 10 mg of *glibenclamide CRS* in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *ethanol (96 per cent) R, glacial acetic acid R, cyclohexane R, methylene chloride R* (5:5:45:45 *V/V/V/V*).

Application 10 µL.

Development Over 1/2 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

E. Dissolve 20 mg in 2 mL of *sulfuric acid R*. The solution is colourless and shows blue fluorescence in ultraviolet light at 365 nm. Dissolve 0.1 g of *chloral hydrate R* in the solution. After about 5 min, the colour changes to deep yellow and, after about 20 min, develops a brownish tinge.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use or store them at 5 °C for not more than 40 h.*

Test solution Dissolve 25.0 mg of the substance to be examined in *methanol R*, with the aid of ultrasound if necessary, and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 3.0 mg of *glibenclamide impurity A CRS* and 3 mg of *glibenclamide impurity B CRS* in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 20.0 mL with *methanol R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (c) Dissolve 12.5 mg of *glibenclamide for peak identification CRS* (containing impurity C) in *methanol R*, with the aid of ultrasound if necessary, and dilute to 5.0 mL with the same solvent.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: spherical *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: mix 20 mL of a 100.0 g/L solution of *triethylamine R2* previously adjusted to pH 3.0 using *phosphoric acid R*, and 50 mL of *acetonitrile R*; dilute to 1000 mL with *water R*;
- *mobile phase B*: mobile phase A, *water R*, *acetonitrile R* (2:6.5:91.5 *V/V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	45	55
15 - 30	45 → 5	55 → 95
30 - 40	5	95

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B; use the chromatogram supplied with *glibenclamide for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity C.

Relative retention With reference to glibenclamide (retention time = about 5 min): impurity A = about 0.5; impurity B = about 0.6; impurity C = about 0.7.

System suitability Reference solution (a):
- *resolution*: minimum 2.0 between the peaks due to impurities A and B.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.8;
- *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: 0.8 per cent;
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

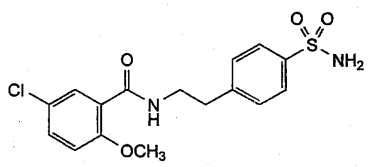
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.400 g with heating in 100 mL of *ethanol (96 per cent) R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

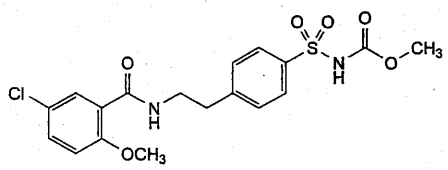
1 mL of *0.1 M sodium hydroxide* is equivalent to 49.40 mg of $C_{23}H_{28}ClN_3O_5S$.

IMPURITIES
Specified impurities *A, C.*

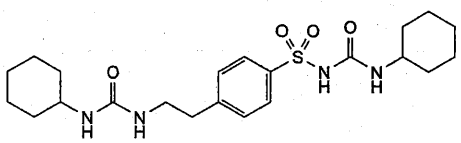
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D, E.*



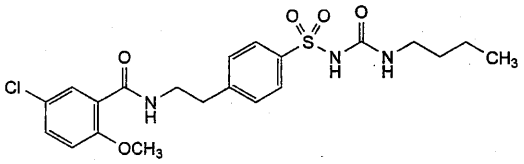
A. 5-chloro-2-methoxy-*N*-[2-(4-sulfamoylphenyl) ethyl]benzamide,



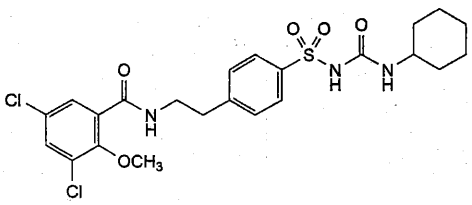
B. methyl [[4-[2-[(5-chloro-2-methoxybenzoyl)amino]ethyl] phenyl]sulfonyl]carbamate,



C. 1-cyclohexyl-3-[[4-[2-[(cyclohexylcarbamoyl)amino]ethyl] phenyl]sulfonyl]urea,



D. 1-butyl-3-[[4-[2-[(5-chloro-2-methoxybenzoyl) amino]ethyl]phenyl]sulfonyl]urea,

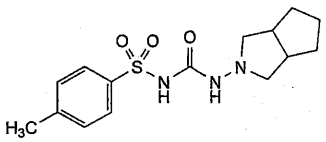


E. 1-cyclohexyl-3-[[4-[2-[(3,5-dichloro-2-methoxybenzoyl)amino]ethyl]phenyl]sulfonyl]urea.

Ph Eur

Gliclazide

(Ph. Eur. monograph 1524)



$C_{15}H_{21}N_3O_3S$ 323.4 *21187-98-4*

Action and use
Inhibition of ATP-dependent potassium channels (sulfonylurea); treatment of diabetes mellitus.

Preparation
Gliclazide Tablets

Ph Eur

DEFINITION
1-(Hexahydrocyclopenta[*c*]pyrrol-2(1*H*)-yl)-3-[(4-methylphenyl)sulfonyl]urea.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in acetone, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *gliclazide CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *acetonitrile R, water R* (45:55 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in 23 mL of *acetonitrile R* and dilute to 50.0 mL with *water R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 10.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 15 mg of *gliclazide impurity F CRS* in 23 mL of *acetonitrile R* and dilute to 50 mL with *water R*. Dilute 1 mL of the solution to 20 mL with the solvent mixture.

Reference solution (c) Dissolve 15.0 mg of *gliclazide impurity F CRS* in 45 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *triethylamine R, trifluoroacetic acid R, acetonitrile R, water R* (0.1:0.1:45:55 *V/V/V/V*).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 µL.

Run time Twice the retention time of gliclazide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity F.

Relative retention With reference to gliclazide (retention time = about 16 min): impurity F = about 0.9.

System suitability Reference solution (b):
— *resolution*: minimum 1.8 between the peaks due to impurity F and gliclazide.

Limits:
— *impurity F*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *sum of impurities other than F*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity B

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Test solution Dissolve 0.400 g of the substance to be examined in 2.5 mL of *dimethyl sulfoxide R* and dilute to 10.0 mL with *water R*. Stir for 10 min, store at 4 °C for 30 min and filter.

Reference solution Dissolve 20.0 mg of *gliclazide impurity B CRS* in *dimethyl sulfoxide R* and dilute to 100.0 mL with the same solvent. To 1.0 mL of the solution, add 12 mL of *dimethyl sulfoxide R* and dilute to 50.0 mL with *water R*. To 1.0 mL of this solution, add 12 mL of *dimethyl sulfoxide R* and dilute to 50.0 mL with *water R*.

Injection 50 µL.

Identification of impurities Use the chromatogram obtained with the reference solution to identify the peak due to impurity B.

Retention time Impurity B = about 8 min.

Limit:
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (2 ppm).

Loss on drying (*2.2.32*)

Maximum 0.25 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

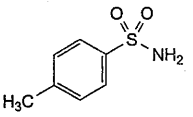
Dissolve 0.250 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 32.34 mg of $C_{15}H_{21}N_3O_3S$.

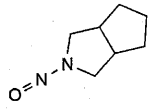
IMPURITIES

Specified impurities *B, F.*

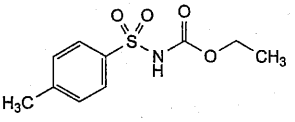
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, G.*



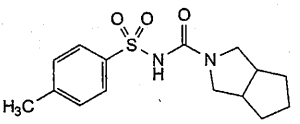
A. 4-methylbenzenesulfonamide,



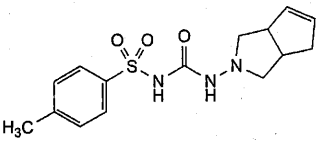
B. 2-nitroso-octahydrocyclopenta[*c*]pyrrole,



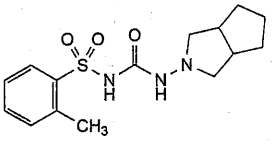
C. ethyl [(4-methylphenyl)sulfonyl]carbamate,



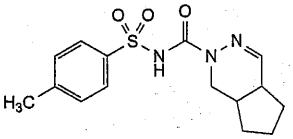
D. *N*-[(4-methylphenyl)sulfonyl]hexahydrocyclopenta[*c*]pyrrol-2(1*H*)-carboxamide,



E. 1-[(4-methylphenyl)sulfonyl]-3-(3,3a,4,6a-tetrahydrocyclopenta[*c*]pyrrol-2(1*H*)-yl)urea,



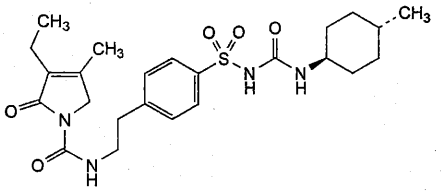
F. 1-(hexahydrocyclopenta[*c*]pyrrol-2(1*H*)-yl)-3-[(2-methylphenyl)sulfonyl]urea,



G. *N*-[(4-methylphenyl)sulfonyl]-1,4a,5,6,7,7a-hexahydro-2*H*-cyclopenta[*d*]pyridazine-2-carboxamide.

Glimepiride

(Ph. Eur. monograph 2223)



$C_{24}H_{34}N_4O_5S$ 490.6 *93479-97-1*

Action and use
Inhibition of ATP-dependent potassium channels (sulfonylurea); treatment of diabetes mellitus.

Preparation
Glimepiride Tablets

Ph Eur

DEFINITION

1-[[4-[2-(3-Ethyl-4-methyl-2-oxo-3-pyrroline-1-carboxamido) ethyl]phenyl]sulfonyl]-3-*trans*-(4-methylcyclohexyl)urea.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Practically insoluble in water, soluble in dimethylformamide, slightly soluble in methylene chloride, very slightly soluble in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glimepiride CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *dimethylformamide R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances
Liquid chromatography (*2.2.29*). *Store the solutions at a temperature not exceeding 12 °C and for not more than 15 h.*

Solvent mixture *water for chromatography R, acetonitrile for chromatography R* (1:4 *V/V*).

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of *glimepiride for system suitability CRS* (containing impurities B, C and D) in 2.0 mL of the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 20.0 mg of *glimepiride CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (4 µm).

Mobile phase Dissolve 0.5 g of *sodium dihydrogen phosphate R* in 500 mL of *water for chromatography R* and adjust to pH 2.5 with *phosphoric acid R*. Add 500 mL of *acetonitrile for chromatography R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 228 nm.

Injection 20 µL of the test solution and reference solutions (a) and (b).

Run time 2.5 times the retention time of glimepiride.

Relative retention With reference to glimepiride (retention time = about 17 min): impurity B = about 0.2; impurity C = about 0.3; impurity D = about 1.1.

System suitability Reference solution (a):
— *resolution*: minimum 4.0 between the peaks due to impurities B and C.

Limits:
— *impurity B*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent),
— *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent),
— *sum of impurities other than B*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity A
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 10.0 mg of the substance to be examined in 5 mL of *methylene chloride R* and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 0.8 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 2.0 mg of *glimepiride for impurity A identification CRS* in 1 mL of *methylene chloride R* and dilute to 4.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 3 mm;
— *stationary phase*: *diol silica gel for chromatography R* (5 µm).

Mobile phase *anhydrous acetic acid R, 2-propanol R, heptane R* (1:100:899 *V/V/V*).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 228 nm.

Injection 10 µL.

Run time 1.5 times the retention time of glimepiride.

Identification of impurities Use the chromatogram supplied with *glimepiride for impurity A identification CRS* and the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to glimepiride (retention time = about 14 min): impurity A = about 0.9.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to glimepiride.

Limit:

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent).

Water (*2.5.32*)

Maximum 0.5 per cent.

Dissolve 0.250 g in *dimethylformamide R* and dilute to 5.0 mL with the same solvent. Carry out the test on 1.0 mL of solution. Carry out a blank test.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

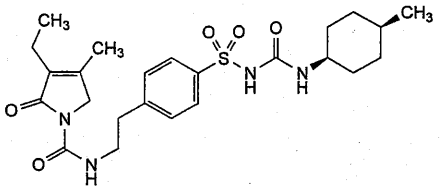
Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{24}H_{34}N_4O_5S$ from the areas of the peaks and the assigned content of *glimepiride CRS*.

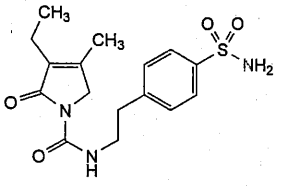
IMPURITIES

Specified impurities *A, B, D.*

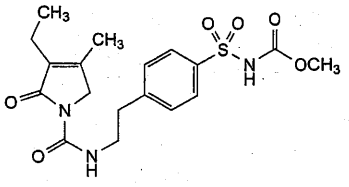
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E, F, G, H, I, J.*



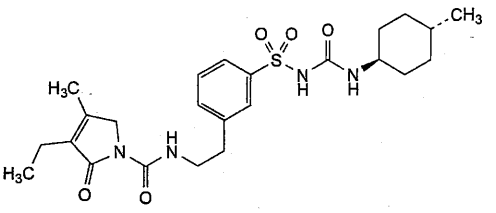
A. 1-[[4-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl]sulfonyl]-3-(*cis*-4-methylcyclohexyl)urea,



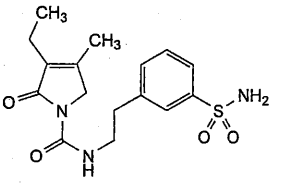
B. 3-ethyl-4-methyl-2-oxo-*N*-[2-(4-sulfamoylphenyl)ethyl]-2,3-dihydro-1*H*-pyrrole-1-carboxamide,



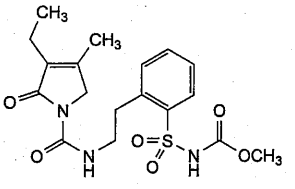
C. methyl [[4-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl] sulfonyl]carbamate,



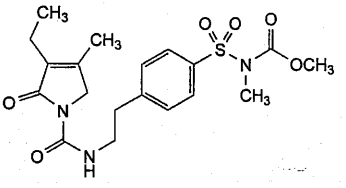
D. 1-[[3-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl]sulfonyl]-3-(*trans*-4-methylcyclohexyl)urea,



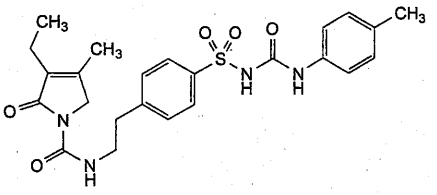
E. 3-ethyl-4-methyl-2-oxo-*N*-[2-(3-sulfamoylphenyl)ethyl]-2,3-dihydro-1*H*-pyrrole-1-carboxamide,



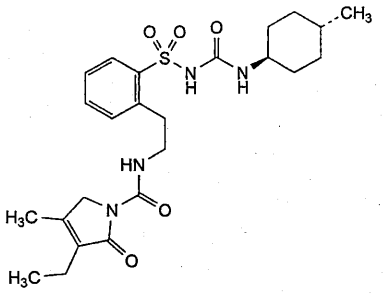
F. methyl [[2-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl] sulfonyl]carbamate,



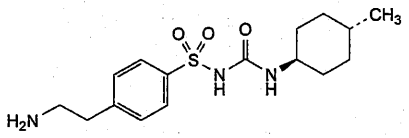
G. methyl [[4-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl] sulfonyl]methylcarbamate,



H. 1-[[4-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl]sulfonyl]-3-(4-methylphenyl)urea,



I. 1-[[2-[2-[[(3-ethyl-4-methyl-2-oxo-2,3-dihydro-1*H*-pyrrol-1-yl)carbonyl]amino]ethyl]phenyl]sulfonyl]-3-(*trans*-4-methylcyclohexyl)urea,

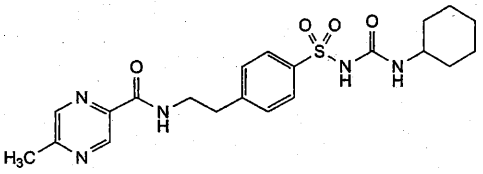


J. 1-[[4-(2-aminoethyl)phenyl]sulfonyl]-3-(*trans*-4-methylcyclohexyl)urea.

Ph Eur

Glipizide

(Ph. Eur. monograph 0906)



$C_{21}H_{27}N_5O_4S$ 445.5 *29094-61-9*

Action and use
Inhibition of ATP-dependent potassium channels (sulfonylurea); treatment of diabetes mellitus.

Preparation
Glipizide Tablets

Ph Eur

DEFINITION

N-[2-[4-[(Cyclohexylcarbamoyl)sulfamoyl]phenyl]ethyl]-5-methylpyrazine-2-carboxamide.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, very slightly soluble in acetone and in methylene chloride, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve about 2 mg in *methanol R* and dilute to 100 mL with the same solvent.

Spectral range 220-350 nm.

Absorption maxima 226 nm and 274 nm.

Absorbance ratio A_{226}/A_{274} = 2.0 to 2.4.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glipizide CRS*.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 10 mg of *glipizide CRS* in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *anhydrous formic acid R, ethyl acetate R, methylene chloride R* (25:25:50 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Mix 40 volumes of *acetonitrile R* and 60 volumes of *water R* previously adjusted to pH 3.5 with *acetic acid R*.

Test solution Dissolve 20.0 mg of the substance to be examined in 20.0 mL of *methanol R* using sonication and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 6.0 mg of *glipizide impurity A CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of *glipizide impurity C CRS* in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL of the solution to 100 mL with the solvent mixture.

Reference solution (d) Dissolve 2 mg of *glipizide impurity D CRS* in the solvent mixture and dilute to 250 mL with the solvent mixture. Dilute 1 mL of the solution to 20 mL with reference solution (b).

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *water for chromatography R* adjusted to pH 3.5 with *acetic acid R*;
— *mobile phase B*: *acetonitrile for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	75	25
5 - 12	75 → 65	25 → 35
12 - 20	65	35
20 - 25	65 → 50	35 → 50
25 - 30	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity D.

Relative retention With reference to glipizide (retention time = about 22 min): impurity A = about 0.25; impurity D = about 0.27; impurity C = about 1.2.

System suitability Reference solution (d):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity A.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity C by 1.7;
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurity C*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: maximum 0.5 per cent;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity B

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 25.0 mg of *decane R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with *methylene chloride R*.

Test solution (a) Dissolve 1.000 g of the substance to be examined in 50 mL of a 12 g/L solution of *sodium hydroxide R* and shake with 2 quantities, each of 5.0 mL, of *methylene chloride R*. Use the combined lower layers.

Test solution (b) Dissolve 1.000 g of the substance to be examined in 50 mL of a 12 g/L solution of *sodium hydroxide R* and shake with 2 quantities, each of 5.0 mL, of the internal standard solution. Use the combined lower layers.

Reference solution Dissolve 10.0 mg of *cyclohexylamine R* (impurity B) in a 17.5 g/L solution of *hydrochloric acid R* and dilute to 100.0 mL with the same acid. To 1.0 mL of this solution add 50 mL of a 12 g/L solution of *sodium hydroxide R* and shake with 2 quantities, each of 5.0 mL, of the internal standard solution. Use the combined lower layers.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.5 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1.8 mL/min.

Split ratio 1:11.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 4	40
	4 - 20	40 → 200
	20 - 25	200
Injection port		250
Detector		270

Detection Flame ionisation.

Injection 1 µL.

Elution order Impurity B, decane.

System suitability:
— *resolution*: minimum 7 between the peaks due to impurity B and the internal standard in the chromatogram obtained with the reference solution;
— there is no peak with the same retention time as that of the internal standard in the chromatogram obtained with test solution (a).

Calculate the ratio (*R*) of the area of the peak due to impurity B to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with test solution (b), calculate the ratio of the area of any peak due to impurity B to the area of the peak due to the internal standard.

Limit:
— *impurity B*: not more than *R* (100 ppm).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

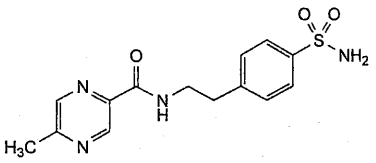
Dissolve 0.400 g in 50 mL of *dimethylformamide R*. Add 0.2 mL of *quinaldine red solution R*. Titrate with *0.1 M lithium methoxide* until the colour changes from red to colourless.

1 mL of *0.1 M lithium methoxide* is equivalent to 44.55 mg of $C_{21}H_{27}N_5O_4S$.

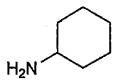
IMPURITIES

Specified impurities *A, B, C.*

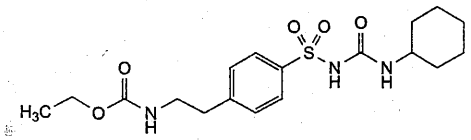
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, E, F, G, H, I, J.*



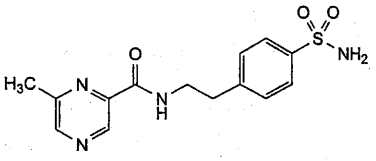
A. 5-methyl-*N*-[2-(4-sulfamoylphenyl)ethyl]pyrazine-2-carboxamide,



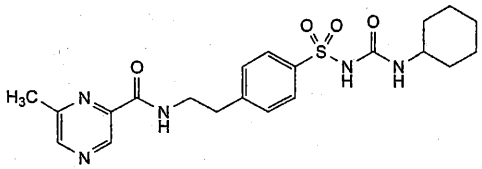
B. cyclohexanamine (cyclohexylamine),



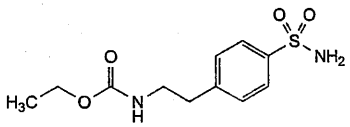
C. ethyl [2-[4-[(cyclohexylcarbamoyl)sulfamoyl]phenyl]ethyl] carbamate,



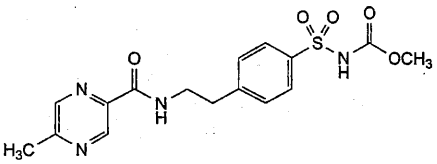
D. 6-methyl-*N*-[2-(4-sulfamoylphenyl)ethyl]pyrazine-2-carboxamide,



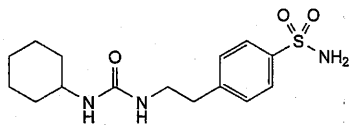
E. *N*-[2-[4-[(cyclohexylcarbamoyl)sulfamoyl]phenyl]ethyl]-6-methylpyrazine-2-carboxamide,



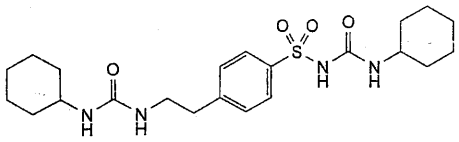
F. ethyl [2-(4-sulfamoylphenyl)ethyl]carbamate,



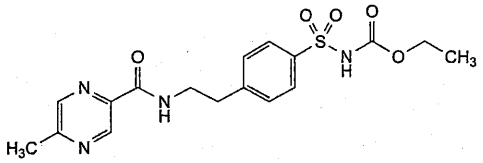
G. methyl [4-[2-(5-methylpyrazine-2-carboxamido) ethyl]benzene-1-sulfonyl]carbamate,



H. 4-[2-[(cyclohexylcarbamoyl)amino]ethyl]benzene-1-sulfonamide,



I. *N*-(cyclohexylcarbamoyl)-4-[2-[(cyclohexylcarbamoyl)amino]ethyl]benzene-1-sulfonamide,



J. ethyl [4-[2-(5-methylpyrazine-2-carboxamido) ethyl]benzene-1-sulfonyl]carbamate.

Ph Eur

Human Glucagon

(Glucagon, Human, Ph. Eur. monograph 1635)

H-His-Ser-Gln-Gly-Thr-Phe-Thr-Ser-Asp-Tyr-
Ser-Lys-Tyr-Leu-Asp-Ser-Arg-Arg-Ala-Gln-
Asp-Phe-Val-Gln-Trp-Leu-Met-Asn-Thr-OH

$C_{153}H_{225}N_{43}O_{49}S$ 3483

Action and use
Hormone; treatment of hypoglycaemia.

Preparation
Human Glucagon Injection

Ph Eur

DEFINITION
Polypeptide having the same structure (29 amino acids) as the hormone produced by the α-cells of the human pancreas, which increases the blood-glucose concentration by promoting rapid breakdown of liver glycogen.

Content
92.5 per cent to 105.0 per cent (anhydrous substance).

PRODUCTION
Human glucagon is produced by a method based on recombinant DNA (rDNA) technology. During the course of product development it must be demonstrated that the manufacturing process produces a product having a biological activity of not less than 1 IU/mg using a suitable validated bioassay.

Host-cell-derived proteins
The limit is approved by the competent authority.

Host-cell- and vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water and in most organic solvents. It is soluble in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Peptide mapping. Liquid chromatography (*2.2.29*).

Test solution Prepare a 5 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid*. Mix 200 µL of this solution with 800 µL of *0.1 M ammonium carbonate buffer solution pH 10.3 R* (diluted stock solution). Prepare a 2 mg/mL solution of *α-chymotrypsin for peptide mapping R* in *0.1 M ammonium carbonate buffer solution pH 10.3 R* and add 25 µL of this solution to the diluted stock solution. Place the solution in a closed vial at 37 °C for 2 h. Remove the vial and stop the reaction immediately by adding 120 µL of *glacial acetic acid R*.

Reference solution Prepare a 1 mg/mL solution of *human glucagon CRS* in *0.1 M ammonium carbonate buffer solution pH 10.3 R* (diluted stock solution) and continue as described for the test solution.

Column:
- *size*: *l* = 0.05 m, Ø = 4 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: mix 500 µL of *trifluoroacetic acid R* and 1000 mL of *water R*;
- *mobile phase B*: mix 500 µL of *trifluoroacetic acid R* with 600 mL of *anhydrous ethanol R* and add 400 mL of *water R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 35	100 → 53	0 → 47
35 - 45	53 → 0	47 → 100
45 - 46	0 → 100	100 → 0
46 - 75	100	0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Equilibration With mobile phase A for at least 15 min.

Injection 20 µL.

System suitability The chromatogram obtained with the reference solution is similar to the chromatogram supplied with *human glucagon CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Related proteins and deamidated forms

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve the substance to be examined in *0.01 M hydrochloric acid* to obtain a concentration of 0.5 mg/mL. *Maintain the solution at 2-8 °C.*

Reference solution (a) Dissolve the contents of a vial of *human glucagon CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 0.5 mg/mL. *Maintain the solution at 2-8 °C.*

Reference solution (b) Dissolve the substance to be examined in *0.01 M hydrochloric acid* to obtain a concentration of about 0.5 mg/mL. Heat at 50 °C for 48 h (*in situ* preparation of all 4 deamidated forms of glucagon at a total concentration of not less than 7 per cent).

Column:
- *size*: *l* = 0.15 m, Ø = 3 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 45 °C.

Mobile phase:
- *mobile phase A*: dissolve 16.3 g of *potassium dihydrogen phosphate R* in 800 mL of *water R*, adjust to pH 2.7 with *phosphoric acid R* and add 200 mL of *acetonitrile for chromatography R*;
- *mobile phase B*: *acetonitrile for chromatography R*, *water R* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	61	39
25 - 29	61 → 12	39 → 88
29 - 30	12	88
30 - 31	12 → 61	88 → 39

NOTE The end time of the isocratic elution may be adjusted so that the gradient begins after elution of the peak due to deamidated glucagon 4 (see relative retention below).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 15 µL.

Relative retention With reference to glucagon (retention time = about 21 min): deamidated glucagon 1 = about 1.1; deamidated glucagon 4 = about 1.4.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to glucagon and deamidated glucagon 1 in the chromatogram obtained with reference solution (b);
- *symmetry factor*: maximum 1.8 for the peak due to glucagon in the chromatogram obtained with reference solution (a);
- *repeatability*: maximum relative standard deviation of 2.0 per cent after 5 injections of reference solution (a);
- 4 peaks eluting after the principal peak, that correspond to the deamidated forms, are clearly visible in the chromatogram obtained with reference solution (b).

Limits:
- *deamidated forms*: maximum 0.8 per cent;
- *total*: maximum 3.0 per cent.

Water (*2.5.32*)

Maximum 10 per cent, determined on 50 mg.

Bacterial endotoxins (*2.6.14*)

Less than 10 IU/mg.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related proteins and deamidated forms with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of human glucagon ($C_{153}H_{225}N_{43}O_{49}S$) taking into account the assigned content of $C_{153}H_{225}N_{43}O_{49}S$ in *human glucagon CRS*.

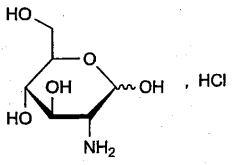
STORAGE

In an airtight container, protected from light, at a temperature lower than −15 °C.

Ph Eur

Glucosamine Hydrochloride

(Ph. Eur. monograph 2446)



$C_6H_{14}ClNO_5$ 215.6 *66-84-2*

Ph Eur

DEFINITION

2-Amino-2-deoxy-D-glucopyranose hydrochloride.

Isolated from natural sources or produced by fermentation.

Content

98.0 per cent to 102.0 per cent (dried substance).

PRODUCTION

The animals from which glucosamine hydrochloride is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glucosamine hydrochloride CRS.*

B. 1 mL of solution S (see Tests) gives reaction (a) of chlorides (*2.3.1*).

C. Specific optical rotation (see Tests).

TESTS

Solution S

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dilute 5.0 mL of solution S to 25.0 mL with *water R.*

pH (*2.2.3*)

3.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 70.0 to + 74.0 (dried substance), determined on solution S.

Examine 3 h after preparation of solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution To 0.300 g of the substance to be examined add 80 mL of the mobile phase and sonicate for 10 min. Cool to room temperature and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *2-methylpyrazine CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *glucosamine for system suitability CRS* (containing impurities B and C) in 1.0 mL of the mobile phase.

Column:

— *size: l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);

— *temperature:* 30 °C.

Mobile phase Dissolve 0.5 g of *sodium heptanesulfonate R* in *water for chromatography R,* add 0.5 mL of *phosphoric acid R* and 4 mL of a 56 g/L solution of *potassium hydroxide R* and dilute to 1000 mL with *water for chromatography R;* to 1000 mL of this solution add 50 mL of *acetonitrile R1.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 195 nm.

Injection 20 µL.

Run time Twice the retention time of 2-methylpyrazine.

Retention time 2-methylpyrazine = about 9 min.

System suitability Reference solution (b):

— *resolution:* minimum 1.5 between the peaks due to impurities B and C.

Limits:

— *unspecified impurities:* for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);

— *total:* not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *disregard limit:* 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

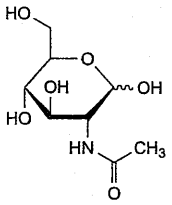
ASSAY

Dissolve 0.200 g in 50 mL of *water R* and add 1.0 mL of *0.1 M hydrochloric acid.* Titrate with *0.1 M sodium hydroxide,* determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

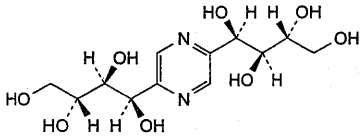
1 mL of *0.1 M sodium hydroxide* is equivalent to 21.56 mg of $C_6H_{14}ClNO_5$.

IMPURITIES

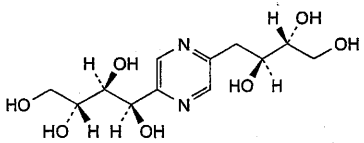
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E.*



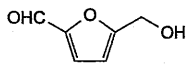
A. 2-(acetylamino)-2-deoxy-D-glucopyranose (*N*-acetylglucosamine),



B. (1*R*,1′*R*,2*S*,2′*S*,3*R*,3′*R*)-1,1′-pyrazine-2,5-diylbis(butane-1,2,3,4-tetrol) (fructosazine),



C. (1*R*,2*S*,3*R*)-1-[5-[(2*S*,3*R*)-2,3,4-trihydroxybutyl]pyrazin-2-yl]butane-1,2,3,4-tetrol (deoxyfructosazine),

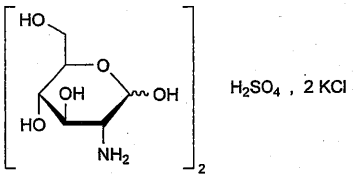


E. 5-(hydroxymethyl)furan-2-carbaldehyde (5-hydroxymethylfurfural).

Ph Eur

Glucosamine Sulfate Potassium Chloride

(Ph. Eur. monograph 2708)



$C_{12}H_{28}Cl_2K_2N_2O_{14}S$ 606 *216699-44-4*

Ph Eur

DEFINITION

Bis(2-amino-2-deoxy-D-glucopyranose) sulfate bis(potassium chloride).

Substance prepared from glucosamine hydrochloride isolated from natural sources or produced by fermentation, and potassium sulfate.

Content

98.0 per cent to 102.0 per cent (dried substance).

PRODUCTION

The animals from which glucosamine sulfate potassium chloride is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glucosamine sulfate potassium chloride CRS.*

C. It gives reaction (a) of chlorides (*2.3.1*).

D. It gives reaction (a) of sulfates (*2.3.1*).

E. 1 mL of solution S (see Tests) gives reaction (a) of potassium (*2.3.1*).

TESTS

Solution S

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dilute 5.0 mL of solution S to 25.0 mL with *water R.*

pH (*2.2.3*)

3.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 47.0 to + 53.0 (dried substance), determined on solution S.

Examine 3 h after preparation of solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution To 0.42 g of the substance to be examined add 80 mL of the mobile phase and sonicate for 10 min. Cool to room temperature and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *2-methylpyrazine CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *glucosamine for system suitability CRS* (containing impurities B and C) in 1 mL of the mobile phase.

Column:

— *size*: *l* = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm) (3 µm);

— *temperature*: 30 °C.

Mobile phase Dissolve 0.5 g of *sodium heptanesulfonate R* in *water for chromatography R*, add 0.5 mL of *phosphoric acid R*, 4 mL of a 56 g/L solution of *potassium hydroxide R* and dilute to 1000 mL with *water for chromatography R*, then add 50 mL of *acetonitrile R1.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 195 nm.

Injection 20 µL.

Run time Twice the retention time of 2-methylpyrazine.

Retention time 2-methylpyrazine = about 9 min.

System suitability Reference solution (b):

— *resolution*: minimum 1.5 between the peaks due to impurities B and C.

Calculation of percentage contents:
— for each impurity, use the concentration of 2-methylpyrazine in reference solution (a).

Limits:
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.2 per cent;
— *reporting threshold*: 0.03 per cent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
27.0 per cent to 31.0 per cent, determined on 1.0 g.

Ignite first at 600 ± 50 °C for 2 h. Do not repeat the moistening with *sulfuric acid R* between any re-ignition.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

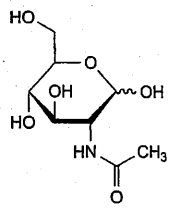
ASSAY

Dissolve 0.280 g in 50 mL of *water R* and add 1.0 mL of *0.1 M hydrochloric acid*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

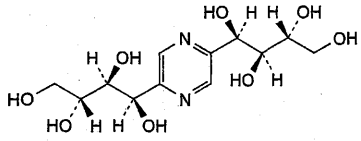
1 mL of *0.1 M sodium hydroxide* is equivalent to 30.28 mg of $C_{12}H_{28}Cl_2K_2N_2O_{14}S$.

IMPURITIES

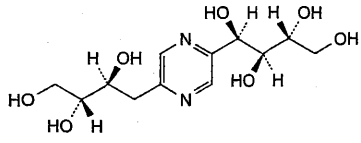
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E.*



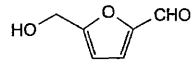
A. 2-(acetylamino)-2-deoxy-D-glucopyranose (*N*-acetylglucosamine),



B. (1*R*,1′*R*,2*S*,2′*S*,3*R*,3′*R*)-1,1′-pyrazine-2,5-diylbis (butane-1,2,3,4-tetrol) (fructosazine),



C. (1*R*,2*S*,3*R*)-1-[5-[(2*S*,3*R*)-2,3,4-trihydroxybutyl]pyrazin-2-yl]butane-1,2,3,4-tetrol (deoxyfructosazine),

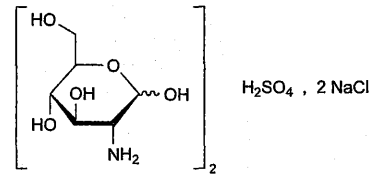


E. 5-(hydroxymethyl)furan-2-carbaldehyde (5-hydroxymethylfurfural).

Ph Eur

Glucosamine Sulfate Sodium Chloride

(Ph. Eur. monograph 2447)



$C_{12}H_{28}Cl_2N_2Na_2O_{14}S$ 573.3 *216447-62-0*

Ph Eur

DEFINITION

Bis(2-amino-2-deoxy-D-glucopyranose) sulfate bis(sodium chloride).

Substance prepared from glucosamine hydrochloride isolated from natural sources or produced by fermentation, and sodium sulfate.

Content
98.0 per cent to 102.0 per cent (dried substance).

PRODUCTION

The animals from which glucosamine sulfate sodium chloride is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in methanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison glucosamine sulfate sodium chloride CRS.

B. It gives reaction (a) of chlorides (*2.3.1*).

C. 1 mL of solution S (see Tests) gives reaction (a) of sodium (*2.3.1*).

D. It gives reaction (a) of sulfates (*2.3.1*).

E. Specific optical rotation (see Tests).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dilute 5.0 mL of solution S to 25.0 mL with *water R*.

pH (*2.2.3*)
3.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)
+ 50.0 to + 55.0 (dried substance), determined on solution S.

Examine 3 h after preparation of solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution To 0.40 g of the substance to be examined add 80 mL of the mobile phase and sonicate for 10 min. Cool to room temperature and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *2-methylpyrazine CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *glucosamine for system suitability CRS* (containing impurities B and C) in 1 mL of the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 30 °C.

Mobile phase Dissolve 0.5 g of *sodium heptanesulfonate R* in *water for chromatography R*, add 0.5 mL of *phosphoric acid R* and 4 mL of a 56 g/L solution of *potassium hydroxide R* and dilute to 1000 mL with *water for chromatography R*; to 1000 mL of this solution add 50 mL of *acetonitrile R1*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 195 nm.

Injection 20 µL.

Run time Twice the retention time of 2-methylpyrazine.

Retention time 2-methylpyrazine = about 9 min.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurities B and C.

Limits:
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
23.5 per cent to 26.0 per cent, determined on 1.0 g.

Ignite first at 600 ± 50 °C for 2 h. Do not repeat the moistening with *sulfuric acid R* between any re-ignition.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

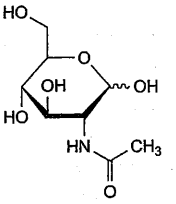
TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

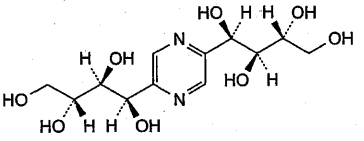
ASSAY
Dissolve 0.250 g in 50 mL of *water R* and add 1.0 mL of *0.1 M hydrochloric acid*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*). Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 28.67 mg of $C_{12}H_{28}Cl_2N_2Na_2O_{14}S$.

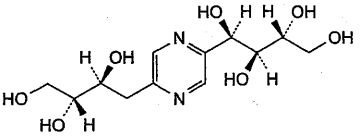
IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, E.*



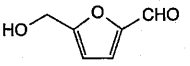
A. 2-(acetylamino)-2-deoxy-D-glucopyranose (*N*-acetylglucosamine),



B. (1*R*,1′*R*,2*S*,2′*S*,3*R*,3′*R*)-1,1′-pyrazine-2,5-diylbis (butane-1,2,3,4-tetrol) (fructosazine),



C. (1*R*,2*S*,3*R*)-1-[5-[(2*S*,3*R*)-2,3,4-trihydroxybutyl] pyrazin-2-yl]butane-1,2,3,4-tetrol (deoxyfructosazine),

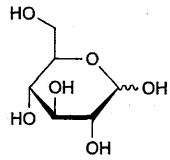


E. 5-(hydroxymethyl)furan-2-carbaldehyde (5-hydroxymethylfurfural).

Glucose[1]

Anhydrous Glucose

(Ph. Eur. monograph 0177)



$C_6H_{12}O_6$ 180.2 *50-99-7*

Preparations

Glucose Infusion

Compound Glucose, Sodium Chloride and Sodium Citrate Oral Solution

Oral Rehydration Salts

Potassium Chloride and Glucose Intravenous Infusion

Potassium Chloride, Sodium Chloride and Glucose Intravenous Infusion

Sodium Chloride and Glucose Intravenous Infusion

Ph Eur

DEFINITION

D-Glucopyranose.

It is derived from starch.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

♦CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).♦

IDENTIFICATION

First identification: ◊A◊, B, E.

◊Second identification: C, D.◊

◊A. Specific optical rotation (*2.2.7*): + 52.5 to + 53.3 (anhydrous substance).

Dissolve 10.0 g in 80 mL of *water R*, add 0.2 mL of *dilute ammonia R1*, allow to stand for 30 min and dilute to 100.0 mL with *water R*.◊

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

◊C. Thin-layer chromatography (*2.2.27*).

Solvent mixture *water R, methanol R* (2:3 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of *glucose monohydrate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg each of *fructose R, glucose R, lactose monohydrate R* and *sucrose R* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Plate *TLC silica gel plate R.*

Mobile phase *water R, methanol R, anhydrous acetic acid R, ethylene chloride R* (10:15:25:50 *V/V/V/V*); measure the volumes accurately since a slight excess of water produces cloudiness.

Application 2 µL; thoroughly dry the points of application.

Development A Over 3/4 of the plate.

Drying A In a current of warm air.

Development B Immediately, over 3/4 of the plate, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Treat with a solution of 0.5 g of *thymol R* in a mixture of 5 mL of *sulfuric acid R* and 95 mL of *ethanol (96 per cent) R*; heat at 130 °C for 10 min.

System suitability Reference solution (b):

— the chromatogram shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 10 mL of *water R*. Add 3 mL of *cupri-tartaric solution R* and heat. A red precipitate is formed.◊

E. Water (see Tests).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 10.0 g in 15 mL of *water R*, heating on a water-bath.

Conductivity (*2.2.38*)

Maximum 20 $\mu S \cdot cm^{-1}$.

Dissolve 20.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.300 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.330 g of *glucose monohydrate CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 250.0 mL with *water R*.

Reference solution (c) Dilute 25.0 mL of reference solution (b) to 200.0 mL with *water R*.

Reference solution (d) Dissolve 5 mg of *fructose R* (impurity D), 5 mg of *maltose monohydrate R* (impurity A) and 5 mg of *maltotriose R* (impurity C) in *water R* and dilute to 50 mL with the same solvent.

Column:

— *size*: *l* = 0.3 m, Ø = 7.8 mm;

— *stationary phase*: *strong cation-exchange resin (calcium form) R* (9 µm);

— *temperature*: 85 ± 1 °C.

Mobile phase Degassed *water R*.

Flow rate 0.3 mL/min.

Detection Refractometer maintained at a constant temperature (40 °C for example).

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time 1.5 times the retention time of glucose.

Relative retention With reference to glucose (retention time = about 21 min): impurity C = about 0.7; impurities A and B = about 0.8; impurity D = about 1.3.

System suitability Reference solution (d):
— *resolution*: minimum 1.3 between the peaks due to impurities C and A.

Limits:
— *sum of impurities A and B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);
— *impurity C*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *impurity D*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Dextrin
To 1 g of the finely powdered substance to be examined add 20 mL of *ethanol (96 per cent) R* and heat under a reflux condenser. The substance dissolves completely.

Soluble starch, sulfite
Maximum 15 ppm.

Dissolve 6.7 g in 15.0 mL of *water R*, heating on a water-bath. Allow to cool and add 25 µL of *iodine solution R5*. The solution is yellow.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 0.50 g.

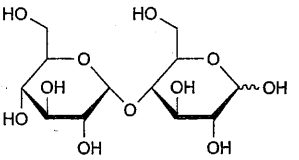
◊Pyrogens (*2.6.8*)
If intended for use in the manufacture of large-volume parental preparations without a further appropriate procedure for the removal of pyrogens, the competent authority may require that it comply with the test for pyrogens. Inject per kilogram of the rabbit's mass 10 mL of a solution in *water for injections R* containing 50 mg of the substance to be examined per millilitre.◊

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

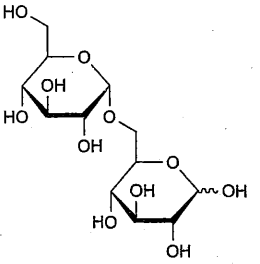
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_6H_{12}O_6$ taking into account the assigned content of *glucose monohydrate CRS*.

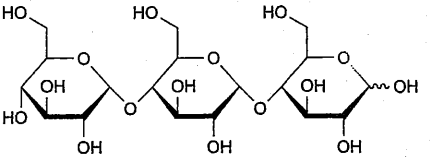
IMPURITIES
Specified impurities *A, B, C, D.*



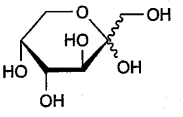
A. 4-*O*-α-D-glucopyranosyl-D-glucopyranose (maltose),



B. 6-*O*-α-D-glucopyranosyl-D-glucopyranose (isomaltose),



C. α-D-glucopyranosyl-(1→4)-α-D-glucopyranosyl-(1→4)-D-glucopyranose (maltotriose),



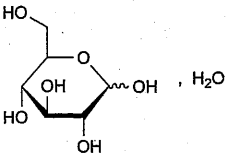
D. D-*arabino*-hex-2-ulopyranose (fructose).

Ph Eur

Glucose Monohydrate[1]

(Ph. Eur. monograph 0178)

NOTE: The name Glucose was formerly used in the United Kingdom.



$C_6H_{12}O_6,H_2O$ 198.2 *77938-63-7*

Preparations
Glucose Infusion
Oral Rehydration Salts
Potassium Chloride and Glucose Intravenous Infusion
Potassium Chloride, Sodium Chloride and Glucose Intravenous Infusion
Sodium Chloride and Glucose Intravenous Infusion

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Ph Eur

DEFINITION

D-Glucopyranose monohydrate.

It is derived from starch.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

♦CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).♦

IDENTIFICATION

First identification: ◊*A*◊, *B, E.*

◊*Second identification: C, D.*◊

◊A. Specific optical rotation (*2.2.7*): + 52.5 to + 53.3 (anhydrous substance).

Dissolve 10.0 g in 80 mL of *water R*, add 0.2 mL of *dilute ammonia R1*, allow to stand for 30 min and dilute to 100.0 mL with *water R.*◊

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

◊C. Thin-layer chromatography (*2.2.27*).

Solvent mixture *water R, methanol R* (2:3 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of *glucose monohydrate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg each of *fructose R, glucose R, lactose monohydrate R* and *sucrose R* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Plate *TLC silica gel plate R.*

Mobile phase *water R, methanol R, anhydrous acetic acid R, ethylene chloride R* (10:15:25:50 *V/V/V/V*); measure the volumes accurately since a slight excess of water produces cloudiness.

Application 2 µL; thoroughly dry the points of application.

Development A Over 3/4 of the plate.

Drying A In a current of warm air.

Development B Immediately, over 3/4 of the plate, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Treat with a solution of 0.5 g of *thymol R* in a mixture of 5 mL of *sulfuric acid R* and 95 mL of *ethanol (96 per cent) R*; heat at 130 °C for 10 min.

System suitability Reference solution (b):

— the chromatogram shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 10 mL of *water R*. Add 3 mL of *cupri-tartaric solution R* and heat. A red precipitate is formed.◊

E. Water (see Tests).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 10.0 g in 15 mL of *water R.*

Conductivity (*2.2.38*)

Maximum 20 $\mu S \cdot cm^{-1}$.

Dissolve 20.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.330 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.330 g of *glucose monohydrate CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 250.0 mL with *water R.*

Reference solution (c) Dilute 25.0 mL of reference solution (b) to 200.0 mL with *water R.*

Reference solution (d) Dissolve 5 mg of *fructose R* (impurity D), 5 mg of *maltose monohydrate R* (impurity A) and 5 mg of *maltotriose R* (impurity C) in *water R* and dilute to 50 mL with the same solvent.

Column:

— *size*: *l* = 0.3 m, Ø = 7.8 mm;

— *stationary phase*: *strong cation-exchange resin (calcium form) R* (9 µm);

— *temperature*: 85 ± 1 °C.

Mobile phase Degassed *water R.*

Flow rate 0.3 mL/min.

Detection Refractometer maintained at a constant temperature (40 °C for example).

Injection 20 µL of the test solution and reference solutions (b), (c) and (d).

Run time 1.5 times the retention time of glucose.

Relative retention With reference to glucose (retention time = about 21 min): impurity C = about 0.7; impurities A and B = about 0.8; impurity D = about 1.3.

System suitability Reference solution (d):

— *resolution*: minimum 1.3 between the peaks due to impurities C and A.

Limits:

— *sum of impurities A and B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— *impurity C*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurity D*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Dextrin

To 1 g of the finely powdered substance to be examined add 20 mL of *ethanol (96 per cent) R* and heat under a reflux condenser. The substance dissolves completely.

Soluble starch, sulfite

Maximum 15 ppm.

Dissolve 7.4 g in 15.0 mL of *water R*, heating on a water-bath. Allow to cool and add 25 µL of *iodine solution R5*. The solution is yellow.

Water (*2.5.12*)

7.5 per cent to 9.5 per cent, determined on 0.25 g.

◊**Pyrogens** (*2.6.8*)

If intended for use in the manufacture of large-volume parenteral preparations without a further appropriate procedure for the removal of pyrogens, the competent authority may require that it comply with the test for pyrogens. Inject per kilogram of the rabbit's mass 10 mL of a solution in *water for injections R* containing 50 mg of the substance to be examined per millilitre.◊

ASSAY

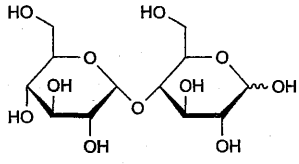
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

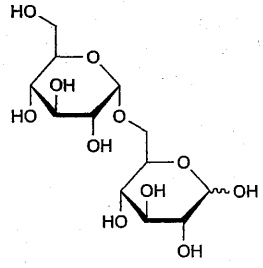
Calculate the percentage content of $C_6H_{12}O_6$ taking into account the assigned content of *glucose monohydrate CRS*.

IMPURITIES

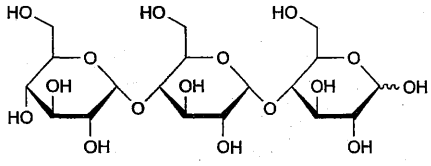
Specified impurities *A, B, C, D.*



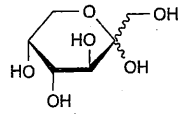
A. 4-*O*-α-D-glucopyranosyl-D-glucopyranose (maltose),



B. 6-*O*-α-D-glucopyranosyl-D-glucopyranose (isomaltose),



C. α-D-glucopyranosyl-(1→4)-α-D-glucopyranosyl-(1→4)-D-glucopyranose (maltotriose),



D. D-*arabino*-hex-2-ulopyranose (fructose).

Ph Eur

Liquid Glucose

(Ph. Eur. monograph 1330)

Action and use

Excipient.

Ph Eur

DEFINITION

Aqueous solution containing a mixture of glucose, oligosaccharides and polysaccharides obtained by hydrolysis of starch.

It contains a minimum of 70.0 per cent dry matter.

The degree of hydrolysis, expressed as dextrose equivalent (DE), is not less than 20 (nominal value).

CHARACTERS

Appearance

Clear, colourless or brown, viscous liquid.

Solubility

Miscible with water.

It may partly or totally solidify at room temperature and liquefies again when heated to 50 °C.

IDENTIFICATION

A. Dissolve 0.1 g in 2.5 mL of *water R* and heat with 2.5 mL of *cupri-tartaric solution R*. A red precipitate is formed.

B. Dip, for 1 s, a suitable stick with a reactive pad containing glucose-oxidase, peroxidase and a hydrogen-donating substance, such as tetramethylbenzidine, in a 5 g/L solution of the substance to be examined. Observe the colour of the reactive pad; within 60 s the colour changes from yellow to green or blue.

C. It is a clear, colourless or brown, viscous liquid, miscible with water. The substance may partly or totally solidify at room temperature and liquefies again when heated to 50 °C.

D. Dextrose equivalent (see Tests).

TESTS

Solution S

Dissolve 25.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

pH (*2.2.3*)

4.0 to 6.0.

Mix 1 mL of a 223.6 g/L solution of *potassium chloride R* and 30 mL of solution S.

Sulfur dioxide (*2.5.29*)

Maximum 20 ppm; maximum 400 ppm if intended for the production of lozenges or pastilles obtained by high boiling techniques, provided that the final product contains maximum 50 ppm of sulfur dioxide.

Loss on drying (*2.2.32*)

Maximum 30.0 per cent, determined on 1.000 g. Triturate the sample with 3.000 g of *kieselguhr G R*, previously dried at 80 °C under high vacuum for 2 h, and dry at 80 °C under high vacuum for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

Dextrose equivalent
(DE): within 10 per cent of the nominal value.

Weigh an amount of the substance to be examined equivalent to 2.85-3.15 g of reducing carbohydrates, calculated as dextrose equivalent, into a 500 mL volumetric flask. Dissolve in *water R* and dilute to 500.0 mL with the same solvent. Transfer the solution to a 50 mL burette.

Pipette 25.0 mL of *cupri-tartaric solution R* into a 250 mL flask and add 18.5 mL of the test solution from the burette, mix and add a few glass beads. Place the flask on a hot plate, previously adjusted so that the solution begins to boil after 2 min ± 15 s. Allow to boil for exactly 120 s, add 1 mL of a 1 g/L solution of *methylene blue R* and titrate with the test solution (V_1) until the blue colour disappears. Maintain the solution at boiling throughout the titration.

Standardise the cupri-tartaric solution using a 6.00 g/L solution of *glucose R* (V_0).

Calculate the dextrose equivalent using the following expression:

$$\frac{300 \times V_0 \times 100}{V_1 \times M \times D}$$

V_0 = total volume of glucose standard solution, in millilitres,
V_1 = total volume of test solution, in millilitres,
M = mass of the sample, in grams,
D = percentage content of dry matter in the substance.

LABELLING
The label states the dextrose equivalent (DE) (= nominal value).

Ph Eur

Spray-dried Liquid Glucose

(Ph. Eur. monograph 1525)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of glucose, oligosaccharides and polysaccharides, obtained by the partial hydrolysis of starch.

The degree of hydrolysis, expressed as dextrose equivalent (DE), is not less than 20 (nominal value).

CHARACTERS
Appearance
White or almost white, slightly hygroscopic powder or granules.

Solubility
Freely soluble in water.

IDENTIFICATION
A. Dissolve 0.1 g in 2.5 mL of *water R* and heat with 2.5 mL of *cupri-tartaric solution R*. A red precipitate is formed.

B. Dip, for 1 s, a suitable stick with a reactive pad containing glucose-oxidase, peroxidase and a hydrogen-donating substance, such as tetramethylbenzidine, in a 5 g/L solution of the substance to be examined. Observe the colour of the reactive pad; within 60 s the colour changes from yellow to green or blue.

C. It is a powder or granules.

D. Dextrose equivalent (see Tests).

TESTS
Solution S
Dissolve 12.5 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 7.0.

Mix 1 mL of a 223.6 g/L solution of *potassium chloride R* and 30 mL of solution S.

Sulfur dioxide (*2.5.29*)
Maximum 20 ppm.

Loss on drying (*2.2.32*)
Maximum 6.0 per cent, determined on 10.00 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.5 per cent, determined on 1.0 g.

Dextrose equivalent
(DE): within 10 per cent of the nominal value.

Weigh an amount of the substance to be examined equivalent to 2.85-3.15 g of reducing carbohydrates, calculated as dextrose equivalent, into a 500 mL volumetric flask. Dissolve in *water R* and dilute to 500.0 mL with the same solvent. Transfer the solution to a 50 mL burette.

Pipette 25.0 mL of *cupri-tartaric solution R* into a 250 mL flask and add 18.5 mL of the test solution from the burette, mix and add a few glass beads. Place the flask on a hot plate, previously adjusted so that the solution begins to boil after 2 min ± 15 s. Allow to boil for exactly 120 s, add 1 mL of a 1 g/L solution of *methylene blue R* and titrate with the test solution (V_1) until the blue colour disappears. Maintain the solution at boiling throughout the titration.

Standardise the cupri-tartaric solution using a 6.00 g/L solution of *glucose R* (V_0).

Calculate the dextrose equivalent using the following expression:

$$\frac{300 \times V_0 \times 100}{V_1 \times M \times D}$$

V_0 = total volume of glucose standard solution, in millilitres;
V_1 = total volume of test solution, in millilitres;
M = mass of the sample, in grams;
D = percentage content of dry matter in the substance.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

LABELLING
The label states the dextrose equivalent (DE) (= nominal value).

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute

to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for spray-dried liquid glucose used as filler or binder for wet granulation.

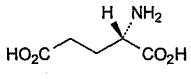
Dextrose equivalent
(see Tests).

Particle-size distribution (*2.9.31* or *2.9.38*)

Ph Eur

Glutamic Acid

(Ph. Eur. monograph 0750)



$C_5H_9NO_4$ 147.1 *56-86-0*

Action and use
Amino acid.

Ph Eur

DEFINITION
Glutamic acid contains not less than 98.5 per cent and not more than the equivalent of 100.5 per cent of (2*S*)-2-aminopentanedioic acid, calculated with reference to the dried substance.

CHARACTERS
A white or almost white, crystalline powder or colourless crystals, freely soluble in boiling water, slightly soluble in cold water, practically insoluble in acetic acid, in acetone and in alcohol.

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *glutamic acid CRS*. Examine the substances prepared as discs. If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum quantity of *water R*, evaporate to dryness at 60 °C and record new spectra using the residues.

C. Examine the chromatograms obtained in the test for ninhydrin-positive substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 2.0 mL of solution S (see Tests) add 0.1 mL of *phenolphthalein solution R* and 3.0 mL to 3.5 mL of *1 M sodium hydroxide* to change the colour of the indicator to red. Add a mixture of 3 mL of *formaldehyde solution R*, 3 mL of *carbon dioxide-free water R* and 0.1 mL of *phenolphthalein solution R*, to which sufficient *1 M sodium hydroxide* has been added to produce a pink colour. The solution is decolourised. Add *1 M sodium hydroxide* until a red colour is produced. The total volume of *1 M sodium hydroxide* used is 4.0 mL to 4.7 mL.

TESTS
Solution S
Dissolve 5.00 g in *1 M hydrochloric acid* with gentle heating, and dilute to 50.0 mL with the same acid.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
+ 30.5 to + 32.5, determined on solution S and calculated with reference to the dried substance.

Ninhydrin-positive substances
Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel plate R*.

Test solution (a) Dissolve 0.10 g of the substance to be examined in 5 mL of *dilute ammonia R2* and dilute to 10 mL with *water R*.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with *water R*.

Reference solution (a) Dissolve 10 mg of *glutamic acid CRS* in *water R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (b) to 20 mL with *water R*.

Reference solution (c) Dissolve 10 mg of *glutamic acid CRS* and 10 mg of *aspartic acid CRS* in *water R* and dilute to 25 mL with the same solvent.

Apply to the plate 5 µL of each solution. Dry the plate in a current of air for 15 min. Develop over a path of 15 cm using a mixture of 20 volumes of *glacial acetic acid R*, 20 volumes of *water R* and 60 volumes of *butanol R*. Allow the plate to dry in air, spray with *ninhydrin solution R* and heat at 100-105 °C for 15 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows 2 clearly separated spots.

Chlorides (*2.4.4*)
Dissolve 0.25 g in 3 mL of *dilute nitric acid R* and dilute to 15 mL with *water R*. The solution, to which 1 mL of *water R* is added instead of *dilute nitric acid R*, complies with the limit test for chlorides (200 ppm).

Sulfates (*2.4.13*)
Dilute 5 mL of solution S to 15 mL with *distilled water R*. The solution complies with the limit test for sulfates (300 ppm).

Ammonium (*2.4.1*)
50 mg complies with limit test B for ammonium (200 ppm). Prepare the standard using 0.1 mL of *ammonium standard solution (100 ppm NH_4) R*.

Iron (*2.4.9*)
In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. The aqueous layer complies with the limit test for iron (10 ppm).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.130 g in 50 mL of *carbon dioxide-free water R* with gentle heating. Cool. Using 0.1 mL of *bromothymol blue solution R1* as indicator, titrate with *0.1 M sodium hydroxide* until the colour changes from yellow to blue.

1 mL of *0.1 M sodium hydroxide* is equivalent to 14.71 mg of $C_5H_9NO_4$.

STORAGE
Protected from light.

Ph Eur

Strong Glutaraldehyde Solution

$C_5H_8O_2$ 100.1 (anhydrous) *111-30-8*

Action and use
Used in treatment of warts.

Preparation
Glutaraldehyde Solution

DEFINITION
Strong Glutaraldehyde Solution is an aqueous solution of glutaraldehyde (pentanedial). It contains not less than 47.0% and not more than 53.0% w/w of glutaraldehyde, $C_5H_8O_2$.

CHARACTERISTICS
A colourless or almost colourless solution.

IDENTIFICATION
A. Heat 1 mL with 10 mL of a solution containing 1 g of *hydroxylamine hydrochloride* and 2 g of *sodium acetate* in *water* on a water bath for 10 minutes, allow to cool and filter. The *melting point* of the residue, after washing with *water* and drying at 105°, is about 178°, Appendix V A.

B. Add 0.05 mL to 2 mL of *ammoniacal silver nitrate solution* and mix gently for a few minutes. Silver is deposited.

TESTS
Acidity
Dilute 10 mL with 10 mL of *carbon dioxide-free water* and titrate with *0.1M sodium hydroxide VS* using *bromothymol blue solution R3* as indicator. Not more than 5.0 mL of *0.1M sodium hydroxide VS* is required to change the colour of the solution.

Clarity and colour of solution
Dilute 1 volume with 4 volumes of *water*. The resulting solution is *clear*, Appendix IV A, and not more intensely coloured than *reference solution B_6*, Appendix IV B, Method I.

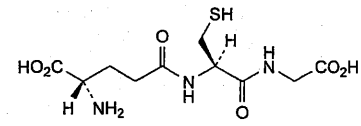
Weight per mL
1.126 to 1.134 g, Appendix V G.

ASSAY
Dissolve 4 g in 100 mL of a 7% w/v solution of *hydroxylamine hydrochloride* previously neutralised to *bromophenol blue solution* with *1M sodium hydroxide VS* and allow to stand for 30 minutes. Add 20 mL of *petroleum spirit (boiling range, 40° to 60°)* and titrate with *1M sodium hydroxide VS* until the colour of the aqueous phase matches that of a 7% w/v solution of *hydroxylamine hydrochloride* previously neutralised to *bromophenol blue solution* with *1M sodium hydroxide VS*. Each mL of *1M sodium hydroxide VS* is equivalent to 50.05 mg of $C_5H_8O_2$.

STORAGE
Strong Glutaraldehyde Solution should be stored at a temperature not exceeding 15°.

Glutathione

(Ph. Eur. monograph 1670)



$C_{10}H_{17}N_3O_6S$ 307.3 *70-18-8*

Ph Eur

DEFINITION
L-γ-Glutamyl-L-cysteinylglycine.
Fermentation product.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison glutathione CRS.

TESTS
Solution S
Dissolve 5.0 g in *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
−17.5 to −15.5 (dried substance).
Dissolve 1.0 g in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Capillary electrophoresis (*2.2.47*). *Prepare the solutions immediately before use.*

Internal standard solution (a) Dissolve 0.100 g of *phenylalanine R* in the electrolyte solution and dilute to 50.0 mL with the same solution.

Internal standard solution (b) Dilute 10.0 mL of internal standard solution (a) to 100.0 mL with the electrolyte solution.

Test solution (a) Dissolve 0.200 g of the substance to be examined in the electrolyte solution and dilute to 10.0 mL with the same solution.

Test solution (b) Dissolve 0.200 g of the substance to be examined in internal standard solution (b) and dilute to 10.0 mL with the same solution.

Reference solution (a) Dissolve 20.0 mg of the substance to be examined in internal standard solution (a) and dilute to 10.0 mL with the same solution.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the electrolyte solution.

Reference solution (c) Dissolve 0.200 g of the substance to be examined in 5 mL of the electrolyte solution. Add 1.0 mL of internal standard solution (a), 0.5 mL of a 2 mg/mL solution of *L-cysteine R* (impurity B) in the electrolyte solution, 0.5 mL of a 2 mg/mL solution of oxidised L-glutathione R (impurity C) in the electrolyte solution and 0.5 mL of a 2 mg/mL solution of *L-γ-glutamyl-L-cysteine R* (impurity D) in the electrolyte solution. Dilute to 10.0 mL with the electrolyte solution.

Capillary:
— *material*: uncoated fused silica;
— *size*: length to the detector cell = 0.5 m; total length = 0.6 m; Ø = 75 µm.

Temperature 25 °C.

Electrolyte solution Dissolve 1.50 g of *anhydrous sodium dihydrogen phosphate R* in 230 mL of *water R* and adjust to pH 1.80 with *phosphoric acid R*. Dilute to 250.0 mL with *water R*. Check the pH and, if necessary, adjust with *phosphoric acid R* or *dilute sodium hydroxide solution R*.

Detection Spectrophotometer at 200 nm.

Preconditioning of a new capillary Rinse the new capillary before the first injection with *0.1 M hydrochloric acid* at 138 kPa for 20 min and with *water R* at 138 kPa for 10 min; for complete equilibration, condition the capillary with the electrolyte solution at 350 kPa for 40 min, and subsequently at a voltage of 20 kV for 60 min.

Preconditioning of the capillary Rinse the capillary with the electrolyte solution at 138 kPa for 40 min.

Between-run rinsing Rinse the capillary with *water R* at 138 kPa for 1 min, with *0.1 M sodium hydroxide* at 138 kPa for 2 min, with *water R* at 138 kPa for 1 min, with *0.1 M hydrochloric acid* at 138 kPa for 3 min and with the electrolyte solution at 138 kPa for 10 min.

Injection Test solutions (a) and (b), reference solutions (b) and (c) and the electrolyte solution (blank): under pressure (3.45 kPa) for 5 s.

Migration Apply a voltage of 20 kV.

Run time 45 min.

Relative migration With reference to the internal standard (about 14 min): impurity A = about 0.77; impurity B = about 1.04; impurity E = about 1.2; impurity C = about 1.26; impurity D = about 1.3.

System suitability:
— *resolution*: minimum 1.5 between the peaks due to the internal standard and impurity B in the electropherogram obtained with reference solution (c); if necessary, increase the pH with *dilute sodium hydroxide solution R*;
— *peak-to-valley ratio*: minimum 2.5, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to glutathione in the electropherogram obtained with reference solution (c); if necessary, lower the pH with *phosphoric acid R*;
— check that in the electropherogram obtained with test solution (a) there is no peak with the same migration time as the internal standard (in such case correct the area of the phenylalanine peak).

Limits Test solution (b):
— *corrected areas*: divide all the peak areas by the corresponding migration times;
— *correction factors*: for the calculation of content, multiply the ratio of time-corrected peak areas of impurity and the internal standard by the corresponding correction factor: impurity B = 3.0; impurity D = 1.4;
— *impurity C*: not more than 1.5 times the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (1.5 per cent);
— *impurity D*: not more than the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (1.0 per cent);
— *impurities A, B, E*: for each impurity, not more than 0.5 times the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (0.5 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (0.2 per cent);
— *total*: not more than 2.5 times the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (2.5 per cent);
— *disregard limit*: 0.05 times the ratio of the area of the peak due to glutathione to the area of the peak due to the internal standard in the electropherogram obtained with reference solution (b) (0.05 per cent).

Chlorides
Maximum 200 ppm.

Dissolve 0.5 g in 5 mL of *dilute nitric acid R* and dilute to 10 mL with the same solvent. Add 10 mL of *strong hydrogen peroxide solution R* and heat on a water-bath for 30 min. Cool and dilute to 50 mL with *water R*. Add 1 mL of *silver nitrate solution R2* and mix. Allow to stand for 5 min protected from light. Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 2 mL of *chloride standard solution (50 ppm Cl) R*. Examine the tubes laterally against a black background.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dilute 5 mL of solution S to 15 mL with *distilled water R*.

Ammonium (*2.4.1, Method B*)
Maximum 200 ppm, determined on 50 mg.

Prepare the standard using 0.1 mL of *ammonium standard solution (100 ppm NH_4) R*.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers, add 10 mL of *water R* and shake for 3 min. The aqueous layer complies with the test.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

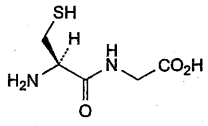
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
In a ground-glass-stoppered flask, dissolve 0.500 g of the substance to be examined and 2 g of *potassium iodide R* in 50 mL of *water R*. Cool the solution in iced water and add 10 mL of *hydrochloric acid R1* and 20.0 mL of *0.05 M iodine*. Stopper the flask and allow to stand in the dark for 15 min. Titrate with *0.1 M sodium thiosulfate* using 1 mL of *starch solution R*, added towards the end of the titration, as indicator. Carry out a blank titration.

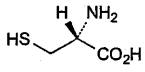
1 mL of *0.05 M iodine* is equivalent to 30.73 mg of $C_{10}H_{17}N_3O_6S$.

STORAGE
Protected from light.

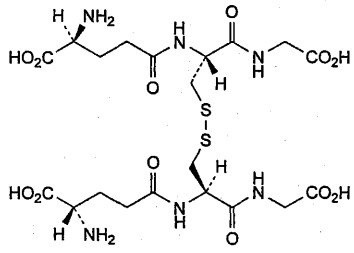
IMPURITIES
Specified impurities *A, B, C, D, E.*



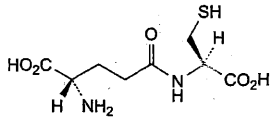
A. L-cysteinylglycine,



B. (2*R*)-2-amino-3-sulfanylpropanoic acid (cysteine),



C. bis(L-γ-glutamyl-L-cysteinylglycine) disulfide (L-glutathione oxidised),



D. L-γ-glutamyl-L-cysteine,

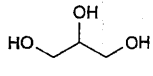
E. unknown structure (product of degradation).

Ph Eur

Glycerol

Glycerin

(Ph. Eur. Monograph 0496)



$C_3H_8O_3$ 92.1 *56-81-5*

Action and use
Lubricant; laxative.

Preparations
Glycerol Eye Drops

Glycerol Suppositories

Ph Eur

DEFINITION
Propane-1,2,3-triol.

Content
98.0 per cent *m/m* to 101.0 per cent *m/m* (anhydrous substance).

CHARACTERS
Aspect Syrupy liquid, unctuous to the touch, colourless or almost colourless, clear, very hygroscopic.

Solubility
Miscible with water and with ethanol (96 per cent), slightly soluble in acetone, practically insoluble in fatty oils and in essential oils.

IDENTIFICATION
First identification: A, B.

Second identification: A, C.

A. Refractive index (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation To 5 mL add 1 mL of *water R* and mix carefully.

Comparison *Ph. Eur. reference spectrum of glycerol (85 per cent)*.

C. Relative density (*2.2.5*): 1.258 to 1.268.

TESTS
Solution S
Dilute 100.0 g to 200.0 mL with *carbon dioxide-free water R*.

Appearance of solution
Solution S is clear (*2.2.1*). Dilute 10 mL of solution S to 25 mL with *water R*. The solution is colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 50 mL of solution S add 0.5 mL of *phenolphthalein solution R*. The solution is colourless. Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Refractive index (*2.2.6*)
1.470 to 1.475.

Aldehydes
Maximum 10 ppm.

Place 7.5 mL of solution S in a ground-glass-stoppered flask and add 7.5 mL of *water R* and 1.0 mL of *decolorised pararosaniline solution R*. Close the flask and allow to stand for 1 h at a temperature of 25 ± 1 °C. The absorbance (*2.2.25*) of the solution measured at 552 nm is not greater than that of a standard prepared at the same time and in the same manner using 7.5 mL of *formaldehyde standard solution (5 ppm CH_2O) R* and 7.5 mL of *water R*. The test is not valid unless the standard is pink.

Esters
Add 10.0 mL of *0.1 M sodium hydroxide* to the final solution obtained in the test for acidity or alkalinity. Boil under a reflux condenser for 5 min. Cool. Add 0.5 mL of *phenolphthalein solution R* and titrate with *0.1 M hydrochloric acid*. Not less than 8.0 mL of *0.1 M hydrochloric acid* is required to change the colour of the indicator.

Impurity A and related substances
Gas chromatography (*2.2.28*).

Test solution Dilute 10.0 mL of solution S to 100.0 mL with *water R*.

Reference solution (a) Dilute 10.0 g of *glycerol R1* to 20.0 mL with *water R*. Dilute 10.0 mL of the solution to 100.0 mL with *water R*.

Reference solution (b) Dissolve 1.000 g of *diethylene glycol R* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with reference solution (a). Dilute 1.0 mL of this solution to 20.0 mL with reference solution (a).

Reference solution (d) Mix 1.0 mL of the test solution and 5.0 mL of reference solution (b) and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (e) Dilute 5.0 mL of reference solution (b) to 100.0 mL with *water R*.

Column:
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: 6 per cent polycyanopropylphenyl siloxane and 94 per cent of polydimethylsiloxane.

Carrier gas *helium for chromatography R*.

Split ratio 1:10.

Linear velocity 38 cm/s.

Temperature:

	Time (min)	Temperature (°C)
Column	0	100
	0 - 16	100 → 220
	16 - 20	220
Injection port		220
Detector		250

Detection Flame ionisation.

Injection 0.5 µL.

Elution order Impurity A, glycerol.

System suitability Reference solution (d):
— *resolution*: minimum 7.0 between the peaks due to impurity A and glycerol.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *any other impurity with a retention time less than the retention time of glycerol*: not more than the area of the peak due to impurity A in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *total of all impurities with retention times greater than the retention time of glycerol*: not more than 5 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *disregard limit*: 0.05 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (e) (0.05 per cent).

Halogenated compounds
Maximum 35 ppm.

To 10 mL of solution S add 1 mL of *dilute sodium hydroxide solution R*, 5 mL of *water R* and 50 mg of *halogen-free nickel-aluminium alloy R*. Heat on a water-bath for 10 min, allow to cool and filter. Rinse the flask and the filter with *water R* until 25 mL of filtrate is obtained. To 5 mL of the filtrate add 4 mL of *ethanol (96 per cent) R*, 2.5 mL of *water R*, 0.5 mL of *nitric acid R* and 0.05 mL of *silver nitrate solution R2* and mix. Allow to stand for 2 min.
Any opalescence in the solution is not more intense than that in a standard prepared at the same time by mixing 7.0 mL of *chloride standard solution (5 ppm Cl) R*, 4 mL of *ethanol (96 per cent) R*, 0.5 mL of *water R*, 0.5 mL of *nitric acid R* and 0.05 mL of *silver nitrate solution R2*.

Sugars
To 10 mL of solution S add 1 mL of *dilute sulfuric acid R* and heat on a water-bath for 5 min. Add 3 mL of carbonate-free *dilute sodium hydroxide solution R* (prepared by the method described for carbonate-free *1 M sodium hydroxide*), mix and add dropwise 1 mL of freshly prepared *copper sulfate solution R*. The solution is clear and blue. Continue heating on the water-bath for 5 min. The solution remains blue and no precipitate is formed.

Chlorides (*2.4.4*)
Maximum 10 ppm.

Dilute 1 mL of solution S to 15 mL with *water R*. Prepare the standard using 1 mL of *chloride standard solution (5 ppm Cl) R* diluted to 15 mL with *water R*.

Water (*2.5.12*)
Maximum 2.0 per cent, determined on 1.000 g.

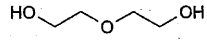
Sulfated ash (*2.4.14*)
Maximum 0.01 per cent, determined on 5.0 g after heating to boiling and ignition.

ASSAY
Thoroughly mix 0.075 g with 45 mL of *water R*.
Add 25.0 mL of a mixture of 1 volume of *0.1 M sulfuric acid* and 20 volumes of *0.1 M sodium periodate*. Allow to stand protected from light for 15 min. Add 5.0 mL of a 500 g/L solution of *ethylene glycol R* and allow to stand protected from light for 20 min. Using 0.5 mL of *phenolphthalein solution R* as indicator, titrate with *0.1 M sodium hydroxide*. Carry out a blank titration.

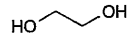
1 mL of *0.1 M sodium hydroxide* is equivalent to 9.21 mg of $C_3H_8O_3$.

STORAGE
In an airtight container.

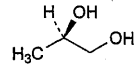
IMPURITIES



A. 2,2′-oxydi(ethan-1-ol) (diethylene glycol),



B. ethane-1,2-diol (ethylene glycol),



and enantiomer

C. (2*RS*)-propane-1,2-diol (propylene glycol).

Ph Eur

Glycerol (85 per cent)

(Ph. Eur. monograph 0497)

Ph Eur

DEFINITION

Aqueous solution of propane-1,2,3-triol.

Content

83.5 per cent *m/m* to 88.5 per cent *m/m* of propane-1,2,3-triol ($C_3H_8O_3$; M_r 92.1).

CHARACTERS

Aspect Syrupy liquid, unctuous to the touch, colourless or almost colourless, clear, very hygroscopic.

Solubility

Miscible with water and with ethanol (96 per cent), slightly soluble in acetone, practically insoluble in fatty oils and in essential oils.

IDENTIFICATION

First identification: A, B.

Second identification: A, C.

A. Refractive index (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of glycerol (85 per cent).*

C. Relative density (*2.2.5*): 1.221 to 1.232.

TESTS

Solution S

Dilute 117.6 g to 200.0 mL with *carbon dioxide-free water R.*

Appearance of solution

Solution S is clear (*2.2.1*). Dilute 10 mL of solution S to 25 mL with *water R.* The solution is colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 50 mL of solution S add 0.5 mL of *phenolphthalein solution R.* The solution is colourless. Not more than 0.2 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator to pink.

Refractive index (*2.2.6*)

1.449 to 1.455.

Aldehydes

Maximum 10 ppm.

Place 7.5 mL of solution S in a ground-glass-stoppered flask and add 7.5 mL of *water R* and 1.0 mL of *decolorised pararosaniline solution R.* Close the flask and allow to stand for 1 h at a temperature of 25 ± 1 °C. The absorbance (*2.2.25*) of the solution measured at 552 nm is not greater than that of a standard prepared at the same time and in the same manner using 7.5 mL of *formaldehyde standard solution (5 ppm CH_2O) R* and 7.5 mL of *water R.* The test is not valid unless the standard is pink.

Esters

Add 10.0 mL of *0.1 M sodium hydroxide* to the final solution obtained in the test for acidity or alkalinity. Boil under a reflux condenser for 5 min. Cool. Add 0.5 mL of *phenolphthalein solution R* and titrate with *0.1 M hydrochloric acid.* Not less than 8.0 mL of *0.1 M hydrochloric acid* is required to change the colour of the indicator.

Impurity A and related substances

Gas chromatography (*2.2.28*).

Test solution Dilute 10.0 mL of solution S to 100.0 mL with *water R.*

Reference solution (a) Dilute 11.8 g of *glycerol (85 per cent) R1* to 20.0 mL with *water R.* Dilute 10.0 mL of the solution to 100.0 mL with *water R.*

Reference solution (b) Dissolve 1.000 g of *diethylene glycol R* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with reference solution (a). Dilute 1.0 mL of this solution to 20.0 mL with reference solution (a).

Reference solution (d) Mix 1.0 mL of the test solution and 5.0 mL of reference solution (b) and dilute to 100.0 mL with *water R.* Dilute 1.0 mL of this solution to 10.0 mL with *water R.*

Reference solution (e) Dilute 5.0 mL of reference solution (b) to 100.0 mL with *water R.*

Column:

— *size*: *l* = 30 m, Ø = 0.53 mm;

— *stationary phase*: 6 per cent polycyanolpropylphenyl siloxane and 94 per cent of polydimethylsiloxane.

Carrier gas *helium for chromatography R.*

Split ratio 1:10.

Linear velocity 38 cm/s.

Temperature:

	Time (min)	Temperature (°C)
Column	0	100
	0 - 16	100 → 220
	16 - 20	220
Injection port		220
Detector		250

Detection Flame ionisation.

Injection 0.5 µL.

Elution order Impurity A, glycerol.

System suitability Reference solution (d):

— *resolution*: minimum 7.0 between the peaks due to impurity A and glycerol.

Limits:

— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

— *any other impurity with a retention time less than the retention time of glycerol*: not more than the area of the peak due to impurity A in the chromatogram obtained with reference solution (c) (0.1 per cent);

— *total of all impurities with retention times greater than the retention time of glycerol*: not more than 5 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *disregard limit*: 0.05 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (e) (0.05 per cent).

Halogenated compounds

Maximum 30 ppm.

To 10 mL of solution S add 1 mL of *dilute sodium hydroxide solution R,* 5 mL of *water R* and 50 mg of *halogen-free nickel-aluminium alloy R.* Heat on a water-bath for 10 min, allow to cool and filter. Rinse the flask and the filter with *water R* until 25 mL of filtrate is obtained. To 5 mL of the filtrate add 4 mL of *ethanol (96 per cent) R,* 2.5 mL of *water R,* 0.5 mL of *nitric acid R* and 0.05 mL of *silver nitrate solution R2* and mix. Allow to stand for 2 min.

Any opalescence in the solution is not more intense than that

in a standard prepared at the same time by mixing 7.0 mL of *chloride standard solution (5 ppm Cl) R*, 4 mL of *ethanol (96 per cent) R*, 0.5 mL of *water R*, 0.5 mL of *nitric acid R* and 0.05 mL of *silver nitrate solution R2*.

Sugars
To 10 mL of solution S add 1 mL of *dilute sulfuric acid R* and heat on a water-bath for 5 min. Add 3 mL of carbonate-free *dilute sodium hydroxide solution R* (prepared by the method described for carbonate-free *1 M sodium hydroxide*), mix and add dropwise 1 mL of freshly prepared *copper sulfate solution R*. The solution is clear and blue. Continue heating on the water-bath for 5 min. The solution remains blue and no precipitate is formed.

Chlorides (*2.4.4*)
Maximum 10 ppm.

Dilute 1 mL of solution S to 15 mL with *water R*. Prepare the standard using 1 mL of *chloride standard solution (5 ppm Cl) R* diluted to 15 mL with *water R*.

Water (*2.5.12*)
12.0 per cent to 16.0 per cent, determined on 0.200 g.

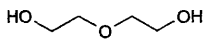
Sulfated ash (*2.4.14*)
Maximum 0.01 per cent, determined on 5.0 g after heating to boiling and ignition.

ASSAY
Thoroughly mix 0.075 g with 45 mL of *water R*.
Add 25.0 mL of a mixture of 1 volume of *0.1 M sulfuric acid* and 20 volumes of *0.1 M sodium periodate*. Allow to stand protected from light for 15 min. Add 5.0 mL of a 500 g/L solution of *ethylene glycol R* and allow to stand protected from light for 20 min. Using 0.5 mL of *phenolphthalein solution R* as indicator, titrate with *0.1 M sodium hydroxide*. Carry out a blank titration.

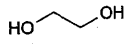
1 mL of *0.1 M sodium hydroxide* is equivalent to 9.21 mg of $C_3H_8O_3$.

STORAGE
In an airtight container.

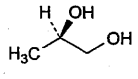
IMPURITIES



A. 2,2′-oxydi(ethan-1-ol) (diethylene glycol),



B. ethane-1,2-diol (ethylene glycol),



and enantiomer

C. (2*RS*)-propane-1,2-diol (propylene glycol).

Ph Eur

Glycerol Dibehenate

(Ph. Eur. monograph 1427)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of diacylglycerols, mainly dibehenylglycerol, together with variable quantities of mono- and triacylglycerols, obtained by esterification of *glycerol (0496)* with behenic (docosanoic) acid.

Content
— *monoacylglycerols*: 15.0 per cent to 23.0 per cent;
— *diacylglycerols*: 40.0 per cent to 60.0 per cent;
— *triacylglycerols*: 21.0 per cent to 35.0 per cent.

CHARACTERS
Appearance
Hard, waxy mass, or powder or white or almost white, unctuous flakes.

Solubility
Practically insoluble in water, soluble in methylene chloride, partly soluble in hot ethanol (96 per cent).

IDENTIFICATION
A. Melting point (*2.2.14*): 65 °C to 77 °C.

B. Composition of fatty acids (see Tests).

C. It complies with the assay (content of diacylglycerols).

TESTS
Acid value (*2.5.1*)
Maximum 4.0, determined on 1.0 g using a mixture of equal volumes of *ethanol (96 per cent) R* and *toluene R* as solvent and with gentle heating.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Saponification value (*2.5.6*)
145 to 165.

Carry out the titration with heating.

Free glycerol
Maximum 1.0 per cent, determined as described under Assay.

Composition of fatty acids (*2.4.22, Method C*)
Raise the temperature of the column to 240 °C and use the mixture of calibrating substances in Table 2.4.22.-3.

Composition of the fatty acid fraction of the substance:
— *palmitic acid*: maximum 3.0 per cent;
— *stearic acid*: maximum 5.0 per cent;
— *arachidic acid*: maximum 10.0 per cent;
— *behenic acid*: minimum 83.0 per cent;
— *erucic acid*: maximum 3.0 per cent;
— *lignoceric acid*: maximum 3.0 per cent.

Nickel (*2.4.31*)
Maximum 1 ppm.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.00 g.
Use *pyridine R* as the solvent.

Total ash (*2.4.16*)
Maximum 0.1 per cent, determined on 1.00 g.

ASSAY

Size-exclusion chromatography (*2.2.30*).

Stock solution Place 0.100 g of *glycerol R* in a flask and dilute to 25.0 mL with *tetrahydrofuran R*.

Test solution In a 15 mL flask, weigh 0.200 g (*m*) of the substance to be examined and add 5.0 mL of *tetrahydrofuran R*. Heat gently, at about 35 °C, and shake to dissolve. Reweigh the flask and calculate the total mass of solvent and substance (*M*); use immediately.

Reference solutions Into four 15 mL flasks, introduce respectively 0.25 mL, 0.5 mL, 1.0 mL and 2.5 mL of the stock solution and add 5.0 mL of *tetrahydrofuran R*. Weigh each flask and calculate the concentration of glycerol in milligrams per gram of each reference solution.

Column:

— *size*: *l* = 0.6 m, Ø = 7 mm;

— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R*.

Flow rate 1 mL/min.

Detection Differential refractive index.

Injection 40 µL; when injecting the test solution, maintain the flask at about 35 °C to avoid precipitation.

Relative retention With reference to glycerol (retention time = about 15 min): triacylglycerols = about 0.73; diacylglycerols = about 0.76; monoacylglycerols = about 0.82.

Calculations:

— *free glycerol*: from the calibration curve obtained with the reference solutions, determine the concentration (*C*) in milligrams per gram in the test solution and calculate the percentage content (*A*) in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *free fatty acids*: calculate the percentage content of free fatty acids (*D*) using the following expression:

$$\frac{I_A \times 340}{561.1}$$

I_A = acid value.

— *monoacylglycerols*: calculate the percentage content of monoacylglycerols using the following expression:

$$\left[\frac{X}{X+Y+Z}(100-A-B)\right]-D$$

A = percentage content of free glycerol (see Tests);
B = percentage content of water (see Tests);
D = percentage content of free fatty acids;
X = area of the peak due to monoacylglycerols;
Y = area of the peak due to diacylglycerols;
Z = area of the peak due to triacylglycerols.

— *diacylglycerols*: calculate the percentage content of diacylglycerols using the following expression:

$$\frac{Y}{X+Y+Z}(100-A-B)$$

— *triacylglycerols*: calculate the percentage content of triacylglycerols using the following expression:

$$\frac{Z}{X+Y+Z}(100-A-B)$$

Ph Eur

Glycerol Distearate

(Ph. Eur. monograph 1428)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of diacylglycerols, mainly distearoylglycerol, together with variable quantities of mono- and triacylglycerols. It is obtained by partial glycerolysis of vegetable oils containing triacylglycerols of palmitic (hexadecanoic) and stearic (octadecanoic) acids or by esterification of glycerol with stearic acid. The fatty acids may be of vegetable or animal origin.

Content

— *monoacylglycerols*: 8.0 per cent to 22.0 per cent;
— *diacylglycerols*: 40.0 per cent to 60.0 per cent;
— *triacylglycerols*: 25.0 per cent to 35.0 per cent.

CHARACTERS

Appearance

Hard, waxy mass or powder, or white or almost white, unctuous flakes.

Solubility

Practically insoluble in water, soluble in methylene chloride, partly soluble in hot ethanol (96 per cent).

IDENTIFICATION

A. Melting point (*2.2.14*): 50 °C to 60 °C (types I and II), 50 °C to 70 °C (type III).

B. Composition of fatty acids (see Tests) according to the type stated on the label.

C. It complies with the limits of the assay (diacylglycerol content).

TESTS

Acid value (*2.5.1*)

Maximum 6.0, determined on 1.0 g.

Use a mixture of equal volumes of *ethanol (96 per cent) R* and *toluene R* as solvent and heat gently.

Iodine value (*2.5.4, Method A*)

Maximum 3.0.

Saponification value (*2.5.6*)

165 to 195, determined on 2.0 g. Carry out the titration with heating.

Free glycerol

Maximum 1.0 per cent, determined as described under Assay.

Composition of fatty acids (*2.4.22, Method C*)

Use the mixture of calibrating substances in Table 2.4.22.-1.

Composition of the fatty-acid fraction of the substance:

Glycerol distearate	Composition of fatty acids
Type I	*Stearic acid*: 40.0 per cent to 60.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 90.0 per cent
Type II	*Stearic acid*: 60.0 per cent to 80.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 90.0 per cent
Type III	*Stearic acid*: 80.0 per cent to 99.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 96.0 per cent

Nickel (*2.4.31*)
Maximum 1 ppm.

Water
(*2.5.12*): maximum 1.0 per cent, determined on 1.00 g. Use *pyridine R* as the solvent.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY
Size-exclusion chromatography (*2.2.30*).

Test solution Into a 15 mL flask, weigh 0.200 g (*m*) of the substance to be examined. Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into three 15 mL flasks, respectively weigh 2.0 mg, 5.0 mg and 10.0 mg of *glycerol R* and add 5.0 mL of *tetrahydrofuran R* to each flask. Into a 4th flask, weigh 2.0 mg of *glycerol R* and add 10.0 mL of *tetrahydrofuran R*. Weigh the flasks again and calculate the concentration of glycerol in milligrams per gram for each reference solution.

Column:
— *size*: *l* = 0.6 m, Ø = 7 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R*.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to glycerol (retention time = about 15 min): triacylglycerols = about 0.75; diacylglycerols = about 0.80; monoacylglycerols and free fatty acids = about 0.85.

Calculations:
— *free glycerol*: from the calibration curve obtained with the reference solutions, determine the concentration (*C*) in milligrams per gram in the test solution and calculate the percentage content (*A*) in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *free fatty acids*: calculate the percentage content of free fatty acids (*D*) using the following expression:

$$\frac{I_A \times 340}{561.1}$$

I_A = acid value.

— *monoacylglycerols*: calculate the percentage content of monoacylglycerols using the following expression:

$$\left[\frac{X}{X+Y+Z}(100-A-B)\right]-D$$

B = percentage content of water (see Tests);
X = area of the peak due to monoacylglycerols and free fatty acids;
Y = area of the peak due to diacylglycerols;
Z = area of the peak due to triacylglycerols.

— *diacylglycerols*: calculate the percentage content of diacylglycerols using the following expression:

$$\frac{Y}{X+Y+Z}(100-A-B)$$

— *triacylglycerols*: calculate the percentage content of triacylglycerols using the following expression:

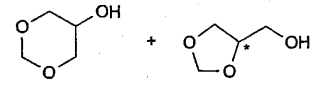
$$\frac{Z}{X+Y+Z}(100-A-B)$$

LABELLING
The label states the type of glycerol distearate.

Ph Eur

Glycerol Formal

(Ph. Eur. monograph 1671)



$C_4H_8O_3$ 104.1

Ph Eur

DEFINITION
Mixture of 1,3-dioxan-5-ol and (1,3-dioxolan-4-yl)methanol.

CHARACTERS
Appearance
Clear, colourless liquid.

Solubility
Miscible with water and with ethanol (96 per cent).

IDENTIFICATION
A. Relative density (see Tests).
B. Refractive index (see Tests).
C. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *glycerol formal CRS*.

TESTS
Appearance
The substance to be examined is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 6.5.
Dilute 1 mL to 10 mL with *carbon dioxide-free water R*.

Relative density (*2.2.5*)
1.210 to 1.220.

Refractive index (*2.2.6*)
1.445 to 1.455.

Peroxide value (*2.5.5*)
Maximum 15.

Formaldehyde
Maximum 200 ppm.
Dilute 0.250 g to 10 mL with *water R*. Add 2.0 mL of *acetylacetone reagent R2*, mix and heat on a water-bath at 60 °C for 20 min. Cool and dilute to 20.0 mL with *water R*. The absorbance (*2.2.25*) of the solution measured at 412 nm is not greater than that of a standard prepared at the same time and in the same manner using 5.0 mL of a dilution of *formaldehyde solution R* containing 10 µg of formaldehyde (CH_2O) per millilitre.

Water (*2.5.12*)
Maximum 0.5 per cent, determined on 5.000 g.

IMPURITIES
Specified impurities *A*.

A. formaldehyde.

STORAGE
Under nitrogen, in an airtight container.

Ph Eur

Glycerol Monocaprylate

(Ph. Eur. monograph 2213)

Ph Eur

DEFINITION
Mixture of monoacylglycerols, mainly mono-*O*-octanoylglycerol, containing variable quantities of di- and triacylglycerols, obtained by direct esterification of glycerol with caprylic (octanoic) acid, followed by a distillation step in the case of glycerol monocaprylate (type II).

Content
— *glycerol monocaprylate (type I)*:
  — *monoacylglycerols*: 45.0 per cent to 75.0 per cent;
  — *diacylglycerols*: 20.0 per cent to 50.0 per cent;
  — *triacylglycerols*: maximum 10.0 per cent;
— *glycerol monocaprylate (type II)*:
  — *monoacylglycerols*: minimum 80.0 per cent;
  — *diacylglycerols*: maximum 20.0 per cent;
  — *triacylglycerols*: maximum 5.0 per cent.

CHARACTERS
Appearance
Colourless or slightly yellow, oily liquid or soft mass.

Solubility
Practically insoluble in water, very soluble in ethanol (96 per cent) and freely soluble in methylene chloride.

IDENTIFICATION
A. Composition of fatty acids (see Tests).

B. It complies with the limits of the assay (monoacylglycerols).

TESTS
Acid value *(2.5.1)*
Maximum 3.0.

Composition of fatty acids *(2.4.22, Method C)*
Use the mixture of calibrating substances in Table 2.4.22.-2.

Composition of the fatty acid fraction of the substance:
— *caproic acid*: maximum 1.0 per cent;
— *caprylic acid*: minimum 90.0 per cent;
— *capric acid*: maximum 10.0 per cent;
— *lauric acid*: maximum 1.0 per cent;
— *myristic acid*: maximum 0.5 per cent.

Free glycerol
Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of *methylene chloride R*. Heat to about 50 °C then allow to cool. Add 100 mL of *water R*. Shake and add 25.0 mL of *periodic acetic acid solution R*. Shake again and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of *potassium iodide R* and allow to stand for 1 min. Add 1 mL of *starch solution R*. Titrate with *0.1 M sodium thiosulfate* until the aqueous phase becomes colourless. Carry out a blank titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 2.3 mg of glycerol.

Water *(2.5.12)*
Maximum 1.0 per cent, determined on 1.00 g.

Total ash *(2.4.16)*
Maximum 0.5 per cent.

ASSAY
Gas chromatography *(2.2.28)*: use the normalisation procedure.

Test solution To 0.25 g of the substance to be examined, add 5.0 mL of *tetrahydrofuran R* and shake to dissolve.

Reference solution (a) To 0.25 g of *glycerol monocaprylate CRS*, add 5.0 mL of *tetrahydrofuran R* and shake to dissolve.

Reference solution (b) To 50 mg of *glycerol 1-octanoate R* and 50 mg of *glycerol 1-decanoate R*, add 2.5 mL of *tetrahydrofuran R* and shake to dissolve.

Column:
— *size*: *l* = 10 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 2.3 mL/min.

Split ratio 1:50.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	60
	3 - 38	60 → 340
	38 - 50	340
Injection port		350
Detector		370

Detection Flame ionisation.

Injection 1 µL.

Identification of peaks Use the chromatogram supplied with *glycerol monocaprylate CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to mono-, di- and triacylglycerols.

System suitability Reference solution (b):
— *resolution*: minimum 5 between the peaks due to glycerol 1-octanoate and glycerol 1-decanoate.

For the calculation of the contents of mono-, di- and triacylglycerols, disregard the peaks with a retention time less than that of the monoacylglycerols, which are due to impurities of the solvent and to the free fatty acids.

Calculate the percentage content of free fatty acids (*C*) using the following expression:

$$\frac{I_A \times 144}{561.1}$$

I_A = acid value of glycerol monocaprylate.

Calculate the content of mono-, di- and triacylglycerols using the following equations:

$$\text{Monoacylglycerols content} = \frac{X \times (100 - A - B - C)}{100}$$

$$\text{Diacylglycerols content} = \frac{Y \times (100 - A - B - C)}{100}$$

$$\text{Triacylglycerols content} = \frac{Z \times (100 - A - B - C)}{100}$$

A = percentage content of free glycerol (see Tests);
B = percentage content of water;
X = monoacylglycerols content obtained by normalisation;
Y = diacylglycerols content obtained by normalisation;
Z = triacylglycerols content obtained by normalisation.

LABELLING
The label states the type of glycerol monocaprylate (type I or II).

Ph Eur

Glycerol Monocaprylocaprate

(Ph. Eur. monograph 2392)

Ph Eur

DEFINITION
Mixture of monoacylglycerols, mainly mono-*O*-octanoylglycerol and mono-*O*-decanoylglycerol, containing variable quantities of di- and triacylglycerols, obtained by direct esterification of glycerol with caprylic (octanoic) and capric (decanoic) acids, followed by a distillation step in the case of glycerol monocaprylocaprate (type II).

Content
— *glycerol monocaprylocaprate (type I)*:
  — *monoacylglycerols*: 45.0 per cent to 75.0 per cent;
  — *diacylglycerols*: 20.0 per cent to 50.0 per cent;
  — *triacylglycerols*: maximum 10.0 per cent;
— *glycerol monocaprylocaprate (type II)*:
  — *monoacylglycerols*: minimum 80.0 per cent;
  — *diacylglycerols*: maximum 20.0 per cent;
  — *triacylglycerols*: maximum 5.0 per cent.

CHARACTERS
Appearance
Colourless or slightly yellow, oily liquid or soft mass.

Solubility
Practically insoluble in water, very soluble in ethanol (96 per cent) and freely soluble in methylene chloride.

IDENTIFICATION
A. Composition of fatty acids (see Tests).

B. It complies with the limits of the assay (monoacylglycerols).

TESTS
Acid value *(2.5.1)*
Maximum 3.0.

Composition of fatty acids *(2.4.22, Method C)*
Use the mixture of calibrating substances in Table 2.4.22.-2.

Composition of the fatty acid fraction of the substance:
— *caproic acid*: maximum 3.0 per cent;
— *caprylic acid*: 50.0 per cent to 90.0 per cent;
— *capric acid*: 10.0 per cent to 50.0 per cent;
— *lauric acid*: maximum 3.0 per cent;
— *myristic acid*: maximum 1.0 per cent.

Free glycerol
Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of *methylene chloride R*. Heat to about 50 °C and allow to cool. Add 100 mL of *water R*, shake and add 25.0 mL of *periodic acetic acid solution R*. Shake again and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of *potassium iodide R* and allow to stand for 1 min. Add 1 mL of *starch solution R*. Titrate with *0.1 M sodium thiosulfate* until the aqueous phase becomes colourless. Carry out a blank titration.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 2.3 mg of glycerol.

Water *(2.5.12)*
Maximum 0.5 per cent, determined on 1.00 g.

Total ash *(2.4.16)*
Maximum 0.5 per cent.

ASSAY
Gas chromatography *(2.2.28)*: use the normalisation procedure.

Test solution To 0.25 g of the substance to be examined, add 5.0 mL of *tetrahydrofuran R* and shake to dissolve.

Reference solution (a) To 0.25 g of *glycerol monocaprylocaprate CRS*, add 5.0 mL of *tetrahydrofuran R* and shake to dissolve.

Reference solution (b) To 50 mg of *glycerol 1-octanoate R* and 50 mg of *glycerol 1-decanoate R*, add 2.5 mL of *tetrahydrofuran R* and shake to dissolve.

Column:
— *size*: l = 10 m, Ø = 0.32 mm;
— *stationary phase*: *poly(dimethyl)(diphenyl)siloxane R* (film thickness 0.1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 2.3 mL/min.

Split ratio 1:50.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	60
	3 - 38	60 → 340
	38 - 50	340
Injection port		350
Detector		370

Detection Flame ionisation.

Injection 1 µL.

Identification of peaks Use the chromatogram supplied with *glycerol monocaprylocaprate CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to mono-, di- and triacylglycerols.

System suitability Reference solution (b):
— *resolution*: minimum 5 between the peaks due to glycerol 1-octanoate and glycerol 1-decanoate.

For the calculation of the contents of mono-, di- and triacylglycerols, disregard the peaks with a retention time less than that of the monoacylglycerols, which are due to the impurities of the solvent and to the free fatty acids.

Calculate the percentage content of free fatty acids *(C)* using the following equations:

$$\frac{I_A \times 144}{561.1}$$

I_A = acid value of the glycerol monocaprylocaprate.

Calculate the content of mono-, di- and triacylglycerols using the following equations:

$$\text{Monoacylglycerols content} = \frac{X \times (100 - A - B - C)}{X + Y + Z}$$

$$\text{Diacylglycerols content} = \frac{Y \times (100 - A - B - C)}{X + Y + Z}$$

$$\text{Triacylglycerols content} = \frac{Z \times (100 - A - B - C)}{X + Y + Z}$$

A = percentage content of free glycerol (see Tests);
B = percentage content of water;
X = area of the peak due to monoacylglycerols;
Y = area of the peak due to diacylglycerols;
Z = area of the peak due to triacylglycerols.

LABELLING
The labelling states the type of glycerol monocaprylocaprate (type I or II).

Ph Eur

Glycerol Monolinoleate

(Ph. Eur. monograph 1429)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of monoacylglycerols, mainly mono-oleoyl- and monolinoleoylglycerol, together with variable quantities of di- and triacylglycerols, obtained by partial glycerolysis of vegetable oils mainly containing triacylglycerols of linoleic ((9*Z*,12*Z*)-octadeca-9,12-dienoic) acid. A suitable antioxidant may be added.

Content
— *monoacylglycerols*: 32.0 per cent to 52.0 per cent;
— *diacylglycerols*: 40.0 per cent to 55.0 per cent;
— *triacylglycerols*: 5.0 per cent to 20.0 per cent.

CHARACTERS
Appearance
Amber, oily liquid which may be partially solidified at room temperature.

Solubility
Practically insoluble in water, freely soluble in methylene chloride.

IDENTIFICATION
A. Iodine value (see Tests).

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 20 mL with the same solvent.

Reference solution Dissolve 1.0 g of *glycerol monolinoleate CRS* in *methylene chloride R* and dilute to 20 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *hexane R, ether R* (30:70 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of *rhodamine B R* in *ethanol (96 per cent) R* and examine in ultraviolet light at 365 nm.

Results The spots in the chromatogram obtained with the test solution are similar in position to those in the chromatogram obtained with the reference solution.

C. Composition of fatty acids (see Tests).

TESTS
Acid value (*2.5.1*)
Maximum 6.0, determined on 1.0 g.

Iodine value (*2.5.4, Method A*)
100 to 140.

Peroxide value (*2.5.5, Method A*)
Maximum 12.0, determined on 2.0 g.

Saponification value (*2.5.6*)
160 to 180, determined on 2.0 g.

Free glycerol
Maximum 6.0 per cent, determined as described in the assay.

Composition of fatty acids (*2.4.22, Method C*)
Composition of the fatty acid fraction of the substance:
— *palmitic acid*: 4.0 per cent to 20.0 per cent;
— *stearic acid*: maximum 6.0 per cent;
— *oleic acid*: 10.0 per cent to 35.0 per cent;
— *linoleic acid*: minimum 50.0 per cent;
— *linolenic acid*: maximum 2.0 per cent;
— *arachidic acid*: maximum 1.0 per cent;
— *eicosenoic acid*: maximum 1.0 per cent.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.00 g. Use as the solvent a mixture of equal volumes of *anhydrous methanol R* and *methylene chloride R*.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY
Size-exclusion chromatography (*2.2.30*).

Test solution Into a 15 mL flask, weigh about 0.2 g (*m*), to the nearest 0.1 mg. Add 5 mL of *tetrahydrofuran R* and shake to dissolve. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into four 15 mL flasks, respectively weigh, to the nearest 0.1 mg, about 2.5 mg, 5 mg, 10 mg and 20 mg of *glycerol R*. Add 5 mL of *tetrahydrofuran R* and shake until well mixed. Weigh the flasks again and calculate the concentration of glycerol in milligrams per gram for each reference solution.

Column:
— *size*: *l* = 0.6 m, Ø = 7 mm,
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R.*

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to glycerol (retention time = about 15.6 min): triacylglycerols = about 0.76; diacylglycerols = about 0.80; monoacylglycerols = about 0.86.

Calculations:
— *free glycerol*: from the calibration curve obtained with the reference solutions, determine the concentration of glycerol (*C*) in milligrams per gram in the test solution and calculate the percentage content of free glycerol (*A*) in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *free fatty acids*: calculate the percentage content of free fatty acids (*D*) using the following expression:

$$\frac{I_A \times 280}{56.11 \times 10}$$

I_A = acid value (see Tests);
280 = rounded molar mass of linoleic acid, in grams per mole;
56.11 = molar mass of potassium hydroxide, in grams per mole.

— *monoacylglycerols*: calculate the percentage content of monoacylglycerols using the following expression:

$$\left[\frac{X}{X+Y+Z}\ (100 - A - B)\right] - D$$

A = percentage content of free glycerol;

B = percentage content of water (see Tests);

D = percentage content of free fatty acids;

X = area of the peak due to monoacylglycerols and free fatty acids;

Y = area of the peak due to diacylglycerols;

Z = area of the peak due to triacylglycerols.

— *diacylglycerols*: calculate the percentage content of diacylglycerols using the following expression:

$$\frac{Y}{X+Y+Z}\ (100 - A - B)$$

— *triacylglycerols*: calculate the percentage content of triacylglycerols using the following expression:

$$\frac{Z}{X+Y+Z}\ (100 - A - B)$$

STORAGE

In an airtight container, protected from light.

Ph Eur

Glycerol Mono-oleate

(Ph. Eur. monograph 1430

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of monoacylglycerols, mainly mono-oleoylglycerol, together with variable quantities of di- and triacylglycerols. It is defined by the nominal content of monoacylglycerols and obtained by partial glycerolysis of vegetable oils mainly containing triacylglycerols of oleic ((9Z)-octadec-9-enoic) acid or by esterification of glycerol by oleic acid, this fatty acid being of vegetable or animal origin. A suitable antioxidant may be added.

Content

	Nominal content of acylglycerol (per cent)		
	40	60	90
Monoacylglycerols	32.0 - 52.0	55.0 - 65.0	90.0 - 101.0
Diacylglycerols	30.0 - 50.0	15.0 - 35.0	< 10.0
Triacylglycerols	5.0 - 20.0	2.0 - 10.0	< 2.0

CHARACTERS

Appearance

Amber, oily liquid which may be partially solidified at room temperature.

Solubility

Practically insoluble in water, freely soluble in methylene chloride.

IDENTIFICATION

A. Iodine value (see Tests).

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *methylene chloride R* and dilute to 20 mL with the same solvent.

Reference solution Dissolve 1.0 g of *glycerol mono-oleate CRS* in *methylene chloride R* and dilute to 20 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *hexane R, ether R* (30:70 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of *rhodamine B R* in *ethanol (96 per cent) R* and examine in ultraviolet light at 365 nm.

Results The spots in the chromatogram obtained with the test solution are similar in position to those in the chromatogram obtained with the reference solution.

C. It complies with the limits of the assay (monoacylglycerol content).

TESTS

Acid value (*2.5.1*)
Maximum 6.0, determined on 1.0 g.

Iodine value (*2.5.4, Method A*)
65.0 to 95.0.

Peroxide value (*2.5.5, Method A*)
Maximum 12.0, determined on 2.0 g.

Saponification value (*2.5.6*)
150 to 175, determined on 2.0 g.

Free glycerol
Maximum 6.0 per cent, determined as described in the assay.

Composition of fatty acids (*2.4.22, Method C*)
Composition of the fatty acid fraction of the substance:
— *palmitic acid*: maximum 12.0 per cent;
— *stearic acid*: maximum 6.0 per cent;
— *oleic acid*: minimum 60.0 per cent;
— *linoleic acid*: maximum 35.0 per cent;
— *linolenic acid*: maximum 2.0 per cent;
— *arachidic acid*: maximum 2.0 per cent;
— *eicosenoic acid*: maximum 2.0 per cent.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.00 g. Use as the solvent a mixture of equal volumes of *anhydrous methanol R* and *methylene chloride R*.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY
Size-exclusion chromatography (*2.2.30*).

Test solution Into a 15 mL flask, weigh about 0.2 g (*m*), to the nearest 0.1 mg. Add 5 mL of *tetrahydrofuran R* and shake to dissolve. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into four 15 mL flasks, respectively weigh, to the nearest 0.1 mg, about 2.5 mg, 5 mg, 10 mg and 20 mg of *glycerol R*. Add 5 mL of *tetrahydrofuran R* and shake until well mixed. Weigh the flasks again and calculate the concentration of glycerol in milligrams per gram for each reference solution.

Column:
— *size*: *l* = 0.6 m, Ø = 7 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R*.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to glycerol (retention time = about 15.6 min): triacylglycerols = about 0.76; diacylglycerols = about 0.79; monoacylglycerols = about 0.85.

Calculations:
— *free glycerol*: from the calibration curve obtained with the reference solutions determine the concentration of glycerol (*C*) in milligrams per gram in the test solution and calculate the percentage content of free glycerol (*A*) in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *free fatty acids*: calculate the percentage content of free fatty acids (*D*) using the following expression:

$$\frac{I_A \times 282}{56.11 \times 10}$$

I_A = acid value (see Tests);
282 = rounded molar mass of oleic acid, in grams per mole;
56.11 = molar mass of potassium hydroxide, in grams per mole.

— *monoacylglycerols*: calculate the percentage content of monoacylglycerols using the following expression:

$$\left[\frac{X}{X+Y+Z}\ (100 - A - B)\right] - D$$

A = percentage content of free glycerol;

B = percentage content of water (see Tests);

D = percentage content of free fatty acids;

X = area of the peak due to monoacylglycerols and free fatty acids;

Y = area of the peak due to diacylglycerols;

Z = area of the peak due to triacylglycerols.

— *diacylglycerols*: calculate the percentage content of diacylglycerols using the following expression:

$$\frac{Y}{X+Y+Z}\ (100 - A - B)$$

— *triacylglycerols*: calculate the percentage content of triacylglycerols using the following expression:

$$\frac{Z}{X+Y+Z}\ (100 - A - B)$$

STORAGE
In an airtight container, protected from light.

LABELLING
The label states the nominal content of monoacylglycerol.

Ph Eur

Glycerol Monostearate 40-55

(Ph. Eur. monograph 0495)

Action and use
Excipient.

Ph Eur

DEFINITION
Mixture of monoacylglycerols, mainly monostearoylglycerol, together with variable quantities of di- and triacylglycerols. It is obtained by partial glycerolysis of vegetable oils mainly containing triacylglycerols of palmitic (hexadecanoic) or stearic (octadecanoic) acid or by esterification of glycerol with stearic acid. The fatty acids may be of vegetable or animal origin.

Content
— *monoacylglycerols*: 40.0 per cent to 55.0 per cent;
— *diacylglycerols*: 30.0 per cent to 45.0 per cent;
— *triacylglycerols*: 5.0 per cent to 15.0 per cent.

CHARACTERS
Appearance
Hard, waxy mass or unctuous powder or flakes, white or almost white.

Solubility
Practically insoluble in water, soluble in ethanol (96 per cent) at 60 °C.

IDENTIFICATION
First identification: C, D.

Second identification: A, B.

A. Melting point (*2.2.15*): 54 °C to 66 °C.

Introduce the melted substance into the capillary tubes and allow to stand for 24 h in a well-closed container.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.5 g of the substance to be examined in *methylene chloride R*, with gentle heating, and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.5 g of *glycerol monostearate 40-55 CRS* in *methylene chloride R*, with gentle heating, and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *hexane R, ether R* (30:70 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Detection Spray with a 0.1 g/L solution of *rhodamine B R* in *ethanol (96 per cent) R* and examine in ultraviolet light at 365 nm.

Suitability system Reference solution:
— the chromatogram shows 4 clearly separated spots.

Results The spots in the chromatogram obtained with the test solution are similar in position to those in the chromatogram obtained with the reference solution.

C. Composition of fatty acids (see Tests) according to the type stated on the label.

D. It complies with the limits of the assay (monoacylglycerol content).

TESTS

Acid value (*2.5.1*)
Maximum 3.0, determined on 1.0 g.

Use a mixture of equal volumes of *ethanol (96 per cent) R* and *toluene R* as solvent and heat gently.

Iodine value (*2.5.4, Method A*)
Maximum 3.0.

Saponification value (*2.5.6*)
158 to 177, determined on 2.0 g. Carry out the titration with heating.

Free glycerol
Maximum 6.0 per cent, determined as described under Assay.

Composition of fatty acids (*2.4.22, Method C*)
Use the mixture of calibrating substances in Table 2.4.22.-1.

Composition of the fatty-acid fraction of the substance:

Glycerol monostearate 40-55	Composition of fatty acids
Type I	*Stearic acid*: 40.0 per cent to 60.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 90.0 per cent
Type II	*Stearic acid*: 60.0 per cent to 80.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 90.0 per cent
Type III	*Stearic acid*: 80.0 per cent to 99.0 per cent *Sum of the contents of palmitic and stearic acids*: minimum 96.0 per cent

Nickel (*2.4.31*)
Maximum 1 ppm.

Water (*2.5.12*)
Maximum 1.0 per cent, determined on 1.00 g.
Use *pyridine R* as the solvent and heat gently.

Total ash (*2.4.16*)
Maximum 0.1 per cent.

ASSAY

Size-exclusion chromatography (*2.2.30*).

Test solution Into a 15 mL flask, weigh 0.200 g (*m*). Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Reweigh the flask and calculate the total mass of solvent and substance (*M*).

Reference solutions Into four 15 mL flasks, respectively weigh 2.5 mg, 5.0 mg, 10.0 mg and 20.0 mg of *glycerol R*, and add 5.0 mL of *tetrahydrofuran R* to each flask. Weigh the flasks again and calculate the concentration of glycerol in milligrams per gram for each reference solution.

Column:
— *size*: *l* = 0.6 m, Ø = 7 mm;
— *stationary phase*: *styrene-divinylbenzene copolymer R* (5 µm) with a pore size of 10 nm.

Mobile phase *tetrahydrofuran R.*

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to glycerol (retention time = about 15 min): triacylglycerols = about 0.75; diacylglycerols = about 0.80; monoacylglycerols = about 0.85.

Calculations:
— *free glycerol*: from the calibration curve obtained with the reference solutions, determine the concentration of glycerol (*C*) in milligrams per gram in the test solution and calculate the percentage content of free glycerol (*A*) in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *free fatty acids*: calculate the percentage content of free fatty acids (*D*) using the following expression:

$$\frac{I_A \times 270}{56.11 \times 10}$$

I_A = acid value (see Tests);
270 = average rounded molar mass of stearic acid and palmitic acid, in grams per mole;
56.11 = molar mass of potassium hydroxide, in grams per mole.

— *monoacylglycerols*: calculate the percentage content of monoacylglycerols using the following expression:

$$\left[\frac{X}{X+Y+Z}\ (100 - A - B)\right] - D$$

A = percentage content of free glycerol;

B = percentage content of water (see Tests);

D = percentage content of free fatty acids;

X = area of the peak due to monoacylglycerols and free fatty acids;

Y = area of the peak due to diacylglycerols;

Z = area of the peak due to triacylglycerols.

— *diacylglycerols*: calculate the percentage content of diacylglycerols using the following expression:

$$\frac{Y}{X+Y+Z}\ (100 - A - B)$$

— *triacylglycerols*: calculate the percentage content of triacylglycerols using the following expression:

$$\frac{Z}{X+Y+Z}\ (100 - A - B)$$

LABELLING
The label states the type of glycerol monostearate 40-55.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for glycerol monostearate 40-55 used as matrix former in prolonged-release oral solid dosage forms.

Composition of fatty acids
(see Tests).

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Thermal analysis (*2.2.34*)
The melting behaviour of the substance as is and after melting followed by solidification when cooling may be considered.

The following characteristics may be relevant for glycerol monostearate 40-55 used as consistency agent in dosage forms for cutaneous application.

Composition of fatty acids
(see Tests).

Thermal analysis (*2.2.34*)
The melting behaviour of the substance as is and after melting followed by solidification when cooling may be considered.

Ph Eur

Self-emulsifying Glyceryl Monostearate
Self-emulsifying Monostearin; Self-emulsifying Mono-and Diglycerides of Food Fatty Acids

Action and use
Excipient.

DEFINITION
Self-emulsifying Glyceryl Monostearate is a mixture consisting principally of mono-, di- and triglycerides of stearic and palmitic acids and of minor proportions of glycerides of other fatty acids; it may also contain free glycerol, free fatty acids and soap. It contains not less than 30.0% of monoglycerides, calculated as $C_{21}H_{42}O_4$, not more than 7.0% of free glycerol, calculated as $C_3H_8O_3$, and not more than 6.0% of soap, calculated as sodium oleate, $C_{18}H_{33}NaO_2$, all calculated with reference to the anhydrous substance.

CHARACTERISTICS
A white to cream coloured, hard, waxy solid.

Dispersible in hot *water*; soluble in hot *absolute ethanol*, in hot *liquid paraffin* and, subject to turbidity at concentrations below 20%, in hot vegetable oils.

TESTS
Acid value
Not more than 6, Appendix X B.

Iodine value
Not more than 3 *(iodine monochloride method)*, Appendix X E.

Alkalinity
Shake 1 g with 20 mL of hot *carbon dioxide-free water* and allow to cool with continuous shaking. The pH of the aqueous layer is 8.0 to 10.0, Appendix V L.

Water
Not more than 2.0% w/w, Appendix IX C. Use 0.5 g and a mixture of 10 mL of *anhydrous methanol* and 10 mL of anhydrous *chloroform* as the solvent.

ASSAY
For free glycerol
Dissolve 0.4 g in 50 mL of *dichloromethane* in a ground-glass-stoppered separating funnel, cool if necessary, add 25 mL of *water* and shake vigorously for 1 minute; add 0.2 mL of *glacial acetic acid*, if necessary, to break the emulsion. Repeat the extraction a further three times using 25-, 20- and 20- mL quantities of *water* and reserve the dichloromethane solution for the Assay for monoglycerides. Filter the combined aqueous extracts through a filter paper moistened with *water*, wash the filter with two 5 mL quantities of *water* and dilute the combined filtrate and washings to 100 mL with *water*. To 50 mL of this solution add 25 mL of *periodic acetic acid solution*, shaking cautiously, allow to stand at 25° to 30° for 30 minutes and add 100 mL of *water* and 12 mL of *potassium iodide solution*. Titrate with 0.1M *sodium thiosulfate VS* using 1 mL of *starch solution* as indicator. Repeat the determination using 50 mL of *water* in place of the 50 mL of the solution being examined. The difference between the titrations represents the amount of sodium thiosulfate required. Each mL of 0.1M *sodium thiosulfate VS* is equivalent to 2.3 mg of glycerol.

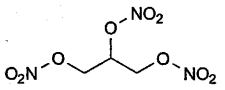
For monoglycerides
Filter the reserved dichloromethane solution obtained in the Assay for free glycerol through absorbent cotton and wash the separating funnel and the filter with three 5 mL quantities of *dichloromethane*. Dilute the combined filtrate and washings to 100 mL with *dichloromethane* and to 50 mL of the solution add 25 mL of *periodic acetic acid solution*, shaking cautiously. Allow to stand at 25° to 30° for 30 minutes and add 100 mL of *water* and 12 mL of *potassium iodide solution*. Titrate the liberated iodine with 0.1M *sodium thiosulfate VS* using 1 mL of *starch solution* as indicator. Repeat the determination using 50 mL of *dichloromethane* in place of the 50 mL of the solution of the substance being examined. The difference between the titrations represents the amount of sodium thiosulfate required. Each mL of 0.1M *sodium thiosulfate VS* is equivalent to 17.9 mg of 1-monoacylglycerols, calculated as $C_{21}H_{42}O_4$. The quantity of 0.1M *sodium thiosulfate VS* used in the assay is not less than 85% of the quantity of sodium thiosulfate used in the blank assay.

For soap
Add 10 g to a mixture of 60 mL of *acetone* and 0.15 mL of a 0.5% w/v solution of *bromophenol blue* in a mixture of 20 mL of *ethanol (20%)* and 80 mL of *water*, the solvent having been previously neutralised with *0.1M hydrochloric acid VS* or *0.1M sodium hydroxide VS*. Warm gently on a water bath until

solution is complete and titrate with *0.1M hydrochloric acid VS* until the blue colour is discharged. Allow to stand for 20 minutes, warm until any solidified matter has redissolved and, if the blue colour reappears, continue the titration. Each mL of *0.1M hydrochloric acid VS* is equivalent to 30.45 mg of $C_{18}H_{33}NaO_2$.

Glyceryl Trinitrate Solution

(Ph. Eur. monograph 1331)



$C_3H_5N_3O_9$ 227.1

Action and use
Vasodilator.

Preparations
Glyceryl Trinitrate Ointment
Glyceryl Trinitrate Sublingual Spray
Glyceryl Trinitrate Tablets
Glyceryl Trinitrate Transdermal Patches

When concentrated glyceryl trinitrate solution is demanded, the intention of the purchaser, with respect to the strength expressed, should be ascertained.

Ph Eur

DEFINITION
Ethanolic solution of glyceryl trinitrate containing 1 per cent *m/m* to 10 per cent *m/m* of propane-1,2,3-triyl trinitrate.

Content
96.5 per cent to 102.5 per cent of the declared content of glyceryl trinitrate stated on the label.

CHARACTERS

Appearance
Clear, colourless or slightly yellow solution.

Solubility
Miscible with acetone and with anhydrous ethanol.

Solubility of pure glyceryl trinitrate: practically insoluble in water, freely soluble in anhydrous ethanol, miscible with acetone.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

Upon diluting glyceryl trinitrate solution, care must be taken to always use anhydrous ethanol, otherwise droplets of pure glyceryl trinitrate may precipitate from the solution.

After examination, the residues and the solutions obtained in both the identification and the test sections must be heated on a water-bath for 5 min with dilute sodium hydroxide solution R.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Place 50 µL of a solution diluted, if necessary, with *anhydrous ethanol R*, to contain 10 g/L of glyceryl trinitrate, on a disc of *potassium bromide R* and evaporate the solvent *in vacuo*.

Comparison *Ph. Eur. reference spectrum of glyceryl trinitrate.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dilute a quantity of the substance to be examined corresponding to 50 mg of glyceryl trinitrate to 100 mL with *acetone R*.

Reference solution Dilute 0.05 mL of *glyceryl trinitrate solution CRS* to 1 mL with *acetone R*.

Plate *TLC silica gel plate R.*

Mobile phase *ethyl acetate R, toluene R* (20:80 *V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with freshly prepared *potassium iodide and starch solution R*; expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It complies with the limits of the assay.

TESTS

Upon diluting glyceryl trinitrate solution, care must be taken to always use anhydrous ethanol, otherwise droplets of pure glyceryl trinitrate may precipitate from the solution.

After examination, the residues and the solutions obtained in both the identification and the test sections must be heated on a water-bath for 5 min with dilute sodium hydroxide solution R.

Appearance of solution
If necessary, dilute the solution to be examined to a concentration of 10 g/L with *anhydrous ethanol R*.
The solution is not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Impurity A (inorganic nitrates)
Thin-layer chromatography (*2.2.27*).

Test solution If necessary, dilute the solution to be examined to a concentration of 10 g/L with *anhydrous ethanol R*.

Reference solution Dissolve 5 mg of *potassium nitrate R* in 1 mL of *water R* and dilute to 100 mL with *ethanol (96 per cent) R*.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, acetone R, toluene R* (15:30:60 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of air until the acetic acid is completely removed.

Detection Spray intensively with freshly prepared *potassium iodide and starch solution R*; expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Limit:
— *nitrate ion*: any spot due to the nitrate ion in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent of the content of glyceryl trinitrate calculated as potassium nitrate).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dilute a quantity of the substance to be examined equivalent to 5.0 mg of glyceryl trinitrate to 10.0 mL with *water for chromatography R*. *NOTE*: for glyceryl trinitrate solutions of more than 1 per cent *m/m*, dilute first to a 1 per cent solution with *anhydrous ethanol R*.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water for chromatography R*.

Reference solution (b) Dilute 25 µL of *glyceryl trinitrate for system suitability CRS* (containing impurities B, C, D and E) to 0.5 mL with *water for chromatography R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: *water for chromatography R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	90	10
2 - 10	90 → 50	10 → 50
10 - 20	50	50

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *glyceryl trinitrate for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D and E.

Relative retention With reference to glyceryl trinitrate (retention time = about 15.5 min): impurity C = about 0.19; impurity B = about 0.21; impurity E = about 0.63; impurity D = about 0.65.

System suitability Reference solution (b):
— *resolution*: minimum 1.5 between the peaks due to impurities C and B; minimum 1.5 between the peaks due to impurities E and D.

Calculation of percentage contents:
— for each impurity, use the concentration of glyceryl trinitrate in reference solution (a).

Limits:
— *impurities B, C, D, E*: for each impurity, maximum 0.5 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

ASSAY

Test solution Prepare a solution containing 1.0 mg of glyceryl trinitrate in 250.0 mL of *methanol R*.

Reference solution Dissolve 70.0 mg of *sodium nitrite R* in *methanol R* and dilute to 250.0 mL with the same solvent. Dilute 5.0 mL of the solution to 500.0 mL with *methanol R*.

Into three 50 mL volumetric flasks introduce 10.0 mL of the test solution, 10.0 mL of the reference solution and 10 mL of *methanol R* as a blank. To each flask add 5 mL of *dilute sodium hydroxide solution R*, close the flask, mix and allow to stand at room temperature for 30 min. Add 10 mL of *sulfanilic acid solution R* and 10 mL of *dilute hydrochloric acid R* and mix. After exactly 4 min, add 10 mL of *naphthylethylenediamine dihydrochloride solution R*, dilute to volume with *water R* and mix. After 10 min read the absorbance (*2.2.25*) of the test solution and the reference solution at 540 nm using the blank solution as the compensation liquid.

Calculate the percentage content of glyceryl trinitrate using the following expression:

$$\frac{A_T \times m_S \times C}{A_R \times m_T \times 60.8}$$

A_T = absorption of the test solution;
m_S = mass of sodium nitrite, in milligrams.
C = percentage content of sodium nitrite used as reference;
A_R = absorption of the reference solution;
m_T = mass of the substance to be examined, in milligrams;

STORAGE

Store the diluted solutions (1 per cent *m/m*) protected from light, at a temperature of 2 °C to 15 °C.

Store the other solutions protected from light, at a temperature of 15 °C to 20 °C.

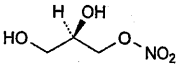
LABELLING

The label states the declared content of glyceryl trinitrate.

IMPURITIES

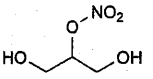
Specified impurities *A, B, C, D, E.*

A. inorganic nitrates,

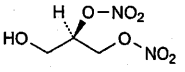


and enantiomer

B. (2*RS*)-2,3-dihydroxypropyl nitrate,

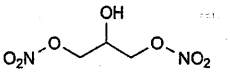


C. 2-hydroxy-1-(hydroxymethyl)ethyl nitrate,



and enantiomer

D. (2*RS*)-3-hydroxypropane-1,2-diyl dinitrate,



E. 2-hydroxypropane-1,3-diyl dinitrate.

Ph Eur

Glycine

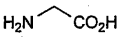
(Ph. Eur. monograph 0614)

$C_2H_5NO_2$ 75.1 *56-40-6*

Action and use
Amino acid used for bladder irrigation during surgery.

Preparation
Glycine Irrigation Solution

Ph Eur



DEFINITION

2-Aminoacetic acid.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glycine CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethanol (60 per cent V/V) R*, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 10 mg of *glycine CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Plate *cellulose for chromatography R* as the coating substance.

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 80 °C for 30 min.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 50 mg in 5 mL of *water R*, add 1 mL of *strong sodium hypochlorite solution R* and boil for 2 min. Add 1 mL of *hydrochloric acid R* and boil for 4-5 min. Add 2 mL of *hydrochloric acid R* and 1 mL of a 20 g/L solution of *resorcinol R*, boil for 1 min and cool. Add 10 mL of *water R* and mix. To 5 mL of the solution add 6 mL of *dilute sodium hydroxide solution R*. The solution is violet with greenish-yellow fluorescence. After a few minutes, the colour becomes orange and then yellow and an intense fluorescence remains.

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)

5.9 to 6.4.

Dilute 10 mL of solution S to 20 mL with *carbon dioxide-free water R*.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture To 500 mL of the mobile phase add 1.5 mL of *phosphoric acid R*.

Test solution Dissolve 0.200 g of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 80 mg of *iminodiacetic acid R* (impurity A) and 80 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of *glycine anhydride R* (impurity B), 50.0 mg of *diglycine R* (impurity H) and 50.0 mg of *triglycine R* (impurity I) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 25 °C.

Mobile phase Dissolve 1.4 g of *sodium pentanesulfonate R* in 900 mL of *water R*, adjust to pH 2.2 with *phosphoric acid R* and dilute to 1 L with *water R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Run time 4 times the retention time of glycine.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, H and I.

Relative retention With reference to glycine (retention time = about 5.5 min): impurity A = about 0.7; impurity B = about 0.75; impurity H = about 1.7; impurity I = about 2.0.

System suitability Reference solution (b):
- *resolution*: minimum 5.0 between the peaks due to impurity A and glycine.

Calculation of percentage contents:
- for each impurity, use the concentration of glycine in reference solution (a);
- for impurities B, H and I, use the concentration of the corresponding impurity in reference solution (c).

Limits:
- *impurities B, H, I*: for each impurity, maximum 0.10 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.2 per cent;
- *reporting threshold*: 0.05 per cent.

Ninhydrin-positive substances

Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *water R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions (a), (b) and (d) into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of glycine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *any ninhydrin-positive substance*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold*: 0.05 per cent.

Chlorides (*2.4.4*)
Maximum 75 ppm.

Dissolve 0.67 g in *water R* and dilute to 15 mL with the same solvent.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

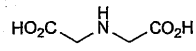
ASSAY
Dissolve 70.0 mg in 5 mL of *anhydrous formic acid R*. Add 50 mL of *anhydrous acetic acid R*. Immediately after dissolution, titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 7.51 mg of $C_2H_5NO_2$.

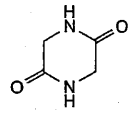
IMPURITIES

Specified impurities *B, H, I.*

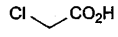
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, D, E, F, G.*



A. 2,2′-iminodiacetic acid,

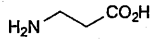


B. piperazine-2,5-dione (glycine anhydride),

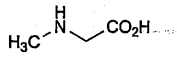


C. 2-chloroacetic acid,

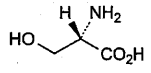
D. 1,3,5,7-tetraazatricyclo[3.3.1.1^{3,7}]decane,



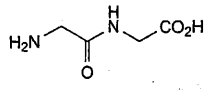
E. 3-aminopropanoic acid (β-alanine),



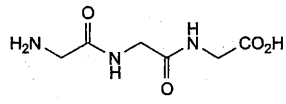
F. 2-(methylamino)acetic acid (sarcosine),



G. (2*S*)-2-amino-3-hydroxypropanoic acid (serine),



H. 2-[(2-aminoacetyl)amino]acetic acid (diglycine),

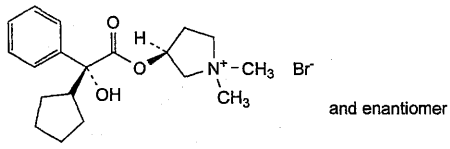


I. 2-[[2-[(2-aminoacetyl)amino]acetyl]amino]acetic acid (triglycine).

Ph Eur

Glycopyrronium Bromide

(Ph. Eur. monograph 1783)



$C_{19}H_{28}BrNO_3$ 398.3 *51186-83-5*

Action and use
Anticholinergic.

Preparation
Glycopyrronium Bromide Oral Solution

Ph Eur

DEFINITION

(3*RS*)-3-[(2*SR*)-(2-Cyclopentyl-2-hydroxy-2-phenylacetyl) oxy]-1,1-dimethylpyrrolidinium bromide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *glycopyrronium bromide CRS.*

B. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.05 mL of *phenolphthalein solution R1*. The solution is colourless. Not more than 0.2 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink. Add 0.4 mL of *0.01 M hydrochloric acid* and 0.05 mL of *methyl red solution R*. The solution is red or orange.

Impurity N
Liquid chromatography (*2.2.29*).

Solution A Dissolve 3.2 g of *sodium dihydrogen phosphate monohydrate R* in 900 mL of *water R*, adjust to pH 6.5 with *dilute sodium hydroxide solution R* and dilute to 1000 mL with *water R*.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of *glycopyrronium impurity N CRS* in 10.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution and 5.0 mL of reference solution (a) to 25.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.0 mm;
— *stationary phase*: *silica gel BC for chiral chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase *acetonitrile R1*, solution A, *methanol R2* (10:40:50 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 222 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 1.5 times the retention time of glycopyrronium.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity N.

Relative retention With reference to glycopyrronium (retention time = about 30 min): impurity N = about 0.9.

System suitability:
— *resolution*: minimum 1.25 between the peaks due to impurity N and glycopyrronium in the chromatogram obtained with reference solution (c);
— *signal-to-noise ratio*: minimum 5 for the peak due to impurity N in the chromatogram obtained with reference solution (b).

Limit:
— *impurity N*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.2 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *glycopyrronium for peak identification CRS* (containing impurities E and I) in 5.0 mL of mobile phase A.

Reference solution (c) Dissolve 10 mg of *benzaldehyde R* (impurity F) in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution and 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: dissolve 0.25 g of *sodium heptanesulfonate R* in 615 mL of a 1.63 g/L solution of *anhydrous sodium sulfate R*; add 3 mL of a 5.15 g/L solution of *sulfuric acid R*, 150 mL of *methanol R2* and 235 mL of *acetonitrile R1*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	100	0
20 - 30	100 → 50	0 → 50
30 - 45	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *glycopyrronium for peak identification CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities E and I; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to glycopyrronium (retention time = about 11 min): impurity E = about 0.7; impurity F = about 0.8; impurity I = about 2.3.

System suitability Reference solution (c):
— *resolution*: minimum 5.0 between the peaks due to impurity F and glycopyrronium.

Limits:
— *impurity I*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurity E*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to the bromide ion appearing close to the peak due to the solvent.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

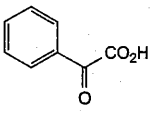
Dissolve 0.300 g in a mixture of 10 mL of *anhydrous acetic acid R* and 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 39.83 mg of $C_{19}H_{28}BrNO_3$.

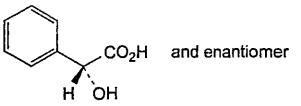
IMPURITIES

Specified impurities *E, I, N.*

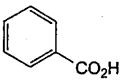
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, F, G, H, J, K, L, M, O.*



B. oxophenylacetic acid (benzoylformic acid),

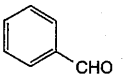


C. (2*RS*)-2-hydroxy-2-phenylacetic acid (mandelic acid),

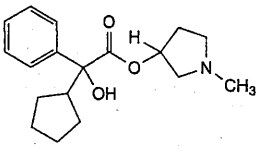


D. benzoic acid,

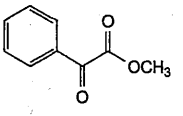
E. unknown structure,



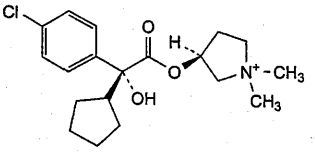
F. benzaldehyde,



G. 1-methylpyrrolidin-3-yl 2-cyclopentyl-2-hydroxy-2-phenylacetate,

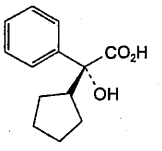


H. methyl 2-oxo-2-phenylacetate,



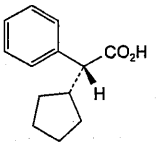
and enantiomer

I. (3*RS*)-3-[(2*SR*)-(2-(4-chlorophenyl)-2-cyclopentyl-2-hydroxyacetyl)oxy]-1,1-dimethylpyrrolidinium,



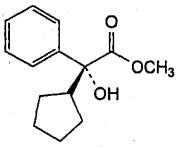
and enantiomer

J. (2*RS*)-2-cyclopentyl-2-hydroxy-2-phenylacetic acid,



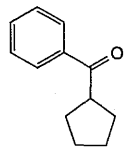
and enantiomer

K. (2*RS*)-2-cyclopentyl-2-phenylacetic acid,

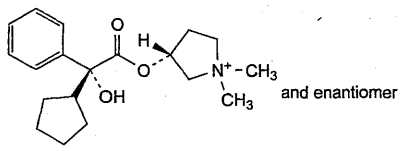


and enantiomer

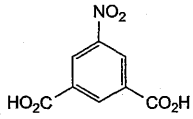
L. methyl (2*RS*)-2-cyclopentyl-2-hydroxy-2-phenylacetate,



M. cyclopentylphenylmethanone,



N. (3*RS*)-3-[(2*RS*)-(2-cyclopentyl-2-hydroxy-2-phenylacetyl) oxy]-1,1-dimethylpyrrolidinium,

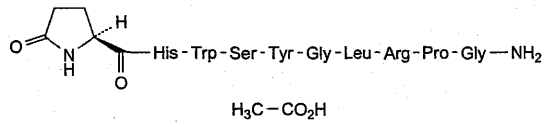


O. 5-nitroisophtalic acid.

Ph Eur

Gonadorelin Acetate

(Ph. Eur. monograph 0827)



$C_{57}H_{79}N_{17}O_{15}$ 1242 *499785-55-6*

Action and use
Gonadotropin-releasing hormone; treatment of prostate cancer.

Ph Eur

DEFINITION
5-Oxo-L-prolyl-L-histidyl-L-tryptophyl-L-seryl-L-tyrosylglycyl-L-leucyl-L-arginyl-L-prolylglycinamide acetate.

Acetate form of a synthetic hypothalamic peptide that stimulates the release of follicle-stimulating hormone and luteinising hormone from the pituitary gland.

Content
95.0 per cent to 102.0 per cent of the peptide $C_{55}H_{75}N_{17}O_{13}$ (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance
White or slightly yellowish powder, hygroscopic.

Solubility
Soluble in water, sparingly soluble in methanol. It dissolves in a 1 per cent *V/V* solution of glacial acetic acid.

IDENTIFICATION
Carry out either tests A, B or tests A, C.

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

B. Nuclear magnetic resonance spectrometry (*2.2.64*).

Preparation 4 mg/mL solution in a mixture of 20 volumes of *deuterated acetic acid R* and 80 volumes of *deuterium oxide R*.

Comparison Dissolve the contents of a vial of *gonadorelin for NMR identification CRS* in a mixture of 20 volumes of *deuterated acetic acid R* and 80 volumes of *deuterium oxide R* to obtain a concentration of 4 mg/mL.

Operating conditions:
— *field strength*: minimum 300 MHz;
— *temperature*: 27 °C.

Results Examine the ^1H NMR spectrum from 0 to 9 ppm. The ^1H NMR spectrum obtained is qualitatively similar to the ^1H NMR spectrum obtained with *gonadorelin for NMR identification CRS*.

C. Amino acid analysis (*2.2.56*). Method 1 for hydrolysis and method 1 for analysis are suitable.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids, taking 1/8 of the sum of the number of moles of histidine, glutamic acid, leucine, proline, glycine, tyrosine and arginine as equal to 1. The values fall within the following limits: serine and tyrosine 0.7 to 1.05; glutamic acid, proline, leucine, histidine and arginine 0.9 to 1.1; glycine 1.8 to 2.2. Lysine and isoleucine are absent, and not more than traces of other amino acids are present.

TESTS

Specific optical rotation (*2.2.7*)
−65 to −58 (anhydrous and acetic acid-free substance).

Dissolve 10.0 mg in 1.0 mL of a 1 per cent *V/V* solution of *glacial acetic acid R*.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of 1.0 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *gonadorelin CRS* in *water R* to obtain a concentration of 1.0 mg/mL.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*.

Reference solution (c) Dissolve the contents of a vial of *gonadorelin for system suitability CRS* (containing impurities C, D, E, F and G) in 1.0 mL of *water R*.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 13 volumes of *acetonitrile R1* and 87 volumes of a 1.18 per cent *V/V* solution of *phosphoric acid R* previously adjusted to pH 2.3 with *triethylamine R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of gonadorelin.

Relative retention With reference to gonadorelin (retention time = about 12-16 min): impurities C and D = about 0.7; impurity E = about 0.8; impurities F and G = about 1.2.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurities C + D, and impurity E.

Limits:
— *impurity E*: maximum 2.0 per cent;

— *sum of impurities F and G*: maximum 1.5 per cent;
— *sum of impurities C and D*: maximum 1.0 per cent;
— *unspecified impurities*: for each impurity, maximum 0.5 per cent;
— *total*: maximum 5.0 per cent;
— *reporting threshold*: 0.1 per cent.

Acetic acid (*2.5.34*)
4.0 per cent to 7.5 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of solvents.

Water (*2.5.32*)
Maximum 7.0 per cent, determined on 20.0 mg.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

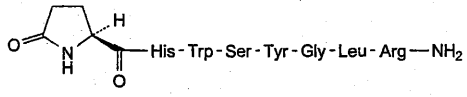
Injection Test solution and reference solution (a).

Calculate the percentage content of gonadorelin ($C_{55}H_{75}N_{17}O_{13}$) taking into account the assigned content of $C_{55}H_{75}N_{17}O_{13}$ in *gonadorelin CRS*.

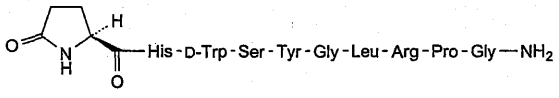
STORAGE
In an airtight container, protected from light at a temperature of 2 °C to 8 °C. If the substance is sterile, the container is also sterile and tamper-proof.

LABELLING
The label states:
— the gonadorelin peptide content;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

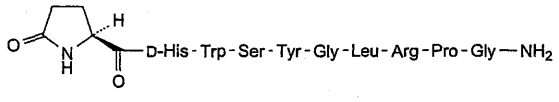
IMPURITIES
Specified impurities *C, D, E, F, G.*



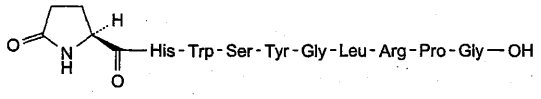
C. des-9-L-proline,10-L-glycine-gonadorelin,



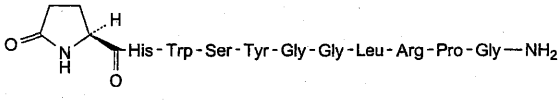
D. [3-D-tryptophan]gonadorelin,



E. [2-D-histidine]gonadorelin,



F. [10-glycine]gonadorelin,

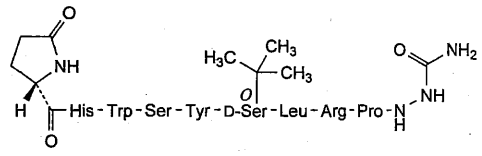


G. endo-5a-glycine-gonadorelin.

Ph Eur

Goserelin

(Ph. Eur. monograph 1636)



$C_{59}H_{84}N_{18}O_{14}$ 1269 *65807-02-5*

Action and use
Gonadotropin-releasing hormone, gonadorelin analogue; treatment of prostate cancer.

Preparation
Goserelin Implants

Ph Eur

DEFINITION
1-Carbamoyl-2-[5-oxo-L-prolyl-L-histidyl-L-tryptophyl-L-seryl-L-tyrosyl-*O*-(1,1-dimethylethyl)-D-seryl-L-leucyl-L-arginyl-L-prolyl]hydrazine.

Synthetic nonapeptide analogue of the hypothalamic decapeptide gonadorelin. It is obtained by chemical synthesis and is available as an acetate.

Content
94.5 per cent to 103.0 per cent of the peptide $C_{59}H_{84}N_{18}O_{14}$ (anhydrous and acetic acid-free substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Soluble in water, freely soluble in glacial acetic acid. It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION
Carry out either tests A and B or tests B and C.

A. Nuclear magnetic resonance spectrometry (*2.2.64*).

Preparation 13 mg/mL solution in *0.2 M deuterated sodium phosphate buffer solution pH 5.0 R* containing 20 µg/mL of *deuterated sodium trimethylsilylpropionate R*.

Comparison 13 mg/mL solution of *goserelin for NMR identification CRS* in *0.2 M deuterated sodium phosphate buffer solution pH 5.0 R* containing 20 µg/mL of *deuterated sodium trimethylsilylpropionate R* (dissolve the contents of a vial of *goserelin for NMR identification CRS* in this solvent to obtain the desired concentration).

Operating conditions:
— *field strength*: minimum 300 MHz;
— *temperature*: 25 °C.

Results Examine the ^{1}H NMR spectrum from 0 ppm to 9 ppm; the ^{1}H NMR spectrum obtained is qualitatively similar to the ^{1}H NMR spectrum obtained with *goserelin for NMR identification CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

C. Amino acid analysis (*2.2.56*). Method 1 for hydrolysis and method 1 for analysis are suitable.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids taking 1/6 of the sum of the number of moles of glutamic acid, histidine, tyrosine, leucine, arginine, proline as equal to 1. The values fall within the following limits: glutamic acid, histidine, tyrosine, leucine, arginine and proline 0.9 to 1.1; serine 1.6 to 2.2. Not more than traces of other amino acids are present, with the exception of tryptophan.

TESTS

Specific optical rotation (*2.2.7*)

−56 to −52 (anhydrous and acetic acid-free substance).

Dissolve the substance to be examined in *water R* to obtain a concentration of 2 mg/mL.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of 1.0 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *goserelin CRS* in *water R* to obtain a concentration of 1.0 mg/mL.

Reference solution (b) Dilute 1.0 mL of the test solution to 100 mL with *water R*.

Reference solution (c) Dilute 1.0 mL of the test solution to 10.0 mL with *water R*.

Resolution solution (a) Dissolve the contents of a vial of *4-D-Ser-goserelin CRS* in *water R* to obtain a concentration of 0.1 mg/mL. Mix equal volumes of this solution and reference solution (c).

Resolution solution (b) Dissolve the contents of a vial of *goserelin validation mixture CRS* in 1.0 mL of *water R*.

Column:
- *size*: *l* = 0.15 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (3.5 µm) with a pore size of 12.5 nm;
- *temperature*: 50-55 °C.

Mobile phase *trifluoroacetic acid R, acetonitrile for chromatography R, water for chromatography R* (0.05:20:80 *V/V/V*).

Flow rate 0.7-1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution, reference solution (b) and resolution solutions (a) and (b).

Run time 90 min.

Relative retention With reference to goserelin: impurity A = about 0.67; impurity C = about 0.78; impurity B = about 0.79; impurity D = about 0.85; impurity E = about 0.89; impurity F = about 0.92; impurity G = about 0.94; impurity H = about 0.98; impurity I = about 1.43; impurity J = about 1.53; impurity K = about 1.67; impurity L = about 1.77.

System suitability:
- *retention time*: goserelin = 40 min to 50 min in the chromatogram obtained with resolution solution (b); adjust the flow rate of the mobile phase if necessary; if adjusting the flow rate does not result in a correct retention time of the principal peak, change the proportion of acetonitrile in the mobile phase to obtain the requested retention time for goserelin;
- *resolution*: minimum 7.0 between the peaks due to impurity A and goserelin in the chromatogram obtained with resolution solution (a);
- *symmetry factor*: 0.8 to 2.5 for the peaks due to impurity A and goserelin in the chromatogram obtained with resolution solution (a);
- the chromatogram obtained with resolution solution (b) is similar to the chromatogram supplied with *goserelin validation mixture CRS*; 2 peaks eluting prior to the principal peak and corresponding to impurities E and G are clearly visible.

Limits:
- *impurity E*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.5 per cent);
- *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Acetic acid (*2.5.34*)

4.5 per cent to 15.0 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Water (*2.5.32*)

Maximum 10.0 per cent.

Bacterial endotoxins (*2.6.14*)

Less than 16 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

Run time 60 min.

Calculate the content of goserelin ($C_{59}H_{84}N_{18}O_{14}$) taking into account the assigned content of $C_{59}H_{84}N_{18}O_{14}$ in *goserelin CRS*.

STORAGE

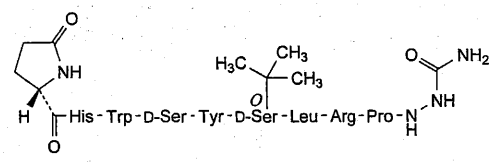
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

LABELLING

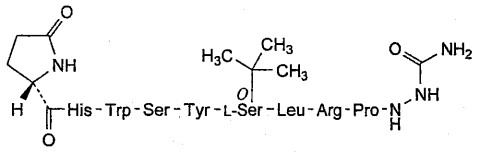
The label states:
- the mass of peptide in the container;
- where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES

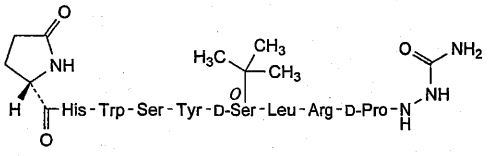
Specified impurities *A, B, C, D, E, F, G, H, I, J, K, L.*



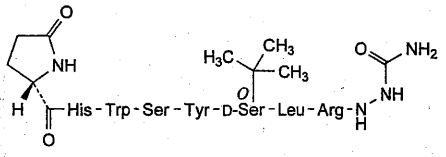
A. [4-D-serine]goserelin,



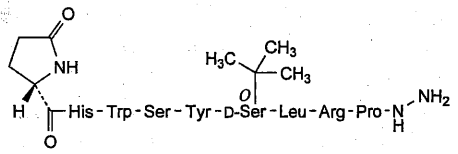
B. [6-[*O*-(1,1-dimethylethyl)-L-serine]]goserelin,



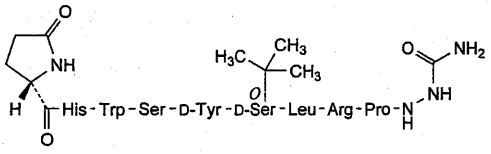
C. [9-D-proline]goserelin,



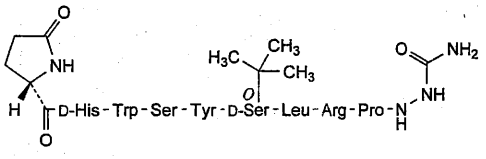
D. des-9-L-proline-goserelin,



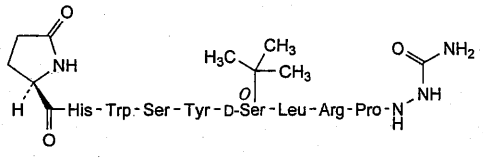
E. goserelin-(1-8)-peptidyl-L-prolinohydrazide,



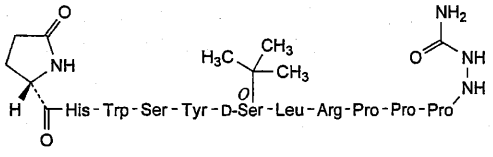
F. [5-D-tyrosine]goserelin,



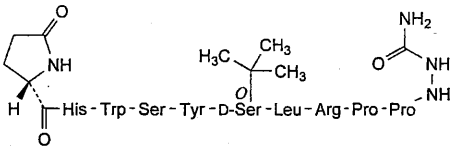
G. [2-D-histidine]goserelin,



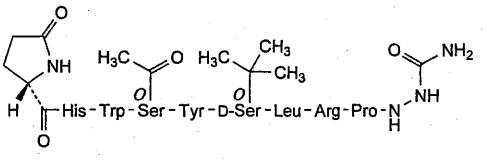
H. [1-(5-oxo-D-proline)]goserelin,



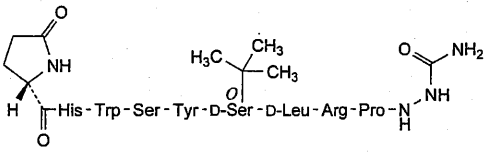
I. endo-8a,8b-di-L-proline-goserelin,



J. endo-8a-L-proline-goserelin,



K. [4-(*O*-acetyl-L-serine)]goserelin,

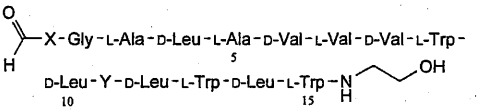


L. [7-D-leucine]goserelin.

Ph Eur

Gramicidin

(Ph. Eur. monograph 0907)



Gramicidin	X	Y	Mol. formula	M_r
A1	L-Val	L-Trp	$C_{99}H_{140}N_{20}O_{17}$	1882
A2	L-Ile	L-Trp	$C_{100}H_{142}N_{20}O_{17}$	1896
B1	L-Val	L-Phe	$C_{97}H_{139}N_{19}O_{17}$	1843
C1	L-Val	L-Tyr	$C_{97}H_{139}N_{19}O_{18}$	1859
C2	L-Ile	L-Tyr	$C_{98}H_{141}N_{19}O_{18}$	1873

Action and use
Polypeptide antibacterial.

Ph Eur

DEFINITION
Gramicidin consists of a family of antimicrobial linear polypeptides, usually obtained by extraction from tyrothricin, the complex isolated from the fermentation broth of *Brevibacillus brevis* Dubos. The main component is gramicidin A1, together with gramicidins A2, B1, C1 and C2 in particular.

Content
Minimum 900 IU/mg (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder, slightly hygroscopic.

Solubility
Practically insoluble in water, soluble in methanol, sparingly soluble in alcohol.

mp
About 230 °C.

IDENTIFICATION
First identification: A, C.
Second identification: A, B.

A. Dissolve 0.100 g in *alcohol R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *alcohol R*. Examined between 240 nm and 320 nm (*2.2.25*), the solution shows 2 absorption maxima, at 282 nm and 290 nm, a shoulder at about 275 nm and an absorption minimum at 247 nm. The specific absorbance at the maximum at 282 nm is 105 to 125.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 5 mg of the substance to be examined in 6.0 mL of *alcohol R*.

Reference solution (a) Dissolve 5 mg of *gramicidin CRS* in 6.0 mL of *alcohol R*.

Reference solution (b) Dissolve 5 mg of *tyrothricin CRS* in 6.0 mL of *alcohol R*.

Plate *TLC silica gel plate R.*

Mobile phase *methanol R, butanol R, water R, glacial acetic acid R, butyl acetate R* (3:9:15:24:49 *V/V/V/V/V*).

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Dip the plate into *dimethylaminobenzaldehyde solution R2*. Heat at 90 °C until the spots appear.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots or 2 clearly separated groups of spots.

Results The principal spot or group of principal spots in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot or group of principal spots in the chromatogram obtained with reference solution (a) and to the spot or group of spots with the highest R_F value in the chromatogram obtained with reference solution (b).

C. Examine the chromatograms obtained in the test for composition.

Results The 3 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 3 principal peaks in the chromatogram obtained with reference solution (a).

TESTS
Composition
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of *methanol R* and dilute to 25 mL with the mobile phase.

Reference solution (a) Dissolve 25 mg of *gramicidin CRS* in 10 mL of *methanol R* and dilute to 25 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 50 °C.

Mobile phase *water R, methanol R* (29:71 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 20 µL.

Run time 2.5 times the retention time of gramicidin A1.

Relative retention With reference to gramicidin A1 (retention time = about 22 min): gramicidin C1 = about 0.7; gramicidin C2 = about 0.8; gramicidin A2 = about 1.2; gramicidin B1 = about 1.9.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to gramicidin A1 and gramicidin A2,
— the chromatogram obtained is concordant with the chromatogram supplied with *gramicidin CRS*.

Limits:
— *sum of gramicidins A1, A2, B1, C1 and C2*: minimum 95.0 per cent,
— *ratio of gramicidin A1 to the sum of gramicidins A1, A2, B1, C1 and C2*: minimum 0.60,
— *disregard limit*: the area of the peak due to gramicidin A1 in the chromatogram obtained with reference solution (b).

Related substances
Liquid chromatography (*2.2.29*) as described in the test for composition.

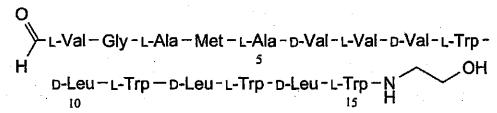
Limit:
— *any impurity*: maximum 2.0 per cent and not more than 1 peak is more than 1.0 per cent; disregard the peaks due to gramicidins A1, A2, B1, C1 and C2.

Loss on drying (*2.2.32*)
Maximum 3.0 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide R* at 60 °C at a pressure not exceeding 0.1 kPa for 3 h.

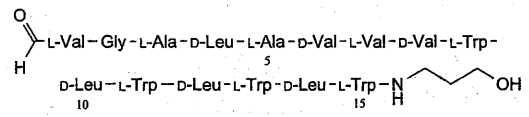
Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.0 g.

ASSAY
Carry out the microbiological assay of antibiotics (*2.7.2*), using the turbidimetric method. Use *gramicidin CRS* as the reference substance.

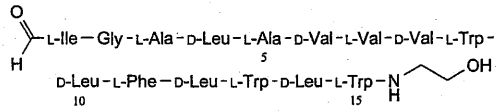
STORAGE
In an airtight container, protected from light.

IMPURITIES


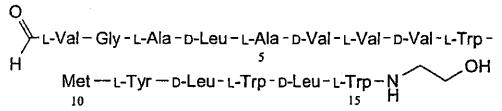
A. [4-methionine]gramicidin A1,



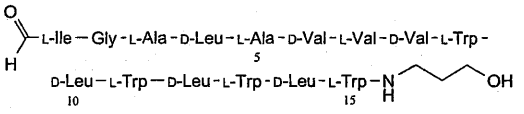
B. gramicidin A1 3-hydroxypropyl,



C. gramicidin B2,



D. [10-methionine]gramicidin C1,

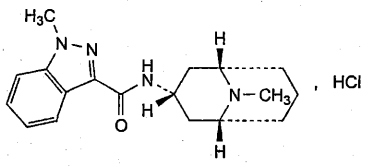


E. gramicidin A2 3-hydroxypropyl.

Ph Eur

Granisetron Hydrochloride

(Ph. Eur. monograph 1695)



$C_{18}H_{25}ClN_4O$ 348.9 *107007-99-8*

Action and use
Serotonin $5HT_3$ receptor antagonist; treatment of nausea and vomiting.

Ph Eur

DEFINITION
1-Methyl-*N*-[(1*R*,3*r*,5*S*)-9-methyl-9-azabicyclo[3.3.1]non-3-yl]-1*H*-indazole-3-carboxamide hydrochloride.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, sparingly soluble in methylene chloride, slightly soluble in methanol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *granisetron hydrochloride CRS.*
B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S
Dissolve 0.2 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.0 to 6.5 for solution S.

Impurity E
Thin-layer chromatography (*2.2.27*).
Solvent mixture *water R, acetonitrile R* (20:80 *V/V*).
Test solution Dissolve 0.25 g of the substance to be examined in the solvent mixture and dilute to 5 mL with the solvent mixture.
Reference solution Dissolve 5.0 mg of *granisetron impurity E CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture.
Plate *TLC silica gel F_{254} plate R.*
Mobile phase *concentrated ammonia R, 2-propanol R, ethyl acetate R* (6.5:30:50 *V/V/V*).
Application 2 µL.
Development Over half of the plate.
Drying In air.
Detection Expose to iodine vapour for 30 min.
Limit:
— *impurity E*: any spot due to impurity E is not more intense than the principal spot in the chromatogram obtained with the reference solution (0.5 per cent).

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.
Reference solution (b) Transfer 2 mL of the test solution to a colourless glass vial, stopper and expose the solution either to sunlight for 4 h or place under a UV lamp for 16 h (partial degradation of granisetron to impurity C). A degradation of at least about 0.3 per cent of granisetron to impurity C must be obtained as shown by appearance of a corresponding peak in the chromatogram. If not, expose the solution once again to sunlight or place under a UV lamp.
Reference solution (c) Dissolve 50.0 mg of *granisetron hydrochloride CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (d) Dissolve the contents of a vial of *granisetron impurity A CRS* in 1 mL of the mobile phase.
Reference solution (e) Dissolve the contents of a vial of *granisetron impurity B CRS* in 1 mL of the mobile phase.
Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: spherical *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.
Mobile phase Dilute 1.6 mL of *phosphoric acid R* to 800 mL with *water for chromatography R*, add 200 mL of *acetonitrile R* and mix. Add 1.0 mL of *hexylamine R* and mix. Adjust to pH 7.5 ± 0.05 with freshly distilled *triethylamine R* (about 4 mL).
Flow rate 1.5 mL/min.
Detection Spectrophotometer at 305 nm.
Injection 10 µL of the test solution and reference solutions (a), (b), (d) and (e).
Run time Twice the retention time of granisetron.
Relative retention With reference to granisetron (retention time = about 7 min): impurity D = about 0.4; impurity B = about 0.5; impurity A = about 0.7; impurity C = about 0.8.
System suitability:
— *resolution*: minimum 3.5 between the peaks due to impurity C and granisetron in the chromatogram obtained with reference solution (b);

— *symmetry factor*: maximum 2.0 for the peak due to granisetron in the chromatogram obtained with reference solution (a).

Limits:

— *correction factor*: for the calculation of content, multiply the peak area of impurity B by 1.7;
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurity C*: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *impurity D*: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

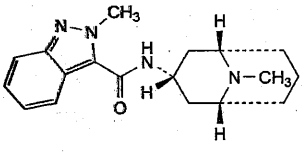
Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{18}H_{25}ClN_4O$ taking into account the assigned content of *granisetron hydrochloride CRS*.

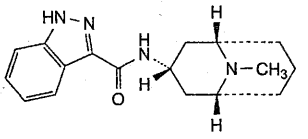
IMPURITIES

Specified impurities *A, B, C, D, E.*

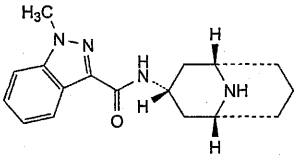
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G, H, I.*



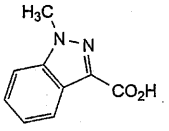
A. 2-methyl-*N*-[(1*R*,3*r*,5*S*)-9-methyl-9-azabicyclo[3.3.1]non-3-yl]-2*H*-indazole-3-carboxamide,



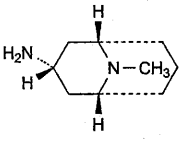
B. *N*-[(1*R*,3*r*,5*S*)-9-methyl-9-azabicyclo[3.3.1]non-3-yl]-1*H*-indazole-3-carboxamide,



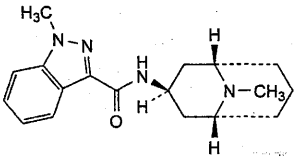
C. *N*-[(1*R*,3*r*,5*S*)-9-azabicyclo[3.3.1]non-3-yl]-1-methyl-1*H*-indazole-3-carboxamide,



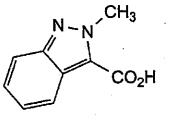
D. 1-methyl-1*H*-indazole-3-carboxylic acid,



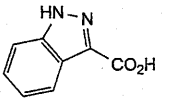
E. (1*R*,3*r*,5*S*)-9-methyl-9-azabicyclo[3.3.1]nonan-3-amine,



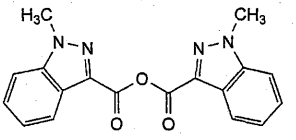
F. 1-methyl-*N*-[(1*R*,3*s*,5*S*)-9-methyl-9-azabicyclo[3.3.1]non-3-yl]-1*H*-indazole-3-carboxamide (*exo*-granisetron),



G. 2-methyl-2*H*-indazole-3-carboxylic acid,



H. 1*H*-indazole-3-carboxylic acid,

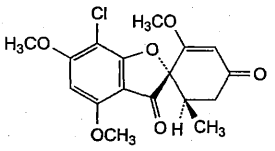


I. 1-methyl-1*H*-indazole-3-carboxylic anhydride.

Ph Eur

Griseofulvin

(Ph. Eur. monograph 0182)



$C_{17}H_{17}ClO_6$ 352.8 *126-07-8*

Action and use
Antifungal.

Preparation
Griseofulvin Tablets

Ph Eur

DEFINITION

(1′*S*,6′*R*)-7-Chloro-2′,4,6-trimethoxy-6′-methyl-3*H*-spiro[1-benzofuran-2,1′-cyclohex[2]ene]-3,4′-dione.

Substance produced by the growth of certain strains of *Penicillium griseofulvum*.

Content
94.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance
White or yellowish-white powder.

Solubility
Practically insoluble in water, freely soluble in dimethylformamide, slightly soluble in anhydrous ethanol and in methanol.

It shows polymorphism (*5.9*).

mp
About 220 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *griseofulvin for LC assay and identification CRS*.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in *acetone R*, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_4 (*2.2.2, Method II*).

Dissolve 0.75 g in *dimethylformamide R* and dilute to 10 mL with the same solvent.

Acidity
Suspend 0.25 g in 20 mL of *ethanol (96 per cent) R* and add 0.1 mL of *phenolphthalein solution R*. Not more than 1.0 mL of *0.02 M sodium hydroxide* is required to change the colour of the indicator.

Specific optical rotation (*2.2.7*)
+ 354 to + 364 (dried substance).

Dissolve 0.250 g in *dimethylformamide R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase B and dilute to 50.0 mL with mobile phase B.

Reference solution (a) Dissolve 25.0 mg of *griseofulvin for LC assay and identification CRS* in mobile phase B and dilute to 50.0 mL with mobile phase B.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase B.

Reference solution (c) Dissolve 5 mg of *griseofulvin for system suitability CRS* (containing impurities A, B and C) in mobile phase B and dilute to 10 mL with mobile phase B.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: mix 20 volumes of a 0.1 per cent *V/V* solution of *anhydrous formic acid R* previously adjusted to pH 4.5 with *dilute ammonia R2* and 80 volumes of *water for chromatography R*;
— *mobile phase B*: mix 15 volumes of *water for chromatography R*, 20 volumes of a 0.1 per cent *V/V* solution of *anhydrous formic acid R* previously adjusted to pH 4.5 with *dilute ammonia R2* and 65 volumes of *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	50	50
3 - 13	50 → 40	50 → 60
13 - 16	40 → 10	60 → 90
16 - 24	10	90

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 290 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to griseofulvin (retention time = about 16 min): impurity A = about 0.4; impurity B = about 0.7; impurity C = about 1.1.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to griseofulvin.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity A by 0.6;
— for each impurity, use the concentration of griseofulvin in reference solution (b).

Limits:
— *impurity B*: maximum 3.0 per cent;
— *impurity A*: maximum 2.0 per cent;
— *impurity C*: maximum 0.75 per cent;
— *unspecified impurities*: for each impurity, maximum 0.15 per cent;
— *total*: maximum 5.0 per cent;
— *reporting threshold*: 0.10 per cent.

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

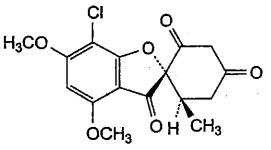
Injection Test solution and reference solution (a).

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 2.0 per cent determined on 6 injections.

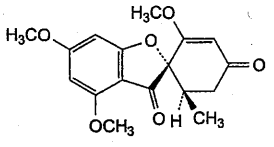
Calculate the percentage content of $C_{17}H_{17}ClO_6$ taking into account the assigned content of *griseofulvin for LC assay and identification CRS*.

IMPURITIES

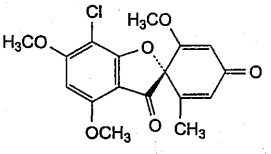
Specified impurities *A, B, C.*



A. (1′*S*,6′*R*)-7-chloro-4,6-dimethoxy-6′-methyl-3*H*-spiro[1-benzofuran-2,1′-cyclohexane]-2′,3,4′-trione,



B. (1′*S*,6′*R*)-2′,4,6-trimethoxy-6′-methyl-3*H*-spiro[1-benzofuran-2,1′-cyclohex[2]ene]-3,4′-dione,

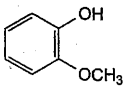


C. (1′*S*)-7-chloro-2′,4,6-trimethoxy-6′-methyl-3*H*-spiro[1-benzofuran-2,1′-cyclohexa[2,5]diene]-3,4′-dione.

Ph Eur

Guaiacol

(Ph. Eur. monograph 1978)



$C_7H_8O_2$ 124.1 *90-05-1*

Ph Eur

DEFINITION

2-Methoxyphenol.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Crystalline mass or colourless or yellowish liquid, hygroscopic.

Solubility
Sparingly soluble in water, very soluble in methylene chloride, freely soluble in ethanol (96 per cent).

mp
About 28 °C.

IDENTIFICATION

First identification: A.

Second identification: B.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *guaiacol CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.5 g of the substance to be examined in *methanol R* and dilute to 25 mL with the same solvent.

Reference solution Dissolve 0.5 g of *guaiacol CRS* in *methanol R* and dilute to 25 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *anhydrous acetic acid R, methanol R, toluene R* (6:14:80 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ferric chloride solution R1.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Solution S
Dissolve 1.00 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method I*).

Acidity or alkalinity
To 5.0 mL of solution S, add 10 mL of *carbon dioxide-free water R* and 0.1 mL of *methyl red mixed solution R*. Not more than 0.05 mL of *0.1 M hydrochloric acid* or *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Impurity A
Liquid chromatography (*2.2.29*).

Solvent mixture *phosphoric acid R, water R, methanol R* (1:499:500 *V/V/V*).

Test solution (a) Dissolve 1.0 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve 0.20 g of *pyrocatechol R* (impurity A) and 0.20 g of *phenol R* (impurity B) in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL of this solution to 10 mL with the solvent mixture.

Reference solution (c) Dissolve 20.0 mg of *guaiacol CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *phosphoric acid R, methanol R, water R* (1:150:849 *V/V/V*);
— *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 28	100	0
28 - 30	100 → 35	0 → 65
30 - 40	35	65

Flow rate 1 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Retention time Guaiacol = about 20 min.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to impurities A (1st peak) and B (2nd peak).

Limit:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Related substances

Gas chromatography (*2.2.28*): use the normalisation procedure.

Test solution Dissolve 1.00 g of the substance to be examined in *acetonitrile R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.20 g of *phenol R* (impurity B) and 0.40 g of *methyl benzoate R* (impurity E) in *acetonitrile R* and dilute to 50 mL with the same solvent. Dilute 1 mL of this solution to 20 mL with *acetonitrile R*.

Reference solution (b) Dilute 0.5 mL of the test solution to 100.0 mL with *acetonitrile R*. Dilute 1.0 mL of this solution to 10.0 mL with *acetonitrile R*.

Reference solution (c) Dissolve 10 mg of *veratrole R* (impurity C) in *acetonitrile R* and dilute to 10 mL with the same solvent.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.53 mm;
— *stationary phase*: *poly(cyanopropyl) (7) (phenyl) (7) (methyl) (86) siloxane R* (film thickness 2 µm).

Carrier gas *helium for chromatography R*.

Flow rate 5 mL/min.

Split ratio 1:5.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 15	90
	15 - 45	90 → 180
Injection port		200
Detector		220

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to guaiacol (retention time = about 25 min): impurity E = about 0.88; impurity B = about 0.92; impurity C = about 1.1.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurities E (1st peak) and B (2nd peak).

Limits:
— *impurity C*: maximum 0.4 per cent;
— *impurity E*: maximum 0.2 per cent;
— *impurity B*: maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.0 per cent;
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 2.000 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for impurity A with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_7H_8O_2$ from the declared content of *guaiacol CRS*.

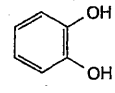
STORAGE

In an airtight container, protected from light.

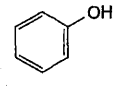
IMPURITIES

Specified impurities *A, B, C, E.*

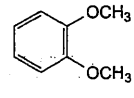
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D, F, G, H.*



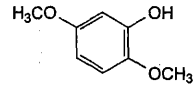
A. benzene-1,2-diol (pyrocatechol),



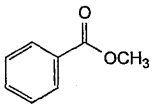
B. phenol,



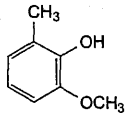
C. 1,2-dimethoxybenzene (veratrole),



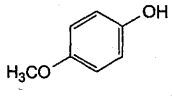
D. 2,5-dimethoxyphenol,



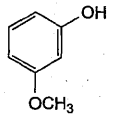
E. methyl benzoate,



F. 2-methoxy-6-methylphenol (6-methylguaiacol),



G. 4-methoxyphenol,

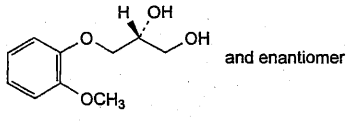


H. 3-methoxyphenol.

Ph Eur

Guaifenesin

(Ph. Eur. monograph 0615)



$C_{10}H_{14}O_4$ 198.2 *93-14-1*

Action and use
Expectorant.

Ph Eur

DEFINITION
(2*RS*)-3-(2-Methoxyphenoxy)propane-1,2-diol.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Melting point (*2.2.14*): 79 °C to 83 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *guaifenesin CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 30 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 30 mg of *guaifenesin CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *methylene chloride R, 2-propanol R* (20:80 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a solution, prepared immediately before use, containing 0.4 g of *dimethylaminobenzaldehyde R* in an ice-cooled mixture of 10 mL of *water R* and 10 mL of *hydrochloric acid R*; allow to dry in air and heat at 105 °C for 10 min; examine under white light.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Solution S
Dissolve 1.0 g in *carbon dioxide-free water R*, heating gently if necessary, and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.05 mL of *phenolphthalein solution R1*. Not more than 0.1 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator. To 10 mL of solution S add 0.15 mL of *methyl red solution R*. Not more than 0.1 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Related substances
Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.100 g of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 50.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dissolve 50.0 mg of *guaiacol R* (impurity A) in *water R* and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 10.0 mL with test solution (a).

Reference solution (c) Dissolve 5.0 mg of *guaifenesin for peak identification CRS* (containing impurities B, C and D) in *water R* and dilute to 5.0 mL with the same solvent.

Reference solution (d) Dissolve 25.0 mg of *guaifenesin CRS* in *water R* and dilute to 50.0 mL with the same solvent.

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A: glacial acetic acid R, water for chromatography R* (1:99 *V/V*);
— *mobile phase B: acetonitrile R;*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	80	20
2 - 34	80 → 50	20 → 50

Flow rate 1 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, C and D.

Relative retention With reference to guaifenesin (retention time = about 8 min): impurity B = about 0.9; impurity A = about 1.4; impurity C = about 3.1; impurity D = about 3.7.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to guaifenesin and impurity A.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity D by 0.7;
— for each impurity, use the concentration of guaifenesin in reference solution (a).

Limits:
— *impurity B*: maximum 0.5 per cent;
— *impurities C, D*: for each impurity, maximum 0.1 per cent;
— *unspecified impurities*: for each impurity, maximum 0.05 per cent;
— *total*: maximum 0.7 per cent;
— *reporting threshold*: 0.03 per cent.

Chlorides and monochlorhydrins
Maximum 250 ppm.

To 10 mL of solution S add 2 mL of *dilute sodium hydroxide solution R* and heat on a water-bath for 5 min. Cool and add 3 mL of *dilute nitric acid R*. The resulting solution complies with the limit test for chlorides (*2.4.4*).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

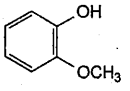
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

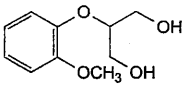
Calculate the percentage content of $C_{10}H_{14}O_4$ taking into account the assigned content of *guaifenesin CRS*.

IMPURITIES
Specified impurities *B, C, D.*

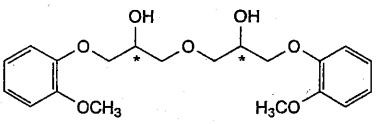
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A.*



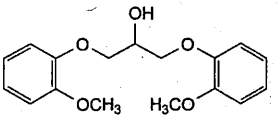
A. 2-methoxyphenol (guaiacol),



B. 2-(2-methoxyphenoxy)propane-1,3-diol (B-isomer),



C. 1,1′-oxybis[3-(2-methoxyphenoxy)propan-2-ol] (bisether),

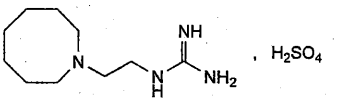


D. 1,3-bis(2-methoxyphenoxy)propan-2-ol.

Ph Eur

Guanethidine Monosulfate

Guanethidine Monosulphate
(Ph. Eur. monograph 0027)



$C_{10}H_{24}N_4O_4S$ 296.4 *645-43-2*

Action and use
Adrenergic neuron blocker.

Preparation
Guanethidine Tablets

Ph Eur

DEFINITION
1-[2-(Hexahydroazocin-1(2*H*)-yl)ethyl]guanidine monosulfate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
Colourless, crystalline powder.

Solubility
Freely soluble in water, practically insoluble in ethanol (96 per cent).

mp
About 250 °C, with decomposition.

IDENTIFICATION
A. Dissolve about 25 mg in 25 mL of *water R*, add 20 mL of *picric acid solution R* and filter. The precipitate, washed with *water R* and dried at 100-105 °C, melts (*2.2.14*) at about 154 °C.

B. Dissolve about 25 mg in 5 mL of *water R*. Add 1 mL of *strong sodium hydroxide solution R*, 1 mL of *α-naphthol solution R* and, dropwise with shaking, 0.5 mL of *strong sodium hypochlorite solution R*. A bright pink precipitate is formed and becomes violet-red on standing.

C. It gives the reactions of sulfates (*2.3.1*).

TESTS

Solution S

Dissolve 0.4 g in *carbon dioxide-free water R* and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).

pH (*2.2.3*)

4.7 to 5.5 for solution S.

Oxidisable substances

In a conical, ground-glass-stoppered flask, dissolve 1.0 g in 25 mL of *water R* and add 25 mL of *dilute sodium hydroxide solution R*. Allow to stand for 10 min and add 1 g of *potassium bromide R* and 1 mL of 0.0083 M potassium bromate. Acidify with 30 mL of *dilute hydrochloric acid R*. Mix and allow to stand in the dark for 5 min. Add 2 g of *potassium iodide R* and shake. Allow to stand for 2 min and titrate the liberated iodine with *0.05 M sodium thiosulfate*, using *starch solution R* as indicator. Not less than 0.3 mL of *0.05 M sodium thiosulfate* is required to decolorise the solution.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.00 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g, warming if necessary, in 30 mL of *anhydrous acetic acid R* and add 15 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 29.64 mg of $C_{10}H_{24}N_4O_4S$.

STORAGE

Protected from light.

Ph Eur

Guar

(Ph. Eur. monograph 1218)

Ph Eur

DEFINITION

Guar is obtained by grinding the endosperms of seeds of *Cyamopsis tetragonolobus* (L.) Taub. It consists mainly of guar galactomannan.

CHARACTERS

Appearance

White or almost white powder.

Solubility

It yields a mucilage of variable viscosity when dissolved in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Examined under a microscope in *glycerol R*, the substance to be examined (125) (*2.9.12*) shows pyriform or ovoid cells, usually isolated, having very thick walls around a central somewhat elongated lumen with granular contents, and smaller polyhedral cells, isolated or in clusters, with thinner walls.

B. In a conical flask place 2 g, add rapidly 45 mL of *water R* and stir vigorously for 30 s. After 5-10 min a stiff gel forms which does not flow when the flask is inverted.

C. Mix a suspension of 0.1 g in 10 mL of *water R* with 1 mL of a 10 g/L solution of *disodium tetraborate R*; the mixture soon gels.

D. Thin-layer chromatography (*2.2.27*).

Test solution To 10 mg of the substance to be examined in a thick-walled centrifuge tube add 2 mL of a 100 g/L solution of *trifluoroacetic acid R*, shake vigorously to dissolve the forming gel, stopper the tube and heat the mixture at 120 °C for 1 h. Centrifuge the hydrolysate, transfer the clear supernatant carefully into a 50 mL flask, add 10 mL of *water R* and evaporate the solution to dryness under reduced pressure. To the resulting clear film add 0.1 mL of *water R* and 0.9 mL of *methanol R*. Centrifuge to separate the amorphous precipitate. Dilute the supernatant, if necessary, to 1 mL with *methanol R*.

Reference solution Dissolve 10 mg of *galactose R* and 10 mg of *mannose R* in 2 mL of *water R*, then dilute to 20 mL with *methanol R*.

Plate *TLC silica gel plate R*.

Mobile phase *water R, acetonitrile R* (15:85 *V/V*).

Application 5 µL, as bands.

Development Over a path of 15 cm.

Detection Spray with *aminohippuric acid reagent R* and dry at 120 °C for 5 min.

Results The chromatogram obtained with the reference solution shows in the lower part 2 clearly separated brownish zones due to galactose and mannose in order of increasing R_F value; the chromatogram obtained with the test solution shows 2 zones due to galactose and mannose.

TESTS

Tragacanth, sterculia gum, agar, alginates, carrageenan

To a small amount of the substance to be examined add 0.2 mL of freshly prepared *ruthenium red solution R*. Examined under a microscope the cell walls do not stain red.

Protein

Maximum 8.0 per cent.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.170 g. Multiply the result by 6.25.

Apparent viscosity (*2.2.10*)

85 per cent to 115 per cent of the value stated on the label.

Moisten a quantity equivalent to 1.00 g of the dried substance with 2.5 mL of *2-propanol R*. While stirring, dilute to 100.0 mL with *water R*. After 1 h, determine the viscosity at 20 °C using a rotating viscometer and a shear rate of 100 s^{-1}.

Loss on drying (*2.2.32*)

Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 5 h.

Total ash (*2.4.16*)

Maximum 1.8 per cent.

Microbial contamination

TAMC: acceptance criterion 10^4 CFU/g (*2.6.12*).

TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

Absence of *Escherichia coli* (*2.6.13*).

Absence of *Salmonella* (*2.6.13*).

LABELLING
The label states the apparent viscosity in millipascal seconds for a 10 g/L solution.

FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for guar used as viscosity-increasing agent or binder.

Apparent viscosity
See Tests.

Ph Eur

Guar Galactomannan

(Ph. Eur. monograph 0908)

Action and use
Excipient.

Ph Eur

DEFINITION
Guar galactomannan is obtained from the seeds of *Cyamopsis tetragonolobus* (L.) Taub. by grinding of the endosperms and subsequent partial hydrolysis. The main components are polysaccharides composed of D-galactose and D-mannose at molar ratios of 1:1.4 to 1:2. The molecules consist of a linear main chain of β-(1→4)-glycosidically linked mannopyranoses and single α-(1→6)-glycosidically linked galactopyranoses.

CHARACTERS
Appearance
Yellowish-white powder.

Solubility
Soluble in cold water and in hot water, practically insoluble in organic solvents.

IDENTIFICATION
A. Mix 5 g of solution S (see Tests) with 0.5 mL of a 10 g/L solution of *disodium tetraborate R*. A gel forms within a short time.

B. Heat 20 g of solution S in a water-bath for 10 min. Allow to cool and adjust to the original mass with *water R*. The solution does not gel.

C. Thin-layer chromatography (*2.2.27*).

Test solution To 10 mg of the substance to be examined in a thick-walled centrifuge tube add 2 mL of a 230 g/L solution of *trifluoroacetic acid R*, shake vigorously to dissolve the forming gel, stopper the tube and heat the mixture at 120 °C for 1 h. Centrifuge the hydrolysate, transfer the clear supernatant carefully into a 50 mL flask, add 10 mL of *water R* and evaporate the solution to dryness under reduced pressure. Take up the residue in 10 mL of *water R* and evaporate again to dryness under reduced pressure. To the resulting clear film, which has no odour of acetic acid, add 0.1 mL of *water R* and 1 mL of *methanol R*. Centrifuge to separate the amorphous precipitate. Dilute the supernatant, if necessary, to 1 mL with *methanol R*.

Reference solution Dissolve 10 mg of *galactose R* and 10 mg of *mannose R* in 2 mL of *water R* and dilute to 10 mL with *methanol R*.

Plate *TLC silica gel G plate R*.

Mobile phase *water R, acetonitrile R* (15:85 *V/V*).

Application 5 µL, as bands of 20 mm by 3 mm.

Development Over a path of 15 cm.

Detection Spray with *aminohippuric acid reagent R* and heat at 120 °C for 5 min.

Results The chromatogram obtained with the reference solution shows in the lower part 2 clearly separated brownish zones due to galactose and mannose in order of increasing R_F value; the chromatogram obtained with the test solution shows 2 zones due to galactose and mannose.

TESTS
Solution S
Moisten 1.0 g with 2 mL of *2-propanol R*. While stirring, dilute to 100 g with *water R* and stir until the substance is uniformly dispersed. Allow to stand for at least 1 h. If the apparent viscosity is below 200 mPa·s, use 3.0 g of substance instead of 1.0 g.

pH (*2.2.3*)
5.5 to 7.5 for solution S.

Apparent viscosity (*2.2.10*)
75 per cent to 140 per cent of the value stated on the label.

Moisten a quantity of the substance to be examined equivalent to 2.00 g of the dried substance with 2.5 mL of *2-propanol R* and, while stirring, dilute to 100.0 mL with *water R*. After 1 h, determine the viscosity at 20 °C using a rotating viscometer and a shear rate of 100 s^{-1}.

Insoluble matter
Maximum 7.0 per cent.

In a 250 mL flask disperse, while stirring, 1.50 g in a mixture of 1.6 mL of *sulfuric acid R* and 150 mL of *water R* and weigh. Immerse the flask in a water-bath and heat under a reflux condenser for 6 h. Adjust to the original mass with *water R*. Filter the hot solution through a tared, sintered-glass filter (160) (*2.1.2*). Rinse the filter with hot *water R* and dry at 100-105 °C. The residue weighs a maximum of 105 mg.

Protein
Maximum 5.0 per cent.

Carry out the determination of nitrogen by sulfuric acid digestion (*2.5.9*), using 0.400 g. Multiply the result by 6.25.

Tragacanth, sterculia gum, agar, alginates and carrageenan
To a small amount of the substance to be examined add 0.2 mL of freshly prepared *ruthenium red solution R*. Examined under a microscope, none of the structures are red.

Loss on drying (*2.2.32*)
Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 5 h.

Total ash (*2.4.16*)
Maximum 1.8 per cent, determined on 1.00 g after wetting with 10 mL of *water R*.

Microbial contamination
TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*).
TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*).
Absence of *Escherichia coli* (*2.6.13*).
Absence of *Salmonella* (*2.6.13*).

LABELLING
The label states the apparent viscosity in millipascal seconds for a 20 g/L solution.

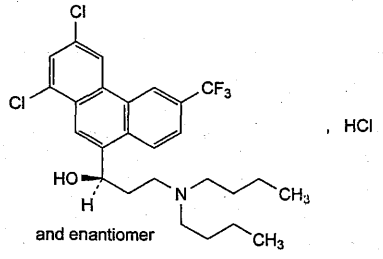
FUNCTIONALITY-RELATED CHARACTERISTICS
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.
The following characteristic may be relevant for guar galactomannan used as viscosity-increasing agent or binder.

Apparent viscosity
See Tests.

Ph Eur

Halofantrine Hydrochloride

(Ph. Eur. monograph 1979)



$C_{26}H_{31}Cl_3F_3NO$ 536.9 *36167-63-2*

Action and use
Antiprotozoal (malaria).

Ph Eur

DEFINITION
(1*RS*)-3-(Dibutylamino)-1-[1,3-dichloro-6-(trifluoromethyl)phenanthren-9-yl]propan-1-ol hydrochloride.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, freely soluble in methanol, sparingly soluble in alcohol.

It shows polymorphism (*5.9*).

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *halofantrine hydrochloride CRS.*
If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methyl ethyl ketone R*, evaporate to dryness and record new spectra using the residues.
B. It gives reaction (b) of chlorides (*2.3.1*).

TESTS
Optical rotation (*2.2.7*)
−0.10° to + 0.10°.
Dissolve 1.00 g in *alcohol R* and dilute to 100.0 mL with the same solvent.

Absorbance (*2.2.25*)
Maximum 0.085 at 450 nm.
Dissolve 0.200 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).
Test solution (a) Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.
Reference solution (a) Dissolve 40.0 mg of *halofantrine hydrochloride CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the mobile phase.
Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.
Reference solution (d) Dissolve 10.0 mg of *halofantrine impurity C CRS* in the mobile phase and dilute to 25 mL with the mobile phase. To 5.0 mL of the solution, add 5.0 mL of reference solution (a) and dilute to 50.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.30 m, Ø = 3.9 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (10 µm) of irregular type, with a specific surface of 330 m^2/g, a pore size of 12.5 nm and a carbon loading of 9.8 per cent .
Mobile phase Mix 250 mL of a 2.0 g/L solution of *sodium hydroxide R*, previously adjusted to pH 2.5 with *perchloric acid R* and 750 mL of *acetonitrile R*.
Flow rate 1 mL/min.
Detection Spectrophotometer at 260 nm.
Injection 20 µL; inject the test solution (a) and reference solutions (c) and (d).
Run time 5 times the retention time of halofantrine which is about 6 min.
System suitability:
— *resolution*: minimum 3.3 between the peaks due to halofantrine and impurity C in the chromatogram obtained with reference solution (d).
Limits:
— *any impurity*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent),

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

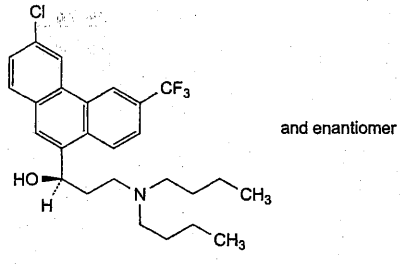
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances.

Injection Test solution (b) and reference solution (b).

Calculate the percentage content of halofantrine hydrochloride.

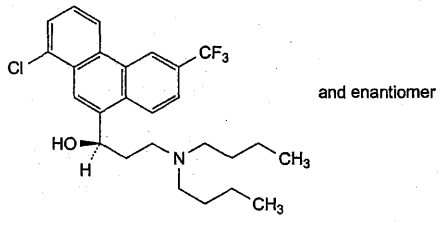
STORAGE
Protected from light.

IMPURITIES



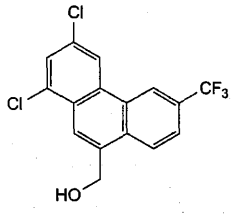
and enantiomer

A. (1*RS*)-1-[3-chloro-6-(trifluoromethyl)phenanthren-9-yl]-3-(dibutylamino)propan-1-ol (1-dechlorohalofantrine),



and enantiomer

B. (1*RS*)-1-[1-chloro-6-(trifluoromethyl)phenanthren-9-yl]-3-(dibutylamino)propan-1-ol (3-dechlorohalofantrine),

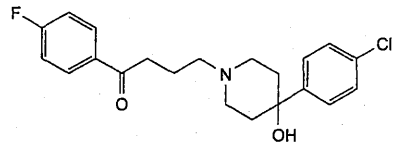


C. [1,3-dichloro-6-(trifluoromethyl)phenanthren-9-yl] methanol.

Ph Eur

Haloperidol

(Ph. Eur. monograph 0616)



$C_{21}H_{23}ClFNO_2$ 375.9 *52-86-8*

Action and use
Dopamine receptor antagonist; neuroleptic.

Preparations
Haloperidol Capsules
Haloperidol Injection
Haloperidol Oral Solution
Strong Haloperidol Oral Solution
Haloperidol Tablets

Ph Eur

DEFINITION
4-[4-(4-Chlorophenyl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, slightly soluble in ethanol (96 per cent), in methanol and in methylene chloride.

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.
A. Melting point (*2.2.14*): 150 °C to 153 °C.
B. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *haloperidol CRS.*
C. Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 10 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.
Reference solution (a) Dissolve 10 mg of *haloperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of *haloperidol CRS* and 10 mg of *bromperidol CRS* in *methanol R* and dilute to 10 mL with the same solvent.
Plate *TLC octadecylsilyl silica gel plate R.*
Mobile phase *tetrahydrofuran R, methanol R*, 58 g/L solution of *sodium chloride R* (10:45:45 *V/V/V*).
Application 1 µL.
Development In an unsaturated tank, over 2/3 of the plate.
Drying In air.
Detection Examine in ultraviolet light at 254 nm.
System suitability Reference solution (b):
— the chromatogram shows 2 spots which may, however, not be completely separated.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 5 mL of *anhydrous ethanol R*. Add 0.5 mL of *dinitrobenzene solution R* and 0.5 mL of *2 M alcoholic potassium hydroxide R*. A violet colour is produced and becomes brownish-red after 20 min.

E. To 0.1 g in a platinum crucible add 0.5 g of *anhydrous sodium carbonate R*. Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 0.2 g in 20 mL of a 1 per cent *V/V* solution of *lactic acid R*.

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of *haloperidol for system suitability CRS* (containing impurities B and D) in 1.0 mL of *methanol R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanol R*.

Reference solution (c) Dissolve 10 mg of *haloperidol for peak identification CRS* (containing impurities G and H) in 1.0 mL of *methanol R*.

Column:
- *size*: *l* = 0.1 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
- *mobile phase A*: 17 g/L solution of *tetrabutylammonium hydrogen sulfate R1*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	90	10
2 - 17	90 → 50	10 → 50
17 - 22	50	50

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *haloperidol for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and D; use the chromatogram supplied with *haloperidol for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities G and H.

Relative retention With reference to haloperidol (retention time = about 8 min): impurity B = about 0.9; impurity D = about 1.6; impurity G = about 1.8; impurity H = about 2.0.

System suitability Reference solution (a):
- *resolution*: minimum 3.0 between the peaks due to impurity B and haloperidol.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity B by 0.7;
- *impurity D*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurities G, H*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

1 mL of *0.1 M perchloric acid* is equivalent to 37.59 mg of $C_{21}H_{23}ClFNO_2$.

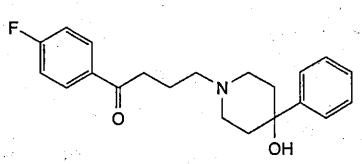
STORAGE

Protected from light.

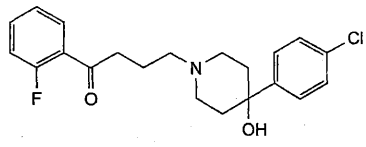
IMPURITIES

Specified impurities *B, D, G, H.*

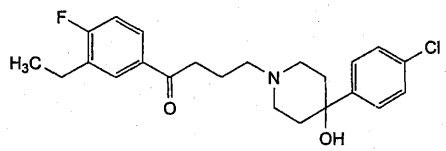
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C, E, F.*



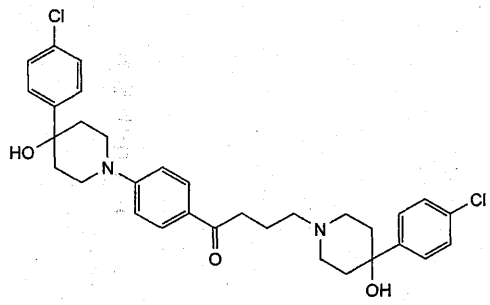
A. 1-(4-fluorophenyl)-4-(4-hydroxy-4-phenylpiperidin-1-yl) butan-1-one,



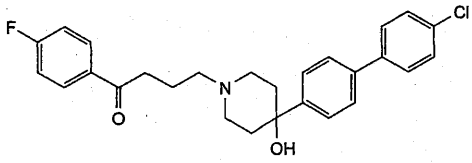
B. 4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]-1-(2-fluorophenyl)butan-1-one,



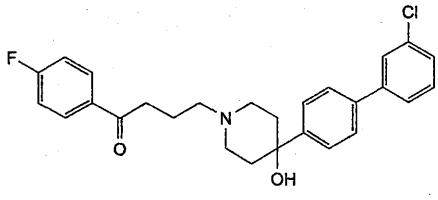
C. 4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]-1-(3-ethyl-4-fluorophenyl)butan-1-one,



D. 4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]-1-[4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]phenyl]butan-1-one,



E. 4-[4-(4′-chlorobiphenyl-4-yl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one,



F. 4-[4-(3′-chlorobiphenyl-4-yl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one,

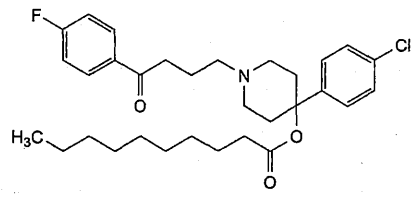
G. unknown structure,

H. unknown structure.

Ph Eur

Haloperidol Decanoate

(Ph. Eur. monograph 1431)



$C_{31}H_{41}ClFNO_3$ 530.1 *74050-97-8*

Action and use
Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION
4-(4-Chlorophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl] piperidin-4-yl decanoate.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, very soluble in ethanol (96 per cent), in methanol and in methylene chloride.

mp
About 42 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *haloperidol decanoate CRS.*

B. To 0.1 g in a porcelain crucible add 0.5 g of *anhydrous sodium carbonate R.* Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R.* The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_5 (*2.2.2, Method II*).

Dissolve 2.0 g in *methylene chloride R* and dilute to 20 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use and protect from light.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of *bromperidol decanoate CRS* and 2.5 mg of *haloperidol decanoate CRS* in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with *methanol R.* Dilute 1.0 mL of this solution to 10.0 mL with *methanol R.*

Column:
— *size: l* = 0.1 m, Ø = 4.0 mm;
— *stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: 27 g/L solution of *tetrabutylammonium hydrogen sulfate R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	80 → 40	20 → 60
30 - 35	40	60
35 - 40	40 → 80	60 → 20

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Relative retention With reference to haloperidol decanoate (retention time = about 24 min): impurity G = about 0.1; impurity L = about 0.2; impurity H = about 0.8; impurity I = about 0.88; impurity A = about 0.9; impurity B = about 0.98; bromperidol decanoate = about 1.02; impurity J = about 1.1; impurity C = about 1.15; impurity D = about 1.2; impurity K = about 1.22; impurity F = about 1.26; impurity E = about 1.28.

System suitability Reference solution (a):
— *resolution*: minimum 1.5 between the peaks due to haloperidol decanoate and bromperidol decanoate.

Limits:
— *impurities A, B, C, D, E, F, G, H, I, J, K*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 30 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.425 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, using 0.2 mL of *naphtholbenzein solution R* as indicator.

1 mL of *0.1 M perchloric acid* is equivalent to 53.01 mg of $C_{31}H_{41}ClFNO_3$.

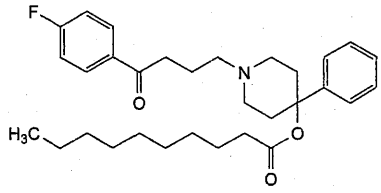
STORAGE

Protected from light, at a temperature below 25 °C.

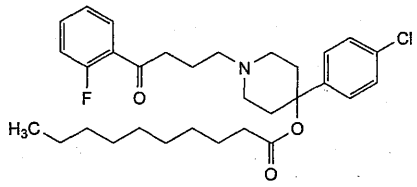
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H, I, J, K.*

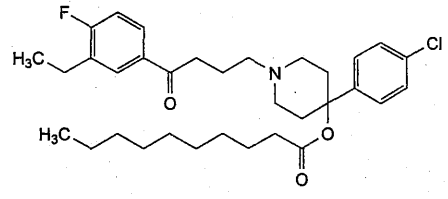
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *L.*



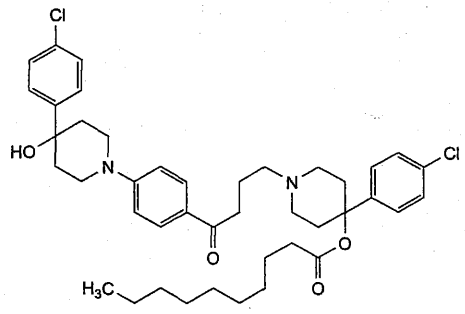
A. 1-[4-(4-fluorophenyl)-4-oxobutyl]-4-phenylpiperidin-4-yl decanoate,



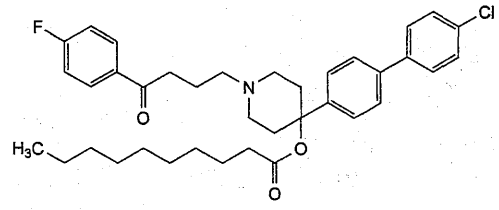
B. 4-(4-chlorophenyl)-1-[4-(2-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate,



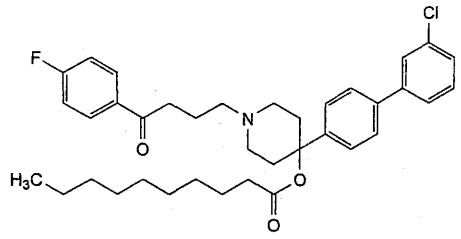
C. 4-(4-chlorophenyl)-1-[4-(3-ethyl-4-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate,



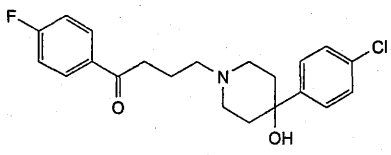
D. 4-(4-chlorophenyl)-1-[4-[4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]phenyl]-4-oxobutyl]piperidin-4-yl decanoate,



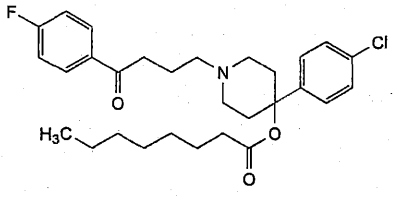
E. 4-(4′-chlorobiphenyl-4-yl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate,



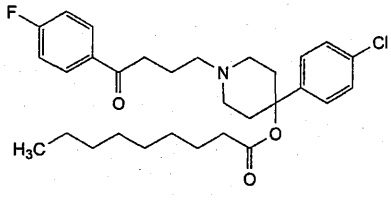
F. 4-(3′-chlorobiphenyl-4-yl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl decanoate,



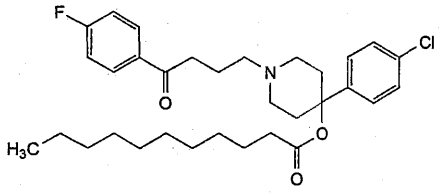
G. 4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]-1-(4-fluorophenyl)butan-1-one (haloperidol),



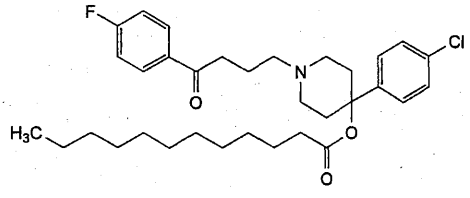
H. 4-(4-chlorophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl octanoate,



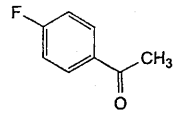
I. 4-(4-chlorophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl nonanoate,



J. 4-(4-chlorophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl undecanoate,



K. 4-(4-chlorophenyl)-1-[4-(4-fluorophenyl)-4-oxobutyl]piperidin-4-yl dodecanoate,

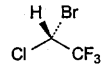


L. 1-(4-fluorophenyl)ethanone.

Ph Eur

Halothane

(Ph. Eur. monograph 0393)


and enantiomer

$C_2HBrClF_3$ 197.4 *151-67-7*

Action and use
General anaesthetic.

Ph Eur

DEFINITION

(*RS*)-2-Bromo-2-chloro-1,1,1-trifluoroethane to which 0.01 per cent *m/m* of thymol has been added.

CHARACTERS

Appearance
Clear, colourless, mobile, heavy, non-flammable liquid.

Solubility
Slightly soluble in water, miscible with anhydrous ethanol.

IDENTIFICATION

First identification: B.

Second identification: A, C.

A. Distillation range (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Examine the substance in a 0.1 mm cell.

Comparison *Ph. Eur. reference spectrum of halothane.*

C. Add 0.1 mL to 2 mL of *2-methyl-2-propanol R* in a test-tube. Add 1 mL of *copper edetate solution R*, 0.5 mL of *concentrated ammonia R* and a mixture of 0.4 mL of *strong hydrogen peroxide solution R* and 1.6 mL of *water R* (solution A). Prepare a blank at the same time (solution B). Place both tubes in a water-bath at 50 °C for 15 min, cool and add 0.3 mL of *glacial acetic acid R*. To 1 mL of each of solutions A and B add 0.5 mL of a mixture of equal volumes of freshly prepared *alizarin S solution R* and *zirconyl nitrate solution R*. Solution A is yellow and solution B is red.

To 1 mL of each of solutions A and B add 1 mL of *buffer solution pH 5.2 R*, 1 mL of *phenol red solution R* diluted 1 to 10 with *water R* and 0.1 mL of *chloramine solution R*. Solution A is bluish-violet and solution B is yellow.

To 2 mL of each of solutions A and B add 0.5 mL of a mixture of 25 volumes of *sulfuric acid R* and 75 volumes of *water R*, 0.5 mL of *acetone R* and 0.2 mL of a 50 g/L solution of *potassium bromate R* and shake. Warm the tubes in a water-bath at 50 °C for 2 min, cool and add 0.5 mL of a mixture of equal volumes of *nitric acid R* and *water R* and 0.5 mL of *silver nitrate solution R2*. Solution A is opalescent and a white precipitate is formed after a few minutes; solution B remains clear.

TESTS

Acidity or alkalinity

To 20 mL add 20 mL of *carbon dioxide-free water R*, shake for 3 min and allow to stand. Separate the aqueous layer and add 0.2 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.01 M sodium hydroxide* or 0.6 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Relative density (*2.2.5*)

1.872 to 1.877.

Distillation range (*2.2.11*)

It distils completely between 49.0 °C and 51.0 °C and 95 per cent distills within a range of 1.0 °C.

Volatile related substances

Gas chromatography (*2.2.28*).

Internal standard *trichlorotrifluoroethane CRS.*

Test solution (a) The substance to be examined.

Test solution (b) Dilute 1.0 mL of *trichlorotrifluoroethane CRS* to 20.0 mL with the substance to be examined. Dilute 1.0 mL of the solution to 100.0 mL with the substance to be examined. Dilute 1.0 mL of this solution to 10.0 mL with the substance to be examined.

Column:

— *size*: *l* = 2.75 m, Ø = 5 mm;

— *stationary phase*: *silanised diatomaceous earth for gas chromatography R1* (180-250 µm), the first 1.8 m being impregnated with 30 per cent *m/m* of *macrogol 400 R* and the remainder with 30 per cent *m/m* of *dinonyl phthalate R*;

— *temperature*: 50 °C.

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Detection Flame ionisation.

Injection 5 µL.

Limit Test solution (b):

— *total*: not more than the area of the peak due to the internal standard, corrected if necessary for any impurity with the same retention time as the internal standard (0.005 per cent).

Thymol

Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 0.10 g of *menthol R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent.

Test solution To 20.0 mL of the substance to be examined add 5.0 mL of the internal standard solution.

Reference solution Dissolve 20.0 mg of *thymol R* in *methylene chloride R* and dilute to 100.0 mL with the same solvent. To 20.0 mL of this solution, add 5.0 mL of the internal standard solution.

Column:

— *material*: fused silica;

— *size*: *l* = 15 m, Ø = 0.53 mm;

— *stationary phase*: *poly(dimethyl)siloxane R* (film thickness 1.5 µm).

Carrier gas *nitrogen for chromatography R.*

Flow rate 15 mL/min.

Temperature:

— *column*: 150 °C;

— *injection port*: 170 °C;

— *detector*: 200 °C.

Detection Flame ionisation.

Injection 1.0 µL.

Limit:

— *thymol*: 0.75 times to 1.15 times the area of the corresponding peak in the chromatogram obtained with the reference solution (0.008 per cent *m/m* to 0.012 per cent *m/m*).

Bromides and chlorides

To 10 mL add 20 mL of *water R* and shake for 3 min. To 5 mL of the aqueous layer add 5 mL of *water R*, 0.05 mL of *nitric acid R* and 0.2 mL of *silver nitrate solution R1*. The solution is not more opalescent than a mixture of 5 mL of the aqueous layer and 5 mL of *water R*.

Bromine and chlorine

To 10 mL of the aqueous layer obtained in the test for bromides and chlorides add 1 mL of *potassium iodide and starch solution R*. No blue colour is produced.

Non-volatile matter

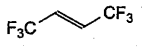
Maximum 20 mg/L.

Evaporate 50 mL to dryness on a water-bath and dry the residue in an oven at 100-105 °C for 2 h. The residue weighs a maximum of 1 mg.

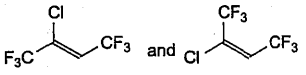
STORAGE

In an airtight container, protected from light, at a temperature not exceeding 25 °C. The choice of material for the container is made taking into account the particular reactivity of halothane with certain metals.

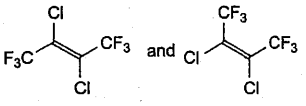
IMPURITIES



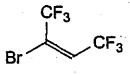
A. (*E*)-1,1,1,4,4,4-hexafluorobut-2-ene,



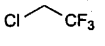
B. (*EZ*)-2-chloro-1,1,1,4,4,4-hexafluorobut-2-ene (*cis* and *trans*),



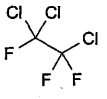
C. (*EZ*)-2,3-dichloro-1,1,1,4,4,4-hexafluorobut-2-ene (*cis* and *trans*),



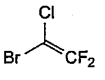
D. (*E*)-2-bromo-1,1,1,4,4,4-hexafluorobut-2-ene,



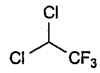
E. 2-chloro-1,1,1-trifluoroethane,



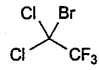
F. 1,1,2-trichloro-1,2,2-trifluoroethane,



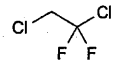
G. 1-bromo-1-chloro-2,2-difluoroethene,



H. 2,2-dichloro-1,1,1-trifluoroethane,



I. 1-bromo-1,1-dichloro-2,2,2-trifluoroethane,



J. 1,2-dichloro-1,1-difluoroethane.

Ph Eur

Helium

(Ph. Eur. monograph 2155)

He 4.00

Ph Eur

DEFINITION

Content

Minimum 99.5 per cent *V/V* of He.

This monograph applies to helium obtained by separation from natural gas and intended for medicinal use.

CHARACTERS

Appearance

Colourless, inert gas.

IDENTIFICATION

Examine the chromatograms obtained in the assay.
The retention time of the principal peak in the chromatogram obtained with the substance to be examined is approximately the same as that of the principal peak in the chromatogram obtained with the reference gas.

TESTS

Methane

Maximum 50.0 ppm *V/V*.

Infrared analyser.

Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena (3 µm filter).

Reference gas (a) *helium for chromatography R.*

Reference gas (b) Mixture containing 50.0 ppm *V/V* of *methane R* in *helium for chromatography R.*

The infrared analyser generally comprises an infrared source emitting broadband infrared radiation, an optical device, a sample cell, a detector and in some analysers a reference cell. The optical device may be positioned either before or after the sample cell. It consists of one or more optical filters, through which the broadband radiation is passed. The optical device is selected for methane determination.
The measurement light beam passes through the sample cell and may also pass through a reference cell if the analyser integrates such a feature. When methane is present in the sample cell, absorption of energy in the measurement light beam will occur according to the Beer-Lambert law, and this produces a change in the detector signal. This measurement signal is compared to a reference signal to generate an output related to the concentration of methane. The generated signal is linearised in order to determine the methane content.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the methane content in the gas to be examined.

Oxygen

Maximum 50.0 ppm *V/V*, determined using an oxygen analyser equipped with an electrochemical cell and a detector scale ranging from 0-100 ppm *V/V*.

The gas to be examined passes through a detection cell containing an aqueous solution of an electrolyte, generally potassium hydroxide. The presence of oxygen in the gas to be examined produces a variation in the electric signal recorded at the outlet of the cell that is proportional to the oxygen content.

Calibrate the analyser according to the instructions of the manufacturer. Pass the gas to be examined through the analyser using a suitable pressure regulator and airtight metal tubes and operating at the prescribed flow rates until constant readings are obtained.

Water *(2.5.28)*

Maximum 67 ppm *V/V*.

ASSAY

Gas chromatography *(2.2.28)*.

Gas to be examined The substance to be examined.

Reference gas *helium for chromatography R.*

Column:
— *size*: *l* = 2 m, Ø = 4.5 mm;
— *stationary phase*: *molecular sieve for chromatography R* (0.5 nm).

Carrier gas *argon for chromatography R.*

Flow rate 60 mL/min.

Temperature:
— *column*: 50 °C;
— *detector*: 150 °C.

Detection Thermal conductivity.

Injection 0.5 mL.

Inject the reference gas. Adjust the injected volumes and operating conditions so that the height of the peak due to helium in the chromatogram obtained is at least 35 per cent of the full scale of the recorder.

System suitability Reference gas:
— *symmetry factor*: minimum 0.6.

Calculate the content of He in the gas to be examined.

STORAGE

As compressed gas or liquid at cryogenic temperature, in appropriate containers, complying with the legal regulations.

IMPURITIES

Specified impurities *A, B, C.*

A. CH_4: methane,

B. O_2: oxygen,

C. H_2O: water.

Ph Eur

Heparin Calcium

(Ph. Eur. monograph 0332)

Action and use
Anticoagulant.

Preparation
Heparin Injection

Ph Eur

DEFINITION

Preparation containing the calcium salt of a sulfated glycosaminoglycan present in mammalian tissues. It is prepared from the intestinal mucosae of pigs. On complete hydrolysis, it liberates D-glucosamine, D-glucuronic acid, L-iduronic acid, acetic acid and sulfuric acid. It has the property of delaying the clotting of blood by catalysing the inhibition of thrombin and factor Xa by antithrombin.

Potency
Minimum 180 IU/mg (dried substance), determined by the assay of anti-factor IIa activity as described under Assay.

PRODUCTION

The animals from which heparin calcium is derived must fulfil the requirements for the health of animals suitable for human consumption. All stages of production and sourcing are subjected to a suitable quality management system. The identity of the source species and the absence of material from possible contaminant species such as cattle, sheep and goats, is verified by appropriate testing during production. The method used to confirm identity of the source species, and the point of application in the process, have been validated and shown to be capable of identifying the presence of material of other species at the level of 0.1 per cent (*m/m*). Species verification by methods based on polymerase chain reaction (PCR) amplification of species-specific DNA sequences has been widely shown to be an appropriate surrogate. If such a method has been chosen, it is also used to test for porcine DNA and to determine that it is present at a consistent level, in line with the manufacturing process used.

It is produced by methods of manufacturing designed to minimise or eliminate substances lowering blood pressure.

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water.

IDENTIFICATION

A. It complies with the requirements described under Assay.

B. Carry out the assay of anti-factor Xa activity of heparin (*2.7.5*). The ratio of anti-factor Xa activity to anti-factor IIa activity determined as described under Assay, ranges between 0.9 and 1.1.

C. Nuclear magnetic resonance spectrometry (*2.2.33*).

Preparation Dissolve 20 mg of the substance to be examined in 0.7 mL of a 20 µg/mL solution of *deuterated sodium trimethylsilylpropionate R* in *deuterium oxide R*.

Comparison Dissolve 20 mg of *heparin calcium for NMR identification CRS* in 0.7 mL of a 20 µg/mL solution of *deuterated sodium trimethylsilylpropionate R* in *deuterium oxide R*.

Apparatus Spectrometer operating at minimum 300 MHz.

Acquisition of ^{1}H-NMR spectra:
- *number of transients*: minimum 16; it is adjusted until the signal-to-noise ratio is at least 1000:1 for the heparin methyl signal at 2.04 ppm;
- *temperature*: about 25 °C; test sample and reference spectra have to be obtained at the same temperature;
- *acquisition time*: minimum 2 s;
- *repetition time* (acquisition time plus delay): minimum 4 s;
- *spectral width*: 10-12 ppm, centred at around 4.5 ppm;
- *pulse width*: to give a flip angle between 30° and 90°.

Processing:
- *exponential line-broadening window function*: 0.3 Hz;
- Fourier transformation;
- trimethylsilylpropionate reference signal set at 0.00 ppm.

Results:
- the large heparin calcium signals must be present: 2.05 ppm, 3.29 ppm (doublet), 4.37 ppm, 5.35 ppm and 5.43 ppm, all within ± 0.03 ppm;
- the ^{1}H-NMR spectrum obtained with the test sample and that obtained with *heparin calcium for NMR identification CRS* are compared qualitatively after the 2 spectra have been normalised so as to have a similar intensity; dermatan sulfate with a methyl signal at 2.08 ± 0.02 ppm may be observed; no unidentified signals larger than 4 per cent compared to the height of the heparin signal at 5.43 ppm are present in the ranges 0.10-2.00 ppm, 2.10-3.10 ppm and 5.70-8.00 ppm; signals from the solvent or process-related substances may be present and have to be identified to be accepted.

D. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (a) and reference solution (c).

Relative retention With reference to heparin (retention time = about 26 min): dermatan sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate = about 1.3.

System suitability Reference solution (c):
- *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the peak due to dermatan sulfate and chondroitin sulfate and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to heparin.

Results The principal peak in the chromatogram obtained with test solution (a) is similar in retention time and shape to the principal peak in the chromatogram obtained with reference solution (c).

E. It gives the reactions of calcium (*2.3.1*).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Dissolve a quantity equivalent to 50 000 IU in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)
5.5 to 8.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Nucleotidic impurities
Dissolve 40 mg in 10 mL of *water R*. The absorbance (*2.2.25*) measured at 260 nm is not greater than 0.15.

Protein
Maximum 0.5 per cent (dried substance).

Solution A Mix 2 volumes of a 10 g/L solution of *sodium hydroxide R* and 2 volumes of a 50 g/L solution of *sodium carbonate R* and dilute to 5 volumes with *water R*.

Solution B Mix 2 volumes of a 12.5 g/L solution of *copper sulfate pentahydrate R* and 2 volumes of a 29.8 g/L solution of *sodium tartrate R* and dilute to 5 volumes with *water R*.

Solution C Mix 1 volume of solution B and 50 volumes of solution A.

Solution D Dilute a phosphomolybdotungstic reagent in *water R*. Suitable dilutions produce solutions of pH 10.25 ± 0.25 after addition of solutions C and D to the test and reference solutions.

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of 5 mg/mL.

Reference solutions Dissolve *bovine albumin R1* in *water R* to obtain a concentration of 100 mg/mL. Prepare dilutions of the solution in *water R* as prescribed in general chapter *2.5.33, method 2*.

Blank *water R*.

Procedure To 1 mL of each reference solution, of the test solution and of the blank, add 5 mL of solution C. Allow to stand for 10 min. Add 0.5 mL of solution D, mix and allow to stand at room temperature for 30 min. Filter through a membrane filter (nominal pore size 0.45 µm). Determine the absorbances (*2.2.25*) of the solutions at 750 nm, using the solution prepared from the blank as compensation liquid.

Calculations As prescribed in general chapter *2.5.33, method 2*.

Related substances
Liquid chromatography (*2.2.29*). *Reference solutions are stable at room temperature for 24 h.*

Test solution (a) Dissolve an accurately weighed quantity of about 50 mg of the substance to be examined in 5.0 mL of *water for chromatography R*. Mix using a vortex mixer until dissolution is complete.

Test solution (b) Dissolve an accurately weighed quantity of about 0.1 g of the substance to be examined in 1.0 mL of *water for chromatography R*. Mix using a vortex mixer until dissolution is complete. Mix 500 µL of the solution and 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Reference solution (a) Dissolve 250 mg of *heparin for physico-chemical analysis CRS* in *water for chromatography R* and dilute to 2.0 mL with the same solvent. Mix using a vortex mixer until dissolution is complete.

Reference solution (b) Add 1200 µL of reference solution (a) to 300 µL of *dermatan sulfate and over-sulfated chondroitin sulfate CRS*. Mix using a vortex mixer to homogenise.

Reference solution (c) Add 100 µL of reference solution (b) to 900 µL of *water for chromatography R*. Mix using a vortex mixer to homogenise.

Reference solution (d) Add 400 µL of reference solution (a) to 100 µL of *water for chromatography R* and mix using a vortex mixer. Add 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Reference solution (e) To 500 µL of reference solution (b), add 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Precolumn:
— *size*: *l* = 0.05 m, Ø = 2 mm;
— *stationary phase*: *anion-exchange resin R* (13 µm).

Column:
— *size*: *l* = 0.25 m, Ø = 2 mm;
— *stationary phase*: *anion-exchange resin R* (9 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R* and adjust to pH 3.0 with *dilute phosphoric acid R*;
— *mobile phase B*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R*, add 140 g of *sodium perchlorate R* and adjust to pH 3.0 with *dilute phosphoric acid R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	75	25
10 - 35	75 → 0	25 → 100
35 - 40	0	100

Flow rate 0.22 mL/min.

Detection Spectrophotometer at 202 nm.

Equilibration At least 15 min.

Injection 20 µL of test solution (b) and reference solutions (d) and (e).

Relative retention With reference to heparin (retention time = about 26 min): dermatan sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate = about 1.3.

System suitability:
— the chromatogram obtained with reference solution (d) shows no peak at the retention time of heparin;
— *resolution*: minimum 3.0 between the peak due to dermatan sulfate and chondroitin sulfate and the peak due to over-sulfated chondroitin sulfate in the chromatogram obtained with reference solution (e).

Limits:
— *sum of dermatan sulfate and chondroitin sulfate*: not more than the area of the peak due to dermatan sulfate and chondroitin sulfate in the chromatogram obtained with reference solution (e) (2.0 per cent);
— *any other impurity*: no peak with an area greater than 0.01 times the area of the peak due to dermatan sulfate and chondroitin sulfate in the chromatogram obtained with reference solution (e) is detected (corresponding to a disregard limit of 0.02 per cent).

Nitrogen (*2.5.9*)
1.5 per cent to 2.5 per cent (dried substance), determined on 0.100 g.

Calcium
9.5 per cent to 11.5 per cent (dried substance), determined on 0.200 g by complexometric titration (*2.5.11*).

Loss on drying (*2.2.32*)
Maximum 8.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 670 Pa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.01 IU per International Unit of heparin, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. The addition of divalent cations may be necessary in order to fulfil the validation criteria.

ASSAY
Carry out the assay of anti-factor IIa activity of heparin (*2.7.5*). The estimated potency is not less than 90 per cent and not more than 111 per cent of the stated potency. The confidence limits of the estimated potency (P = 0.95) are not less than 80 per cent and not more than 125 per cent of the stated potency.

STORAGE
In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

LABELLING
The label states the number of International Units per milligram.

Ph Eur

Heparin Sodium

(Ph. Eur. monograph 0333)

Action and use
Anticoagulant.

Preparations
Heparin Flush Solution
Heparin Injection

Ph Eur

DEFINITION
Preparation containing the sodium salt of a sulfated glycosaminoglycan present in mammalian tissues. It is prepared from the intestinal mucosae of pigs. On complete hydrolysis, it liberates D-glucosamine, D-glucuronic acid, L-iduronic acid, acetic acid and sulfuric acid. It has the property of delaying the clotting of blood by catalysing the inhibition of thrombin and factor Xa by antithrombin.

Potency
Minimum 180 IU/mg (dried substance), determined by the assay of anti-factor IIa activity as described under Assay.

PRODUCTION
The animals from which heparin sodium is derived must fulfil the requirements for the health of animals suitable for human consumption. All stages of production and sourcing are subjected to a suitable quality management system. The identity of the source species and the absence of material from possible contaminant species such as cattle, sheep and goats, is verified by appropriate testing during production. The method used to confirm identity of the source species, and the point of application in the process, have been validated and shown to be capable of identifying the presence of material of other species at the level of 0.1 per cent (*m/m*). Species verification by methods based on polymerase chain reaction (PCR) amplification of species-specific DNA sequences has been widely shown to be an appropriate surrogate. If such a method has been chosen, it is also used to test for porcine DNA and to determine that it is present at a consistent level, in line with the manufacturing process used.

It is produced by methods of manufacturing designed to minimise or eliminate substances lowering blood pressure.

CHARACTERS
Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water.

IDENTIFICATION
A. It complies with the requirements described under Assay.

B. Carry out the assay of anti-factor Xa activity of heparin (*2.7.5*). The ratio of anti-factor Xa activity to anti-factor IIa activity determined as described under Assay, ranges between 0.9 and 1.1.

C. Nuclear magnetic resonance spectrometry (*2.2.33*).

Solution A A solution in *deuterium oxide R* containing 20 µg/mL of *deuterated sodium trimethylsilylpropionate R* and if the signal at 5.22 ppm is smaller than 80 per cent of the signal at 5.44 ppm, 12 µg/mL of *sodium edetate R*.

Preparation Dissolve 20 mg of the substance to be examined in 0.7 mL of solution A.

Comparison Dissolve 20 mg of *heparin sodium for NMR identification CRS* in 0.7 mL of solution A.

If stored, the sodium edetate and deuterated sodium trimethylsilylpropionate solutions must be kept in high-density, natural polyethylene bottles.

Apparatus Spectrometer operating at minimum 300 MHz.

Acquisition of ^{1}H-NMR spectra:
- *number of transients*: minimum 16; it is adjusted until the signal-to-noise ratio is at least 1000:1 for the heparin methyl signal at 2.04 ppm;
- *temperature*: about 25 °C; test sample and reference spectra have to be obtained at the same temperature;
- *acquisition time*: minimum 2 s;
- *repetition time* (acquisition time plus delay): minimum 4 s;
- *spectral width*: 10-12 ppm, centred at around 4.5 ppm;
- *pulse width*: to give a flip angle between 30° and 90°.

Processing:
- *exponential line-broadening window function*: 0.3 Hz;
- Fourier transformation;
- trimethylsilylpropionate reference signal set at 0.00 ppm.

Results:
- the large heparin sodium signals must be present: 2.04 ppm, 3.27 ppm (doublet), 4.34 ppm, 5.22 ppm and 5.42 ppm, all within ± 0.03 ppm;
- the ^{1}H-NMR spectrum obtained with the test sample and that obtained with *heparin sodium for NMR identification CRS* are compared qualitatively after the 2 spectra have been normalised so as to have a similar intensity; dermatan sulfate with a methyl signal at 2.08 ± 0.02 ppm may be observed; no unidentified signals larger than 4 per cent compared to the height of the heparin signal at 5.42 ppm are present in the ranges 0.10-2.00 ppm, 2.10-3.10 ppm and 5.70-8.00 ppm; signals from the solvent or process-related substances may be present and have to be identified to be accepted; variations in the intensity of

some signal regions of the spectrum of heparin may occur: the intensity-variable regions are between 3.35 ppm and 4.55 ppm, where the signal pattern is approximately kept but intensity varies.

D. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (a) and reference solution (c).

Relative retention With reference to heparin (retention time = about 26 min): dermatan sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate = about 1.3.

System suitability Reference solution (c):

— *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the peak due to dermatan sulfate and chondroitin sulfate and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to heparin.

Results The principal peak in the chromatogram obtained with test solution (a) is similar in retention time and shape to the principal peak in the chromatogram obtained with reference solution (c).

E. It complies with the test for sodium (see Tests).

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Dissolve a quantity equivalent to 50 000 IU in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)

5.5 to 8.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Nucleotidic impurities

Dissolve 40 mg in 10 mL of *water R*. The absorbance (*2.2.25*) measured at 260 nm is not greater than 0.15.

Protein

Maximum 0.5 per cent (dried substance).

Solution A Mix 2 volumes of a 10 g/L solution of *sodium hydroxide R* and 2 volumes of a 50 g/L solution of *sodium carbonate R* and dilute to 5 volumes with *water R*.

Solution B Mix 2 volumes of a 12.5 g/L solution of *copper sulfate pentahydrate R* and 2 volumes of a 29.8 g/L solution of *sodium tartrate R* and dilute to 5 volumes with *water R*.

Solution C Mix 1 volume of solution B and 50 volumes of solution A.

Solution D Dilute a phosphomolybdotungstic reagent in *water R*. Suitable dilutions produce solutions of pH 10.25 ± 0.25 after addition of solutions C and D to the test and reference solutions.

Test solution Dissolve the substance to be examined in *water R* to obtain a concentration of 5 mg/mL.

Reference solutions Dissolve *bovine albumin R1* in *water R* to obtain a concentration of 100 mg/mL. Prepare dilutions of the solution in *water R* as prescribed in general chapter *2.5.33, method 2*.

Blank *water R*.

Procedure To 1 mL of each reference solution, of the test solution and of the blank, add 5 mL of solution C. Allow to stand for 10 min. Add 0.5 mL of solution D, mix and allow to stand at room temperature for 30 min. Determine the absorbances (*2.2.25*) of the solutions at 750 nm, using the solution prepared from the blank as compensation liquid.

Calculations As prescribed in general chapter *2.5.33, method 2*.

Related substances

Liquid chromatography (*2.2.29*). *Reference solutions are stable at room temperature for 24 h.*

Test solution (a) Dissolve an accurately weighed quantity of about 50 mg of the substance to be examined in 5.0 mL of *water for chromatography R*. Mix using a vortex mixer until dissolution is complete.

Test solution (b) Dissolve an accurately weighed quantity of about 0.1 g of the substance to be examined in 1.0 mL of *water for chromatography R*. Mix using a vortex mixer until dissolution is complete. Mix 500 µL of the solution and 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Reference solution (a) Dissolve 250 mg of *heparin for physico-chemical analysis CRS* in *water for chromatography R* and dilute to 2.0 mL with the same solvent. Mix using a vortex mixer until dissolution is complete.

Reference solution (b) Add 1200 µL of reference solution (a) to 300 µL of *dermatan sulfate and over-sulfated chondroitin sulfate CRS*. Mix using a vortex mixer to homogenise.

Reference solution (c) Add 100 µL of reference solution (b) to 900 µL of *water for chromatography R*. Mix using a vortex mixer to homogenise.

Reference solution (d) Add 400 µL of reference solution (a) to 100 µL of *water for chromatography R* and mix using a vortex mixer. Add 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Reference solution (e) To 500 µL of reference solution (b), add 250 µL of *1 M hydrochloric acid*, then add 50 µL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to stand at room temperature for 40 min before adding 200 µL of *1 M sodium hydroxide* to stop the reaction.

Precolumn:

— *size*: *l* = 0.05 m, Ø = 2 mm;

— *stationary phase*: *anion-exchange resin R* (13 µm).

Column:

— *size*: *l* = 0.25 m, Ø = 2 mm;

— *stationary phase*: *anion-exchange resin R* (9 µm);

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R* and adjust to pH 3.0 with *dilute phosphoric acid R*;

— *mobile phase B*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R*, add 140 g of *sodium perchlorate R* and adjust to pH 3.0 with *dilute phosphoric acid R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 10	75	25
10 - 35	75 → 0	25 → 100
35 - 40	0	100

Flow rate 0.22 mL/min.
Detection Spectrophotometer at 202 nm.
Equilibration At least 15 min.
Injection 20 µL of test solution (b) and reference solutions (d) and (e).
Relative retention With reference to heparin (retention time = about 26 min): dermatan sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate = about 1.3.
System suitability:
— the chromatogram obtained with reference solution (d) shows no peak at the retention time of heparin;
— *resolution*: minimum 3.0 between the peak due to dermatan sulfate and chondroitin sulfate and the peak due to over-sulfated chondroitin sulfate in the chromatogram obtained with reference solution (e).
Limits:
— *sum of dermatan sulfate and chondroitin sulfate*: not more than the area of the peak due to dermatan sulfate and chondroitin sulfate in the chromatogram obtained with reference solution (e) (2.0 per cent);
— *any other impurity*: no peak with an area greater than 0.01 times the area of the peak due to dermatan sulfate and chondroitin sulfate in the chromatogram obtained with reference solution (e) is detected (corresponding to a disregard limit of 0.02 per cent).

Nitrogen (*2.5.9*)
1.5 per cent to 2.5 per cent (dried substance), determined on 0.100 g.

Sodium
10.5 per cent to 13.5 per cent (dried substance).
Atomic absorption spectrometry (*2.2.23, Method I*).
Test solution Dissolve 50 mg of the substance to be examined in a 1.27 mg/mL solution of *caesium chloride R* in *0.1 M hydrochloric acid* and dilute to 100.0 mL with the same solvent.
Reference solutions Prepare reference solutions containing 25 ppm, 50 ppm and 75 ppm of Na, using *sodium standard solution (200 ppm Na) R* diluted with a 1.27 mg/mL solution of *caesium chloride R* in *0.1 M hydrochloric acid*.
Source Sodium hollow-cathode lamp.
Wavelength 330.3 nm.
Atomisation device Flame of suitable composition (for example 11 L of air and 2 L of acetylene per minute).

Loss on drying (*2.2.32*)
Maximum 8.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 670 Pa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.01 IU per International Unit of heparin, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Carry out the assay of anti-factor IIa activity of heparin (*2.7.5*). The estimated potency is not less than 90 per cent and not more than 111 per cent of the stated potency. The confidence limits of the estimated potency ($P = 0.95$) are not less than 80 per cent and not more than 125 per cent of the stated potency.

STORAGE
In an airtight container. If the substance is sterile, the container is also sterile and tamper-proof.

LABELLING
The label states the number of International Units per milligram.

Ph Eur

Low-molecular-weight Heparins

(Low-molecular-mass Heparins, Ph. Eur. monograph 0828)

Action and use
Anticoagulant.

Ph Eur

DEFINITION
Salts of sulfated glycosaminoglycans having a mass-average relative molecular mass less than 8000 and for which at least 60 per cent of the total mass has a relative molecular mass less than 8000. Low-molecular-mass heparins display different chemical structures at the reducing, or the non-reducing end of the polysaccharide chains.
The potency is not less than 70 IU of anti-factor Xa activity per milligram, calculated with reference to the dried substance. The ratio of anti-factor Xa activity to anti-factor IIa activity, determined as described under Assay, is not less than 1.5.

PRODUCTION
Low-molecular-mass heparins are obtained by fractionation or depolymerisation of heparin of natural origin that complies with the monograph *Heparin sodium (0333)* or *Heparin calcium (0332)*, whichever is appropriate, unless otherwise justified and authorised. For each type of low-molecular-mass heparin the batch-to-batch consistency is ensured by demonstrating, for example, that the mass-average relative molecular mass and the mass percentage within defined relative molecular-mass ranges lower than 8000 are not less than 75 per cent and not more than 125 per cent of the mean value stated as type specification. The same limits apply also to the ratio of anti-factor Xa activity to anti-factor IIa activity.

CHARACTERS
Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water.

IDENTIFICATION
A. Nuclear magnetic resonance spectrometry (*2.2.33*).
Preparation Dissolve 0.200 g of the substance to be examined in a mixture of 0.2 mL of *deuterium oxide R* and 0.8 mL of *water R*.
Comparison Dissolve 0.200 g of the appropriate specific low-molecular-mass heparin reference standard in a mixture of 0.2 mL of *deuterium oxide R* and 0.8 mL of *water R*.
Operating conditions:
— *field strength*: 75 MHz;
— *temperature*: 40 °C;
— *cell diameter*: 5 mm.
Processing:
— Fourier transformation;
— deuterated methanol reference signal set at 50.0 ppm.

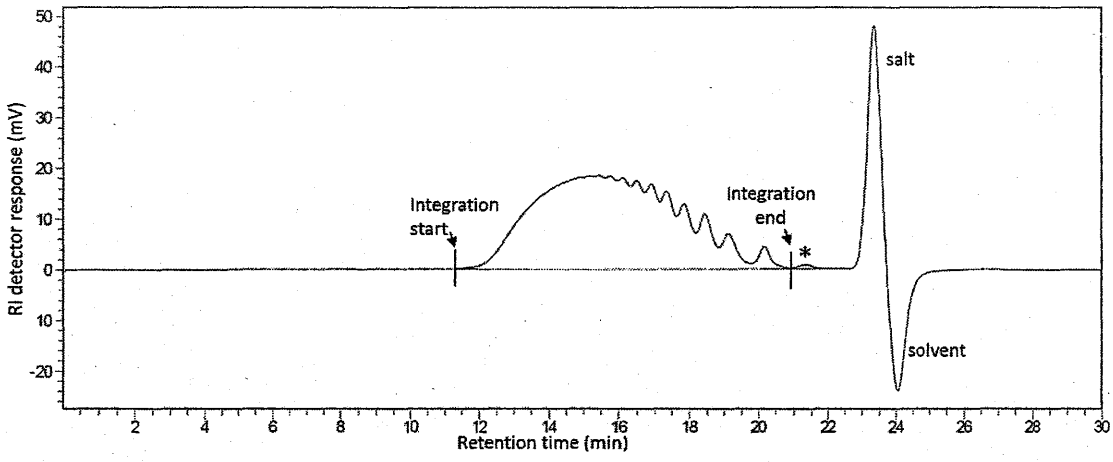


Figure 0828.-1. – *Example chromatogram of a low-molecular-mass heparin sample analysed using the method described in identification test C indicating peak integration*

Results The ^{13}C NMR spectrum obtained is similar to that obtained with the appropriate specific low-molecular-mass heparin reference standard.

B. The ratio of anti-factor Xa activity to anti-factor IIa activity, determined as described under Assay, is not less than 1.5.

C. Size-exclusion chromatography (*2.2.30*).

Test solution Dissolve 20 mg of the substance to be examined in 2 mL of the mobile phase.

Reference solution Dissolve 20 mg of *heparin low-molecular-mass for calibration CRS* in 2 mL of the mobile phase.

Column:

- *size*: l = 0.30 m, Ø = 7.8 mm;
- *stationary phase*: appropriate porous silica beads (5 µm) with a fractionation range for proteins of approximately 15 000 to 100 000;
- *number of theoretical plates*: minimum of 20 000 per metre.

Mobile phase 7.7 g/L solution of *ammonium acetate R*.

Flow rate 0.5 mL/min.

Detection Differential refractometer.

Injection 25 µL.

Calibration Inject 25 µL of the reference solution and record the chromatogram for a period of time, ensuring complete elution of sample, salt and solvent peaks.

Calculate the total area under the RI curve (designated ΣRI) by numerical integration over the range of interest (i.e. excluding salt and solvent peaks at the end of the chromatogram as well as the artefact peak (designated * on the chromatogram), that if present elutes at the same retention time as the peak that would be obtained after injection of sodium sulfate) as shown in Figure 0828.-1.

Calculate the area under the curve up to each point as a percentage of the total area.

Using the Broad Standard Table provided in the leaflet supplied with *heparin low-molecular-mass for calibration CRS*, identify those points in the chromatogram for which the percentage of the total area under the curve is closest to the percentage mass fractions listed in the Broad Standard Table, and assign the molecular mass in the table to the corresponding retention time in the chromatogram.

A calibration curve for the chromatographic system is derived by fitting a suitable mathematical relationship to the set of retention times and logarithms of the molecular masses. A polynomial of the 3rd degree is recommended.

Table 0828.-1. – *Broad Standard Table for heparin low-molecular-mass for calibration CRS*

Point	$\log_{10}(M)$	Molecular mass (M)	mass % > M
1	2.78	600	99.60
2	3.08	1200	96.13
3	3.26	1800	91.06
4	3.38	2400	85.51
5	3.48	3000	79.32
6	3.56	3600	72.80
7	3.62	4200	66.11
8	3.68	4800	59.51
9	3.73	5400	53.17
10	3.78	6000	47.08
11	3.82	6600	41.41
12	3.86	7200	36.11
13	3.92	8400	27.04
14	3.98	9600	19.91
15	4.08	12000	10.79
16	4.13	13600	7.04
17	4.19	15600	4.05
18	4.26	18000	2.23

Inject 25 µL of the test solution and record the chromatogram for a period of time, ensuring complete elution of sample, salt and solvent peaks.

The mass-average relative molecular mass is defined by the following expression:

$$\frac{\sum(RI_i M_i)}{\sum RI_i}$$

RI_i = refractive index of the substance eluting in the fraction i (proportional to the mass of the substance);
M_i = relative molecular mass corresponding to fraction i.

Any low-molecular-mass heparin covered by an individual monograph complies with the requirements for molecular mass parameters assessed in identification test C in the corresponding monograph.

Where no individual monograph exists for the low-molecular-mass heparin to be examined, the mass-average relative molecular mass is not greater than 8000 and at least 60 per cent of the total mass has a relative molecular mass lower than 8000. In addition, the molecular mass parameters (mass-average molecular mass and mass percentages of chains comprised between specified values) correspond to those of the manufacturer's reference preparation.

D. It gives reaction (a) of sodium or the reactions of calcium (as appropriate) (*2.3.1*).

TESTS

pH (*2.2.3*)
5.5 to 8.0.

Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Nitrogen (*2.5.9*)
1.5 per cent to 2.5 per cent (dried substance).

Calcium (*2.5.11*)
9.5 per cent to 11.5 per cent (dried substance), if prepared from heparin complying with the monograph *Heparin calcium (0332)*. Use 0.200 g.

Sodium
10.5 per cent to 13.5 per cent (dried substance), if prepared from heparin complying with the monograph *Heparin sodium (0333)*.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 50 mg in *0.1 M hydrochloric acid* containing 1.27 mg of *caesium chloride R* per millilitre and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare reference solutions (25 ppm, 50 ppm and 75 ppm) using *sodium standard solution (200 ppm Na) R* diluted with *0.1 M hydrochloric acid* containing 1.27 mg of *caesium chloride R* per millilitre.

Source Sodium hollow-cathode lamp.

Wavelength 330.3 nm.

Atomisation device Flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Molar ratio of sulfate ions to carboxylate ions (*2.2.38*)
Minimum 1.8.

The sample of heparin used in this titration must be free from ionisable impurities, particularly salts.

Weigh 0.100 g of the substance to be examined taking the necessary measures to avoid the problems linked to hygroscopicity.

Take up into about 20 mL of double-glass-distilled *water R*. Cool to 4 °C and apply 2.0 mL of this solution to a pre-cooled column (approximately 10 × 1 cm), packed with a suitable *cation-exchange resin R*. Wash through with double-glass-distilled *water R* into the titration vessel up to a final volume of about 10-15 mL (*the titration vessel must be just large enough to hold the electrodes from the conductivity meter, a small stirrer bar and a fine flexible tube from the outlet of a 2 mL burette*). Stir magnetically. When the conductivity reading is constant, note it and titrate with *0.05 M sodium hydroxide* added in approximately 50 µL portions. Record the burette level and the conductivity meter reading a few seconds after each addition until the end-point is reached.

For each measured figure, calculate the number of milliequivalents of sodium hydroxide added from the volume and the known concentration of the sodium hydroxide solution. Plot on a graph the figures for conductivity (as y-axis) against the figures of milliequivalent of sodium hydroxide (as x-axis). The graph will have 3 approximately linear sections: an initial steep downward slope, a middle slight rise and a final steep rise. Estimate the best straight lines through these 3 parts of the graph. At the points where the 1st and 2nd lines intersect, and where the 2nd and 3rd lines intersect, draw perpendiculars to the x-axis to estimate the milliequivalents of sodium hydroxide taken up by the sample at those points. The point where the 1st and 2nd lines intersect will give the number of milliequivalents of sodium hydroxide taken up by the sulfate groups, and the point where the 2nd and 3rd lines intersect will give the number of milliequivalents taken up by the sulfate and carboxylate groups together. The difference between the 2 will therefore give the number of milliequivalents taken up by the carboxylate groups.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying at 60 °C over *diphosphorus pentoxide R* at a pressure not exceeding 0.67 kPa for 3 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.01 IU per International Unit of anti-Xa activity, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. The addition of divalent cations may be necessary to fulfil the validation criteria.

ASSAY

The anticoagulant activity of low-molecular-mass heparins is determined *in vitro* by 2 assays which determine its ability to accelerate the inhibition of factor Xa (anti-Xa assay) and thrombin, factor IIa (anti-IIa assay), by antithrombin III.

The International Units for anti-Xa and anti-IIa activity are the activities contained in a stated amount of the International Standard for low-molecular-mass heparin.

Heparin low-molecular-mass for assay BRP, calibrated in International Units by comparison with the International Standard using the 2 assays given below, is used as reference preparation.

ANTI-FACTOR XA ACTIVITY

Reference and test solutions
Prepare 4 independent series of 4 dilutions each, of the substance to be examined and of the reference preparation of low-molecular-mass heparin in *tris(hydroxymethyl) aminomethane sodium chloride buffer solution pH 7.4 R*; the concentration range should be within 0.025 IU to 0.2 IU of anti-factor Xa activity per millilitre and the dilutions chosen should give a linear response when results are plotted as absorbance against log concentration.

Procedure
Label 16 tubes for the dilutions of the substance to be examined and 16 tubes for the dilutions of the reference preparation: T_1, T_2, T_3, T_4 for each of the 4 series of dilutions of the substance to be examined and S_1, S_2, S_3, S_4 for each of the 4 series of dilutions of the reference preparation. To each of the 32 tubes add 50 µL of *antithrombin III solution R1* and 50 µL of the appropriate dilution of the substance to be examined, or the reference preparation. After each addition, mix but do not allow

bubbles to form. Treating the tubes in 2 subsequent series in the order S_1, S_2, S_3, S_4, T_1, T_2, T_3, T_4, T_1, T_2, T_3, T_4, S_1, S_2, S_3, S_4, allow to equilibrate at 37 °C (water-bath or heating block) for 1 min and add to each tube 100 µL of *bovine factor Xa solution R*. Incubate for exactly 1 min and add 250 µL of *chromogenic substrate R1*. Stop the reaction after exactly 4 min by adding 375 µL of *acetic acid R*. Transfer the mixtures to semi-micro cuvettes and measure the absorbance (*2.2.25*) at 405 nm. Determine the blank amidolytic activity at the beginning and at the end of the procedure in a similar manner, using *tris(hydroxymethyl) aminomethane sodium chloride buffer solution pH 7.4 R* instead of the reference and test solutions; the 2 blank values do not differ significantly. Calculate the regression of the absorbance on log concentrations of the solutions of the substance to be examined and of the reference preparation of low-molecular-mass heparins and calculate the potency of the substance to be examined in International Units of anti-factor Xa activity per millilitre using the usual statistical methods for parallel-line assays.

ANTI-FACTOR IIA ACTIVITY

Reference and test solutions

Prepare 4 independent series of 4 dilutions each, of the substance to be examined and of the reference preparation of low molecular-mass heparin in *tris(hydroxymethyl) aminomethane sodium chloride buffer solution pH 7.4 R*; the concentration range should be within 0.015 IU to 0.075 IU of anti-factor IIa activity per millilitre and the dilutions chosen should give a linear response when results are plotted as absorbance against log concentration.

Procedure

Label 16 tubes for the dilutions of the substance to be examined and 16 tubes for the dilutions of the reference preparation: T_1, T_2, T_3, T_4 for each of the 4 series of dilutions of the substance to be examined and S_1, S_2, S_3, S_4 for each of the 4 series of dilutions of the reference preparation. To each of the 32 tubes add 50 µL of *antithrombin III solution R2* and 50 µL of the appropriate dilution of the substance to be examined or the reference preparation. After each addition, mix but do not allow bubbles to form. Treating the tubes in 2 subsequent series in the order S_1, S_2, S_3, S_4, T_1, T_2, T_3, T_4, T_1, T_2, T_3, T_4, S_1, S_2, S_3, S_4, allow to equilibrate at 37 °C (water-bath or heating block) for 1 min and add to each tube 100 µL of *human thrombin solution R*. Incubate for exactly 1 min and add 250 µL of *chromogenic substrate R2*. Stop the reaction after exactly 4 min by adding 375 µL of *acetic acid R*. Transfer the mixtures to semi-micro cuvettes and measure the absorbance (*2.2.25*) at 405 nm. Determine the blank amidolytic activity at the beginning and at the end of the procedure in a similar manner, using *tris(hydroxymethyl) aminomethane sodium chloride buffer solution pH 7.4 R* instead of the reference and test solutions; the 2 blank values do not differ significantly. Calculate the regression of the absorbance on log concentrations of the solutions of the substance to be examined and of the reference preparation of low-molecular-mass heparins, and calculate the potency of the substance to be examined in International Units of anti-factor IIa activity per millilitre using the usual statistical methods for parallel-line assays.

LABELLING

The label states:

— the number of International Units of anti-factor Xa activity per milligram;
— the number of International Units of anti-factor IIa activity per milligram;
— the mass-average molecular mass and the percentage of molecules within defined molecular mass ranges;
— where applicable, that the contents are the sodium salt;
— where applicable, that the contents are the calcium salt.

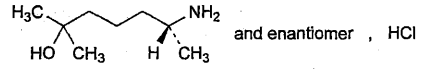
STORAGE

In an airtight tamper-proof container. If the product is sterile and free of bacterial endotoxins, store in a sterile and apyrogenic container.

Ph Eur

Heptaminol Hydrochloride

(Ph. Eur. monograph 1980)



$C_8H_{20}ClNO$ 181.7 *543-15-7*

Action and use

Non-selective phosphodiesterase inhibitor; treatment of reversible airways obstruction.

Ph Eur

DEFINITION

(6*RS*)-6-Amino-2-methylheptan-2-ol hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. To 1 mL of solution S (see Tests) add 4 mL of *water R* and 2 mL of a 200 g/L solution of *ammonium and cerium nitrate R* in *4 M nitric acid*. An orange-brown colour develops.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *heptaminol hydrochloride CRS*.

C. Examine the chromatograms obtained in the test for related substances.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (b).

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Acidity or alkalinity
To 10 mL of solution S add 0.1 mL of *methyl red solution R* and 0.3 mL of *0.01 M hydrochloric acid*. The solution is red. Add 0.6 mL of *0.01 M sodium hydroxide*. The solution is yellow.

Related substances
Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.50 g of the substance to be examined in *methanol R* and dilute to 5.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10 mL with *methanol R*.

Reference solution (a) Dilute 3.0 mL of test solution (a) to 10.0 mL with *methanol R*. Dilute 1.0 mL of this solution to 50.0 mL with *methanol R*.

Reference solution (b) Dissolve 0.10 g of *heptaminol hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution (c) Dissolve 10.0 mg of *heptaminol impurity A CRS* in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 10.0 mL with *methanol R*.

Reference solution (e) To 2.5 mL of reference solution (c) add 0.5 mL of test solution (b) and dilute to 5 mL with *methanol R*.

Plate *TLC silica gel G plate R*.

Mobile phase *concentrated ammonia R, dioxan R, 2-propanol R* (10:50:50 *V/V/V*).

Application 10 µL; apply test solutions (a) and (b) and reference solutions (a), (b), (d) and (e).

Development Over 2/3 of the plate.

Drying In air.

Detection Expose the plate to iodine vapour for at least 15 h.

System suitability The chromatogram obtained with reference solution (e) shows 2 clearly separated principal spots and the chromatogram obtained with reference solution (a) shows a single principal spot.

Limits In the chromatogram obtained with test solution (a):
— *impurity A*: any spot corresponding to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (d) (0.2 per cent),
— *any other impurity*: any spot, apart from the principal spot and any spot corresponding to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.6 per cent).

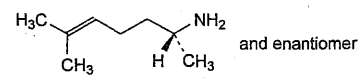
Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.140 g in 50 mL of *alcohol R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 18.17 mg of $C_8H_{20}ClNO$.

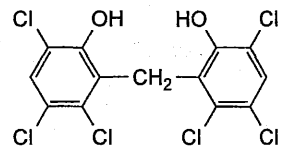
IMPURITIES



A. (2*RS*)-6-methylhept-5-en-2-amine.

Ph Eur

Hexachlorophene



$C_{13}H_6Cl_6O_2$ 406.9 *70-30-4*

Action and use
Antiseptic.

Preparation
Hexachlorophene Dusting Powder

DEFINITION
Hexachlorophene is 2,2-methylenebis(3,4,6-trichlorophenol). It contains not less than 98.0% and not more than 100.5% of $C_{13}H_6Cl_6O_2$, calculated with reference to the dried substance.

PRODUCTION
A suitable test is carried out to demonstrate that the level of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin present does not exceed 2 ppb.

CHARACTERISTICS
A white or pale buff, crystalline powder.

Practically insoluble in *water*; very soluble in *acetone*; freely soluble in *ethanol (96%)*. It dissolves in dilute solutions of the alkali hydroxides.

IDENTIFICATION
The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of hexachlorophene *(RS 174)*.

TESTS

Chloride
Dissolve 0.50 g in 2 mL of *ethanol (96%)*, dilute to 25 mL with *water* and filter. 5 mL of the clear filtrate diluted to 15 mL with *water* complies with the *limit test for chlorides*, Appendix VII (500 ppm).

Non-phenolic substances
Dissolve 5 g in 38 mL of *methanol*, add 125 mL of 0.25M *sodium hydroxide* and extract with three 15 mL quantities of n-*pentane*, retaining any foamy interphase with the aqueous layer. Dry the combined extracts over *anhydrous sodium sulfate* and evaporate to dryness at a pressure not exceeding 2 kPa. The residue weighs not more than 37.5 mg when dried to constant weight (0.75%).

Related substances
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions in *methanol*.

(1) 1.0% w/v of the substance being examined.

(2) 0.020% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (20 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography,* (10 µm) (Spherisorb ODS 1 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 300 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

1 volume of *glacial acetic acid,* 20 volumes of *water* and 100 volumes of *methanol.*

LIMITS

In the chromatogram obtained with solution (1):

the sum of the areas of any *secondary peaks* with a retention time not more than 3 times that of the principal peak is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (4.0%) and not more than one such peak has an area greater than half of the area of the principal peak in the chromatogram obtained with solution (2) (1.0%).

Loss on drying

When dried to constant weight at 105°, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

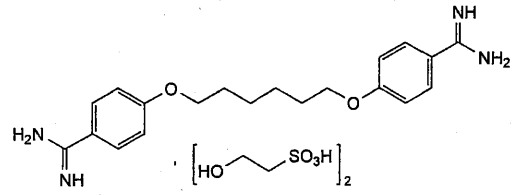
Dissolve 1.000 g in 25 mL of *ethanol (96%)* previously adjusted to pH 9.0 and titrate with *0.1M sodium hydroxide VS* determining the end point potentiometrically. Each mL of *0.1M sodium hydroxide VS* is equivalent to 40.69 mg of $C_{13}H_6Cl_6O_2$.

STORAGE

Hexachlorophene should be protected from light.

Hexamidine Isetionate

(Hexamidine Diisetionate, Ph. Eur. monograph 1436)



$C_{24}H_{38}N_4O_{10}S_2$ 607 *659-40-5*

Action and use

Antiprotozoal.

Ph Eur

DEFINITION

4,4′-[Hexane-1,6-diylbis(oxy)]dibenzimidamide bis(2-hydroxyethanesulfonate).

Content

98.5 per cent to 101.5 per cent (dried substance).

PRODUCTION

It is considered that alkyl 2-hydroxyethanesulfonate esters are potential impurities in hexamidine diisetionate.

The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation including, where necessary, demonstration that alkyl 2-hydroxyethanesulfonate esters are not detectable in the final product.

CHARACTERS

Appearance

White or slightly yellow powder, hygroscopic.

Solubility

Sparingly soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison hexamidine diisetionate CRS.

B. Dissolve about 40 mg in 5 mL of *water R* and add dropwise with shaking 1 mL of a 100 g/L solution of *sodium chloride R.* Allow to stand for 5 min. An abundant, shimmering white precipitate is slowly formed.

TESTS

Appearance of solution

Dissolve 0.50 g in *carbon dioxide-free water R,* heating at about 70 °C and dilute to 10 mL with the same solvent. Allow to cool to room temperature for 10-15 min.

The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (*2.2.2, Method II*).

Acidity or alkalinity

Dissolve 2.0 g in *water R* heating at about 50 °C and dilute to 20 mL with *water R* heating at about 50 °C. Allow to cool to about 35 °C, add 0.1 mL of *methyl red solution R.*

Not more than 0.25 mL of *0.05 M hydrochloric acid* or *0.05 M sodium hydroxide* is required to change the colour of the indicator.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 20.0 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 5 mg of the substance to be examined and 5 mg of *pentamidine diisetionate CRS* in mobile phase A and dilute to 100 mL with mobile phase A. Dilute 2 mL of the solution to 5 mL with mobile phase A.

Column:

— *size: l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase: styrene-divinylbenzene copolymer R* (8 µm).

Mobile phase:

— *mobile phase A*: mix 20 volumes of *acetonitrile R* and 80 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *phosphoric acid R,*

— *mobile phase B*: mix equal volumes of *acetonitrile R* and of a 6.8 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *phosphoric acid R,*

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	100 → 0	0 → 100
30 - 35	0	100
35 - 40	0 → 100	100 → 0

Flow rate 1 mL/min.

Detection Spectrophotometer at 263 nm.

Injection 20 µL.

Relative retention With reference to hexamidine (retention time = about 6 min): impurity B = about 1.7; impurity A = about 2.0; impurity C = about 3.7; impurity D = about 4.7.

System suitability Reference solution (c):
— *resolution*: minimum 5.0 between the peaks due to hexamidine and pentamidine.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *impurity B*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurities C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *any other impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

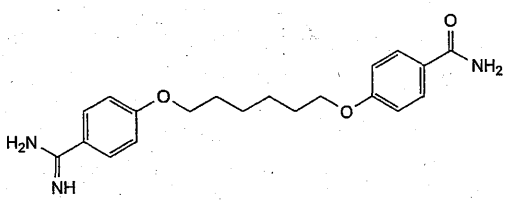
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.250 g in 50 mL of *dimethylformamide R*. Titrate with 0.1 M tetrabutylammonium hydroxide under a current of *nitrogen R*, determining the end-point potentiometrically (*2.2.20*).

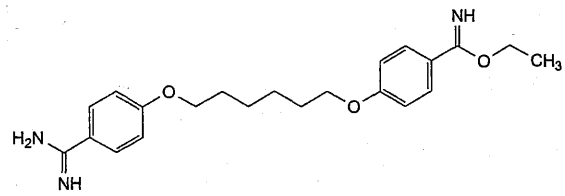
1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 30.35 mg of $C_{24}H_{38}N_4O_{10}S_2$.

STORAGE
In an airtight container.

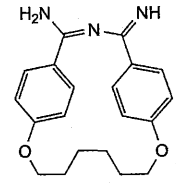
IMPURITIES
Specified impurities *A, B, C, D.*



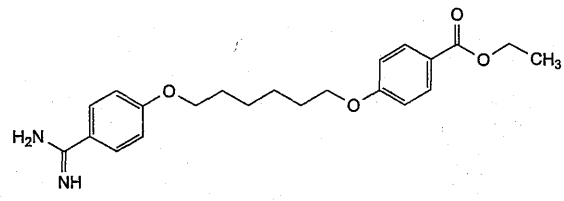
A. 4-[[6-(4-carbamimidoylphenoxy)hexyl]oxy]benzamide,



B. ethyl 4-[[6-(4-carbamimidoylphenoxy)hexyl] oxy]benzimidoate,



C. 4-imino-9,16-dioxa-3-azatricyclo[15.2.2.2^{5,8}]tricosa-1(19),2,5,7,17,20,22-heptaen-2-amine,

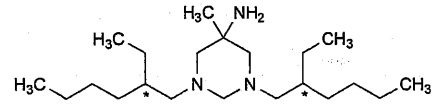


D. ethyl 4-[[6-(4-carbamimidoylphenoxy)hexyl]oxy]benzoate.

Ph Eur

Hexetidine

(Ph. Eur. monograph 1221)



$C_{21}H_{45}N_3$ 339.6 *141-94-6*

Action and use
Antiseptic.

Ph Eur

DEFINITION
Hexetidine contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of 1,3-bis(2-ethylhexyl)]-5-methylhexahydropyrimidin-5-amine.

CHARACTERS
An oily liquid, colourless or slightly yellow, very slightly soluble in water, very soluble in acetone, in alcohol and in methylene chloride. It dissolves in dilute mineral acids.

IDENTIFICATION
First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *hexetidine CRS*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour

and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 0.2 mL add 2 mL of *sulfuric acid R* and 2 mg of *chromotropic acid, sodium salt R*. Heat in a water-bath at 60 °C. A violet colour develops.

D. Dissolve 0.2 mL in 1 mL of *methylene chloride R*. Add 0.5 mL of *copper sulfate solution R*, 0.05 mL of *0.25 M alcoholic sulfuric acid R* and 5 mL of *water R*. Shake, then allow to stand. The lower layer becomes deep blue.

TESTS

Appearance
The substance to be examined is clear (*2.2.1*)and not more intensely coloured than reference solution Y_5 or reference solution GY_5 (*2.2.2, Method II*).

Relative density (*2.2.5*)
0.864 to 0.870.

Refractive index (*2.2.6*)
1.461 to 1.467.

Optical rotation (*2.2.7*)
Dissolve 1.0 g in *ethanol R* and dilute to 10.0 mL with the same solvent. The angle of optical rotation is −0.10° to + 0.10°.

Absorbance (*2.2.25*)
Dissolve 0.50 g in *heptane R* and dilute to 50.0 mL with the same solvent. At wavelengths from 270 nm to 350 nm, the absorbance of the solution is not greater than 0.1.

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel H R* as the coating substance. *Prepare the solutions immediately before use.*

Test solution (a) Dissolve 2.0 g of the substance to be examined in *heptane R* and dilute to 20 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with *heptane R*.

Reference solution (a) Dissolve 20 mg of *hexetidine CRS* in *heptane R* and dilute to 2 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (a) to 100 mL with *heptane R*.

Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with *heptane R*.

Reference solution (d) Dissolve 10 mg of *dehydrohexetidine CRS* in test solution (a) and dilute to 10 mL with the same solution.

Apply separately to the plate 1 µL of each solution. At the bottom of a chromatographic tank, place an evaporating dish containing *concentrated ammonia R1*. Place the dried plate in the tank and close the tank. Leave the plate in contact with the ammonia vapour for 15 min. Withdraw the plate and place it in a current of air to remove the ammonia vapour. Develop over a path of 15 cm using a mixture of 20 volumes of *methanol R* and 80 volumes of *toluene R*. Allow the plate to dry in air. Expose the plate to iodine vapour for 30 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1 per cent) and at most two such spots are more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (d) shows two clearly separated spots.

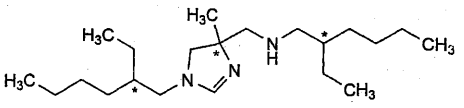
Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.150 g in 80 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

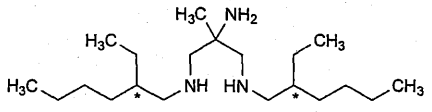
1 mL of *0.1 M perchloric acid* is equivalent to 16.98 mg of $C_{21}H_{45}N_3$.

STORAGE
Store protected from light.

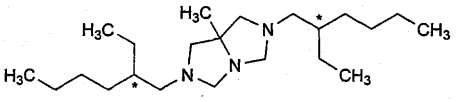
IMPURITIES



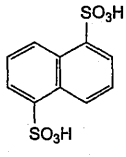
A. 2-ethyl-*N*-[[1-(2-ethylhexyl)-4-methyl-4,5-dihydro-1*H*-imidazol-4-yl]methyl]hexan-1-amine (dehydrohexetidine),



B. N^1,N^3-bis(2-ethylhexyl)-2-methylpropane-1,2,3-triamine (triamine),



C. 2,6-bis(2-ethylhexyl)-7a-methylhexahydro-1*H*-imidazo[1,5-*c*]imidazole (hexedine),

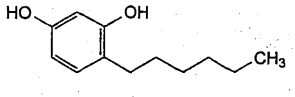


D. naphthalene-1,5-disulfonic acid.

Ph Eur

Hexylresorcinol

(Ph. Eur. monograph 1437)



$C_{12}H_{18}O_2$ 194.3 *136-77-6*

Action and use
Antihelminthic.

Ph Eur

DEFINITION
4-Hexylbenzene-1,3-diol.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Colourless, yellowish or reddish, crystalline powder or needles, turning brownish-pink on exposure to light or air.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 66 °C to 68 °C, melting may occur at about 60 °C, followed by solidification and a second melting between 66 °C and 68 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hexylresorcinol CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dilute 0.1 mL of solution S (see Tests) to 10 mL with *ethanol (96 per cent) R.*

Reference solution (a) Dissolve 10 mg of *hexylresorcinol CRS* in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *hexylresorcinol CRS* and 10 mg of *resorcinol R* in *ethanol (96 per cent) R*, then dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *methyl ethyl ketone R, pentane R* (50:50 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air for 5 min.

Detection Spray with 3 mL of *anisaldehyde solution R* and heat at 100-105 °C for 5 min.

System suitability Reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 1 mL of *ethanol (96 per cent) R.* Add one drop of *ferric chloride solution R1.* A green colour is produced. Add *dilute ammonia R1.* The solution changes to brown.

TESTS

Solution S

Dissolve 1.0 g in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*).

Acidity

Dissolve 0.5 g in a mixture of 25 mL of *carbon dioxide-free water R* and 25 mL of *ether R* previously neutralised to *phenolphthalein solution R1* and titrate with *0.1 M sodium hydroxide*, shaking vigorously after each addition. Not more than 0.4 mL is required to change the colour of the solution.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of *hexylresorcinol for system suitability CRS* (containing impurities C and D) in 1.0 mL of the mobile phase.

Column:

— *size*: l = 0.15 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);

— *temperature*: 30 °C.

Mobile phase Mix 30 volumes of a 3.0 g/L solution of *glacial acetic acid R* previously adjusted to pH 5.9 with *dilute ammonia R1*, and 70 volumes of *methanol R.*

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 281 nm.

Injection 20 µL.

Run time 2.5 times the retention time of hexylresorcinol.

Identification of impurities Use the chromatogram supplied with *hexylresorcinol for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and D.

Relative retention With reference to hexylresorcinol (retention time = about 10 min): impurity C = about 0.7; impurity D = about 1.1.

System suitability Reference solution (b):

— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to hexylresorcinol.

Calculation of percentage contents:

— *correction factor*: multiply the peak area of impurity D by 0.3;

— for each impurity, use the concentration of hexylresorcinol in reference solution (a).

Limits:

— *impurities C, D*: for each impurity, maximum 0.2 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;

— *total*: maximum 0.5 per cent;

— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 10 mL of *methanol R* in a ground-glass-stoppered flask, add 30.0 mL of *0.0167 M potassium bromate* and 2 g of *potassium bromide R.* Shake to dissolve the substance and add 15 mL of *dilute sulfuric acid R.* Stopper the flask, shake and allow to stand in the dark for 15 min, stirring continuously. Add 5 mL of *methylene chloride R* and a solution of 1 g of *potassium iodide R* in 10 mL of *water R*, allow to stand in the dark for 15 min, stirring continuously. Titrate with *0.1 M sodium thiosulfate*, using 1 mL of *starch solution R*, shaking thoroughly. Carry out a blank titration under the same conditions.

1 mL of *0.0167 M potassium bromate* is equivalent to 4.857 mg of $C_{12}H_{18}O_2$.

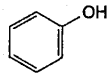
STORAGE

In an airtight container, protected from light.

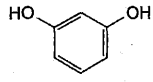
IMPURITIES

Specified impurities C, D.

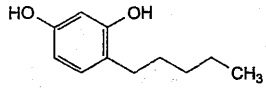
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A, B.



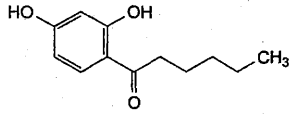
A. phenol,



B. benzene-1,3-diol (resorcinol),



C. 4-pentylbenzene-1,3-diol,

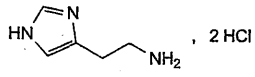


D. 1-(2,4-dihydroxyphenyl)hexan-1-one.

Ph Eur

Histamine Dihydrochloride

(Ph. Eur. monograph 0143)



$C_5H_{11}Cl_2N_3$ 184.1 *56-92-8*

Ph Eur

DEFINITION

Histamine dihydrochloride contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of 2-(1*H*-imidazol-4-yl)ethan-1-amine dihydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, hygroscopic, very soluble in water, soluble in alcohol.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *histamine dihydrochloride CRS*. Examine as discs prepared using 1 mg of substance.

B. Examine the chromatograms obtained in the test for histidine. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in 7 mL of *water R* and add 3 mL of a 200 g/L solution of *sodium hydroxide R*. Dissolve 50 mg of *sulfanilic acid R* in a mixture of 0.1 mL of *hydrochloric acid R* and 10 mL of *water R* and add 0.1 mL of *sodium nitrite solution R*. Add the second solution to the first and mix. A red colour is produced.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 0.5 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of solution S is 2.85 to 3.60.

Histidine

Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel G plate R*.

Test solution (a) Dissolve 0.5 g of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 2 mL of test solution (a) to 10 mL with *water R*.

Reference solution (a) Dissolve 0.1 g of *histamine dihydrochloride CRS* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 50 mg of *histidine monohydrochloride R* in *water R* and dilute to 100 mL with the same solvent.

Reference solution (c) Mix 1 mL of test solution (a) and 1 mL of reference solution (b).

Apply to the plate 1 µL of test solution (a), 1 µL of test solution (b), 1 µL of reference solution (a), 1 µL of reference solution (b) and 2 µL of reference solution (c). Develop over a path of 15 cm using a mixture of 5 volumes of *concentrated ammonia R*, 20 volumes of *water R* and 75 volumes of *acetonitrile R*. Dry the plate in a current of air. Repeat the development in the same direction, dry the plate in a current of air and spray with *ninhydrin solution R1*. Heat the plate at 110 °C for 10 min. Any spot corresponding to histidine in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (b) (1 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows 2 clearly separated spots.

Sulfates (*2.4.13*)

3 mL of solution S diluted to 15 mL with *distilled water R* complies with the limit test for sulfates (0.1 per cent).

Loss on drying (*2.2.32*)

Not more than 0.5 per cent, determined on 0.20 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 0.5 g.

ASSAY
Dissolve 0.080 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the first and third points of inflexion.

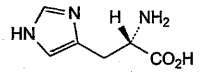
1 mL of *0.1 M sodium hydroxide* is equivalent to 9.203 mg of $C_5H_{11}Cl_2N_3$.

STORAGE
Store in an airtight container, protected from light.

Ph Eur

Histidine

(Ph. Eur. monograph 0911)



$C_6H_9N_3O_2$ 155.2 *71-00-1*

Action and use
Amino acid.

Ph Eur

DEFINITION
(2*S*)-2-Amino-3-(1*H*-imidazol-4-yl)propanoic acid.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *histidine CRS.*

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *water R*, evaporate to dryness at 60 °C and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of *histidine CRS* in *water R* and dilute to 50 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 0.1 g in 7 mL of *water R* and add 3 mL of a 200 g/L solution of *sodium hydroxide R*. Dissolve 50 mg of *sulfanilic acid R* in a mixture of 0.1 mL of *hydrochloric acid R* and 10 mL of *water R* and add 0.1 mL of *sodium nitrite solution R*. Add the second solution to the first and mix. An orange-red colour develops.

TESTS
Solution S
Dissolve 2.5 g in *distilled water R*, heating in a water-bath, and dilute to 50 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Specific optical rotation (*2.2.7*)
+ 11.4 to + 12.4 (dried substance).

Dissolve 2.75 g in 12.0 mL of *hydrochloric acid R1* and dilute to 25.0 mL with *water R*.

Ninhydrin-positive substances
Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *water R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of histidine in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

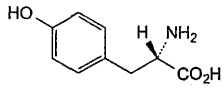
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.130 g in 50 mL of *water R*. Titrate with *0.1 M hydrochloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M hydrochloric acid* is equivalent to 15.52 mg of $C_6H_9N_3O_2$.

STORAGE
Protected from light.

IMPURITIES
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A.

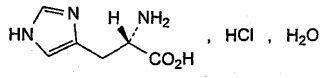


A. (2*S*)-2-amino-3-(4-hydroxyphenyl)propanoic acid (tyrosine).

Ph Eur

Histidine Hydrochloride Monohydrate

(Ph. Eur. monograph 0910)



$C_6H_{10}ClN_3O_2,H_2O$ 209.6 *5934-29-2*

Action and use
Amino acid.

Ph Eur

DEFINITION
(2*S*)-2-Amino-3-(1*H*-imidazol-4-yl)propanoic acid hydrochloride monohydrate.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B, C, F.

Second identification: A, B, D, E, F.

A. Specific optical rotation (see Tests).

B. pH (see Tests).

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *histidine hydrochloride monohydrate CRS.*

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in *water R* and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of *histidine hydrochloride monohydrate CRS* in *water R* and dilute to 50 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to

the principal spot in the chromatogram obtained with the reference solution.

E. Dissolve 0.1 g in 7 mL of *water R* and add 3 mL of a 200 g/L solution of *sodium hydroxide R*. Dissolve 50 mg of *sulfanilic acid R* in a mixture of 0.1 mL of *hydrochloric acid R* and 10 mL of *water R* and add 0.1 mL of *sodium nitrite solution R*. Add the second solution to the first and mix. An orange-red colour develops.

F. About 20 mg gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 2.5 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)

3.0 to 5.0 for solution S.

Specific optical rotation (*2.2.7*)

+ 9.2 to + 10.6 (dried substance).

Dissolve 2.75 g in 12.0 mL of *hydrochloric acid R1* and dilute to 25.0 mL with *water R*.

Ninhydrin-positive substances

Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *water R* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):

— *resolution*: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:

— for any ninhydrin-positive substance detected at 570 nm, use the concentration of histidine in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:

— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;

— *total*: maximum 0.5 per cent;

— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Sulfates (*2.4.13*)

Maximum 300 ppm.

Dilute 10 mL of solution S to 15 mL with *distilled water R*.

Ammonium

Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:

— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)

7.0 per cent to 10.0 per cent, determined on 1.000 g by drying in an oven at 145-150 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.160 g in 50 mL of *carbon dioxide-free water R*. Titrate with *0.1 M sodium hydroxide*, determining the end-point potentiometrically (*2.2.20*).

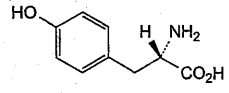
1 mL of *0.1 M sodium hydroxide* is equivalent to 19.16 mg of $C_6H_{10}ClN_3O_2$.

STORAGE

Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A*.

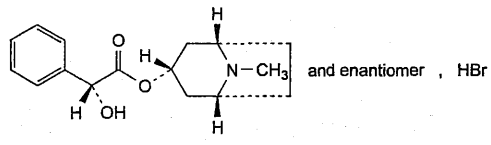


A. (2*S*)-2-amino-3-(4-hydroxyphenyl)propanoic acid (tyrosine).

Ph Eur

Homatropine Hydrobromide

(Ph. Eur. monograph 0500)



$C_{16}H_{22}BrNO_3$ 356.3 *51-56-9*

Action and use
Anticholinergic.

Preparation
Homatropine Eye Drops

Ph Eur

DEFINITION

(1*R*,3*r*,5*S*)-8-Methyl-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-2-hydroxy-2-phenylacetate hydrobromide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, sparingly soluble in alcohol.

mp
About 215 °C, with decomposition.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *homatropine hydrobromide CRS.*

B. Dissolve 50 mg in 1 mL of *water R* and add 2 mL of *dilute acetic acid R*. Heat and add 4 mL of *picric acid solution R*. Allow to cool, shaking occasionally. Collect the crystals, wash with 2 quantities, each of 3 mL, of iced *water R* and dry at 100-105 °C. The crystals melt (*2.2.14*) at 182 °C to 186 °C.

C. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 1.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
5.0 to 6.5 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 25.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of *hyoscine hydrobromide CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. To 10.0 mL of this solution add 0.5 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Column:
— *size: l* = 0.1 m, Ø = 4.6 mm,
— *stationary phase: octadecylsilyl silica gel for chromatography R* (3 µm),
— *temperature*: 40 °C.

Mobile phase Mix 33 volumes of *methanol R2* and 67 volumes of a solution prepared as follows: dissolve 6.8 g of *potassium dihydrogen phosphate R* and 7.0 g of *sodium heptanesulfonate monohydrate R* in 1000 mL of *water R* and adjust to pH 2.7 with a 330 g/L solution of *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Run time 3 times the retention time of homatropine.

Relative retention With reference to homatropine (retention time = about 6.8 min): impurity C = about 0.2; impurity A = about 0.9; impurity B = about 1.1; impurity D = about 1.9.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to homatropine and impurity B,
— *symmetry factor*: maximum 2.5 for the peak due to homatropine.

Limits:
— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *impurities B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent); disregard the peak due to the bromide ion which appears close to the peak due to the solvent,
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *alcohol R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

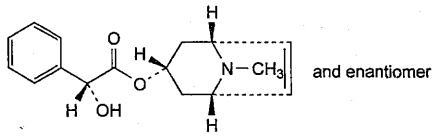
1 mL of *0.1 M sodium hydroxide* is equivalent to 35.63 mg of $C_{16}H_{22}BrNO_3$.

STORAGE

Protected from light.

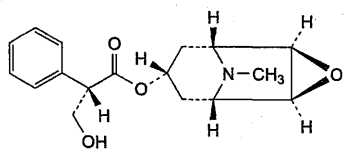
IMPURITIES

Specified impurities *A, B, C, D.*

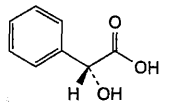


and enantiomer

A. (1*R*,3*s*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-6-en-3-yl (2*RS*)-2-hydroxy-2-phenylacetate (dehydrohomatropine),

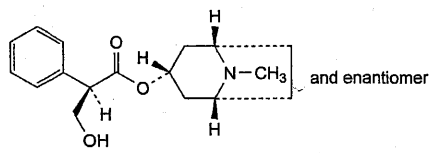


B. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$] non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscine),



and enantiomer

C. (2*RS*)-2-hydroxy-2-phenylacetic acid (mandelic acid),



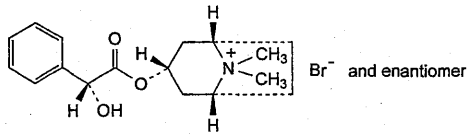
and enantiomer

D. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate (atropine).

Ph Eur

Homatropine Methylbromide

(Ph. Eur. monograph 0720)



and enantiomer

$C_{17}H_{24}BrNO_3$ 370.3 *80-49-9*

Action and use
Anticholinergic.

Ph Eur

DEFINITION
(1*R*,3*r*,5*S*)-3-[[(2*RS*)-2-Hydroxy-2-phenylacetyl]oxy]-8,8-dimethyl-8-azoniabicyclo[3.2.1]octane bromide.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, soluble in ethanol 96 per cent.

mp
About 190 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *homatropine methylbromide CRS.*

B. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 1.25 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)
4.5 to 6.5 for solution S.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1*, mobile phase A (9:41 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 25.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of *homatropine hydrobromide CRS* (impurity B) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. To 10.0 mL of the solution add 0.5 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 2.0 mg of *homatropine methylbromide for system suitability CRS* (containing impurity A) in 1.0 mL of the solvent mixture.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 25 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.4 g of *potassium dihydrogen phosphate R* and 5.0 g of *sodium pentanesulfonate monohydrate R* in 980 mL of *water for chromatography R*, adjust to pH 3.0 with a 330 g/L solution of *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*;
— *mobile phase B*: mix 400 mL of mobile phase A and 600 mL of *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	70	30
2 - 15	70 → 30	30 → 70

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Relative retention With reference to homatropine methylbromide (retention time = about 5 min): impurity A = about 0.9; impurity B = about 1.2.

Identification of impurities Use the chromatogram supplied with *homatropine methylbromide for system suitability CRS* and the chromatogram obtained with reference solution (d) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B.

System suitability:
— *resolution*: minimum 2.5 between the peaks due to homatropine methylbromide and impurity B in the chromatogram obtained with reference solution (c);
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to homatropine methylbromide in the chromatogram obtained with reference solution (d).

Limits:
— *impurities A, B*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent); disregard the peak due to the bromide ion which appears close to the peak due to the solvent;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

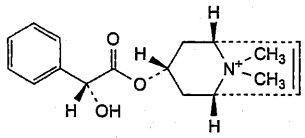
ASSAY
Dissolve 0.300 g in 10 mL of *water R*. Titrate with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*), using a silver indicator electrode and a silver-silver chloride reference electrode.

1 mL of *0.1 M silver nitrate* is equivalent to 37.03 mg of $C_{17}H_{24}BrNO_3$.

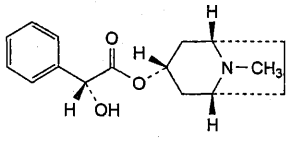
STORAGE
Protected from light.

IMPURITIES
Specified impurities *A, B.*

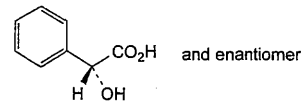
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, F.*



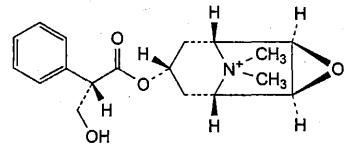
A. (1*R*,3*s*,5*S*)-3-[[(2*RS*)-2-hydroxy-2-phenylacetyl]oxy]-8,8-dimethyl-8-azoniabicyclo[3.2.1]oct-6-ene (methyldehydrohomatropine),



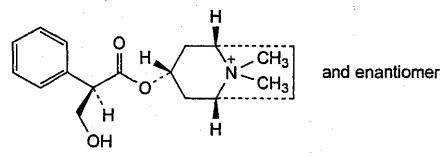
B. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-2-hydroxy-2-phenylacetate (homatropine),



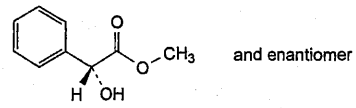
C. (2*RS*)-2-hydroxy-2-phenylacetic acid (mandelic acid),



D. (1*R*,2*R*,4*S*,5*S*,7*s*)-7-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-9,9-dimethyl-3-oxa-9-azoniatricyclo[3.3.1.$0^{2,4}$]nonane (methylhyoscine),



E. (1*R*,3*r*,5*S*)-3-[[(2*RS*)-3-hydroxy-2-phenylpropanoyl]oxy]-8,8-dimethyl-8-azoniabicyclo[3.2.1]octane (methylatropine),



F. methyl (2*RS*)-2-hydroxy-2-phenylacetate (methyl mandelate).

Ph Eur

Honey

(Ph. Eur. monograph 2051)

Ph Eur

DEFINITION
Honey is produced by bees (*Apis mellifera* L.) from the nectar of plants or from secretions of living parts of plants which the bees collect, transform by combining with specific substances of their own, deposit, dehydrate, store and leave in the honey comb to ripen and mature.

PRODUCTION
If the bee has been exposed to treatment to prevent or cure diseases or to any substance intended for preventing, destroying or controlling any pest, unwanted species of plants or animals, appropriate measures are taken to ensure that the levels of residues are as low as possible.

CHARACTERS
Appearance
Viscous liquid which may be partly crystalline, almost white to dark brown.

IDENTIFICATION
Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 0.6 g of the substance to be examined in 50 mL of *ethanol (30 per cent V/V) R*.

Reference solution Dissolve 0.5 g of *fructose R*, 0.5 g of *glucose R* and 0.1 g of *sucrose R* in 100 mL of *ethanol (30 per cent V/V) R*.

Plate *TLC silica gel plate R.*

Mobile phase *water R, acetonitrile R* (13:87 *V/V*).

Application 5 µL as bands.

Development 3 times over a path of 15 cm.

Drying In warm air.

Detection Spray with a solution prepared as follows: dissolve 2 g of *diphenylamine R* and 2 mL of *aniline R* in 100 mL of *acetone R*. Add a 850 g/L solution of *phosphoric acid R* until the precipitate formed dissolves again (about 15-20 mL). Examine in daylight after heating at 100-105 °C for 5-10 min.

Results See below the sequence of the zones present in the chromatograms obtained with the reference solution and the test solution. Furthermore, the weak brown zone due to sucrose in the chromatogram obtained with the reference solution may be present in the chromatogram obtained with the test solution. One or more other weak zones may be present in the chromatogram obtained with the test solution.

Top of the plate	
Fructose: an intense brown zone	An intense brown zone (fructose)
Glucose: an intense greyish-blue zone	An intense greyish-blue zone (glucose)
Sucrose: a brown zone	2 to 3 brownish-grey zones
Reference solution	**Test solution**

TESTS

Refractive index (*2.2.6*)

Minimum 1.487 (equivalent to a maximum water content of 20 per cent).

Homogenise 100 g and transfer into a flask. Close tightly and place in a water-bath at 50 ± 0.2 °C until all sugar crystals have dissolved. Cool the solution to 20 °C and rehomogenise. Immediately after rehomogenisation, cover the surface of the refractometer prism evenly with the sample. Determine the refractive index after 2 min if using an Abbe refractometer and after 4 min if using a digital refractometer. Use the average value of 2 determinations.

Conductivity (*2.2.38*)

Maximum 800 µS·cm^{-1}.

Using the value obtained for the refractive index, determine the water content of the substance to be examined from Table 2051.-1. Using this information, dissolve an amount of the substance to be examined equivalent to 20.0 g of honey dry solids, in *water R* to produce 100.0 mL.

Optical rotation (*2.2.7*)

Maximum + 0.6°.

Using the value obtained for the refractive index, determine the water content of the substance to be examined from Table 2051.-1. Using this information, dissolve an amount of the substance to be examined, equivalent to 20.0 g of honey dry solids, in 50 mL of *water R*. Add 0.2 mL of *concentrated ammonia R* and dilute to 100.0 mL with *water R*. If necessary decolourise the solution with *activated charcoal R*.

Table 2051.-1. – *Relationship of water content of honey to refractive index*

Water content (per cent *m/m*)	Refractive index at 20 °C
15.0	1.4992
15.2	1.4987
15.4	1.4982
15.6	1.4976
15.8	1.4971
16.0	1.4966
16.2	1.4961
16.4	1.4956
16.6	1.4951
16.8	1.4946
17.0	1.4940
17.2	1.4935
17.4	1.4930
17.6	1.4925
17.8	1.4920
18.0	1.4915
18.2	1.4910
18.4	1.4905
18.6	1.4900
18.8	1.4895
19.0	1.4890
19.2	1.4885
19.4	1.4880
19.6	1.4875
19.8	1.4870
20.0	1.4865

5-Hydroxymethylfurfural

Maximum 80 ppm, calculated on dry solids.

Using the value obtained for the refractive index, determine the water content of the substance to be examined from Table 2051.-1. Using this information, dissolve an amount of the substance to be examined, equivalent to 5.0 g of honey dry solids, in 25 mL of *water R* and transfer to a 50.0 mL volumetric flask with the same solvent. Add 0.5 mL of a 150 g/L solution of *potassium ferrocyanide R* and mix. Add 0.5 mL of a 300 g/L solution of *zinc acetate R*, mix and dilute to 50.0 mL with *water R* (a drop of *anhydrous ethanol R* may be added to avoid foaming). Filter. Transfer 5.0 mL of the filtered solution into each of 2 tubes. To one tube add 5.0 mL of *water R* (test solution). To the other tube add 5.0 mL of a 2.0 g/L solution of *sodium hydrogensulfite R* (reference solution). Determine the absorbance (*2.2.25*) of the test solution against the reference solution at 284 nm and 336 nm within 60 min. If the absorbance at 284 nm is greater than 0.8, dilute to the same extent the test solution with *water R* and the reference solution with a 2.0 g/L solution of *sodium hydrogensulfite R* so as to obtain an absorbance of less than 0.8.

Calculate the content of 5-hydroxymethylfurfural from the expression:

$$(A_1 - A_2) \times D \times 149.7$$

A_1 = absorbance at 284 nm,
A_2 = absorbance at 336 nm,
D = dilution factor, where applicable.

Chlorides (*2.4.4*)

Maximum 350 ppm, determined on 15 mL of a 10 g/L solution.

Sulfates (*2.4.13*)
Maximum 250 ppm, determined on 15 mL of a 40 g/L solution.

Ph Eur

Hyaluronidase

(Ph. Eur. monograph 0912)

9001-54-1

Action and use
Used to promote absorption of fluid into tissues.

Preparation
Hyaluronidase Injection

Ph Eur

DEFINITION
Enzyme extracted from mammalian testes (for example bovine testes) and capable of hydrolysing mucopolysaccharides of the hyaluronic acid type. It may contain a suitable stabiliser.

Potency
Minimum 300 IU of hyaluronidase activity per milligram (dried substance).

PRODUCTION
The animals from which hyaluronidase is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS
Appearance
White or yellowish-white, amorphous powder.

Solubility
Soluble in water, practically insoluble in acetone and in anhydrous ethanol.

IDENTIFICATION
A solution containing the equivalent of 100 IU of hyaluronidase in 1 mL of a 9 g/L solution of *sodium chloride R* depolymerises an equal volume of a 10 g/L solution of *sodium hyaluronate BRP* in 1 min at 20 °C as shown by a pronounced decrease in viscosity. This action is destroyed by heating the hyaluronidase at 100 °C for 30 min.

TESTS
Appearance of solution
The solution is clear (*2.2.1*).

Dissolve 0.10 g in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)
4.5 to 7.5.

Dissolve 30 mg in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 0.500 g by drying at 60 °C at a pressure not exceeding 670 Pa for 2 h.

Bacterial endotoxins (*2.6.14*)
Less than 0.2 IU per IU of hyaluronidase.

ASSAY
The activity of hyaluronidase is determined by comparing the rate at which it hydrolyses *sodium hyaluronate BRP* with the rate obtained with the International Standard, or a reference preparation calibrated in International Units, using a slope-ratio assay.

Substrate solution To 0.10 g of *sodium hyaluronate BRP* in a 25 mL conical flask add slowly 20.0 mL of *water R* at 4 °C. The rate of addition must be slow enough to allow the substrate particles to swell (about 5 min). Maintain at 4 °C and stir for at least 12 h. Store at 4 °C and use within 4 days.

For the test solution and the reference solution, prepare the solution and carry out the dilution at 0 °C to 4 °C.

Test solution Dissolve a suitable amount of the substance to be examined in *hyaluronidase diluent R* so as to obtain a solution containing 0.6 ± 0.3 IU of hyaluronidase per millilitre.

Reference solution Dissolve a suitable amount of *hyaluronidase BRP* in *hyaluronidase diluent R* so as to obtain a solution containing 0.6 IU of hyaluronidase per millilitre.

In a reaction vessel, mix 1.50 mL of *phosphate buffer solution pH 6.4 R* and 1.0 mL of the substrate solution and equilibrate at 37 ± 0.1 °C. At time $t_1 = 0$ (first chronometer) add 0.50 mL of the test solution containing E_t mg of the enzyme to be examined, mix, measure the viscosity of the solution using a suitable viscometer maintained at 37 ± 0.1 °C and record the outflow time t_2 using a second chronometer (graduated in 0.1 second intervals), several times during about 20 min (read on the first chronometer). The following viscometer has been found suitable: Ubbelohde microviscometer (DIN 51 562, Part 2), capillary type MII, viscometer constant about 0.1 mm^2/s^2.

Repeat the procedure using 0.50 mL of the reference solution containing E_r mg of *hyaluronidase BRP*.

Calculate the viscosity ratio from the expression:

$$\eta_r = \frac{k \times t_2}{0.6915}$$

k = the viscometer constant in mm^2/s^2 (indicated on the viscometer);
t_2 = the outflow time (in seconds) of the solution;
0.6915 = the kinematic viscosity in mm^2/s of the buffer solution at 37 °C.

Since the enzymatic reaction continues during the outflow time measurements, the real reaction time equals $t_1 + t_2/2$, half of the outflow time ($t_2/2$) for which a certain measurement is valid being added to the time t_1 at which the measurement is started. Plot $(\ln \eta_r)^{-1}$ as a function of the reaction time ($t_1 + t_2/2$) in seconds. A linear relationship is obtained. Calculate the slope for the substance to be examined (b_t) and the reference preparation (b_r).

Calculate the specific activity in International Units per milligram from the expression:

$$\frac{b_t}{b_r} \times \frac{E_r}{E_t} \times A$$

A = the specific activity of *hyaluronidase BRP* in International Units per milligram.

Carry out the complete procedure at least three times and calculate the average activity of the substance to be examined.

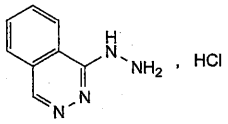
STORAGE
Store in an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, tamper-proof container.

LABELLING
The label states the activity in International Units per milligram.

Ph Eur

Hydralazine Hydrochloride

(Ph. Eur. monograph 0829)



$C_8H_9ClN_4$ 196.6 *304-20-1*

Action and use
Vasodilator; treatment of hypertension.

Preparations
Hydralazine Injection
Hydralazine Tablets

Ph Eur

DEFINITION
1-Hydrazinophthalazine hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

mp
About 275 °C, with decomposition.

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).
Test solution Dissolve 50 mg in *water R* and dilute to 100 mL with the same solvent. Dilute 2 mL of this solution to 100 mL with *water R*.
Spectral range 220-350 nm.
Absorption maxima At 240 nm, 260 nm, 303 nm and 315 nm.
Absorbance ratio A_{240}/A_{303} = 2.0 to 2.2.

B. Infrared absorption spectrophotometry (*2.2.24*).
Preparation Discs.
Comparison *hydralazine hydrochloride CRS*.

C. Dissolve 0.5 g in a mixture of 8 mL of *dilute hydrochloric acid R* and 100 mL of *water R*. Add 2 mL of *sodium nitrite solution R*, allow to stand for 10 min and filter. The precipitate, washed with *water R* and dried at 100-105 °C, melts (*2.2.14*) at 209 °C to 212 °C.

D. Dissolve about 10 mg in 2 mL of *water R*. Add 2 mL of a 20 g/L solution of *nitrobenzaldehyde R* in *ethanol (96 per cent) R*. An orange precipitate is formed.

E. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_6 (*2.2.2, Method II*).
Dilute 4 mL of solution S to 20 mL with *water R*.

pH (*2.2.3*)
3.5 to 4.2 for solution S.

Hydrazine
Thin-layer chromatography (*2.2.27*).
Test solution Dissolve 0.12 g of the substance to be examined in 4 mL of *water R* and add 4 mL of a 150 g/L solution of *salicylaldehyde R* in *methanol R* and 0.2 mL of *hydrochloric acid R*. Mix and keep at a temperature not exceeding 25 °C for 2-4 h, until the precipitate formed has sedimented. Add 4 mL of *toluene R*, shake vigorously and centrifuge. Transfer the clear supernatant to a 100 mL separating funnel and shake vigorously, each time for 3 min, with 2 quantities, each of 20 mL, of a 200 g/L solution of *sodium metabisulfite R* and with 2 quantities, each of 50 mL, of *water R*. Separate the upper toluene layer which is the test solution.
Reference solution (a) Dissolve 12 mg of *hydrazine sulfate R* in *dilute hydrochloric acid R* and dilute to 100.0 mL with the same acid. Dilute 1.0 mL of this solution to 100.0 mL with *dilute hydrochloric acid R*.
Reference solution (b) Prepare the solution at the same time and in the same manner as for the test solution, using 1.0 mL of reference solution (a) and 3 mL of *water R*.
Plate *TLC silica gel G plate R*.
Mobile phase *ethanol (96 per cent) R, toluene R* (10:90 *V/V*).
Application 20 µL of the test solution and reference solution (b).
Development Over a path of 10 cm.
Drying In air.
Detection Examine in ultraviolet light at 365 nm.
Limit:
— *hydrazine*: any yellow fluorescent spot due to hydrazine is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (10 ppm).

Related substances
Liquid chromatography (*2.2.29*). *The solutions must be injected within one working day.*
Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.
Reference solution (b) Dilute 10.0 mL of reference solution (a) to 50.0 mL with the mobile phase.
Reference solution (c) Dissolve 25.0 mg of *phthalazine R* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 4.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (d) Dilute a mixture of 4.0 mL of the test solution and 10.0 mL of reference solution (c) to 100.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *nitrile silica gel for chromatography R1* (10 µm).

Mobile phase Mix 22 volumes of *acetonitrile R* and 78 volumes of a solution containing 1.44 g/L of *sodium laurilsulfate R* and 0.75 g/L of *tetrabutylammonium bromide R*, then adjust to pH 3.0 with *dilute sulfuric acid R1*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 3 times the retention time of hydralazine.

Retention time Hydralazine = about 10 min to 12 min; if necessary, adjust the concentration of acetonitrile in the mobile phase.

System suitability:
— the chromatogram obtained with reference solution (d) shows 2 principal peaks;
— *resolution*: minimum 2.5 between the peaks due to hydralazine and phtalazine in the chromatogram obtained with reference solution (d);
— *signal-to-noise ratio*: minimum 3 for the principal peak in the chromatogram obtained with reference solution (b).

Limit:
— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo*.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 80.0 mg in 25 mL of *water R*. Add 35 mL of *hydrochloric acid R* and titrate with *0.05 M potassium iodate*, determining the end-point potentiometrically (*2.2.20*) using a platinum indicator electrode.

1 mL of *0.05 M potassium iodate* is equivalent to 9.832 mg of $C_8H_9ClN_4$.

STORAGE
Protected from light.

Ph Eur

Hydrochloric Acid

(Concentrated Hydrochloric Acid, Ph. Eur. monograph 0002)

HCl	36.46	*7647-01-0*

Preparation
Dilute Hydrochloric Acid

Ph Eur

DEFINITION
Content
35.0 per cent *m/m* to 39.0 per cent *m/m*.

CHARACTERS
Appearance
Clear, colourless, fuming liquid.

Solubility
Miscible with water.

Relative density
About 1.18.

IDENTIFICATION
A. Dilute with *water R*. The solution is strongly acid (*2.2.4*).

B. It gives the reactions of chlorides (*2.3.1*).

C. It complies with the limits of the assay.

TESTS
Appearance of solution
To 2 mL add 8 mL of *water R*. The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Free chlorine
Maximum 4 ppm.

To 15 mL add 100 mL of *carbon dioxide-free water R*, 1 mL of a 100 g/L solution of *potassium iodide R* and 0.5 mL of *iodide-free starch solution R*. Allow to stand in the dark for 2 min. Any blue colour disappears on the addition of 0.2 mL of *0.01 M sodium thiosulfate*.

Sulfates (*2.4.13*)
Maximum 20 ppm.

To 6.4 mL add 10 mg of *sodium hydrogen carbonate R* and evaporate to dryness on a water-bath. Dissolve the residue in 15 mL of *distilled water R*.

Residue on evaporation
Maximum 0.01 per cent.

Evaporate 100.0 g to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 10 mg.

ASSAY
Weigh accurately a ground-glass-stoppered flask containing 30 mL of *water R*. Introduce 1.5 mL of the acid to be examined and weigh again. Titrate with *1 M sodium hydroxide*, using *methyl red solution R* as indicator.

1 mL of *1 M sodium hydroxide* is equivalent to 36.46 mg of HCl.

STORAGE
In a stoppered container made of glass or another inert material, at a temperature not exceeding 30 °C.

Ph Eur

Dilute Hydrochloric Acid

(Ph. Eur. monograph 0003)

Ph Eur

DEFINITION
Content
9.5 per cent *m/m* to 10.5 per cent *m/m* of HCl (M_r 36.46).

PREPARATION
To 726 g of *water R* add 274 g of concentrated hydrochloric acid and mix.

IDENTIFICATION
A. It is strongly acid (*2.2.4*).

B. It gives the reactions of chlorides (*2.3.1*).

C. It complies with the limits of the assay.

TESTS
Appearance
It is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Free chlorine
Maximum 1 ppm.

To 60 mL add 50 mL of *carbon dioxide-free water R*, 1 mL of a 100 g/L solution of *potassium iodide R* and 0.5 mL of *iodide-*

free starch solution R. Allow to stand in the dark for 2 min. Any blue colour disappears on the addition of 0.2 mL of *0.01 M sodium thiosulfate*.

Sulfates (*2.4.13*)
Maximum 5 ppm.

To 26 mL add 10 mg of *sodium hydrogen carbonate R* and evaporate to dryness on a water-bath. Dissolve the residue in 15 mL of *distilled water R*.

Residue on evaporation
Maximum 0.01 per cent.

Evaporate 100.0 g to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 10 mg.

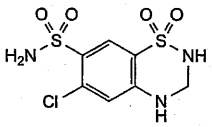
ASSAY
To 6.00 g add 30 mL of *water R*. Titrate with *1 M sodium hydroxide*, using *methyl red solution R* as indicator.

1 mL of *1 M sodium hydroxide* is equivalent to 36.46 mg of HCl.

Ph Eur

Hydrochlorothiazide

(Ph. Eur. monograph 0394)



$C_7H_8ClN_3O_4S_2$ 297.7 *58-93-5*

Action and use
Thiazide diuretic.

Preparations
Co-amilozide Oral Solution
Co-amilozide Tablets
Co-triamterzide Tablets
Hydrochlorothiazide Tablets

Ph Eur

DEFINITION
6-Chloro-3,4-dihydro-2*H*-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide.

Content
97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Very slightly soluble in water, soluble in acetone, sparingly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (*5.9*).

IDENTIFICATION
First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in 10 mL of *0.1 M sodium hydroxide* and dilute to 100.0 mL with *water R*. Dilute 2.0 mL of this solution to 100.0 mL with *0.01 M sodium hydroxide*.

Spectral range 250-350 nm.

Absorption maxima At 273 nm and 323 nm.

Absorbance ratio A_{273}/A_{323} = 5.4 to 5.7.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydrochlorothiazide CRS*.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *ethanol R1*, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *hydrochlorothiazide CRS* in *acetone R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of *chlorothiazide R* in reference solution (a) and dilute to 5 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R*.

Mobile phase *ethyl acetate R*.

Application 2 µL.

Development Over 1/2 of the plate.

Drying In a current of air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Gently heat about 1 mg with 2 mL of a freshly prepared 0.5 g/L solution of *chromotropic acid, sodium salt R* in a cooled mixture of 35 volumes of *water R* and 65 volumes of *sulfuric acid R*. A violet colour develops.

TESTS
Acidity or alkalinity
Shake 0.5 g of the powdered substance to be examined with 25 mL of *water R* for 2 min and filter. To 10 mL of the filtrate, add 0.2 mL of *0.01 M sodium hydroxide* and 0.15 mL of *methyl red solution R*. The solution is yellow. Not more than 0.4 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red.

Related substances
Liquid chromatography (*2.2.29*).

Solvent mixture Dilute 50.0 mL of a mixture of equal volumes of *acetonitrile R1* and *methanol R2* to 200.0 mL with *phosphate buffer solution pH 3.2 R1*.

Test solution (a) Dissolve 30.0 mg of the substance to be examined in 5 mL of a mixture of equal volumes of *acetonitrile R1* and *methanol R2*, using sonication if necessary, and dilute to 20.0 mL with *phosphate buffer solution pH 3.2 R1*.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20.0 mL with *phosphate buffer solution pH 3.2 R1*.

Reference solution (a) Dissolve 3 mg of *chlorothiazide CRS* (impurity A) and 3 mg of *hydrochlorothiazide CRS* in 5 mL of a mixture of equal volumes of *acetonitrile R1* and

methanol R2, using sonication if necessary, and dilute to 20.0 mL with *phosphate buffer solution pH 3.2 R1*. Dilute 5.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 30.0 mg of *hydrochlorothiazide CRS* in 5 mL of a mixture of equal volumes of *acetonitrile R1* and *methanol R2*, using sonication if necessary, and dilute to 20.0 mL with *phosphate buffer solution pH 3.2 R1*. Dilute 1.0 mL of this solution to 20.0 mL with *phosphate buffer solution pH 3.2 R1*.

Reference solution (d) Dissolve 3 mg of *hydrochlorothiazide for peak identification CRS* (containing impurities B and C) in 0.5 mL of a mixture of equal volumes of *acetonitrile R1* and *methanol R2*, using sonication if necessary, and dilute to 2.0 mL with *phosphate buffer solution pH 3.2 R1*.

Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase:
— *mobile phase A*: to 940 mL of *phosphate buffer solution pH 3.2 R1* add 60.0 mL of *methanol R2* and 10.0 mL of *tetrahydrofuran R* and mix;
— *mobile phase B*: to a mixture of 500 mL of *methanol R2* and 500 mL of *phosphate buffer solution pH 3.2 R1* add 50.0 mL of *tetrahydrofuran R* and mix;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 17	100 → 55	0 → 45
17 - 30	55	45

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 224 nm.

Injection 10 µL of test solution (a) and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A; use the chromatogram supplied with *hydrochlorothiazide for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B and C.

Relative retention With reference to hydrochlorothiazide (retention time = about 8 min): impurity B = about 0.7; impurity A = about 0.9; impurity C = about 2.8.

System suitability Reference solution (a):
— *resolution*: minimum 2.5 between the peaks due to impurity A and hydrochlorothiazide.

Limits:
— *impurities A, B, C*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (*2.4.4*)
Maximum 100 ppm.

Dissolve 1.0 g in 25 mL of *acetone R* and dilute to 30 mL with *water R*. Prepare the standard using 5 mL of *acetone R* containing 15 per cent *V/V* of *water R* and 10 mL of *chloride standard solution (5 ppm Cl) R*.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Mobile phase:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 4	80	20
4 - 10	80 → 20	20 → 80

Flow rate 1.6 mL/min.

Injection Test solution (b) and reference solutions (a) and (c).

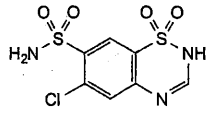
Relative retention With reference to hydrochlorothiazide (retention time = about 2.2 min): impurity A = about 0.9.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to impurity A and hydrochlorothiazide.

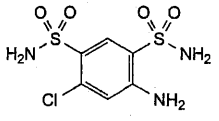
Calculate the percentage content of $C_7H_8ClN_3O_4S_2$ taking into account the assigned content of *hydrochlorothiazide CRS*.

IMPURITIES

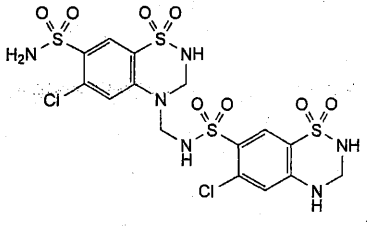
Specified impurities *A, B, C.*



A. 6-chloro-2*H*-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide (chlorothiazide),



B. 4-amino-6-chlorobenzene-1,3-disulfonamide (salamide),

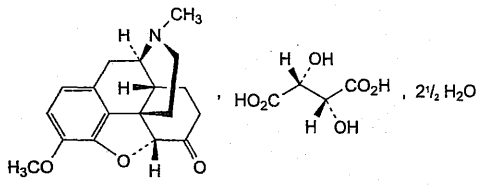


C. 6-chloro-*N*-[(6-chloro-7-sulfamoyl-2,3-dihydro-4*H*-1,2,4-benzothiadiazin-4-yl 1,1-dioxide)methyl]-3,4-dihydro-2*H*-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide.

Ph Eur

Hydrocodone Hydrogen Tartrate Hydrate

(Hydrocodone Hydrogen Tartrate 2.5-Hydrate, Ph. Eur. monograph 1784)



$C_{22}H_{27}NO_9,2.5H_2O$ 494.5

Action and use
Opioid receptor agonist; antitussive.

Ph Eur

DEFINITION

4,5α-Epoxy-3-methoxy-17-methylmorphinan-6-one hydrogen (2*R*,3*R*)-2,3-dihydroxybutanedioate 2.5-hydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble or soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydrocodone hydrogen tartrate 2.5-hydrate CRS.*

If the spectra obtained in the solid state show differences, dry the substance to be examined and the reference substance at 105 °C and record new spectra using the residues.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_5 (*2.2.2, Method II*).

Dissolve 0.5 g in *water R* and dilute to 10 mL with the same solvent.

pH (*2.2.3*)
3.2 to 3.8.

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−87 to −91 (anhydrous substance).

Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of *oxycodone hydrochloride CRS* (impurity D) in mobile phase A, add 0.5 mL of the test solution and dilute to 5.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 20 mg of *benzophenone CRS* (impurity H) in 50.0 mL of *methanol R*. Dilute 1.0 mL of this solution to 20.0 mL with mobile phase A.

Reference solution (d) Dissolve the contents of a vial of *hydrocodone for peak identification CRS* (containing impurities B, C, D, E, F and I) in 1.0 mL of mobile phase A.

Reference solution (e) Dissolve 5 mg of *morphine sulfate CRS* (impurity A) in 5 mL of mobile phase A.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 1.08 g of *sodium octanesulfonate R* in *water R*, adjust to pH 2.0 with *phosphoric acid R* and dilute to 1000 mL with *water R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	80	20
15 - 30	80 → 70	20 → 30
30 - 40	70 → 40	30 → 60
40 - 42	40	60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 283 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *hydrocodone for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B, C, D, E, F and I; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity A.

Relative retention With reference to hydrocodone (retention time = about 14 min): impurity A = about 0.3; impurity K = about 0.43; impurity B = about 0.57; impurity C = about 0.61; impurity D = about 0.9; impurity E = about 1.1; impurity F = about 1.5; impurity I = about 2.0; impurity H = about 2.9.

System suitability Reference solution (a):
— *resolution*: minimum 3.0 between the peaks due to impurity D and hydrocodone.

Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity I by 0.2;
— *impurity I*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *impurity H*: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— *impurities A, B, C, D, E, F, K*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
7.0 per cent to 12.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 44.95 mg of $C_{22}H_{27}NO_9$.

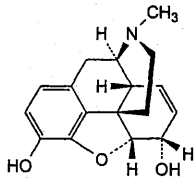
STORAGE

In an airtight container, protected from light.

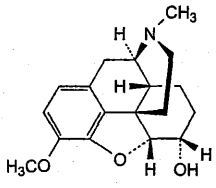
IMPURITIES

Specified impurities *A, B, C, D, E, F, H, I, K.*

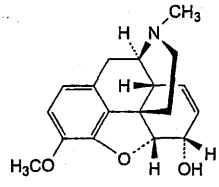
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *G, J.*



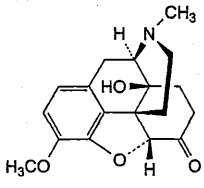
A. morphine,



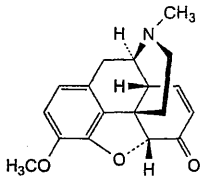
B. 4,5α-epoxy-3-methoxy-17-methylmorphinan-6α-ol (dihydrocodeine),



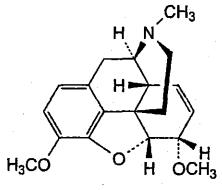
C. codeine,



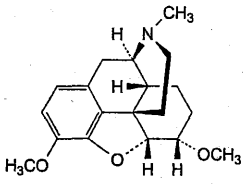
D. 4,5α-epoxy-14-hydroxy-3-methoxy-17-methylmorphinan-6-one (oxycodone),



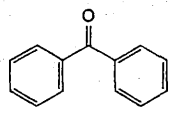
E. 7,8-didehydro-4,5α-epoxy-3-methoxy-17-methylmorphinan-6-one (codeinone),



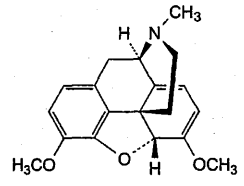
F. 7,8-didehydro-4,5α-epoxy-3,6α-dimethoxy-17-methylmorphinan (methylcodeine),



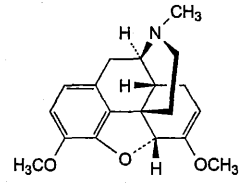
G. 4,5α-epoxy-3,6α-dimethoxy-17-methylmorphinan (tetrahydrothebaine),



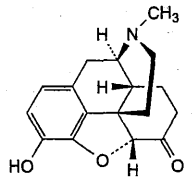
H. diphenylmethanone (benzophenone),



I. 6,7,8,14-tetradehydro-4,5α-epoxy-3,6-dimethoxy-17-methylmorphinan (thebaine),



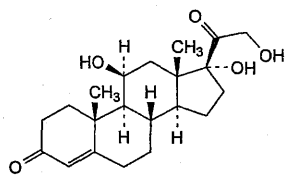
J. 6,7-didehydro-4,5α-epoxy-3,6-dimethoxy-17-methylmorphinan,



K. 4,5α-epoxy-3-hydroxy-17-methylmorphinan-6-one.

Hydrocortisone

(Ph. Eur. monograph 0335)



$C_{21}H_{30}O_5$ 362.5 *50-23-7*

Action and use
Corticosteroid.

Preparations
Hydrocortisone Cream
Hydrocortisone and Clioquinol Cream
Hydrocortisone and Neomycin Cream
Hydrocortisone Ointment
Hydrocortisone and Clioquinol Ointment
Miconazole and Hydrocortisone Cream
Miconazole and Hydrocortisone Ointment
Tretinoin, Hydrocortisone and Hydroquinone Cream

Ph Eur

DEFINITION

11β,17,21-Trihydroxypregn-4-ene-3,20-dione.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, sparingly soluble in acetone and in ethanol (96 per cent), slightly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

First identification: A, B.

Second identification: C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydrocortisone CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of *acetone R*, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

C. Thin-layer chromatography (*2.2.27*).

Solution A Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Solution B Dissolve 25 mg of *hydrocortisone CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Test solution (a) Dilute 2 mL of solution A to 10 mL with *methylene chloride R.*

Test solution (b) Transfer 0.4 mL of solution A to a glass tube 100 mm long and 20 mm in diameter and fitted with a ground-glass stopper or a polytetrafluoroethylene cap. Evaporate the solvent with gentle heating under a stream of *nitrogen R.* Add 2 mL of a 15 per cent *V/V* solution of *glacial acetic acid R* and 50 mg of *sodium bismuthate R.* Stopper the tube and shake the suspension in a mechanical shaker, protected from light, for 1 h. Add 2 mL of a 15 per cent *V/V* solution of *glacial acetic acid R* and filter into a 50 mL separating funnel, washing the filter with 2 quantities, each of 5 mL, of *water R.* Shake the clear filtrate with 10 mL of *methylene chloride R.* Wash the organic layer with 5 mL of *1 M sodium hydroxide* and then with 2 quantities, each of 5 mL, of *water R.* Dry over *anhydrous sodium sulfate R.*

Reference solution (a) Dilute 2 mL of solution B to 10 mL with *methylene chloride R.*

Reference solution (b) Transfer 0.4 mL of solution B to a glass tube 100 mm long and 20 mm in diameter and fitted with a ground-glass stopper or a polytetrafluoroethylene cap. Evaporate the solvent with gentle heating under a stream of *nitrogen R.* Add 2 mL of a 15 per cent *V/V* solution of *glacial acetic acid R* and 50 mg of *sodium bismuthate R.* Stopper the tube and shake the suspension in a mechanical shaker, protected from light, for 1 h. Add 2 mL of a 15 per cent *V/V* solution of *glacial acetic acid R* and filter into a 50 mL separating funnel, washing the filter with 2 quantities, each of 5 mL, of *water R.* Shake the clear filtrate with 10 mL of *methylene chloride R.* Wash the organic layer with 5 mL of *1 M sodium hydroxide* and then with 2 quantities, each of 5 mL, of *water R.* Dry over *anhydrous sodium sulfate R.*

Plate *TLC silica gel F_{254} plate R.*

Mobile phase A Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Mobile phase B *butanol R* saturated with *water R*, *toluene R*, *ether R* (5:15:80 *V/V/V*).

Application 5 µL of test solution (a) and reference solution (a), 25 µL of test solution (b) and reference solution (b), applying the latter 2 in small quantities to obtain small spots.

Development Over a path of 15 cm with mobile phase A, and then over a path of 15 cm with mobile phase B.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with test solutions (a) and (b) is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R* and heat at 120 °C for 10 min or until the spots appear; allow to cool, and examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with test solutions (a) and (b) is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution; the principal spots in the chromatograms obtained with test solution (b) and reference solution (b) have an R_F value distinctly higher than that of the principal spots in the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, an intense brownish-red colour develops with a green fluorescence that is particularly intense when examined in ultraviolet light at 365 nm. Add the solution to 10 mL of *water R* and mix. The colour fades and a clear solution remains. The fluorescence in ultraviolet light does not disappear.

TESTS

Specific optical rotation (*2.2.7*)

+ 162 to + 168 (dried substance).

Dissolve 0.200 g in *methanol R*, dilute to 25.0 mL with the same solvent and sonicate for 10 min.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (40:60 *V/V*).

Test solution Dissolve 20 mg of the substance to be examined in the solvent mixture, dilute to 10.0 mL with the solvent mixture and sonicate for 10 min.

Reference solution (a) Dissolve 4 mg of *prednisolone CRS* (impurity A), 2 mg of *cortisone R* (impurity B), 8 mg of *hydrocortisone acetate CRS* (impurity C) and 6 mg of Reichstein's substance S R (impurity F) in 40 mL of *acetonitrile R* and dilute to 100.0 mL with *water R*. Dilute 0.5 mL of the solution to 5.0 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of *hydrocortisone CRS* in 1.0 mL of the solvent mixture and sonicate for 10 min.

Reference solution (d) Dissolve 2 mg of *hydrocortisone for peak identification CRS* (containing impurities D, E, G, H, I and N) in 1.0 mL of the solvent mixture and sonicate for 10 min.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:

— *mobile phase A*: *water R*;

— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 18	74	26
18 - 32	74 → 55	26 → 45
32 - 48	55 → 30	45 → 70

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram supplied with *hydrocortisone for peak identification CRS* and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities D, E, G, H, I and N; use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C and F.

Relative retention With reference to hydrocortisone (retention time = about 24 min): impurity D = about 0.2; impurity H = about 0.3; impurity I = about 0.5; impurity G = about 0.8; impurity E = about 0.86; impurity A = about 0.96; impurity B = about 1.1; impurity F = about 1.4; impurity C = about 1.5; impurity N = about 1.7.

System suitability Reference solution (a):

— *peak-to-valley ratio*: minimum 3.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to hydrocortisone.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 1.8; impurity E = 2.7;

— *impurities C, D, E, I*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurity G*: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— *impurity F*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurities A, B*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurities H, N*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 20 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 241.5 nm.

Calculate the content of $C_{21}H_{30}O_5$ taking the specific absorbance to be 440.

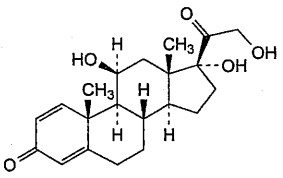
STORAGE

Protected from light.

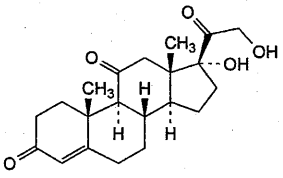
IMPURITIES

Specified impurities *A, B, C, D, E, F, G, H, I, N.*

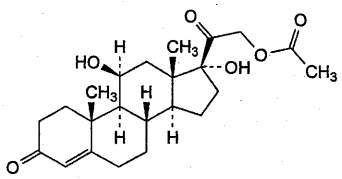
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *J, K, L, M, O.*



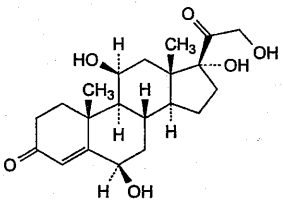
A. 11β,17,21-trihydroxypregna-1,4-diene-3,20-dione (prednisolone),



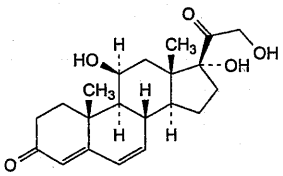
B. 17,21-dihydroxypregn-4-ene-3,11,20-trione (cortisone),



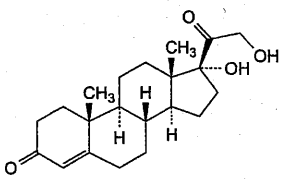
C. 11β,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate (hydrocortisone acetate),



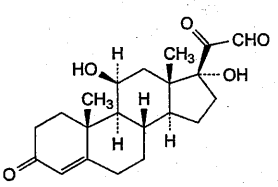
D. 6β,11β,17,21-tetrahydroxypregn-4-ene-3,20-dione (6β-hydroxyhydrocortisone),



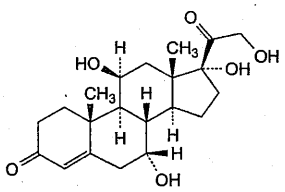
E. 11β,17,21-trihydroxypregna-4,6-diene-3,20-dione (Δ6-hydrocortisone),



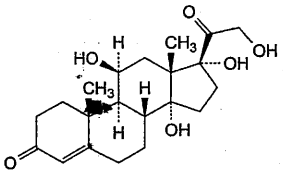
F. 17,21-dihydroxypregn-4-ene-3,20-dione (Reichstein's substance S),



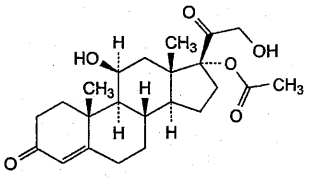
G. 11β,17-dihydroxy-3,20-dioxopregn-4-en-21-al (hydrocortisone-21-aldehyde),



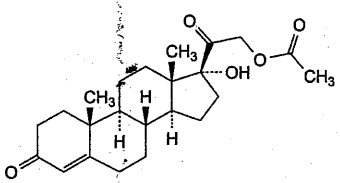
H. 7α,11β,17,21-tetrahydroxypregn-4-ene-3,20-dione (7α-hydroxyhydrocortisone),



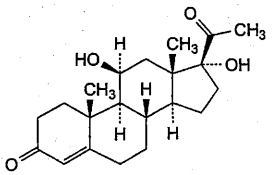
I. 11β,14,17,21-tetrahydroxypregn-4-ene-3,20-dione (14α-hydroxyhydrocortisone),



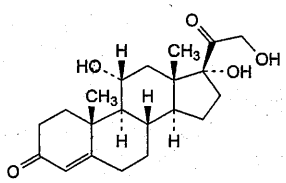
J. 11β,21-dihydroxy-3,20-dioxopregn-4-en-17-yl acetate (hydrocortisone-17-acetate),



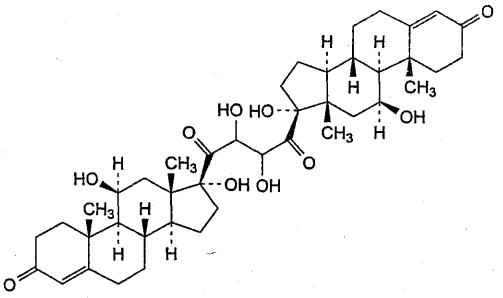
K. 17-hydroxy-3,20-dioxopregn-4-en-21-yl acetate (Reichstein's substance S-21-acetate),



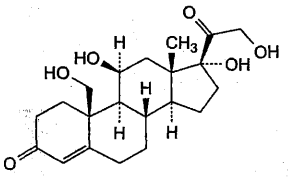
L. 11β,17-dihydroxypregn-4-ene-3,20-dione (oxenol),



M. 11α,17,21-trihydroxypregn-4-ene-3,20-dione (*epi*-hydrocortisone),



N. 11β,17,21-trihydroxy-21-(11β,17,21-trihydroxy-3,20-dioxopregn-4-en-21-yl)pregn-4-ene-3,20-dione (hydrocortisone dimer),

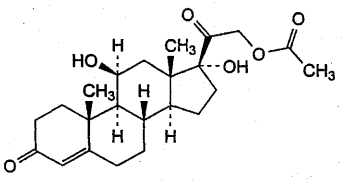


O. 11β,17,19,21-tetrahydroxypregn-4-ene-3,20-dione (19-hydroxyhydrocortisone).

Ph Eur

Hydrocortisone Acetate

(Ph. Eur. monograph 0334)



$C_{23}H_{32}O_6$ 404.5 *50-03-3*

Action and use
Corticosteroid.

Preparations
Clotrimazole and Hydrocortisone Acetate Cream
Gentamicin and Hydrocortisone Acetate Ear Drops
Hydrocortisone Acetate Cream
Hydrocortisone Acetate and Neomycin Ear Drops
Hydrocortisone Acetate and Neomycin Eye Drops
Hydrocortisone Acetate and Neomycin Eye Ointment
Hydrocortisone Acetate Injection
Hydrocortisone Acetate Ointment
Hydrocortisone Acetate Oral Suspension
Miconazole and Hydrocortisone Acetate Cream

Ph Eur

DEFINITION
11β,17-Dihydroxy-3,20-dioxopregn-4-en-21-yl acetate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Practically insoluble in water, slightly soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION
First identification: A, B.

Second identification: C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydrocortisone acetate CRS.*

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (d).

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 25 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of the solution to 10 mL with *methylene chloride R.*

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C protected from light for 2 h 30 min. Allow to cool.

Reference solution (a) Dissolve 25 mg of *hydrocortisone acetate CRS* in *methanol R* and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of the solution to 10 mL with *methylene chloride R.*

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of *saturated methanolic potassium hydrogen carbonate solution R* and immediately pass a stream of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C protected from light for 2 h 30 min. Allow to cool.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R.*

Application 5 μL.

Development Over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R* and heat at 120 °C for 10 min or until the spots appear and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spots in the chromatograms obtained with test solution (b) and reference solution (b) have an R_F value distinctly lower than that of the principal spots in the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min an intense brownish-red colour develops with a green fluorescence which is particularly intense when viewed in ultraviolet light at 365 nm. Add this solution to 10 mL of *water R* and mix. The colour fades and the fluorescence in ultraviolet light does not disappear.

E. About 10 mg gives the reaction of acetyl (*2.3.1*).

TESTS

Specific optical rotation (*2.2.7*)

+ 158 to + 167 (dried substance).

Dissolve 0.250 g in *dioxan R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetic acid R, water R, methanol R* (1:10:90 *V/V/V*).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of *hydrocortisone acetate CRS* and 2 mg of *prednisolone acetate CRS* (impurity C) in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of *hydrocortisone acetate for peak identification CRS* (containing impurities A, B, D, E and G) in 5 mL of the solvent mixture.

Reference solution (d) Dissolve 25.0 mg of *hydrocortisone acetate CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 400 mL of *acetonitrile R* with 550 mL of *water for chromatography R* and allow to equilibrate; dilute to 1000 mL with *water for chromatography R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b) and (c).

Run time 4 times the retention time of hydrocortisone acetate.

Identification of impurities Use the chromatogram supplied with *hydrocortisone acetate for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, D, E and G; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to hydrocortisone acetate (retention time = about 10 min): impurity A = about 0.4; impurity B = about 0.7; impurity C = about 0.9; impurity D = about 1.2; impurity G = about 1.8; impurity E = about 2.3.

System suitability Reference solution (a):

— *resolution*: minimum 1.5 between the peaks due to impurity C and hydrocortisone acetate.

Limits:

— *impurity C*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.6 per cent);

— *impurity A*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurities B, D, E*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurity G*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (d).

Run time 1.5 times the retention time of hydrocortisone acetate.

Retention time Hydrocortisone acetate = about 10 min.

Calculate the percentage content of $C_{23}H_{32}O_6$ taking into account the assigned content of *hydrocortisone acetate CRS*.

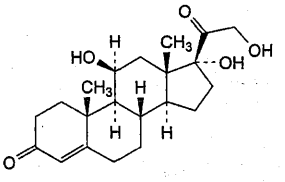
STORAGE

Protected from light.

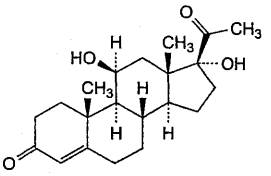
IMPURITIES

Specified impurities *A, B, C, D, E, G.*

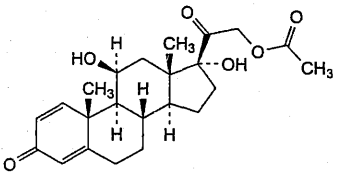
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F.*



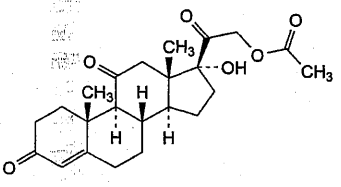
A. 11β,17,21-trihydroxypregn-4-ene-3,20-dione (hydrocortisone),



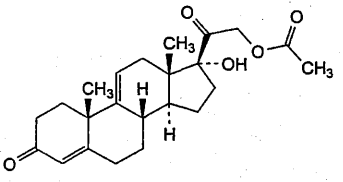
B. 11β,17-dihydroxypregn-4-ene-3,20-dione (oxenol),



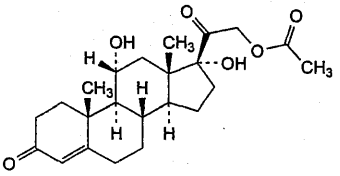
C. 11β,17-dihydroxy-3,20-dioxopregna-1,4-dien-21-yl acetate (prednisolone acetate),



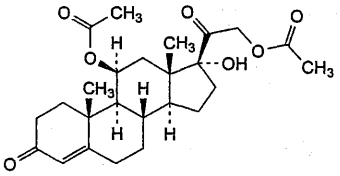
D. 17-hydroxy-3,11,20-trioxopregn-4-en-21-yl acetate (cortisone acetate),



E. 17-hydroxy-3,20-dioxopregna-4,9(11)-dien-21-yl acetate,



F. 11α,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate (*epi*-hydrocortisone acetate),

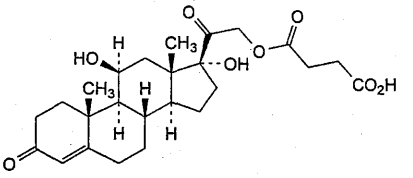


G. 17-hydroxy-3,20-dioxopregn-4-ene-11β,21-diyl diacetate.

Ph Eur

Hydrocortisone Hydrogen Succinate

(Ph. Eur. monograph 0768)



$C_{25}H_{34}O_8$ 462.5 *2203-97-6*

Action and use
Corticosteroid.

Preparations
Hydrocortisone Sodium Succinate Injection
Hydrocortisone Oromucosal Tablets

Ph Eur

DEFINITION

11β,17-Dihydroxy-3,20-dioxopregn-4-en-21-yl hydrogen butanedioate.

Content
97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Practically insoluble in water, freely soluble in acetone and in anhydrous ethanol. It dissolves in dilute solutions of alkali carbonates and alkali hydroxides.

IDENTIFICATION

First identification: A, B.

Second identification: C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dry the substances before use at 100-105 °C for 3 h.

Comparison *hydrocortisone hydrogen succinate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (1:9 *V/V*).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of *hydrocortisone hydrogen succinate CRS* in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of *methylprednisolone hydrogen succinate CRS* in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *anhydrous formic acid R, anhydrous ethanol R, methylene chloride R* (0.1:1:15 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability Reference solution (b):
— the chromatogram shows 2 spots which may, however, not be completely separated.

C. Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 25 mg of the substance to be examined in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of this solution to 10 mL with *methylene chloride R*.

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of a 0.8 g/L solution of *sodium hydroxide R* in *methanol R* and immediately pass a stream of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 30 min. Allow to cool.

Reference solution (a) Dissolve 25 mg of *hydrocortisone hydrogen succinate CRS* in *methanol R* with gentle heating and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of this solution to 10 mL with *methylene chloride R*.

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a polytetrafluoroethylene cap. Add 10 mL of a 0.8 g/L solution of *sodium hydroxide R* in *methanol R* and immediately pass a stream of *nitrogen R* briskly through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 30 min. Allow to cool.

Plate *TLC silica gel F_{254} plate R*.

Mobile phase Add a mixture of 1.2 volumes of *water R* and 8 volumes of *methanol R* to a mixture of 15 volumes of *ether R* and 77 volumes of *methylene chloride R*.

Application 5 µL.

Developpement Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with *alcoholic solution of sulfuric acid R*. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spot in each of the chromatograms obtained with test solution (b) and reference solution (b) has an R_F value distinctly higher than that of the principal spot in each of the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of *sulfuric acid R* and shake to dissolve. Within 5 min, an intense brownish-red colour develops with a green fluorescence which is particularly intense when viewed in ultraviolet light at 365 nm. Add this solution to 10 mL of *water R* and mix. The colour fades and a clear solution remains. The fluorescence in ultraviolet light does not disappear.

TESTS

Appearance of solution

The solution is clear (*2.2.1*).

Dissolve 0.10 g in 5 mL of *sodium hydrogen carbonate solution R*.

Specific optical rotation (*2.2.7*)

+ 147 to + 153 (dried substance).

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in a mixture of equal volumes of *acetonitrile R* and *water R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 2 mg of *hydrocortisone hydrogen succinate CRS* and 2 mg of *dexamethasone CRS* in 50 mL of *acetonitrile R*, then dilute to 100.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of equal volumes of *acetonitrile R* and *water R*.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase In a 1000 mL volumetric flask mix 330 mL of *acetonitrile R* with 600 mL of *water R* and 1.0 mL of *phosphoric acid R*, then allow to equilibrate; dilute to 1000 mL with *water R* and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 30 min.

Injection 20 µL.

Run time Twice the retention time of hydrocortisone hydrogen succinate.

Retention time Dexamethasone = about 12.5 min; hydrocortisone hydrogen succinate = about 15 min.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to dexamethasone and hydrocortisone hydrogen succinate; if necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:
— *impurities A, B*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *total*: not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.75 per cent);
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 4.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

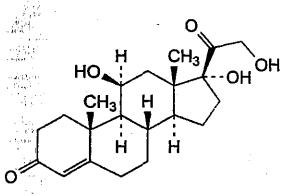
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.100 g in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with *ethanol (96 per cent) R*. Measure the absorbance (*2.2.25*) at the absorption maximum at 241.5 nm.

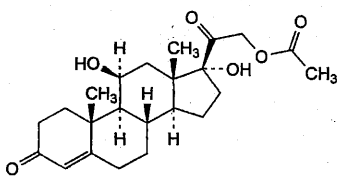
Calculate the content of $C_{25}H_{34}O_8$ taking the specific absorbance to be 353.

STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities A, B.



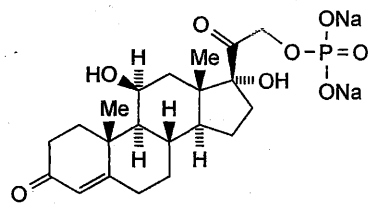
A. 11β,17,21-trihydroxypregn-4-ene-3,20-dione (hydrocortisone),



B. 11β,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate (hydrocortisone acetate).

Ph Eur

Hydrocortisone Sodium Phosphate



$C_{21}H_{29}Na_2O_8P$ 486.4 *6000-74-4*

Action and use
Corticosteroid.

Preparations
Hydrocortisone Sodium Phosphate Injection
Hydrocortisone Sodium Phosphate Oral Solution

DEFINITION
Hydrocortisone Sodium Phosphate is disodium 11β,17α-dihydroxy-3,20-dioxopregn-4-en-21-yl orthophosphate. It contains not less than 96.0% and not more than 103.0% of $C_{21}H_{29}Na_2O_8P$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
A white or almost white powder; hygroscopic.

Freely soluble in *water*; practically insoluble in *absolute ethanol*.

IDENTIFICATION
Test A may be omitted if tests B, C and D are carried out.
Tests B and C may be omitted if tests A and D are carried out.

A. The *infrared absorption spectrum*, Apppendix II A, is concordant with the *reference spectrum* of hydrocortisone sodium phosphate (*RS 386*).

B. Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and a freshly prepared mixture of 60 volumes of *butan-1-ol*, 20 volumes of *acetic anhydride* and 20 volumes of *water* as the mobile phase. Apply separately to the plate 2 µL of each of the following solutions. Solution (1) contains 0.25% w/v of the substance being examined in *methanol*. Solution (2) contains 0.25% w/v of *hydrocortisone sodium phosphate BPCRS* in *methanol*. Solution (3) is a mixture of equal volumes of solutions (1) and (2). Solution (4) is a mixture of equal volumes of solution (1) and a 0.25% w/v solution of *betamethasone sodium phosphate BPCRS* in *methanol*. After removal of the plate, allow it to dry in air until the solvent has evaporated, spray with *ethanolic sulfuric acid (20%)*, heat at 120° for 10 minutes and examine under *ultraviolet light (365 nm)*. The principal spot in the chromatogram obtained with solution (1) corresponds to that in the chromatogram obtained with solution (2). The principal spot in the chromatogram obtained with solution (3) appears as a single, compact spot and the chromatogram obtained with solution (4) shows two principal spots with almost identical R_f values.

C. Dissolve 2 mg in 2 mL of *sulfuric acid*. A yellowish green fluorescence is produced immediately (distinction from betamethasone sodium phosphate, dexamethasone sodium phosphate and prednisolone sodium phosphate).

D. Heat gently 40 mg with 2 mL of *sulfuric acid* until white fumes are evolved, add *nitric acid* dropwise until oxidation is complete and cool. Add 2 mL of *water*, heat until white fumes are again evolved, cool, add 10 mL of *water* and neutralise to *litmus paper* with 5M *ammonia*. The resulting solution yields reaction A characteristic of *sodium salts* and reaction B characteristic of *phosphates*, Appendix VI.

TESTS
Alkalinity
pH of a 0.5% w/v solution, 7.5 to 9.0, Appendix V L.

Specific optical rotation
In a 1% w/v solution, +121 to +129, calculated with reference to the anhydrous substance, Appendix V F.

Inorganic phosphate
Dissolve 25 mg in 10 mL of *water*, add 4 mL of 1M *sulfuric acid*, 1 mL of a 10% w/v solution of *ammonium molybdate* and 2 mL of *methylaminophenol-sulfite reagent* and allow to stand for 15 minutes. Add sufficient *water* to produce 25 mL and allow to stand for a further 15 minutes. The *absorbance* of a 4-cm layer of the resulting solution at 730 nm, Appendix II B, is not more than that of a 4-cm layer of a solution prepared by treating 10 mL of a 0.0036% w/v solution of *potassium dihydrogen orthophosphate* in the same manner, beginning at the words 'add 4 mL ...'.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel* GF_{254} as the coating substance and a mixture of 77 volumes of *dichloromethane*, 15 volumes of *ether*, 8 volumes of *methanol* and 1.2 volumes

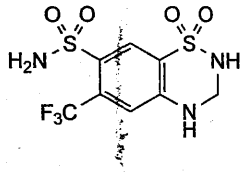
of *water* as the mobile phase. Apply separately to the plate 2 µL of each of three solutions in *methanol* containing (1) 1.0% w/v of the substance being examined, (2) 1.0% w/v of *hydrocortisone sodium phosphate BPCRS* and (3) 0.020% w/v of *hydrocortisone BPCRS*. After removal of the plate, allow it to dry in air for 5 minutes and examine under *ultraviolet light (254 nm)*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3) (2%).

Water
Not more than 10.0%, Appendix IX C. Use 0.4 g.

ASSAY
Dissolve 0.1 g in sufficient *water* to produce 200 mL. Dilute 5 mL to 100 mL with *water* and measure the *absorbance* of the resulting solution at the maximum at 248 nm, Appendix II B. Calculate the content of $C_{21}H_{29}Na_2O_8P$ taking 333 as the value of A(1%, 1 cm) at the maximum at 248 nm.

STORAGE
Hydrocortisone Sodium Phosphate should be protected from light.

Hydroflumethiazide



$C_8H_8F_3N_3O_4S_2$ 331.3 *135-09-1*

Action and use
Thiazide diuretic.

DEFINITION
Hydroflumethiazide is 3,4-dihydro-6-trifluoromethyl-2*H*-1,2,4-benzothiadiazine-7sulfonamide 1,1-dioxide. It contains not less than 98.0% and not more than 102.0% of $C_8H_8F_3N_3O_4S_2$, calculated with reference to the dried substance.

CHARACTERISTICS
White or almost white, glistening crystals or crystalline powder.

Practically insoluble in *water*; soluble in *ethanol (96%)*; practically insoluble in *ether*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of hydroflumethiazide *(RS 181)*.

B. Dissolve 10 mg in 10 mL of 0.1M *sodium hydroxide*, add sufficient *water* to produce 100 mL and dilute 10 mL to 50 mL with 0.01M *sodium hydroxide*. The *light absorption* of the resulting solution, Appendix II B, in the range 230 to 350 nm exhibits two maxima, at 274 nm and 333 nm. The *absorbance* at the maxima is about 0.92 and about 0.19 respectively.

C. Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel* GF_{254} as the coating substance and *ethyl acetate* as the mobile phase. Apply separately to the plate 5 µL of each of two solutions in *acetone* containing (1) 0.1% w/v of the substance being examined and (2) 0.1% w/v of *hydroflumethiazide BPCRS*. After removal of the plate, dry it in a current of air, examine under *ultraviolet light (254 nm)* and then reveal the spots by *Method I* and examine again. By each method of visualisation the principal spot in the chromatogram obtained with solution (1) corresponds in colour and intensity to that in the chromatogram obtained with solution (2).

TESTS
Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and *ethyl acetate* as the mobile phase. Apply separately to the plate 10 µL of each of two solutions of the substance being examined in *acetone* containing (1) 1.0% w/v and (2) 0.010% w/v. After removal of the plate, dry it in a current of air and reveal the spots by *Method I*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Loss on drying
When dried to constant weight at 105°C, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.3 g in 50 mL of *anhydrous pyridine* and carry out Method II for *non-aqueous titration*, Appendix VIII A, using 0.1M *tetrabutylammonium hydroxide VS* as titrant and determining the end point potentiometrically. Each mL of 0.1M *tetrabutylammonium hydroxide VS* is equivalent to 16.56 mg of $C_8H_8F_3N_3O_4S_2$.

Hydrogen Peroxide Solution (3 per cent)

Dilute Hydrogen Peroxide Solution
(Ph. Eur. monograph 0395)

Action and use
Antiseptic; deodorant.

When hydrogen peroxide is prescribed or demanded, Hydrogen Peroxide Solution (6 per cent) shall be dispensed or supplied.

Ph Eur

DEFINITION
Content
2.5 per cent *m/m* to 3.5 per cent *m/m* of H_2O_2 (M_r 34.01).

1 volume of hydrogen peroxide solution (3 per cent) corresponds to about 10 times its volume of oxygen.
A suitable stabiliser may be added.

CHARACTERS
Appearance
Colourless, clear liquid.

IDENTIFICATION
A. To 2 mL, add 0.2 mL of *dilute sulfuric acid R* and 0.2 mL of *0.02 M potassium permanganate*. The solution becomes colourless or slightly pink within 2 min.

B. To 1 mL, add 0.1 mL of *dilute hydrochloric acid R* and 0.1 mL of *potassium iodide solution R*. A brown colour appears. Black particles may be formed.

C. It complies with the requirement for the content of H_2O_2.

TESTS

Acidity

To 10 mL, add 20 mL of *water R* and 0.25 mL of *methyl red solution R*. Not less than 0.05 mL and not more than 1.0 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Organic stabilisers

Maximum 250 ppm.

Shake 20 mL with 10 mL of *chloroform R* and then with 2 quantities, each of 5 mL, of *chloroform R*. Evaporate the combined chloroform layers under reduced pressure at a temperature not exceeding 25 °C and dry in a desiccator. The residue weighs a maximum of 5 mg.

Non-volatile residue

Maximum 2 g/L.

Allow 10 mL to stand in a platinum dish until all effervescence has ceased. Evaporate to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 20 mg.

ASSAY

Dilute 10.0 g to 100.0 mL with *water R*. To 10.0 mL of this solution add 20 mL of *dilute sulfuric acid R*. Titrate with *0.02 M potassium permanganate* until a pink colour is obtained.

1 mL of *0.02 M potassium permanganate* is equivalent to 1.701 mg of H_2O_2 or 0.56 mL of oxygen.

STORAGE

Protected from light, and if the solution does not contain a stabiliser, at a temperature below 15 °C.

LABELLING

If the solution contains a stabiliser, the label states that the contents are stabilised. The competent authority may require that the name of the stabiliser be stated on the label.

CAUTION

It decomposes in contact with oxidisable organic matter and with certain metals and if allowed to become alkaline.

Ph Eur

Hydrogen Peroxide Solution (6 per cent)

Hydrogen Peroxide Solution

Action and use

Antiseptic; deodorant.

Preparations

Hydrogen Peroxide Mouthwash

When hydrogen peroxide is prescribed or demanded, Hydrogen Peroxide Solution (6 per cent) shall be dispensed or supplied.

DEFINITION

Hydrogen Peroxide Solution (6 per cent) is an aqueous solution of hydrogen peroxide containing not less than 5.0% w/v and not more than 7.0% w/v of H_2O_2 (34.01), corresponding to about 20 times its volume of available oxygen. It may contain a suitable stabilising agent.

CHARACTERISTICS

A clear, colourless liquid. It decomposes in contact with oxidisable organic matter and with certain metals and if allowed to become alkaline.

IDENTIFICATION

A. To 1 mL add 0.2 mL of 1M *sulfuric acid* and 0.25 mL of 0.02M *potassium permanganate*. The solution becomes colourless with evolution of gas.

B. Shake 0.05 mL with 2 mL of 1M *sulfuric acid*, 2 mL of *ether* and 0.05 mL of *potassium chromate solution*. The ether layer is blue.

C. Complies with the requirement for the content of H_2O_2.

TESTS

Acidity

Dilute 10 mL with 20 mL of *water* and add 0.25 mL of *methyl red solution*. Not less than 0.05 mL and not more than 1.0 mL of *0.1M sodium hydroxide VS* is required to change the colour of the solution.

Organic stabilisers

Shake 20 mL with successive quantities of 10, 5 and 5 mL of *chloroform*. Evaporate the combined chloroform extracts at a temperature not exceeding 25° at a pressure of 2 kPa and dry in a desiccator. Any residue weighs not more than 5 mg (250 ppm).

Non-volatile matter

Place 10 mL in a platinum dish and allow to stand until effervescence has ceased, cooling if necessary. Evaporate the solution on a water bath. Any residue, when dried at 100° to 105°, weighs not more than 20 mg (0.2% w/v).

ASSAY

Dilute 10 mL to 100 mL with *water*. To 10 mL of the resulting solution add 20 mL of 1M *sulfuric acid* and titrate with 0.02M *potassium permanganate VS*. Each mL of 0.02M *potassium permanganate VS* is equivalent to 1.701 mg of H_2O_2 or 0.56 mL of oxygen.

STORAGE

Hydrogen Peroxide Solution (6 per cent) should be protected from light. If the solution does not contain a stabilising agent, it should be stored at a temperature not exceeding 15°. It should not be stored for long periods.

LABELLING

The label states, where applicable, that the solution contains a stabilising agent.

Hydrogen Peroxide Solution (30 per cent)

(Ph. Eur. monograph 0396)

7722-84-1

Action and use

Antiseptic; deodorant.

When hydrogen peroxide is prescribed or demanded, Hydrogen Peroxide Solution (6 per cent) shall be dispensed or supplied.

Ph Eur

DEFINITION

Content

29.0 per cent *m/m* to 31.0 per cent *m/m* of H_2O_2 (M_r 34.01).

1 volume of hydrogen peroxide solution (30 per cent) corresponds to about 110 times its volume of oxygen.

A suitable stabiliser may be added.

CHARACTERS
Appearance
Colourless, clear liquid.

IDENTIFICATION
A. To 1 mL, add 0.2 mL of *dilute sulfuric acid R* and 0.25 mL of *0.02 M potassium permanganate*. The solution becomes colourless with evolution of gas.

B. To 1 mL, add 0.1 mL of *dilute hydrochloric acid R* and 0.1 mL of *potassium iodide solution R*. A brown colour appears. Black particles may be formed.

C. It complies with the requirement for the content of H_2O_2.

TESTS
Acidity
To 10 mL, add 100 mL of *water R* and 0.25 mL of *methyl red solution R*. Not less than 0.05 mL and not more than 0.5 mL of *0.1 M sodium hydroxide* is required to change the colour of the indicator.

Organic stabilisers
Maximum 500 ppm.

Shake 20 mL with 10 mL of *chloroform R* and then with 2 quantities, each of 5 mL, of *chloroform R*. Evaporate the combined chloroform layers under reduced pressure at a temperature not exceeding 25 °C and dry in a desiccator. The residue weighs a maximum of 10 mg.

Non-volatile residue
Maximum 2 g/L.

Allow 10 mL to stand in a platinum dish until all effervescence has ceased, cooling if necessary. Evaporate to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 20 mg.

ASSAY
Dilute 1.00 g to 100.0 mL with *water R*. To 10.0 mL of this solution add 20 mL of *dilute sulfuric acid R*. Titrate with *0.02 M potassium permanganate* until a pink colour is obtained.

1 mL of *0.02 M potassium permanganate* is equivalent to 1.701 mg of H_2O_2 or 0.56 mL of oxygen.

STORAGE
Protected from light, and if the solution does not contain a stabiliser, at a temperature below 15 °C.

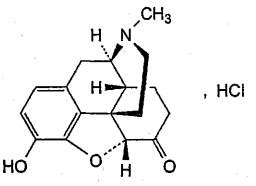
LABELLING
If the solution contains a stabiliser, the label states that the contents are stabilised. The competent authority may require that the name of the stabiliser be stated on the label.

CAUTION
It decomposes vigorously in contact with oxidisable organic matter and with certain metals and if allowed to become alkaline.

Ph Eur

Hydromorphone Hydrochloride

(Ph. Eur. monograph 2099)



$C_{17}H_{20}ClNO_3$ 321.8 *71-68-1*

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION
4,5α-Epoxy-3-hydroxy-17-methylmorphinan-6-one hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydromorphone hydrochloride CRS*.

B. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 1.250 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_5 (*2.2.2, Method II*).

Acidity or alkalinity
To 2 mL of solution S add 0.1 mL of *methyl red solution R*. The solution is not yellow. To 2 mL of solution S add 0.05 mL of *bromocresol green solution R*. The solution is not yellow.

Specific optical rotation (*2.2.7*)
−136 to −140 (dried substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in *water R*, sonicating if necessary and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) To 5 mL of the test solution add 5 mg of *naloxone hydrochloride dihydrate CRS* and dilute to 50 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Dissolve 18.29 g of *diethylamine R* and 2.88 g of *sodium laurilsulfate R* in *water R* and dilute to 1000 mL with the same solvent. Adjust 800 mL of this solution to pH 3.0 with *phosphoric acid R*. Add 100 mL of *acetonitrile R* and 100 mL of *methanol R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 284 nm.

Injection 20 µL.

Run time 4 times the retention time of hydromorphone.

Relative retention With reference to hydromorphone (retention time = about 9 min): impurity D = about 0.72; impurity B = about 0.77; impurity C = about 0.82; impurity A = about 3.2.

System suitability Reference solution (b):
— *resolution*: minimum 4.0 between the peaks due to hydromorphone and naloxone.

Limits:
— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *impurities B, C, D*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

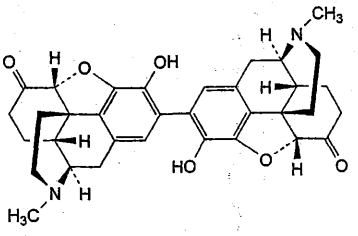
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY
Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

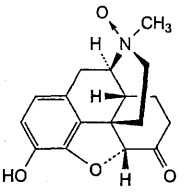
1 mL of *0.1 M sodium hydroxide* is equivalent to 32.18 mg of $C_{17}H_{20}ClNO_3$.

STORAGE
Protected from light.

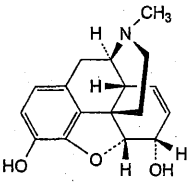
IMPURITIES
Specified impurities *A, B, C, D.*



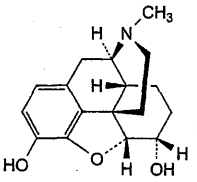
A. 4,5α:4′,5′α-diepoxy-3,3′-dihydroxy-17,17′-dimethyl-2,2′-bimorphinanyl-6,6′-dione (pseudohydromorphone),



B. 4,5α-epoxy-3-hydroxy-17-methylmorphinan-6-one 17-oxide (hydromorphone *N*-oxide),



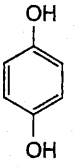
C. 7,8-didehydro-4,5α-epoxy-17-methylmorphinan-3,6α-diol (morphine),



D. 4,5α-epoxy-17-methylmorphinan-3,6α-diol (dihydromorphine).

Ph Eur

Hydroquinone



$C_6H_6O_2$ 110.11 *123-31-9*

Action and use
Depigmenting agent.

Preparation
Tretinoin, Hydrocortisone and Hydroquinone Cream

DEFINITION
Hydroquinone is 1,4-benzenediol. It contains not less than 99.0% and not more than 100.5% of $C_6H_6O_2$, calculated with reference to the anhydrous substance.

CHARACTERISTICS
Fine, white or almost white needles, which darken on exposure to light and air.

Freely soluble in *water*, in *ethanol (96%)* and in *ether*.

IDENTIFICATION
A. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of hydroquinone *(RS 495)*.

B. Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions.

(1) Dissolve the substance being examined in *methanol* to produce a solution containing 0.1% w/v of Hydroquinone.

(2) 0.1% w/v of *hydroquinone BPCRS* in *methanol.*

(3) Equal volumes of solutions (1) and (2).

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating *silica gel* F_{254} (Merck silica gel 60 F_{254} plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 5 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, allow it to dry in air until the solvent has evaporated and heat at 105° for 15 minutes. Allow to cool and examine under *ultraviolet light (254 nm).*

MOBILE PHASE

Equal volumes of *dichloromethane* and *methanol.*

SYSTEM SUITABILITY

The test is not valid unless the principal spot in the chromatogram obtained with solution (3) appears as a single, compact spot.

CONFIRMATION

The principal spot in the chromatogram obtained with solution (1) corresponds to that in the chromatogram obtained with solution (2).

TESTS

Melting point
172° to 174°, Appendix V A.

Water
Not more than 0.5%, Appendix IX C, Method I.

Sulfated ash
Not more than 0.5% w/w, Appendix IX A.

ASSAY

Dissolve 0.25 g in a mixture of 100 mL of *water* and 10 mL of 0.1M *sulfuric acid,* add 3 drops of *diphenylamine* and titrate with 0.1M *cerium(IV) sulfate VS* until a red-violet colour is reached. Each mL of 0.1M *cerium(IV) sulfate VS* is equivalent to 5.506 mg of $C_6H_6O_2$.

STORAGE

Hydroquinone should be protected from light.

Hydrotalcite

$Mg_6Al_2(OH)_{16}CO_3,4H_2O$ 604.0 *12304-65-3*

Action and use
Antacid.

Preparation
Hydrotalcite Tablets

DEFINITION

Hydrotalcite is a hydrated form of an aluminium magnesium basic carbonate corresponding to the formula $Mg_6Al_2(OH)_{16}CO_3,4H_2O$. It contains not less than 15.3% and not more than 18.7% of Al_2O_3 and not less than 36.0% and not more than 44.0% of MgO. The ratio of the content of Al_2O_3 to the content of MgO is not less than 0.40 and not more than 0.45.

CHARACTERISTICS

A white or almost white, free-flowing, granular powder.

Practically insoluble in *water.* It dissolves in dilute mineral acids with slight effervescence.

IDENTIFICATION

A. Dissolve 1.0 g in 20 mL of 2M *hydrochloric acid.* Effervescence occurs. Add 30 mL of *water,* boil, add 2M *ammonia* until just alkaline to *methyl red solution,* continue boiling for 2 minutes and filter, reserving the filtrate for test B. Wash the precipitate with 50 mL of a hot 2% w/v solution of *ammonium chloride* and dissolve in 15 mL of 2M *hydrochloric acid.* The resulting solution yields the reaction characteristic of *aluminium salts,* Appendix VI.

B. Dilute 1 mL of the filtrate obtained in test A to 10 mL with *water.* The resulting solution yields the reactions characteristic of *magnesium salts,* Appendix VI.

TESTS

Alkalinity
pH of a 4% w/v suspension in *carbon dioxide-free water,* 8.0 to 10.0, Appendix V L.

Neutralising capacity
Mix 0.2 g with a small quantity of *water* to give a smooth paste and gradually add sufficient further quantities of *water* to produce 100 mL. Warm at 37°, add 100 mL of *0.1M hydrochloric acid VS* previously heated to 37° and stir continuously for 1 hour using a paddle stirrer at a rate of about 200 revolutions per minute, maintaining the temperature at 37°, and titrate with *0.1M sodium hydroxide VS* to pH 3.5. Subtract the volume of *0.1M sodium hydroxide VS* from 100 mL to obtain the number of mL of *0.1M hydrochloric acid VS* required for neutralisation. Not less than 260 mL of *0.1M hydrochloric acid VS* is required to neutralise 1 g.

Arsenic
Dissolve 0.33 g in 5 mL of 2M *hydrochloric acid.* The resulting solution complies with the *limit test for arsenic,* Appendix VII (3 ppm).

Sodium
Not more than 0.1% of Na when determined by Method II for *atomic emission spectrophotometry,* Appendix II D, measuring at 589 nm. To prepare the test solution dissolve 0.1 g in 4 mL of *5M hydrochloric acid,* dilute to 200 mL with *water* and use *sodium standard solution (200 ppm Na),* diluted if necessary with *0.1M hydrochloric acid,* to prepare the standard solutions.

Chloride
Dissolve 0.18 g in 10 mL of 2M *nitric acid,* boil, allow to cool and dilute to 100 mL with *water.* To 10 mL add 5 mL of *water.* The resulting solution complies with the *limit test for chlorides,* Appendix VII (0.3%).

Sulfate
Dissolve 0.14 g in 15 mL of *1M hydrochloric acid* and dilute to 100 mL with *water.* 15 mL of the resulting solution complies with the *limit test for sulfates,* Appendix VII (0.7%).

Loss on ignition
When ignited at 800°, loses 40.0 to 50.0% of its weight. Use 1 g.

ASSAY

For Al_2O_3
Dissolve 0.3 g in 2 mL of *7M hydrochloric acid,* add 250 mL of *water* and 50 mL of 0.05M *disodium edetate VS* and neutralise with 1M *sodium hydroxide* using *methyl red solution* as indicator. Heat the solution on a water bath for 30 minutes and allow to cool. Add 3 g of *hexamine* and titrate the excess of disodium edetate with 0.05M *lead nitrate*

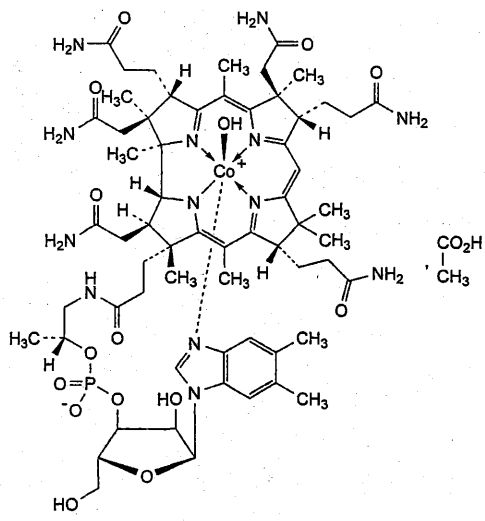
VS using *xylenol orange solution* as indicator. Each mL of 0.05M *disodium edetate VS* is equivalent to 2.549 mg of Al_2O_3.

For MgO

Dissolve 0.125 g in the minimum volume of *7M hydrochloric acid*, add 30 mL of *water*, 1 g of *ammonium chloride*, 10 mL of *triethanolamine*, 150 mL of *water* and 5 mL of *ammonia buffer pH 10.9* and titrate immediately with 0.05M *disodium edetate VS* using *mordant black 11 solution* as indicator. Each mL of 0.05M *disodium edetate VS* is equivalent to 2.015 mg of MgO.

Hydroxocobalamin Acetate

(Ph. Eur. monograph 0913)



$C_{64}H_{93}CoN_{13}O_{17}P$ 1406 *22465-48-1*

Action and use

Vitamin B_{12} analogue.

Preparation

Hydroxocobalamin Injection

Ph Eur

DEFINITION

Coα-[α-(5,6-dimethylbenzimidazolyl)]-Coβ-hydroxocobamide acetate.

Content

96.0 per cent to 102.0 per cent (dried substance).

This monograph applies to hydroxocobalamin acetate produced by fermentation.

CHARACTERS

Appearance

Dark red, crystalline powder or dark red crystals, very hygroscopic.

Solubility

Soluble in water.

Some decomposition may occur on drying.

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 2.5 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R*, then dilute to 100 mL with the same solution.

Spectral range 260-610 nm.

Absorption maxima At 274 nm, 351 nm and 525 nm.

Absorbance ratio:

— A_{274}/A_{351} = 0.75 to 0.83;

— A_{525}/A_{351} = 0.31 to 0.35.

B. Thin-layer chromatography (*2.2.27*). *Carry out the test protected from light.*

Test solution Dissolve 2 mg of the substance to be examined in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Reference solution Dissolve 2 mg of *hydroxocobalamin CRS* in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Plate *TLC silica gel G plate R.*

Mobile phase *dilute ammonia R1, methanol R* (25:75 *V/V*).

Application 10 µL.

Development In an unlined tank, over a path of 12 cm.

Drying In air.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of acetates (*2.3.1*).

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions and protect them from bright light.*

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 25 mg of the substance to be examined in 10 mL of *water R*, warming if necessary. Allow to cool and add 1 mL of a 20 g/L solution of *chloramine R* and 0.5 mL of *0.05 M hydrochloric acid*. Dilute this solution to 25 mL with *water R*. Shake and allow to stand for 5 min. Inject immediately.

Column:

— *size*: *l* = 0.25 m, Ø = 4 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 19.5 volumes of *methanol R* and 80.5 volumes of a solution containing 15 g/L of *citric acid monohydrate R* and 8.1 g/L of *disodium hydrogen phosphate dodecahydrate R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 351 nm.

Injection 20 µL.

Run time 4 times the retention time of hydroxocobalamine.

System suitability:

— the chromatogram obtained with reference solution (c) shows 3 principal peaks;

— *resolution*: minimum 3.0 between each pair of adjacent peaks in the chromatogram obtained with reference solution (c);
— *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (5 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying (*2.2.32*)
8.0 per cent to 12.0 per cent, determined on 0.400 g by drying at 105 °C at a pressure not exceeding 0.7 kPa.

ASSAY
Protect the solutions from light throughout the assay Dissolve 25.0 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R*, then dilute to 1000.0 mL with the same solution. Measure the absorbance (*2.2.25*) at the absorption maximum at 351 nm.

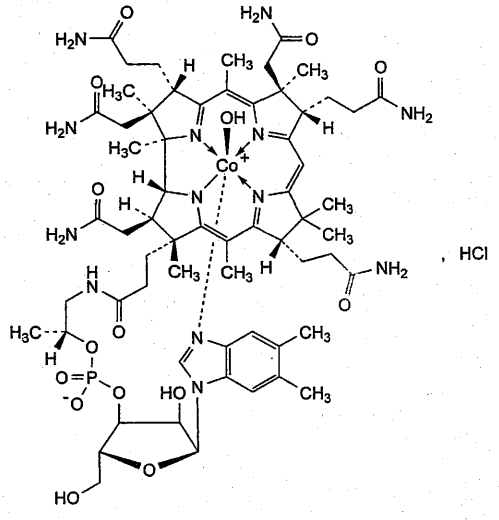
Calculate the content of $C_{64}H_{93}CoN_{13}O_{17}P$ taking the specific absorbance to be 187.

STORAGE
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

Ph Eur

Hydroxocobalamin Chloride

(Ph. Eur. monograph 0914)



$C_{62}H_{90}ClCoN_{13}O_{15}P$ 1383 *58288-50-9*

Action and use
Vitamin B_{12} analogue.

Preparation
Hydroxocobalamin Injection

Ph Eur

DEFINITION
Coα-[α-(5,6-dimethylbenzimidazolyl)]-Coβ-hydroxocobamide chloride.

Fermentation product.

Content
96.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Dark red crystalline powder or dark red crystals, very hygroscopic.

Solubility
Soluble in water.

Some decomposition may occur on drying.

IDENTIFICATION
A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 2.5 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R*, then dilute to 100 mL with the same solution.

Spectral range 260-610 nm.

Absorption maxima At 274 nm, 351 nm and 525 nm.

Absorbance ratio:
— A_{274}/A_{351} = 0.75 to 0.83;
— A_{525}/A_{351} = 0.31 to 0.35.

B. Thin-layer chromatography (*2.2.27*). *Carry out the identification test protected from light.*

Test solution Dissolve 2 mg of the substance to be examined in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Reference solution Dissolve 2 mg of *hydroxocobalamin CRS* in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Plate *TLC silica gel G plate R*.

Mobile phase *dilute ammonia R1, methanol R* (25:75 *V/V*).

Application 10 µL.

Development In an unlined tank, over a path of 12 cm.

Drying In air.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Use freshly prepared solutions and protect them from bright light.*

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 25 mg of the substance to be examined in 10 mL of *water R*, warming if necessary. Allow to cool and add 1 mL of a 20 g/L solution of *chloramine R* and 0.5 mL of *0.05 M hydrochloric acid*. Dilute to 25 mL with *water R*. Shake and allow to stand for 5 min. Inject immediately.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;

— *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 19.5 volumes of *methanol R* and 80.5 volumes of a solution containing 15 g/L of *citric acid monohydrate R* and 8.1 g/L of *disodium hydrogen phosphate dodecahydrate R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 351 nm.

Injection 20 µL.

Run time 4 times the retention time of hydroxocobalamin.

System suitability:

— the chromatogram obtained with reference solution (c) shows 3 principal peaks;

— *resolution*: minimum 3.0 between each pair of adjacent peaks in the chromatogram obtained with reference solution (c);

— *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:

— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (5.0 per cent);

— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying *(2.2.32)*

8.0 per cent to 12.0 per cent, determined on 0.400 g by drying at 105 °C at a pressure not exceeding 0.7 kPa.

ASSAY

Protect the solutions from light throughout the assay Dissolve 25.0 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R*, then dilute to 1000.0 mL with the same solution. Measure the absorbance *(2.2.25)* at the absorption maximum at 351 nm.

Calculate the content of $C_{62}H_{90}ClCoN_{13}O_{15}P$ taking the specific absorbance to be 190.

STORAGE

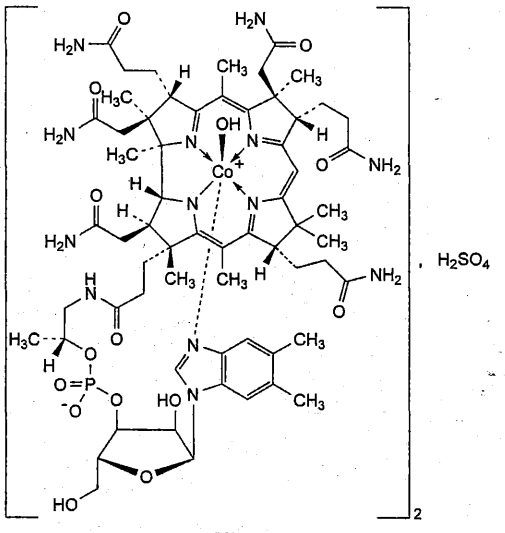
In an airtight container protected from light, at a temperature of 2 °C to 8 °C.

Ph Eur

Hydroxocobalamin Sulfate

Hydroxocobalamin Sulphate

(Ph. Eur. monograph 0915)



$C_{124}H_{180}Co_2N_{26}O_{34}P_2S$ 2791

Action and use

Vitamin B_{12} analogue.

Preparation

Hydroxocobalamin Injection

Ph Eur

DEFINITION

Di-(Coα-[α-(5,6-dimethylbenzimidazolyl)]-Coβ-hydroxocobamide) sulfate.

Fermentation product.

Content

96.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Dark red crystalline powder or dark red crystals, very hygroscopic.

Solubility

Soluble in water.

Some decomposition may occur on drying.

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry *(2.2.25)*.

Test solution Dissolve 2.5 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R*, then dilute to 100 mL with the same solution.

Spectral range 260-610 nm.

Absorption maxima At 274 nm, 351 nm and 525 nm.

Absorbance ratios:

— A_{274}/A_{351} = 0.75 to 0.83;

— A_{525}/A_{351} = 0.31 to 0.35.

B. Thin-layer chromatography *(2.2.27)*. *Carry out the test protected from light.*

Test solution Dissolve 2 mg of the substance to be examined in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Reference solution Dissolve 2 mg of *hydroxocobalamin CRS* in 1 mL of a mixture of equal volumes of *ethanol (96 per cent) R* and *water R*.

Plate *TLC silica gel G plate R.*

Mobile phase *dilute ammonia R1, methanol R* (25:75 *V/V*).

Application 10 µL.

Development In an unlined tank, over a path of 12 cm.

Drying In air.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions and protect them from bright light.*

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 25 mg of the substance to be examined in 10 mL of *water R*, warming if necessary. Allow to cool and add 1 mL of a 20 g/L solution of *chloramine R* and 0.5 mL of *0.05 M hydrochloric acid*. Dilute to 25 mL with *water R*. Shake and allow to stand for 5 min. Inject immediately.

Column:
- *size*: *l* = 0.25 m, Ø = 4 mm;
- *stationary phase*: *octylsilyl silica gel for chromatography R* (5 µm).

Mobile phase Mix 19.5 volumes of *methanol R* and 80.5 volumes of a solution containing 15 g/L of *citric acid monohydrate R* and 8.1 g/L of *disodium hydrogen phosphate dodecahydrate R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 351 nm.

Injection 20 µL.

Run time 4 times the retention time of hydroxocobalamin.

System suitability:
- the chromatogram obtained with reference solution (c) shows 3 principal peaks;
- *resolution*: minimum 3.0 between each pair of adjacent peaks in the chromatogram obtained with reference solution (c);
- *signal-to-noise ratio*: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:
- *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (5.0 per cent);
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Loss on drying (*2.2.32*)

8.0 per cent to 16.0 per cent, determined on 0.400 g by drying at 105 °C at a pressure not exceeding 0.7 kPa.

ASSAY

Protect the solutions from light throughout the assay Dissolve 25.0 mg in a solution containing 0.8 per cent *V/V* of *glacial acetic acid R* and 10.9 g/L of *sodium acetate R* and dilute to 1000.0 mL with the same solution. Measure the absorbance (*2.2.25*) at the absorption maximum at 351 nm.

Calculate the content of $C_{124}H_{180}Co_2N_{26}O_{34}P_2S$ taking the specific absorbance to be 188.

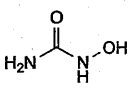
STORAGE

In an airtight container protected from light, at a temperature of 2 °C to 8 °C.

Ph Eur

Hydroxycarbamide

(Ph. Eur. monograph 1616)



$CH_4N_2O_2$ 76.1 *127-07-1*

Action and use

Cytotoxic alkylating drug.

Preparation

Hydroxycarbamide Capsules

Ph Eur

DEFINITION

N-Hydroxyurea.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydroxycarbamide CRS.*

If the spectra obtained in the solid state show differences dissolve the substance to be examined and the reference substance separately in *ethanol (96 per cent) R*, evaporate to dryness and record new spectra using the residues.

B. Examine the chromatograms obtained in the test for urea.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (c).

TESTS

Urea

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *water R* and dilute to 1.0 mL with the same solvent.

Reference solution (a) Dissolve 12.5 mg of *urea R* in *water R* and dilute to 50 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *urea R* in *water R* and dilute to 20 mL with the same solvent.

Reference solution (c) Dissolve 50 mg of *hydroxycarbamide CRS* in *water R* and dilute to 1 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *pyridine R, water R, ethyl acetate R* (2:2:10 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 10 g/L solution of *dimethylaminobenzaldehyde R* in *1 M hydrochloric acid.*

System suitability The test is not valid unless the chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Limit:

— *urea*: any spot corresponding to urea in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with the reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Test solution (a) Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the same mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 0.100 g of *hydroxylamine hydrochloride R* and 5 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase. Prepare immediately before use.

Reference solution (b) Dilute 0.1 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 0.100 g of *hydroxycarbamide CRS* in the mobile phase and dilute to 10.0 mL with the same solvent. Dilute 5.0 mL to 50.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm,

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R, water R* (5:95 *V/V*).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL; inject test solution (a) and reference solutions (a) and (b).

Run time 3 times the retention time of hydroxycarbamide which is about 5 min.

System suitability Reference solution (a):

— *resolution*: minimum of 1.0 between the peaks due to impurity A and to hydroxycarbamide.

Limits:

— *any impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),

— *total*: not more than 2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),

— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent).

Chlorides (*2.4.4*)

Maximum 50 ppm.

Dissolve 1.0 g in *water R* and dilute to 15 mL with the same solvent.

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances.

Injection Test solution (b) and reference solution (c).

STORAGE

In an airtight container, protected from light.

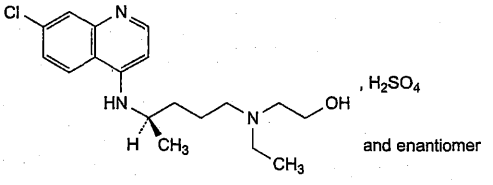
IMPURITIES

A. H_2N-OH: hydroxylamine.

Ph Eur

Hydroxychloroquine Sulfate

(Ph. Eur. monograph 2849)



$C_{18}H_{28}ClN_3O_5S$ 434.0 *747-36-4*

Action and use

Antiprotozoal (malaria).

Ph Eur

DEFINITION

2-[[(4*RS*)-4-[(7-Chloroquinolin-4-yl)amino]pentyl](ethyl)amino]ethan-1-ol sulfate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydroxychloroquine sulfate CRS.*

B. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

Appearance of solution

The solution is clear and not more intensely coloured than reference solution Y_7 (*2.2.2, Method I*).

Dissolve 1.0 g in *water R* and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture Mix 500 mL of *water R*, 500 mL of *methanol R* and 4 mL of a 10 per cent *V/V* solution of *sulfuric acid R*.

Buffer solution Dissolve 1.36 g of *potassium dihydrogen phosphate R* in 900 mL of *water R*. Add 0.15 g of *sodium heptanesulfonate R*, adjust to pH 7.0 with *triethylamine R* and dilute to 1 L with *water R*.

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of *hydroxychloroquine for system suitability CRS* (containing impurities B and C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of *hydroxychloroquine sulfate CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.05 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped ethylene-bridged octadecylsilyl silica gel for chromatography (hybrid material) R* (1.7 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: *methanol R*, buffer solution (10:90 *V/V*);
- *mobile phase B*: buffer solution, *methanol R* (15:85 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	100	0
1 - 11	100 → 0	0 → 100

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 4 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *hydroxychloroquine for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and C.

Relative retention With reference to hydroxychloroquine (retention time = about 6 min): impurity B = about 0.8; impurity C = about 0.9.

System suitability Reference solution (a):
- *resolution*: minimum 3.0 between the peaks due to impurity C and hydroxychloroquine; minimum 3.0 between the peaks due to impurities B and C.

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity B by 1.6;
- for each impurity, use the concentration of hydroxychloroquine in reference solution (b).

Limits:
- *impurity C*: maximum 0.4 per cent;
- *impurity B*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.6 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

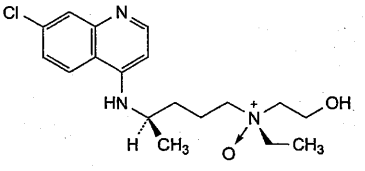
Injection 4 µL of test solution (b) and reference solution (c).

Calculate the percentage content of $C_{18}H_{28}ClN_3O_5S$ taking into account the assigned content of *hydroxychloroquine sulfate CRS*.

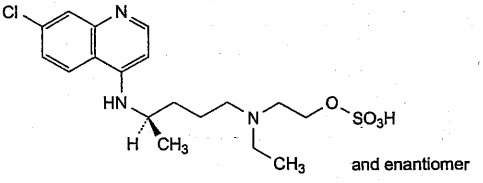
IMPURITIES

Specified impurities *B, C.*

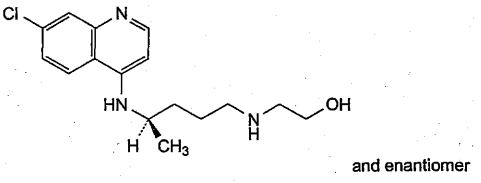
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D, E, F, G.*



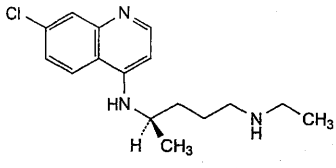
A. mixture of diastereoisomers of 4-[(7-chloroquinolin-4-yl) amino]-*N*-ethyl-*N*-(hydroxyethyl)pentan-1-amine *N*-oxide,



B. 2-[[(4*RS*)-4-[(7-chloroquinolin-4-yl)amino]pentyl](ethyl) amino]ethyl hydrogen sulfate,

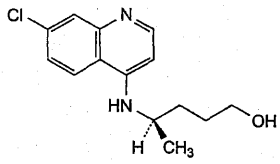


C. 2-[[(4*RS*)-4-[(7-chloroquinolin-4-yl)amino]pentyl]amino] ethan-1-ol,



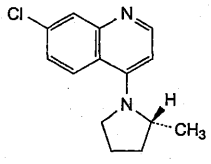
and enantiomer

D. (4*RS*)-N^4-(7-chloroquinolin-4-yl)-N^1-ethylpentane-1,4-diamine,



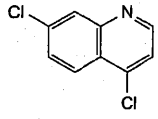
and enantiomer

E. (4*RS*)-4-[(7-chloroquinolin-4-yl)amino]pentan-1-ol,



and enantiomer

F. 7-chloro-4-[(2*RS*)-2-methylpyrrolidin-1-yl]quinoline,

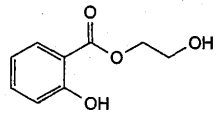


G. 4,7-dichloroquinoline.

Ph Eur

Hydroxyethyl Salicylate

(Ph. Eur. monograph 1225)



$C_9H_{10}O_4$ 182.2 *87-28-5*

Ph Eur

DEFINITION

2-Hydroxyethyl 2-hydroxybenzoate.

Content

98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

Oily, colourless or almost colourless liquid, or colourless crystals.

Solubility

Sparingly soluble in water, very soluble in acetone and in methylene chloride, freely soluble in ethanol (96 per cent).

Mp: about 21 °C.

IDENTIFICATION

First identification: A, B.

Second identification: A, C, D, E.

A. Refractive index (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Thin films.

Comparison *hydroxyethyl salicylate CRS.*

C. Examine the chromatograms obtained in the test for related substances.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 1 mL of solution S (see Tests), add 1 mL of *water R* and 0.2 mL of *ferric chloride solution R2*. A violet-red colour appears which disappears immediately after the addition of 2 mL of *dilute acetic acid R*. A very faint violet colour may remain.

E. In a test tube 160 mm long, mix 1.0 g with 2.0 g of finely powdered manganese sulfate R. Insert 2 cm into the test-tube a strip of filter paper impregnated with a freshly prepared mixture of 1 volume of a 20 per cent *V/V* solution of *diethanolamine R* and 11 volumes of a 50 g/L solution of *sodium nitroprusside R* adjusted to pH 9.8 with *1 M hydrochloric acid*. Heat the test-tube over a naked flame for 1-2 min. The filter paper becomes blue.

TESTS

Solution S

Dissolve 2.5 g in 40 mL of *ethanol (96 per cent) R* and dilute to 50 mL with *distilled water R*.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

To 2 mL of solution S add 0.1 mL of *methyl red solution R* and 0.2 mL of *0.01 M sodium hydroxide*. The solution is yellow. Add 0.3 mL of *0.01 M hydrochloric acid*. The solution is red.

Relative density (*2.2.5*)

1.252 to 1.257.

Refractive index (*2.2.6*)

1.548 to 1.551.

Related substances

Thin-layer chromatography (*2.2.27*).

Test solution (a) Dissolve 0.50 g of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 2 mL of test solution (a) to 50 mL with *methanol R*.

Reference solution (a) Dissolve 50.0 mg of *hydroxyethyl salicylate CRS* in *methanol R* and dilute to 25 mL with the same solvent.

Reference solution (b) Dilute 2.5 mL of test solution (b) to 10 mL with *methanol R*.

Reference solution (c) Dissolve 0.10 g of *ethylene glycol R* in *methanol R* and dilute to 50 mL with the same solvent. Dilute 1.25 mL of the solution to 10 mL with *methanol R*.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *ethyl acetate R, glacial acetic acid R, cyclohexane R* (20:20:60 *V/V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of cold air.

Detection A In ultraviolet light at 254 nm.

Limits A:

— *any impurity*: any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1 per cent).

Detection B Spray the plate with ammonium vanadate solution R and heat at 100 °C for 10 min. Allow to cool for 10 min and examine in daylight.

Limits B In the chromatogram obtained with test solution (a):

— *impurity B*: any spot corresponding to impurity B is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *any other impurity*: any spot, apart from the principal spot and any spot corresponding to impurity B is not more intense than the spot in the chromatogram obtained with reference solution (b) (1 per cent).

System suitability The chromatogram obtained with reference solution (c) shows a clearly visible spot.

Chlorides (*2.4.4*)
Maximum 100 ppm.

Dilute 10 mL of solution S to 15 mL with *water R*.

Sulfates (*2.4.13*)
Maximum 250 ppm.

Dilute 12 mL of solution S to 15 mL with *distilled water R*.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

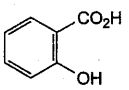
In a flask with a ground-glass stopper, dissolve 0.125 g in 30 mL of *glacial acetic acid R*. Add 10 mL of *dilute sulfuric acid R*, 1.5 g of *potassium bromide R* and 50.0 mL of *0.0167 M potassium bromate*. Immediately close the flask and allow to stand protected from light for 15 min. Add 1.5 g of *potassium iodide R* immediately after removing the stopper and titrate with *0.1 M sodium thiosulfate*, adding 1 mL of *starch solution R* towards the end of the titration. Carry out a blank titration.

1 mL of *0.0167 M potassium bromate* is equivalent to 4.555 mg of $C_9H_{10}O_4$.

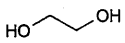
STORAGE

Protected from light.

IMPURITIES



A. 2-hydroxybenzenecarboxylic acid (salicylic acid),



B. ethane-1,2-diol (ethylene glycol).

Ph Eur

Hydroxyethylcellulose[1]

(Ph. Eur. monograph 0336)

9004-62-0

Action and use

Excipient.

Ph Eur

DEFINITION

Partly *O*-(2-hydroxyethylated) cellulose. Suitable pH-stabilisers, such as phosphates, may be added.

Content
30.0 per cent to 70.0 per cent of hydroxyethoxy ($-OC_2H_4OH$) groups (dried substance).

♦CHARACTERS

Appearance
White, yellowish-white or greyish-white, hygroscopic powder or granules.

Solubility
Soluble in hot and cold water giving a colloidal solution, practically insoluble in acetone, in ethanol (96 per cent) and in toluene.♦

IDENTIFICATION

First identification: A, B.

◊Second identification: B, C, D, E.◊

A. Infrared absorption spectrophotometry (*2.2.24*). Examine by attenuated total reflectance (ATR).

Comparison hydroxyethylcellulose CRS.

B. Heat 10 mL of solution S (see Tests) to boiling. The solution remains clear.

◊C. To 10 mL of solution S add 0.3 mL of *dilute acetic acid R* and 2.5 mL of a 100 g/L solution of *tannic acid R*. A yellowish-white, flocculent precipitate is formed which dissolves in *dilute ammonia R1*.

D. In a test-tube about 160 mm in length, thoroughly mix 1 g with 2 g of finely powdered manganese sulfate R. Introduce to a depth of 2 cm into the upper part of the tube a strip of filter paper impregnated with a freshly prepared mixture of 1 volume of a 200 g/L solution of *diethanolamine R* and 11 volumes of a 50 g/L solution of *sodium nitroprusside R*, adjusted to about pH 9.8 with *1 M hydrochloric acid*. Insert the tube 8 cm into a silicone-oil bath and heat at 190-200 °C. The filter paper becomes blue within 10 min. Carry out a blank test.

E. Dissolve 0.2 g completely, without heating, in 15 mL of a 700 g/L solution of *sulfuric acid R*. Pour the solution with stirring into 100 mL of iced *water R* and dilute to 250 mL with iced *water R*. In a test-tube, mix thoroughly while cooling in iced water 1 mL of the solution with 8 mL of *sulfuric acid R*, added dropwise. Heat on a water-bath for exactly 3 min and immediately cool in iced water. While the mixture is cold, carefully add 0.6 mL of *ninhydrin solution R2* and mix well. Allow to stand at 25 °C. A pink colour is produced immediately and does not become violet within 100 min.◊

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8. Pharmacopoeial harmonisation.*

TESTS

Solution S

Disperse 1.0 g (dried substance) in 50 mL of *carbon dioxide-free water R*. After 10 min, dilute to 100 mL with *carbon dioxide-free water R* and stir until dissolution is complete.

pH (*2.2.3*)

5.5 to 8.5 for solution S.

♦**Viscosity** (*2.2.10*)

75 per cent to 140 per cent of the value stated on the label.

While stirring, introduce a quantity of the substance to be examined equivalent to 2.00 g of the dried substance into 50 g of *water R*. Dilute to 100.0 g with *water R* and stir until dissolution is complete. Determine the viscosity using a rotating viscometer at 25 °C and at a shear rate of 100 s^{-1} for substances with an expected viscosity up to 100 mPa·s, at a shear rate of 10 s^{-1} for substances with an expected viscosity between 100 mPa·s and 20 000 mPa·s and at a shear rate of 1 s^{-1} for substances with an expected viscosity above 20 000 mPa·s. If it is impossible to obtain a shear rate of exactly 1 s^{-1}, 10 s^{-1} or 100 s^{-1} respectively, use a rate slightly higher and a rate slightly lower and interpolate.♦

Chlorides (*2.4.4*)

Maximum 1.0 per cent.

Dilute 1 mL of solution S to 30 mL with *water R*.

Nitrates

Maximum 3.0 per cent (dried substance) if hydroxyethylcellulose has a viscosity of 1000 mPa·s or less and maximum 0.2 per cent (dried substance) if hydroxyethylcellulose has a viscosity greater than 1000 mPa·s.

Determine potentiometrically (*2.2.36, Method I*) using as indicator a nitrate-selective electrode and a silver-silver chloride electrode with a 13.2 g/L solution of *ammonium sulfate R* as reference electrolyte.

Prepare the solutions immediately before use.

Buffer solution To a mixture of 50 mL of *dilute sulfuric acid R* and 800 mL of *water R*, add 135 g of *potassium dihydrogen phosphate R* and dilute to 1000 mL with *water R*.

Buffered water Dilute 80 mL of the buffer solution to 2000 mL with *water R*.

Nitrate standard solution (500 ppm NO_3) Dissolve 0.8154 g of *potassium nitrate R* in 500 mL of the buffered water and dilute to 1000.0 mL with the same solvent.

Test solution Dissolve 0.50 g of the substance to be examined in the buffered water and dilute to 100.0 mL with the same solvent.

Reference solutions If hydroxyethylcellulose has a viscosity of 1000 mPa·s or less, dilute 10.0 mL, 20.0 mL and 40.0 mL of the nitrate standard solution (500 ppm NO_3) to 100.0 mL with the buffered water and mix.

If hydroxyethylcellulose has a viscosity greater than 1000 mPa·s, dilute 1.0 mL, 2.0 mL and 4.0 mL of the nitrate standard solution (500 ppm NO_3) to 100.0 mL with the buffered water and mix.

Carry out the measurements for each solution. Calculate the concentration of nitrates using the calibration curve.

Aldehydes

Maximum 20 ppm, expressed as glyoxal.

Introduce 1.0 g into a test-tube with a ground-glass stopper and add 10.0 mL of *anhydrous ethanol R*. Stopper the tube and stir mechanically for 30 min. Centrifuge. To 2.0 mL of the supernatant add 5.0 mL of a 4 g/L solution of *methylbenzothiazolone hydrazone hydrochloride R* in an 80 per cent *V/V* solution of *glacial acetic acid R*. Shake to homogenise. After 2 h, the solution is not more intensely coloured than a standard prepared at the same time and in the same manner using 2.0 mL of *glyoxal standard solution (2 ppm $C_2H_2O_2$) R* instead of the 2.0 mL of supernatant.

◊**Ethylene oxide**

Head-space gas chromatography (*2.4.25*).

Test preparation Place 1.00 g of the substance to be examined in a 5 mL vial (other sizes may be used depending on the operating conditions) and add 1 mL of *water R*. It swells in water but does not dissolve.

Reference preparation (a) Place 1.00 g of the substance to be examined in an identical 5 mL vial. Add 0.1 mL of cooled *ethylene oxide solution R2* and 0.9 mL of *water R*. It swells in water but does not dissolve.

Reference preparation (b) To 0.1 mL of *ethylene oxide solution R2* in a 5 mL vial add 0.1 mL of a freshly prepared 10 mg/L solution of *acetaldehyde R*.

Close the vials immediately with a butyl rubber membrane stopper, coated with aluminium or polytetrafluoroethylene and secured with an aluminium crimped cap.

Limit:

— *ethylene oxide*: maximum 1 ppm.

2-Chloroethanol

Head-space gas chromatography (*2.2.28*).

Test preparation To 50 mg of the substance to be examined in a 10 mL vial (other sizes may be used depending on the operating conditions), add 2 µL of *2-propanol R*. Seal the flask and mix.

Reference preparation (a) Dissolve 0.125 g of *2-chloroethanol R* and dilute to 50.0 mL with *2-propanol R*. Dilute 1.0 mL of the solution to 10.0 mL with *2-propanol R*.

Reference preparation (b) To 50 mg of the substance to be examined in an identical 10 mL vial, add 2 µL of reference solution (a). Seal the flask and mix.

Close the vials immediately with a butyl rubber membrane stopper, coated with aluminium or polytetrafluoroethylene and secured with an aluminium crimped cap.

Column:

— *size*: *l* = 50 m, Ø = 0.32 mm,

— *stationary phase*: *poly(dimethyl)siloxane R* (1.2 µm).

Carrier gas *helium for chromatography R*.

Flow rate 25-35 cm/s.

Split ratio 1:10.

Static head-space conditions that may be used:

— *equilibration temperature*: 110 °C,

— *equilibration time*: 20 min,

— *temperature of injection system*: 115 °C.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 6	60
	6 - 16	60 → 110
	16 - 31	110 → 230
	31 - 36	230
Injection port		150
Detector		250

Detection Flame ionisation.

Injection 2 mL.

Retention time 2-chloroethanol = about 7.8 min.

Limit:

— *2-chloroethanol*: not more than 0.5 times the area of the peak due to 2-chloroethanol in the chromatogram obtained with reference solution (b) (10 ppm).◊

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (*2.4.14*)

Maximum 4.0 per cent if hydroxyethylcellulose has a viscosity of 1000 mPa·s or less and maximum 1.0 per cent if hydroxyethylcellulose has a viscosity greater than 1000 mPa·s, determined on 1.0 g.

ASSAY

Gas chromatography (*2.2.28*)

Internal standard solution To 10 mL of *o-xylene R* add 0.5 mL of *octane R* and dilute to 100.0 mL with *o-xylene R*.

Test solution To 30.0 mg (dried substance) add 60 mg of *adipic acid R* in a 5 mL pressure-tight reaction vial equipped with a pressure-tight membrane stopper coated with polytetrafluoroethylene and secured with an aluminium crimped cap or another sealing system providing a sufficient air-tightness. Add 2.00 mL of the internal standard solution and 1.0 mL of *hydriodic acid R* and close immediately. Accurately weigh the vial (total mass before heating). Do not mix the contents of the vial by hand before placing in the oven or the heater. Place the vial in an oven or heat in a suitable heater with continuous mechanical agitation, maintaining an internal temperature of the vial of 165 ± 2 °C for 2.5 h. Allow to cool and weigh accurately the vial (total mass after heating). If the difference between the total mass before heating and the total mass after heating is more than 10 mg, prepare a new test solution. After phase separation, pierce through the septum of the vial with a cooled syringe and withdraw a sufficient volume of the upper layer as test solution.

Reference solution Place 60 mg of *adipic acid R* and 2.00 mL of the internal standard solution in another 5 mL reaction vial, add 1.0 mL of *hydriodic acid R* and close immediately with a septum. Weigh accurately the vial then inject 55 µL of *iodoethane R* through the septum in the vial, weigh again accurately and mix. After phase separation, pierce through the septum of the vial with a cooled syringe and withdraw a sufficient volume of the upper layer as reference solution.

Column:

— *material*: fused silica;

— *size*: *l* = 30 m, Ø = 0.53 mm;

— *stationary phase*: *poly(dimethyl)siloxane R* (3 µm).

Carrier gas *helium for chromatography R*.

Flow rate 4.2 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	50
	3 - 8	50 → 100
	8 - 12	100 → 250
	12 - 20	250
Injection port		250
Detector		280

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to octane (retention time = about 10 min): iodoethane = about 0.6.

System suitability Reference solution:

— *resolution*: minimum 5.0 between the peaks due to iodoethane and octane;

— *repeatability*: maximum relative standard deviation of 2.0 per cent for the response factor of the principal peak determined on 6 injections.

Calculate the response factor (*R*) using the following expression:

$$\frac{A_1 \times m_1 \times C}{A_2 \times 100}$$

A_1 = area of the peak due to the internal standard in the chromatogram obtained with the reference solution;

A_2 = area of the peak due to iodoethane in the chromatogram obtained with the reference solution;

m_1 = mass of *iodoethane R* in the reference solution, in milligrams;

C = percentage content of *iodoethane R*.

Calculate the percentage content *m/m* of the hydroxyethoxy groups using the following expression:

$$\frac{A_4 \times R \times M_1 \times 100}{A_3 \times m_2 \times M_2}$$

A_3 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;

A_4 = area of the peak due to iodoethane in the chromatogram obtained with the test solution;

R = response factor;

M_1 = molar mass of hydroxyethoxy group (61.1);

M_2 = molar mass of iodoethane (156.0);

m_2 = mass of the sample (dried substance) in the test solurion, in milligrams.

♦LABELLING

The label states:

— the viscosity, in millipascal seconds for a 2 per cent *m/m* solution;

— the name and concentration of any added pH-stabiliser.♦

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hydroxyethylcellulose used as viscosity-increasing agent.

Viscosity

(see Tests).

Degree of substitution

(see Assay).

Hydroxyethylmethylcellulose

(Methylhydroxyethylcellulose, Ph. Eur. monograph 0346)

Action and use
Excipient.

Ph Eur

DEFINITION
Partly *O*-methylated and *O*-(2-hydroxyethylated) cellulose.

CHARACTERS

Appearance
White, yellowish-white or greyish-white powder or granules, hygroscopic after drying.

Solubility
Practically insoluble in hot water, in acetone, in anhydrous ethanol and in toluene. It dissolves in cold water giving a colloidal solution.

IDENTIFICATION
A. Heat 10 mL of solution S (see Tests) in a water-bath while stirring. At a temperature above 50 °C, the solution becomes cloudy or a flocculent precipitate is formed. The solution becomes clear again on cooling.

B. To 10 mL of solution S add 0.3 mL of *dilute acetic acid R* and 2.5 mL of a 100 g/L solution of *tannic acid R*. A yellowish-white flocculent precipitate is formed which dissolves in *dilute ammonia R1*.

C. In a test-tube about 160 mm long, thoroughly mix 1 g with 2 g of finely powdered manganese sulfate R. Introduce to a depth of 2 cm into the upper part of the tube a strip of filter paper impregnated with a freshly prepared mixture of 1 volume of a 20 per cent *V/V* solution of *diethanolamine R* and 11 volumes of a 50 g/L solution of *sodium nitroprusside R*, adjusted to about pH 9.8 with *1 M hydrochloric acid*. Insert the tube 8 cm into a silicone-oil bath at 190-200 °C. The filter paper becomes blue within 10 min. Carry out a blank test.

D. Dissolve completely 0.2 g without heating in 15 mL of a 70 per cent *m/m* solution of *sulfuric acid R*. Pour the solution with stirring into 100 mL of iced *water R* and dilute to 250 mL with iced *water R*. In a test-tube, mix thoroughly while cooling in iced water 1 mL of this solution with 8 mL of *sulfuric acid R* added dropwise. Heat in a water-bath for exactly 3 min and immediately cool in iced water. While the mixture is cold, carefully add 0.6 mL of *ninhydrin solution R2* and mix well. Allow to stand at 25 °C. A pink colour is produced immediately and does not become violet within 100 min.

E. Place 1 mL of solution S on a glass plate. After evaporation of the water a thin film is formed.

TESTS

Solution S
While stirring, introduce a quantity of the substance to be examined equivalent to 1.0 g of the dried substance into 50 g of *carbon dioxide-free water R* heated to 90 °C. Allow to cool, adjust the mass of the solution to 100 g with *carbon dioxide-free water R* and stir until dissolution is complete.

Appearance of solution
Solution S is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
5.5 to 8.0 for solution S.

Apparent viscosity (*2.2.10*)
75 per cent to 140 per cent of the value stated on the label.

While stirring, introduce a quantity of the substance to be examined equivalent to 6.00 g of the dried substance into 150 g of *water R* heated to 90 °C. Stir with a propeller-type stirrer for 10 min, place the flask in a bath of iced water, continue the stirring and allow to remain in the bath of iced water for 40 min to ensure that dissolution is complete. Adjust the mass of the solution to 300 g and centrifuge the solution to expel any entrapped air. Adjust the temperature of the solution to 20 ± 0.1 °C. Determine the viscosity with a rotating viscometer at 20 °C and a shear rate of 10 s^{-1}.

Chlorides (*2.4.4*)
Maximum 0.5 per cent.

Dilute 1 mL of solution S to 15 mL with *water R*.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

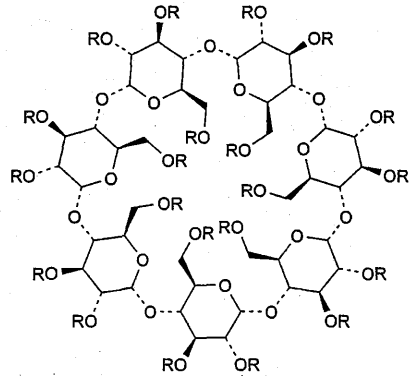
Sulfated ash (*2.4.14*)
Maximum 1.0 per cent, determined on 1.000 g.

LABELLING
The label states the apparent viscosity in millipascal seconds for a 2 per cent *m/m* solution.

Ph Eur

Hydroxypropylbetadex

(Ph. Eur. monograph 1804)



$C_{42}H_{70}O_{35}(C_3H_6O)_x$ with $x = 7$ *MS*

Action and use
Excipient.

Ph Eur

DEFINITION
Hydroxypropylbetadex (β-cyclodextrin, 2-hydroxypropyl ether) is a partially substituted poly(hydroxypropyl) ether of betadex.

Content
— *hydroxypropyl groups per anhydroglucose unit, expressed as molar substitution (MS)*: 0.40 to 1.50 and content within 10 per cent of the value stated on the label.

CHARACTERS

Appearance

White or almost white, amorphous or crystalline powder.

Solubility

Freely soluble in water and in propylene glycol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydroxypropylbetadex CRS.*

Results The spectrum obtained with the substance to be examined shows the same absorption bands as the spectrum obtained with *hydroxypropylbetadex CRS*. Due to differences in the substitution of the substance, the intensity of some absorption bands can vary.

B. Appearance of solution (see Tests).

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* prepared from *distilled water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*), and remains so after cooling to room temperature.

Dissolve 5.0 g in 10.0 mL of *water R*, with heating.

Conductivity (*2.2.38*)

Maximum 200 $\mu S \cdot cm^{-1}$.

Measure the conductivity of solution S, while gently stirring with a magnetic stirrer.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.600 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 60.0 mg of *betadex CRS* (impurity A) in *water R* and dilute to 50.0 mL with the same solvent.

Reference solutions (b), (c), (d), (e), (f) Dilute reference solution (a) with *water R* to obtain 5 reference solutions containing respectively 0.03 mg/mL, 0.09 mg/mL, 0.45 mg/mL, 0.90 mg/mL and 1.20 mg/mL of *betadex CRS*.

Reference solution (g) Dissolve 0.15 g of *hydroxypropylbetadex CRS* (containing impurity A) in *water R* and dilute to 10 mL with the same solvent.

Column:
- *size*: l = 0.25 m, Ø = 4.0 mm;
- *stationary phase*: *4-nitrophenylcarbamidesilyl silica gel for chromatography R* (5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: *water for chromatography R*;
- *mobile phase B*: *water for chromatography R, methanol R* (10:90 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	52	48
5 - 15	52 → 0	48 → 100
15 - 20	0	100

Flow rate 1.0 mL/min.

Detection Evaporative light-scattering detector; the following settings have been found to be suitable; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criteria. The use of a 2-port/6-way valve is advisable for 'heart-cutting' hydroxypropylbetadex peaks to save the detector from the huge amount of injected hydroxypropylbetadex:
- *carrier gas*: *nitrogen R*;
- *flow rate*: 1.5 L/min;
- *evaporator temperature*: 70 °C.

Injection 20 µL.

Retention time Impurity A = about 4.2 min.

Hydroxypropylbetadex elutes as a very wide peak or as several peaks after impurity A. Other typical impurities elute together as a wide peak or as a group of several peaks before impurity A.

System suitability:
- *resolution*: minimum 2.0 between the peak due to impurity A and the 1st peak due to hydroxypropylbetadex in the chromatogram obtained with reference solution (g); if necessary, adjust the column temperature (decreasing the temperature improves the resolution);
- plot a curve representing the logarithm of the concentration of impurity A in reference solutions (b), (c), (d), (e) and (f) as the abscissa and the logarithm of the corresponding peak areas as ordinates taking the assigned content of *betadex CRS* into account; the coefficient of correlation is not less than 0.950.

Calculate the percentage content of impurities with reference to the dried substance using the curve.

Limits:
- *impurity A*: maximum 1.5 per cent;
- *sum of impurities other than A*: maximum 1.0 per cent;
- *reporting threshold*: 0.05 per cent; disregard any peak eluting after impurity A.

Impurity B

Gas chromatography (*2.2.28*).

Internal standard solution To 62.5 mg of *ethylene glycol R*, add *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with *ethanol (96 per cent) R*.

Test solution Dissolve 50.0 mg of the substance to be examined in *ethanol (96 per cent) R* and dilute to 10.0 mL with the same solvent. To 1.0 mL of the solution, add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *ethanol (96 per cent) R*.

Reference solution Dissolve 62.5 mg of *propylene glycol CRS* (impurity B) in *ethanol (96 per cent) R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *ethanol (96 per cent) R*. To 1.0 mL of this solution, add 1.0 mL of the internal standard solution and dilute to 10.0 mL with *ethanol (96 per cent) R*.

Column:
- *material*: fused silica;
- *size*: l = 30 m, Ø = 0.32 mm;
- *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1.4 mL/min.

Split ratio 1:35.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 10	150 → 200
	10 - 11	200 → 240
Injection port		220
Detector		240

Detection Flame ionisation.

Injection 2 µL; wash the syringe thoroughly with *ethanol (96 per cent) R* to avoid occlusion in the needle.

Relative retention With reference to ethylene glycol (retention time = about 7.5 min): impurity B = about 0.9.

System suitability Reference solution:
— *resolution*: minimum 4.0 between the peaks due to impurity B and ethylene glycol;
— *symmetry factor*: maximum 2.0 for the peak due to propylene glycol.

Calculation of percentage contents Use the internal standard method.

Limit:
— *impurity B*: maximum 2.5 per cent.

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 120 °C for 2 h.

Microbial contamination
If intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion 10^2 CFU/g (*2.6.12*).

If not intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion 10^3 CFU/g (*2.6.12*);
— TYMC: acceptance criterion 10^2 CFU/g (*2.6.12*);
— absence of *Escherichia coli* (*2.6.13*);
— absence of *Salmonella* (*2.6.13*).

Bacterial endotoxins (*2.6.14*)
Less than 10 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Nuclear magnetic resonance spectrometry (*2.2.33*).

The molar substitution (*MS*) is calculated from the ratio between the signal from the 3 protons of the methyl group that is part of the hydroxypropyl group and the signal from the proton attached to the C1 carbon (glycosidic proton) of the anhydroglucose units.

Test solution Introduce not less than the equivalent of 10.0 mg of the substance to be examined, previously dried, into a 5 mm NMR tube equipped with a spinner in order to record the spectrum in rotation. Add approximately 0.75 mL of *deuterium oxide R1*. Cap the tube, mix thoroughly and adapt the spinner.

Apparatus FT-NMR spectrometer operating at minimum 250 MHz, suited to record a proton spectrum and to carry out quantitative analysis, at a temperature of at least 25 °C.

Acquisition of ^{1}H NMR spectra Use the appropriate instrument settings (frequency, gain, digital resolution, sample rotation, shims, probe tuning, resolution/data point, receiver gain, etc.) so as to obtain a suitable spectrum for quantitative analysis (good FID (Free Induction Decay), no distortion of the spectrum after Fourier transform and phase corrections). The relaxation delay must be adapted to the pulse angle in order to have sufficient relaxation of the protons of interest between 2 pulses (for example: 10 s for a 90° pulse).

Record the FID signal with at least 8 scans so as to obtain a spectral window comprised, at least, between 0 ppm and + 6.2 ppm, referring to the signal of exchangeable protons (solvent) at + 4.8 ppm (25 °C).

Make a zero filling at least 3-fold in size relative to the acquisition data file and transform the FID to the spectrum without any correction of Gaussian broadening factor (GB = 0) and with a line broadening factor not greater than 0.2 Hz (LB ≤ 0.2).

Call the integration sub-routine after phase corrections and baseline correction between + 0.5 ppm and + 6.2 ppm.

Measure the peak areas of the doublet from the methyl groups at + 1.2 ppm (A_1), and of the signals of the glycosidic protons between + 5 ppm and + 5.4 ppm (A_2).

Calculate the molar substitution (*MS*) using the following expression:

$$\frac{A\,1}{(3 \times A\,2)}$$

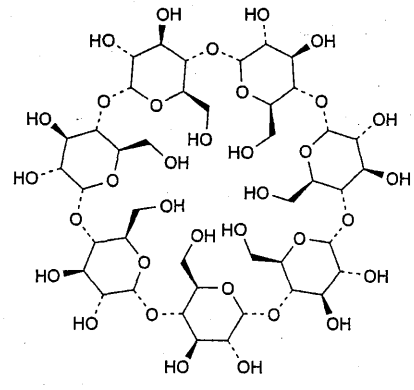
A_1 = area of the signal due to the 3 protons of the methyl groups that are part of the hydroxypropyl groups;
A_2 = area of the signals due to the glycosidic protons (protons attached to the C1 carbon) of the anhydroglucose units.

The degree of substitution is the number of hydroxypropyl groups per molecule of β-cyclodextrin and is obtained by multiplying the *MS* by 7.

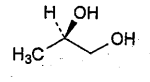
LABELLING

The label states:
— the molar substitution (*MS*);
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES



A. cycloheptakis-(1→4)-(α-D-glucopyranosyl) (betadex or cyclomaltoheptaose or β-cyclodextrin),



and enantiomer

B. (2*RS*)-propane-1,2-diol (propylene glycol).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality

criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for hydroxypropylbetadex used as solubility-increasing agent.

Degree of substitution
(see Assay).

Ph Eur

Hydroxypropylcellulose[1]

(Ph. Eur. monograph 0337)

9004-64-2

Action and use
Excipient.

Ph Eur

DEFINITION

Partly *O*-(2-hydroxypropylated) cellulose.

Content
53.4 per cent to 80.5 per cent of hydroxypropoxy groups (dried substance).

It may contain suitable anticaking agents, such as silica.

♦CHARACTERS

Appearance
White or yellowish-white powder or granules, slightly hygroscopic.

Solubility
Soluble in cold water, in ethanol (96 per cent) and in propylene glycol giving colloidal solutions, practically insoluble in hot water.♦

IDENTIFICATION

A. Dissolve 1 g in *water R* and dilute to 100 mL with the same solvent. Place 1 mL of this solution on a glass plate. After evaporation of the water a thin film is formed.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydroxypropylcellulose CRS.*

Disregard any peak at about 1719 cm^{-1}.

◊C. Evenly distribute 1.0 g into 100 mL of boiling *water R*, and stir the mixture using a magnetic stirrer with a bar about 25 mm long. Add 50 mL of this solution to 50 mL of *water R* in a beaker. Insert a thermometer into the solution. Stir the solution using a magnetic stirrer on a hot plate and heat at a rate of 2-5 °C/min. At a temperature above 40 °C the solution becomes cloudy or a flocculent precipitate is formed. The solution becomes clear again on cooling.◊

TESTS

pH (*2.2.3*)
5.0 to 8.0.

Evenly distribute 1.0 g into 100 mL of boiling *carbon dioxide-free water R*, and stir the mixture using a magnetic stirrer.

♦Viscosity (*2.2.10*)
75 per cent to 140 per cent of the value stated on the label.

While stirring, introduce a quantity of the substance to be examined equivalent to 6.00 g of the dried substance into 150 g of *water R* heated to 90 °C. Stir with a propeller-type stirrer for 10 min, place the flask in a bath of iced water, continue the stirring and allow to remain in the bath of iced water for 40 min to ensure that dissolution is complete. Adjust the mass of the solution to 300 g and centrifuge the solution to expel any entrapped air. Adjust the temperature of the solution to 20 ± 0.1 °C. Determine the viscosity with a rotating viscometer at 20 °C and a shear rate of 10 s^{-1}.

For a product of low viscosity, use a quantity of the substance to be examined sufficient to prepare a solution of the concentration stated on the label.♦

Silica
Maximum 0.6 per cent.

If the addition of silica is stated on the label and if more than 0.2 per cent is found in the test for sulfated ash, moisten the residue obtained in the test for sulfated ash with *water R* and add 5 mL of *hydrofluoric acid R* in small portions. Evaporate to dryness at 95-105 °C, taking care to avoid loss from sputtering. Cool and rinse the wall of the platinum crucible with 5 mL of *hydrofluoric acid R*. Add 0.5 mL of *sulfuric acid R* and evaporate to dryness. Progressively increase the temperature until all the acids have been volatised and ignite at 1000 ± 25 °C. Allow to cool in a desiccator and weigh. The difference between the mass of the residue obtained in the test for sulfated ash and the mass of the final residue is equal to the amount of silica in the substance to be examined.

Loss on drying
(*2.2.32*)Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.8 per cent, determined on 1.0 g using a platinum crucible.

ASSAY

Gas chromatography (*2.2.28*).

Internal standard solution To 10 mL of *o-xylene R* add 1.0 mL of *methylcyclohexane R* and dilute to 50.0 mL with *o-xylene R*.

Test solution To 30.0 mg (dried substance) add 60 mg of *adipic acid R* in a 5.0 mL reaction vial. Add 2.00 mL of the internal standard solution and 1.0 mL of *hydriodic acid R*. Close with a septum and weigh accurately the reaction vial (total mass before heating). Place the vial in an oven or heat in a suitable heater capable of maintaining an internal temperature of 115 ± 2 °C for 70 min with continuous stirring. Allow the vial to cool and weigh accurately the reaction vial (total mass after heating). If the difference of the total mass before heating to the total mass after heating is more than 10 mg, prepare a new solution. After phase separation, pierce through the septum of the vial with a cooled syringe and withdraw a sufficient volume of the upper phase as the test solution.

Reference solution Place 60 mg of *adipic acid R* and 2.00 mL of the internal standard solution in a 5.0 mL reaction vial. Add 1.0 mL of *hydriodic acid R*, close with a septum and weigh accurately. Inject 25 µL of isopropyl iodide R through the septum and weigh again accurately. Mix well. After phase separation, pierce through the septum of the vial with a

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

cooled syringe and withdraw a sufficient volume of the upper phase as the reference solution.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (3 µm).

Carrier gas *helium for chromatography R.*

Flow rate 7 mL/min.

Split ratio 1:50.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	40
	3 - 9	40 → 100
	9 - 12	100 → 250
	12 - 15	250
Injection port		180
Detector		280

Detection Flame ionisation.

Injection 2 µL.

Relative retention With reference to methylcyclohexane (retention time = about 8 min): isopropyl iodide = about 0.8.

System suitability Reference solution:
— *resolution*: minimum 2.0 between the peaks due to isopropyl iodide and methylcyclohexane in the chromatogram obtained with the reference solution;
— *repeatability*: maximum relative standard deviation of 2.0 per cent of the response factor of the principal peak determined on 6 injections.

Calculate the response factor *R* using the following expression:

$$\frac{A_1 \times W_1 \times C}{A_2 \times 100}$$

A_1 = area of the peak due to the internal standard in the chromatogram obtained with the reference solution;
A_2 = area of the peak due to isopropyl iodide in the chromatogram obtained with the reference solution;
W_1 = mass of isopropyl iodide R in the reference solution, in milligrams;
C = percentage content of isopropyl iodide R.

Calculate the percentage content *m/m* of the hydroxypropoxy groups using the following expression:

$$\frac{1.15 \times A_4 \times R \times M_1 \times 100}{A_3 \times W_2 \times M_2}$$

A_3 = area of the peak due to the internal standard in the chromatogram obtained with the test solution;
A_4 = area of the peak due to isopropyl iodide in the chromatogram obtained with the test solution;
R = response factor;
M_1 = molar mass of hydroxypropoxy group (75.1);
M_2 = molar mass of isopropyl iodide (170.0);
W_2 = mass of sample (dried substance) in the test solution, in milligrams;
1.15 = conversion factor.

♦LABELLING

The label states:
— the viscosity in millipascal seconds for a 2 per cent *m/m* solution;
— for a product of low viscosity, the concentration of the solution to be used and the viscosity in millipascal seconds;
— where applicable, that the substance contains silica.♦

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hydroxypropylcellulose used as binder, viscosity-increasing agent or film former:

Viscosity
(see Tests).

Degree of substitution
(see Assay).

The following characteristics may be relevant for hydroxypropylcellulose used as matrix former in prolonged-release tablets.

Viscosity
(see Tests).

Degree of substitution
(see Assay).

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Low-substituted Hydroxypropylcellulose [1]

(Ph. Eur. monograph 2083)

9004-64-2

Action and use
Excipient.

Ph Eur

DEFINITION

Low-substituted *O*-(2-hydroxypropylated) cellulose.

Content
5.0 per cent to 16.0 per cent of hydroxypropoxy groups (dried substance).

♦CHARACTERS

Appearance
White or yellowish-white, hygroscopic powder or granules.

Solubility
Practically insoluble in ethanol (96 per cent). It dissolves in a dilute solution of sodium hydroxide producing a viscous

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

solution. It swells in water, in a 106 g/L solution of sodium carbonate and in a 206 g/L solution of *hydrochloric acid R*.♦

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *low-substituted hydroxypropylcellulose CRS*.

B. Shake thoroughly 0.1 g with 10 mL of *water R*. It does not dissolve.

C. To the suspension obtained in Identification B add 1 g of *sodium hydroxide R* and shake until it becomes homogeneous. Transfer 5 mL of the solution to a suitable container, add 10 mL of a mixture of 1 volume of *methanol R* and 4 volumes of *acetone R* and shake; a white, flocculent precipitate is formed.

TESTS

pH (*2.2.3*)

5.0 to 7.5.

Evenly distribute 1.0 g onto the surface of 100 mL of *carbon dioxide-free water R* and stir using a magnetic stirrer.

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (*2.4.14*)

Not more than 0.8 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (*2.2.28*).

Apparatus:

— *reaction vial*: a 5 mL pressure-tight vial, 50 mm in height, 20 mm in external diameter and 13 mm in internal diameter at the mouth, equipped with a pressure-tight butyl rubber membrane stopper coated with polytetrafluoroethylene and secured with an aluminium crimped cap or another sealing system providing a sufficient air-tightness;

— *heater*: a heating module with a square aluminium block having holes 20 mm in diameter and 32 mm in depth, so that the reaction vials fit; mixing of the contents of the vial is effected using a magnetic stirrer equipped in the heating module or using a reciprocal shaker that performs approximately 100 cycles/min.

Internal standard solution 30 g/L solution of *octane R* in *o-xylene R*.

Test solution Weigh 65.0 mg of the substance to be examined, place in a reaction vial, add 0.06-0.10 g of *adipic acid R*, 2.0 mL of the internal standard solution and 2.0 mL of *hydriodic acid R*, immediately cap and seal the vial, and weigh accurately. Mix the contents of the vial continuously for 60 min while heating the block so that the temperature of the contents is maintained at 130 ± 2 °C. If a reciprocal shaker or magnetic stirrer cannot be used, shake the vial thoroughly by hand at 5 min intervals during the initial 30 min of the heating time. Allow the vial to cool, and again weigh accurately. If the loss of mass is less than 26 mg and there is no evidence of a leak, use the upper layer of the mixture as the test solution.

Reference solution Place 0.06-0.10 g of *adipic acid R*, 2.0 mL of the internal standard solution and 2.0 mL of *hydriodic acid R* in another reaction vial, cap and seal the vial, and weigh accurately. Add 15-22 µL of isopropyl iodide R through the septum with a syringe, and weigh accurately. Shake the reaction vial thoroughly and use the upper layer as the reference solution.

Column:

— *material*: fused silica;

— *size*: *l* = 30 m, Ø = 0.53 mm;

— *stationary phase*: *poly(dimethyl)siloxane R* (3 µm).

Use a precolumn if needed.

Carrier gas *helium for chromatography R*.

Flow rate 4.3 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	50
	3 - 8	50 → 100
	8 - 12	100 → 250
	12 - 20	250
Injection port		250
Detector		280

Detection Flame ionisation or thermal conductivity.

Injection 1-2 µL of the test solution and the reference solution.

Relative retention With reference to octane (retention time = about 8 min): isopropyl iodide = about 0.8.

System suitability Reference solution:

— *resolution*: minimum 5.0 between the peaks due to isopropyl iodide and octane;

— *repeatability*: maximum relative standard deviation of 2.0 per cent for the ratio of the area of the peak due to isopropyl iodide to that due to octane determined on 6 injections.

Calculate the ratio (Q) of the area of the peak due to isopropyl iodide to the area of the peak due to the internal standard from the chromatogram obtained with the test solution, and the ratio (Q_1) of the area of the peak due to isopropyl iodide to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution.

Calculate the percentage content of hydroxypropoxy groups using the following expression:

$$\frac{Q \times m_1}{Q_1 \times m} \times \frac{M_1}{M_2} \times 100$$

m_1 = mass of isopropyl iodide in the reference solution, in milligrams;
m = mass of the sample (dried substance), in milligrams;
M_1 = molar mass of hydroxypropoxy group (75.1);
M_2 = molar mass of isopropyl iodide (170.0).

STORAGE

In an airtight container.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for low-substituted hydroxypropylcellulose used as disintegrant.

Settling volume
20.0 mL to 35.0 mL.

To 20 mL of *2-propanol R* in a 100 mL graduated cylinder add 5.0 g of the substance to be examined and shake vigorously. Dilute to 30 mL with *2-propanol R*, then dilute to 50 mL with *water R* and shake vigorously. Within 15 min, repeat the shaking 3 times. Seal the cylinder to avoid evaporation of the solvent. Allow to stand for 2 h and determine the volume of the settled mass.

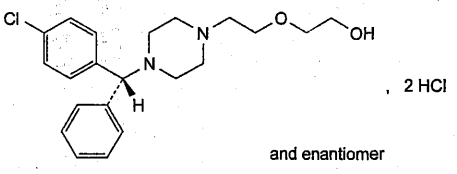
Degree of substitution
(see Assay).

Particle-size distribution (*2.9.31*)

Ph Eur

Hydroxyzine Hydrochloride

(Ph. Eur. monograph 0916)



$C_{21}H_{29}Cl_3N_2O_2$ 447.8 *2192-20-3*

Action and use
Histamine H_1 receptor antagonist.

Preparations
Hydroxyzine Oral Solution
Hydroxyzine Tablets

Ph Eur

DEFINITION
(*RS*)-2-[2-[4-[(4-Chlorophenyl)phenylmethyl]piperazin-1-yl] ethoxy]ethanol dihydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water and in ethanol (96 per cent), very slightly soluble in acetone.

mp
About 200 °C, with decomposition.

IDENTIFICATION
First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hydroxyzine hydrochloride CRS.*

B. Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, methylene chloride R* (50:50 *V/V*).

Test solution Dissolve 0.50 g of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 0.50 g of *hydroxyzine hydrochloride CRS* in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 0.50 g of meclozine dihydrochloride R in the solvent mixture and dilute to 10 mL with the solvent mixture. Dilute 1 mL of the solution to 2 mL with reference solution (a).

Plate *TLC silica gel G plate R.*

Mobile phase *concentrated ammonia R, ethanol (96 per cent) R, toluene R* (1:24:75 *V/V/V*).

Application 2 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with *potassium iodobismuthate solution R2.*

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in *ethanol (96 per cent) R* and dilute to 15 mL with the same solvent. Add 15 mL of a saturated solution of *picric acid R* in *ethanol (96 per cent) R*. Allow to stand for 15 min. A precipitate is formed. Filter. Recrystallise from *ethanol (96 per cent) R*. Initiate crystallisation, if necessary, by scratching the wall of the tube with a glass rod. The crystals melt (*2.2.14*) at 189 °C to 192 °C.

D. It gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 2.0 g in *water R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *hydroxyzine hydrochloride CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 3.0 mL of the test solution to 200.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 25.0 mL with the mobile phase.

Column:
— *size: l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Dissolve 0.5 g of *sodium methanesulfonate R* in a mixture of 14 mL of *triethylamine R*, 300 mL of *acetonitrile for chromatography R* and 686 mL of *water for chromatography R*, then adjust to pH 2.7 with *sulfuric acid R.*

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 2.5 times the retention time of hydroxyzine.

System suitability Reference solution (a):
— *peak-to-valley ratio*: minimum 10, where H_p = height above the baseline of the peak immediately before the peak due to hydroxyzine and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to hydroxyzine.

Limits:
— *any impurity*: for each impurity, not more than 1/3 of the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

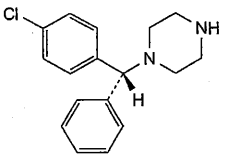
Loss on drying (*2.2.32*)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 10 mL of *anhydrous acetic acid R*. Add 40 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

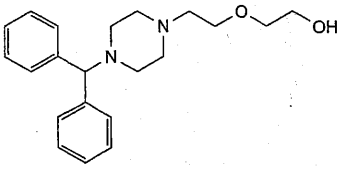
1 mL of *0.1 M perchloric acid* is equivalent to 22.39 mg of $C_{21}H_{29}Cl_3N_2O_2$.

STORAGE
In an airtight container, protected from light.

IMPURITIES


and enantiomer

A. (*RS*)-1-[(4-chlorophenyl)phenylmethyl]piperazine,

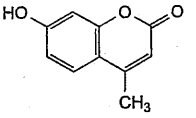


B. 2-[2-[4-(diphenylmethyl)piperazin-1-yl]ethoxy]ethanol (decloxizine).

Ph Eur

Hymecromone

(Ph. Eur. monograph 1786)



$C_{10}H_8O_3$ 176.2 *90-33-5*

Action and use
Choleretic; antispasmodic.

Ph Eur

DEFINITION
7-Hydroxy-4-methyl-2*H*-1-benzopyran-2-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
Almost white crystalline powder.

Solubility
Very slightly soluble in water, sparingly soluble in methanol, slightly soluble in methylene chloride. It dissolves in dilute solutions of ammonia.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *hymecromone CRS*.

TESTS
Absorbance (*2.2.25*)
Dissolve 50 mg in 10 mL of *ammonium chloride buffer solution pH 10.4 R* and dilute to 100.0 mL with *water R*. To 1.0 mL of the solution, add 10 mL of *ammonium chloride buffer solution pH 10.4 R* and dilute to 100.0 mL with *water R*. Examined between 200 nm and 400 nm, the solution shows 2 absorption maxima, at 229 nm and 360 nm, and an absorption minimum at 276 nm. The specific absorbance at the maximum at 360 nm is 1020 to 1120.

Related substances
Liquid chromatography (*2.2.29*).
Buffer solution To 280 mL of a 1.56 g/L solution of *sodium dihydrogen phosphate R*, add 720 mL of a 3.58 g/L solution of *disodium hydrogen phosphate dodecahydrate R*. Adjust to pH 7 with a 100 g/L solution of *phosphoric acid R*.
Test solution Dissolve 10 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.
Reference solution (a) Dissolve 20 mg of *hymecromone CRS*, 10 mg of *hymecromone impurity A CRS* and 10 mg of *hymecromone impurity B CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of reference solution (a) to 200.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.25 m, Ø = 4 mm,
— *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (10 µm).
Mobile phase *methanol R*, buffer solution (465:535 *V/V*).
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 270 nm.
Injection 20 µL.

Run time 1.5 times the retention time of hymecromone.

Relative retention With reference to hymecromone (retention time = about 6 min): impurity A = about 0.5; impurity B = about 0.7.

System suitability Reference solution (a):
— *resolution*: minimum of 2 between the peaks due to impurity A and to impurity B and minimum of 3 between the peaks due to impurity B and to hymecromone.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.05 per cent),
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.05 per cent),
— *unspecified impurities*: for each impurity, not more than the area of the peak due to hymecromone in the chromatogram obtained with reference solution (b) (0.10 per cent),
— *total*: not more than twice the area of the peak due to hymecromone in the chromatogram obtained with reference solution (b) (0.2 per cent),
— *disregard limit*: 0.1 times the area of the peak due to hymecromone in the chromatogram obtained with reference solution (b) (0.01 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 80 mL of *2-propanol R*. Titrate with *0.1 M tetrabutylammonium hydroxide in 2-propanol* determining the end-point potentiometrically (*2.2.20*). Carry out a blank titration.

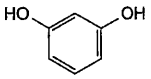
1 mL of *0.1 M tetrabutylammonium hydroxide in 2-propanol* is equivalent to 17.62 mg of $C_{10}H_8O_3$.

STORAGE

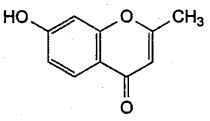
Protected from light.

IMPURITIES

Specified impurities *A, B.*



A. benzene-1,3-diol (resorcinol),



B. 7-hydroxy-2-methyl-4*H*-1-benzopyran-4-one.

Ph Eur

Hymenoptera Venoms for Allergen Products

(Ph. Eur. monograph 2623)

Ph Eur

DEFINITION

Hymenoptera venoms for allergen products are obtained from stinging insects mainly belonging to the families Apidae or Vespidae. They contain soluble substances, including proteins from which they derive their antigenic and allergenic properties, e.g. phospholipase and hyaluronidase.

Hymenoptera venoms for allergen products are unprocessed liquid venom, the venom sac, or venom processed (e.g. by dissolution, filtration and/or drying) using qualified methods.

PRODUCTION

The species of Hymenoptera from which the venom is extracted is identified and specified e.g. by morphological features. The origin, quality and traceability of the source material must be demonstrated. The methods of insect collection and venom extraction are described and must ensure that the venom is of appropriate quality. Electrostimulation may be used to collect the venom.

Pesticides and/or chemical attractants are not used in the collection of insects.

Where major changes to the production of the Hymenoptera venom source material take place (e.g. when a new process or supplier of Hymenoptera venom source material is introduced), such changes are qualified for each Hymenoptera venom source material.

Microbial contamination of the Hymenoptera venom source material may be unavoidable and should be monitored on a representative number of batches of venom source material according to a justified sampling plan and each time a new supplier and/or a new process for the venom source material production is introduced; if a determination of microbial contamination is not applicable (e.g. when the source material is bactericidal), this must be justified. Microbial contamination values and potential increases in microbial contamination are monitored during stability studies, in order to assess this aspect along with the venom source material characteristics upon storage.

Control methods and acceptance criteria relating to identity and purity (e.g. absence of foreign matter) of the Hymenoptera venom source material are established. The acceptance criteria must ensure the consistency of the venom source material from a qualitative and quantitative point of view. The venom source material is stored under controlled conditions justified by stability data.
The collection and production, as well as the handling of the venom source material, are such that consistent composition is ensured from batch to batch.

HYMENOPTERA VENOMS FOR ALLERGEN PRODUCTS REFERENCE BATCH

An appropriate reference batch is established for each Hymenoptera venom source material. The nature of the reference batch depends on the testing approach to verify batch-to-batch consistency and to establish acceptable quality. The reference batch may be, for example, an internal reference preparation (if available), a venom or venom sac source material, or a sample of a production batch.
Its characterisation must be described. The extent of characterisation of the reference batch depends on the nature

of the venom source material. The reference batch is stored under controlled conditions ensuring its stability.

BATCH-TO-BATCH CONSISTENCY

To establish batch-to-batch consistency, one or more of the following tests are performed on each batch. The choice of tests and the production step at which they are carried out must be justified.

Total protein (*2.5.33*)

Protein profile

Determined by using suitable electrophoresis methods (*2.2.31*, *2.2.54*).

Enzyme activity

(e.g. hyaluronidase or phospholipase activity, as appropriate). Determined by using a suitable assay.

Allergen profile

Relevant allergenic components are identified by means of suitable techniques using allergen-specific antibodies.

Major allergen content

Determined by using suitable immunochemical methods (*2.7.1*) such as enzyme-linked immunosorbent assay (ELISA).

Total allergenic activity

Determined by testing inhibition of the binding capacity of specific immunoglobulin E antibodies or by a suitable equivalent *in vitro* method.

CHARACTERS

Hymenoptera venoms for allergen products are colourless or yellowish sacs or liquid, or white to yellowish powders.

IDENTIFICATION

Where possible, tests on identity are performed on each individual batch of venom using adequate methods, e.g. polyacrylamide gel electrophoresis analysis (*2.2.31*), liquid chromatography (*2.2.29*) and/or enzyme activity.

TESTS

Foreign matter

Foreign matter is defined as any particles that are not part of the venom or venom sac. Foreign matter is detected using appropriate methods (e.g. microscopic methods); a limit for the amount of foreign matter has to be established based on historical data.

Water (*2.5.12* or *2.5.32*) or **loss on drying** (*2.2.32*)

The water content of dried material is determined; specification limits must be supported by batch analysis and stability data.

STORAGE

The Hymenoptera venom source material is stored under controlled conditions justified by stability data.

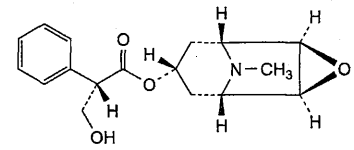
LABELLING

The label states the species of the source insect(s).

Ph Eur

Hyoscine

(Ph. Eur. monograph 2167)



$C_{17}H_{21}NO_4$ 303.4 *51-34-3*

Action and use

Anticholinergic.

Ph Eur

DEFINITION

(1*R*,2*R*,4*S*,5*S*,7*s*)-9-Methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$] non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Soluble in water, freely soluble in ethanol (96 per cent).

mp

66 °C to 70 °C.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hyoscine CRS*.

TESTS

Specific optical rotation (*2.2.7*)

−33 to −39 (anhydrous substance).

Dissolve 1.00 g in *dilute hydrochloric acid R* and dilute to 25.0 mL with the same acid.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of *hyoscine impurity A CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 25.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (d) Mix 2.0 mL of reference solution (b) and 1.0 mL of the test solution and dilute to 10.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.125 m, Ø = 4.0 mm,

— *stationary phase*: *octylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Mix 33 volumes of *acetonitrile R* and 67 volumes of a 2.5 g/L solution of *sodium dodecyl sulfate R*

previously adjusted to pH 2.5 with a 346 g/L solution of *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Run time 3 times the retention time of hyoscine.

Relative retention With reference to hyoscine (retention time = about 5 min): impurity C = about 0.2; impurity A = about 0.9; impurity D = about 1.3; impurity B = about 2.5.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to impurity A and hyoscine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.6; impurity C = 0.3;
— *impurity A*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurities B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

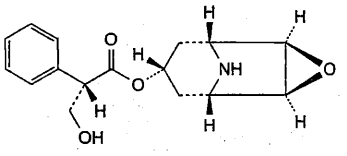
Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

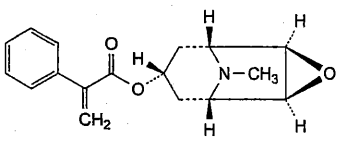
ASSAY
Dissolve 0.250 g in 60 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 30.34 mg of $C_{17}H_{21}NO_4$.

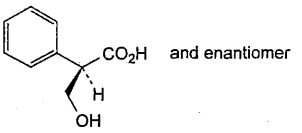
IMPURITIES
Specified impurities *A, B, C, D.*



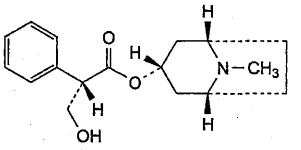
A. (1*R*,2*R*,4*S*,5*S*,7*s*)-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (norhyoscine),



B. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo [3.3.1.$0^{2,4}$]non-7-yl 2-phenylprop-2-enoate (apohyoscine),



C. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (DL-tropic acid),

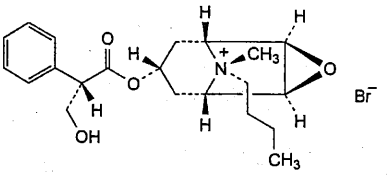


D. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscyamine).

Ph Eur

Hyoscine Butylbromide

(Ph. Eur. monograph 0737)



$C_{21}H_{30}BrNO_4$ 440.4 *149-64-4*

Action and use
Anticholinergic.

Preparations
Hyoscine Butylbromide Injection
Hyoscine Butylbromide Tablets

Ph Eur

DEFINITION
(1*R*,2*R*,4*S*,5*S*,7*s*,9*r*)-9-Butyl-7-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-9-methyl-3-oxa-9-azatricyclo [3.3.1.$0^{2,4}$]nonan-9-ium bromide.

Content
98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water and in methylene chloride, sparingly soluble in anhydrous ethanol.

IDENTIFICATION
First identification: A, C, F.
Second identification: A, B, D, E, F.

A. Specific optical rotation (see Tests).

B. Melting point (*2.2.14*): 139 °C to 141 °C.

C. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *hyoscine butylbromide CRS.*

D. To about 1 mg add 0.2 mL of *nitric acid R* and evaporate to dryness on a water-bath. Dissolve the residue in 2 mL of *acetone R* and add 0.1 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*. A violet colour develops.

E. To 5 mL of solution S (see Tests) add 2 mL of *dilute sodium hydroxide solution R*. No precipitate is formed.

F. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S

Dissolve 1.25 g in *water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Acidity or alkalinity

Dissolve 1.0 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent. Add 0.25 mL of *0.01 M sodium hydroxide* and 0.2 mL of *methyl red mixed solution R*. The solution is green. Not more than 0.5 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to reddish-violet.

Specific optical rotation (*2.2.7*)

−20 to −18 (dried substance), determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase B and dilute to 10.0 mL with mobile phase B.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase B.

Reference solution (b) Dissolve 5 mg of *hyoscine butylbromide for system suitability CRS* (containing impurities A and B) in mobile phase B and dilute to 10.0 mL with mobile phase B.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *silica gel for chromatography, alkyl-bonded for use with highly aqueous mobile phases R* (1.8 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R1*, 0.2 per cent *V/V* solution of *perchloric acid R* (5:95 *V/V*);
- *mobile phase B*: 0.2 per cent *V/V* solution of *perchloric acid R, acetonitrile R1* (30:70 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	91	9
1 - 4.2	91 → 75	9 → 25
4.2 - 5.5	75 → 66	25 → 34
5.5 - 10	66 → 15	34 → 85
10 - 11	15	85

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 2 µL.

Identification of impurities Use the chromatogram supplied with *hyoscine butylbromide for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to hyoscine butylbromide (retention time = about 6 min): bromide = about 0.1; impurity B = about 0.28; impurity A = about 0.37.

System suitability Reference solution (b):
- *resolution*: minimum 2.0 between the peaks due to impurities B and A.

Calculation of percentage contents:
- for each impurity, use the concentration of hyoscine butylbromide in reference solution (a).

Limits:
- *impurity A*: maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.4 per cent;
- *reporting threshold*: 0.05 per cent; disregard the peak due to bromide.

Loss on drying (*2.2.32*)

Maximum 2.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 0.5 g.

ASSAY

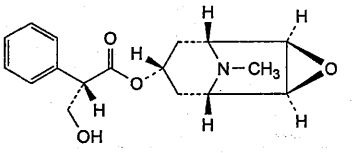
Dissolve 0.400 g in 50 mL of *water R*. Titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*) using a silver indicator electrode and a silver-silver chloride reference electrode.

1 mL of *0.1 M silver nitrate* is equivalent to 44.04 mg of $C_{21}H_{30}BrNO_4$.

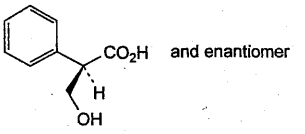
IMPURITIES

Specified impurities *A.*

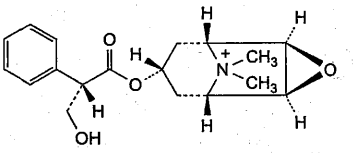
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, F, G, H.*



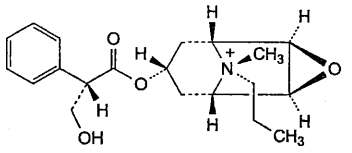
A. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo [3.3.1.$0^{2,4}$]nonan-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscine),



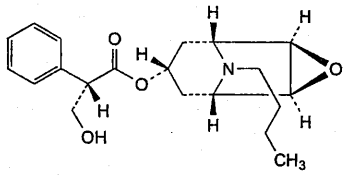
B. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (DL-tropic acid),



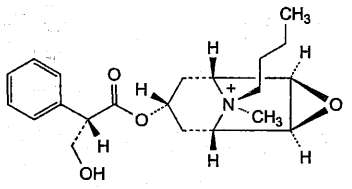
C. (1*R*,2*R*,4*S*,5*S*,7s)-7-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-9,9-dimethyl-3-oxa-9-azatricyclo [3.3.1.$0^{2,4}$]nonan-9-ium (methylhyoscine),



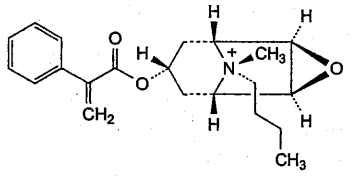
D. (1*R*,2*R*,4*S*,5*S*,7*s*,9*r*)-7-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-9-methyl-9-propyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]nonan-9-ium (propylhyoscine),



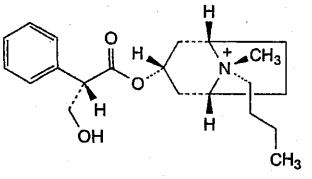
E. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-butyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]nonan-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (*N*-butylnorhyoscine),



F. (1*R*,2*R*,4*S*,5*S*,7*s*,9*s*)-9-butyl-7-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-9-methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]nonan-9-ium,



G. (1*R*,2*R*,4*S*,5*S*,7*s*,9*r*)-9-butyl-9-methyl-7-[(2-phenylprop-2-enoyl)oxy]-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]nonan-9-ium (apo-*N*-butylhyoscine),

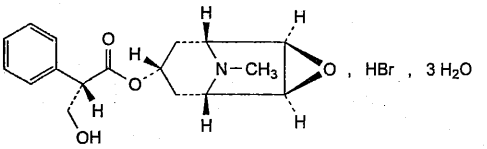


H. (1*R*,3*r*,5*S*,8*s*)-8-butyl-3-[[(2*S*)-3-hydroxy-2-phenylpropanoyl]oxy]-8-methyl-8-azabicyclo[3.2.1]octan-8-ium (*N*-butylhyoscyamine).

Ph Eur

Hyoscine Hydrobromide

(Ph. Eur. monograph 0106)



$C_{17}H_{22}BrNO_4,3H_2O$ 438.3 *6533-68-2*

Action and use
Anticholinergic.

Preparations
Hyoscine Eye Drops
Hyoscine Injection
Hyoscine Tablets

Ph Eur

DEFINITION

(1*R*,2*R*,4*S*,5*S*,7*s*)-9-Methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate hydrobromide trihydrate.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, efflorescent.

Solubility
Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: B, E.

Second identification: A, C, D, E.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison hyoscine hydrobromide CRS.

If the spectra obtained in the solid state show differences, proceed as follows: dissolve 3 mg of the substance to be examined in 1 mL of *ethanol (96 per cent) R* and evaporate to dryness on a water-bath; dissolve the residue in 0.5 mL of *methylene chloride R* and add 0.2 g of *potassium bromide R* and 15 mL of *ether R*; allow to stand for 5 min shaking frequently; decant; dry the residue on a water-bath until the solvents have evaporated; using the residue prepare a disc and dry at 100-105 °C for 3 h. Repeat the procedure with *hyoscine hydrobromide CRS* and record the spectra.

C. Dissolve about 50 mg in 5 mL of *water R* and add 5 mL of *picric acid solution R* dropwise and with shaking. The precipitate, washed with *water R* and dried at 100-105 °C for 2 h, melts (*2.2.14*) at 188 °C to 193 °C.

D. To about 1 mg add 0.2 mL of *fuming nitric acid R* and evaporate to dryness on a water-bath. Dissolve the residue in 2 mL of *acetone R* and add 0.1 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*. A violet colour develops.

E. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

pH (*2.2.3*)
4.0 to 5.5 for solution S.

Specific optical rotation (*2.2.7*)
−24 to −27 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 70.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 25.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of *hyoscine hydrobromide impurity B CRS* in the mobile phase, add 5.0 mL of the test solution and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.125 m, Ø = 4.0 mm,
— *stationary phase*: *octylsilyl silica gel for chromatography R* (3 µm),
— *temperature*: 25 ± 1 °C.

Mobile phase Mix 330 mL of *acetonitrile R* with 670 mL of a 2.5 g/L solution of *sodium dodecyl sulfate R* previously adjusted to pH 2.5 with a 345 g/L solution of *phosphoric acid R*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 5 µL.

Run time 3 times the retention time of hyoscine.

Relative retention With reference to hyoscine (retention time = about 5.0 min): impurity D = about 0.2; impurity B = about 0.9; impurity A = about 1.3; impurity C = about 2.4.

System suitability Reference solution (c):
— *resolution*: minimum 1.5 between the peaks due to impurity B and hyoscine,
— *symmetry factor*: maximum 2.5 for the peak due to hyoscine.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.3; impurity C = 0.6;
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *impurities A, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— *total*: not more than 1.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent); disregard any peak due to the bromide ion which appears close to the solvent peak;
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)
10.0 per cent to 13.0 per cent, determined on 0.20 g.

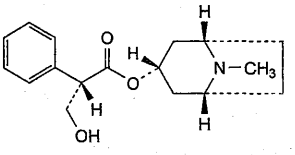
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in a mixture of 5.0 mL of *0.01 M hydrochloric acid* and 50 mL of *ethanol (96 per cent) R*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide* free from carbonate. Read the volume added between the 2 points of inflexion.

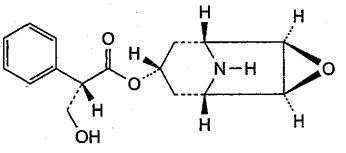
1 mL of *0.1 M sodium hydroxide* is equivalent to 38.43 mg of $C_{17}H_{22}BrNO_4$.

STORAGE
In a well-filled, airtight container of small capacity, protected from light.

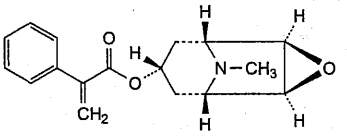
IMPURITIES
Specified impurities *A, B, C, D.*



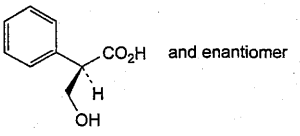
A. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscyamine),



B. (1*R*,2*R*,4*S*,5*S*,7*s*)-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$]non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (norhyoscine),



C. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo [3.3.1.$0^{2,4}$]non-7-yl 2-phenylprop-2-enoate (apohyoscine),

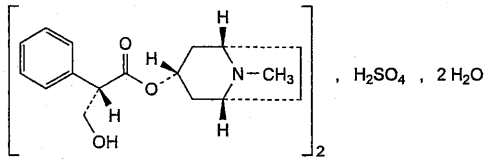


D. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (DL-tropic acid).

Ph Eur

Hyoscyamine Sulfate

(Ph. Eur. monograph 0501)



$C_{34}H_{48}N_2O_{10}S,2H_2O$ 713 *620-61-1*

Action and use
Anticholinergic.

Ph Eur

DEFINITION

Bis[(1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate] sulfate dihydrate.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless needles.

Solubility
Very soluble in water, sparingly soluble or soluble in ethanol (96 per cent).

IDENTIFICATION

First identification: A, B, E.

Second identification: C, D, E.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *hyoscyamine sulfate CRS.*

C. To 0.5 mL of solution S (see Tests) add 2 mL of *dilute acetic acid R* and heat. To the hot solution add 4 mL of *picric acid solution R*. Allow to cool, shaking occasionally. Collect the crystals, wash with 2 quantities, each of 3 mL, of iced *water R* and dry at 100-105 °C. The crystals melt (*2.2.14*) at 164 °C to 168 °C.

D. To about 1 mg add 0.2 mL of *fuming nitric acid R* and evaporate to dryness on a water-bath. Dissolve the residue in 2 mL of *acetone R* and add 0.2 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*. A violet colour develops.

E. It gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S
Dissolve 2.50 g in *water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution
Solution S is not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)
4.5 to 6.2.

Dissolve 0.5 g in *carbon dioxide-free water R* and dilute to 25 mL with the same solvent.

Specific optical rotation (*2.2.7*)
−24 to −29 (anhydrous substance), determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 60.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A. Dilute 10.0 mL of the solution to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 50.0 mL with mobile phase A.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 25.0 mL with mobile phase A.

Reference solution (c) Dissolve 5.0 mg of *hyoscyamine impurity E CRS* in the test solution and dilute to 20.0 mL with the test solution. Dilute 5.0 mL of this solution to 25.0 mL with mobile phase A.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 25 ± 1 °C.

Mobile phase:
— *mobile phase A*: dissolve 3.5 g of *sodium dodecyl sulfate R* in 606 mL of a 7.0 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.3 with a 0.58 g/L solution of *phosphoric acid R* and mix with 320 mL of *acetonitrile R*;
— *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2.0	95	5
2.0 - 20.0	95 → 70	5 → 30
20.0 - 20.1	70 → 95	30 → 5
20.1 - 25.0	95	5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Relative retention With reference to hyoscyamine (retention time = about 10.5 min): impurity A = about 0.2; impurity B = about 0.67; impurity C = about 0.72; impurity D = about 0.8; impurity E = about 0.9; impurity F = about 1.1; impurity G = about 1.8.

System suitability Reference solution (c):
— *resolution*: minimum 2.5 between the peaks due to hyoscyamine and impurity E;
— *symmetry factor*: maximum 2.5 for the peak due to hyoscyamine.

Limits:
— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.3; impurity G = 0.6;
— *impurity E*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *impurities A, B, C, D, F, G*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (*2.5.12*)
2.0 per cent to 5.5 per cent, determined on 0.500 g.

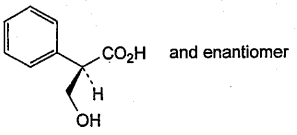
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.500 g in 25 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

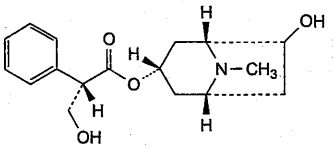
1 mL of *0.1 M perchloric acid* is equivalent to 67.7 mg of $C_{34}H_{48}N_2O_{10}S$.

STORAGE
In an airtight container, protected from light.

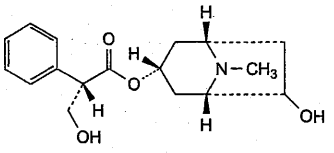
IMPURITIES
Specified impurities *A, B, C, D, E, F, G.*



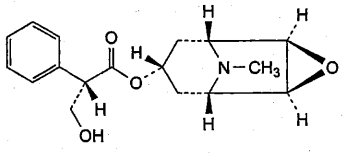
A. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (DL-tropic acid),



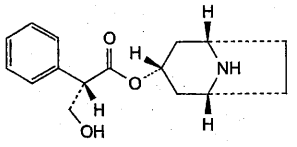
B. (1*R*,3*S*,5*R*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[3.2.1] oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (7-hydroxyhyoscyamine),



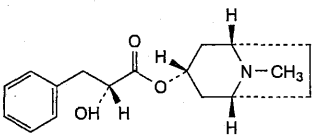
C. (1*S*,3*R*,5*S*,6*RS*)-6-hydroxy-8-methyl-8-azabicyclo[3.2.1] oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (6-hydroxyhyoscyamine),



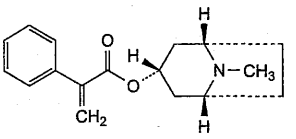
D. (1*R*,2*R*,4*S*,5*S*,7*s*)-9-methyl-3-oxa-9-azatricyclo[3.3.1.$0^{2,4}$] non-7-yl (2*S*)-3-hydroxy-2-phenylpropanoate (hyoscine),



E. (1*R*,3*r*,5*S*)-8-azabicyclo[3.2.1]oct-3-yl (2*S*)-3-hydroxy-2-phenylpropanoate (norhyoscyamine),



F. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl (2*R*)-2-hydroxy-3-phenylpropanoate (littorine),



G. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-3-yl 2-phenylprop-2-enoate (apoatropine).

Ph Eur

Hypromellose[1]

(Ph. Eur. monograph 0348)

9004-65-3

Action and use
Artificial tears.

Preparation
Hypromellose Eye Drops

Ph Eur

DEFINITION
Hydroxypropylmethylcellulose. Cellulose, 2-hydroxypropylmethyl ether.

Partly *O*-methylated and *O*-(2-hydroxypropylated) cellulose.

Content
Methoxy (-OCH_3; M_r 31.03) and hydroxypropoxy (-OC_3H_6OH; M_r 75.09) groups (dried substance) conforming to the types of hypromellose set forth in the accompanying table.

Substitution type	Methoxy (per cent)	Hydroxypropoxy (per cent)
1828	16.5 to 20.0	23.0 to 32.0
2208	19.0 to 24.0	4.0 to 12.0
2906	27.0 to 30.0	4.0 to 7.5
2910	28.0 to 30.0	7.0 to 12.0

♦CHARACTERS
Appearance
White, yellowish-white or greyish-white powder or granules, hygroscopic after drying.

Solubility
Practically insoluble in hot water, in acetone, in anhydrous ethanol and in toluene. It dissolves in cold water giving a colloidal solution.♦

IDENTIFICATION
A. Evenly distribute 1.0 g onto the surface of 100 mL of *water R* in a beaker, tapping the top of the beaker gently if necessary to ensure a uniform layer on the surface. Allow to

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

stand for 1-2 min: the powdered material aggregates on the surface.

B. Evenly distribute 1.0 g into 100 mL of boiling *water R*, and stir the mixture using a magnetic stirrer with a bar 25 mm long: a slurry is formed and the particles do not dissolve. Allow the slurry to cool to 10 °C and stir using a magnetic stirrer: a clear or slightly turbid solution occurs with its thickness dependent on the viscosity grade.

C. To 0.1 mL of the solution obtained in identification test B add 9 mL of a 90 per cent *V/V* solution of *sulfuric acid R*, shake, heat on a water-bath for exactly 3 min, immediately cool in an ice-bath, carefully add 0.6 mL of a 20 g/L solution of *ninhydrin R*, shake and allow to stand at 25 °C: a red colour develops at first and changes to purple within 100 min.

D. Place 2-3 mL of the solution obtained in identification test B onto a glass slide as a thin film and allow the water to evaporate: a coherent, clear film forms on the glass slide.

E. Add 50.0 mL of the solution obtained in identification test B to 50.0 mL of *water R* in a beaker. Insert a thermometer into the solution. Stir the solution on a magnetic stirrer/hot plate and begin heating, increasing the temperature at a rate of 2-5 °C per minute. Determine the temperature at which a turbidity increase begins to occur and designate the temperature as the flocculation temperature: the flocculation temperature is higher than 50 °C.

TESTS

◊Appearance of solution

The solution is not more opalescent than reference suspension III (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

While stirring, introduce a quantity of the substance to be examined equivalent to 1.0 g of the dried substance into 50 g of *carbon dioxide-free water R* heated to 90 °C. Allow to cool, adjust the mass of the solution to 100 g with *carbon dioxide-free water R* and stir until dissolution is complete.◊

pH (*2.2.3*)

5.0 to 8.0 for the solution prepared as described under Viscosity.

Read the indicated pH value after the probe has been immersed for 5 ± 0.5 min.

Viscosity

80 per cent to 120 per cent of the nominal value for samples with a viscosity less than 600 mPa·s (Method 1); 75 per cent to 140 per cent of the nominal value for samples with a viscosity of 600 mPa·s or higher (Method 2).

Method 1, to be applied to samples with a viscosity of less than 600 mPa·s Weigh a quantity of the substance to be examined equivalent to 4.000 g of the dried substance. Transfer into a wide-mouthed bottle, and adjust the total mass of the sample and the water to 200.0 g with hot *water R* (90-99 °C). Capping the bottle, stir by mechanical means at 400 ± 50 r/min for 10-20 min until the particles are thoroughly dispersed and wetted. Scrape down the insides of the bottle with a spatula if necessary, to ensure that there is no undissolved material on the sides of the bottle, and continue the stirring in a cooling water-bath maintained at a temperature below 10 °C for another 20-40 min. Adjust the solution mass if necessary to 200.0 g using cold *water R*. Centrifuge the solution if necessary to expel any entrapped air bubbles. Using a spatula, remove any foam. Determine the kinematic viscosity (ν) of this solution using the capillary viscometer method (*2.2.9*). Separately determine the density (ρ) (*2.2.5*) of the solution and calculate the dynamic viscosity (η), as $\eta = \rho\nu$.

Method 2, to be applied to samples with a viscosity of 600 mPa·s or higher Weigh a quantity of the substance to be examined equivalent to 10.00 g of the dried substance. Transfer into a wide-mouthed bottle, and adjust the total mass of the sample and the water to 500.0 g with hot *water R* (90-99 °C). Capping the bottle, stir by mechanical means at 400 ± 50 r/min for 10-20 min until the particles are thoroughly dispersed and wetted. Scrape down the insides of the bottle with a spatula if necessary, to ensure that there is no undissolved material on the sides of the bottle, and continue the stirring in a cooling water-bath maintained at a temperature below 10 °C for another 20-40 min. Adjust the solution mass if necessary to 500.0 g using cold *water R*. Centrifuge the solution if necessary to expel any entrapped air bubbles. Using a spatula, remove any foam. Determine the viscosity (*2.2.10*) of this solution at 20 ± 0.1 °C using a rotating viscometer.

Apparatus Single-cylinder type spindle viscometer.

Rotor number, revolution and calculation multiplier Apply the conditions specified in Table 0348.-1.

Table 0348.-1.

Nominal viscosity* (mPa·s)	Rotor number	Revolution (r/min)	Calculation multiplier
600 to less than 1400	3	60	20
1400 to less than 3500	3	12	100
3500 to less than 9500	4	60	100
9500 to less than 99 500	4	6	1000
99 500 or more	4	3	2000

* the nominal viscosity is based on the manufacturer's specifications.

Allow the spindle to rotate for 2 min before taking the measurement. Allow a rest period of at least 2 min between subsequent measurements. Repeat the measurement twice and determine the mean of the 3 readings.

Loss on drying (*2.2.32*)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (*2.4.14*)

Maximum 1.5 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (*2.2.28*).

Apparatus:

- *reaction vial*: a 5 mL pressure-tight vial, 50 mm in height, 20 mm in external diameter and 13 mm in internal diameter at the mouth, equipped with a pressure-tight butyl rubber membrane stopper coated with polytetrafluoroethylene and secured with an aluminium crimped cap or another sealing system providing a sufficient air-tightness;
- *heater*: a heating module with a square aluminium block having holes 20 mm in diameter and 32 mm in depth, so that the reaction vials fit; mixing of the contents of the vial is effected using a magnetic stirrer equipped in the heating module or using a reciprocal shaker that performs approximately 100 cycles/min.

Internal standard solution 30 g/L solution of *octane R* in *o-xylene R*.

Test solution Weigh 65.0 mg of the substance to be examined, place in a reaction vial, add 0.06-0.10 g of *adipic acid R*, 2.0 mL of the internal standard solution and 2.0 mL of *hydriodic acid R*, immediately cap and seal the vial, and weigh accurately. Mix the contents of the vial continuously for 60 min while heating the block so that the temperature of the contents is maintained at 130 ± 2 °C. If a reciprocal shaker or magnetic stirrer cannot be used, shake the vial thoroughly by hand at 5 min intervals during the initial 30 min of the heating time. Allow the vial to cool, and again weigh accurately. If the loss of mass is less than 26 mg and there is no evidence of a leak, use the upper layer of the mixture as the test solution.

Reference solution Place 0.06-0.10 g of *adipic acid R*, 2.0 mL of the internal standard solution and 2.0 mL of *hydriodic acid R* in another reaction vial, cap and seal the vial, and weigh accurately. Add 15-22 µL of *isopropyl iodide R* through the septum with a syringe, weigh accurately, add 45 µL of *methyl iodide R* in the same manner, and weigh accurately. Shake the reaction vial thoroughly and use the upper layer as the reference solution.

Use a precolumn if needed.

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *poly(dimethyl)siloxane R* (3 µm).

Carrier gas *helium for chromatography R.*

Flow rate 4.3 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 3	50
	3 - 8	50 → 100
	8 - 12.3	100 → 250
	12.3 - 20.3	250
Injection port		250
Detector		280

Detection Flame ionisation or thermal conductivity.

Injection 1-2 µL.

Relative retention With reference to octane (retention time = about 10 min): methyl iodide = about 0.4; isopropyl iodide = about 0.7.

System suitability Reference solution:
— *resolution*: minimum 5.0 between the peaks due to methyl iodide and isopropyl iodide and between the peaks due to isopropyl iodide and octane;
— *repeatability*: maximum relative standard deviation of 2.0 per cent for the ratios of the areas of the peaks respectively due to methyl iodide and isopropyl iodide to the area of the peak due to octane, determined on 6 injections.

Calculate the ratios (Q_1 and Q_2) of the areas of the peaks due to methyl iodide and isopropyl iodide to the area of the peak due to the internal standard in the chromatogram obtained with the test solution, and the ratios (Q_3 and Q_4) of the areas of the peaks due to methyl iodide and isopropyl iodide to the area of the peak due to the internal standard in the chromatogram obtained with the reference solution.

Calculate the percentage content of methoxy groups using the following expression:

$$\frac{Q_1}{Q_3} \times \frac{m_1}{m} \times 21.864$$

Calculate the percentage content of hydroxypropoxy groups using the following expression:

$$\frac{Q_2}{Q_4} \times \frac{m_2}{m} \times 44.17$$

m_1 = mass of methyl iodide in the reference solution, in milligrams;
m_2 = mass of isopropyl iodide in the reference solution, in milligrams;
m = mass of the sample (dried substance), in milligrams.

LABELLING

The label states:
— the nominal viscosity in millipascal seconds (mPa·s);
— the substitution type.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hypromellose used as binder, viscosity-increasing agent or film former.

Viscosity
See Tests.

Degree of substitution
See Assay.

The following characteristics may be relevant for hypromellose used as matrix former in prolonged-release tablets.

Viscosity
See Tests.

Degree of substitution
See Assay.

Molecular mass distribution (*2.2.30*)

Particle-size distribution (*2.9.31* or *2.9.38*)

Powder flow (*2.9.36*)

Ph Eur

Hypromellose Phthalate

(Ph. Eur. monograph 0347)

Action and use
Artificial tears.

Ph Eur

DEFINITION

Hydroxypropylmethylcellulose phthalate.

Monophthalic acid ester of hypromellose, containing methoxy (-OCH_3), 2-hydroxypropoxy (-$OCH_2CHOHCH_3$) and phthaloyl (*o*-carboxybenzoyl $C_8H_5O_3$) groups.

CHARACTERS

Appearance

White or almost white, free-flowing flakes or granular powder.

Solubility

Practically insoluble in water, soluble in a mixture of equal volumes of acetone and methanol and in a mixture of equal volumes of methanol and methylene chloride, very slightly soluble in acetone and in toluene, practically insoluble in anhydrous ethanol.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Preparation Dissolve 40 mg in 1 mL of a mixture of equal volumes of *methanol R* and *methylene chloride R*; spread 2 drops of this solution between 2 sodium chloride plates, then remove one of the plates to evaporate the solvent.

Comparison *hypromellose phthalate CRS*.

TESTS

Free phthalic acid

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.20 g of the substance to be examined in about 50 mL of *acetonitrile R* with the aid of ultrasound. Add 10 mL of *water R*, cool to room temperature, dilute to 100.0 mL with *acetonitrile R* and mix.

Reference solution Dissolve 12.5 mg of *phthalic acid R* in 125 mL of *acetonitrile R*. Add 25 mL of *water R*, dilute to 250.0 mL with *acetonitrile R* and mix.

Column:

— *size*: l = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5-10 µm).

Mobile phase *acetonitrile R*, 1 g/L solution of *trifluoroacetic acid R* (1:9 *V/V*).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 10 µL.

System suitability Reference solution:

— *repeatability*: maximum relative standard deviation of 1.0 per cent after 2 injections.

Limit:

— *phthalic acid*: not more than 0.4 times the area of the corresponding peak in the chromatogram obtained with the reference solution (1.0 per cent).

Chlorides

Maximum 0.07 per cent.

Dissolve 1.0 g in 40 mL of *0.2 M sodium hydroxide*, add 0.05 mL of *phenolphthalein solution R* and add *dilute nitric acid R* dropwise, with stirring, until the red colour disappears. Add an additional 20 mL of *dilute nitric acid R* with stirring. Heat on a water-bath with stirring until the gel-like precipitate formed becomes granular. Cool and centrifuge. Separate the liquid phase and wash the residue with 3 quantities, each of 20 mL, of *water R*, separating the washings by centrifugation. Combine the liquid phases, dilute to 200 mL with *water R*, mix and filter. To 50 mL of this solution, add 1 mL of *0.1 M silver nitrate*. The solution is not more opalescent than a standard prepared by mixing 0.5 mL of *0.01 M hydrochloric acid* with 10 mL of *0.2 M sodium hydroxide*, adding 7 mL of *dilute nitric acid R* and 1 mL of *0.1 M silver nitrate*, and diluting to 50 mL with *water R*.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.2 per cent, determined on 1.0 g.

STORAGE

In an airtight container.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for hypromellose phthalate used as a gastro-resistant coating agent.

Apparent viscosity (*2.2.9*)

80 per cent to 120 per cent of the nominal value.

Dissolve 10 g, previously dried at 105 °C for 1 h, in 90 g of a mixture of equal masses of *methanol R* and *methylene chloride R* by mixing and shaking.

Solubility

0.2 g does not dissolve in *0.1 M hydrochloric acid* but dissolves quickly and completely in 100 mL of phosphate buffer solution pH 6.8 R with stirring.

Phthaloyl groups

Typically 21.0 per cent to 35.0 per cent (anhydrous substance).

Dissolve 1.000 g in 50 mL of a mixture of 1 volume of *water R*, 2 volumes of *acetone R* and 2 volumes of *ethanol (96 per cent) R*. Add 0.1 mL of *phenolphthalein solution R* and titrate with *0.1 M sodium hydroxide* until a faint pink colour is obtained. Carry out a blank titration.

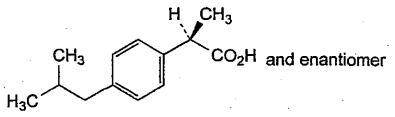
Calculate the percentage content of phthaloyl groups using the following expression:

$$\frac{149n}{(100-a)m} - 1.795S$$

a = percentage content of water;
m = mass of the substance to be examined, in grams;
n = volume of *0.1 M sodium hydroxide* used, in millilitres;
S = percentage content of free phthalic acid (see Tests).

Ibuprofen

(Ph. Eur. monograph 0721)



$C_{13}H_{18}O_2$ 206.3 *15687-27-1*

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Ibuprofen Capsules
Ibuprofen Gel
Ibuprofen Oral Suspension
Ibuprofen Orodispersible Tablets
Ibuprofen Prolonged-release Capsules
Ibuprofen Prolonged-release Tablets
Ibuprofen Tablets

Ph Eur

DEFINITION
(2*RS*)-2-[4-(2-Methylpropyl)phenyl]propanoic acid.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Practically insoluble in water, freely soluble in acetone, in methanol and in methylene chloride. It dissolves in dilute solutions of alkali hydroxides and carbonates.

IDENTIFICATION
First identification: A, C.

Second identification: A, B, D.

A. Melting point (*2.2.14*): 75 °C to 78 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in a 4 g/L solution of *sodium hydroxide R* and dilute to 100.0 mL with the same alkaline solution.

Spectral range 240-300 nm, using a spectrophotometer with a band width of 1.0 nm and a scan speed of not more than 50 nm/min.

Absorption maxima At 264 nm and 272 nm.

Shoulder At 258 nm.

Absorbance ratio:
— A_{264} / A_{258} = 1.20 to 1.30;
— A_{272} / A_{258} = 1.00 to 1.10.

C. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ibuprofen CRS.*

D. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *methylene chloride R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of *ibuprofen CRS* in *methylene chloride R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel plate R.*

Mobile phase *anhydrous acetic acid R, ethyl acetate R, hexane R* (5:24:71 *V/V/V*).

Application 5 µL.

Development Over a path of 10 cm.

Drying At 120 °C for 30 min.

Detection Lightly spray with a 10 g/L solution of *potassium permanganate R* in *dilute sulfuric acid R* and heat at 120 °C for 20 min; examine in ultraviolet light at 365 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS
Solution S
Dissolve 2.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.05° to + 0.05°.

Dissolve 0.50 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 20 mg of the substance to be examined in 2 mL of *acetonitrile R* and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of *ibuprofen impurity B CRS* to 10.0 mL with *acetonitrile R* (solution A). Dissolve 20 mg of *ibuprofen CRS* in 2 mL of *acetonitrile R*, add 1.0 mL of solution A and dilute to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of *ibuprofen for peak identification CRS* (mixture of impurities A, J and N) in 1 mL of *acetonitrile R* and dilute to 5 mL with mobile phase A.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: mix 0.5 volumes of *phosphoric acid R*, 340 volumes of *acetonitrile R1* and 600 volumes of *water for chromatography R*; allow to equilibrate and dilute to 1000 volumes with *water for chromatography R*;
— *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 25	100	0
25 - 55	100 → 15	0 → 85
55 - 70	15	85

Flow rate 2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *ibuprofen for peak identification CRS* and the

chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, J and N.

Relative retention With reference to ibuprofen (retention time = about 21 min): impurity J = about 0.2; impurity N = about 0.3; impurity A = about 0.9; impurity B = about 1.1.

System suitability Reference solution (b):
— *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity B, and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to ibuprofen. If necessary, adjust the concentration of acetonitrile in mobile phase A.

Limits:
— *impurities A, J, N*: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Impurity F

Gas chromatography (*2.2.28*): use the normalisation procedure.

Methylating solution Dilute 1 mL of *N,N-dimethylformamide dimethylacetal R* and 1 mL of *pyridine R* to 10 mL with *ethyl acetate R*.

Test solution Weigh about 50.0 mg of the substance to be examined into a sealable vial, dissolve in 1.0 mL of *ethyl acetate R*, add 1 mL of the methylating solution, seal and heat at 100 °C in a block heater for 20 min. Allow to cool. Remove the reagents under a stream of nitrogen at room temperature. Dissolve the residue in 5 mL of *ethyl acetate R*.

Reference solution (a) Dissolve 0.5 mg of *ibuprofen impurity F CRS* in *ethyl acetate R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Weigh about 50.0 mg of *ibuprofen CRS* into a sealable vial, dissolve in 1.0 mL of reference solution (a), add 1 mL of the methylating solution, seal and heat at 100 °C in a block heater for 20 min. Allow to cool. Remove the reagents under a stream of nitrogen at room temperature. Dissolve the residue in 5 mL of *ethyl acetate R*.

Column:
— *material*: fused silica;
— *size*: *l* = 25 m, Ø = 0.53 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 2 µm).

Carrier gas *helium for chromatography R*.

Flow rate 5.0 mL/min.

Temperature:
— *column*: 150 °C;
— *injection port*: 200 °C;
— *detector*: 250 °C.

Detection Flame ionisation.

Injection 1 µL of the test solution and reference solution (b).

Run time Twice the retention time of ibuprofen.

System suitability:
— *relative retention* with reference to ibuprofen (retention time = about 17 min): impurity F = about 1.5.

Limit:
— *impurity F*: maximum 0.1 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo*.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

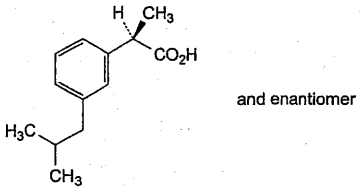
Dissolve 0.450 g in 50 mL of *methanol R*. Add 0.4 mL of *phenolphthalein solution R1*. Titrate with *0.1 M sodium hydroxide* until a red colour is obtained. Carry out a blank titration.

1 mL of *0.1 M sodium hydroxide* is equivalent to 20.63 mg of $C_{13}H_{18}O_2$.

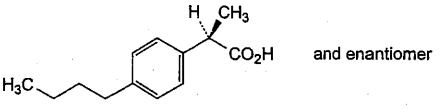
IMPURITIES

Specified impurities *A, F, J, N.*

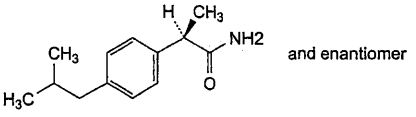
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, E, G, H, I, K, L, M, O, P, Q, R.*



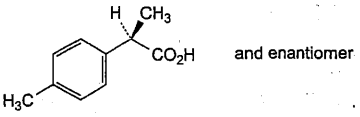
A. (2*RS*)-2-[3-(2-methylpropyl)phenyl]propanoic acid,



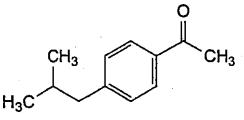
B. (2*RS*)-2-(4-butylphenyl)propanoic acid,



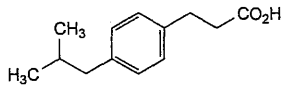
C. (2*RS*)-2-[4-(2-methylpropyl)phenyl]propanamide,



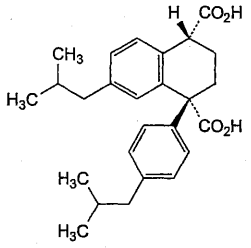
D. (2*RS*)-2-(4-methylphenyl)propanoic acid,



E. 1-[4-(2-methylpropyl)phenyl]ethanone,

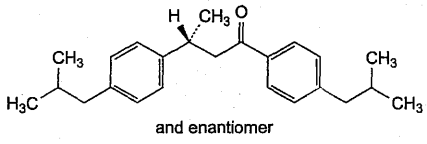


F. 3-[4-(2-methylpropyl)phenyl]propanoic acid,



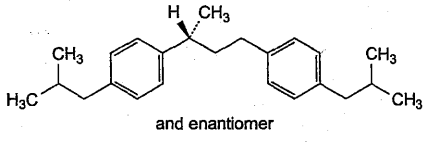
and enantiomer

G. (1*RS*,4*RS*)-7-(2-methylpropyl)-1-[4-(2-methylpropyl)phenyl]-1,2,3,4-tetrahydronaphthalene-1,4-dicarboxylic acid,



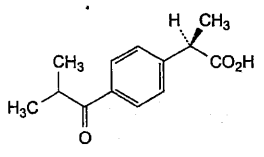
and enantiomer

H. (3*RS*)-1,3-bis[4-(2-methylpropyl)phenyl]butan-1-one,



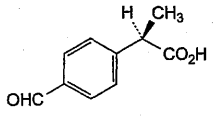
and enantiomer

I. 1-(2-methylpropyl)-4-[(3*RS*)-3-[4-(2-methylpropyl)phenyl]butyl]benzene,



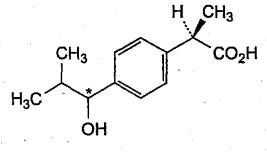
and enantiomer

J. (2*RS*)-2-[4-(2-methylpropanoyl)phenyl]propanoic acid,



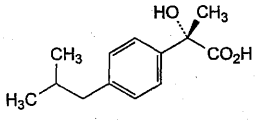
and enantiomer

K. (2*RS*)-2-(4-formylphenyl)propanoic acid,



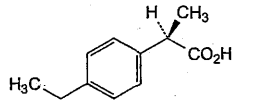
and enantiomer

L. (2*RS*)-2-[4-(1-hydroxy-2-methylpropyl)phenyl]propanoic acid,



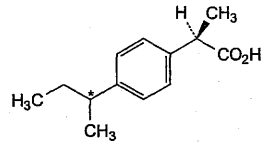
and enantiomer

M. (2*RS*)-2-hydroxy-2-[4-(2-methylpropyl)phenyl]propanoic acid,



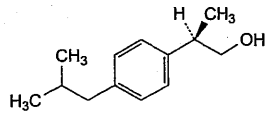
and enantiomer

N. (2*RS*)-2-(4-ethylphenyl)propanoic acid,



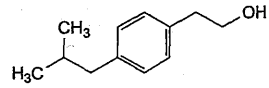
and enantiomer

O. (2*RS*)-2-[4-(1-methylpropyl)phenyl]propanoic acid,

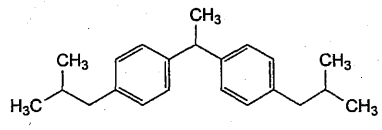


and enantiomer

P. (2*RS*)-2-[4-(2-methylpropyl)phenyl]propan-1-ol,



Q. 2-[4-(2-methylpropyl)phenyl]ethanol,



R. 1,1′-(ethane-1,1-diyl)-4,4′-(2-methylpropyl)dibenzene.

Ph Eur

Ichthammol

Ammonium Ichthosulphonate

(Ph. Eur. monograph 0917)

Action and use
Chronic lichenified eczema.

Preparation
Zinc and Ichthammol Cream

Ph Eur

DEFINITION

Ichthammol is obtained by distillation from certain bituminous schists, sulfonation of the distillate and neutralisation of the product with ammonia.

Content

— *dry matter*: 50.0 per cent *m/m* to 56.0 per cent *m/m*;
— *total ammonia* (NH_3; M_r 17.03): 4.5 per cent *m/m* to 7.0 per cent *m/m* (dried substance);
— *organically combined sulfur*: minimum 10.5 per cent *m/m* (dried substance);
— *sulfur in the form of sulfate*: maximum 20.0 per cent *m/m* of the total sulfur.

CHARACTERS

Appearance
Dense, blackish-brown liquid.

Solubility
Miscible with water and with glycerol, slightly soluble in ethanol (96 per cent), in fatty oils and in liquid paraffin.

It forms homogeneous mixtures with wool fat and soft paraffin.

IDENTIFICATION

A. Dissolve 1.5 g in 15 mL of *water R* (solution A). To 2 mL of solution A add 2 mL of *hydrochloric acid R*. A resinous precipitate is formed. Decant the supernatant.
The precipitate is partly soluble in *ether R*.

B. 2 mL of solution A, obtained in identification test A, gives the reaction of ammonium salts and salts of volatile bases (*2.3.1*).

C. Evaporate and ignite the mixture of solution A and *dilute sodium hydroxide solution R* obtained in identification test B. Take up the residue with 5 mL of *dilute hydrochloric acid R*. Gas is evolved which turns *lead acetate paper R* brown or black. Filter the solution. The filtrate gives reaction (a) of sulfates (*2.3.1*).

TESTS

Acidity or alkalinity

To 10.0 mL of the clear filtrate obtained in the assay of total ammonia add 0.05 mL of *methyl red solution R*. Not more than 0.2 mL of *0.02 M hydrochloric acid* or *0.02 M sodium hydroxide* is required to change the colour of the indicator.

Relative density (*2.2.5*)

1.040 to 1.085, determined on a mixture of equal volumes of the substance to be examined and *water R*.

Sulfated ash (*2.4.14*)

Maximum 0.3 per cent, determined on 1.00 g.

ASSAY

Dry matter

Weigh 1.000 g in a tared flask containing 2 g of *sand R*, previously dried to constant mass, and a small glass rod. Heat on a water-bath for 2 h with frequent stirring and dry in an oven at 100-105 °C until 2 consecutive weighings do not differ by more than 2.0 mg; the 2nd weighing is carried out after drying again for 1 h.

Total ammonia

Dissolve 2.50 g in 25 mL of warm *water R*. Rinse the solution into a 250 mL volumetric flask, add 200 mL of *sodium chloride solution R* and dilute to 250.0 mL with *water R*. Filter the solution, discarding the first 20 mL of filtrate. To 100.0 mL of the clear filtrate add 25 mL of *formaldehyde solution R*, neutralised to *phenolphthalein solution R1*. Titrate with *0.1 M sodium hydroxide* until a faint pink colour is obtained.

1 mL of *0.1 M sodium hydroxide* is equivalent to 1.703 mg of NH_3.

Organically combined sulfur

Mix 0.500 g with 4 g of *anhydrous sodium carbonate R* and 3 mL of *methylene chloride R* in a porcelain crucible of about 50 mL capacity, warm and stir until all the methylene chloride has evaporated. Add 10 g of coarsely powdered copper nitrate R, mix thoroughly and heat the mixture very gently using a small flame. When the initial reaction has subsided, increase the temperature slightly until most of the material has blackened. Cool, place the crucible in a large beaker, add 20 mL of *hydrochloric acid R* and, when the reaction has ceased, add 100 mL of *water R* and boil until all the copper oxide has dissolved. Filter the solution, add 400 mL of *water R*, heat to boiling and add 20 mL of *barium chloride solution R1*. Allow to stand for 2 h, filter, wash with *water R*, dry and ignite at about 600 ± 50 °C until 2 successive weighings do not differ by more than 0.2 per cent of the mass of the residue.

1 g of residue is equivalent to 0.1374 g of total sulfur.

Calculate the percentage content of total sulfur and subtract the percentage content of sulfur in the form of sulfate.

Sulfur in the form of sulfate

Dissolve 2.000 g in 100 mL of *water R*, add 2 g of *cupric chloride R* dissolved in 80 mL of *water R* and dilute to 200.0 mL with *water R*. Shake and filter. Heat 100.0 mL of the filtrate almost to boiling, add 1 mL of *hydrochloric acid R* and 5 mL of *barium chloride solution R1* dropwise and heat on a water-bath. Filter, wash the precipitate with *water R*, dry and ignite at about 600 ± 50 °C until 2 successive weighings do not differ by more than 0.2 per cent of the mass of the residue.

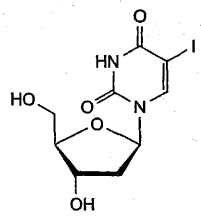
1 g of residue is equivalent to 0.1374 g of sulfur present in the form of sulfate.

Calculate the percentage content of sulfur in the form of sulfate.

Ph Eur

Idoxuridine

(Ph. Eur. monograph 0669)



$C_9H_{11}IN_2O_5$ 354.1 *54-42-2*

Action and use

Pyrimidine nucleoside analogue; antiviral (herpes viruses).

Ph Eur

DEFINITION

Idoxuridine contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of 5-iodo-1-(2-deoxy-β-D-*erythro*-pentofuranosyl)pyrimidine-2,4(1*H*,3*H*)-dione, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, slightly soluble in water and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

It melts at about 180 °C, with decomposition.

IDENTIFICATION

First identification: A.

Second identification: B, C, D.

A. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *idoxuridine CRS*. Examine the substances as discs prepared using 1 mg of the substance to be examined and of the reference substance each in 0.3 g of *potassium bromide R*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (c).

C. Heat about 5 mg in a test-tube over a naked flame. Violet vapour is evolved.

D. Disperse about 2 mg in 1 mL of *water R* and add 2 mL of *diphenylamine solution R2*. Heat in a water-bath for 10 min. A persistent light-blue colour develops.

TESTS

Solution S

Dissolve 0.500 g in *1 M sodium hydroxide* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

Dissolve 0.10 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent. The pH of the solution is 5.5 to 6.5.

Specific optical rotation (*2.2.7*)

+ 28 to + 32, determined on solution S and calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (*2.2.27*), using as coating substance a suitable silica gel with a fluorescent indicator having an optimal intensity at 254 nm.

Test solution (a) Dissolve 0.20 g of the substance to be examined in a mixture of 1 volume of *concentrated ammonia R* and 5 volumes of *methanol R* and dilute to 5 mL with the same mixture of solvents.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of *concentrated ammonia R* and 5 volumes of *methanol R*.

Reference solution (a) Dissolve 20 mg of *5-iodouracil R*, 20 mg of *2′-deoxyuridine R* and 20 mg of *5-bromo-2′-deoxyuridine R* in a mixture of 1 volume of *concentrated ammonia R* and 5 volumes of *methanol R* and dilute to 100 mL with the same mixture of solvents.

Reference solution (b) Dissolve 0.20 g of the substance to be examined in 5 mL of reference solution (a).

Reference solution (c) Dissolve 20 mg of *idoxuridine CRS* in a mixture of 1 volume of *concentrated ammonia R* and 5 volumes of *methanol R* and dilute to 5 mL with the same mixture of solvents.

Reference solution (d) Dilute 1 mL of test solution (b) to 20 mL with a mixture of 1 volume of *concentrated ammonia R* and 5 volumes of *methanol R*.

Apply separately to the plate 5 µL of each solution. Develop twice over a path of 15 cm using a mixture of 10 volumes of *concentrated ammonia R*, 40 volumes of *chloroform R* and 50 volumes of *2-propanol R*, drying the plate in a current of cold air after each development. Examine in ultraviolet light at 254 nm. In the chromatogram obtained with test solution (a): any spots corresponding to 5-iodouracil, 2′-deoxyuridine and 5-bromo-2′-deoxyuridine are not more intense than the corresponding spots in the chromatogram obtained with reference solution (a) (0.5 per cent); any spot, apart from the principal spot and the spots corresponding to 5-iodouracil, 2′-deoxyuridine and 5-bromo-2′-deoxyuridine, is not more intense than the spot in the chromatogram obtained with reference solution (d) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (b) shows four clearly separated spots.

Iodide

Dissolve 0.25 g in 25 mL of *0.1 M sodium hydroxide*, add 5 mL of *dilute hydrochloric acid R* and dilute to 50 mL with *water R*. Allow to stand for 10 min and filter. To 25 mL of the filtrate add 5 mL of *dilute hydrogen peroxide solution R* and 10 mL of *chloroform R* and shake. Any pink colour in the organic layer is not more intense than that in a standard prepared at the same time in the same manner using 1 mL of a 0.33 g/L solution of *potassium iodide R* instead of the substance to be examined (0.1 per cent).

Loss on drying (*2.2.32*)

Not more than 1.0 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

Sulfated ash (*2.4.14*)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.3000 g in 20 mL of *dimethylformamide R*. Titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (*2.2.20*).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 35.41 mg of $C_9H_{11}IN_2O_5$.

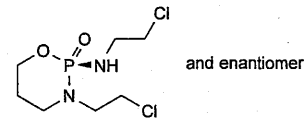
STORAGE

Store protected from light.

Ph Eur

Ifosfamide

(Ph. Eur. monograph 1529)



and enantiomer

$C_7H_{15}Cl_2N_2O_2P$ 261.1 *3778-73-2*

Action and use

Cytotoxic alkylating agent.

Preparation

Ifosfamide Injection

Ph Eur

DEFINITION

Ifosfamide contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of (*RS*)-*N*,3-bis(2-chloroethyl)-1,3,2-oxazaphosphinan-2-amine 2-oxide, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, fine, crystalline powder, hygroscopic, soluble in water, freely soluble in methylene chloride.

IDENTIFICATION

Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the *Ph. Eur. reference spectrum of ifosfamide*. Examine the substance prepared as a disc.

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Acidity or alkalinity
Dilute 5 mL of solution S to 50 mL with *carbon dioxide-free water R*. To 10 mL of this solution add 0.1 mL of *methyl red solution R*. Not more than 0.1 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator to red. To another 10 mL of the solution add 0.1 mL of *phenolphthalein solution R*. Not more than 0.3 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pink.

Optical rotation (*2.2.7*)
The angle of optical rotation, determined on solution S, is −0.10° to + 0.10°.

Related substances
A. Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel plate R*.

Test solution Dissolve 1.00 g of the substance to be examined in a mixture of equal volumes of *methanol R* and *water R* and dilute to 10 mL with the same mixture of solvents.

Reference solution (a) Dissolve 25 mg of *ifosfamide impurity A CRS* and 25 mg of *chloroethylamine hydrochloride R* (impurity C) ina mixture of equal volumes of *methanol R* and *water R* and dilute to 100 mL with the same mixture of solvents.

Reference solution (b) Dissolve 15 mg of *ifosfamide impurity B CRS* ina mixture of equal volumes of *methanol R* and *water R* and dilute to 100 mL with the same mixture of solvents.

Reference solution (c) Dissolve 5 mg of *ethanolamine R* (impurity D), 20 mg of *ifosfamide impurity A CRS* and 80 mg of *chloroethylamine hydrochloride R* (impurity C) in a mixture of equal volumes of *methanol R* and *water R* and dilute to 100 mL with the same mixture of solvents.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of *water R*, 15 volumes of *methanol R*, 25 volumes of *anhydrous acetic acid R* and 50 volumes of *methylene chloride R*. Dry the plate at 115 °C for 45 min. At the bottom of a chromatographic tank, place an evaporating dish containing a 3.2 g/L solution of *potassium permanganate R* and add an equal volume of *dilute hydrochloric acid R*, close the tank and allow to stand for 10 min. Place the plate whilst still hot in the tank, avoiding contact of the stationary phase with the solution, and close the tank. Leave the plate in contact with the chlorine vapour for 20 min. Withdraw the plate and place it in a current of cold air until the excess of chlorine is removed (about 20 min) and an area of coating below the points of application does not give a blue colour with a drop of *potassium iodide and starch solution R*. Avoid prolonged exposure to cold air. Immerse the plate in a 1 g/L solution of *tetramethylbenzidine R* in *alcohol R* for 5 s. Allow the plate to dry and examine. In the chromatogram obtained with the test solution: any spot corresponding to impurity A or impurity C is not more intense than the corresponding spot in the chromatogram obtained with reference solution (a) (0.25 per cent); any spot corresponding to impurity B is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.15 per cent); any other spot is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.15 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows 3 clearly separated spots.

B. Examine by thin-layer chromatography (*2.2.27*), using a *TLC silica gel plate R*.

Test solution Dissolve 0.200 g of the substance to be examined in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 10 mL with the same mixture of solvents.

Reference solution (a) Dissolve 5 mg of *ifosfamide impurity E CRS* and 5 mg of *ifosfamide impurity F CRS* in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 100 mL with the same mixture of solvents.

Reference solution (b) Dissolve 10 mg of *ifosfamide impurity E CRS* and 10 mg of *ifosfamide CRS* in a mixture of equal volumes of *methanol R* and *methylene chloride R* and dilute to 100 mL with the same mixture of solvents.

Apply to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of *methylene chloride R* and 10 volumes of *acetone R*. Dry the plate at 115 °C for 45 min. Proceed as described in test A for related substances. Any spot corresponding to impurity E or impurity F in the chromatogram obtained with the test solution is not more intense than the corresponding spot in the chromatogram obtained with reference solution (a) (0.25 per cent). The test is not valid unless the chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Chlorides (*2.4.4*)
Dilute 5 mL of solution S to 15 mL with *water R*.
The freshly prepared solution complies with the limit test for chlorides (100 ppm).

Water (*2.5.12*)
Not more than 0.5 per cent, determined on 1.00 g by the semi-micro determination of water.

ASSAY
Examine by liquid chromatography (*2.2.29*). *Use the solutions within 24 h.*

Solution A Dissolve 50.0 mg of *ethyl parahydroxybenzoate R* in 25 mL of *alcohol R*, dilute to 100.0 mL with *water R* and mix.

Test solution To 0.150 g of the substance to be examined add 10.0 mL of solution A and dilute to 250.0 mL with *water R*.

Reference solution To 15.0 mg of *ifosfamide CRS* add 1.0 mL of solution A and dilute to 25.0 mL with *water R*.

The chromatography may be carried out using:
— a stainless steel column 0.25 m long and 4.6 mm in internal diameter packed with *octadecylsilyl silica gel for chromatography R* (5 µm),
— as mobile phase at a flow rate of 1.5 mL/min a mixture of 30 volumes of *acetonitrile R* and 70 volumes of *water R*,
— as detector a spectrophotometer set at 195 nm.

Inject 1 µL of the reference solution six times. The assay is not valid unless the resolution between the peaks due to ifosfamide and to ethyl parahydroxybenzoate is not less than 6.0 and the relative standard deviation of the peak area for ifosfamide is at most 2.0 per cent.

Inject 1 µL of the test solution. Calculate the percentage content of $C_7H_{15}Cl_2N_2O_2P$ from the area of the corresponding peak in the chromatogram obtained and the declared content of *ifosfamide CRS*.

STORAGE
Store in an airtight container.

IMPURITIES

Test A for related substances

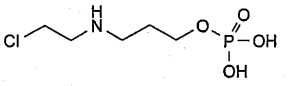
A, B, C, D.

Test B for related substances

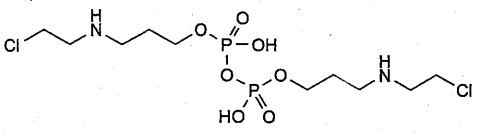
E, F.

Specified impurities *A, B, C, E, F.*

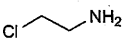
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *D.*



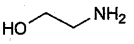
A. 3-[(2-chloroethyl)amino]propyl dihydrogen phosphate,



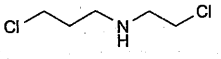
B. bis[3-[(2-chloroethyl)amino]propyl] dihydrogen diphosphate,



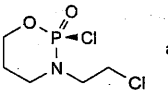
C. 2-chloroethanamine,



D. 2-aminoethanol.



E. 3-chloro-*N*-(2-chloroethyl)propan-1-amine,



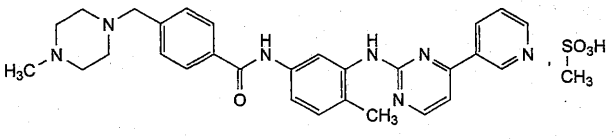
and enantiomer

F. (*RS*)-2-chloro-3-(2-chloroethyl)-1,3,2-oxazaphosphinane 2-oxide.

Ph Eur

Imatinib Mesilate

(Ph. Eur. monograph 2736)



$C_{30}H_{35}N_7SO_4$ 589.7 *220127-57-1*

Action and use

Cytotoxic.

Ph Eur

DEFINITION

4-[(4-Methylpiperazin-1-yl)methyl]-*N*-[4-methyl-3-[[4-(pyridin-3-yl)pyrimidin-2-yl]amino]phenyl]benzamide methanesulfonate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

It is considered that alkyl methanesulfonate esters are genotoxic and are potential impurities in imatinib mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods *2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid*, *2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances* and *2.5.39. Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance

White or almost white, slightly brownish or yellowish powder; yellow or pale yellow, very hygroscopic, for the amorphous form.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *imatinib mesilate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *anhydrous ethanol R*, evaporate to dryness and record new spectra using the residues.

TESTS

Impurity F

Liquid chromatography (*2.2.29*) coupled with mass spectrometry (*2.2.43*).

Solvent mixture *acetonitrile R1, water for chromatography R* (30:70 *V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution Dissolve 2.0 mg of *imatinib impurity F CRS* in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 200.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— *size*: *l* = 0.15 m, Ø = 3.0 mm;

— *stationary phase*: *end-capped octadecylsilyl amorphous organosilica polymer for chromatography R* (3.5 µm);

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: 1.26 g/L solution of *ammonium formate R* in *water for chromatography R* adjusted to pH 3.4-3.5 with *anhydrous formic acid R*;

— *mobile phase B*: 0.05 per cent *V/V* solution of *anhydrous formic acid R* in *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 6	80	20
6 - 10	80 → 20	20 → 80
10 - 15	20	80

NOTE: MS acquisition can be started at 3.5 min and stopped at 6 min; during non-acquisition the eluent is directed to waste.

Flow rate 0.5 mL/min.

Detection Mass detector: the following settings have been found to be suitable and are given as examples; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criterion:
— *ionisation*: ESI-positive;
— *detection m/z (SIM)*: 278.2;
— *gas temperature*: 350 °C;
— *drying gas flow*: 12 L/min;
— *nebuliser pressure*: 414 kPa;
— *capillary voltage (Vcap)*: 3 kV.

Injection 10 µL.

System suitability Reference solution:
— *signal-to-noise ratio*: minimum 20 for the principal peak;
— *repeatability*: maximum relative standard deviation of 10 per cent determined on 6 injections.

Calculation of percentage content:
— for impurity F, use the concentration of impurity F in the reference solution.

Limit:
— *impurity F*: maximum 20 ppm.

Impurity H

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water for chromatography R* (30:70 *V/V*).

Test solution Dissolve 75.0 mg of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of *imatinib impurity A CRS* in 1.0 mL of the solvent mixture.

Reference solution (b) Dissolve 60.0 mg of *imatinib impurity H CRS* in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 50.0 mL with the solvent mixture.

Reference solution (d) Dissolve 0.150 g of the substance to be examined in the solvent mixture, add 1.0 mL each of reference solutions (a) and (b) and dilute to 10.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: dissolve 2.3 g of *sodium octanesulfonate monohydrate R* in 700 mL of *water for chromatography R* and add 300 mL of *acetonitrile R1* and 1.2 mL of *dilute phosphoric acid R*;
— *mobile phase B*: dissolve 2.3 g of *sodium octanesulfonate monohydrate R* in 100 mL of *water for chromatography R* and add 900 mL of *acetonitrile R1* and 1.2 mL of *dilute phosphoric acid R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 6	98	2
6 - 8	98 → 20	2 → 80
8 - 10	20	80

Flow rate 2.3 mL/min.

Detection Spectrophotometer at 227 nm.

Injection 10 µL of the test solution and reference solutions (c) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A and H.

Relative retention With reference to imatinib (retention time = about 8 min): impurity A = about 0.17; impurity H = about 0.2.

System suitability Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to impurities A and H.

Calculation of percentage content:
— for impurity H, use the concentration of impurity H in reference solution (c).

Limit:
— *impurity H*: maximum 0.02 per cent.

Related substances

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R1, water for chromatography R* (30:70 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 1 mg of *imatinib for system suitability CRS* (containing impurities A, B, C, D and J) in the solvent mixture and dilute to 2 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of *imatinib mesilate CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 35 °C.

Mobile phase:
— *mobile phase A*: dissolve 2.3 g of *sodium octanesulfonate monohydrate R* in 700 mL of *water for chromatography R* and add 300 mL of *acetonitrile R1* and 1.2 mL of *dilute phosphoric acid R*;
— *mobile phase B*: dissolve 2.3 g of *sodium octanesulfonate monohydrate R* in 100 mL of *water for chromatography R* and add 900 mL of *acetonitrile R1* and 1.2 mL of *dilute phosphoric acid R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 16	98	2
16 - 30	98 → 50	2 → 50

Flow rate 2.3 mL/min.

Detection Spectrophotometer at 267 nm.

Injection 10 µL of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with *imatinib for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and J.

Relative retention With reference to imatinib (retention time = about 11 min): impurity A = about 0.2; impurity B = about 0.6; impurity J = about 0.9; impurity C = about 1.2; impurity D = about 2.3.

System suitability:
- *resolution*: minimum 3.0 between the peaks due to imatinib and impurity C in the chromatogram obtained with reference solution (b);
- *signal-to-noise ratio*: minimum 45 for the principal peak in the chromatogram obtained with reference solution (a);
- *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the peak due to impurity J and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to imatinib in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 2.2; impurity B = 2.0;
- for each impurity, use the concentration of imatinib mesilate in reference solution (a).

Limits:
- *impurity C*: maximum 0.3 per cent;
- *impurity D*: maximum 0.2 per cent;
- *impurities A, B*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.8 per cent;
- *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)

Maximum 3.0 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

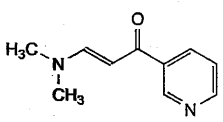
Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{30}H_{35}N_7SO_4$ taking into account the assigned content of *imatinib mesilate CRS*.

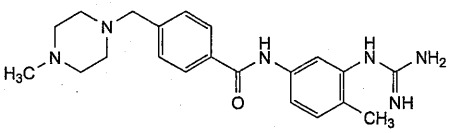
IMPURITIES

Specified impurities *A, B, C, D, F, H.*

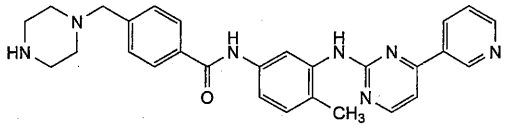
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *J.*



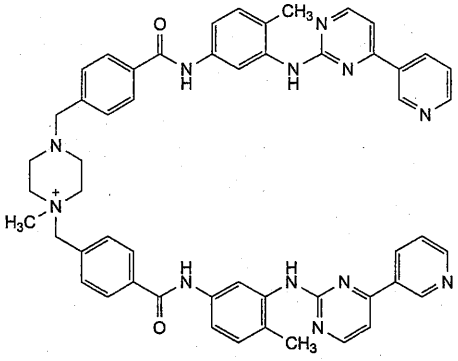
A. (2*E*)-3-(dimethylamino)-1-(pyridin-3-yl)prop-2-en-1-one,



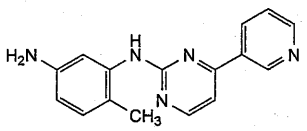
B. *N*-(3-carbamimidamido-4-methylphenyl)-4-[(4-methylpiperazin-1-yl)methyl]benzamide,



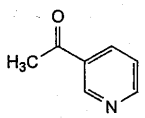
C. *N*-[4-methyl-3-[[4-(pyridin-3-yl)pyrimidin-2-yl]amino]phenyl]-4-(piperazin-1-ylmethyl)benzamide (desmethylimatinib),



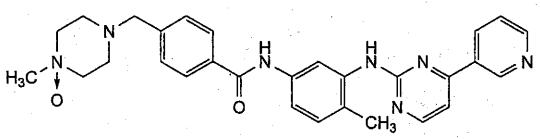
D. 1-methyl-1,4-bis[4-[[4-methyl-3-[[4-(pyridin-3-yl)pyrimidin-2-yl]amino]phenyl]carbamoyl]benzyl]piperazin-1-ium (imatinib dimer),



F. 4-methyl-N^3-[4-(pyridin-3-yl)pyrimidin-2-yl]benzene-1,3-diamine,



H. 1-(pyridin-3-yl)ethan-1-one,



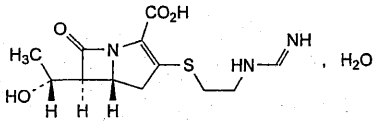
J. 4-[(4-methyl-4-oxidopiperazin-1-yl)methyl]-*N*-[4-methyl-3-[[4-(pyridin-3-yl)pyrimidin-2-yl]amino]phenyl]benzamide.

Ph Eur

Imipenem Monohydrate

Imipenem

(Ph. Eur. monograph 1226)



$C_{12}H_{17}N_3O_4S,H_2O$ 317.4 *74431-23-5*

Action and use
Carbapenem antibacterial.

Preparation
Cilastatin and Imipenem for Infusion

Ph Eur

DEFINITION

(5*R*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-3-[[2-[(iminomethyl) amino]ethyl]sulfanyl]-7-oxo-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid monohydrate.

Semi-synthetic product derived from a fermentation product or obtained by any other means.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white or pale yellow powder, slightly hygroscopic.

Solubility
Slightly soluble in water and in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *imipenem CRS.*

TESTS

Appearance of solution
The solution is not more opalescent than reference suspension II (*2.2.1*) and not more intensely coloured than intensity 6 of the range of the reference solutions of the most appropriate colour (*2.2.2, Method II*).

Dissolve 0.500 g in *phosphate buffer solution pH 7.0 R3* and dilute to 50 mL with the same solution.

pH (*2.2.3*)
4.5 to 7.5.

Dissolve 0.500 g in *carbon dioxide-free water R* and dilute to 100.0 mL with the same solvent.

Specific optical rotation (*2.2.7*)
+ 90 to + 95 (anhydrous substance), measured at 25 °C.
Prepare the solutions immediately before use.

Dissolve 0.125 g in *phosphate buffer solution pH 7.0 R3* and dilute to 25.0 mL with the same solution.

Related substances
Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Buffer solution A Dissolve 0.32 g of *anhydrous sodium dihydrogen phosphate R* and 1.04 g of *anhydrous disodium hydrogen phosphate R* in 900 mL of *water for chromatography R*. Adjust to pH 7.3 with *dilute phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Buffer solution B Dissolve 0.11 g of *anhydrous disodium hydrogen phosphate R* in 900 mL of *water R*. Adjust to pH 6.8 with *dilute phosphoric acid R* and dilute to 1000 mL with *water R*.

Solvent mixture *acetonitrile R*, buffer solution B (0.7:99.3 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of *imipenem CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of the substance to be examined in 8 mL of a mixture of 1 volume of *dilute sulfuric acid R* and 200 volumes of *water R*. After 5 min, add 10 mg of *sodium carbonate R* and dilute to 10.0 mL with *water R*.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (3 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: *acetonitrile R1*, buffer solution A (0.7:99.3 *V/V*);
— *mobile phase B*: *acetonitrile R1*, buffer solution A (25:75 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 9	100	0
9 - 24	100 → 68	0 → 32
24 - 24.5	68 → 50	32 → 50
24.5 - 29	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Autosampler Set at 5 °C.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to the epimers of impurity B.

Relative retention With reference to imipenem (retention time = about 8 min): epimer I of impurity B = about 0.33; epimer II of impurity B = about 0.35; impurity A = about 0.8.

System suitability Reference solution (c):
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to epimer I of impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to epimer II of impurity B.

Calculation of percentage contents:
— *correction factor*: multiply the peak area of impurity A by 2.4;
— for each impurity, use the concentration of imipenem in reference solution (b).

Limits:
— *impurity A*: maximum 1.0 per cent;
— *impurity B*: for each epimer, maximum 0.3 per cent;

— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 1.5 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
5.0 per cent to 8.0 per cent, determined on 0.100 g. Use an iodosulfurous reagent containing imidazole instead of pyridine and a clean container for each determination.

Sulfated ash (*2.4.14*)
Maximum 0.2 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.17 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

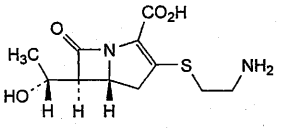
ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

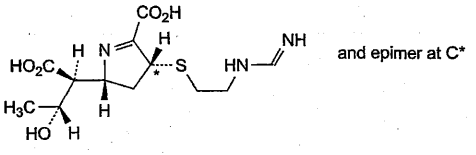
Calculate the percentage content of $C_{12}H_{17}N_3O_4S$ taking into account the assigned content of *imipenem CRS*.

STORAGE
In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES
Specified impurities *A, B.*



A. (5*R*,6*S*)-3-[(2-aminoethyl)sulfanyl]-6-[(*R*)-1-hydroxyethyl]-7-oxo-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid (thienamycin),

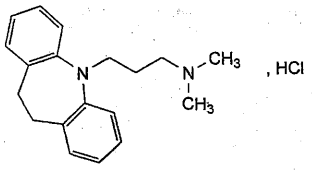


B. (2*R*,4*RS*)-2-[(1*S*,2*R*)-1-carboxy-2-hydroxypropyl]-4-[[2-[(iminomethyl)amino]ethyl]sulfanyl]-3,4-dihydro-2*H*-pyrrole-5-carboxylic acid (imipenemoic acid).

Ph Eur

Imipramine Hydrochloride

(Ph. Eur. monograph 0029)



$C_{19}H_{25}ClN_2$ 316.9 *113-52-0*

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparation
Imipramine Tablets

Ph Eur

DEFINITION
3-(10,11-Dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine hydrochloride.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or slightly yellow, crystalline powder.

Solubility
Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION
First identification: B, D.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 170 °C to 174 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *imipramine hydrochloride CRS.*

C. Dissolve about 5 mg in 2 mL of *nitric acid R*. An intense blue colour develops.

D. About 20 mg gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
To 3.0 g add 20 mL of *carbon dioxide-free water R*, dissolve rapidly by shaking and triturating with a glass rod and dilute to 30 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*). Immediately after preparation, dilute solution S with an equal volume of *water R*. This solution is not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

pH (*2.2.3*)
3.6 to 5.0 for solution S, measured immediately after preparation.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of *imipramine for system suitability CRS* (containing impurity B) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 40 volumes of *acetonitrile R1* with 60 volumes of a 5.2 g/L solution of *dipotassium hydrogen phosphate R* previously adjusted to pH 7.0 with *phosphoric acid R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 2.5 times the retention time of imipramine.

Relative retention With reference to imipramine (retention time = about 7 min): impurity B = about 0.7.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to impurity B and imipramine.

Limits:
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the peak due to imipramine in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 3 times the area of the peak due to imipramine in the chromatogram obtained with reference solution (b) (0.3 per cent);
— *disregard limit*: 0.5 times the area of the peak due to imipramine in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

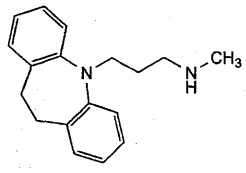
ASSAY
Dissolve 0.250 g in 50 mL of *ethanol (96 per cent) R* and add 5.0 mL of *0.01 M hydrochloric acid*. Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide*. Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 31.69 mg of $C_{19}H_{25}ClN_2$.

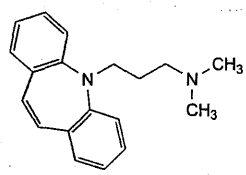
STORAGE
Protected from light.

IMPURITIES
Specified impurities *B.*

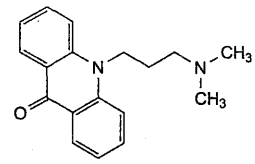
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, C.*



A. 3-(10,11-dihydro-5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*-methylpropan-1-amine (desipramine),



B. 3-(5*H*-dibenzo[*b*,*f*]azepin-5-yl)-*N*,*N*-dimethylpropan-1-amine (depramine),

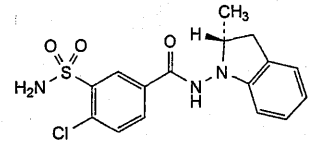


C. 10-[3-(dimethylamino)propyl]acridin-9(10*H*)-one.

Ph Eur

Indapamide

(Ph. Eur. monograph 1108)



and enantiomer

$C_{16}H_{16}ClN_3O_3S$ 365.8 *26807-65-8*

Action and use
Thiazide-like diuretic.

Preparations
Indapamide Tablets
Indapamide Prolonged-release Tablets

Ph Eur

DEFINITION
4-Chloro-*N*-[(2*RS*)-2-methyl-2,3-dihydro-1*H*-indol-1-yl]-3-sulfamoylbenzamide.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: B.

Second identification: A, C.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *ethanol (96 per cent) R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with *ethanol (96 per cent) R*.

Spectral range 220-350 nm.

Absorption maximum At 242 nm.

Shoulders At 279 nm and 287 nm.

Specific absorbance at the absorption maximum 590 to 630.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *indapamide CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *indapamide CRS* in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *indometacin R* in 5 mL of reference solution (a) and dilute to 10 mL with *ethanol (96 per cent) R*.

Plate *TLC silica gel GF_{254} plate R.*

Mobile phase *glacial acetic acid R, acetone R, toluene R* (1:20:79 *V/V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Optical rotation (*2.2.7*)
−0.02° to + 0.02°.

Dissolve 0.250 g in *anhydrous ethanol R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use or maintain them at 4 °C.*

Solvent mixture *acetonitrile R, methanol R* (50:50 *V/V*).

Test solution Dissolve 20.0 mg of the substance to be examined in 7 mL of the solvent mixture and dilute to 20.0 mL with a 0.2 g/L solution of *sodium edetate R*.

Reference solution (a) Dissolve 3.0 mg of *indapamide impurity B CRS* in 3.5 mL of the solvent mixture and dilute to 10.0 mL with a 0.2 g/L solution of *sodium edetate R*. To 1.0 mL of the solution add 35 mL of the solvent mixture and dilute to 100.0 mL with a 0.2 g/L solution of *sodium edetate R*.

Reference solution (b) To 1.0 mL of the test solution add 17.5 mL of the solvent mixture and dilute to 50.0 mL with a 0.2 g/L solution of *sodium edetate R*. To 1.0 mL of this solution add 7 mL of the solvent mixture and dilute to 20.0 mL with a 0.2 g/L solution of *sodium edetate R*.

Reference solution (c) Dissolve 20.0 mg of *indapamide CRS* in 7 mL of the solvent mixture and dilute to 20.0 mL with a 0.2 g/L solution of *sodium edetate R*.

Reference solution (d) Dissolve 25.0 mg of *indapamide CRS* and 45.0 mg of *methylnitrosoindoline CRS* (impurity A) in 17.5 mL of the solvent mixture and dilute to 50.0 mL with a 0.2 g/L solution of *sodium edetate R*.

Column:
— *size*: *l* = 0.20 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase *glacial acetic acid R, acetonitrile R, methanol R,* 0.2 g/L solution of *sodium edetate R* (0.1:17.5:17.5:65 *V/V/V/V*).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a), (b) and (d).

Run time 2.5 times the retention time of indapamide.

Retention time Indapamide = about 11 min.

System suitability:
— *resolution*: minimum 4.0 between the peaks due to indapamide and impurity A in the chromatogram obtained with reference solution (d).

Limits:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity A
Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Test solution Dissolve 25.0 mg of the substance to be examined in 1 mL of *acetonitrile R* and dilute to 10.0 mL with *water R*. Shake for 15 min. Allow to stand at 4 °C for 1 h and filter.

Reference solution Dissolve 25.0 mg of the substance to be examined in 1.0 mL of a 0.125 mg/L solution of *methylnitrosoindoline CRS* (impurity A) in *acetonitrile R* and dilute to 10.0 mL with *water R*. Shake for 15 min. Allow to stand at 4 °C for 1 h and filter.

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 30 °C.

Mobile phase Mix 7 volumes of *acetonitrile R*, 20 volumes of *tetrahydrofuran R* and 73 volumes of a 1.5 g/L solution of *triethylamine R* adjusted to pH 2.8 with *phosphoric acid R*.

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 305 nm.

Injection 0.1 mL.

System suitability Reference solution:
— *signal-to-noise ratio*: minimum 3 for the peak due to impurity A appearing just before the peak due to indapamide;
— *peak-to-valley-ratio*: minimum 6.7, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to indapamide.

Limit:
— *impurity A*: not more than the difference between the areas of the peaks due to impurity A in the chromatograms obtained with the reference solution and the test solution (5 ppm).

Impurity C
Liquid chromatography (*2.2.29*). *Maintain the solutions at 10 °C after preparation.*

Solution A Dissolve 0.20 g of *sodium edetate R* in *water R*, add 1.5 mL of *anhydrous acetic acid R* and dilute to 1 L with *water R*.

Test solution Dissolve 75.0 mg of the substance to be examined in 7.5 mL of *acetonitrile for chromatography R* and dilute to 25.0 mL with *water R*.

Reference solution (a) Dissolve 9.0 mg of *indapamide impurity C CRS* in 1.0 mL of *water R*, add 6.0 mL of *acetonitrile for chromatography R* and dilute to 20.0 mL with *water R*. To 1.0 mL of the solution add 7.5 mL of *acetonitrile for chromatography R* and dilute to 25.0 mL with *water R*.

Reference solution (b) To 1.0 mL of reference solution (a) add 3.0 mL of *acetonitrile for chromatography R* and dilute to 10.0 mL with *water R*.

Reference solution (c) To 1.0 mL of reference solution (a) add 3.0 mL of *acetonitrile for chromatography R* and dilute to 10.0 mL with the test solution.

Column:
— *size*: *l* = 0.05 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (1.8 µm);
— *temperature*: 50 °C.

Mobile phase *acetonitrile for chromatography R*, solution A (30:70 *V/V*).

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 2 µL of the test solution and reference solutions (b) and (c).

Run time 3 times the retention time of indapamide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to indapamide (retention time = about 1.3 min): impurity C = about 0.5.

System suitability Reference solution (c):
— *resolution*: minimum 4.0 between the peaks due to impurity C and indapamide;
— *signal-to-noise ratio*: minimum 20 for the peak due to impurity C.

Calculation of content:
— for impurity C, use the concentration of impurity C in reference solution (b).

Limit:
— *impurity C*: maximum 600 ppm.

Water (*2.5.12*)
Maximum 3.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

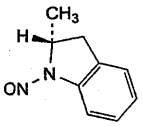
Calculate the percentage content of $C_{16}H_{16}ClN_3O_3S$ taking into account the assigned content of *indapamide CRS*.

STORAGE

Protected from light.

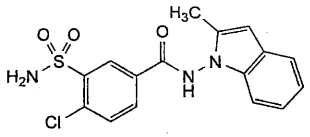
IMPURITIES

Specified impurities *A, B, C.*

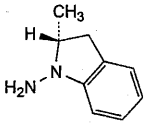


and enantiomer

A. (2*RS*)-2-methyl-1-nitroso-2,3-dihydro-1*H*-indole,



B. 4-chloro-*N*-(2-methyl-1*H*-indol-1-yl)-3-sulfamoylbenzamide,



and enantiomer

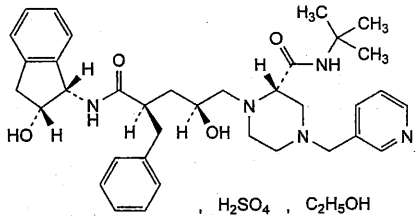
C. (2*RS*)-2-methyl-2,3-dihydro-1*H*-indol-1-amine.

Ph Eur

Indinavir Sulfate

Indinavir Sulphate

(Ph. Eur. monograph 2214)



$C_{36}H_{49}N_5O_8S,C_2H_6O$ 758 *157810-81-6*

Action and use
Protease inhibitor; antiviral (HIV).

Ph Eur

DEFINITION

(2*S*)-1-[(2*S*,4*R*)-4-Benzyl-2-hydroxy-5-[[(1*S*,2*R*)-2-hydroxy-2,3-dihydro-1*H*-inden-1-yl]amino]-5-oxopentyl]-*N*-(1,1-dimethylethyl)-4-(pyridin-3-ylmethyl)piperazine-2-carboxamide sulfate ethanolate.

Content
98.0 per cent to 102.0 per cent (anhydrous and ethanol-free substance).

PRODUCTION

A test for enantiomeric purity is carried out unless it has been demonstrated that the manufacturing process is enantioselective for the substance.

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Freely soluble in water, soluble in methanol, practically insoluble in heptane.

IDENTIFICATION

A. Specific optical rotation (*2.2.7*): + 122 to + 129 (anhydrous and ethanol-free substance), determined at 365 nm and at 25 °C.

Dissolve 0.500 g in *water R* and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *Ph. Eur. reference spectrum of indinavir sulfate.*

C. It gives reaction (a) of sulfates (*2.3.1*).

D. Ethanol (see Tests).

TESTS

Related substances

Liquid chromatography (*2.2.29*).

Solution A Thoroughly mix equal volumes of mobile phase A and *acetonitrile R1*.

Test solution Dissolve 50.0 mg of the substance to be examined in solution A and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 4 mg of *indinavir for system suitability CRS* (containing impurities B, C and E) in solution A and dilute to 10 mL with the same solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (c) Dissolve 5.0 mg of *cis-aminoindanol R* (impurity A) in solution A and dilute to 10.0 mL with the same solution. Dilute 1.0 mL of the solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (d) To 30 mg of the substance to be examined add 0.25 mL of *2 M hydrochloric acid R* and allow to stand at room temperature for 1 h. Dilute to 100 mL with a mixture of 2 volumes of *acetonitrile R1* and 3 volumes of mobile phase A and mix (*in situ* degradation to obtain impurity D).

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: solution containing 0.27 g/L of *potassium dihydrogen phosphate R* and 1.40 g/L of *dipotassium hydrogen phosphate R*; filter and degas;
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	80	20
5 - 40	80 → 30	20 → 70
40 - 45	30	70
45 - 47	30 → 80	70 → 20
47 - 52	80	20

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *indinavir for system suitability CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C and E; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity D.

Relative retention With reference to indinavir (retention time = about 25 min): impurity A = about 0.2; impurity B = about 0.8; impurity C = about 0.98; impurity D = about 1.1; impurity E = about 1.3.

System suitability Reference solution (a):
- *resolution*: minimum 1.8 between the peaks due to impurity C and indinavir.

Limits:
- *correction factor*: for the calculation of content, multiply the peak area of impurity D by 1.8;
- *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
- *impurity D*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *impurities B, C, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
- *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

Ethanol

Gas chromatography (*2.2.28*).

Internal standard solution Dilute 1.0 mL of *propanol R* to 200.0 mL with *water R*.

Test solution Dissolve 0.400 g of the substance to be examined in 50.0 mL of *water R*, add 8.0 mL of the internal standard solution and dilute to 100.0 mL with *water R*.

Reference solution Dilute 1.0 mL of *anhydrous ethanol R* to 200.0 mL. Dilute 2.0 mL of this solution and 2.0 mL of the internal standard solution to 25.0 mL with *water R*.

Column:
- *material*: fused silica;
- *size*: *l* = 30 m, Ø = 0.53 mm;
- *stationary phase*: *macrogol 20 000 R* (film thickness 1.0 µm).

Carrier gas *helium for chromatography R*.

Flow rate 10 mL/min.

Split ratio 1:10.

Temperature:
- *column*: 35 °C;
- *injection port*: 140 °C;
- *detector*: 220 °C.

Detection Flame ionisation.

Injection 1.0 µL.

System suitability Reference solution:
- *retention time*: ethanol = 2 min to 4 min;
- *resolution*: minimum 5.0 between the peaks due to ethanol and propanol.

Calculate the percentage content of ethanol taking the density (*2.2.5*) to be 0.790 g/mL.

Limit:
- *ethanol*: 5.0 per cent to 8.0 per cent *m/m*.

Water (*2.5.12*)

Maximum 1.5 per cent, determined on 0.500 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*).

Solution B Add 20 mL of *dibutylammonium phosphate for ion-pairing R* to 1000 mL of *water R*. Adjust to pH 6.5 with *1 M sodium hydroxide*.

Test solution Dissolve 60.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution Dissolve 50.0 mg of *indinavir CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *base-deactivated octylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase *acetonitrile R*, solution B (45:55 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 µL.

Run time Twice the retention time of indinavir.

Retention time Indinavir = about 10 min.

Calculate the percentage content of $C_{36}H_{49}N_5O_8S$ using the declared content of *indinavir CRS* and multiplying by a correction factor of 1.1598.

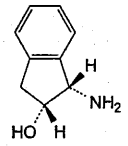
STORAGE

In an airtight container, protected from light.

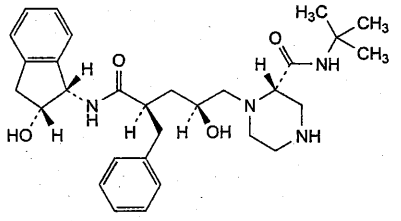
IMPURITIES

Specified impurities *A, B, C, D, E.*

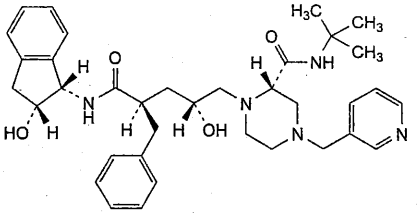
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F.*



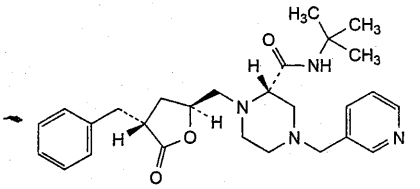
A. (1*S*,2*R*)-1-amino-2,3-dihydro-1*H*-inden-2-ol (*cis*-aminoindanol),



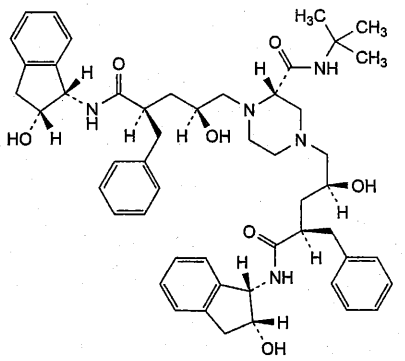
B. (2*S*)-1-[(2*S*,4*R*)-4-benzyl-2-hydroxy-5-[[(1*S*,2*R*)-2-hydroxy-2,3-dihydro-1*H*-inden-1-yl]amino]-5-oxopentyl]-*N*-(1,1-dimethylethyl)piperazine-2-carboxamide,



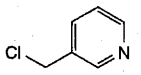
C. (2*S*)-1-[(2*R*,4*R*)-4-benzyl-2-hydroxy-5-[[(1*S*,2*R*)-2-hydroxy-2,3-dihydro-1*H*-inden-1-yl]amino]-5-oxopentyl]-*N*-(1,1-dimethylethyl)-4-(pyridin-3-ylmethyl)piperazine-2-carboxamide,



D. (3*R*,5*S*)-3-benzyl-5-[[(2*S*)-2-[(1,1-dimethylethyl)carbamoyl]-4-(pyridin-3-ylmethyl)piperazin-1-yl]methyl]-4,5-dihydrofuran-2(3*H*)-one,



E. (2*S*)-1,4-bis[(2*S*,4*R*)-4-benzyl-2-hydroxy-5-[[(1*S*,2*R*)-2-hydroxy-2,3-dihydro-1*H*-inden-1-yl]amino]-5-oxopentyl]-*N*-(1,1-dimethylethyl)piperazine-2-carboxamide,

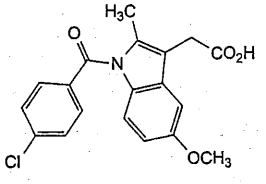


F. 3-(chloromethyl)pyridine (nicotinyl chloride).

Ph Eur

Indometacin

(Ph. Eur. monograph 0092)



$C_{19}H_{16}ClNO_4$ 357.8 *53-86-1*

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations

Indometacin Capsules

Indometacin Suppositories

Ph Eur

DEFINITION

[1-(4-Chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl] acetic acid.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellow, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (*5.9*). The acceptable crystalline form corresponds to *indometacin CRS*.

IDENTIFICATION

First identification: A, C.

Second identification: A, B, D, E.

A. Melting point (*2.2.14*): 158 °C to 162 °C.

B. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 25 mg in a mixture of 1 volume of a 103.0 g/L solution of *hydrochloric acid R* and 9 volumes of *methanol R* and dilute to 100.0 mL with the same mixture of solvents. Dilute 10.0 mL of the solution to 100.0 mL with a mixture of 1 volume of a 103.0 g/L solution of *hydrochloric acid R* and 9 volumes of *methanol R*.

Spectral range 300-350 nm.

Absorption maximum At 318 nm.

Specific absorbance at the absorption maximum 170 to 190.

C. Infrared absorption spectrophotometry (*2.2.24*), without recrystallisation.

Comparison *indometacin CRS*.

D. Dissolve 0.1 g in 10 mL of *ethanol (96 per cent) R*, heating slightly if necessary. To 0.1 mL of the solution add 2 mL of a freshly prepared mixture of 1 volume of a 250 g/L solution of *hydroxylamine hydrochloride R* and 3 volumes of *dilute sodium hydroxide solution R*. Add 2 mL of *dilute hydrochloric acid R* and 1 mL of *ferric chloride solution R2* and mix. A violet-pink colour develops.

E. To 0.5 mL of the solution in ethanol (96 per cent) prepared in identification test D, add 0.5 mL of *dimethylaminobenzaldehyde solution R2*. A precipitate is formed that dissolves on shaking. Heat on a water-bath. A bluish-green colour is produced. Continue to heat for 5 min and cool in iced water for 2 min. A precipitate is formed and the colour changes to light greyish-green. Add 3 mL of *ethanol (96 per cent) R*. The solution is clear and violet-pink in colour.

TESTS

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Solvent mixture *acetonitrile R, water R* (50:50 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture, using sonication if necessary, and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *indometacin impurity mixture CRS* (impurities I and J) in 1 mL of the solvent mixture.

Column:
- *size*: *l* = 0.10 m, Ø = 2.1 mm;
- *stationary phase*: *end-capped ethylene-bridged phenylsilyl silica gel for chromatography (hybrid material) R* (1.7 µm);
- *temperature*: 50 °C.

Mobile phase:
- *mobile phase A*: 5 g/L solution of *anhydrous formic acid R*;
- *mobile phase B*: 5 g/L solution of *anhydrous formic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5.5	60	40
5.5 - 5.6	60 → 30	40 → 70
5.6 - 9	30	70

Flow rate 0.3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 1.4 µL.

Identification of impurities Use the chromatogram supplied with *indometacin impurity mixture CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities I and J.

Relative retention With reference to indometacin (retention time = about 6 min): impurity I = about 1.3; impurity J = about 1.4.

System suitability Reference solution (b):
- *resolution*: minimum 3.0 between the peaks due to impurities I and J.

Calculation of percentage contents:
- for each impurity, use the concentration of indometacin in reference solution (a).

Limits:
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.3 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*).

Solvent mixture *acetonitrile R, water R* (50:50 *V/V*).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution Dissolve 25.0 mg of *indometacin CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped solid core octadecylsilyl silica gel for chromatography R* (2.6 µm);
- *temperature*: 50 °C.

Mobile phase:
— *mobile phase A*: 10 g/L solution of *acetic acid R*;
— *mobile phase B*: 10 g/L solution of *acetic acid R* in *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	50	50
5 - 5.5	50 → 0	50 → 100
5.5 - 8	0	100

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Retention time Indometacin = about 4 min.

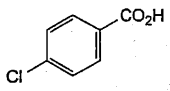
Calculate the percentage content of $C_{19}H_{16}ClNO_4$ taking into account the assigned content of *indometacin CRS*.

STORAGE

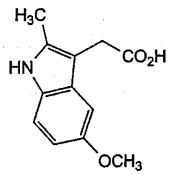
Protected from light.

IMPURITIES

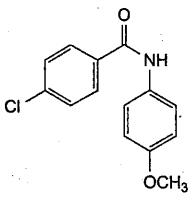
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, C, D, E, F, G, H, I, J.*



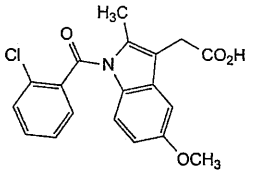
A. 4-chlorobenzoic acid,



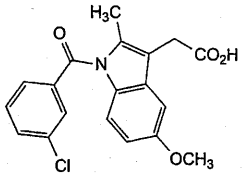
B. (5-methoxy-2-methyl-1*H*-indol-3-yl)acetic acid,



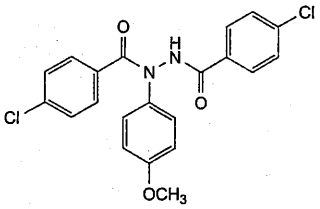
C. 4-chloro-*N*-(4-methoxyphenyl)benzamide,



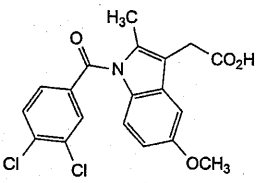
D. [1-(2-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl] acetic acid,



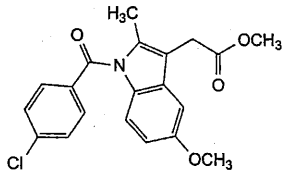
E. [1-(3-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl] acetic acid,



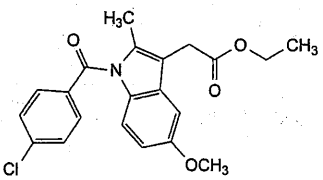
F. 4-chloro-*N'*-(4-chlorobenzoyl)-*N*-(4-methoxyphenyl)benzohydrazide,



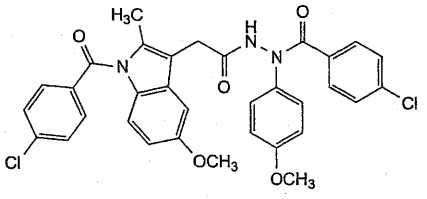
G. [1-(3,4-dichlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetic acid,



H. methyl [1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetate,



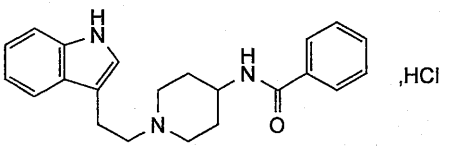
I. ethyl [1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetate,



J. 4-chloro-*N'*-[[1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]acetyl]-*N*-(4-methoxyphenyl)benzohydrazide.

Ph Eur

Indoramin Hydrochloride



$C_{22}H_{25}N_3O,HCl$ 383.9 *33124-53-7*

Action and use
Alpha$_1$-adrenoceptor antagonist.

Preparation
Indoramin Tablets

DEFINITION
Indoramin Hydrochloride is *N*-1-[2-(indol-3-yl)ethyl]-4-piperidylbenzamide hydrochloride. It contains not less than 98.5% and not more than 100.5% of $C_{22}H_{25}N_3O,HCl$, calculated with reference to the dried substance.

CHARACTERISTICS
A white or almost white powder. It exhibits polymorphism.

Slightly soluble in *water*; sparingly soluble in *ethanol (96%)*; soluble in *methanol*; very slightly soluble in *ether*.

IDENTIFICATION
A. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.0045% w/v solution in *ethanol (96%)* exhibits three maxima, at 273, 280 and 290 nm. The *absorbances* at the maxima are about 0.76, 0.77 and 0.64, respectively.

B. Dissolve 50 mg in 30 mL of *water*, make the solution alkaline by the addition of 5M *ammonia* and shake with 50 mL of *dichloromethane*. Dry the dichloromethane layer with *anhydrous sodium sulfate*, filter and evaporate the filtrate to dryness using a rotary evaporator. The *infrared absorption spectrum* of the residue, Appendix II A, is concordant with the *reference spectrum* of indoramin *(RS 188)*.

C. Yields reaction A characteristic of *chlorides*, Appendix VI.

TESTS
Acidity
pH of a 2% w/v suspension in *water*, 4.0 to 5.5, Appendix V L.

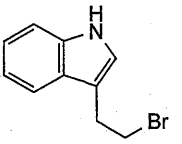
Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using a *silica gel* F_{254} precoated plate (Merck silica gel 60 F_{254} plates are suitable)and a mixture of 1 volume of 18M *ammonia*, 20 volumes of *absolute ethanol* and 79 volumes of *toluene* as the mobile phase. Apply separately to the plate 10 µL of each of three solutions of the substance being examined in *ethanol (96%)* containing (1) 1.0% w/v, (2) 0.0050% w/v and (3) 0.0010% w/v. After removal of the plate, allow it to dry in a current of warm air and examine under *ultraviolet light (254 nm)*. Any *secondary spot* in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.5%) and not more than one such spot is more intense than the spot in the chromatogram obtained with solution (3) (0.1%).

Loss on drying
When dried at 100° to 105° for 4 hours, loses not more than 0.5% of its weight. Use 1 g.

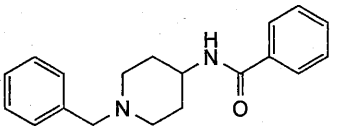
Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Dissolve 0.2 g in 30 mL of *anhydrous acetic acid*, add 6 mL of *acetic anhydride* and 6 mL of *mercury(II) acetate* solution. Titrate with *0.1M perchloric acid VS* determining the end point potentiometrically. Each mL of *0.1M perchloric acid VS* is equivalent to 38.39 mg of $C_{22}H_{25}N_3O,HCl$.

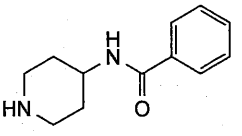
STORAGE
Indoramin Hydrochloride should be protected from light.

IMPURITIES


A. 3-(2-bromoethyl)indole



B. *N*-(1-benzyl-4-piperidyl)benzamide



C. *N*-(4-piperidyl)benzamide

Infliximab Concentrated Solution

(Ph. Eur. monograph 2928)

heavy chain

```
EVKLEESGGG  LVQPGGSMKL  SCVASGFIFS  NHWMNWVRQS   40
PEKGLEWVAE  IRSKSINSAT  HYAESVKGRF  TISRDDSKSA   80
VYLQMTDLRT  EDTGVYYCSR  NYYGSTYDYW  GQGTTLTVSS  120
ASTKGPSVFP  LAPSSKSTSG  GTAALGCLVK  DYFPEPVTVS  160
WNSGALTSGV  HTFPAVLQSS  GLYSLSSVVT  VPSSSLGTQT  200
YICNVNHKPS  NTKVDKKVEP  KSCDKTHTCP  PCPAPELLGG  240
PSVFLFPPKP  KDTLMISRTP  EVTCVVVDVS  HEDPEVKFNW  280
YVDGVEVHNA  KTKPREEQYN  STYRVVSVLT  VLHQDWLNGK  320
EYKCKVSNKA  LPAPIEKTIS  KAKGQPREPQ  VYTLPPSRDE  360
LTKNQVSLTC  LVKGFYPSDI  AVEWESNGQP  ENNYKTTPPV  400
LDSDGSFFLY  SKLTVDKSRW  QQGNVFSCSV  MHEALHNHYT  440
QKSLSLSPGK                                    450
```

light chain

```
DILLTQSPAI  LSVSPGERVS  FSCRASQFVG  SSIHWYQQRT   40
NGSPRLLIKY  ASESMSGIPS  RFSGSGSGTD  FTLSINTVES   80
EDIADYYCQQ  SHSWPFTFGS  GTNLEVKRTV  AAPSVFIFPP  120
SDEQLKSGTA  SVVCLLNNFY  PREAKVQWKV  DNALQSGNSQ  160
ESVTEQDSKD  STYSLSSTLT  LSKADYEKHK  VYACEVTHQG  200
LSSPVTKSFN  RGEC                              214
```

disulfide bridges:

Intra-H	22-98	147-203	264-324	370-428
	22"-98"	147"-203"	264"-324"	370"-428"
Intra-L	23'-88'	134'-194'		
	23'"-88'"	134'"-194'"		
Inter-H-L	223-214'			
	223"-214'"			
Inter-H-H	229-229"	232-232"		

N-glycosylation sites:
300, 300"

$C_{6462}H_{9960}N_{1728}O_{2036}S_{44}$ (dimer without glycosylation)

M_r approx. 145 kDa
(dimer without glycosylation)

Action and use

Monoclonal antibody (TNF alfa).

Ph Eur

DEFINITION

Solution of a monoclonal antibody consisting of a bisdisulfide dimer of 1328 amino acid residues with a molecular weight of approximately 145 kDa, which binds with high affinity to both soluble and transmembrane forms of TNF-α.

Infliximab is a chimeric human-murine IgG1 kappa monoclonal antibody representing a glycosylated immunoglobulin with 1 *N*-linked glycosylation site (Asn 300) in the CH2 domain of each heavy chain. The detected oligosaccharides are mostly G0F (absence of terminal galactose) and G1F (1 terminal galactose) structures. Each heavy chain consists of 450 amino acids with 11 cysteine residues, and each light chain consists of 214 amino acids with 5 cysteine residues. All cysteine residues in heavy and light chains are involved in either intra- or inter-disulfide bonding.

Content (milligrams of protein per millilitre) As approved by the competent authority.

Potency

8×10^3 to 12×10^3 IU per milligram of protein.

PRODUCTION

Infliximab is produced in a suitable mammalian cell expression system by a method based on recombinant DNA (rDNA) technology. During the course of product development, it must be demonstrated that the manufacturing process consistently produces a product with the expected *N*-glycan occupancy and Fc-effector functions (antibody-dependent cellular cytotoxicity (ADCC), complement-dependent cytotoxicity (CDC)) using suitably qualified assay(s).

Prior to release, the following tests are carried out on each batch of infliximab concentrated solution, unless an exemption has been granted by the competent authority.

Host-cell-derived proteins *(2.6.34)*
The limit is approved by the competent authority.

Host-cell- and vector-derived DNA
The limit is approved by the competent authority.

Residual Protein A
Use a suitable immunochemical method *(2.7.1)* based on an enzyme-linked immunosorbent assay (ELISA). To determine residual Protein A, the preparation to be examined is transferred to an ELISA microplate coated with an anti-protein A capture antibody. Peroxidase-conjugated polyclonal anti-protein A antibody is added followed by a peroxidase substrate. Optical density is measured and the amount of residual Protein A is calculated using a standard curve and the usual statistical methods (for example, *5.3*).

Limit As approved by the competent authority.

Glycan analysis
Use a suitable method developed according to general chapter *2.2.59. Glycan analysis of glycoproteins*, section 2-3:
— after desalting, release the glycans using 1 of the agents described in Table 2.2.59.-1, for example peptide *N*-glycosidase F (PNGase F);
— if needed, label the released glycans with 1 of the fluorescent labelling agents described in Table 2.2.59.-2;
— analyse the labelled or unlabelled glycans using a suitable technique.

The following procedure is given as an example.

Solution A Dissolve 0.17 g of *anhydrous sodium dihydrogen phosphate R* and 0.53 g of *anhydrous disodium hydrogen phosphate R* in *water R*. Adjust to pH 7.6 with *sodium hydroxide solution R* or *hydrochloric acid R* and dilute to 1000.0 mL with *water R*.

Test solution Desalt a volume of the preparation to be examined by a suitable method (for example, using a suitable centrifugal filter with *water for chromatography R* as elution buffer), and dilute with solution A to obtain a concentration of about 1 mg/mL. To 200 µL of this solution add 0.3 µL of a 500 000 U/mL solution of *peptide N-glycosidase F R*. Incubate at 37 °C for at least 16 hours.

Reference solution (a) Dissolve the contents of a vial of *infliximab CRS* in *water R*. Desalt a volume of this preparation and carry out the glycan release at the same time and in the same manner as for the test solution.

Reference solution (b) Use a suitable infliximab in-house reference preparation shown to be representative of batches tested clinically and batches used to demonstrate consistency of production. Desalt a volume of this preparation and carry out the glycan release at the same time and in the same manner as for the test solution.

Blank solution Use 200 µL of solution A to proceed to glycan release.

Analyse the native glycans by liquid chromatography *(2.2.29)*.

Precolumn:
— *size*: *l* = 0.05 m, Ø = 3.0 mm;

— *stationary phase*: *strongly basic anion-exchange resin for chromatography R2* (5.5 µm).

Column:
— *size*: *l* = 0.25 m, Ø = 3.0 mm;
— *stationary phase*: *strongly basic anion-exchange resin for chromatography R2* (5.5 µm);
— *temperature*: 30 °C.

Mobile phase:
— *mobile phase A*: 2 g/L solution of *sodium hydroxide R*;
— *mobile phase B*: solution containing 2 g/L of *sodium hydroxide R* and 10.25 g/L of *anhydrous sodium acetate R*;
— *mobile phase C*: solution containing 2 g/L of *sodium hydroxide R* and 41 g/L of *anhydrous sodium acetate R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Mobile phase C (per cent *V/V*)	Flow rate (mL/min)
0 - 5	99.2	0.8	0	0.35
5 - 45	99.2 → 95.2	0.8 → 4.8	0	0.35
45 - 50	95.2 → 72	4.8 → 28	0	0.35
50 - 77	72 → 4	28 → 96	0	0.35
77 - 77.1	4 → 0	96 → 0	0 → 100	0.35
77.1 - 87	0	0	100	0.40

Detection Electrochemical detector (pulsed amperometry).

Autosampler Set at 2-8 °C.

Injection 25 µL.

Identification of peaks Use the chromatogram in Figure 2928.-1 to identify the 7 peaks corresponding to fucosylated (peaks 1, 4 and 5), afucosylated (peaks 2 and 3) and sialylated (peaks 6 and 7) glycans; record the retention time of each peak.

System suitability:
— the chromatogram obtained with reference solution (a) is qualitatively similar to the chromatogram supplied with *infliximab CRS* and peaks 1 to 7 are clearly visible;
— no significant peaks are observed in the chromatogram obtained with the blank solution.

Results:
— the profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with reference solution (b);
— the retention times of the peaks in the chromatogram obtained with the test solution correspond to those in the chromatogram obtained with reference solution (b);
— no additional peaks are observed in the chromatogram obtained with the test solution in comparison with the chromatogram obtained with reference solution (b).

Calculate the relative peak areas of the individual peaks corresponding to fucosylated, afucosylated and sialylated glycans with reference to the sum of the areas of all retained glycan peaks.

Calculate the percentage contents of fucosylated, afucosylated and sialylated glycans, using the following expressions:

$$\frac{A}{A+B+C} \times 100$$

$$\frac{B}{A+B+C} \times 100$$

$$\frac{C}{A+B+C} \times 100$$

A = sum of the areas of the peaks due to fucosylated glycans;
B = sum of the areas of the peaks due to afucosylated glycans;
C = sum of the areas of the peaks due to sialylated glycans.

NOTE: sialylated glycans elute as peak clusters and are integrated as such.

Limits:
— *percentage of fucosylated glycans*: as authorised by the competent authority;
— *percentage of afucosylated glycans*: as authorised by the competent authority;
— *percentage of sialylated glycans*: as authorised by the competent authority.

Charged variants

A. Isoelectric focusing (*2.2.54*): use suitable agarose gels.

The following procedure is given as an example.

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 0.5 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *infliximab CRS* in *water R* to obtain a concentration of 0.5 mg/mL.

Reference solution (b) Use a suitable infliximab in-house reference preparation shown to be representative of batches tested clinically and batches used to demonstrate consistency of production. Dilute with *water R* to obtain a concentration of 0.5 mg/mL.

Reference solution (c) Use an isoelectric point (pI) calibration solution, in the pI range of 3-10, prepared according to the manufacturer's instructions. Adjust the concentration to allow detection of all marker proteins.

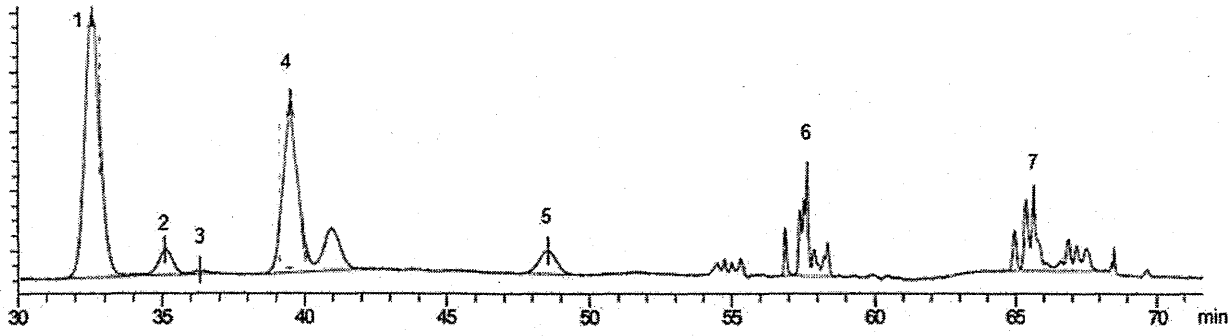
Focusing:
— *pH gradient*: 3-10;
— *catholyte*: 40 g/L solution of *sodium hydroxide R*;
— *anolyte*: 2.87 per cent *V/V* solution of *glacial acetic acid R*;
— *application*: 15 µL.

For each gel, run 2 lanes with reference solution (a), 2 lanes with reference solution (b) and 2 lanes with reference solution (c); use the remaining lanes for the test solution. Proceed with the isoelectric focusing by applying the electrical parameters according to the manufacturer's instructions (the following parameters have been found suitable: 1500 V, 7 mA, 25 W and 80 min).

Detection As described in general chapter *2.2.54*, with the following modifications.

Immerse the gel in a solution containing 36 g/L of *sulfosalicylic acid R*, 60 g/L of *trichloroacetic acid R* and 285 g/L of *methanol R*. Incubate with gentle shaking at room temperature for 30 min. Drain off the solution and transfer the gel to a mixture of 80 volumes of *glacial acetic acid R*, 250 volumes of *ethanol R* and 670 volumes of *water R* (mixture A); while shaking, rinse for 10 min at room temperature. Immerse the gel in a staining solution (1 g/L solution of *acid blue 83 R* in mixture A) and incubate for 20 min at room temperature. Destain the gel by passive diffusion (for at least 6 h) with mixture A until the bands are well visualised against a clear background. Soak and wash the gel for 1 h with *water R*.

System suitability:
— in the electropherogram obtained with reference solution (a), 7 bands (4 major and 3 minor) in the pI region 7.35-8.30 are clearly visible;
— in the electropherogram obtained with reference solution (c), the relevant pI markers are distributed along the entire length of the gel.•



Peak	Charged	Glycoform	Peak	Charged	Glycoform
1.	No	Fucosylated G0F (absence of terminal galactose)	5.	No	Fucosylated G2F (2 terminal galactoses)
2.	No	Afucosylated Man5 (addition of 2 mannoses instead of terminal *N*-acetylglucosamine)	6.	Yes	Sialylated SA1 (addition of 1 sialic acid)
3.	No	Afucosylated G0 (absence of fucose)	7.	Yes	Sialylated SA2 (addition of 2 sialic acids)
4.	No	Fucosylated G1F (1 terminal galactose)			

Figure 2928.-1. – *Chromatogram for glycan analysis of infliximab*

Results:
- — the electropherogram obtained with the test solution is similar to the electropherogram obtained with reference solution (b). Plot the migration distances of the relevant pI markers versus their pI and determine the isoelectric points of the principal components of the test solution and reference solution (b); they do not differ by more than 0.05 pI units;
- — no additional bands are observed in the electropherogram obtained with the test solution in comparison to the electropherogram obtained with reference solution (b).

Alternatively, use a suitable capillary isoelectric focusing method developed according to general chapter *2.2.47. Capillary electrophoresis*.

B. Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dilute the preparation to be examined with mobile phase A to obtain a concentration of about 1 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *infliximab CRS* in mobile phase A to obtain a concentration of about 1 mg/mL.

Reference solution (b) Use a suitable infliximab in-house reference preparation shown to be representative of batches tested clinically and batches used to demonstrate consistency of production. Dilute with mobile phase A to obtain a concentration of about 1 mg/mL.

Precolumn:
- — *size*: *l* = 0.05 m, Ø = 4 mm;
- — *stationary phase*: *weak cation-exchange resin R* (10 µm).

Column:
- — *size*: *l* = 0.25 m, Ø = 4 mm;
- — *stationary phase*: *weak cation-exchange resin R* (10 µm).

Mobile phase:
- — *mobile phase A*: dissolve 0.56 g of *sodium dihydrogen phosphate R* and 1.14 g of *disodium hydrogen phosphate dihydrate R* in 800 mL of *water for chromatography R*, and adjust to pH 7.25, if necessary, with *sodium hydroxide solution R* or *hydrochloric acid R*; dilute to 1000.0 mL with *water for chromatography R* and degas;
- — *mobile phase B*: dissolve 0.56 g of *sodium dihydrogen phosphate R*, 1.14 g of *disodium hydrogen phosphate dihydrate R* and 58.44 g of *sodium chloride R* in 800 mL of *water for chromatography R*, and adjust to pH 7.25, if necessary, with *sodium hydroxide solution R* or *hydrochloric acid R*; dilute to 1000.0 mL with *water for chromatography R* and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 15	100 → 92	0 → 8
15 - 16	92 → 60	8 → 40
16 - 21	60	40

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 214 nm.

Autosampler Set at 10 °C.

Injection 50 µL.

Relative retention With reference to the peak due to isoform 6 (retention time = about 9.2 min): isoform 1 = 0.68; isoform 2 = 0.74; isoform 3 = 0.80; isoform 4 = 0.87; isoform 5 = 0.95.

System suitability Reference solution (a):
- — the chromatogram obtained is similar to the chromatogram supplied with *infliximab CRS*;
- — *resolution*: minimum 1.5 between the peaks due to isoforms 3 and 4.

Results:
- — the profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with reference solution (b);
- — the relative retentions of the peaks due to isoforms 2, 3 and 4 in the chromatogram obtained with the test solution are within 1 per cent of those of the corresponding peaks in the chromatogram obtained with reference solution (b).

Calculate the relative peak areas of the individual peaks due to isoforms with reference to the total area of all peaks eluting between 3 min and 11 min.

Limits:
- — *sum of isoforms 1 and 2*: as authorised by the competent authority;
- — *sum of isoforms 3, 4 and 6*: as authorised by the competent authority;
- — *isoform 5*: as authorised by the competent authority.

CHARACTERS

Appearance

Opalescent or slightly opalescent, colourless or light yellow liquid.

IDENTIFICATION

A. It complies with the limits of the assay (potency).

B. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Dilution buffer Dissolve 50.00 g of *sucrose R*, 0.22 g of *sodium dihydrogen phosphate monohydrate R*, 0.61 g of *disodium hydrogen phosphate dihydrate R* and 0.05 g of *polysorbate 80 R* in *water R*. Adjust to pH 7.2, if necessary, and dilute to 500.0 mL with *water R*.

Test solution Dilute the preparation to be examined with the dilution buffer to obtain a concentration of about 5 mg/mL.

Reference solution Dissolve the contents of a vial of *infliximab CRS* in the dilution buffer to obtain a concentration of about 5 mg/mL.

Reduction and alkylation Dilute the test solution with *guanidine-tris(hydroxymethyl)aminomethane-EDTA buffer solution pH 8.6 R* to obtain a concentration of 2 mg/mL. To 1 mL of this solution add 10 µL of a 154 g/L solution of *dithiothreitol R* and incubate at 37 °C for 1 h. Add 20 µL of a freshly prepared 185 g/L solution of *iodoacetamide R* and incubate at room temperature for 15 min, protected from light. Add 10 µL of a 154 g/L of solution of *dithiothreitol R* and mix well.

Digestion Desalt a volume of the reduced solution prepared previously by a suitable method (for example, using a suitable centrifugal filter unit with *tris(hydroxymethyl) aminomethane buffer solution pH 7.5 R1* as elution buffer), and adjust the concentration to 1 mg/mL with *tris(hydroxymethyl) aminomethane buffer solution pH 7.5 R1*. Prepare a 0.5 mg/mL solution of *trypsin for peptide mapping R*, and add 10 µL of the solution to 100 µL of the desalted solution. Incubate at 37 °C for 16 h. Add 2 µL of a 150 g/L solution of *trifluoroacetic acid R* and mix gently using a vortex mixer.

NOTE: a protease/protein ratio of 1:20 (w/w) is recommended.

Carry out the reduction/alkylation and digestion steps for the reference solution in the same manner as for the test solution.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (*2.2.29*).

Column:
- — *size*: *l* = 0.25 m, Ø = 4.6 mm;
- — *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm;
- — *temperature*: 30 °C.

Mobile phase:
- — *mobile phase A*: add 0.6 mL of *trifluoroacetic acid R* to 1000 mL of *water for chromatography R*; degas;
- — *mobile phase B*: add 0.6 mL of *trifluoroacetic acid R* to a mixture of 100 mL of *water for chromatography R* and 900 mL of *acetonitrile R1*; degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 3	100	0
3 - 115	100 → 50	0 → 50
115 - 115.5	50 → 10	50 → 90
115.5 - 135	10	90

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Autosampler Set at 2-8 °C.

Injection 80 µL.

Identification of peaks Use the chromatogram supplied with *infliximab CRS* to identify peaks 1 to 20.

System suitability Reference solution:
- — the chromatogram obtained is qualitatively similar to the chromatogram supplied with *infliximab CRS* and peaks 1 to 20 are clearly visible;
- — peaks 5 and 6 are separated as shown in the chromatogram supplied with *infliximab CRS*.

Results:
- — the profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution;
- — no additional peak in the chromatogram obtained with the test solution has an area greater than 0.5 per cent of the sum of the areas of peaks 1 to 20.

TESTS

pH (*2.2.3*)

As approved by the competent authority.

Related proteins

Capillary electrophoresis (*2.2.47*) under both reducing and non-reducing conditions.

Sample buffer Dissolve 1 g of *sodium dodecyl sulfate R* in *tris (hydroxymethyl)aminomethane buffer solution pH 9.0 R1* and dilute to 100.0 mL with the same solution.

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of 2 mg/mL. Mix 27 µL of this solution and 30 µL of sample buffer.
- — *reducing conditions*: add 3 µL of *2-mercaptoethanol R* and incubate at 80 °C for 10 min; allow to cool for 5 min and transfer 60 µL to the autosampler vial;
- — *non-reducing conditions*: add 3 µL of a 46.3 g/L solution of *iodoacetamide R* and incubate at 60-65 °C for 5 min; allow to cool for 5 min and transfer 60 µL to the autosampler vial.

Reference solution Dissolve the contents of a vial of *infliximab CRS* in *water R* to obtain a concentration of 2 mg/mL. Mix 27 µL of the solution and 30 µL of sample buffer. Proceed at the same time and in the same manner as for the test solution.

Capillary:
- — *material*: uncoated fused silica;
- — *size*: total length = about 30 cm, effective length = 20 cm, Ø = 50 µm.

Temperature 25 °C.

Gel buffer Use a formulation suitable for a sieving range of approximately 10-225 kDa.

Acidic wash solution *dilute hydrochloric acid R3.*

Basic wash solution 4 g/L solution of *sodium hydroxide R.*

Detection Spectrophotometer at 220 nm.

Autosampler Set at 25 °C.

Preconditioning of the capillary Rinse the capillary with the basic wash solution for 10 min at 138 kPa, with the acidic wash solution for 5 min at 138 kPa, with *water R* for 2 min at 138 kPa and with the gel buffer for 10 min at 483 kPa. Apply voltage for 10 min (15 kV reversed polarity).

Between-run rinsing Rinse the capillary with the basic wash solution for 3 min at 483 kPa, with the acidic wash solution for 1 min at 483 kPa, with *water R* for 1 min at 483 kPa and with the gel buffer for 10 min at 483 kPa.

Injection Electrokinetically at 5 kV reversed polarity for 20 s.

Migration Apply a voltage of 15 kV reversed polarity for 35 min using the gel buffer as the electrolyte in both buffer reservoirs.

Migration time:
- *reducing conditions*: light chain = 14 min to 17 min; non-glycosylated heavy chain = 17 min to 20 min; heavy chain = 18 min to 21 min;
- *non-reducing conditions*: intact IgG = 26 min to 32 min.

Calculate corrected areas of all peaks with a migration time greater than 11 min, using the following expression:

$$\frac{L_d \times A}{t}$$

L_d = capillary length to detector;
A = uncorrected peak area;
t = migration time.

System suitability Reference solution:
- *reducing conditions*: the electropherogram obtained is qualitatively similar to the electropherogram supplied with *infliximab CRS*;
- *non-reducing conditions*: the electropherogram obtained is qualitatively similar to the electropherogram supplied with *infliximab CRS*.

Result:
- the profile of the electropherogram obtained with the test solution corresponds to that of the electropherogram obtained with the reference solution, except for minor peaks, that may be absent in the electropherogram obtained with the test solution.

Calculate individual peak areas expressed as a percentage of the sum of all corrected peak areas with a migration time greater than 11 min.

Limits:

Reducing conditions:
- *sum of all peaks other than heavy chain and light chain*: maximum 2 per cent;

Non-reducing conditions:
- *sum of all peaks other than the principal peak*: maximum 8 per cent.

Impurities with molecular masses differing from that of infliximab

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Dilute the preparation to be examined with the mobile phase to obtain a concentration of 8 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *infliximab CRS* in the mobile phase to obtain a concentration of 8 mg/mL.

Reference solution (b) Reconstitute a mixture of thyroglobulin, gamma-globulin, ovalbumin, myoglobin and vitamin B_{12} in *water R* to obtain an 18 mg/mL solution of molecular mass markers suitable for calibration in the range of 1350-670 000 Da. Further dilute 10 µL of the solution with *water for chromatography R* to obtain a concentration of 0.9 mg/mL.

Column:
- *size*: *l* = 0.30 m, Ø = 7.8 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5 µm) with a pore size of 25 nm and of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 10 000 to 500 000.

Mobile phase Dissolve 0.78 g of *anhydrous sodium dihydrogen phosphate R*, 1.92 g of *anhydrous disodium hydrogen phosphate R* and 8.77 g of *sodium chloride R* in 900 mL of *water for chromatography R* and dilute to 1000.0 mL with *water for chromatography R*; filter and degas.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 214 nm.

Autosampler Set at 10 °C.

Injection 10 µL.

Relative retention With reference to infliximab monomer (retention time = about 8 min): high molecular weight species = about 0.88; low molecular weight species = about 1.28.

Calculate individual peak areas expressed as a percentage relative to the sum of all peak areas eluting between 5 min and 11 min. Individual relative per cent peak areas are calculated as the average of 3 injections.

NOTE Protein species that elute between 5 min and the monomer peak are classified as high molecular weight species, while those that elute after the monomer peak and before 11 min are classified as low molecular weight species.

System suitability:
- the chromatogram obtained with reference solution (a) is qualitatively similar to the chromatogram supplied with *infliximab CRS*;
- *resolution*: minimum 1.2 between the peaks due to gamma-globulin and ovalbumin in the chromatogram obtained with reference solution (b).

Result:
- the retention time of the principal peak in the chromatogram obtained with the test solution corresponds to that of the principal peak in the chromatogram obtained with reference solution (a).

Limit:
- *sum of all peaks other than the monomer peak*: maximum 2 per cent.

ASSAY

Protein (*2.5.33, Method 1*)

Test solution Dilute the preparation to be examined with a suitable buffer to obtain a concentration of about 1 mg/mL. Prepare and analyse each preparation in duplicate.

Record the UV spectrum between 280 nm and 350 nm. Measure the value at the absorbance maximum of 280 nm, after correction for any light scattering measured up to 350 nm. Calculate the protein content, taking the specific absorbance to be 14.5.

Potency

The potency of infliximab is determined by comparison of dilutions of the test preparation with dilutions of *infliximab BRP* using a suitable cell-based assay based on the inhibitory action of infliximab on the biological activity of TNF-α with a suitable readout for assessing this inhibitory effect.

The following procedure is given as an example.

Carry out a cell proliferation assay based on the ability of infliximab to block TNF-induced inhibition of murine fibrosarcoma WEHI-164 cell proliferation. The WEHI-164 cells (ATCC No. CRL-1751) are incubated with varying dilutions of test and reference preparations of infliximab in the presence of TNF-α. Cell growth is assessed by a staining method using the tetrazolium salt WST-8 ([2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2*H*-tetrazolium, monosodium salt]), which is converted by cellular dehydrogenases to a coloured formazan product. The amount of released formazan is then measured spectrophotometrically and is directly proportional to the number of living cells.

Assay medium RPMI 1640 containing 4.5 g/L *glucose R*, 1.5 g/L *sodium hydrogen carbonate R*, 0.11 g/L *sodium pyruvate R*, 2.4 g/L *HEPES R* (10 mM), 300 g/L *L-glutamine R*, heat-inactivated foetal bovine serum (10 per cent *V/V*) and a penicillin/streptomycin solution (1 per cent *V/V*) containing 10 000 U/mL of *benzylpenicillin sodium R* and 10 000 µg/mL of *streptomycin sulfate R* in an 8.5 g/L solution of *sodium chloride R*.

Test solution Dilute the preparation to be examined with assay medium to obtain a concentration of about 640 ng/mL. Analyse in duplicate.

Reference solution Dissolve the contents of a vial of *infliximab BRP* in assay medium to obtain a concentration of about 640 ng/mL. Analyse in duplicate.

TNF-α working solutions Dissolve the contents of a vial of TNF-α according to the supplier's instructions. Further dilute with assay medium to obtain 2 suitable working concentrations in the range of 400-50 000 pg/mL (working solution A and working solution B (dilution of A)). As the biological activity of TNF-α is likely to vary between different suppliers and also between different batches from the same supplier, this should be controlled by use of an appropriate standard (e.g. WHO International Standard for TNF-α).

Method.

Plate preparation Add 150 µL of assay medium to the wells designated for 'cell only control' (columns 1-6, row H) and for blanks (columns 2-12, row A) on a 96-well microplate. Add 187.5 µL of TNF-α working solution A (column 1, row A) and further perform 5-fold dilutions (columns 2-12, row A) to generate the 'TNF-α control curve'. Add 100 µL of assay medium and 50 µL of TNF-α working solution B to the wells designated for 'cell + TNF-α control' (columns 7-12, row H). Add 100 µL of assay medium to the sample wells (columns 2-12, rows B-G) and 200 µL of the test or reference solutions (column 1, rows B-G). Further prepare a series of 2-fold dilutions (columns 2-12, rows B-G). Then, add 50 µL of TNF-α working solution B (columns 1-12, rows B-G). Incubate at 36.0-38.0 °C for 1 h in an incubator using 5 ± 2 per cent CO_2.

Cell preparation Prepare a suspension of WEHI-164 cells containing 1 × 10^6 cells per millilitre, using assay medium containing 2 µg/mL of actinomycin D.

Plating cells Add 50 µL of the cell suspension to each well maintaining the cells in a uniform suspension during addition. Incubate at 36.0-38.0 °C for 20-24 h in an incubator using 5 ± 2 per cent CO_2.

Addition of tetrazolium salt Remove 100 µL of medium from each well, add 10 µL of reconstituted WST-8 mixture to each well and reincubate for 3-4 h. Estimate the quantity of formazan produced using a microtitre well plate reader at 450 nm and 650 nm. Subtract the reading at 650 nm from the reading at 450 nm.

Calculate the potency of the preparation to be examined using the four-parameter logistic curve model (see general chapter *5.3. Statistical analysis*).

System suitability:
- — the 'TNF-α control curve' corresponds to a sigmoid curve;
- — the coefficient of determination calculated for the 'TNF-α control curve' (r^2) is not less than 0.97;
- — the standard curve corresponds to a sigmoid curve with upper and lower plateaus corresponding to 'cell only control' and 'cell + TNF-α control' respectively;
- — the coefficient of determination calculated for the standard curve (r^2) is not less than 0.97;
- — *maximum value (cell only) to minimum value (TNF-α control) ratio*: minimum 3.0.

Calculate the potency of the preparation to be examined using a suitable statistical method (see general chapter *5.3. Statistical analysis*).

Result The estimated potency is not less than 80 per cent and not more than 120 per cent relative to the reference solution. The confidence limits (P = 0.95) are not less than 80 per cent and not more than 125 per cent of the estimated potency.

STORAGE

In an airtight container, under approved conditions.

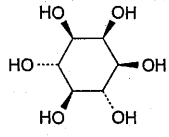
LABELLING

The label states the content in milligrams of protein per millilitre.

Ph Eur

myo-Inositol

(Ph. Eur. monograph 1805)



$C_6H_{12}O_6$ 180.2 *87-89-8*

Action and use

Vasodilator.

Ph Eur

DEFINITION

Cyclohexane-1,2,3,5/4,6-hexol.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *myo-inositol CRS*.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 10.0 g in *distilled water R* and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Conductivity (*2.2.38*)

Maximum 30 $\mu S \cdot cm^{-1}$.

Dissolve 10.0 g in *carbon dioxide-free water R* prepared from *distilled water R*, with gentle warming if necessary, and dilute to 50.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.500 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.500 g of *myo-inositol CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with *water R*. Dilute 5.0 mL of this solution to 100.0 mL with *water R*.

Reference solution (c) Dissolve 0.5 g of *myo-inositol R* and 0.5 g of *mannitol R* in *water R* and dilute to 10 mL with the same solvent.

Column:
- *size*: *l* = 0.3 m, Ø = 7.8 mm;
- *stationary phase*: *strong cation-exchange resin (calcium form) R* (9 µm);
- *temperature*: 85 ± 2 °C.

Mobile phase *water for chromatography R*.

Flow rate 0.5 mL/min.

Detection Differential refractometer maintained at a constant temperature (e.g. 35 °C).

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time Twice the retention time of *myo*-inositol.

Relative retention With reference to *myo*-inositol (retention time = about 17.5 min): impurity A = about 1.3; impurity B = about 1.4.

System suitability Reference solution (c):
- *resolution*: minimum 4.0 between the peaks due to *myo*-inositol and impurity A.

Limits:
- *impurities A, B*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (*2.5.12*)

Maximum 0.5 per cent, determined on 1.00 g.

ASSAY

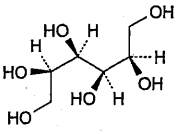
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

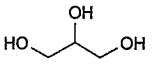
Calculate the percentage content of $C_6H_{12}O_6$ taking into account the assigned content of *myo-inositol CRS*.

IMPURITIES

Specified impurities *A, B.*



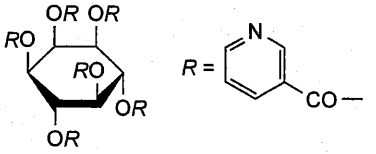
A. D-mannitol,



B. propane-1,2,3-triol (glycerol).

Ph Eur

Inositol Nicotinate



$C_{42}H_{30}N_6O_{12}$ 810.7 *6556-11-2*

Action and use

Vasodilator.

Preparation

Inositol Nicotinate Tablets

DEFINITION

Inositol Nicotinate is *myo*-inositol hexanicotinate. It contains not less than 98.0% and not more than 101.0% of $C_{42}H_{30}N_6O_{12}$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.

Practically insoluble in *water*; practically insoluble in *acetone*, in *ethanol (96%)* and in *ether*. It dissolves in dilute mineral acids.

IDENTIFICATION

The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of inositol nicotinate *(RS 190)*.

TESTS

Clarity and colour of solution

A 5.0% w/v solution in 0.5M *sulfuric acid* is *clear*, Appendix IV A, and not more intensely coloured than *reference solution* BY_6, Appendix IV B, Method II.

Chloride

Dissolve 0.14 g in a sufficient quantity of 2M *nitric acid* and dilute to 16 mL with *water*. The resulting solution complies

with the *limit test for chlorides*, Appendix VII, beginning at the words 'pour the mixture as a single addition...' (350 ppm).

Free nicotinic acid
To 1 g add 75 mL of *water*, shake for 15 minutes and titrate with *0.02M sodium hydroxide VS* using *phenolphthalein solution R1* as indicator. Not more than 0.8 mL of *0.02M sodium hydroxide VS* is required to produce the first pink colour.

Related substances
Carry out the method for *thin-layer chromatography*, Appendix III A, using a plate 200 mm × 200 mm in size and *silica gel* GF_{254} as the coating substance. For the first development use a mixture of 90 volumes of *chloroform* and 10 volumes of *methanol* as the mobile phase. Apply to the bottom right-hand corner of the plate 5 µL of solution (1) containing 5.0% w/v of the substance being examined in a mixture of 9 volumes of *chloroform* and 1 volume of *methanol* and develop over a path of 12 cm. After removal of the plate, allow it to dry in air and turn the plate through 90° in a clockwise direction. Apply separately to the bottom right-hand corner of the plate, and to the right of the solvent front, 5 µL of each of two solutions of the substance being examined in a mixture of 9 volumes of *chloroform* and 1 volume of *methanol* containing (2) 0.075% w/v and (3) 0.050% w/v. For the second development use a mixture of 50 volumes of *ethyl acetate* and 5 volumes each of *glacial acetic acid*, *ethanol (96%)* and *water* as the mobile phase. After removal of the plate, allow it to dry in air and examine under *ultraviolet light (254 nm)*. In the chromatogram obtained with solution (1) any *secondary spot* is not more intense than the spot in the chromatogram obtained with solution (2) (1.5%) and not more than one such spot is more intense than the spot in the chromatogram obtained with solution (3) (1%).

Acetone
Prepare a 0.020% v/v solution of *butan-2-one* (internal standard) in *dimethylformamide* (solution A). Carry out the method for *gas chromatography*, Appendix III B, using the following solutions. Solution (1) contains 0.020% v/v of *acetone* in solution A. For solution (2) add 5 mL of *dimethylformamide* to 0.20 g of the substance being examined contained in a suitable vessel, stopper securely, suspend in a water bath until solution is complete and allow to cool. Prepare solution (3) in the same manner as solution (2) but using 5 mL of solution A in place of the dimethylformamide.

The chromatographic procedure may be carried out using a glass column (1.5 m × 4 mm) packed with *acid-washed, silanised diatomaceous support* coated with 10% w/w of polyethylene glycol 1000 and maintained at 60°.

In the chromatogram obtained with solution (3) the ratio of the area of any peak corresponding to acetone to the area of the peak due to the internal standard is not greater than the corresponding ratio in the chromatogram obtained with solution (1).

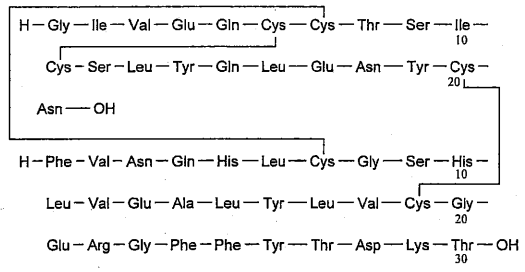
Loss on drying
When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Carry out Method I for *non-aqueous titration*, Appendix VIII A, using 0.2 g and *1-naphtholbenzein solution* as indicator. Each mL of *0.1M perchloric acid VS* is equivalent to 13.51 mg of $C_{42}H_{30}N_6O_{12}$.

Insulin Aspart

(Ph. Eur. monograph 2084)



$C_{256}H_{381}N_{65}O_{79}S_6$ 5826

Action and use
Hormone; treatment of diabetes mellitus.

Preparation
Biphasic Insulin Aspart Injection

Ph Eur

DEFINITION
28^B-L-Aspartate insulin (human).

Insulin aspart is a 2-chain peptide containing 51 amino acids. The A-chain is composed of 21 amino acids and the B-chain is composed of 30 amino acids. It is identical in primary structure to human insulin, except that it has aspartic acid instead of proline at position 28 of the B-chain. As in human insulin, insulin aspart contains 2 interchain disulfide bonds and 1 intrachain disulfide bond.

Content
90.0 per cent to 104.0 per cent of insulin aspart $C_{256}H_{381}N_{65}O_{79}S_6$ plus A21Asp insulin aspart, B3Asp insulin aspart, B3isoAsp insulin aspart and B28isoAsp insulin aspart (dried substance).

By convention, for the purpose of labelling insulin aspart preparations, 0.0350 mg of insulin aspart is equivalent to 1 unit.

PRODUCTION
Insulin aspart is produced by a method based on recombinant DNA (rDNA) technology under conditions designed to minimise the degree of microbial contamination.

Prior to release the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins
The limit is approved by the competent authority.

Single-chain precursor
The limit is approved by the competent authority. Use a suitably sensitive method.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in ethanol (96 per cent), in methanol and in aqueous solutions with a pH around 5.1. In aqueous solutions below pH 3.5 or above pH 6.5, the solubility is greater than or equal to 25 mg/mL.

IDENTIFICATION
A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the

principal peak in the chromatogram obtained with reference solution (a).

B. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a 2.0 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid* and transfer 25 µL of this solution to a clean tube. Add 100 µL of *HEPES buffer solution pH 7.5 R* and 20 µL of a 1 mg/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R*. Cap the tube and incubate at 25 °C for 6 h. Stop the reaction by adding 145 µL of *sulfate buffer solution pH 2.0 R*.

Reference solution Prepare at the same time and in the same manner as for the test solution, but using *insulin aspart CRS* instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION. Liquid chromatography (*2.2.29*).

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm) with a pore size of 8 nm,
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: mix 100 mL of *acetonitrile for chromatography R*, 200 mL of *sulfate buffer solution pH 2.0 R* and 700 mL of *water R*; filter and degas;
- *mobile phase B*: mix 200 mL of *sulfate buffer solution pH 2.0 R*, 400 mL of *acetonitrile for chromatography R* and 400 mL of *water R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	90 → 30	10 → 70
60 - 65	30 → 0	70 → 100
65 - 70	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability:
- the chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of insulin aspart digest supplied with *insulin aspart CRS*,
- in the chromatogram obtained with the reference solution, identify the peaks due to digest fragments I, II and III:

  symmetry factor Maximum 1.5, for the peaks due to fragments II and III,

  resolution Minimum 8.0, between the peaks due to fragments II and III.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: the retention times of fragments I, II and IV are the same as for human insulin. The retention time of fragment III differs from human insulin due to substitution of proline by aspartic acid.

TESTS

Impurities with molecular masses greater than that of insulin aspart

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Prepare a solution containing 4 mg/mL of the substance to be examined in *0.01 M hydrochloric acid*. Maintain the solution at 2-8 °C and use within 48 h.

Resolution solution Use a solution of insulin (about 4 mg/mL), containing more than 0.4 per cent of high molecular mass proteins. An injectable insulin preparation, whether a solution or a suspension, that has been clarified with a sufficient amount of *6 M hydrochloric acid R*, containing the indicated percentage of high molecular mass proteins, or a solution prepared from insulin, dissolved in *0.01 M hydrochloric acid* may be used. Insulin containing the indicated percentage of high molecular mass proteins may be prepared by allowing insulin powder to stand at room temperature for about 10 days. Maintain the solution at 2-8 °C and use within 7 days.

Column:
- *size*: *l* = 0.3 m, Ø = 7.8 mm,
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5-10 µm) with a pore size of 12-12.5 nm, of a grade suitable for the separation of insulin monomer from dimer and polymers.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile for chromatography R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Equilibration At least 3 injections of the resolution solution; the column is equilibrated when repeatable results are obtained from 2 subsequent injections.

Injection 100 µL.

Run time About 35 min.

Retention time Insulin aspart polymers = 13-17 min; insulin aspart dimer = about 17.5 min; insulin aspart monomer = about 20 min; salts = about 22 min.

System suitability Resolution solution:
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer.

Limits The sum of the areas of the peaks with a retention time less than that of the principal peak is not more than 0.5 per cent of the total area of the peaks. Disregard any peak with a retention time greater than that of the peak due to insulin aspart monomer.

Related proteins

Liquid chromatography (*2.2.29*) as described under Assay: use the normalisation procedure.

Limits:
- *B28isoAsp insulin aspart*: maximum 1.0 per cent,
- *total of the peaks due to A21Asp insulin aspart, B3Asp insulin aspart and B3isoAsp insulin aspart*: maximum 2.0 per cent,
- *total of other impurities*: maximum 1.5 per cent.

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 0.200 g by drying in an oven at 105 °C for 24 h.

Sulfated ash (*2.4.14*)

Maximum 6.0 per cent, determined on 0.200 g (dried substance).

Bacterial endotoxins (*2.6.14*)
Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution Dissolve the substance to be examined in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL. Maintain the solution at 2-8 °C and use within 24 h.

Reference solution Dissolve the contents of a vial of *insulin aspart CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL. Maintain the solution at 2-8 °C and use within 48 h.

Resolution solution Use an appropriate solution with a content of B3Asp insulin aspart and A21Asp insulin aspart of not less than 1 per cent. This may be achieved by storing reference solution at room temperature for about 1-3 days. Maintain the solution at 2-8 °C and use within 72 h.

Column:
— *size*: l = 0.25 m, Ø = 4 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: dissolve 142.0 g of *anhydrous sodium sulfate R* in *water R*; add 13.5 mL of *phosphoric acid R* and dilute to 5000 mL with *water R*; adjust to pH 3.6, if necessary, with *strong sodium hydroxide solution R*; filter and degas; mix 9 volumes of the solution with 1 volume of *acetonitrile for chromatography R*; filter and degas;
— *mobile phase B*: mix equal volumes of *water R* and *acetonitrile for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 35	58	42
35 - 40	58 → 20	42 → 80
40 - 45	20	80
45 - 46	20 → 58	80 → 42
46 - 60	58	42

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 10 µL.

Relative retention With reference to insulin aspart (retention time = 20-24 min): B28isoAsp insulin aspart = about 0.9; B3Asp insulin aspart plus A21Asp insulin aspart (generally coeluted) = about 1.3; B3isoAsp insulin aspart = about 1.5.

System suitability Resolution solution:
— *resolution*: minimum 2.0 between the peak due to insulin aspart and the peak due to A21Asp insulin aspart and to B3Asp insulin aspart.

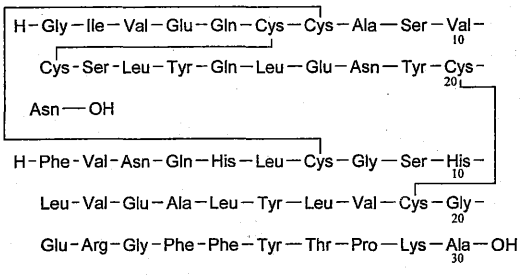
Calculate the content of insulin aspart $C_{256}H_{381}N_{65}O_{79}S_6$, plus B28isoAsp insulin aspart, A21Asp insulin aspart, B3Asp insulin aspart and B3isoAsp insulin aspart using the areas of the corresponding peaks in the chromatograms obtained with the test solution and reference solution and the declared content of insulin aspart plus B28isoAsp insulin aspart, A21Asp insulin aspart, B3Asp insulin aspart and B3isoAsp insulin aspart in *insulin aspart CRS*.

STORAGE
In an airtight container, protected from light, at or below −18 °C until released by the manufacturer. When thawed, insulin aspart is stored at 5 ± 3 °C and used for manufacturing preparations within a short period of time. To avoid absorption of humidity from the air during weighing, insulin aspart must be at room temperature before opening the container.

Ph Eur

Bovine Insulin

(Ph. Eur. monograph 1637)



$C_{254}H_{377}N_{65}O_{75}S_6$ 5734

Action and use
Hormone; treatment of diabetes mellitus.

Preparation
Insulin Preparations

Ph Eur

DEFINITION
Bovine insulin is the natural antidiabetic principle obtained from beef pancreas and purified.

Content
— *sum of bovine insulin ($C_{254}H_{377}N_{65}O_{75}S_6$) and A21 desamido bovine insulin*: 93.0 per cent to 105.0 per cent (dried substance).

By convention, for the purpose of labelling insulin preparations, 0.0342 mg of bovine insulin is equivalent to 1 IU of insulin.

PRODUCTION
The animals from which bovine insulin is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids and with decomposition in dilute solutions of alkali hydroxides.

IDENTIFICATION
A. Examine the chromatograms obtained in the assay.

Results The retention time of the principal peak in the chromatogram obtained with the test solution corresponds to that of the principal peak in the chromatogram obtained with reference solution (c).

B. Peptide mapping

Test solution Prepare a 2.0 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid* and transfer 500 µL of this solution to a clean tube. Add 2.0 mL of *HEPES buffer solution pH 7.5 R* and 400 µL of a 1 mg/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R*. Cap the tube and incubate at 25 °C for 6 h. Stop the reaction by adding 2.9 mL of *sulfate buffer solution pH 2.0 R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *bovine insulin CRS* instead of the substance to be examined.

Examine the digests by liquid chromatography (*2.2.29*).

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm);
- *temperature*: 40 °C.

Mobile phase:
- *mobile phase A*: mix 100 mL of *acetonitrile for chromatography R*, 700 mL of *water for chromatography R* and 200 mL of *sulfate buffer solution pH 2.0 R*; filter and degas;
- *mobile phase B*: mix 400 mL of *acetonitrile for chromatography R*, 400 mL of *water for chromatography R* and 200 mL of *sulfate buffer solution pH 2.0 R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	90 → 30	10 → 70
60 - 65	30 → 0	70 → 100
65 - 70	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability The chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of bovine insulin digest supplied with *bovine insulin CRS*. In the chromatogram obtained with the reference solution, identify the peaks due to digest fragments I, II and III. The symmetry factor of the peaks due to fragments II and III is not greater than 1.5, and the resolution between the 2 peaks is at least 1.9.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: The retention time of fragment I is the same for porcine insulin and for human insulin. The retention times of fragments II and IV are the same for all insulins. The retention time of fragment III is the same for bovine insulin and for porcine insulin.

TESTS

Impurities with molecular masses greater than that of insulin

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure. *Maintain the solutions at 2-10 °C and use within 7 days. If an automatic injector is used, maintain the temperature at 2-10 °C.*

Test solution Dissolve 4 mg of the substance to be examined in 1.0 mL of *0.01 M hydrochloric acid*.

Resolution solution Use a solution of insulin (approximately 4 mg/mL), containing more than 0.4 per cent of high molecular mass proteins. An injectable insulin preparation, whether a solution or a suspension, that has been clarified with a sufficient amount of *6 M hydrochloric acid R*, containing the indicated percentage of high molecular mass proteins, or a solution prepared from insulin, dissolved in *0.01 M hydrochloric acid*, may be used. Insulin containing the indicated percentage of high molecular mass proteins may be prepared by allowing insulin powder to stand at room temperature for about 10 days.

Column:
- *size*: *l* = 0.3 m, Ø = at least 7.5 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5-10 µm), of a grade suitable for the separation of insulin monomer from dimer and polymers.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Equilibration Before using a new column for chromatographic analysis, equilibrate by repeated injections of an insulin solution containing high molecular mass proteins. This can be done by at least 3 injections of the resolution solution. The column is equilibrated when repeatable results are obtained from 2 subsequent injections.

Injection 100 µL.

Run time About 35 min.

Retention times Polymeric insulin complexes = 13 min to 17 min; covalent insulin dimer = about 17.5 min; insulin monomer = about 20 min; salts = about 22 min.

System suitability Resolution solution:
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer.

Limits The sum of the areas of any peaks with a retention time less than that of the principal peak is not greater than 1.0 per cent of the total area of the peaks; disregard any peak with a retention time greater than that of the insulin peak.

Related proteins

Liquid chromatography (*2.2.29*) as described under Assay, following the elution conditions as described in the table below.

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	42	58
30 - 44	42 → 11	58 → 89
44 - 50	11	89

Maintain the solutions at 2-10 °C and use within 24 h

Perform a system suitability test (resolution, linearity) as described under Assay. If necessary, the relative proportions of the mobile phases may be adjusted to ensure complete elution of A21 desamido porcine insulin before commencement of the gradient. The profile of the gradient may also be adjusted to ensure complete elution of all insulin related impurities.

Inject 20 µL of reference solution (c) and 20 µL of the test solution. If necessary, adjust the injection volume to between 10 µL and 20 µL in accordance with the results obtained in the test for linearity as described under Assay. Record the chromatograms for approximately 50 min. In the

chromatogram obtained with reference solution (c), A21 desamido bovine insulin appears as a small peak after the principal peak and has a relative retention of about 1.3 with reference to the principal peak. In the chromatogram obtained with the test solution, the area of the peak due to A21 desamido bovine insulin is not greater than 3.0 per cent of the total area of the peaks; the sum of the areas of all the peaks, apart from those due to bovine insulin and A21 desamido bovine insulin, is not greater than 3.0 per cent of the total area of the peaks.

Bovine proinsulin-like immunoreactivity (PLI)
Maximum 10 ppm (dried substance).

Use a suitably sensitive immunochemical method (*2.7.1*) such as radio-immunoassay, using the International Reference Reagent for bovine proinsulin to calibrate the method.

Zinc
Maximum 1.0 per cent (dried substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 50.0 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 25.0 mL with the same acid. Dilute if necessary to a suitable concentration (for example, 0.4 µg to 1.6 µg of Zn per millilitre) with *0.01 M hydrochloric acid*.

Reference solutions Use solutions containing 0.40 µg, 0.80 µg, 1.00 µg, 1.20 µg and 1.60 µg of Zn per millilitre, freshly prepared by diluting *zinc standard solution (5 mg/mL Zn) R* with *0.01 M hydrochloric acid*.

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Flame Air-acetylene flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 0.200 g by drying in an oven at 105 °C for 24 h.

Sulfated ash (*2.4.14*)
Maximum 2.5 per cent, determined on 0.200 g (dried substance).

Bacterial endotoxins (*2.6.14*)
Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve a suitable amount of the substance to be examined in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (a) Dissolve the contents of a vial of *human insulin CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (b) Dissolve the contents of a vial of *insulin porcine for system suitability CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4 mg/mL.

Reference solution (c) Dissolve the contents of a vial of *bovine insulin CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 10.0 mL with *0.01 M hydrochloric acid*.

Resolution solution Mix 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b).

Maintain the solutions at 2-10 °C and use within 48 h. If an automatic injector is used, maintain the temperature at 2-10 °C.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 42 volumes of mobile phase A and 58 volumes of mobile phase B, adjusting the composition of the mixture if necessary.

Prepare and maintain the following solutions at a temperature of at least 20 °C:
— *mobile phase A*: dissolve 28.4 g of *anhydrous sodium sulfate R* in *water for chromatography R* and dilute to 1000 mL with the same solvent; add 2.7 mL of *phosphoric acid R*; adjust to pH 2.3, if necessary, with *ethanolamine R*; filter and degas;
— *mobile phase B*: mix 550 mL of mobile phase A with 450 mL of *acetonitrile R*; warm the solution to a temperature of at least 20 °C in order to avoid precipitation (mixing of mobile phase A with acetonitrile is endothermic); filter and degas.

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

System suitability:
— *resolution*: inject 20 µL of the resolution solution and 20 µL of reference solution (b). Record the chromatogram of the resolution solution until the peak corresponding to the principal peak in the chromatogram obtained with reference solution (b) is clearly visible. In the chromatogram obtained with the resolution solution, identify the peaks due to porcine insulin and human insulin. The test is not valid unless the resolution between the peaks due to human insulin and porcine insulin is at least 1.2. If necessary, adjust the concentration of acetonitrile in the mobile phase until this resolution is achieved;
— *linearity*: inject 20 µL each of reference solutions (c) and (d). The test is not valid unless the area of the principal peak in the chromatogram obtained with reference solution (c) is 10 ± 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d). If this test fails, adjust the injection volume to between 10 µL and 20 µL, in order that the responses are within the linearity range of the detector.

Injection 20 µL of the test solution.

Calculate the content of bovine insulin $C_{254}H_{377}N_{65}O_{75}S_6$ plus A21 desamido bovine insulin from the area of the principal peak and the area of the peak due to A21 desamido bovine insulin in the chromatograms obtained with the test solution and reference solution (c) and the declared content of bovine insulin plus A21 desamido bovine insulin in *bovine insulin CRS*.

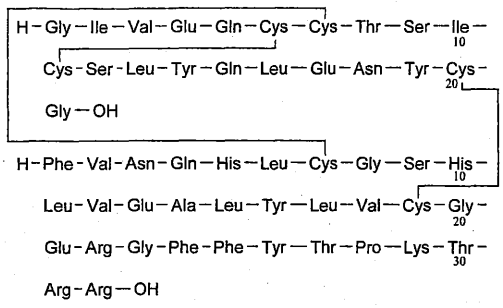
STORAGE

In an airtight container, protected from light, at −20 °C until released by the manufacturer. When thawed, insulin may be stored at 5 ± 3 °C and used for manufacturing preparations within a short period of time. To avoid absorption of humidity from the air during weighing, the insulin must be at room temperature.

Ph Eur

Insulin Glargine

(Ph. Eur. monograph 2571)



$C_{267}H_{404}N_{72}O_{78}S_6$ 6063

Action and use
Hormone; treatment of diabetes mellitus.

Preparation
Insulin Glargine Injection

Ph Eur

DEFINITION

21^{A}-Glycine-30^{B}a-L-arginine-30^{B}b-L-arginine-insulin (human).

2-chain peptide containing 53 amino acids. The A-chain is composed of 21 amino acids and the B-chain is composed of 32 amino acids. It is identical in primary structure to human insulin, only differing in amino acid sequence at position 21 in the A-chain and at the C-terminal end of the B-chain where it contains 2 additional amino acids. Human insulin is Asn(A21), whereas insulin glargine is Gly(A21), Arg(B31), Arg(B32). As in human insulin, insulin glargine contains 2 interchain disulfide bonds and 1 intrachain disulfide bond.

Content
94.0 per cent to 105.0 per cent (anhydrous substance).

By convention, for the purpose of labelling insulin glargine preparations, 0.0364 mg of insulin glargine is equivalent to 1 unit.

PRODUCTION

Insulin glargine is produced by a method based on recombinant DNA (rDNA) technology under conditions designed to minimise the degree of microbial contamination.

Prior to release, the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins
The limit is approved by the competent authority.

Single-chain precursor
The limit is approved by the competent authority. Use a suitably sensitive method.

CHARACTERS

Appearance
White or almost white, hygroscopic powder.

Solubility
Practically insoluble in water and in anhydrous ethanol, soluble in dilute mineral acids.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

B. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a 10.0 mg/mL solution of the substance to be examined in a 1 g/L solution of *hydrochloric acid R* and transfer 5 µL of the solution to a clean tube. Add 1.0 mL of *1 M tris-hydrochloride buffer solution pH 7.5 R* and 100 µL of a 20 U/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R* in *1 M tris-hydrochloride buffer solution pH 7.5 R*. Mix and incubate at 45 °C for about 2 h. Stop the reaction by adding 2 µL of *phosphoric acid R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *insulin glargine CRS* instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION. Liquid chromatography (*2.2.29*).

Buffer solution Dissolve 11.6 g of *phosphoric acid R* and 42.1 g of *sodium perchlorate R* in 1600 mL of *water for chromatography R*, adjust to pH 2.3 with *triethylamine R* and dilute to 2000 mL with *water for chromatography R*.

Column:
- *size*: *l* = 0.125 m, Ø = 3.0 mm;
- *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (4 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: *acetonitrile R1*, buffer solution (7:93 *V/V*);
- *mobile phase B*: buffer solution, *acetonitrile R1* (43:57 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	90 → 20	10 → 80
30 - 35	20	80

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min.

Injection 50 µL.

Retention time Insulin glargine fragment II = about 14 min; insulin glargine fragment III = about 15 min.

System suitability:
- the chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of insulin glargine digest supplied with *insulin glargine CRS*;
- in the chromatogram obtained with the reference solution, identify the peaks due to digest fragments II and III:

  symmetry factor Maximum 1.5 for the peaks due to fragments II and III;

  resolution Minimum 3.4 between the peaks due to fragments II and III.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: the retention times of fragments I and IV are the same as for human insulin; the retention times of fragments II and III differ from human insulin due to the difference in the sequence at position 21 of the A-chain and to the 2 additional amino acids of the B-chain.

TESTS

Impurities with molecular masses greater than that of insulin glargine

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Prepare a 4 mg/mL solution of the substance to be examined in a 1 g/L solution of *hydrochloric acid R*.

Resolution solution Dry about 200 mg of the substance to be examined in an oven at 100 °C for 1.5-3 h. Dissolve 15.0 mg of the dried substance in 1.5 mL of a 1 g/L solution of *hydrochloric acid R* and dilute to 10.0 mL with *water R*.

Reference solution Dilute 1.0 mL of the test solution to 100.0 mL with *water R*. Dilute 3.0 mL of this solution to 20.0 mL with *water R*.

Column:
- *size*: l = 0.3 m, Ø = 7.8 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (10 µm) with a pore size of 12.5 nm, of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 5000 to 150 000.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 100 µL.

Run time About 35 min.

Retention time Insulin glargine = about 18 min.

System suitability:
- *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with the reference solution;
- *symmetry factor*: maximum 2.0 for the peak due to insulin glargine in the chromatogram obtained with the resolution solution;
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to high molecular mass proteins and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to insulin glargine in the chromatogram obtained with the resolution solution.

Limits:
- *total of impurities with a retention time less than that of insulin glargine*: maximum 0.3 per cent of the total area of the peaks; disregard any peak with a retention time greater than that of the peak due to insulin glargine.

Related proteins

Liquid chromatography (*2.2.29*): use the normalisation procedure. *Maintain the solutions at 2-8 °C.*

Test solution Dissolve 15.0 mg of the substance to be examined in 1.5 mL of a 1 g/L solution of *hydrochloric acid R* and dilute to 10.0 mL with *water R*.

Reference solution Dissolve the contents of a vial of *insulin glargine CRS* in 1.5 mL of a 1 g/L solution of *hydrochloric acid R*, transfer the solution with *water R* to a 10 mL volumetric flask and dilute to 10.0 mL with *water R*.

Resolution solution Dissolve the contents of a vial of *insulin glargine for peak identification CRS* (containing 0^A-Arg-insulin glargine) in 0.3 mL of a 1 g/L solution of *hydrochloric acid R* and add 1.7 mL of *water R*.

Buffer solution Dissolve 20.7 g of *anhydrous sodium dihydrogen phosphate R* in 900 mL of *water for chromatography R*, adjust to pH 2.5 with *phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*.

Column:
- *size*: l = 0.25 m, Ø = 3.0 mm;
- *stationary phase*: spherical *end-capped octadecylsilyl silica gel for chromatography R* (4 µm);
- *temperature*: 35 °C.

Mobile phase:
- *mobile phase A*: dissolve 18.4 g of *sodium chloride R* in 250 mL of the buffer solution, add 250 mL of *acetonitrile R1* and mix; dilute to 1000 mL with *water for chromatography R*;
- *mobile phase B*: dissolve 3.2 g of *sodium chloride R* in 250 mL of the buffer solution, add 650 mL of *acetonitrile R1* and mix; dilute to 1000 mL with *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	96 → 83	4 → 17
20 - 30	83 → 63	17 → 37
30 - 33	63 → 96	37 → 4
33 - 40	96	4

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 5 µL of the test solution and the resolution solution.

Retention time Insulin glargine = about 20 min.

System suitability Resolution solution:
- *peak-to-valley ratio*: minimum 2, where H_p = height above the baseline of the peak due to 0^A-Arg-insulin glargine and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to insulin glargine.

Limits:
- *any impurity*: for each impurity, maximum 0.4 per cent;
- *total*: maximum 1.0 per cent.

Zinc

Maximum 0.80 per cent.

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 45.0 mg of the substance to be examined in a 1 g/L solution of *hydrochloric acid R* and dilute to 50.0 mL with the same solution. Dilute 10.0 mL of the solution to 100.0 mL with a 1 g/L solution of *hydrochloric acid R*.

Reference solutions Prepare reference solutions containing 0.2 µg, 0.4 µg and 0.6 µg of zinc per millilitre by diluting *zinc standard solution (10 ppm Zn) R* with a 1 g/L solution of *hydrochloric acid R*.

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Water (*2.5.32*)

Maximum 8.0 per cent, determined on 30.0 mg.

Bacterial endotoxins (*2.6.14, Method D*)

Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related proteins with the following modification.

Injection 5 µL of the test solution and the reference solution.

Calculate the content of insulin glargine ($C_{267}H_{404}N_{72}O_{78}S_6$) taking into account the assigned content of *insulin glargine CRS*.

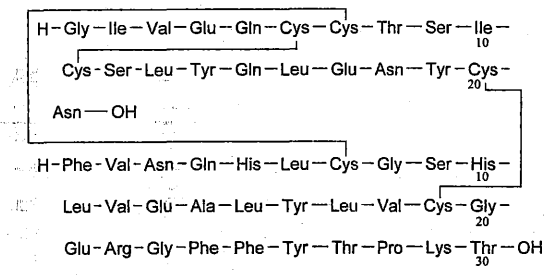
STORAGE

In an airtight container, protected from light, at a temperature of −20 ± 5 °C.

Ph Eur

Human Insulin

(Ph. Eur. monograph 0838)



$C_{257}H_{383}N_{65}O_{77}S_6$ 5808

Action and use

Hormone; treatment of diabetes mellitus.

Preparation

Insulin Preparations

Ph Eur

DEFINITION

Human insulin is a 2-chain peptide having the structure of the antidiabetic hormone produced by the human pancreas.

Content

95.0 per cent to 105.0 per cent of human insulin $C_{257}H_{383}N_{65}O_{77}S_6$ plus A21 desamido human insulin (dried substance).

By convention, for the purpose of labelling insulin preparations, 0.0347 mg of human insulin is equivalent to 1 IU of insulin.

PRODUCTION

Human insulin is produced either by enzymatic modification and suitable purification of insulin obtained from the pancreas of the pig or by a method based on recombinant DNA (rDNA) technology.

Where applicable, the animals from which human insulin is derived must fulfil the requirements for the health of animals suitable for human consumption.

Human insulin is produced under conditions designed to minimise the degree of microbial contamination.

For human insulin produced by enzymatic modification of insulin obtained from the pancreas of the pig, the manufacturing process is validated to demonstrate removal of any residual proteolytic activity. The competent authority may require additional tests.

For human insulin produced by a method based on rDNA technology, prior to release the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins

The limit is approved by the competent authority.

Single chain precursor

The limit is approved by the competent authority. Use a suitably sensitive method.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids and with decomposition in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a 2.0 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid* and transfer 500 µL of this solution to a clean tube. Add 2.0 mL of *HEPES buffer solution pH 7.5 R* and 400 µL of a 1 mg/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R*. Cap the tube and incubate at 25 °C for 6 h. Stop the reaction by adding 2.9 mL of *sulfate buffer solution pH 2.0 R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *human insulin CRS* instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION. Liquid chromatography (*2.2.29*).

Column:

- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm) with a pore size of 8 nm;
- *temperature*: 40 °C.

Mobile phase:

- *mobile phase A*: mix 100 mL of *acetonitrile for chromatography R*, 200 mL of *sulfate buffer solution pH 2.0 R* and 700 mL of *water R*; filter and degas;
- *mobile phase B*: mix 200 mL of *sulfate buffer solution pH 2.0 R*, 400 mL of *acetonitrile for chromatography R* and 400 mL of *water R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	90 → 30	10 → 70
60 - 65	30 → 0	70 → 100
65 - 70	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability:

- the chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of human insulin digest supplied with *human insulin CRS*;

— in the chromatogram obtained with the reference solution, identify the peaks due to digest fragments I, II and III;

symmetry factor Maximum 1.5 for the peaks due to fragments II and III;

resolution Minimum 3.4 between the peaks due to fragments II and III;

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: the retention time of fragment I is the same for porcine insulin and for human insulin. The retention times of fragments II and IV are the same for all insulins. The retention time of fragment III is the same for bovine insulin and for porcine insulin.

TESTS

Impurities with molecular masses greater than that of insulin

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Prepare a solution containing 4 mg/mL of the substance to be examined in *0.01 M hydrochloric acid.*

Resolution solution Use a solution of insulin (about 4 mg/mL), containing more than 0.4 per cent of high molecular mass proteins. An injectable insulin preparation, whether a solution or a suspension, that has been clarified with a sufficient amount of *6 M hydrochloric acid R*, containing the indicated percentage of high molecular mass proteins, or a solution prepared from insulin, dissolved in *0.01 M hydrochloric acid*, may be used. Insulin containing the indicated percentage of high molecular mass proteins may be prepared by allowing insulin powder to stand at room temperature for about 10 days.

Maintain the solutions at 2-8 °C and use within 7 days. If an automatic injector is used, maintain the temperature at 2-8 °C.

Column:

— *size*: *l* = 0.3 m, Ø = minimum 7.5 mm;

— *stationary phase*: *hydrophilic silica gel for chromatography R* (5-10 µm) with a pore size of 12-12.5 nm, of a grade suitable for the separation of insulin monomer from dimer and polymers.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Equilibration Before using a new column for chromatographic analysis, equilibrate by repeated injections of an insulin solution containing high molecular mass proteins. This can be done by at least 3 injections of the resolution solution. The column is equilibrated when repeatable results are obtained from 2 subsequent injections.

Injection 100 µL.

Run time About 35 min.

Retention time Polymeric insulin complexes = 13-17 min; covalent insulin dimer = about 17.5 min; insulin monomer = about 20 min; salts = about 22 min.

System suitability Resolution solution:

— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer.

Limits The sum of the areas of any peaks with a retention time less than that of the principal peak is not greater than 1.0 per cent of the total area of the peaks. Disregard any peak with a retention time greater than that of the peak due to insulin.

Related proteins

Liquid chromatography (*2.2.29*) as described under Assay, following the elution conditions as described below:

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	42	58
30 - 44	42 → 11	58 → 89
44 - 50	11	89

Maintain the solutions at 2-8 °C and use within 24 h. Perform a system suitability test (resolution, linearity) as described in the assay. If necessary, the relative proportions of the mobile phases may be adjusted to ensure complete elution of A21 desamido porcine insulin before commencement of the gradient. The profile of the gradient may also be adjusted to ensure complete elution of all insulin related impurities.

Inject 20 µL of reference solution (a), 20 µL of reference solution (b), 20 µL of reference solution (c) and 20 µL of the test solution. If necessary, adjust the injection volume to a volume between 10 µL and 20 µL in accordance with the results obtained in the test for linearity as described in the assay. Record the chromatograms for approximately 50 min. In the chromatogram obtained with reference solution (a), A21 desamido human insulin appears as a small peak after the principal peak and has a retention time of about 1.3 relative to the principal peak. In the chromatogram obtained with the test solution, the area of the peak due to A21 desamido human insulin is not greater than 2.0 per cent of the total area of the peaks; the sum of the areas of all peaks, apart from those due to human insulin and that due to A21 desamido human insulin, is not greater than 2.0 per cent of the total area of the peaks. For semi-synthetic human insulin only: in the chromatogram obtained with the test solution, the area of any peak corresponding to the peak due to porcine insulin in the chromatogram obtained with reference solution (c) is not greater than 1.0 per cent of the total area of the peaks.

The following test applies only to human insulin produced by enzymatic modification of porcine insulin.

Proinsulin-like immunoreactivity (PLI)

Maximum 10 ppm, calculated with reference to the dried substance and determined by a suitably sensitive immunochemical method (*2.7.1*) such as radio-immunoassay. Use the International Reference Reagent for porcine proinsulin to calibrate the method.

Zinc

Maximum 1.0 per cent (dried substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 50.0 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 25.0 mL with the same acid. Dilute if necessary to a suitable concentration (for example, 0.4-1.6 µg of Zn per millilitre) with *0.01 M hydrochloric acid.*

Reference solutions Use solutions containing 0.40 µg, 0.80 µg, 1.00 µg, 1.20 µg and 1.60 µg of Zn per millilitre, freshly prepared by diluting *zinc standard solution (5 mg/mL Zn) R* with *0.01 M hydrochloric acid.*

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 0.200 g by drying in an oven at 105 °C for 24 h.

Sulfated ash (*2.4.14*)
Maximum 2.5 per cent, determined on 0.200 g (dried substance).

Bacterial endotoxins (*2.6.14*)
Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of *human insulin CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (b) Dissolve the contents of a vial of *insulin porcine for system suitability CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4 mg/mL.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 50.0 mL with *0.01 M hydrochloric acid.* To 1.0 mL of this solution add 1.0 mL of reference solution (a).

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with *0.01 M hydrochloric acid.*

Resolution solution Mix 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b).

Maintain the solutions at 2-8 °C and use within 48 h. If an automatic injector is used, maintain at 2-8 °C.

Column:
— *size*: *l* = 0.25, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 42 volumes of mobile phase A and 58 volumes of mobile phase B, adjusting the composition of the mixture if necessary.

Prepare and maintain the following solutions at a temperature of at least 20 °C:
— *mobile phase A*: dissolve 28.4 g of *anhydrous sodium sulfate R* in *water R* and dilute to 1000 mL with the same solvent; add 2.7 mL of *phosphoric acid R*; adjust to pH 2.3, if necessary, with *ethanolamine R*; filter and degas;
— *mobile phase B*: mix 550 mL of mobile phase A with 450 mL of *acetonitrile R*. Warm the solution to a temperature of at least 20 °C in order to avoid precipitation (mixing of mobile phase A with acetonitrile is endothermic); filter and degas.

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

System suitability:
— *resolution*: inject 20 µL of the resolution solution and 20 µL of reference solution (b). Record the chromatogram of the resolution solution until the peak corresponding to the principal peak in the chromatogram obtained with reference solution (b) is clearly visible. In the chromatogram obtained with the resolution solution, identify the peaks due to porcine insulin and human insulin. The test is not valid unless the resolution between the peaks due to human insulin and porcine insulin is at least 1.2. If necessary, adjust the concentration of acetonitrile in the mobile phase until this resolution is achieved.
— *linearity*: inject 20 µL each of reference solutions (a) and (d). The test is not valid unless the area of the principal peak in the chromatogram obtained with reference solution (a) is 10 ± 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d). If this test fails, adjust the injection volume to between 10 µL and 20 µL, in order that the responses are within the linearity range of the detector.

Injection 20 µL of the test solution and reference solution (a).

Calculate the content of human insulin $C_{257}H_{383}N_{65}O_{77}S_6$ plus A21 desamido human insulin using the areas of the corresponding peaks in the chromatograms obtained with the test solution and reference solution (a) and the declared content of human insulin plus A21 desamido human insulin in *human insulin CRS*.

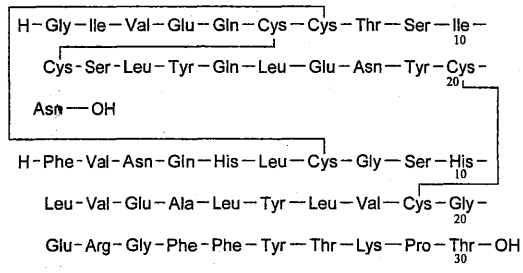
STORAGE
In an airtight container, protected from light, at −18 °C or below, until released by the manufacturer. When thawed, insulin is stored at 5 ± 3 °C and used for manufacturing preparations within a short period of time. To avoid absorption of humidity from the air during weighing, the insulin must be at room temperature.

LABELLING
The label states whether the substance is produced by enzymatic modification of porcine insulin or by rDNA technology.

Ph Eur

Insulin Lispro

(Ph. Eur. monograph 2085)



$C_{257}H_{383}N_{65}O_{77}S_6$ 5808

Action and use
Hormone; treatment of diabetes mellitus.

Preparation
Biphasic Insulin Lispro Injection

Ph Eur

DEFINITION
28^B-L-Lysine-29^B-L-proline insulin (human).

Insulin lispro is a 2-chain peptide containing 51 amino acids. The A-chain is composed of 21 amino acids and the B-chain is composed of 30 amino acids. It is identical in primary

structure to human insulin, only differing in amino acid sequence at positions 28 and 29 of the B-chain. Human insulin is Pro(B28), Lys(B29), whereas insulin lispro is Lys (B28), Pro(B29). As in human insulin, insulin lispro contains 2 interchain disulfide bonds and 1 intrachain disulfide bond.

Content

94.0 per cent to 104.0 per cent (dried substance).

By convention, for the purpose of labelling insulin lispro preparations, 0.0347 mg of insulin lispro is equivalent to 1 unit.

PRODUCTION

Insulin lispro is produced by a method based on recombinant DNA (rDNA) technology under conditions designed to minimise the degree of microbial contamination.

Prior to release the following tests are carried out on each batch of final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins

The limit is approved by the competent authority.

Single-chain precursor

The limit is approved by the competent authority. Use a suitably sensitive method.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids and with decomposition in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

B. Peptide mapping (*2.2.55*).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a 2.0 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid* and transfer 500 µL of this solution to a clean tube. Add 2.0 mL of *HEPES buffer solution pH 7.5 R* and 400 µL of a 1 mg/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R*. Cap the tube and incubate at 25 °C for 6 h. Stop the reaction by adding 2.9 mL of *sulfate buffer solution pH 2.0 R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *insulin lispro CRS* instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION. Liquid chromatography (*2.2.29*).

Column:

- *size*: *l* = 0.10 m, Ø = 4.6 mm,
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm) with a pore size of 8 nm,
- *temperature*: 40 °C.

Mobile phase:

- *mobile phase A*: mix 100 mL of *acetonitrile for chromatography R*, 200 mL of *sulfate buffer solution pH 2.0 R* and 700 mL of *water R*; filter and degas;
- *mobile phase B*: mix 200 mL of *sulfate buffer solution pH 2.0 R*, 400 mL of *acetonitrile for chromatography R* and 400 mL of *water R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	90 → 30	10 → 70
60 - 65	30 → 0	70 → 100
65 - 70	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability:

- the chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of insulin lispro digest supplied with *insulin lispro CRS*,
- in the chromatogram obtained with the reference solution, identify the peaks due to digest fragments I, II and III:

symmetry factor Maximum 1.5 for the peaks due to fragments II and III,

resolution Minimum 8.0 between the peaks due to fragments II and III.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: the retention times of fragments I, II and IV are the same as for human insulin. The retention time of fragment III differs from human insulin due to differences in sequence at positions 28 and 29 of the B-chain.

TESTS

Impurities with molecular masses greater than that of insulin lispro

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure.

Test solution Prepare a solution containing 4 mg/mL of the substance to be examined in *0.01 M hydrochloric acid*. Maintain the solution at 2-8 °C and use within 48 h.

Resolution solution Use a solution of insulin (about 4 mg/mL), containing more than 0.4 per cent of high molecular mass proteins. An injectable insulin preparation, whether a solution or a suspension, that has been clarified with a sufficient amount of *6 M hydrochloric acid R*, containing the indicated percentage of high molecular mass proteins, or a solution prepared from insulin, dissolved in *0.01 M hydrochloric acid*, may be used. Insulin containing the indicated percentage of high molecular mass proteins may be prepared by allowing insulin powder to stand at room temperature for about 10 days. Maintain the solution at 2-8 °C and use within 8 days.

Column:

- *size*: *l* = 0.30 m, Ø = 7.8 mm,
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5-10 µm) with a pore size of 12-12.5 nm, of a grade suitable for the separation of insulin monomer from dimer and polymers.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile for chromatography R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Equilibration At least 3 injections of the resolution solution; the column is equilibrated when repeatable results are obtained for 2 subsequent injections.

Injection 100 µL.

Run time About 35 min.

Retention time Insulin lispro polymers = 13-17 min; insulin lispro dimer = about 17.5 min; insulin lispro monomer = about 20 min; salts = about 22 min.

System suitability Resolution solution:
— *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer,
— *symmetry factor*: maximum 2.0 for the peak due to insulin lispro.

Limits The sum of the areas of the peaks with a retention time less than that of the principal peak is not more than 0.25 per cent of the total area of the peaks. Disregard any peak with a retention time greater than that of the peak due to insulin lispro monomer.

Related proteins
Liquid chromatography (*2.2.29*): use the normalisation procedure.

Test solution Dissolve 3.5 mg of the substance to be examined in 1.0 mL of *0.01 M hydrochloric acid*. Maintain the solution at 2-8 °C and use within 56 h.

Resolution solution Dissolve 3.5 mg of the substance to be examined in 1.0 mL of *0.01 M hydrochloric acid*. Allow to stand at room temperature to obtain a solution containing between 0.8 per cent and 11 per cent of A21 desamido insulin lispro.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm,
— *temperature*: 40 °C.

Mobile phase:
— *mobile phase A*: mix 82 volumes of a 28.4 g/L solution of *anhydrous sodium sulfate R* adjusted to pH 2.3 with *phosphoric acid R* and 18 volumes of *acetonitrile for chromatography R*; filter and degas;
— *mobile phase B*: mix equal volumes of a 28.4 g/L solution of *anhydrous sodium sulfate R* adjusted to pH 2.3 with *phosphoric acid R* and *acetonitrile for chromatography R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	81	19
60 - 83	81 → 51	19 → 49
83 - 84	51 → 81	49 → 19
84 - 94	81	19

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Retention time Adjust the mobile phase composition to obtain a retention time of about 41 min for insulin lispro; A21 desamido insulin lispro elutes near the start of the gradient elution.

System suitability Resolution solution:
— *resolution*: minimum 1.5 between the 1st peak (insulin lispro) and the 2nd peak (A21 desamido insulin lispro),
— *symmetry factor*: maximum 2.0 for the peak due to insulin lispro.

Limits:
— *A21 desamido insulin lispro*: maximum 1.0 per cent,
— *any other impurity*: maximum 0.50 per cent,
— *total (excluding A21)*: maximum 2.0 per cent.

Zinc
Maximum 1.0 per cent (dried substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve at least 50 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 25 mL with the same acid. Dilute if necessary to a suitable concentration (for example 0.4-0.6 µg of Zn per millilitre) with *0.01 M hydrochloric acid*.

Reference solutions Use solutions of concentrations which bracket the expected zinc concentration of the samples, for example, 0.2-0.8 µg of Zn per millilitre, freshly prepared by diluting *zinc standard solution (5 mg/mL Zn) R* with *0.01 M hydrochloric acid*.

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Loss on drying (*2.2.32*)
Maximum 10.0 per cent, determined on 0.200 g by drying in an oven at 105 °C for 16 h.

Sulfated ash (*2.4.14*)
Maximum 2.5 per cent, determined on 0.200 g (dried substance).

Bacterial endotoxins (*2.6.14, Method D*)
Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
Liquid chromatography (*2.2.29*).

Test solution Dissolve the substance to be examined in *0.01 M hydrochloric acid* to obtain a concentration of 0.8 mg/mL. Maintain the solution at 2-8 °C and use within 48 h.

Reference solution Dissolve the contents of a vial of *insulin lispro CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 0.8 mg/mL. Maintain the solution at 2-8 °C and use within 48 h.

Resolution solution Dissolve about 10 mg of the substance to be examined in 10 mL of *0.01 M hydrochloric acid*. Allow to stand at room temperature to obtain a solution containing between 0.8 per cent and 11 per cent of A21 desamido insulin lispro. Maintain the solution at 2-8 °C and use within 14 days.

Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm) with a pore size of 8 nm,
— *temperature*: 40 °C.

Mobile phase Mix 745 volumes of a 28.4 g/L solution of *anhydrous sodium sulfate R* adjusted to pH 2.3 with *phosphoric acid R* and 255 volumes of *acetonitrile for chromatography R*; filter and degas.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Retention time Insulin lispro = about 24 min.

System suitability:

— *resolution*: minimum 1.8 between the 1st peak (insulin lispro) and the 2nd peak (A21 desamido insulin lispro), in the chromatogram obtained with the resolution solution,

— *repeatability*: maximum relative standard deviation of 1.1 per cent after 3 injections of the reference solution.

Calculate the content of insulin lispro $C_{257}H_{383}N_{65}O_{77}S_6$ using the chromatograms obtained with the test solution and the reference solution and the declared content of $C_{257}H_{383}N_{65}O_{77}S_6$ in *insulin lispro CRS*.

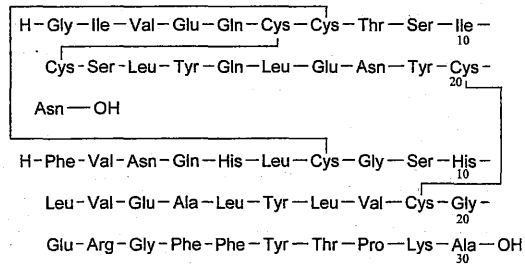
STORAGE

In an airtight container, protected from light, at or below −18 °C. When thawed, insulin lispro is stored and weighed under conditions defined by the manufacturer to maintain the quality attributes of the drug substance and is used for manufacturing preparations within a short period of time. To avoid absorption of humidity from the air during weighing, insulin lispro must be at room temperature before opening the container.

Ph Eur

Porcine Insulin

(Ph. Eur. monograph 1638)



$C_{256}H_{381}N_{65}O_{76}S_6$ 5778

Action and use

Hormone; treatment of diabetes mellitus.

Preparation

Insulin Preparations

Ph Eur

DEFINITION

Porcine insulin is the natural antidiabetic principle obtained from pork pancreas and purified.

Content

— *sum of porcine insulin ($C_{256}H_{381}N_{65}O_{76}S_6$) and A21 desamido porcine insulin*: 95.0 per cent to 105.0 per cent (dried substance).

By convention, for the purpose of labelling insulin preparations, 0.0345 mg of porcine insulin is equivalent to 1 IU of insulin.

PRODUCTION

The animals from which porcine insulin is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids and with decomposition in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The retention time of the principal peak in the chromatogram obtained with the test solution corresponds to that of the principal peak in the chromatogram obtained with reference solution (b).

B. Peptide mapping.

Test solution Prepare a 2.0 mg/mL solution of the substance to be examined in *0.01 M hydrochloric acid* and transfer 500 µL of this solution to a clean tube. Add 2.0 mL of *HEPES buffer solution pH 7.5 R* and 400 µL of a 1 mg/mL solution of *Staphylococcus aureus strain V8 protease, type XVII-B R*. Cap the tube and incubate at 25 °C for 6 h. Stop the reaction by adding 2.9 mL of *sulfate buffer solution pH 2.0 R*.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *insulin porcine for system suitability CRS* instead of the substance to be examined.

Examine the digests by liquid chromatography (*2.2.29*).

Column:

— *size*: *l* = 0.10 m, Ø = 4.6 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm);

— *temperature*: 40 °C.

Mobile phase:

— *mobile phase A*: mix 100 mL of *acetonitrile for chromatography R*, 700 mL of *water for chromatography R* and 200 mL of *sulfate buffer solution pH 2.0 R*; filter and degas;

— *mobile phase B*: mix 400 mL of *acetonitrile for chromatography R*, 400 mL of *water for chromatography R* and 200 mL of *sulfate buffer solution pH 2.0 R*; filter and degas;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	90 → 30	10 → 70
60 - 65	30 → 0	70 → 100
65 - 70	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Equilibration At initial conditions for at least 15 min. Carry out a blank run using the above-mentioned gradient.

Injection 50 µL.

System suitability The chromatograms obtained with the test solution and the reference solution are qualitatively similar to the chromatogram of porcine insulin digest supplied with *insulin porcine for system suitability CRS*. In the chromatogram obtained with the reference solution, identify the peaks due to digest fragments I, II and III. The symmetry factor of the peaks due to fragments II and III is not greater than 1.5, and the resolution between the 2 peaks is at least 1.9.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

NOTE: the retention time of fragment I is the same for porcine insulin and for human insulin. The retention times of fragments II and IV are the same for all insulins. The retention time of fragment III is the same for bovine insulin and for porcine insulin.

TESTS

Impurities with molecular masses greater than that of insulin

Size-exclusion chromatography (*2.2.30*): use the normalisation procedure. *Maintain the solutions at 2-10 °C and use within 7 days. If an automatic injector is used, maintain the temperature at 2-10 °C.*

Test solution Dissolve 4 mg of the substance to be examined in 1.0 mL of *0.01 M hydrochloric acid.*

Resolution solution Use a solution of insulin (approximately 4 mg/mL), containing more than 0.4 per cent of high molecular mass proteins. An injectable insulin preparation, whether a solution or a suspension, that has been clarified with a sufficient amount of *6 M hydrochloric acid R*, containing the indicated percentage of high molecular mass proteins, or a solution prepared from insulin, dissolved in *0.01 M hydrochloric acid*, may be used. Insulin containing the indicated percentage of high molecular mass proteins may be prepared by allowing insulin powder to stand at room temperature for about 10 days.

Column:
- *size*: l = 0.3 m, Ø = at least 7.5 mm;
- *stationary phase*: *hydrophilic silica gel for chromatography R* (5-10 µm), of a grade suitable for the separation of insulin monomer from dimer and polymers.

Mobile phase Mix 15 volumes of *glacial acetic acid R*, 20 volumes of *acetonitrile R* and 65 volumes of a 1.0 g/L solution of *arginine R*; filter and degas.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 276 nm.

Equilibration Before using a new column for chromatographic analysis, equilibrate by repeated injections of an insulin solution containing high molecular mass proteins. This can be done by at least 3 injections of the resolution solution. The column is equilibrated when repeatable results are obtained from 2 subsequent injections.

Injection 100 µL.

Run time About 35 min.

Retention times Polymeric insulin complexes = 13 min to 17 min; covalent insulin dimer = about 17.5 min; insulin monomer = about 20 min; salts = about 22 min.

System suitability Resolution solution:
- *peak-to-valley ratio*: minimum 2.0, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer.

Limits The sum of the areas of any peaks with a retention time less than that of the principal peak is not greater than 1.0 per cent of the total area of the peaks; disregard any peak with a retention time greater than that of the insulin peak.

Related proteins

Liquid chromatography (*2.2.29*) as described under Assay, following the elution conditions as described in the table below.

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	42	58
30 - 44	42 → 11	58 → 89
44 - 50	11	89

Maintain the solutions at 2-10 °C and use within 24 h.

Perform a system suitability test (resolution, linearity) as described under Assay. If necessary, the relative proportions of the mobile phases may be adjusted to ensure complete elution of A21 desamido porcine insulin before commencement of the gradient. The profile of the gradient may also be adjusted to ensure complete elution of all insulin related impurities.

Inject 20 µL of reference solution (b) and 20 µL of the test solution. If necessary, adjust the injection volume to between 10 µL and 20 µL in accordance with the results obtained in the test for linearity as described under Assay. Record the chromatograms for approximately 50 min. In the chromatogram obtained with reference solution (b), A21 desamido porcine insulin appears as a small peak after the principal peak and has a relative retention of about 1.3 with reference to the principal peak. In the chromatogram obtained with the test solution, the area of the peak due to A21 desamido porcine insulin is not greater than 2.0 per cent of the total area of the peaks; the sum of the areas of all the peaks, apart from those due to porcine insulin and A21 desamido porcine insulin, is not greater than 2.0 per cent of the total area of the peaks.

Porcine proinsulin-like immunoreactivity (PLI)

Maximum 10 ppm (dried substance).

Use a suitably sensitive immunochemical method (*2.7.1*) such as radio-immunoassay, using the International Reference Reagent for porcine proinsulin to calibrate the method.

Zinc

Maximum 1.0 per cent (dried substance).

Atomic absorption spectrometry (*2.2.23, Method I*).

Test solution Dissolve 50.0 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 25.0 mL with the same acid. Dilute if necessary to a suitable concentration (for example, 0.4 µg to 1.6 µg of Zn per millilitre) with *0.01 M hydrochloric acid.*

Reference solutions Use solutions containing 0.40 µg, 0.80 µg, 1.00 µg, 1.20 µg and 1.60 µg of Zn per millilitre, freshly prepared by diluting *zinc standard solution (5 mg/mL Zn) R* with *0.01 M hydrochloric acid.*

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Flame Air-acetylene flame of suitable composition (for example, 11 L of air and 2 L of acetylene per minute).

Loss on drying (*2.2.32*)

Maximum 10.0 per cent, determined on 0.200 g by drying in an oven at 105 °C for 24 h.

Sulfated ash (*2.4.14*)

Maximum 2.5 per cent, determined on 0.200 g (dried substance).

Bacterial endotoxins (*2.6.14*)

Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in *0.01 M hydrochloric acid* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of *human insulin CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (b) Dissolve the contents of a vial of *porcine insulin CRS* in *0.01 M hydrochloric acid* to obtain a concentration of 4.0 mg/mL.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with *0.01 M hydrochloric acid.*

Resolution solution Mix 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b).

Maintain the solutions at 2-10 °C and use within 48 h. If an automatic injector is used, maintain the temperature at 2-10 °C.

Column:
— *size*: l = 0.25, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase Mix 42 volumes of mobile phase A and 58 volumes of mobile phase B, adjusting the composition of the mixture if necessary.

Prepare and maintain the following solutions at a temperature of at least 20 °C:
— *mobile phase A*: dissolve 28.4 g of *anhydrous sodium sulfate R* in *water for chromatography R* and dilute to 1000 mL with the same solvent; add 2.7 mL of *phosphoric acid R*; adjust to pH 2.3, if necessary, with *ethanolamine R*; filter and degas;
— *mobile phase B*: mix 550 mL of mobile phase A with 450 mL of *acetonitrile R*; warm the solution to a temperature of at least 20 °C in order to avoid precipitation (mixing of mobile phase A with acetonitrile is endothermic); filter and degas.

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

System suitability:
— *resolution*: inject 20 µL of the resolution solution and 20 µL of reference solution (b). Record the chromatogram of the resolution solution until the peak corresponding to the principal peak in the chromatogram obtained with reference solution (b) is clearly visible. In the chromatogram obtained with the resolution solution, identify the peaks due to porcine insulin and human insulin. The test is not valid unless the resolution between the peaks due to human insulin and porcine insulin is at least 1.2. If necessary, adjust the concentration of acetonitrile in the mobile phase until this resolution is achieved.
— *linearity*: inject 20 µL each of reference solutions (b) and (c). The test is not valid unless the area of the principal peak in the chromatogram obtained with reference solution (b) is 10 ± 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c). If this test fails, adjust the injection volume to between 10 µL and 20 µL, in order that the responses are within the linearity range of the detector.

Injection 20 µL of the test solution.

Calculate the content of porcine insulin $C_{256}H_{381}N_{65}O_{76}S_6$ plus A21 desamido porcine insulin from the area of the principal peak and the area of the peak due to A21 desamido porcine insulin in the chromatograms obtained with the test solution and reference solution (b) and the declared content of porcine insulin plus A21 desamido porcine insulin in *porcine insulin CRS*.

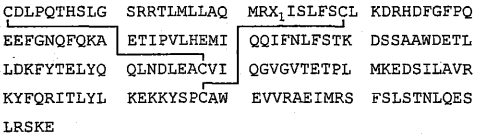
STORAGE

In an airtight container, protected from light, at −20 °C until released by the manufacturer. When thawed, insulin may be stored at 5 ± 3 °C and used for manufacturing preparations within a short period of time. To avoid absorption of humidity from the air during weighing, the insulin must be at room temperature.

Ph Eur

Interferon Alfa-2 Concentrated Solution

(Ph. Eur. monograph 1110)



Action and use

Cytokine.

Preparation

Interferon Alfa-2a Injection

Ph Eur

DEFINITION

Interferon alfa-2 concentrated solution is a solution of a protein that is produced according to the information coded by the alfa-2 sub-species of interferon alfa gene and that exerts non-specific antiviral activity, at least in homologous cells, through cellular metabolic processes involving synthesis of both ribonucleic acid and protein. Interferon alfa-2 concentrated solution also exerts antiproliferative activity. Different types of alfa-2 interferon, varying in the amino acid residue at position 23, are designated by a letter in lower case.

Designation	Residue at position 23 (X_1)
alfa-2a	Lys
alfa-2b	Arg

This monograph applies to interferon alfa-2a and -2b concentrated solutions.

The potency of interferon alfa-2 concentrated solution is not less than 1.4×10^8 IU per milligram of protein. Interferon alfa-2 concentrated solution contains not less than 2×10^8 IU of interferon alfa-2 per millilitre.

PRODUCTION

Interferon alfa-2 concentrated solution is produced by a method based on recombinant DNA (rDNA) technology using bacteria as host cells. It is produced under conditions designed to minimise microbial contamination of the product.

Interferon alfa-2 concentrated solution complies with the following additional requirements.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell- or vector-derived DNA
The limit is approved by the competent authority.

CHARACTERS

A clear, colourless or slightly yellowish liquid.

IDENTIFICATION

A. It shows the expected biological activity (see Assay).

B. Examine by isoelectric focusing.

Test solution Dilute the preparation to be examined with *water R* to a protein concentration of 1 mg/mL.

Reference solution Prepare a 1 mg/mL solution of *interferon alfa-2b CRS* in *water R*.

Isoelectric point calibration solution pI range 3.0 to 10.0
Prepare and use according to the manufacturer's instructions.

Use a suitable apparatus connected with a recirculating temperature controlled water-bath set at 10 °C and gels for isoelectric focusing with a pH gradient from 3.5 to 9.5. Operate the apparatus in accordance with the manufacturer's instructions. Use as the anode solution *phosphoric acid R* (98 g/L H_3PO_4) and as the cathode solution *1 M sodium hydroxide*. Samples are applied to the gel by filter papers. Place sample application filters on the gel close to the cathode.

Apply 15 µL of the test solution and 15 µL of the reference solution. Start the isoelectric focusing at 1500 V and 50 mA. Turn off the power after 30 min, remove the application filters and reconnect the power supply for 1 h. Keep the power constant during the focusing process. After focusing, immerse the gel in a suitable volume of a solution containing 115 g/L of *trichloroacetic acid R* and 34.5 g/L of *sulfosalicylic acid R* in *water R* and agitate the container gently for 60 min. Transfer the gel to a mixture of 32 volumes of *glacial acetic acid R*, 100 volumes of *anhydrous ethanol R* and 268 volumes of *water R*, and soak for 5 min. Immerse the gel for 10 min in a staining solution prewarmed to 60 °C in which 1.2 g/L of *acid blue 83 R* has been added to the previous mixture of glacial acetic acid, ethanol and water. Wash the gel in several containers with the previous mixture of glacial acetic acid, ethanol and water and keep the gel in this mixture until the background is clear (12 h to 24 h). After adequate destaining, soak the gel for 1 h in a 10 per cent *V/V* solution of *glycerol R* in the previous mixture of glacial acetic acid, ethanol and water.

The principal bands of the electropherogram obtained with the test solution correspond in position to the principal bands of the electropherogram obtained with the reference solution. Plot the migration distances of the isoelectric point markers versus their isoelectric points and determine the isoelectric points of the principal components of the test solution and the reference solution. They do not differ by more than 0.2 pI units. The test is not valid unless the isoelectric point markers are distributed along the entire length of the gel and the isoelectric points of the principal bands in the electropherogram obtained with the reference solution are between 5.8 and 6.3.

C. Examine the electropherograms obtained under reducing conditions in the test for impurities of molecular masses differing from that of interferon alfa-2. The principal band in the electropherogram obtained with test solution (a) corresponds in position to the principal band in the electropherogram obtained with reference solution (a).

D. Examine by peptide mapping.

Test solution Dilute the preparation to be examined in *water R* to a protein concentration of 1.5 mg/mL. Transfer 25 µL to a polypropylene or glass tube of 1.5 mL capacity. Add 1.6 µL of 1 M phosphate buffer solution pH 8.0 R, 2.8 µL of a freshly prepared 1.0 mg/mL solution of *trypsin for peptide mapping R* in *water R* and 3.6 µL of *water R* and mix vigorously. Cap the tube and place it in a water-bath at 37 °C for 18 h, then add 100 µL of a 573 g/L solution of *guanidine hydrochloride R* and mix well. Add 7 µL of 154.2 g/L solution of *dithiothreitol R* and mix well. Place the capped tube in boiling water for 1 min. Cool to room temperature.

Reference solution Prepare at the same time and in the same manner as for the test solution but use a 1.5 mg/mL solution of *interferon alfa-2b CRS* in *water R*.

Examine by liquid chromatography (*2.2.29*).

The chromatographic procedure may be carried out using:
- a stainless steel column 0.10 m long and 4.6 mm in internal diameter packed with *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm,
- as mobile phase at a flow rate of 1.0 mL/min:

Mobile phase A Dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water R*,

Mobile phase B To 100 mL of *water R* add 1 mL of *trifluoroacetic acid R* and dilute to 1000 mL with *acetonitrile for chromatography R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Comment
0 - 8	100	0	isocratic
8 - 68	100 → 40	0 → 60	linear gradient
68 - 72	40	60	isocratic
72 - 75	40 → 100	60 → 0	linear gradient
75 - 80	100	0	re-equilibration

- as detector a spectrophotometer set at 214 nm,

maintaining the temperature of the column at 30 °C.

Equilibrate the column with mobile phase A for at least 15 min.

Inject 100 µL of the test solution and 100 µL of the reference solution. The test is not valid unless the chromatogram obtained with each solution is qualitatively similar to the chromatogram of interferon alfa-2 digest supplied with *interferon alfa-2b CRS*. The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

TESTS

Impurities of molecular masses differing from that of interferon alfa-2
Examine by SDS polyacrylamide gel electrophoresis (*2.2.31*). The test is performed under both reducing and non-reducing conditions, using resolving gels of 14 per cent acrylamide and silver staining as the detection method.

Sample buffer (non-reducing conditions) Mix equal volumes of *water R* and *concentrated SDS-PAGE sample buffer R*.

Sample buffer (reducing conditions) Mix equal volumes of *water R* and *concentrated SDS-PAGE sample buffer for reducing conditions R* containing 2-mercaptoethanol as the reducing agent.

Test solution (a) Dilute the preparation to be examined in sample buffer to a protein concentration of 0.5 mg/mL.

Test solution (b) Dilute 0.20 mL of test solution (a) to 1 mL with sample buffer.

Reference solution (a) Prepare a 0.625 mg/mL solution of *interferon alfa-2b CRS* in sample buffer.

Reference solution (b) Dilute 0.20 mL of reference solution (a) to 1 mL with sample buffer.

Reference solution (c) Dilute 0.20 mL of reference solution (b) to 1 mL with sample buffer.

Reference solution (d) Dilute 0.20 mL of reference solution (c) to 1 mL with sample buffer.

Reference solution (e) Dilute 0.20 mL of reference solution (d) to 1 mL with sample buffer.

Reference solution (f) Use a solution of molecular mass standards suitable for calibrating SDS-PAGE gels in the range 15 kDa to 67 kDa.

Place test and reference solutions, contained in covered test-tubes, on a water-bath for 2 min.

Apply 10 µL of reference solution (f) and 50 µL of each of the other solutions to the stacking gel wells. Perform the electrophoresis under the conditions recommended by the manufacturer of the equipment. Detect proteins in the gel by silver staining.

The test is not valid unless: the validation criteria are met (*2.2.31*); a band is seen in the electropherogram obtained with reference solution (e); and a gradation of intensity of staining is seen in the electropherograms obtained, respectively, with test solution (a) and test solution (b) and with reference solutions (a) to (e).

The electropherogram obtained with test solution (a) under reducing conditions may show, in addition to the principal band, less intense bands with molecular masses lower than the principal band. No such band is more intense than the principal band in the electropherogram obtained with reference solution (d) (1.0 per cent) and not more than 3 such bands are more intense than the principal band in the electropherogram obtained with reference solution (e) (0.2 per cent).

The electropherogram obtained with test solution (a) under non-reducing conditions may show, in addition to the principal band, less intense bands with molecular masses higher than the principal band. No such band is more intense than the principal band in the electropherogram obtained with reference solution (d) (1.0 per cent) and not more than 3 such bands are more intense than the principal band in the electropherogram obtained with reference solution (e) (0.2 per cent).

Related proteins

Examine by liquid chromatography (*2.2.29*).

Test solution Dilute the preparation to be examined with *water R* to a protein concentration of 1 mg/mL.

0.25 per cent m/m hydrogen peroxide solution Dilute *dilute hydrogen peroxide solution R* in *water R* in order to obtain a 0.25 per cent *m/m* solution.

Reference solution To a volume of the test solution, add a suitable volume of 0.25 per cent *m/m* hydrogen peroxide solution to give a final hydrogen peroxide concentration of 0.005 per cent *m/m*, and allow to stand at room temperature for 1 h, or for the length of time that will generate about 5 per cent oxidised interferon. Add 12.5 mg of *L-methionine R* per millilitre of solution. Allow to stand at room temperature for 1 h. Store the solutions for not longer than 24 h at a temperature of 2-8 °C.

The chromatographic procedure may be carried out using:

— a stainless steel column 0.25 m long and 4.6 mm in internal diameter packed with *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm,

— as mobile phase at a flow rate of 1.0 mL/min:

Mobile phase A To 700 mL of *water R* add 2 mL of *trifluoroacetic acid R* and 300 mL of *acetonitrile for chromatography R*,

Mobile phase B To 200 mL of *water R* add 2 mL of *trifluoroacetic acid R* and 800 mL of *acetonitrile for chromatography R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Comment
0 - 1	72	28	isocratic
1 - 5	72 → 67	28 → 33	linear gradient
5 - 20	67 → 63	33 → 37	linear gradient
20 - 30	63 → 57	37 → 43	linear gradient
30 - 40	57 → 40	43 → 60	linear gradient
40 - 42	40	60	isocratic
42 - 50	40 → 72	60 → 28	linear gradient
50 - 60	72	28	re-equilibration

— as detector a spectrophotometer set at 210 nm.

Equilibrate the column with the mobile phases in the initial gradient ratio for at least 15 min. Inject 50 µL of each solution.

In the chromatograms obtained, interferon alfa-2 elutes at a retention time of about 20 min. In the chromatogram obtained with the reference solution a peak related to oxidised interferon appears at a retention time of about 0.9 relative to the principal peak. The test is not valid unless the resolution between the peaks due to oxidised interferon and interferon is at least 1.0. Consider only the peaks whose retention time is 0.7 to 1.4 relative to that of the principal peak. In the chromatogram obtained with the test solution, the area of any peak, apart from the principal peak, is not greater than 3.0 per cent of the total area of all of the peaks. The sum of the areas of any peaks other than the principal peak is not greater than 5.0 per cent of the total area of all of the peaks.

Bacterial endotoxins (*2.6.14*)

Less than 100 IU in the volume that contains 1.0 mg of protein.

ASSAY

Protein

Test solution Dilute the preparation to be examined with *water R* to obtain a concentration of about 0.5 mg/mL of interferon alfa-2.

Reference solutions Prepare a stock solution of 0.5 mg/mL of *bovine albumin R*. Prepare 8 dilutions of the stock solution containing between 3 µg/mL and 30 µg/mL of *bovine albumin R*.

Prepare 30-fold and 50-fold dilutions of the test solution. Add 1.25 mL of a mixture prepared the same day by combining 2.0 mL of a 20 g/L solution of *copper sulfate pentahydrate R* in *water R*, 2.0 mL of a 40 g/L solution of *sodium tartrate R* in *water R* and 96.0 mL of a 40 g/L solution of *sodium carbonate R* in *0.2 M sodium hydroxide* to test-tubes containing 1.5 mL of *water R* (blank), 1.5 mL of the different dilutions of the test solution or 1.5 mL of the reference solutions. Mix after each addition. After approximately 10 min, add to each test-tube 0.25 mL of a mixture of equal volumes of *water R* and *phosphomolybdotungstic reagent R*. Mix after each addition.

After approximately 30 min, measure the absorbance (*2.2.25*) of each solution at 750 nm using the blank as the compensation liquid. Draw a calibration curve from the absorbances of the 8 reference solutions and the corresponding protein contents and read from the curve the content of protein in the test solution.

Potency

The potency of interferon alfa-2 is estimated by comparing its effect to protect cells against a viral cytopathic effect with the same effect of the appropriate International Standard of human recombinant interferon alfa-2 or of a reference preparation calibrated in International Units.

The International Unit is the activity contained in a stated amount of the appropriate International Standard.

The equivalence in International Units of the International Standard is stated by the World Health Organization.

Carry out the assay by a suitable method, based on the following design.

Use, in standard culture conditions, an established cell line sensitive to the cytopathic effect of a suitable virus (a human diploid fibroblast cell line, free of microbial contamination, responsive to interferon and sensitive to encephalomyocarditis virus, is suitable).

The following cell cultures and virus have been shown to be suitable: MDBK cells (ATCC No. CCL22), or Mouse L cells (NCTC clone 929; ATCC No. CCL 1) as the cell culture and vesicular stomatitis virus VSV, Indiana strain (ATCC No. VR-158) as the infective agent; or A-549 cells (ATCC No. CCL-185) responsive to interferon as the cell culture, and encephalomyocarditis virus (ATCC No. VR-129B) as the infective agent.

Incubate in at least 4 series, cells with 3 or more different concentrations of the preparation to be examined and the reference preparation in a microtitre plate and include in each series appropriate controls of untreated cells. Choose the concentrations of the preparations such that the lowest concentration produces some protection and the largest concentration produces less than maximal protection against the viral cytopathic effect. Add at a suitable time the cytopathic virus to all wells with the exception of a sufficient number of wells in all series, which are left with uninfected control cells. Determine the cytopathic effect of virus quantitatively with a suitable method. Calculate the potency of the preparation to be examined by the usual statistical methods for a parallel line assay.

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.

The confidence limits of the estimated potency ($P = 0.95$) are not less than 64 per cent and not more than 156 per cent of the stated potency.

STORAGE

Store in an airtight container, protected from light, at or below −20 °C.

LABELLING

The label states:

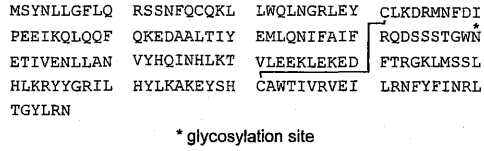
— the type of interferon (alfa-2a or alfa-2b),

— the type of production.

Ph Eur

Interferon Beta-1a Concentrated Solution

(Ph. Eur. monograph 1639)



$C_{908}H_{1406}N_{246}O_{252}S_7$ M_r approx. 22 500

Action and use

Cytokine.

Ph Eur

DEFINITION

Solution of a glycosylated protein having the same amino acid sequence and disulfide bridge and a similar glycosylation pattern as interferon beta produced by human diploid fibroblasts in response to viral infections and various other inducers. It exerts antiviral, antiproliferative and immunomodulatory activity.

Content

Minimum 0.20 mg of protein per millilitre.

Potency

Minimum 1.5×10^8 IU per milligram of protein.

It may contain buffer salts.

PRODUCTION

Interferon beta-1a concentrated solution is produced by a method based on recombinant DNA (rDNA) technology, using mammalian cells in culture.

Prior to release, the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell or vector-derived DNA

The limit is approved by the competent authority.

***N*-terminal truncated forms**

Examination for specific *N*-terminal truncated forms should be performed using a suitable technique such as *N*-terminal sequence determination. The limits are approved by the competent authority.

Dimer and related substances of higher molecular mass

Not more than the amount approved by the competent authority, using an appropriate validated liquid chromatography method.

CHARACTERS

Appearance

Clear or slightly opalescent, colourless or slightly yellowish liquid.

IDENTIFICATION

A. It shows the expected biological activity (see Assay).

B. Isoform distribution. Mass spectrometry (*2.2.43*).

Introduction of the sample Direct inflow of a desalted preparation to be examined or liquid chromatography-mass spectrometry combination.

Mode of ionisation Electrospray.

Signal acquisition Complete spectrum mode from 1100 to 2400.

Calibration Use myoglobin in the *m/z* range of 600-2400; set the instrument within validated instrumental settings and analyse the sample; the deviation of the measured mass does not exceed 0.02 per cent of the reported mass.

Interpretation of results A typical spectrum consists of 6 major glycoforms (A to F), which differ in their degree of sialylation and/or antennarity type as shown in Table 1639.-1.

Table 1639.-1.

MS peak	Glycoform*	Expected M_r	Sialylation level
A	2A2S1F	22 375	Disialylated
B	2A1S1F	22 084	Monosialylated
C	3A2S1F and/or 2A2S1F + 1 HexNacHex repeat	22 739	Disialylated
D	3A3S1F	23 031	Trisialylated
E	4A3S1F and/or 3A3S1F + 1 HexNacHex repeat	23 400	Trisialylated
F	2A0S1F	21 793	Non-sialylated

* 2A = biantennary complex type oligosaccharide; 3A = triantennary complex type oligosaccharide; 4A = tetraantennary complex type oligosaccharide; 0S = non-sialylated; 1S = monosialylated; 2S = disialylated; 3S = trisialylated; 1F = fucosylated.

Results The mass spectrum obtained with the preparation to be examined corresponds, with respect to the 6 major peaks, to the mass spectrum obtained with *interferon beta-1a CRS*.

C. Peptide mapping (*2.2.55*) and liquid chromatography (*2.2.29*).

Test solution Add 5 µL of a 242 g/L solution of *tris (hydroxymethyl) aminomethane R* and a volume of the preparation to be examined containing 20 µg of protein to a polypropylene tube of 0.5 mL capacity. Add 4 µL of a 1 mg/mL solution of *endoprotease LysC R* in *0.05 M tris-hydrochloride buffer solution pH 9.0 R*. Mix gently and incubate at 30 °C for 2 h. Add 10 µL of a 15.4 g/L solution of *dithiothreitol R*. Dilute the solution with the same volume of a 573 g/L solution of *guanidine hydrochloride R*. Incubate at 4 °C for 3-4 h.

Reference solution Prepare at the same time and in the same manner as for the test solution but using *interferon beta-1a CRS* instead of the preparation to be examined.

Precolumn:
- — *size*: *l* = 0.02 m, Ø = 2.1 mm;
- — *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Column:
- — *size*: *l* = 0.25 m, Ø = 2.1 mm;
- — *stationary phase*: spherical *octadecylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
- — *mobile phase A*: dilute 1 mL of *trifluoroacetic acid R* to 1000 mL with *water for chromatography R*;
- — *mobile phase B*: dilute 1 mL of *trifluoroacetic acid R* in 700 mL of *acetonitrile R1*, then dilute to 1000 mL with *water for chromatography R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	100 → 64	0 → 36
30 - 45	64 → 55	36 → 45
45 - 50	55 → 40	45 → 60
50 - 70	40 → 0	60 → 100
70 - 83	0	100
83 - 85	0 → 100	100 → 0

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection Volume that contains 20 µg of digested protein.

System suitability The chromatogram obtained with the reference solution is qualitatively similar to the chromatogram of interferon beta-1a digest supplied with *interferon beta-1a CRS*.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

TESTS

Impurities of molecular masses differing from that of interferon beta-1a

Polyacrylamide gel electrophoresis (*2.2.31*) under reducing conditions.

Resolving gel 12 per cent acrylamide.

Concentrated sample buffer *concentrated SDS-PAGE sample buffer for reducing conditions R* containing 2-mercaptoethanol as the reducing agent.

Sample buffer Mixture of equal volumes of *concentrated SDS-PAGE sample buffer for reducing conditions R* and *water R*.

Test solution (a) Concentrate the preparation to be examined using a suitable method to obtain a protein concentration of 1.5 mg/mL.

Test solution (b) Mixture of equal volumes of test solution (a) and the concentrated sample buffer.

Test solution (c) Dilute test solution (a) to obtain a protein concentration of 0.6 mg/mL. Mix equal volumes of this solution and the concentrated sample buffer.

Test solution (d) Mix 8 µL of test solution (c) and 40 µL of the sample buffer.

Test solution (e) Mix 15 µL of test solution (d) and 35 µL of the sample buffer.

Test solution (f) Mix 18 µL of test solution (e) and 18 µL of the sample buffer.

Test solution (g) Mix 12 µL of test solution (f) and 12 µL of the sample buffer.

Reference solution Solution of relative molecular mass markers suitable for calibrating SDS-PAGE gels in the range of 15-67 kDa. Dissolve in the sample buffer.

Sample treatment Boil for 3 min.

Application 20 µL of test solutions (b) to (g) and the reference solution.

Detection Coomassie staining, carried out as follows: immerse the gel in *Coomassie staining solution R1* at 33-37 °C for 90 min with gentle shaking, then remove the staining solution; destain the gel with a large excess of a mixture of 1 volume of *glacial acetic acid R*, 1 volume of *2-propanol R* and 8 volumes of *water R*.

Apparent molecular masses Interferon beta-1a = about 23 000; underglycosylated interferon beta-1a = about 21 000; deglycosylated interferon beta-1a = about 20 000; interferon beta-1a dimer = about 46 000.

Identification of bands Use the electropherogram provided with *interferon beta-1a CRS*.

System suitability:
— the validation criteria are met (*2.2.31*);
— a band is seen in the electropherogram obtained with test solution (g);
— a gradation of intensity of staining is seen in the electropherograms obtained with test solutions (b) to (g).

Limits:
— in the electropherogram obtained with test solution (c), the band corresponding to underglycosylated interferon beta-1a is not more intense than the principal band in the electropherogram obtained with test solution (e) (5 per cent);
— in the electropherogram obtained with test solution (b), the band corresponding to deglycosylated interferon beta-1a is not more intense than the principal band in the electropherogram obtained with test solution (e) (2 per cent); any other band corresponding to an impurity of a molecular mass lower than that of interferon beta-1a, apart from the band corresponding to underglycosylated interferon beta-1a is not more intense than the principal band in the electropherogram obtained with test solution (f) (1 per cent).

Oxidised interferon beta-1a

Maximum 6 per cent.

Use the chromatogram obtained with the test solution in identification C. Locate the peaks due to the peptide fragment comprising amino acids 34-45 and its oxidised form using the chromatogram of oxidised interferon beta-1a digest supplied with *interferon beta-1a CRS*.

Calculate the percentage of oxidation of interferon beta-1a using the following expression:

$$\frac{A_{34-45ox}}{A_{34-45} + A_{34-45ox}} \times 100$$

$A_{34-45ox}$ = area of the peak due to the oxidised peptide fragment 34-45;

A_{34-45} = area of the peak due to the peptide fragment 34-45.

Bacterial endotoxins (*2.6.14*)

Less than 0.7 IU in the volume that contains 1×10^6 IU of interferon beta-1a, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

ASSAY

Protein

Liquid chromatography (*2.2.29*). Prepare 3 independent dilutions for each solution.

Test solution Dilute the preparation to be examined to obtain a concentration of 100 µg/mL.

Reference solution Dissolve the contents of a vial of *interferon beta-1a CRS* to obtain a concentration of 100 µg/mL.

Precolumn:
— *size*: *l* = 0.02 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Column:
— *size*: *l* = 0.25 m, Ø = 2.1 mm;
— *stationary phase*: *end-capped butylsilyl silica gel for chromatography R* (5 µm) with a pore size of 30 nm.

Mobile phase:
— *mobile phase A*: 0.1 per cent *V/V* solution of *trifluoroacetic acid R*;
— *mobile phase B*: to 300 mL of *water for chromatography R*, add 1 mL of *trifluoroacetic acid R* and dilute to 1000 mL with *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 20	100 → 0	0 → 100
20 - 25	0	100
25 - 26	0 → 100	100 → 0
26 - 40	100	0

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 50 µL.

Retention time Interferon beta-1a = about 20 min.

System suitability Reference solution:
— *symmetry factor*: 0.8 to 2.0 for the peak due to interferon beta-1a;
— *repeatability*: maximum relative standard deviation of 3.0 per cent between the peak areas obtained after injection of the 3 independent dilutions.

Calculate the content of interferon beta-1a ($C_{908}H_{1406}N_{246}O_{252}S_7$) taking into account the assigned content of $C_{908}H_{1406}N_{246}O_{252}S_7$ in *interferon beta-1a CRS*.

Potency

The potency of interferon beta-1a is estimated by comparing its ability to protect cells against a viral cytopathic effect with the same ability of the appropriate International Standard of human recombinant interferon beta-1a or of a reference preparation calibrated in International Units.

The International Unit is the activity contained in a stated amount of the appropriate International Standard.
The equivalence in International Units of the International Standard is stated by the World Health Organization.

Carry out the assay using a suitable method, based on the following design.

Use, in standard culture conditions, an established cell line sensitive to the cytopathic effect of a suitable virus and responsive to interferon. The cell cultures and viruses that have been shown to be suitable include the following:
— WISH cells (ATCC No. CCL-25) and vesicular stomatitis virus VSV, Indiana strain (ATCC No. VR-158) as infective agent;
— A549 cells (ATCC No. CCL-185) and encephalomyocarditis virus EMC (ATCC No. VR-129B) as infective agent.

Incubate in at least 4 series, cells with 3 or more different concentrations of the preparation to be examined and the reference preparation in a microtitre plate and include in each series appropriate controls of untreated cells. Choose the concentrations of the preparations such that the lowest concentration produces some protection and the largest concentration produces less than maximal protection against the viral cytopathic effect. Add at a suitable time the cytopathic virus to all wells with the exception of a sufficient number of wells in all series, which are left with uninfected control cells. Determine the cytopathic effect of the virus quantitatively with a suitable method. Calculate the potency of the preparation to be examined by the usual statistical methods (for example, *5.3*).

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency.
The confidence limits (P = 0.95) are not less than

64 per cent and not more than 156 per cent of the estimated potency.

STORAGE

In an airtight container, protected from light, at a temperature below −70 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

The label states:

— the interferon beta-1a content, in milligrams per millilitre;
— the antiviral activity, in International Units per millilitre;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Interferon Gamma-1b Concentrated Solution

(Ph. Eur. monograph 1440)

$C_{734}H_{1166}N_{204}O_{216}S_5$ 16 465

Action and use

Cytokine.

Ph Eur

DEFINITION

Interferon gamma-1b concentrated solution is a solution of the *N*-terminal methionyl form of interferon gamma, a protein which is produced and secreted by human antigen-stimulated T lymphocytes in response to viral infections and various other inducers. It has specific immunomodulatory properties, such as potent phagocyte-activating effects.
The protein consists of non-covalent dimers of two identical monomers. The formula of the monomer is as follows:

```
                                                  M
QDPYVKEAEN  LKKYFNAGHS  DVADNGTLFL  GILKNWKEES
DRKIMQSQIV  SFYFKLFKNF  KDDQSIQKSV  ETIKEDMNVK
FFNSNKKKRD  DFEKLTNYSV  TDLNVQRKAI  HELIQVMAEL
SPAAKTGKRK  RSQMLFRGR
```

The potency of interferon gamma-1b is not less than 20×10^6 IU per milligram of protein. Interferon gamma-1b concentrated solution contains not less than 30×10^6 IU of interferon gamma-1b per millilitre.

PRODUCTION

Interferon gamma-1b concentrated solution is produced by a method based on recombinant DNA technology, using bacteria as host-cells. It is produced under conditions designed to minimise microbial contamination.

Interferon gamma-1b concentrated solution complies with the following additional requirements.

Host-cell derived proteins

The limit is approved by the competent authority.

Host-cell- and vector-derived DNA

The limit is approved by the competent authority.

CHARACTERS

A clear, colourless or slightly yellowish liquid.

IDENTIFICATION

A. It shows the expected biological activity when tested as prescribed in the assay.

B. Examine the electropherograms obtained in the test for impurities of molecular masses differing from that of interferon gamma-1b. The principal bands in the electropherogram obtained with the test solution correspond in position to the principal bands in the electropherogram obtained with reference solution (a).

C. Examine by peptide mapping.

Solution A Prepare a solution containing 1.2 g/L of *tris (hydroxymethyl) aminomethane R*, 8.2 g/L of *anhydrous sodium acetate R*, 0.02 g/L of *calcium chloride R* and adjust to pH 8.3 with *dilute acetic acid R*. Add *polysorbate 20 R* to a concentration of 0.1 per cent *V/V*.

Test solution Desalt a volume of the preparation to be examined containing 1 mg of protein by a suitable procedure. For example, filter in a microcentrifuge tube and reconstitute with 500 µL of solution A. Add 10 µL of a freshly prepared 1 mg/mL solution of *trypsin for peptide mapping R* in *water R* and mix gently by inversion. Incubate at 30 °C to 37 °C for 24 h, add 100 µL of *phosphoric acid R* per millilitre of digested sample and mix by inversion.

Reference solution Dilute *interferon gamma-1b CRS* in *water R* to obtain a concentration of 1 mg/mL. Prepare as for the test solution, ensuring that all procedures are carried out simultaneously and under identical conditions.

Examine by liquid chromatography (*2.2.29*).

The chromatographic procedure may be carried out using:
— a stainless steel column, 0.15 m long and 4.6 mm in internal diameter packed with *octadecylsilyl silica gel for chromatography R* (10 µm),
— as mobile phase at a flow rate of 1.0 mL/min:

Mobile phase A (*0.05 M sodium phosphate buffer solution pH 3.3*). Solution I: dissolve 7.80 g of *sodium dihydrogen phosphate R* in *water R* and dilute to 1000.0 mL with the same solvent. Solution II: dilute 0.33 mL of *phosphoric acid R* to 100.0 mL with *water R*. Mix 920 mL of solution I and 80 mL of solution II. Adjust the pH if necessary,

Mobile phase B *Acetonitrile for chromatography R*, with the following elution conditions (if necessary, the gradient may be modified to improve the separation of the digest):

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 30	100 → 80	0 → 20
30 - 50	80 → 60	20 → 40
50 - 51	60 → 30	40 → 70
51 - 59	30	70

— as detector a spectrophotometer set at 214 nm,

maintaining the temperature of the column at 40 °C.

Equilibrate the column for at least 15 min at the initial elution composition. Carry out a blank run using the above-mentioned gradient.

Inject 100 µL of the test solution and 100 µL of the reference solution. The test is not valid unless the chromatogram obtained with each solution is qualitatively similar to the chromatogram of interferon gamma-1b digest supplied with *interferon gamma-1b CRS*. The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

D. Examine by *N*-terminal sequence analysis.

Use an automated solid-phase sequencer, operated in accordance with the manufacturer's instructions.

Equilibrate by a suitable procedure the equivalent of 100 µg of interferon gamma-1b in a 10 g/L solution of *ammonium hydrogen carbonate R*, pH 9.0.

Identify the phenylthiohydantoin (PTH)-amino acids released at each sequencing cycle by reverse-phase liquid chromatography. The procedure may be carried out using the column and reagents recommended by the manufacturer of the sequencing equipment for the separation of PTH-amino acids.

The separation procedure is calibrated using:
- — the mixture of PTH-amino acids provided by the manufacturer, with the gradient conditions adjusted as indicated to achieve optimum resolution of all amino acids,
- — a sample from a blank sequencing cycle, obtained as recommended by the equipment manufacturer.

The first fifteen amino acids are:

Met-Gln-Asp-Pro-Tyr-Val-Lys-Glu-Ala-Glu-Asn-Leu-Lys-Lys-Tyr.

TESTS

Appearance

The preparation to be examined is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

pH (*2.2.3*)

The pH of the preparation to be examined is 4.5 to 5.5.

Covalent dimers and oligomers

Not greater than 2 per cent, determined by size-exclusion chromatography (*2.2.30*).

Test solution Dilute the preparation to be examined with the mobile phase to a protein concentration of 0.1 mg/mL.

Reference solution (a) Dilute *interferon gamma-1b CRS* with the mobile phase to a protein concentration of 0.1 mg/mL.

Reference solution (b) Prepare a mixture of the following molecular mass standards: bovine albumin, ovalbumin, trypsinogen, lysozyme, at a concentration of 0.1 mg/mL to 0.2 mg/mL for each standard.

The chromatographic procedure may be carried out using:
- — a stainless steel column 0.3 m long and 7.8 mm in internal diameter packed with *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the molecular weight range of 10 000 to 500 000 (5 µm),
- — as mobile phase at a flow rate of 1.0 mL/min a mixture prepared as follows (0.2 M sodium phosphate buffer solution pH 6.8). Solution I: dissolve 31.2 g of *sodium dihydrogen phosphate R* and 1.0 g of *sodium dodecyl sulfate R* in *water R* and dilute to 1000.0 mL with the same solvent. Solution II: dissolve 28.4 g of *anhydrous disodium hydrogen phosphate R* and 1.0 g of *sodium dodecyl sulfate R* in *water R* and dilute to 1000.0 mL with the same solvent. Mix 450 mL of solution I and 550 mL of solution II. Adjust the pH if necessary,
- — as detector a spectrophotometer set at 210 nm to 214 nm.

Inject 200 µL of each solution. The test is not valid unless: the molecular mass standards in reference solution (b) are well separated; the retention time of the principal peak in the chromatogram obtained with reference solution (a) is between the retention time of trypsinogen and lysozyme in the chromatogram obtained with reference solution (b).

Compare the chromatograms obtained with the test solution and with reference solution (a). There are no additional shoulders or peaks in the chromatogram obtained with the test solution compared with the chromatogram obtained with reference solution (a).

Calculate the percentage content of covalent dimers and oligomers.

Monomer and aggregates

Examine by size-exclusion chromatography (*2.2.30*).

The content of monomer and aggregates is not greater than 2 per cent.

Solution A Prepare a solution of the following composition: 0.59 g/L of *succinic acid R* and 40 g/L of *mannitol R*, adjusted to pH 5.0 with *sodium hydroxide solution R*.

Test solution Dilute the preparation to be examined with solution A to a protein concentration of 1 mg/mL.

Reference solution Dilute *interferon gamma-1b CRS* with solution A to a protein concentration of 1 mg/mL.

Resolution solution Prepare 500 µL of a mixture consisting of 0.04 mg/mL of *bovine albumin R* and 0.2 mg/mL of *interferon gamma-1b CRS* in solution A. Use this solution within 24 h of preparation.

The chromatographic procedure may be carried out using:
- — a stainless steel column 0.3 m long and 7.8 mm in internal diameter packed with *hydrophilic silica gel for chromatography R*, of a grade suitable for fractionation of globular proteins in the molecular weight range of 10 000 - 300 000 (5 µm),
- — as mobile phase at a flow rate of 0.8 mL/min a 89.5 g/L solution of *potassium chloride R* (1.2 M),
- — as detector a spectrophotometer set at 214 nm.

Inject 20 µL of the resolution solution. In the chromatogram obtained, the retention time of the principal peak, corresponding to the native interferon gamma-1b dimer, is about 10 min. Bovine albumin elutes at a relative retention time of about 0.85, relative to the main peak. The test is not valid unless the resolution between the peaks due to bovine albumin and interferon gamma-1b is at least 1.5.

Inject 20 µL of the test solution and 20 µL of the reference solution. The chromatograms obtained show principal peaks with identical retention times. Calculate the percentage content of monomer and aggregates from the peak area of the monomer peak and of peaks which elute prior to the native interferon gamma-1b peak in the chromatogram obtained with the test solution, by the normalisation procedure, disregarding any peak due to the solvent.

Deamidated and oxidised forms and heterodimers

Examine by liquid chromatography (*2.2.29*). The content of deamidated and oxidised forms is not greater than 10 per cent. The content of heterodimers is not greater than 3 per cent.

Test solution Dilute the preparation to be examined with *water R* to a protein concentration of 1 mg/mL.

Reference solution (a) Dilute *interferon gamma-1b CRS* with *water R* to a protein concentration of 1 mg/mL.

Reference solution (b) Dissolve the contents of a vial of *interferon gamma-1b for system suitability CRS* in *water R* to obtain a protein concentration of about 1 mg/mL.

The chromatographic procedure may be carried out using:
- — a stainless steel column 0.075 m long and 7.5 mm in internal diameter packed with an appropriate hydrophilic polymethacrylate, strong cation-exchange gel (10 µm, 100 nm),
- — as mobile phase at a flow rate of 1.2 mL/min:

Mobile phase A *(0.05 M ammonium acetate buffer pH 6.5).* A 3.86 g/L solution of *ammonium acetate R*, adjusted to pH 6.5 with *dilute acetic acid R*,

Mobile phase B *(1.2 M ammonium acetate buffer pH 6.5).* A 92.5 g/L solution of *ammonium acetate R*, adjusted to pH 6.5 with *dilute acetic acid R*,

with the following elution conditions (if necessary, the slope of the gradient may be modified to improve the separation).

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 1	100	0
2 - 30	100 → 0	0 → 100
31 - 35	0	100

— as detector a spectrophotometer set at 280 nm,

maintaining the temperature of the column at 35 °C.

Inject 25 µL of reference solution (b). In the chromatogram obtained, the retention time of the principal peak is about 26 min. Deamidated and oxidised forms co-elute at a relative retention time of about 0.95, relative to the principal peak. The test is not valid unless the resolution, defined by the ratio of the height of the peak corresponding to the deamidated and oxidised forms to the height above the baseline of the valley separating the two peaks, is at least 1.2.

Inject 25 µL of the test solution and 25 µL of reference solution (a). The chromatograms obtained show principal peaks with identical retention times. Calculate the percentage content of deamidated and oxidised interferon gamma-1b as a percentage of the area of the main peak. Heterodimers have relative retention times of 0.7 and 0.85 relative to the main peak. Calculate the percentage of heterodimers as a percentage of the sum of the areas of all peaks.

Impurities of molecular masses differing from that of interferon gamma-1b

Examine by polyacrylamide gel electrophoresis (*2.2.31*). The test is performed under both reducing and non-reducing conditions, using resolving gels of 15 per cent acrylamide and silver staining as the detection method.

Sample buffer (non-reducing conditions) Dissolve 3.78 g of *tris (hydroxymethyl) aminomethane R*, 10.0 g of *sodium dodecyl sulfate R* and 0.100 g of *bromophenol blue R* in *water R*. Add 50.0 mL of *glycerol R* and dilute to 80 mL with *water R*. Adjust the pH to 6.8 with *hydrochloric acid R* and dilute to 100 mL with *water R*.

Sample buffer (reducing conditions) Dissolve 3.78 g of *tris (hydroxymethyl) aminomethane R*, 10.0 g of *sodium dodecyl sulfate R* and 0.100 g of *bromophenol blue R* in *water R*. Add 50.0 mL of *glycerol R* and dilute to 80 mL with *water R*. Adjust the pH to 6.8 with *hydrochloric acid R* and dilute to 100 mL with *water R*. Immediately before use, add *dithiothreitol R* to a final concentration of 250 mM.

Test solution Dilute the preparation to be examined in *water R* to a protein concentration of 1 mg/mL. Dilute 150 µL of the solution with 38 µL of sample buffer.

Reference solution (a) Prepare in the same manner as for the test solution, but using *interferon gamma-1b CRS* instead of the preparation to be examined.

Reference solution (b) (5 ng control) Mix 50 µL of a 0.01 mg/mL solution of *bovine albumin R* with 2000 µL of *water R* and 450 µL of sample buffer.

Reference solution (c) (2 ng control) Mix 20 µL of a 0.01 mg/mL solution of *bovine albumin R* with 2000 µL of *water R* and 450 µL of sample buffer.

Reference solution (d) Use a solution of molecular mass standards suitable for calibrating SDS-polyacrylamide gels in the range of 10 kDa to 70 kDa.

Leave each solution, contained in a test tube, at ambient temperature for 15 min, then store on ice.

Apply 25 µL of each solution to the stacking gel wells. Perform the electrophoresis under the conditions recommended by the manufacturer of the equipment. Detect proteins in the gel by silver staining.

The test is not valid unless: the validation criteria are met (*2.2.31*); a band is seen in the electropherograms obtained with reference solutions (b) and (c).

The principal band in the electropherogram obtained with the test solution is similar in intensity to the principal band in the electropherogram obtained with reference solution (a). In the electropherogram obtained with the test solution, no significant bands are observed that are not present in the electropherogram obtained with reference solution (a) (0.01 per cent). A significant band is defined as any band whose intensity is greater than or equal to that of the band in the electropherogram obtained with reference solution (c).

Norleucine

Not more than 0.2 mole of norleucine per mole of interferon gamma-1b, determined by amino acid analysis.

Test solution Add 2.5 mL of the preparation to be examined onto a column suitable for the desalting of proteins previously equilibrated with 25 mL of a 10 per cent *V/V* solution of *acetic acid R*. Elute the sample with another 2.5 mL of a 10 per cent *V/V* solution of *acetic acid R*. Determine the protein content by measuring the absorbance of this solution as described under Protein, in the Assay section. Pipette a volume containing the equivalent of 100 µg of interferon gamma-1b into each of three reaction vials. Evaporate to dryness under reduced pressure.

Perform the hydrolysis of the three samples as follows. Add to each reaction vial 200 µL of a 50 per cent *V/V* solution of *hydrochloric acid R* containing 1 per cent *V/V* of *phenol R*, evacuate the samples, purge with nitrogen and hydrolyse in the gas phase. Heat the reaction vials at 110 °C for 22 h. After hydrolysis evaporate to dryness under reduced pressure.

Perform the derivatisation of the samples as follows. Prepare immediately before use a mixture consisting of two volumes of *ethanol R*, one volume of *water R* and one volume of *triethylamine R*. Add 50 µL of this solution to each reaction vial and shake lightly. Evaporate to dryness under reduced pressure. Add to each vial 50 µL of a mixture consisting of 7 volumes of *ethanol R*, one volume of *water R*, one volume of *triethylamine R* and one volume of *phenyl isothiocyanate R*. Shake lightly and allow to stand at room temperature for about 15 min. Evaporate to dryness under reduced pressure. Reconstitute the samples in 250 µL of mobile phase A.

Norleucine stock solution Prepare a 250 nmol/mL solution of *DL-norleucine R* in *0.01 M hydrochloric acid. This solution may be kept for two months at 4 °C.*

Leucine stock solution Prepare a 250 nmol/mL solution of *leucine R* in *0.01 M hydrochloric acid. This solution may be kept at 4 °C for two months.*

Reference solution Mix 10 µL of norleucine stock solution with 100 µL of leucine stock solution in each of the three reaction vials. Evaporate to dryness under reduced pressure. Perform the derivatisation of the samples as described for the preparation of the test solution.

Examine by liquid chromatography (*2.2.29*).

The chromatographic procedure may be carried out using:
— a stainless steel column 0.15 m long and 3.9 mm in diameter packed with *octadecylsilyl silica gel for chromatography R* (4 µm),
— as mobile phase at a flow rate of 1.0 mL/min:

Mobile phase A Mix 70 volumes of a 19 g/L solution of *sodium acetate R* containing 0.05 per cent *V/V* of *triethylamine R* and adjusted to pH 6.4 with *dilute acetic acid R* and 30 volumes of mobile phase B,

Mobile phase B Mix 40 volumes of *water R* and 60 volumes of *acetonitrile R*,

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)	Comment
0 - 7	100	0	isocratic
7 - 7.1	100 → 0	0 → 100	linear gradient
7.1 - 10	0	100	washing step
10 - 10.1	0 → 100	100 → 0	linear gradient
10.1 - 15	100	0	re-equilibration

— as detector a spectrophotometer set at 254 nm,

maintaining the temperature of the column at 43 °C.

Inject 50 µL of each solution.

In the chromatograms obtained with the test solution, identify the peaks corresponding to leucine and norleucine. The retention time of norleucine is 6.2 min to 7 min.

Calculate the content of norleucine (in moles of norleucine per mole of interferon gamma-1b) from the peak areas of leucine and norleucine in the chromatograms obtained with the reference and test solutions, considering that there are 10 moles of leucine per mole of interferon gamma-1b.

Bacterial endotoxins (*2.6.14*)
Less than 5 IU in the volume that contains 20 × 10^6 IU of interferon gamma-1b.

ASSAY

Protein (*2.2.25*)
Dilute the substance to be examined in *water R* to obtain a concentration of 1 mg/mL. Record the absorbance spectrum between 220 nm and 340 nm. Measure the value at the absorbance maximum of 280 nm, after correction for any light scattering due to turbidity measured at 316 nm. Calculate the concentration of interferon gamma-1b using a specific absorbance value of 7.5.

Potency
The potency of interferon gamma-1b is estimated by evaluating the increase of the expression of human-leukocyte-antigen-DR (HLA-DR) due to the interferon gamma-1b present in test solutions during cultivation of the cells, and comparing this increase with the same effect of the appropriate International Standard of human recombinant interferon gamma or of a reference preparation calibrated in International Units.

The International Unit is the activity contained in a stated amount of the appropriate International Standard. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Carry out the assay by a suitable method, based on the following design.

Use COLO 205 cells under standard culture conditions. Trypsinise a 3- to 5-day-old flask of COLO 205 cells and prepare a cell suspension at a concentration of 1.0 × 10^6 cells/mL.

Add 100 µL of the dilution medium to all wells of a 96-well microtitre plate. Add an additional 100 µL of this solution to the wells designed for the blanks. Add 100 µL of each solution to be tested onto the plate and carry out a series of twofold dilution steps in order to obtain a standard curve. Then add 100 µL of the cell suspension to all wells and incubate the plate under appropriate conditions for cell cultivation.

After cultivation remove the growth medium and wash and fix cells to the plate. Add an antibody able to detect HLA-DR expressed due to the presence of interferon gamma-1b and incubate under appropriate conditions. After washing the plate, incubate with an antibody conjugated to a marker enzyme which is able to detect the anti-HLA-DR antibody. After this incubation step, wash the plate and add an appropriate substrate solution. Stop the reaction. Measure the absorbance of the solution and calculate the potency of the preparation to be examined by the usual statistical methods.

The estimated specific activity is not less than 80 per cent and not more than 125 per cent of the stated potency. The confidence limits (P = 0.95) are not less than 70 per cent and not more than 140 per cent of the estimated potency.

STORAGE

Store in an airtight container, protected from light and at a temperature of −70 °C.

Ph Eur

Iodine

(Ph. Eur. monograph 0031)

I_2 253.8 *7553-56-2*

Action and use
Antiseptic; antithyroid.

Preparations
Alcoholic Iodine Solution
Aqueous Iodine Oral Solution
Povidone-Iodine Eye Drops
Povidone-Iodine Mouthwash
Povidone-Iodine Solution

Ph Eur

DEFINITION

Content
99.5 per cent to 100.5 per cent of I.

CHARACTERS

Appearance
Greyish-violet, brittle plates or fine crystals with a metallic sheen.

Solubility
Very slightly soluble in water, very soluble in concentrated solutions of iodides, soluble in ethanol (96 per cent), slightly soluble in glycerol.

It volatilises slowly at room temperature.

IDENTIFICATION

A. Heat a few fragments in a test-tube. Violet vapour is evolved and a bluish-black crystalline sublimate is formed.

B. To a saturated solution add *starch solution R*. A blue colour is produced. Heat until decolourised. On cooling, the colour reappears.

TESTS

Solution S

Triturate 3.0 g with 20 mL of *water R*, filter, wash the filter with *water R* and dilute the filtrate to 30 mL with the same solvent. To the solution add 1 g of *zinc powder R*. When the solution is decolourised, filter, wash the filter with *water R* and dilute to 40 mL with the same solvent.

Bromides and chlorides

Maximum 250 ppm.

To 10 mL of solution S add 3 mL of *ammonia R* and 6 mL of *silver nitrate solution R2*. Filter, wash the filter with *water R* and dilute the filtrate to 20 mL with the same solvent. To 10 mL of the solution add 1.5 mL of *nitric acid R*. After 1 min, any opalescence in the solution is not more intense than that in a standard prepared at the same time by mixing 10.75 mL of *water R*, 0.25 mL of *0.01 M hydrochloric acid*, 0.2 mL of *dilute nitric acid R* and 0.3 mL of *silver nitrate solution R2*.

Non-volatile substances

Maximum 0.1 per cent.

Heat 1.00 g in a porcelain dish on a water-bath until the iodine has volatilised. Dry the residue at 100-105 °C. The residue weighs a maximum of 1 mg.

ASSAY

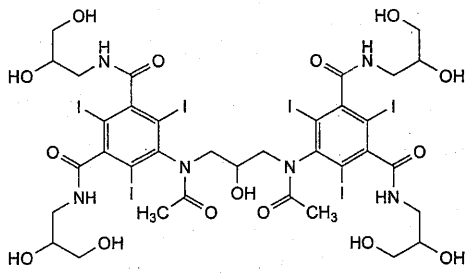
Introduce 0.200 g into a flask containing 1 g of *potassium iodide R* and 2 mL of *water R* and add 1 mL of *dilute acetic acid R*. When dissolution is complete, add 50 mL of *water R* and titrate with *0.1 M sodium thiosulfate*, using *starch solution R* as indicator.

1 mL of *0.1 M sodium thiosulfate* is equivalent to 12.69 mg of I.

Ph Eur

Iodixanol

(Ph. Eur. monograph 2215)



$C_{35}H_{44}I_6N_6O_{15}$ 1550 *92339-11-2*

Action and use

Iodinated contrast medium.

Ph Eur

DEFINITION

Mixture of stereoisomers of 5,5′-[(2-hydroxypropane-1,3-diyl)bis(acetylimino)]bis[*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide].

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder, hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *iodixanol CRS*.

B. Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (b).

Results The 3 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 3 principal peaks in the chromatogram obtained with reference solution (b).

TESTS

Solution S

Dissolve 5.0 g in *water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Heat solution S at about 98 °C for 30 min without boiling then allow to cool to room temperature. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Impurities E and H

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.250 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*.

Reference solution (b) Dissolve 5 mg of *iodixanol impurity E CRS* and 5 mg of *iodixanol impurity H CRS* in *water R* and dilute to 20.0 mL with the same solvent.

Reference solution (c) Mix 5.0 mL of the test solution with 5.0 mL of reference solution (b) and dilute to 50.0 mL with *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *aminopropylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: *acetonitrile R*, *water R* (50:50 *V/V*);
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	30	70
2 - 27	30 → 68	70 → 32

Flow rate 1.7 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities E and H.

Relative retention With reference to iodixanol (1st peak) (retention time = about 16 min): impurity E

(1st peak) = about 0.7; impurity E (2nd peak) = about 0.8; impurity H = about 1.4.

System suitability Reference solution (c):
— *resolution*: minimum 5.0 between the 1st peak due to impurity E and the 1st peak due to iodixanol.

Limits:
— *correction factor*: for the calculation of total content of impurity E, multiply the peak area of the 1st peak due to impurity E by 1.7;
— *impurity H*: not more than 0.6 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.6 per cent);
— *impurity E*: not more than 0.3 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.3 per cent).

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.250 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*.

Reference solution (b) Dissolve 25 mg of *iodixanol CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (c) Dissolve 5 mg of *iodixanol impurity C CRS* and 5 mg of *iopentol CRS* in *water R* and dilute to 10.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with *water R*.

Reference solution (d) Mix 5.0 mL of the test solution with 5.0 mL of reference solution (c) and dilute to 50.0 mL with *water R*.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
— *mobile phase A*: *water R*;
— *mobile phase B*: *acetonitrile R*, *water R* (50:50 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	94	6
2 - 32	94 → 80	6 → 20
32 - 72	80 → 0	20 → 100
72 - 82	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurity C and iopentol.

Relative retention With reference to iodixanol (1st peak) (retention time = about 27 min): iopentol (1st peak) = about 0.8; iopentol (2nd peak) = about 0.9; impurity C (1st peak) = about 1.04; overalkylated impurities (a group of peaks) = 1.33-1.70.

System suitability Reference solution (d):
— *resolution*: baseline separation between the 2 peaks due to iopentol;
— *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the 1st peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the 1st peak due to iodixanol.

Limits:
— *correction factor*: for the calculation of total content of impurity C, multiply the peak area of the 1st peak due to impurity C by 1.3;
— *impurity C*: not more than 0.4 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.4 per cent);
— *overalkylated impurities (such as impurity I)*: not more than the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (1.0 per cent);
— *unspecified impurities*: for each impurity, not more than 0.1 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than 1.5 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (1.5 per cent);
— *disregard limit*: 0.05 times the sum of the areas of the 2 principal peaks in the chromatogram obtained with reference solution (a) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Free aromatic amine
Maximum 500 ppm.

Test solution Transfer 0.200 g of the substance to be examined to a 25 mL volumetric flask and dissolve in 15.0 mL of *water R*.

Reference solution Dissolve 5.0 mg of *iohexol impurity J CRS* in *water R* and dilute to 5.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *water R*. Mix 10.0 mL of this solution with 5.0 mL of *water R* in a 25 mL volumetric flask.

Blank solution Transfer 15.0 mL of *water R* to a 25 mL volumetric flask.

In conducting the following steps, keep the flasks in iced water and protected as much as possible from light until all the reagents have been added.

Place the 3 flasks containing respectively the test solution, the reference solution and the blank solution in iced water, protected from light, for 5 min. Add 1.5 mL of *hydrochloric acid R1* and mix by swirling. Add 1.0 mL of a 20 g/L solution of *sodium nitrite R*, mix and allow to stand for 4 min. Add 1.0 mL of a 40 g/L solution of *sulfamic acid R*, swirl gently until gas liberation has ceased and allow to stand for 1 min. (*CAUTION: considerable pressure is produced*). Add 1.0 mL of a freshly prepared 3 g/L solution of *naphthylethylenediamine dihydrochloride R* in a mixture of 30 volumes of *water R* and 70 volumes of *propylene glycol R* and mix. Remove the flasks from the iced water, dilute to 25.0 mL with *water R*, mix and examine the solutions after 5 min. The solution obtained from the test solution is less coloured than the solution obtained from the reference solution. If the solution obtained from the test solution is about the same colour or darker than the solution obtained from the reference solution, proceed as follows. Concomitantly determine the absorbance (*2.2.25*) at 495 nm of the solution obtained from the test solution and the reference solution in 5 cm cells, using the blank solution as the compensation liquid. The absorbance of the solution

obtained from the test solution is not greater than that of the solution obtained from the reference solution.

Free iodine

Transfer 2.0 g to a glass-stoppered tube, add 20 mL of *water R*, 5 mL of *toluene R* and 5 mL of *dilute sulfuric acid R*, shake vigorously and allow the phases to separate: the toluene layer shows no red or pink colour.

Iodide

Maximum 10 ppm.

Dissolve 5.000 g in *water R* and dilute to 20.0 mL with the same solvent. Titrate with *0.001 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*) using a silver indicator electrode and an appropriate reference electrode.

1 mL of *0.001 M silver nitrate* is equivalent to 126.9 µg of iodide.

Ionic compounds (*2.2.38*)

Maximum 0.02 per cent *m/m* calculated as sodium chloride.

Rinse all glassware with distilled water R 5 times before use.

Test solution Dissolve 1.0 g of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 20.0 mg of *sodium chloride R* in *water R* and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *water R*.

Measure the specific conductivity of the test solution and the reference solution using a suitable conductivity meter. The specific conductivity of the test solution is not greater than that of the reference solution.

Water (*2.5.12*)

Maximum 4.0 per cent, determined on 0.500 g.

ASSAY

In a 125 mL round-bottomed flask, dissolve 0.200 g in 25 mL of a 50 g/L solution of *sodium hydroxide R*, add 0.5 g of *zinc powder R* and a few glass beads. Boil under a reflux condenser for 1 h. Allow to cool and rinse the condenser with 20 mL of *water R*, adding the rinsings to the flask. Filter through a sintered-glass filter (40) (*2.1.2*) and wash the filter with several quantities of *water R*. Collect the filtrate and washings. Add 5 mL of *glacial acetic acid R* and titrate immediately with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 25.84 mg of $C_{35}H_{44}I_6N_6O_{15}$.

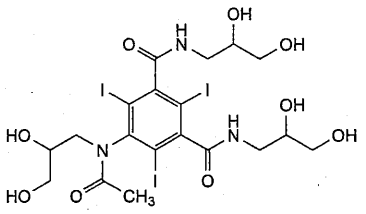
STORAGE

In an airtight container, protected from light.

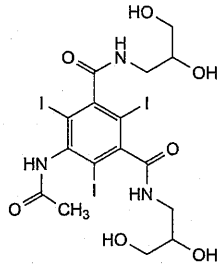
IMPURITIES

Specified impurities *C, E, H, overalkylated impurities.*

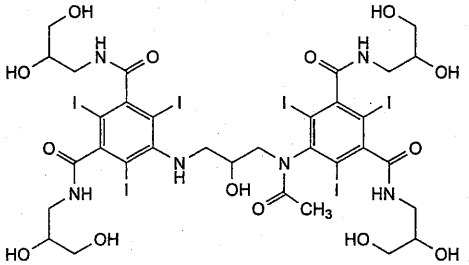
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also *5.10. Control of impurities in substances for pharmaceutical use: A, B, F, G.*



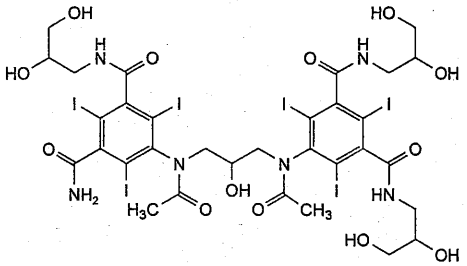
A. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide (iohexol),



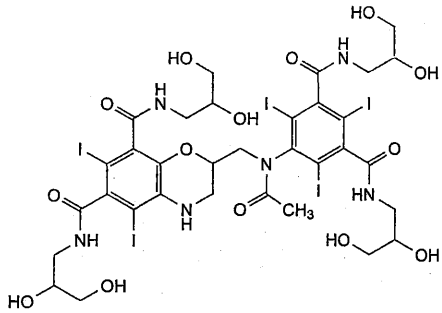
B. 5-acetamido-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



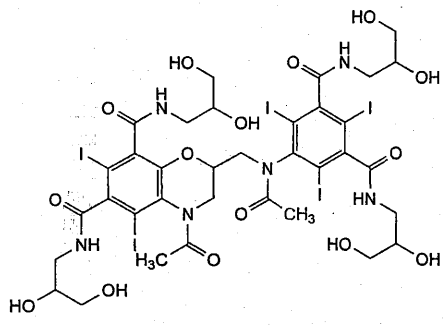
C. 5-[acetyl[3-[[3,5-bis[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]-2-hydroxypropyl]amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



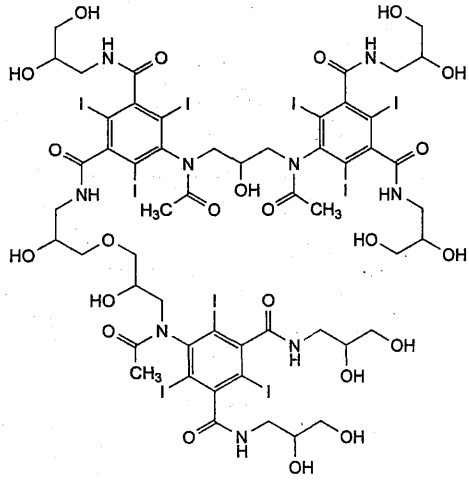
E. 5-[acetyl[3-[acetyl[3-carbamoyl-5-[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]-2-hydroxypropyl]amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



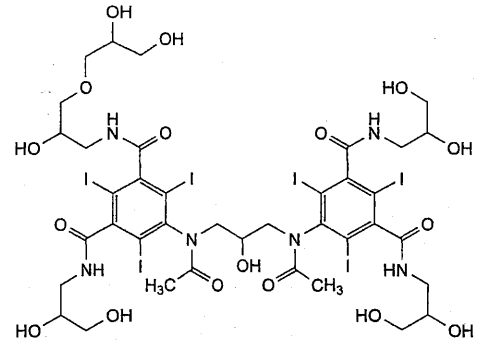
F. 2-[[acetyl[3,5-bis[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]methyl]-*N*,*N*′-bis(2,3-dihydroxypropyl)-5,7-diiodo-3,4-dihydro-2*H*-1,4-benzoxazine-6,8-dicarboxamide,



G. 4-acetyl-2-[[acetyl[3,5-bis[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]methyl]-*N*,*N*′-bis(2,3-dihydroxypropyl)-5,7-diiodo-3,4-dihydro-2*H*-1,4-benzoxazine-6,8-dicarboxamide,



H. 5-[acetyl[3-[acetyl[3-[[3-[3-[acetyl[3,5-bis[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]-2-hydroxypropoxy]-2-hydroxypropyl]carbamoyl]-5-[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]-2-hydroxypropyl]amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide.

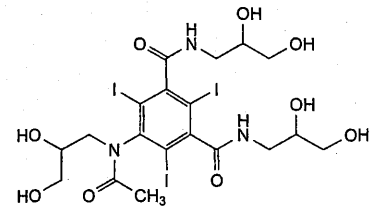


I. overalkylated impurities (an example): 5-[acetyl[3-[acetyl[3,5-bis[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodophenyl]amino]-2-hydroxypropyl]amino]-*N*-[3-(2,3-dihydroxypropoxy)-2-hydroxypropyl]-*N*′-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide.

Ph Eur

Iohexol

(Ph. Eur. monograph 1114)



$C_{19}H_{26}I_3N_3O_9$ 821 *66108-95-0*

Action and use
Iodinated contrast medium.

Ph Eur

DEFINITION

5-[Acetyl(2,3-dihydroxypropyl)amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide.

The substance is a mixture of diastereoisomers and atropisomers.

Content
98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or greyish-white, hygroscopic powder.

Solubility
Very soluble in water, freely soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *iohexol CRS*.

B. Examine the chromatograms obtained in test A for related substances.

Results The principal peaks in the chromatogram obtained with reference solution (b) are similar in retention time and size to the peaks due to iohexol in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 5.0 g in *water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Related substances

A. Liquid chromatography (*2.2.29*).

NOTE: iohexol gives rise to 2 non-resolved peaks in the chromatogram due to endo-exo isomerism. In addition, a small peak (also due to iohexol) usually appears at the leading edge of the 1st principal peak. This small peak has a retention time about 1.2 min less than the 1st principal peak.

Test solution Dissolve 0.150 g of the substance to be examined in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 15.0 mg of *iohexol CRS* and 15.0 mg of *iohexol impurity A CRS* in a mixture of 0.05-0.1 mL of *dilute sodium hydroxide solution R* and 10 mL of *water R* and dilute to 100.0 mL with *water R*. Dilute 1.0 mL of this solution to 10.0 mL with *water R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *water R*.

Reference solution (c) Dissolve 5.0 mg of *iohexol for peak identification CRS* (containing impurities B, C, D and E) in *water R* and dilute to 5.0 mL with the same solvent.

Blank solution *water R*.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: *water R*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 60	99 → 87	1 → 13

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Retention time Impurities A and H = about 17 min; iohexol (peaks corresponding to *endo-exo* isomerism) = about 20 min.

System suitability Reference solution (a):
- *resolution*: minimum 5.0 between the peak due to impurity A and the 2nd and greater peak due to iohexol.

Limits:
- *sum of impurities B, C, D and E* (relative retention with reference to the 2nd and greater peak due to iohexol between 1.1 and 1.4): not more than 0.6 times the total area of the principal peaks in the chromatogram obtained with reference solution (b) (0.6 per cent); use the chromatogram obtained with reference solution (c) to identify the corresponding peaks;
- *sum of impurities A and H*: not more than 0.5 times the total area of the principal peaks in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *unspecified impurities*: for each impurity, not more than 0.1 times the total area of the principal peaks in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than 1.5 times the total area of the principal peaks in the chromatogram obtained with reference solution (b) (1.5 per cent);
- *disregard limit*: 0.03 times the total area of the principal peaks in the chromatogram obtained with reference solution (b) (0.03 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 1.0 g of the substance to be examined in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *iohexol impurity J CRS* and 50 mg of *iohexol CRS* in *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with *water R*. Dilute 1.0 mL of this solution to 50.0 mL with *water R*.

Plate *TLC silica gel F_{254} plate R*.

Pretreatment Wash the plate with the mobile phase, dry at room temperature for 30 min, then at 90 °C for 1 h.

Mobile phase *concentrated ammonia R, methanol R, 2-propanol R, acetone R* (16:16:28:40 *V/V/V/V*).

Application 10 µL.

Development Over 1/2 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability Reference solution (a):
- the chromatogram shows 2 clearly separated spots.

Limits:
- *any impurity*: any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

3-Chloropropane-1,2-diol

Gas chromatography (*2.2.28*).

Test solution Dissolve 1.0 g of the substance to be examined in 1.0 mL of *water R*. Shake with 4 quantities, each of 2 mL, of *methyl acetate R*. Dry the combined upper layers over *anhydrous sodium sulfate R*. Filter and concentrate to about 0.7 mL using a warm water-bath at 60 °C and a stream of nitrogen and dilute to 1.0 mL with *methyl acetate R*.

Reference solution Dissolve 0.25 g of *3-chloropropane-1,2-diol R* in 100.0 mL of *methyl acetate R*. Dilute 1.0 mL of this solution to 100.0 mL with *methyl acetate R*.

Column:
- *material*: fused silica;
- *size*: *l* = 25 m, Ø = 0.33 mm;
- *stationary phase*: *polymethylphenylsiloxane R* (film thickness 1 µm).

Carrier gas *helium for chromatography R*.

Flow rate 1 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	80
	2 - 8	80 → 170
	8 - 10	170
Injection port		230
Detector		250

Detection Flame ionisation.

Injection 2 µL (splitless for 30 s).

System suitability Reference solution:
— *retention time*: 3-chloropropane-1,2-diol = about 8 min.

Limit:
— *3-chloropropane-1,2-diol*: not more than the area of the principal peak in the chromatogram obtained with the reference solution (25 ppm).

Free aromatic amine
Maximum 500 ppm.

Test solution Transfer 0.200 g of the substance to be examined to a 25 mL volumetric flask and dissolve in 15.0 mL of *water R*.

Reference solution Dissolve 5.0 mg of *iohexol impurity J CRS* in *water R* and dilute to 5.0 mL with *water R*. Dilute 1.0 mL of the solution to 100.0 mL with *water R*. Mix 10.0 mL of this solution with 5.0 mL of *water R* in a 25 mL volumetric flask.

Blank solution Transfer 15.0 mL of *water R* to a 25 mL volumetric flask.

In conducting the following steps, keep the flasks in iced water and protected as much as possible from light until all of the reagents have been added.

Place the 3 flasks containing respectively the test solution, the reference solution and the blank solution in iced water, protected from light, for 5 min. Add 1.5 mL of *hydrochloric acid R1* and mix by swirling. Add 1.0 mL of a 20 g/L solution of *sodium nitrite R*, mix and allow to stand for 4 min. Add 1.0 mL of a 40 g/L solution of *sulfamic acid R*, swirl gently until gas liberation has ceased and allow to stand for 1 min. *(CAUTION: considerable pressure is produced)*. Add 1.0 mL of a freshly prepared 3 g/L solution of *naphthylethylenediamine dihydrochloride R* in a mixture of 30 volumes of *water R* and 70 volumes of *propylene glycol R* and mix. Remove the flasks from the iced water, dilute to 25.0 mL with *water R*, mix and allow to stand for 5 min. Simultaneously determine the absorbance (*2.2.25*) at 495 nm of the solutions obtained from the test solution and the reference solution in 5 cm cells, using the blank as the compensation liquid. The absorbance of the test solution is not greater than that of the reference solution.

Iodide
Maximum 10 ppm.

Dissolve 6.000 g in *water R* and dilute to 20 mL with the same solvent. Add 2.0 mL of 0.001 M potassium iodide. Titrate with *0.001 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*), using a silver indicator electrode and an appropriate reference electrode. Subtract the volume of titrant corresponding to the 2.0 mL of 0.001 M potassium iodide, determined by titrating a blank to which is added 2.0 mL of 0.001 M potassium iodide and use the residual value to calculate the iodide content.

1 mL of *0.001 M silver nitrate* is equivalent to 126.9 µg of I^-.

Ionic compounds (*2.2.38*)
Maximum 0.01 per cent *m/m* calculated as sodium chloride.

Rinse all glassware with distilled water R 5 times before use.

Test solution Dissolve 1.0 g of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 20.0 mg of *sodium chloride R* in *water R* and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with *water R*.

Measure the conductivity of the test solution and the reference solution using a suitable conductivity meter. The conductivity of the test solution is not greater than that of the reference solution.

Water (*2.5.12*)
Maximum 4.0 per cent, determined on 1.00 g.

ASSAY

To 0.500 g in a 125 mL round-bottomed flask add 25 mL of a 50 g/L solution of *sodium hydroxide R*, 0.5 g of *zinc powder R* and a few glass beads. Boil under a reflux condenser for 30 min. Allow to cool and rinse the condenser with 20 mL of *water R*, adding the rinsings to the flask. Filter through a sintered-glass filter (*2.1.2*) and wash the filter with several quantities of *water R*. Collect the filtrate and washings. Add 5 mL of *glacial acetic acid R* and titrate immediately with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 27.37 mg of $C_{19}H_{26}I_3N_3O_9$.

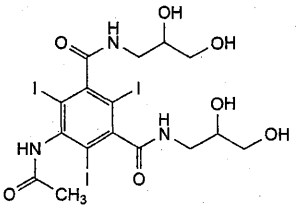
STORAGE

In an airtight container, protected from light and moisture.

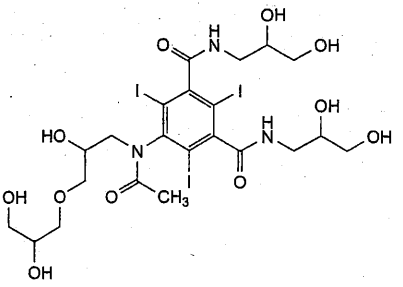
IMPURITIES

Specified impurities *A, B, C, D, E, H.*

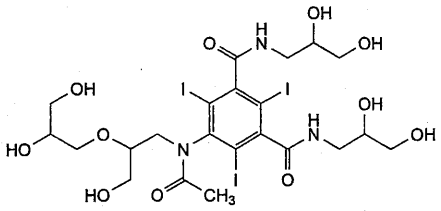
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *F, G, I, J, K, L, M, N, O, P, Q.*



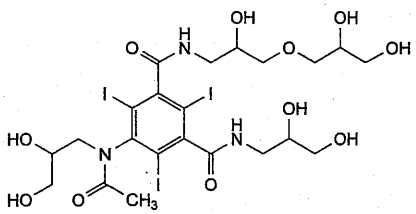
A. 5-(acetylamino)-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



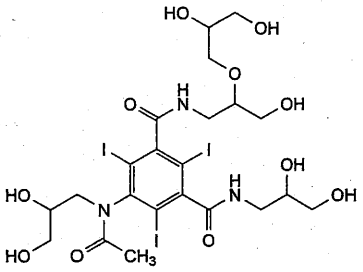
B. 5-[acetyl[3-(2,3-dihydroxypropoxy)-2-hydroxypropyl]amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



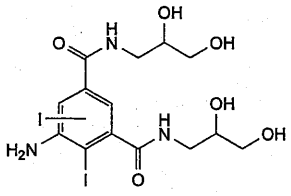
C. 5-[acetyl[2-(2,3-dihydroxypropoxy)-3-hydroxypropyl]amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



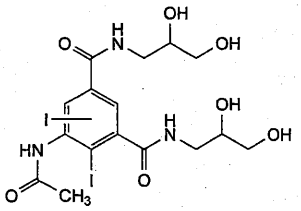
D. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*-[3-(2,3-dihydroxypropoxy)-2-hydroxypropyl]-*N*′-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



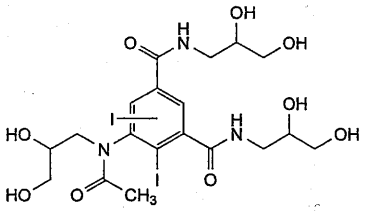
E. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*-[2-(2,3-dihydroxypropoxy)-3-hydroxypropyl]-*N*′-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



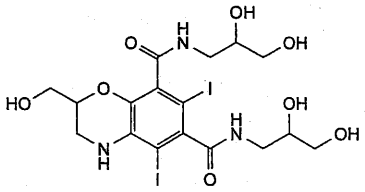
F. 5-amino-*N*,*N*′-bis(2,3-dihydroxypropyl)diiodobenzene-1,3-dicarboxamide,



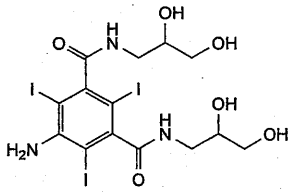
G. 5-(acetylamino)-*N*,*N*′-bis(2,3-dihydroxypropyl) diiodobenzene-1,3-dicarboxamide,



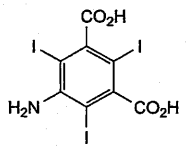
H. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*,*N*′-bis(2,3-dihydroxypropyl)diiodobenzene-1,3-dicarboxamide,



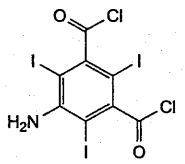
I. *N*,*N*′-bis(2,3-dihydroxypropyl)-2-(hydroxymethyl)-5,7-diiodo-3,4-dihydro-2*H*-1,4-benzoxazine-6,8-dicarboxamide,



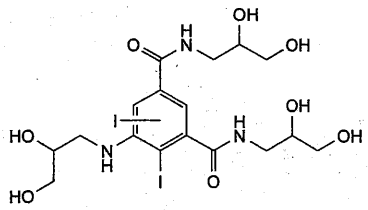
J. 5-amino-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



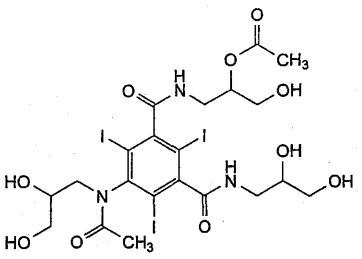
K. 5-amino-2,4,6-triiodobenzene-1,3-dicarboxylic acid,



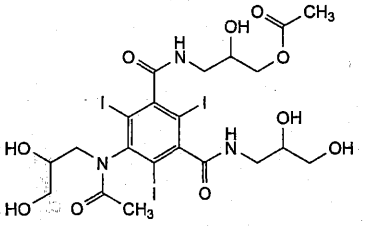
L. 3,5-bis(chlorocarbonyl)-2,4,6-triiodobenzenamine,



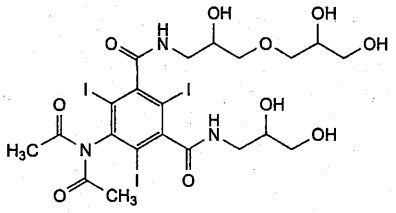
M. *N*,*N*′-bis(2,3-dihydroxypropyl)-5-[(2,3-dihydroxypropyl) amino]diiodobenzene-1,3-dicarboxamide,



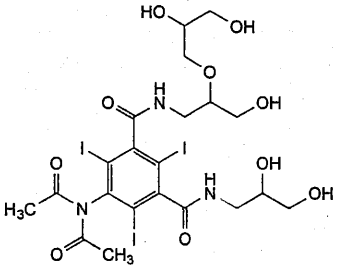
N. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*-[2-(acetyloxy)-3-hydroxypropyl]-*N'*-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



O. 5-[acetyl(2,3-dihydroxypropyl)amino]-*N*-[3-(acetyloxy)-2-hydroxypropyl]-*N'*-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,



P. 5-(diacetylamino)-*N*-[3-(2,3-dihydroxypropoxy)-2-hydroxypropyl]-*N'*-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide,

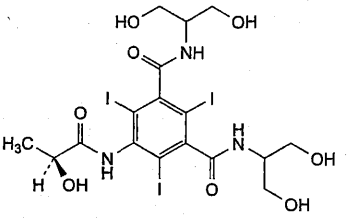


Q. 5-(diacetylamino)-*N*-[2-(2,3-dihydroxypropoxy)-3-hydroxypropyl]-*N'*-(2,3-dihydroxypropyl)-2,4,6-triiodobenzene-1,3-dicarboxamide.

Ph Eur

Iopamidol

(Ph. Eur. monograph 1115)



$C_{17}H_{22}I_3N_3O_8$ 777 *60166-93-0*

Action and use
Iodinated contrast medium.

Preparations
Iopamidol Injection
Iopamidol Oral Solution

Ph Eur

DEFINITION

N,N'-Bis[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4,6-triiodobenzene-1,3-dicarboxamide.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility
Freely soluble in water, very slightly soluble in methanol, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *iopamidol CRS*.

B. Loss on drying (see Tests).

C. Specific optical rotation (see Tests).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

Dissolve 1 g in *water R* and dilute to 50 mL with the same solvent.

Acidity or alkalinity
Dissolve 10.0 g in *carbon dioxide-free water R* and dilute to 100 mL with the same solvent. Not more than 0.75 mL of *0.01 M hydrochloric acid* or 1.4 mL of *0.01 M sodium hydroxide* is required to adjust to pH 7.0 (*2.2.3*).

Specific optical rotation (*2.2.7*)
−4.6 to −5.2 (dried substance), determined at 436 nm.

Dissolve 10.0 g, with heating if necessary, in *water R* and dilute to 25.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.50 g of the substance to be examined in *water R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *iopamidol impurity H CRS* in *water R* and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 20.0 mL with *water R*. Dilute 1.0 mL of this solution to 50.0 mL with *water R*.

Reference solution (c) Add 0.1 mL of the test solution to 20 mL of reference solution (a) and dilute to 50 mL with *water R*.

Column 2 columns coupled in series,
— *size*: *l* = 0.25 m, Ø = 4.6 mm,
— *stationary phase*: *phenylsilyl silica gel for chromatography R* (5 µm),
— *temperature*: 60 °C.

Mobile phase:
— *mobile phase A*: *water R*,
— *mobile phase B*: *acetonitrile R*, *water R* (50:50 *V/V*),

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 18	100	0
18 - 40	100 - 62	0 - 38
40 - 45	62 - 50	38 - 50
45 - 50	50 - 100	50 - 0
50 - 60	100	0

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL.

Relative retention With reference to iopamidol (retention time = about 14.6 min): impurity D = about 0.1; impurity B = about 0.6; impurities I and H = about 0.9; impurity G = about 1.1; impurity K = about 1.2; impurity C = about 1.3; impurity J = about 1.5; impurity A = about 1.8; impurity E = about 2.2; impurity F = about 2.3.

System suitability Reference solution (c):
— *resolution*: minimum 2.0 between the peaks due to impurity H and iopamidol.

Limits:
— *sum of impurities H and I*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— *impurities A, B, C, D, E, F, G, J, K*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *any other impurity*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— *sum of impurities other than H and I*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),
— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.01 per cent).

Free aromatic amines
Maximum 200 ppm.

Keep the solutions and reagents in iced water, protected from bright light.

Test solution In a 25 mL volumetric flask, dissolve 0.500 g of the substance to be examined in 20.0 mL of *water R*.

Reference solution In a 25 mL volumetric flask, mix 4.0 mL of a 25.0 mg/L solution of *iopamidol impurity A CRS* with 16.0 mL of *water R*.

Blank solution Place 20.0 mL of *water R* in a 25 mL volumetric flask.

Place the flasks in iced water, protected from light, for 5 min. Add 1.0 mL of *hydrochloric acid R* to each flask, mix and allow to stand for 5 min. Add 1.0 mL of a 20 g/L solution of *sodium nitrite R* prepared immediately before use, mix and allow to stand for 5 min. Add 1.0 mL of a 120 g/L solution of *ammonium sulfamate R*, swirl gently until gas liberation has ceased, and allow to stand for 5 min. *(CAUTION: considerable pressure is produced)*. Add 1.0 mL of a freshly prepared 1 g/L solution of *naphthylethylenediamine dihydrochloride R* and mix. Remove the flasks from the iced water and allow to stand for 10 min. Dilute to 25.0 mL with *water R* and mix. Measure immediately the absorbance (*2.2.25*) at 500 nm of the solutions obtained from the test solution and the reference solution using, as the compensation liquid, the solution obtained from the blank solution.

The absorbance of the test solution is not greater than that of the reference solution.

Free iodine
Maximum 10 ppm.

Dissolve 2.0 g in 25 mL of *water R* in a ground-glass stoppered centrifuge tube. Add 5 mL of *toluene R* and 5 mL of *dilute sulfuric acid R*. Shake and centrifuge. Any red colour of the upper layer is not more intense than that of the upper phase obtained in the same way from 22 mL of *water R*, 2 mL of *iodide standard solution (10 ppm I) R*, 5 mL of *dilute sulfuric acid R*, 1 mL of *strong hydrogen peroxide solution R* and 5 mL of *toluene R*.

Iodide
Maximum 10 ppm.

Dissolve 6.000 g in *water R* and dilute to 20 mL with the same solvent. Add 2.0 mL of 0.001 M potassium iodide. Carry out a potentiometric titration (*2.2.20*) with *0.001 M silver nitrate* using a silver indicator electrode and an appropriate reference electrode. Subtract the volume of titrant corresponding to the 2.0 mL of 0.001 M potassium iodide, determined by titrating a blank to which is added 2.0 mL of 0.001 M potassium iodide and use the residual value to calculate the iodide content.

1 mL of *0.001 M silver nitrate* is equivalent to 126.9 µg of iodide.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 1.4 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
To 0.300 g in a 250 mL round-bottomed flask add 5 mL of *strong sodium hydroxide solution R*, 20 mL of *water R*, 1 g of *zinc powder R* and a few glass beads. Boil under a reflux condenser for 30 min. Allow to cool and rinse the condenser with 20 mL of *water R*, adding the rinsings to the flask. Filter through a sintered-glass filter (*2.1.2*) and wash the filter with several quantities of *water R*. Collect the filtrate and

washings. Add 5 mL of *glacial acetic acid R* and titrate immediately with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*) using a suitable electrode system such as silver-silver chloride.

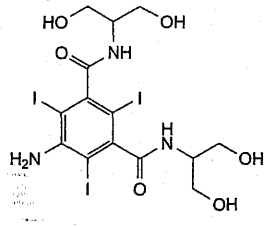
1 mL of *0.1 M silver nitrate* is equivalent to 25.90 mg of $C_{17}H_{22}I_3N_3O_8$.

STORAGE

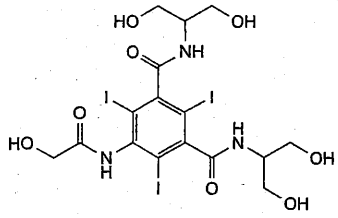
Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

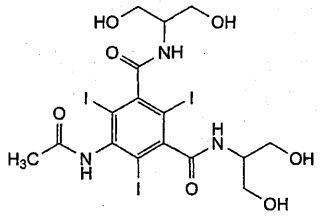
Specified impurities *A, B, C, D, E, F, G, H, I, J, K.*



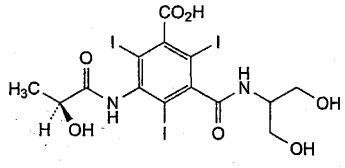
A. 5-amino-*N*,*N*'-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-2,4,6-triiodobenzene-1,3-dicarboxamide,



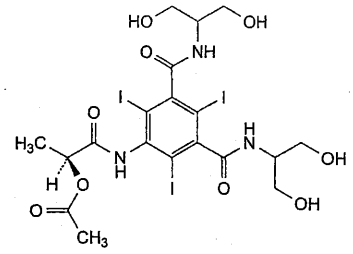
B. 5-[(hydroxyacetyl)amino]-*N*,*N*'-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-2,4,6-triiodobenzene-1,3-dicarboxamide,



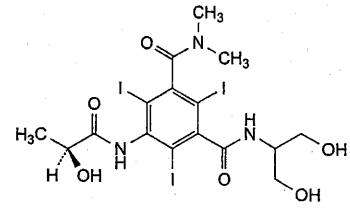
C. 5-(acetylamino)-*N*,*N*'-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-2,4,6-triiodobenzene-1,3-dicarboxamide,



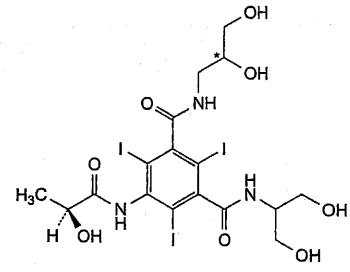
D. 3-[[2-hydroxy-1-(hydroxymethyl)ethyl]carbamoyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4,6-triiodobenzoic acid,



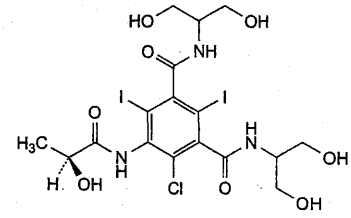
E. (1*S*)-2-[[3,5-bis[[2-hydroxy-1-(hydroxymethyl)ethyl]carbamoyl]-2,4,6-triiodophenyl]amino]-1-methyl-2-oxoethyl acetate,



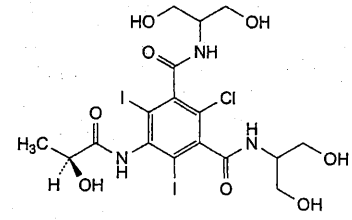
F. *N*'-[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4,6-triiodo-*N*,*N*-dimethylbenzene-1,3-dicarboxamide,



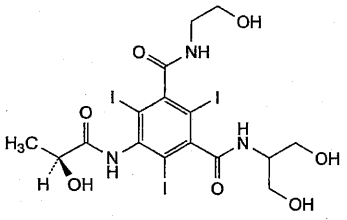
G. *N*-(2,3-dihydroxypropyl)-*N*'-[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4,6-triiodobenzene-1,3-dicarboxamide,



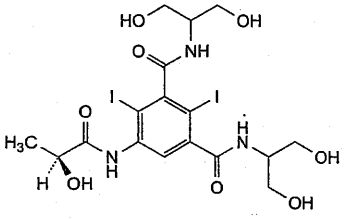
H. 4-chloro-*N*,*N*'-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,6-diiodobenzene-1,3-dicarboxamide,



I. 2-chloro-*N*,*N*'-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-4,6-diiodobenzene-1,3-dicarboxamide,



J. *N*-(2-hydroxyethyl)-*N'*-[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4,6-triiodobenzene-1,3-dicarboxamide,

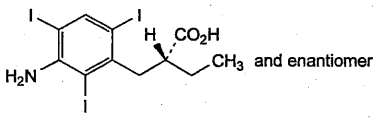


K. *N,N'*-bis[2-hydroxy-1-(hydroxymethyl)ethyl]-5-[[(2*S*)-2-hydroxypropanoyl]amino]-2,4-diiodobenzene-1,3-dicarboxamide.

Ph Eur

Iopanoic Acid

(Ph. Eur. monograph 0700)



$C_{11}H_{12}I_3NO_2$ 571 *96-83-3*

Action and use
Iodinated contrast medium.

Ph Eur

DEFINITION
Iopanoic acid contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of (*RS*)-2-(3-amino-2,4,6-tri-iodobenzyl)butanoic acid, calculated with reference to the dried substance.

CHARACTERS
A white or yellowish-white powder, practically insoluble in water, soluble in anhydrous ethanol and in methanol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION
First identification: B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): about 155 °C, with decomposition.

B. Examine by infrared absorption spectrophotometry (*2.2.24*), comparing with the spectrum obtained with *iopanoic acid CRS*.

C. Examine the chromatograms obtained in the test for related substances (see Tests). Spray the plate with a 1 g/L solution of *4-dimethylaminocinnamaldehyde R* in a mixture of 1 volume of *hydrochloric acid R* and 99 volumes of *ethanol (96 per cent) R*. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Heat 50 mg carefully in a small porcelain dish over a flame. Violet vapour is evolved.

TESTS
Appearance of solution
Dissolve 1.0 g in *1 M sodium hydroxide* and dilute to 20 mL with the same solvent. The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_3 (*2.2.2, Method II*).

Related substances
Examine by thin-layer chromatography (*2.2.27*), using *silica gel* GF_{254} *R* as the coating substance.

Test solution (a) Dissolve 1.0 g of the substance to be examined in a mixture of 3 volumes of *ammonia R* and 97 volumes of *methanol R* and dilute to 10 mL with the same mixture of solvents.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 3 volumes of *ammonia R* and 97 volumes of *methanol R*.

Reference solution (a) Dissolve 50 mg of *iopanoic acid CRS* in a mixture of 3 volumes of *ammonia R* and 97 volumes of *methanol R* and dilute to 5 mL with the same mixture of solvents.

Reference solution (b) Dilute 1 mL of test solution (b) to 50 mL with a mixture of 3 volumes of *ammonia R* and 97 volumes of *methanol R*.

Apply separately to the plate 5 µL of each solution. Develop over a path of 10 cm using a mixture of 10 volumes of *concentrated ammonia R*, 20 volumes of *methanol R*, 20 volumes of *toluene R* and 50 volumes of *dioxan R*. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Halides
To 0.46 g add 10 mL of *nitric acid R* and 15 mL of *water R*. Shake for 5 min and filter. 15 mL of the filtrate complies with the limit test for chlorides (*2.4.4*) (180 ppm, expressed as chloride).

Loss on drying (*2.2.32*)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (*2.4.14*)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
To 0.150 g in a 250 mL round-bottomed flask add 5 mL of *strong sodium hydroxide solution R*, 20 mL of *water R*, 1 g of *zinc powder R* and a few glass beads. Boil under a reflux condenser for 60 min. Allow to cool and rinse the condenser with 20 mL of *water R*, adding the rinsings to the flask. Filter through a sintered-glass filter (*2.1.2*) and wash the filter with several quantities of *water R*. Collect the filtrate and washings. Add 40 mL of *dilute sulfuric acid R* and titrate immediately with *0.1 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*).

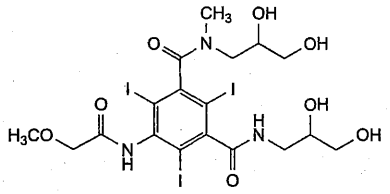
1 mL of *0.1 M silver nitrate* is equivalent to 19.03 mg of $C_{11}H_{12}I_3NO_2$.

STORAGE
Store protected from light.

Ph Eur

Iopromide

(Ph. Eur. monograph 1753)



$C_{18}H_{24}I_3N_3O_8$ 791 *73334-07-3*

Action and use
Iodinated contrast medium.

Ph Eur

DEFINITION

N,N′-Bis(2,3-dihydroxypropyl)-2,4,6-triiodo-5-[(methoxyacetyl)amino]-*N*-methylbenzene-1,3-dicarboxamide.

Mixture of diastereoisomers and atropisomers.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly yellowish powder.

Solubility
Freely soluble in water and in dimethyl sulfoxide, practically insoluble in ethanol (96 per cent) and in acetone.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *iopromide CRS*.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solutions BY_6, B_6 and Y_6 (*2.2.2, Method I*).

Dissolve 16.5 g in 20 mL of *carbon dioxide-free water R* while heating on a water-bath at a temperature not exceeding 70 °C. Allow to cool to room temperature.

Conductivity (*2.2.38*)
Maximum 50 $\mu S \cdot cm^{-1}$.

Dissolve 1.000 g in *water R* and dilute to 50.0 mL with the same solvent.

Impurity A and related primary aromatic amines
Maximum 0.01 per cent.

Protect the solutions from light throughout the test. All given times are critical for the test results. The test solution, reference solution and blank solution must be processed in parallel.

Test solution Dissolve 0.500 g of the substance to be examined in 20.0 mL of *water R* in a 25 mL volumetric flask.

Reference solution Dissolve the contents of a vial of *iopromide impurity A CRS* in 5.0 mL of *water R*. Transfer 2.0 mL of this solution to a 25 mL volumetric flask and add 18.0 mL of *water R*.

Blank solution Place 20.0 mL of *water R* in a 25 mL volumetric flask.

Cool the test solution, reference solution and blank solution in a bath of iced water for 5 min. Add 1.0 mL of *hydrochloric acid R1* to each solution and cool again for 5 min in a bath of iced water. Add 1.0 mL of a 20 g/L solution of *sodium nitrite R*, shake vigorously and cool for another 5 min in a bath of iced water. To each solution add 0.50 mL of an 80 g/L solution of *sulfamic acid R*. Over the next 5 min, shake vigorously several times, raising the stoppers to vent the gas that evolves. Afterwards, add to each solution 1.0 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* in a mixture of 300 volumes of *water R* and 700 volumes of *propylene glycol R*, shake, allow to cool to room temperature for 10 min and dilute to 25.0 mL with *water R*. Degas the solutions in an ultrasonic bath for 1 min and measure the absorbance (*2.2.25*) of the test solution and the reference solution at 495 nm against the blank, within 5 min. The test is not valid unless the absorbance of the reference solution is at least 0.08. The absorbance of the test solution is not greater than the absorbance of the reference solution.

Impurity B
Liquid chromatography (*2.2.29*).

Solvent mixture *methanol R, water R* (50:50 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 40.0 mg of *iopromide CRS* in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Introduce several millilitres of reference solution (a) into a vial sealed with a crimp-top. Heat at 121 °C for 15 min.

Reference solution (c) Dilute 1.5 mL of the test solution to 100.0 mL with the solvent mixture.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 20 °C.

Mobile phase Mix 6 g of *chloroform R* with 59 g of *methanol R*. Add 900 g of *water for chromatography R* in small portions to the chloroform/methanol mixture and stir for at least 2 h to obtain a homogeneous solution.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Run time 50 min.

Identification of impurities Use the chromatogram supplied with *iopromide CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurity B isomers Y_1 and Y_2.

Relative retention With reference to iopromide isomer Z_2 (retention time = about 34 min): impurity B isomer Y_1 = about 0.28; impurity B isomer Y_2 = about 0.31.

System suitability Reference solution (a):
— the chromatogram obtained shows 2 peaks due to impurity B isomers Y_1 and Y_2.

Limit:
— *sum of impurity B isomers Y_1 and Y_2*: not more than the sum of the areas of the 2 principal peaks due to the iopromide in the chromatogram obtained with reference solution (c) (1.5 per cent).

Related substances
Thin-layer chromatography (*2.2.27*).

Solvent mixture *methanol R, water R* (50:50 *V/V*).

Test solution Dissolve 1.0 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 2.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (e) Dissolve the contents of a vial of *iopromide for system suitability 1 CRS* (containing impurities B and E) in 50 µL of the solvent mixture.

Reference solution (f) Dissolve the contents of a vial of *iopromide for system suitability 2 CRS* (containing impurities B, C, D and F) in 50 µL of the solvent mixture.

Plates *TLC silica gel F_{254} plate R* (2 plates).

A. *Mobile phase*: *concentrated ammonia R, water R, dioxan R* (4:15:85 *V/V/V*).

Application 2 µL of the test solution and reference solutions (b), (d) and (e).

Development Over 3/4 of the plate.

Drying In a current of air, until complete evaporation of the solvents, then at 120 °C for 30 min.

Detection Examine immediately in ultraviolet light at 254 nm; expose to ultraviolet light for 2-5 min until the principal spots appear clearly as yellow spots, then spray with *ferric chloride-ferricyanide-arsenite reagent R* and examine immediately in daylight.

Retardation factors Impurity B = about 0.26; iopromide = about 0.34; impurity E = about 0.41.

System suitability Reference solution (e):
- the chromatogram shows 3 clearly separated spots.

Limits:
- *impurity E*: any spot due to impurity E is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *unspecified impurities*: any other spot is not more intense than the principal spot in the chromatogram obtained with reference solution (d) (0.10 per cent); disregard any spot due to impurity B.

B. *Mobile phase*: *anhydrous formic acid R, water R, methanol R, chloroform R* (2:6:32:62 *V/V/V/V*).

Application 2 µL of the test solution and reference solutions (a), (b) ,(c), (d) and (f).

Development Over 3/4 of the plate.

Drying In a current of air, until complete evaporation of the solvents, then at 120 °C for 30 min.

Detection Examine immediately in ultraviolet light at 254 nm; expose to an ammonia vapour for 30 min, dry in a current of air for 10 min, then expose to ultraviolet light for 2-5 min until the principal spots appear clearly as yellow spots, then spray with *ferric chloride-ferricyanide-arsenite reagent R* and examine immediately in daylight.

Retardation factors Impurity C = about 0.23; impurity D = about 0.29; impurity B = about 0.36; iopromide = about 0.43; impurity F = about 0.71.

System suitability Reference solution (f):
- the chromatogram shows 5 clearly separated spots.

Limits:
- *impurity D*: any spot due to impurity D is not more intense than the principal spot in the chromatogram obtained with reference solution (a) (1.0 per cent);
- *impurity C*: any spot due to impurity C is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurity F*: any spot due to impurity F is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.2 per cent);
- *unspecified impurities*: any other spot is not more intense than the principal spot in the chromatogram obtained with reference solution (d) (0.10 per cent); disregard any spot due to impurity B.

Isomer distribution

Liquid chromatography (*2.2.29*) as described in the test for impurity B with the following modifications.

Calculate the percentage content of the isomer groups with reference to the total area of all the peaks due to the 4 iopromide isomers, using the chromatogram obtained with the test solution.

Limits:
- *sum of iopromide isomers E_1 and Z_1*: 40.0 per cent to 51.0 per cent;
- *sum of iopromide isomers E_2 and Z_2*: 49.0 per cent to 60.0 per cent.

Free iodine

Dissolve 2.0 g in 20 mL of *water R* in a glass-stoppered test tube. Add 2 mL of *dilute sulfuric acid R* and 2 mL of *toluene R*, close and shake vigorously. The upper layer remains colourless (*2.2.2, Method II*).

Iodide

Maximum 2 ppm.

Dissolve 10.0 g in 50 mL of *carbon dioxide-free water R*. Adjust to pH 3-4 adding about 0.15 mL of *0.1 M sulfuric acid*. Titrate with *0.001 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*) using a combined metal electrode. Not more than 0.15 mL of *0.001 M silver nitrate* is required to reach the end-point.

Water (*2.5.12*)

Maximum 1.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)

Less than 1.0 IU/g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for impurity B with the following modifications.

Injection Test solution and reference solutions (a) and (b).

Identification of the isomers The 2 principal peaks in the chromatogram obtained with reference solution (a) are due to iopromide isomers Z_1 and Z_2 . The 2 peaks that have an increased size in the chromatogram obtained with reference solution (b) in comparison to the chromatogram obtained with reference solution (a), are due to iopromide isomers E_1 and E_2.

Relative retention With reference to iopromide isomer Z_2 (retention time = about 34 min): iopromide isomer E_1 = about 0.70; iopromide isomer E_2 = about 0.75; iopromide isomer Z_1 = about 0.85.

System suitability Reference solution (a):
- *resolution*: minimum 2.0 between the peaks due to iopromide isomers Z_1 and Z_2.

Calculate the percentage content of iopromide from the declared content of *iopromide CRS* and from the sum of the areas of all of the peaks due to isomer groups E and Z.

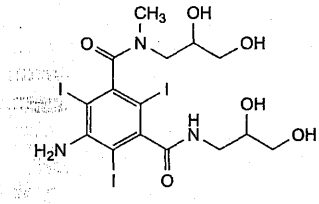
STORAGE

Protected from light.

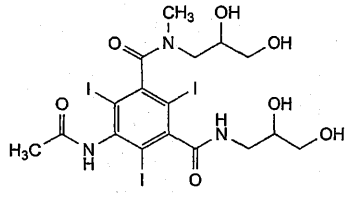
IMPURITIES

Specified impurities A, B, C, D, E, F.

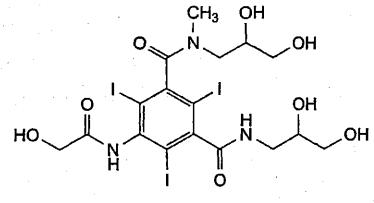
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) G, H.



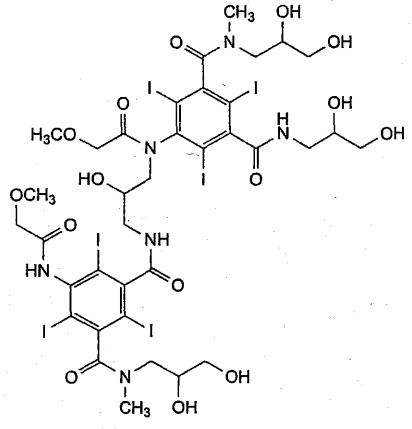
A. 5-amino-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodo-*N*-methylbenzene-1,3-dicarboxamide,



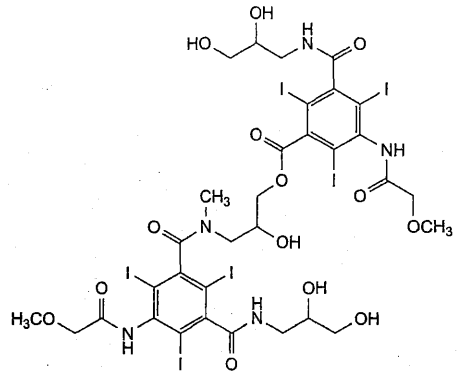
B. 5-(acetylamino)-*N*,*N*′-bis(2,3-dihydroxypropyl)-2,4,6-triiodo-*N*-methylbenzene-1,3-dicarboxamide,



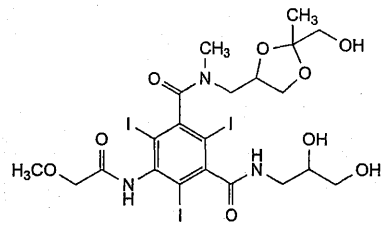
C. *N*,*N*′-bis(2,3-dihydroxypropyl)-5-[(hydroxyacetyl)amino]-2,4,6-triiodo-*N*-methylbenzene-1,3-dicarboxamide,



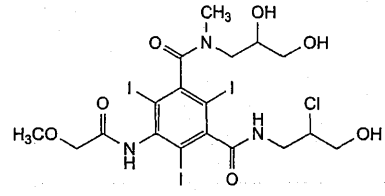
D. *N*-(2,3-dihydroxypropyl)-*N*′-[3-[[3-[(2,3-dihydroxypropyl)carbamoyl]-5-[(2,3-dihydroxypropyl)methylcarbamoyl]-2,4,6-triiodophenyl](methoxyacetyl)amino]-2-hydroxypropyl]-2,4,6-triiodo-5-[(methoxyacetyl)amino]-*N*-methylbenzene-1,3-dicarboxamide,



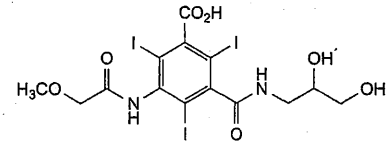
E. 3-[[3-[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodo-5-[(methoxyacetyl)amino]benzoyl]methylamino]-2-hydroxypropyl 3-[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodo-5-[(methoxyacetyl)amino]benzoate,



F. *N*′-(2,3-dihydroxypropyl)-*N*-[[2-(hydroxymethyl)-2-methyl-1,3-dioxolan-4-yl]methyl]-2,4,6-triiodo-5-[(methoxyacetyl)amino]-*N*-methylbenzene-1,3-dicarboxamide,



G. *N*′-(2-chloro-3-hydroxypropyl)-*N*-(2,3-dihydroxypropyl)-2,4,6-triiodo-5-[(methoxyacetyl)amino]-*N*-methylbenzene-1,3-dicarboxamide,

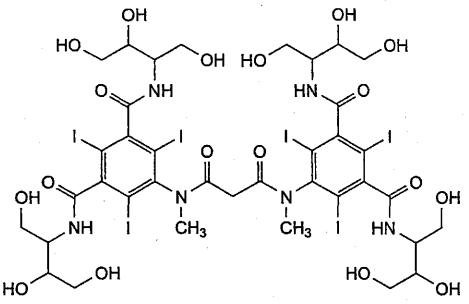


H. 3-[(2,3-dihydroxypropyl)carbamoyl]-2,4,6-triiodo-5-[(methoxyacetyl)amino]benzoic acid.

Ph Eur

Iotrolan

(Ph. Eur. monograph 1754)



$C_{37}H_{48}I_6N_6O_{18}$ 1626 *79770-24-4*

Action and use
Iodinated contrast medium.

Ph Eur

DEFINITION

Mixture of stereoisomers of 5,5′-[propanedioylbis (methylimino)]bis[*N*,*N*′-bis[2,3-dihydroxy-1-(hydroxymethyl)propyl]2,4,6-triiodobenzene-1,3-dicarboxamide].

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or yellowish-white powder, hygroscopic.

Solubility
Very soluble in water, freely soluble in dimethyl sulfoxide, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *iotrolan CRS.*

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 18.0 g in *carbon dioxide-free water R* and dilute to 20.0 mL with the same solvent.

Conductivity (*2.2.38*)
Maximum 25 µS·cm^{-1}.

Dissolve 1.000 g in *water R* and dilute to 50.0 mL with the same solvent.

Primary aromatic amines
Protect the solutions from light throughout the test. All given times are critical for the test results. The test solution, the reference solution and the blank solution must be processed in parallel.

Test solution Dissolve 0.500 g of the substance to be examined in 20.0 mL of *water R* in a 25 mL volumetric flask.

Reference solution Dissolve 5.0 mg of *iopamidol impurity A CRS* in *water R* and dilute to 20.0 mL with the same solvent. Transfer 1.0 mL of this solution to a 25 mL volumetric flask and add 19.0 mL of *water R*.

Blank solution Place 20.0 mL of *water R* in a 25 mL volumetric flask.

Procedure Cool the test solution, reference solution and blank solution in a bath of iced water for 5 min. Add 1.0 mL of *hydrochloric acid R1* to each solution and cool again for 5 min in a bath of iced water. Add 1.0 mL of a 20 g/L solution of *sodium nitrite R*, shake vigorously and cool for another 5 min in a bath of iced water. To each solution add 0.50 mL of an 80 g/L solution of *sulfamic acid R*. Over the next 5 min, shake vigorously several times, raising the stoppers to vent the gas that evolves. Afterwards add to each solution 1.0 mL of a 1 g/L solution of *naphthylethylenediamine dihydrochloride R* in a mixture of 300 volumes of *water R* and 700 volumes of *propylene glycol R*, shake, allow to cool to room temperature for 10 min and dilute to 25.0 mL with *water R*. Degas the solutions in an ultrasonic bath for 1 min and measure the absorbance (*2.2.25*) of the test solution and the reference solution at 495 nm against the blank, within 5 min.

System suitability:
— *absorbance of the reference solution*: minimum 0.40.

Limit:
— *absorbance of the test solution*: not more than the absorbance of the reference solution (0.05 per cent).

Related substances
Thin-layer chromatography (*2.2.27*). *Prepare the solutions immediately before use.*

Test solution Dissolve 1.0 g of the substance to be examined in a mixture of equal volumes of *methanol R* and *water R* and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with a mixture of equal volumes of *methanol R* and *water R*.

Reference solution (b) Dilute 2.0 mL of reference solution (a) to 10.0 mL with a mixture of equal volumes of *methanol R* and *water R*.

Reference solution (c) Dissolve the contents of a vial of *iotrolan for system suitability CRS* (containing about 0.05 per cent of each of impurities A and B) in 50 µL of a mixture of equal volumes of *methanol R* and *water R*.

Plate *TLC silica gel F_{254} plate R.*

Pretreatment Over 3/4 of the plate with *methylene chloride R.*

Mobile phase *concentrated ammonia R, water R, dioxan R* (4:20:80 *V/V/V*).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In a current of air until the solvents have evaporated.

Detection Examine in ultraviolet light at 254 nm. Expose the plate to the ultraviolet light for 2-5 min until the principal spots appear clearly as yellow spots. Spray with *ferric chloride-ferricyanide-arsenite reagent R* and examine in daylight.

R_F values Iotrolan = about 0.25; impurity A = about 0.4; impurity B = about 0.5.

System suitability Reference solution (c):
— the chromatogram shows 3 clearly separated spots.

Limits:
— *impurities A, B*: any spot due to impurity A or B is not more intense than the principal spot in the chromatogram obtained with reference solution (a) (0.5 per cent);
— *unspecified impurities*: any other spot is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.10 per cent).

Isomer distribution
Liquid chromatography (*2.2.29*) as described under Assay. Use the normalisation procedure.

Identification of peaks Use the chromatogram supplied with *iotrolan CRS* and the chromatogram obtained with the reference solution to identify the peaks due to the 3 isomer groups.

Calculate the percentage content of each of the isomer groups G1, G2 and G3, with reference to the total area of all of the peaks due to the 3 isomer groups, using the chromatogram obtained with the test solution.

Limits:
— *isomer group G1*: 53.0 per cent to 70.0 per cent;
— *isomer group G2*: 3.0 per cent to 11.0 per cent;
— *isomer group G3*: 25.0 per cent to 39.0 per cent.

Free iodine
Dissolve 0.20 g in 1 mL of *water R* in a glass-stoppered test tube. Add 4 mL of a 370 g/L solution of *sulfuric acid R* and 5 mL of *toluene R*, close and shake vigorously. The upper layer remains colourless (*2.2.2, Method II*).

Iodide
Maximum 20 ppm.

Dissolve 10.0 g in 50 mL of *carbon dioxide-free water R*. Adjust to pH 3-4 adding about 0.15 mL of *dilute sulfuric acid R*. Titrate with *0.001 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*). Not more than 1.5 mL of *0.001 M silver nitrate* is required to reach the end-point.

Water (*2.5.12*)
Maximum 3.5 per cent, determined on 0.250 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (*2.6.14*)
Less than 0.7 IU/g.

ASSAY

Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in *water R* and dilute to 25.0 mL with the same solvent.

Reference solution Dissolve 40.0 mg of *iotrolan CRS* in *water R* and dilute to 25.0 mL with the same solvent.

Column:
— *size*: *l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm);
— *temperature*: 40 °C.

Mobile phase *methanol R, water for chromatography R* (10:90 *V/V*).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 40 min.

Retention time Isomer group G1 = about 8 min to 12 min; isomer group G2 = about 15 min to 22 min; isomer group G3 = about 22 min to 32 min.

System suitability Reference solution:
— the chromatogram obtained is similar to the chromatogram supplied with *iotrolan CRS*.

Calculate the percentage content of iotrolan from the total area of all of the peaks of the 3 isomer groups G1, G2 and G3 and the declared content of *iotrolan CRS*.

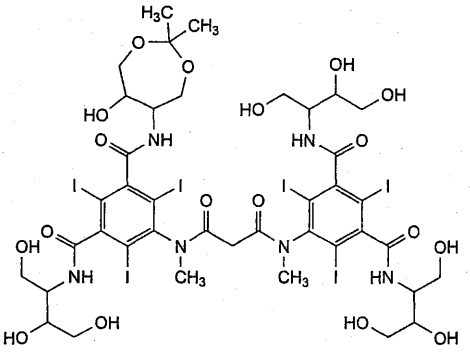
STORAGE

In an airtight container, protected from light.

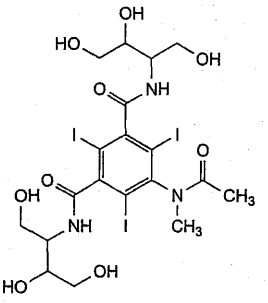
IMPURITIES

Specified impurities *A, B.*

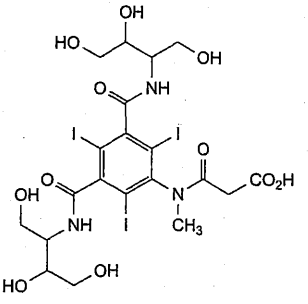
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D, E, F, G, H, I, J.*



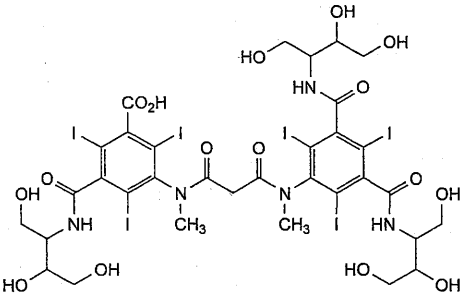
A. *N,N′*-bis[2,3-dihydroxy-1-(hydroxymethyl)propyl]-5-[[3-[[3-[[2,3-dihydroxy-1-(hydroxymethyl)propyl]carbamoyl]-5-[(6-hydroxy-2,2-dimethyl-1,3-dioxepan-5-yl)carbamoyl]-2,4,6-triiodophenyl]methylamino]-3-oxopropanoyl]methylamino]-2,4,6-triiodobenzene-1,3-dicarboxamide,



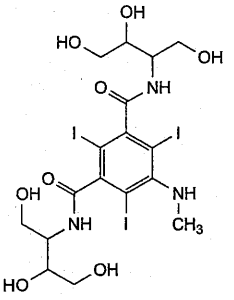
B. 5-(acetylmethylamino)-*N,N′*-bis[2,3-dihydroxy-1-(hydroxymethyl)propyl]-2,4,6-triiodobenzene-1,3-dicarboxamide,



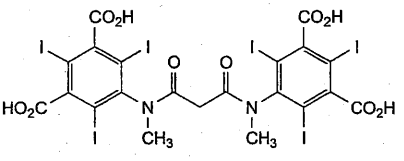
C. 3-[[3,5-bis[[2,3-dihydroxy-1-(hydroxymethyl)propyl]carbamoyl]-2,4,6-triiodophenyl]methylamino]-3-oxopropanoic acid,



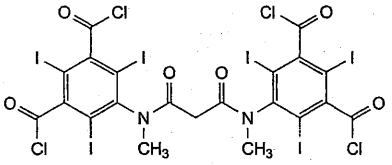
D. 3-[[3-[[3,5-bis[[2,3-dihydroxy-1-(hydroxymethyl)propyl] carbamoyl]-2,4,6-triiodophenyl]methylamino]-3-oxopropanoyl]methylamino]-5-[[2,3-dihydroxy-1-(hydroxymethyl)propyl]carbamoyl]-2,4,6-triiodobenzoic acid,



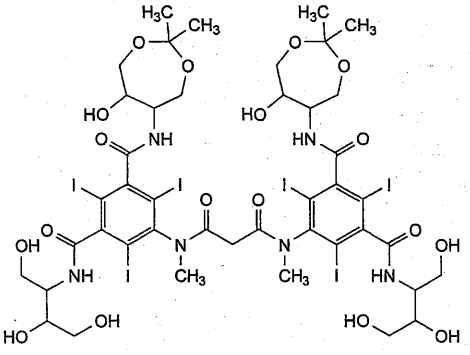
E. *N,N′*-bis[2,3-dihydroxy-1-(hydroxymethyl)propyl]-2,4,6-triiodo-5-(methylamino)benzene-1,3-dicarboxamide,



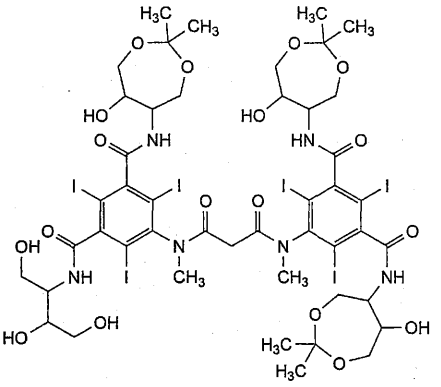
F. 5,5′-[propanedioylbis(methylimino)]bis[2,4,6-triiodobenzene-1,3-dicarboxylic] acid,



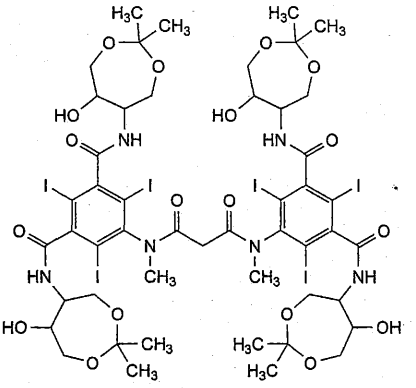
G. 5,5′-[propanedioylbis(methylimino)]bis[2,4,6-triiodobenzene-1,3-dicarbonyl] tetrachloride,



H. 5,5′-[propanedioylbis(methylimino)]bis[*N*-[2,3-dihydroxy-1-(hydroxymethyl)propyl]-*N′*-(6-hydroxy-2,2-dimethyl-1,3-dioxepan-5-yl)-2,4,6-triiodobenzene-1,3-dicarboxamide],



I. 5-[[3-[[3-[[2,3-dihydroxy-1-(hydroxymethyl) propyl]carbamoyl]-5-[(6-hydroxy-2,2-dimethyl-1,3-dioxepan-5-yl)carbamoyl]-2,4,6-triiodophenyl] methylamino]-3-oxopropanoyl]methylamino]-*N,N′*-bis(6-hydroxy-2,2-dimethyl-1,3-dioxepan-5-yl)-2,4,6-triiodobenzene-1,3-dicarboxamide,

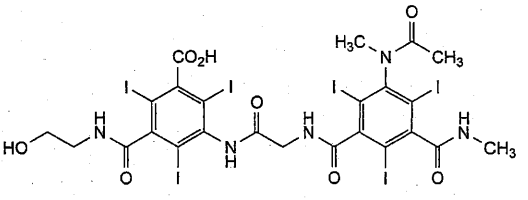


J. 5,5′-[propanedioylbis(methylimino)]bis[*N,N′*-bis(6-hydroxy-2,2-dimethyl-1,3-dioxepan-5-yl)-2,4,6-triiodobenzene-1,3-dicarboxamide].

Ph Eur

Ioxaglic Acid

(Ph. Eur. monograph 2009)



$C_{24}H_{21}I_6N_5O_8$ 1269 *59017-64-0*

Action and use
Iodinated contrast medium.

Ph Eur

DEFINITION
3-[[[[3-(Acetylmethylamino)-2,4,6-triiodo-5-(methylcarbamoyl)benzoyl]amino]acetyl]amino]-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic acid.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Very slightly soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ioxaglic acid CRS.*

TESTS

Appearance of solution

The solution is clear (*2.2.1*).

Dissolve 1.0 g in a 40 g/L solution of *sodium hydroxide R* and dilute to 20 mL with the same solution.

Absorbance (*2.2.25*)

Maximum 0.18, calculated for a solution containing 40 per cent of anhydrous ioxaglic acid.

Dissolve 10.0 g in about 8 mL of a 40 g/L solution of *sodium hydroxide R*. Adjust to pH 7.2-7.6 with a 40 g/L solution of *sodium hydroxide R* or *1 M hydrochloric acid*. Dilute to 25 mL with *water R*. Filter through a membrane filter (nominal pore size 0.45 µm). Measure the absorbance at 450 nm using *water R* as the compensation liquid.

Related substances

Liquid chromatography (*2.2.29*): use the normalisation procedure.

Solvent mixture *acetonitrile R, water R* (5:95 *V/V*).

Test solution Dissolve 0.10 g of the substance to be examined in about 40 mL of the solvent mixture. Add 0.5 ± 0.1 mL of a 4 g/L solution of *sodium hydroxide R* and dilute to 50.0 mL with the solvent mixture. Shake until dissolution is complete, using ultrasound if necessary.

Reference solution (a) Dissolve 0.10 g of *ioxaglic acid CRS* in about 40 mL of the solvent mixture. Add 0.5 ± 0.1 mL of a 4 g/L solution of *sodium hydroxide R* and dilute to 50.0 mL with the solvent mixture. Shake until dissolution is complete, using ultrasound if necessary.

Reference solution (b) Dissolve 5 mg of *ioxaglic acid impurity A CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octylsilyl silica gel for chromatography R* (5 µm);
- *temperature*: 25 °C.

Mobile phase:
- *mobile phase A*: 0.136 g/L solution of *potassium dihydrogen phosphate R* previously adjusted to pH 3.0 with *phosphoric acid R*;
- *mobile phase B*: *acetonitrile R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 5	95 → 90	5 → 10
5 - 40	90	10
40 - 85	90 → 70	10 → 30
85 - 115	70	30
115 - 120	70 → 50	30 → 50
120 - 125	50	50

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 242 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *ioxaglic acid CRS* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C, D1, D2, D3, D4, E and F; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to ioxaglic acid (retention time = about 65 min): impurity A = about 0.3; impurity B = about 0.7; impurity C = about 0.9; impurity D1 = about 1.09; impurity E = about 1.12; impurity D2 = about 1.20; impurity D3 = about 1.26; impurity D4 = about 1.28; impurity F = about 1.6.

System suitability Reference solution (a):
- *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to ioxaglic acid.

Limits:
- *impurity D (sum of the peaks due to impurities D1, D2, D3 and D4)*: maximum 0.7 per cent;
- *impurity E*: maximum 0.7 per cent;
- *impurity F*: maximum 0.4 per cent;
- *impurity B*: maximum 0.3 per cent;
- *impurity C*: maximum 0.3 per cent;
- *impurity A*: maximum 0.1 per cent;
- *unspecified impurities*: for each impurity, maximum 0.2 per cent;
- *total*: maximum 2.0 per cent;
- *reporting threshold*: 0.05 per cent; disregard any peak with a retention time greater than 125 min.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Iodides

Maximum 50 ppm.

Disperse 10.0 g in 50 mL of *water R*. Add 8 mL of *1 M sodium hydroxide*. After dissolution and homogenisation, add 1.0 mL of *glacial acetic acid R*. Immediately titrate with *0.001 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*), using a silver indicator electrode and a suitable reference electrode.

1 mL of *0.001 M silver nitrate* is equivalent to 0.1269 mg of iodides.

Water (*2.5.12*)

Maximum 5.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

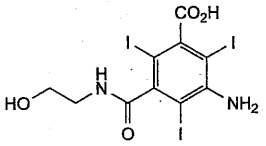
ASSAY

In a round-bottomed flask place 0.100 g of the substance to be examined and add 5 mL of *strong sodium hydroxide solution R*, 20 mL of *water R*, 1 g of *zinc powder R* and a few glass beads. Fit the flask with a reflux condenser and boil for 30 min. Cool and rinse the condenser with 20 mL of *water R*. Add the rinsings to the contents of the flask. Filter, wash the filter with 3 quantities, each of 15 mL, of *water R* and add the washings to the filtrate. Add 40 mL of *dilute sulfuric acid R* and titrate immediately with *0.05 M silver nitrate*. Determine the end-point potentiometrically (*2.2.20*).

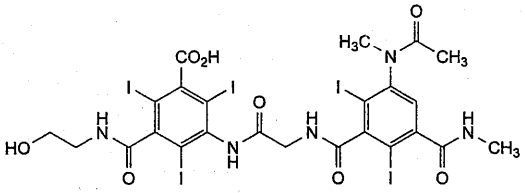
1 mL of *0.05 M silver nitrate* is equivalent to 10.58 mg of $C_{24}H_{21}I_6N_5O_8$.

STORAGE
In an airtight container, protected from light.

IMPURITIES
Specified impurities A, B, C, D, E, F.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) G, H.

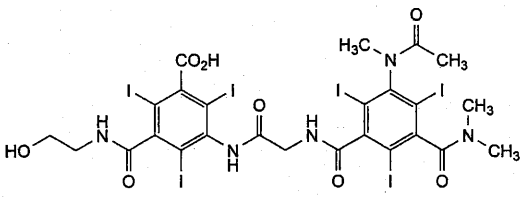


A. 3-amino-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic acid,

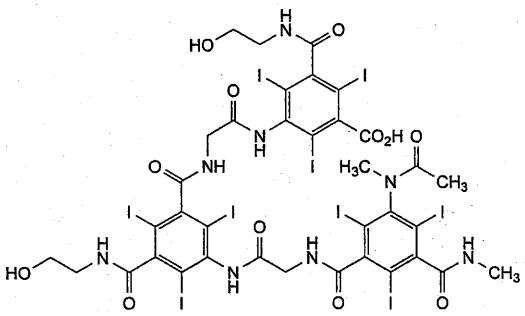


B. 3-[[[[3-(acetylmethylamino)-2,6-diiodo-5-(methylcarbamoyl)benzoyl]amino]acetyl]amino]-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic acid,

C. unknown structure,

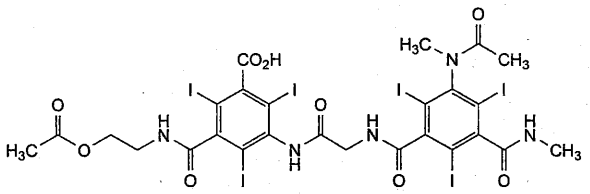


D. D1, D2, D3 and D4: 3-[[[[3-(acetylmethylamino)-5-(dimethylcarbamoyl)-2,4,6-triiodobenzoyl]amino]acetyl]amino]-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic acid,

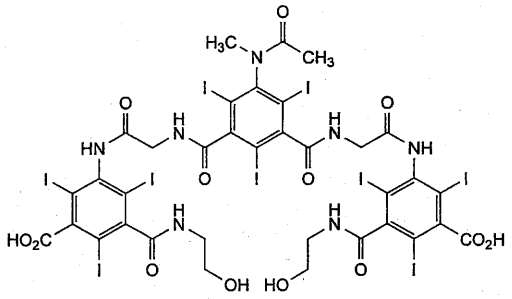


E. 3-[[[[3-[[[[3-(acetylmethylamino)-2,4,6-triiodo-5-(methylcarbamoyl)benzoyl]amino]acetyl]amino]-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoyl]amino]acetyl]amino]-5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic acid,

F. unknown structure,



G. 3-[[[[3-(acetylmethylamino)-2,4,6-triiodo-5-(methylcarbamoyl)benzoyl]amino]acetyl]amino]-5-[[2-(acetyloxy)ethyl]carbamoyl]-2,4,6-triiodobenzoic acid,

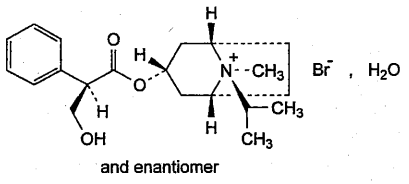


H. 3,3′-[[5-(acetylmethylamino)-2,4,6-triiodo-1,3-phenylene]bis(carbonyliminomethylenecarbonylimino)]bis[5-[(2-hydroxyethyl)carbamoyl]-2,4,6-triiodobenzoic] acid.

Ph Eur

Ipratropium Bromide

(Ph. Eur. monograph 0919)



$C_{20}H_{30}BrNO_3,H_2O$ 430.4 *66985-17-9*

Action and use
Anticholinergic (antimuscarinic) bronchodilator.

Preparations
Ipratropium Nebuliser Solution
Ipratropium Pressurised Inhalation

Ph Eur

DEFINITION
(1*R*,3*r*,5*S*,8*r*)-3-[[(2*RS*)-3-Hydroxy-2-phenylpropanoyl]oxy]-8-methyl-8-(1-methylethyl)-8-azoniabicyclo[3.2.1]octane bromide monohydrate.

Content
99.0 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Soluble in water, freely soluble in methanol, slightly soluble in ethanol (96 per cent).

mp
About 230 °C, with decomposition.

IDENTIFICATION

First identification: A, E.

Second identification: B, C, D, E.

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ipratropium bromide CRS.*

B. Examine the chromatograms obtained in the test for impurity A.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 5 mL of solution S (see Tests), add 2 mL of *dilute sodium hydroxide solution R*. No precipitate is formed.

D. To about 1 mg add 0.2 mL of *nitric acid R* and evaporate to dryness on a water-bath. Dissolve the residue in 2 mL of *acetone R* and add 0.1 mL of a 30 g/L solution of *potassium hydroxide R* in *methanol R*. A violet colour develops.

E. It gives reaction (a) of bromides (*2.3.1*).

TESTS

Solution S

Dissolve 0.50 g in *carbon dioxide-free water R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution GY_7 (*2.2.2, Method II*).

pH (*2.2.3*)

5.0 to 7.5 for solution S.

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 1.0 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *ipratropium bromide CRS* in *methanol R* and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of *methylatropine bromide CRS* in 1.0 mL of reference solution (a).

Reference solution (c) Dissolve 5 mg of *ipratropium impurity A CRS* in 100.0 mL of *methanol R*. Dilute 2.0 mL of the solution to 5.0 mL with *methanol R*.

Plate *TLC silica gel plate R* (2-10 µm).

Mobile phase *anhydrous formic acid R, water R, ethanol (96 per cent) R, methylene chloride R* (1:3:18:18 *V/V/V/V*).

Application 1 µL.

Development Over a path of 6 cm.

Drying At 60 °C for 15 min.

Detection Spray with *potassium iodobismuthate solution R5*, allow the plate to dry in air, spray with a 50 g/L solution of *sodium nitrite R* and protect immediately with a sheet of glass.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated principal spots.

Limit:

— *impurity A*: any spot due to impurity A is not more intense than the principal spot in the chromatogram obtained with reference solution (c) (0.1 per cent).

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.200 g of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of *ipratropium bromide CRS* in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *ipratropium bromide CRS* and 5 mg of *ipratropium impurity B CRS* in 1 mL of *methanol R* and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 20.0 mL with the mobile phase.

Column:

— *size*: *l* = 0.15 m, Ø = 3.9 mm;

— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm);

— *temperature*: 30 °C.

Mobile phase Dissolve 12.4 g of *sodium dihydrogen phosphate R* and 1.7 g of *tetrapropylammonium chloride R* in 870 mL of *water for chromatography R*; adjust to pH 5.5 with a 180 g/L solution of *disodium hydrogen phosphate dodecahydrate R* and add 130 mL of *methanol R1*.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 5 µL.

Run time 6 times the retention time of ipratropium.

Relative retention With reference to ipratropium (retention time = about 4.9 min): impurity C = about 0.7; impurity B = about 1.2; impurity D = about 1.8; impurity E = about 2.3; impurity F = about 5.1.

System suitability Reference solution (b):

— *resolution*: minimum 3.0 between the peaks due to impurity B and ipratropium;

— *symmetry factor*: maximum 2.5 for the principal peak.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.3; impurity D = 0.2; impurity F = 0.5;

— *impurity D*: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);

— *impurities B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);

— *disregard limit*: one-third of the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent); disregard the peak due to the bromide ion.

Water (*2.5.12*)

3.9 per cent to 4.4 per cent, determined on 0.50 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

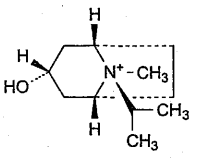
Dissolve 0.350 g in 50 mL of *water R* and add 3 mL of *dilute nitric acid R*. Titrate with *0.1 M silver nitrate*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M silver nitrate* is equivalent to 41.24 mg of $C_{20}H_{30}BrNO_3$.

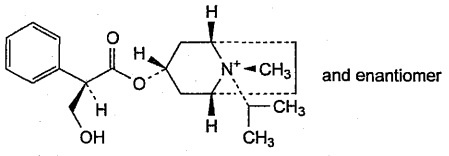
IMPURITIES

Specified impurities *A, B, C, D.*

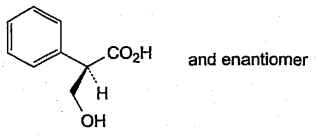
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *E, F.*



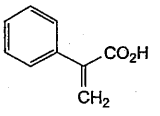
A. (1*R*,3*r*,5*S*,8*r*)-3-hydroxy-8-methyl-8-(1-methylethyl)-8-azoniabicyclo[3.2.1]octane,



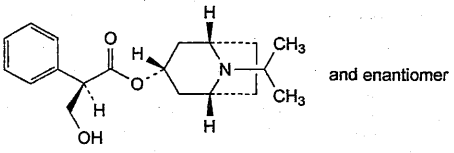
B. (1*R*,3*r*,5*S*,8*s*)-3-[[(2*RS*)-3-hydroxy-2-phenylpropanoyl]oxy]-8-methyl-8-(1-methylethyl)-8-azoniabicyclo[3.2.1]octane,



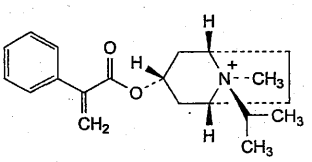
C. (2*RS*)-3-hydroxy-2-phenylpropanoic acid (DL-tropic acid),



D. 2-phenylpropenoic acid (atropic acid),



E. (1*R*,3*r*,5*S*)-8-(1-methylethyl)-8-azabicyclo[3.2.1]oct-3-yl (2*RS*)-3-hydroxy-2-phenylpropanoate,

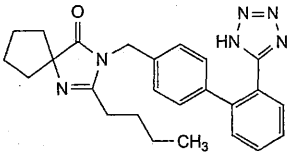


F. (1*R*,3*r*,5*S*,8*r*)-8-methyl-8-(1-methylethyl)-3-[(2-phenylpropenoyl)oxy]-8-azoniabicyclo[3.2.1]octane.

Ph Eur

Irbesartan

(Ph. Eur. monograph 2465)



$C_{25}H_{28}N_6O$ 428.5 *138402-11-6*

Action and use

Angiotensis II (AT_1) receptor antagonist.

Preparation

Irbesartan Tablets

Ph Eur

DEFINITION

2-Butyl-3-[[2′-(1*H*-tetrazol-5-yl)[1,1′-biphenyl]-4-yl]methyl]-1,3-diazaspiro[4.4]non-1-en-4-one.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methanol, slightly soluble in methylene chloride.

It shows polymorphism (*5.9*).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *irbesartan CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness at 60 °C and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 (*2.2.2, Method II*).

Dissolve 0.50 g in a mixture of 1 volume of *2 M sodium hydroxide R* and 9 volumes of *methanol R2* and dilute to 10 mL with the same mixture of solvents.

Impurity B

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution Dissolve 25.0 mg of *sodium azide R* (sodium salt of impurity B) in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 0.25 mL of the solution to 200.0 mL with the mobile phase.

Precolumn (used to prevent saturation of the column with irbesartan):

— *size*: *l* = 0.05 m, Ø = 4 mm;

— *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (8.5 µm).

Column:

— *size*: *l* = 0.25 m, Ø = 4 mm;

— *stationary phase*: *strongly basic anion-exchange resin for chromatography R* (8.5 µm).

Mobile phase 4.2 g/L solution of *sodium hydroxide R* in *carbon dioxide-free water R*.

Flow rate 1.0 mL/min.

Detection Conductivity detector with a sensitivity of 3 µS; use a self-regenerating anion suppressor.

Neutralisation of the eluent Either chemical or electrochemical:
— *chemical*: by continuous countercurrent circulation in a neutralising micromembrane, performed before detection:
  — *neutralising solvent*: *0.025 M sulfuric acid*;
  — *flow rate*: 10 mL/min;
  — *pressure*: about 100 kPa.
— *electrochemical*: 300 mA (for example).

Injection 200 µL; after each injection of the test solution, rinse the precolumn with a mixture of mobile phase and *methanol R* (40:60 *V/V*) for 10 min; equilibrate to initial conditions as necessary; a switch valve can be used to avoid disconnecting the precolumn from the column.

Run time 25 min.

Retention time Impurity B = about 14 min.

System suitability Reference solution:
— *signal-to-noise ratio*: minimum 10 for the peak due to impurity B.

Limit:
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (10 ppm).

Related substances
Liquid chromatography (*2.2.29*).

Buffer solution pH 3.2 Mix 5.5 mL of *phosphoric acid R* and 950 mL of *water R* and adjust to pH 3.2 with *triethylamine R*.

Test solution Dissolve 50 mg of the substance to be examined in *methanol R2* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 20.0 mL with *methanol R2*. Dilute 1.0 mL of this solution to 50.0 mL with *methanol R2*.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of *irbesartan impurity A CRS* in *methanol R2* and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with *methanol R2*.

Column:
— *size*: *l* = 0.25 m, Ø = 4 mm;
— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *acetonitrile R1*, buffer solution pH 3.2 (33:67 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Run time 1.4 times the retention time of irbesartan.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to irbesartan (retention time = about 23 min): impurity A = about 0.7.

System suitability Reference solution (b):
— *resolution*: minimum 3.0 between the peaks due to impurity A and irbesartan.

Limits:
— *impurity A*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

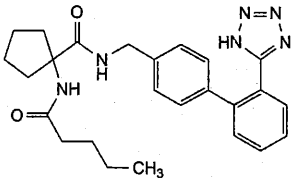
Water (*2.5.12*)
Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 50 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 42.85 mg of $C_{25}H_{28}N_6O$.

IMPURITIES
Specified impurities *A, B.*



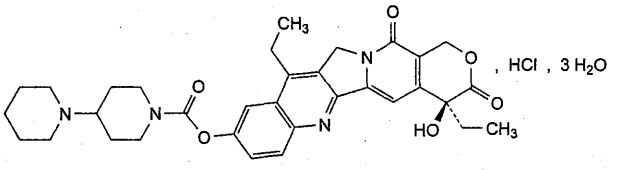
A. 1-(pentanoylamino)-*N*-[[2′-(1*H*-tetrazol-5-yl)[1,1′-biphenyl]-4-yl]methyl]cyclopentane-1-carboxamide,

B. N_3^-: trinitride (azide).

Ph Eur

Irinotecan Hydrochloride Trihydrate

(Ph. Eur. monograph 2675)



$C_{33}H_{39}ClN_4O_6,3H_2O$ 677 *136572-09-3*

Action and use
Inhibitor of DNA topoisomerase type I; antineoplastic.

Ph Eur

DEFINITION
(4*S*)-4,11-Diethyl-4-hydroxy-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate hydrochloride trihydrate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Pale yellow or yellow, crystalline powder.

Solubility

Sparingly soluble in water, in ethanol (96 per cent) and in methanol.

It shows polymorphism (*5.9*).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *irinotecan hydrochloride trihydrate CRS.*

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol R*, evaporate to dryness and record new spectra using the residues.

B. Water (see Tests).

C. It gives reaction (a) of chlorides (*2.3.1*).

Dissolve 0.10 g in *water R* and dilute to 50 mL with the same solvent.

TESTS

Appearance of solution

The solution is clear (*2.2.1*) and not more intensely coloured than reference solution GY_2 (*2.2.2, Method II*).

Dissolve 0.200 g in *water R* with heating at 80 °C and dilute to 20 mL with the same solvent.

Enantiomeric Purity

Liquid chromatography (*2.2.29*).

Solvent mixture *diethylamine R, anhydrous ethanol R* (0.4:100 *V/V*).

Test solution Dissolve 15.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of *irinotecan for system suitability 1 CRS* (containing impurity L) in 1.0 mL of the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *cellulose derivative of silica gel for chiral separation R* (10 µm).

Mobile phase *diethylamine R, anhydrous ethanol R, hexane R* (0.2:50:50 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 370 nm.

Injection 20 µL.

Run time 1.5 times the retention time of irinotecan.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity L.

Relative retention With reference to irinotecan (retention time = about 15 min): impurity L = about 0.7.

System suitability:
- *resolution*: minimum 1.5 between the peaks due to impurity L and irinotecan in the chromatogram obtained with reference solution (b).

Calculation of percentage content:
- for impurity L, use the concentration of irinotecan in reference solution (a).

Limit:
- *impurity L*: maximum 0.15 per cent.

Related substances

Liquid chromatography (*2.2.29*). *Carry out the test protected from light.*

Solvent mixture *acetonitrile R1, methanol R2,* mobile phase A (25:25:50 *V/V/V*).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of *irinotecan hydrochloride trihydrate CRS* in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of *irinotecan for peak identification CRS* (containing impurities C and E) in 1.0 mL of the solvent mixture.

Reference solution (d) Dissolve 5 mg of *irinotecan for system suitability 2 CRS* (containing impurity M) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography with polar incorporated groups R* (5 µm).

Mobile phase:
- *mobile phase A*: dissolve 2.72 g of *potassium dihydrogen phosphate R* in 950 mL of *water for chromatography R*, adjust to pH 3.5 with *dilute phosphoric acid R* and dilute to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *methanol R2, acetonitrile R1* (40:60 *V/V*);

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	80	20
2 - 42	80 → 30	20 → 70
42 - 47	30	70

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL of the test solution and reference solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with *irinotecan for peak identification CRS* and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and E; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity M.

Relative retention With reference to irinotecan (retention time = about 17 min): impurity M = about 0.9; impurity C = about 1.3; impurity E = about 1.5.

System suitability:
- *resolution*: minimum 2.0 between the peaks due to impurity M and irinotecan in the chromatogram obtained with reference solution (d).

Calculation of percentage contents:
- *correction factor*: multiply the peak area of impurity M by 1.3;
- for each impurity, use the concentration of irinotecan in reference solution (b).

Limits:
— *impurities C, E, M*: for each impurity, maximum 0.15 per cent;
— *unspecified impurities*: for each impurity, maximum 0.10 per cent;
— *total*: maximum 0.5 per cent;
— *reporting threshold*: 0.05 per cent.

Water (*2.5.12*)
7.0 per cent to 9.0 per cent, determined on 0.100 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the percentage content of $C_{33}H_{39}ClN_4O_6$ taking into account the assigned content of *irinotecan hydrochloride trihydrate CRS*.

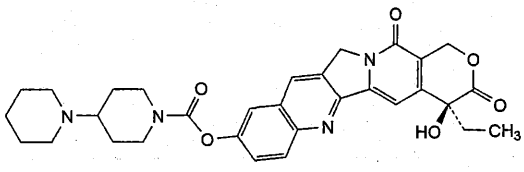
STORAGE

In an airtight container, protected from light.

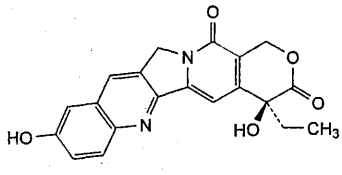
IMPURITIES

Specified impurities *C, E, L, M.*

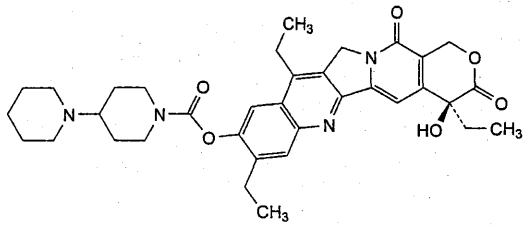
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, B, D, F, G, H, K.*



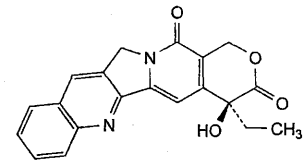
A. (4*S*)-4-ethyl-4-hydroxy-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate (11-desethyl irinotecan),



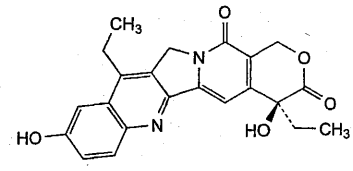
B. (4*S*)-4-ethyl-4,9-dihydroxy-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinoline-3,14(4*H*,12*H*)-dione (9-hydroxycamptothecin),



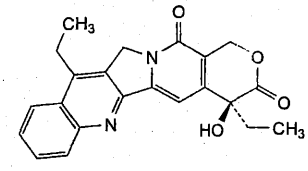
C. (4*S*)-4,8,11-triethyl-4-hydroxy-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate (8-ethyl irinotecan),



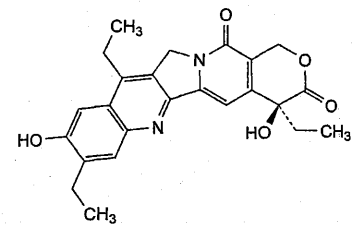
D. (4*S*)-4-ethyl-4-hydroxy-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinoline-3,14(4*H*,12*H*)-dione (camptothecin),



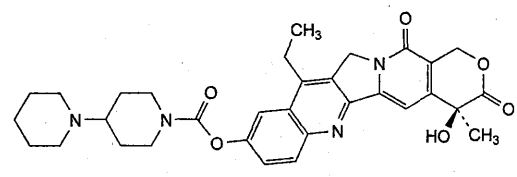
E. (4*S*)-4,11-diethyl-4,9-dihydroxy-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinoline-3,14(4*H*,12*H*)-dione (11-ethyl-9-hydroxycamptothecin),



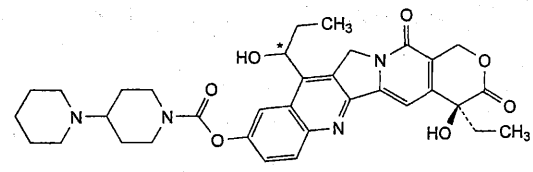
F. (4*S*)-4,11-diethyl-4-hydroxy-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinoline-3,14(4*H*,12*H*)-dione (11-ethylcamptothecin),



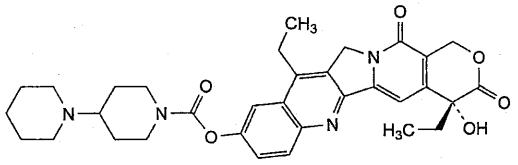
G. (4*S*)-4,8,11-triethyl-4,9-dihydroxy-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinoline-3,14(4*H*,12*H*)-dione (8,11-diethyl-9-hydroxycamptothecin),



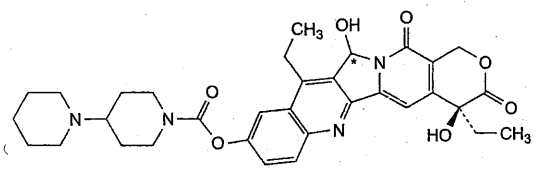
H. (4*S*)-11-ethyl-4-hydroxy-4-methyl-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate (4-methyl irinotecan analogue),



K. (4*S*)-4-ethyl-4-hydroxy-11-(1-hydroxypropyl)-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate,



L. (4*R*)-4,11-diethyl-4-hydroxy-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate (irinotecan enantiomer).

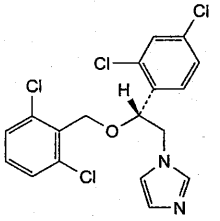


M. (4*S*)-4,11-diethyl-4,12-dihydroxy-3,14-dioxo-3,4,12,14-tetrahydro-1*H*-pyrano[3′,4′:6,7]indolizino[1,2-*b*]quinolin-9-yl 1,4′-bipiperidine-1′-carboxylate (12-hydroxy irinotecan).

Ph Eur

Isoconazole

(Ph. Eur. monograph 1018)



and enantiomer

$C_{18}H_{14}Cl_4N_2O$ 416.1 *27523-40-6*

Action and use
Antifungal.

Ph Eur

DEFINITION
1-[(2*RS*)-2-[(2,6-Dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Practically insoluble in water, very soluble in methanol, freely soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 111 °C to 115 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *isoconazole CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 30 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 30 mg of *isoconazole CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 30 mg of *isoconazole CRS* and 30 mg of *econazole nitrate CRS* in *methanol R*, then dilute to 5 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *ammonium acetate solution R, dioxan R, methanol R* (20:40:40 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To about 30 mg in a porcelain crucible add 0.3 g of *anhydrous sodium carbonate R*. Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of *dilute nitric acid R* and filter. To 1 mL of the filtrate add 1 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS
Solution S
Dissolve 0.20 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in 3.2 mL of *methanol R*. Add 3.0 mL of *acetonitrile R* and dilute to 10.0 mL with a solution of *ammonium acetate R* (6.0 g in 380 mL of *water R*).

Reference solution (a) Dissolve 2.5 mg of *isoconazole CRS* and 2.5 mg of *econazole nitrate CRS* in the mobile phase, then dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Dissolve 6.0 g of *ammonium acetate R* in a mixture of 300 mL of *acetonitrile R*, 320 mL of *methanol R* and 380 mL of *water R*.

Flow rate 2 ml/min.

Detection Spectrophotometer at 235 nm.

Equilibration With the mobile phase for about 30 min.

Injection 10 µL.

Run time 1.5 times the retention time of isoconazole.

Retention time Econazole = about 10 min; isoconazole = about 14 min.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to econazole and isoconazole; if necessary, adjust the composition of the mobile phase.

Limits:
— *impurities B, C, D*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

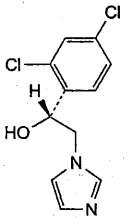
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 50 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Using 0.2 mL of *naphtholbenzein solution R* as indicator, titrate with *0.1 M perchloric acid* until the colour changes from orange-yellow to green.

1 mL of *0.1 M perchloric acid* is equivalent to 41.61 mg of $C_{18}H_{14}Cl_4N_2O$.

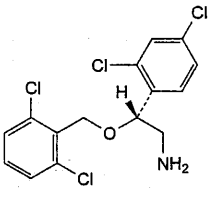
STORAGE
Protected from light.

IMPURITIES
Specified impurities *B, C, D.*



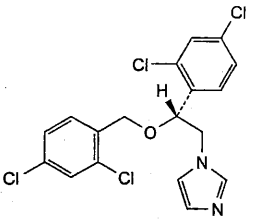
and enantiomer

B. (1*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl) ethanol,



and enantiomer

C. (2*RS*)-2-[(2,6-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethanamine,



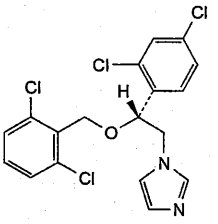
and enantiomer

D. 1-[(2*RS*)-2-[(2,4-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole.

Ph Eur

Isoconazole Nitrate

(Ph. Eur. monograph 1017)



and enantiomer , HNO_3

$C_{18}H_{15}Cl_4N_3O_4$ 479.1 *24168-96-5*

Action and use
Antifungal.

Ph Eur

DEFINITION
1-[(2*RS*)-2-[(2,6-Dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole nitrate.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white powder.

Solubility
Very slightly soluble in water, soluble in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B.

Second identification: A, C, D.

A. Melting point (*2.2.14*): 178 °C to 182 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *isoconazole nitrate CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 30 mg of the substance to be examined in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 30 mg of *isoconazole nitrate CRS* in *methanol R* and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 30 mg of *isoconazole nitrate CRS* and 30 mg of *econazole nitrate CRS* in *methanol R*, then dilute to 5 mL with the same solvent.

Plate *TLC octadecylsilyl silica gel plate R.*

Mobile phase *ammonium acetate solution R, dioxan R, methanol R* (20:40:40 *V/V/V*).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability Reference solution (b):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives the reaction of nitrates (*2.3.1*).

TESTS

Solution S

Dissolve 0.20 g in *methanol R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Optical rotation (*2.2.7*)

−0.10° to + 0.10°, determined on solution S.

Related substances

Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.5 mg of *isoconazole nitrate CRS* and 2.5 mg of *econazole nitrate CRS* in the mobile phase, then dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— *size*: *l* = 0.1 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase Dissolve 6.0 g of *ammonium acetate R* in a mixture of 300 mL of *acetonitrile R*, 320 mL of *methanol R* and 380 mL of *water R*.

Flow rate 2 mL/min.

Detection Spectrophotometer at 235 nm.

Equilibration With the mobile phase for about 30 min.

Injection 10 µL.

Run time 1.5 times the retention time of isoconazole.

Retention time Econazole = about 10 min; isoconazole = about 14 min.

System suitability Reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to econazole and isoconazole; if necessary, adjust the composition of the mobile phase.

Limits:
— *impurities A, B, C*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to the nitrate ion.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 75 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

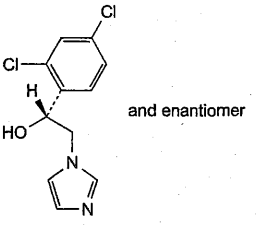
1 mL of *0.1 M perchloric acid* is equivalent to 47.91 mg of $C_{18}H_{15}Cl_4N_3O_4$.

STORAGE

Protected from light.

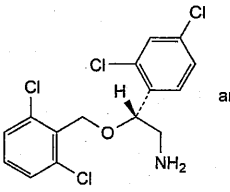
IMPURITIES

Specified impurities *A, B, C.*



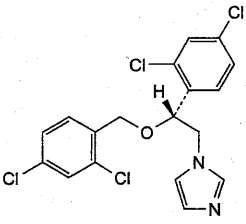
and enantiomer

A. (1*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl) ethanol,



and enantiomer

B. (2*RS*)-2-[(2,6-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl) ethanamine,



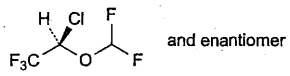
and enantiomer

C. 1-[(2*RS*)-2-[(2,4-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole.

Ph Eur

Isoflurane

(Ph. Eur. monograph 1673)



and enantiomer

$C_3H_2ClF_5O$ 184.5 *26675-46-7*

Action and use
General anaesthetic.

Ph Eur

DEFINITION

(2*RS*)-2-Chloro-2-(difluoromethoxy)-1,1,1-trifluoroethane.

CHARACTERS

Appearance
Clear, colourless, mobile, heavy liquid.

Solubility
Practically insoluble in water, miscible with ethanol.

bp
About 48 °C.

It is non-flammable.

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Preparation Examine the substance in the gaseous state.

Comparison *Ph. Eur. reference spectrum of isoflurane.*

TESTS

Acidity or alkalinity
To 20 mL add 20 mL of *carbon dioxide-free water R*, shake for 3 min and allow to stand. Collect the upper layer and add 0.2 mL of *bromocresol purple solution R*. Not more than 0.1 mL of *0.01 M sodium hydroxide* or 0.6 mL of *0.01 M hydrochloric acid* is required to change the colour of the indicator.

Related substances
Gas chromatography (*2.2.28*).

Test solution The substance to be examined.

Reference solution To 80 mL of *anhydrous ethanol R*, add 1.0 mL of the substance to be examined and 1.0 mL of *acetone R*, avoiding loss by evaporation. Dilute to 100.0 mL with *anhydrous ethanol R*. Dilute 1.0 mL of the solution to 100.0 mL with *anhydrous ethanol R*.

Column:
— *material*: fused silica,
— *size*: *l* = 30 m, Ø = 0.32 mm,
— *stationary phase*: *macrogol 20 000 R* (film thickness 0.25 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1.0 mL/min.

Split ratio 1:25.

Temperature:
— *column*: 35 °C,
— *injection port*: 150 °C,
— *detector*: 250 °C.

Detection Flame ionisation.

Injection 1.0 µL of each solution and 1.0 µL of *anhydrous ethanol R* as a blank.

Run time Until elution of the ethanol peak in the chromatogram obtained with the reference solution.

Relative retention With reference to isoflurane (retention time = about 3.8 min): acetone = about 0.75.

System suitability Reference solution:
— *resolution*: minimum of 5 between the peaks due to acetone and to isoflurane,
— *repeatability*: maximum relative standard deviation 15.0 per cent for the peak due to isoflurane after 3 injections.

Limits:
— *acetone*: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.01 per cent),
— *any other impurity*: not more than the area of the peak due to isoflurane in the chromatogram obtained with the reference solution (0.01 per cent),
— *total*: not more than 3 times the area of the peak due to isoflurane in the chromatogram obtained with the reference solution (0.03 per cent),
— *disregard limit*: 0.1 times the area of the peak due to isoflurane in the chromatogram obtained with the reference solution (0.001 per cent).

Chlorides (*2.4.4*)
Maximum 10 ppm.

To 10 mL add 10 mL of *0.01 M sodium hydroxide* and shake for 3 min. To 5 mL of the upper layer add 10 mL of *water R*.

Fluorides
Maximum 10 ppm.

Determine by potentiometry (*2.2.36, Method I*) using a fluoride-selective indicator-electrode and a silver-silver chloride reference electrode.

Test solution To 10.0 mL in a separating funnel, add 10 mL of a mixture of 30.0 mL of *dilute ammonia R2* and 70.0 mL of *distilled water R*. Shake for 1 min and collect the upper layer. Repeat this extraction procedure twice collecting the upper layer each time. Adjust the combined upper layers to pH 5.2 using *dilute hydrochloric acid R*. Add 5.0 mL of *fluoride standard solution (1 ppm F) R* and dilute to 50.0 mL with *distilled water R*. To 20.0 mL of the solution add 20.0 mL of *total-ionic-strength-adjustment buffer R* and dilute to 50.0 mL with *distilled water R*.

Reference solutions To each of 5.0 mL, 4.0 mL, 3.0 mL, 2.0 mL and 1.0 mL of *fluoride standard solution (10 ppm F) R* add 20.0 mL of *total-ionic-strength-adjustment buffer R* and dilute to 50.0 mL with *distilled water R*.

Carry out the measurements on 20 mL of each solution. Calculate the concentration of fluorides using the calibration curve, taking into account the addition of fluoride to the test solution.

Non-volatile matter
Maximum 200 mg/L.

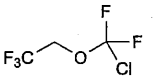
Evaporate 10.0 mL to dryness with the aid of a stream of cold air and dry the residue at 50 °C for 2 h. The residue weighs a maximum of 2.0 mg.

Water (*2.5.12*)
Maximum 1.0 mg/mL, determined on 10.0 mL.

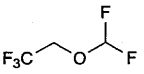
STORAGE

In an airtight container, protected from light.

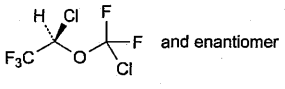
IMPURITIES



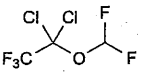
A. 2-(chlorodifluoromethoxy)-1,1,1-trifluoroethane,



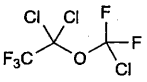
B. 2-(difluoromethoxy)-1,1,1-trifluoroethane,



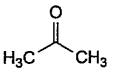
C. (2*RS*)-2-chloro-2-(chlorodifluoromethoxy)-1,1,1-trifluoroethane,



D. 1,1-dichloro-1-(difluoromethoxy)-2,2,2-trifluoroethane,



E. 1,1-dichloro-1-(chlorodifluoromethoxy)-2,2,2-trifluoroethane,

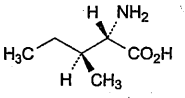


F. propanone (acetone).

Ph Eur

Isoleucine

(Ph. Eur. monograph 0770)



$C_6H_{13}NO_2$ 131.2 *73-32-5*

Action and use
Amino acid.

Ph Eur

DEFINITION

(2*S*,3*S*)-2-Amino-3-methylpentanoic acid.

Product of fermentation or of protein hydrolysis.

Content
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or flakes.

Solubility
Sparingly soluble in water, slightly soluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification: A, B.

Second identification: A, C.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *isoleucine CRS.*

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 10 mg of the substance to be examined in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 50 mL with the same solution.

Reference solution Dissolve 10 mg of *isoleucine CRS* in a 10.3 g/L solution of *hydrochloric acid R* and dilute to 50 mL with the same solution.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, water R, butanol R* (20:20:60 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with *ninhydrin solution R* and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_6 (*2.2.2, Method II*).

Dissolve 0.5 g in a 103 g/L solution of *hydrochloric acid R* and dilute to 10 mL with the same solution.

Specific optical rotation (*2.2.7*)
+ 40.0 to + 43.0 (dried substance).

Dissolve 1.00 g in *hydrochloric acid R1* and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances
Amino acid analysis (*2.2.56*). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter *2.2.46* are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A *dilute hydrochloric acid R1* or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *valine R* (impurity A) in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (d) Dissolve 30.0 mg of *leucine R* (impurity C) in solution A and dilute to 100.0 mL with

solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (e) Dilute 6.0 mL of *ammonium standard solution (100 ppm NH_4) R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (f) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* (impurity C) in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test solution, blank solution and reference solutions (a), (b), (c), (d) and (f) into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (f):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and impurity C.

Calculation of percentage contents:
— for impurity A, use the concentration of impurity A in reference solution (b);
— for impurity C, use the concentration of impurity C in reference solution (d);
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of isoleucine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (c); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:
— *impurities A and C at 570 nm*: for each impurity, maximum 0.3 per cent;
— *any ninhydrin-positive substance*: for each impurity, maximum 0.2 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold*: 0.05 per cent.

The thresholds indicated under Related Substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use (2034)* do not apply.

Chlorides (*2.4.4*)
Maximum 200 ppm.

Dissolve 0.25 g in *water R* and dilute to 15 mL with the same solvent.

Sulfates (*2.4.13*)
Maximum 300 ppm.

Dissolve 0.5 g in 3 mL of *dilute hydrochloric acid R* and dilute to 15 mL with *distilled water R*.

Ammonium
Amino acid analysis (*2.2.56*) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (e) and blank solution.

Limit:
— *ammonium at 570 nm*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (e) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (*2.4.9*)
Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of *dilute hydrochloric acid R*. Shake with 3 quantities, each of 10 mL, of *methyl isobutyl ketone R1*, shaking for 3 min each time. To the combined organic layers add 10 mL of *water R* and shake for 3 min. Use the aqueous layer.

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of *anhydrous formic acid R*. Add 30 mL of *anhydrous acetic acid R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 13.12 mg of $C_6H_{13}NO_2$.

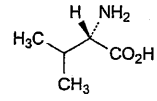
STORAGE

Protected from light.

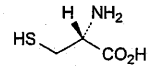
IMPURITIES

Specified impurities *A, C.*

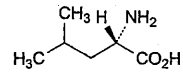
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, D.*



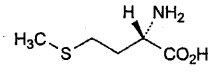
A. (2*S*)-2-amino-3-methylbutanoic acid (valine),



B. (2*R*)-2-amino-3-sulfanylpropanoic acid (cysteine),



C. (2*S*)-2-amino-4-methylpentanoic acid (leucine),

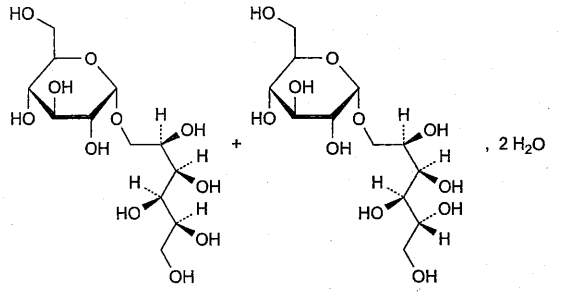


D. (2*S*)-2-amino-4-(methylsulfanyl)butanoic acid (methionine).

Ph Eur

Isomalt[1]

(Ph. Eur. monograph 1531)



$C_{12}H_{24}O_{11}$ 344.3

$C_{12}H_{24}O_{11},2H_2O$ 380.3Anhydrous isomalt *64519-82-0*

Action and use
Sweetening agent.

Ph Eur

DEFINITION
Mixture of 6-*O*-α-D-glucopyranosyl-D-glucitol (6-*O*-α-D-glucopyranosyl-D-sorbitol; 1,6-GPS) and 1-*O*-α-D-glucopyranosyl-D-mannitol (1,1-GPM).

Content
98.0 per cent to 102.0 per cent for the mixture of 1,6-GPS and 1,1-GPM and neither of the 2 components is less than 3.0 per cent (anhydrous substance).

♦CHARACTERS
Appearance
White or almost white powder or granules.

Solubility
Freely soluble in water, practically insoluble in anhydrous ethanol.♦

IDENTIFICATION
First identification: A.

◊Second identification: B, C.◊

A. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 2 principal peaks in the chromatogram obtained with reference solution (a).

◊B. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 50 mg of the substance to be examined in *water R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of *isomalt CRS* in *water R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel F_{254} plate R.*

Mobile phase *acetic acid R, propionic acid R, water R, ethyl acetate R, pyridine R* (5:5:10:50:50 *V/V/V/V/V*).

Application 1 µL; thoroughly dry the points of application in warm air.

Development Over 1/2 of the plate.

Drying In a current of warm air.

Detection Dip for 3 s in a 1 g/L solution of *sodium periodate R* and dry in a current of hot air; dip for 3 s in a mixture of 1 volume of *acetic acid R*, 1 volume of *anisaldehyde R*, 5 volumes of *sulfuric acid R* and 90 volumes of *anhydrous ethanol R*; dry in a current of hot air until coloured spots become visible; the background colour may be brightened in warm steam; examine in daylight.

Results The chromatogram obtained with the reference solution shows 2 blue-grey spots with R_F values of about 0.13 (1,6-GPS) and 0.16 (1,1-GPM). The chromatogram obtained with the test solution shows principal spots similar in position and colour to the principal spots in the chromatogram obtained with the reference solution.

C. To 3 mL of a freshly prepared 100 g/L solution of *pyrocatechol R* add 6 mL of *sulfuric acid R* while cooling in iced water. To 3 mL of the cooled mixture add 0.3 mL of a 100 g/L solution of the substance to be examined. Heat gently over a naked flame for about 30 s. A pink colour develops.◊

TESTS
Conductivity (*2.2.38*)
Maximum 20 µS·cm^{-1}.

Dissolve 20.0 g in *carbon dioxide-free water R* with gentle heating (40-50 °C) and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Reducing sugars
Maximum 0.3 per cent, expressed as glucose equivalent.

Dissolve 3.3 g in 10 mL of *water R* with gentle heating. Cool and add 20 mL of *cupri-citric solution R* and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent *V/V* solution of *glacial acetic acid R* and 20.0 mL of *0.025 M iodine*. With continuous shaking, add 25 mL of a mixture of 6 volumes of *hydrochloric acid R* and 94 volumes of *water R*. When the precipitate has dissolved, titrate the excess of iodine with *0.05 M sodium thiosulfate* using 1 mL of *starch solution R* as indicator, added towards the end of the titration. Not less than 12.8 mL of *0.05 M sodium thiosulfate* is required.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 0.200 g of the substance to be examined in 4 mL of *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.200 g of *isomalt CRS* in 4 mL of *water R* and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dissolve 10.0 mg of *sorbitol CRS* (impurity C) and 10.0 mg of *mannitol CRS* (impurity B) in 20 mL of *water R* and dilute to 100.0 mL with the same solvent.

Precolumn:
— *size*: *l* = 30 mm, Ø = 4.6 mm;
— *stationary phase*: *strong cation-exchange resin (calcium form) R* (9 µm);
— *temperature*: 80 ± 3 °C.

Column:
— *size*: *l* = 0.3 m, Ø = 7.8 mm;
— *stationary phase*: *strong cation-exchange resin (calcium form) R* (9 µm);
— *temperature*: 80 ± 3 °C.

Mobile phase Degassed *water for chromatography R*.

Flow rate 0.5 mL/min.

[1] *This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial harmonisation.*

Detection Differential refractometer maintained at a constant temperature (e.g. 40 °C).

Injection 20 µL.

Run time 2.5 times the retention time of 1,1-GPM.

Relative retention With reference to 1,1-GPM (retention time = about 14 min): 1,6-GPS = about 1.2; impurity B = about 1.6; impurity C = about 2.0.

System suitability Reference solution (a):
— *resolution*: minimum 2.0 between the peaks due to 1,1-GPM and 1,6-GPS.

Limits:
— *impurities B, C*: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the peak due to impurity C in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *total*: not more than 4 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (b) (2.0 per cent);
— *disregard limit*: 0.2 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (b) (0.1 per cent).

Water (*2.5.12*)
Maximum 7.0 per cent, determined on 0.300 g. As solvent, use a mixture of 20 mL of *anhydrous methanol R* and 20 mL of *formamide R1* at 50 ± 5 °C.

ASSAY
Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modification.

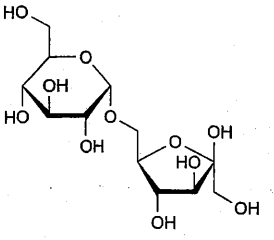
Injection Test solution and reference solution (a).

Calculate the percentage content of isomalt (1,1-GPM and 1,6-GPS) taking into account the assigned contents of 1,1-GPM and 1,6-GPS in *isomalt CRS*.

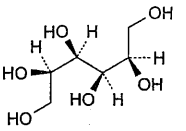
LABELLING
The label states the percentage contents of 1,1-GPM and 1,6-GPS.

IMPURITIES
Specified impurities *B, C.*

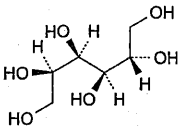
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, D.*



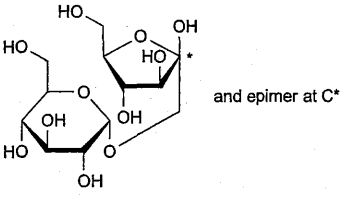
A. 6-*O*-α-D-glucopyranosyl-β-D-*arabino*-hex-2-ulofuranose (isomaltulose),



B. D-mannitol,



C. D-glucitol (D-sorbitol),

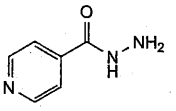


D. 1-*O*-α-D-glucopyranosyl-D-*arabino*-hex-2-ulofuranose (trehalulose).

Ph Eur

Isoniazid

(Ph. Eur. monograph 0146)



$C_6H_7N_3O$ 137.1 *54-85-3*

Action and use
Antituberculosis drug.

Preparations
Isoniazid Injection
Isoniazid Oral Solution
Isoniazid Tablets

Ph Eur

DEFINITION
Pyridine-4-carbohydrazide.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS
Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION
First identification: A, B.

Second identification: A, C.

A. Melting point (*2.2.14*): 170 °C to 174 °C.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *isoniazid CRS.*

C. Dissolve 0.1 g in 2 mL of *water R* and add 10 mL of a warm 10 g/L solution of *vanillin R*. Allow to stand and scratch the wall of the test tube with a glass rod. A yellow precipitate is formed, which, after recrystallisation from 5 mL of *ethanol (70 per cent V/V) R* and drying at 100-105 °C, melts (*2.2.14*) at 226 °C to 231 °C.

TESTS

Solution S

Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

pH (*2.2.3*)

6.0 to 8.0 for solution S.

Impurity E

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions.*

Solvent mixture *acetonitrile R, water R* (50:50 *V/V*).

Solution A To 2.0 mL of *benzaldehyde R* add *acetonitrile R* and dilute to 100.0 mL with the same solvent. Use the solution within 4 h.

Test solution Dissolve 50.0 mg of the substance to be examined in 1.0 mL of *water R*, add 5.0 mL of solution A and shake well. Wait for 45 min for completion of derivatisation and dilute to 10.0 mL with the solvent mixture.

Reference solution Dissolve 20.0 mg of *hydrazine sulfate R* (equivalent to 4.925 mg of impurity E) in *water R* and dilute to 50.0 mL with the same solvent. Dilute 2.5 mL of the solution to 100.0 mL with *water R*. To 1.0 mL of this solution add 2.5 mL of solution A and shake well. Wait for 45 min for completion of derivatisation and dilute to 25.0 mL with the solvent mixture. Dilute 7.5 mL of this solution to 10.0 mL with the solvent mixture.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water for chromatography R, acetonitrile R* (40:60 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 300 nm.

Injection 10 µL.

Run time 1.5 times the retention time of benzaldehyde azine.

Retention time Benzaldehyde azine = about 13 min.

System suitability Reference solution:
- *repeatability*: maximum relative standard deviation of 5.0 per cent determined on 6 injections.

Calculation of percentage content:
- for impurity E, use the concentration of hydrazine sulfate in the reference solution.

Limit:
- *impurity E*: maximum 15 ppm.

Related substances

Liquid chromatography (*2.2.29*). *Use freshly prepared solutions.*

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 25.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of *isonicotinic acid R* (impurity A), 5 mg of *isonicotinamide R* (impurity B) and 5 mg of *nicotinoyl hydrazide R* (impurity D) in mobile phase A and dilute to 50.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Column:
- *size*: *l* = 0.25 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase:
- *mobile phase A*: mix 3 volumes of *methanol R* and 97 volumes of a buffer solution prepared as follows: dissolve 13.6 g of *potassium dihydrogen phosphate R* in 950 mL of *water for chromatography R*, adjust to pH 6.9 with *strong sodium hydroxide solution R*, add 30 mg of *triethanolamine R* and dilute to 1000 mL with *water for chromatography R*;
- *mobile phase B*: *methanol R*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 12	100	0
12 - 20	100 → 85	0 → 15
20 - 28	85	15

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and D.

Relative retention With reference to isoniazid (retention time = about 8.4 min): impurity A = about 0.5; impurity D = about 1.15; impurity B = about 1.4.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 1.8, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to isoniazid.

Calculation of percentage contents:
- *correction factors*: multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.4; impurity B = 1.5;
- for each impurity, use the concentration of isoniazid in reference solution (a).

Limits:
- *impurities A, B*: for each impurity, maximum 0.15 per cent;
- *unspecified impurities*: for each impurity, maximum 0.10 per cent;
- *total*: maximum 0.5 per cent;
- *reporting threshold*: 0.05 per cent.

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in *water R* and dilute to 100.0 mL with the same solvent. To 20.0 mL of the solution add 100 mL of *water R*, 20 mL of *hydrochloric acid R*, 0.2 g of *potassium*

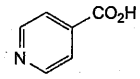
bromide R and 0.05 mL of *methyl red solution R*. Titrate dropwise with *0.0167 M potassium bromate*, shaking continuously, until the red colour disappears.

1 mL of *0.0167 M potassium bromate* is equivalent to 3.429 mg of $C_6H_7N_3O$.

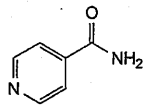
IMPURITIES

Specified impurities *A, B, E.*

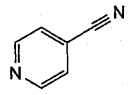
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, D.*



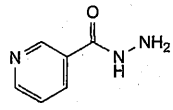
A. pyridine-4-carboxylic acid (isonicotinic acid),



B. pyridine-4-carboxamide (isonicotinamide),



C. pyridine-4-carbonitrile (isonicotinonitrile),



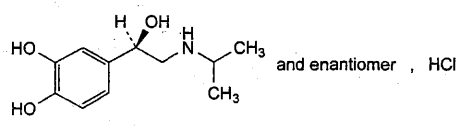
D. pyridine-3-carbohydrazide (nicotinoyl hydrazide),

E. hydrazine.

Ph Eur

Isoprenaline Hydrochloride

(Ph. Eur. monograph 1332)



$C_{11}H_{18}ClNO_3$ 247.7 *51-30-9*

Action and use
Adrenoceptor agonist.

Preparation
Isoprenaline Injection

Ph Eur

DEFINITION

(1*RS*)-1-(3,4-Dihydroxyphenyl)-2-[(1-methylethyl) amino]ethanol hydrochloride.

Content
98.0 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: B, C, E.

Second identification: A, C, D, E.

A. Melting point (*2.2.14*): 165 °C to 170 °C, with decomposition.

B. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *isoprenaline hydrochloride CRS.*

C. Optical rotation (see Tests).

D. To 0.1 mL of solution S (see Tests) add 0.05 mL of *ferric chloride solution R1* and 0.9 mL of *water R*. A green colour is produced. Add dropwise *sodium hydrogen carbonate solution R*. The colour becomes blue and then red.

E. To 0.5 mL of solution S add 1.5 mL of *water R*. The solution gives reaction (a) of chlorides (*2.3.1*).

TESTS

Prepare the solutions immediately before use.

Solution S
Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to 25.0 mL with the same solvent.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution B_7 or BY_7 (*2.2.2, Method II*).

pH (*2.2.3*)
4.3 to 5.5.

Mix 5 mL of solution S and 5 mL of *carbon dioxide-free water R*.

Optical rotation (*2.2.7*)
−0.10° to + 0.10°, determined on solution S.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2.5 mg of *orciprenaline sulfate CRS* in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (c) To 5.0 mL of reference solution (a) add 5.0 mL of reference solution (b).

Reference solution (d) Dissolve 6.0 mg of *isoprenaline impurity A CRS* in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 50.0 mL with the mobile phase.

Column:
— *size:* l = 0.125 m, Ø = 4.0 mm;

— *stationary phase*: *end-capped octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *methanol R*, 11.5 g/L solution of *phosphoric acid R* (5:95 *V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 7 times the retention time of isoprenaline.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (b) to identify the peak due to orciprenaline.

Relative retention With reference to isoprenaline (retention time = about 3 min): orciprenaline = about 1.5; impurity A = about 1.8. If necessary, adjust the concentration of methanol in the mobile phase.

System suitability Reference solution (c):
— *resolution*: minimum 3.0 between the peaks due to isoprenaline and orciprenaline.

Limits:
— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— *total*: maximum 1.0 per cent;
— *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 1.0 per cent, determined on 1.000 g by drying *in vacuo* at 15-25 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.150 g in 10 mL of *anhydrous formic acid R* and add 50 mL of *acetic anhydride R*. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically (*2.2.20*).

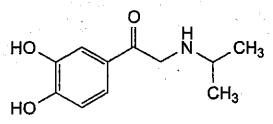
1 mL of *0.1 M perchloric acid* is equivalent to 24.77 mg of $C_{11}H_{18}ClNO_3$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities *A*.



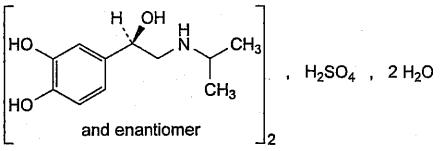
A. 1-(3,4-dihydroxyphenyl)-2-[(1-methylethyl) amino]ethanone.

Ph Eur

Isoprenaline Sulfate

Isoprenaline Sulphate

(Ph. Eur. monograph 0502)



$C_{22}H_{36}N_2O_{10}S,2H_2O$ 556.6 *6700-39-6*

Action and use
Adrenoceptor agonist.

Ph Eur

DEFINITION

Bis[(1*RS*)-1-(3,4-dihydroxyphenyl)-2-[(1-methylethyl)amino] ethanol] sulfate dihydrate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Freely soluble in water, very slightly soluble in ethanol (96 per cent).

mp
About 128 °C, with decomposition.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Dissolve 0.5 g in 1.5 mL of *water R* and add 3.5 mL of *2-propanol R*. Scratch the wall of the tube with a glass rod to initiate crystallisation. Collect the crystals and dry *in vacuo* at 60 °C over *diphosphorus pentoxide R*.

Comparison Repeat the operations using 0.5 g of *isoprenaline sulfate CRS*.

B. To 0.1 mL of solution S (see Tests) add 0.9 mL of *water R* and 0.05 mL of *ferric chloride solution R1*. A green colour is produced. Add dropwise *sodium hydrogen carbonate solution R*. The colour becomes blue and then red.

C. Dilute 1 mL of solution S to 10 mL with *water R* and add 0.25 mL of *silver nitrate solution R1*. A shining, grey, fine precipitate is formed within 10 min and the solution becomes pink.

D. Solution S gives reaction (a) of sulfates (*2.3.1*).

TESTS

Solution S
Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to 50 mL with the same solvent. Use within 2 h of preparation.

Appearance of solution
Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution Y_6 (*2.2.2, Method II*).

pH (*2.2.3*)
4.3 to 5.5.

Dilute 5 mL of solution S to 10 mL with *carbon dioxide-free water R*.

Isoprenalone
The absorbance (*2.2.25*) is not greater than 0.20 at 310 nm.
Dissolve 0.20 g in *0.005 M sulfuric acid* and dilute to 100.0 mL with the same acid.

Water (*2.5.12*)
5.0 per cent to 7.5 per cent, determined on 0.200 g.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

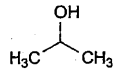
ASSAY
Dissolve 0.400 g in 20 mL of *anhydrous acetic acid R*, warming gently if necessary and add 20 mL of *methyl isobutyl ketone R*. Titrate with *0.1 M perchloric acid* determining the end-point potentiometrically (*2.2.20*).
1 mL of *0.1 M perchloric acid* is equivalent to 52.06 mg of $C_{22}H_{36}N_2O_{10}S$.

STORAGE
In an airtight container, protected from light.

Ph Eur

Isopropyl Alcohol

(Ph. Eur. monograph 0970)



C_3H_8O 60.1 *67-63-0*

Ph Eur

DEFINITION
Propan-2-ol.

CHARACTERS
Appearance
Clear, colourless liquid.

Solubility
Miscible with water and with ethanol (96 per cent).

IDENTIFICATION
First identification: B, C.
Second identification: A, B, D.
A. Relative density (*2.2.5*): 0.785 to 0.789.
B. Refractive index (*2.2.6*): 1.376 to 1.379.
C. Infrared absorption spectrophotometry (*2.2.24*).
Comparison *isopropyl alcohol CRS.*
D. To 1 mL add 4 mL of *water R* and mix. Carefully add 2 mL of a 10 g/L solution of *dimethylaminobenzaldehyde R* in *sulfuric acid R*, ensuring that the liquids do not mix; a bright reddish-violet ring forms immediately at the junction of the 2 liquids. After 2-5 min, the entire sulfuric acid layer turns violet.

TESTS
Appearance
The substance to be examined is clear (*2.2.1*) and colourless (*2.2.2, Method II*). Dilute 1 mL to 20 mL with *water R*. After 5 min, the solution is clear (*2.2.1*).

Acidity or alkalinity
Gently boil 25 mL for 5 min. Add 25 mL of *carbon dioxide-free water R* and allow to cool protected from carbon dioxide in the air. Add 0.1 mL of *phenolphthalein solution R*.
The solution is colourless. Not more than 0.6 mL of *0.01 M sodium hydroxide* is required to change the colour of the indicator to pale pink.

Absorbance (*2.2.25*)
Maximum 0.30 at 230 nm, 0.10 at 250 nm, 0.03 at 270 nm, 0.02 at 290 nm and 0.01 at 310 nm.
The absorbance is measured between 230 nm and 310 nm using *water R* as the compensation liquid. The spectrum shows a steadily descending curve with no observable peaks or shoulders.

Benzene and related substances
Gas chromatography (*2.2.28*).
Test solution (a) The substance to be examined.
Test solution (b) Dilute 1.0 mL of *2-butanol R1* to 50.0 mL with test solution (a). Dilute 5.0 mL of this solution to 100.0 mL with test solution (a).
Reference solution (a) Dilute 0.5 mL of *2-butanol R1* and 0.5 mL of *propanol R* to 50.0 mL with test solution (a). Dilute 5.0 mL of this solution to 50.0 mL with test solution (a).
Reference solution (b) Dilute 100 µL of *benzene R* to 100.0 mL with test solution (a). Dilute 0.20 mL of this solution to 100.0 mL with test solution (a).
Column:
— *material:* fused silica;
— *size:* l = 30 m, Ø = 0.32 mm;
— *stationary phase: poly[(cyanopropyl)(phenyl)][dimethyl] siloxane R* (film thickness 1.8 µm).
Carrier gas *helium for chromatography R.*
Auxiliary gas *nitrogen for chromatography R* or *helium for chromatography R.*
Linear velocity 35 cm/s.
Split ratio 1:5.
Temperature:

	Time (min)	Temperature (°C)
Column	0 - 12	40
	12 - 32	40 → 240
	32 - 42	240
Injection port		280
Detector		280

Detection Flame ionisation.
Injection 1 µL.
Retention time Benzene = about 10 min.
System suitability Reference solution (a):
— *resolution:* minimum 10 between the 1st peak (propanol) and the 2nd peak (2-butanol).
Limits:
— *benzene* (test solution (a)): not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (2 ppm), after the sensitivity has been adjusted so that the height of the peak due to benzene in the chromatogram obtained with reference solution (b) represents at least 10 per cent of the full scale of the recorder;
— *total of impurities apart from 2-butanol* (test solution (b)): not more than 3 times the area of the peak due to 2-butanol in the chromatogram obtained with test solution (b) (0.3 per cent), after the sensitivity has been adjusted so that the height of the 2 peaks following the principal peak in the chromatogram obtained with

reference solution (a) represents at least 50 per cent of the full scale of the recorder.

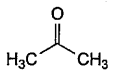
Peroxides
In a 12 mL test-tube with a ground-glass stopper and a diameter of about 15 mm, introduce 8 mL of *potassium iodide and starch solution R*. Fill completely with the substance to be examined. Shake vigorously and allow to stand protected from light for 30 min. No colour develops.

Non-volatile substances
Maximum 20 ppm.

Evaporate 100 g to dryness on a water-bath *after having verified that it complies with the test for peroxides* and dry in an oven at 100-105 °C. The residue weighs a maximum of 2 mg.

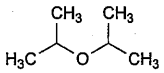
Water (*2.5.12*)
Maximum 0.5 per cent, determined on 5.0 g.

STORAGE
Protected from light.

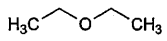
IMPURITIES


A. propanone (acetone),

B. benzene,

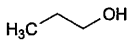


C. 2-(1-methylethoxy)propane (diisopropyl ether),



D. ethoxyethane (diethyl ether),

E. methanol,



F. propan-1-ol (*n*-propanol).

Ph Eur

Isopropyl Isostearate

(Ph. Eur. monograph 2867)

Ph Eur

DEFINITION
Product obtained by esterification of isostearic acid (regioisomers mainly of octadecanoic acid, a complex mixture of branched and straight-chain fatty acids) with isopropyl alcohol. The monograph applies to isopropyl isostearate for cutaneous use.

CHARACTERS
Appearance
Clear, colourless or pale yellow liquid.

Solubility
Practically insoluble in water, miscible with ethanol (96 per cent) and with fatty oils.

Viscosity
About 11 mPa·s at 20 °C.

Relative density
About 0.866 at 20 °C.

Refractive index
About 1.447 at 20 °C.

IDENTIFICATION
A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison isopropyl isostearate CRS.

An additional weak absorption band at 1780 cm^{-1} may be present.

B. Saponification value (see Tests).

C. Composition of fatty acids (see Tests).

TESTS
Appearance of solution
The solution is not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 10.0 g in *ethanol (96 per cent) R* and dilute to 20 mL with the same solvent.

Acid value (*2.5.1*)
Maximum 1.0, determined on 20.0 g.

Iodine value (*2.5.4, Method B*)
Maximum 3.0, determined on 3.0 g.

Peroxide value (*2.5.5, Method B*)
Maximum 5.0.

Saponification value (*2.5.6*)
165 to 180, determined on 2.0 g.

Composition of fatty acids (*2.4.22, Method B*)
Use the mixture of calibrating substances in Table 2.4.22.-1.

Composition of the fatty-acid fraction of the substance:
— *sum of the contents of the fatty acids eluting between palmitic acid and stearic acid*: minimum 65.0 per cent;
— *sum of the contents of myristic acid, palmitic acid and stearic acid*: maximum 11.0 per cent.

Water (*2.5.12*)
Maximum 0.1 per cent, determined on 5.00 g.

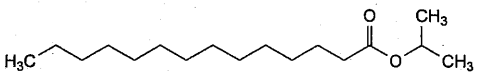
Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

STORAGE
In an airtight container.

Ph Eur

Isopropyl Myristate

(Ph. Eur. monograph 0725)



$C_{17}H_{34}O_2$ 270.5

Action and use
Excipient.

Ph Eur

DEFINITION
1-Methylethyl tetradecanoate together with variable amounts of other fatty acid isopropyl esters.

Content
Minimum 90.0 per cent of $C_{17}H_{34}O_2$.

CHARACTERS

Appearance
Clear, colourless, oily liquid.

Solubility
Immiscible with water, miscible with ethanol (96 per cent), with methylene chloride, with fatty oils and with liquid paraffin.

Relative density
About 0.853.

IDENTIFICATION
First identification: B.
Second identification: A, C.
A. Saponification value (see Tests).
B. Examine the chromatograms obtained in the assay.
Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.
C. Superpose 2 mL of a 1 g/L solution in *ethanol (96 per cent) R* on a freshly prepared solution of 20 mg of *dimethylaminobenzaldehyde R* in 2 mL of *sulfuric acid R*. After 2 min, a yellowish-red colour appears at the junction of the 2 liquids and gradually becomes red.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).
Dissolve 2.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Refractive index (*2.2.6*)
1.434 to 1.437.

Viscosity (*2.2.9*)
5 mPa·s to 6 mPa·s.

Acid value (*2.5.1*)
Maximum 1.0.

Iodine value (*2.5.4*)
Maximum 1.0.

Saponification value (*2.5.6*)
202 to 212.

Water (*2.5.12*)
Maximum 0.1 per cent, determined on 5.0 g.

Total ash (*2.4.16*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*).
Internal standard solution Dissolve 50.0 mg of *tricosane R* in *heptane R* and dilute to 250.0 mL with the same solvent.
Test solution Dissolve 20.0 mg of the substance to be examined in the internal standard solution and dilute to 100.0 mL with the same solution.
Reference solution Dissolve 20.0 mg of *isopropyl tetradecanoate CRS* in the internal standard solution and dilute to 100.0 mL with the same solution.
Column:
— *material*: fused silica,
— *size*: *l* = 50 m, Ø = 0.2 mm,
— *stationary phase*: *poly(cyanopropyl)siloxane R* (film thickness 0.2 µm).
Carrier gas *helium for chromatography R.*
Flow rate 1 mL/min.
Split ratio 1:40.
Temperature:

	Time (min)	Temperature (°C)
Column	0 - 6	125 → 185
	6 - 16	185
Injection port		250
Detector		250

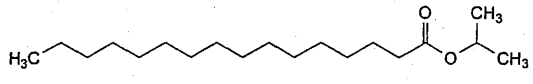
Detection Flame ionisation.
Injection 2 µL.
Calculate the percentage content of $C_{17}H_{34}O_2$ in the substance to be examined.

STORAGE
Protected from light.

Ph Eur

Isopropyl Palmitate

(Ph. Eur. monograph 0839)



$C_{19}H_{38}O_2$ 298.5

Action and use
Excipient.

Ph Eur

DEFINITION
1-Methylethyl hexadecanoate together with varying amounts of other fatty acid isopropyl esters.

Content
Minimum 90.0 per cent of $C_{19}H_{38}O_2$.

CHARACTERS

Appearance
Clear, colourless, oily liquid.

Solubility
Immiscible with water, miscible with ethanol (96 per cent), with methylene chloride, with fatty oils and with liquid paraffin.

Relative density
About 0.854.

IDENTIFICATION
First identification: B.
Second identification: A, C.
A. Saponification value (see Tests).
B. Examine the chromatograms obtained in the assay.
Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.
C. Superpose 2 mL of a 1 g/L solution in *ethanol (96 per cent) R* on a freshly prepared solution of 20 mg of *dimethylaminobenzaldehyde R* in 2 mL of *sulfuric acid R*. After

2 min, a yellowish-red colour appears at the junction of the 2 liquids which gradually becomes red.

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution Y_7 (*2.2.2, Method II*).

Dissolve 2.0 g in *methanol R* and dilute to 20 mL with the same solvent.

Refractive index (*2.2.6*)
1.436 to 1.440.

Viscosity (*2.2.9*)
5 mPa·s to 10 mPa·s.

Acid value (*2.5.1*)
Maximum 1.0.

Iodine value (*2.5.4*)
Maximum 1.0.

Saponification value (*2.5.6*)
183 to 193.

Water (*2.5.12*)
Maximum 0.1 per cent, determined on 5.0 g.

Total ash (*2.4.16*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Gas chromatography (*2.2.28*).

Internal standard solution Dissolve 50.0 mg of *tricosane R* in *heptane R* and dilute to 250.0 mL with the same solvent.

Test solution Dissolve 20.0 mg of the substance to be examined in the internal standard solution and dilute to 100.0 mL with the same solution.

Reference solution Dissolve 20.0 mg of *isopropyl hexadecanoate CRS* in the internal standard solution and dilute to 100.0 mL with the same solution.

Column:
— *material*: fused silica,
— *size*: *l* = 50 m, Ø = 0.2 mm,
— *stationary phase*: *poly(cyanopropyl)siloxane R* (film thickness 0.2 µm).

Carrier gas *helium for chromatography R.*

Flow rate 1 mL/min.

Split ratio 1:40.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 6	125 → 185
	6 - 16	185
Injection port		250
Detector		250

Detection Flame ionisation.

Injection 2 µL.

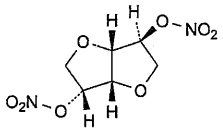
Calculate the percentage content of $C_{19}H_{38}O_2$ in the substance to be examined.

STORAGE
Protected from light.

Ph Eur

Diluted Isosorbide Dinitrate

(Ph. Eur. monograph 1117)



$C_6H_8N_2O_8$ 236.1

Action and use
Nitric acid analogue; treatment of angina pectoris.

Preparation
Isosorbide Dinitrate Tablets

Ph Eur

DEFINITION
Dry mixture of isosorbide dinitrate and *Lactose monohydrate (0187)* or *Mannitol (0559)*.

Content
95.0 per cent *m/m* to 105.0 per cent *m/m* of the content of 1,4:3,6-dianhydro-D-glucitol 2,5-dinitrate stated on the label.

CAUTION: undiluted isosorbide dinitrate may explode if subjected to percussion or excessive heat. Appropriate precautions must be taken and only very small quantities handled.

CHARACTERS

Appearance
Undiluted isosorbide dinitrate is a fine, white or almost white, crystalline powder.

Solubility
Undiluted isosorbide dinitrate is very slightly soluble in water, very soluble in acetone, sparingly soluble in ethanol (96 per cent).

The solubility of the diluted product depends on the diluent and its concentration.

IDENTIFICATION
First identification: A, C, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs prepared with the residue obtained in identification test D.

Comparison *isosorbide dinitrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 10 mg of isosorbide dinitrate with 10 mL of *ethanol (96 per cent) R* for 5 min and filter.

Reference solution Shake a quantity of *isosorbide dinitrate CRS* corresponding to 10 mg of isosorbide dinitrate with 10 mL of *ethanol (96 per cent) R* for 5 min and filter.

Plate *TLC silica gel G plate R.*

Mobile phase *methanol R, methylene chloride R* (5:95 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of air.

Detection Spray with freshly prepared *potassium iodide and starch solution R*; expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to

the principal spot in the chromatogram obtained with the reference solution.

C. Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 0.10 g of lactose or mannitol with 10 mL of *water R*. Filter if necessary.

Reference solution (a) Dissolve 0.10 g of *lactose monohydrate R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.10 g of *mannitol R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (c) Mix equal volumes of reference solutions (a) and (b).

Plate *TLC silica gel G plate R.*

Mobile phase *water R, methanol R, anhydrous acetic acid R, ethylene chloride R* (10:15:25:50 *V/V/V/V*); measure the volumes accurately since a slight excess of water produces cloudiness.

Application 1 µL; thoroughly dry the points of application.

Development A Over a path of 15 cm.

Drying A In a current of warm air.

Development B Immediately, over a path of 15 cm, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with *4-aminobenzoic acid solution R*, dry in a current of cold air until the acetone is removed, then heat at 100 °C for 15 min; allow to cool, spray with a 2 g/L solution of *sodium periodate R*, dry in a current of cold air, and heat at 100 °C for 15 min.

System suitability Reference solution (c):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a) for lactose or to the principal spot in the chromatogram obtained with reference solution (b) for mannitol.

D. Shake a quantity of the substance to be examined corresponding to 25 mg of isosorbide dinitrate with 10 mL of *acetone R* for 5 min. Filter, evaporate to dryness at a temperature below 40 °C and dry the residue over *diphosphorus pentoxide R* at a pressure of 0.7 kPa for 16 h. The melting point (*2.2.14*) of the residue is 69 °C to 72 °C.

TESTS

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 0.10 g of isosorbide dinitrate with 5 mL of *ethanol (96 per cent) R* and filter.

Reference solution Dissolve 10 mg of *potassium nitrate R* in 1 mL of *water R* and dilute to 100 mL with *ethanol (96 per cent) R*.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, acetone R, toluene R* (15:30:60 *V/V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of air until the acetic acid is completely removed.

Detection Spray copiously with freshly prepared *potassium iodide and starch solution R*; expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Limit:

— *impurity A*: any spot due to impurity A is not more intense than the principal spot in the chromatogram obtained with the reference solution (0.5 per cent, calculated as potassium nitrate).

Impurities B and C

Liquid chromatography (*2.2.29*).

Test solution (a) Sonicate a quantity of the substance to be examined corresponding to 25.0 mg of isosorbide dinitrate with 20 mL of the mobile phase for 15 min and dilute to 25.0 mL with the mobile phase. Filter the solution through a suitable membrane filter.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Sonicate a quantity of *isosorbide dinitrate CRS* corresponding to 25.0 mg of isosorbide dinitrate with 20 mL of the mobile phase for 15 min and dilute to 25.0 mL with the mobile phase. Filter the solution through a suitable membrane filter.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 10.0 mg of *isosorbide 2-nitrate CRS* (impurity B) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 0.1 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (d) Dissolve 10.0 mg of *isosorbide mononitrate CRS* (impurity C) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 0.1 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (e) Dissolve 5 mg of *isosorbide 2-nitrate CRS* (impurity B) in the mobile phase and dilute to 10 mL with the mobile phase. To 1 mL of this solution add 0.5 mL of reference solution (a) and dilute to 10 mL with the mobile phase.

Column:

— *size*: *l* = 0.25 m, Ø = 4.6 mm;

— *stationary phase*: *aminopropylmethylsilyl silica gel for chromatography R* (10 µm).

Mobile phase *anhydrous ethanol R, trimethylpentane R* (15:85 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 210-215 nm.

Injection 10 µL of test solution (a) and reference solutions (c), (d) and (e).

Retention time Isosorbide dinitrate = about 5 min; impurity B = about 8 min; impurity C = about 11 min.

System suitability Reference solution (e):

— *resolution*: minimum 6.0 between the peaks due to isosorbide dinitrate and impurity B.

Limits:

— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent).

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for impurities B and C with the following modifications.

Detection Spectrophotometer at 230 nm.

Injection 20 µL of test solution (b) and reference solution (b).

If the areas of the peaks from 2 successive injections of reference solution (b) do not agree to within 1.0 per cent, then inject a further 4 times and calculate, for the 6 injections, the relative standard deviation.

System suitability Reference solution (b):
— *repeatability*: maximum relative standard deviation of 2.0 per cent after 6 injections.

Calculate the content of isosorbide dinitrate as a percentage of the declared content.

STORAGE

Protected from light.

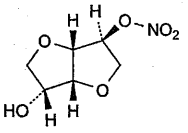
LABELLING

The label states the percentage content of isosorbide dinitrate.

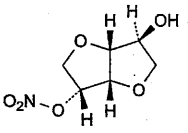
IMPURITIES

Specified impurities *A, B, C.*

A. inorganic nitrates,



B. 1,4:3,6-dianhydro-D-glucitol 2-nitrate (isosorbide 2-nitrate),

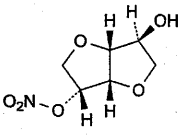


C. 1,4:3,6-dianhydro-D-glucitol 5-nitrate (isosorbide 5-nitrate, isosorbide mononitrate).

Ph Eur

Diluted Isosorbide Mononitrate

(Ph. Eur. monograph 1118)



$C_6H_9NO_6$ 191.1

Action and use
Nitric acid analogue; treatment of angina pectoris.

Preparations
Isosorbide Mononitrate Tablets
Isosorbide Mononitrate Prolonged-release Capsules
Isosorbide Mononitrate Prolonged-release Tablets

Ph Eur

DEFINITION

Dry mixture of isosorbide mononitrate and *Lactose monohydrate (0187)* or *Mannitol (0559)*.

Content

95.0 per cent *m/m* to 105.0 per cent *m/m* of the content of 1,4:3,6-dianhydro-D-glucitol 5-nitrate stated on the label.

CHARACTERS

Appearance
Undiluted isosorbide mononitrate is a white or almost white, crystalline powder.

Solubility
Undiluted isosorbide mononitrate is freely soluble in water, in acetone, in ethanol (96 per cent) and in methylene chloride.

The solubility of the diluted product depends on the diluent and its concentration.

IDENTIFICATION

First identification: A, C, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs prepared with the residue obtained in identification test D.

Comparison *isosorbide mononitrate CRS.*

B. Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 10 mg of isosorbide mononitrate with 10 mL of *ethanol (96 per cent) R* for 5 min and filter.

Reference solution Dissolve 10 mg of *isosorbide mononitrate CRS* in *ethanol (96 per cent) R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *methanol R, methylene chloride R* (5:95 *V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of air.

Detection Spray with freshly prepared *potassium iodide and starch solution R.* Expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 0.10 g of lactose or mannitol with 10 mL of *water R*; filter if necessary.

Reference solution (a) Dissolve 0.10 g of *lactose monohydrate R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.10 g of *mannitol R* in *water R* and dilute to 10 mL with the same solvent.

Reference solution (c) Mix equal volumes of reference solutions (a) and (b).

Plate *TLC silica gel G plate R.*

Mobile phase *water R, methanol R, anhydrous acetic acid R, ethylene chloride R* (10:15:25:50 *V/V/V/V*); measure the volumes accurately since a slight excess of water produces cloudiness.

Application 1 µL; thoroughly dry the points of application.

Development A Over a path of 15 cm.

Drying A In a current of warm air.

Development B Immediately, over a path of 15 cm, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with *4-aminobenzoic acid solution R* and dry in a current of cold air until the acetone is removed; heat at 100 °C for 15 min and allow to cool; spray with a 2 g/L solution of *sodium periodate R* and dry in a current of cold air; heat at 100 °C for 15 min.

System suitability Reference solution (c):
— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a) for lactose or to the principal spot in the chromatogram obtained with reference solution (b) for mannitol.

D. Shake a quantity of the substance to be examined corresponding to 25 mg of isosorbide mononitrate with 10 mL of *acetone R* for 5 min. Filter, evaporate to dryness at a temperature below 40 °C and dry the residue over *diphosphorus pentoxide R* at a pressure of 0.7 kPa for 16 h. The melting point (*2.2.14*) of the residue is 89 °C to 91 °C.

TESTS

Impurity A

Thin-layer chromatography (*2.2.27*).

Test solution Shake a quantity of the substance to be examined corresponding to 0.10 g of isosorbide mononitrate with 5 mL of *ethanol (96 per cent) R* and filter.

Reference solution Dissolve 10 mg of *potassium nitrate R* in 1 mL of *water R* and dilute to 100 mL with *ethanol (96 per cent) R*.

Plate *TLC silica gel plate R.*

Mobile phase *glacial acetic acid R, acetone R, toluene R* (15:30:60 *V/V/V*).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of air until the acetic acid is completely removed.

Detection Spray copiously with freshly prepared *potassium iodide and starch solution R*; expose to ultraviolet light at 254 nm for 15 min and examine in daylight.

Limit:
— *impurity A*: any spot due to impurity A is not more intense than the principal spot in the chromatogram obtained with the reference solution (0.5 per cent, calculated as potassium nitrate).

Impurities B and C

Liquid chromatography (*2.2.29*).

Test solution (a) Sonicate a quantity of the substance to be examined corresponding to 25.0 mg of isosorbide mononitrate with 20 mL of the mobile phase for 15 min and dilute to 25.0 mL with the mobile phase. Filter the solution through a suitable membrane filter.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of *isosorbide mononitrate CRS* in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10.0 mg of *isosorbide-2-nitrate CRS* (impurity C) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 0.1 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Sonicate a quantity of *isosorbide dinitrate CRS* (impurity B) corresponding to 10.0 mg of isosorbide dinitrate in 15 mL of the mobile phase for 15 min and dilute to 20.0 mL with the mobile phase. Filter the solution through a suitable membrane filter. Dilute 0.1 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 5 mg of *isosorbide mononitrate CRS* and 5 mg of *isosorbide-2-nitrate CRS* (impurity C) in the mobile phase and dilute to 10 mL with the mobile phase. Dilute 1 mL of this solution to 10 mL with the mobile phase.

Column:
— *size*: l = 0.25 m, Ø = 4.6 mm;
— *stationary phase*: *aminopropylmethylsilyl silica gel for chromatography R* (10 µm).

Mobile phase *anhydrous ethanol R, trimethylpentane R* (15:85 *V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 210-215 nm.

Injection 10 µL of test solution (a) and reference solutions (b), (c) and (d).

Retention time Impurity B = about 5 min; impurity C = about 8 min; isosorbide 5-nitrate = about 11 min.

System suitability Reference solution (d):
— *resolution*: minimum 4.0 between the peaks due to impurity C and isosorbide 5-nitrate.

Limits:
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent).

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for impurities B and C with the following modifications.

Detection Spectrophotometer at 230 nm.

Injection 20 µL of test solution (b) and reference solution (a).

If the areas of the peaks from 2 successive injections of reference solution (a) do not agree to within 1.0 per cent, then inject a further 4 times and calculate, for the 6 injections, the relative standard deviation.

System suitability Reference solution (a):
— *repeatability*: maximum relative standard deviation of 2.0 per cent after 6 injections.

Calculate the content of isosorbide mononitrate as a percentage of the declared content.

STORAGE

Protected from light.

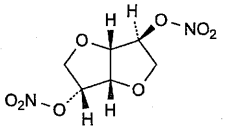
LABELLING

The label states the percentage content of isosorbide mononitrate.

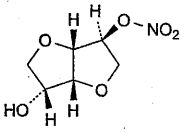
IMPURITIES

Specified impurities *A, B, C.*

A. inorganic nitrates,



B. 1,4:3,6-dianhydro-D-glucitol 2,5-dinitrate (isosorbide dinitrate),

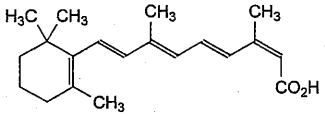


C. 1,4:3,6-dianhydro-D-glucitol 2-nitrate (isosorbide 2-nitrate).

Ph Eur

Isotretinoin

(Ph. Eur. monograph 1019)



$C_{20}H_{28}O_2$ 300.4 *4759-48-2*

Action and use
Vitamin A analogue (retinoid); treatment of acne.

Preparations
Isotretinoin Capsules
Isotretinoin Gel

Ph Eur

DEFINITION
(2*Z*,4*E*,6*E*,8*E*)-3,7-Dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid.

Content
98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Yellow or light orange, crystalline powder.

Solubility
Practically insoluble in water, soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

It is sensitive to air, heat and light, especially in solution.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).

Comparison *isotretinoin CRS.*

TESTS
Related substances
Liquid chromatography (*2.2.29*). *Carry out the test protected from light and prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of *tretinoin CRS* (impurity A) in *methanol R* and dilute to 5.0 mL with the same solvent.

Reference solution (b) Mix 0.5 mL of the test solution and 1 mL of reference solution (a) and dilute to 25 mL with *methanol R.*

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol R.* Dilute 1.0 mL of this solution to 10.0 mL with *methanol R.*

Column:
— *size*: *l* = 0.15 m, Ø = 4.6 mm;
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (3 µm).

Mobile phase *glacial acetic acid R, water for chromatography R, methanol R* (5:225:770 *V/V/V*).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 355 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Run time 1.6 times the retention time of isotretinoin.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to isotretinoin (retention time = about 26 min): impurity A = about 1.3.

System suitability Reference solution (b):
— *resolution*: minimum 5.0 between the peaks due to isotretinoin and impurity A.

Limits:
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* for 16 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 70 mL of *acetone R.* Titrate with *0.1 M tetrabutylammonium hydroxide in 2-propanol,* determining the end-point potentiometrically (*2.2.20*).

1 mL of *0.1 M tetrabutylammonium hydroxide in 2-propanol* is equivalent to 30.04 mg of $C_{20}H_{28}O_2$.

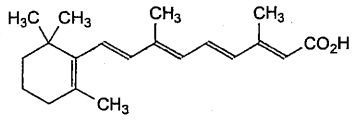
STORAGE
Under an inert gas, in an airtight container, protected from light.

It is recommended that the contents of an opened container be used as soon as possible and any unused part be protected by an atmosphere of inert gas.

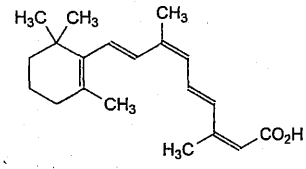
IMPURITIES
Specified impurities *A.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is

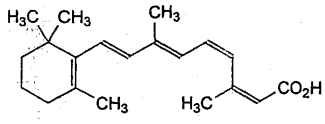
therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *B, C, D, F, G, H, I.*



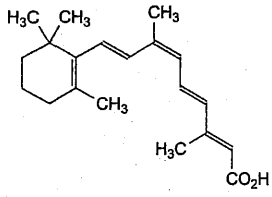
A. (2*E*,4*E*,6*E*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (tretinoin),



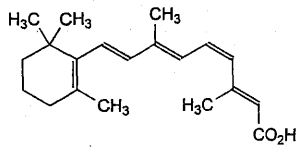
B. (2*Z*,4*E*,6*Z*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (9,13-di*cis*-retinoic acid),



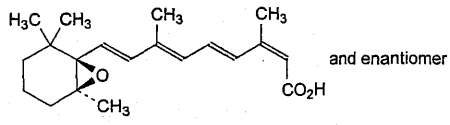
C. (2*Z*,4*Z*,6*E*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (11,13-di*cis*-retinoic acid),



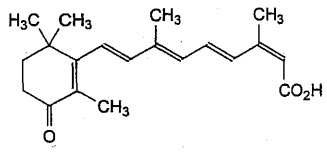
D. (2*E*,4*E*,6*Z*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (9-*cis*-retinoic acid),



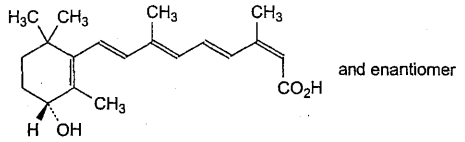
F. (2*E*,4*Z*,6*E*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (11-*cis*-retinoic acid),



G. (2*Z*,4*E*,6*E*,8*E*)-3,7-dimethyl-9-[(1*RS*,6*SR*)-2,2,6-trimethyl-7-oxabicyclo[4.1.0]heptan-1-yl]nona-2,4,6,8-tetraenoic acid (13-*cis*-5,6-dihydro-5,6-epoxyretinoic acid),



H. (2*Z*,4*E*,6*E*,8*E*)-3,7-dimethyl-9-(2,6,6-trimethyl-3-oxocyclohex-1-en-1-yl)nona-2,4,6,8-tetraenoic acid (13-*cis*-4-oxoretinoic acid),

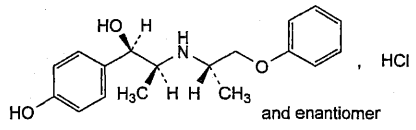


I. (2*Z*,4*E*,6*E*,8*E*)-9-[(3*RS*)-3-hydroxy-2,6,6-trimethylcyclohex-1-en-1-yl]-3,7-dimethylnona-2,4,6,8-tetraenoic acid (13-*cis*-4-hydroxyretinoic acid).

Ph Eur

Isoxsuprine Hydrochloride

(Ph. Eur. monograph 1119)



$C_{18}H_{24}ClNO_3$ 337.8 *579-56-6*

Action and use
$Beta_2$-adrenoceptor agonist.

Ph Eur

DEFINITION
(1*RS*,2*SR*)-1-(4-Hydroxyphenyl)-2-[[(1*SR*)-1-methyl-2-phenoxyethyl]amino]propan-1-ol hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility
Sparingly soluble in water and in ethanol (96 per cent), practically insoluble in methylene chloride.

mp
About 205 °C, with decomposition.

IDENTIFICATION
First identification: B, E.
Second identification: A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (*2.2.25*).

Test solution Dissolve 50.0 mg in *0.1 M hydrochloric acid* and dilute to 50.0 mL with the same acid. Dilute 10.0 mL of this solution to 100.0 mL with *0.1 M hydrochloric acid.*

Spectral range 230-350 nm.

Absorption maxima At 269 nm and 275 nm.

Resolution (*2.2.25*): minimum 1.7 for the absorbance ratio.

Specific absorbance at the absorption maxima:
— at 269 nm: 71 to 74;
— at 275 nm: 70 to 73.

B. Infrared absorption spectrophotometry (*2.2.24*).

Preparation Discs.

Comparison *isoxsuprine hydrochloride CRS.*

If the spectra obtained show differences, dissolve 50 mg of the substance to be examined and of the reference substance separately in 2 mL of *methanol R*, add 15 mL of *methylene*

chloride R, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (*2.2.27*).

Test solution Dissolve 20 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of *isoxsuprine hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.

Plate *TLC silica gel G plate R.*

Mobile phase *concentrated ammonia R, methanol R, methylene chloride R* (0.25:15:85 *V/V/V*).

Application 10 µL.

Development Over a path of 12 cm.

Drying In a current of warm air.

Detection Spray with a 10 g/L solution of *potassium permanganate R.*

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 1 mL of solution S (see Tests) add 0.05 mL of *copper sulfate solution R* and 0.5 mL of *strong sodium hydroxide solution R*. The solution becomes blue. Add 1 mL of *ether R* and shake. Allow to separate. The upper layer remains colourless.

E. 2 mL of solution S gives reaction (a) of chlorides (*2.3.1*).

TESTS

Solution S

Dissolve 0.50 g, with gentle heating if necessary, in *carbon dioxide-free water R*, cool and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and colourless (*2.2.2, Method II*).

pH (*2.2.3*)

4.5 to 6.0 for solution S.

Optical rotation (*2.2.7*)

−0.05° to + 0.05°, determined on solution S.

Phenones

Maximum 1.0 per cent, calculated as impurity B.

Dissolve 10.0 mg in *water R* and dilute to 100.0 mL with the same solvent. The absorbance (*2.2.25*) of the solution measured at the absorption maximum at 310 nm is not greater than 0.10.

Related substances

Gas chromatography (*2.2.28*). *Prepare the solutions immediately before use.*

Internal standard solution (a) Dissolve 0.1 g of *hexacosane R* in *trimethylpentane R* and dilute to 20 mL with the same solvent.

Internal standard solution (b) Dilute 1 mL of internal standard solution (a) to 50 mL with *trimethylpentane R.*

Test solution To 10.0 mg of the substance to be examined, add 0.5 mL of *N-trimethylsilylimidazole R.* Heat to 65 °C for 10 min. Allow to cool, then add 2.0 mL of the internal standard solution (b) and 2.0 mL of *water R.* Shake. Use the upper layer.

Reference solution (a) To 10.0 mg of the substance to be examined, add 0.5 mL of *N-trimethylsilylimidazole R.* Heat to 65 °C for 10 min. Allow to cool, then add 2.0 mL of the internal standard solution (a) and 2.0 mL of *water R.* Shake. Dilute 1.0 mL of the upper layer to 50.0 mL with *trimethylpentane R.*

Reference solution (b) To 10.0 mg of the substance to be examined, add 0.5 mL of *N-trimethylsilylimidazole R.* Heat to 65 °C for 10 min. Allow to cool, then add 2.0 mL of *trimethylpentane R* and 2.0 mL of *water R.* Shake. Use the upper layer.

Column:
- *material*: glass;
- *size*: *l* = 1.5 m, Ø = 4 mm;
- *stationary phase*: *silanised diatomaceous earth for gas chromatography R* (125-135 µm) impregnated with 3 per cent *m/m* of *poly(dimethyl)siloxane R.*

Carrier gas *nitrogen for chromatography R.*

Flow rate 30 mL/min.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 25	195
	25 - 29	195 → 215
	29 - 39	215
Injection port		225
Detector		225

Detection Flame ionisation.

Injection 1 µL.

Elution order Isoxsuprine, hexacosane.

System suitability:
- *resolution*: minimum 5.0 between the peaks due to isoxsurpine and hexacosane in the chromatogram obtained with reference solution (a);
- in the chromatogram obtained with reference solution (b), there is no peak with the same retention time as the internal standard.

Limit:
- *total*: calculate the ratio (*R*) of the area of the peak due to the trimethylsilyl derivative of isoxsuprine to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (a); from the chromatogram obtained with the test solution, calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard: this ratio is not greater than *R* (2.0 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 80 mL of *ethanol (96 per cent) R* and add 1.0 mL of *0.1 M hydrochloric acid.* Carry out a potentiometric titration (*2.2.20*), using *0.1 M sodium hydroxide.* Read the volume added between the 2 points of inflexion.

1 mL of *0.1 M sodium hydroxide* is equivalent to 33.78 mg of $C_{18}H_{24}ClNO_3$.

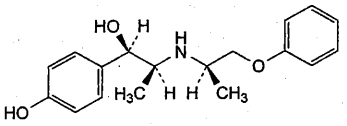
STORAGE

Protected from light.

IMPURITIES

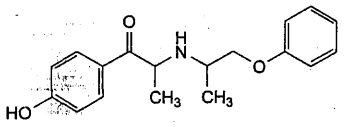
Specified impurities *B.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) A.



and enantiomer

A. (1*RS*,2*SR*)-1-(4-hydroxyphenyl)-2-[[(1*RS*)-1-methyl-2-phenoxyethyl]amino]propan-1-ol,

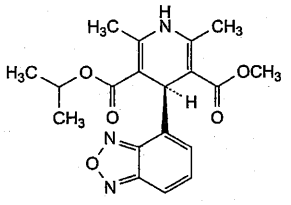


B. 1-(4-hydroxyphenyl)-2-[(1-methyl-2-phenoxyethyl)amino]propan-1-one.

Ph Eur

Isradipine

(Ph. Eur. monograph 2110)



and enantiomer

$C_{19}H_{21}N_3O_5$ 371.4 *75695-93-1*

Action and use
Calcium channel blocker.

Preparation
Isradipine Tablets

Ph Eur

DEFINITION
Methyl 1-methylethyl (4*RS*)-4-(2,1,3-benzoxadiazol-4-yl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate.

Content
97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS
Appearance
Yellow, crystalline powder.

Solubility
Practically insoluble in water, freely soluble in acetone, soluble in methanol.

mp
About 168 °C.

IDENTIFICATION
Infrared absorption spectrophotometry (*2.2.24*).
Comparison *isradipine CRS*.

TESTS
Related substances
Liquid chromatography (*2.2.29*).
Test solution (a) Dissolve 50.0 mg of the substance to be examined in 1 mL of *methanol R*, using an ultrasonic bath if necessary, and dilute to 25.0 mL with the mobile phase.
Test solution (b) Dissolve 50.0 mg of the substance to be examined in 2 mL of *methanol R* and dilute to 250.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 2 mg of the substance to be examined and 2 mg of *isradipine impurity D CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 50.0 mg of *isradipine CRS* in 2 mL of *methanol R* and dilute to 250.0 mL with the mobile phase.
Column:
— *size*: *l* = 0.10 m, Ø = 4.6 mm,
— *stationary phase*: *octadecylsilyl silica gel for chromatography R* (5 µm).
Mobile phase *acetonitrile R, tetrahydrofuran R, water R* (125:270:625 *V/V/V*).
Flow rate 1.2 mL/min.
Detection Spectrophotometer at 230 nm.
Injection 20 µL of test solution (a) and reference solutions (a) and (b).
Run time 5 times the retention time of isradipine.
Identification of impurities Use the chromatogram supplied with *isradipine CRS* to identify the peaks due to impurities A and B.
Relative retention With reference to isradipine (retention time = about 7 min): impurity A = about 0.8; impurity D = about 0.9; impurity B = about 1.8.
System suitability Reference solution (b):
— *resolution*: minimum 2.0 between the peaks due to isradipine and impurity D.
Limits:
— *correction factor*: for the calculation of content, multiply the peak area of impurity D by 1.4,
— *impurity A*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),
— *impurity B*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent),
— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— *total*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent),
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)
Maximum 0.2 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 326 nm.

Injection Test solution (b) and reference solution (c).

Run time Twice the retention time of isradipine.

Calculate the percentage content of isradipine from the areas of the peaks and the declared content of *isradipine CRS*.

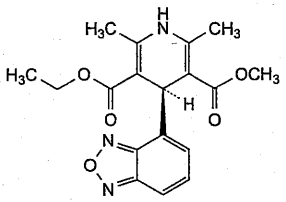
STORAGE

Protected from light.

IMPURITIES

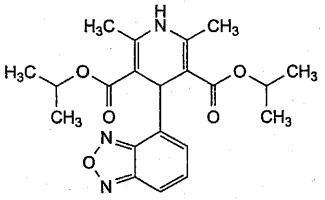
Specified impurities *A, B, D.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *C, E.*

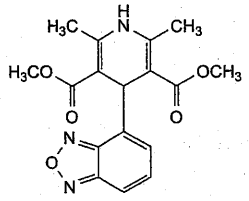


and enantiomer

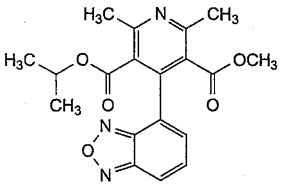
A. ethyl methyl (4*RS*)-4-(2,1,3-benzoxadiazol-4-yl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate,



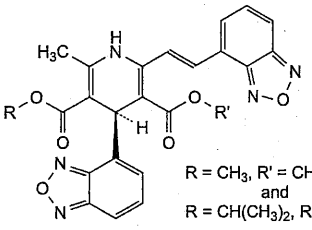
B. bis(1-methylethyl) (4*RS*)-4-(2,1,3-benzoxadiazol-4-yl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate,



C. dimethyl (4*RS*)-4-(2,1,3-benzoxadiazol-4-yl)-2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate,



D. methyl 1-methylethyl 4-(2,1,3-benzoxadiazol-4-yl)-2,6-dimethylpyridine-3,5-dicarboxylate,



its (*Z*) isomer and their enantiomers

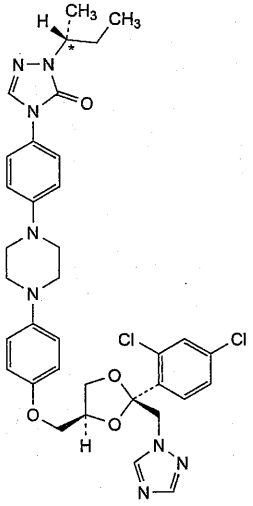
R = CH_3, R' = $CH(CH_3)_2$
and
R = $CH(CH_3)_2$, R' = CH_3

E. methyl 1-methylethyl (4*RS*)-4-(2,1,3-benzoxadiazol-4-yl)-2-[(*EZ*)-2-(2,1,3-benzoxadiazol-4-yl)ethenyl]-6-methyl-1,4-dihydropyridine-3,5-dicarboxylate.

Ph Eur

Itraconazole

(Ph. Eur. monograph 1335)



its epimer at C*

and their enantiomers

$C_{35}H_{38}Cl_2N_8O_4$ 706 *84625-61-6*

Action and use

Antifungal.

Preparations

Itraconazole Capsules

Itraconazole Oral Solution

Itraconazole Sterile Concentrate

Ph Eur

DEFINITION

4-[4-[4-[[*cis*-2-(2,4-Dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-[(1*RS*)-1-methylpropyl]-2,4-dihydro-3*H*-1,2,4-triazol-3-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (*2.2.24*).

Comparison *itraconazole CRS*.

TESTS

Solution S

Dissolve 2.0 g in *methylene chloride R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (*2.2.1*) and not more intensely coloured than reference solution R_6 or B_6 (*2.2.2, Method II*).

Related substances

Liquid chromatography (*2.2.29*). *Prepare the solutions immediately before use.*

Test solution Dissolve 0.100 g of the substance to be examined in *methanolic hydrochloric acid R* and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with *methanolic hydrochloric acid R*. Dilute 1.0 mL of this solution to 10.0 mL with *methanolic hydrochloric acid R*.

Reference solution (b) Dissolve 10 mg of *itraconazole for system suitability CRS* (containing impurities B, C, D, E, F and G) in 1.0 mL of *methanolic hydrochloric acid R*.

Column:
- *size*: *l* = 0.10 m, Ø = 4.6 mm;
- *stationary phase*: *base-deactivated end-capped octadecylsilyl silica gel for chromatography R* (3 µm or 3.5 µm);
- *temperature*: 30 °C.

Mobile phase:
- *mobile phase A*: 27.2 g/L solution of *tetrabutylammonium hydrogen sulfate R1*;
- *mobile phase B*: *acetonitrile R1*;

Time (min)	Mobile phase A (per cent *V/V*)	Mobile phase B (per cent *V/V*)
0 - 2	80	20
2 - 22	80 → 50	20 → 50
22 - 27	50	50

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with *itraconazole for system suitability CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, D, E, F and G.

Relative retention With reference to itraconazole (retention time = about 14 min): impurity B = about 0.7; impurities C and D = about 0.8; impurity E = about 0.9; impurity F = about 1.05; impurity G = about 1.3.

System suitability Reference solution (b):
- *peak-to-valley ratio*: minimum 1.5, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to itraconazole.

Limits:
- *impurities B, G*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *impurity E*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- *sum of impurities C and D*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- *total*: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (*2.2.32*)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 70 mL of a mixture of 1 volume of *anhydrous acetic acid R* and 7 volumes of *methyl ethyl ketone R* by vigorous stirring for at least 10 min. Titrate with *0.1 M perchloric acid*, determining the end-point potentiometrically at the second point of inflexion (*2.2.20*).

1 mL of *0.1 M perchloric acid* is equivalent to 35.3 mg of $C_{35}H_{38}Cl_2N_8O_4$.

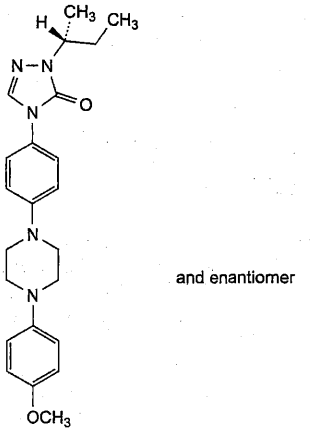
STORAGE

Protected from light.

IMPURITIES

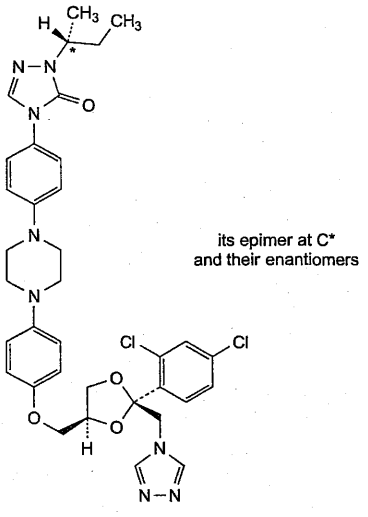
Specified impurities *B, C, D, E, G.*

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use) *A, F.*

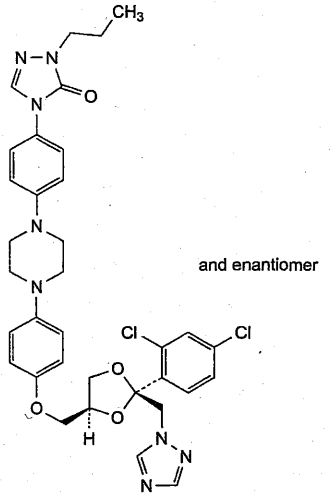


and enantiomer

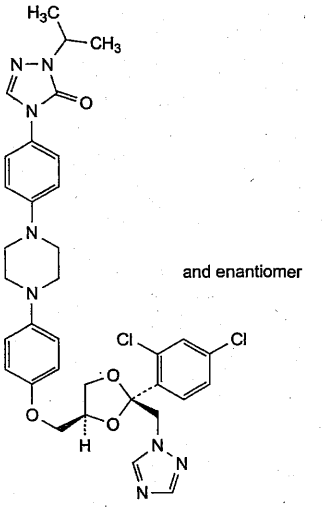
A. 4-[4-[4-(4-methoxyphenyl)piperazin-1-yl]phenyl]-2-[(1*RS*)-1-methylpropyl]-2,4-dihydro-3*H*-1,2,4-triazol-3-one,



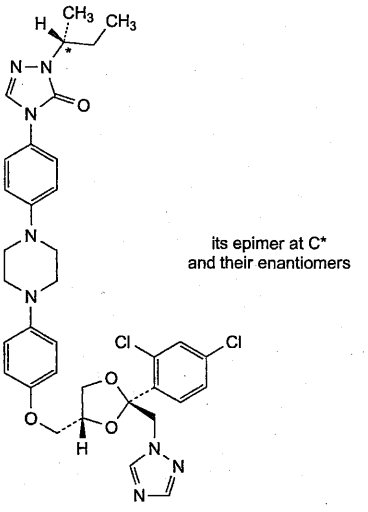
B. 4-[4-[4-[4-[[*cis*-2-(2,4-dichlorophenyl)-2-(4*H*-1,2,4-triazol-4-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-[(1*RS*)-1-methylpropyl]-2,4-dihydro-3*H*-1,2,4-triazol-3-one,



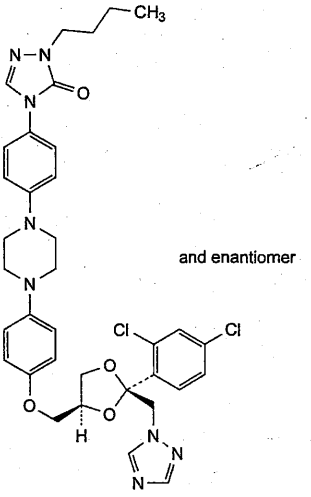
C. 4-[4-[4-[4-[[*cis*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-propyl-2,4-dihydro-3*H*-1,2,4-triazol-3-one,



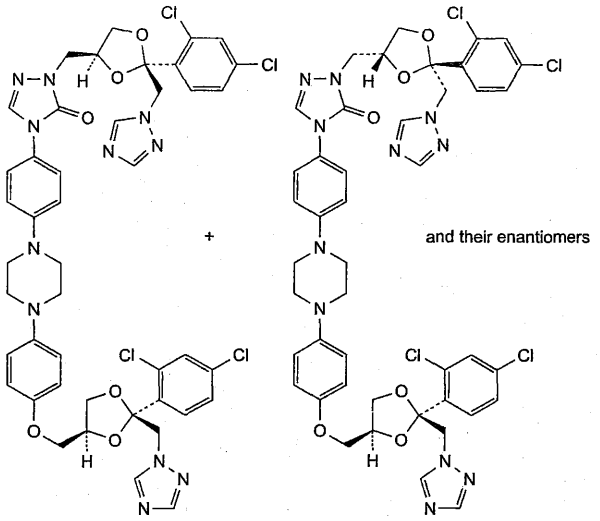
D. 4-[4-[4-[4-[[*cis*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-(1-methylethyl)-2,4-dihydro-3*H*-1,2,4-triazol-3-one,



E. 4-[4-[4-[4-[[*trans*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-[(1*RS*)-1-methylpropyl]-2,4-dihydro-3*H*-1,2,4-triazol- 3-one,



F. 2-butyl-4-[4-[4-[4-[[*cis*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2,4-dihydro-3*H*-1,2,4-triazol-3-one,

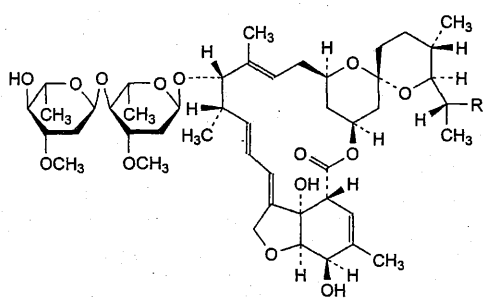


G. 4-[4-[4-[4-[[*cis*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methoxy]phenyl]piperazin-1-yl]phenyl]-2-[[*cis*-2-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-ylmethyl)-1,3-dioxolan-4-yl]methyl]-2,4-dihydro-3*H*-1,2,4-triazol-3-one.

Ph Eur

Ivermectin

(Ph. Eur. monograph 1336)



Component	R	Molecular formula	M_r
H_2B_{1a}	CH_2-CH_3	$C_{48}H_{74}O_{14}$	875
H_2B_{1b}	CH_3	$C_{47}H_{72}O_{14}$	861

Ivermectin B1a *71827-03-7*

Ivermectin B1b *70209-81-3*

Action and use
Antihelminthic.

Ph Eur

DEFINITION

Mixture of (2a*E*,4*E*,5′*S*,6*S*,6′*R*,7*S*,8*E*,11*R*,13*R*,15*S*,17a*R*,20*R*,20a*R*,20b*S*)-7-[[2,6-dideoxy-4-*O*-(2,6-dideoxy-3-*O*-methyl-α-L-*arabino*-hexopyranosyl)-3-*O*-methyl-α-L-*arabino*-hexopyranosyl]oxy]-20,20b-dihydroxy-5′,6,8,19-tetramethyl-6′-[(1*S*)-1-methylpropyl]-3′,4′,5′,6,6′,7,10,11,14,15,17a,20,20a,20b-tetradecahydrospiro[11,15-methano-2*H*,13*H*,17*H*-furo[4,3,2-*pq*][2,6]benzodioxacyclooctadecene-13,2′-[2*H*]pyran]-17-one (or 5-*O*-demethyl-22,23-dihydroavermectin A_{1a}) (component H_2B_{1a}) and (2a*E*,4*E*,5′*S*,6*S*,6′*R*,7*S*,8*E*,11*R*,13*R*,15*S*,17a*R*,20*R*,20a*R*,20b*S*)-7-[[2,6-dideoxy-4-*O*-(2,6-dideoxy-3-*O*-methyl-α-L-*arabino*-hexopyranosyl)-3-*O*-methyl-α-L-*arabino*-hexopyranosyl]oxy]-20,20b-dihydroxy-5′,6,8,19-tetramethyl-6′-(1-methylethyl)-3′,4′,5′,6,6′,7,10,11,14,15,17a,20,20a,20b-tetradecahydrospiro[11,15-methano-2*H*,13*H*,17*H*-furo[4,3,2-*pq*][2,6]benzodioxacyclooctadecene-13,2′-[2*H*]pyran]-17-one (or 5-*O*-demethyl-25-de(1-methylpropyl)-25-(1-methylethyl)-22,23-dihydroavermectin A_{1a}) (component H_2B_{1b}).

Semi-synthetic product derived from a fermentation product.

Content
— *ivermectin* (H_2B_{1a} + H_2B_{1b}): 95.0 per cent to 102.0 per cent (anhydrous substance);
— *ratio* H_2B_{1a}/(H_2B_{1a} + H_2B_{1b}) (areas by liquid chromatography): minimum 90.0 per cent.

CHARACTERS

Appearance
White or yellowish-white, crystalline powder, slightly hygroscopic.

Solubility
Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (*2.2.24*).

Comparison *ivermectin CRS.*

B. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time to the 2 principal peaks in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution
The solution is clear (*2.2.1*) and not more intensely coloured than reference solution BY_7 (*2.2.2, Method II*).

Dissolve 1.0 g in 50 mL of *toluene R.*

Specific optical rotation (*2.2.7*)
−20 to −17 (anhydrous substance).

Dissolve 0.250 g in *methanol R* and dilute to 10.0 mL with the same solvent.

Related substances
Liquid chromatography (*2.2.29*).

Test solution Dissolve 40.0 mg of the substance to be examined in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 40.0 mg of *ivermectin CRS* in *methanol R* and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *methanol R.*

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 100.0 mL with *methanol R.*

Reference solution (d) Dilute 5.0 mL of reference solution (a) to 100.0 mL with *methanol R.*

Column:
— *size: l* = 0.25 m, Ø = 4.6 mm;
— *stationary phase: octadecylsilyl silica gel for chromatography R* (5 µm).

Mobile phase *water R, methanol R, acetonitrile R* (15:34:51 *V/V/V*).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

System suitability:
— *resolution*: minimum 3.0 between the 1st peak (component H_2B_{1b}) and the 2nd peak (component H_2B_{1a}) in the chromatogram obtained with reference solution (a);
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (c);
— *symmetry factor*: maximum 2.5 for the principal peak in the chromatogram obtained with reference solution (a).

Limits:
— *impurity with a relative retention of 1.3 to 1.5* with reference to the principal peak: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (2.5 per cent);
— *any other impurity* (apart from the 2 principal peaks): not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);
— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5 per cent);
— *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

Ethanol and formamide

Gas chromatography (*2.2.28*).

Internal standard solution Dilute 0.5 mL of *propanol R* to 100 mL with *water R*.

Test solution In a centrifuge tube, dissolve 0.120 g of the substance to be examined in 2.0 mL of *m-xylene R* (if necessary heat in a water-bath at 40-50 °C). Add 2.0 mL of *water R*, mix thoroughly and centrifuge. Remove the upper layer and extract it with 2.0 mL of *water R*. Discard the upper layer and combine the aqueous layers. Add 1.0 mL of the internal standard solution. Centrifuge and discard any remaining *m*-xylene.

Reference solution (a) Dilute 3.0 g of *anhydrous ethanol R* to 100.0 mL with *water R*.

Reference solution (b) Dilute 1.0 g of *formamide R* to 100.0 mL with *water R*.

Reference solution (c) Dilute 5.0 mL of reference solution (a) and 5.0 mL of reference solution (b) to 50.0 mL with *water R*. Introduce 2.0 mL of this solution into a centrifuge tube, add 2.0 mL of *m-xylene R*, mix thoroughly and centrifuge. Remove the upper layer and extract it with 2.0 mL of *water R*. Discard the upper layer and combine the aqueous layers. Add 1.0 mL of the internal standard solution. Centrifuge and discard any remaining *m*-xylene.

Reference solution (d) Dilute 10.0 mL of reference solution (a) and 10.0 mL of reference solution (b) to 50.0 mL with *water R*. Treat as prescribed for reference solution (c) (from "Introduce 2.0 mL of this solution...").

Column:
— *material*: fused silica;
— *size*: *l* = 30 m, Ø = 0.53 mm;
— *stationary phase*: *macrogol 20 000 R* (film thickness 1 µm).

Carrier gas *helium for chromatography R*.

Flow rate 7.5 mL/min.

Split ratio 1:10.

Temperature:

	Time (min)	Temperature (°C)
Column	0 - 2	50 → 80
	2 - 8	80 → 240
Injection port		220
Detector		280

Detection Flame ionisation.

Injection 1 µL of the test solution and reference solutions (c) and (d).

Limits:
— *ethanol*: maximum 5.0 per cent;
— *formamide*: maximum 3.0 per cent.

Water (*2.5.12*)

Maximum 1.0 per cent, determined on 0.50 g.

Sulfated ash (*2.4.14*)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (*2.2.29*) as described in the test for related substances.

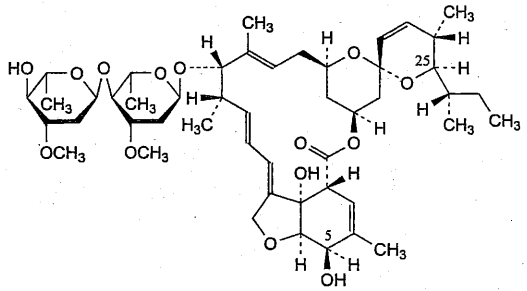
Injection 20 µL of the test solution and reference solutions (a) and (d).

Calculate the percentage content of ivermectin (H_2B_{1a} + H_2B_{1b}) and the ratio $H_2B_{1a}/(H_2B_{1a} + H_2B_{1b})$ taking into account the assigned content of component H_2B_{1a} in *ivermectin CRS*. Determine the content of ivermectin component H_2B_{1a} by comparing with the peak area due to component H_2B_{1a} in the chromatogram obtained with reference solution (a). Determine the content of ivermectin component H_2B_{1b} by comparing with the peak area due to component H_2B_{1a} in the chromatogram obtained with reference solution (d).

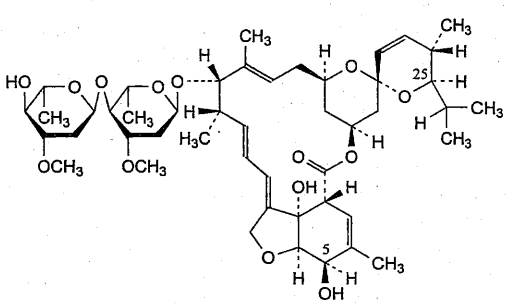
STORAGE

In an airtight container.

IMPURITIES



A. 5-*O*-demethylavermectin A_{1a} (avermectin B_{1a}),



B. 5-*O*-demethyl-25-de(1-methylpropyl)-25-(1-methylethyl) avermectin A_{1a} (avermectin B_{1b}),